

Supporting Information

C-H Bond Cleavage-Enabled Aerobic Ring-Opening Reaction of in Situ Formed 2-Aminobenzofuran-3(2H)-ones

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Corresponding Author

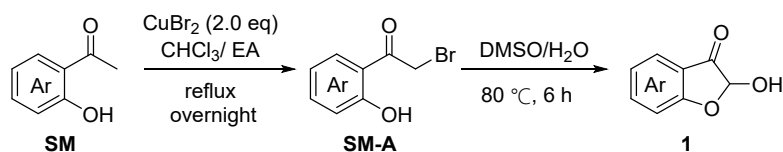
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zhjiang@must.edu.mo

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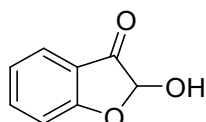
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1. Synthesis and characterization of substrates 1a-1g



The intermediates **SM-A** were synthesized according the published methods^[1] and this final material was used without further purification.

To a solution of **SM-A** in DMSO (1 M) was added H₂O (1 M) at room temperature. After stirring 6 hours at 80 °C, the reaction mixture was allowed to cooled to room temperature and subsequently diluted with saturated aqueous NaHCO₃. The crude products were extracted with EtOAc and the organic extracts were washed with brine, dried over anhydrous sodium sulfate and concentrated in vacuo. The residue was purified by column chromatography to afford **1a-1g**.



2-hydroxybenzofuran-3(2H)-one (**1a**)^[2]

*R*_f: 0.27 (petroleum ether/EtOAc = 2:1)

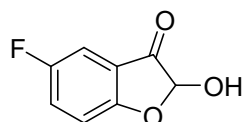
Light yellow solid.

Yield: 72% (from 2'-hydroxyacetophenone)

¹H NMR (600 MHz, CDCl₃) δ 7.64-7.67 (m, 2H), 7.09-7.12 (m, 2H), 5.58 (s, 1H), 4.03 (br, 1H).

¹³C NMR (151 MHz, CDCl₃) δ 197.72, 171.35, 139.29, 125.04, 122.61, 119.11, 113.54, 96.73.

HRMS (ESI) *m/z* calcd for C₈H₇O₃ (M + H)⁺: 151.0390; found: 151.0314.



5-fluoro-2-hydroxybenzofuran-3(2H)-one (**1b**)^[2]

*R*_f: 0.29 (petroleum ether/EtOAc = 2:1)

Light white solid.

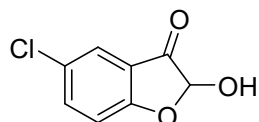
Yield: 75% (from 1-(5-fluoro-2-hydroxyphenyl)ethan-1-one)

¹H NMR (600 MHz, DMSO) δ 8.12 (d, *J* = 9.4 Hz, 1H), 7.61-7.64 (m, 1H), 7.45 (m, 1H), 7.23 (m, 1H), 5.64 (d, *J* = 9.4 Hz, 1H).

¹³C NMR (151 MHz, DMSO) δ 198.78 (d, *J*_{CF} = 2.8 Hz), 167.47, 157.45 (d, *J*_{CF} = 240.1 Hz), 126.91 (d, *J*_{CF} = 25.7 Hz), 120.30 (d, *J*_{CF} = 7.8 Hz), 115.36, 109.93 (d, *J*_{CF} = 23.7 Hz), 99.21.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ -119.67.

HRMS (ESI) *m/z* calcd for C₈H₆FO₃ (M + H)⁺: 169.0230; found: 169.0291.



1-(5-chloro-2-hydroxyphenyl)ethan-1-one (**1c**)

m.p. 185 °C

R_f: 0.32 (petroleum ether/EtOAc = 2:1)

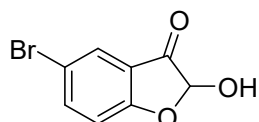
Light white solid.

Yield: 68% (from 1-(5-chloro-2-hydroxyphenyl)ethan-1-one).

¹H NMR (600 MHz, DMSO) δ 8.18 (d, *J* = 9.5 Hz, 1H), 7.75-7.77 (m, 1H), 7.67 (d, *J* = 2.3 Hz, 1H), 7.23 (d, *J* = 8.8 Hz, 1H), 5.66 (d, *J* = 9.4 Hz, 1H).

¹³C NMR (151 MHz, DMSO) δ 197.97, 169.58, 138.99, 126.43, 123.91, 121.21, 115.76, 99.15.

HRMS (ESI) *m/z* calcd for C₈H₆ClO₃ (M + H)⁺: 185.0000; found: 184.9996.



5-bromo-2-hydroxybenzofuran-3(2H)-one (**1d**) ^[2]

R_f: 0.33 (petroleum ether/EtOAc = 2:1)

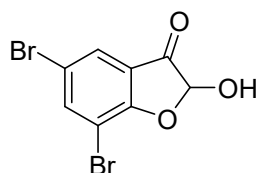
White solid.

Yield: 57% (from 1-(5-bromo-2-hydroxyphenyl)ethan-1-one).

¹H NMR (600 MHz, DMSO) δ 8.18 (d, *J* = 9.4 Hz, 1H), 7.86-7.88 (m, 1H), 7.78 (d, *J* = 2.2 Hz, 1H), 7.18 (d, *J* = 8.8 Hz, 1H), 5.65 (d, *J* = 9.4 Hz, 1H).

¹³C NMR (151 MHz, DMSO) δ 197.29, 169.46, 141.19, 126.44, 121.33, 115.72, 113.42, 98.53.

HRMS (ESI) *m/z* calcd for C₈H₄BrO₃ (M - H)⁻: 228.9323; found: 228.9353.



5,7-dibromo-2-hydroxybenzofuran-3(2H)-one (**1e**)

m.p. 174 °C

R_f: 0.44 (petroleum ether/EtOAc = 2:1)

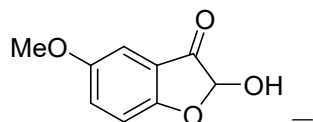
Yellow solid.

Yield: 63%.

¹H NMR (600 MHz, DMSO) δ 8.40 (d, *J* = 8.1 Hz, 1H), 8.23 (d, *J* = 2.0 Hz, 1H), 7.83 (d, *J* = 2.0 Hz, 1H), 5.79 (d, *J* = 8.0 Hz, 1H).

¹³C NMR (151 MHz, DMSO) δ 196.64, 166.20, 142.29, 125.94, 122.47, 113.73, 107.11, 99.59.

HRMS (ESI) *m/z* calcd for C₈H₃Br₂O₃ (M - H)⁻: 306.8449; found: 306.8417.



2-hydroxy-5-methoxybenzofuran-3(2H)-one (**1f**)^[2]

*R*_f: 0.21 (petroleum ether/EtOAc = 2:1)

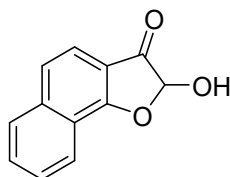
Yellow solid.

Yield: 69%.

¹H NMR (600 MHz, DMSO) δ 7.99 (d, *J* = 9.3 Hz, 1H), 7.34-7.36 (m, 1H), 7.12 (d, *J* = 9.0 Hz, 1H), 7.06 (d, *J* = 2.9 Hz, 1H), 5.56 (d, *J* = 9.3 Hz, 1H).

¹³C NMR (151 MHz, DMSO) δ 198.65, 165.74, 154.38, 127.99, 119.26, 114.31, 104.91, 98.29, 55.84.

HRMS (ESI) *m/z* calcd for C₉H₉O₄ (M + H)⁺: 181.0495; found: 181.0491.



2-hydroxynaphtho[1,2-b]furan-3(2H)-one (**1g**)^[2]

*R*_f: 0.27 (petroleum ether/EtOAc = 2:1)

Yellow solid.

Yield: 42%.

¹H NMR (600 MHz, DMSO) δ 8.29 (d, *J* = 9.6 Hz, 1H), 8.19 (d, *J* = 8.1 Hz, 1H), 8.05 (d, *J* = 8.2 Hz, 1H), 7.82 (t, *J* = 7.6 Hz, 1H), 7.70 (t, *J* = 7.6 Hz, 1H), 7.59 (d, *J* = 8.5 Hz, 1H), 7.51 (d, *J* = 8.5 Hz, 1H), 5.84 (d, *J* = 9.6 Hz, 1H).

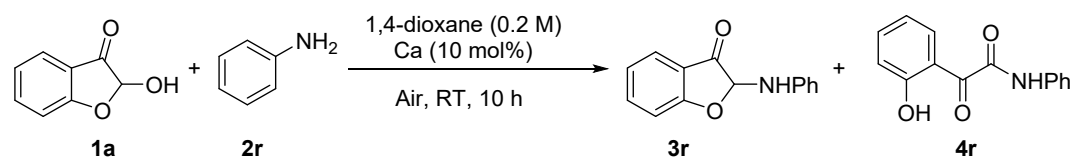
¹³C NMR (151 MHz, DMSO) δ 197.38, 171.06, 138.53, 131.15, 128.70, 127.23, 122.14, 121.96, 120.79, 118.67, 113.94, 99.43.

HRMS (ESI) *m/z* calcd for C₁₂H₉O₃ (M + H)⁺: 201.0546; found: 201.0549.

2. Optimization studies for the reaction of **1** with arylamines

To smoothly promote conversion of **1** with arylamines into *o*-hydroxyaryl glyoxylamides, the step-wise screening of reaction conditions was operated. For condensation reaction to access *N,O*-acetals, reactions of **1a** with aniline **2r** were performed in the presence of a catalytic amount of a Lewis or Brønsted acid (10 mol%) at room temperature in 1,4-dioxane under open air (Table S1). Among a few acid catalysts tested (Table S1, Entries 2-5), DPP (diphenyl phosphate) was found to be the most effective (Table S1, Entry 4), and the *N,O*-acetal **3r** was obtained in 93% yield, but the ring-opening product **4r** did not be observed.

Table S1. Condition optimization for condensation reactions. ^[a]



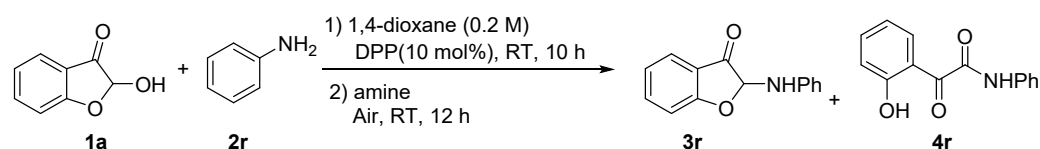
Entry	Ca.	Yield ^[b] of 3r (%)	Yield ^[b] of 4r (%)
1 ^[c]	None	21	ND ^[e]

2	TfOH	72	ND
3	TsOH.H ₂ O	87	ND
4	DPP ^[d]	93	ND
5	BF ₃ .Et ₂ O	85	ND

[a] Reaction conditions: **1a** (30 mg, 0.2 mmol), **2r** (19 mg, 0.2 mmol), catalyst (0.02 mmol), and 1,4-dioxane (1 mL) were stirred at room temperature (RT) for 10 h under open air. [b] Determined by ¹H NMR analysis of the crude reaction mixture with dibromomethane as an internal standard. [c] Incomplete conversion was observed. [d] DPP: Diphenyl phosphate. [e] Not detected.

With the optimized catalyst for condensation reactions at hand, various amines, including primary alkylamines, secondary alkylamines and arylamine, were allowed to direct the ring-opening reaction of *N,O*-acetal **3r**. As summarized in Table S2, alkylamines rather than arylamines facilitated the transformation, in which the desired product **4r** was obtained in the highest yield when diethylamine was used as the promoter (Table S2, Entry 2). Furthermore, the yield of **4r** decreased to 30% with the amount of diethylamine reduced to 0.5 equivalent (Table S2, Entry 6).

Table S2. Condition optimization for ring-opening reactions. ^[a]



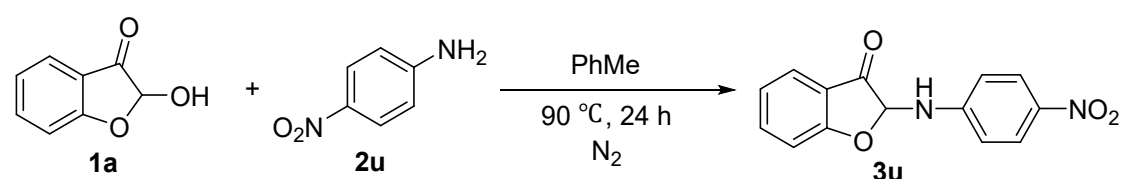
Entry	Amines	Yield ^[b] of 3r	Yield ^[b] of 4r
1	PhNH ₂	37%	ND ^[c]
2	Diethylamine	ND	76 (73 ^[d])
3	Pyrrolidine	ND	60
4	<i>tert</i> -Butylamine	ND	73
5	Propylamine	ND	47
6 ^[e]	Diethylamine	ND	30

[a] Reaction conditions: **1a** (30 mg, 0.2 mmol), **2r** (19mg, 0.2 mmol), DPP (5 mg, 0.02 mmol) and 1,4-dioxane (1 mL) at RT for 10 h followed by the addition of amine (0.2 mmol), stirring continued under open air at RT for 12 h. [b] Determined by ¹H NMR analysis of the crude reaction mixture with dibromomethane as an internal standard. [c] Not detected. [d] Isolated yield. [e] 0.5 equivalents of diethylamine were used.

3. Mechanistic studies

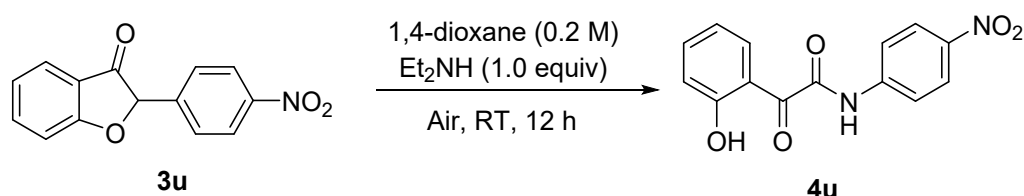
1) The control experiment of semicyclic *N,O*-acetal **3u**

Synthesis of **3u**:



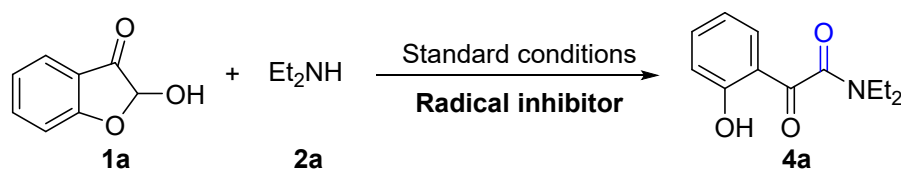
A reaction tube was charged with the 2-hydroxybenzofuran-3(2H)-one **1a** (75 mg, 0.5 mmol, 1.0 eq.), *p*-nitroaniline **2u** (69 mg, 0.5 mmol, 1.0eq) and PhMe (3 mL). The mixture was heated to 90 °C oil bath temperature for 24 hours while stirring under nitrogen atmosphere. After cooling the precipitate was filtered and rinsed with a minimum amount of cold PhMe. The yellow solid was dried under vacuum to afford semicyclic *N,O*-acetal **3u** (121 mg, 90%). m.p. 154 °C. *R*_f: 0.38 (petroleum ether/EtOAc = 4:1). ¹H NMR (600 MHz, DMSO) δ 8.38 (d, *J* = 9.5 Hz, 1H), 8.15 (d, *J* = 9.1 Hz, 2H), 7.79-7.82 (m, 1H), 7.74 (d, *J* = 7.4 Hz, 1H), 7.26 (d, *J* = 8.3 Hz, 1H), 7.21 (t, *J* = 7.5 Hz, 1H), 6.99 (d, *J* = 9.1 Hz, 2H), 6.31 (d, *J* = 9.5 Hz, 1H). ¹³C NMR (151 MHz, DMSO) δ 196.05, 170.91, 152.02, 139.44, 139.10, 125.97, 124.33, 122.27, 119.76, 113.71, 113.16, 86.67. HRMS (ESI) *m/z* calcd for C₁₄H₁₀N₂O₄ (M + H)⁺: 271.0713; found: 271.0716.

The control experiment of **3u**:



To a solution of **3u** (54 mg, 0.2 mmol) in 1,4-dioxane (1 mL) was diethylamine (15 mg, 0.2 mmol) at RT, then the reaction mixture was stirred at RT under open air. After 12 hours, the mixture was diluted with HCl aqueous solution (0.2 N, 5 mL) and extracted with ethyl acetate (2×5 mL). The combined organic extracts were washed with brine (5 ml), dried over anhydrous sodium sulfate, filtered and concentrated in vacuo. The residue was purified by column chromatography on silica gel using petroleum ether/EtOAc (4:1) as the eluent to give the desired product **4u** in 57% yield.

2) Radical-trapping experiments



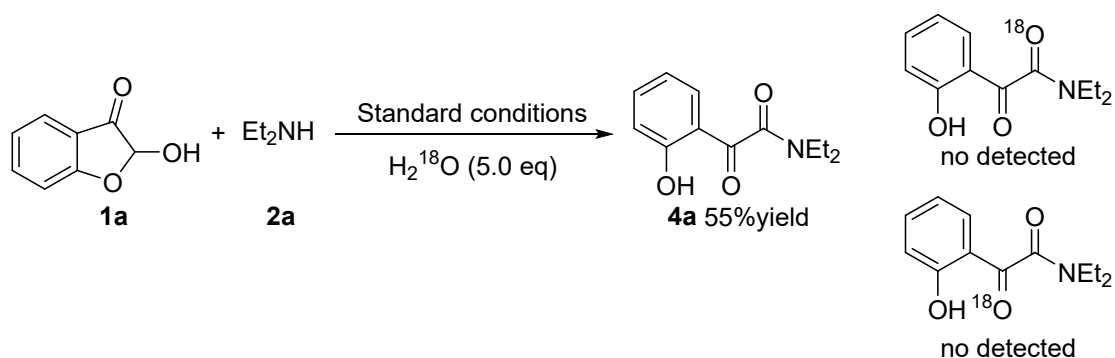
Radical inhibitor	Yield of 4a (%)
None	96
TEMPO	98
BHT	95
TRION	94

Reaction conditions: **1a** (30 mg, 0.2 mmol), **2a** (29 mg, 0.4 mmol), Radical inhibitor (0.4 mmol), in 1,4-dioxane (1.0 mL), at RT, under open air, for 14 h. Isolated yields.

The reactions performed well in the TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy) or BHT (2,6-di-tert-butyl-4-methylphenol) producing the desired product **4a** in 98% and 95% yields, respectively, which indicate that a radical process might not be involved in the present reaction

system. The superoxide radical anion scavenger TIRON (sodium 4,5-dihydroxybenzene-1,3-disulfonate)^[3] was added to the model reaction. However, the reaction did not be inhibited and **4a** was obtained in 94% isolated yield, which may exclude reactive oxygen species in this transformation.

3) Control experiment of ¹⁸O-labeled water



Reaction conditions: **1a** (30 mg, 0.2 mmol), **2a** (29 mg, 0.4 mmol), H^{18}O (20 mg, 1.0 mmol), in 1,4-dioxane (1.0 mL), at RT, under open air, for 14 h. Isolated yield. The ^{18}O in product **4a** was determined by HRMS and no ^{18}O incorporation was observed.

4) The consumption of Et_2NH detected by ^1H NMR.

At the end of model reaction of **1a** with Et_2NH , the solvent was concentrated in vacuo under room temperature and appropriate amount of residue was dissolved CDCl_3 for ^1H NMR analysis, as shown in Figure S1a. By comparing the ^1H NMR spectra of the reaction mixture with that of **4a** (Figure S1b) and diethylamine (Figure S1c), we found excessive diethylamine was consumed completely.

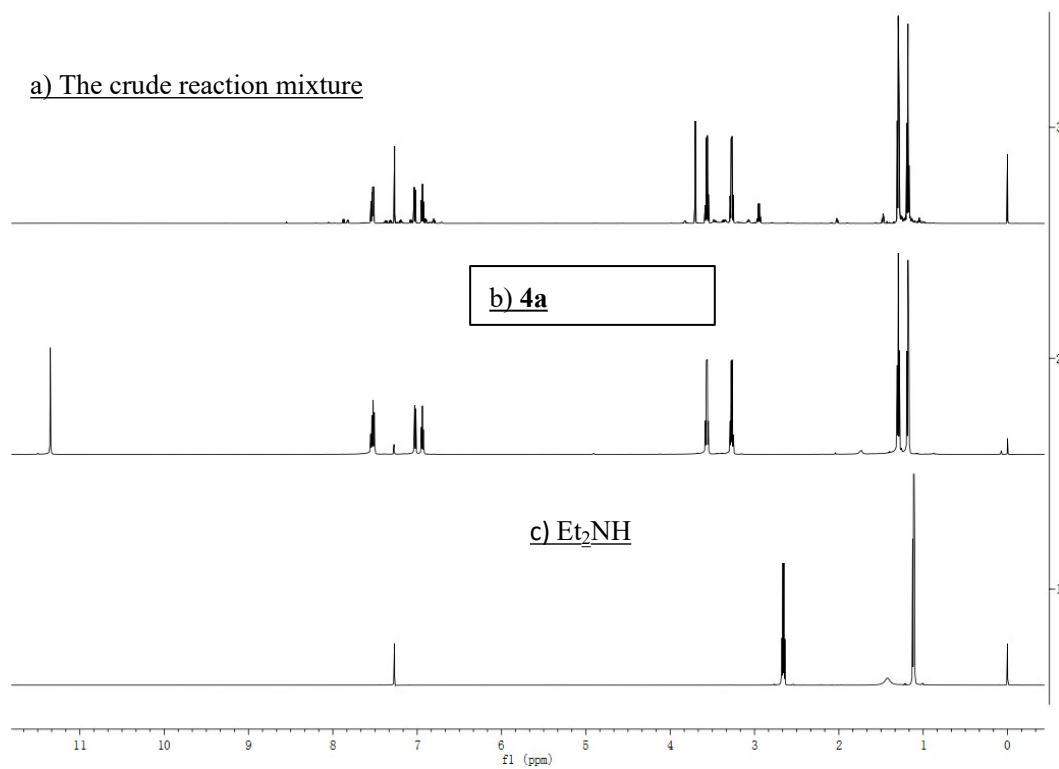
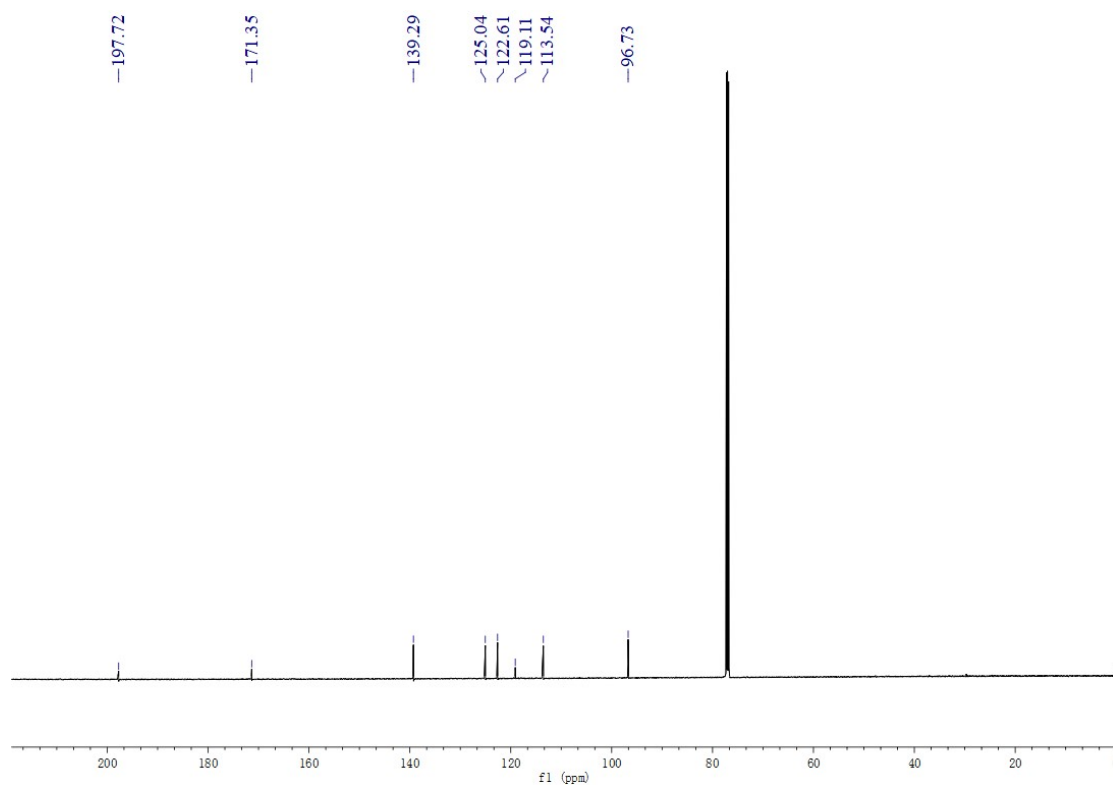
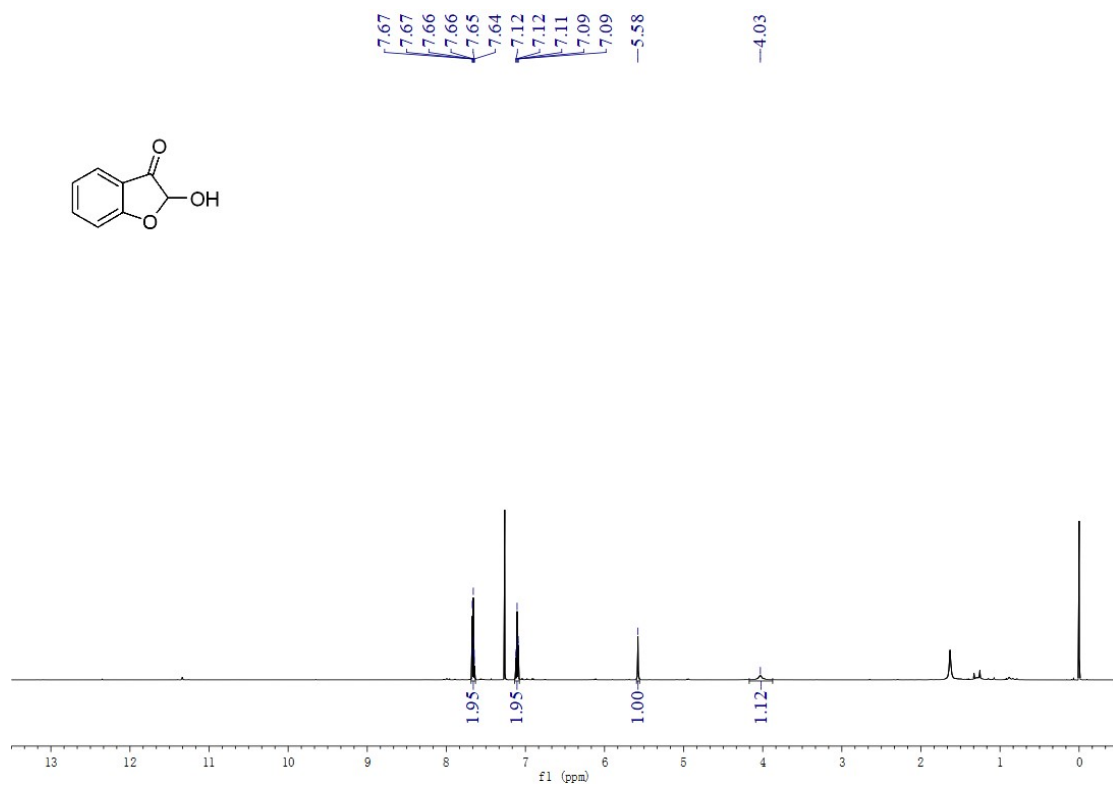
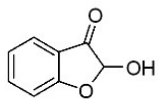


Figure S1. ¹H NMR spectra of a) reaction mixture, b) **4a** and c) **2a**

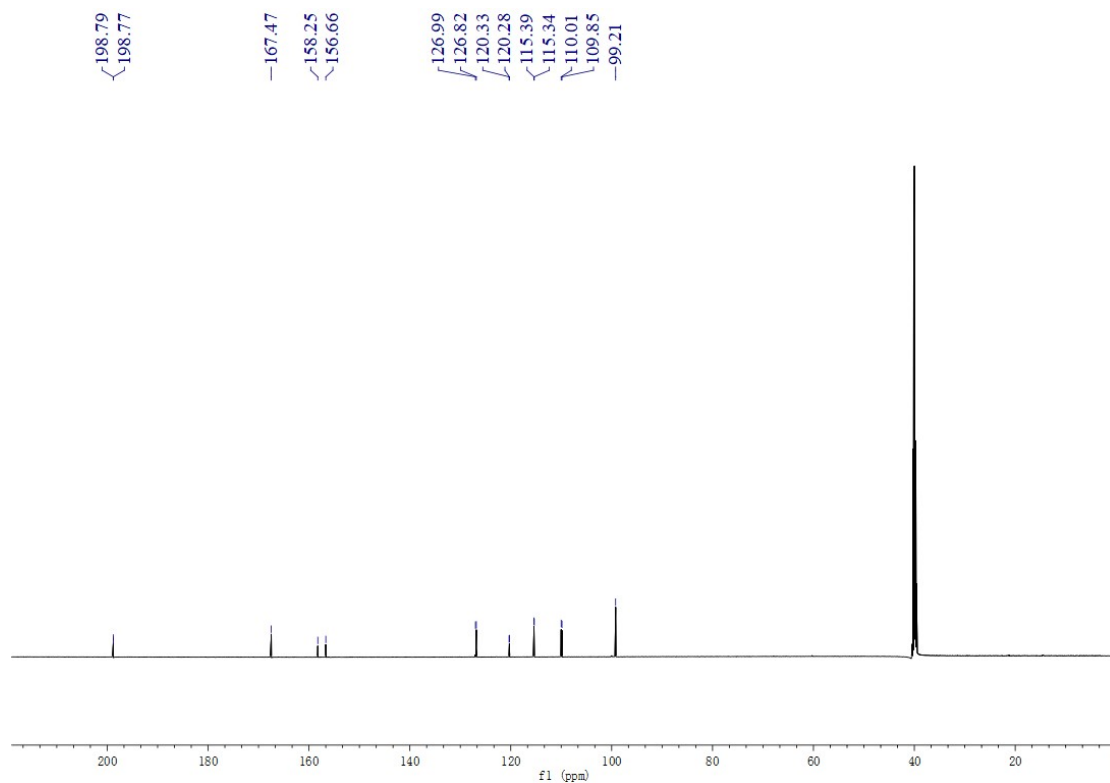
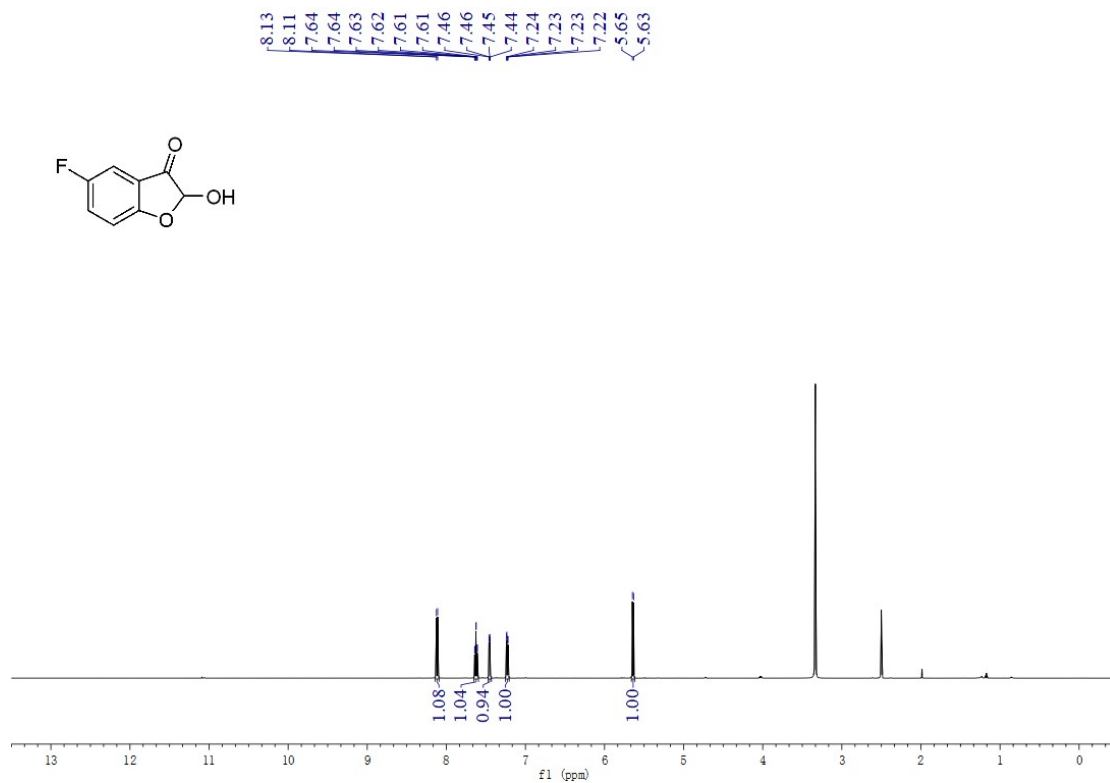
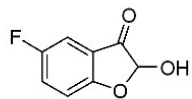
4. References

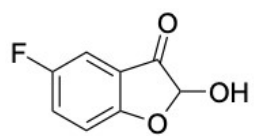
- [1] D. S. Black, N. Kumar, D. B. McConnell, *Tetrahedron* **2001**, *57*, 2203.
- [2] F. F. Gao, W. J. Xue, J. G. Wang, A. X. Wu, *Tetrahedron* **2014**, *70*, 4331.
- [3] W. D. Castro-Godoy, L. C. Schmidt, J. E. Arguello, *Eur. J. Org. Chem.* **2019**, *2019*, 3035.

¹H and ¹³C NMR Spectra for **1a**

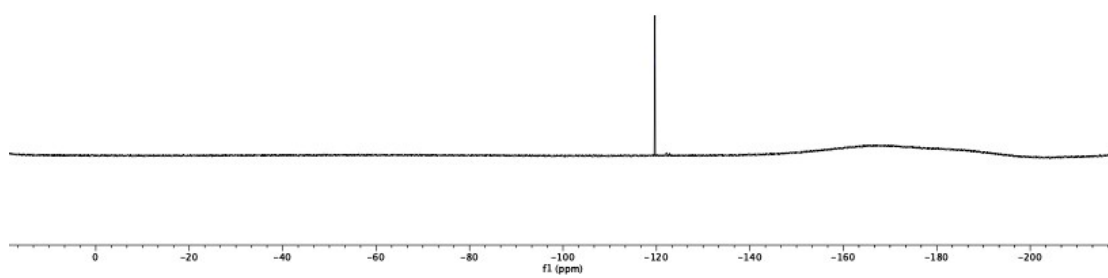


^1H , ^{13}C and ^{19}F NMR Spectra for **1b**

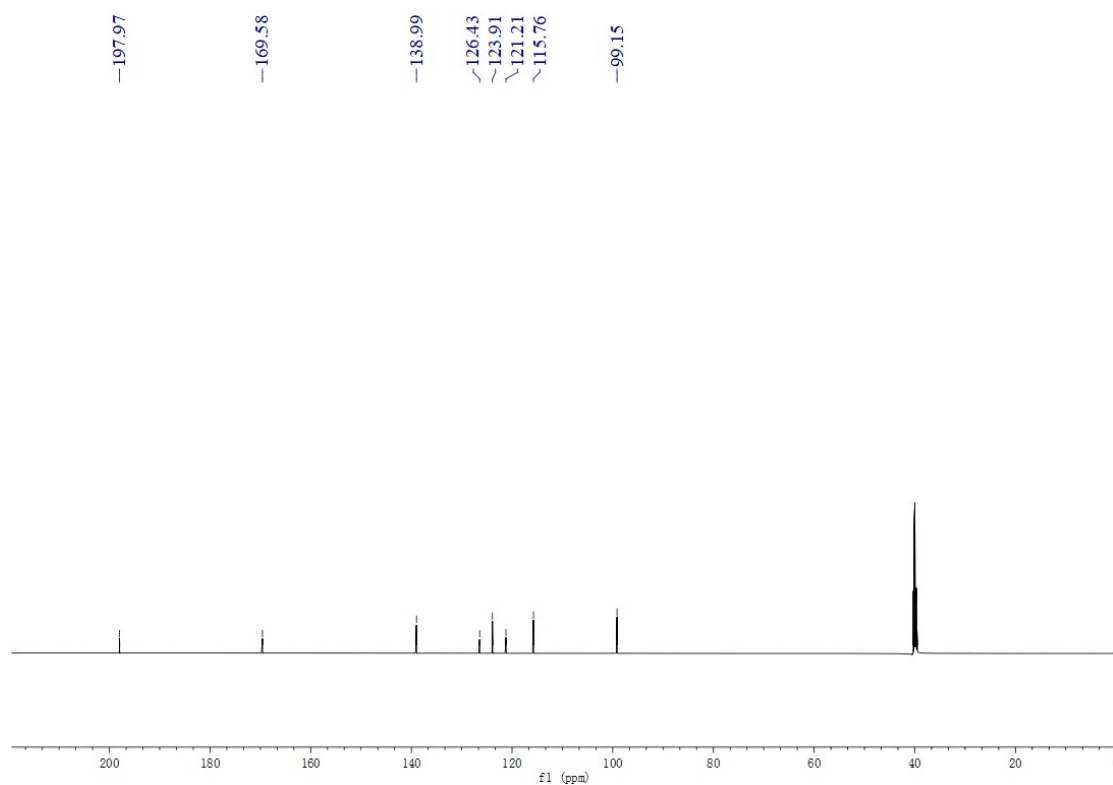
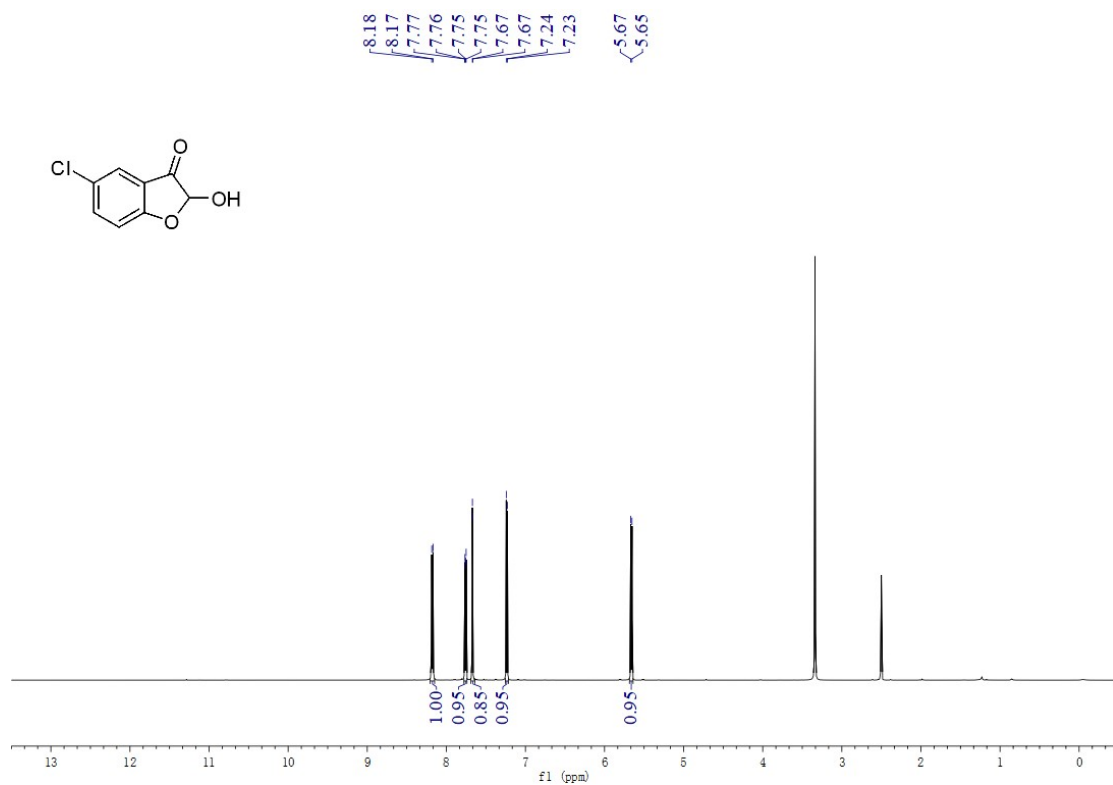
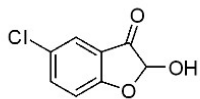




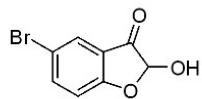
--119.67



¹H and ¹³C NMR Spectra for 1c

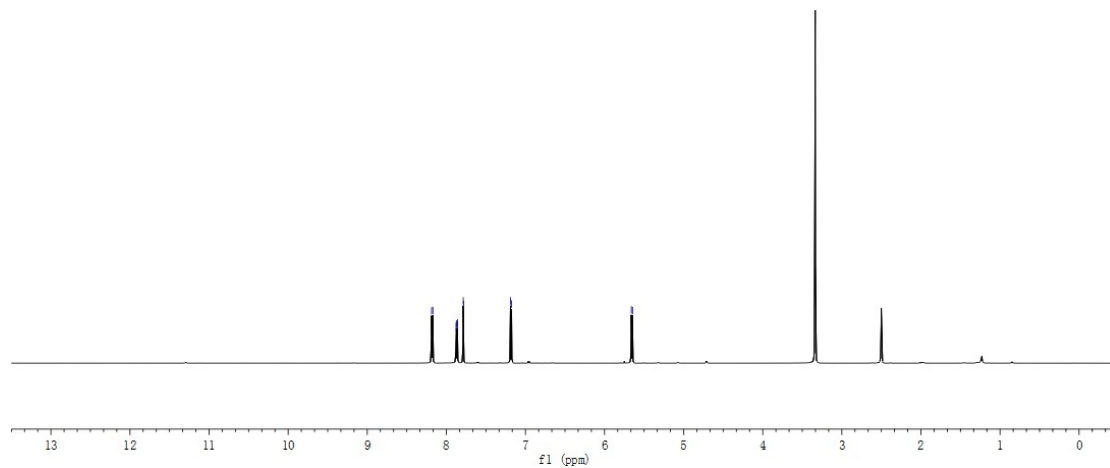


¹H and ¹³C NMR Spectra for **1d**

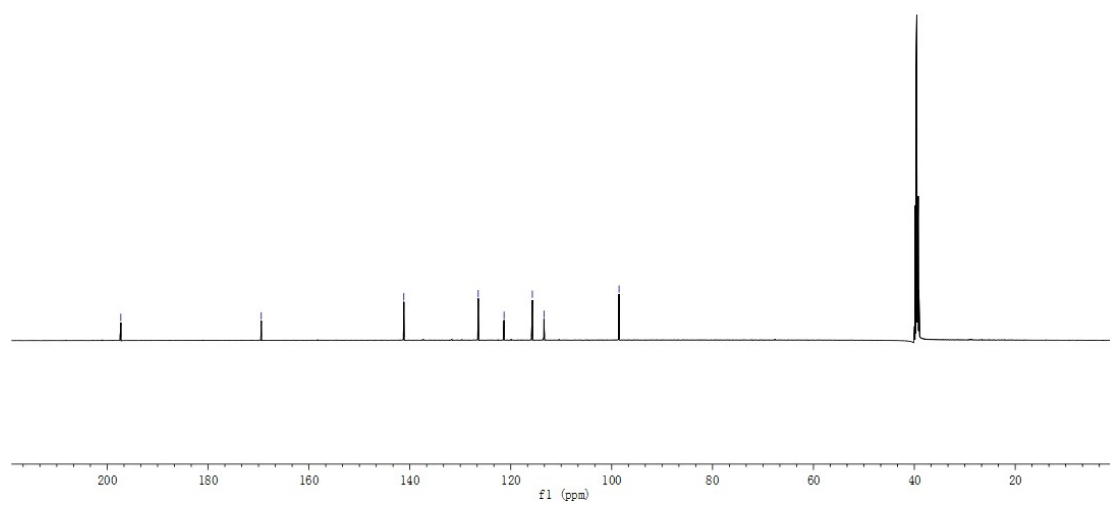


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8.17
7.88
7.87
7.86
7.79
7.78
7.19
7.18

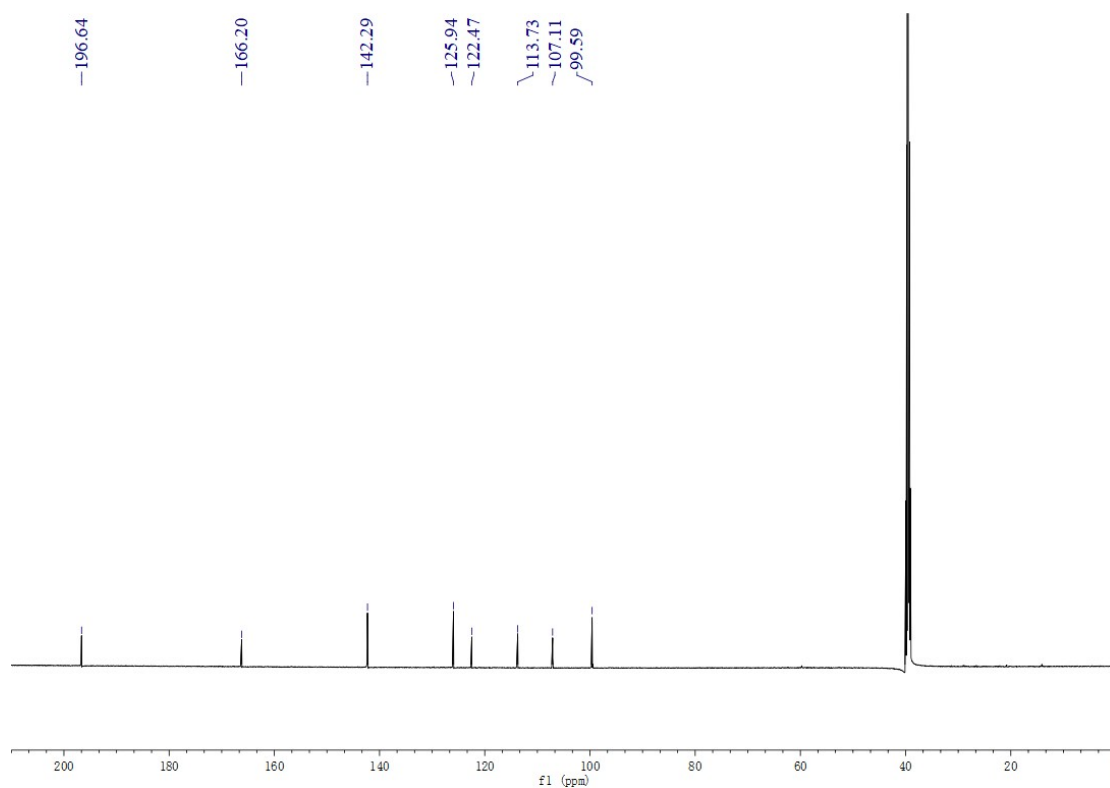
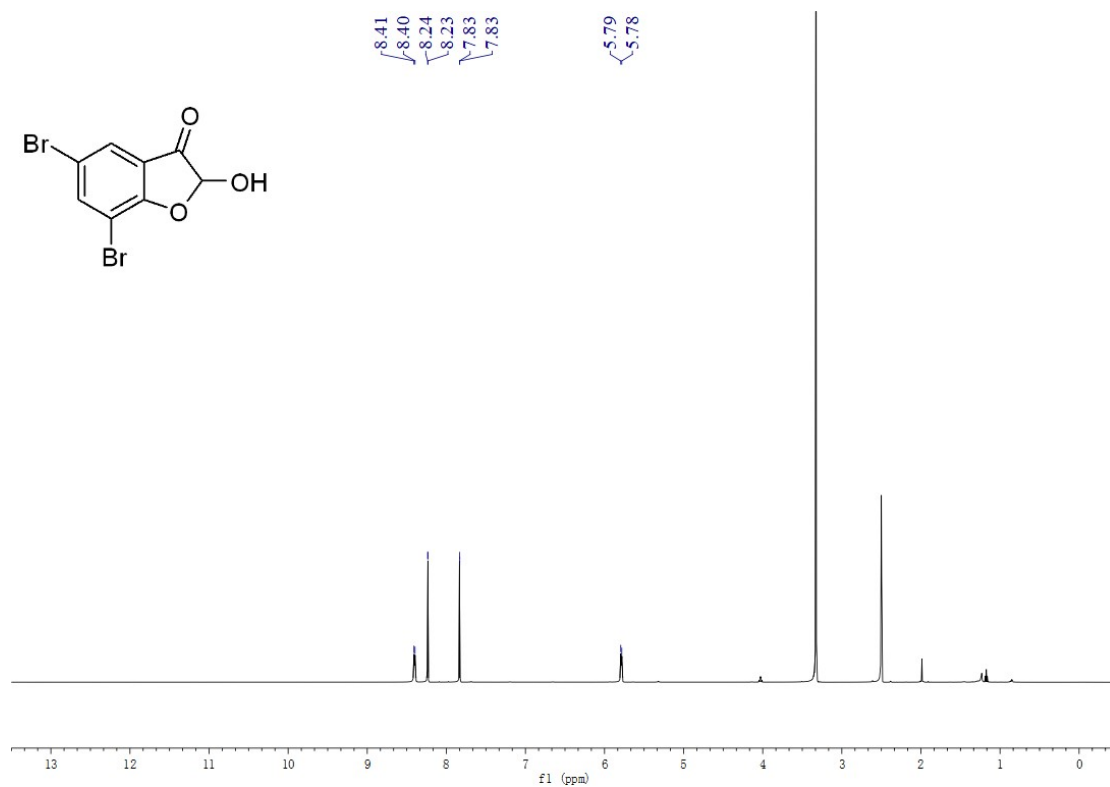
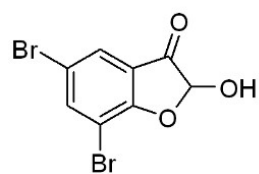
5.66
5.65



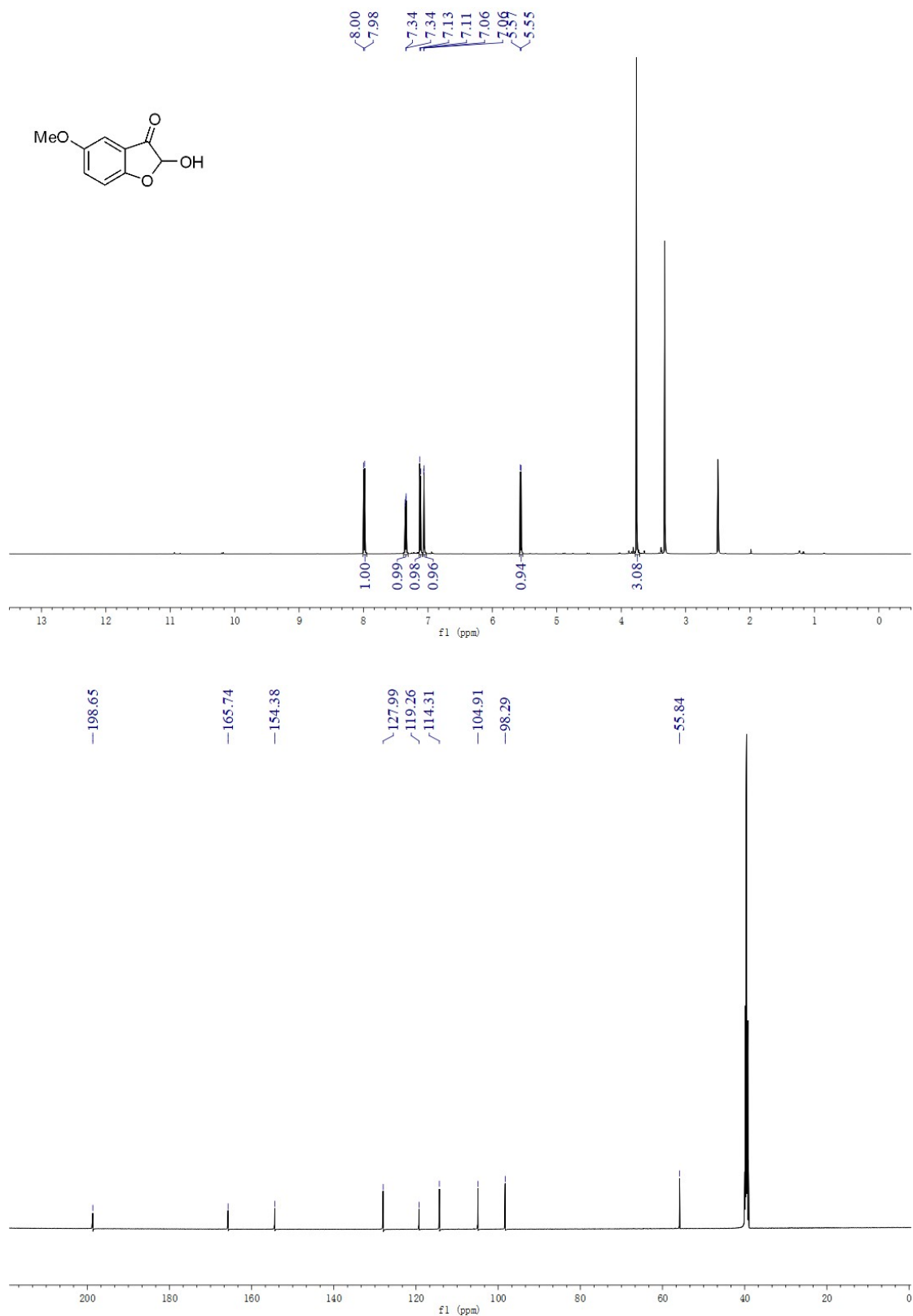
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113.42
98.53



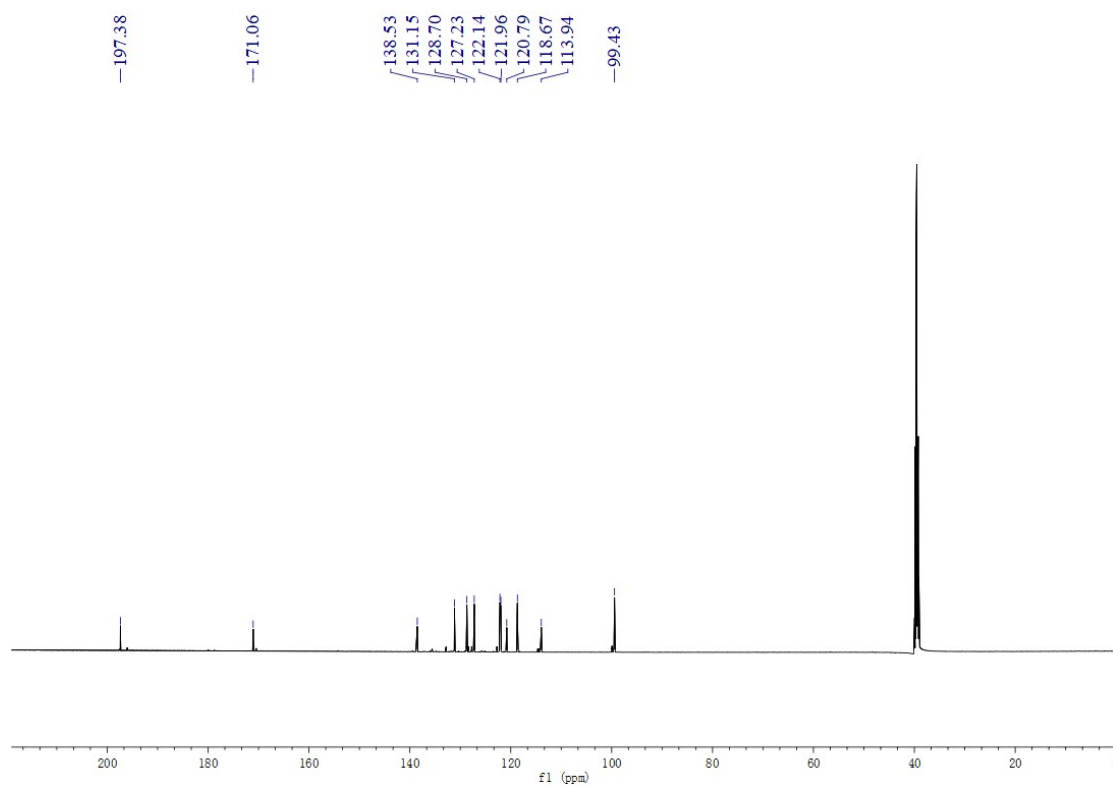
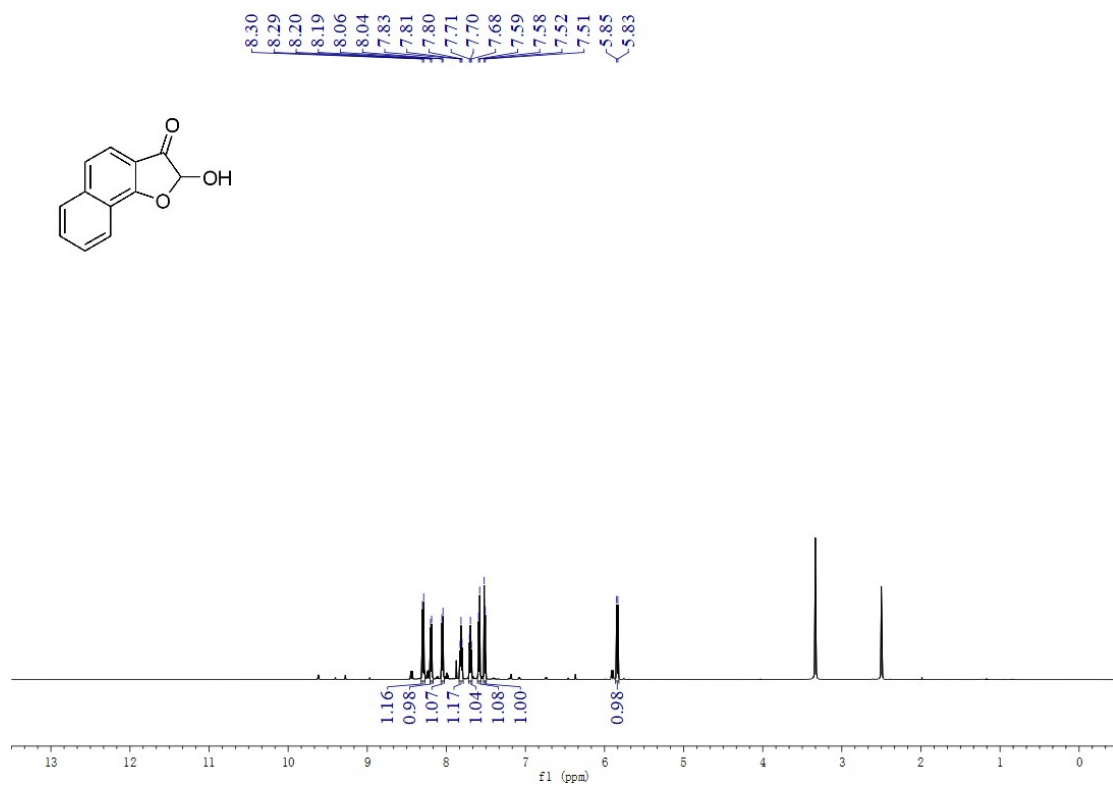
¹H and ¹³C NMR Spectra for **1e**



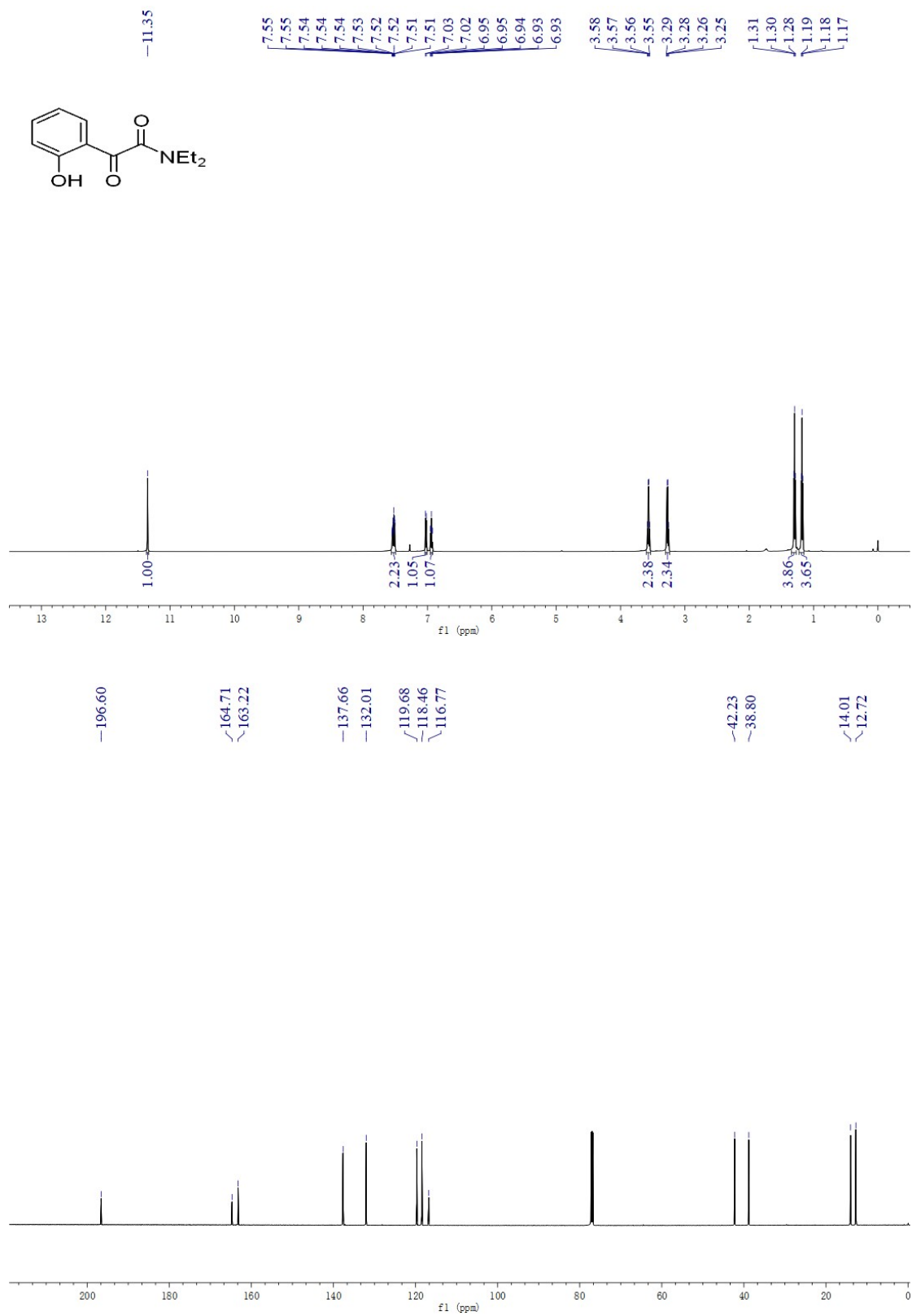
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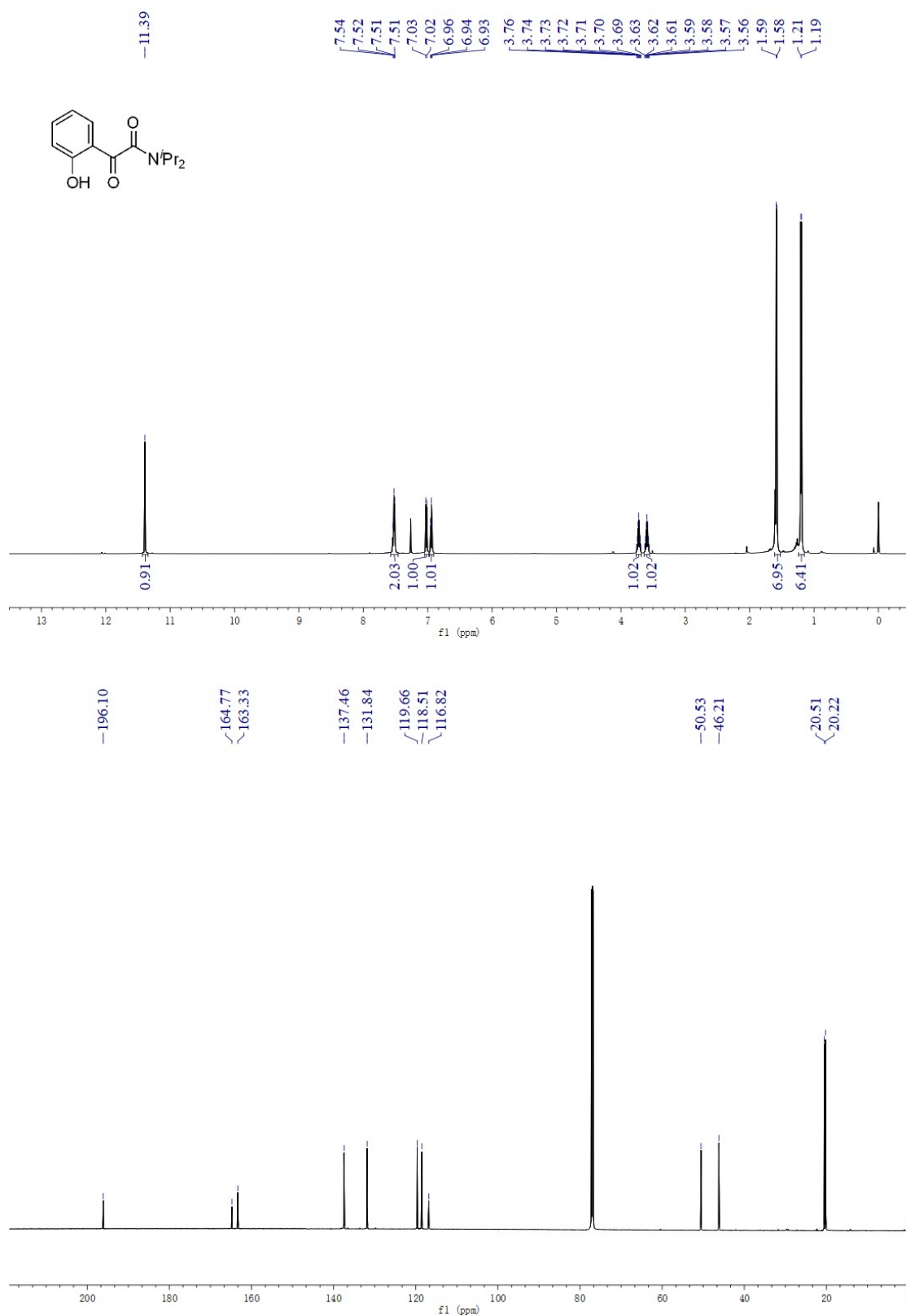
^1H and ^{13}C NMR Spectra for **1g**



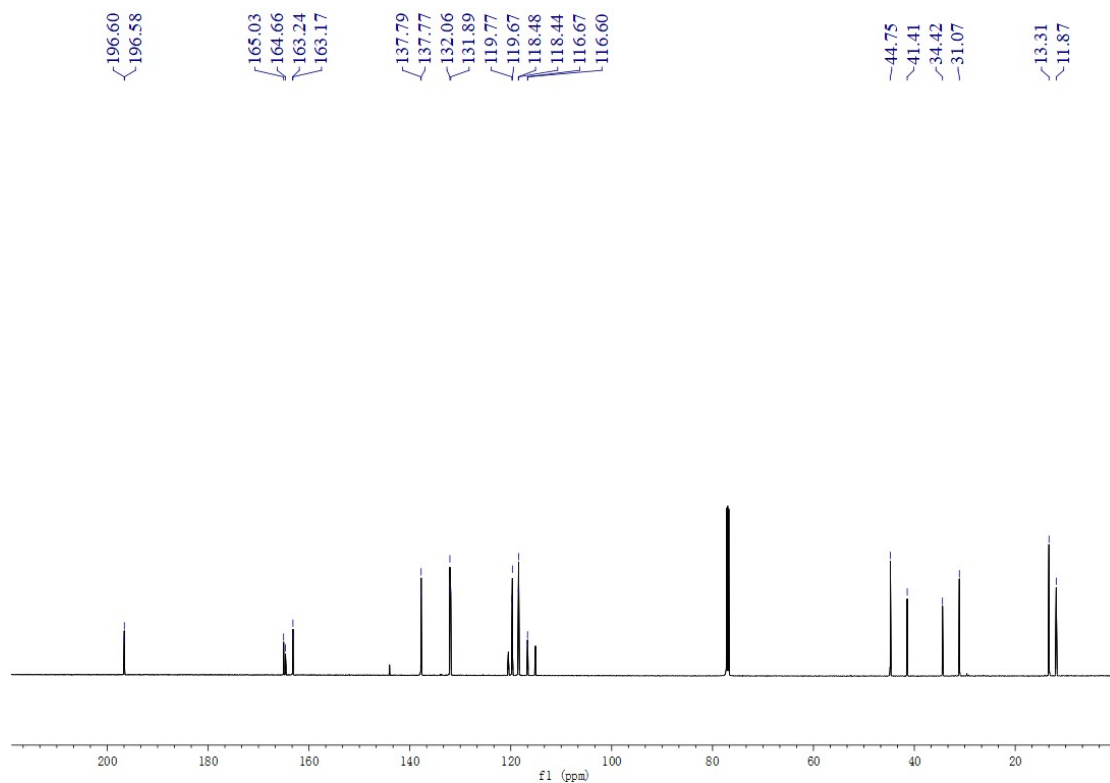
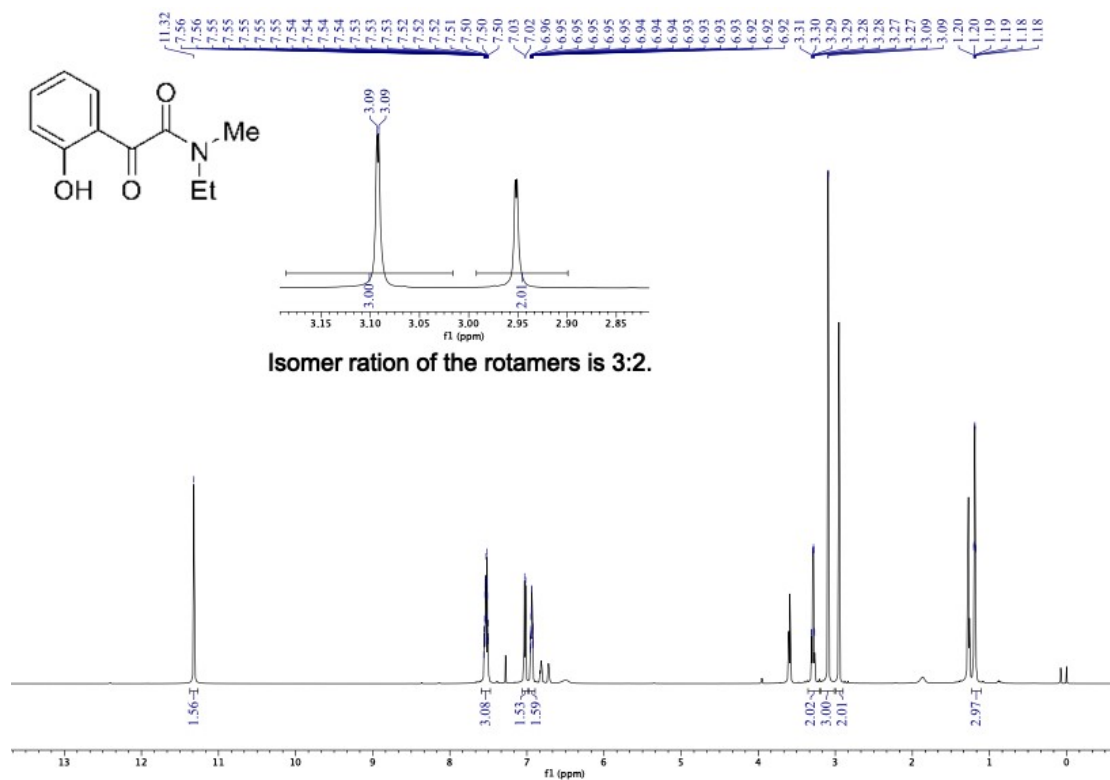
¹H and ¹³C NMR Spectra for 4a



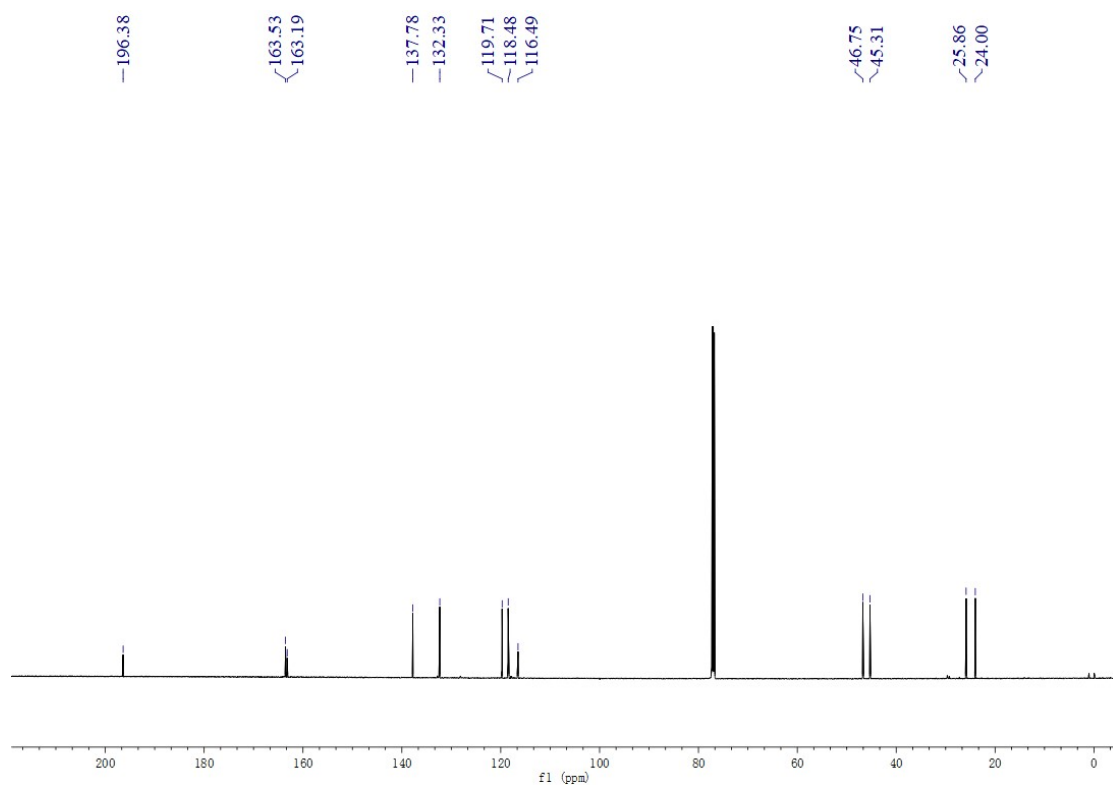
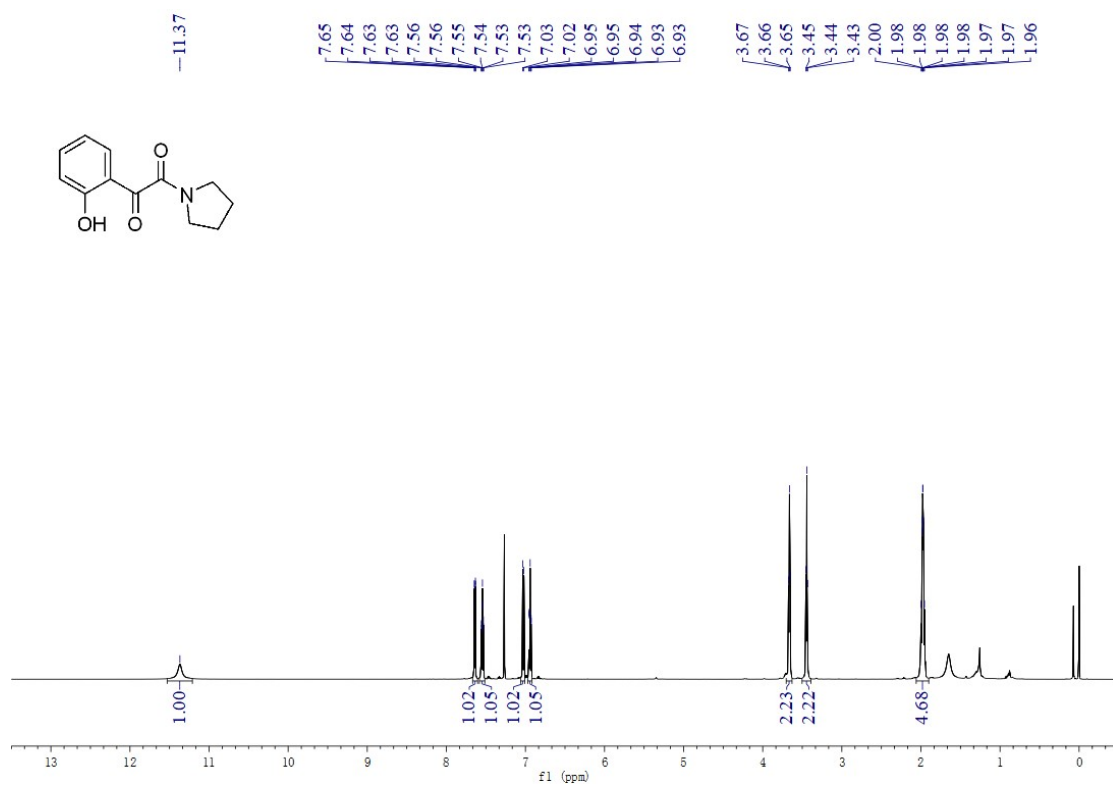
^1H and ^{13}C NMR Spectra for **4b**



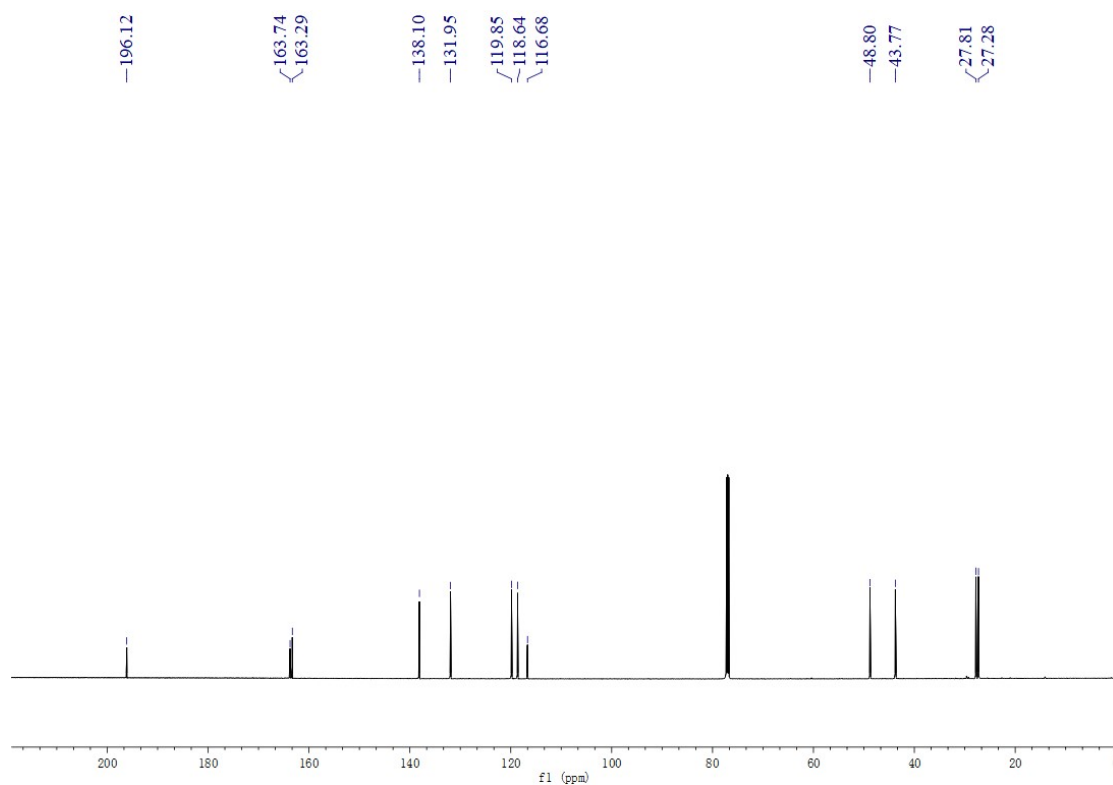
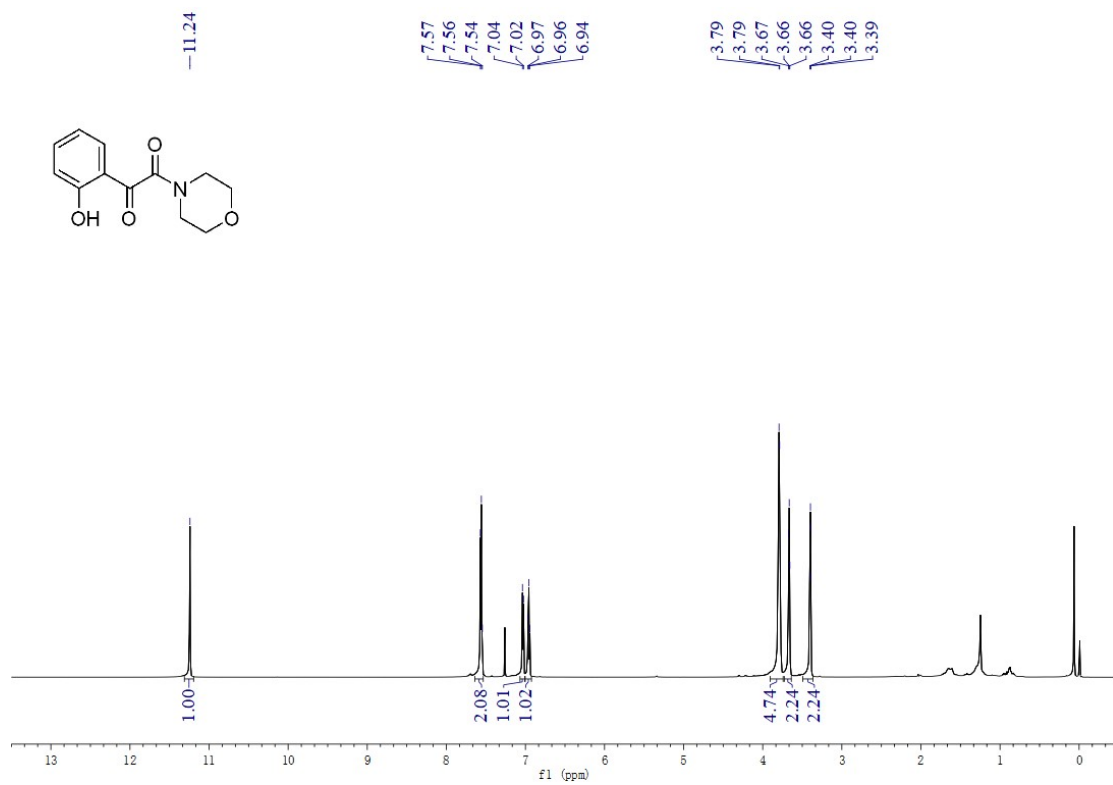
¹H and ¹³C NMR Spectra for 4c



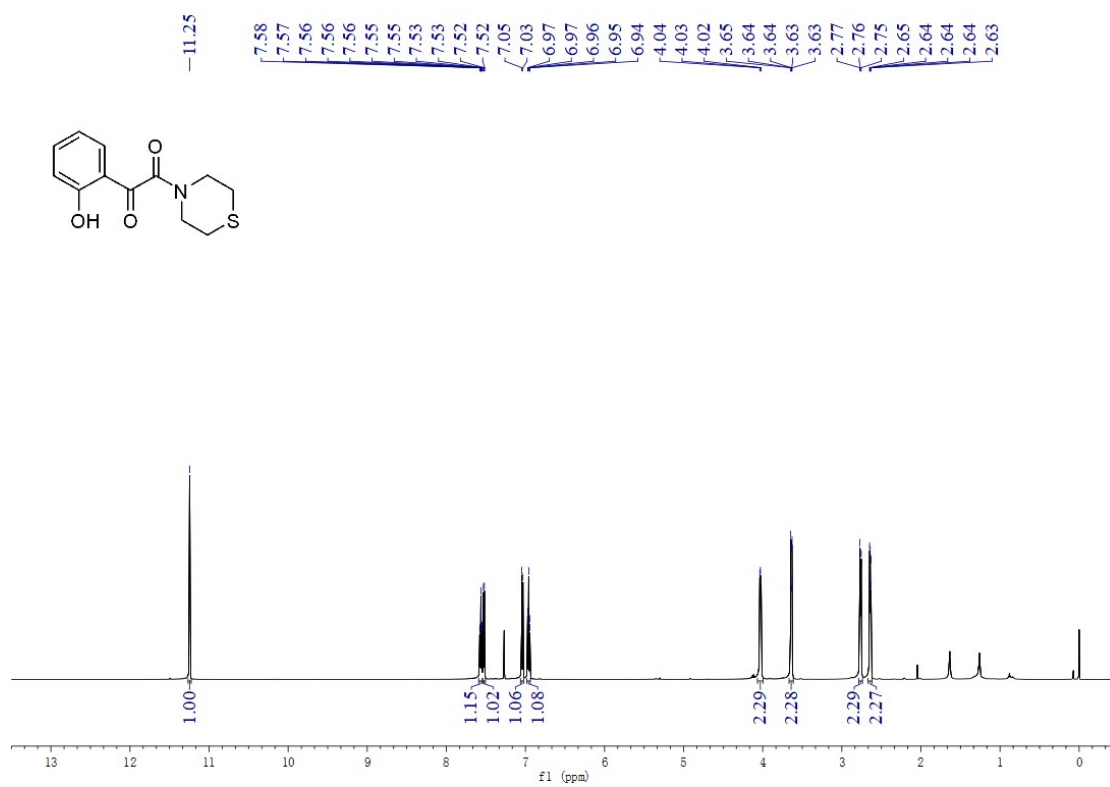
¹H and ¹³C NMR Spectra for 4d



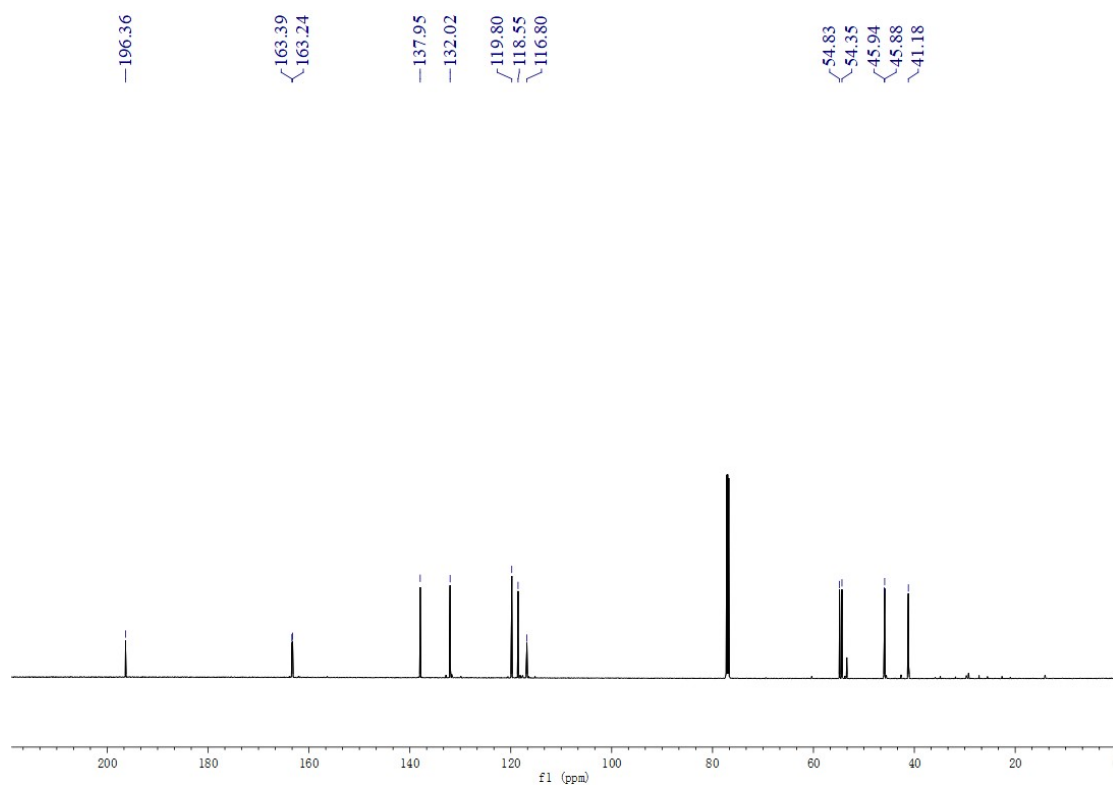
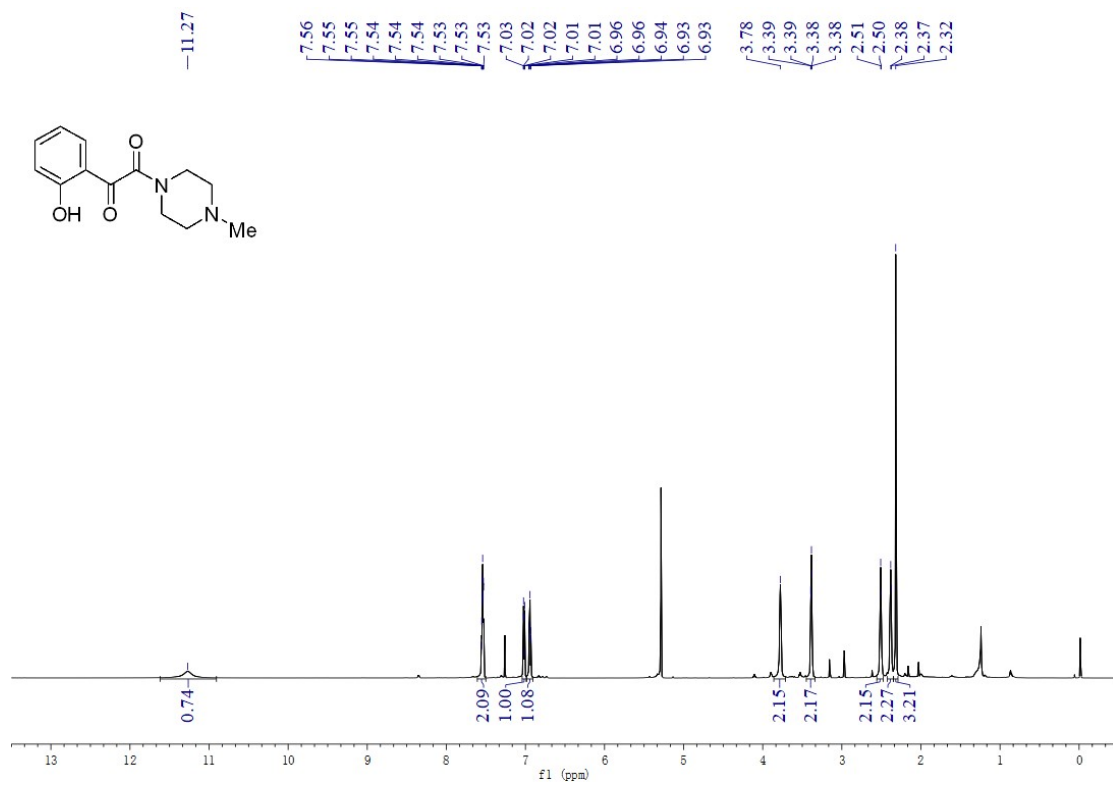
¹H and ¹³C NMR Spectra for 4e



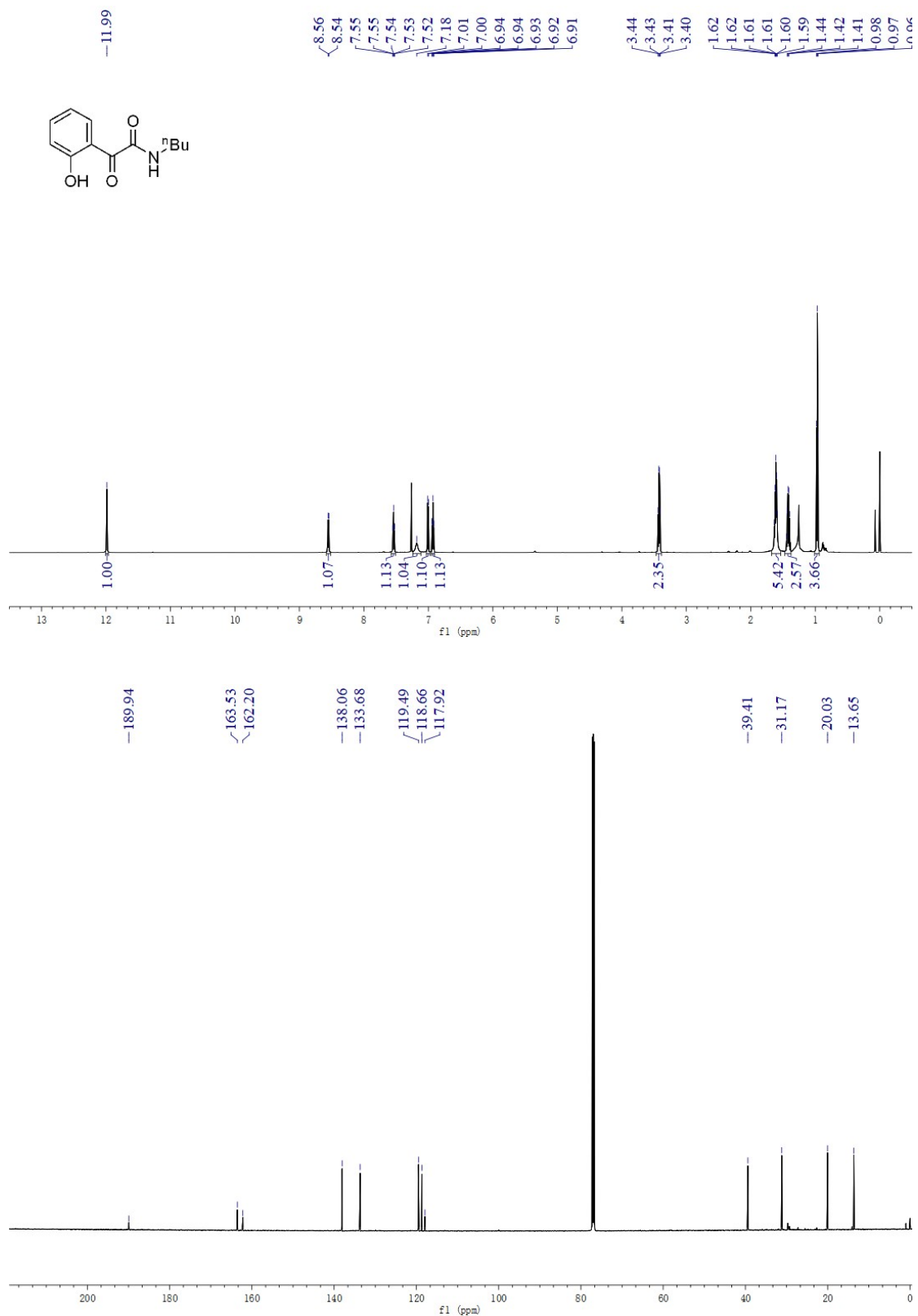
^1H and ^{13}C NMR Spectra for **4f**



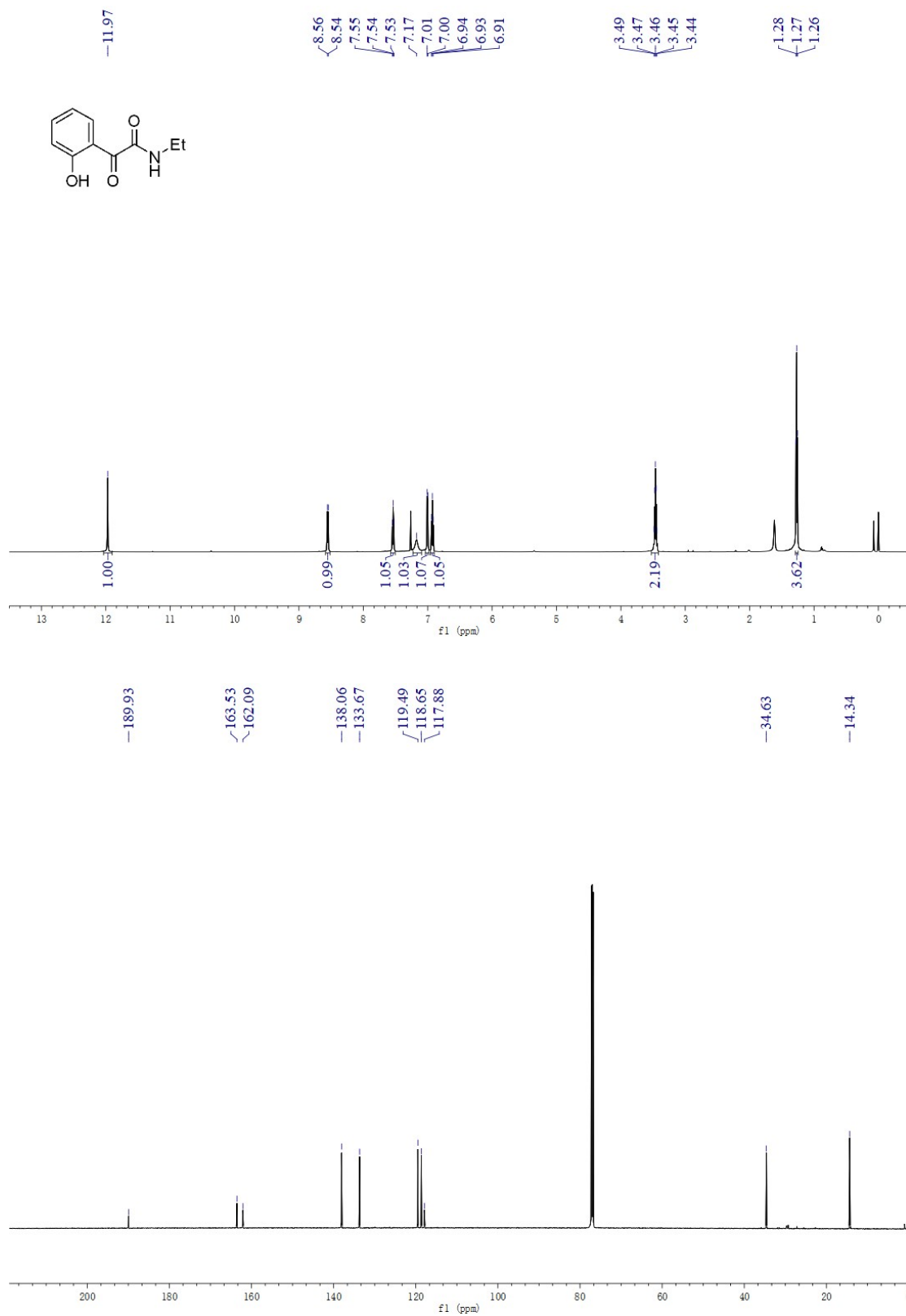
¹H and ¹³C NMR Spectra for 4g



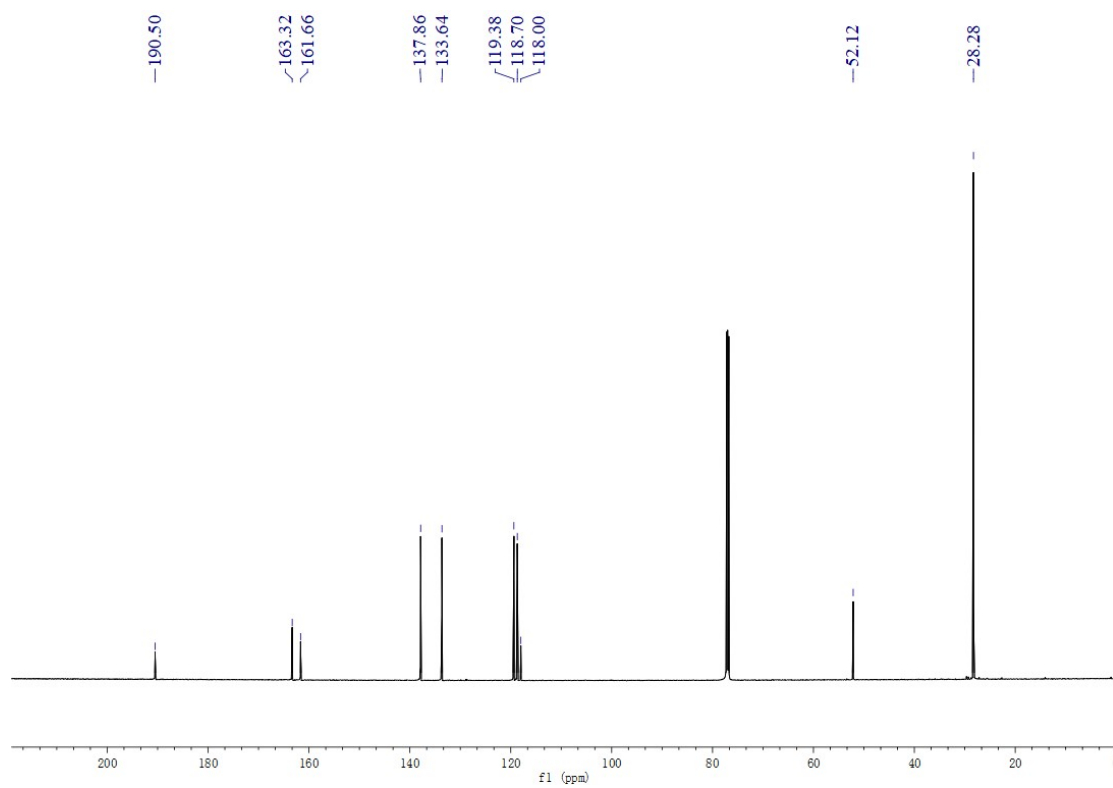
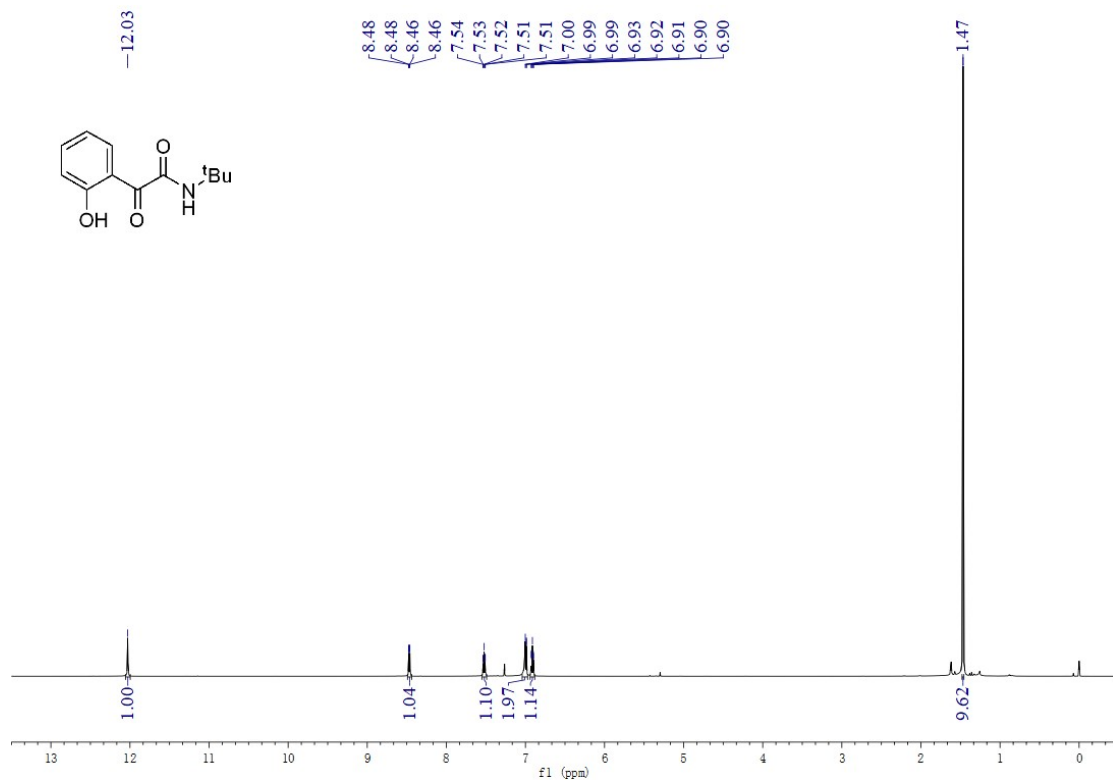
¹H and ¹³C NMR Spectra for 4h



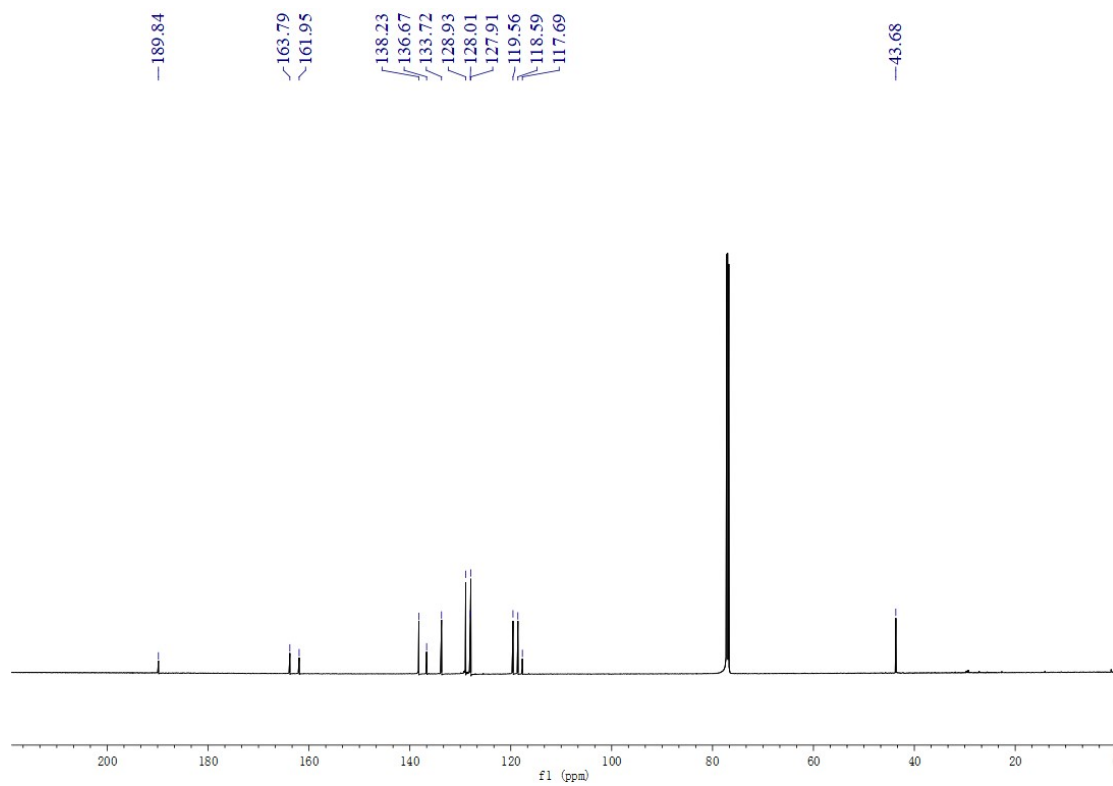
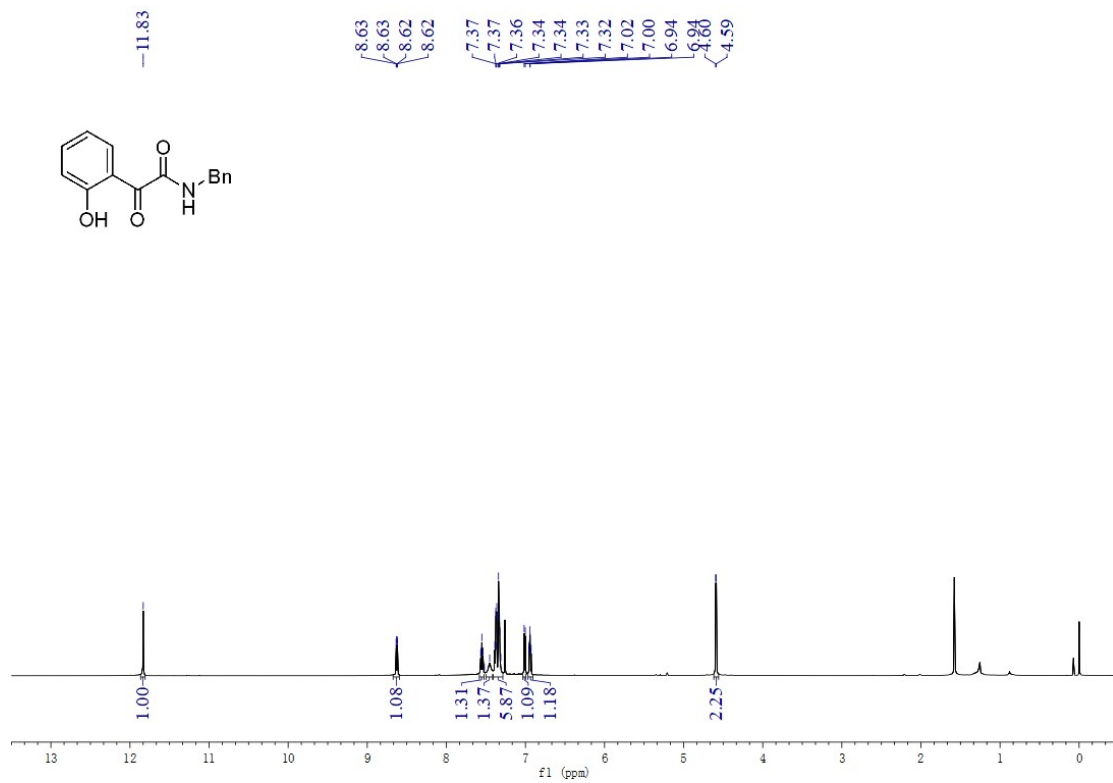
¹H and ¹³C NMR Spectra for 4i



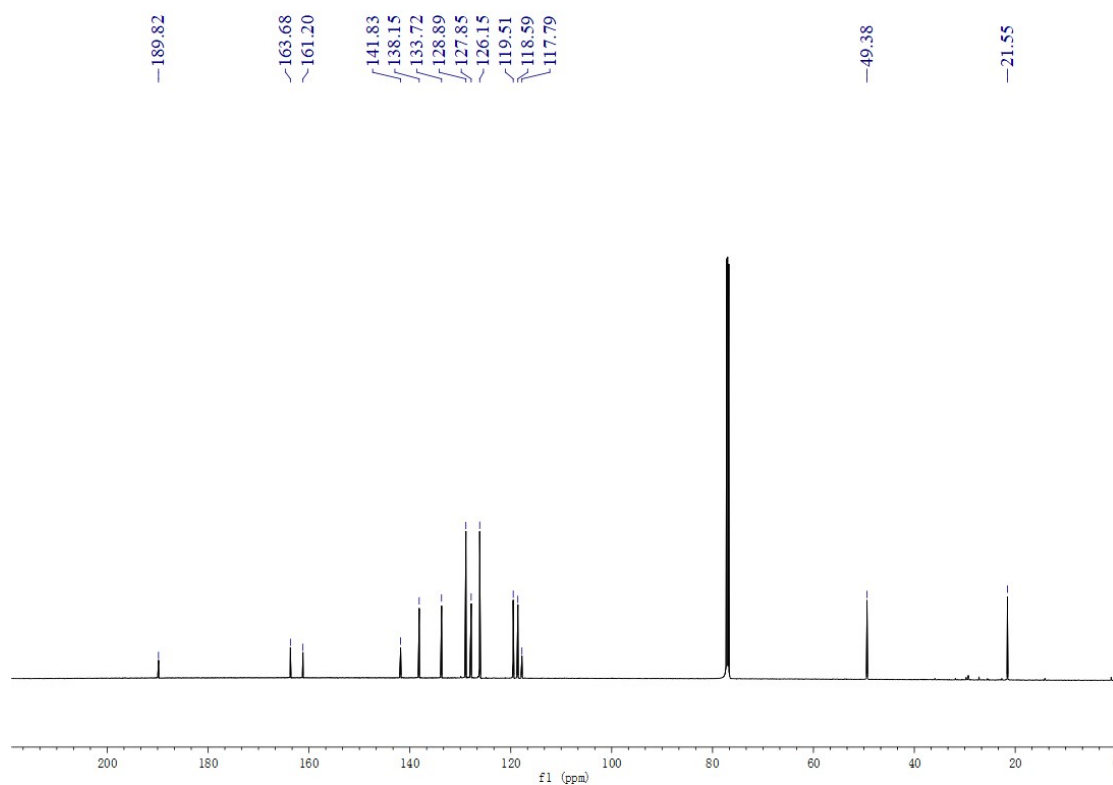
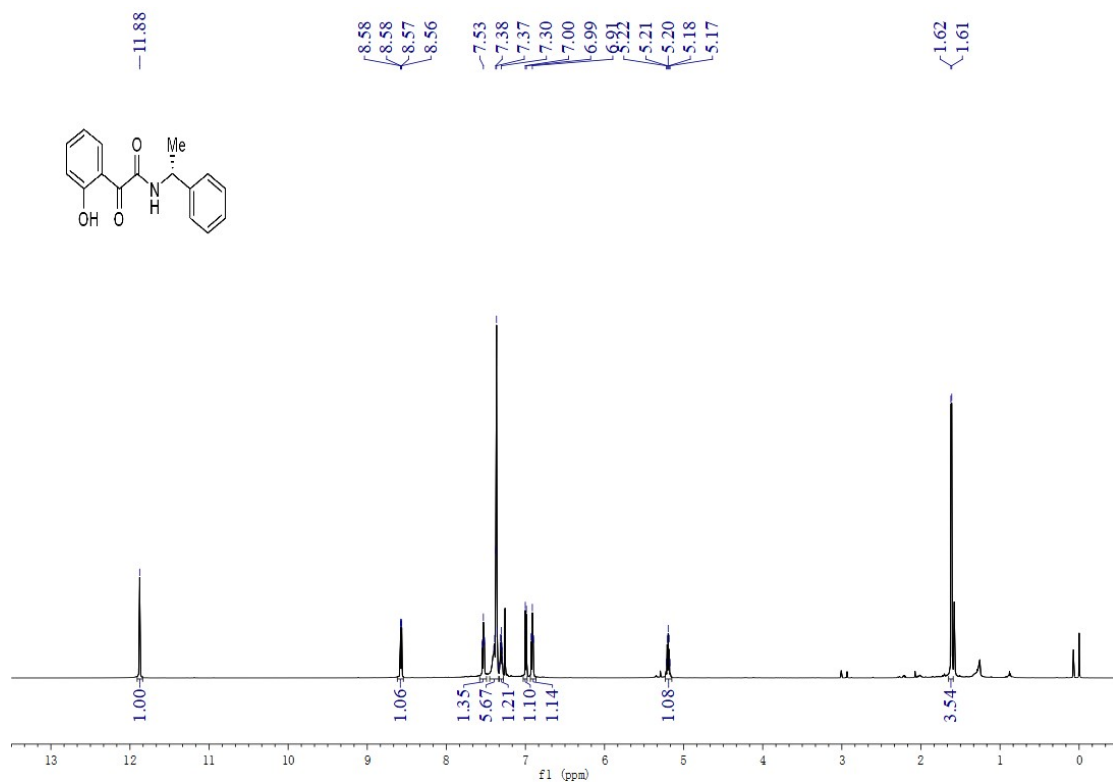
¹H and ¹³C NMR Spectra for 4j



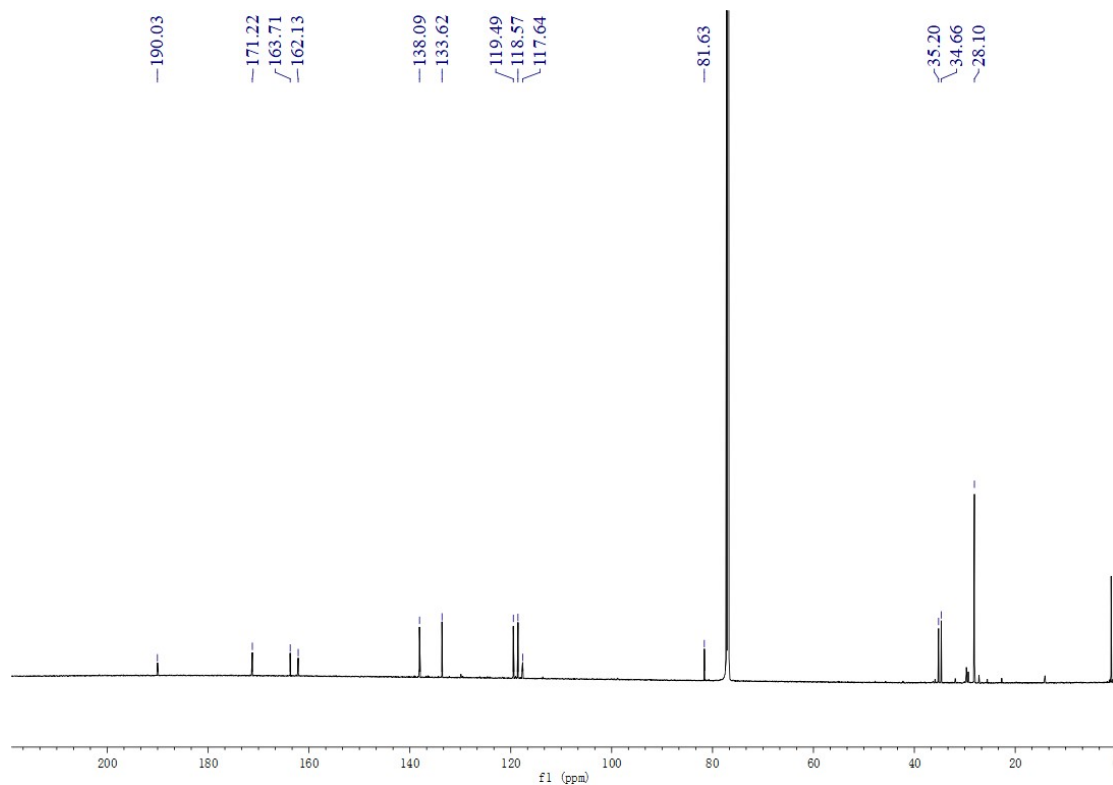
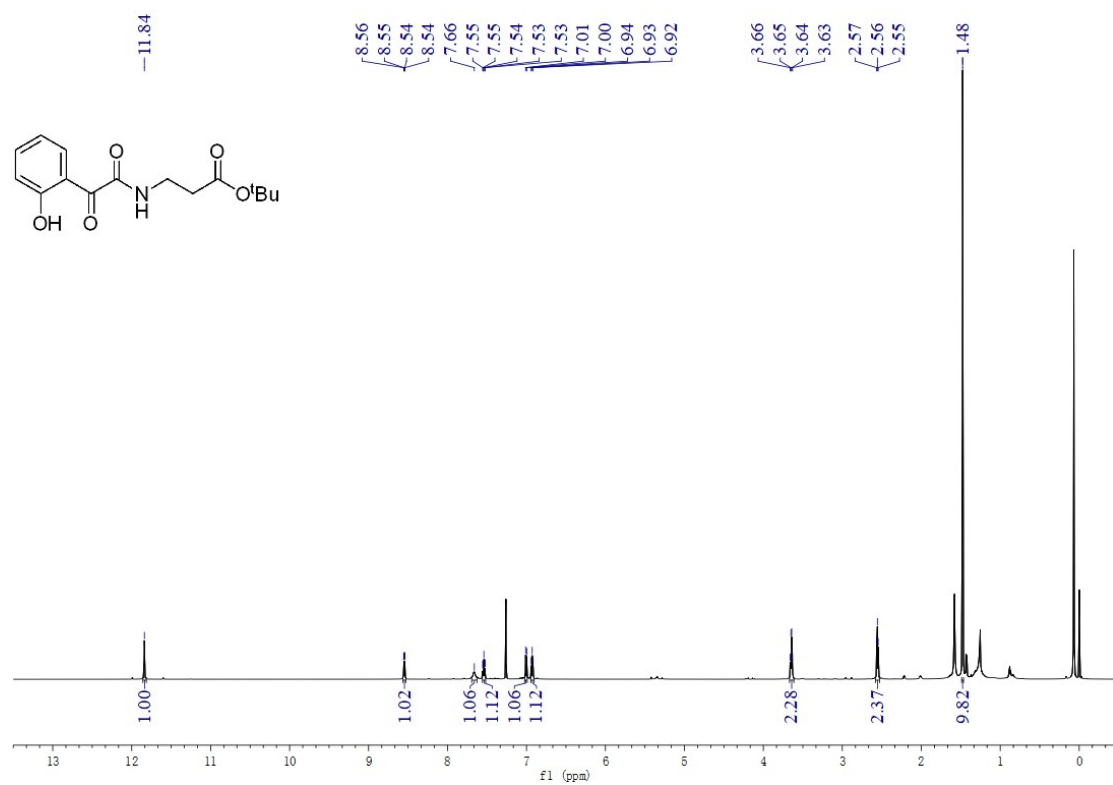
¹H and ¹³C NMR Spectra for 4k



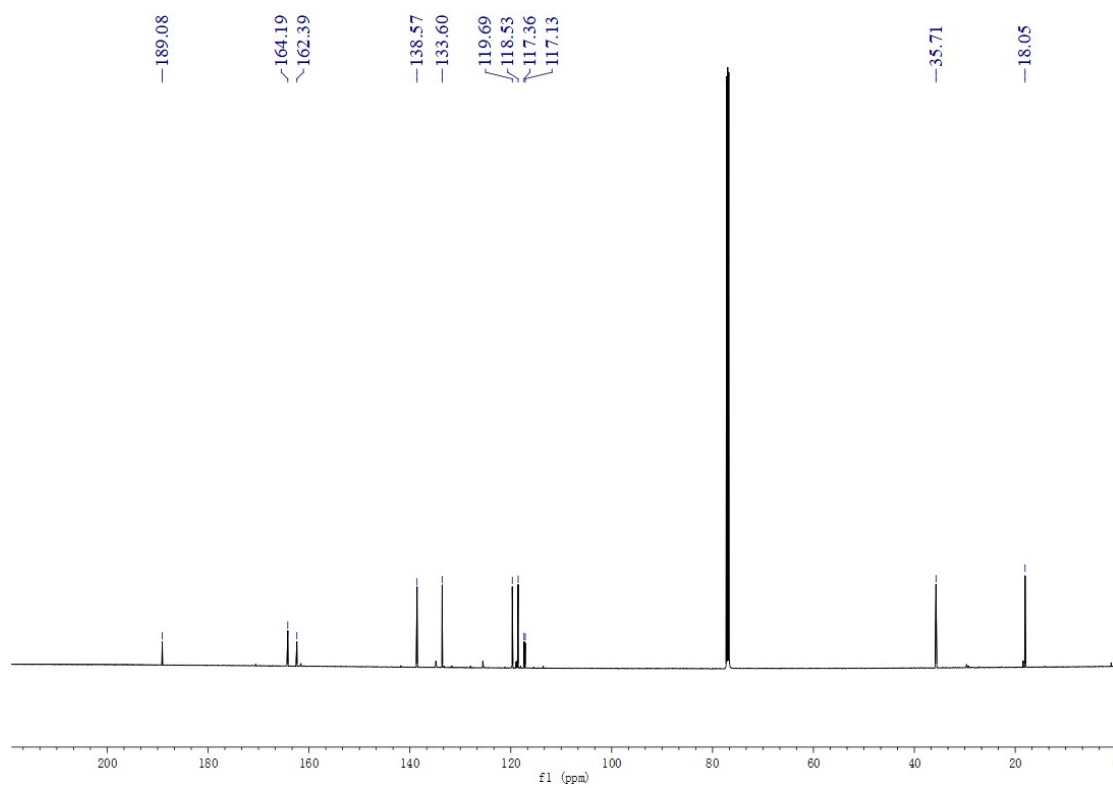
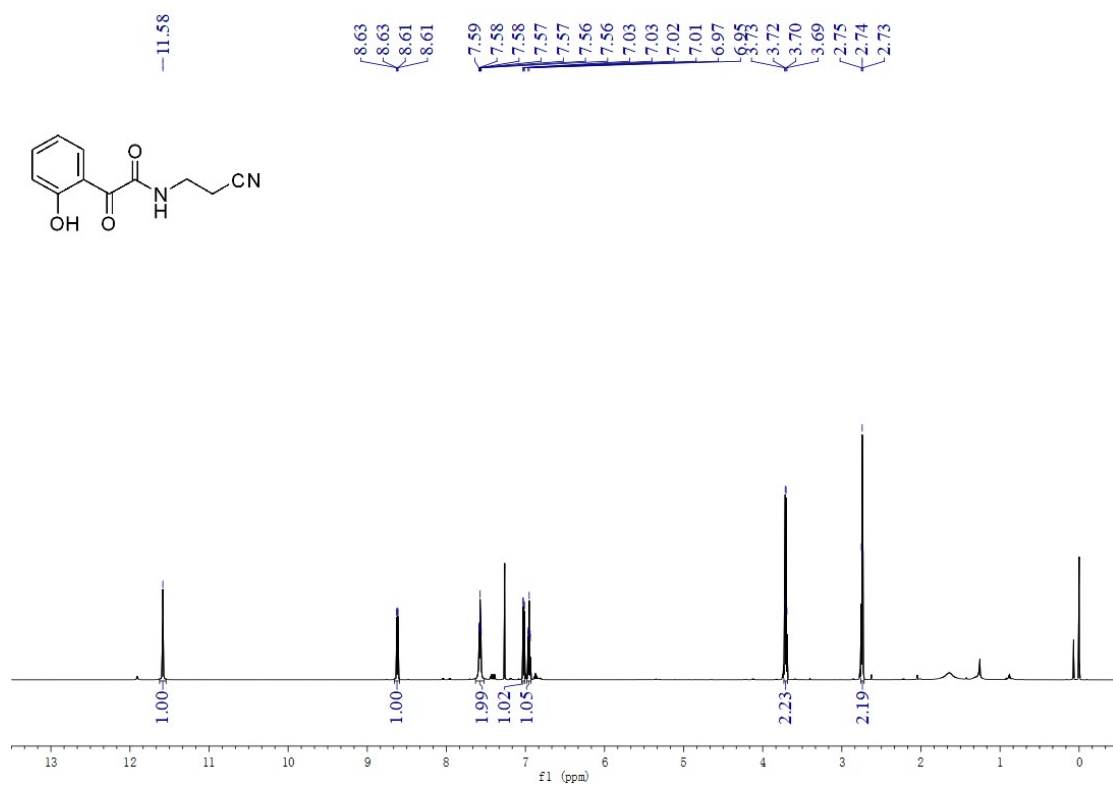
¹H and ¹³C NMR Spectra for **41**



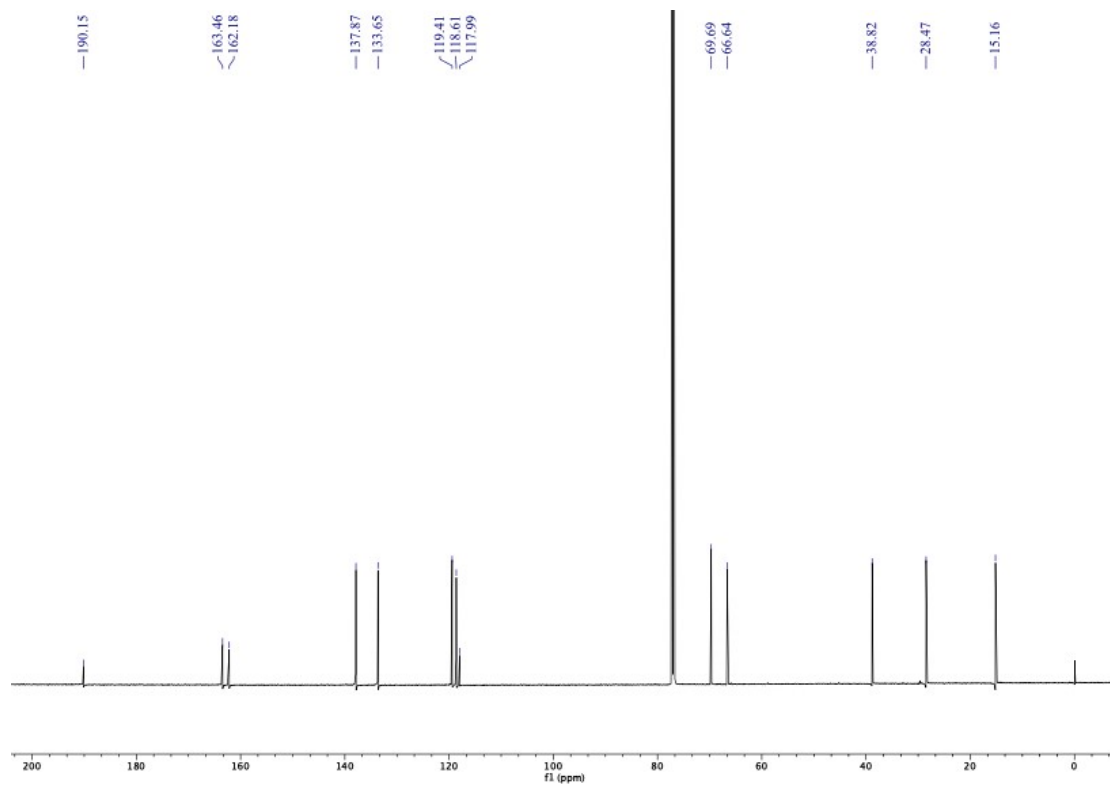
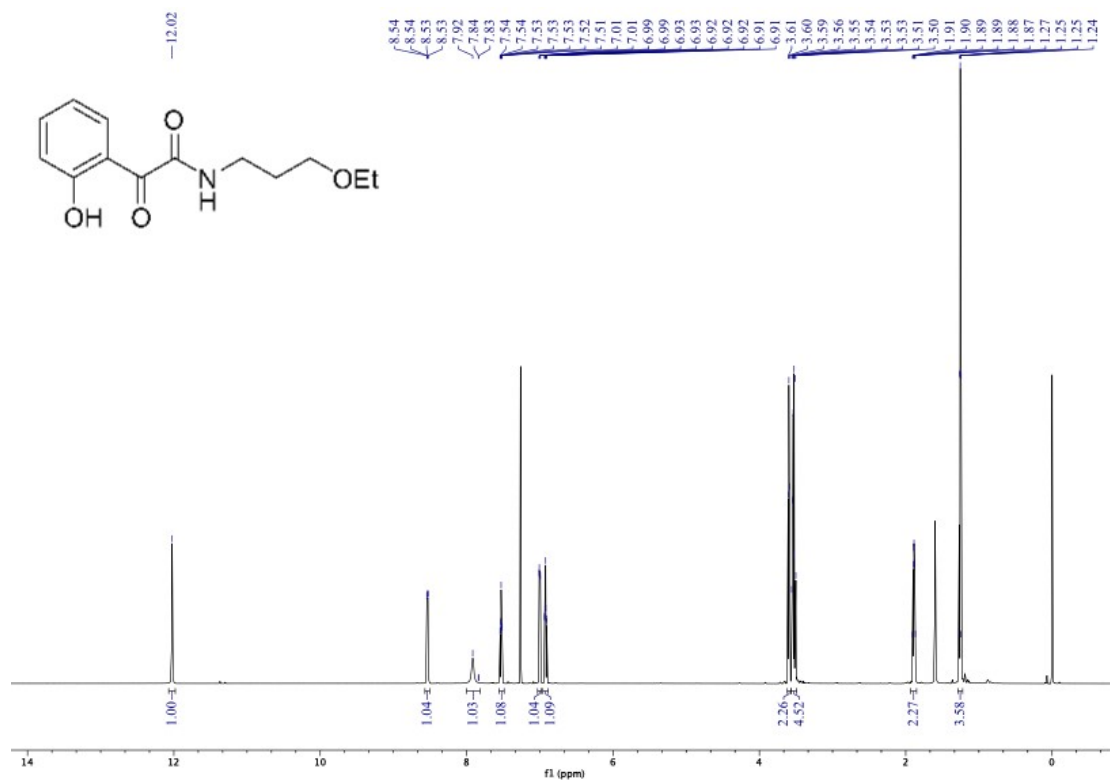
¹H and ¹³C NMR Spectra for 4m



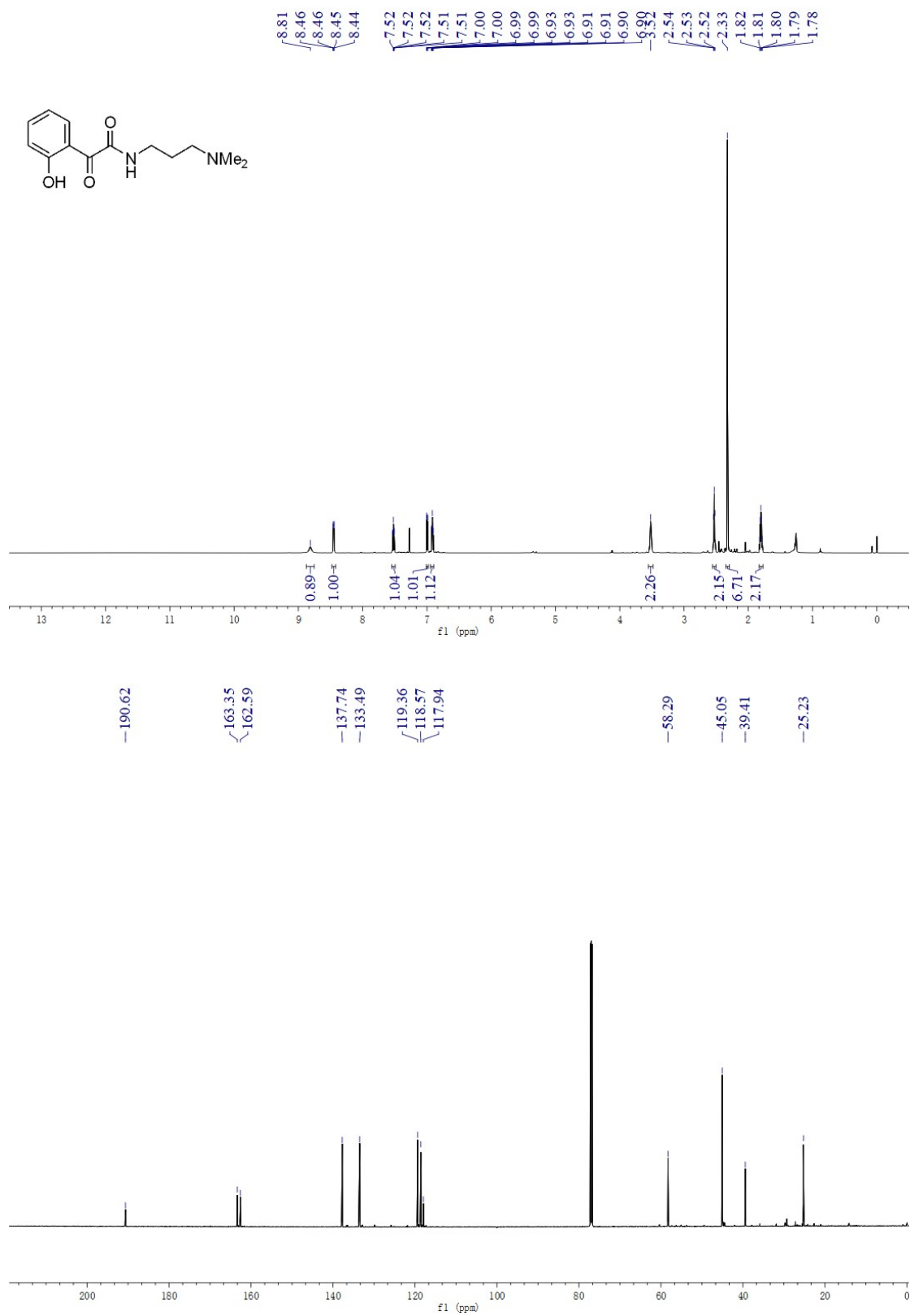
¹H and ¹³C NMR Spectra for 4n



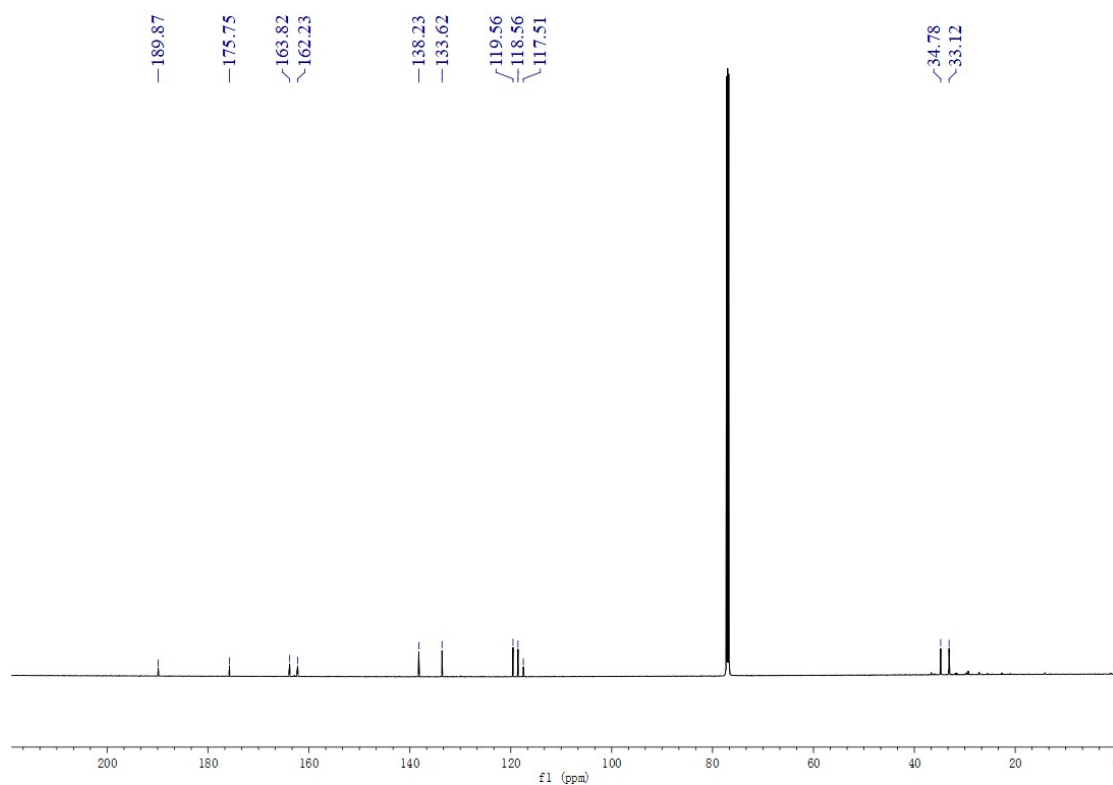
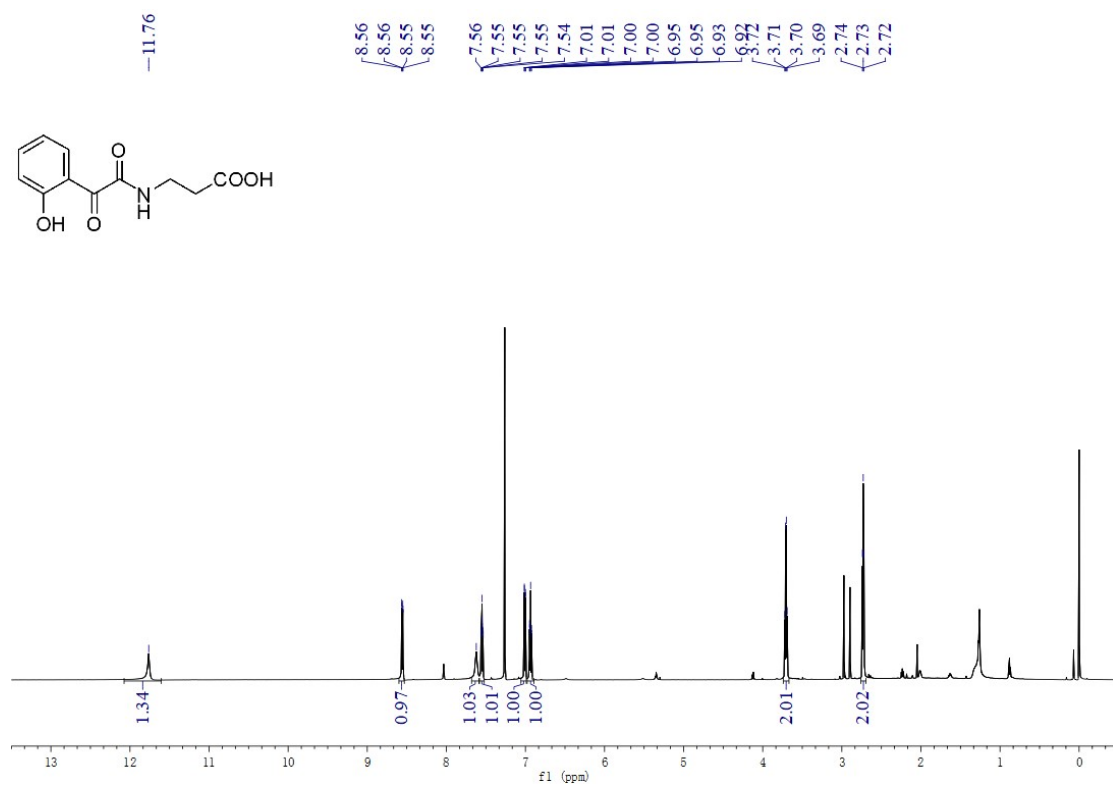
¹H and ¹³C NMR Spectra for **40**



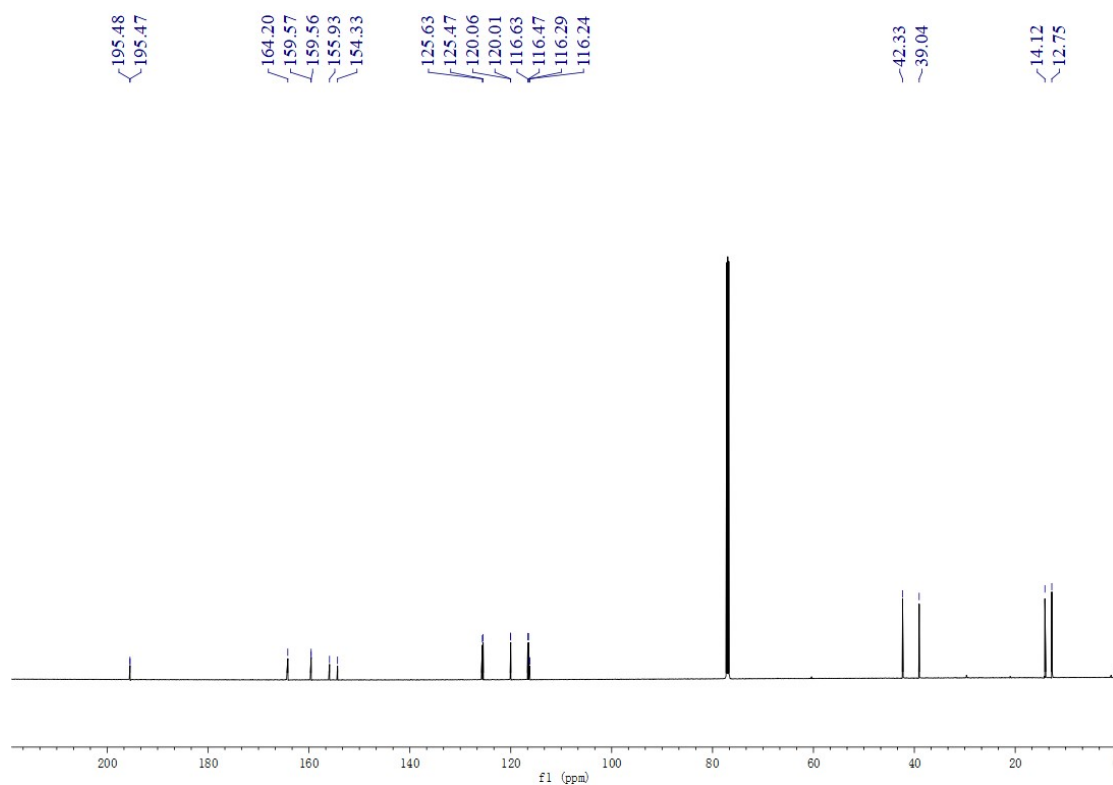
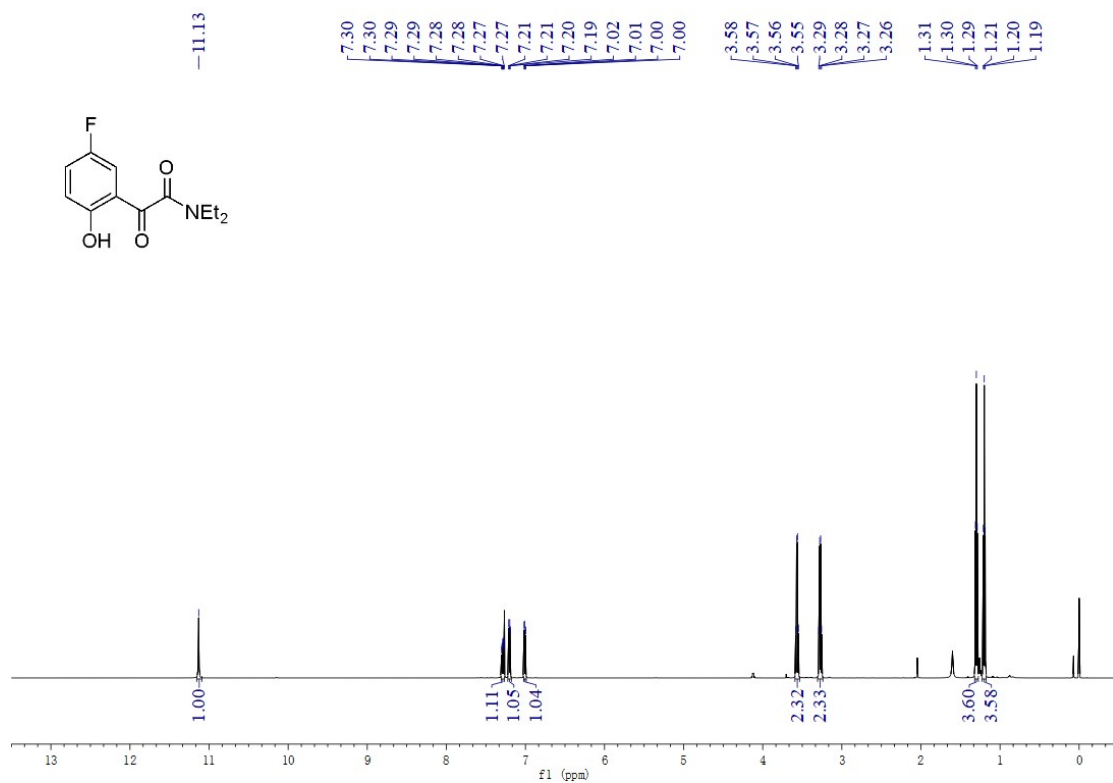
¹H and ¹³C NMR Spectra for 4p

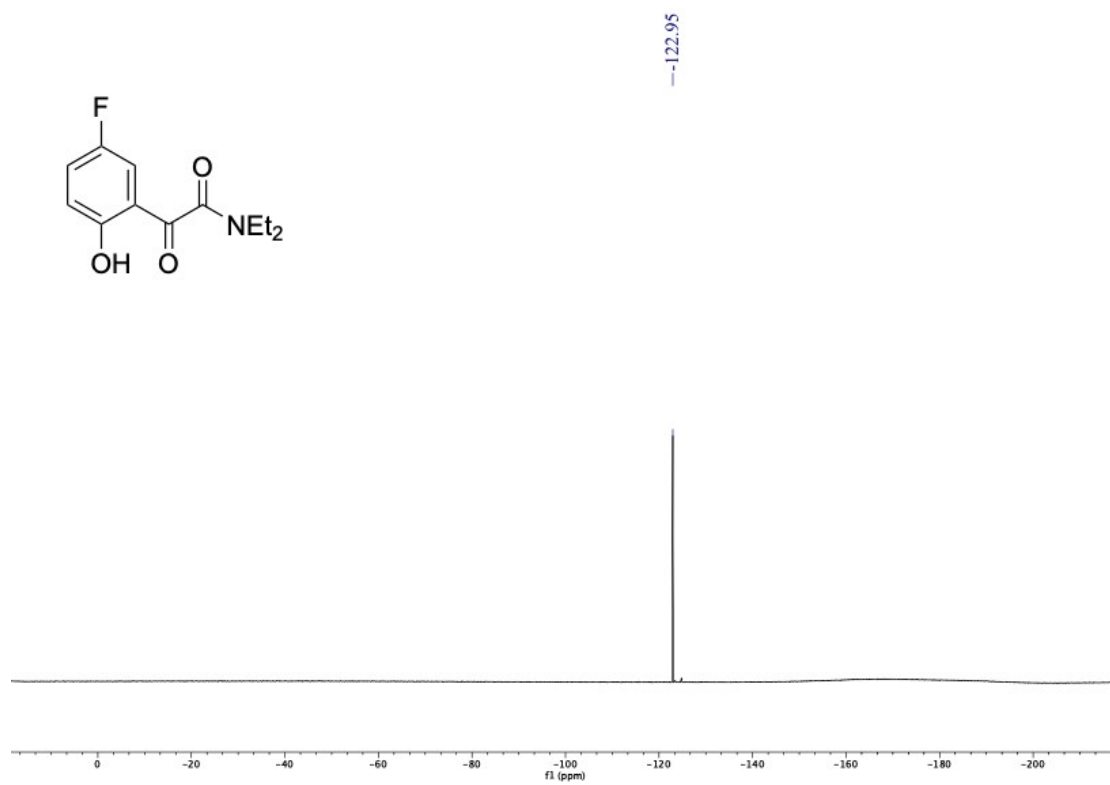


¹H and ¹³C NMR Spectra for 4q

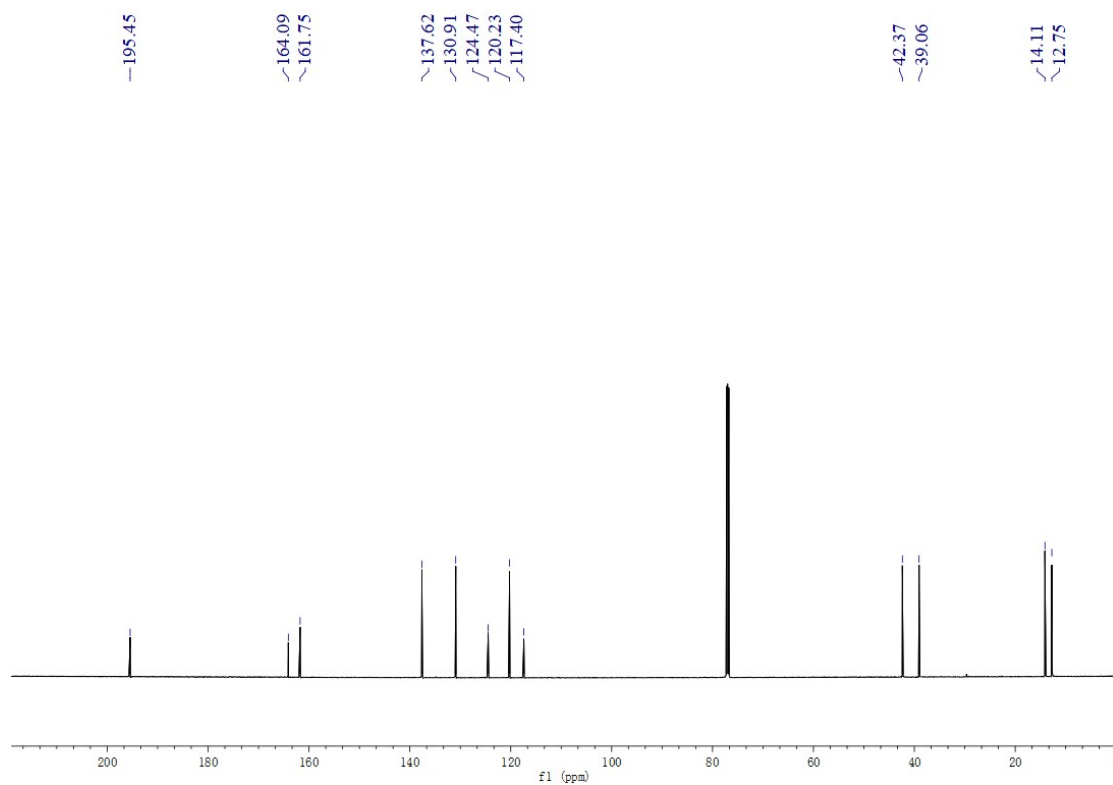
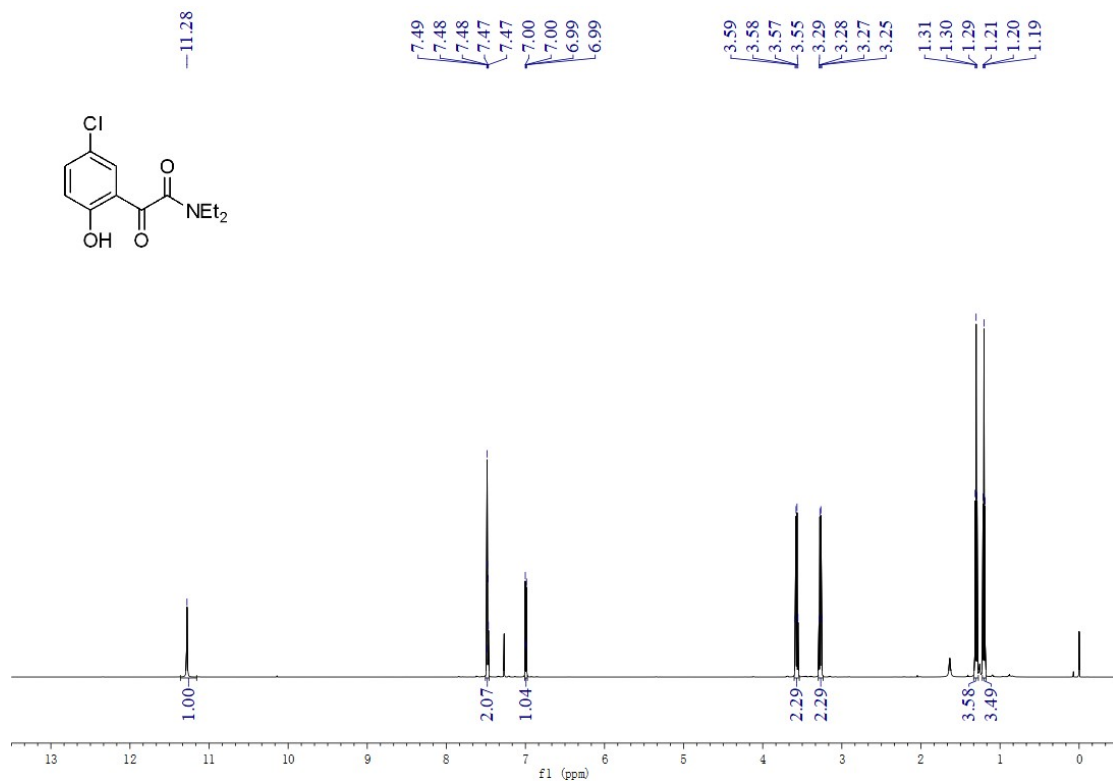


¹H, ¹³C and ¹⁹F NMR Spectra for **4ab**

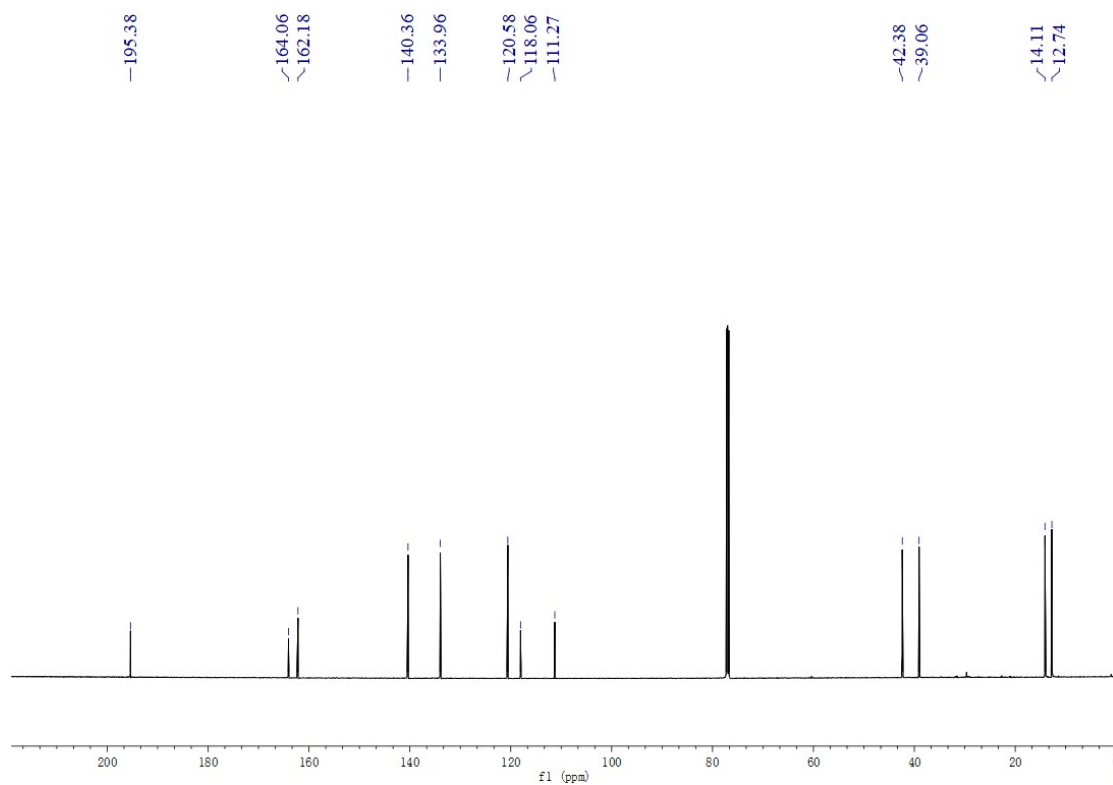
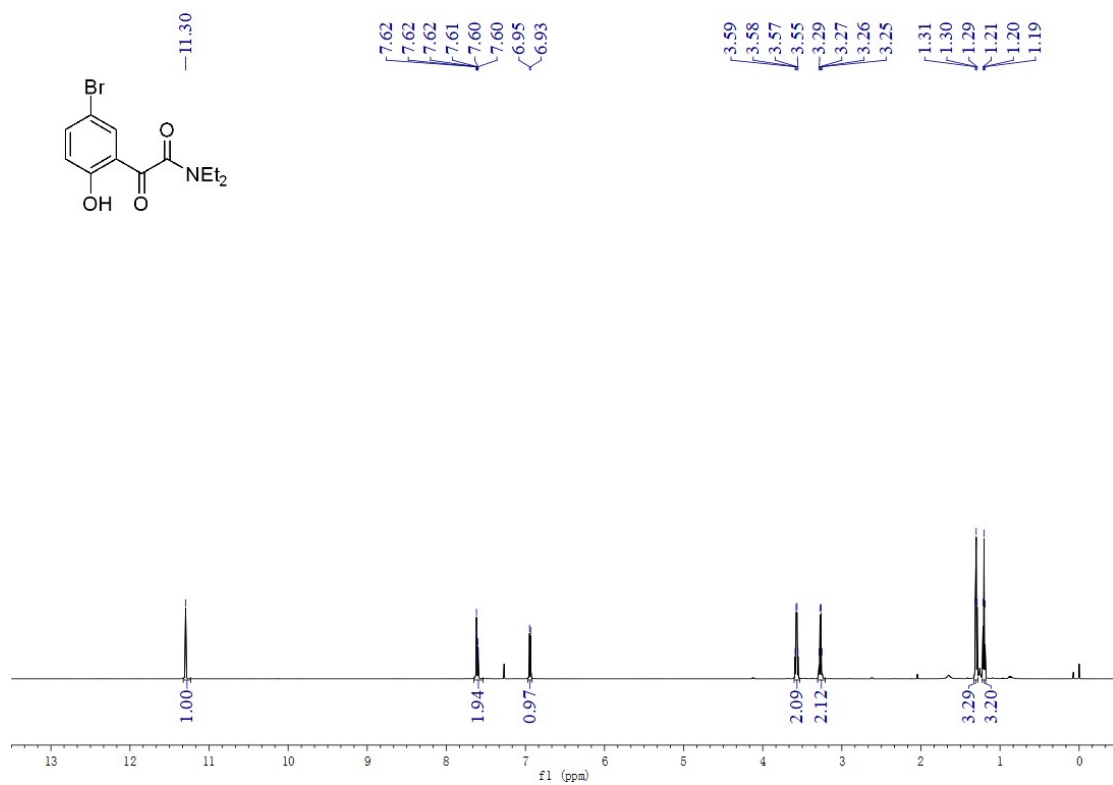




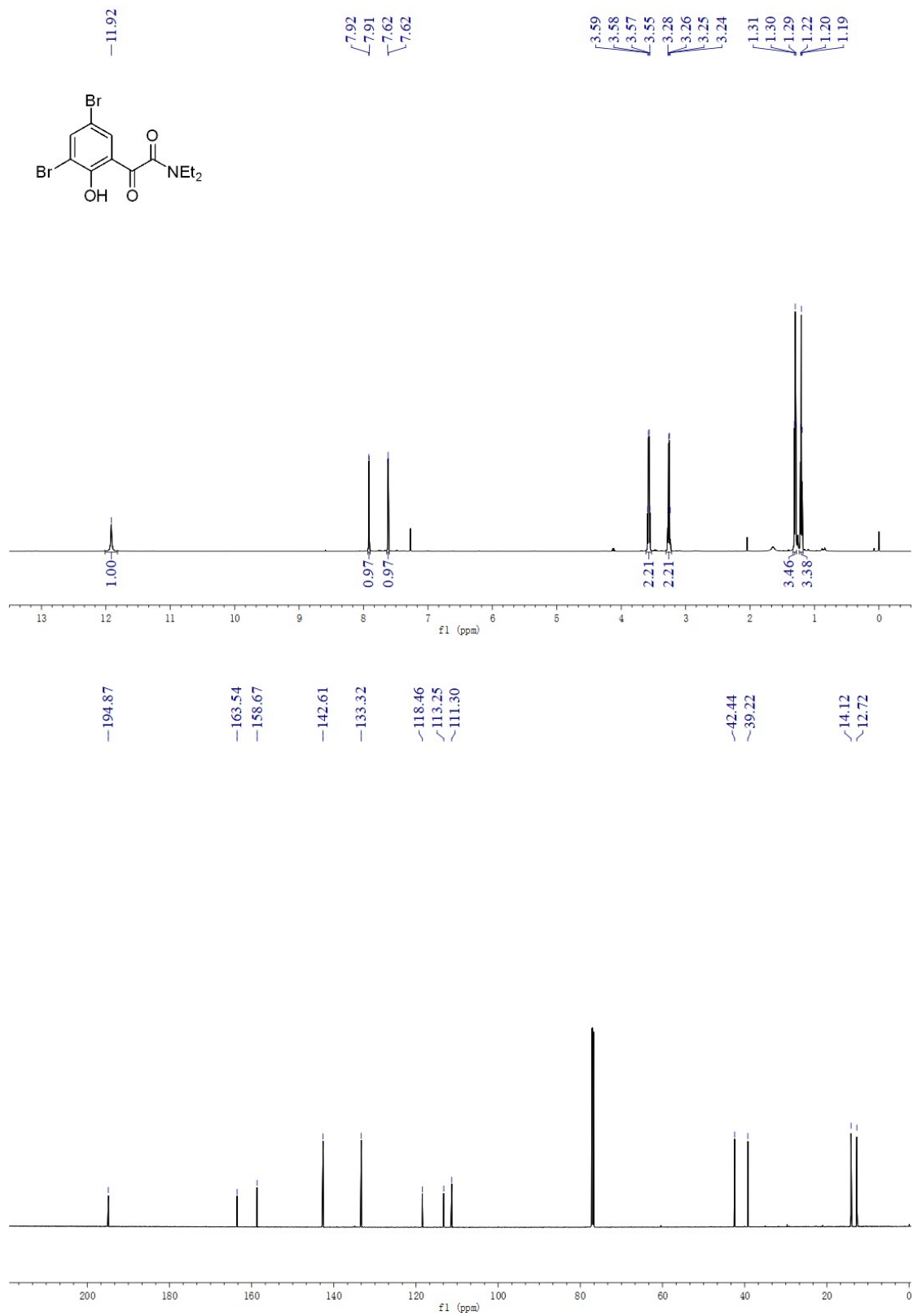
^1H and ^{13}C NMR Spectra for **4ac**



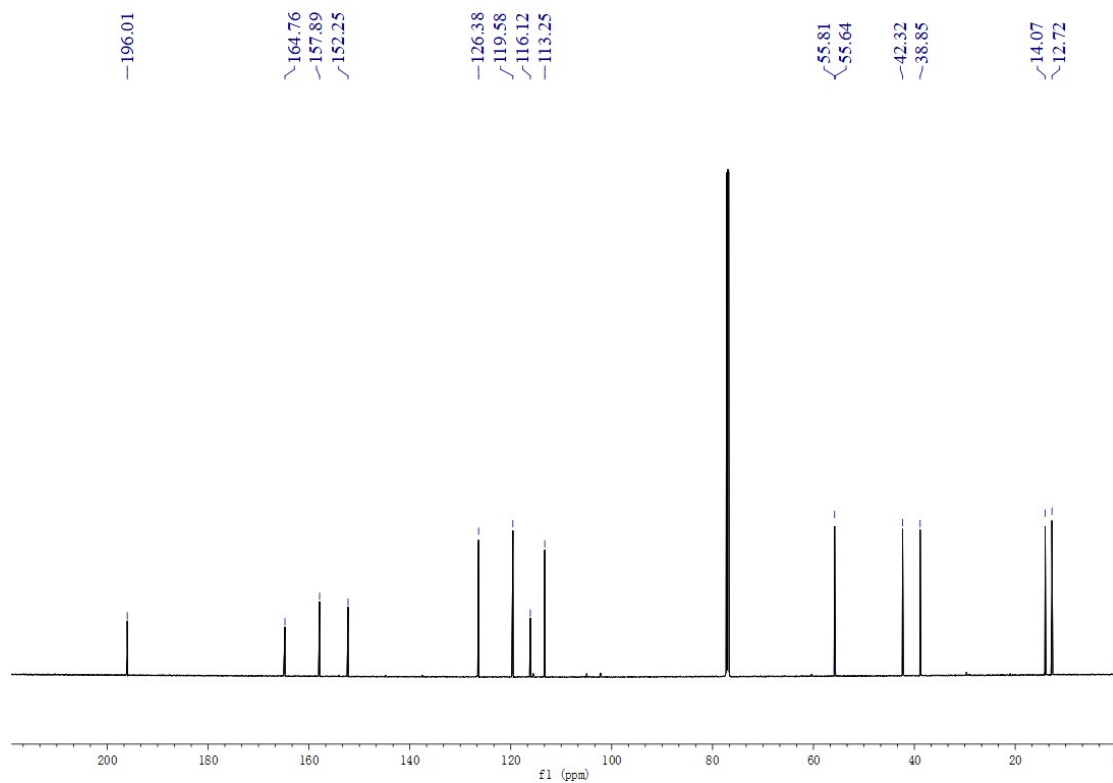
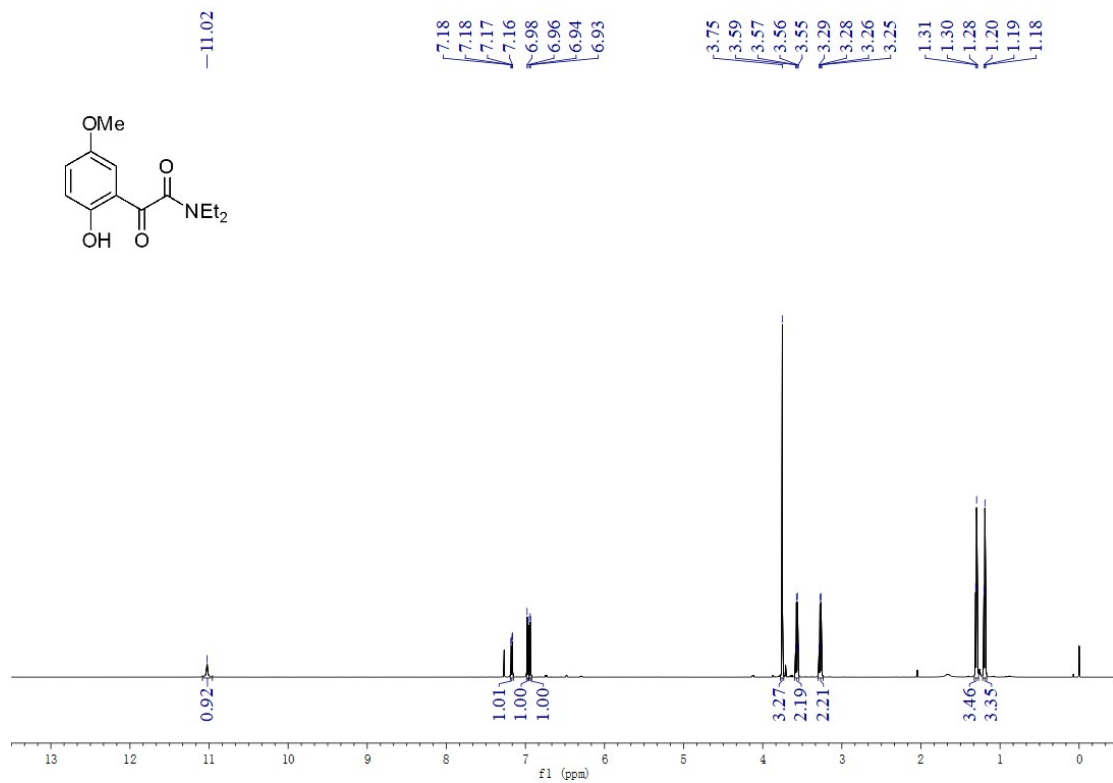
^1H and ^{13}C NMR Spectra for **4ad**



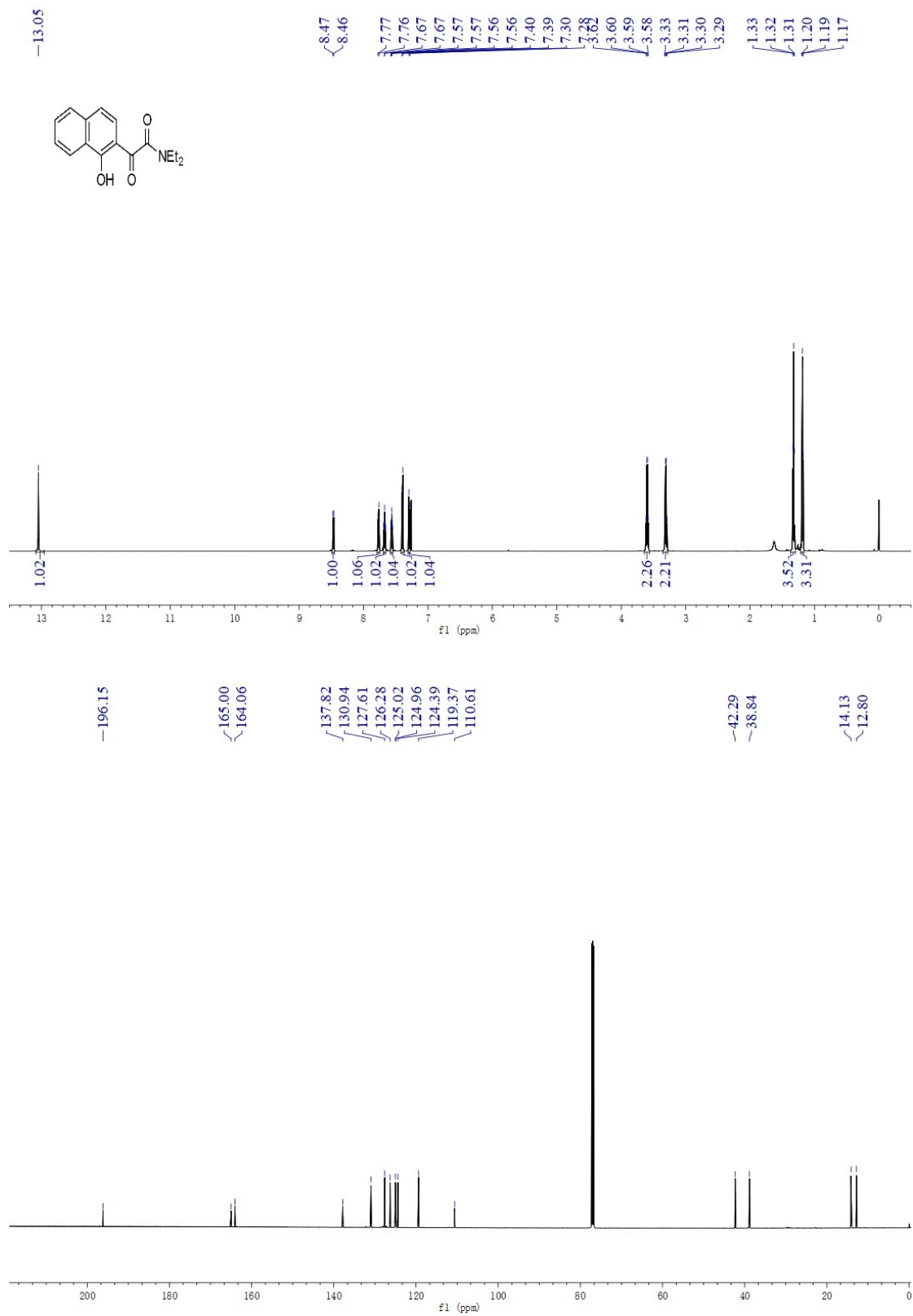
^1H and ^{13}C NMR Spectra for 4ae



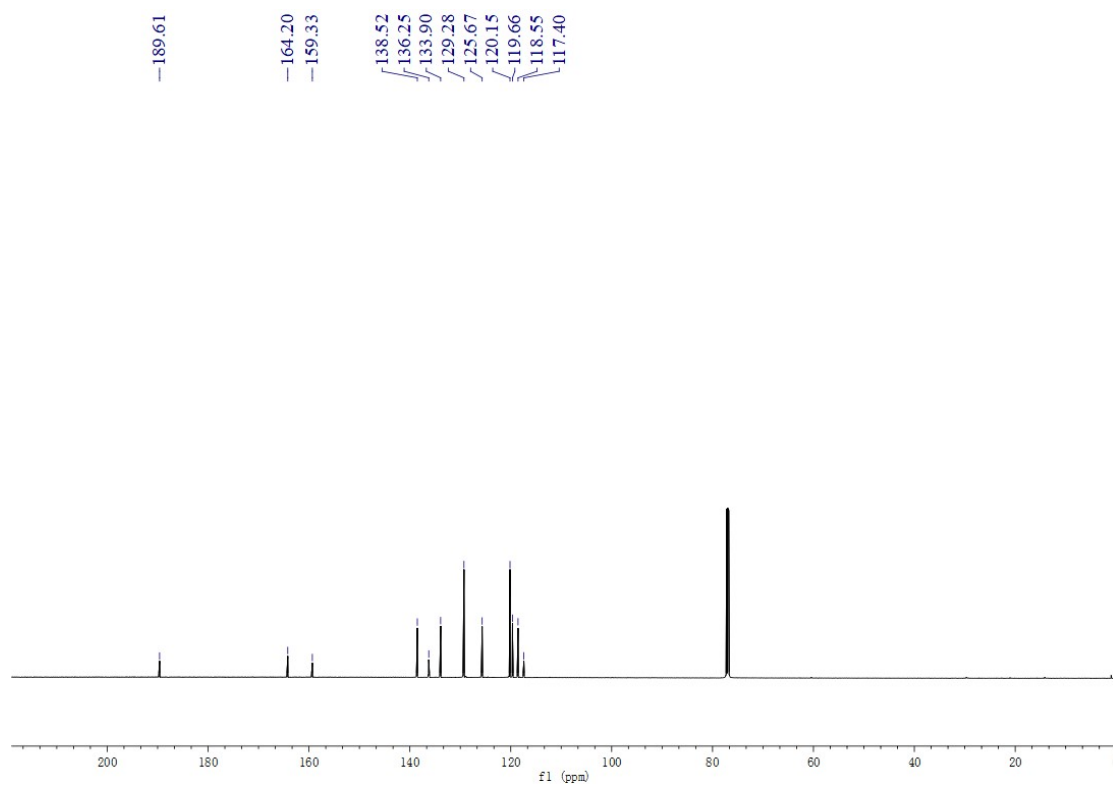
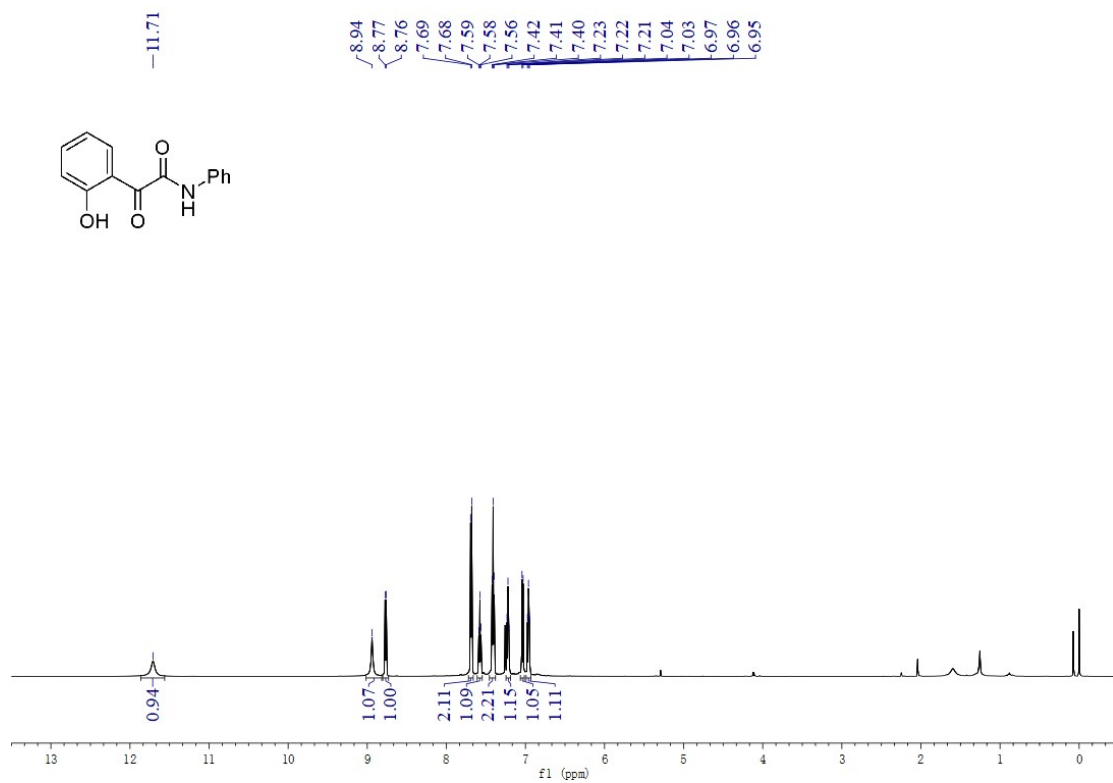
¹H and ¹³C NMR Spectra for **4f**



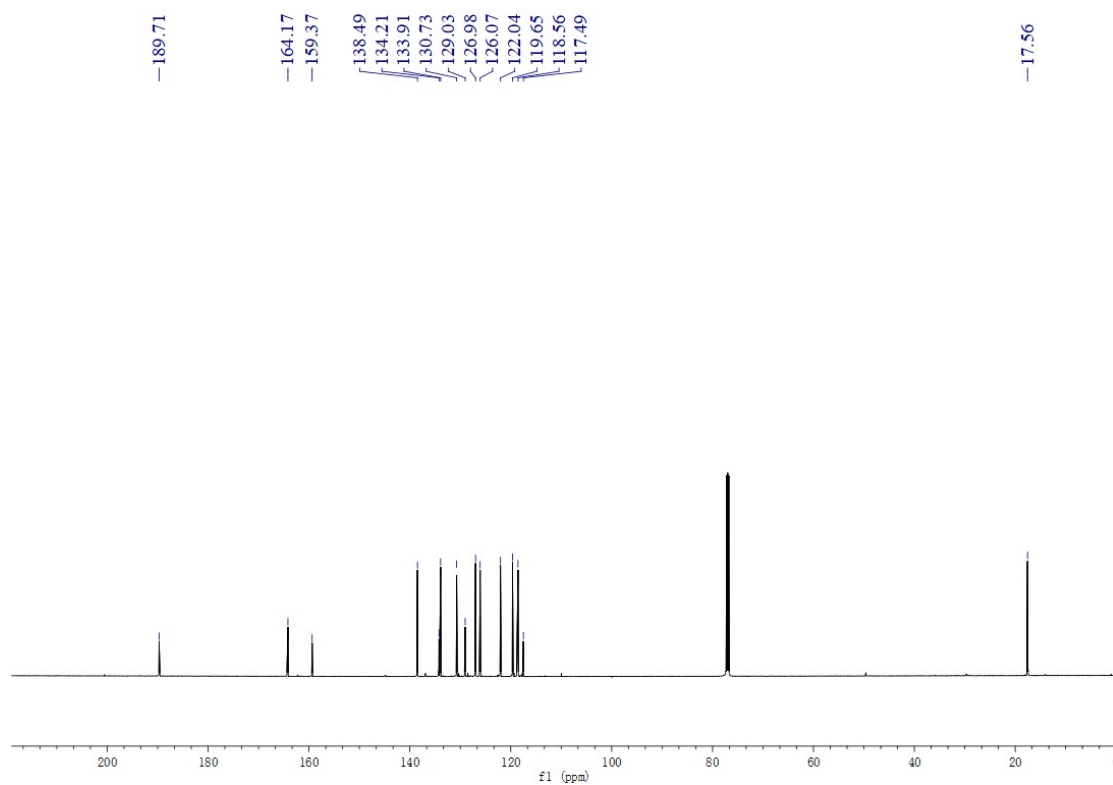
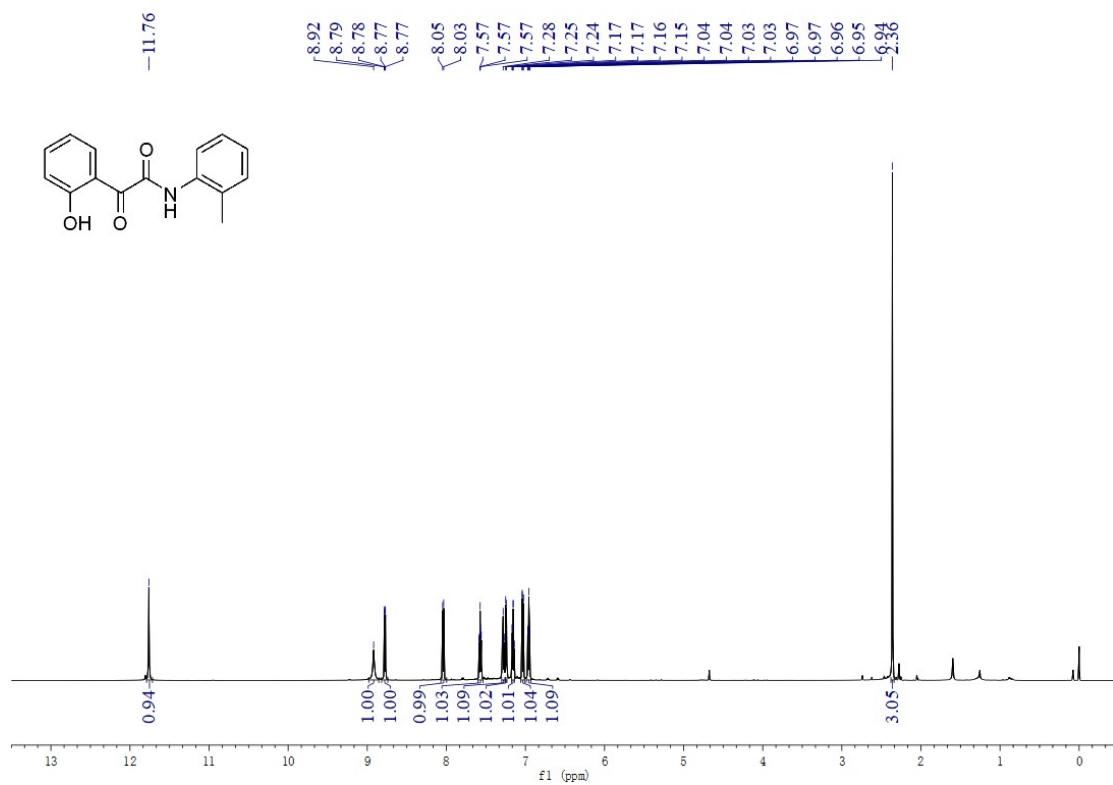
^1H and ^{13}C NMR Spectra for **4ag**



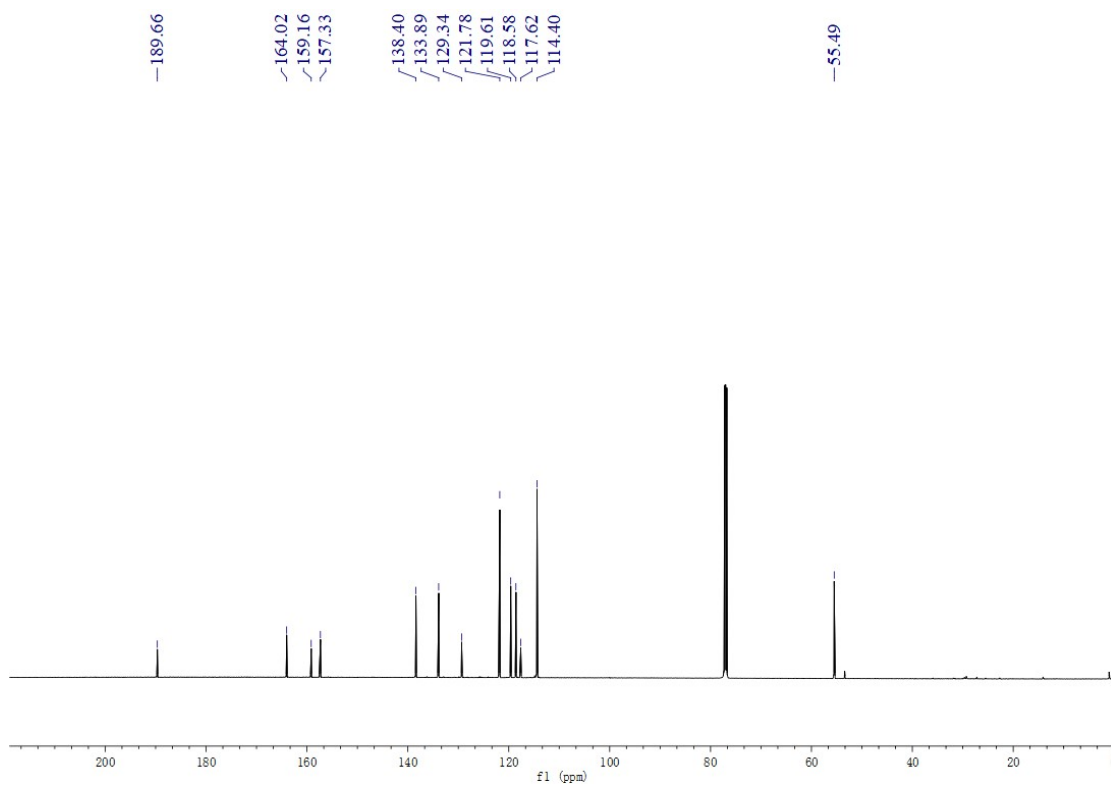
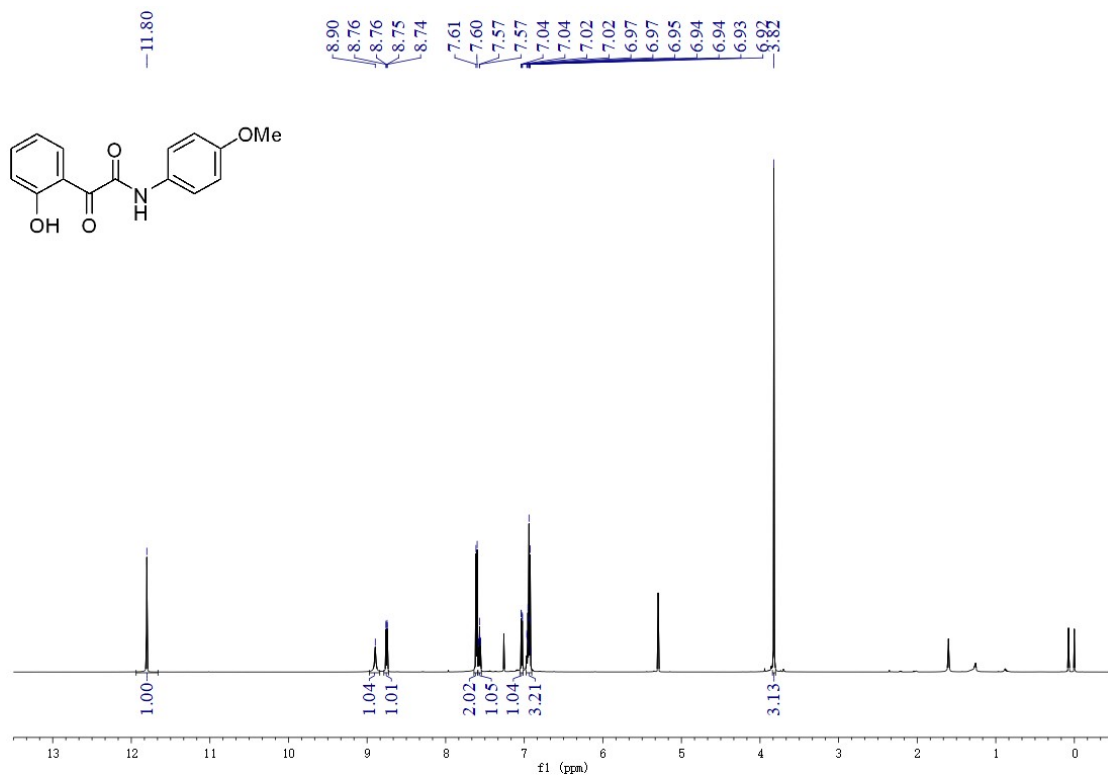
¹H and ¹³C NMR Spectra for **4r**



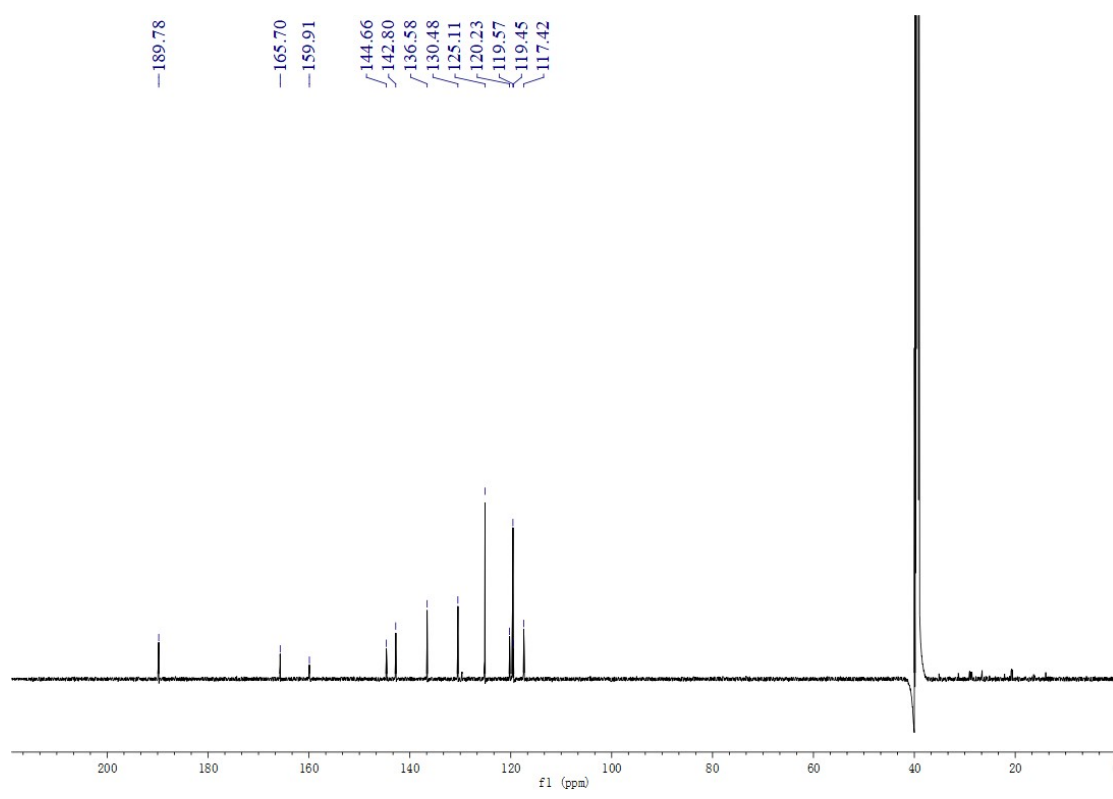
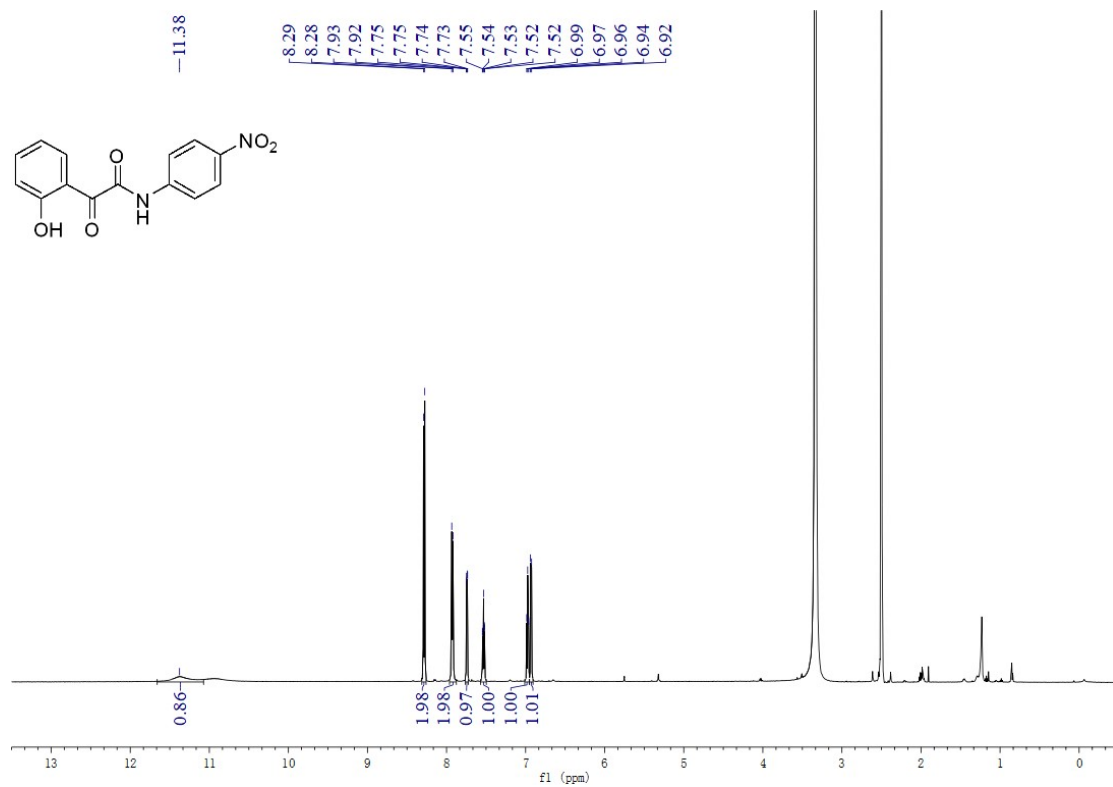
¹H and ¹³C NMR Spectra for 4s



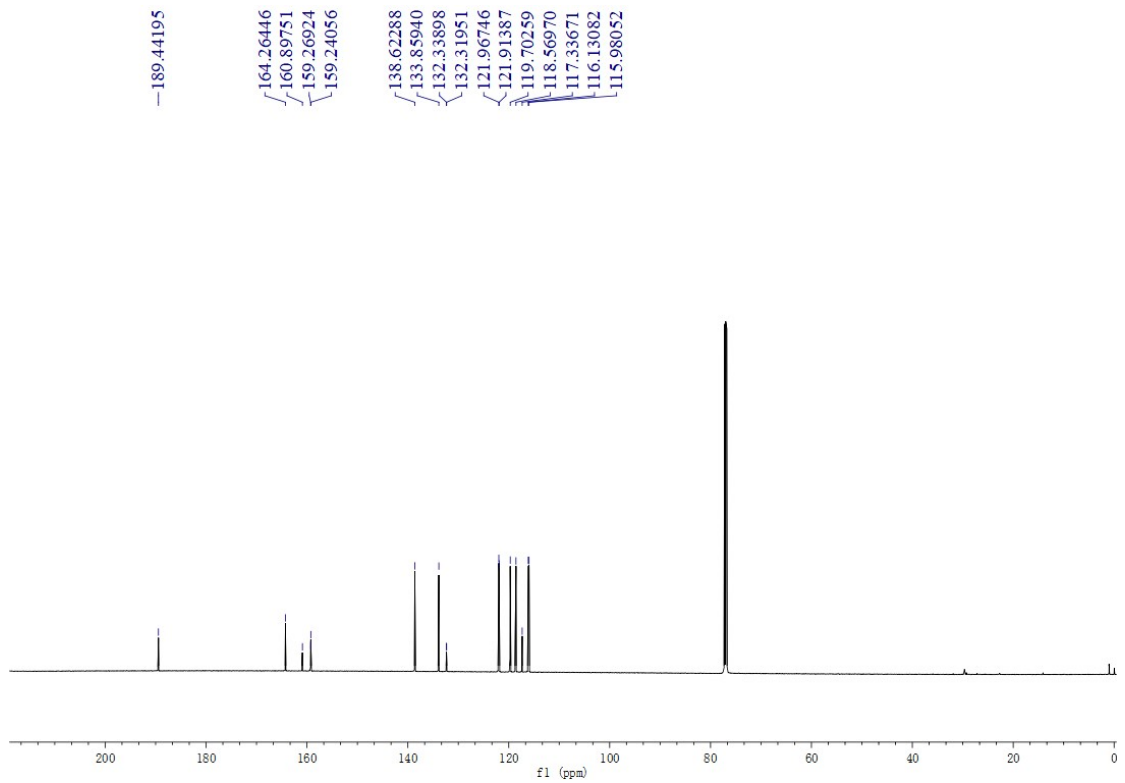
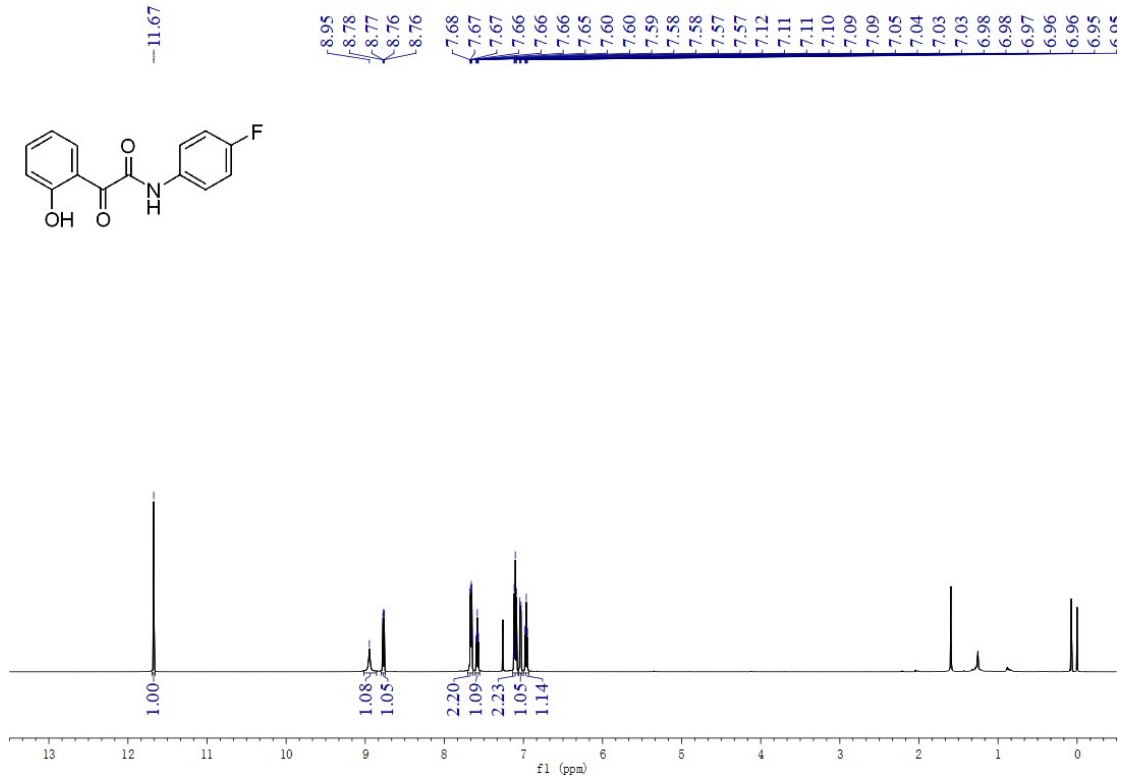
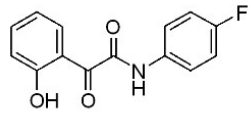
¹H and ¹³C NMR Spectra for 4t

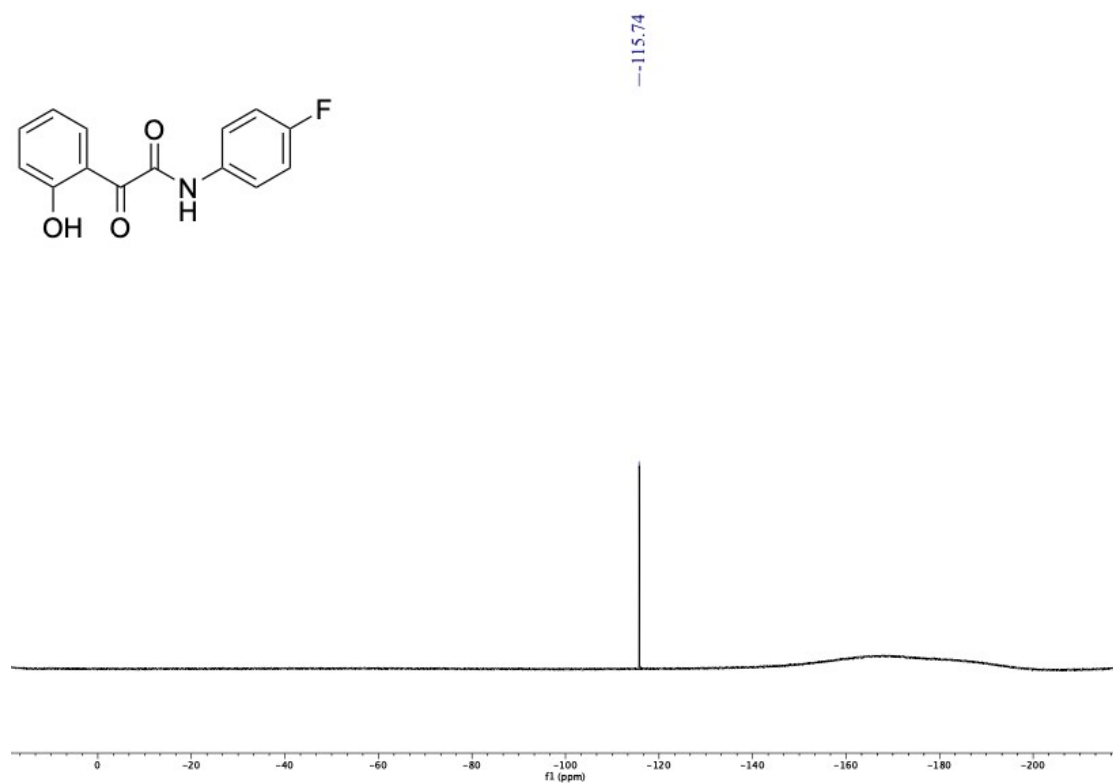
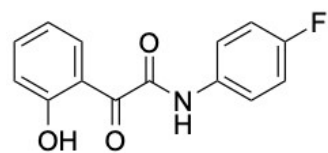


¹H and ¹³C NMR Spectra for 4u

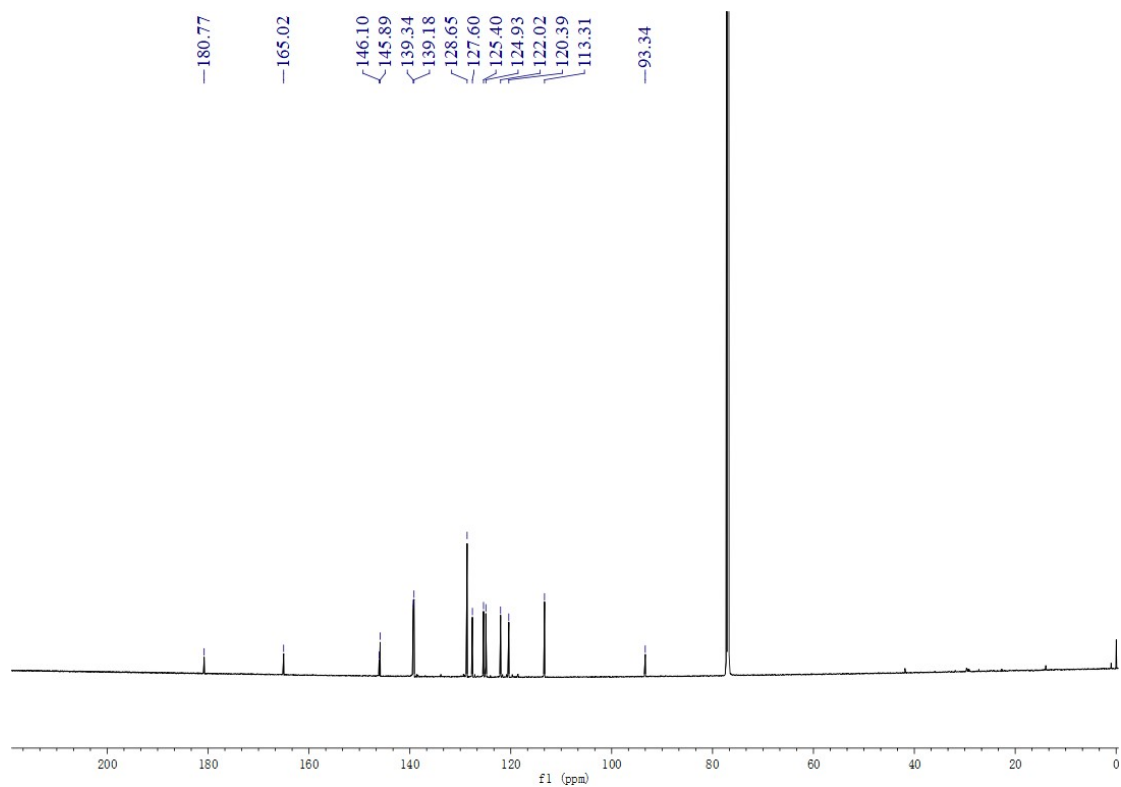
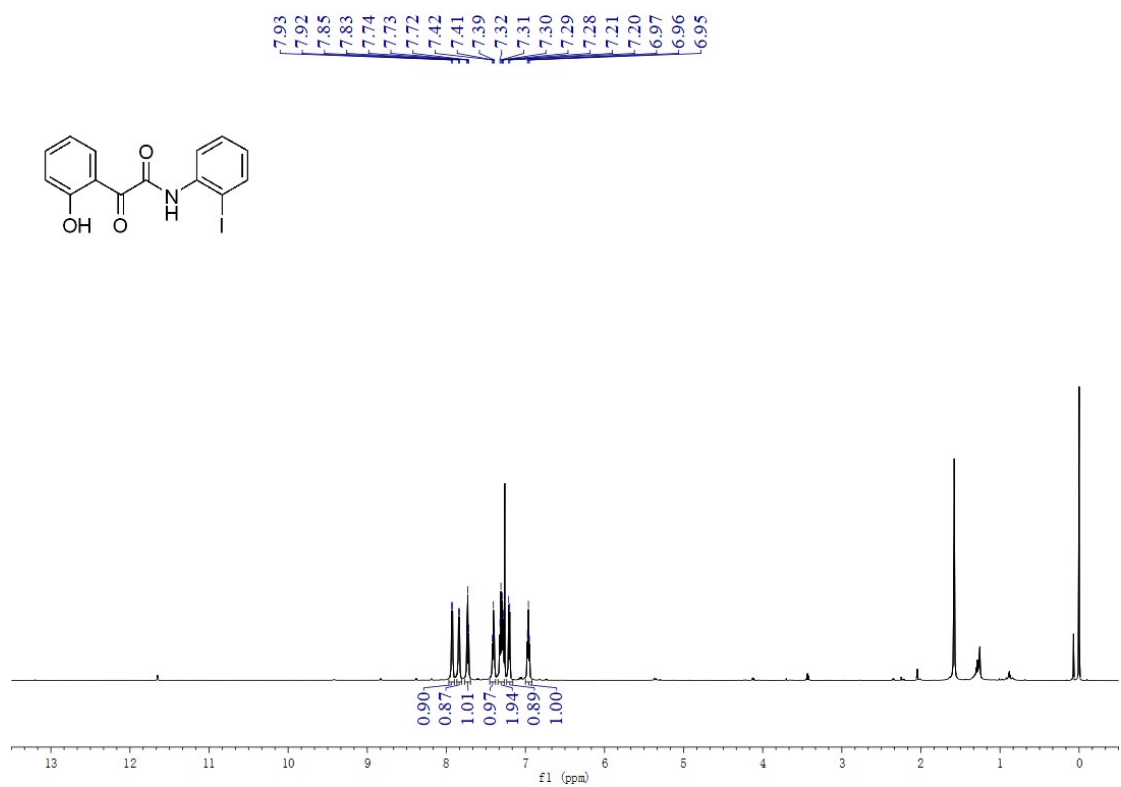


¹H ¹³C and ¹⁹F NMR Spectra for **4v**

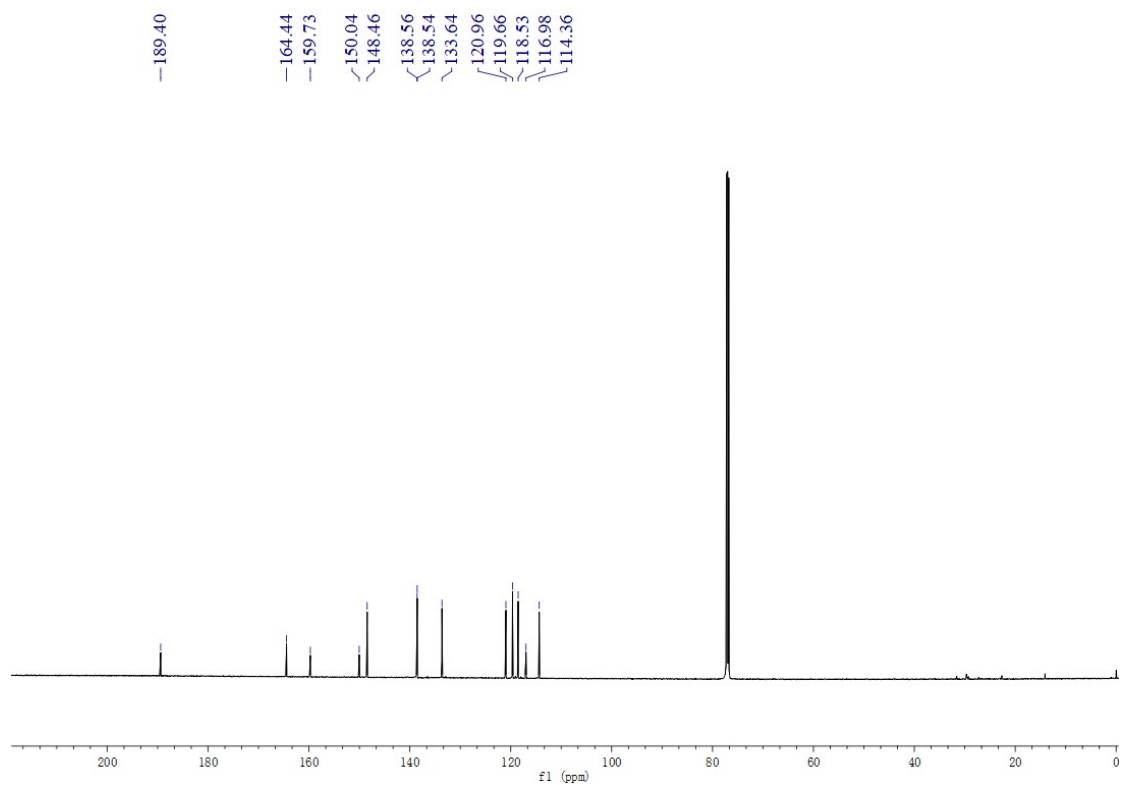
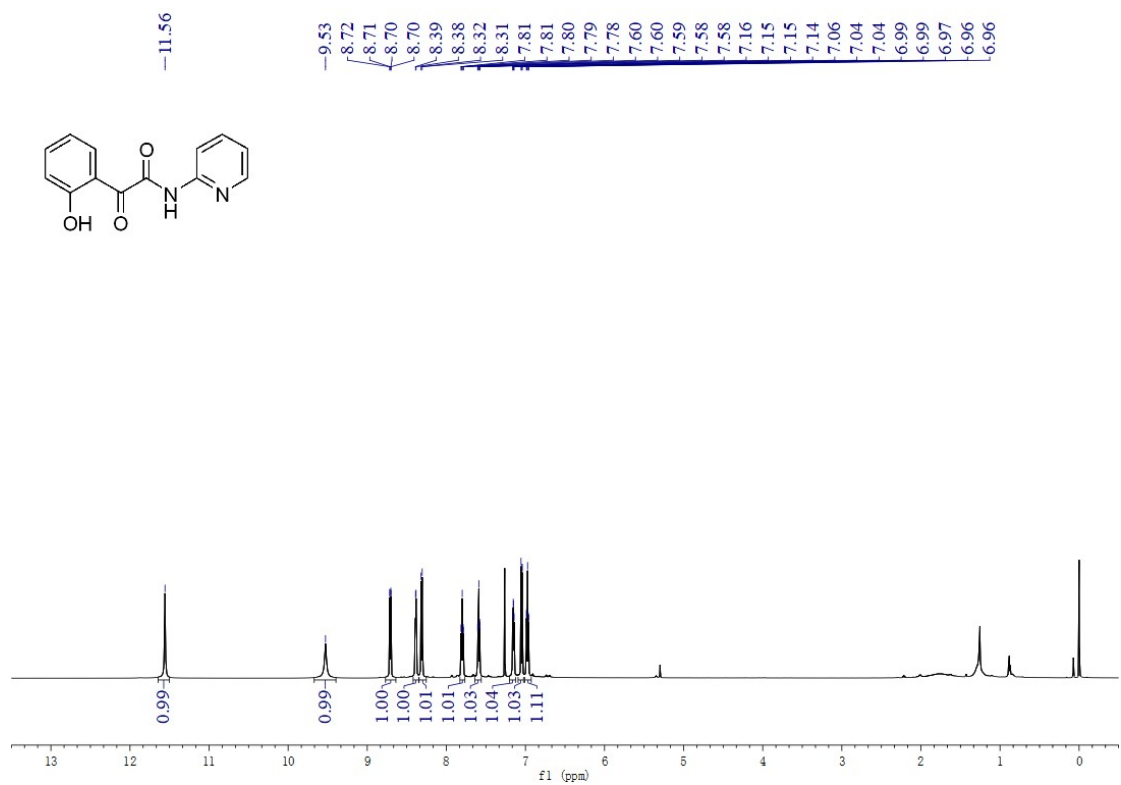




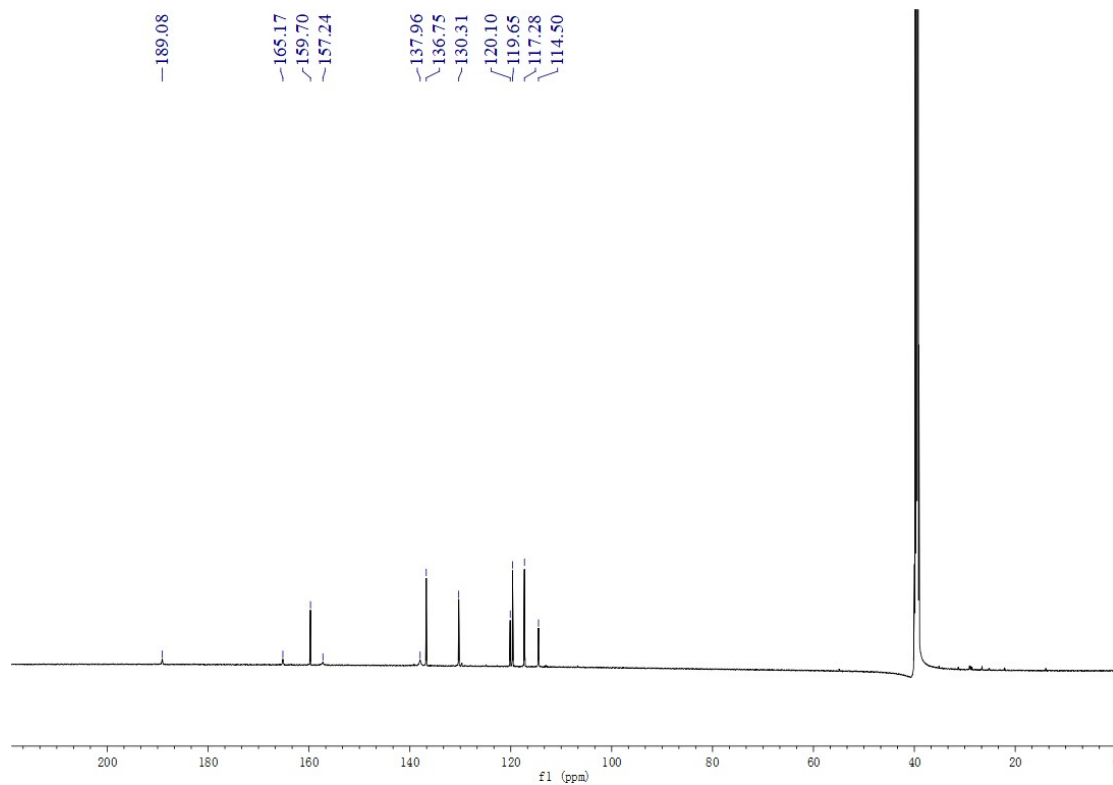
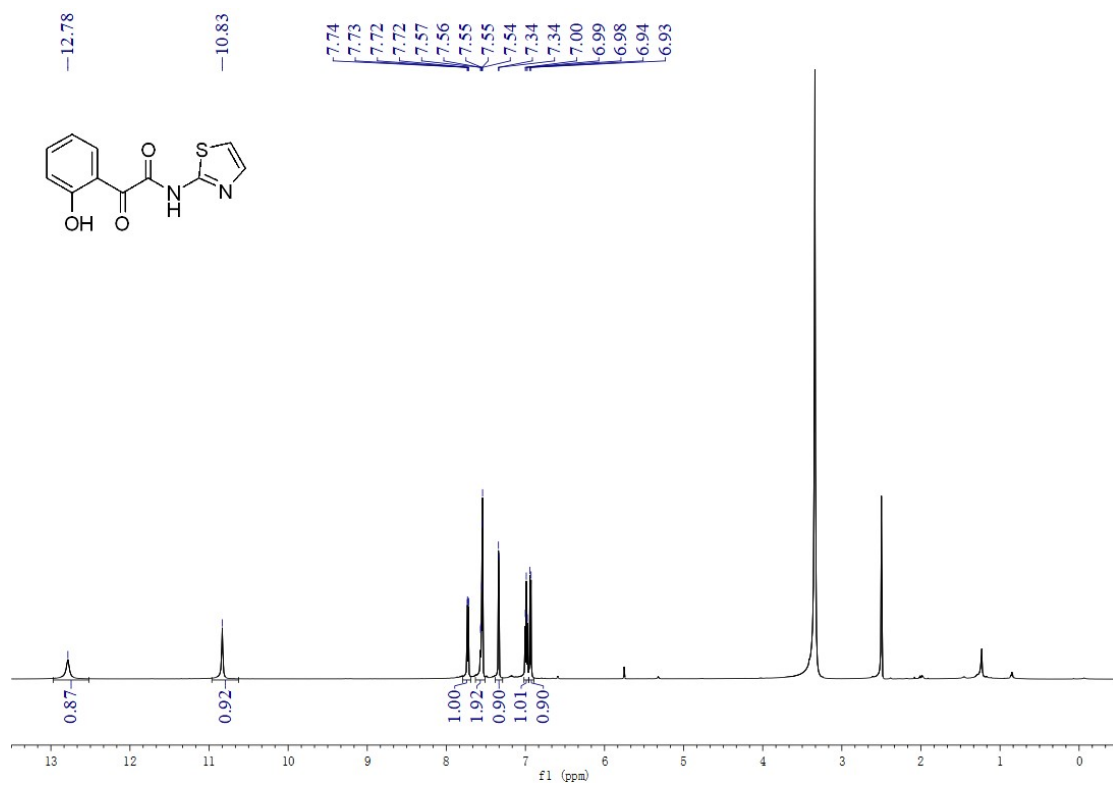
^1H and ^{13}C NMR Spectra for **4w**



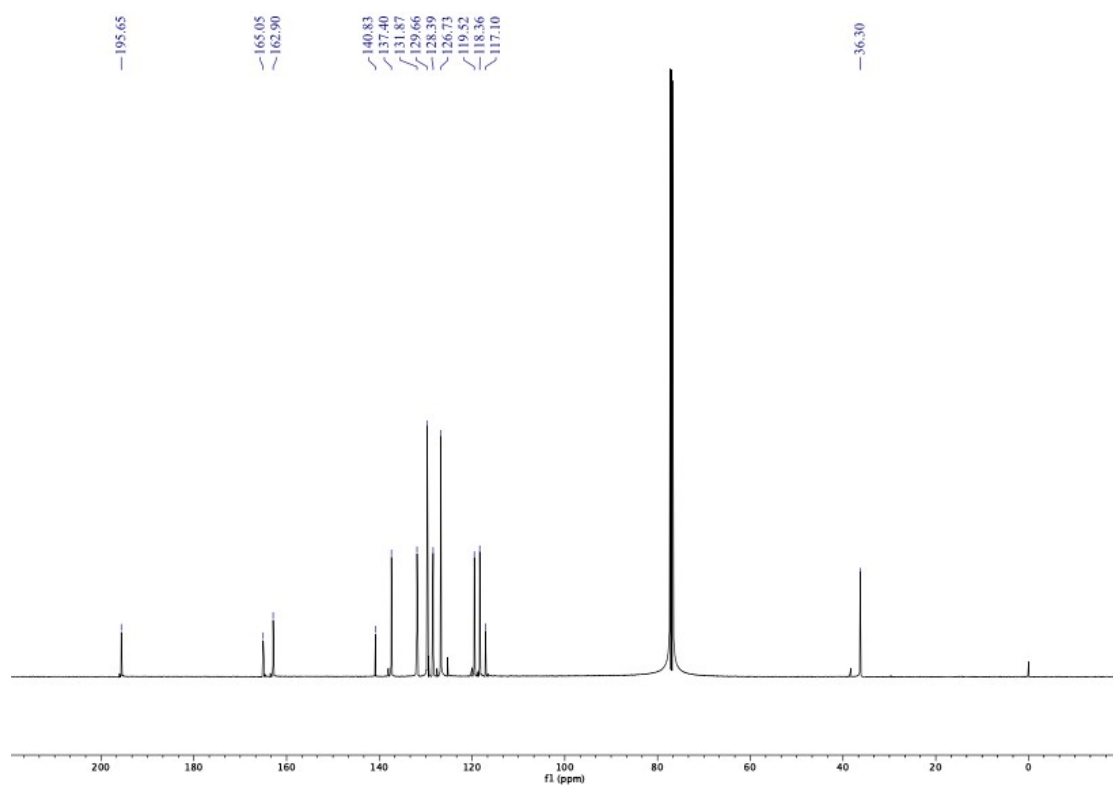
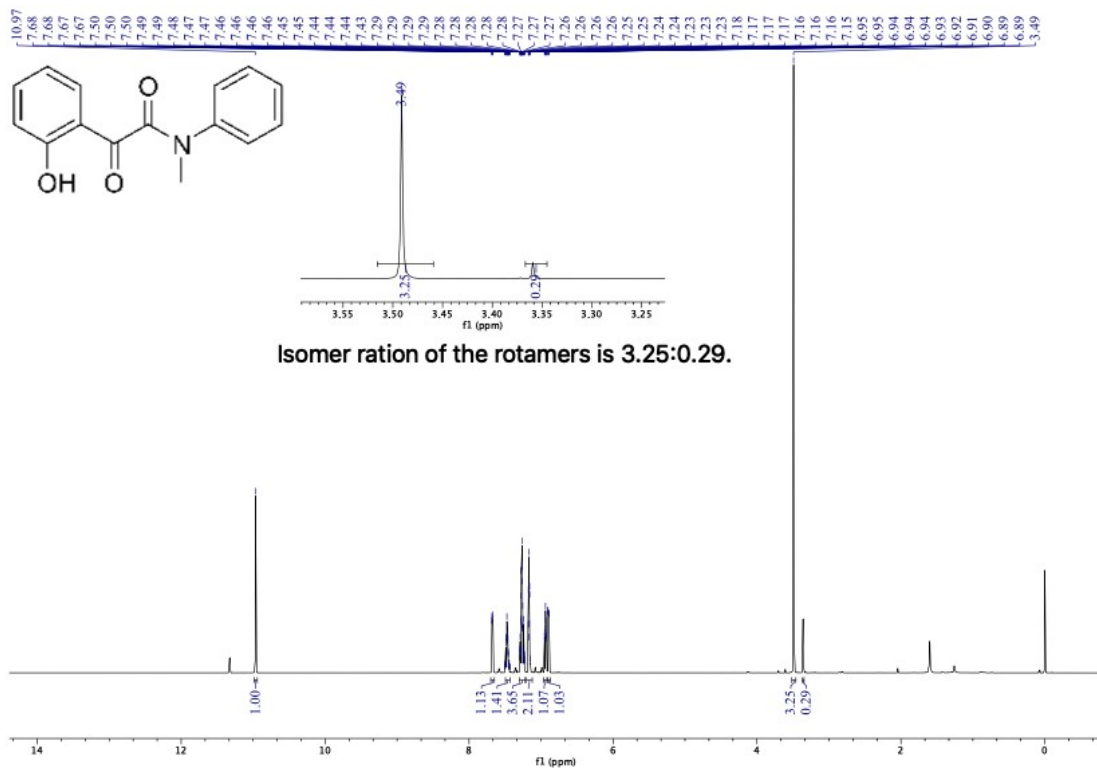
¹H and ¹³C NMR Spectra for 4x



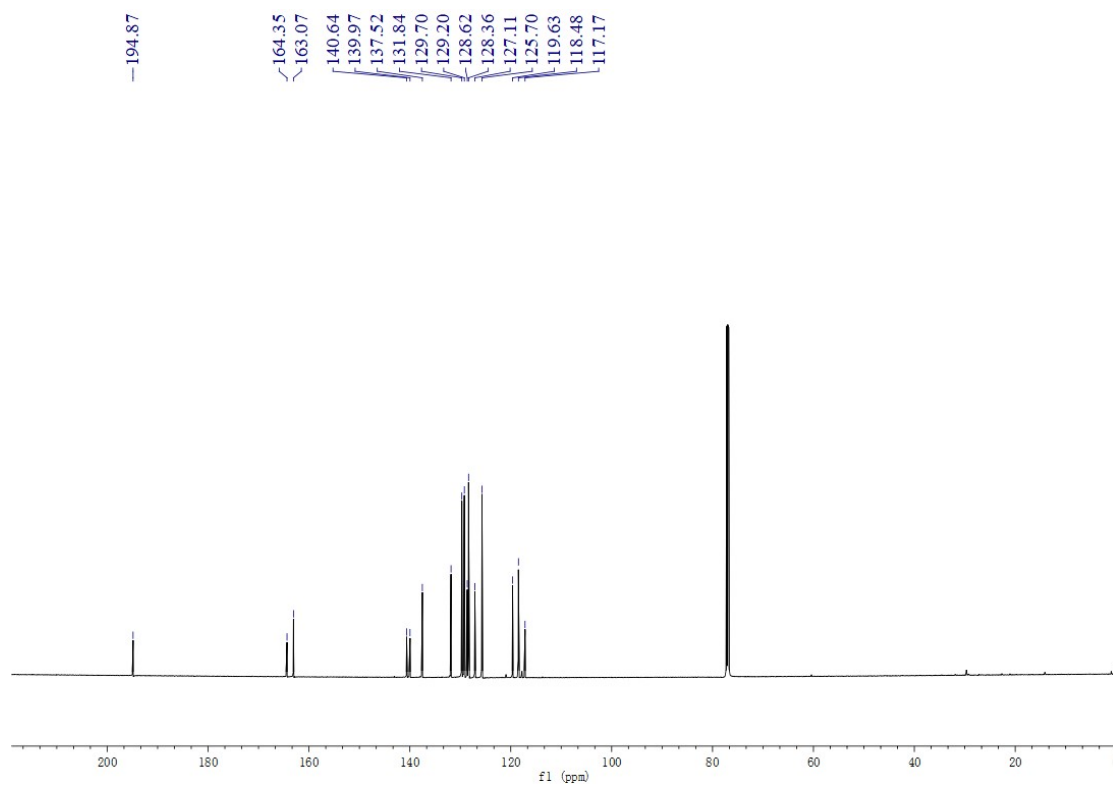
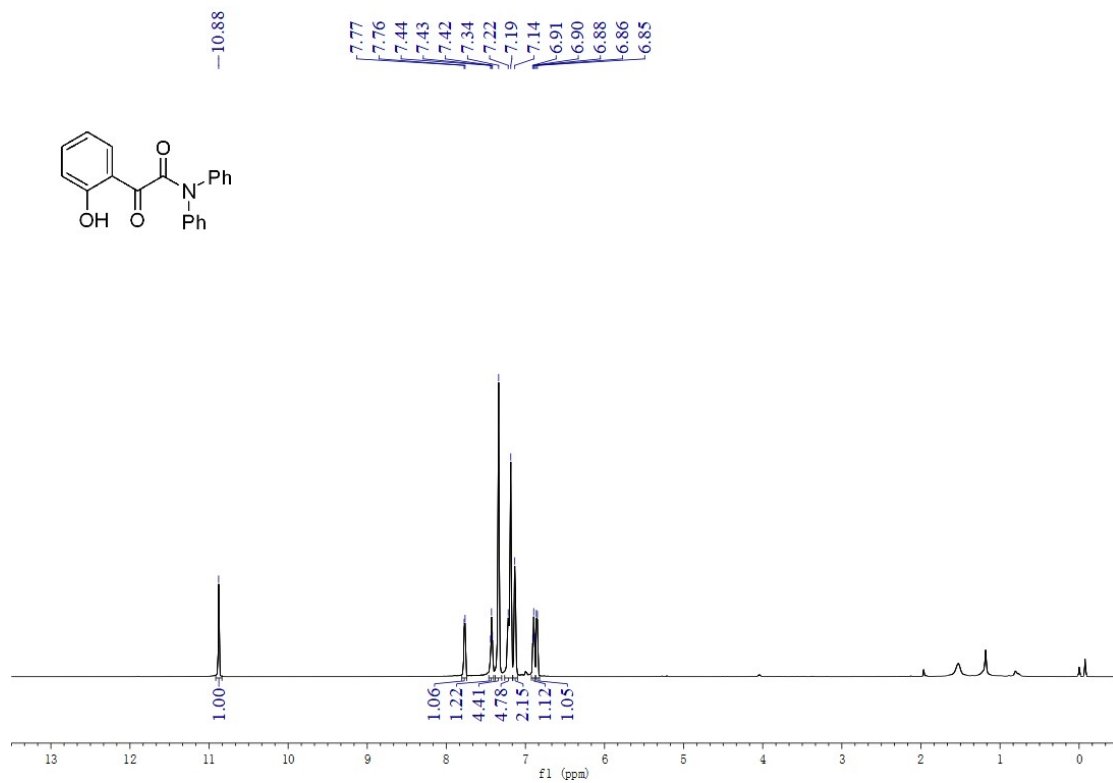
¹H and ¹³C NMR Spectra for 4y



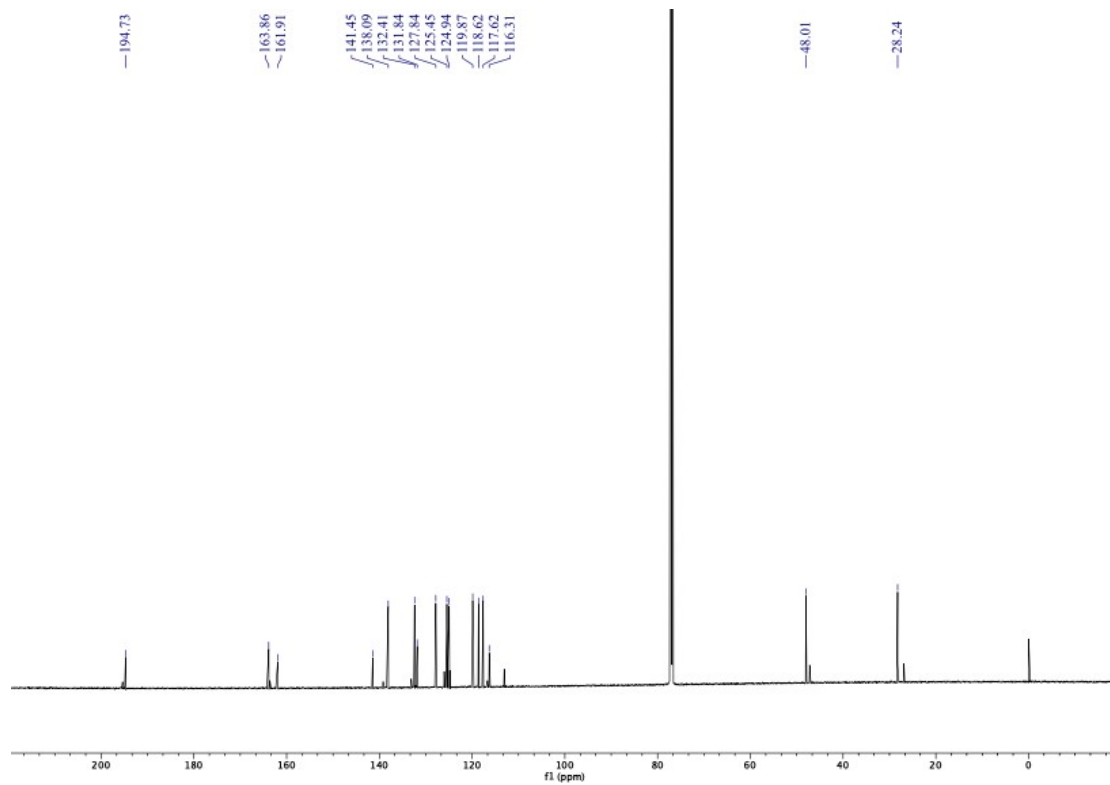
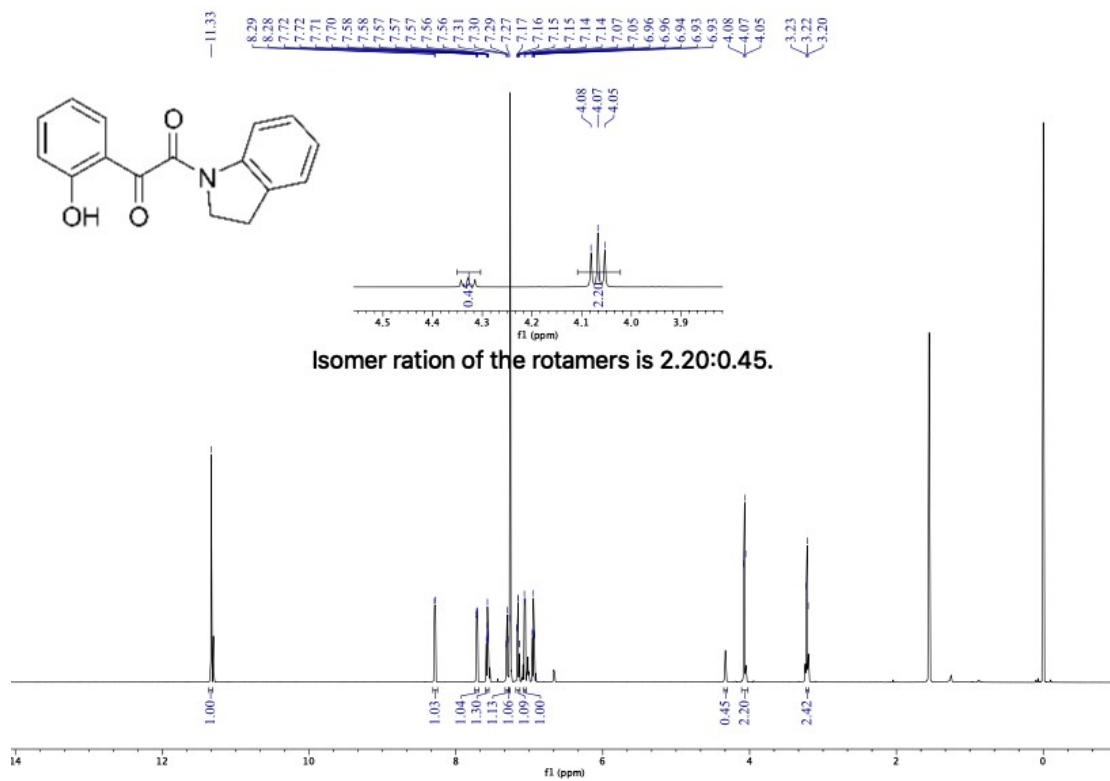
¹H and ¹³C NMR Spectra for 4z



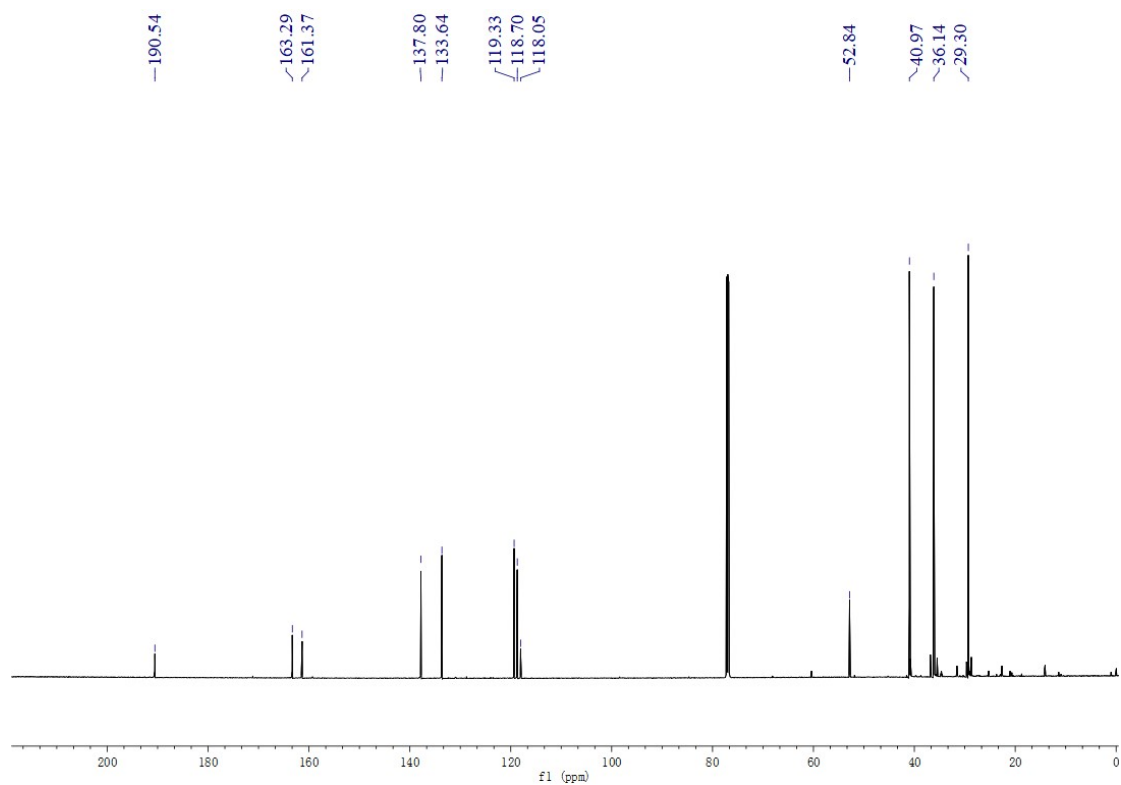
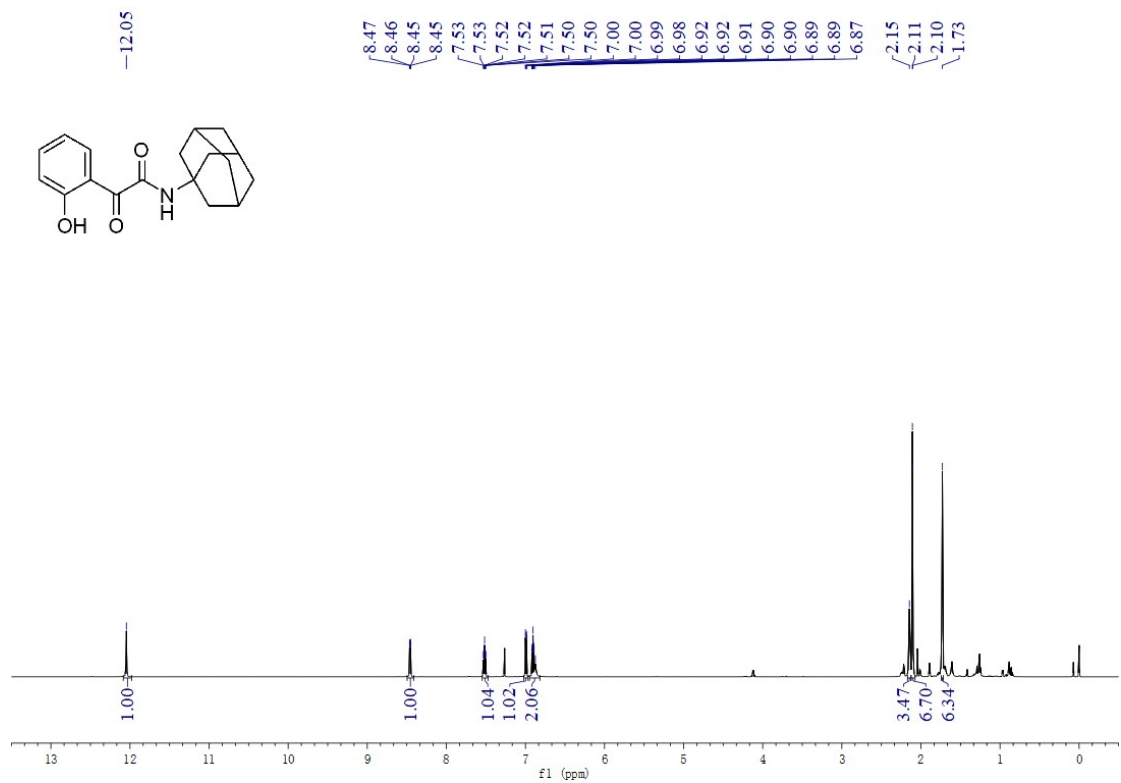
¹H and ¹³C NMR Spectra for **4za**



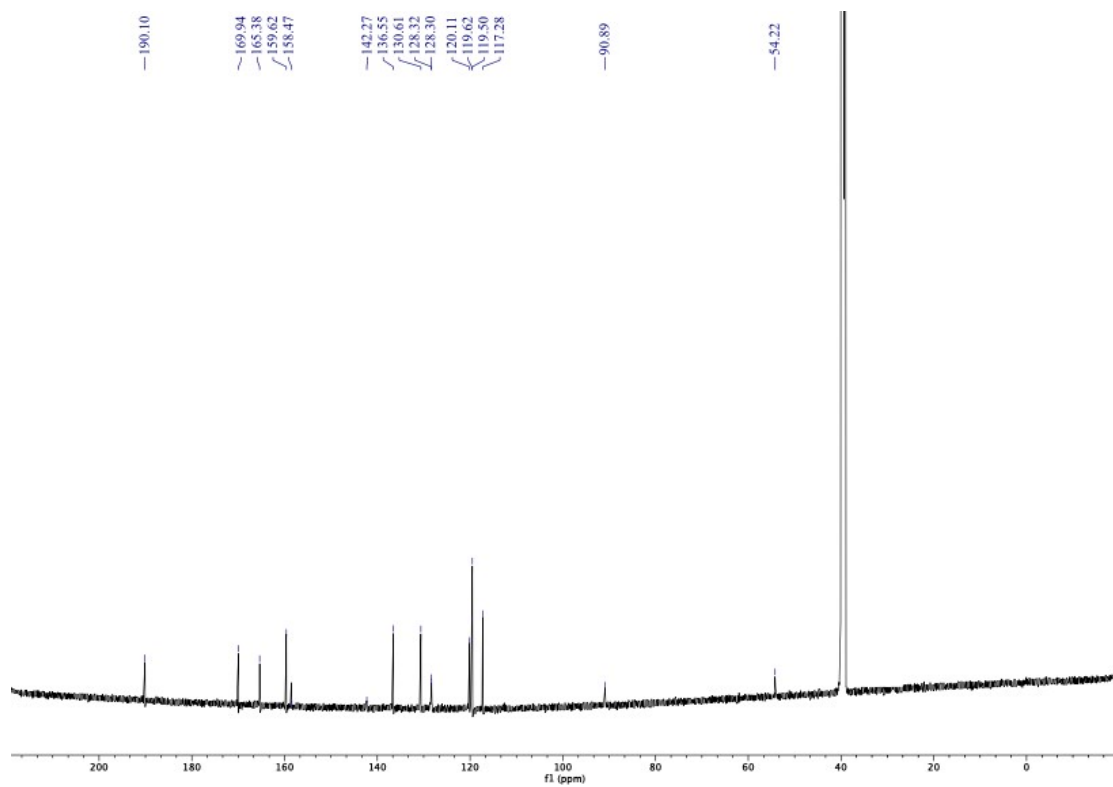
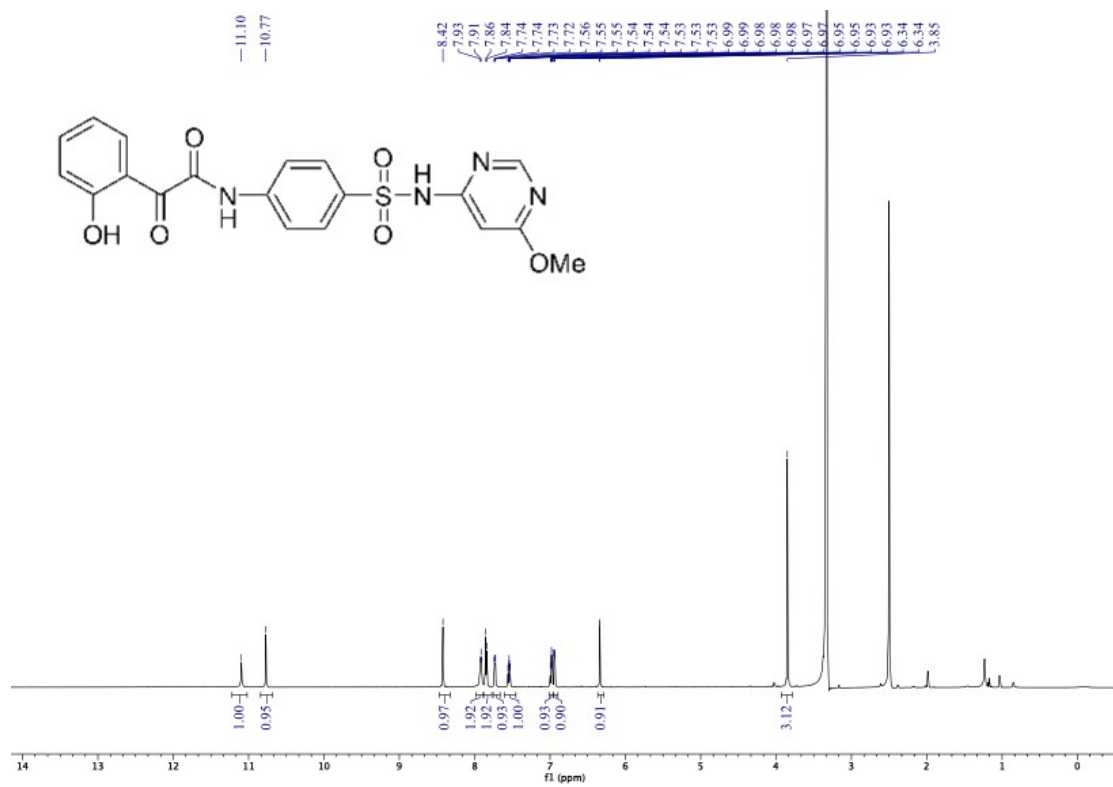
¹H and ¹³C NMR Spectra for **4zb**



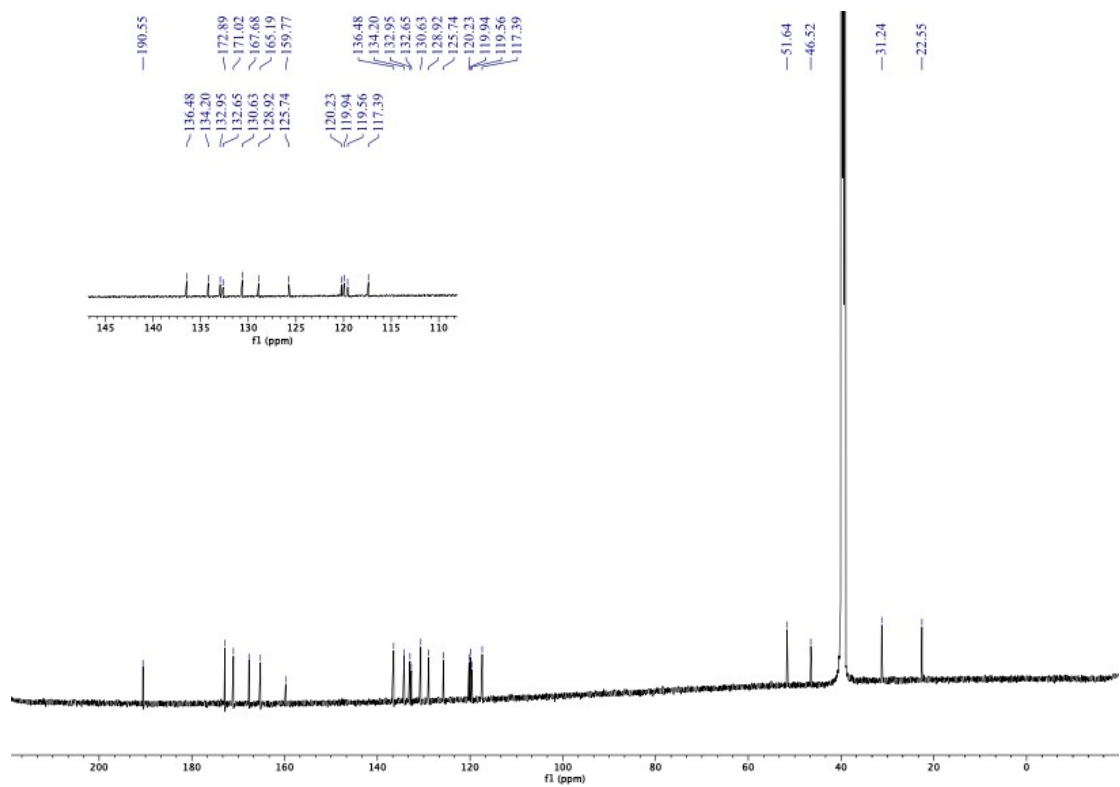
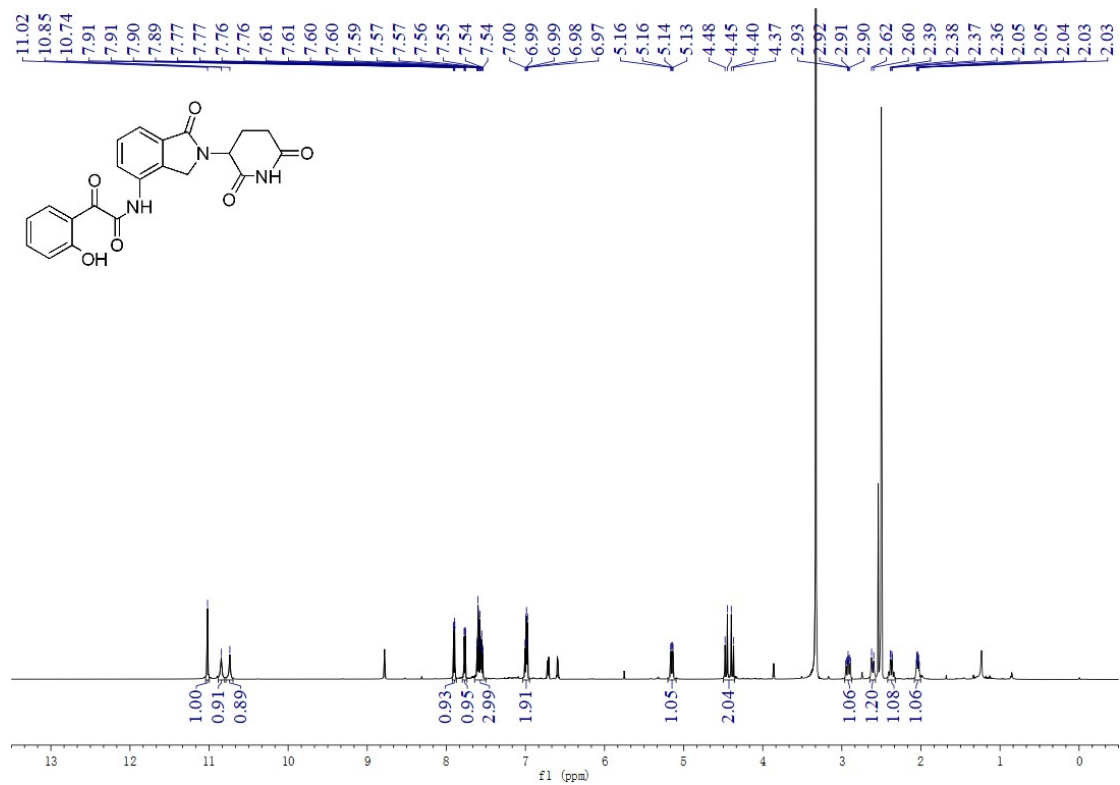
¹H and ¹³C NMR Spectra for 5a



^1H and ^{13}C NMR Spectra for **5b**



¹H and ¹³C NMR Spectra for **5c**



¹H and ¹³C NMR Spectra for **3u**

