# **Supporting Information**

# Polymersomes with smectic liquid crystal structure and AIE fluorescence

Nian Zhang, <sup>a,†</sup> Yujiao Fan, <sup>a,†</sup> Hui Chen, <sup>a,b</sup> Sylvain Trépout, <sup>c</sup> Annie Brûlet, <sup>d</sup> and Min-Hui Li\*a

<sup>a</sup> Chimie ParisTech, PSL Université Paris, CNRS, Institut de Recherche de Chimie Paris, UMR8247, 11 rue Pierre et Marie Curie, 75005 Paris, France.

<sup>b</sup> Beijing Advanced Innovation Center for Soft Matter Science and Engineering, Beijing University of Chemical Technology, 15 North Third Ring Road, Chaoyang District, 100029 Beijing, China.

<sup>c</sup> Institut Curie, Université Paris-Saclay, Inserm US43, CNRS UMS2016, Centre Universitaire, Bât. 101B-110-111-112, Rue Henri Becquerel, CS 90030, 91401 ORSAY Cedex, France.

<sup>d</sup> Laboratoire Léon Brillouin, Université Paris-Saclay, UMR12 CEA-CNRS, CEA Saclay, 91191 Gif sur Yvette cedex, France.

Email: min-hui.li@chimieparistech.psl.eu

<sup>†</sup>These authors contributed equally to this work.

### 1. Characterizations

### Nuclear magnetic resonance (NMR)

<sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on Bruker Avance III HD 400 MHz spectrometer at 298 K. Deuterated chloroform (CDCl<sub>3</sub>) was used as the solvent. NMR chemical shifts were recorded in parts per million referenced to the residual solvent proton ( $\delta$  = 7.26 ppm) for <sup>1</sup>H NMR and carbon ( $\delta$  = 77.1 ppm) for <sup>13</sup>C NMR.

## Size exclusion chromatography (SEC)

The number-average molecular weights (Mn) and molecular weight distributions of polymers (polydispersity index, *D*) were evaluated by size exclusion chromatography (SEC), using Agilent 1260 Infinity Series GPC (ResiPore 3  $\mu$ m, 300 × 7.5 mm, 1.0 mL/min, UV (250 nm) and refractive index (RI, PLGPC 220) detector. All measurements were performed with THF as the eluent at a flow rate of 1.0 mL/min at 35 °C. Monodisperse polystyrene polymers were used as calibration standards.

#### Small-angle X-ray scattering (SAXS)

SAXS experiments were carried out on the Xeuss 2.0 apparatus of Laboratoire Léon Brillouin installed in the SWAXS Lab (CEA Saclay, France). The instrument uses a micro-focused Cu K $\alpha$  source (wavelength of 1.54 Å, 8 keV) and a Pilatus3 1M detector (Dectris, Switzerland). One configuration was chosen to cover the broad q range, from 0.09 to 4 nm<sup>-1</sup>, with the sample to detector distance set to 1.18m. The sample is a fiber drawn by hand with a needle from a molten copolymer at a temperature just below the isotropic-liquid crystal phase.

### Polarizing optical microscopy (POM) and differential scanning calorimetry (DSC)

The mesomorphic properties were studied by thermal optical polarizing microscopy using a Leitz Ortholux microscope equipped with a Mettler FP82 hot stage, and differential scanning calorimetry using a Perkin-Elmer DSC7 at the heating and cooling rates of 10 °C min<sup>-1</sup>.

#### Fluorescence emission spectroscopy

The fluorescence emission spectroscopy characterization was carried out on a FluoroMax spectrofluorometer. Samples were added to a 1cm quartz cuvette with all flanks transparent.

### Ultraviolet-visible spectroscopy (UV-Vis)

UV-vis absorption spectra were measured on a Milton Ray Spectronic 3000 Array spectrophotometer. Photoluminescence (PL) spectra were recorded on a PerkinElmer LS55 and Spex FluoroMax Spectrofluorometer.

### Dynamic light scattering (DLS)

Hydrodynamic diameters ( $D_h$ ) of the self-assemblies and their size distributions in deionized water were measured at 25 °C by dynamic light scattering (DLS, Malvern zetasizer 3000HS, UK) with a 633 nm laser. All measurements were performed with a 90° scattering angle. The sample solution in the scattering cell was equilibrated for 10 min before measurement.

### Scanning electron microscopy (SEM)

SEM observations were performed on a Zeiss Leo 1530 FEG-SEM at 10kV. SEM samples were prepared by dropping a drop of the aggregate dispersions onto silicon wafers, followed by freeze-drying or vacuum drying for 3 hours. Before dropping the samples, the silicon wafers were hydrophilized in a bath of 100 mL mixture (70%  $H_2SO_4$ : 30%  $H_2O_2$ ) and then and then cleaned by 20 mL Milli-Q water, Acetone and Ethanol at room temperature respectively. The silicon surface was then dried with liquid nitrogen before use.

#### Cryo-electron microscopy (Cryo-EM)

Morphologies of the copolymer colloids were characterized by cryo-EM. Images were acquired on a JEOL 2200FS energy-filtered (20 eV) field emission gun electron microscopy operating at 200 kV using a Gatan ssCCD 2048  $\times$  2048 pixels. Samples were prepared by deposition of 5  $\mu$ L sample solution onto a 200-mesh holey carbon copper grid (Ted Pella Inc., U.S.A.) and the samples were flash-frozen in liquid ethane cooled down at liquid nitrogen.

#### 2. Synthesis of AChol monomer and mPEG-TPE-CTA

The synthesis of AChol monomer was according to the reported procedures.<sup>1</sup> As shown in Figure S1, AChol was synthesized by a one-step reaction between 2-hydroxyethyl acrylate and cholesteryl chloroformate using pyridine as the acid-binding agent. After purification by column chromatography with chloroform and *n*-hexane as the eluents, pure AChol was obtained with a yield of about 72%. AChol has been characterized carefully by NMR. Its <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra are shown in Figure S2. All the NMR signals could be assigned to the corresponding protons or carbons of AChol, indicating that pure AChol was prepared.



Figure S1. Synthetic route to AChol monomer.



Figure S2. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra of AChol. CDCl<sub>3</sub>, 400 MHz, 297 K.



Figure S3. <sup>1</sup>H NMR spectrum of mPEG-TPE-CTA. CDCl<sub>3</sub>, 400 MHz, 297 K.



Figure S4. GPC traces of mPEG-OH and mPEG-TPE-CTA with THF as the eluent.



Figure S5. <sup>1</sup>H NMR spectrum of PEG<sub>45</sub>-TPE-*b*-PAChol<sub>16</sub> copolymer. CDCl<sub>3</sub>, 400 MHz, 297 K.

Table S1. Synthesis of LC amphiphilic PEG-TPE-b-PAChol diblock copolymer.<sup>a</sup>

	Sample	[AChol]/ [mPEG-TPE-CTA]/[AIBN] <sup>b</sup>	Yield <sup>c</sup>	
_	PEG <sub>45</sub> -TPE- <i>b</i> -PAChol <sub>11</sub>	5:1:0.2	57%	
	PEG <sub>45</sub> -TPE- <i>b</i> -PAChol <sub>14</sub>	7:1:0.2	64%	
	PEG <sub>45</sub> -TPE- <i>b</i> -PAChol <sub>16</sub>	10:1:0.2	67%	
	PEG <sub>45</sub> -TPE- <i>b</i> -PAChol <sub>22</sub>	15:1:0.2	76%	
	PEG <sub>45</sub> -TPE- <i>b</i> -PAChol <sub>43</sub>	30:1:0.2	51%	

*a*. All the polymerization trials were performed using mPEG-TPE-CTA ( $M_{n,NMR} = 2680$  Da, D = 1.16) as the chain transfer agent in 1,4-dioxane at 80 °C; polymerization time: 24 h; concentration of mPEG-TPE-CTA: 0.028 M.

*b.* Molar ratio.

c. Degree of polymerization (DP),  $M_n$  and yield were calculated by <sup>1</sup>H NMR.

3. Supplementary data of liquid crystal properties of PEG<sub>45</sub>-TPE-*b*-PAChol<sub>n</sub> (n =11, 14, 16)



**Figure S6.** The DCS curves of  $PEG_{45}$ -TPE-*b*-PAChol<sub>16</sub>. The scan rate was  $\pm 10$  °C/min. The vertical dotted line indicates the smectic-isotropic (SmA-I) transition position of the hydrophobic block PAChol<sub>16</sub>.

Process	PEG	PAChol <sub>16</sub>	PAChol <sub>16</sub>	PAChol <sub>16</sub>	
	Melting (T <sub>m</sub> ) or	SmA-I	$T_{g}$	melting	
	Recrystallization				
	(T <sub>cr</sub> )				
1 <sup>st</sup> Heating	45.7°C		66.9°C	143.2 °C, 152.4 °C (24.58 J/g)	
	(0.58 J/g)		(2.25 J/g)		
1 <sup>st</sup> Cooling	-34.7 °C	139.8 °C			
	(-3.66 J/g)	(-0.11 J/g)			
2 <sup>nd</sup> Heating	40.2 °C	Signal too flat			
	(0.79 J/g)				
2 <sup>nd</sup> Cooling	-31.5 °C	137.0 °C			
	(-4.00 J/g)	(-0.11 J/g)			

Table S2. Phase transition temperatures of PEG<sub>45</sub>-TPE-*b*-PAChol<sub>16</sub> obtained by DSC (±10 °C/min)



**Figure S7.** POM textures of PEG<sub>45</sub>-TPE-*b*-PAChol<sub>16</sub> at 25°C.



**Figure S8.** POM textures of  $PEG_{45}$ -TPE-*b*-PAChol<sub>n</sub>. (a-b) n = 14 at 120°C (a) and 25°C (b). (c-d) n = 11 at 117°C (c) and 25°C (d).



**Figure S9.** SAXS of  $PEG_{45}$ -TPE-*b*-PAChol<sub>n</sub> (n = 11 left, n = 14 right) at 25°C. The red arrow indicates the fiber drawing direction.



	Low q peak	Low q Bump	q1		q2	
	Å-1	Å-1	Å-1	d (nm)	Å-1	q2/q1
n = 11		0.69	0.1419	4.42789288	0.283	1.99436223
n = 14	0.040	0.63	0.1429	4.39690693	0.2861	2.00209937
n = 16		0.57	0.1447	4.34221147	0.2878	1.98894264

**Figure S10.** Data analyses of SAXS of  $PEG_{45}$ -TPE-*b*-PAChol<sub>n</sub> (n = 11, 14, 16) at 25°C. q1 and q2 correspond to the signals of aligned smectic A phase. Low q bump and peak correspond the microphase separation of PEG and PAChol blocks (bump signals give lengths of about 9nm, 10nm and 11nm for n = 11, 14 and 16, respectively.

#### 4. Supplementary data of self-assembly of PEG<sub>45</sub>-TPE-*b*-PAChol<sub>16</sub>



**Figure S11.** Supplemental SEM images of the self-assemblies formed from the diblock copolymer  $PEG_{45}$ -TPE-*b*-PAChol<sub>16</sub> in THF/water system at a concentration of 2 mg/mL (a) and of 1mg/mL (b), and their intensity profile of size distribution by DLS at 2 mg/mL (c) and of 1mg/mL (d).



**Figure S12.** The DLS profile (a) and SEM pictures (b-d) of the self-assemblies formed from the diblock copolymer  $PEG_{45}$ -TPE-*b*-PAChol<sub>16</sub> in THF/water system at a concentration of 10 mg/mL.

#### 5. Fluorescence quantum yield measurement

Quantum yield of the AIE polymersomers in aqueous solution were obtained by comparing the integrated fluorescence spectra of the self-assembly solution with the fluorescence spectrum of a standard fluorogen, 9,10-diphenylanthracene, in ethanol (excitation wavelength: 355 nm) with corrections of refractive index differences using equation:<sup>2</sup>

$$\phi_2 = \phi_1 \times \frac{n_2^2}{n_1^2} \times \frac{F_2}{F_1} \times \frac{A_1}{A_2}$$

 temperature (1  $\mu g/mL;$   $\lambda_{ex}$  = 355 nm).

### References

1. Piñol, R.; Jia, L.; Gubellini, F.; Lévy, D.; Albouy, P.-A.; Keller, P.; Cao, A.; Li, M.-H. *Macromolecules* **2007**, *40*, 5625-5627.

2. Lu, H.; Su, F.; Mei, Q.; Tian, Y.; Tian, W.; Johnson, R. H.; Meldrum, D. R. J. Mater. Chem. 2012, 22, 9890-9900.