Supporting Information

For

Unusual C₃-Acetylation of Quinoxalin-2(1*H*)-one via Oxidative C–C and C–O Bond Cleavages of PEG-400.

Vishal Suresh Kudale, ^a Mohana Reddy Mutra, ^a Ching-Piao Chu ^a and Jeh-Jeng Wang*^{ab}

^{*a*} Department of Medicinal and Applied Chemistry, Kaohsiung Medical University, No. 100, Shiquan 1st Rd, Sanmin District, Kaohsiung City, 807, Taiwan.

^b Department of Medical Research, Kaohsiung Medical University Hospital, No. 100 Tzyou 1st Rd, Sanmin District, Kaohsiung City 807, Taiwan.

E-mail: jjwang@kmu.edu.tw

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(1) General Information

¹H, ¹³C, and DEPT NMR spectra were recorded on a 400 MHz Varian Unity Plus or Varian Mercury plus spectrometer. The chemical shift (δ) values are reported in parts per million (ppm), and the coupling constants (J) are given in Hz. The spectra were recorded using CDCl₃ as a solvent. ¹ H NMR chemical shifts are referenced to tetramethylsilane (TMS) (0 ppm). ¹³C NMR was referenced to CDCl₃ (77.0 ppm) or DMSO-d₆ (39.51 ppm). The abbreviations used are as follows: s, singlet; d, doublet; t, triplet; q, quartet; dd, doublet of doublet; ddd, doublet of doublet; dt, doublet of triplets; td, triplet of doublet; m, multiplet. Mass spectra and high-resolution mass spectra (HRMS) were measured using the ESI (FT-MS solariX) at National Sun Yat-Sen University, Kaohsiung, Taiwan. Melting points were determined on an EZ-Melt (Automated melting point apparatus). All products reported showed ¹H NMR spectra in agreement with the assigned structures. Reaction progress and product mixtures were routinely monitored by TLC using Merck TLC aluminum sheets (silica gel 60 F254). Column chromatography was carried out with 230–400 mesh silica gel 60 (Merck) and a mixture of hexane/ethyl acetate or hexane as eluent. Preparative TLC was run on Merck TLC aluminum sheets (silica gel 60 F254).



(2) Mechanistic studies:

Fig S1: GC-MS with different retention times generated from PEG-400 in the presence of oxidant & radical scavenger.





44.82

50-

40를

30

20-

10

0-

40

58.85

60

60.86 85.86

45.82

88.82

102.81

100

132.81

146.82

140

176.80

180

194.79 206.78 220.83

200

220

m/z

162.80

160

130.80

120.86

120

86.84

80





238.83 250.81

240

272.19

280

260

296.91

300

310.84 318.91

320

342.79 350.21 370.94

360

340

388.21

400

380

(3) Experimental Procedures

(i) General Experimental Procedure and Spectral Characterization for the Synthesis of 1quinoxalin-2(1H)-one acetylation with PEG-400 as "CH₃CO" Source



To an oven-dried sealed tube was charged with 1-methylquinoxalin-2(1H)-one **1a-1w**⁻¹ (0.25 mmol), PEG-400 (0.25 M), and AgNO₃ (0.25 mmol) and allowed to stir at 100° C until the completion of the reaction (7 ~ 24 h) by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 5.0 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate, and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford pure heteroaryl acetylation **2a-2u** in 48%-76% yields.

(4) Spectral Characterization

3-acetyl-1-methylquinoxalin-2(1*H***)-one (2a)²:** Following the general procedure, a 15 mL reaction tube was charged with 1-methylquinoxalin-2(1*H*)-one (1b) (35 mg, 0.2 mmol), PEG-400 (0.20 M), AgNO₃ (25 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with

 $(\overline{3X10 \text{ mL}})$ of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-methylquinoxalin-2(1*H*)-one (2a) as a yellow solid (30 mg, yield = 76 %); Mp. 116.2-116.9 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.97-7.94 (m, 1H), 7.69 (ddd, *J* = 8.4, 7.2, 1.6 Hz, 1H), 7.41 (ddd, *J* = 8.4, 7.2, 1.2 Hz, 1H), 7.36 (dd, *J* = 8.4, 0.8 Hz, 1H), 3.73 (s, 3H), 2.72 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.16, 152.83, 151.77, 134.42, 132.77, 131.88, 131.52, 124.15, 113.84, 29.02, 28.51.

3-acetyl-1-ethylquinoxalin-2(1H)-one (2b): Following the general procedure, a 15 mL reaction tube was



charged with 1-ethylquinoxalin-2(1*H*)-one (1b) (37 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with

(3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-ethylquinoxalin-2(1*H*)-one (2b) as a yellow solid (31 mg, yield = 73 %); Mp. 122-122.5 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.70-7.65 (m, 1H), 7.41-7.37 (m, 2H), 4.35 (q, *J* = 7.2 Hz, 1H), 2.72 (s, 3H), 1.40 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.22, 152.34, 151.69, 133.42, 132.71, 132.17, 131.75, 123.93, 113.67, 37.39, 28.52, 12.34. HRMS (ESI) calcd for Cl₁₂ H₁₂ N₂ O₂ Na [M + Na] +: 239.0796; found: 239.0799.

3-acetyl-1-allylquinoxalin-2(1H)-one (2c): Following the general procedure, a 15 mL reaction tube was



charged with 1-allylquinoxalin-2(1*H*)-one (1c) (40 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with

(3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-allylquinoxalin-2(1*H*)-one (2c) as a yellow solid (29 mg, yield = 65 %); Mp. 136.1-137 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96 (ddd, *J* = 8.0, 1.2, 0.4 Hz, 1H), δ 7.65 (ddd, *J* = 8.4, 7.2, 1.2 Hz, 1H), 7.42-7.37 (m, 1H), 7.34 (dd, *J* = 8.4, 1.2, 1H), 5.97-5.90 (m, 1H), 5.32-5.28 (m, 1H), 5.23-5.18 (m, 1H), 4.93 (dt, *J* = 3.6, 2.0, 2H), 2.72 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.01, 152.36, 151.61, 133.69, 132.68, 132.00, 131.56, 130.13, 124.11, 118.57, 114.39, 44.44, 28.51. HRMS (ESI) calcd for C₁₃ H₁₂ N₂ O₂ Na [M + Na]⁺: 251.0796; found: 251.0799.

3-acetyl-1-(but-3-en-1-yl)quinoxalin-2(1H)-one (2d): Following the general procedure, a 15 mL reaction



tube was charged with 1-(but-3-en-1-yl)quinoxalin-2(1*H*)-one (1d) (43 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was

extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10

mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(but-3-en-1-yl)quinoxalin-2(1*H*)-one (2d) as a yellow solid (30 mg, yield = 63 %); Mp. 140.5-141.2°C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96 (dd, *J* = 8.0, 1.2 Hz, 1H), δ 7.68 (ddd, *J* = 8.4, 7.8, 1.6 Hz, 1H), 7.42-7.35 (m, 2H), 5.93-5.83 (m, 1H), 5.15-5.09 (m, 2H), 4.39-4.33 (m, 2H), 2.72 (s, 3H) 2.56-2.50 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.11, 152.53, 151.68, 133.56, 132.70, 132.14, 131.81, 124.01, 117.92, 113.78, 41.60, 31.42, 28.51.HRMS (ESI) calcd for C₁₂ H₁₂ N₂ O₂ Na [M + Na] ⁺: 265.0796; found: 265.0799.

3-acetyl-1-(2-methylallyl)quinoxalin-2(1H)-one (2e): Following the general procedure, a 15 mL reaction



tube was charged with 1-(2-methylallyl)quinoxalin-2(1*H*)-one (1e) (43 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was

extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(2-methylallyl)quinoxalin-2(1*H*)-one (2e) as a yellow solid (28 mg, yield = 58 %); Mp. 132.5-132.9 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96 (ddd, *J* = 8.0, 1.2, 0.4 Hz, 1H), δ 7.62 (ddd, *J* = 6.8, 3.6, 1.6 Hz, 1H), 7.41-7.37 (m, 1H), 7.28-7.26 (m, 1H), 4.84 (s, 2H), 4.62 (dt, *J* = 1.6, 0.8 Hz 1H), 2.73 (s, 3H), 1.83 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.03, 152.56, 151.82, 137.82, 133.93, 132.61, 132.01, 131.50, 124.15, 114.68, 112.20, 77.31, 76.99, 76.68, 47.55, 28.58, 20.17.HRMS (ESI) calcd for C₁₂ H₁₂ N₂ O₂ Na [M + Na] ⁺: 266.1693; found: 266.1696.

3-acetyl-1-(prop-2-yn-1-yl)quinoxalin-2(1H)-one (2f): Following the general procedure, a 15 mL reaction



tube was charged with 1-(prop-2-yn-1-yl)quinoxalin-2(1*H*)-one (1f) (40 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was

extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(prop-2-yn-1-yl)quinoxalin-2(1*H*)-one (2f) as a yellow solid (32 mg, yield = 71 %); Mp. 106.2-106.7 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.97 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.74-7.69 (m, 1H), 7.52 (d, *J* = 8.8, 1H), 7.46-7.42 (m, 1H), 5.08 (d, *J* = 2.8, 2H), 2.72 (s, 3H); ¹³C NMR

 $(CDCl_3, 100 \text{ MHz}) \delta 13C \text{ NMR} (101 \text{ MHz}, cdcl3) \delta 197.64, 151.75, 151.42, 132.95, 132.91, 132.05, 131.66, 124.52, 114.39, 76.27, 73.58, 31.33, 28.46. HRMS (ESI) calcd for C₁₃ H₁₀ N₂ O₂ Na [M + Na] +: 249.0640; found: 249.0642.$

ethyl 2-(3-acetyl-2-oxoquinoxalin-1(2H)-yl)acetate (2g)²: Following the general procedure, a 15 mL



reaction tube was charged with ethyl 2-(2-oxoquinoxalin-1(2*H*)-yl)acetate (1g) (49 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate

layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding ethyl 2-(3-acetyl-2-oxoquinoxalin-1(2*H*)-yl)acetate (2g) as a yellow solid (37 mg, yield = 67 %); Mp. 107.1-107.6 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.98 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.65 (ddd, *J* = 8.4, 7.2, 1.6 Hz, 1H), 7.42 (ddd, *J* = 8.47.6, 1.2 Hz, 1H), 7.12 (dd, *J* = 8.4, 0.8 Hz, 1H), 5.02 (s, 2H), 4.26 (q, *J* = 7.2 Hz, 2H), 2.72 (s, 3H), 1.28 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.57, 166.65, 152.30, 151.14, 133.68, 133.00, 131.90, 131.85, 124.42, 113.34, 62.24, 43.30, 28.41, 14.06.

3-acetyl-1-(2-oxo-2-phenylethyl)quinoxalin-2(1H)-one (2h): Following the general procedure, a 15 mL



reaction tube was charged with 1-(2-oxo-2-phenylethyl)quinoxalin-2(1*H*)-one (1h) (55 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate

layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(2-oxo-2-phenylethyl)quinoxalin-2(1*H*)-one (2h) as a yellow solid (29 mg, yield = 48 %); Mp. 138.2-139.1 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.07-8.05 (m, 1H), 8.02-8.00 (m, 2H), 7.68-7.57 (m, 4H), 7.53-7.50 (m, 2H), 5.86 (s, 2H), 2.85 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.39, 192.77, 153.89, 141.67, 140.96, 137.93, 134.46, 133.74, 131.99, 129.65, 128.81, 127.80, 127.56, 126.79, 68.02, 28.43. HRMS (ESI) calcd for C₁₈H₁₄ N2 O₃ Na [M + Na] ⁺: 329.0902; found: 329.0905.

3-acetyl-1-benzylquinoxalin-2(1H)-one (2i): Following the general procedure, a 15 mL reaction tube was



charged with 1-benzylquinoxalin-2(1*H*)-one (1i) (50 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash

(1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-benzylquinoxalin-2(1*H*)-one (2i) as a yellow solid (38 mg, yield = 69 %); Mp. 129.7-130.1 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96 (ddd, *J* = 8.0, 1.6, 0.4 Hz, 1H), 7.55 (ddd, *J* = 8.8, 7.2, 1.6 Hz, 1H), 7.38-7.35 (m, 1H), 7.34-7.31 (m, 2H), 7.29-7.25 (m, 4H), 5.51 (s, 2H) 2.75 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.01, 152.90, 151.70, 134.74, 133.82, 132.75, 132.12, 131.61, 128.96, 127.85, 126.96, 124.18, 114.63, 45.83, 28.52. HRMS (ESI) calcd for C₁₇ H₁₄ N₂ O₂ Na [M + Na] ⁺: 301.0953; found: 301.0956.

3-acetyl-1-(4-methylbenzyl)quinoxalin-2(1H)-one (2j): Following the general procedure, a 15 mL reaction



tube was charged with 1-(4-methylbenzyl)quinoxalin-2(1*H*)-one (1j) (53 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and $K_2S_2O_8$ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate

layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(4-methylbenzyl)quinoxalin-2(1*H*)-one (2j) as a yellow solid (38 mg, yield = 66 %); Mp. 132.4-133.6 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96-7.93 (m, 1H), 7.55 (ddd, *J* = 8.8, 7.6, 1.6 Hz, 1H), 7.37-7.32 (m, 2H), 7.18-7.11 (m, 4H), 5.47 (s, 2H), 2.75 (s, 3H), 2.30 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) 13C NMR (101 MHz, cdcl3) δ 198.07, 152.93, 151.76, 137.65, 133.86, 132.71, 132.14, 131.75, 131.59, 129.61, 127.01, 124.12, 114.66, 45.64, 28.55, 21.05. HRMS (ESI) calcd for C₁₈ H₁₆ N₂ O₂ Na [M + Na] ⁺: 315.1109; found: 315.1109.

3-acetyl-1-(3-methoxybenzyl)quinoxalin-2(1H)-one (2k): Following the general procedure, a 15 mL



reaction tube was charged with 1-(3-methoxybenzyl)quinoxalin-2(1*H*)-one (1k) (56 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water

layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(3-methoxybenzyl)quinoxalin-2(1*H*)-one (2k) as a yellow solid (34 mg, yield = 55 %); Mp. 142.1-143 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.96 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.57-7.53 (m, 1H), 7.38-7.34 (m, 1H), 7.31-7.29 (m, 1H),), 7.24-7.22 (m, 1H),), 6.84- 6.78 (m, 3H), 5.48 (s, 2H), 3.76 (s, 3H), 2.75 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.00, 160.05, 151.73, 136.36, 133.88, 132.78, 132.15, 131.64, 130.08, 124.21, 119.14, 114.68, 113.04, 112.79, 77.31, 76.99, 76.67, 55.26, 45.81, 28.56. HRMS (ESI) calcd for C₁₈ H₁₆ N₂ O₃ Na [M + Na] +: 331.1059; found: 331.1060.

3-acetyl-1-(3-chlorobenzyl)quinoxalin-2(1H)-one (2l): Following the general procedure, a 15 mL reaction



tube was charged with 1-(3-chlorobenzyl)quinoxalin-2(1*H*)-one (11) (57 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate

and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(3-chlorobenzyl)quinoxalin-2(1*H*)-one (2l) as a yellow solid (38 mg, yield = 61 %); Mp. 136.4-137 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.98-7.95 (m, 1H), 7.59-7.54 (m, 1H), 7.40-7.36 (m, 1H), 7.30-7.26 (m, 3H), 7.23-7.20 (m, 2H), 5.47 (s, 2H), 2.75 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) 13C NMR (101 MHz, cdcl₃) ¹³C NMR (101 MHz, cdcl₃) δ 197.84, 152.80, 151.58, 133.83, 133.66, 133.29, 132.86, 132.13, 131.79, 129.18, 128.49, 124.35, 114.37, 45.26. 28.45. HRMS (ESI) calcd for C₁₇ H₁₃ N₂ O₂ Cl Na [M + Na] ⁺: 335.0563; found: 335.0567.

3-acetyl-1-(4-fluorobenzyl)quinoxalin-2(1H)-one (2m): Following the general procedure, a 15 mL reaction



tube was charged with 1-(4-fluorobenzyl)quinoxalin-2(1*H*)-one (1m) (54 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate

layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(4-

fluorobenzyl)quinoxalin-2(1*H*)-one (2m) as a yellow solid (32 mg, yield = 54 %); Mp. 129.1-130 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.97 (dd, *J* = 8.0, 0.4 Hz, 1H), 7.58 (ddd, *J* = 8.8, 7.2, 0.4 Hz, 1H), 7.38 (ddd, *J* = 8.0, 7.2, 0.8 Hz, 1H), 7.31-7.25 (m, 3H), 7.03-6.99 (m, 2H), 5.47 (s, 2H), 2.74 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.89, 163.51, (d, *J_F* = 245 Hz), 161.06, 152.84 (d, *J_F* = 119 Hz), 151.65, 133.72, 132.82, 132.15, 131.77, 130.56, 130.53, 128.97, (d, *J_F* = 8 Hz), 128.89, 124.30, 116.06, (d, *J_F* = 21.7 Hz),115.85, 114.40, 45.22, 28.47. HRMS (ESI) calcd for C₁₇ H₁₃ N₂ O₂ F Na [M + Na] +: 319.0859; found: 319.0863

3-acetyl-1-(4-(trifluoromethyl)benzyl)quinoxalin-2(1H)-one (2n): Following the general procedure, a 15



mL reaction tube was charged with 1-(4-(trifluoromethyl)benzyl)quinoxalin-2(1*H*)one (1n) (64 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl

acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(4-(trifluoromethyl)benzyl)quinoxalin-2(1*H*)-one (2n) as a yellow solid (36 mg, yield = 52 %); Mp. 130.1-131.3 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.99 (dd, *J* = 8.0, 0.4 Hz, 1H), 7.58 (ddd, *J* = 8.4, 7.2, 1.2 Hz, 3H), 7.40 (ddd, *J* = 8.0, 7.2, 1.2 Hz, 3H), 7.23 (dd, *J* = 8.8, 1.2 Hz, 1H), 5.56 (s, 2H), 2.75 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.74, 152.76, 151.51, 138.78, 133.61, 132.98, 132.12, 131.86, 130.44, 130.11(q, *J* = 28.6 Hz), 127.32, 126.05, 126.01, 125.98, 125.94 (q, *J* = 3.7 Hz), 124.47, 122.449 (q, *J* = 270.4 Hz), 114.25, 45.45, 29.67, 28.41. HRMS (ESI) calcd for C₁₈ H₁₃ N₂ F₃ O₂ Na [M + Na] +: 369.0827; found: 369.0831.

3-acetyl-1-(2,4-dichlorobenzyl)quinoxalin-2(1H)-one (2o): Following the general procedure, a 15 mL



reaction tube was charged with 1-(2,4-dichlorobenzyl)quinoxalin-2(1*H*)-one (10) (63 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl

acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(2,4-dichlorobenzyl)quinoxalin-2(1*H*)-one (20) as a yellow solid (42 mg, yield = 61 %); Mp. 125.2-126 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.00-7.98 (m, 1H), 7.57 (ddd, *J* = 8.4,

7.2, 1.6 Hz, 1H), 7.48 (d, J = 2.0 Hz, 1H), 7.40 (ddd, J = 8.4, 7.6, 1.2 Hz, 1H), 7.09 (ddd, J = 8.4, 7.2, 2.4 Hz, 2H), 6.75 (dd, J = 8.4 Hz, 1H), 5.53 (s, 2H), 2.75 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.67, 152.81, 151.48, 134.28, 133.40, 133.29, 133.12, 132.10, 131.78, 130.54, 129.67, 128.04, 127.75, 124.58, 114.31, 77.31, 76.99, 76.67, 43.00, 28.44. HRMS (ESI) calcd for C₁₂ H₁₂ N₂ O₂ Cl₂ Na [M + Na] ⁺: 369.0174; found 369.0173.

3-acetyl-1-(naphthalen-2-ylmethyl)quinoxalin-2(1H)-one (2p): Following the general procedure, a 15 mL



CI

Мe

2q

reaction tube was charged with 1-(naphthalen-2-ylmethyl)quinoxalin-2(1H)-one (10) (60 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined

ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-(naphthalen-2-ylmethyl)quinoxalin-2(1*H*)-one (2p) as a yellow solid (43 mg, yield = 65 %); Mp 167.9-168.3 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.98-7.95 (m, 1H), 7.82-7.79 (m, 2H), 7.75 (dd, J = 8.8, 3.2 Hz, 1H), 7.66 (s, 1H) 7.54-7.49 (m, 1H), 7.47-7.45 (m, 2H), 7.41 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.37 (s, 1H), 7.35 (d, *J* = 1.2 Hz, 1H), 5. 67(s, 2H) 2.78 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.03, 153.02, 151.75, 133.89, 133.26, 132.80, 132.25, 132.20, 131.67, 129.01, 127.71, 126.51, 126.24, 125.82, 124.71, 124.23. 46.10, 28.55. HRMS (ESI) calcd for C₂₁ H₁₆ N₂ O₂ Na [M + Na] ⁺: 351.1109; found: 351.1108.

3-acetyl-7-chloro-1-methylquinoxalin-2(1H)-one (2q): Following the general procedure, a 15 mL reaction Ο tube was charged with 7-chloro-1-methylquinoxalin-2(1H)-one (1p) (41 mg, 0.2 Me mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed Ν to stir at 100° C until the completion of reaction by TLC. After completion, the

reaction mixture was cooled to room temperature and diluted with 10 mL of water.

The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-7-chloro-1methylquinoxalin-2(1*H*)-one (2q) as a yellow solid (28 mg, yield = 59 %); Mp. 191.4-192 °C; ¹H NMR $(CDCl_3, 400 \text{ MHz}) \delta 7.94 \text{ (d, } J = 2.4 \text{ Hz}, 1\text{H}), 7.63 \text{ (dd, } J = 9.2, 2.4 \text{ Hz}, 1\text{H}), 7.30 \text{ (d, } J = 8.8 \text{ Hz}, 1\text{H}), 3.71 \text{ (s, } J = 0.2 \text{ Hz}, 100 \text{ Hz})$ 3H), 2.69 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.72, 152.88, 152.38, 133.07, 132.68, 132.28, 130.51, 129.54, 115.03, 29.22, 28.38. HRMS (ESI) calcd for C_{11} H₀₉ N₂ O₂ Cl Na [M + Na] ⁺: 259.0250; found: 259.0252.

3-acetyl-7-Bromo-1-methylquinoxalin-2(1H)-one (2r): Following the general procedure, a 15 mL reaction



tube was charged with 7-Bromo-1-methylquinoxalin-2(1H)-one (1q) (51 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water.

The water layer was extracted with (3X10 mL) of ethyl acetate, and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-7-Bromo-1-methylquinoxalin-2(1*H*)-one (2r) as a yellow solid (35 mg, yield = 63 %); Mp 188-189.2 °C;¹H NMR (CDCl₃, 400 MHz) δ 8.10 (d, *J* = 2.4 Hz, 1H), 7.76 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.24 (d, *J* = 9.2 Hz, 1H), 3.70 (s, 3H), 2.69 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.65, 152.80, 152.38, 135.39, 133.62, 133.53, 132.59, 116.71, 115.29, 29.19, 28.35. HRMS (ESI) calcd for C₁₁ H₀₉ N₂ O₂ Br Na [M + Na] ⁺: 302.9745; found: 302.9746.

3-acetyl-1-methyl-7-(trifluoromethyl)quinoxalin-2(1H)-one (2s): Following the general procedure, a 15



Me 2t mL reaction tube was charged with 1-methyl-7-(trifluoromethyl)quinoxalin-2(1H)-one (1r) (48 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and

diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-methyl-7-(trifluoromethyl)quinoxalin-2(1*H*)-one (2s) as a yellow solid (32 mg, yield = 59 %); Mp. 170-170.6 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.23 (d, *J* = 1.6 Hz, 1H), 7.89 (dd, *J* = 8.8, 2.0 Hz, 1H), 7.48 (d, *J* = 8.8 Hz, 1H), 3.75 (s, 3H), 2.70 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 197.36, 153.07, 152.44, 136.61, 131.09, 128.85, 128.81, 128.77, 128.73 (q, *J* = 3.4 Hz), 127.44(q, *J* = 41.6 Hz), 126.66, 126.33, 124.72,125.99 (q, *J* = 33.7 Hz), 122.02(q, *J* = 270.1 Hz), 114.66, 29.30, 28.25. HRMS (ESI) calcd for C₁₂ H₁₀ N₂ O₂ F₃ [M + H] ⁺: 271.0694; found: 271.0697.

3-acetyl-1-methyl-7-phenylquinoxalin-2(1*H*)-one (2t): Following the general procedure, a 15 mL reaction tube was charged with 1-methyl-7-phenylquinoxalin-2(1*H*)-one (1s) (50 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) allowed to 12

stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL) of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-methyl-7-phenylquinoxalin-2(1*H*)-one (2t) as a yellow solid (32 mg, yield = 58 %); Mp 178-178.9 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.19 (d, *J* = 2.4 Hz, 1H), 7.93 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.67-7.64 (m, 2H), 7.52-7.47 (m, 2H), 7.44-7.38 (m, 2H), 3.76 (s, 3H), 2.73 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.07, 152.78, 152.12, 138.72, 137.34, 133.55, 132.17, 131.63, 129.29, 129.11, 127.95, 126.89, 114.28, 29.15, 28.54. HRMS (ESI) calcd for C₁₂ H₁₂ N₂ O₂ Na [M + Na] ⁺: 301.1106; found: 301.1107.

3-acetyl-1-methyl-7-(p-tolyl)quinoxalin-2(1H)-one (2u): Following the general procedure, a 15 mL reaction



tube was charged with 1-methyl-7-(p-tolyl)quinoxalin-2(1*H*)-one (1t) (53 mg, 0.2 mmol), PEG-400 (0.25 M), AgNO₃ (20 mol %) and K₂S₂O₈ (2.0 equiv) and allowed to stir at 100° C until the completion of reaction by TLC. After completion, the reaction mixture was cooled to room temperature and diluted with 10 mL of water. The water layer was extracted with (3X10 mL)

of ethyl acetate and the combined ethyl acetate layer was given brine wash (1X10 mL). The final ethyl acetate layer was dried over sodium sulfate and concentrated under reduced pressure to get the crude compound. The obtained crude was purified using column chromatography by eluting from (Hex/EA = 8/2) to afford the corresponding 3-acetyl-1-methyl-7-(p-tolyl)quinoxalin-2(1*H*)-one (2u) as a yellow solid (35 mg, yield = 60 %); Mp 183-184.6 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.16 (d, *J* = 2.4 Hz, 1H), 7.91 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.56-7.54 (m, 2H), 7.40 (d, *J* = 8.8 Hz, 1H), 7.31-7.28 (m, 2H), 3.75 (s, 3H), 2.73 (s, 3H), 2.42 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 198.09, 152.76, 152.04, 137.89, 137.28, 135.79, 133.32, 132.17, 131.48, 129.81, 128.95, 126.68, 114.20, 29.11, 28.54, 21.10. HRMS (ESI) calcd for C₁₈ H₁₆ N₂ O₂ Na [M + Na]⁺: 315.1109; found: 315.1111.

(5) References:

1. M. I. Shahin, D. A. Abou El Ella, N. S. M. Ismail, K. A. M. Abouzid. Bioorg. Chem. 2014, 56, 16-26.

2. X. Zeng, C. Liu, X. Wang, J. Zhang, X. Wang, Y. Hu. Organic & Biomolecular Chemistry. 2017, 15, 8929-8935.







Spectrometer Frequency 100.69







































---0,000 -1.589 7 7, 982 7 7, 984 7 7, 985 7 7, 985 7 7, 985 7 7 7.829 7.816 7.816 7.7808 7.792 7.755 7.755 7,550 7,5517 7.982 7.964 7.964 7.958 Solvent CDC13 Spectrometer Frequency 400.40 55 M 1.00 1.00 2p -00'1 1.01 à 00.3 7.70 7.65 f1 (ppm) 7.45 3.00 7.95 7.90 7.85 7.80 7.75 7.55 7.50 7.40 7.60 7.35 ¥ 2.00 2,86-7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 f1 (ppm) ..5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0 -1 133.261 132.261 132.247 132.247 132.247 131.668 131.668 123.001 127.00 1 ~153.015 ~151.747 77.307 76.990 76.673 -129.014 133.894 133.261 132.800 132.800 132.247 132.202 --126.506 --126.237 --125.819 Solvent CDC13 Spectrometer Frequency 100.69 2p 133 131 129 f1 (ppm) 127 125











checkCIF/PLATON report

Structure factors have been supplied for datablock(s) I

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No syntax errors found. CIF dictionary Interpreting this report

Datablock: I

Bond precision: C-C = 0.0021 A Wavelength=0.71073 c=14.5759(4) Cell: a=4.6921(1) b=21.3139(7) alpha=90 beta=97.032(3) gamma=90 Temperature: 113 K Calculated Reported Volume 1446.73(7) 1446.73(7) Space group P 21/c P 1 21/c 1 -P 2ybc Hall group -P 2ybc Moiety formula C18 H16 N2 O2 C18 H16 N2 O2 Sum formula C18 H16 N2 O2 C18 H16 N2 O2 Mr 292.33 292.33 1.342 1.342 Dx,g cm-3 Ζ 4 4 Mu (mm-1) 0.089 0.089 F000 616.0 616.0 F000′ 616.26 h,k,lmax 5,25,17 5,25,17 Nref 2512 2555 Tmin,Tmax 0.978,0.982 0.832,1.000 Tmin′ 0.978 Correction method= # Reported T Limits: Tmin=0.832 Tmax=1.000 AbsCorr = MULTI-SCAN Data completeness= 0.983 Theta(max) = 24.997 R(reflections) = 0.0404(2068) wR2(reflections) = 0.1052(2512) S = 1.094Npar= 201

The following ALERTS were generated. Each ALERT has the format **test-name_ALERT_alert-type_alert-level**. Click on the hyperlinks for more details of the test.

Alert level C		
PLAT911_ALERT_3_C Missing FCF Refl Between Thmin & STh/L= 0.595	43 Rep	ort
PLAT918_ALERT_3_C Reflection(s) with I(obs) much Smaller I(calc) .	1 Che	ck
Alert level G		
PLAT909_ALERT_3_G Percentage of I>2sig(I) Data at Theta(Max) Still	65% Not	.e
PLAT933_ALERT_2_G Number of OMIT Records in Embedded .res File	3 Not	.e
PLAT941_ALERT_3_G Average HKL Measurement Multiplicity	3.8 Low	ŗ
PLAT978 ALERT 2 G Number C-C Bonds with Positive Residual Density.	5 Inf	0

0 ALERT level A = Most likely a serious problem - resolve or explain 0 ALERT level B = A potentially serious problem, consider carefully 2 ALERT level C = Check. Ensure it is not caused by an omission or oversight 4 ALERT level G = General information/check it is not something unexpected 0 ALERT type 1 CIF construction/syntax error, inconsistent or missing data 2 ALERT type 2 Indicator that the structure model may be wrong or deficient 4 ALERT type 3 Indicator that the structure quality may be low 0 ALERT type 4 Improvement, methodology, query or suggestion 0 ALERT type 5 Informative message, check

checkCIF publication errors

```
Alert level A
PUBL004_ALERT_1_A The contact author's name and address are missing,
    ______publ_contact_author_name and __publ_contact_author_address.
PUBL005_ALERT_1_A __publ_contact_author_email, __publ_contact_author_fax and
    ______publ_contact_author_phone are all missing.
    At least one of these should be present.
PUBL006_ALERT_1_A __publ_requested_journal is missing
    e.g. 'Acta Crystallographica Section C'
PUBL008_ALERT_1_A __publ_section_title is missing. Title of paper.
PUBL009_ALERT_1_A __publ_author_name is missing. List of author(s) name(s).
PUBL010_ALERT_1_A __publ_author_address is missing. Author(s) address(es).
PUBL012_ALERT_1_A __publ_section_abstract is missing.
```

7 ALERT level A = Data missing that is essential or data in wrong format 0 ALERT level G = General alerts. Data that may be required is missing

Publication of your CIF

You should attempt to resolve as many as possible of the alerts in all categories. Often the minor alerts point to easily fixed oversights, errors and omissions in your CIF or refinement strategy, so attention to these fine details can be worthwhile. In order to resolve some of the more serious problems it may be necessary to carry out additional measurements or structure refinements. However, the nature of your study may justify the reported deviations from journal submission requirements and the more serious of these should be commented upon in the discussion or experimental section of a paper or in the "special_details" fields of the CIF. *checkCIF* was carefully designed to identify outliers and unusual parameters, but every test has its limitations and alerts that are not important in a particular case may appear. Conversely, the absence of alerts does not guarantee there are no aspects of the results needing attention. It is up to the individual to critically assess their own results and, if necessary, seek expert advice.

If level A alerts remain, which you believe to be justified deviations, and you intend to submit this CIF for publication in a journal, you should additionally insert an explanation in your CIF using the Validation Reply Form (VRF) below. This will allow your explanation to be considered as part of the review process.

Validation response form

Please find below a validation response form (VRF) that can be filled in and pasted into your CIF.

```
# start Validation Reply Form
vrf PUBL004 GLOBAL
;
PROBLEM: The contact author's name and address are missing,
RESPONSE: ...
;
_vrf_PUBL005 GLOBAL
PROBLEM: _publ_contact_author_email, _publ_contact_author_fax and
RESPONSE: ...
vrf PUBL006 GLOBAL
PROBLEM: publ requested journal is missing
RESPONSE: ...
_vrf_PUBL008 GLOBAL
PROBLEM: _publ_section_title is missing. Title of paper.
RESPONSE: ...
_vrf_PUBL009 GLOBAL
;
PROBLEM: publ author name is missing. List of author(s) name(s).
RESPONSE: ...
vrf PUBL010 GLOBAL
;
PROBLEM: publ author address is missing. Author(s) address(es).
RESPONSE: ...
vrf PUBL012 GLOBAL
```

```
PROBLEM: _publ_section_abstract is missing.
RESPONSE: ...
;
# end Validation Reply Form
```

If you wish to submit your CIF for publication in Acta Crystallographica Section C or E, you should upload your CIF via the web. If you wish to submit your CIF for publication in IUCrData you should upload your CIF via the web. If your CIF is to form part of a submission to another IUCr journal, you will be asked, either during electronic submission or by the Co-editor handling your paper, to upload your CIF via our web site.

PLATON version of 10/08/2020; check.def file version of 06/08/2020

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Datablock I - ellipsoid plot
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checkCIF/PLATON report

Structure factors have been supplied for datablock(s) I

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No syntax errors found. CIF dictionary Interpreting this report

Datablock: I

Bond precision: C-C = 0.0020 A Wavelength=0.71073 c=22.5179(2) Cell: a=7.7070(1) b=17.6134(2) alpha=90 beta=93.088(1) gamma=90 Temperature: 113 K Calculated Reported Volume 3052.29(6) 3052.29(6) Space group P 21/c P 1 21/c 1 Hall group -P 2ybc -P 2ybc 2(C17 H12 Cl2 N2 O2) Moiety formula C17 H12 Cl2 N2 O2 Sum formula C17 H12 C12 N2 O2 C34 H24 C14 N4 O4 Mr 347.19 694.37 1.511 1.511 Dx,g cm-3 Ζ 8 4 Mu (mm-1) 0.436 0.436 F000 1424.0 1424.0 F000′ 1426.89 h,k,lmax 9,22,28 9,22,28 Nref 6693 6459 Tmin,Tmax 0.877,0.897 0.638,1.000 Tmin′ 0.877 Correction method= # Reported T Limits: Tmin=0.638 Tmax=1.000 AbsCorr = MULTI-SCAN Data completeness= 0.965 Theta(max) = 27.039 R(reflections) = 0.0305(5716) wR2(reflections) = 0.0777(6459) S = 1.059Npar= 417

The following ALERTS were generated. Each ALERT has the format **test-name_ALERT_alert-type_alert-level**. Click on the hyperlinks for more details of the test.

Alert level C PLAT911_ALERT_3_C Missing FCF Refl Between Thmin & STh/L= 0.600	5	Report
Alert level G		
PLAT042 ALERT 1 G Calc. and Reported MoietyFormula Strings Differ	Please	Check
PLAT045 ALERT 1 G Calculated and Reported Z Differ by a Factor	2.00	Check
PLAT143 ALERT 4 G s.u. on c - Axis Small or Missing	0.00020	Ang.
PLAT380 ALERT 4 G Incorrectly? Oriented X(sp2)-Methyl Moiety	C17	Check
PLAT432_ALERT_2_G Short Inter XY Contact 01C32	2.98	Ang.
x,y,z =	1_555 Chec	ck
PLAT910_ALERT_3_G Missing # of FCF Reflection(s) Below Theta(Min).	2	Note
PLAT912_ALERT_4_G Missing # of FCF Reflections Above STh/L= 0.600	227	Note
PLAT933_ALERT_2_G Number of OMIT Records in Embedded .res File	3	Note
PLAT978_ALERT_2_G Number C-C Bonds with Positive Residual Density.	15	Info
<pre>0 ALERT level A = Most likely a serious problem - resolve or exp 0 ALERT level B = A potentially serious problem, consider carefu 1 ALERT level C = Check. Ensure it is not caused by an omission 9 ALERT level G = General information/check it is not something</pre>	lain lly or oversigh unexpected	nt

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2 ALERT type 1 CIF construction/syntax error, inconsistent or missing data
3 ALERT type 2 Indicator that the structure model may be wrong or deficient
2 ALERT type 3 Indicator that the structure quality may be low
3 ALERT type 4 Improvement, methodology, query or suggestion
0 ALERT type 5 Informative message, check
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checkCIF publication errors

🔩 Alert level A

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PUBL004_ALERT_1_A The contact author's name and address are missing,
    ______publ_contact_author_name and __publ_contact_author_address.
PUBL005_ALERT_1_A __publ_contact_author_email, __publ_contact_author_fax and
    ______publ_contact_author_phone are all missing.
    ______At least one of these should be present.
PUBL006_ALERT_1_A __publ_requested_journal is missing
    ______e.g. 'Acta Crystallographica Section C'
PUBL008_ALERT_1_A __publ_section_title is missing. Title of paper.
PUBL009_ALERT_1_A __publ_author_name is missing. List of author(s) name(s).
PUBL010_ALERT_1_A __publ_author_address is missing. Author(s) address(es).
PUBL012_ALERT_1_A __publ_section_abstract is missing.
    ______Abstract of paper in English.
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7 ALERT level A = Data missing that is essential or data in wrong format 0 ALERT level G = General alerts. Data that may be required is missing

Publication of your CIF

You should attempt to resolve as many as possible of the alerts in all categories. Often the minor alerts point to easily fixed oversights, errors and omissions in your CIF or refinement strategy, so attention to these fine details can be worthwhile. In order to resolve some of the more serious problems it may be necessary to carry out additional measurements or structure refinements. However, the nature of your study may justify the reported deviations from journal submission requirements and the more serious of these should be commented upon in the discussion or experimental section of a paper or in the "special_details" fields of the CIF. *checkCIF* was carefully designed to identify outliers and unusual parameters, but every test has its limitations and alerts that are not important in a particular case may appear. Conversely, the absence of alerts does not guarantee there are no aspects of the results needing attention. It is up to the individual to critically assess their own results and, if necessary, seek expert advice.

If level A alerts remain, which you believe to be justified deviations, and you intend to submit this CIF for publication in a journal, you should additionally insert an explanation in your CIF using the Validation Reply Form (VRF) below. This will allow your explanation to be considered as part of the review process.

Validation response form

Please find below a validation response form (VRF) that can be filled in and pasted into your CIF.

```
# start Validation Reply Form
vrf PUBL004 GLOBAL
;
PROBLEM: The contact author's name and address are missing,
RESPONSE: ...
;
_vrf_PUBL005 GLOBAL
PROBLEM: _publ_contact_author_email, _publ_contact_author_fax and
RESPONSE: ...
vrf PUBL006 GLOBAL
PROBLEM: publ requested journal is missing
RESPONSE: ...
_vrf_PUBL008 GLOBAL
PROBLEM: _publ_section_title is missing. Title of paper.
RESPONSE: ...
_vrf_PUBL009 GLOBAL
;
PROBLEM: publ author name is missing. List of author(s) name(s).
RESPONSE: ...
vrf PUBL010 GLOBAL
;
PROBLEM: publ author address is missing. Author(s) address(es).
RESPONSE: ...
vrf PUBL012 GLOBAL
```

```
PROBLEM: _publ_section_abstract is missing.
RESPONSE: ...
;
# end Validation Reply Form
```

If you wish to submit your CIF for publication in Acta Crystallographica Section C or E, you should upload your CIF via the web. If you wish to submit your CIF for publication in IUCrData you should upload your CIF via the web. If your CIF is to form part of a submission to another IUCr journal, you will be asked, either during electronic submission or by the Co-editor handling your paper, to upload your CIF via our web site.

PLATON version of 22/04/2020; check.def file version of 09/03/2020

```
Datablock I - ellipsoid plot
```

