

Supporting Information for

Ruthenium-Catalyzed Enantioselective Hydrogenation of Quinoxalinones and Quinazolinones

Chenghao Li,[‡] Shuxin Zhang,[‡] Shan Li, Yu Feng*, and Qing-Hua Fan*

Beijing National Laboratory for Molecular Sciences, CAS Key Laboratory of Molecular Recognition and Function, Institute of Chemistry, Chinese Academy of Sciences (CAS) and University of Chinese Academy of Sciences, Beijing 100190, P. R. China;

E-mail: fengyu211@iccas.ac.cn; fanqh@iccas.ac.cn

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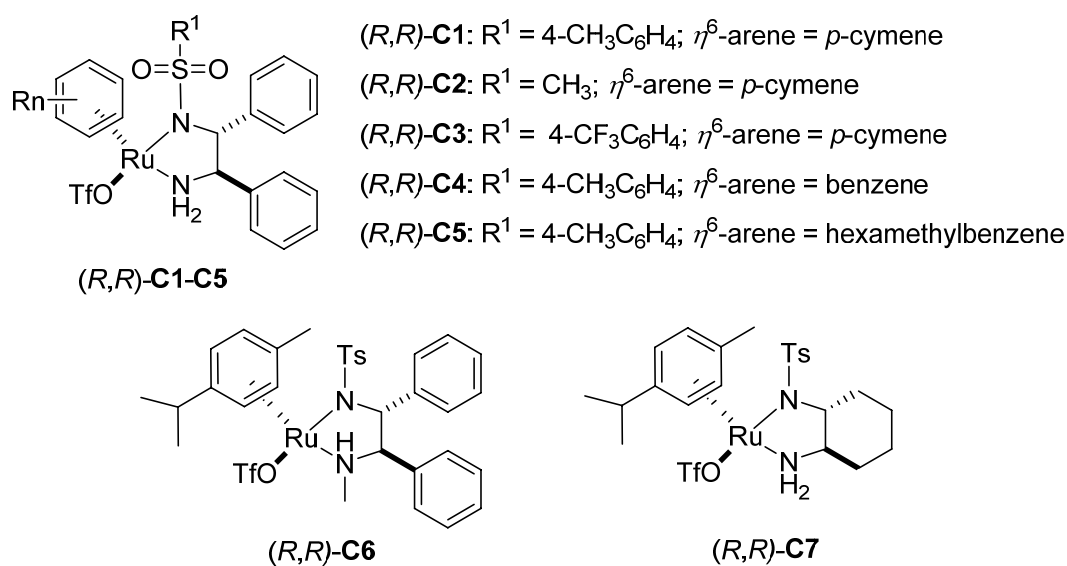
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1. General information

Unless otherwise noted, all experiments were carried out under an atmosphere of nitrogen using standard Schlenk techniques or in a nitrogen-filled glovebox. ^1H NMR and ^{13}C NMR spectra were recorded on Bruker Model Avance DMX 300 Spectrometer (^1H 300 MHz and ^{13}C 75 MHz, respectively), Bruker Model Avance DMX 400 Spectrometer (^1H 400 MHz and ^{13}C 100 MHz, respectively), Bruker Model Avance DMX 500 Spectrometer (^1H 500 MHz and ^{13}C 125 MHz, respectively) and Bruker Model Avance DMX 600 Spectrometer (^1H 600 MHz and ^{13}C 150 MHz, respectively). Chemical shifts (δ) were given in ppm and were referenced to residual solvent or TMS peaks. Optical rotations were measured with Rudolph Autopl VI polarimeter. High resolution ESI mass spectra (P-ESI HRMS) were obtained on Thermo Fisher Q Exactive Mass Spectrometer. HPLC analyses were performed on a Varian Prostar 210 liquid chromatography. All organic solvents were dried using standard, published methods and were distilled before use. All other chemicals were used as received from Aldrich or Acros without further purification. The catalysts^[1] and substrates^[2] were synthesized according to the modified literature methods.

2. Optimization of conditions for asymmetric hydrogenation reactions

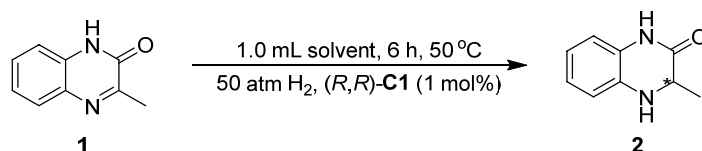
For this study, we chose the chiral diamines DPEN and CYDN as the ligand skeletons and prepared a series of Ru-complexes bearing chloride as anion according to the previous method.^[1] Then, the anion Cl⁻ was exchanged to the weakly coordinating counteranions, giving the corresponding cationic ruthenium catalysts in quantitative yields, which did not require further purification. Their structures are shown in Scheme S1.



Scheme S1. Chiral ruthenium diamine catalysts used in this study

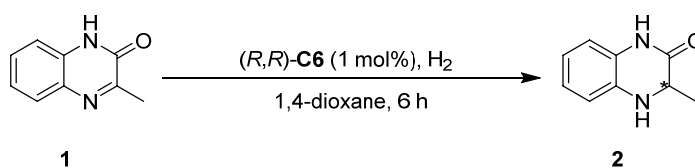
Optimization of conditions for asymmetric hydrogenation reactions are shown in Table S1-S3.

Table S1. Optimization of reaction condition: solvents^[a]



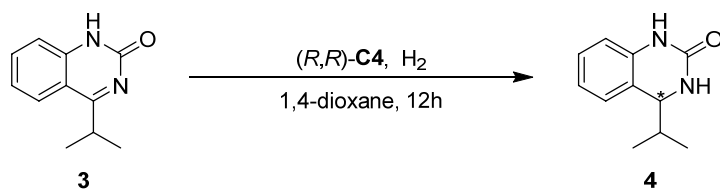
Entry	Solvent	Conv. (%) ^[b]	<i>ee</i> (%) ^[c]
1	MeOH	97	17
2	EtOH	>99	25
3	isopropanol	>99	28
4	CF ₃ CH ₂ OH	79	25
5	HFIP	18	29
6	dichloromethane	95	61
8	ethyl acetate	>99	71
9	toluene	>99	63
10	tetrahydrofuran	97	84
11	1,4-dioxane	96	84

^aReaction conditions: **1a** (0.1 mmol) in 1.0 mL solvent, (*R,R*)-**C1** (1.0 mol%), H₂ (50 atm), stirred at 50 °C for 6 h. ^bThe conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^cThe enantiomeric excesses (*ee*) were determined by HPLC with a chiral OD-H column. HFIP = 1,1,1,3,3,3-hexafluoro-2-propanol.

Table S2. Optimization of other reaction conditions^[a]

Entry	T (°C)	H ₂ (atm)	Conv. (%) ^[b]	ee (%) ^[c]
1	25	50	>99	98
2	50	50	>99	98
3	80	50	>99	98
4	25	80	>99	98
5	25	10	40	98

^aReaction conditions: **1a** (0.1 mmol) in 1.0 mL 1,4-dioxane, (*R,R*)-**C6** (1.0 mol%), stirred for 6 h. ^bThe conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^cThe enantiomeric excesses (*ee*) were determined by HPLC with a chiral OD-H column.

Table S3. Optimization of other reaction conditions^[a]

Entry	T (°C)	H ₂ (atm)	Conv. (%) ^[b]	ee (%) ^[c]
1	25	50	>99	86
2	50	50	>99	87
3	80	50	>99	86
4	25	20	>99	86
5	25	70	>99	86
6^[d]	25	50	>99	87
7 ^[e]	25	50	<5	-

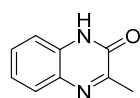
^aReaction conditions: **3b** (0.1 mmol) in 1.0 mL solvent, (*R,R*)-**C4** (10 mol%), H₂, stirred for 12 h. ^bThe conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^cThe enantiomeric excesses (*ee*) were determined by HPLC with a chiral IC-H column. ^d(*R,R*)-**C4** (5 mol%), ^e(*R,R*)-**C4** (1 mol%).

3. General procedure for the synthesis of substrates

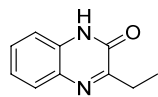
3.1 Synthesis of substrate quinoxalin-2(1H)-ones (1)



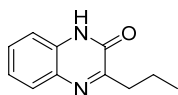
General procedure: In a 250 mL flask, 1,2-diaminobenzene (0.540 g, 5.0 mmol) was dissolved in ethyl alcohol (100 mL). Then, ethyl pyruvate (1.277 g, 11.0 mmol) was added, and the reaction mixture was stirred at room temperature until the reaction was completed. Then, the mixture was filtered and recrystallized from ethyl alcohol to afford 3-methylquinoxalin-2(1H)-one (**1a**) as white solid. The analytical data of the products are summarized below.



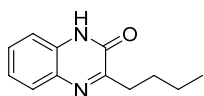
3-methylquinoxalin-2(1H)-one (1a): Known compound (Xia, Q.-H.; Hu, W.; Li, C.; Wu, J.-F.; Yang, L.; Han, X.-M.; Shen, Y.-M.; Li, Z.-Y.; Li, X. *Eur. J. Med. Chem.* **2016**, *124*, 311-325). White solid, 0.736 g, isolated yield 92%. ¹H NMR (500 MHz, DMSO-*d*₆): δ (ppm) 12.29 (bs, 1H), 7.69 (d, *J* = 8.0 Hz, 1H), 7.46 (t, *J* = 7.5 Hz, 1H), 7.28-7.24 (m, 2H), 2.40 (s, 3H); ¹³C NMR (125 MHz, DMSO-*d*₆): δ (ppm) 159.2, 154.9, 131.9, 131.6, 129.3, 127.9, 123.0, 115.2, 20.5. HRMS-ESI exact mass calcd. for C₉H₇N₂O⁺ ([M-H]⁺) requires *m/z* 159.05639, found *m/z* 159.05539.



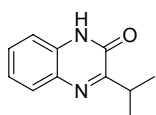
3-ethylquinoxalin-2(1H)-one (1b): Known compound (Blocher, R.; Ramirez, A. R.; Castro-Escarpulli, G.; Curiel-Quesada, E.; Reyes-Arellano, A. *Molecules* **2018**, *23*, 3075). Pale yellow solid, 0.783 g, isolated yield 90%. ¹H NMR (500 MHz, DMSO-*d*₆): δ (ppm) 12.28 (bs, 1H), 7.70 (d, *J* = 7.5 Hz, 1H), 7.45 (t, *J* = 7.5 Hz, 1H), 7.28-7.24 (m, 2H), 2.79 (q, *J* = 7.3 Hz, 2H), 1.21 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (125 MHz, DMSO-*d*₆): δ (ppm) 162.5, 154.6, 131.7, 131.6, 129.3, 128.0, 123.0, 115.2, 26.0, 10.5. HRMS-ESI exact mass calcd. for C₁₀H₉N₂O⁺ ([M-H]⁺) requires *m/z* 173.07204, found *m/z* 173.07117.



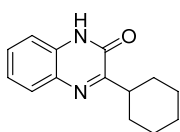
3-propylquinoxalin-2(1H)-one (1c): Known compound (see: Gobec, S. and Urleb, U. *Sci. Synth.* **2004**, *16*, 845-911). White solid, 0.828 g, isolated yield 88%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.28 (bs, 1H), 7.70 (d, $J = 7.5$ Hz, 1H), 7.45 (t, $J = 7.8$ Hz, 1H), 7.28-7.24 (m, 2H), 2.75 (t, $J = 7.5$ Hz, 2H), 1.75-1.69 (m, 2H), 0.95 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 161.7, 154.6, 131.7, 131.6, 129.3, 128.1, 123.0, 115.2, 34.7, 19.4, 13.9. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{13}\text{N}_2\text{O}^+$ ($[\text{M}-\text{H}]^+$) requires m/z 189.10224, found m/z 189.10223.



3-butylquinoxalin-2(1H)-one (1d): Known compound (see: Kalinin, A. A.; Mamedov, V. A. *Russian J. Org. Chem.* **2009**, *45*, 1098-1101). White solid, 0.960 g, isolated yield 95%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.29 (bs, 1H), 7.71 (d, $J = 8.0$ Hz, 1H), 7.46 (t, $J = 7.5$ Hz, 1H), 7.28-7.24 (m, 2H), 2.77 (t, $J = 7.8$ Hz, 2H), 1.70-1.64 (m, 2H), 1.41-1.34 (m, 2H), 0.91 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 161.9, 154.6, 131.7, 131.7, 129.3, 128.0, 123.0, 115.2, 32.4, 28.2, 22.1, 13.8. HRMS-ESI exact mass calcd. for $\text{C}_{12}\text{H}_{13}\text{N}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 201.10334, found m/z 201.10263.

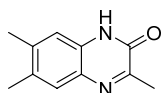


3-isopropylquinoxalin-2(1H)-one (1e): Known compound (see: Briguglio, I.; Piras, S.; Corona, P.; Pirisi, M. A.; Burrari, L.; Boatto, G.; Gavini, E.; Rassu, G. *J. Heterocycl. Chem.* **2016**, *53*, 1721-1737). White solid, 0.818 g, isolated yield 87%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.30 (bs, 1H), 7.71 (d, $J = 8.0$ Hz, 1H), 7.47-7.44 (m, 1H), 7.28-7.24 (m, 2H), 3.48-3.43 (m, 1H), 1.21 (d, $J = 6.5$ Hz, 6H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 165.5, 154.2, 131.7, 131.5, 129.4, 128.2, 123.0, 115.2, 29.9, 20.0. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{11}\text{N}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 187.08769, found m/z 187.08681.

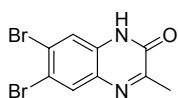


3-cyclohexylquinoxalin-2(1H)-one (1f): Known compound (see: Lian, F.; Xu, K.; Meng, W.; Zhang, H.; Tan, Z.; Zeng, C. *Chem. Commun.* **2019**, *55*, 14685-14688). White solid, 1.061 g, isolated yield 93%. ^1H NMR (400 MHz,

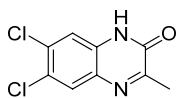
DMSO-*d*₆): δ (ppm) 12.30 (bs, 1H), 7.70 (d, $J = 8.4$ Hz, 1H), 7.46 (t, $J = 7.6$ Hz, 1H), 7.27-7.23 (m, 2H), 3.17 (t, $J = 11.0$ Hz, 1H), 1.88-1.69 (m, 5H), 1.49-1.18 (m, 5H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ (ppm) 164.8, 154.2, 131.7, 131.5, 129.3, 128.2, 123.0, 115.1, 39.4, 30.0, 25.8, 25.8. HRMS-ESI exact mass calcd. for C₁₄H₁₅N₂O⁻ ([M-H]⁻) requires m/z 227.11899, found m/z 227.11838.



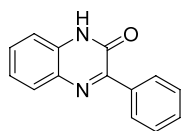
3,6,7-trimethylquinoxalin-2(1H)-one (1h): Known compound (see: unez-Rico, J. L.; Vidal-Ferran, A. *Org. Lett.* **2013**, *15*, 2066-2069). Pale yellow solid, 0.865 g, isolated yield 92%. ¹H NMR (500 MHz, DMSO-*d*₆): δ (ppm) 12.15 (bs, 1H), 7.45 (s, 1H), 7.02 (s, 1H), 2.36 (s, 3H), 2.28-2.26 (m, 6H); ¹³C NMR (125 MHz, DMSO-*d*₆): δ (ppm) 157.7, 155.0, 138.6, 131.5, 130.2, 129.9, 127.8, 115.3, 20.4, 19.7, 18.9. HRMS-ESI exact mass calcd. for C₁₁H₁₃N₂O⁺ ([M-H]⁺) requires m/z 189.10224, found m/z 189.10223.



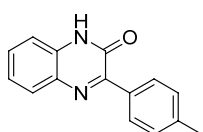
6,7-dibromo-3-methylquinoxalin-2(1H)-one (1i): (New compound). White solid, m.p. 290-292 °C, 1.279 g, isolated yield 81%. ¹H NMR (500 MHz, DMSO-*d*₆): δ (ppm) 12.38 (bs, 1H), 8.00 (s, 1H), 7.54 (s, 1H), 2.38 (s, 3H); ¹³C NMR (125 MHz, DMSO-*d*₆): δ (ppm) 161.4, 154.4, 132.3, 131.8, 131.7, 123.9, 119.3, 116.5, 20.6. HRMS-ESI exact mass calcd. for C₉H₅Br₂N₂O⁻ ([M-H]⁻) requires m/z 314.87741, found m/z 314.87659.



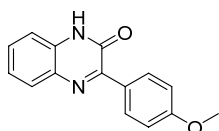
6,7-dichloro-3-methylquinoxalin-2(1H)-one (1j): Known compound (see: Mondieig, D.; Negrier, P.; Massip, S.; Leger, J. M.; Jarmoumi, C.; Lakhrissi, B. *J. Phy. Org. Chem.* **2011**, *24*, 1193-1200). White solid, 0.946 g, isolated yield 83%. ¹H NMR (300 MHz, DMSO-*d*₆): δ (ppm) 12.42 (bs, 1H), 7.93 (s, 1H), 7.40 (s, 1H), 2.39 (s, 3H); ¹³C NMR (75 MHz, DMSO-*d*₆): δ (ppm) 161.4, 154.5, 131.9, 131.2, 131.1, 128.9, 124.7, 116.2, 20.6. HRMS-ESI exact mass calcd. for C₉H₅Cl₂N₂O⁻ ([M-H]⁻) requires m/z 226.97844, found m/z 226.97760.



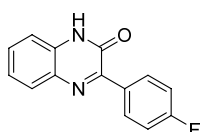
3-phenylquinoxalin-2(1H)-one (1k): Known compound (see: Rueping, M.; Tato, F.; Schoepke, F. R. *Chem. Eur. J.* **2010**, *16*, 2688-2691). Yellow solid, 0.966 g, isolated yield 87%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.57 (bs, 1H), 8.31-8.29 (m, 2H), 7.84 (d, $J = 7.5$ Hz, 1H), 7.56-7.47 (m, 4H), 7.35-7.31 (m, 2H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 154.6, 154.1, 135.6, 132.1, 132.0, 130.3, 130.2, 129.2, 128.8, 127.9, 123.4, 115.1. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_9\text{N}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 221.07204, found m/z 221.07145.



3-(*p*-tolyl)quinoxalin-2(1H)-one (1l): Known compound (see: unez-Rico, J. L.; Vidal-Ferran, A. *Org. Lett.* **2013**, *15*, 2066-2069). Yellow solid, 0.992 g, isolated yield 84%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.53 (bs, 1H), 8.26 (d, $J = 8.0$ Hz, 2H), 7.81 (d, $J = 7.5$ Hz, 2H), 7.52 (t, $J = 7.3$ Hz, 1H), 7.33-7.28 (m, 4H), 2.38 (s, 3H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 154.6, 153.8, 140.1, 132.9, 132.0, 131.9, 130.1, 129.2, 128.6, 128.5, 123.3, 115.0, 21.1. HRMS-ESI exact mass calcd. for $\text{C}_{15}\text{H}_{11}\text{N}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 235.08659, found m/z 235.08684.

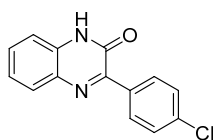


3-(4-methoxyphenyl)quinoxalin-2(1H)-one (1m): Known compound (see: Rueping, M.; Tato, F.; Schoepke, F. R. *Chem. Eur. J.* **2010**, *16*, 2688-2691). Yellow solid, 1.122 g, isolated yield 89%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.50 (bs, 1H), 8.40 (d, $J = 9.0$ Hz, 2H), 7.80 (d, $J = 8.0$ Hz, 1H), 7.52-7.49 (m, 1H), 7.33-7.29 (m, 2H), 7.05 (d, $J = 8.5$ Hz, 2H), 3.84 (s, 3H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 161.0, 154.7, 153.1, 132.1, 131.8, 131.0, 129.7, 128.5, 128.1, 123.3, 115.0, 113.3, 55.3. HRMS-ESI exact mass calcd. for $\text{C}_{15}\text{H}_{11}\text{N}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 251.08260, found m/z 251.08179.



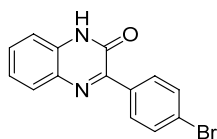
3-(4-fluorophenyl)quinoxalin-2(1H)-one (1n): Known compound (see: Yuan, J. W.; Liu, S. N.; Qu, L. B. *Adv. Synth. Catal.* **2017**, *359*, 4197-4207). Yellow solid, 0.890 g, isolated yield 74%. ^1H NMR (300 MHz, DMSO- d_6): δ (ppm) 12.61 (bs, 1H), 8.42 (dd, $J = 6.0$ Hz, $J = 8.7$ Hz, 2H), 7.84 (d, $J = 7.8$ Hz, 1H), 7.55 (t,

$J = 7.5$ Hz, 1H), 7.36-7.30 (m, 4H), ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 163.3 (d, $J_{\text{C-F}} = 246.3$ Hz), 154.6, 152.8, 132.1 (d, $J_{\text{C-F}} = 2.5$ Hz), 132.0, 131.9, 131.7 (d, $J_{\text{C-F}} = 8.7$ Hz), 130.3, 128.7, 123.4, 115.1, 114.8 (d, $J_{\text{C-F}} = 21.2$ Hz). HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_{10}\text{FN}_2\text{O}^+$ $[\text{M}+\text{H}]^+$ requires m/z 241.07717, found m/z 241.07640.



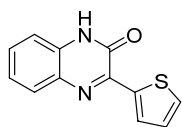
3-(4-chlorophenyl)quinoxalin-2(1H)-one (1o): Known compound (see: Nagaraj, M.; Sathiyamoorthy, S.; Boominathan, M.; Muthusubramanian, S.; Bhuvanesh, N. *J. Heterocycl. Chem.* **2013**, *50*, 1146-1151). Yellow solid,

1.191 g, isolated yield 93%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.62 (bs, 1H), 8.37 (d, $J = 8.5$ Hz, 2H), 7.83 (d, $J = 8.0$ Hz, 1H), 7.56-7.53 (m, 3H), 7.34-7.31 (m, 2H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 154.5, 152.7, 135.1, 134.4, 132.1, 131.9, 131.0, 130.5, 128.8, 128.0, 123.5, 115.1. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_8\text{ClN}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 255.03306, found m/z 255.03224.



3-(4-bromophenyl)quinoxalin-2(1H)-one (1p): Known compound (see: Nagaraj, M.; Sathiyamoorthy, S.; Boominathan, M.; Muthusubramanian, S.; Bhuvanesh, N. *J. Heterocycl. Chem.* **2013**, *50*, 1146-1151). Yellow

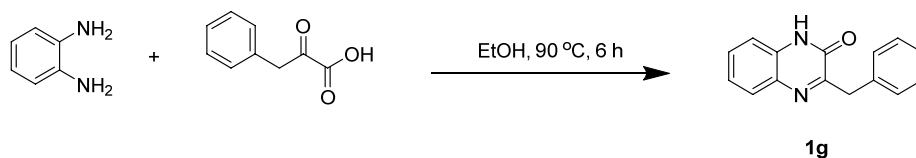
solid, 1.365 g, isolated yield 91%. ^1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.63 (bs, 1H), 8.31 (d, $J = 9.0$ Hz, 2H), 7.84 (d, $J = 8.5$ Hz, 1H), 7.70 (d, $J = 9.0$ Hz, 2H), 7.56 (t, $J = 7.8$ Hz, 1H), 7.35-7.32 (m, 2H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 155.0, 153.3, 135.2, 132.6, 132.4, 131.7, 131.4, 131.1, 129.3, 124.5, 124.0, 115.6. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_8\text{BrN}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 298.98255, found m/z 298.98181.



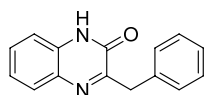
3-(thiophen-2-yl)quinoxalin-2(1H)-one (1q): Known compound (see: Xue, Z.-Y.; Jiang, Y.; Peng, X.-Z.; Yuan, W.-C.; Zhang, X.-M. *Adv. Synth. Catal.* **2010**, *352*, 2132-2136). Yellow solid, 0.775 g, isolated yield 68%. ^1H NMR

(500 MHz, DMSO- d_6): δ (ppm) 12.71 (bs, 1H), 8.41-8.40 (m, 1H), 7.83 (d, $J = 5.0$ Hz, 1H), 7.72 (d, $J = 7.5$ Hz, 1H), 7.51 (t, $J = 7.8$ Hz, 1H), 7.35-7.30 (m, 2H), 7.23 (t, $J = 4.3$ Hz, 1H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 153.5, 148.9, 139.0, 132.1, 131.9, 131.5, 131.4,

129.8, 128.2, 128.1, 123.7, 115.3. HRMS-ESI exact mass calcd. for $C_{12}H_7N_2OS^-$ ($[M-H]^-$) requires m/z 227.02846, found m/z 227.02803.

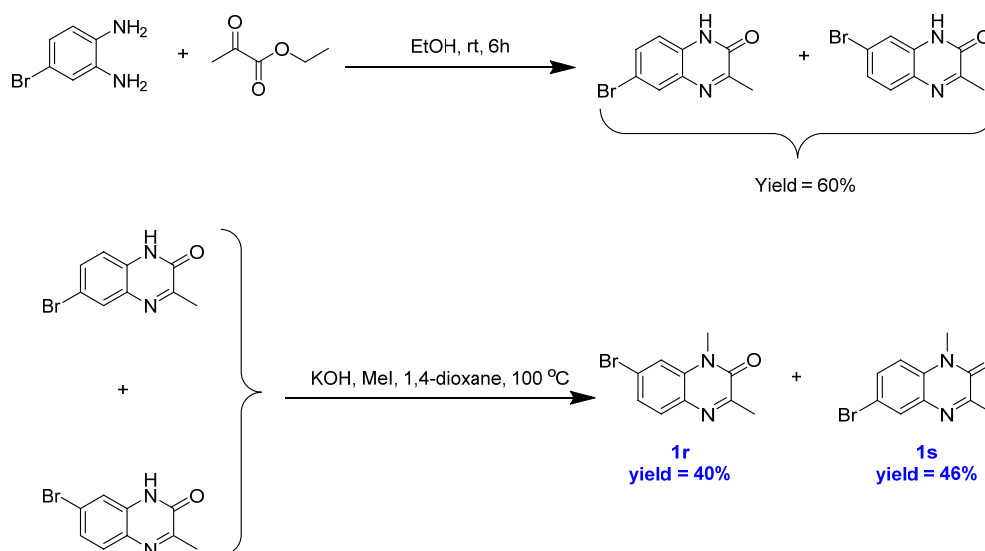


General procedure: In a 250 mL flask, 1,2-diaminobenzene (0.540 g, 5.0 mmol) was dissolved in ethyl alcohol (100 mL). Then, 3-phenylpyruvic acid (1.806 g, 11.0 mmol) was added, and the reaction mixture was stirred at 90 °C for 6h. The reaction mixture was cooled to room temperature, the crude product was recrystallized by ethyl alcohol to afford benzylquinoxalin-2(1H)-one (**1g**) as pale yellow solid. The analytical data of the products are summarized below.



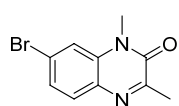
3-benzylquinoxalin-2(1H)-one (1g): Known compound (see: Piras, S.; Loriga, M.; Carta, A.; Paglietti, G.; Costi, M. P.; Ferrari, S. *J. Heterocycl. Chem.* **2006**, *43*, 541-548). Pale yellow solid, 1.003 g, isolated yield 85%.

1H NMR (500 MHz, DMSO- d_6): δ (ppm) 12.38 (bs, 1H), 7.71 (d, $J = 8.0$ Hz, 1H), 7.48-7.44 (m, 1H), 7.34-7.17 (m, 7H), 4.12 (s, 2H); ^{13}C NMR (125 MHz, DMSO- d_6): δ (ppm) 160.3, 154.5, 137.4, 132.0, 131.6, 129.7, 129.1, 128.3, 128.2, 126.3, 123.1, 115.3, 39.0. HRMS-ESI exact mass calcd. for $C_{15}H_{11}N_2O^-$ ($[M-H]^-$) requires m/z 235.08769, found m/z 235.08710.



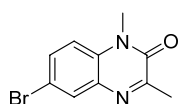
General procedure: In a 250 mL flask, 4-bromo-1,2-benzenediamine (0.540 g, 5.000 mmol) was dissolved in ethyl alcohol (100 mL). Then, ethyl pyruvate (1.277 g, 11.0 mmol) was added, and the reaction mixture was stirred at room temperature until the reaction was completed which was monitored by TLC. Then, the mixture was filtered to afford 7-bromo-3-methylquinoxalin-2(1*H*)-one and 6-bromo-3-methylquinoxalin-2(1*H*)-one (0.717 g, 3.0 mmol).

In a 250 mL flask, 7-bromo-3-methylquinoxalin-2(1*H*)-one and 6-bromo-3-methylquinoxalin-2(1*H*)-one (0.717 g, 3.0 mmol) were dissolved in 1,4-dioxane (100 mL). Potassium hydroxide (0.336 mg, 6.0 mmol) were added and the reaction mixture was stirred at 100 °C for 5 min. The reaction mixture was cooled to room temperature and iodomethane (0.511 g, 3.6 mmol) was added slowly and the reaction mixture was stirred at 100 °C until the reaction was completed which was monitored by TLC. Then, the solvent was removed under reduced pressure. The crude product was purified by flash column chromatography on silica gel (eluent: ethyl acetate/petroleum ether) to yield the desired 7-bromo-1,3-dimethylquinoxalin-2(1*H*)-one (**1q**) and 6-bromo-1,3-dimethylquinoxalin-2(1*H*)-one (**1r**) as white solid. The analytical data of the products are summarized below.



7-bromo-1,3-dimethylquinoxalin-2(1*H*)-one (1r): Known compound (see: Zhao, Z.-B.; Li, X.; Chen, M.-W.; Zhao Z. K.; Zhou, Y.-G. *Chem. Commun.*

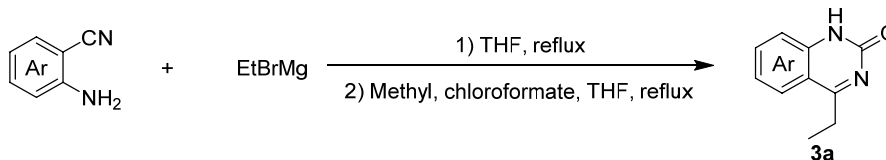
2020, 56, 7309-7312). White solid, 0.152 g, isolated yield 40%. ^1H NMR (500 MHz, CDCl_3): δ (ppm) 7.63 (d, $J = 8.5$ Hz, 1H), 7.43-7.41 (m, 2H), 3.65 (s, 3H), 2.56 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 158.9, 154.9, 134.4, 131.5, 130.8, 127.0, 123.7, 116.8, 29.3, 21.7. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{10}\text{BrN}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 252.99710, found m/z 252.99701.



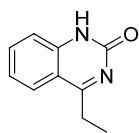
6-bromo-1,3-dimethylquinoxalin-2(1H)-one (1s): Known compound (see: Zhao, Z.-B.; Li, X.; Chen, M.-W.; Zhao Z. K.; Zhou, Y.-G. *Chem. Commun.*

2020, 56, 7309-7312). White solid, 0.175 g, isolated yield 46%. ^1H NMR (500 MHz, CDCl_3): δ (ppm) 7.95 (s, 1H), 7.61-7.59 (m, 1H), 7.16 (d, $J = 9.0$ Hz, 1H), 3.67 (s, 3H), 2.59 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 160.0, 155.0, 133.6, 132.5, 132.5, 132.0, 116.3, 115.2, 29.3, 21.8. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{10}\text{BrN}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 252.99710, found m/z 252.99699.

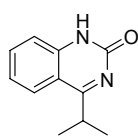
3.2 Synthesis of substrate quinazolin-2(1H)-ones (3)



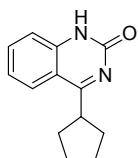
General procedure: In a 250 mL flask, 2-aminobenzonitrile (0.118 g, 1.0 mmol) was dissolved in dry tetrahydrofuran (100 mL). Then, a solution of ethylmagnesium bromide (0.7 mL, 2.0 mmol) was added slowly. After refluxing for 2 h, methyl chloroformate (0.142 g, 1.5 mol) was added dropwise at room temperature, and the resulting mixture was refluxed until completed which was monitored by TLC. The mixture was cooled to room temperature and poured into a hydrochloric acid solution (2 M). The mixture was neutralized with 10% sodium bicarbonate aqueous solution and extracted with dichloromethane (30 mL \times 3). The combined organic layer was dried over anhydrous sodium sulfate, concentrated in vacuo. The crude product was purified by flash column chromatography on silica gel (eluent: ethyl acetate/petroleum ether) to yield the desired ethylquinazolin-2(1H)-one (**3a**) as yellow solid



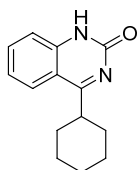
4-ethylquinazolin-2(1H)-one (3a): Known compound (see: Bergman, J.; Brynolf, A.; Elman, B.; Vuorinen, E. *Tetrahedron* **1986**, *42*, 3697-3706). Yellow solid, 0.117 g, isolated yield 67%. ^1H NMR (400 MHz, CD_2Cl_2): δ (ppm) 12.51 (bs, 1H), 7.93-7.91 (m, 1H), 7.70-7.66 (m, 1H), 7.45 (d, $J = 8.0$ Hz, 1H), 7.30-7.26 (m, 1H), 3.18 (q, $J = 7.3$ Hz, 2H), 1.41 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (100 MHz, CD_2Cl_2): δ (ppm) 181.0, 158.4, 142.4, 135.3, 126.2, 123.3, 116.7, 116.0, 28.9, 11.5. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{11}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 175.08659, found m/z 175.08510.



4-isopropylquinazolin-2(1H)-one (3b): Known compound (see: Bergman, J.; Brynolf, A.; Elman, B.; Vuorinen, E. *Tetrahedron* **1986**, *42*, 3697-3706). White solid, 0.122 g, isolated yield 65%. ^1H NMR (400 MHz, CD_2Cl_2): δ (ppm) 12.80 (bs, 1H), 7.97 (d, $J = 8.0$ Hz, 1H), 7.72-7.67 (m, 1H), 7.51 (d, $J = 8.0$ Hz, 1H), 7.29 (t, $J = 7.6$ Hz, 1H), 3.79-3.72 (m, 1H), 1.39 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (100 MHz, CD_2Cl_2): δ (ppm) 184.7, 158.7, 142.8, 135.3, 126.0, 123.4, 117.1, 115.5, 32.3, 21.4. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{13}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 189.10224, found m/z 189.10063.

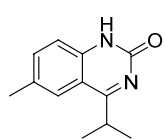


4-cyclopentylquinazolin-2(1H)-one (3c): (New compound). Pale yellow solid, m.p. 199-201 $^\circ\text{C}$, 0.124 g, isolated yield 58%. ^1H NMR (300 MHz, CDCl_3): δ (ppm) 12.97 (bs, 1H), 7.96 (d, $J = 8.1$ Hz, 1H), 7.66 (t, $J = 7.5$ Hz, 1H), 7.57 (d, $J = 7.8$ Hz, 1H), 7.29-7.24 (m, 1H), 3.90-3.79 (m, 1H), 2.23-2.07 (m, 4H), 1.97-1.85 (m, 2H), 1.80-1.73 (m, 2H); ^{13}C NMR (125 MHz, CD_2Cl_2): δ (ppm) 183.3, 158.8, 142.3, 134.9, 126.1, 123.1, 117.0, 116.2, 43.5, 32.3, 26.4. HRMS-ESI exact mass calcd. for $\text{C}_{13}\text{H}_{15}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 215.11789, found m/z 215.11811.

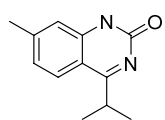


4-cyclohexylquinazolin-2(1H)-one (3d): Known compound (see: Milton, L. H.; Ann, H. US 3305553, **1967**). White solid, 0.162 g, isolated yield 71%. ^1H NMR (400 MHz, CD_2Cl_2): δ (ppm) 12.92 (s, 1H), 7.97-7.29 (m, 4H), 3.43-3.37 (m, 1H), 1.93-1.38 (m, 10H); ^{13}C NMR (100 MHz, CD_2Cl_2): δ (ppm) 183.7, 158.6, 142.6, 135.0, 125.8, 123.1, 116.9, 115.3, 42.4, 31.7, 26.6, 26.3. HRMS-ESI exact mass calcd.

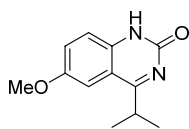
for $C_{14}H_{17}N_2O^+$ ($[M+H]^+$) requires m/z 229.13354, found m/z 229.13296.



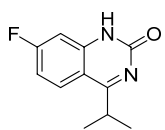
4-isopropyl-6-methylquinazolin-2(1H)-one (3e): (New compound). Pale yellow solid, m.p. 188-190 °C, 0.131 g, isolated yield 65%. 1H NMR (400 MHz, CD_2Cl_2): δ (ppm) 12.92 (bs, 1H), 7.74 (s, 1H), 7.53 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.41 (d, $J = 8.4$ Hz, 1H), 3.79-3.69 (m, 1H), 2.45 (s, 3H), 1.38 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (100 MHz, CD_2Cl_2): δ (ppm) 184.2, 158.8, 140.9, 136.8, 133.1, 125.3, 116.9, 115.4, 32.2, 21.4, 21.3. HRMS-ESI exact mass calcd. for $C_{12}H_{15}N_2O^+$ ($[M+H]^+$) requires m/z 203.11789, found m/z 203.11914.



7-isopropyl-6-methylquinazolin-2(1H)-one (3f): (New compound). Pale yellow solid, m.p. 186-188 °C, 0.126 g, isolated yield 63%. 1H NMR (400 MHz, $CDCl_3$): δ (ppm) 13.32 (bs, 1H), 7.80 (d, $J = 8.4$, 1H), 7.42 (s, 1H), 7.06 (d, $J = 8.4$ Hz, 1H), 3.73-3.66 (m, 1H), 2.44 (s, 3H), 1.40 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ (ppm) 184.1, 159.3, 146.7, 142.8, 125.3, 124.9, 116.8, 113.4, 31.9, 21.9, 21.3. HRMS-ESI exact mass calcd. for $C_{12}H_{13}N_2O^-$ ($[M-H]^-$) requires m/z 201.10334, found m/z 201.10278.

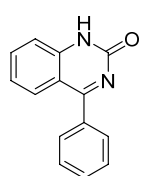


6-methoxy-4-isopropylquinazolin-2(1H)-one (3g): (New compound). Pale yellow solid, m.p. 198-200 °C, 0.122 g, isolated yield 56%. 1H NMR (400 MHz, $CDCl_3$): δ (ppm) 13.36 (bs, 1H), 7.56 (d, $J = 8.8$, 1H), 7.34-7.28 (m, 2H), 3.87 (s, 3H), 3.71-3.64 (m, 1H), 1.43 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ (ppm) 183.5, 158.8, 155.5, 137.5, 125.0, 118.6, 115.6, 106.4, 56.0, 32., 21.2. HRMS-ESI exact mass calcd. for $C_{12}H_{13}N_2O_2^-$ ($[M-H]^-$) requires m/z 217.09825, found m/z 217.09796.



7-fluoro-4-isopropylquinazolin-2(1H)-one (3h): (New compound). Pale yellow solid, m.p. 206-208 °C, 0.128 g, isolated yield 62%. 1H NMR (400 MHz, $CDCl_3$): δ (ppm) 13.35 (bs, 1H), 7.97 (dd, $J = 5.6, J = 9.2$, 1H), 7.31-7.28 (m, 1H), 7.03-6.98 (m, 1H), 3.73-3.66 (m, 1H), 1.43 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$): δ (ppm) 183.5, 158.8, 155.5, 137.5, 125.0, 118.6, 115.6, 106.4, 56.0, 32., 21.2.

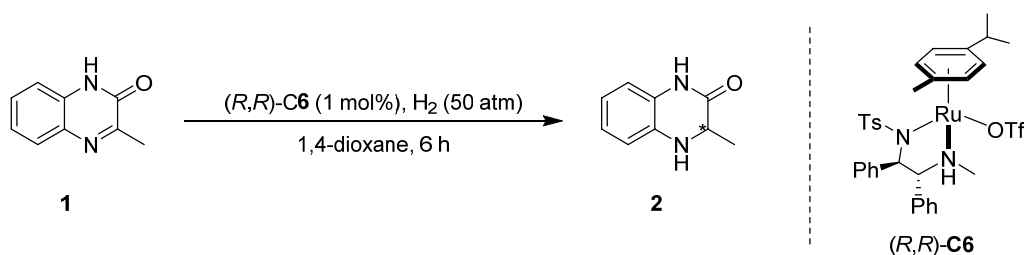
CDCl₃): δ (ppm) 184.1, 166.7 (d, J_{C-F} = 256.0 Hz), 159.1, 144.6 (d, J_{C-F} = 13.0 Hz), 128.5 (d, J_{C-F} = 11.0 Hz), 112.3 (d, J_{C-F} = 24.0 Hz), 112.3 (d, J_{C-F} = 2.0 Hz), 103.1 (d, J_{C-F} = 25.0 Hz),. HRMS-ESI exact mass calcd. for C₁₁H₁₀FN₂O⁻ ([M-H]⁻) requires m/z 205.07826, found m/z 205.07782.



4-phenylquinazolin-2(1H)-one (3i): Known compound (see: Bergman, J.; Brynolf, A.; Elman, B.; Vuorinen, E. *Tetrahedron* **1986**, *42*, 3697-3706). White solid, 0.140 g, isolated yield 63%. ¹H NMR (400 MHz, CD₂Cl₂): δ (ppm) 12.83 (bs, 1H), 7.86-7.71 (m, 4H), 7.60-7.58 (m, 4H), 7.25 (t, J = 7.2 Hz, 1H); ¹³C NMR (100 MHz, CD₂Cl₂): δ (ppm) 177.0, 158.3, 143.8, 137.1, 135.7, 131.1, 130.0, 129.2, 128.8, 123.4, 116.9, 115.8. HRMS-ESI exact mass calcd. for C₁₄H₁₁N₂O⁺ ([M+H]⁺) requires m/z 223.08659, found m/z 223.08466.

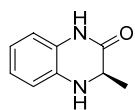
4. General procedure for asymmetric hydrogenation

4.1 Asymmetric hydrogenation of substrate quinoxalin-2(1H)-ones (1)



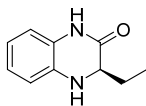
General procedure: A 15 mL glass-lined stainless-steel reactor equipped with a magnetic stirrer bar was charged with substrate **1** (0.2 mmol), Ru-catalyst (*R,R*)-**C6** (0.002 mmol) in 2.0 mL of solvent under N₂ atmosphere in a glove box. The autoclave was closed, and the final pressure of the hydrogen gas was adjusted to 50 atm after purging the autoclave with hydrogen gas several times. The reaction mixture was stirred at 25 °C for 6 h. Then the hydrogen gas was carefully released and the conversion was determined by ¹H NMR

spectroscopy. The reaction mixture was filtered through a short pad of silica eluted with ethyl acetate and petroleum ether to give the chiral product 3,4-dihydroquinoxalin-2(1*H*)-one (**2**). The enantiomeric excess of the product was determined by HPLC with a chiral column.



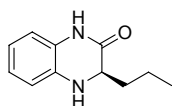
(R)-3-methyl-3,4-dihydroquinoxalin-2(1*H*)-one (2a): Known compound (see: Li, D.; Ollevier, T. *Eur. J. Org. Chem.* **2019**, *6*, 1273-1280). White solid, m.p. 114-116 °C, 31.8 mg, isolated yield 98%, 98% *ee*. $[\alpha]_D^{20} = -77.6$ ($c = 1.0$, CDCl_3 , 98% *ee*); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 9.20 (bs, 1H), 6.90-6.66 (m, 4H), 4.02 (q, $J = 6.2$ Hz, 1H), 3.88 (s, 1H), 1.46 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 169.9, 133.6, 125.8, 123.9, 119.7, 115.7, 114.2, 52.0, 18.0. HRMS-ESI exact mass calcd. for $\text{C}_9\text{H}_{11}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 163.08659, found m/z 163.08650.

The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm), $t_{\text{R}1} = 17.9$ min (minor), $t_{\text{R}2} = 19.1$ min (major).



(R)-3-ethyl-3,4-dihydroquinoxalin-2(1*H*)-one (2b): Known compound (see: Pan, Y.; Chen, C.; Xu, X.; Zhao, H.; Han, J.; Li, H.; Xu, L.; Fan, Q.; Xiao, J. *Green Chem.* **2018**, *20*, 403-411). Pale yellow oil, 34.2 mg, isolated yield 97%, 98% *ee*. $[\alpha]_D^{20} = -54.0$ ($c = 1.0$, CDCl_3 , 98% *ee*); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 8.68 (bs, 1H), 6.91-6.84 (m, 1H), 6.77-6.67 (m, 3H), 3.96 (s, 1H), 3.87 (q, $J = 5.1$ Hz, 1H), 1.95-1.72 (m, 2H), 1.04 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ (ppm) 169.1, 133.2, 125.4, 124.0, 119.4, 115.5, 114.2, 57.7, 25.2, 9.8. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{11}\text{N}_2\text{O}^+$ ($[\text{M}_- \text{H}]^+$) requires m/z 175.08659, found m/z 175.08659.

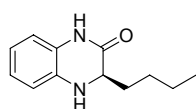
The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 220$ nm), $t_{\text{R}1} = 8.0$ min (minor), $t_{\text{R}2} = 9.4$ min (major).



(R)-3-propyl-3,4-dihydroquinoxalin-2(1*H*)-one (2c): Known compound (see: Kamila, S.; Biehl, E. R. *Heterocycles* **2006**, *68*, 1931-1939). colourless oil,

34.2 mg, isolated yield 90%, 97% *ee*. $[\alpha]_{\text{D}}^{20} = -36.0$ ($c = 1.0$, CDCl_3 , 97% *ee*); ^1H NMR (500 MHz, CDCl_3): δ (ppm) 9.19 (s, 1H), 6.88 (t, $J = 7.3$ Hz, 4H), 6.78-6.73 (m, 2H), 6.67 (t, $J = 8.0$ Hz, 1H), 3.97-3.91 (m, 2H), 1.85-1.70 (m, 2H), 1.55-1.41 (m, 2H), 0.96 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 169.6, 133.1, 125.5, 123.9, 119.4, 115.6, 114.2, 56.3, 34.1, 18.7, 13.9. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 191.11789, found m/z 191.11806.

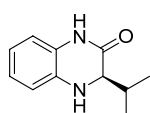
The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{\text{R}1} = 14.8$ min (minor), $t_{\text{R}2} = 19.2$ min (major).



(R)-3-butyl-3,4-dihydroquinoxalin-2(1H)-one (2d): Known compound (see: Kamila, S.; Biehl, E. R. *Heterocycles* **2006**, *68*, 1931-1939). Pale yellow oil, 37.2 mg, isolated yield 91%, 97% *ee*. $[\alpha]_{\text{D}}^{20} = -22.8$ ($c = 1.0$,

CDCl_3 , 97% *ee*); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 8.80 (bs, 1H), 6.91-6.86 (m, 1H), 6.75-6.66 (m, 3H), 3.95-3.89 (m, 2H), 1.89-1.68 (m, 2H), 1.53-1.26 (m, 4H), 0.91 (t, $J = 6.9$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 169.3, 133.2, 125.5, 123.9, 119.4, 115.5, 114.2, 56.6, 31.7, 27.6, 22.6, 14.1. HRMS-ESI exact mass calcd. for $\text{C}_{12}\text{H}_{15}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 203.11789, found m/z 203.11794.

The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{\text{R}1} = 6.6$ min (major), $t_{\text{R}2} = 7.5$ min (minor).

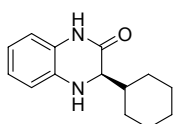


(R)-3-isopropyl-3,4-dihydroquinoxalin-2(1H)-one (2e): Known compound (see: Li, D.; Ollevier, T. *Eur. J. Org. Chem.* **2019**, *6*, 1273-1280). Pale yellow viscous solid, 34.2 mg, isolated yield 90%, 94% *ee*. $[\alpha]_{\text{D}}^{20} = -69.2$ ($c = 1.0$,

CDCl_3 , 94% *ee*); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 8.66 (bs, 1H), 6.90-6.84 (m, 1H), 6.71-6.64 (m, 3H), 4.00 (s, 1H), 3.78-3.76 (m, 1H), 2.30-2.19 (m, 1H), 1.04 (d, $J = 6.9$ Hz, 3H), 0.98 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 168.3, 133.4, 124.9, 124.0, 119.0, 115.3, 113.6, 62.0, 30.9, 19.1, 17.6. HRMS-ESI exact mass calcd. for

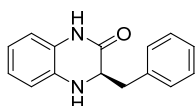
$C_{11}H_{15}N_2O^+$ ($[M+H]^+$) requires m/z 191.11789, found m/z 191.11781.

The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 11.5$ min (minor), $t_{R2} = 16.0$ min (major).



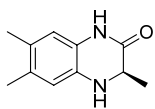
(R)-3-cyclohexyl-3,4-dihydroquinoxalin-2(1H)-one (2f): Known compound (see: Zhang, L.; Qiu, R.; Xue, X.; Pan, Y.; Xu, C.; Li, H.; Xu, L. *Adv. Synth. Catal.* **2015**, 357, 3529-3537). White solid, m.p. 61-63 °C, 45.1 mg, isolated yield 98%, 94% *ee*. $[\alpha]_D^{20} = -28.4$ ($c = 1.0$, $CDCl_3$, 94% *ee*); 1H NMR (300 MHz, $CDCl_3$): δ (ppm) 9.14 (bs, 1H), 6.89-6.84 (m, 1H), 6.75-6.62 (m, 3H), 4.06 (s, 1H), 3.76 (d, $J = 5.4$ Hz, 1H), 1.86-1.63 (m, 6H), 1.28-1.09 (m, 5H); ^{13}C NMR (75 MHz, $CDCl_3$): δ (ppm) 168.4, 133.4, 125.0, 124.0, 118.8, 115.5, 113.5, 61.6, 40.7, 29.6, 27.9, 26.0, 25.7. HRMS-ESI exact mass calcd. for $C_{14}H_{19}N_2O^+$ ($[M+H]^+$) requires m/z 231.14919, found m/z 231.114919.

The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 6.3$ min (major), $t_{R2} = 7.6$ min (minor).



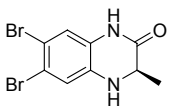
(R)-3-benzyl-3,4-dihydroquinoxalin-2(1H)-one (2g): Known compound (see: Li, D.; Ollevier, T. *Eur. J. Org. Chem.* **2019**, 6, 1273-1280). White viscous solid, 46.2 mg, isolated yield 97%, 94% *ee*. $[\alpha]_D^{20} = 82.0$ ($c = 1.0$, $CDCl_3$, 94% *ee*); 1H NMR (300 MHz, $DMSO-d_6$): δ (ppm) 10.24 (s, 1H), 7.29-7.16 (m, 5H), 6.77-6.54 (m, 4H), 5.82 (s, 1H), 4.01 (t, $J = 5.1$ Hz, 1H), 2.97-2.82 (m, 2H); ^{13}C NMR (75 MHz, $DMSO-d_6$): δ (ppm) 166.9, 137.5, 133.5, 129.6, 128.1, 126.2, 125.7, 122.8, 117.6, 114.6, 113.6, 56.9, 37.4. HRMS-ESI exact mass calcd. for $C_{15}H_{15}N_2O^+$ ($[M+H]^+$) requires m/z 239.11798, found m/z 239.11783.

The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 85 : 15, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 240$ nm), $t_{R1} = 12.2$ min (minor), $t_{R2} = 13.6$ min (major).



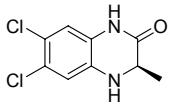
(R)-3,6,7-trimethyl-3,4-dihydroquinoxalin-2(1H)-one (2h): Known compound (see: Pan, Y.; Chen, C.; Xu, X.; Zhao, H.; Han, J.; Li, H.; Xu, L.; Fan, Q.; Xiao, J. *Green Chem.* **2018**, *20*, 403-411). Yellow solid, m.p. 180-182 °C, 33.8 mg, isolated yield 89%, 97% *ee*. $[\alpha]_D^{20} = -115.2$ ($c = 1.0$, CDCl_3 , 97% *ee*); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ (ppm) 8.27 (bs, 1H), 6.52 (s, 1H), 6.49 (s, 1H), 3.95 (q, $J = 6.7$ Hz, 1H), 3.66 (s, 1H), 2.15 (s, 6H), 1.44 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ (ppm) 169.6, 131.9, 131.3, 127.9, 123.6, 116.7, 115.8, 52.4, 19.4, 19.0, 17.9. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 191.11789, found m/z 191.11809.

The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 9.4$ min (minor), $t_{R2} = 11.9$ min (major).



(R)-6,7-dibromo-3-methyl-3,4-dihydroquinoxalin-2(1H)-one (2i): New compound. White solid, m.p. 162-164 °C, 61.0 mg, isolated yield 96%, 97% *ee*. $[\alpha]_D^{20} = -31.2$ ($c = 1.0$, CDCl_3 , 97% *ee*); $^1\text{H NMR}$ (300 MHz, $\text{DMSO}-d_6$): δ (ppm) 10.39 (s, 1H), 6.98 (s, H), 6.97 (s, 1H), 6.43 (s, 1H), 3.89-3.82 (m, 1H), 1.24 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C NMR}$ (75 MHz, $\text{DMSO}-d_6$): δ (ppm) 167.9, 135.4, 127.2, 118.3, 116.9, 115.8, 109.6, 50.4, 17.7. HRMS-ESI exact mass calcd. for $\text{C}_9\text{H}_7\text{Br}_2\text{N}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 316.89306, found m/z 316.89236.

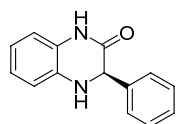
The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 15.5$ min (minor), $t_{R2} = 18.2$ min (major).



(R)-6,7-dichloro-3-methyl-3,4-dihydroquinoxalin-2(1H)-one (2j): Known compound (see: Krchnak, V.; Smith, J.; Vagner, J. *Collect. Czech. Chem. Commun.* **2001**, *66*, 1078-1106). White solid, m.p. 190-192 °C, 42.8 mg, isolated yield 93%, 98% *ee*. $[\alpha]_D^{20} = -52.8$ ($c = 1.0$, CDCl_3 , 98% *ee*); $^1\text{H NMR}$ (300 MHz, $\text{DMSO}-d_6$): δ (ppm) 10.40 (bs, 1H), 6.85 (s, 1H), 6.82 (s, 1H), 6.42 (s, 1H), 3.85 (q, $J = 6.4$ Hz, 1H), 1.25 (d, $J = 6.9$ Hz, 3H); $^{13}\text{C NMR}$ (75 MHz, $\text{DMSO}-d_6$): δ (ppm) 167.9, 134.8,

126.5, 123.8, 118.3, 115.3, 113.8, 50.4, 17.7. HRMS-ESI exact mass calcd. for $C_9H_7Cl_2N_2O^-$ ($[M-H]^-$) requires m/z 228.99409, found m/z 228.99327.

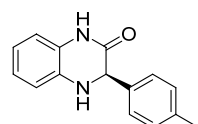
The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 10.9$ min (minor), $t_{R2} = 12.1$ min (major).



(R)-3-phenyl-3,4-dihydroquinoxalin-2(1H)-one (2k): Known compound (see:

Li, D.; Ollevier, T. *Eur. J. Org. Chem.* **2019**, *6*, 1273-1280). White solid, m.p. 150-152 °C, 41.7 mg, isolated yield 93%, 97% *ee*. $[\alpha]_D^{20} = -110.0$ ($c = 1.0$, $CDCl_3$, 97% *ee*); 1H NMR (500 MHz, $CDCl_3$): δ (ppm) 8.91 (bs, 1H), 7.42-7.41 (m, 2H), 7.35-7.29 (m, 3H), 6.93-6.90 (m, 1H), 6.77-6.69 (m, 3H), 5.07 (s, 1H), 4.30 (s, 1H); ^{13}C NMR (125 MHz, $CDCl_3$): δ (ppm) 167.3, 139.2, 133.0, 128.9, 128.6, 127.3, 124.9, 124.2, 119.5, 115.8, 113.8, 60.8. HRMS-ESI exact mass calcd. for $C_{14}H_{13}N_2O^+$ ($[M+H]^+$) requires m/z 225.10224, found m/z 225.10225.

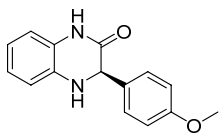
The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 210$ nm), $t_{R1} = 15.8$ min (minor), $t_{R2} = 27.1$ min (major).



(R)-3-(p-tolyl)-3,4-dihydroquinoxalin-2(1H)-one (2l): Known compound

(see: unez-Rico, J. L.; Vidal-Ferran, A. *Org. Lett.* **2013**, *15*, 2066-2069). Yellow solid, m.p. 143-145 °C, 45.2 mg, isolated yield 95%, 97% *ee*. $[\alpha]_D^{20} = -128.8$ ($c = 1.0$, $CDCl_3$, 97% *ee*); 1H NMR (300 MHz, $CDCl_3$): δ (ppm) 9.44 (bs, 1H), 7.26 (d, $J = 7.8$ Hz, 2H), 7.09 (d, $J = 7.8$ Hz, 2H), 6.88-6.83 (m, 1H), 6.69-6.61 (m, 3H), 4.97 (s, 1H), 4.27 (s, 1H), 2.29 (s, 3H); ^{13}C NMR (75 MHz, $CDCl_3$): δ (ppm) 167.8, 138.3, 136.3, 133.1, 129.6, 127.2, 124.9, 124.0, 119.3, 115.9, 113.7, 60.4, 21.2. HRMS-ESI exact mass calcd. for $C_{15}H_{15}N_2O^+$ ($[M+H]^+$) requires m/z 239.11789, found m/z 239.11784.

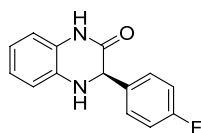
The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 19.8$ min (minor), $t_{R2} = 22.4$ min (major).



(R)-3-(4-methoxyphenyl)-3,4-dihydroquinoxalin-2(1H)-one (2m):

Known compound (see: Rueping, M.; Tato, F.; Schoepke, F. R. *Chem. Eur. J.* **2010**, *16*, 2688-2691). White solid, m.p. 124-126 °C, 46.8 mg, isolated yield 92%, 96% *ee*. $[\alpha]_D^{20} = -124.0$ ($c = 1.0$, CDCl_3 , 96% *ee*); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ (ppm) 8.78 (bs, 1H), 7.32 (d, $J = 8.4$ Hz, 2H), 6.93-6.84 (m, 3H), 6.77-6.67 (m, 3H), 5.01 (s, 1H), 4.25 (s, 1H), 3.77 (s, 3H); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ (ppm) 167.6, 159.8, 133.2, 131.3, 128.6, 125.0, 124.2, 119.5, 115.7, 114.4, 113.9, 60.3, 55.4. HRMS-ESI exact mass calcd. for $\text{C}_{15}\text{H}_{14}\text{N}_2\text{O}_2^+$ ($[\text{M}+\text{H}]^+$) requires m/z 255.11280, found m/z 255.11299.

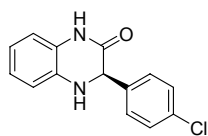
The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{R1} = 28.5$ min (major), $t_{R2} = 35.6$ min (minor).



(R)-3-(4-fluorophenyl)-3,4-dihydroquinoxalin-2(1H)-one (2n): Known

compound (see: Ogino, E.; Nakamura, A.; Kuwano, S.; Arai, T. *Org. Lett.* **2021**, *23*, 1980-1985). White solid, m.p. 174-176 °C, 45.5mg, isolated yield 94%, 96% *ee*. $[\alpha]_D^{20} = -132.0$ ($c = 1.0$, CDCl_3 , 96% *ee*); $^1\text{H NMR}$ (400 MHz, $\text{CDCl}_3/\text{Methanol-}d_4 = 10:1$): δ (ppm) 7.35-7.31 (m, 2H), 6.96 (t, $J = 8.6$ Hz, 2H), 6.89-6.85 (m, 1H), 6.72-6.66 (m, 3H), 4.97 (s, 1H), $^{13}\text{C NMR}$ (100 MHz, $\text{CDCl}_3/\text{Methanol-}d_4 = 10:1$): δ (ppm) 167.2, 162.8 (d, $J_{\text{C-F}} = 245.0$ Hz), 134.9 (d, $J_{\text{C-F}} = 3.0$ Hz), 133.0, 129.0 (d, $J_{\text{C-F}} = 8.0$ Hz), 124.8, 124.2, 119.5, 115.6 (d, $J_{\text{C-F}} = 22.0$ Hz), 115.6, 113.8, 59.9. ESI exact mass calcd. for $\text{C}_{14}\text{H}_{11}\text{FN}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 243.09282, found m/z 243.09214.

The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm), $t_{R1} = 15.3$ min (major), $t_{R2} = 26.8$ min (minor).

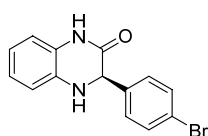


(R)-3-(4-chlorophenyl)-3,4-dihydroquinoxalin-2(1H)-one (2o): Known

compound (see: Kristoffersen, T.; Elumalai, V.; Starck, E.; Cousin, E.; Wagner, L. J.; Hansen, S. R.; Hansen, J. H. *Eur. J. Org. Chem.* **2020**, *45*, 7069-7078). Yellow solid, m.p. 70-72 °C, 49.0 mg, isolated yield 95%, 96% *ee*. $[\alpha]_D^{20} =$

-104.8 ($c = 1.0$, CDCl_3 , 96% *ee*); ^1H NMR (300 MHz, $\text{DMSO}-d_6$): δ (ppm) 10.46 (s, 1H), 7.42-7.34 (m, 4H), 6.82-6.58 (m, 5H), 4.96 (s, 1H); ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$): δ (ppm) 165.7, 139.1, 133.7, 132.4, 128.9, 128.3, 125.3, 123.1, 117.9, 114.9, 113.5, 58.8. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_{10}\text{ClN}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 257.04871, found m/z 257.04797.

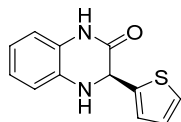
The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{\text{R}1} = 16.2$ min (major), $t_{\text{R}2} = 19.1$ min (minor).



(R)-3-(4-bromophenyl)-3,4-dihydroquinoxalin-2(1H)-one (2p): Known compound (see: Kristoffersen, T.; Elumalai, V.; Starck, E.; Cousin, E.; Wagner, L. J.; Hansen, S. R.; Hansen, J. H. *Eur. J. Org. Chem.* **2020**, 45,

7069-7078). White solid, m.p. 176-178 °C, 56.8 mg, isolated yield 94%, 92% *ee*. $[\alpha]_{\text{D}}^{20} = -106.4$ ($c = 1.0$, CDCl_3 , 92% *ee*); ^1H NMR (300 MHz, $\text{DMSO}-d_6$): δ (ppm) 10.46 (bs, 1H), 7.53 (d, $J = 8.4$ Hz, 2H), 7.30 (d, $J = 8.4$ Hz, 2H), 6.82-6.58 (m, 5H), 4.94 (s, 1H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 165.5, 139.5, 133.6, 131.2, 129.2, 125.3, 123.1, 120.8, 117.9, 114.9, 113.4, 58.7. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_{10}\text{BrN}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 300.99820, found m/z 300.99759.

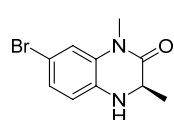
The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 230$ nm), $t_{\text{R}1} = 48.7$ min (minor), $t_{\text{R}2} = 60.6$ min (major).



(S)-3-(thiophen-2-yl)-3,4-dihydroquinoxalin-2(1H)-one (2q): Known compound (see: Petasis, N.A.; Patel, D. A. *Tetrahedron. Lett.* **2000**, 41, 9607-9611). White solid, m.p. 182-184 °C, 43.2 mg, isolated yield 94%, 97%

ee. $[\alpha]_{\text{D}}^{20} = -46.3$ ($c = 1.0$, CDCl_3 , 97% *ee*); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 8.77 (bs, 1H), 7.22 (dd, $J = 5.1$ Hz, $J = 1.2$ Hz, 2H), 7.06 (d, $J = 3.6$ Hz, 1H), 6.97-6.91 (m, 2H), 6.83-6.73 (m, 3H), 5.33 (s, 1H), 4.40 (s, 1H); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 166.4, 141.4, 132.2, 127.0, 126.0, 125.9, 125.0, 124.4, 120.2, 115.9, 114.6, 56.9. HRMS-ESI exact mass calcd. for $\text{C}_{12}\text{H}_9\text{N}_2\text{OS}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 229.04411, found m/z 229.04369.

The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm), $t_{R1} = 16.3$ min (minor), $t_{R2} = 23.2$ min (major).

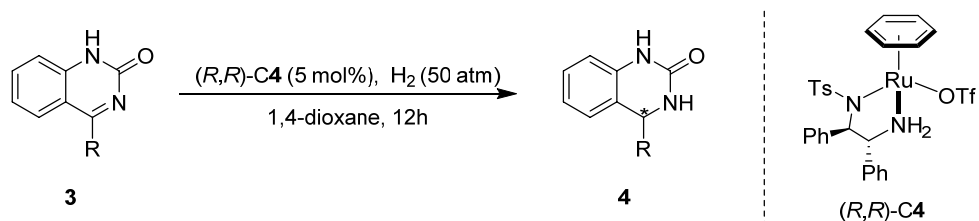


(*R*)-7-bromo-1,3-dimethyl-3,4-dihydroquinoxalin-2(1*H*)-one (2r): New compound. White solid, m.p. 124-126 °C, 47.2 mg, isolated yield 93%, 97% *ee*.

$[\alpha]_D^{20} = -50.4$ ($c = 1.0$, CDCl_3 , 97% *ee*); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ (ppm) 7.02-6.99 (m, 2H), 6.57 (d, $J = 8.7$ Hz, 1H), 3.97-3.91 (m, 2H), 3.33 (s, 3H), 1.42 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ (ppm) 168.1, 134.1, 130.6, 126.0, 117.7, 115.5, 111.5, 52.2, 29.3, 18.0. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{10}\text{BrN}_2\text{O}^+$ ($[\text{M}-\text{H}]^+$) requires m/z 252.99820, found m/z 252.99759.

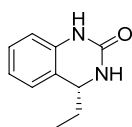
The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 220$ nm), $t_{R1} = 7.7$ min (major), $t_{R2} = 14.5$ min (minor).

5.2 Asymmetric hydrogenation of substrate quinazolin-2(1*H*)-one (3)



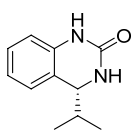
General procedure: A 15 mL glass-lined stainless-steel reactor equipped with a magnetic stirrer bar was charged with substrate **3** (0.1 mmol), Ru-catalyst (*R,R*)-**C4** (0.005 mmol) in 1.0 mL of solvent under N_2 atmosphere in a glove box. The autoclave was closed, and the final pressure of the hydrogen gas was adjusted to 50 atm after purging the autoclave with hydrogen gas several times. The reaction mixture was stirred at 25 °C for 12 h. Then the hydrogen gas was carefully released and the conversion was determined by $^1\text{H NMR}$ spectroscopy. The reaction mixture was filtered through a short pad of silica eluted with ethyl

acetate and petroleum ether to give the chiral product 3,4-dihydroquinazolin-2(1*H*)-one (**4**). The enantiomeric excess of the product was determined by HPLC with a chiral column.



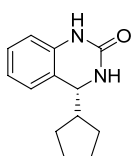
(R)-4-ethyl-3,4-dihydroquinazolin-2(1*H*)-one (4a): (New compound). White solid, m.p. 188-190 °C, 16.2 mg, isolated yield 92%, 41% *ee*. $[\alpha]_{\text{D}}^{20} = -5.6$ ($c = 1.0$, CHCl_3 , 41% *ee*); ^1H NMR (400 MHz, $\text{DMSO-}d_6$): δ (ppm) 9.02 (s, 1H), 7.11-7.04 (m, 2H), 6.95 (s, 1H), 6.85 (t, $J = 7.2$ Hz, 1H), 6.77 (d, $J = 7.6$ Hz, 1H), 4.32 (s, 1H), 1.61-1.57 (m, 2H), 0.78 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$): δ (ppm) 154.0, 137.7, 127.5, 126.0, 121.3, 120.8, 113.6, 54.0, 31.1, 8.5. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{13}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 177.10224, found m/z 177.10132.

The enantiomeric excess was determined by HPLC on the connection of Chiralcel IC-H column and Chiralcel IA-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 30 °C, UV detection at $\lambda = 254$ nm), $t_{\text{R}1} = 15.3$ min (minor), $t_{\text{R}2} = 16.5$ min (major).



(R)-4-isopropyl-3,4-dihydroquinazolin-2(1*H*)-one (4b): Known compound (see: Feng, G.-S.; Zhao, Z.-B.; Shi, L.; Zhou, Y.-G. *Org. Chem. Front.* **2019**, *6*, 2250-2253). White solid, 17.9 mg, isolated yield 94%, 87% *ee*. $[\alpha]_{\text{D}}^{20} = -29.0$ ($c = 1.0$, MeOH, 87% *ee*); ^1H NMR (400 MHz, CDCl_3): δ (ppm) 7.84 (s, 1H), 7.18-7.14 (m, 1H), 7.03 (d, $J = 7.2$ Hz, 1H), 6.97-6.93 (m, 1H), 6.72 (d, $J = 7.6$ Hz, 1H), 5.50 (s, 1H), 4.37-4.35 (m, 1H), 1.99-1.94 (m, 1H), 0.99 (d, $J = 6.8$ Hz, 3H), 0.87 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ (ppm) 155.4, 136.8, 128.2, 126.7, 122.1, 120.4, 114.3, 60.2, 36.8, 18.6, 16.1. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 191.11789, found m/z 191.11661.

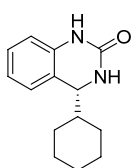
The enantiomeric excess was determined by HPLC on the Chiralcel IC-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 30 °C, UV detection at $\lambda = 254$ nm), $t_{\text{R}1} = 13.4$ min (minor), $t_{\text{R}2} = 14.4$ min (major).



(R)-4-cyclopentyl-3,4-dihydroquinazolin-2(1H)-one (4c): (New compound).

White solid, m.p. 230-232 °C, 19.4 mg, isolated yield 90%, 79% *ee*. $[\alpha]_D^{20} = -30.0$ ($c = 1.0$, MeOH, 79% *ee*); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ (ppm) 8.55 (s, 1H), 7.13 (t, $J = 7.6$ Hz, 1H), 7.05 (d, $J = 7.6$ Hz, 1H), 6.93 (t, $J = 7.2$ Hz, 1H), 6.76 (d, $J = 8.0$ Hz, 1H), 6.0 (s, 1H), 4.35-4.33 (m, 1H), 2.24-2.16 (m, 1H), 1.77-1.26 (m, 8H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ (ppm) 156.2, 136.6, 128.1, 126.5, 122.0, 121.6, 114.5, 58.1, 48.6, 29.1, 27.9, 25.2, 25.0. HRMS-ESI exact mass calcd. for $\text{C}_{13}\text{H}_{17}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 217.13354, found m/z 217.13232.

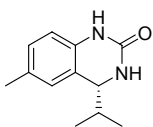
The enantiomeric excess was determined by HPLC on the Chiralcel IA-H column (*n*-hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 30 °C, UV detection at $\lambda = 254$ nm), $t_{R1} = 15.9$ min (minor), $t_{R2} = 17.7$ min (major).



(R)-4-cyclohexyl-3,4-dihydroquinazolin-2(1H)-one (4d): Known compound

(see: Feng, G.-S.; Zhao, Z.-B.; Shi, L.; Zhou, Y.-G. *Org. Chem. Front.* **2019**, *6*, 2250-2253). White solid, 21.4 mg, isolated yield 93%, 88% *ee*. $[\alpha]_D^{20} = -19.0$ ($c = 1.0$, MeOH, 88% *ee*); $^1\text{H NMR}$ (400 MHz, $\text{DMSO}-d_6$): δ (ppm) 8.98 (s, 1H), 7.10 (t, $J = 7.4$ Hz, 1H), 7.03-6.99 (m, 2H), 6.84 (t, $J = 6.6$ Hz, 1H), 6.76 (d, $J = 7.2$ Hz, 1H), 4.10 (s, 1H), 1.65-1.41 (m, 6H), 1.11-0.93 (m, 5H); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ (ppm) 154.3, 138.1, 127.5, 126.7, 120.5, 120.4, 113.4, 58.1, 46.4, 28.2, 26.7, 25.9, 25.7, 25.6. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_{19}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 231.14919, found m/z 231.14942.

The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 30 °C, UV detection at $\lambda = 254$ nm), $t_{R1} = 4.2$ min (minor), $t_{R2} = 4.9$ min (major).

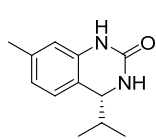


(R)-4-isopropyl-6-methyl-3,4-dihydroquinazolin-2(1H)-one (4e): (New

compound). White solid, m.p. 232-234 °C, 18.6 mg, isolated yield 91%, 87% *ee*. $[\alpha]_D^{20} = -61.6$ ($c = 1.0$, MeOH, 87% *ee*); $^1\text{H NMR}$ (300 MHz, $\text{DMSO}-d_6$): δ (ppm) 8.86 (s, 1H), 6.92-6.85 (m, 3H), 6.65 (d, $J = 7.8$ Hz, 1H), 4.11 (s, 1H), 2.20 (s, 3H), 1.80-1.74 (m, 1H), 0.84 (d, $J = 6.6$ Hz, 3H), 0.73 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C NMR}$ (75 MHz,

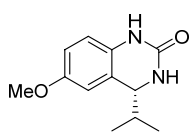
CDCl₃): δ (ppm) 154.3, 135.6, 129.2, 128.0, 127.0, 120.3, 113.3, 58.5, 36.5, 20.4, 18.3, 16.4. HRMS-ESI exact mass calcd. for C₁₂H₁₇N₂O⁺ ([M+H]⁺) requires m/z 205.13354, found m/z 205.13509.

The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 95 : 5, flowing rate = 1.0 mL/min, 25 °C, UV detection at λ = 254 nm), t_{R1} = 8.0 min (major), t_{R2} = 9.6 min (minor).



(R)-4-isopropyl-7-methyl-3,4-dihydroquinazolin-2(1H)-one (4f): (New compound). White solid, m.p. 225-227 °C, 18.8 mg, isolated yield 92%, 89% *ee*. $[\alpha]_D^{20}$ = -33.0 (*c* = 1.0, MeOH, 89% *ee*); ¹H NMR (300 MHz, CDCl₃/Methanol-d₄ = 10:1): δ (ppm) 6.85 (d, *J* = 7.8 Hz, 1H), 6.72 (d, *J* = 7.5 Hz, 1H), 6.50 (s, 1H), 4.25 (d, *J* = 3.6 Hz, 1H), 2.23 (s, 3H), 1.89-1.83 (m, 1H), 0.90 (d, *J* = 6.9 Hz, 3H), 0.78 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃/Methanol-d₄ = 10:1): δ (ppm) 155.9, 138.2, 136.4, 126.5, 123.0, 117.4, 114.7, 59.7, 36.8, 21.1, 18.4, 16.1. HRMS-ESI exact mass calcd. for C₁₂H₁₇N₂O⁺ ([M+H]⁺) requires m/z 205.13354, found m/z 205.13322.

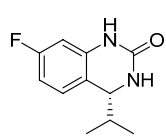
The enantiomeric excess was determined by HPLC on the Chiralcel AD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at λ = 254 nm), t_{R1} = 7.7 min (minor), t_{R2} = 30.2 min (major).



(R)-4-isopropyl-5-methoxy-3,4-dihydroquinazolin-2(1H)-one (4g): (New compound). White solid, m.p. 220-222 °C, 20.7mg, isolated yield 94%, 90% *ee*. $[\alpha]_D^{20}$ = -36.0 (*c* = 1.0, MeOH, 90% *ee*); ¹H NMR (400 MHz, CDCl₃): δ (ppm) 7.84 (s, 1H), 6.74-6.71 (m, 1H), 6.66 (d, *J* = 8.4 Hz, 1H), 6.58 (d, *J* = 2.4 Hz, 1H), 5.46 (s, 1H), 4.32 (t, *J* = 3.2 Hz, 1H), 3.76 (s, 3H), 2.00-1.93 (m, 1H), 0.99 (d, *J* = 7.2 Hz, 3H), 0.86 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ (ppm) 155.6, 155.0, 130.5, 121.6, 115.1, 113.5, 112.5, 60.3, 55.8, 36.8, 18.7, 16.1. HRMS-ESI exact mass calcd. for C₁₂H₁₇N₂O₂⁺ ([M+H]⁺) requires m/z 221.12845, found m/z 221.12801.

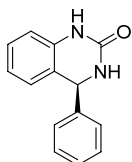
The enantiomeric excess was determined by HPLC on the Chiralcel AD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at λ = 254

nm), t_{R1} = 26.2 min (minor), t_{R2} = 33.1 min (major).



(R)-7-fluoro-4-isopropyl-3,4-dihydroquinazolin-2(1H)-one (4h): (New compound). White solid, m.p. 225-227 °C, 18.9mg, isolated yield 91%, 86% *ee*. $[\alpha]_D^{20}$ = -28.0 (c = 1.0, MeOH, 86% *ee*); ^1H NMR (500 MHz, $\text{CDCl}_3/\text{Methanol-}d_4$ = 10:1): δ (ppm) 6.94-6.91 (m, 1H), 6.45 (d, J = 9.5 Hz, 1H), 4.26 (d, J = 3.0 Hz, 1H), 1.88-1.85 (m, 1H), 0.92 (d, J = 6.5 Hz, 3H), 0.80 (d, J = 6.5 Hz, 3H). ^{13}C NMR (125 MHz, $\text{CDCl}_3/\text{Methanol-}d_4$ = 10:1): δ (ppm). 162.5(d, $J_{\text{C-F}}$ = 243.8 Hz), 155.4, 138.2 (d, $J_{\text{C-F}}$ = 11.3 Hz), 128.0 (d, $J_{\text{C-F}}$ = 10.0 Hz), 116.0 (d, $J_{\text{C-F}}$ = 3.8 Hz), 108.9 (d, $J_{\text{C-F}}$ = 22.5 Hz), 101.5 (d, $J_{\text{C-F}}$ = 25.0 Hz), 59.4, 36.9, 18.2, 16.0. HRMS-ESI exact mass calcd. for $\text{C}_{11}\text{H}_{14}\text{FN}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 209.10847, found m/z 209.10803.

The enantiomeric excess was determined by HPLC on the Chiralcel AD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at λ = 254 nm), t_{R1} = 7.9 min (minor), t_{R2} = 20.7 min (major).

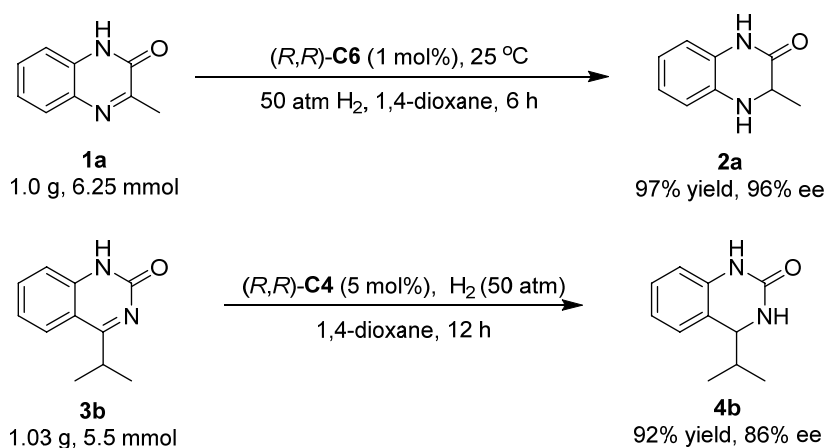


(S)-4-phenyl-3,4-dihydroquinazolin-2(1H)-one (4i): Known compound (see: Feng, G.-S.; Zhao, Z.-B.; Shi, L.; Zhou, Y.-G. *Org. Chem. Front.* **2019**, *6*, 2250-2253). White solid, 20.3 mg, isolated yield 91%, 9% *ee*. ^1H NMR (400 MHz, CDCl_3): δ (ppm) 7.88 (bs, 1H), 7.36-7.29 (m, 5H), 7.15 (t, J = 7.6 Hz, 1H), 6.87 (t, J = 7.4 Hz, 1H), 6.82-6.77 (m, 2H), 5.64 (s, 1H), 5.45 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ (ppm) 154.4, 142.8, 135.9, 129.2, 128.6, 128.5, 127.4, 127.2, 122.6, 121.4, 114.6, 58.9. HRMS-ESI exact mass calcd. for $\text{C}_{14}\text{H}_{13}\text{N}_2\text{O}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 225.10224, found m/z 225.10200.

The enantiomeric excess was determined by HPLC on the Chiralcel OJ-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 30 °C, UV detection at λ = 254 nm), t_{R1} = 7.8 min (major), t_{R2} = 12.3 min (minor).

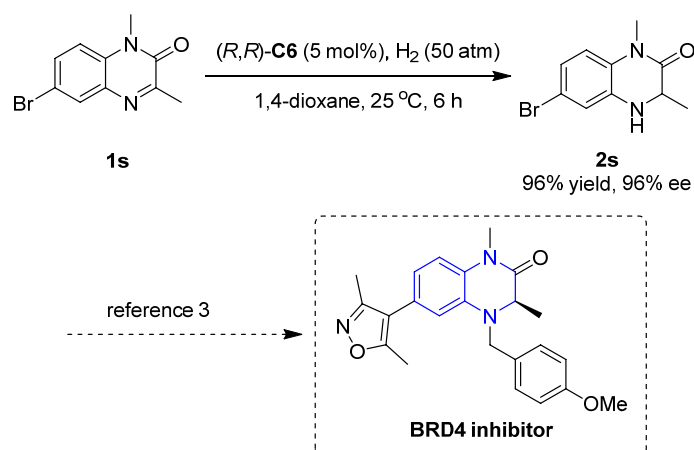
5. Scale-up syntheses and synthetic applications

5.1 Scale-up synthesis

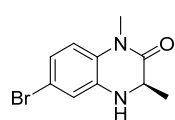


Scheme S3. Scale-up synthesis

General procedure: A 100 mL glass-lined stainless-steel reactor equipped with a magnetic stirrer bar was charged with substrate **1a** (6.25 mmol), Ru-catalyst *(R,R)*-**C6** (0.0625 mmol) in 62.5 mL of 1,4-dioxane under N₂ atmosphere in a glove box. The autoclave was closed, and the final pressure of the hydrogen gas was adjusted to 50 atm after purging the autoclave with hydrogen gas several times. The reaction mixture was stirred at 25 °C for 6 h. Then the hydrogen gas was carefully released and the conversion was determined by ¹H NMR spectroscopy. The reaction mixture was filtered through a short pad of silica eluted with ethyl acetate and petroleum ether to give the chiral product (*S,S*)-**2a**. The enantiomeric excess of the product was determined by HPLC with a chiral column.



Scheme S4. Synthesis of a key intermediate of a bioactive BRD4 inhibitor



(R)-6-bromo-1,3-dimethyl-3,4-dihydroquinoxalin-2(1H)-one (2s): Known compound (see: Zhao, Z.-B.; Li, X.; Chen, M.-W.; Zhao Z. K.; Zhou, Y.-G. *Chem. Commun.* **2020**, 56, 7309-7312). White solid, m.p. 110-112 °C, 48.3 mg, isolated yield 95%, 96% *ee*. $[\alpha]_{\text{D}}^{20} = -76.8$ ($c = 1.0$, CDCl_3 , 96% *ee*); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ (ppm) 6.97-6.93 (m, 1H), 6.82 (d, $J = 2.1$ Hz, 1H), 6.75 (d, $J = 8.7$ Hz, 1H), 3.99-3.93 (m, 2H), 3.33 (s, 3H), 1.42 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ (ppm) 167.9, 136.3, 128.4, 122.4, 116.9, 116.1, 116.1, 52.2, 29.3, 18.2. HRMS-ESI exact mass calcd. for $\text{C}_{10}\text{H}_{10}\text{BrN}_2\text{O}^-$ ($[\text{M}-\text{H}]^-$) requires m/z 253.09715, found m/z 253.09695.

The enantiomeric excess was determined by HPLC on the Chiralcel OD-H column (*n*-hexane : isopropanol = 80 : 20, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 220$ nm), $t_{\text{R}1} = 7.4$ min (major), $t_{\text{R}2} = 17.5$ min (minor).

6. References

1. (a) Ohkuma, T.; Utsumi, N.; Tsutsumi, K.; Murata, K.; Sandoval, C.; Noyori, R. *J. Am. Chem. Soc.* **2006**, *128*, 8724. (b) Wang, T. L.; Zhuo, L.-G.; Li, Z.; Chen, F.; Ding, Z.; He, Y.-M.; Fan, Q.-H.; Xiang, J.-F.; Yu, Z.-X.; Chan, A. S. C. *J. Am. Chem. Soc.* **2011**, *133*, 9878.
2. (a) Rueping, M.; Tato, F.; Schoepke, F. R. *Chem. Eur. J.* **2010**, *16*, 2688-2691. (b) Shi, F.; Tan, W.; Zhang, H.-H.; Li, M.; Ye, Q.; Ma, G.-H.; Tu, S.-J.; Li, G. *Adv. Synth. Catal.* **2013**, *355*, 3715-3726. (c) Núñez-Rico, J. L.; Vidal-Ferran, A. *Org. Lett.* **2013**, *15*, 2066-2069. (d) Zhao, Z.-B.; Li, X.; Chen, M.-W.; Zhao, Z. K.; Zhou, Y.-G. *Chem. Commun.* **2020**, *56*, 7309-7312. (e) Bergman, J.; Brynolf, A.; Elman, B.; Vuorinen, E. *Tetrahedron.* **1986**, *42*, 3697-3706.
3. Yang, Y.; Zhao, L.; Xu, B.; Yang, L.; Zhang, J.; Zhang, H.; Zhou, J. *Bioorg. Chem.* **2016**, *68*, 236-244.

7. Copy of NMR spectra for all substrates and products

Figure S1. ^1H NMR and ^{13}C NMR spectra of **1a** (CDCl_3)

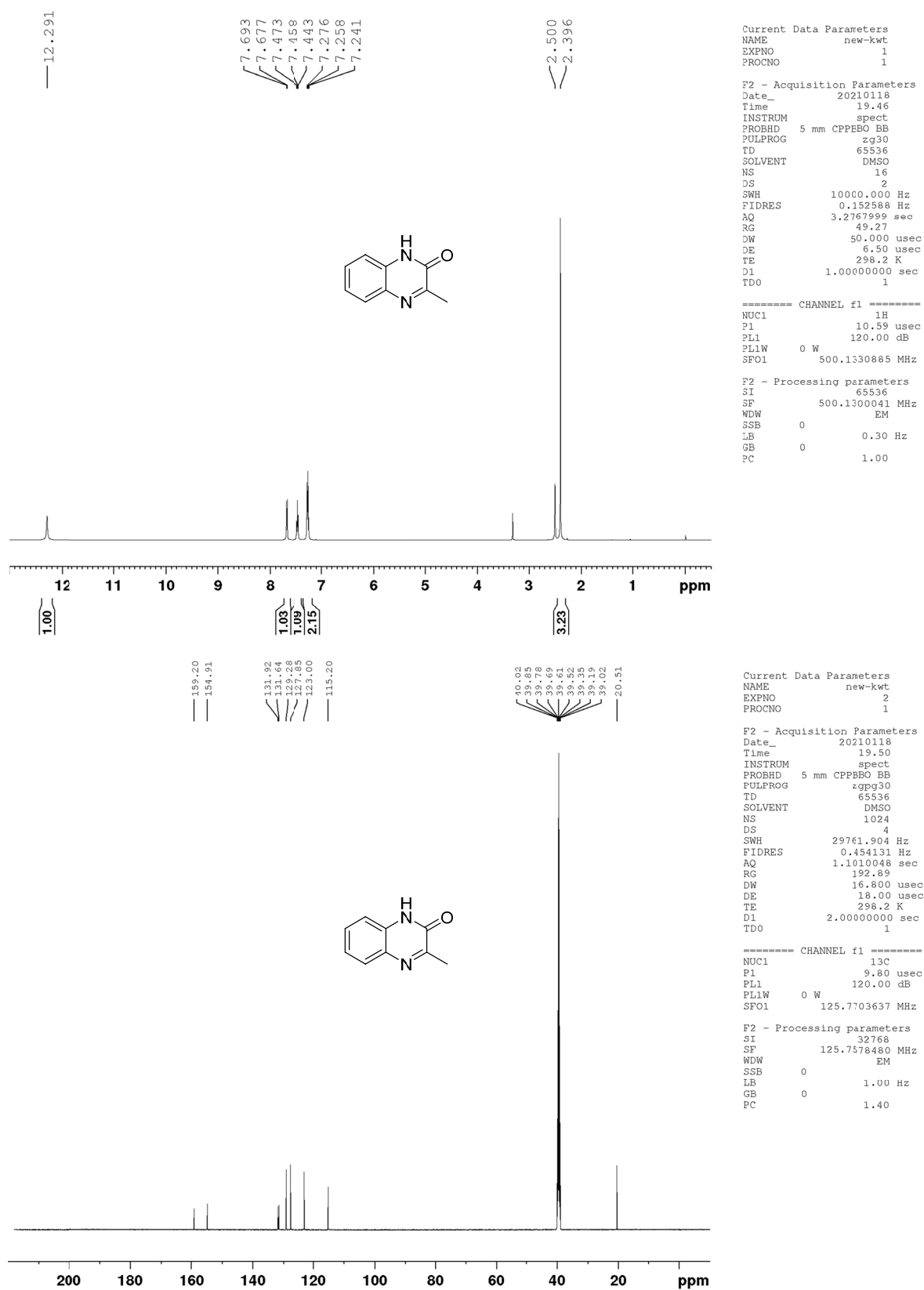


Figure S2. ¹H NMR and ¹³C NMR spectra of **1b**

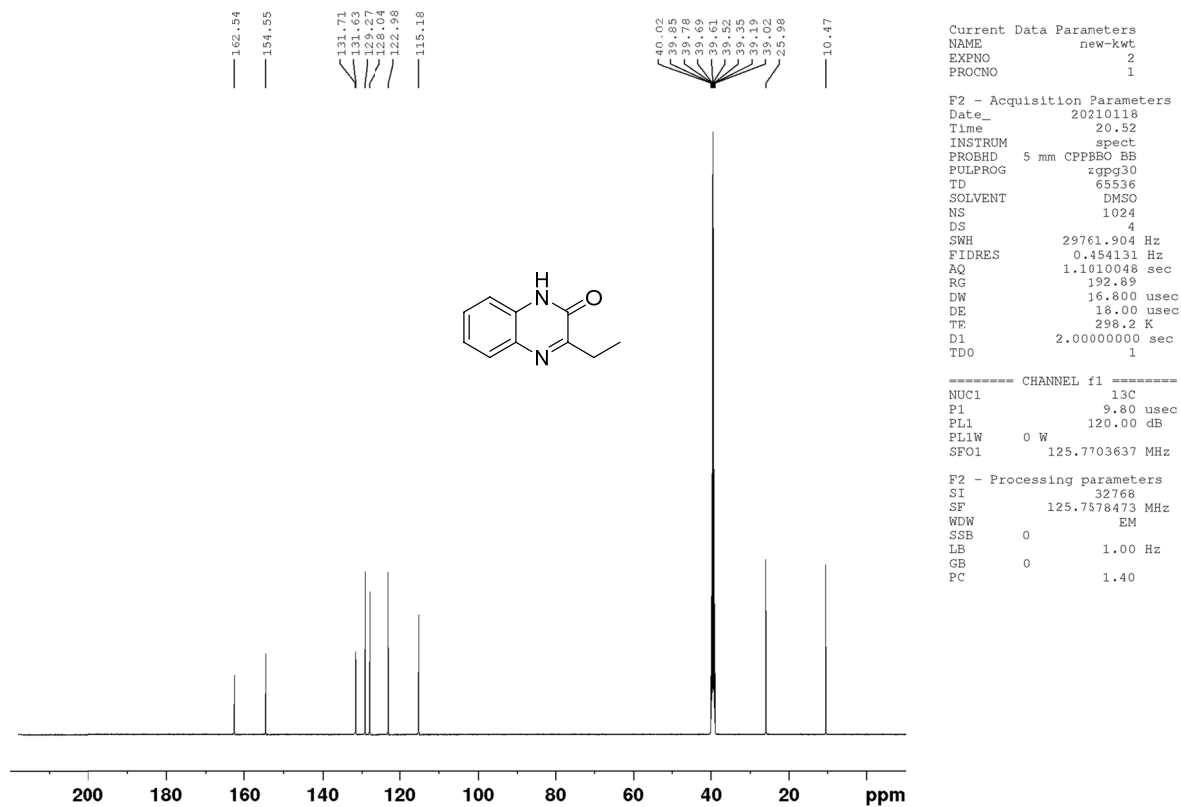
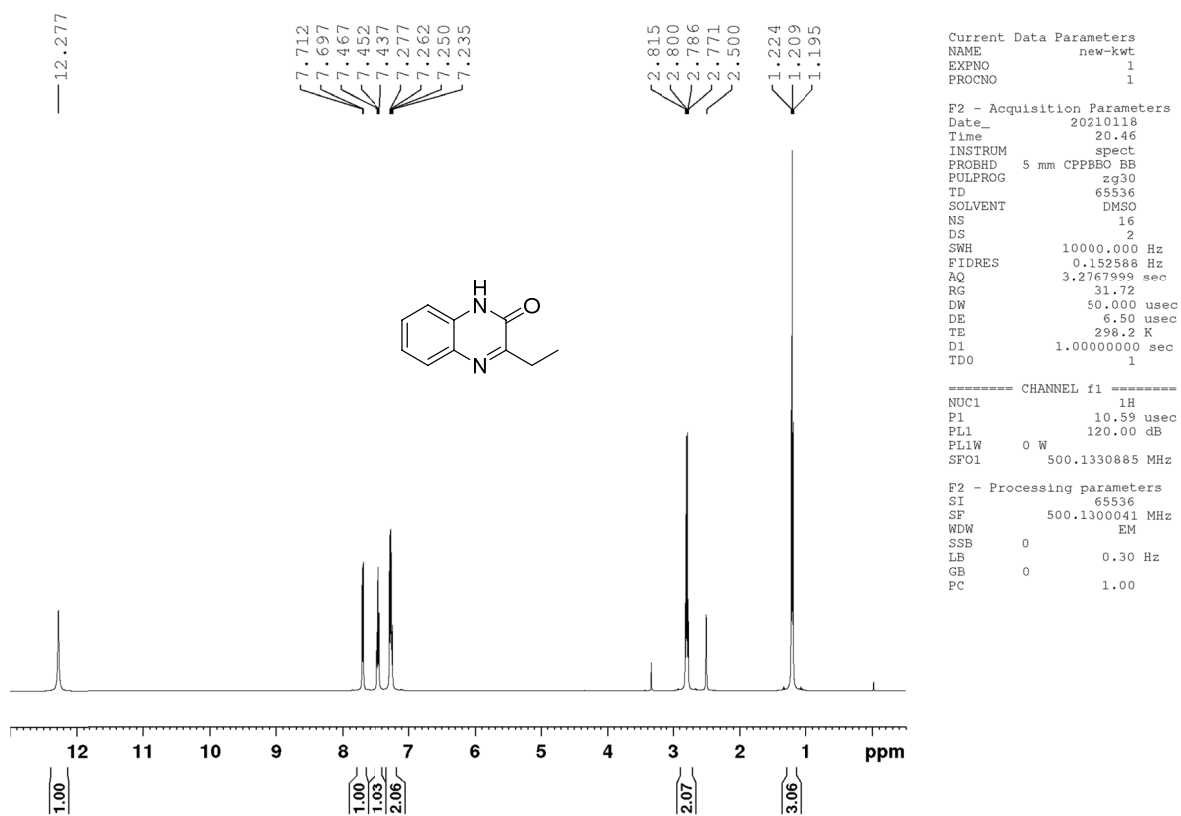


Figure S3. ¹H NMR and ¹³C NMR spectra of **1c**

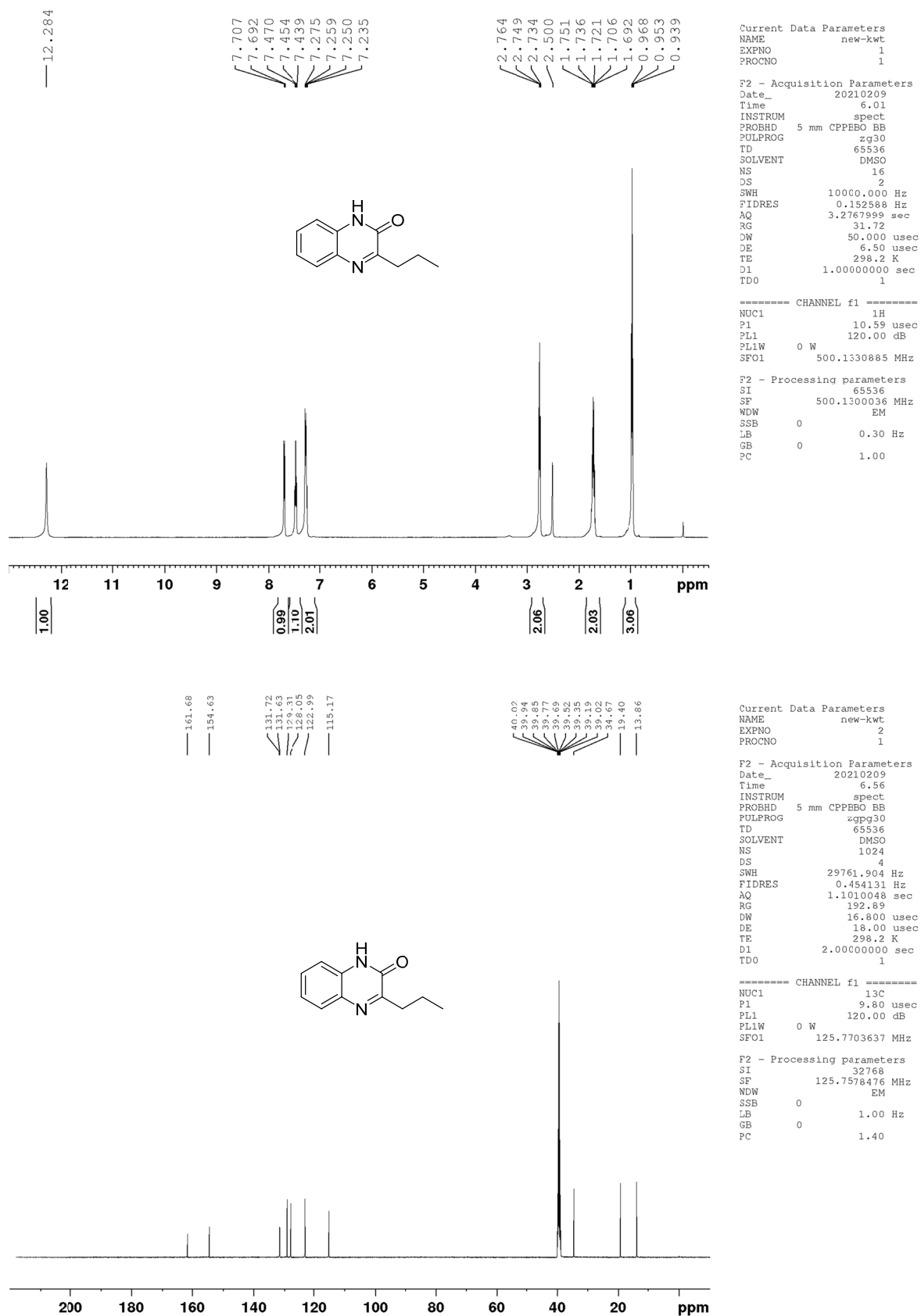


Figure S4. ¹H NMR and ¹³C NMR spectra of **1d**

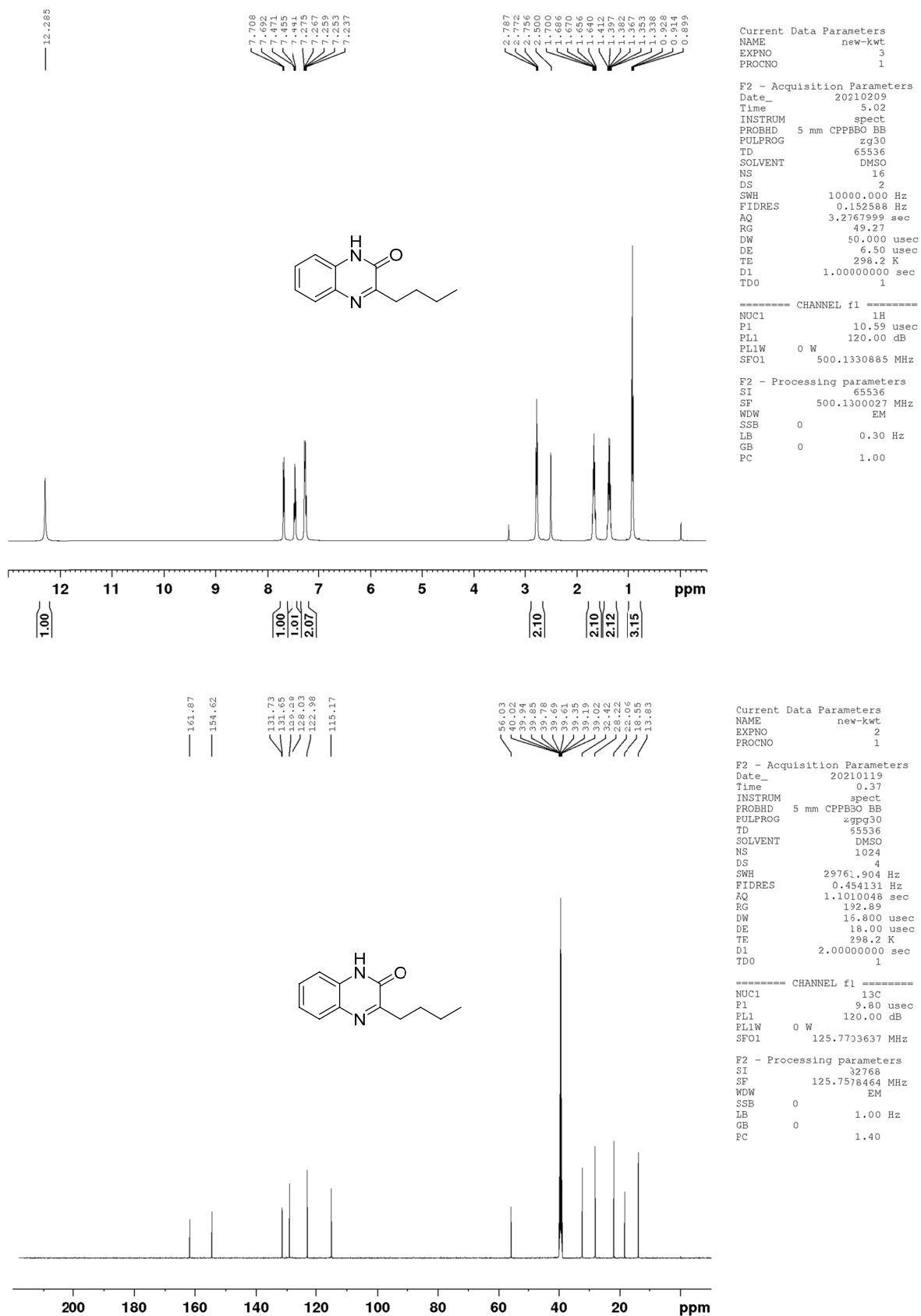


Figure S5. ¹H NMR and ¹³C NMR spectra of **1e**

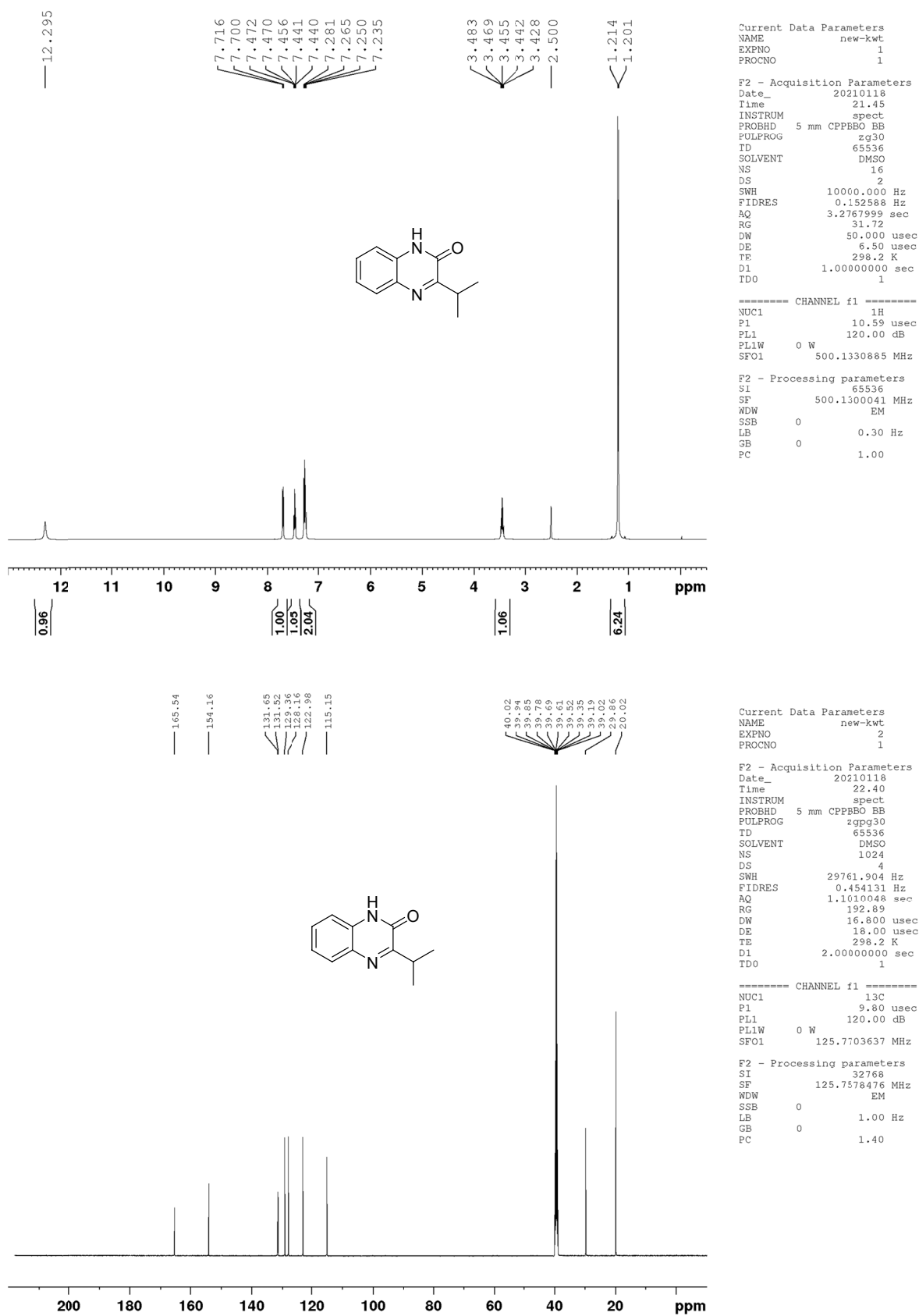


Figure S6. ^1H NMR and ^{13}C NMR spectra of **1f**

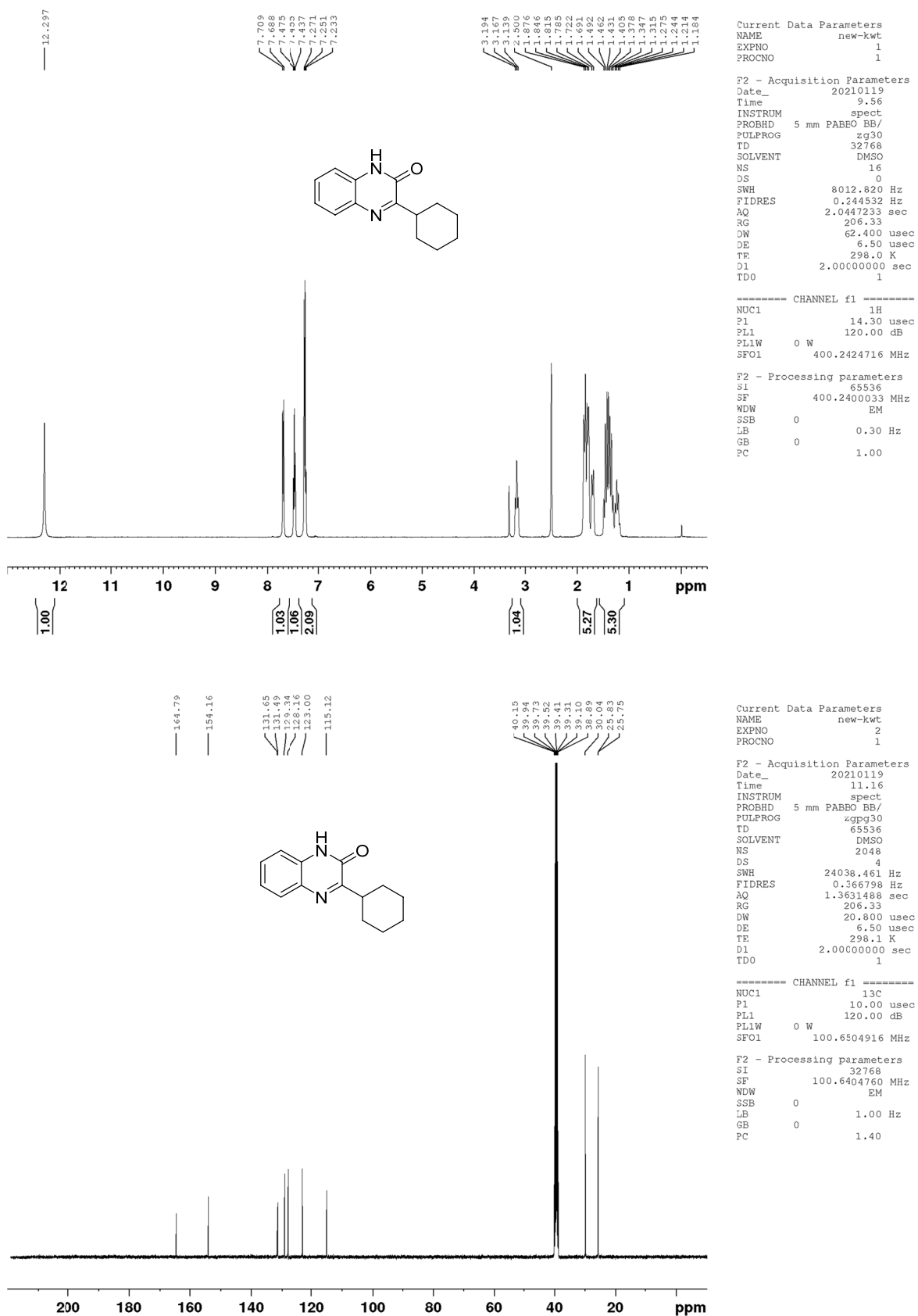
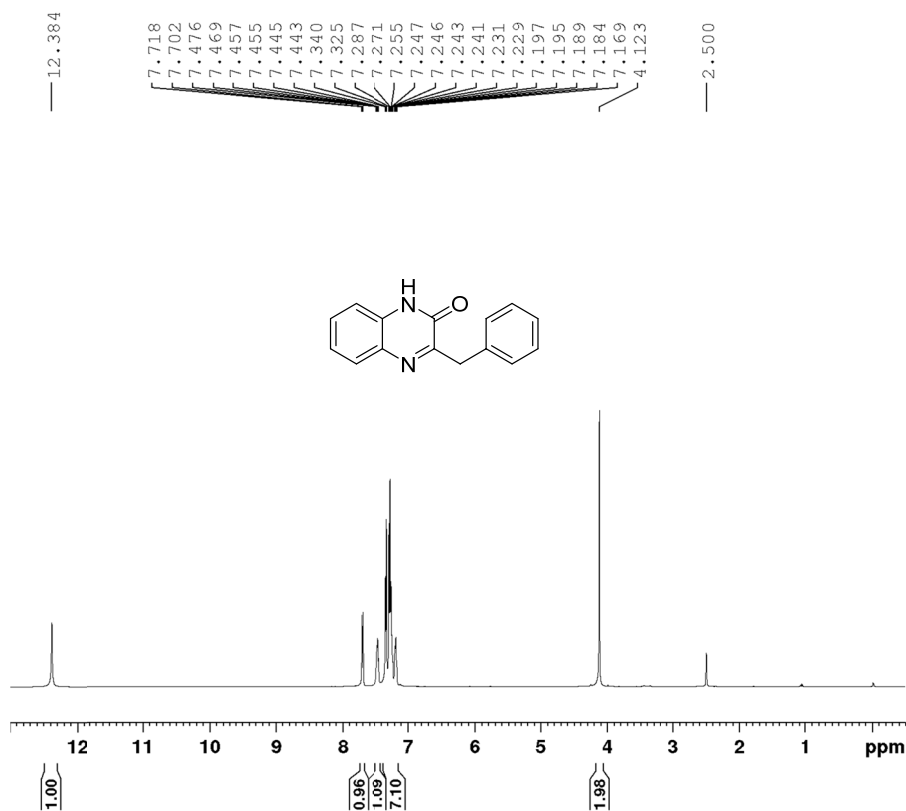


Figure S7. ¹H NMR and ¹³C NMR spectra of **1g**

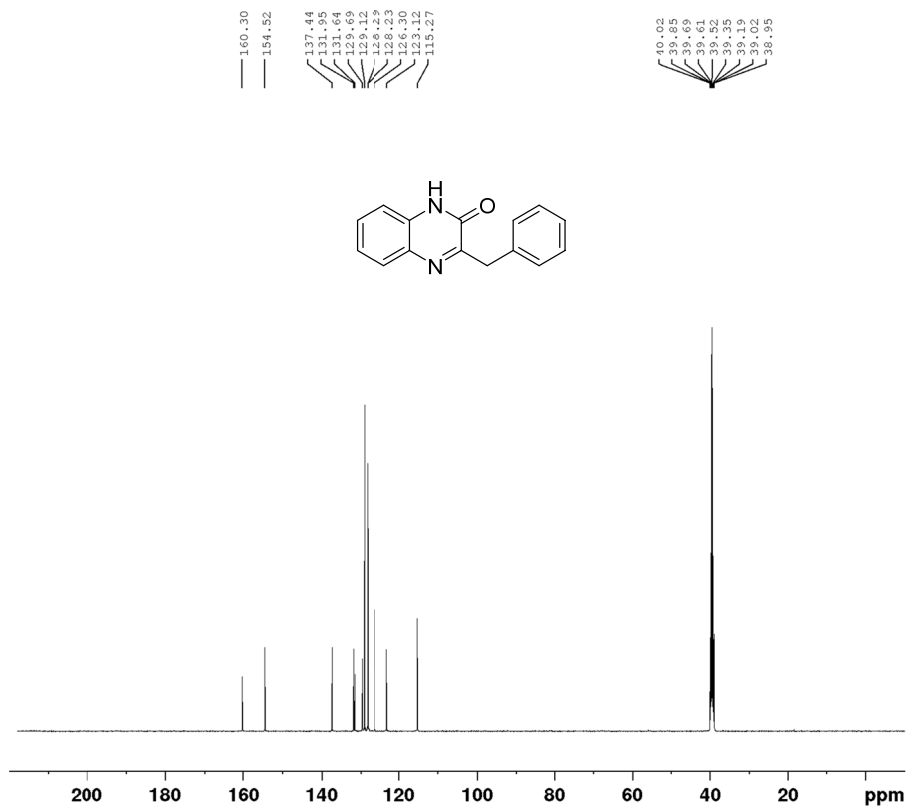


Current Data Parameters
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 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210119
 Time 0.41
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 ID 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152568 Hz
 AQ 3.2767999 sec
 RG 31.72
 DW 50.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.59 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300045 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME new-kwt
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210119
 Time 1.36
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 1024
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 192.89
 DW 16.800 usec
 DE 18.00 usec
 TE 298.2 K
 D1 2.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.80 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 125.7703637 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7578480 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S8. ¹H NMR and ¹³C NMR spectra of **1h**

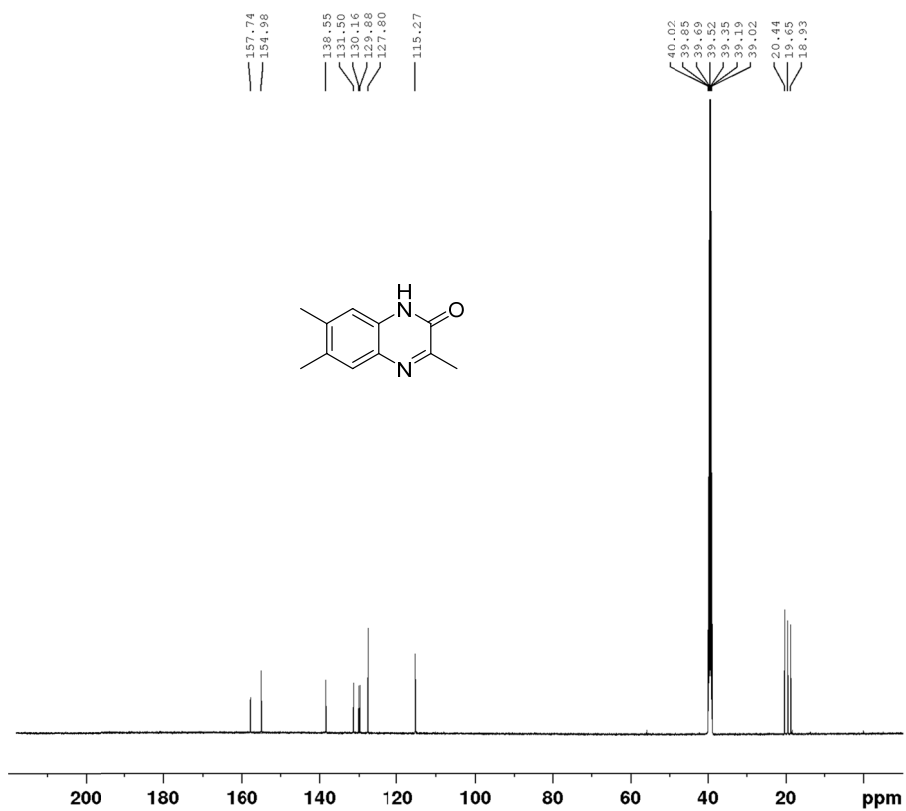
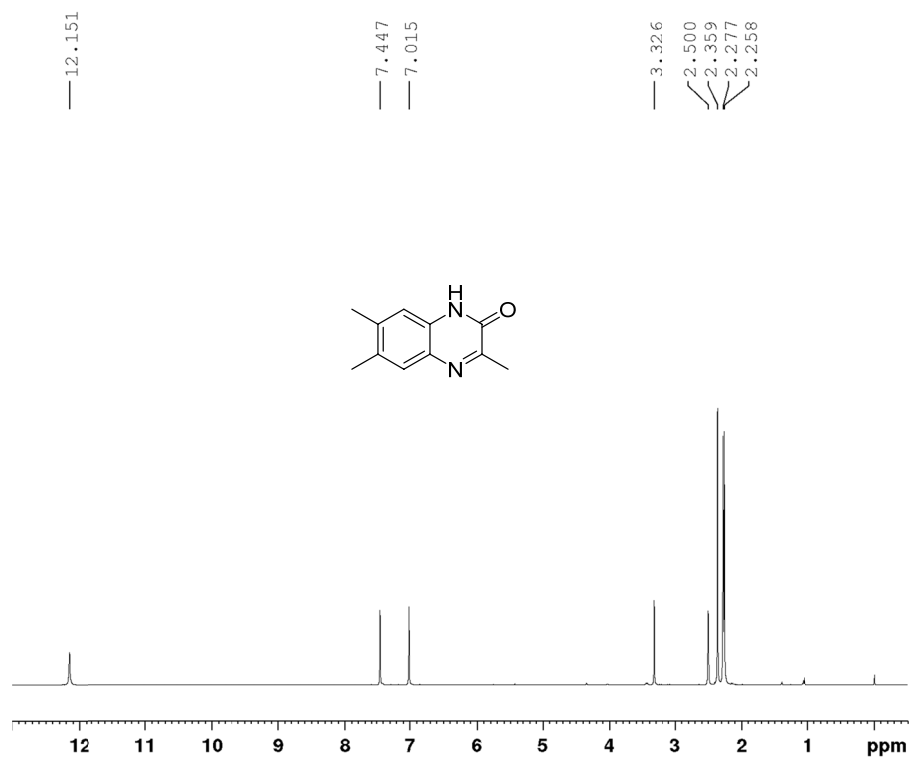


Figure S9. ¹H NMR and ¹³C NMR spectra of **1i**

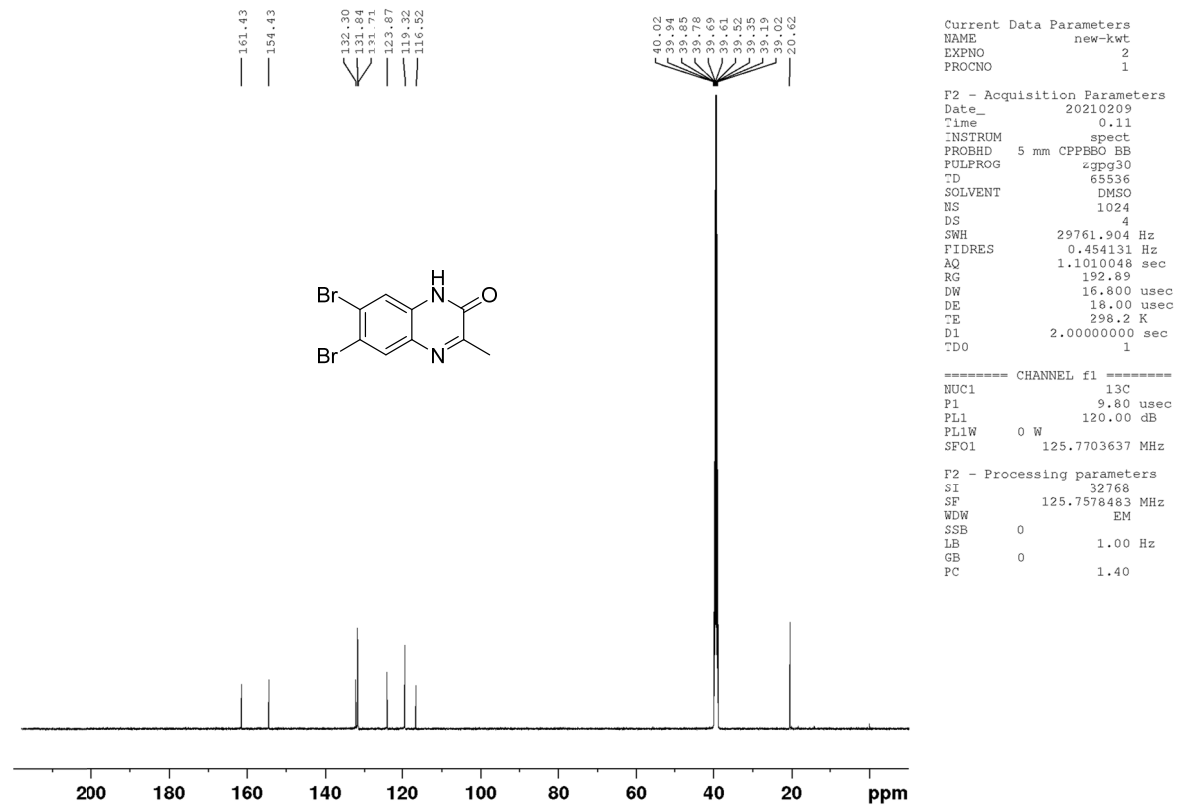
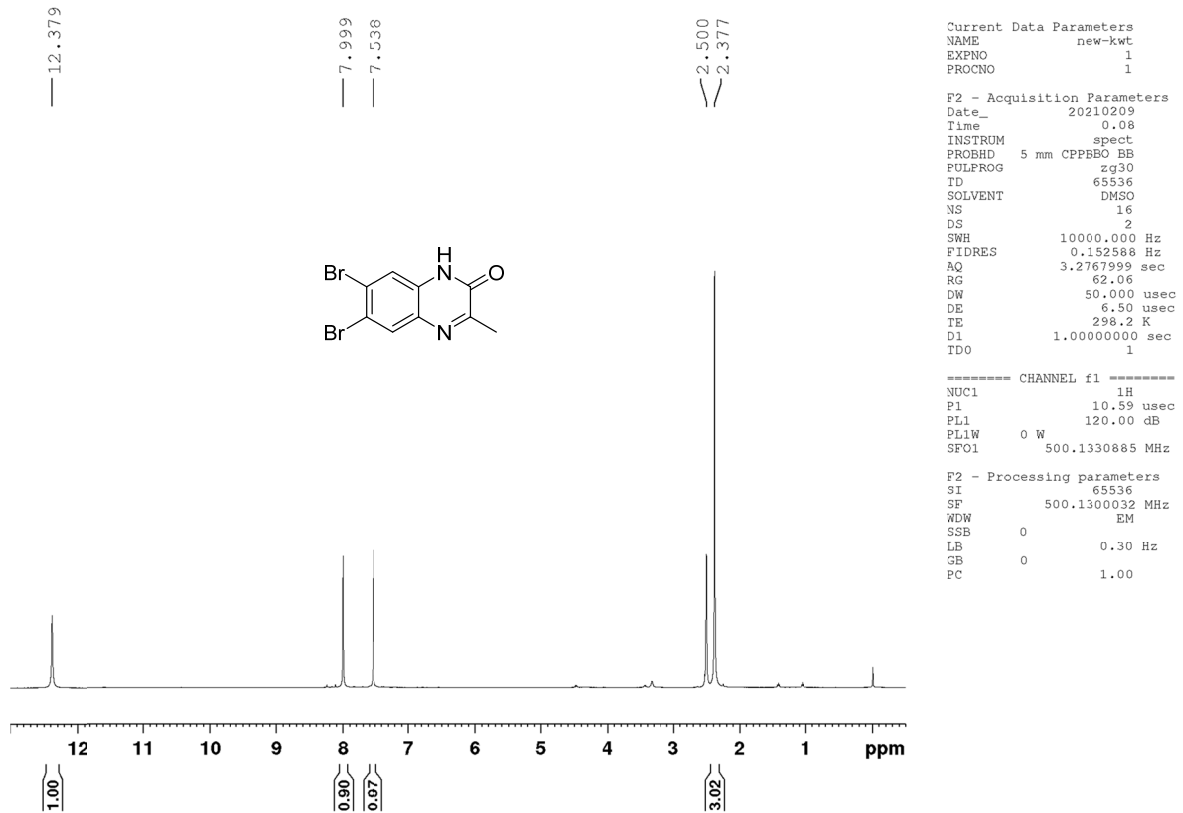


Figure S10. ¹H NMR and ¹³C NMR spectra of **1j**

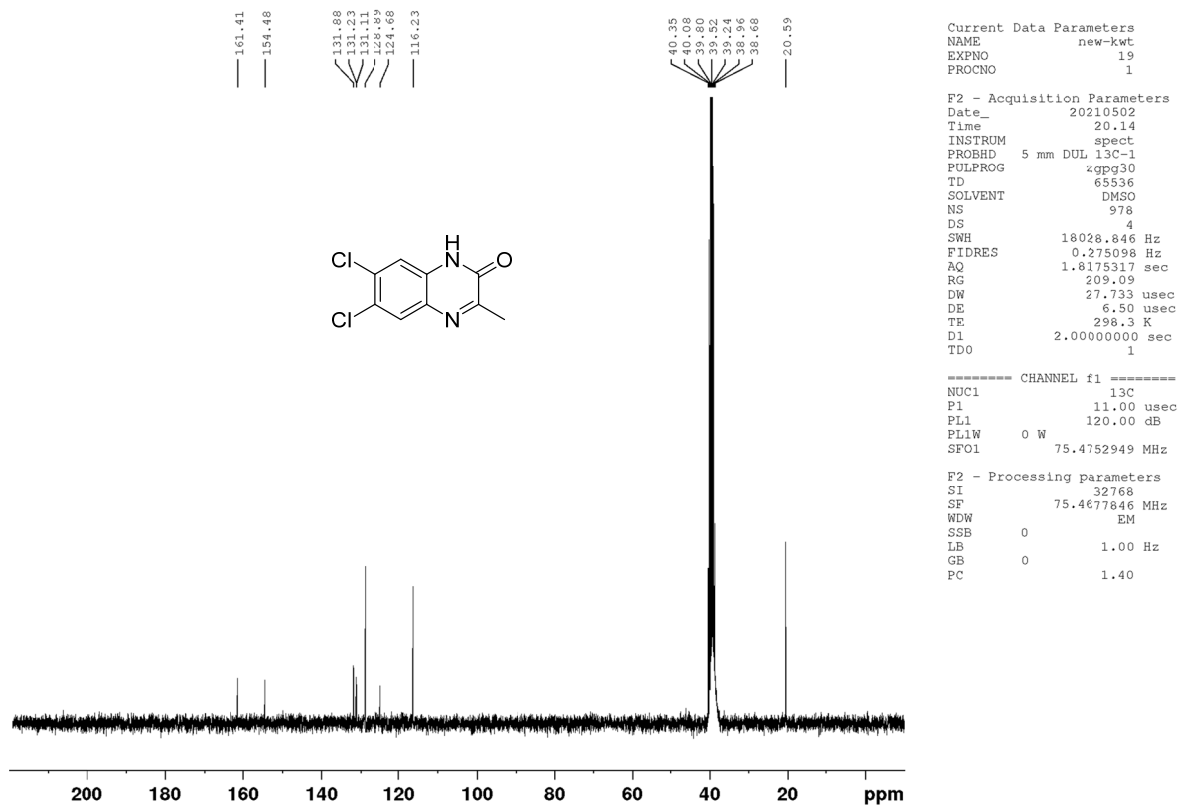
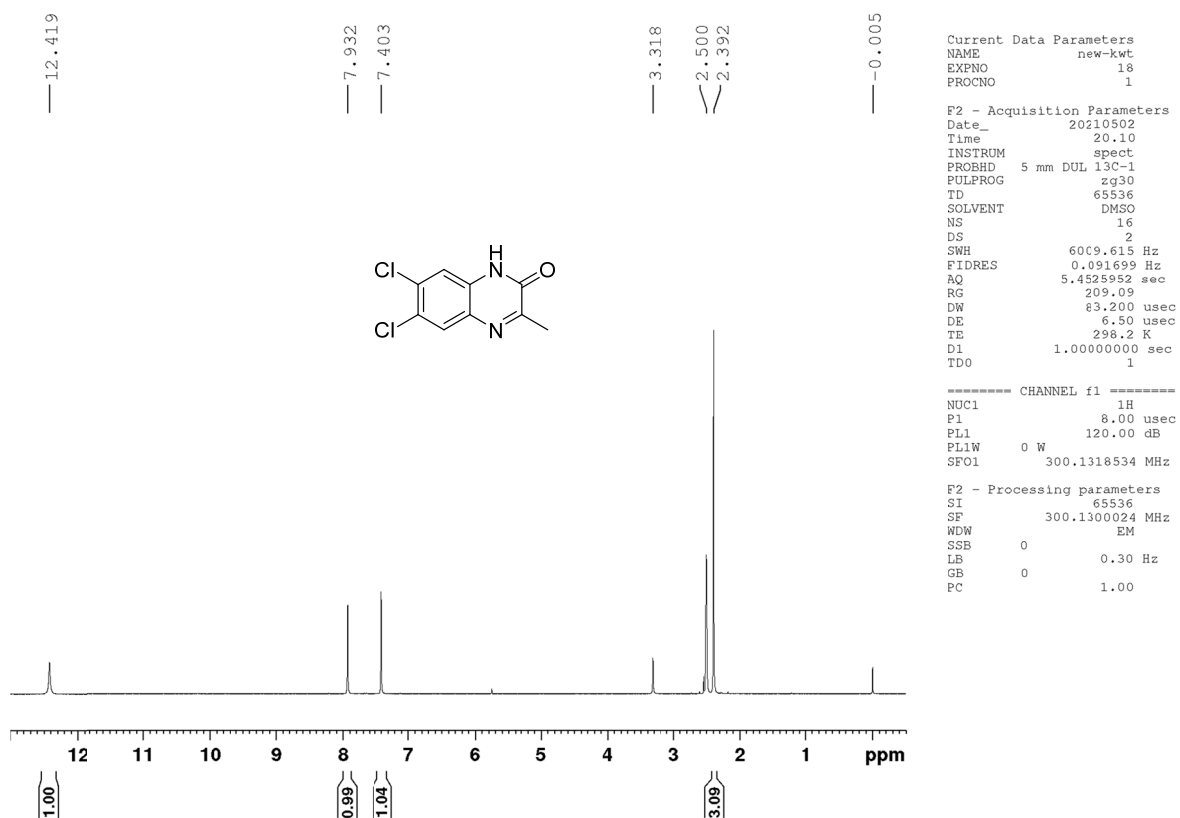


Figure S11. ¹H NMR and ¹³C NMR spectra of 1k

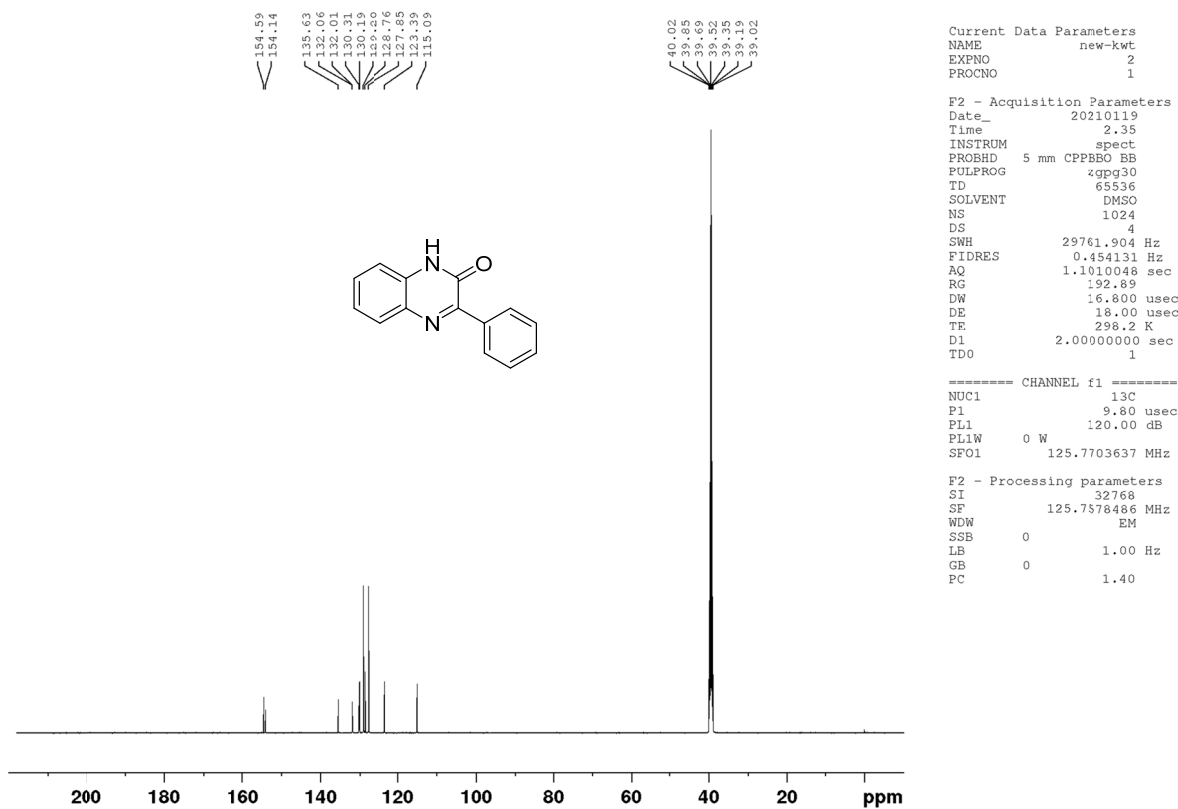
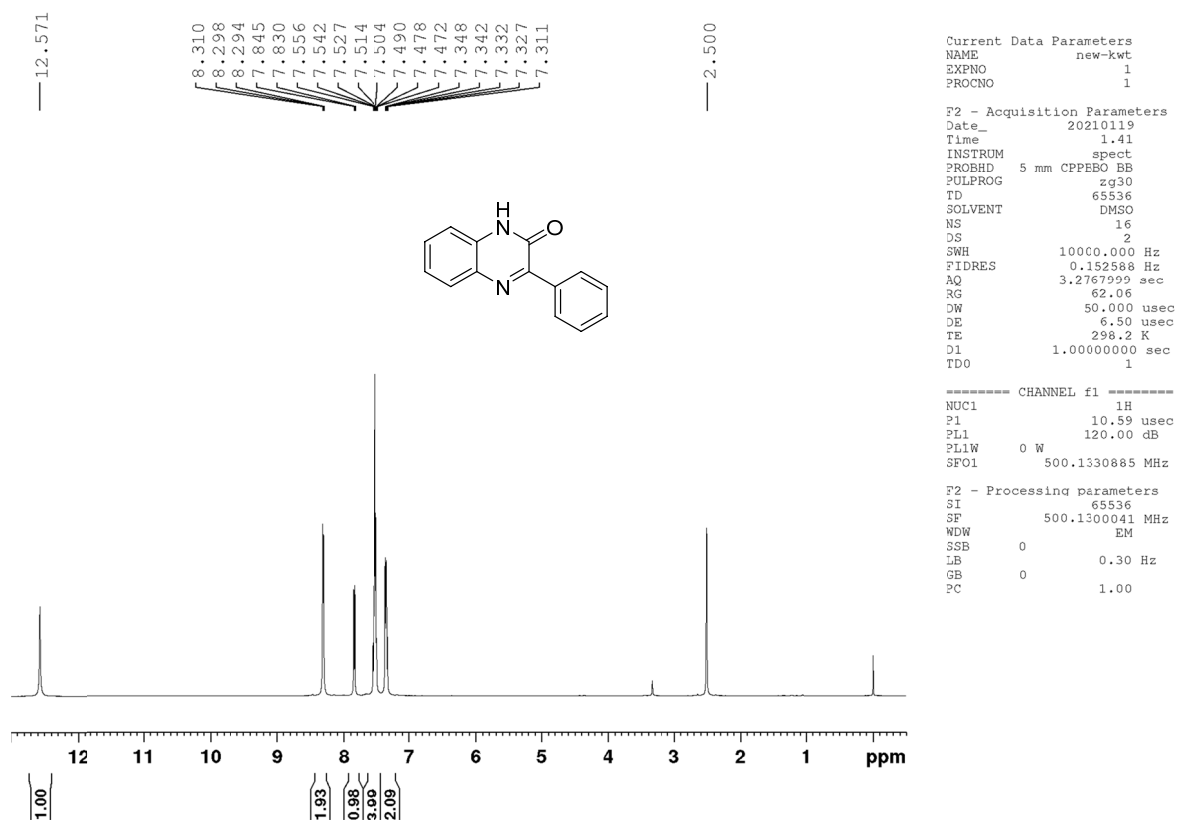


Figure S12. ^1H NMR and ^{13}C NMR spectra of **11**

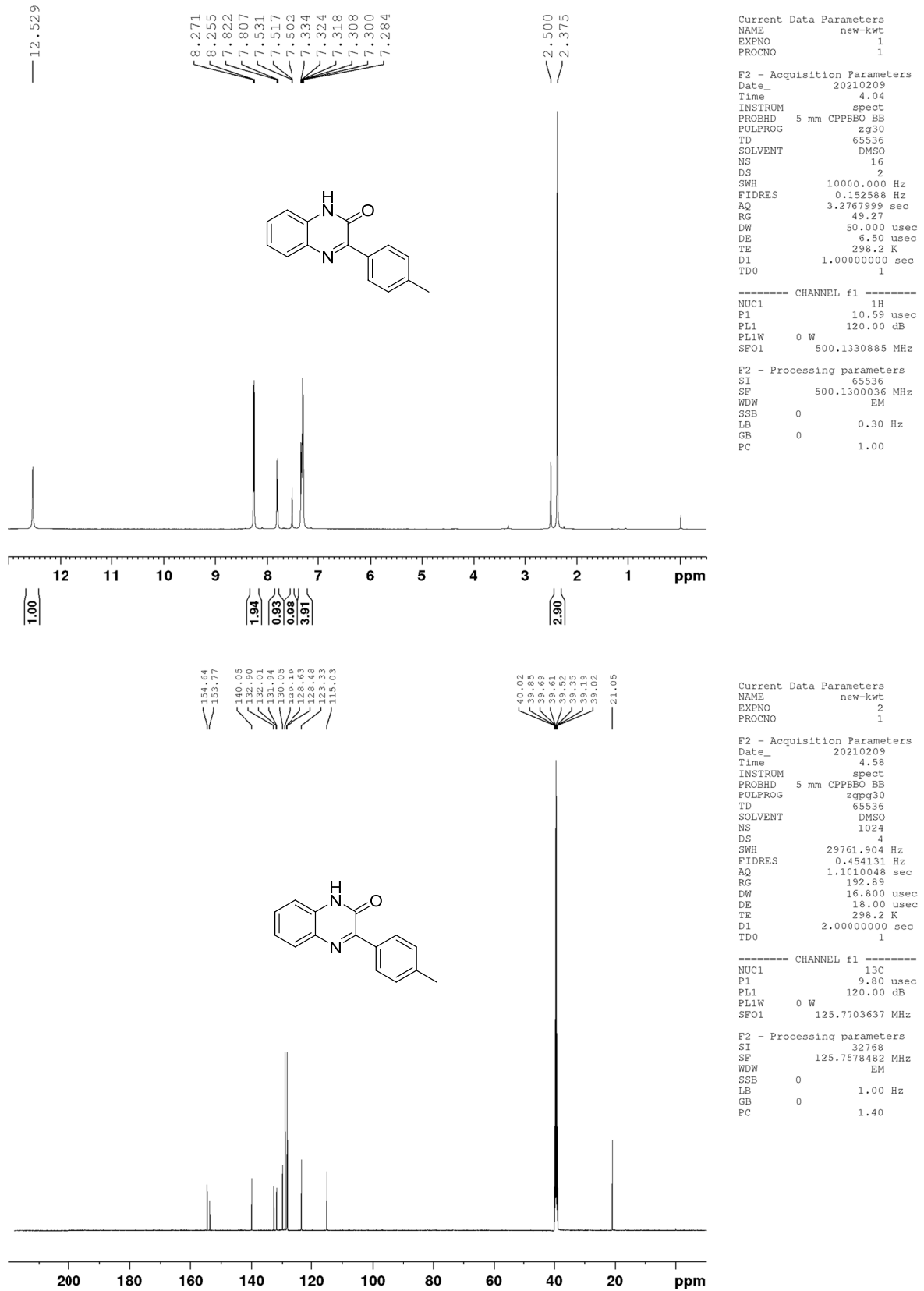


Figure S13. ¹H NMR and ¹³C NMR spectra of **1m**

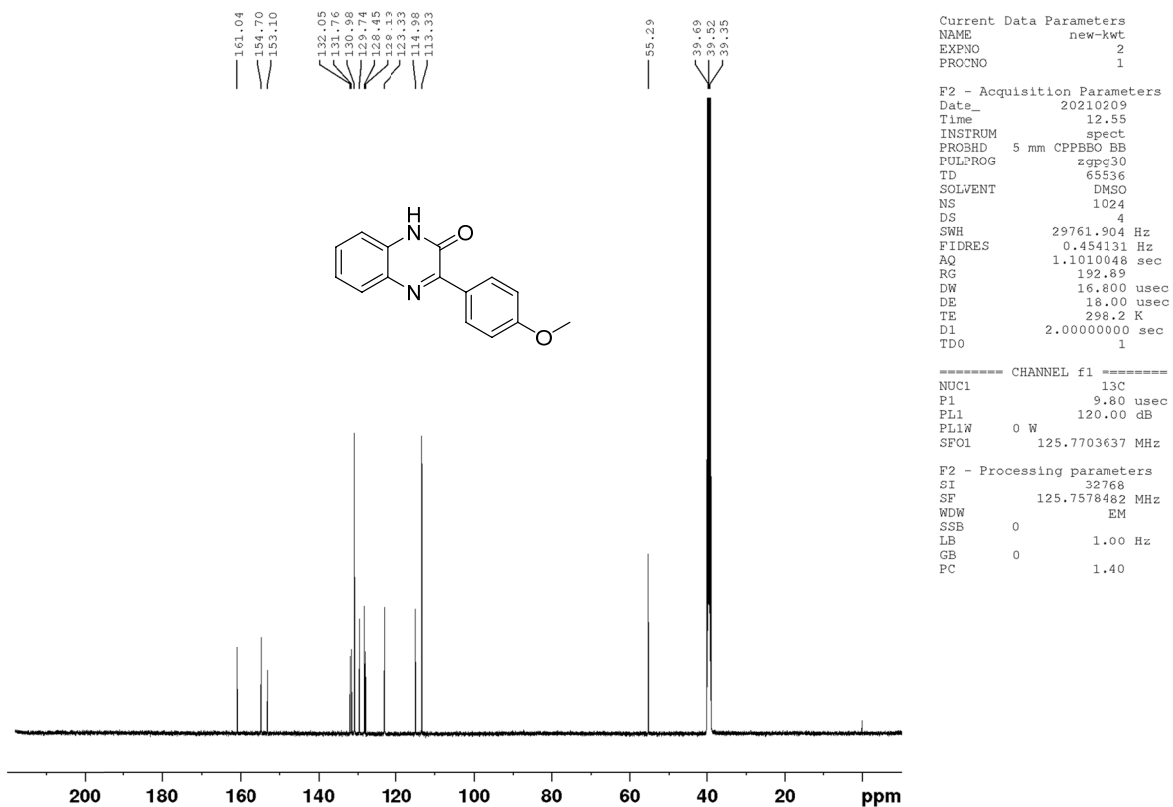
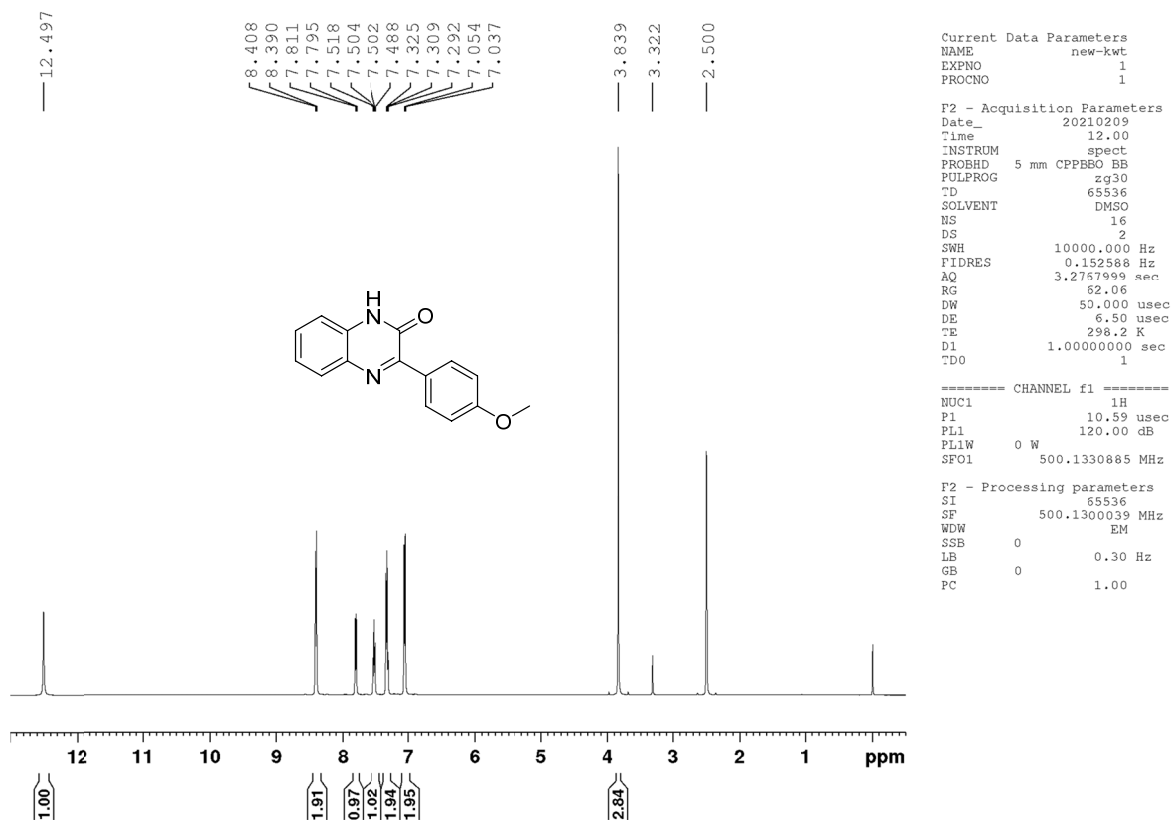


Figure S14. ¹H NMR and ¹³C NMR spectra of **1n**

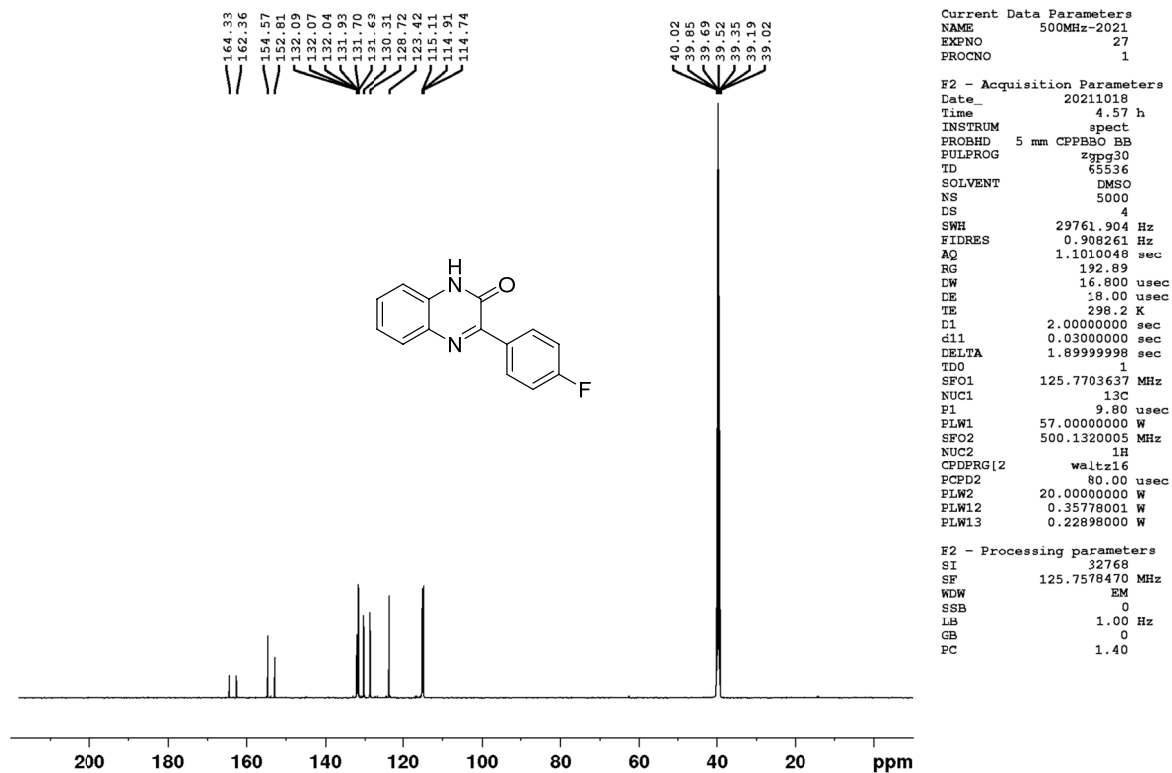
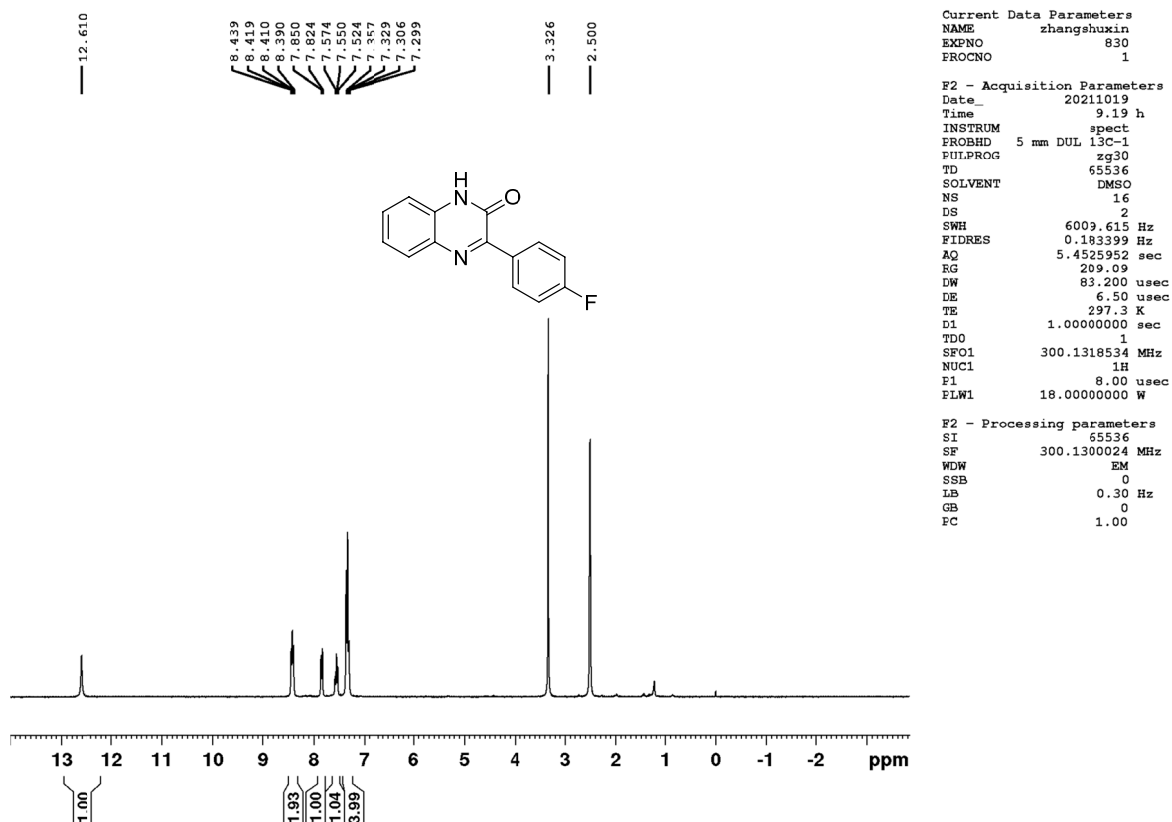
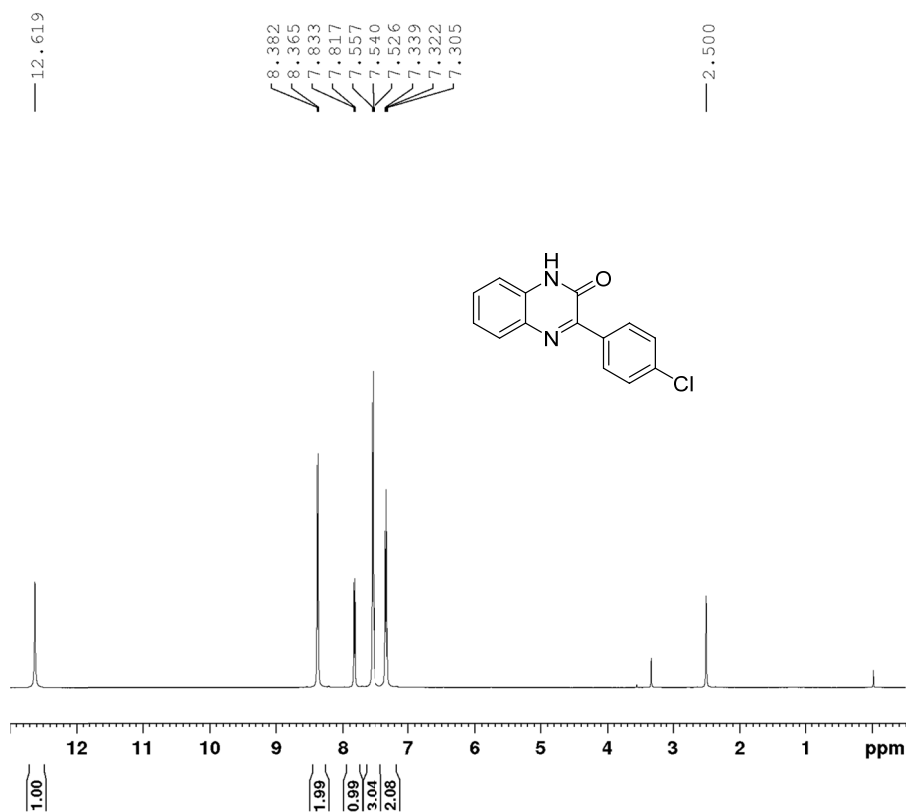


Figure S15. ¹H NMR and ¹³C NMR spectra of **1o**



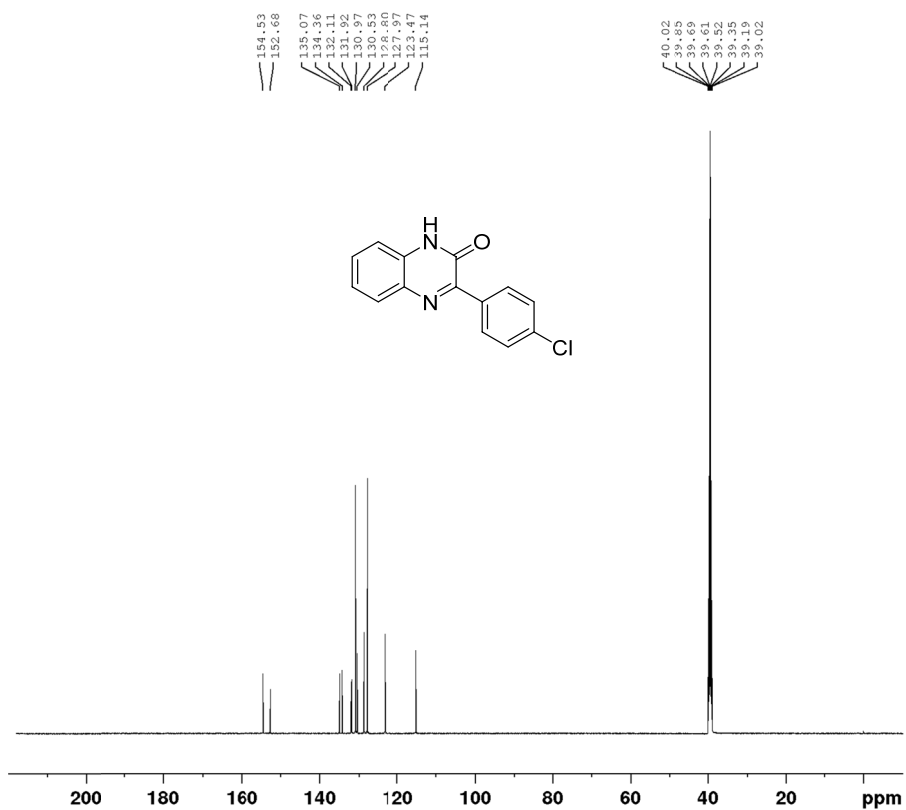
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Current Data Parameters
NAME      new-kwt
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20210209
Time     2.06
INSTRUM  spect
PROBHD   5 mm CPPBBO BB
FULPROG  zg30
TD       65536
SOLVENT  DMSO
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152588 Hz
AQ       3.2767999 sec
RG       55.37
DW       50.000 usec
DE       6.50 usec
TE       298.2 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       10.59 usec
PL1     120.00 dB
PL1W    0 W
SFO1    500.1330885 MHz

F2 - Processing parameters
SI       65536
SF       500.1300037 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```



```

Current Data Parameters
NAME      new-kwt
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20210209
Time     3.01
INSTRUM  spect
PROBHD   5 mm CPPBBO BB
FULPROG  zgpg30
TD       65536
SOLVENT  DMSO
NS       1024
DS       4
SWH      29761.904 Hz
FIDRES   0.454131 Hz
AQ       1.1010048 sec
RG       192.89
DW       16.800 usec
DE       13.00 usec
TE       298.2 K
D1       2.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1       9.80 usec
PL1     120.00 dB
PL1W    0 W
SFO1    125.7703637 MHz

F2 - Processing parameters
SI       32768
SF       125.7578482 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
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Figure S16. ¹H NMR and ¹³C NMR spectra of **1p**

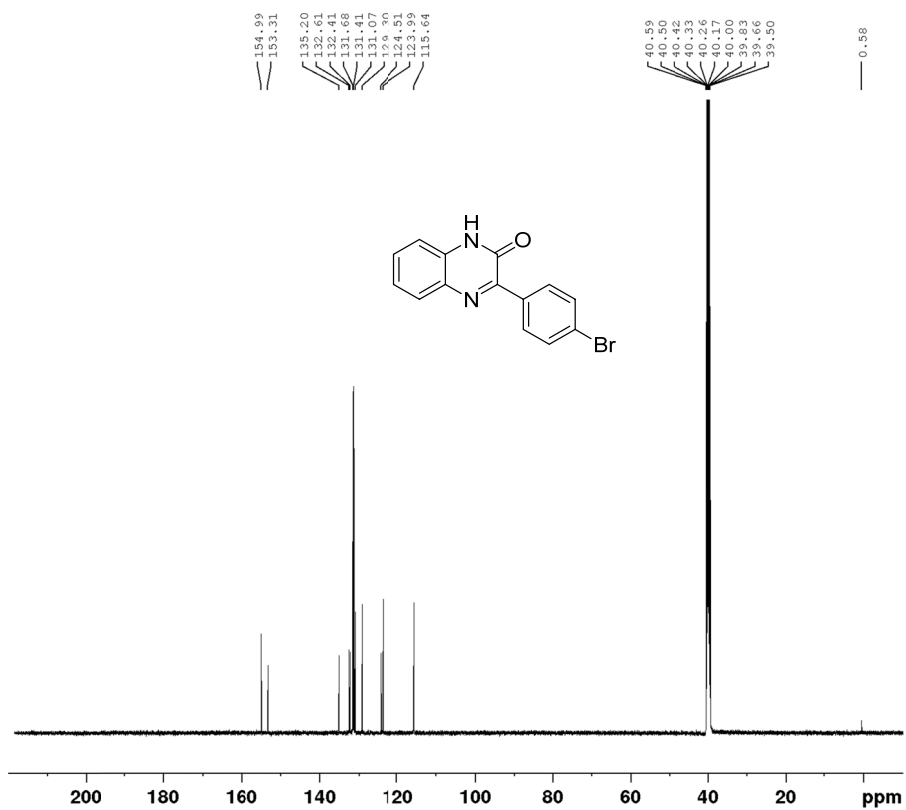
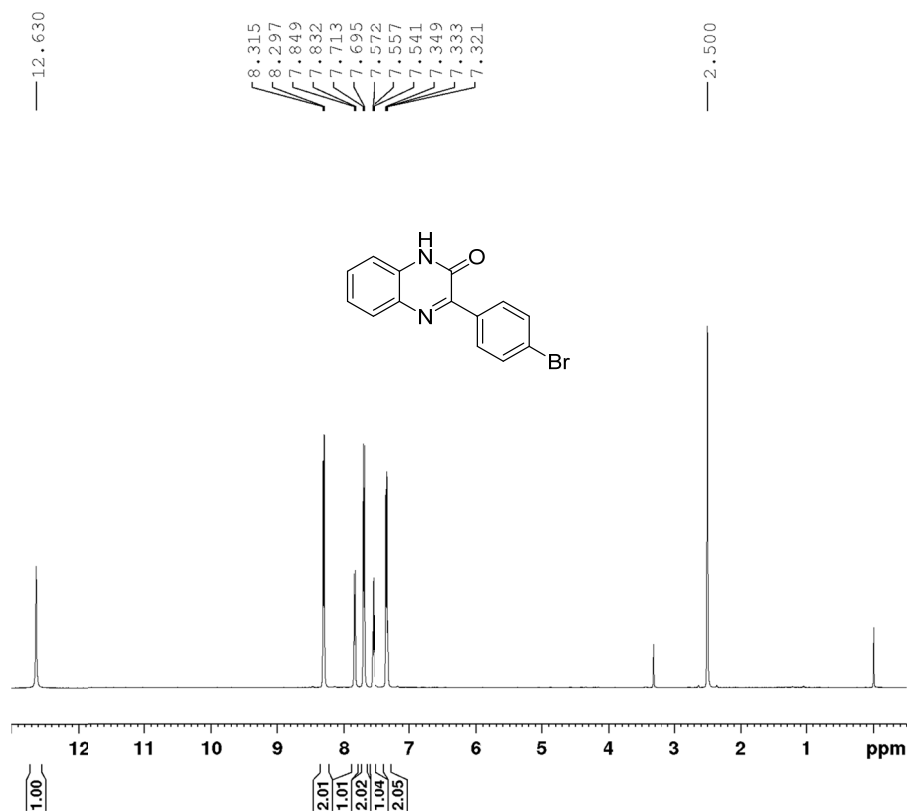
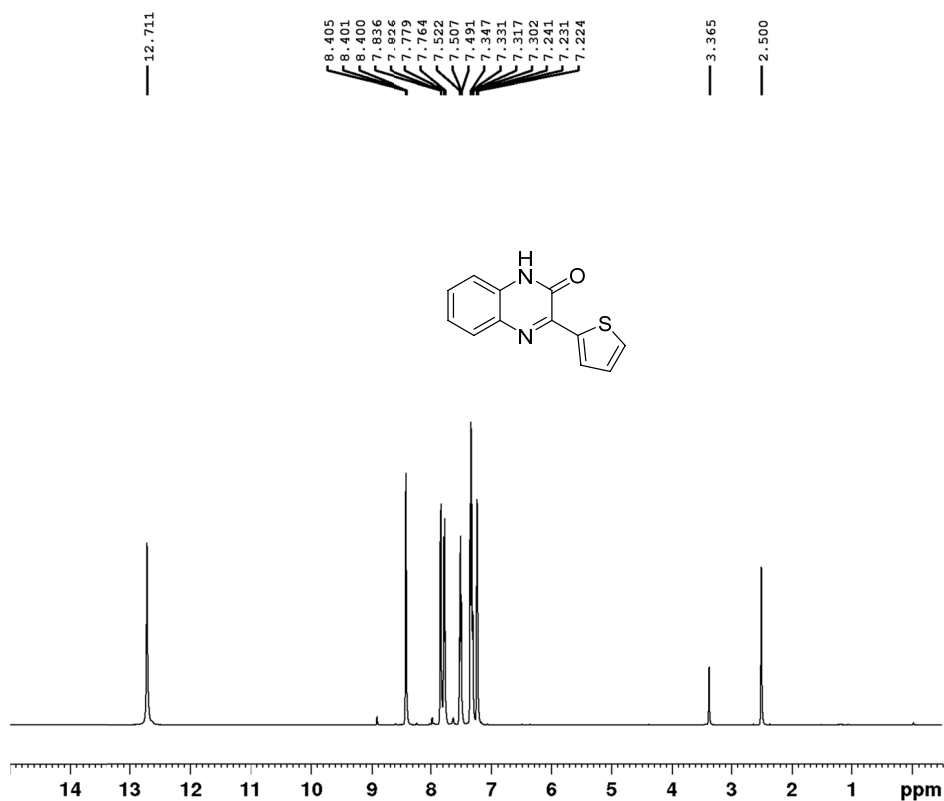


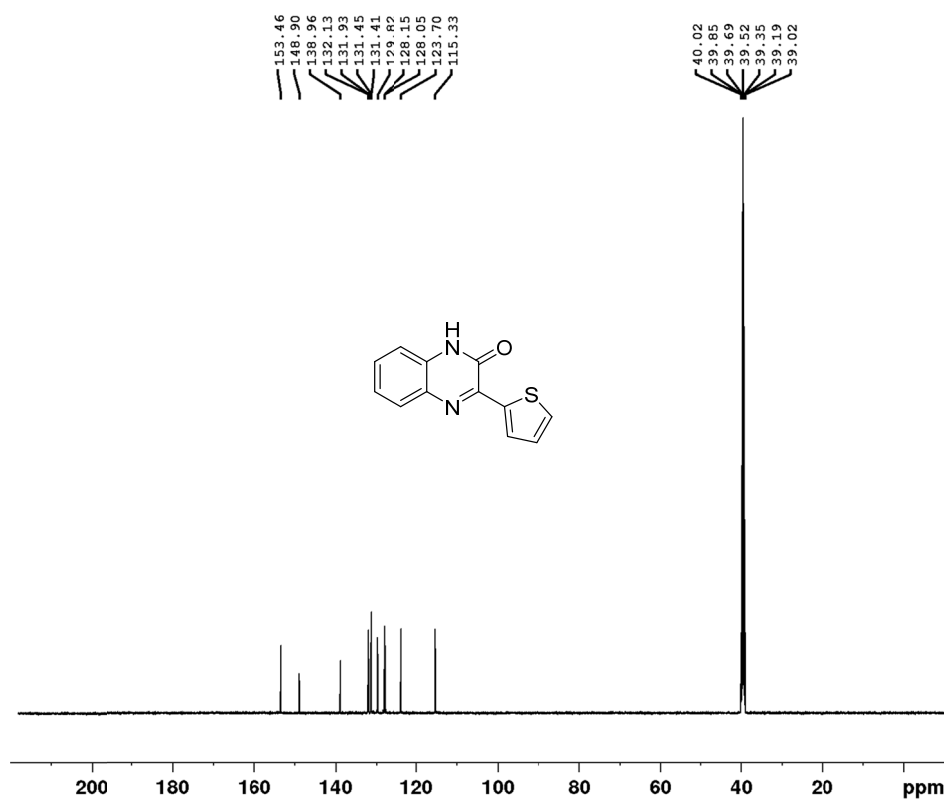
Figure S17. ¹H NMR and ¹³C NMR spectra of **1q**



Current Data Parameters
 NAME 500MHz-2021
 EXPNO 28
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211123
 Time 23.18 h
 INSTRUM spect
 PROBHDD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.305176 Hz
 AQ 3.2767999 sec
 RG 62.06
 DW 50.000 usec
 DE 6.50 usec
 TE 0 K
 D1 1.00000000 sec
 TD0 1
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 10.59 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 65536
 SF 500.1300008 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

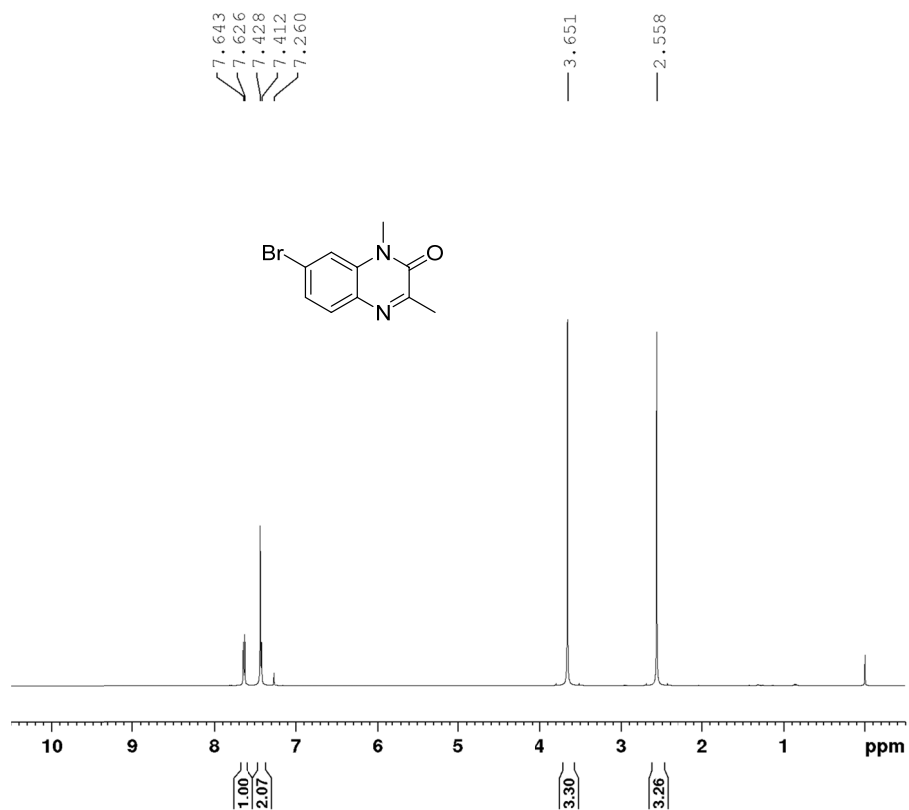


Current Data Parameters
 NAME 500MHz-2021
 EXPNO 29
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211123
 Time 23.23 h
 INSTRUM spect
 PROBHDD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 167
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.908261 Hz
 AQ 1.1010048 sec
 RG 192.89
 DW 16.800 usec
 DE 18.00 usec
 TE 0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.899999998 sec
 TD0 1
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 9.80 usec
 PLW1 57.00000000 W
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 20.00000000 W
 PLW12 0.35778001 W
 PLW13 0.22898000 W

F2 - Processing parameters
 SI 32768
 SF 125.7576422 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S18. ¹H NMR and ¹³C NMR spectra of **1r**



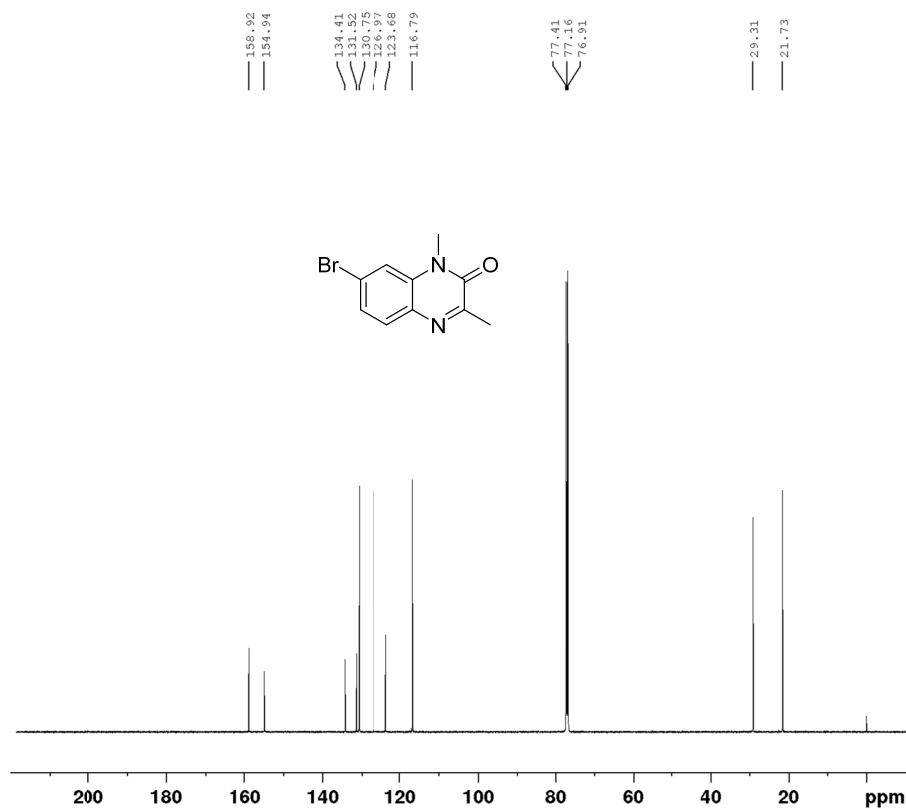
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Current Data Parameters
NAME          new-kwt
EXPNO        1
PROCNO       1

F2 - Acquisition Parameters
Date_        20210209
Time         13.58
INSTRUM      spect
PROBHD       5 mm CPPBBO BB
PULPROG      zg30
TD           65536
SOLVENT      CDCl3
NS           16
DS           2
SWH          10000.000 Hz
FIDRES       0.152588 Hz
AQ           3.2767999 sec
RG           49.27
DW           50.000 usec
DE           6.50 usec
TE           298.2 K
D1           1.00000000 sec
TD0          1

===== CHANNEL f1 =====
NUC1         1H
P1           10.59 usec
PL1          120.00 dB
PL1W         0 W
SF01         500.1330865 MHz

F2 - Processing parameters
SI           65536
SF           500.130123 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           1.00
    
```



```

Current Data Parameters
NAME          new-kwt
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20210209
Time         14.52
INSTRUM      spect
PROBHD       5 mm CPPBBO BB
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           1024
DS           4
SWH          29761.904 Hz
FIDRES       0.454131 Hz
AQ           1.1010048 sec
RG           192.89
DW           16.800 usec
DE           18.00 usec
TE           298.2 K
D1           2.00000000 sec
TD0          1

===== CHANNEL f1 =====
NUC1         13C
P1           9.80 usec
PL1          120.00 dB
PL1W         0 W
SF01         125.7703637 MHz

F2 - Processing parameters
SI           32768
SF           125.7577737 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
    
```

Figure S19. ¹H NMR and ¹³C NMR spectra of **3a**

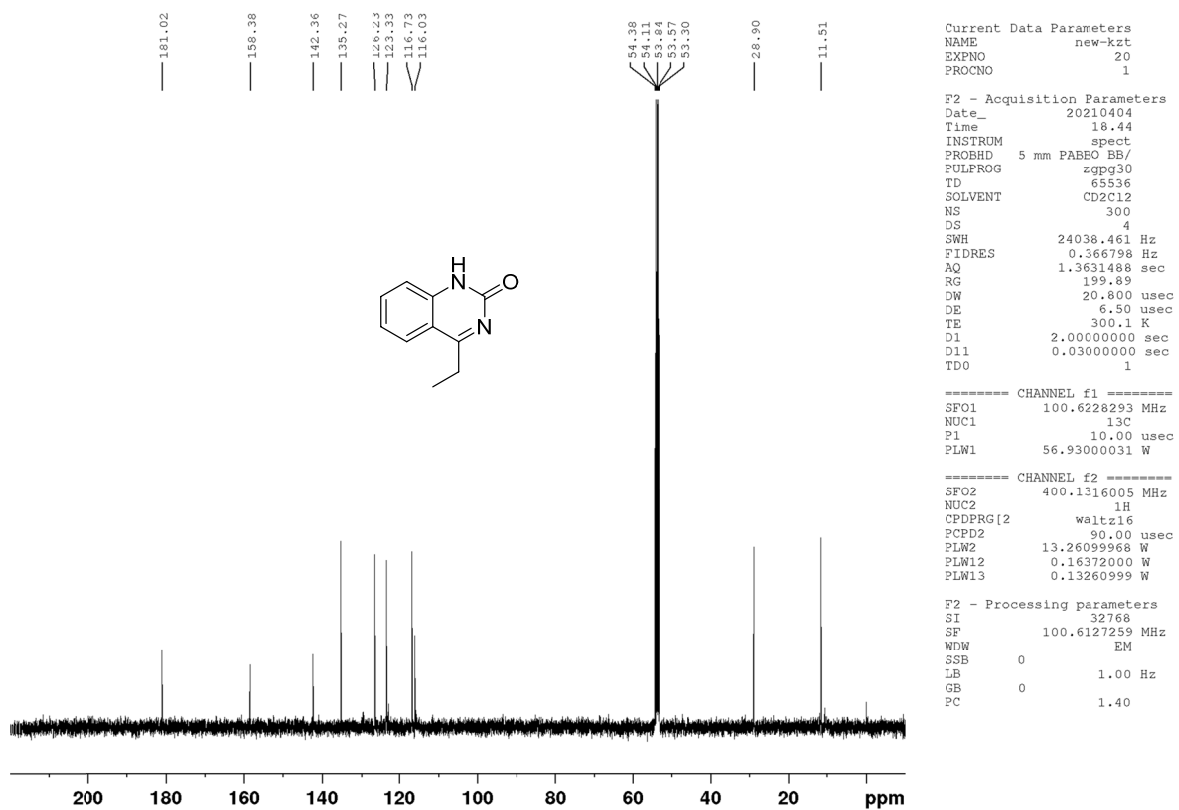
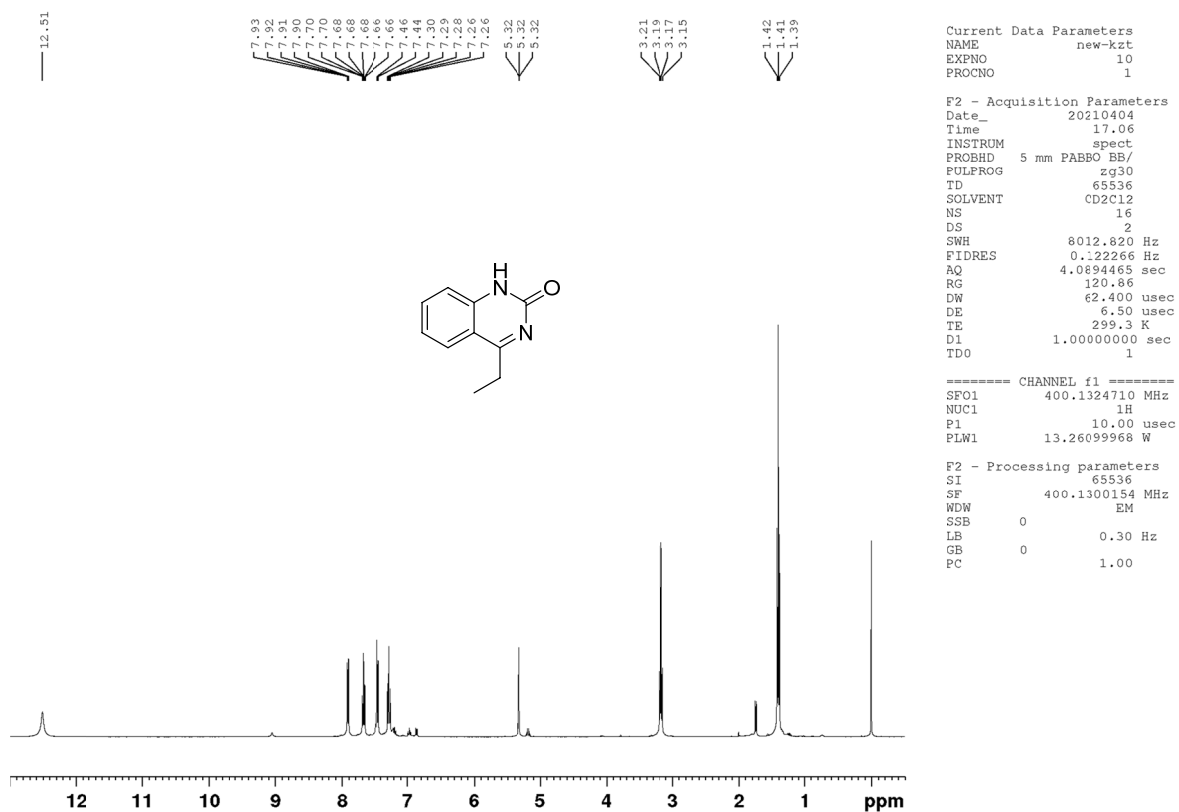


Figure S20. ¹H NMR and ¹³C NMR spectra of **3b**

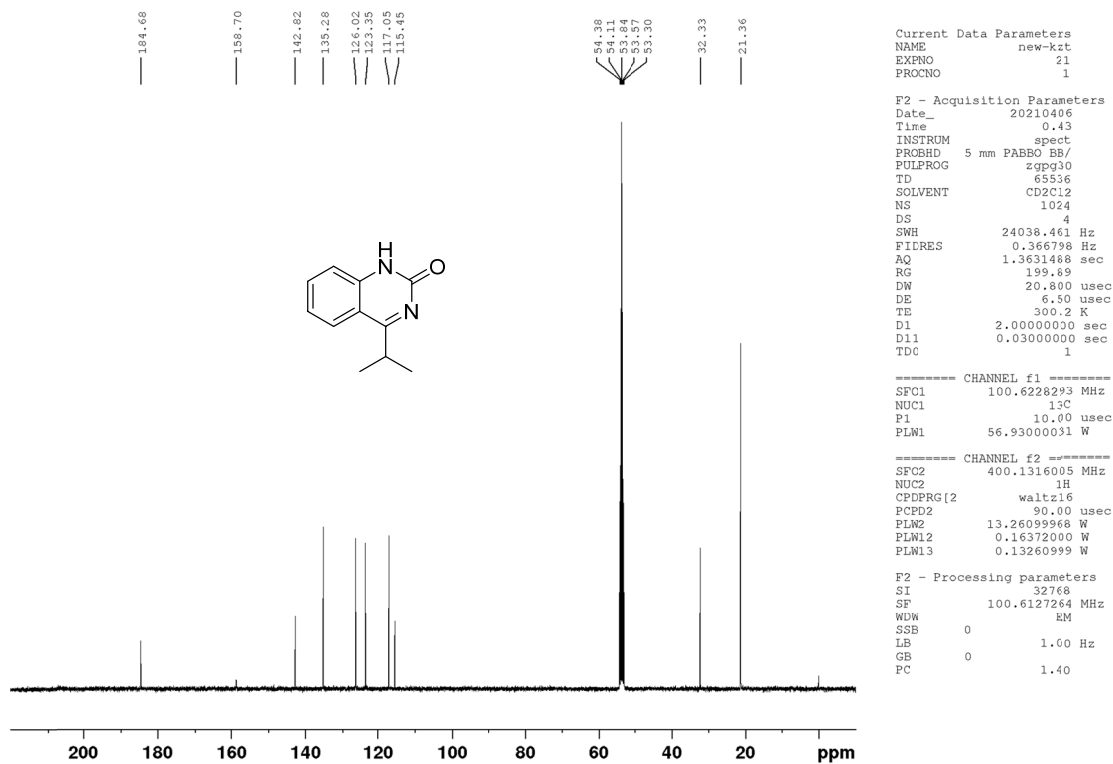
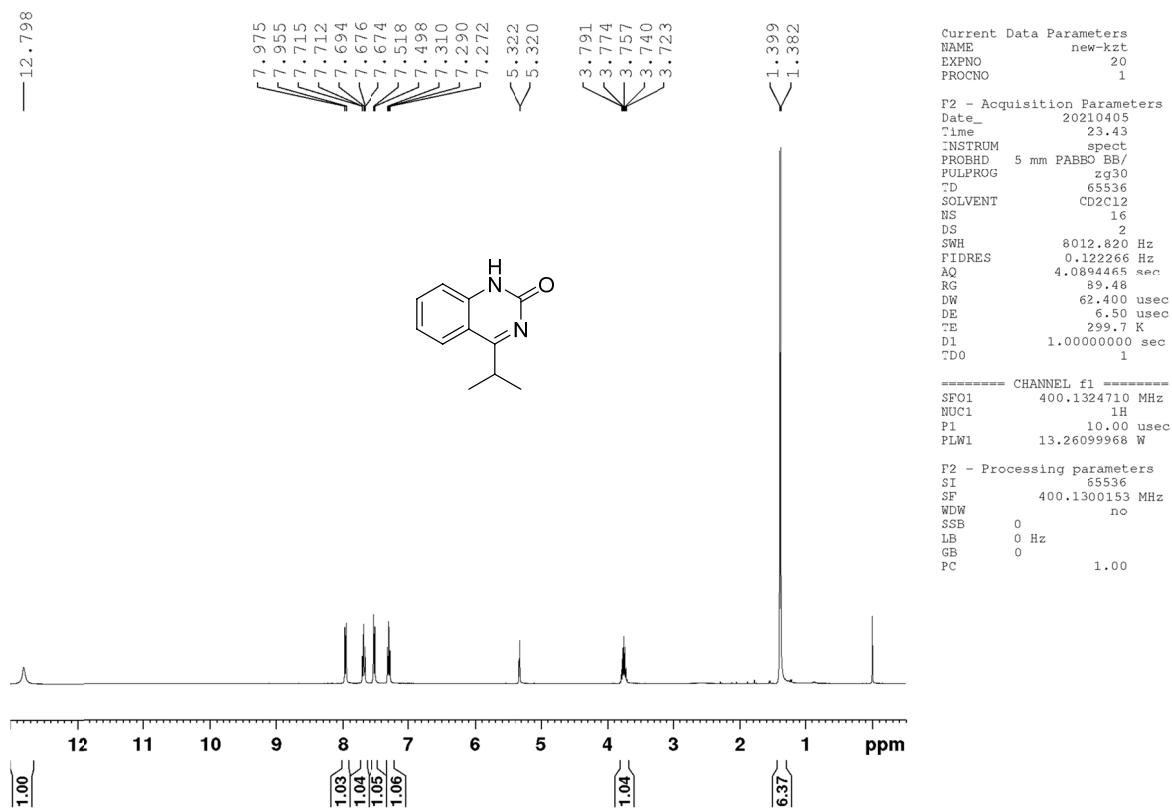


Figure S21. ¹H NMR and ¹³C NMR spectra of **3c**

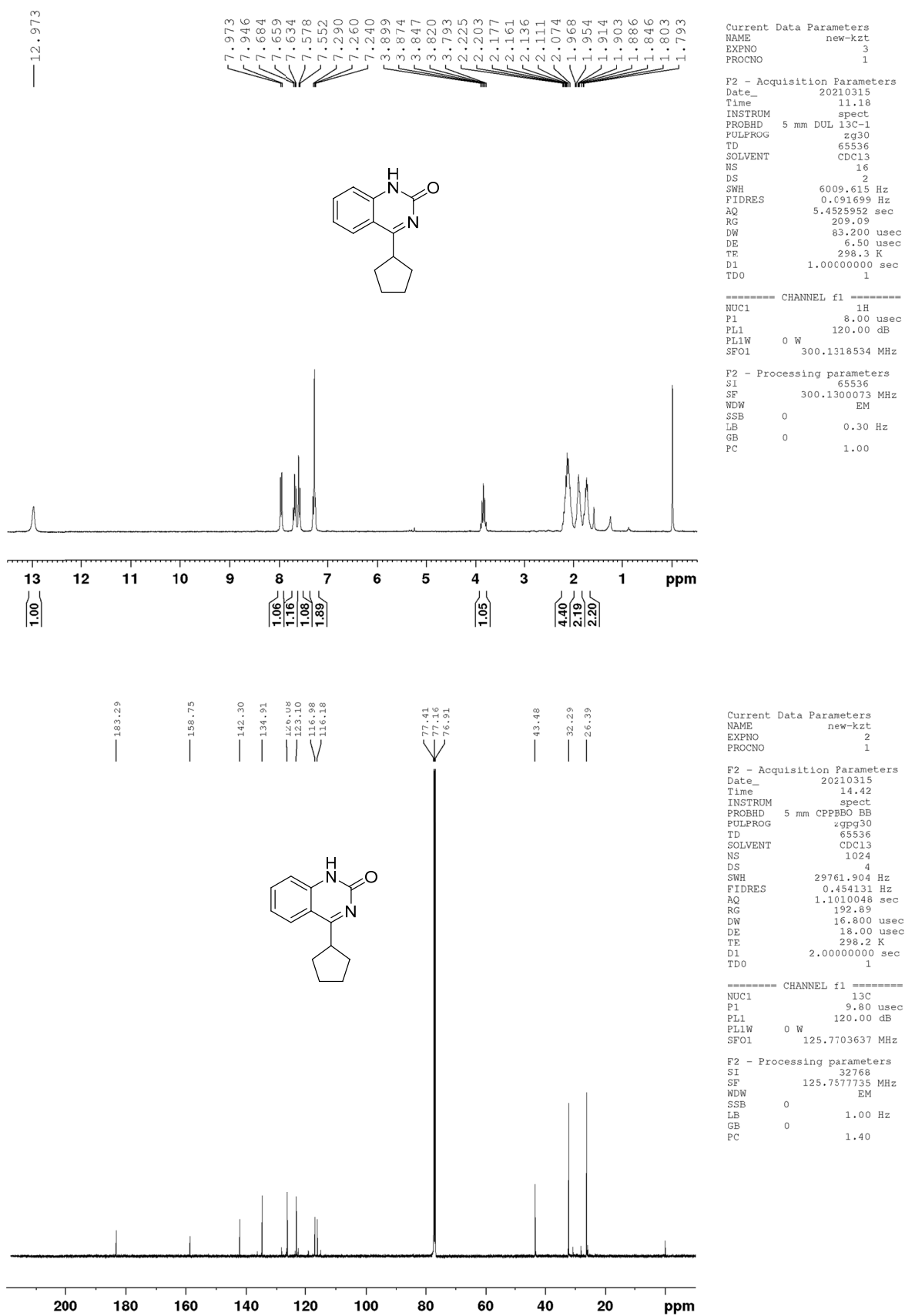
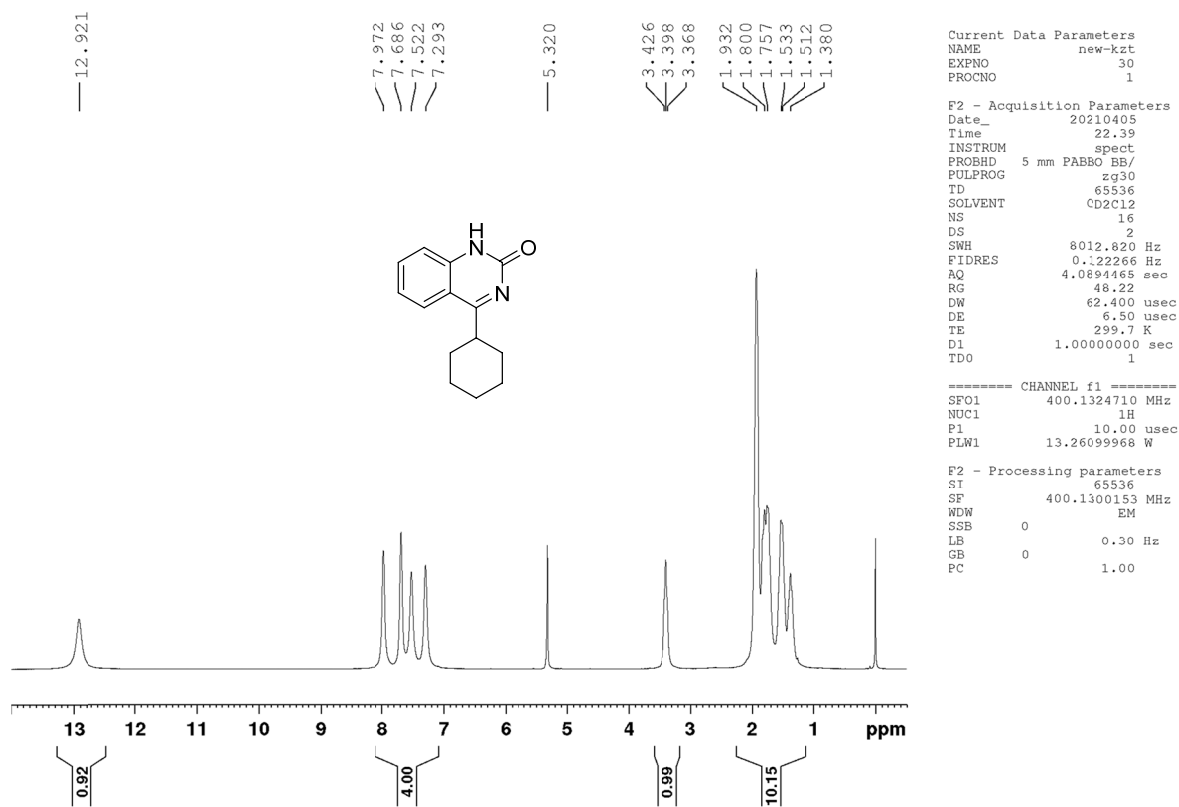


Figure S22. ¹H NMR and ¹³C NMR spectra of **3d**

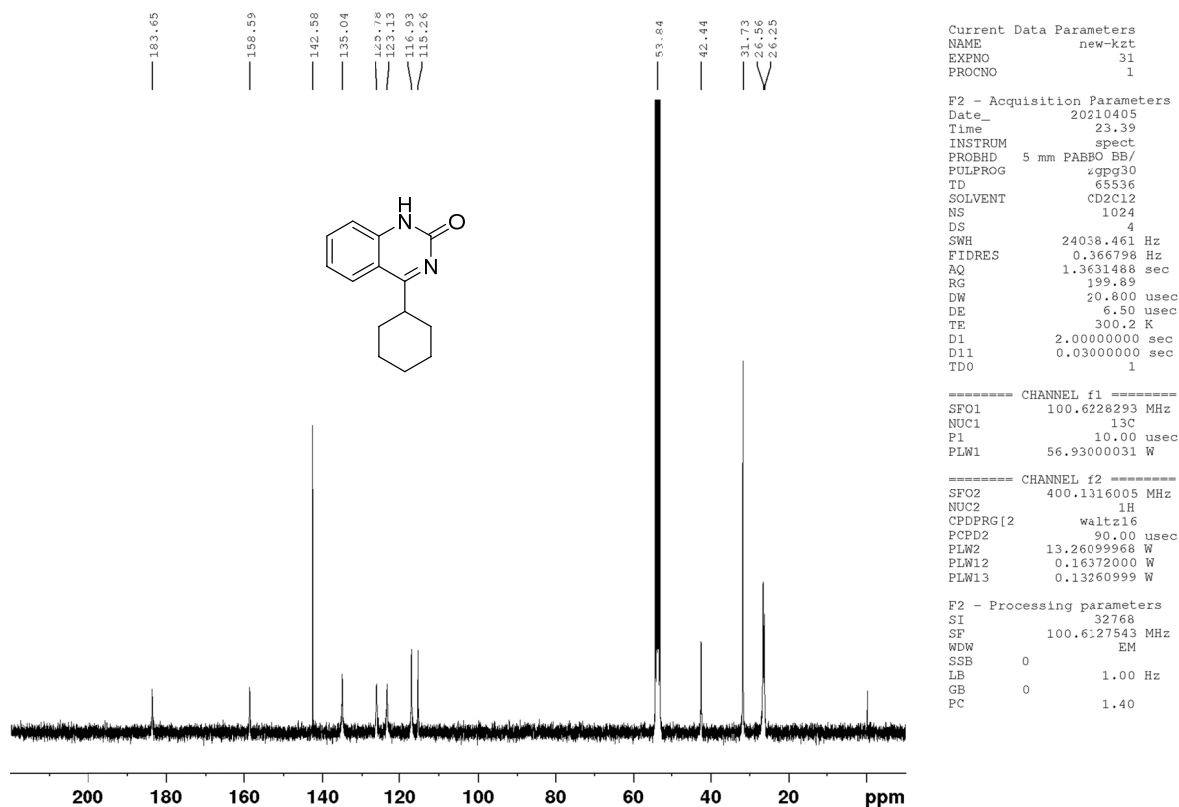


Current Data Parameters
 NAME new-kzt
 EXPNO 30
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210405
 Time 22.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CD2Cl2
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 48.22
 DW 62.400 usec
 DE 6.50 usec
 TE 299.7 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 13.26099968 W

F2 - Processing parameters
 SI 65536
 SF 400.1300153 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME new-kzt
 EXPNO 31
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210405
 Time 23.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT CD2Cl2
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 199.89
 DW 20.800 usec
 DE 6.50 usec
 TE 300.2 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 100.6228293 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 56.93000031 W

===== CHANNEL f2 =====
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 90.00 usec
 PLW2 13.26099968 W
 PLW12 0.16372000 W
 PLW13 0.13260999 W

F2 - Processing parameters
 SI 32768
 SF 100.627543 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S23. ¹H NMR and ¹³C NMR spectra of 3e

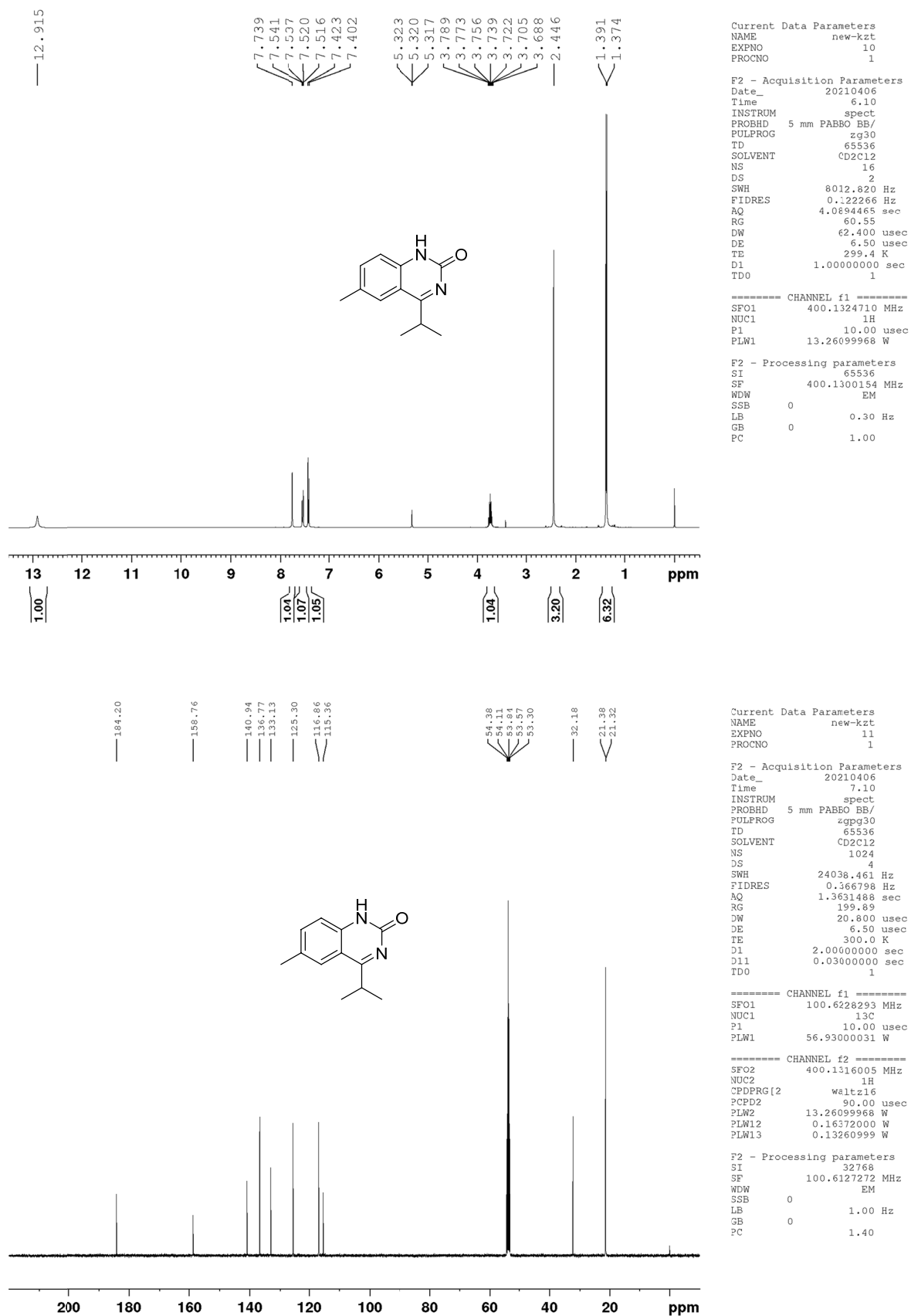
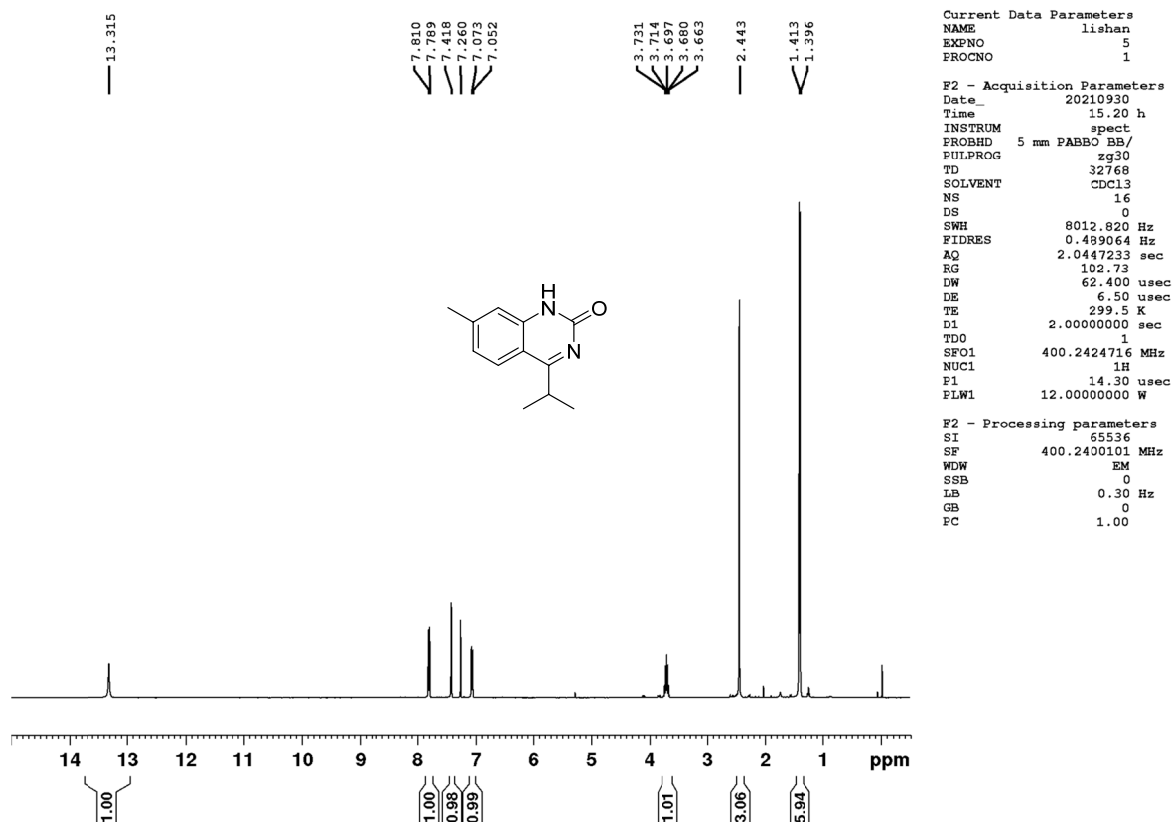


Figure S24. ¹H NMR and ¹³C NMR spectra of **3f**

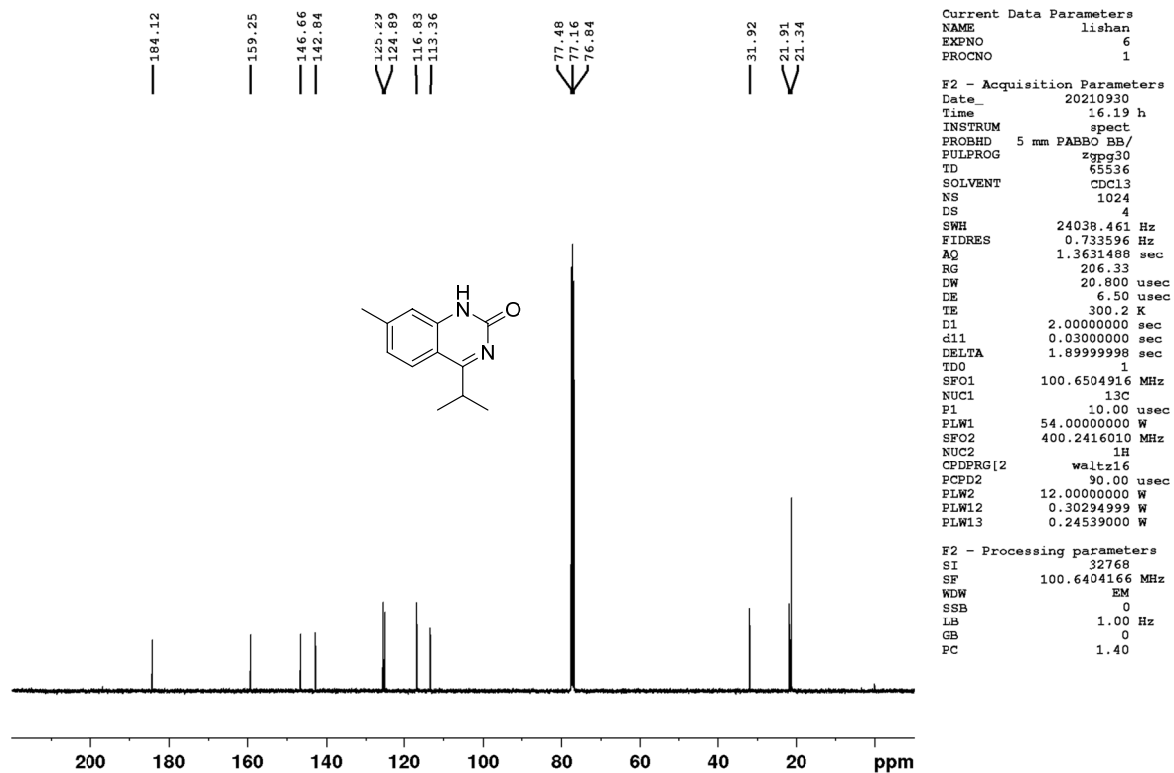


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Current Data Parameters
NAME      lishan
EXPNO     5
PROCNO    1

F2 - Acquisition Parameters
Date_     20210930
Time      15.20 h
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         0
SWH        8012.820 Hz
FIDRES     0.489064 Hz
AQ         2.0447233 sec
RG         102.73
EW         62.400 usec
DE         6.50 usec
TE         299.5 K
D1         2.0000000 sec
TDO        1
SFO1       400.2424716 MHz
NUC1       1H
F1         14.30 usec
FLW1       12.0000000 W

F2 - Processing parameters
SI         65536
SF         400.2400101 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
    
```



```

Current Data Parameters
NAME      lishan
EXPNO     6
PROCNO    1

F2 - Acquisition Parameters
Date_     20210930
Time      16.19 h
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg30
TD         55536
SOLVENT   CDCl3
NS         1024
DS         4
SWH        24038.461 Hz
FIDRES     0.733596 Hz
AQ         1.3631488 sec
RG         206.33
EW         20.800 usec
DE         6.50 usec
TE         300.2 K
D1         2.0000000 sec
d11        0.0300000 sec
DELTA      1.899999998 sec
TDO        1
SFO1       100.6504916 MHz
NUC1       13C
F1         10.00 usec
PLW1       54.0000000 W
SFO2       400.2416010 MHz
NUC2       1H
CPDPRG2    waltz16
ECPD2      90.00 usec
PLW2       12.0000000 W
PLW12      0.30294999 W
PLW13      0.24539000 W

F2 - Processing parameters
SI         32768
SF         100.6404166 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
    
```


Figure S25. ¹H NMR and ¹³C NMR spectra of **3g**

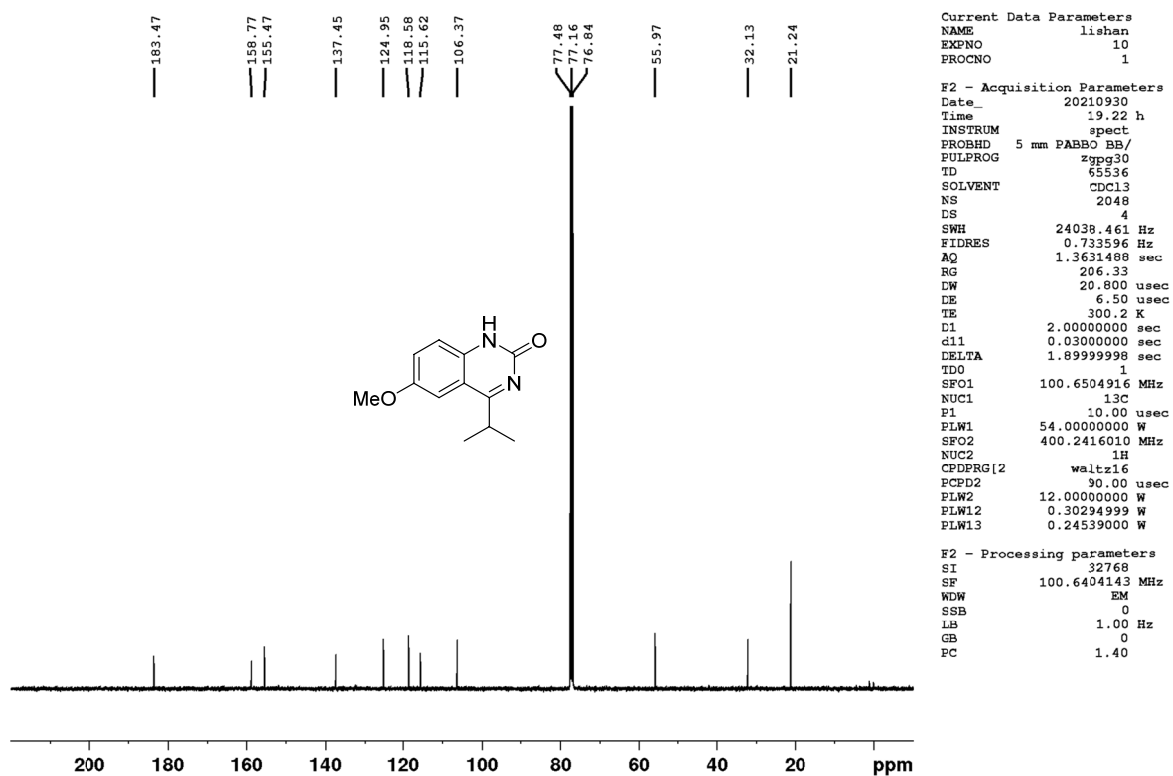
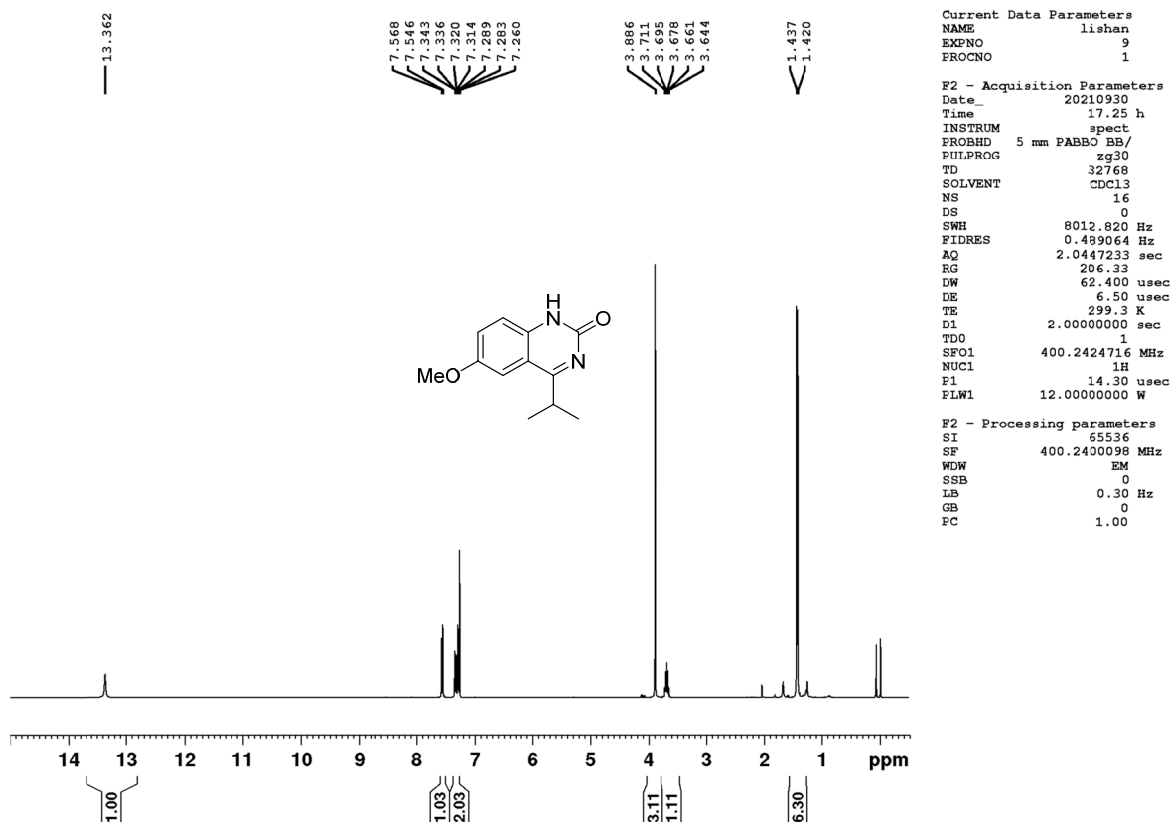
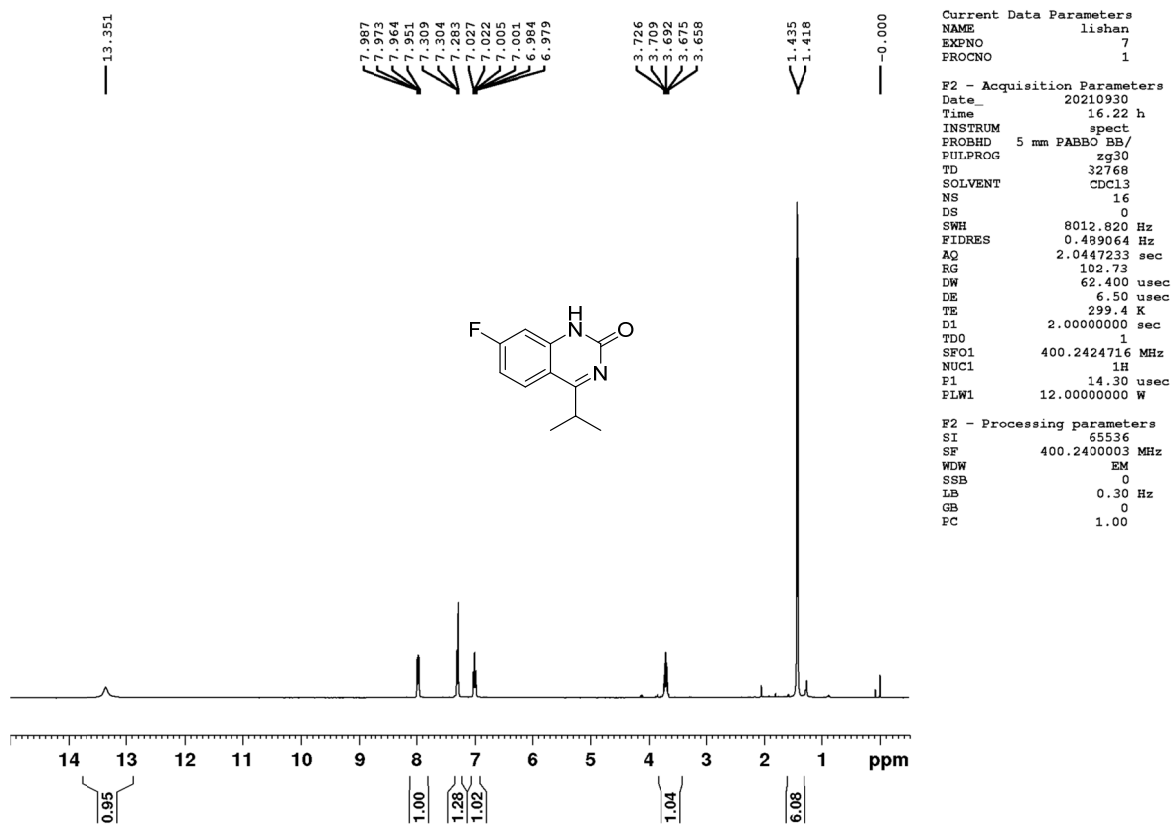


Figure S26. ¹H NMR and ¹³C NMR spectra of **3h**

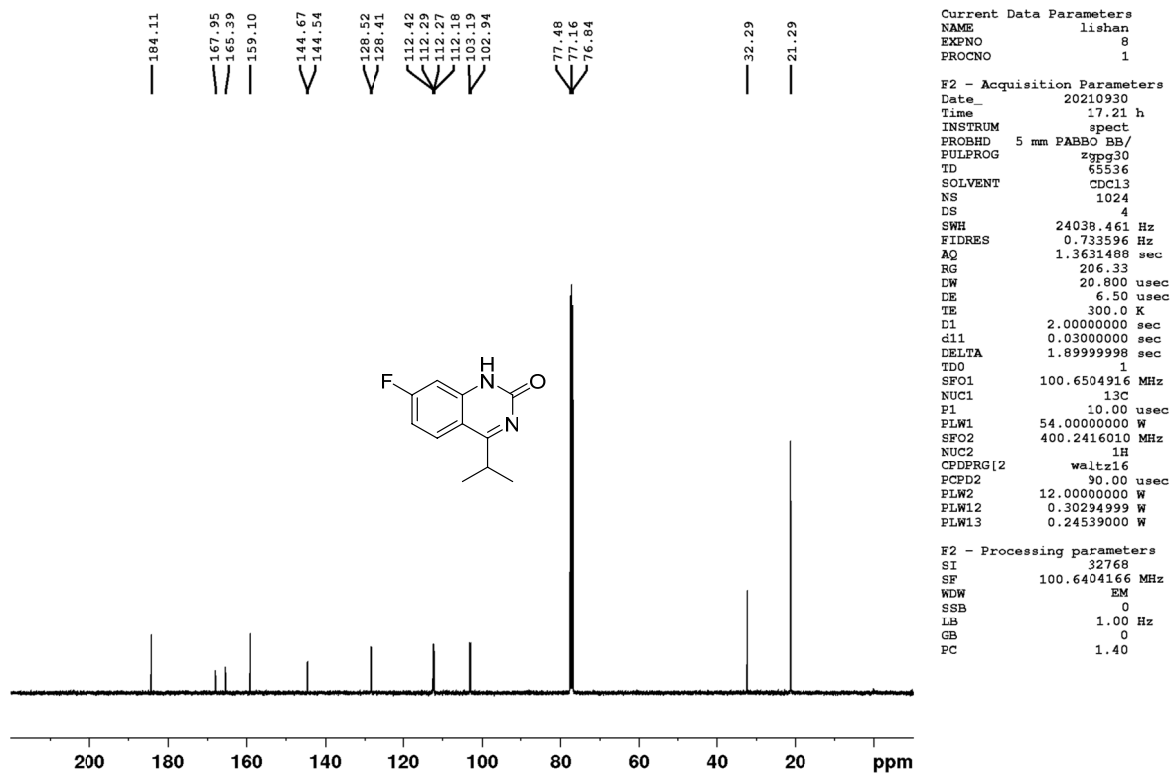


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Current Data Parameters
NAME      lishan
EXPNO    7
PROCNO   1

F2 - Acquisition Parameters
Date_    20210930
Time     16.22 h
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD       32768
SOLVENT  CDCl3
NS       16
DS       0
SWH      8012.820 Hz
FIDRES   0.489064 Hz
AQ       2.0447233 sec
RG       102.73
EW       62.400 usec
DE       6.50 usec
TE       299.4 K
D1       2.0000000 sec
TD0      1
SFO1     400.2424716 MHz
NUC1     1H
F1       14.30 usec
FLW1     12.0000000 W

F2 - Processing parameters
SI       65536
SF       400.2400003 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
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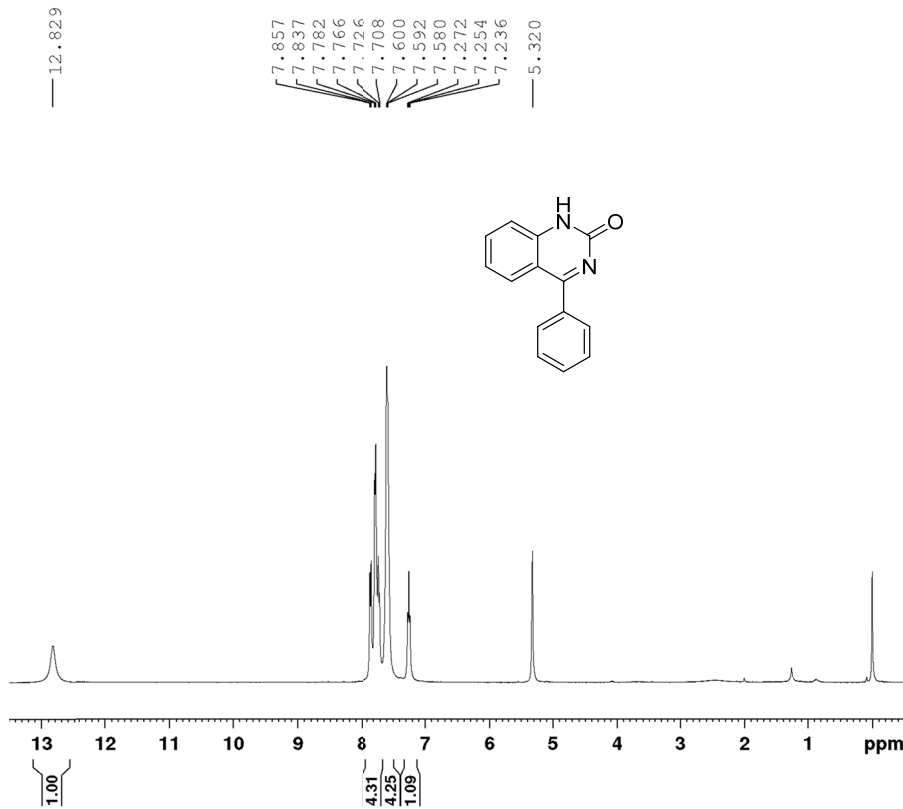
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Current Data Parameters
NAME      lishan
EXPNO    8
PROCNO   1

F2 - Acquisition Parameters
Date_    20210930
Time     17.21 h
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg30
TD       55536
SOLVENT  CDCl3
NS       1024
DS       4
SWH      24038.461 Hz
FIDRES   0.733596 Hz
AQ       1.3631488 sec
RG       206.33
EW       20.800 usec
DE       6.50 usec
TE       300.0 K
D1       2.0000000 sec
d11      0.0300000 sec
DELTA    1.89999999 sec
TD0      1
SFO1     100.6504916 MHz
NUC1     13C
P1       10.00 usec
PLW1     54.0000000 W
SFO2     400.2416010 MHz
NUC2     1H
CPDPRG2  waltz16
ECPD2    90.00 usec
ELW2     12.0000000 W
ELW12    0.30294999 W
ELW13    0.24539000 W

F2 - Processing parameters
SI       32768
SF       100.6404166 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```

Figure S27. ¹H NMR and ¹³C NMR spectra of **3i**



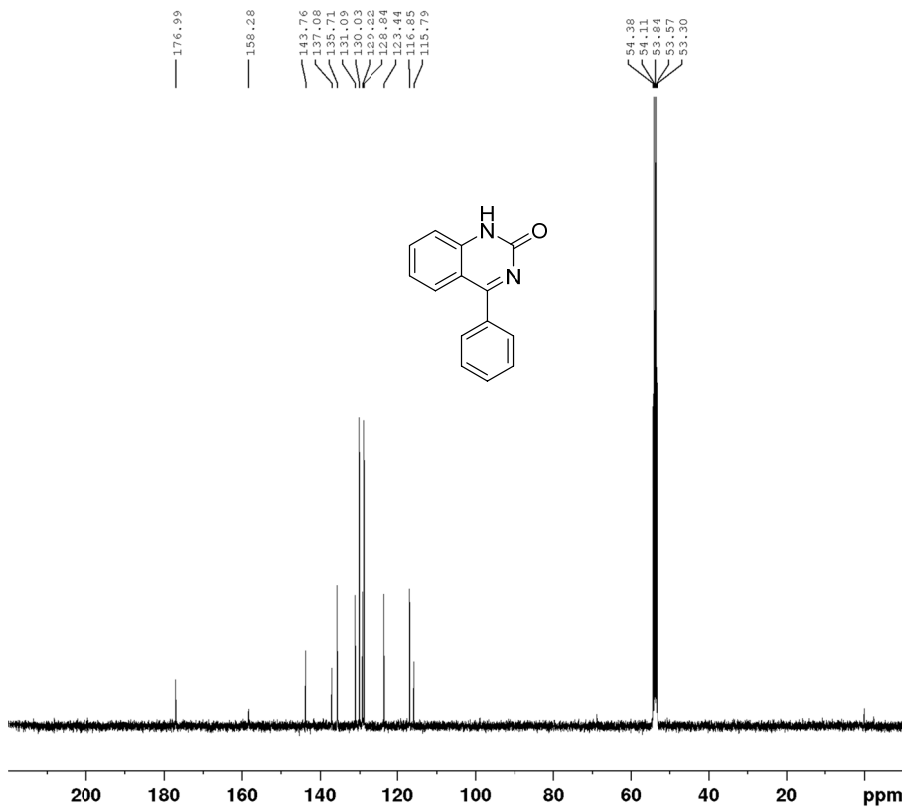
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Current Data Parameters
NAME          new-kzt
EXPNO        10
PROCNO       1

F2 - Acquisition Parameters
Date_        20210406
Time         7.15
INSTRUM     spect
PROBHD      5 mm PABBO BB/
PULPROG     zg30
TD           65536
SOLVENT     CD2C12
NS           16
DS           2
SWH          8012.820 Hz
FIDRES      0.122266 Hz
AQ           4.0894465 sec
RG           112.4
DW           62.400 usec
DE           6.50 usec
TE           299.5 K
D1           1.0000000 sec
TDO         1

----- CHANNEL f1 -----
SFO1        400.1324710 MHz
NUC1         1H
P1           10.00 usec
PLW1        13.26099968 W

F2 - Processing parameters
SI           65536
SF           400.1300157 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           1.00
    
```



```

Current Data Parameters
NAME          new-kzt
EXPNO        11
PROCNO       1

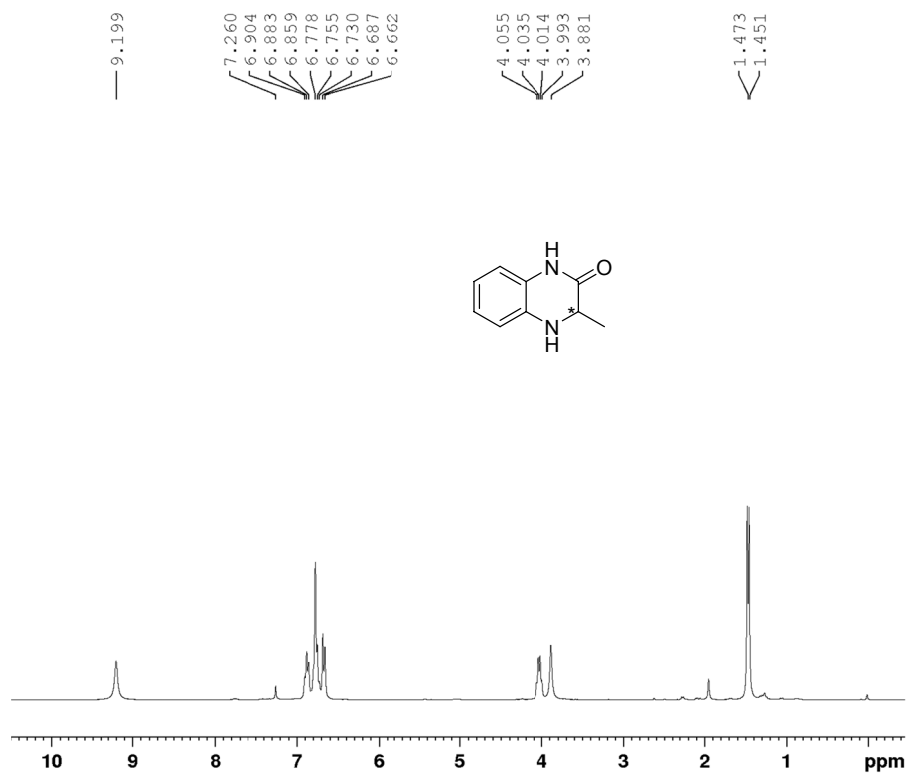
F2 - Acquisition Parameters
Date_        20210406
Time         8.15
INSTRUM     spect
PROBHD      5 mm PABBO BB/
PULPROG     zgpg30
TD           65536
SOLVENT     CD2C12
NS           1024
DS           4
SWH          24038.461 Hz
FIDRES      0.366798 Hz
AQ           1.3631488 sec
RG           199.89
DW           20.800 usec
DE           6.50 usec
TE           300.0 K
D1           2.0000000 sec
D11          0.03000000 sec
TDO         1

----- CHANNEL f1 -----
SFO1        100.6228293 MHz
NUC1         13C
P1           10.00 usec
PLW1        56.93000031 W

----- CHANNEL f2 -----
SFO2        400.1316005 MHz
NUC2         1H
PCPDPRG[2]  waltz16
PCPD2       90.00 usec
PLW2        13.26099968 W
PLW12       0.16372000 W
PLW13       0.13260999 W

F2 - Processing parameters
SI           32768
SF           100.6127266 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
    
```

Figure S28. ¹H NMR and ¹³C NMR spectra of 2a

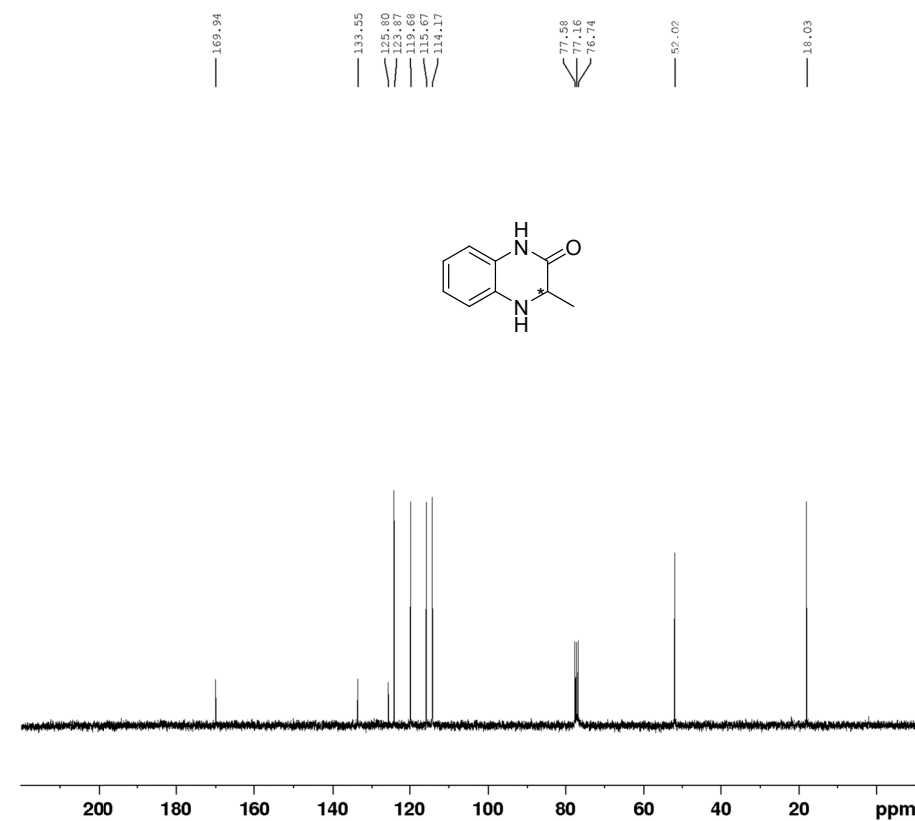


Current Data Parameters
 NAME lichenghao
 EXPNO 31
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 18.58
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 166.41
 DW 83.200 usec
 DE 6.50 usec
 TE 298.3 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300072 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



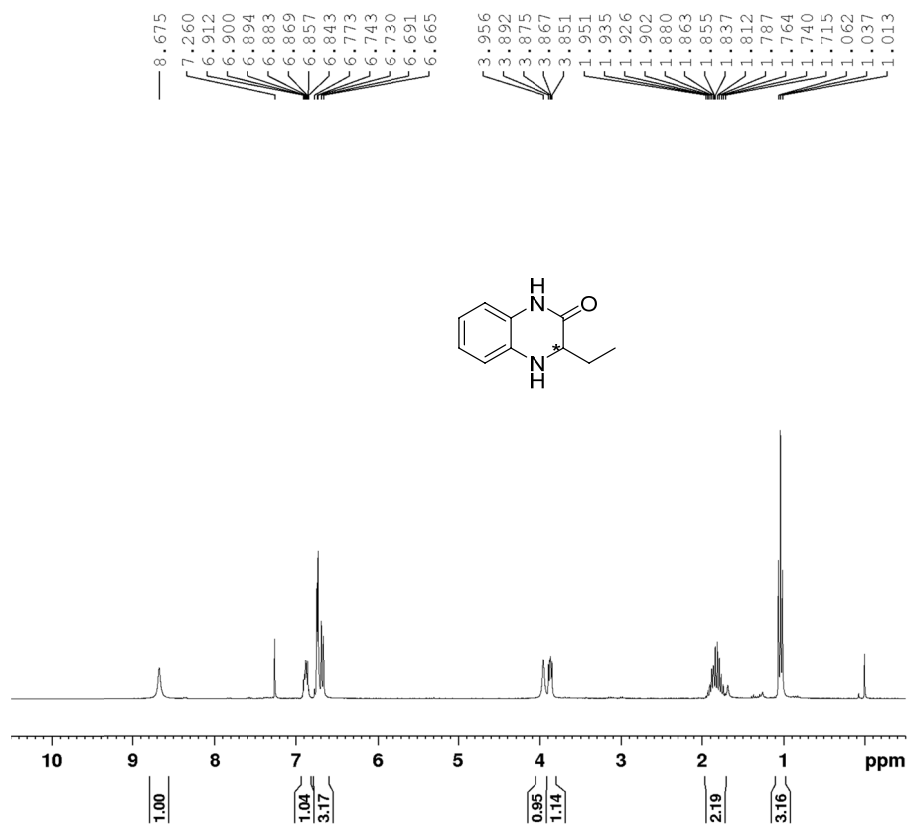
Current Data Parameters
 NAME lichenghao
 EXPNO 32
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 19.08
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 83
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175317 sec
 RG 209.09
 DW 27.733 usec
 DE 6.50 usec
 TE 298.2 K
 D1 2.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677421 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S29 ¹H NMR and ¹³C NMR spectra of **2b**



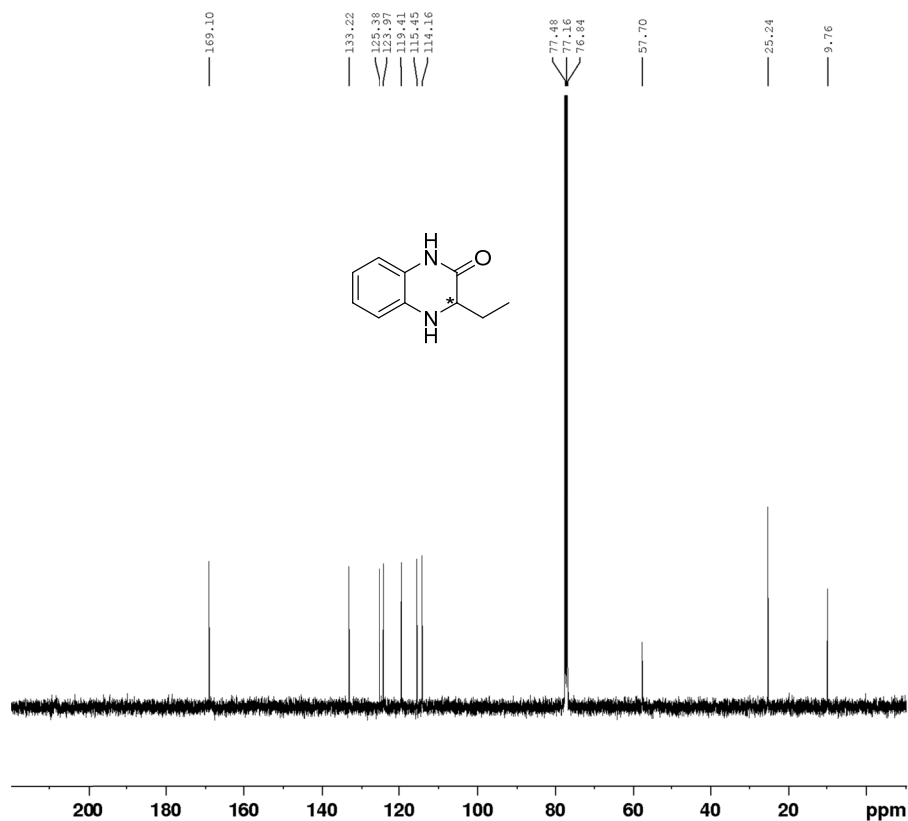
```

Current Data Parameters
NAME          KWT-Et-gh
EXPNO        3
PROCNO       1

F2 - Acquisition Parameters
Date_        20210427
Time         22.04
INSTRUM      spect
PROBHD       5 mm DUL 13C-1
PULPROG      zg30
TD           65536
SOLVENT      CDC13
NS           16
DS           2
SWH          6009.615 Hz
FIDRES       0.091699 Hz
AQ           5.4525952 sec
RG           209.09
DW           83.200 usec
DE           6.50 usec
TE           298.4 K
D1           1.00000000 sec
TD0          1

===== CHANNEL f1 =====
NUC1         1H
P1           8.00 usec
PL1         120.00 dB
PL1W        0 W
SFO1        300.1318534 MHz

F2 - Processing parameters
SI           65536
SF           300.1300072 MHz
WDW         EM
SSB         0
LB          0.30 Hz
GB          0
PC          1.00
    
```



```

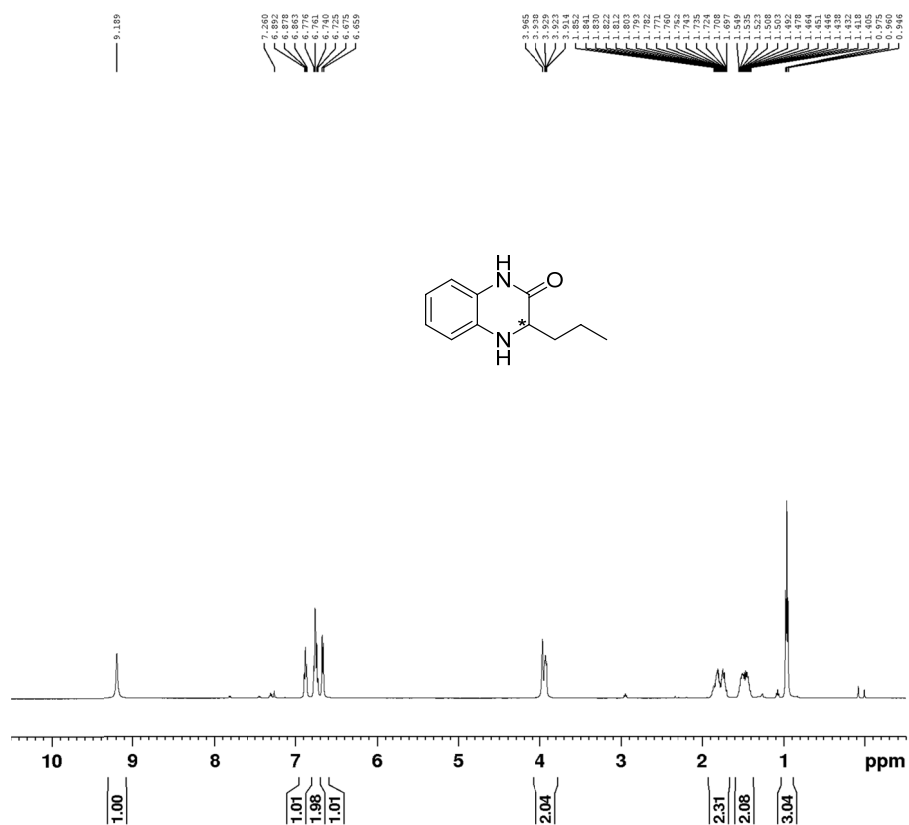
Current Data Parameters
NAME          KWT-Et-gh-C
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20210428
Time         1.14
INSTRUM      spect
PROBHD       5 mm PABBO BB/
PULPROG      zgpg30
TD           65536
SOLVENT      CDC13
NS           1024
DS           4
SWH          24038.461 Hz
FIDRES       0.366798 Hz
AQ           1.3531488 sec
RG           206.33
DW           20.800 usec
DE           6.50 usec
TE           298.9 K
D1           2.00000000 sec
TD0          1

===== CHANNEL f1 =====
NUC1         13C
P1           10.00 usec
PL1         120.00 dB
PL1W        0 W
SFO1        100.6504916 MHz

F2 - Processing parameters
SI           32768
SF           100.6404153 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          1.40
    
```

Figure S30. ¹H NMR and ¹³C NMR spectra of 2c

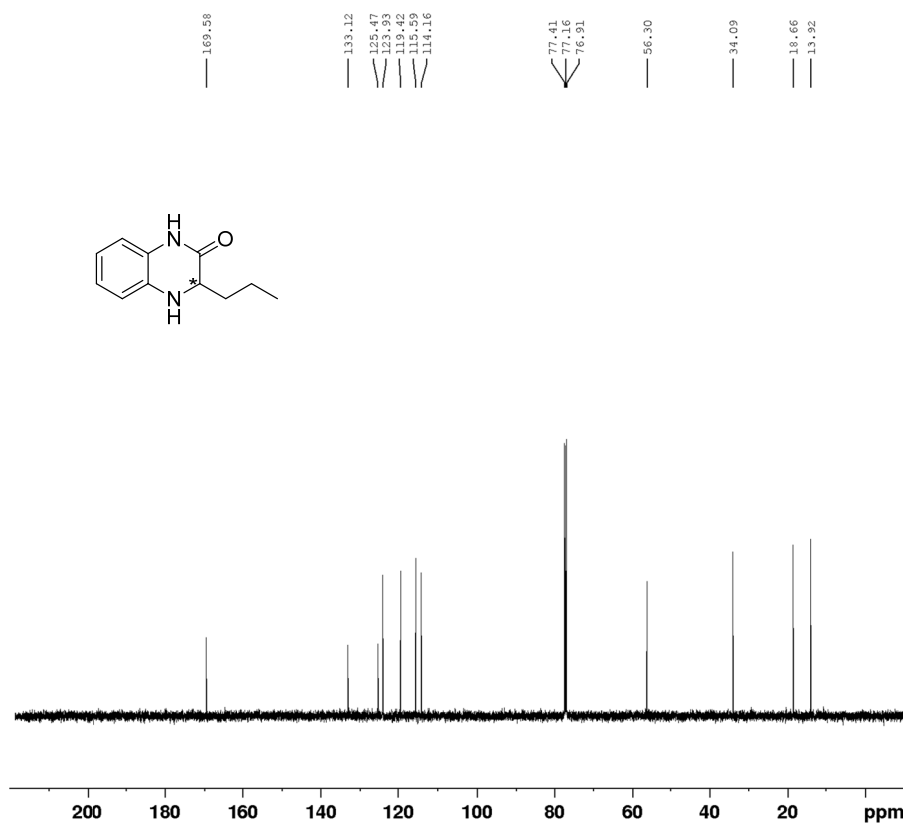


Current Data Parameters
 NAME KWI-npr-gh
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 13.24
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 31.72
 DW 50.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.59 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300121 MHz
 WDW EM
 SSB 0
 LB 0.3 Hz
 GB 0
 PC 1.00



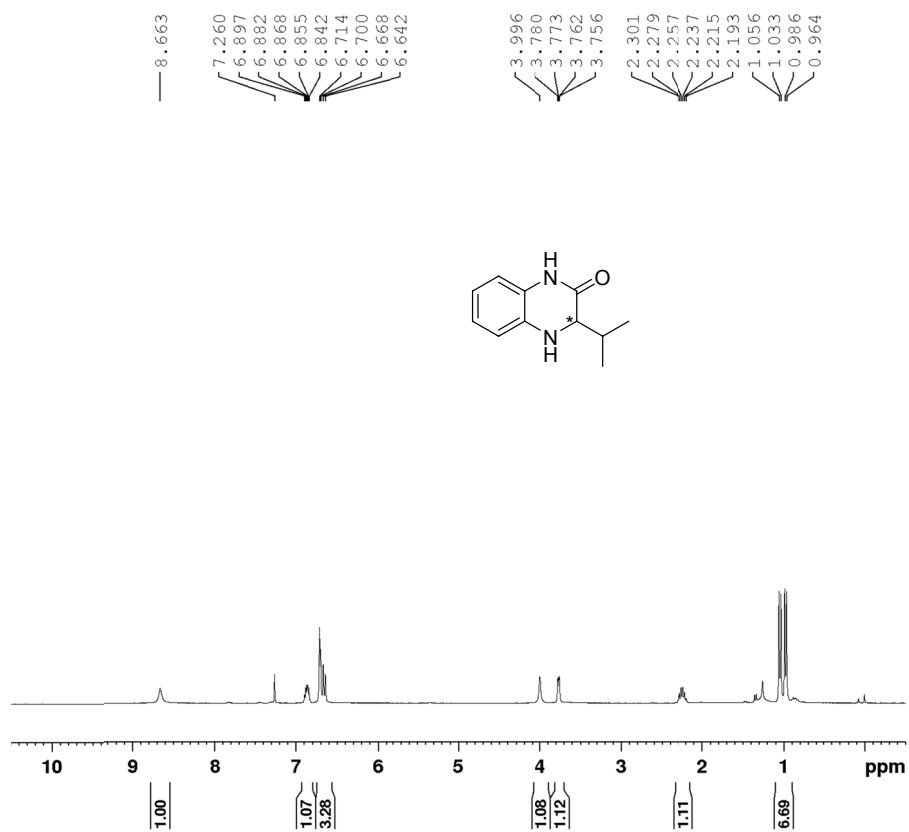
Current Data Parameters
 NAME KWI-npr-gh
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 14.10
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 6
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 192.89
 DW 16.800 usec
 DE 18.00 usec
 TE 298.2 K
 D1 2.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.80 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 125.7703637 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577746 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S32. ¹H NMR and ¹³C NMR spectra of 2e

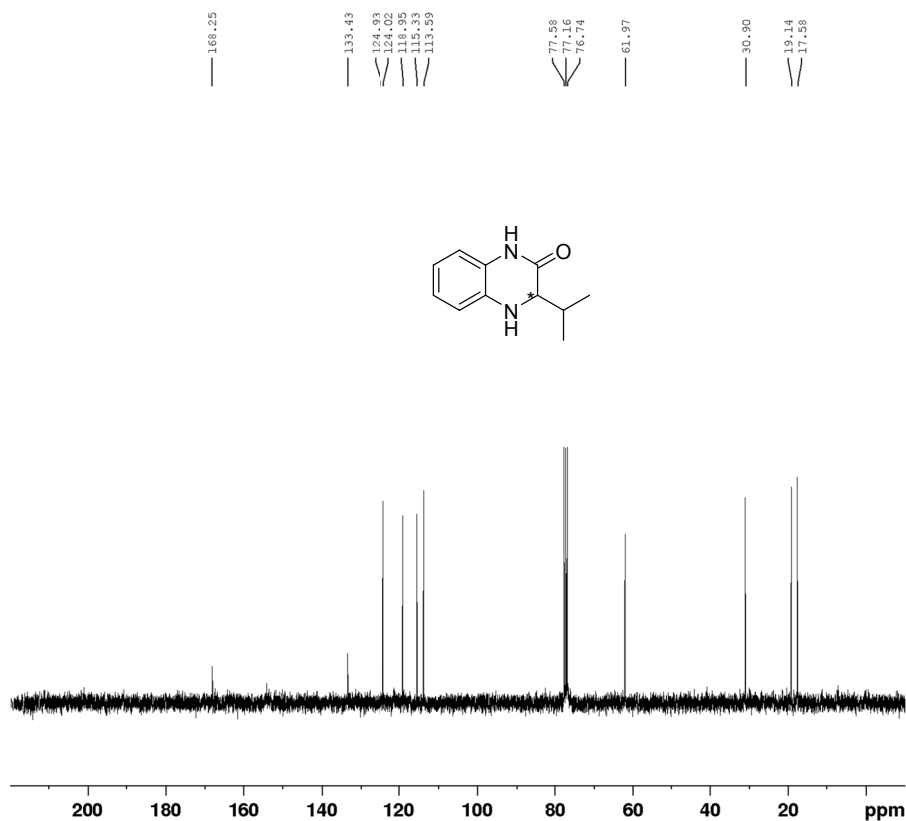


```
Current Data Parameters
NAME      KWT-1pr-qh
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20210502
Time     12.18
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
PULPROG  zg30
TD       55536
SOLVENT  CDCl3
NS       16
DS       2
SWH      6009.615 Hz
FIDRES   0.091699 Hz
AQ       5.4525952 sec
RG       209.09
DW       83.200 usec
DE       6.50 usec
TE       298.2 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       8.00 usec
PL1      120.00 dB
PL1W     0 W
SFO1     300.1318534 MHz

F2 - Processing parameters
SI       55536
SF       300.1300073 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
```



```
Current Data Parameters
NAME      KWT-1pr-qh-C
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20210502
Time     13.16
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       128
DS       4
SWH      18028.846 Hz
FIDRES   0.275098 Hz
AQ       1.8175317 sec
RG       209.09
DW       27.733 usec
DE       6.50 usec
TE       298.2 K
D1       2.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1       11.00 usec
PL1      120.00 dB
PL1W     0 W
SFO1     75.4752949 MHz

F2 - Processing parameters
SI       32768
SF       75.4677388 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
```


Figure S33. ¹H NMR and ¹³C NMR spectra of **2f**

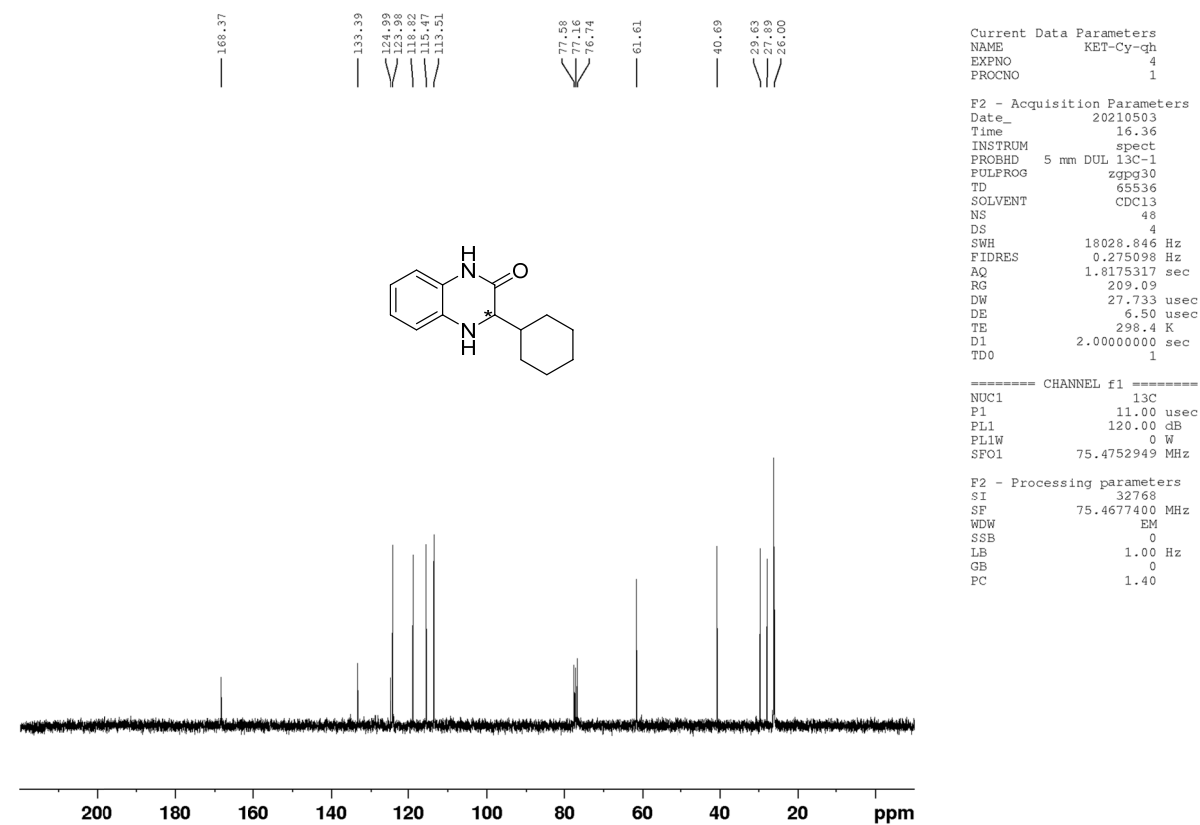
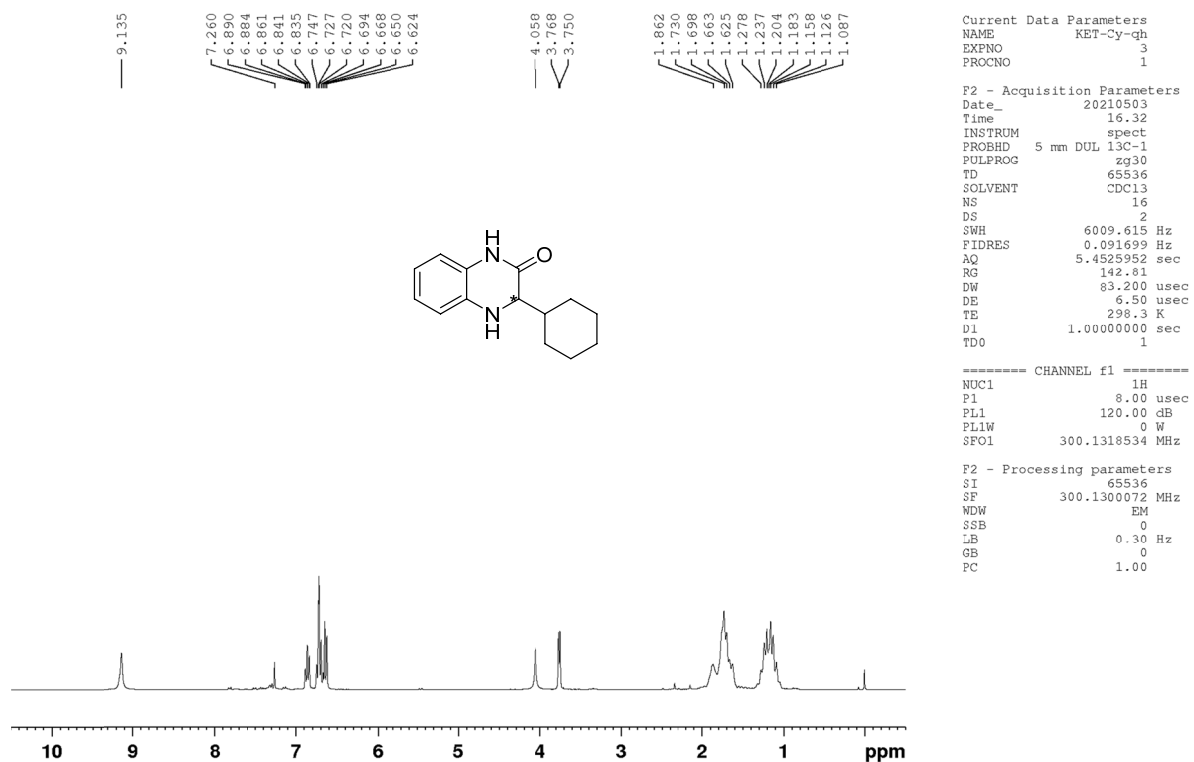
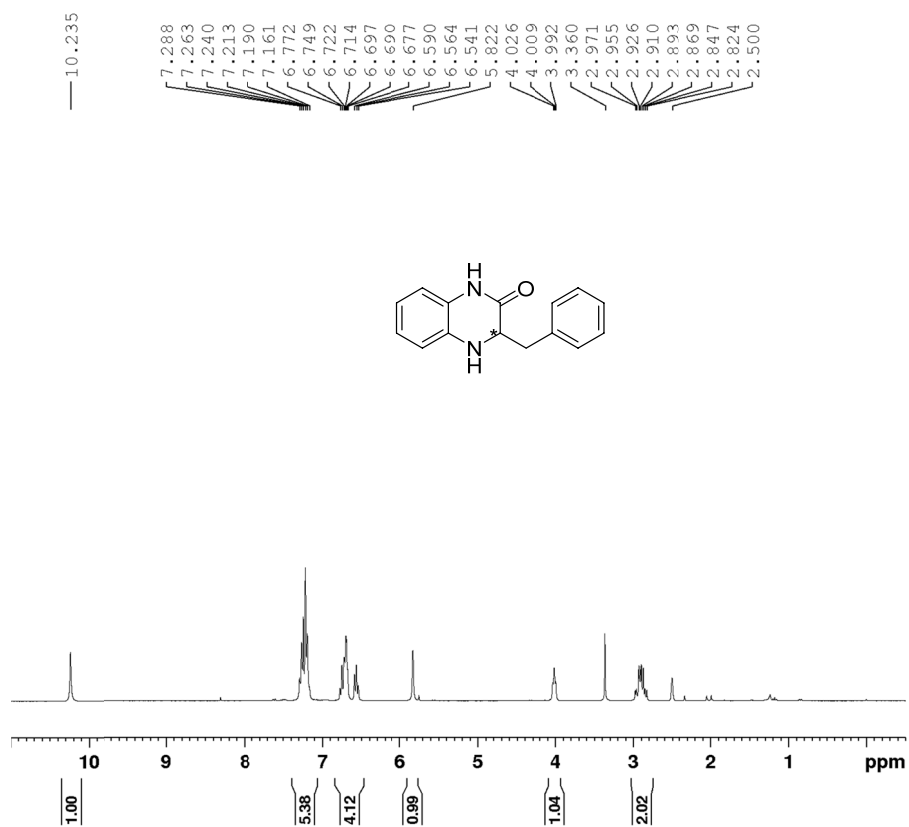


Figure S34. ¹H NMR and ¹³C NMR spectra of **2g**



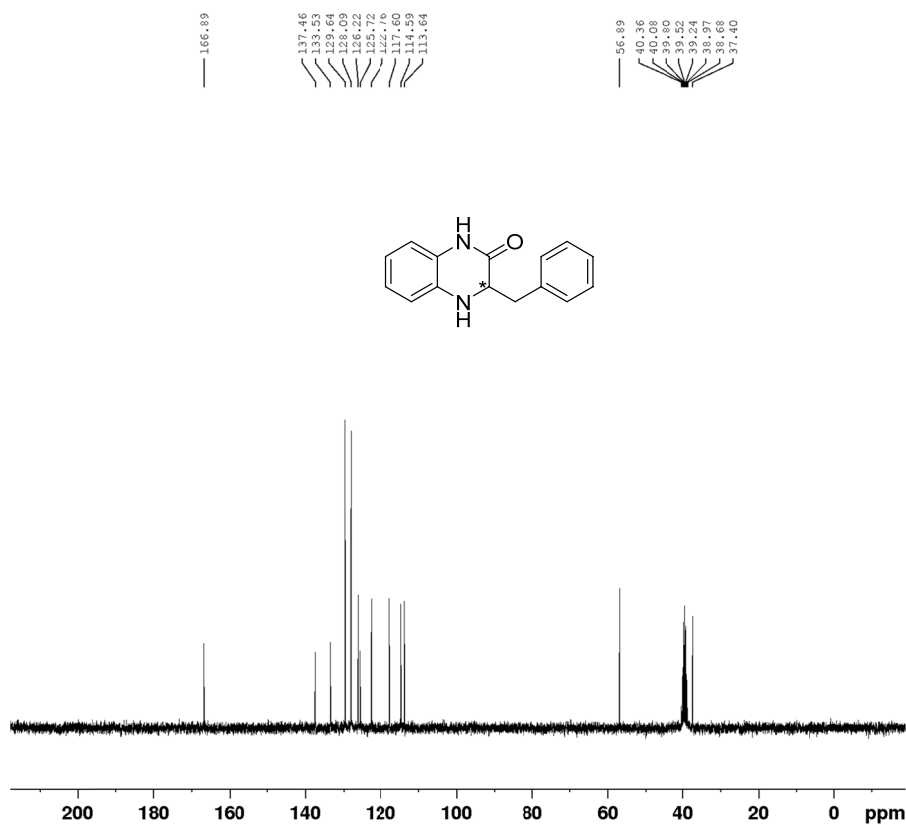
```

Current Data Parameters
NAME      lichanghao
EXPNO    41
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     20.59
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
PULPROG  zg30
TD        65536
SOLVENT  DMSO
NS        16
DS        2
SWH       6009.615 Hz
FIDRES    0.091699 Hz
AQ        5.4525952 sec
RG        142.81
DW        83.200 usec
DE        6.50 usec
TE        298.3 K
D1        1.0000000 sec
TD0       1

===== CHANNEL f1 =====
NUC1      1H
P1        8.00 usec
PL1       120.00 dB
PL1W      0 W
SFO1      300.1318534 MHz

F2 - Processing parameters
SI        65536
SF        300.1300025 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```



```

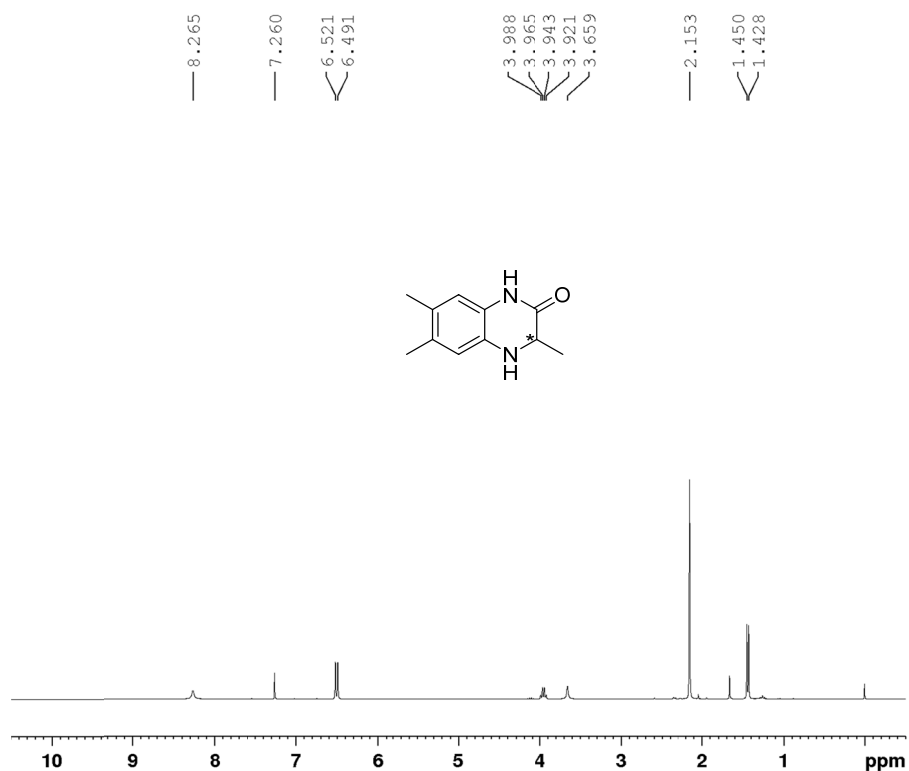
Current Data Parameters
NAME      lichanghao
EXPNO    42
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     21.02
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        27
DS        4
SWH       18028.846 Hz
FIDRES    0.275098 Hz
AQ        1.8175317 sec
RG        209.09
DW        27.733 usec
DE        6.50 usec
TE        298.3 K
D1        2.0000000 sec
TD0       1

===== CHANNEL f1 =====
NUC1      13C
P1        11.00 usec
PL1       120.00 dB
PL1W      0 W
SFO1      75.4752949 MHz

F2 - Processing parameters
SI        32768
SF        75.4577841 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```

Figure S35. ¹H NMR and ¹³C NMR spectra of **2h**

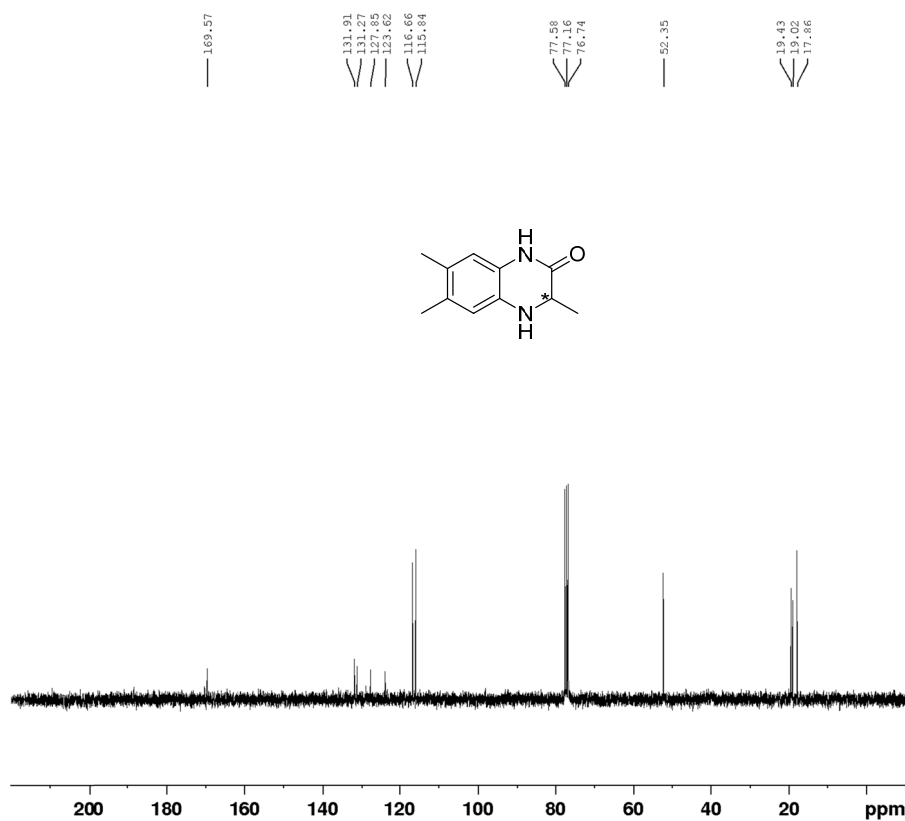


```
Current Data Parameters
NAME      KWT-3Me-qh
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     16.18
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
FULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      6009.615 Hz
FIDRES   0.091699 Hz
AQ       5.4525952 sec
RG       209.09
DW       83.200 usec
DE       6.50 usec
TE       298.3 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       8.00 usec
PL1      120.00 dB
PL1W     0 W
SFO1     300.1318534 MHz

F2 - Processing parameters
SI       65536
SF       300.1300072 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
FC       1.00
```



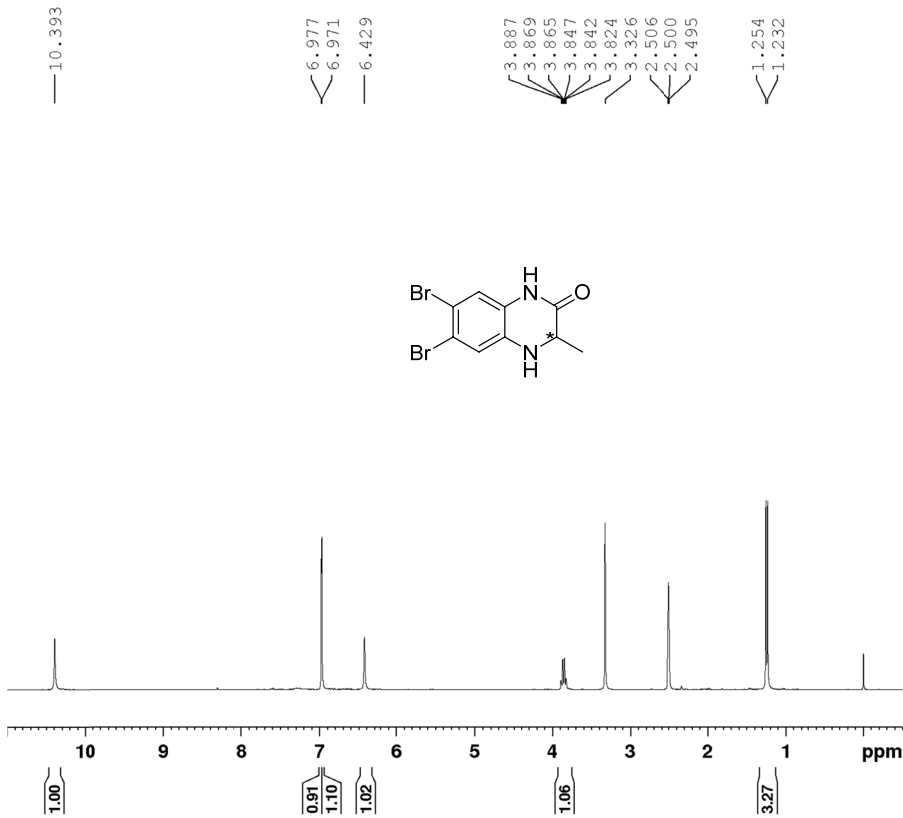
```
Current Data Parameters
NAME      KWT-3Me-qh
EXPNO    3
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     16.21
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
FULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       149
DS       4
SWH      18028.846 Hz
FIDRES   0.275098 Hz
AQ       1.8175317 sec
RG       209.09
DW       27.733 usec
DE       6.50 usec
TE       298.3 K
D1       2.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1       11.00 usec
PL1      120.00 dB
PL1W     0 W
SFO1     75.4752949 MHz

F2 - Processing parameters
SI       32768
SF       75.4677388 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
FC       1.40
```

Figure S36. ¹H NMR and ¹³C NMR spectra of **2i**

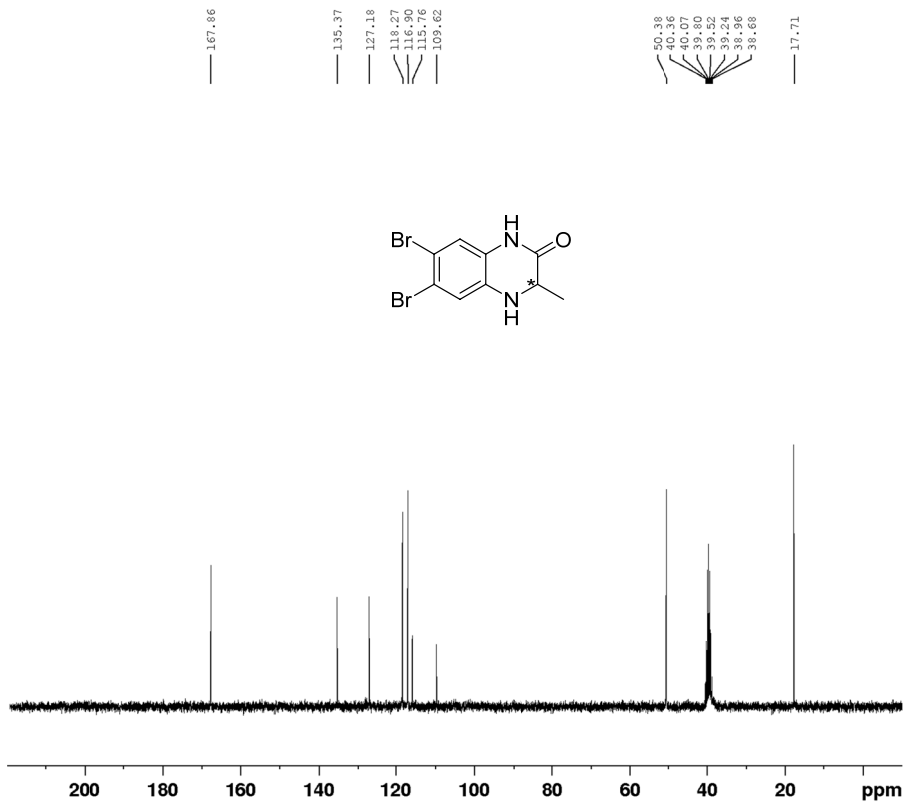


Current Data Parameters
 NAME lichenghao
 EXPNO 53
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 23.29
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 209.09
 DW 33.200 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300025 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



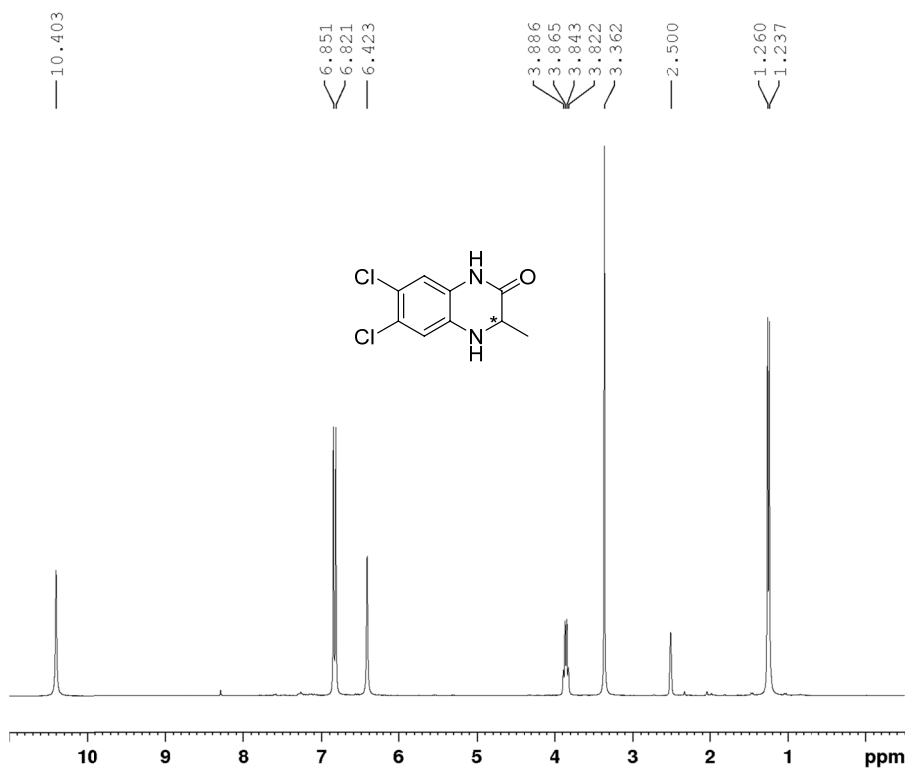
Current Data Parameters
 NAME lichenghao
 EXPNO 44
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 22.32
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 41
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175317 sec
 RG 209.09
 DW 27.733 usec
 DE 6.50 usec
 TE 298.3 K
 D1 2.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677820 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S37. ¹H NMR and ¹³C NMR spectra of **2j**



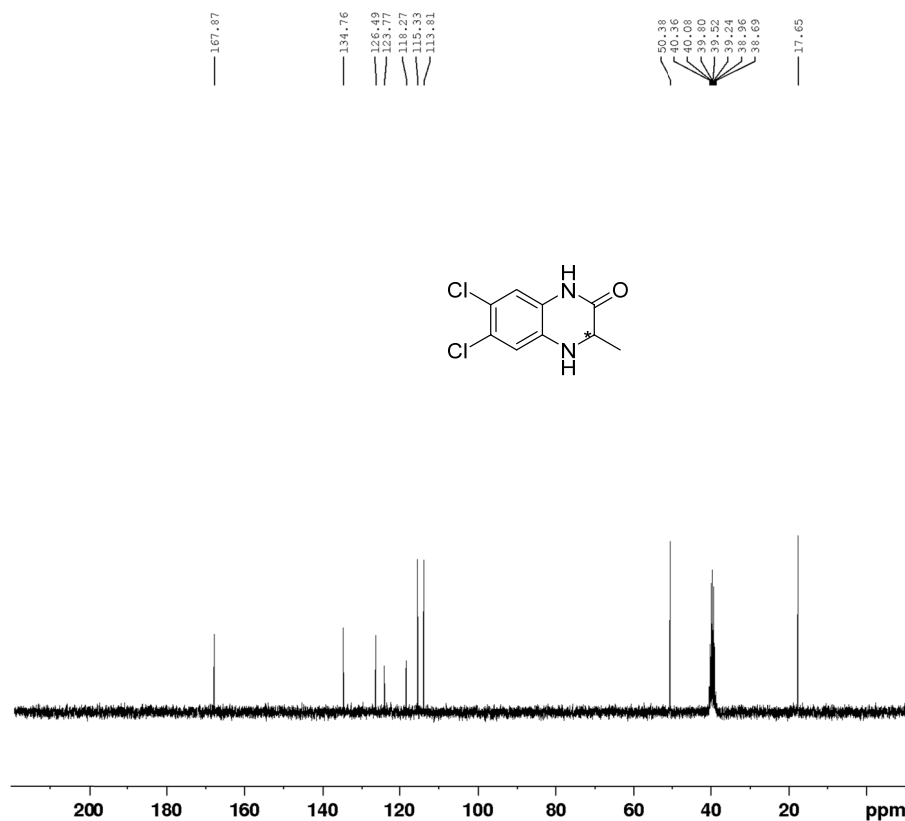
```

Current Data Parameters
NAME      lichenghao
EXPNO    35
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     20.00
INSTRUM spect
PROBHD   5 mm DUL 13C-1
PULPROG zg30
TD       65536
SOLVENT DMSO
NS       16
DS       2
SWH      6009.615 Hz
FIDRES   0.091699 Hz
AQ       5.4525952 sec
RG       209.09
DW       83.200 usec
DE       6.50 usec
TE       298.2 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       8.00 usec
PL1     120.00 dB
PL1W    0 W
SFO1    300.1318534 MHz

F2 - Processing parameters
SI       65536
SF      300.1300025 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00
    
```



```

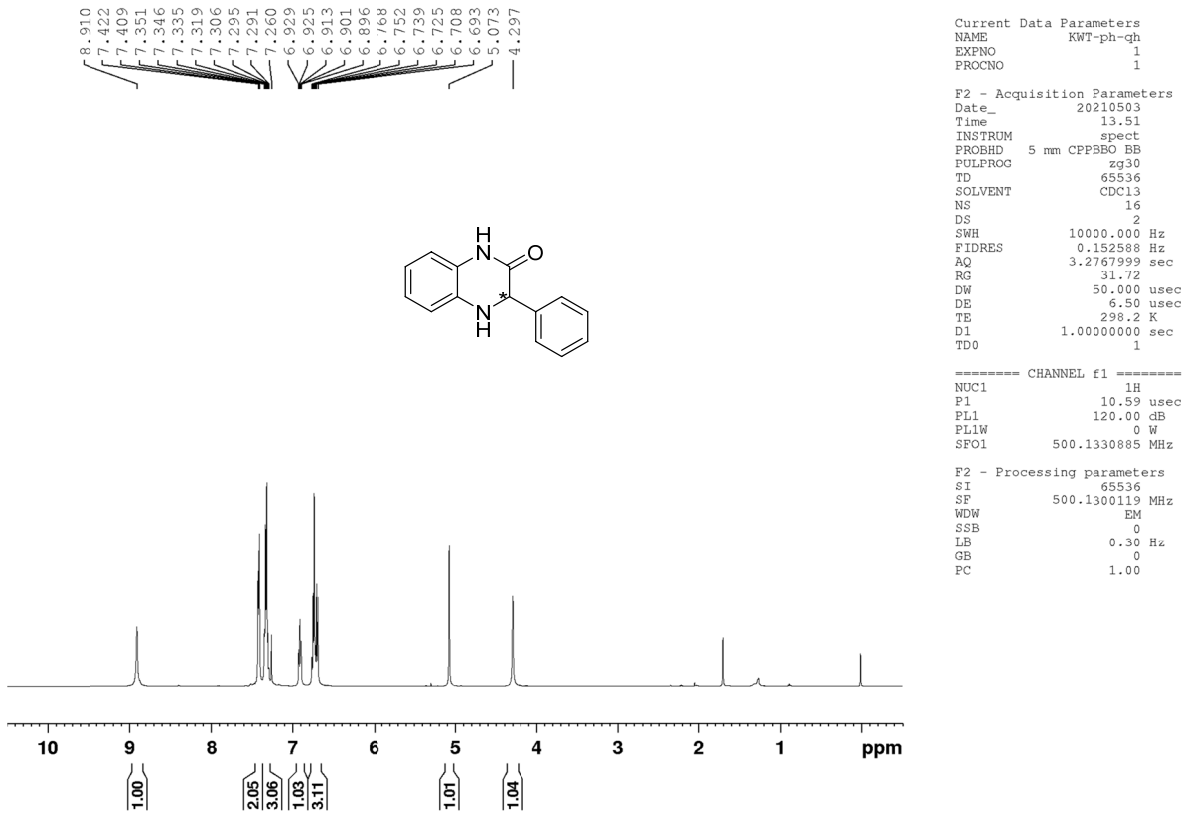
Current Data Parameters
NAME      lichenghao
EXPNO    36
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     20.03
INSTRUM spect
PROBHD   5 mm DUL 13C-1
PULPROG zgpg30
TD       65536
SOLVENT DMSO
NS       32
DS       4
SWH      18028.846 Hz
FIDRES   0.275098 Hz
AQ       1.8175317 sec
RG       209.09
DW       27.733 usec
DE       6.50 usec
TE       298.3 K
D1       2.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1      11.00 usec
PL1     120.00 dB
PL1W    0 W
SFO1    75.4752949 MHz

F2 - Processing parameters
SI      32768
SF     75.4677823 MHz
WDW     EM
SSB     0
LB      1.00 Hz
GB      0
PC      1.40
    
```

Figure S38. ¹H NMR and ¹³C NMR spectra of **2k**

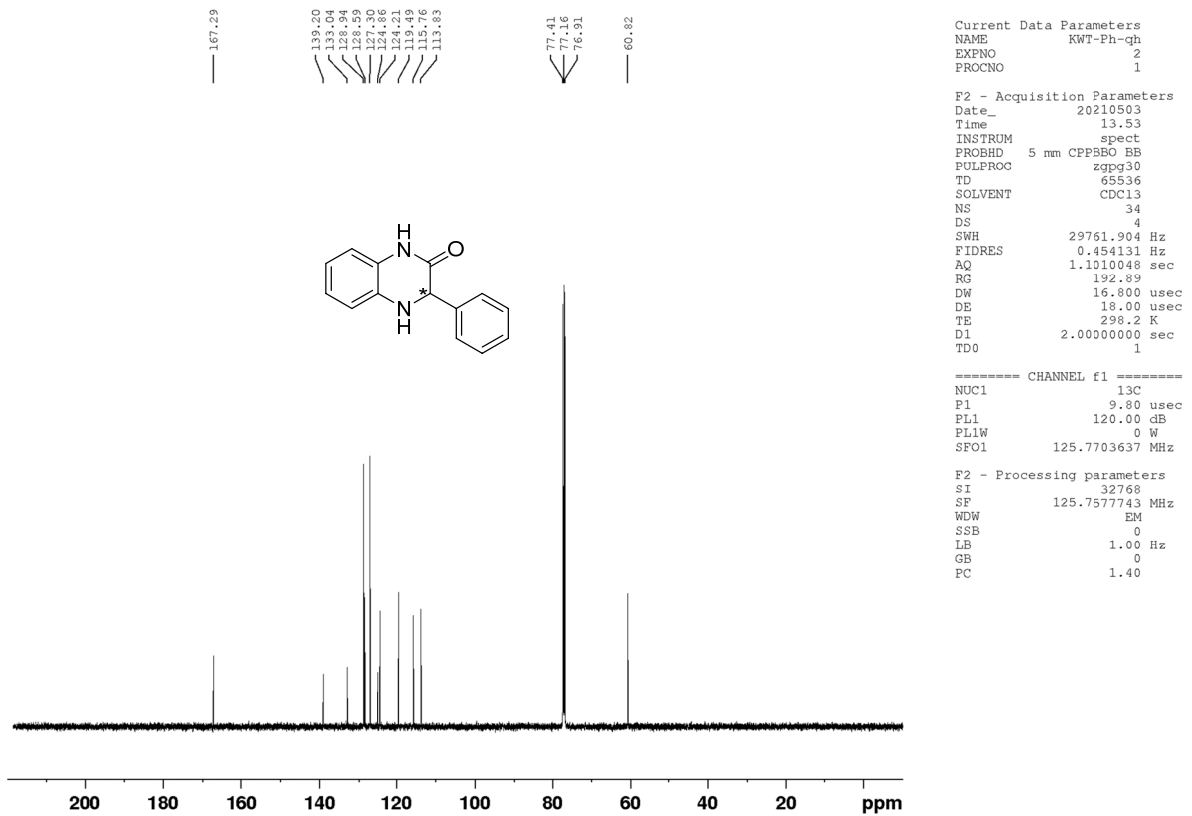


Current Data Parameters
 NAME KWT-Ph-qh
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 13.51
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 1000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 31.72
 DW 50.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.59 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 500.1330885 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300119 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



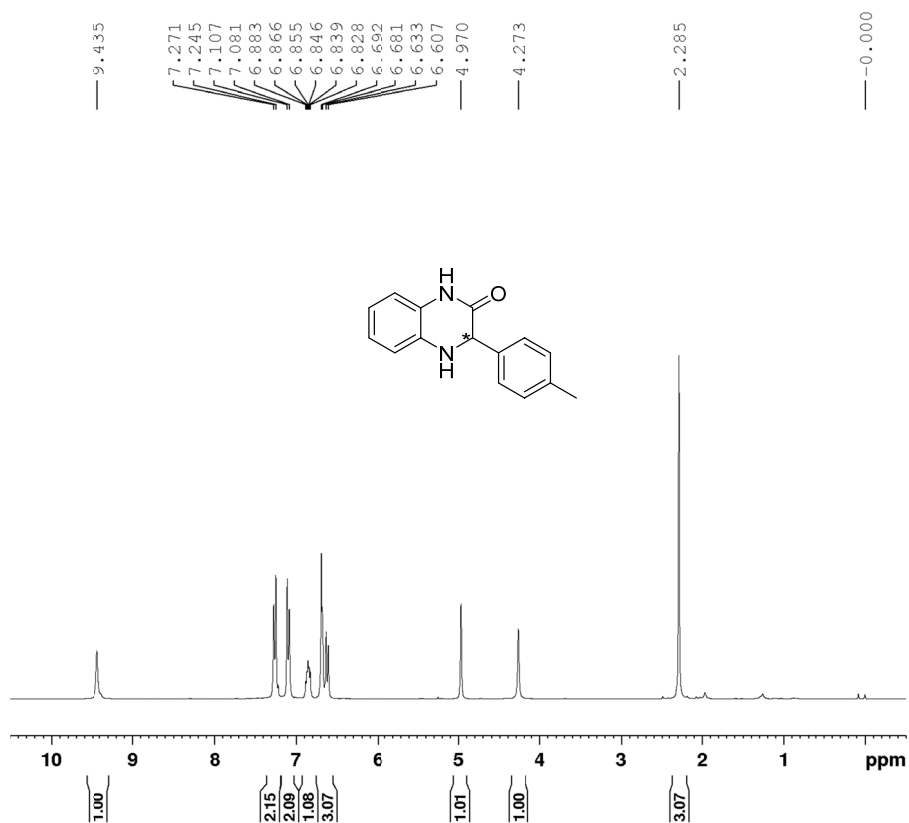
Current Data Parameters
 NAME KWT-Ph-qh
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 13.53
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 34
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 192.89
 DW 16.800 usec
 DE 18.00 usec
 TE 298.2 K
 D1 2.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.80 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 125.7703637 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577743 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S39. ¹H NMR and ¹³C NMR spectra of 2I

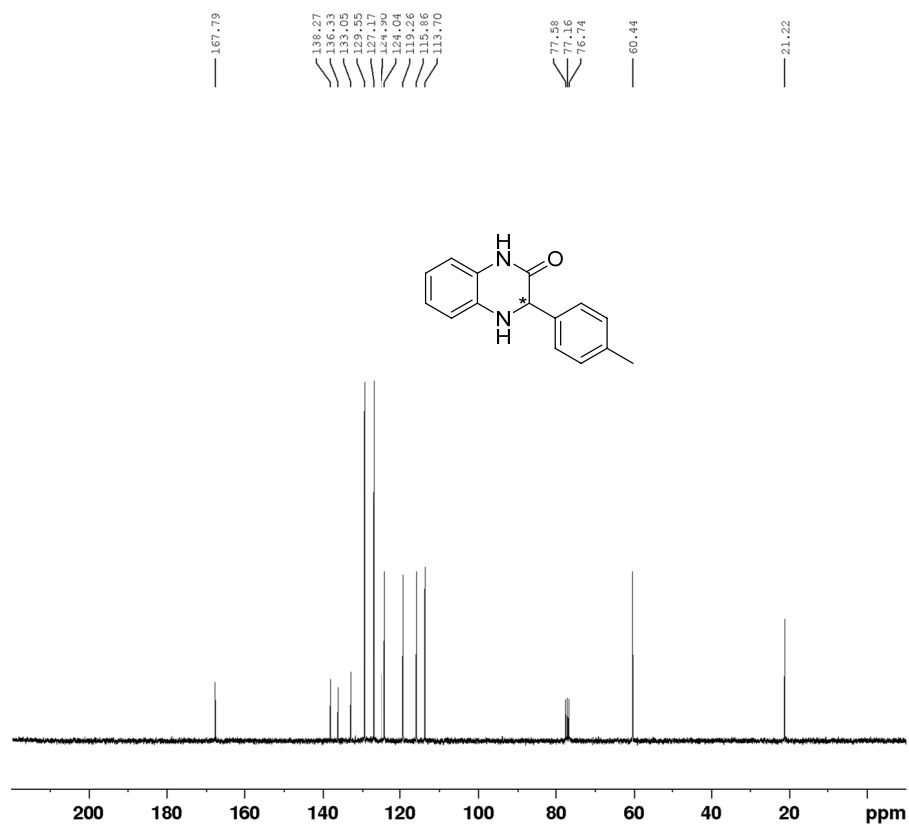


```
Current Data Parameters
NAME      KWT-4MePh-qh
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     15.13
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
FULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      6009.615 Hz
FIDRES   0.091699 Hz
AQ       5.4525952 sec
RG       103.3/
DW       83.200 usec
DE       6.50 usec
TE       298.3 K
D1       1.0000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       8.00 usec
PL1     120.00 dB
PL1W    0 W
SFO1    300.1318534 MHz

F2 - Processing parameters
SI       65536
SF      300.1300196 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00
```



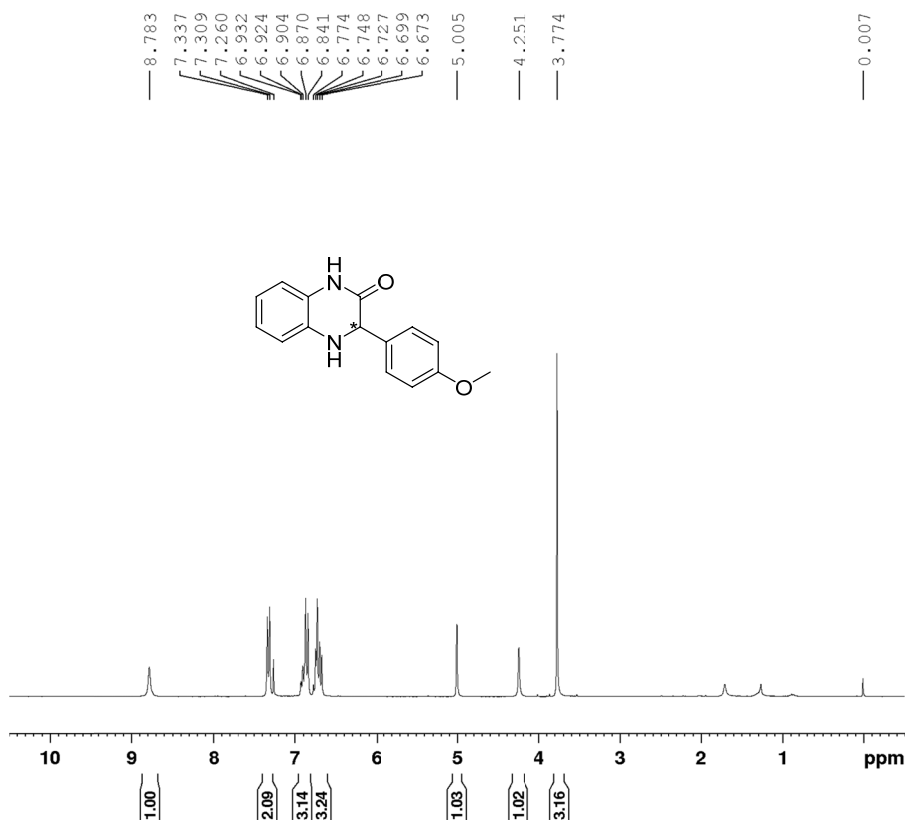
```
Current Data Parameters
NAME      KWT-4MePh-qh
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20210503
Time     15.18
INSTRUM  spect
PROBHD   5 mm DUL 13C-1
FULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       64
DS       4
SWH      18028.846 Hz
FIDRES   0.275098 Hz
AQ       1.8175317 sec
RG       209.09
DW       27.733 usec
DE       6.50 usec
TE       298.3 K
D1       2.0000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1       11.00 usec
PL1     120.00 dB
PL1W    0 W
SFO1    75.4752949 MHz

F2 - Processing parameters
SI       32768
SF      75.4677458 MHz
WDW     EM
SSB     0
LB      1.00 Hz
GB      0
PC      1.40
```

Figure S40. ^1H NMR and ^{13}C NMR spectra of **2m**

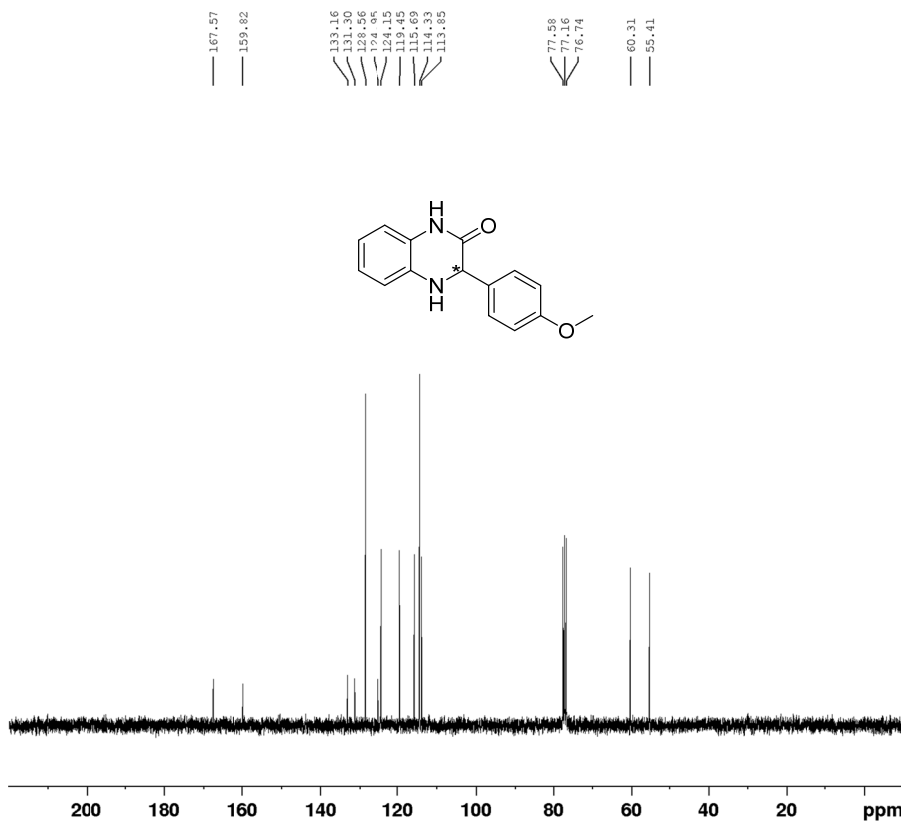


Current Data Parameters
 NAME lichenghao
 EXPNO 29
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 18.36
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 209.09
 DW 83.200 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300072 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME lichenghao
 EXPNO 30
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 18.44
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 93
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175317 sec
 RG 209.09
 DW 27.733 usec
 DE 6.50 usec
 TE 298.3 K
 D1 2.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4577400 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S41. ¹H NMR and ¹³C NMR spectra of **2n**

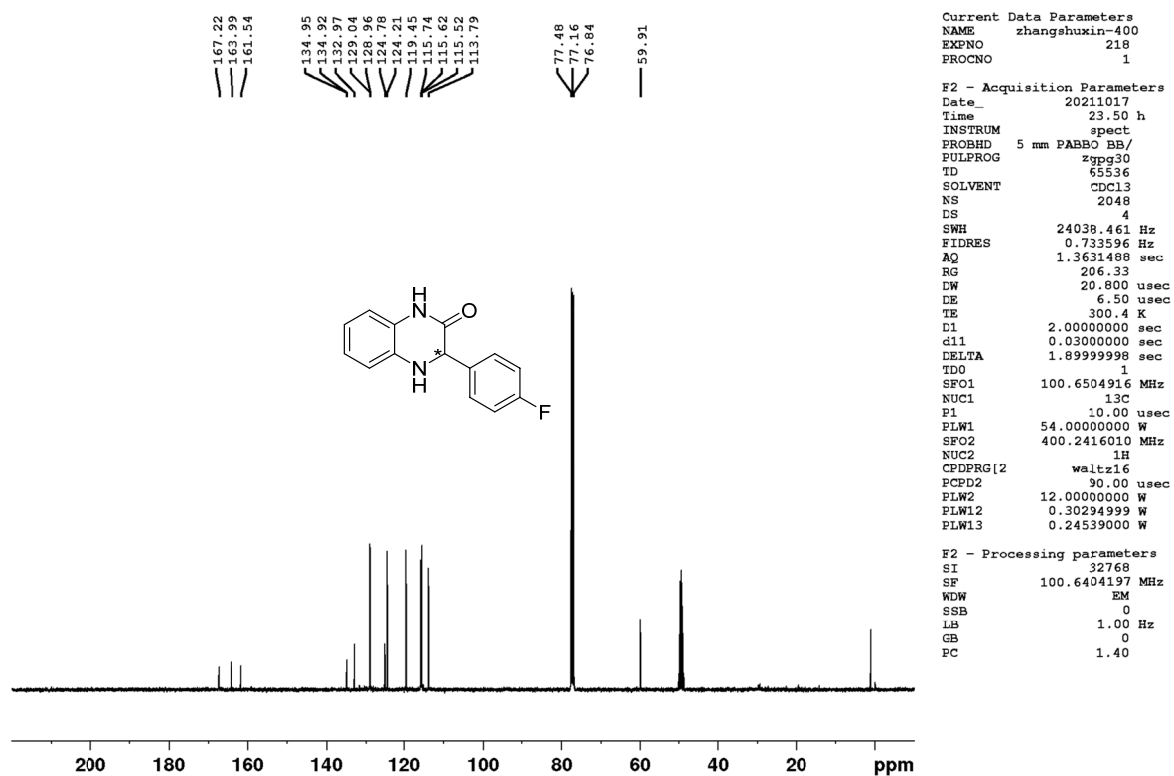
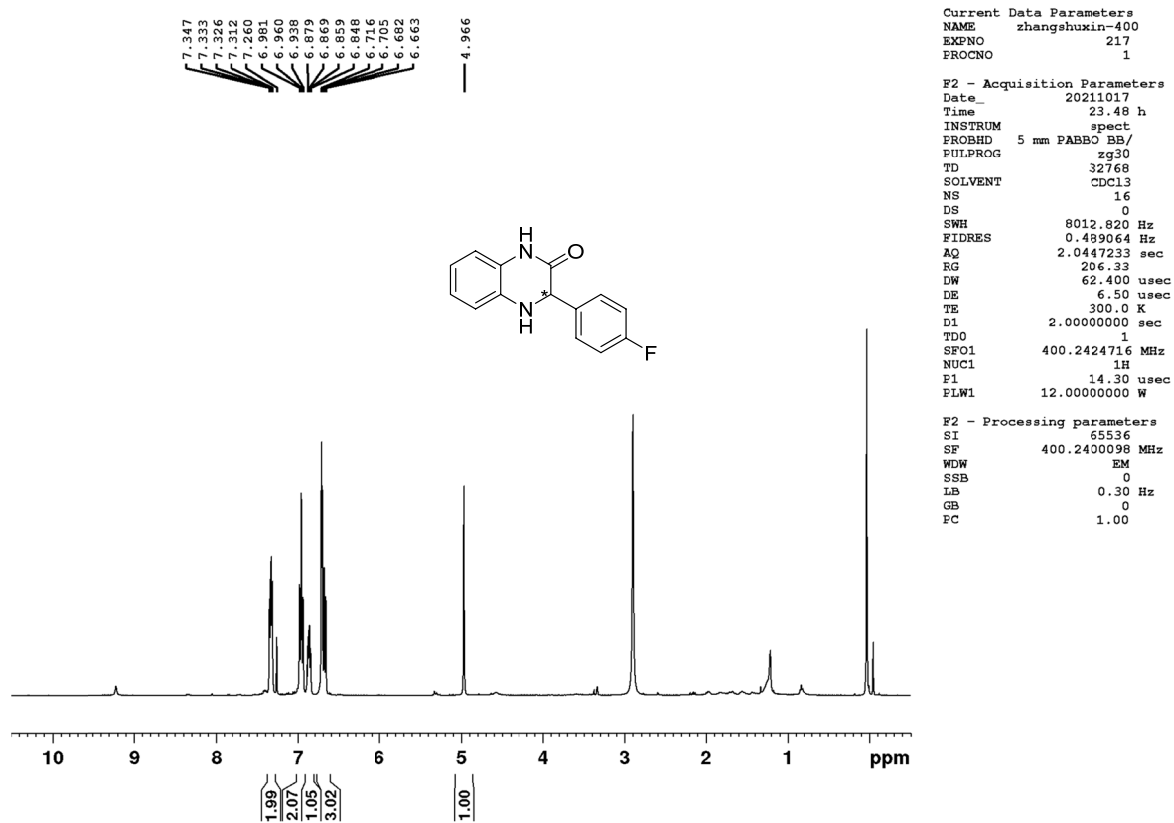
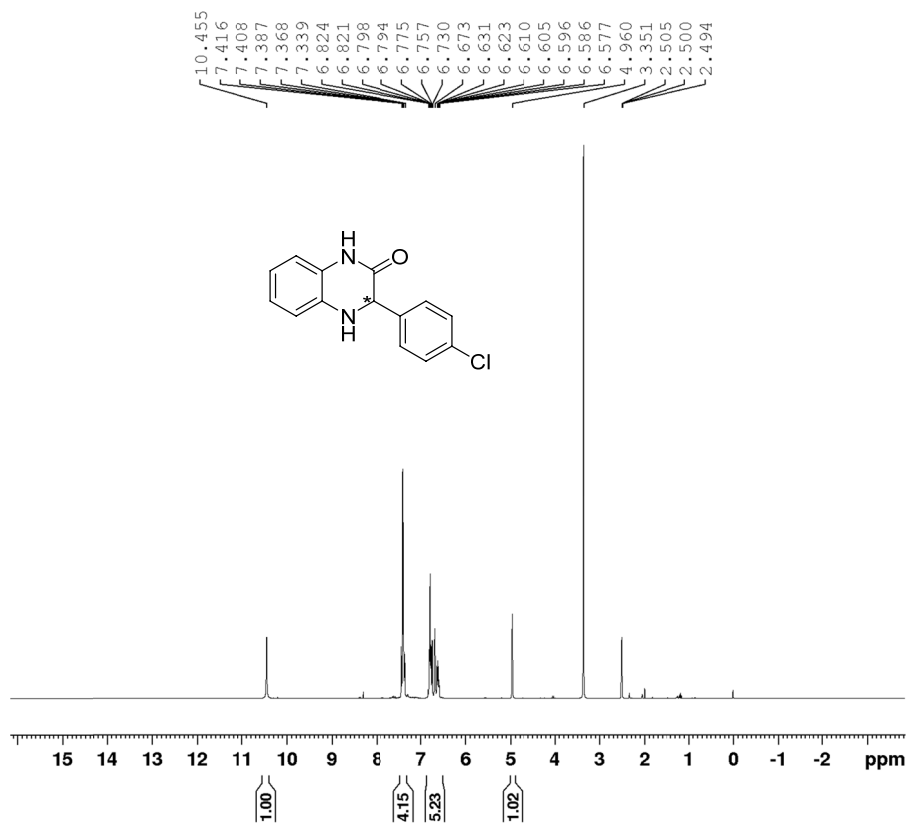


Figure S42. ¹H NMR and ¹³C NMR spectra of 2o

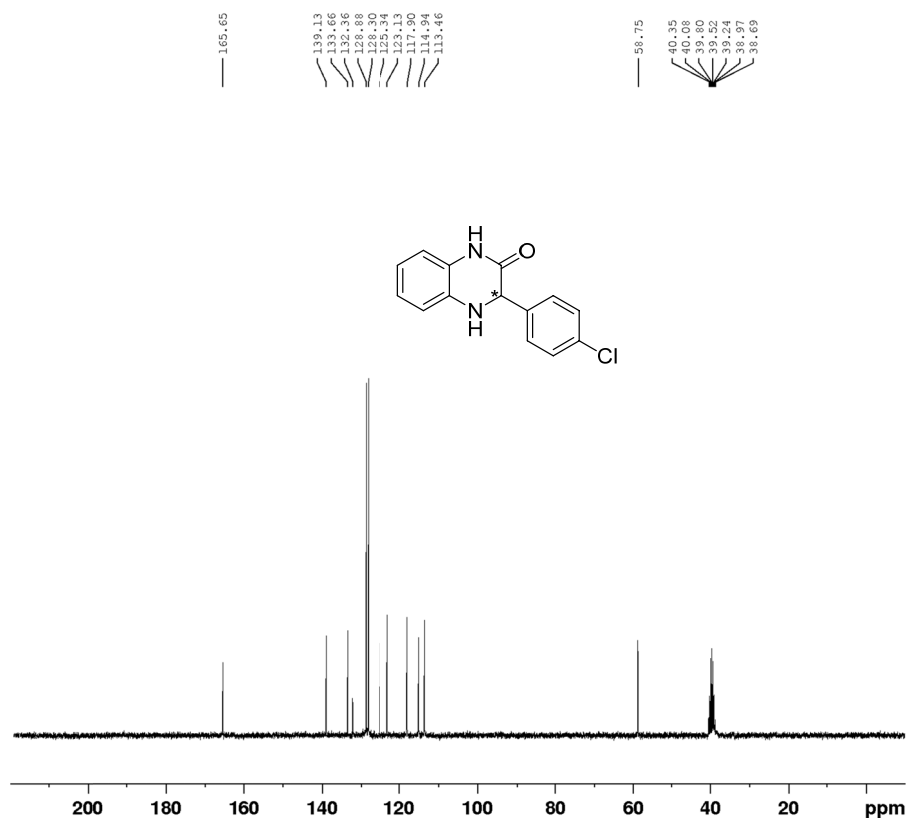


Current Data Parameters
 NAME lichenghao
 EXPNO 49
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 23.07
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 6079.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 209.09
 DW 33.200 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00100000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300024 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



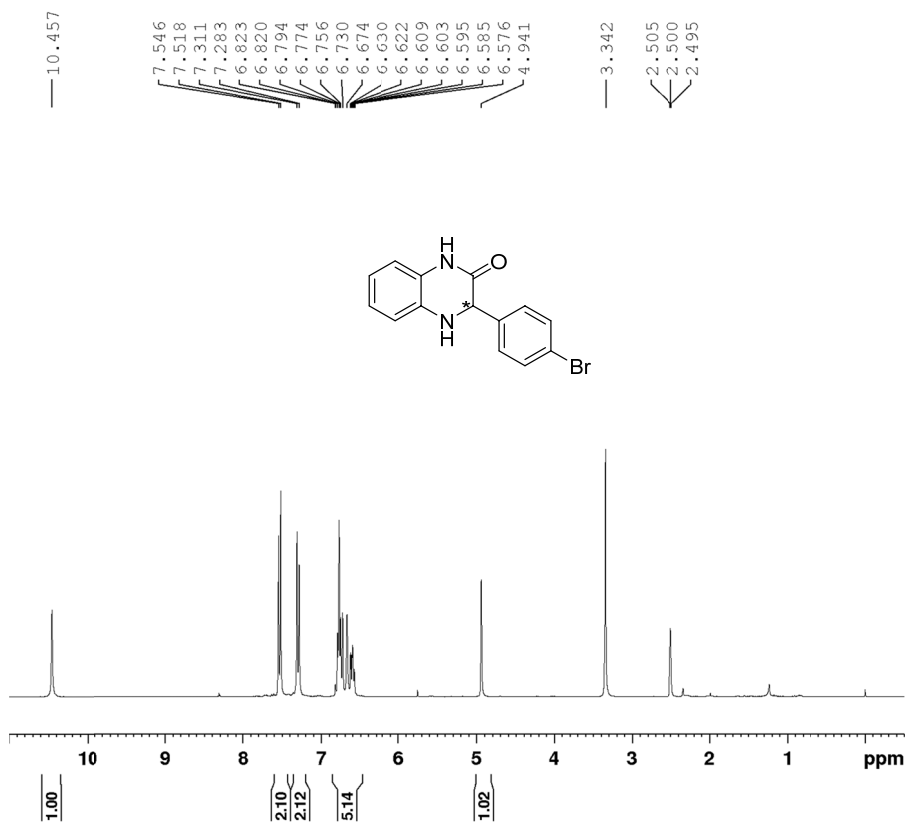
Current Data Parameters
 NAME lichenghao
 EXPNO 47
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 22.27
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 43
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.6175317 sec
 RG 209.09
 DW 27.733 usec
 DE 6.50 usec
 TE 298.3 K
 D1 2.00100000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4577817 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S43. ¹H NMR and ¹³C NMR spectra of **2p**

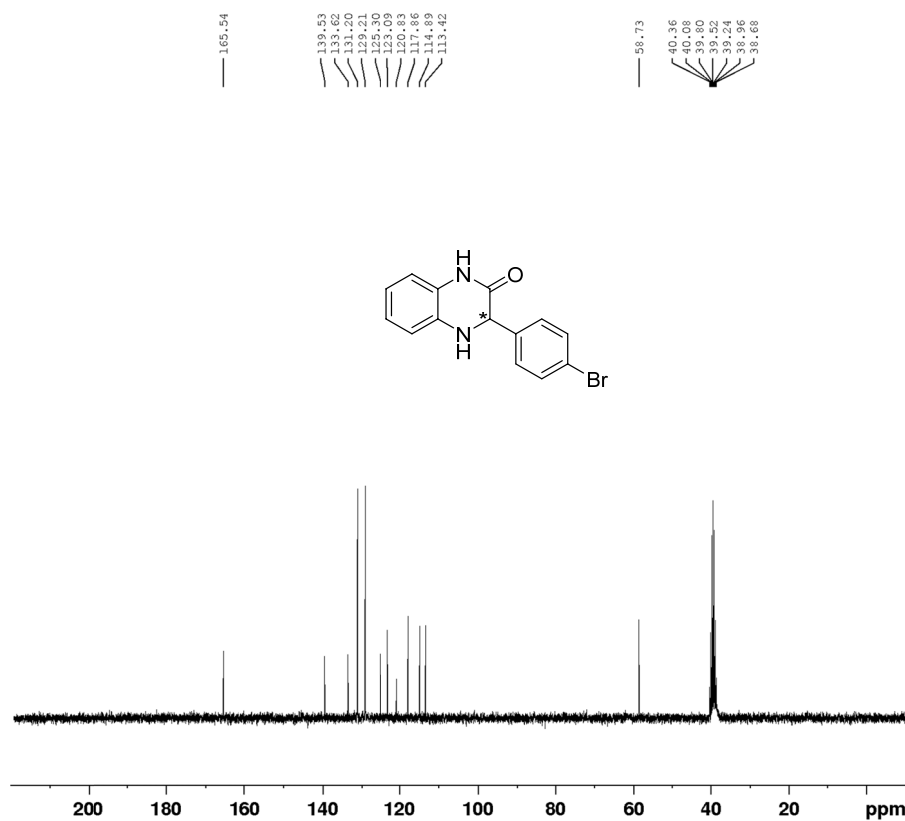


Current Data Parameters
 NAME lichenghao
 EXPNO 51
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 23.15
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 209.09
 DW 83.200 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300015 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME lichenghao
 EXPNO 52
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 23.17
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 68
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175317 sec
 RG 209.09
 DW 27.733 usec
 DE 6.50 usec
 TE 298.3 K
 D1 2.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677830 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S44. ¹H NMR and ¹³C NMR spectra of **2q**

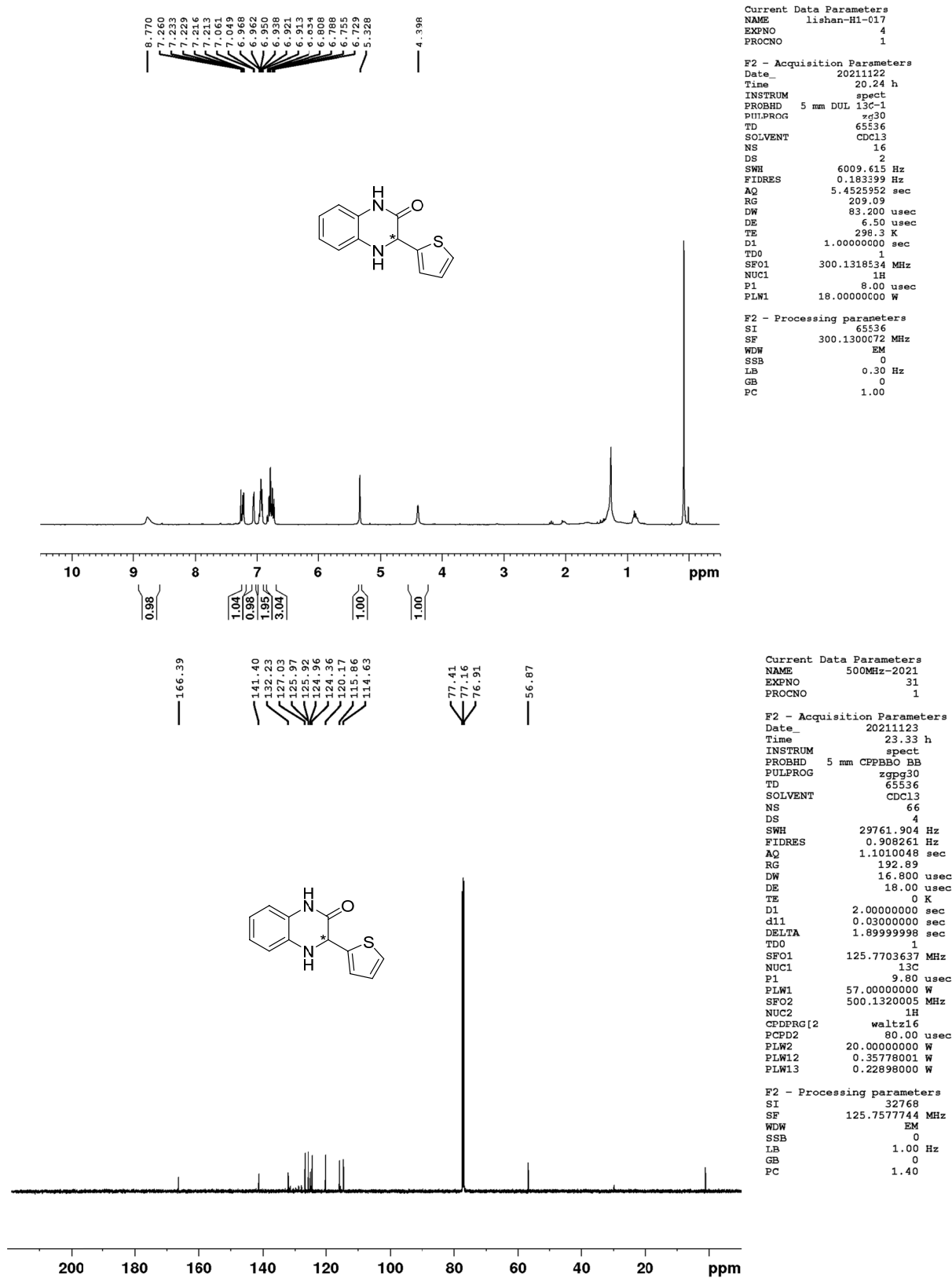


Figure S45. ¹H NMR and ¹³C NMR spectra of **2r**

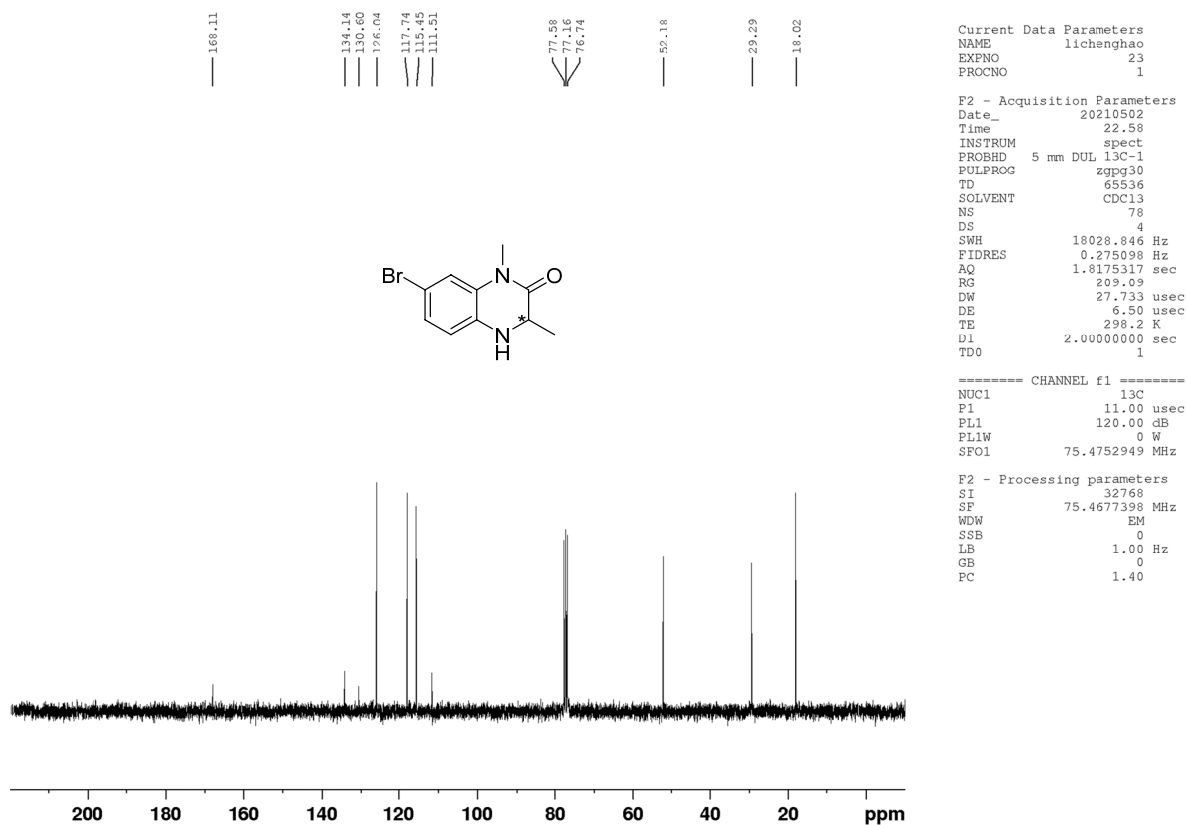
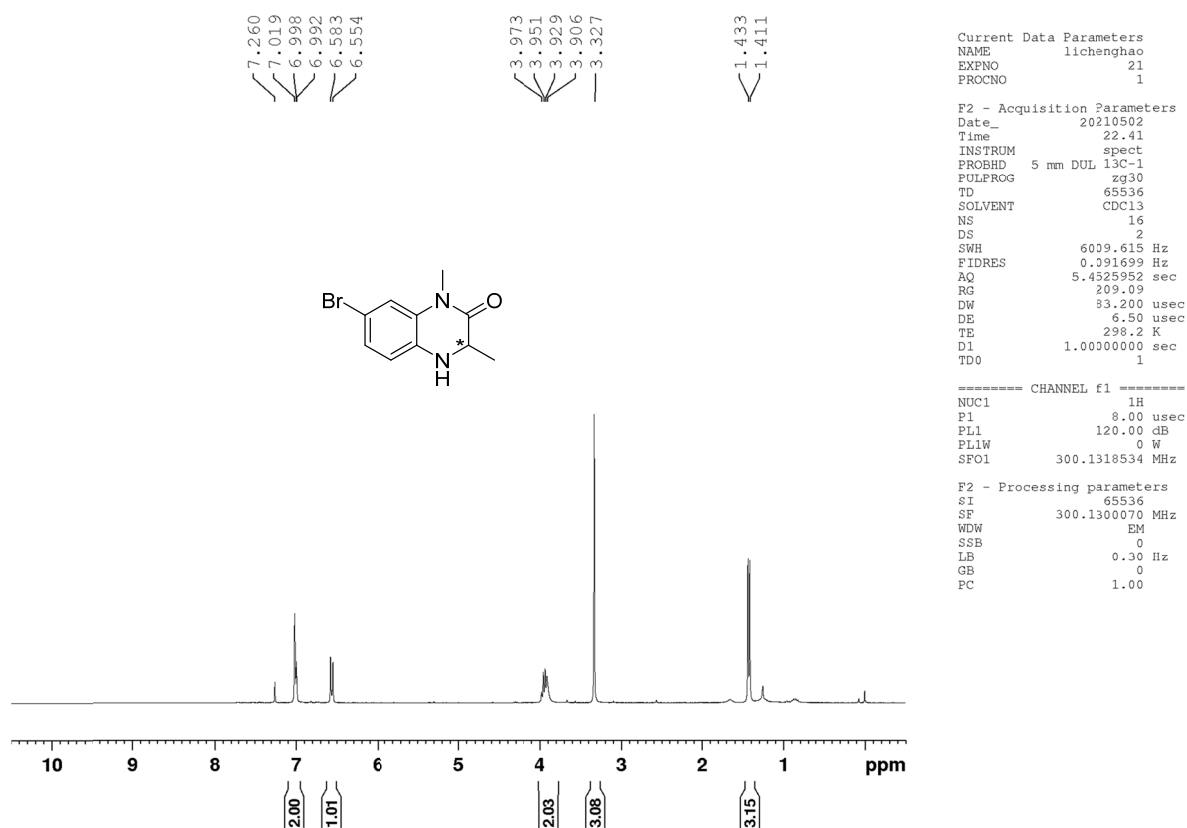
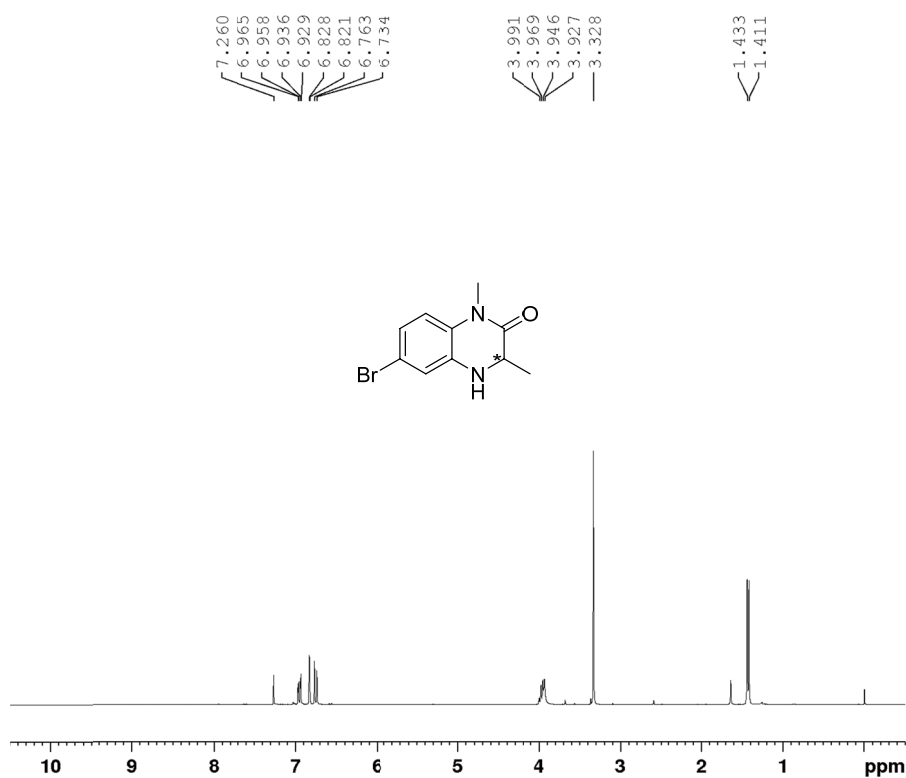


Figure S46. ¹H NMR and ¹³C NMR spectra of 2s

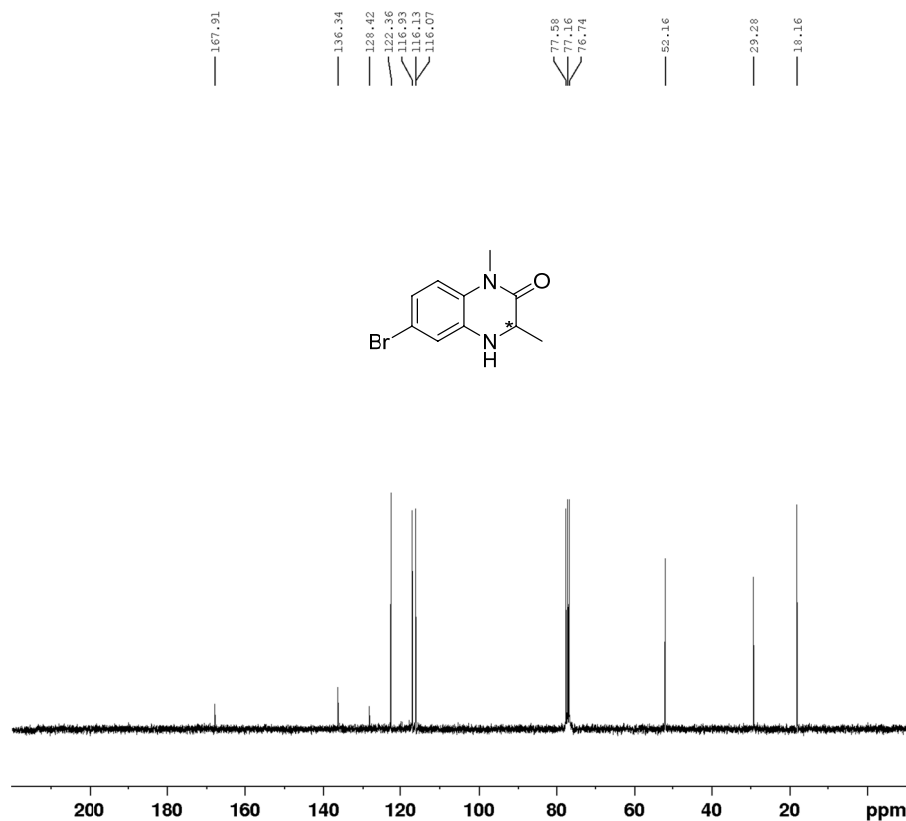


Current Data Parameters
 NAME KWT-6-Br-Me-qh
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 17.10
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 209.09
 DW 83.200 usec
 DE 6.50 usec
 TE 298.3 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300072 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



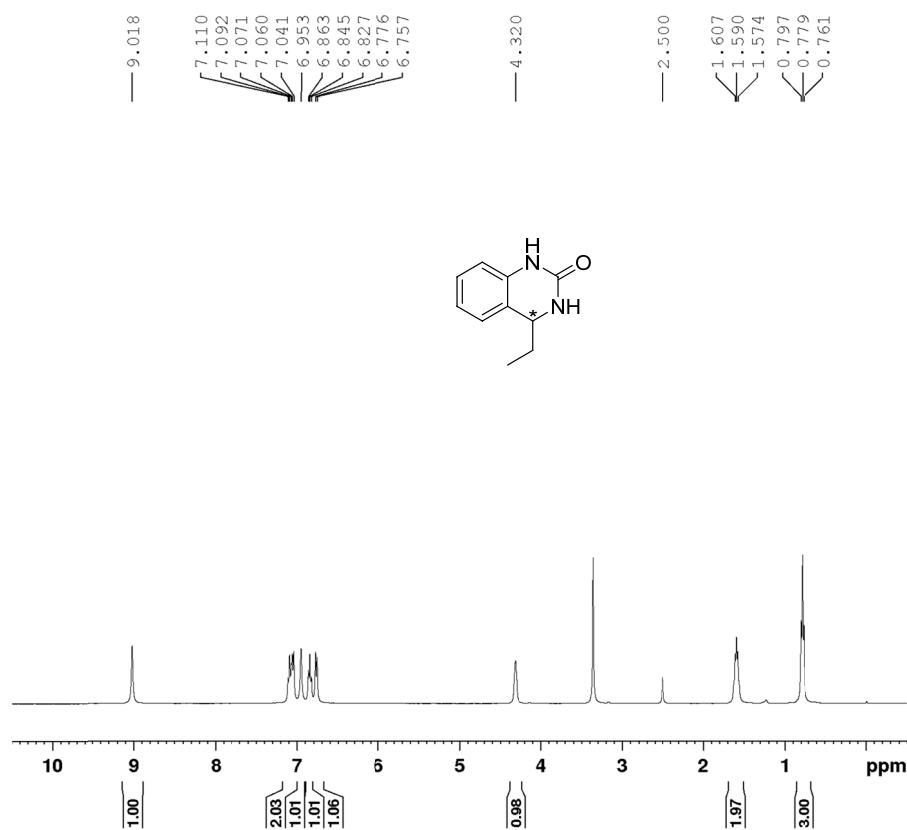
Current Data Parameters
 NAME KWT-6-Br-Me-qh
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210503
 Time 17.12
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 512
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175317 sec
 RG 209.09
 DW 27.733 usec
 DE 6.50 usec
 TE 298.4 K
 D1 2.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4577392 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S47. ¹H NMR and ¹³C NMR spectra of 4a

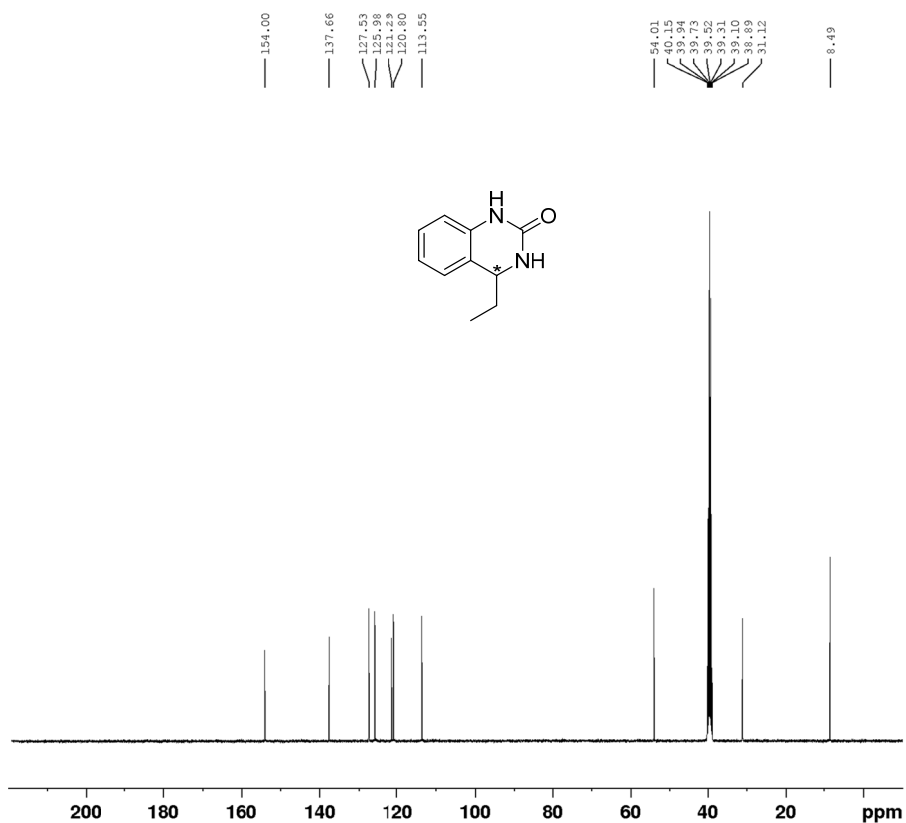


Current Data Parameters
 NAME 20210411-K2T-ET-QH
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210411
 Time 15.19
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 60.55
 DW 62.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 13.26099968 W

F2 - Processing parameters
 SI 65536
 SF 400.1300032 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME 20210411-K2T-ET-QH
 EXPNO 20
 PROCNO 1

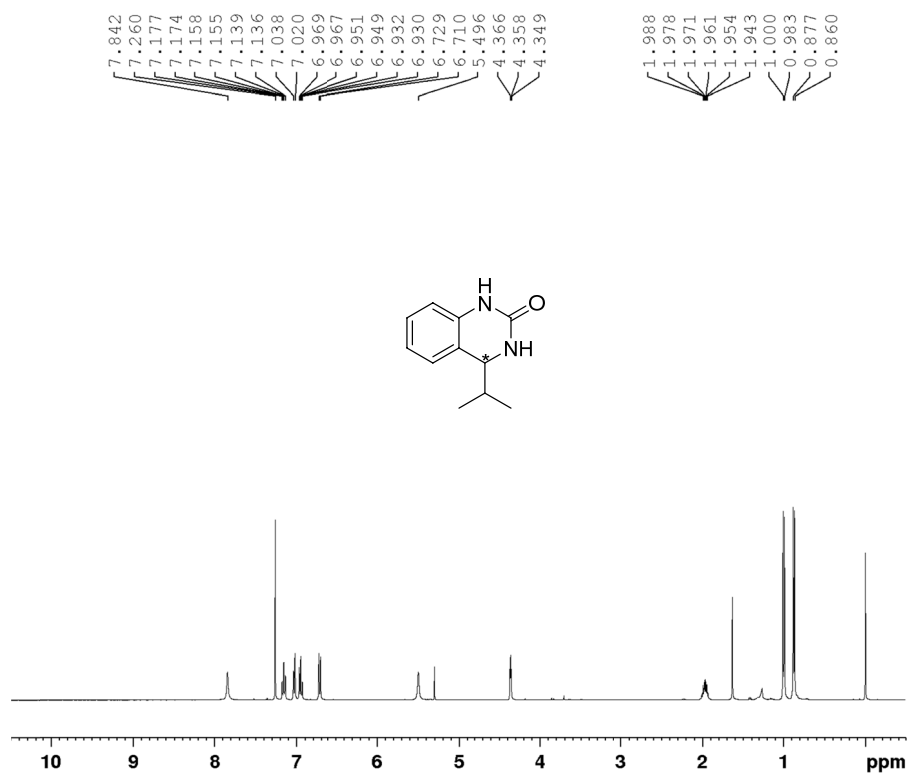
F2 - Acquisition Parameters
 Date_ 20210411
 Time 16.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TE 65536
 SOLVENT DMSO
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 199.89
 DW 20.800 usec
 DE 6.50 usec
 TE 298.1 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 100.6228293 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 56.93000031 W

===== CHANNEL f2 =====
 SFO2 400.1316005 MHz
 NUC2 1H
 CDEPRG[2] waltz16
 PCPD2 90.00 usec
 PLW2 13.26099968 W
 PLW12 0.16372000 W
 PLW13 0.13260999 W

F2 - Processing parameters
 SI 32768
 SF 100.6128139 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S48. ¹H NMR and ¹³C NMR spectra of **4b**

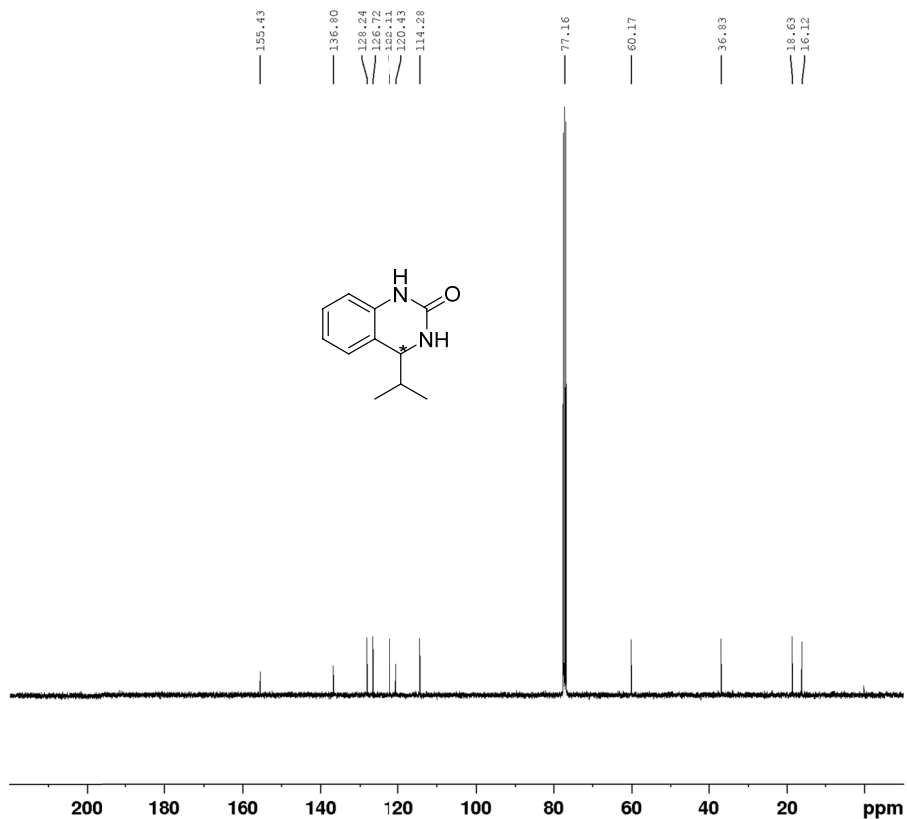


Current Data Parameters
 NAME 20210411-KZT-IPR-QH
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210411
 Time 20.54
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 136.06
 DW 62.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 13.26099968 W

F2 - Processing parameters
 SI 65536
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME 20210411-KZT-IPR-QH-1024
 EXPNO 11
 PROCNO 1

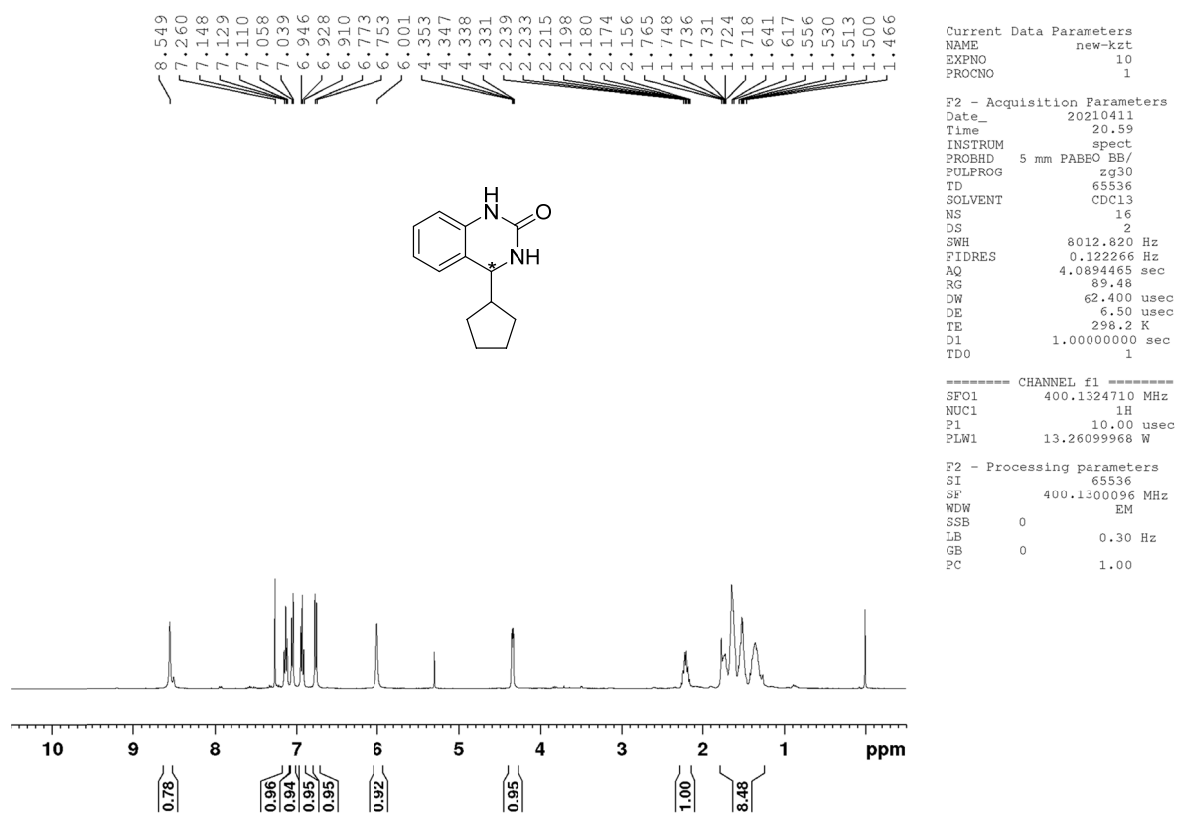
F2 - Acquisition Parameters
 Date_ 20210412
 Time 0.14
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT cdcl3
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 199.89
 DW 20.800 usec
 DE 6.50 usec
 TE 298.2 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 100.6228293 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 56.93000031 W

===== CHANNEL f2 =====
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 90.00 usec
 PLW2 13.26099968 W
 PLW1:2 0.16372000 W
 PLW1:3 0.13260999 W

F2 - Processing parameters
 SI 32768
 SF 100.6127547 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S49. ¹H NMR and ¹³C NMR spectra of **4c**



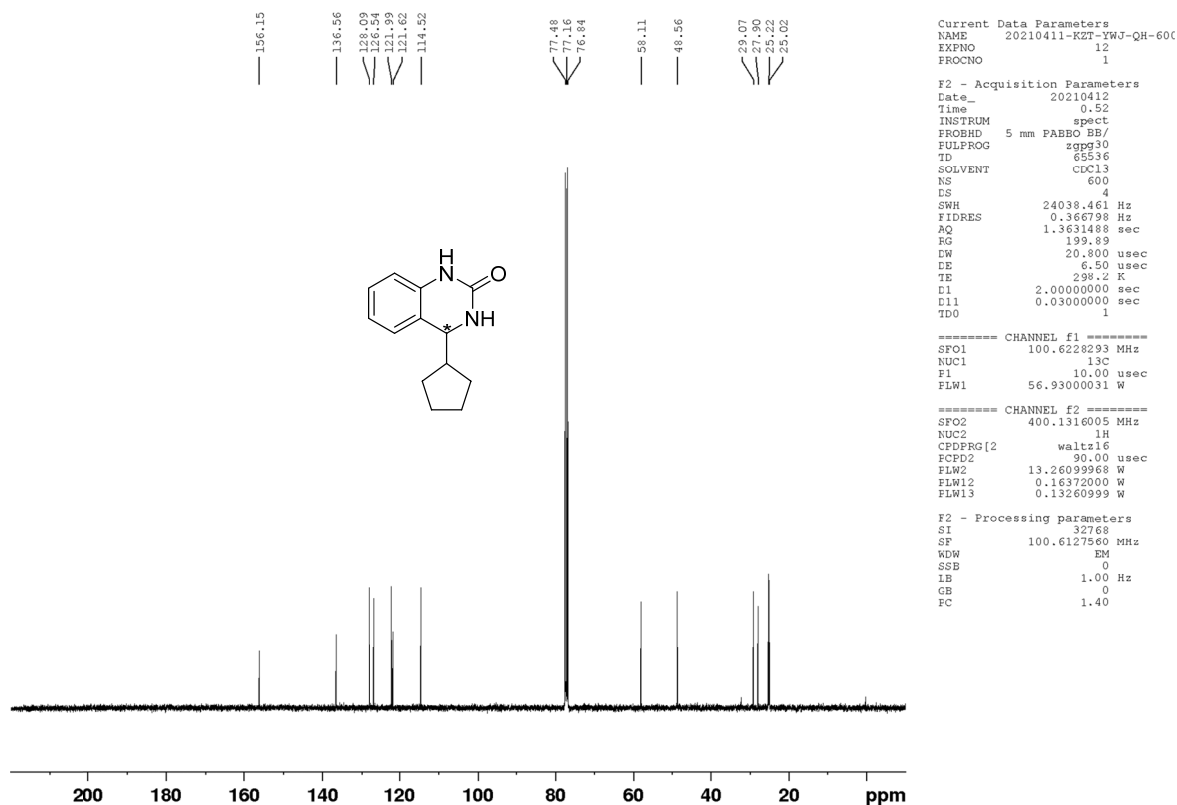
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Current Data Parameters
NAME          new-kt
EXPNO        10
PROCNO       1

F2 - Acquisition Parameters
Date_        20210411
Time         20.59
INSTRUM      spect
PROBHD       5 mm PABBO BB/
PULPROG      zg30
TD           65536
SOLVENT      CDCl3
NS           16
DS           2
SWH          8012.820 Hz
FIDRES      0.122266 Hz
AQ          4.0894455 sec
RG          89.48
DW          62.400 usec
DE          6.50 usec
TE          298.2 K
D1          1.0000000 sec
TDO         1

===== CHANNEL f1 =====
SFO1         400.1324710 MHz
NUC1         1H
P1           10.00 usec
PLW1         13.26099968 W

F2 - Processing parameters
SI           65536
SF           400.1300096 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           1.00
    
```



```

Current Data Parameters
NAME          20210411-KZT-YWJ-QH-60C
EXPNO        12
PROCNO       1

F2 - Acquisition Parameters
Date_        20210412
Time         0.52
INSTRUM      spect
PROBHD       5 mm PABBO BB/
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           600
DS           4
SWH          24038.461 Hz
FIDRES      0.366798 Hz
AQ          1.3631488 sec
RG          199.89
DW          20.800 usec
DE          6.50 usec
TE          298.2 K
D1          2.0000000 sec
F11         0.0300000 sec
TDO         1

===== CHANNEL f1 =====
SFO1         100.6228293 MHz
NUC1         13C
P1           10.00 usec
PLW1         56.93000031 W

===== CHANNEL f2 =====
SFO2         400.1316005 MHz
NUC2         1H
CPDPRG2      waltz16
F2PD2        90.00 usec
ELW2         13.26099968 W
ELW12        0.16372000 W
ELW13        0.13260999 W

F2 - Processing parameters
SI           32768
SF           100.6127560 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
    
```

Figure S50. ¹H NMR and ¹³C NMR spectra of **4d**

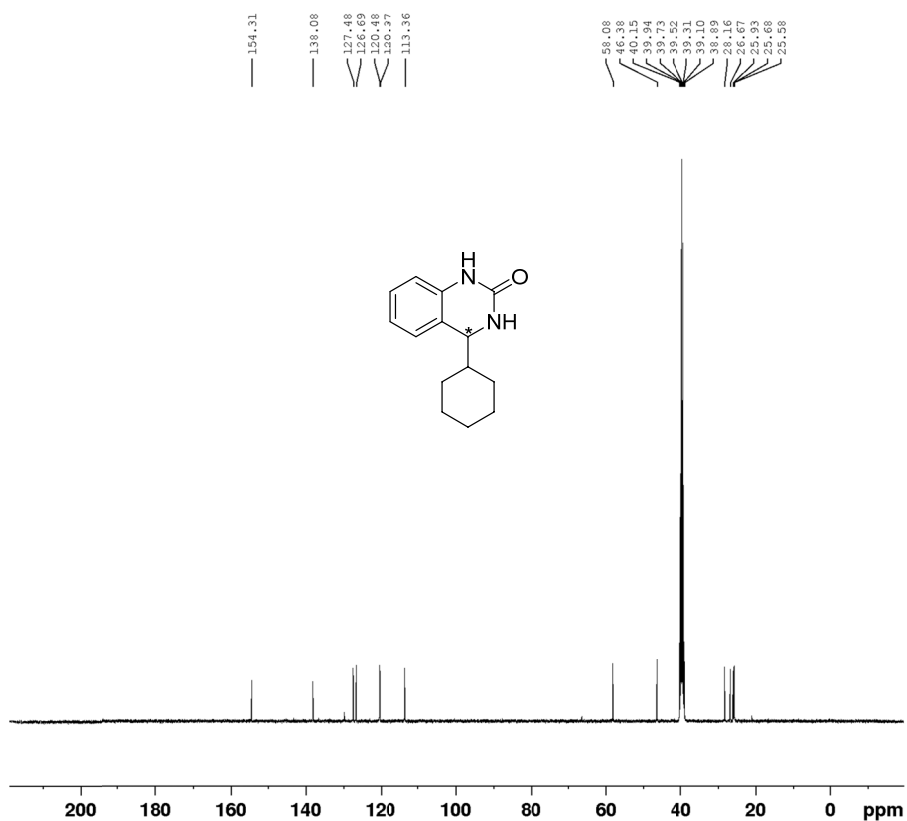
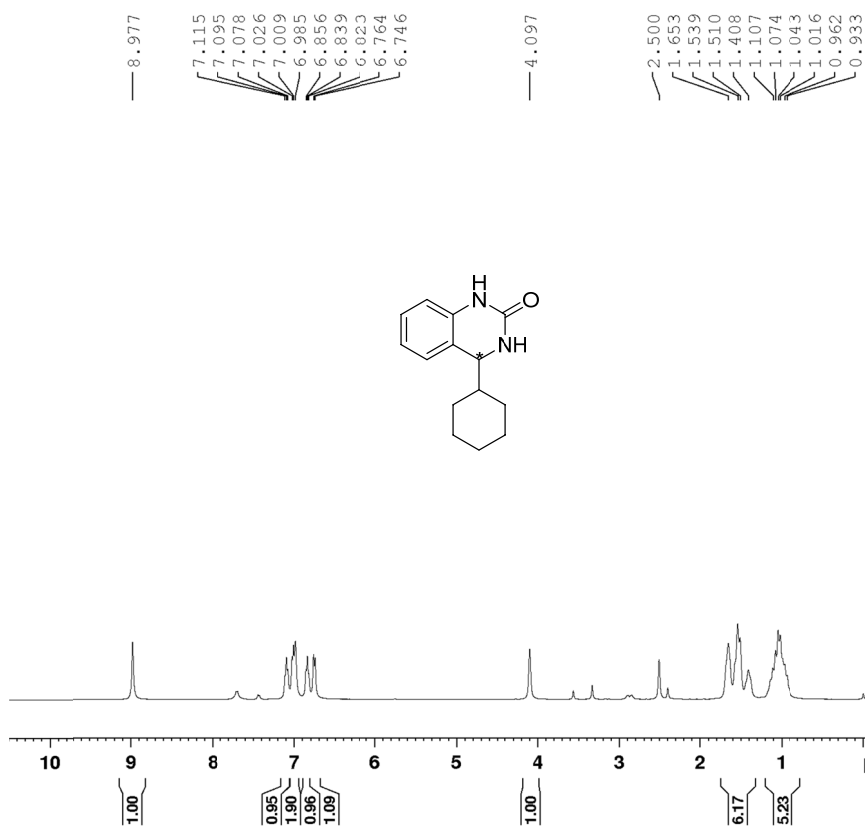
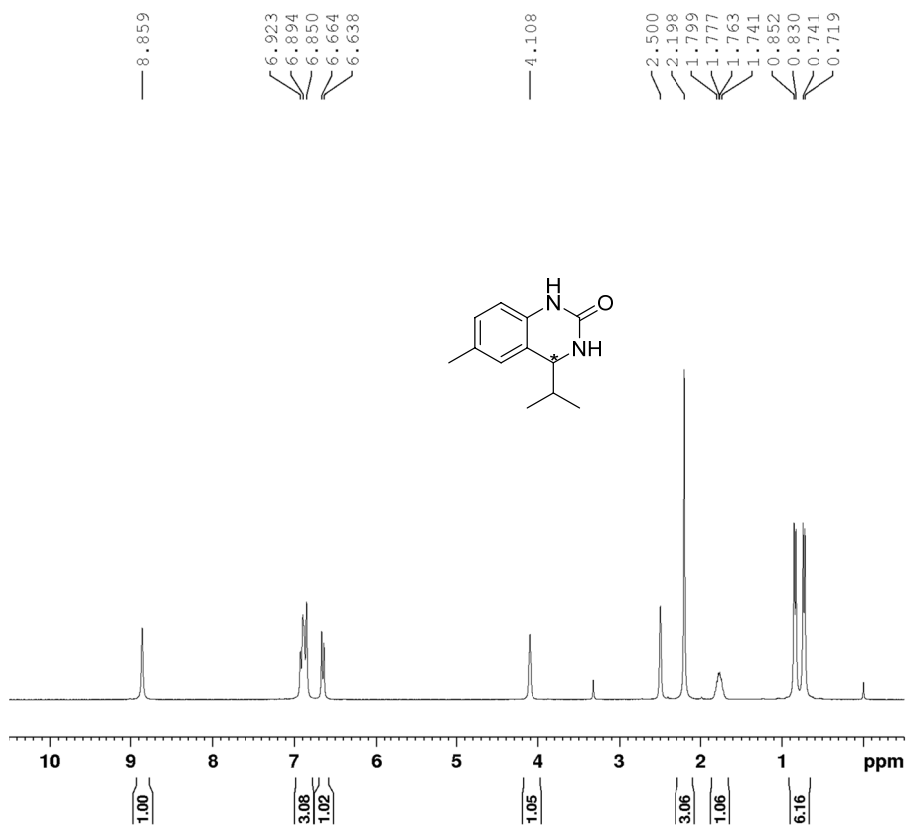


Figure S51. ¹H NMR and ¹³C NMR spectra of 4e

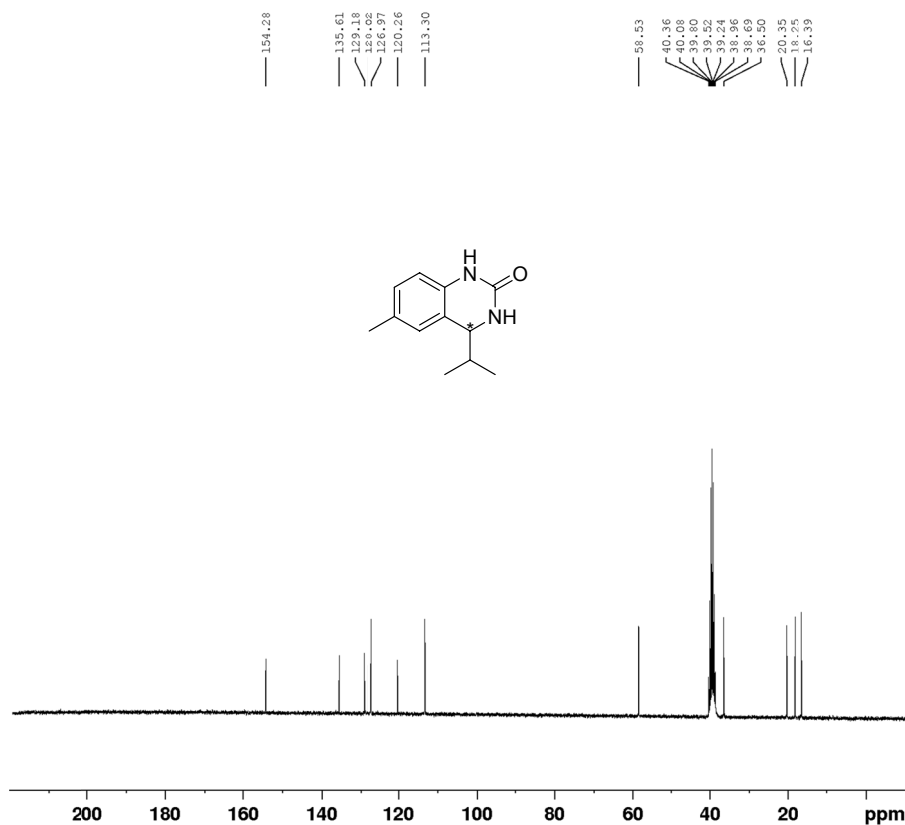


Current Data Parameters
 NAME KZT-Me-IPR-Qh
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210411
 Time 22.43
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 209.09
 DW 83.200 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 65536
 SF 300.1300024 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME KZT-Me-IPR-Qh
 EXPNO 5
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210411
 Time 23.49
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 1024
 DS 4
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175317 sec
 RG 209.09
 DW 21.733 usec
 DE 6.50 usec
 TE 298.3 K
 D1 2.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 11.00 usec
 PL1 120.00 dB
 PL1W 0 W
 SFO1 75.4752949 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677843 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure S52. ¹H NMR and ¹³C NMR spectra of 4f

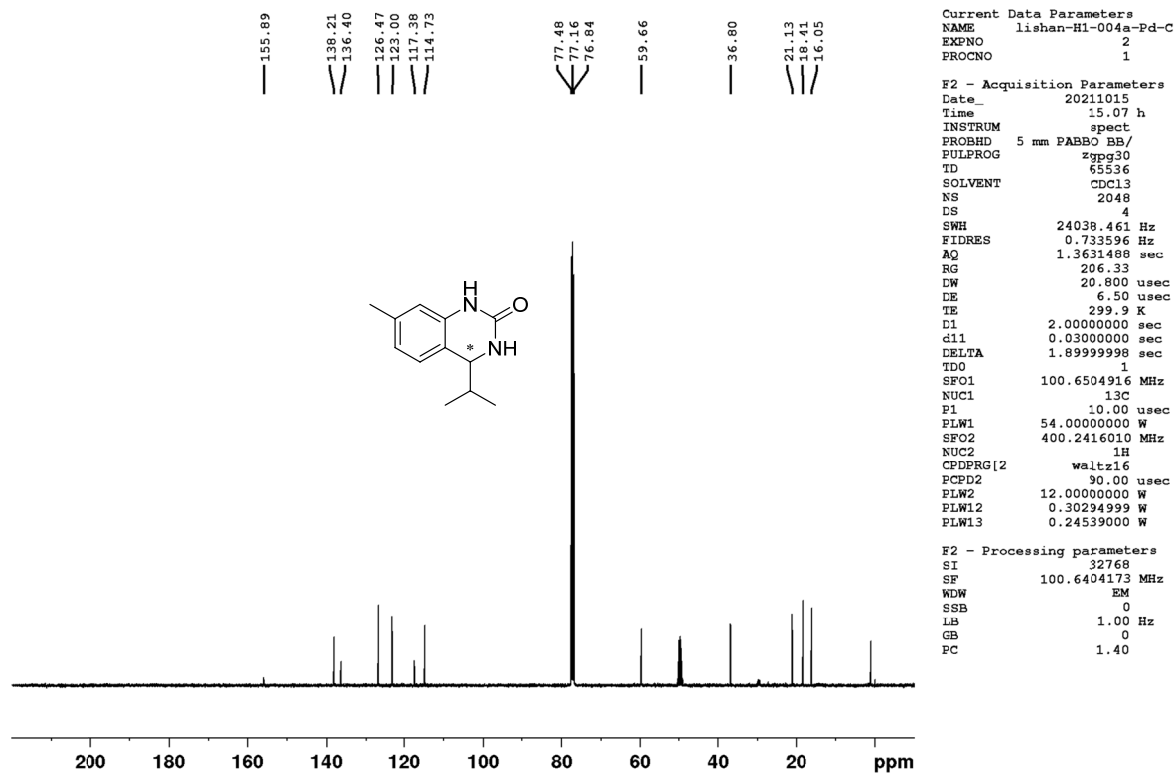
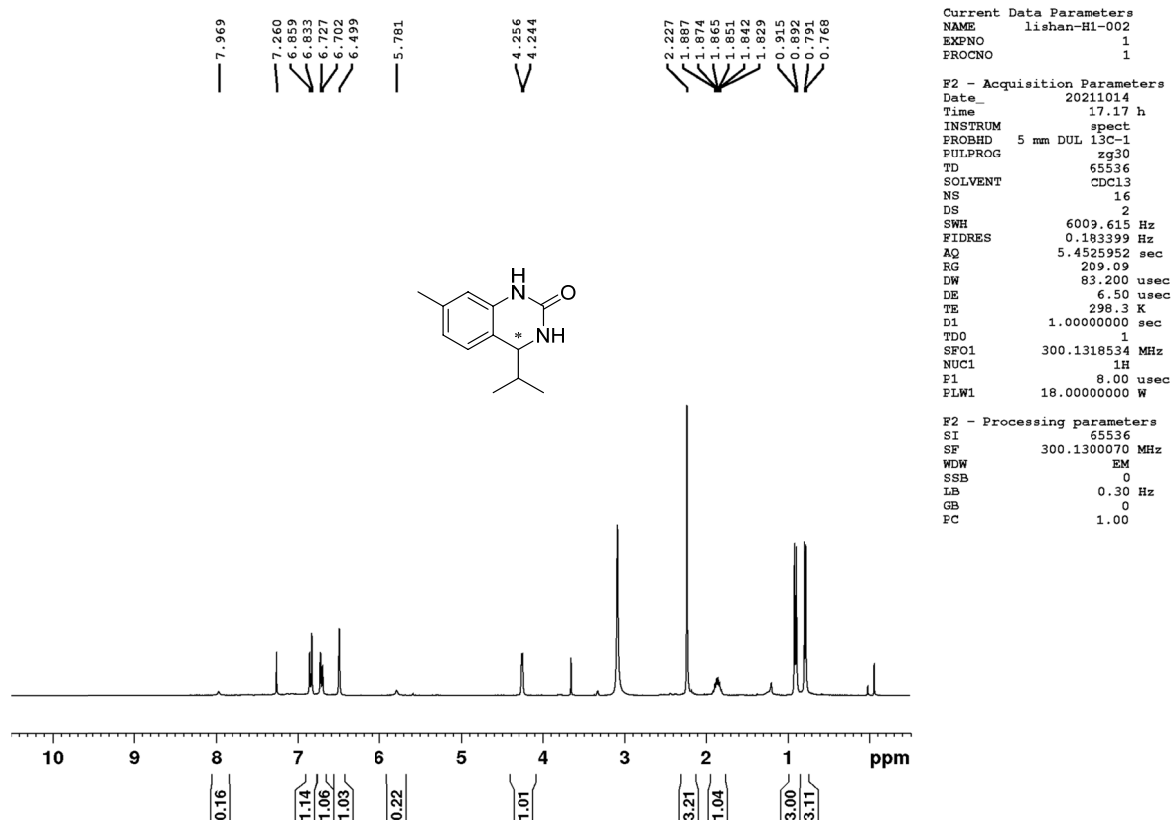


Figure S53. ¹H NMR and ¹³C NMR spectra of **4g**

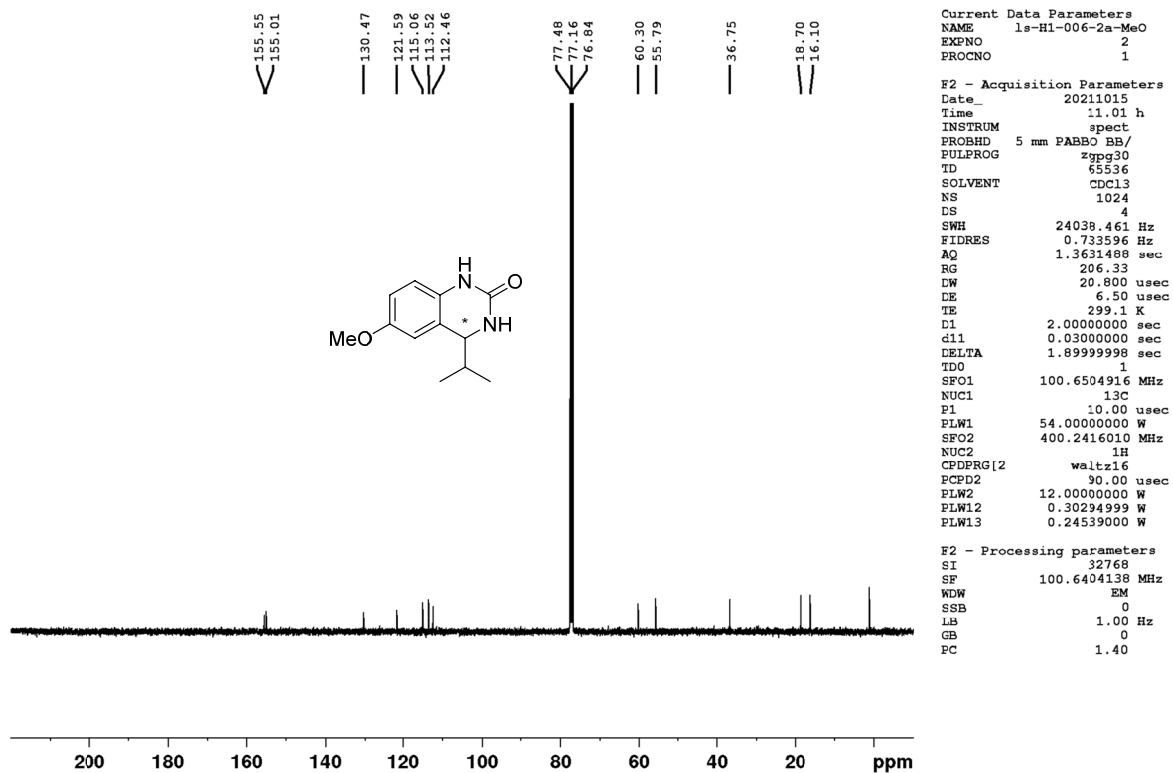
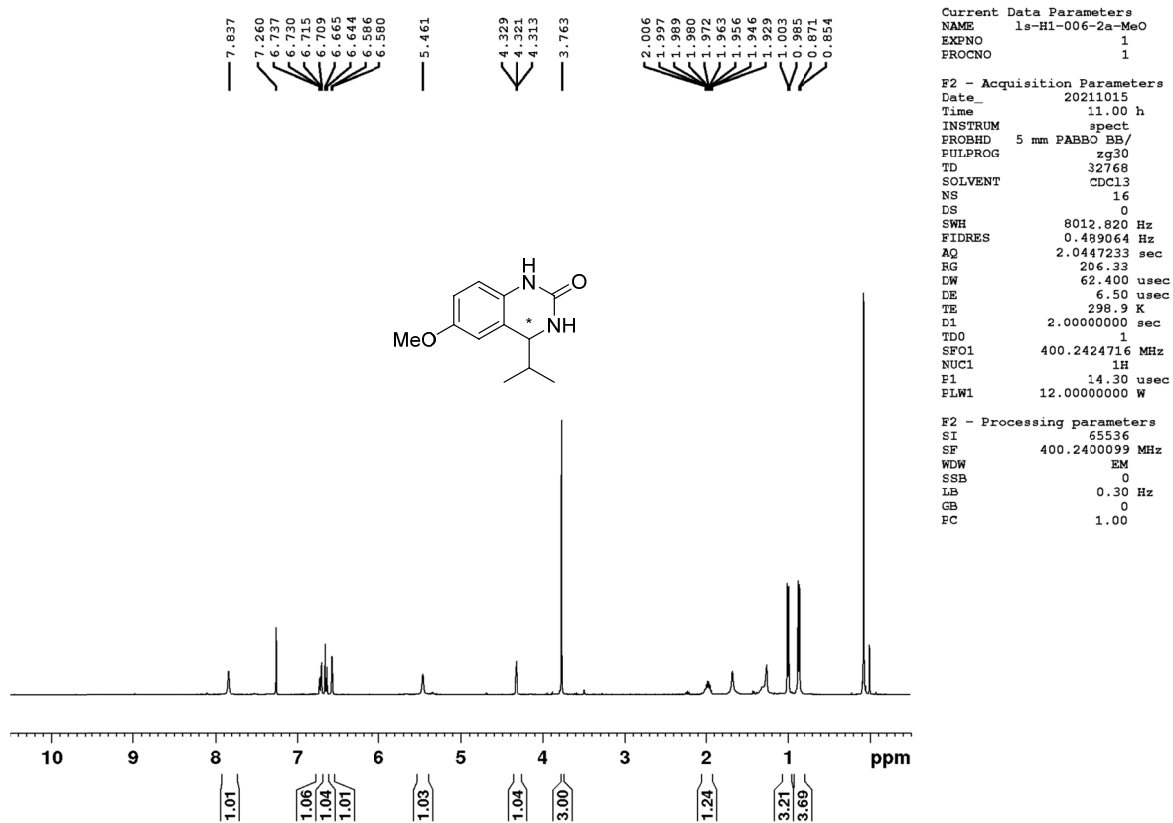


Figure S54. ¹H NMR and ¹³C NMR spectra of **4h**

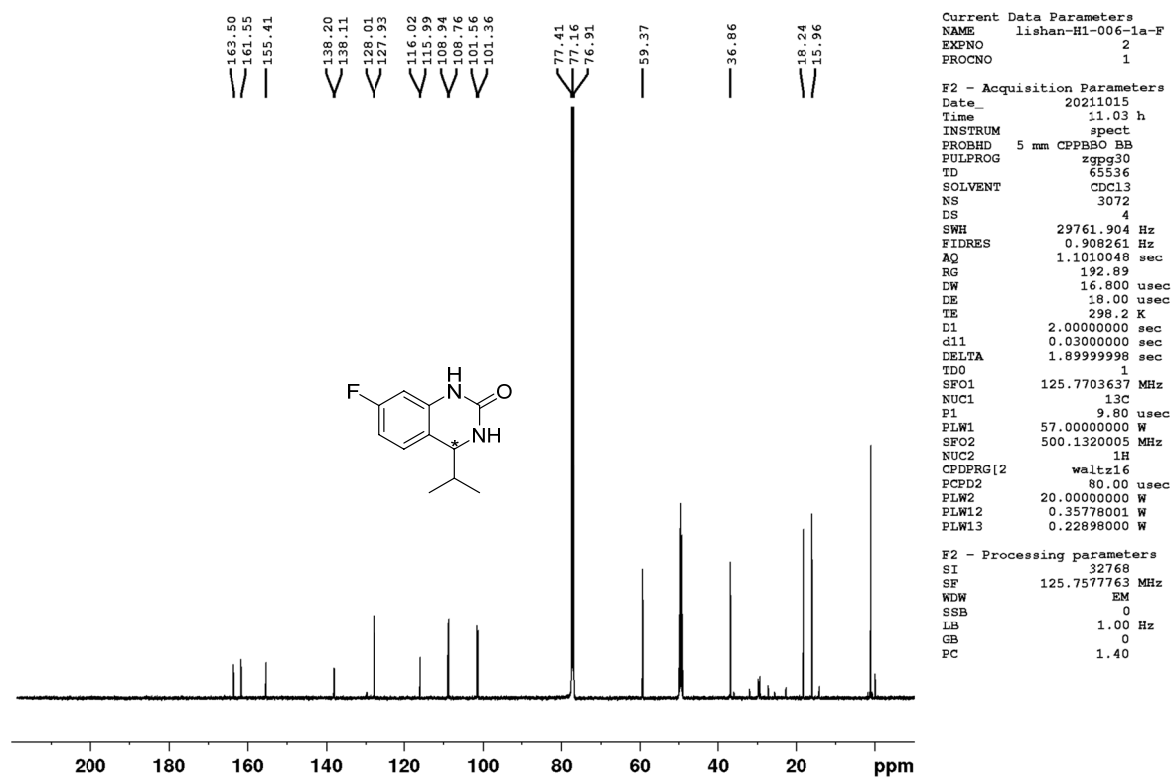
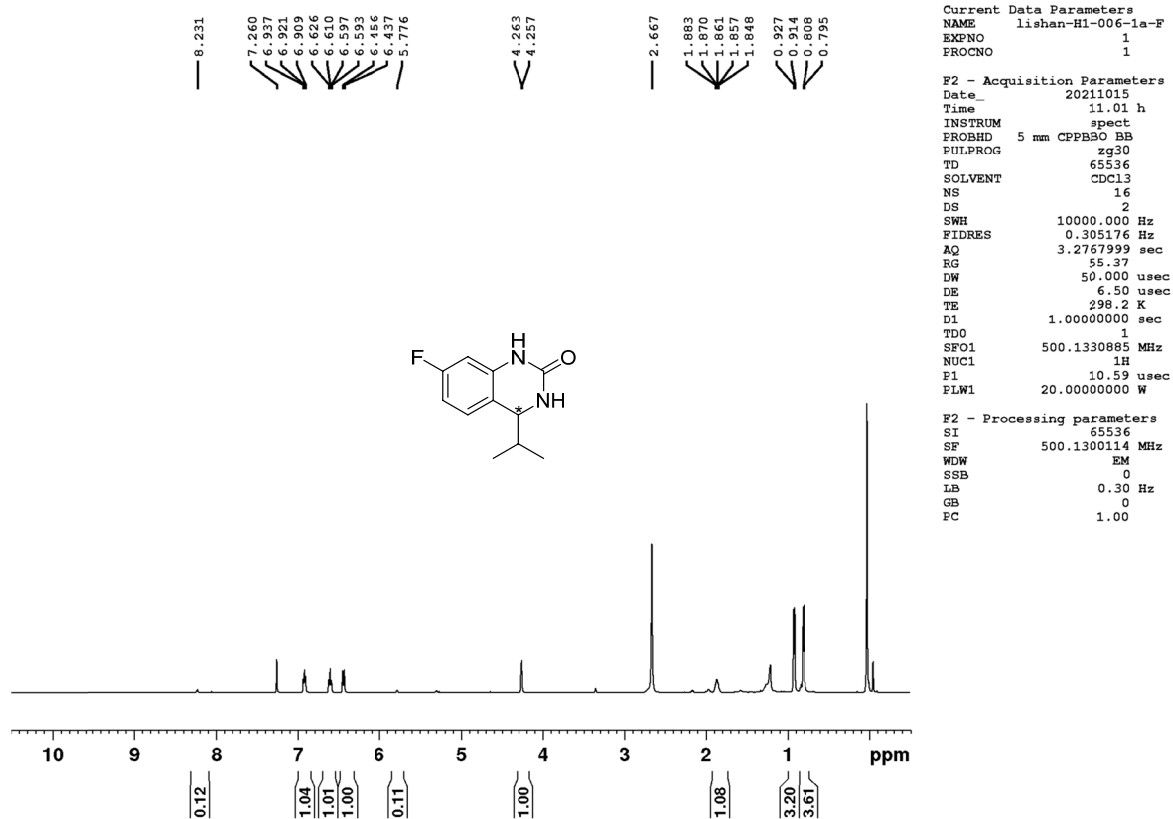
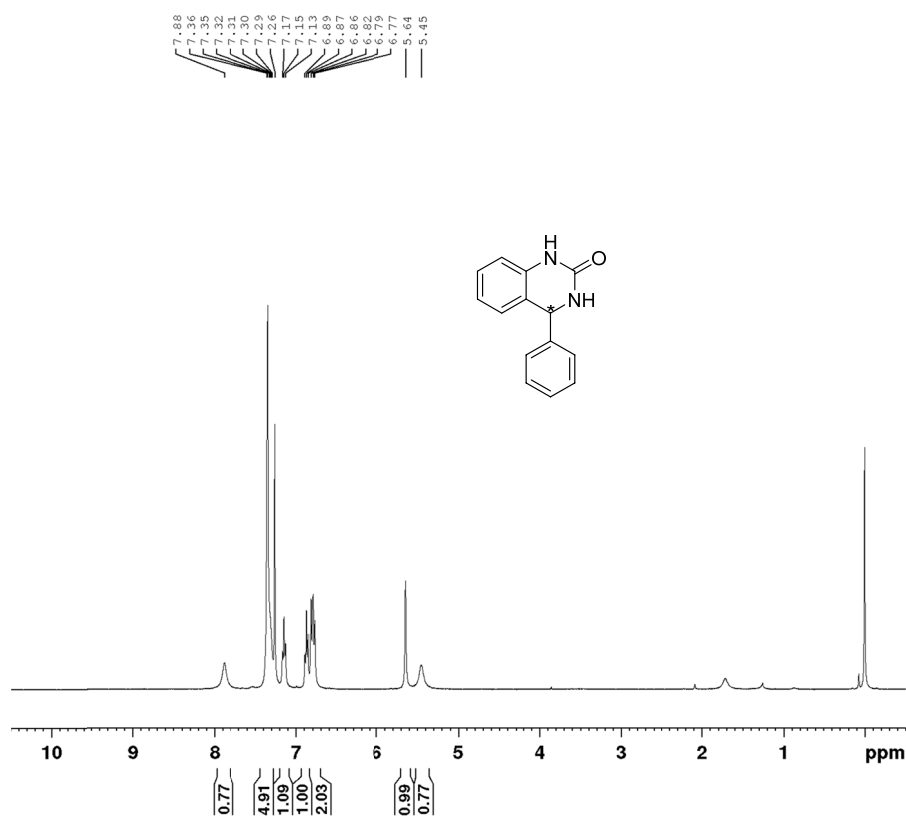


Figure S55. ¹H NMR and ¹³C NMR spectra of **4i**



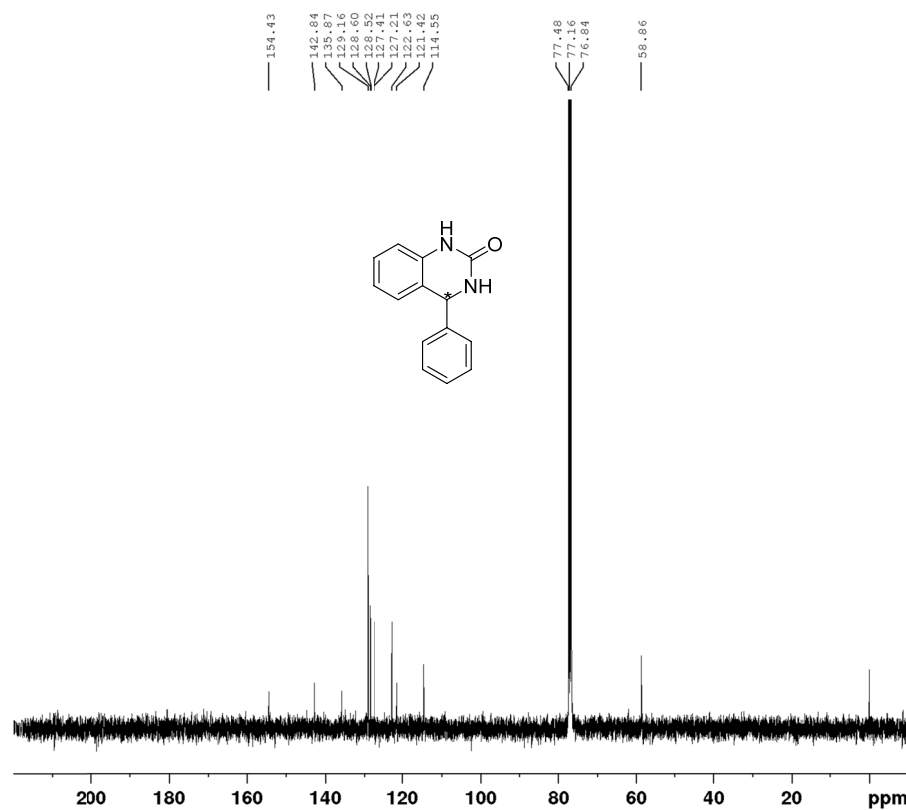
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Current Data Parameters
NAME          new-kzt
EXPNO         20
PROCNO        1

F2 - Acquisition Parameters
Date_         20210405
Time          14.49
INSTRUM       spect
PROBHD        5 mm PABBO BB/
PULPROG       zg30
TD            65536
SOLVENT       CDCl3
NS            16
DS            2
SWH           8012.820 Hz
FIDRES        0.122266 Hz
AQ            4.0694465 sec
RG            158.77
DW            62.400 usec
DE            6.50 usec
TE            299.2 K
D1            1.00000000 sec
TD0           1

----- CHANNEL f1 -----
SF01          400.1324710 MHz
NUC1           1H
P1            10.00 usec
PLW1          13.26099968 W

F2 - Processing parameters
SI            65536
SF            400.1300098 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```



```

Current Data Parameters
NAME          new-kzt
EXPNO         10
PROCNO        1

F2 - Acquisition Parameters
Date_         20210405
Time          22.34
INSTRUM       spect
PROBHD        5 mm PABBO BB/
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            1024
DS            4
SWH           24038.461 Hz
FIDRES        0.366798 Hz
AQ            1.3631488 sec
RG            199.89
DW            20.800 usec
DE            6.50 usec
TE            300.2 K
D1            2.00000000 sec
D11           0.03000000 sec
TD0           1

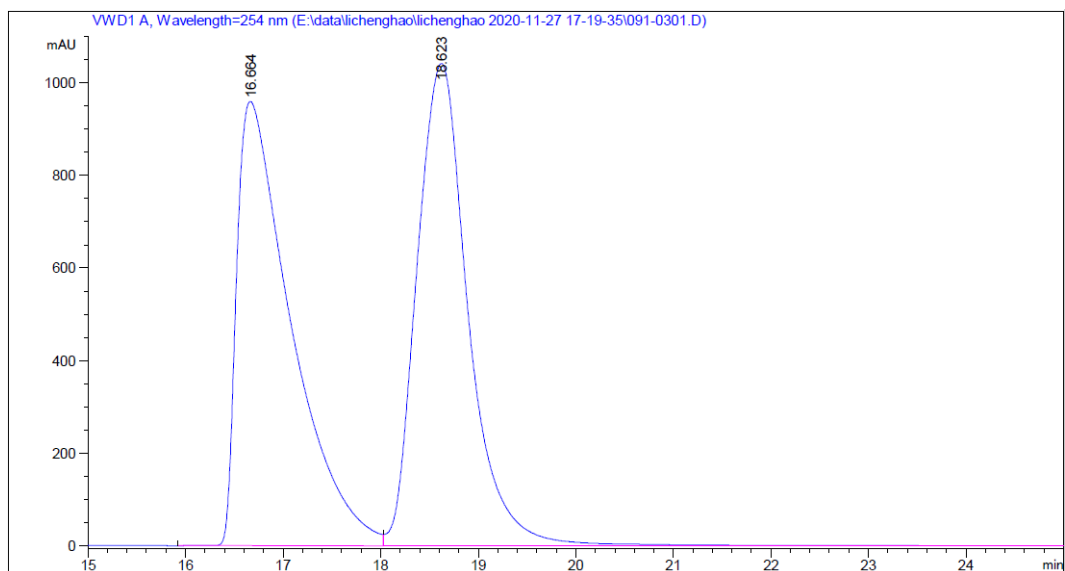
----- CHANNEL f1 -----
SF01          100.6228293 MHz
NUC1           13C
P1            10.00 usec
PLW1          56.93000031 W

----- CHANNEL f2 -----
SF02          400.1316005 MHz
NUC2           1H
PCPDPRG[2]    waltz16
PCPD2         90.00 usec
PLW2          13.26099968 W
PLW12         0.16372000 W
PLW13         0.13260999 W

F2 - Processing parameters
SI            32768
SF            100.6127540 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40
    
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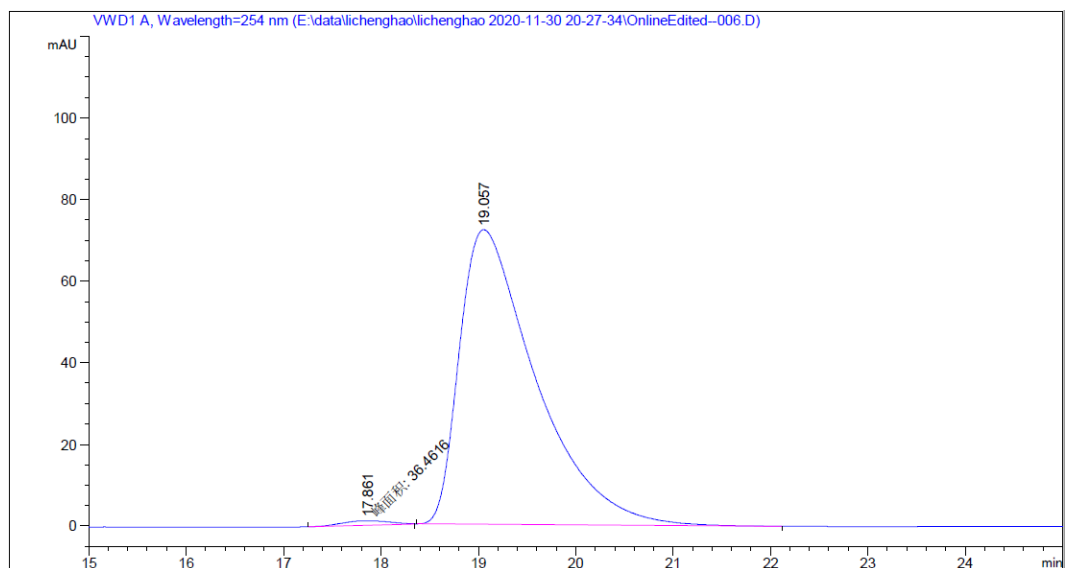
8. Copy of HPLC spectra

Figure S56. Copy of HPLC spectra of 2a



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	16.664	BV	0.5657	3.70550e4	959.17773	49.3659
2	18.623	VB	0.5709	3.80069e4	1041.13806	50.6341

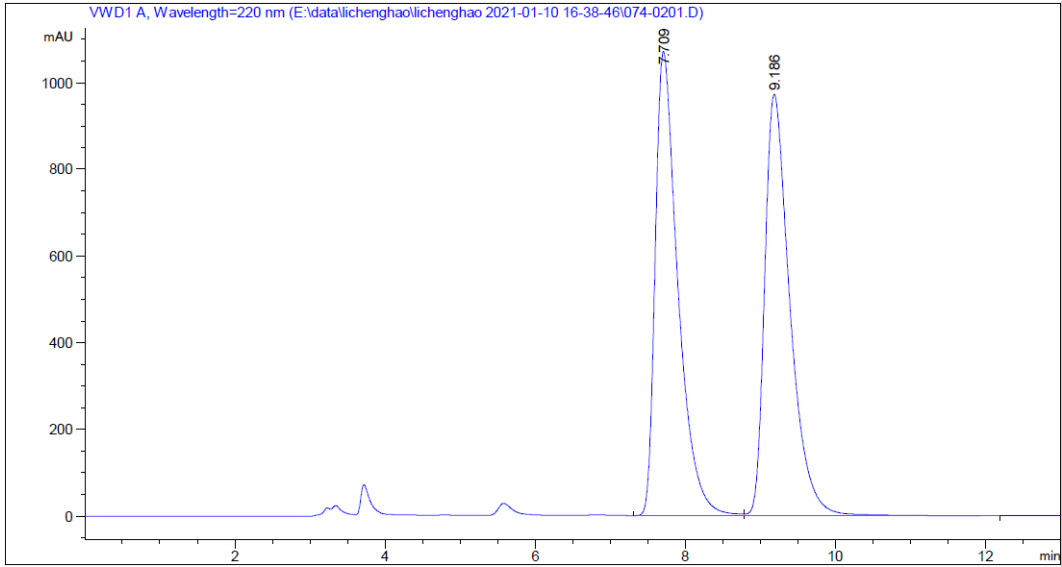
总量 : 7.50619e4 2000.31580



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	17.861	MM	0.5632	36.46165	1.07902	0.9182
2	19.057	BB	0.8105	3934.53931	72.23414	99.0818

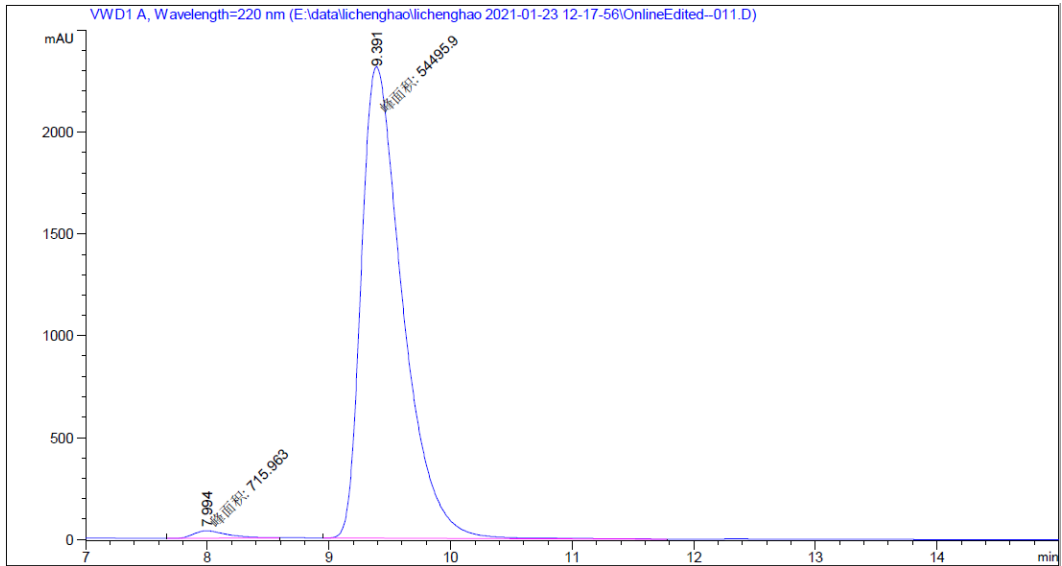
总量 : 3971.00095 73.31316

Figure S57. Copy of HPLC spectra of 2b



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.709	BV	0.3128	2.24610e4	1070.52710	49.7725
2	9.186	VB	0.3510	2.26663e4	972.14868	50.2275

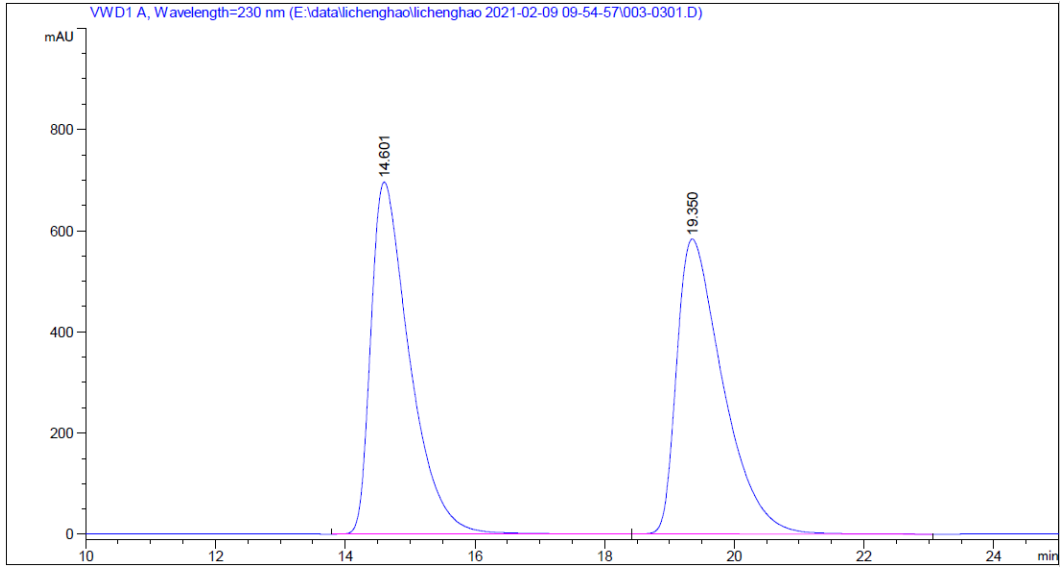
总量 : 4.51273e4 2042.67578



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.994	MM	0.3240	715.96295	36.82597	1.2968
2	9.391	MM	0.3923	5.44959e4	2315.45776	98.7032

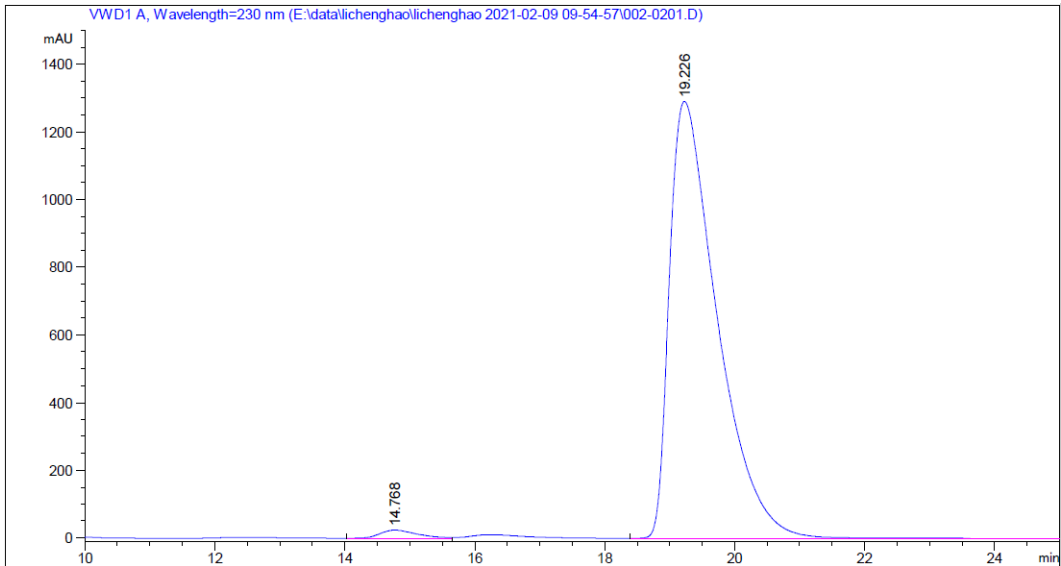
总量 : 5.52119e4 2352.28373

Figure S58. Copy of HPLC spectra of 2c



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	14.601	BB	0.6184	2.87642e4	695.66376	50.0060
2	19.350	BB	0.7444	2.87573e4	583.12433	49.9940

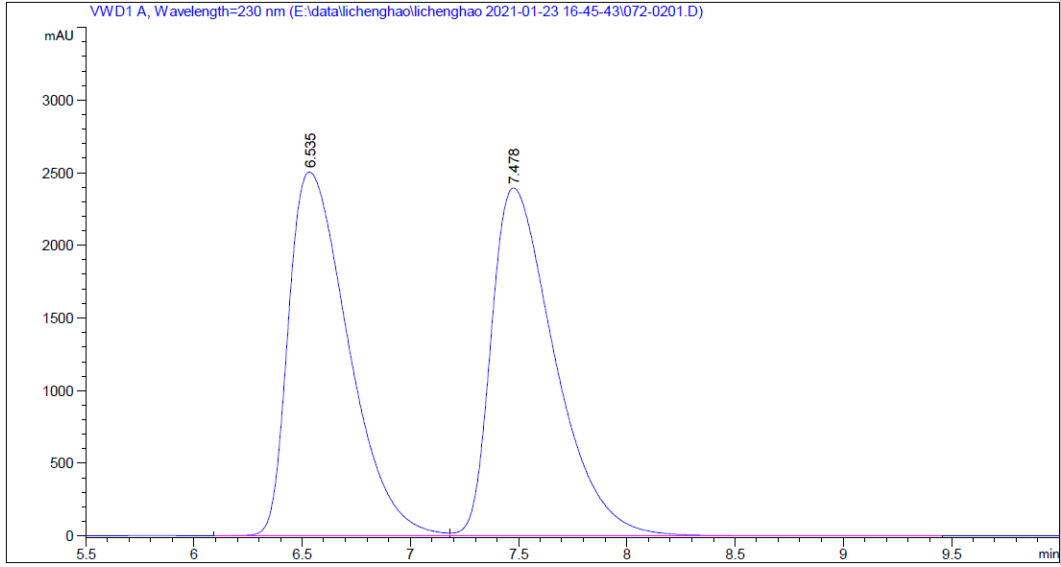
总量 : 5.75216e4 1278.78809



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	14.768	BV	0.6019	956.61603	24.05832	1.4341
2	19.226	BBA	0.7590	6.57505e4	1291.30261	98.5659

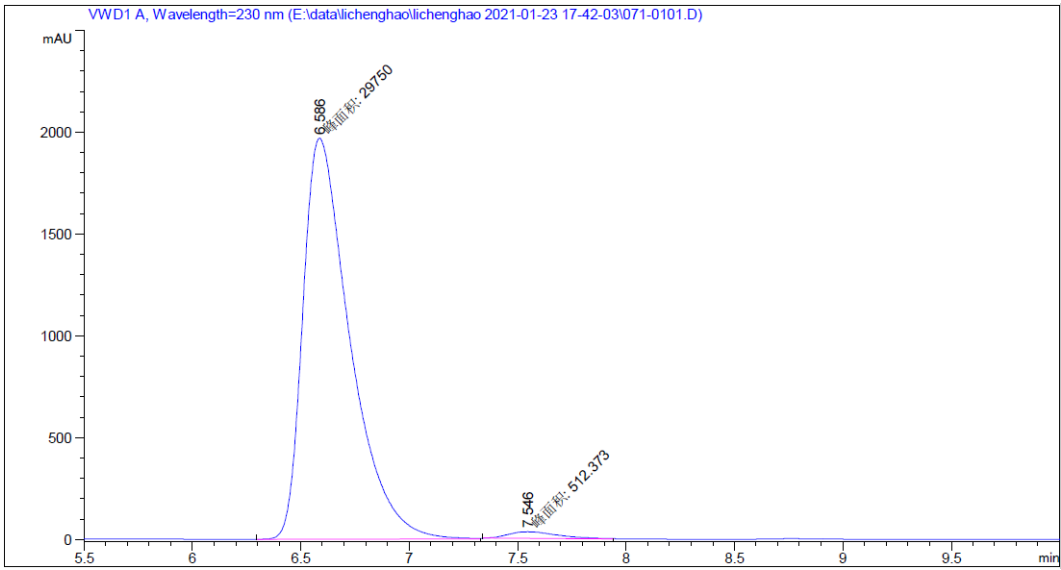
总量 : 6.67071e4 1315.36094

Figure S59. Copy of HPLC spectra of 2d



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	6.535	BV	0.2983	4.78979e4	2502.41284	49.6181
2	7.478	VB	0.3100	4.86353e4	2394.54688	50.3819

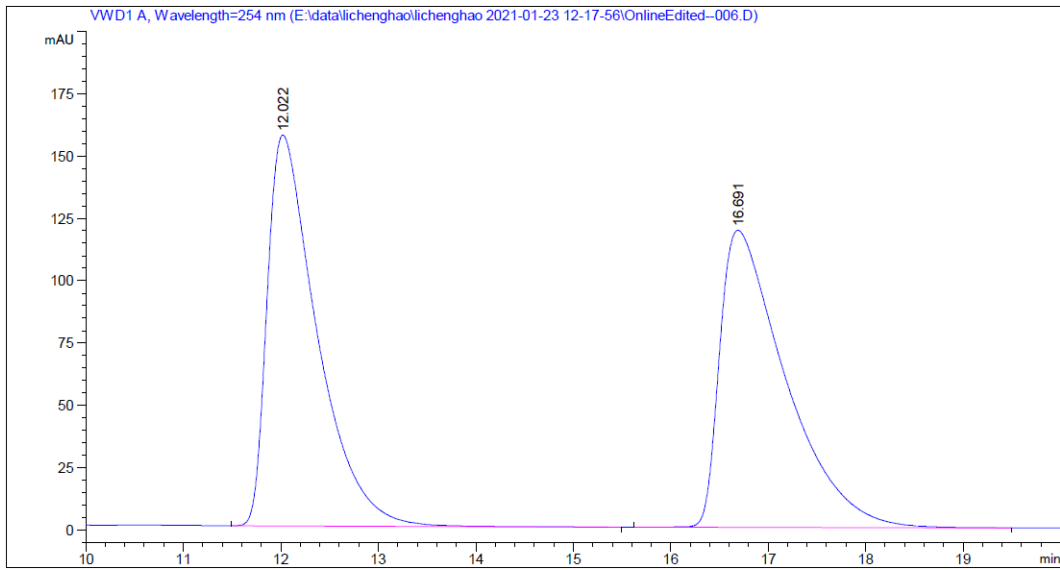
总量 : 9.65333e4 4896.95972



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	6.586	MM	0.2515	2.97500e4	1971.41284	98.3069
2	7.546	MM	0.2574	512.37317	33.17017	1.6931

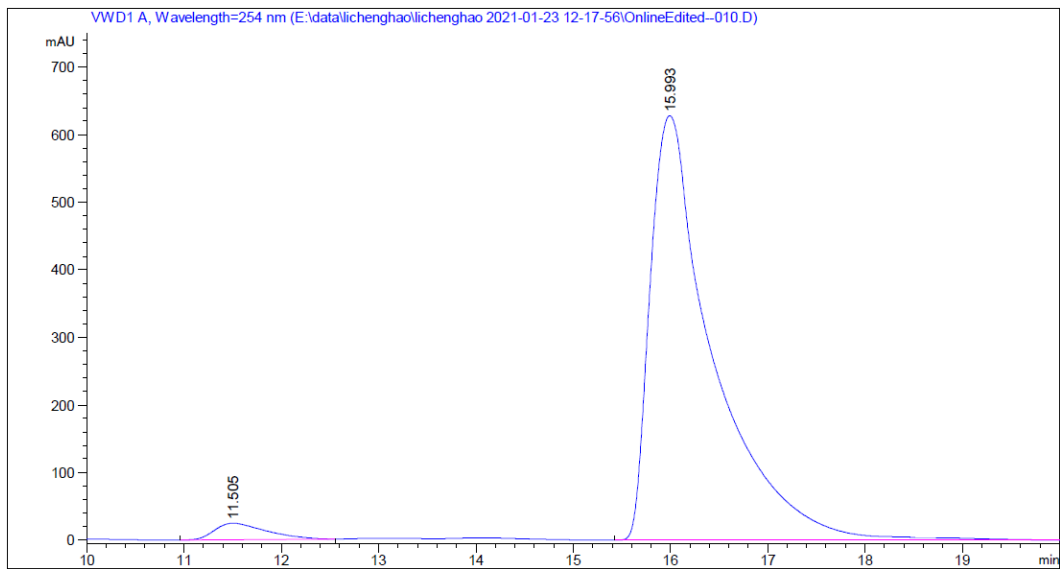
总量 : 3.02624e4 2004.58301

Figure S60. Copy of HPLC spectra of 2e



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	12.022	BB	0.5273	5570.41895	156.86319	49.9314
2	16.691	BB	0.6956	5585.72998	119.22216	50.0686

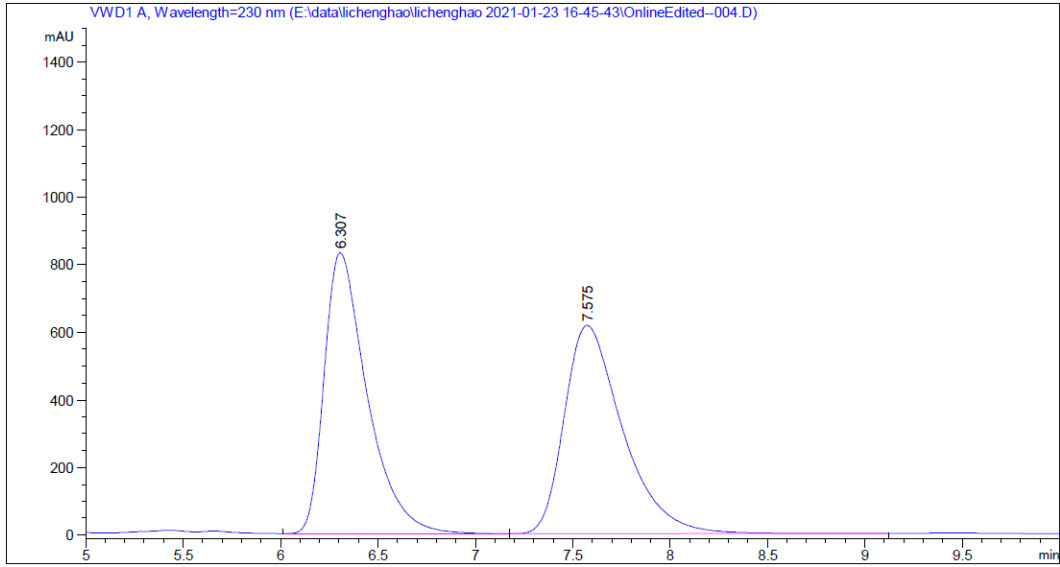
总量 : 1.11561e4 276.08535



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	11.505	BB	0.5529	899.69263	24.36135	3.0961
2	15.993	BBA	0.6397	2.81591e4	628.45605	96.9039

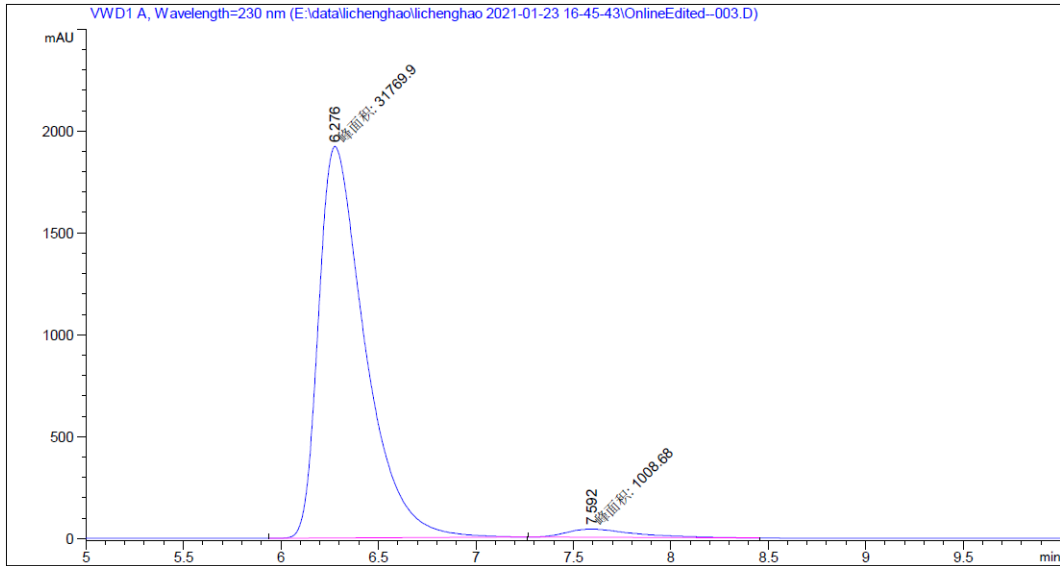
总量 : 2.90588e4 652.81741

Figure S61. Copy of HPLC spectra of 2f



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	6.307	BB	0.2300	1.27568e4	831.37976	49.8210
2	7.575	BB	0.3141	1.28484e4	616.63257	50.1790

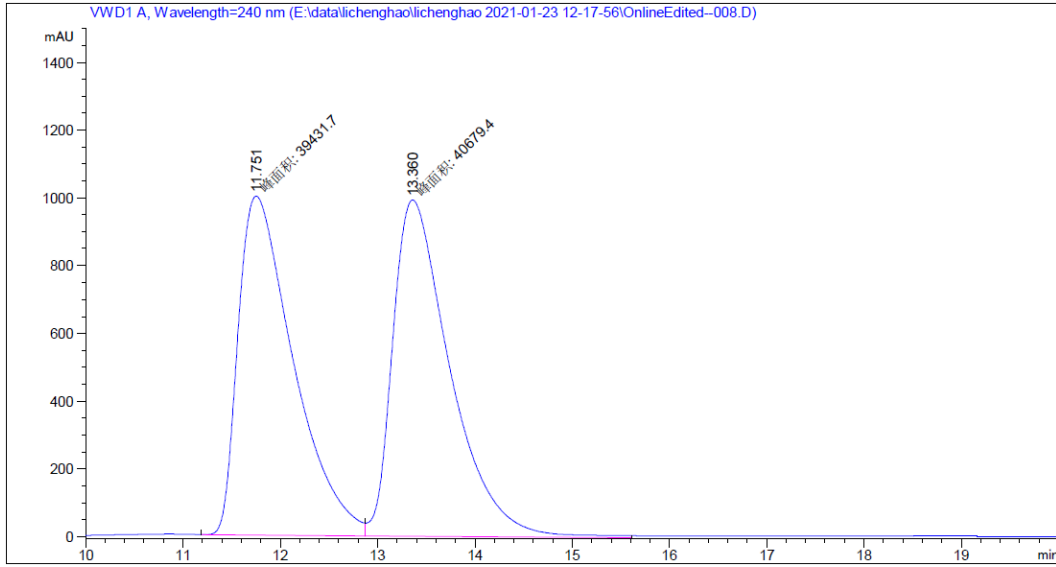
总量 : 2.56052e4 1448.01233



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	6.276	MM	0.2753	3.17699e4	1923.47571	96.9227
2	7.592	MM	0.4132	1008.68262	40.68394	3.0773

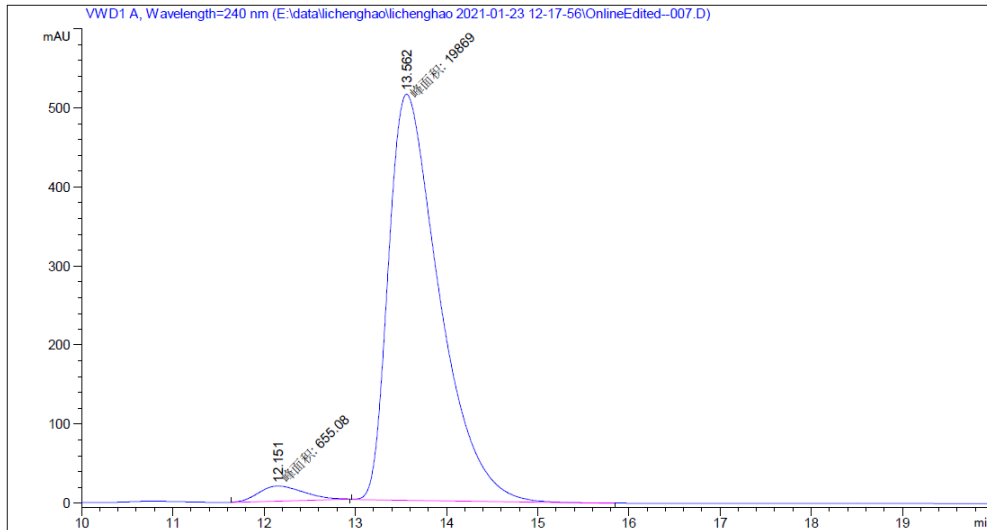
总量 : 3.27785e4 1964.15965

Figure S62. Copy of HPLC spectra of 2g



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	11.751	MF	0.6567	3.94317e4	1000.70264	49.2213
2	13.360	FM	0.6831	4.06794e4	992.52240	50.7787

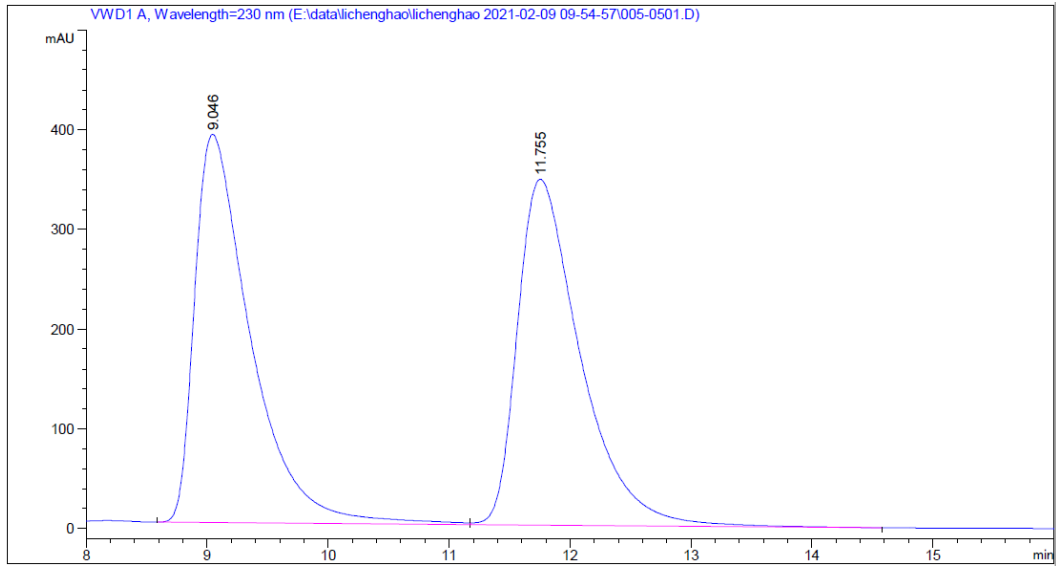
总量 : 8.01110e4 1993.22504



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	12.151	MM	0.5730	655.07996	19.05336	3.1918
2	13.562	MM	0.6449	1.98690e4	513.45728	96.8082

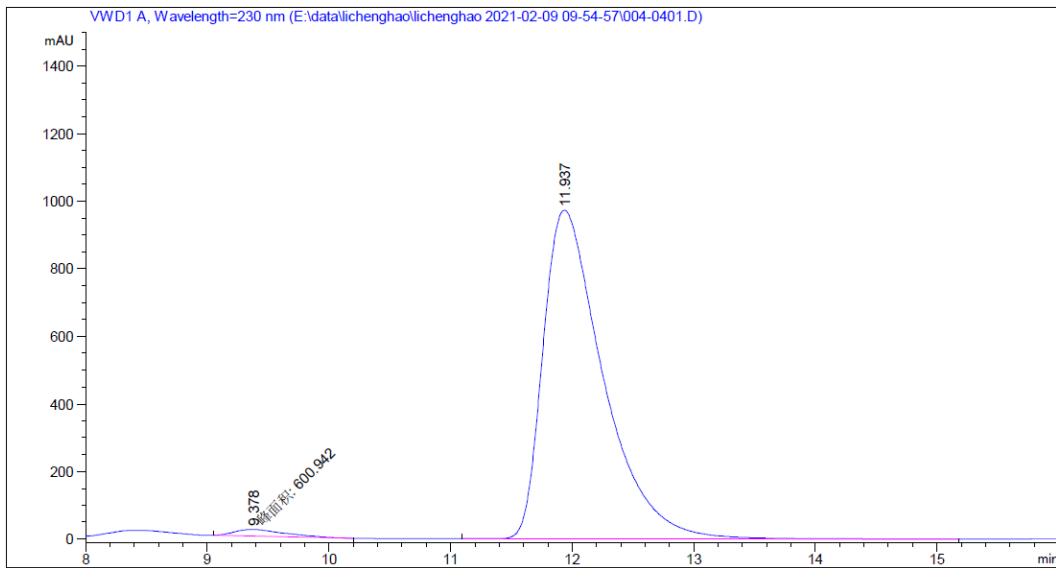
总量 : 2.05241e4 532.51063

Figure S63. Copy of HPLC spectra of 2h



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	9.046	BV	0.4759	1.25135e4	389.44415	49.9214
2	11.755	VB	0.5431	1.25529e4	346.99478	50.0786

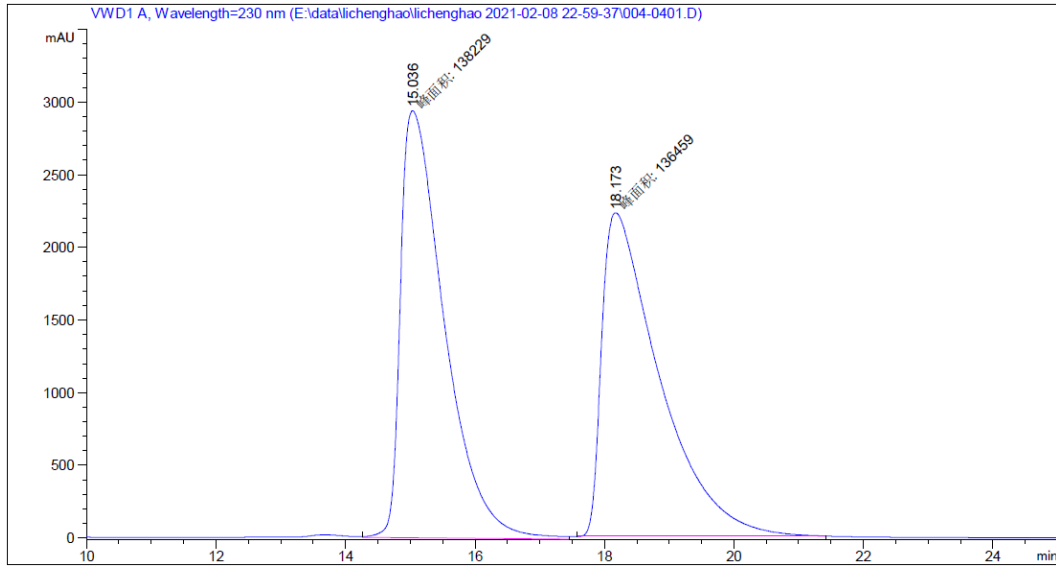
总量 : 2.50663e4 736.43893



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	9.378	MM	0.4917	600.94196	20.36783	1.7631
2	11.937	BB	0.5173	3.34829e4	973.38226	98.2369

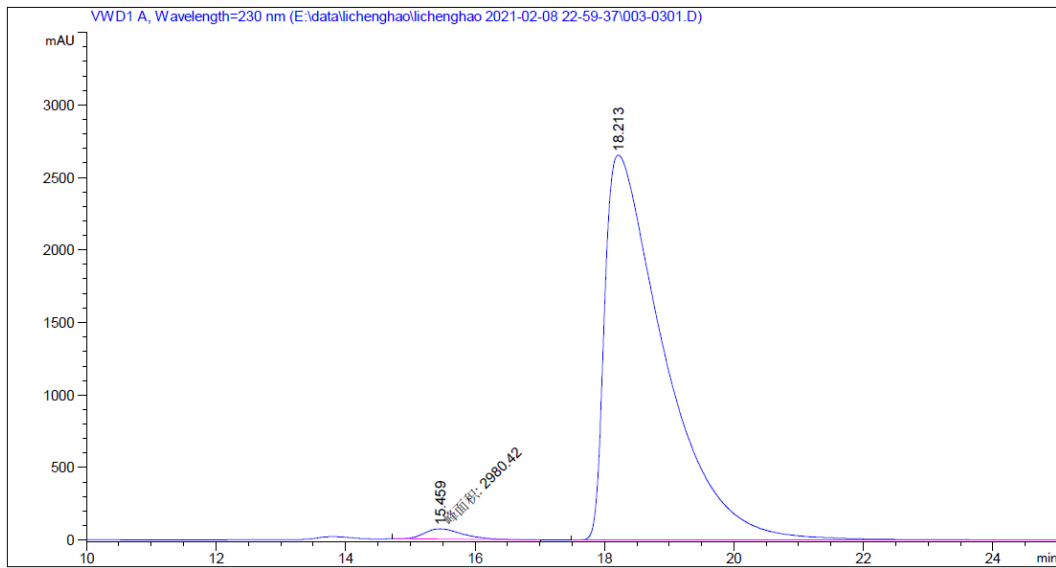
总量 : 3.40838e4 993.75009

Figure S64. Copy of HPLC spectra of 2i



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.036	MM	0.7842	1.38229e5	2937.87061	50.3221
2	18.173	MM	1.0239	1.36459e5	2221.29224	49.6779

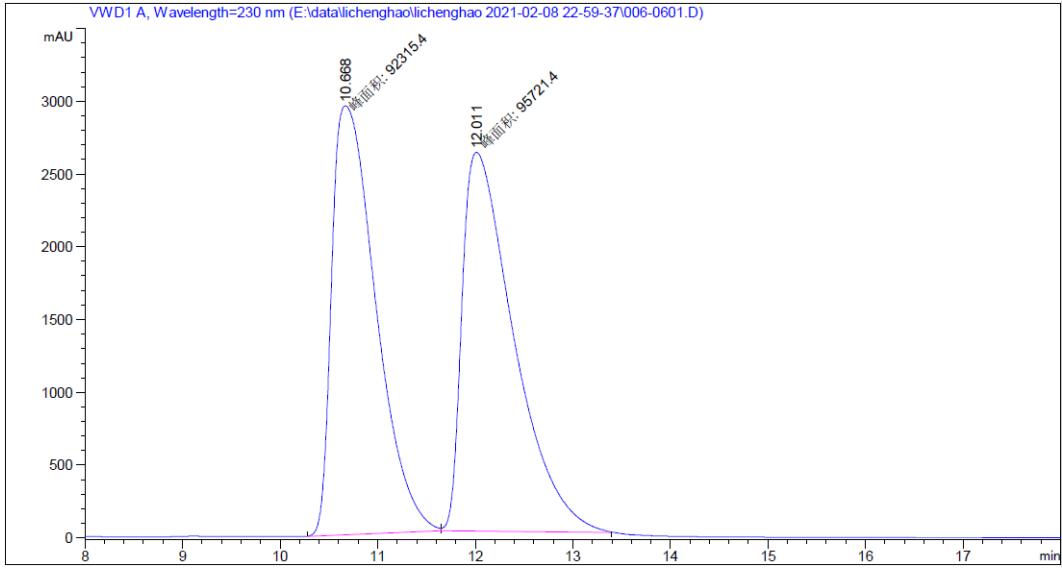
总量 : 2.74688e5 5159.16284



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.459	MM	0.6927	2980.42456	71.70898	1.7345
2	18.213	BB	0.9238	1.68853e5	2651.99976	98.2655

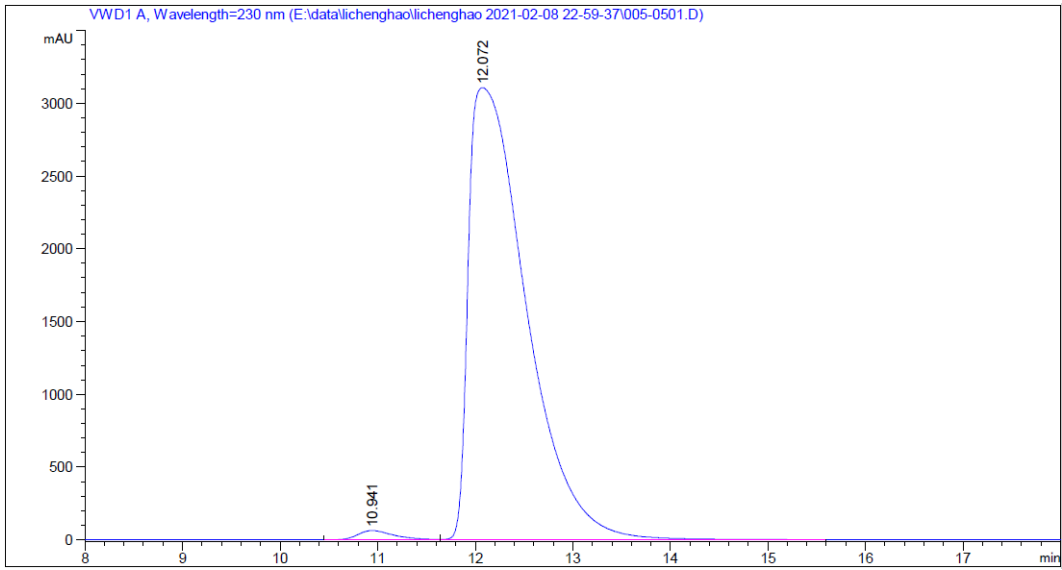
总量 : 1.71834e5 2723.70874

Figure S65. Copy of HPLC spectra of 2j



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	10.668	MM	0.5221	9.23154e4	2946.71924	49.0943
2	12.011	MM	0.6130	9.57214e4	2602.74683	50.9057

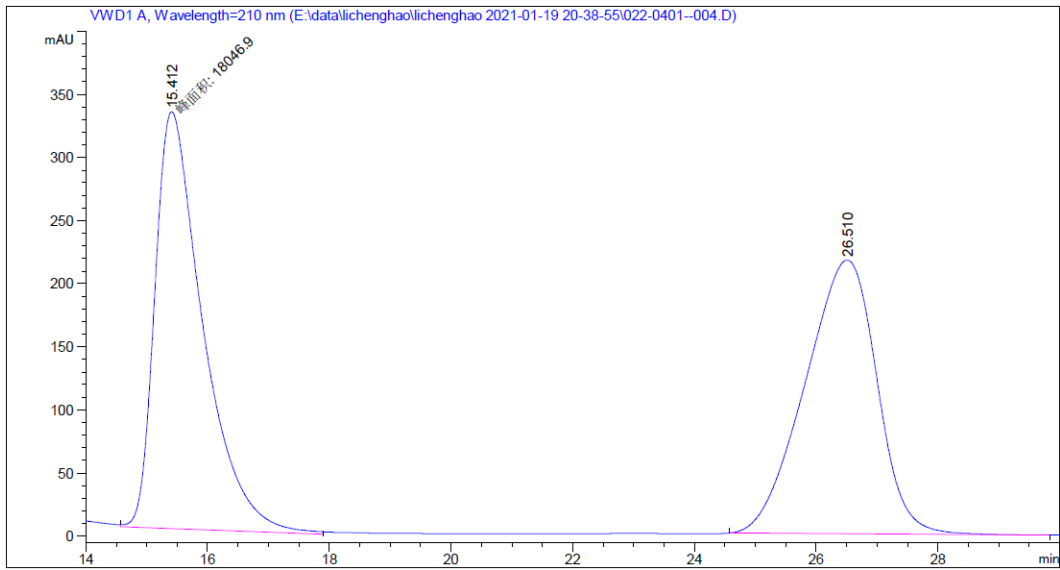
总量 : 1.88037e5 5549.46606



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	10.941	BV	0.3809	1613.04980	63.38098	1.2479
2	12.072	VB	0.6266	1.27651e5	3105.09277	98.7521

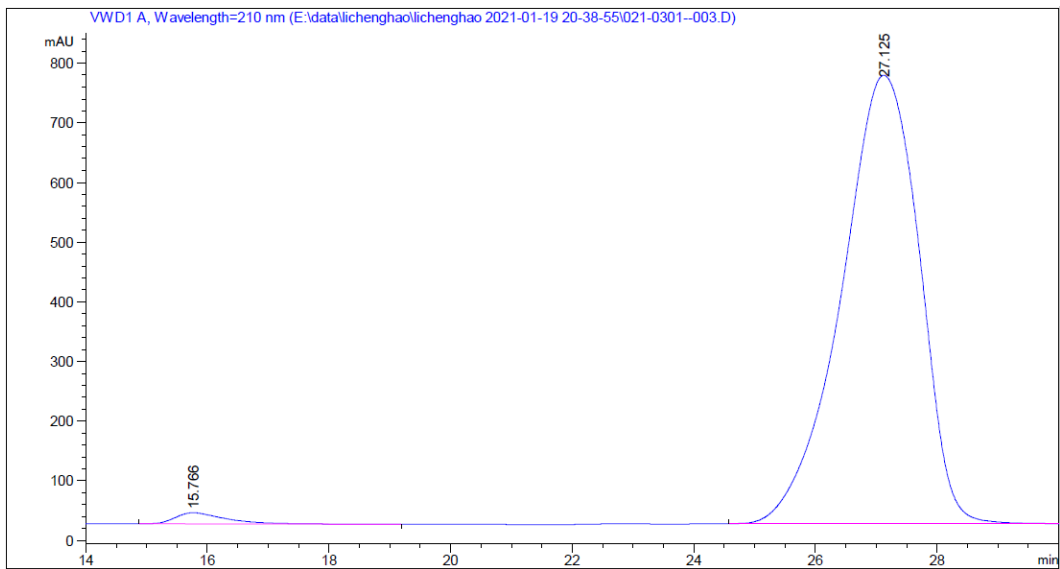
总量 : 1.29264e5 3168.47375

Figure S66. Copy of HPLC spectra of 2k



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.412	MM	0.9099	1.80469e4	330.55215	50.3990
2	26.510	BB	1.2169	1.77612e4	217.01056	49.6010

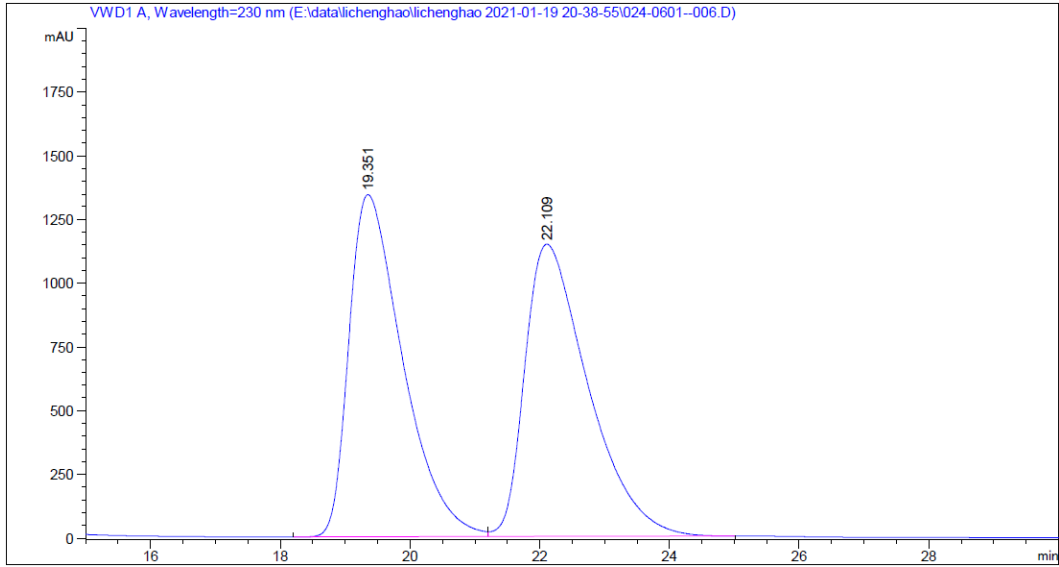
总量 : 3.58082e4 547.56271



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.766	BB	0.8515	1098.01587	18.74114	1.6127
2	27.125	BB	1.4039	6.69890e4	752.26044	98.3873

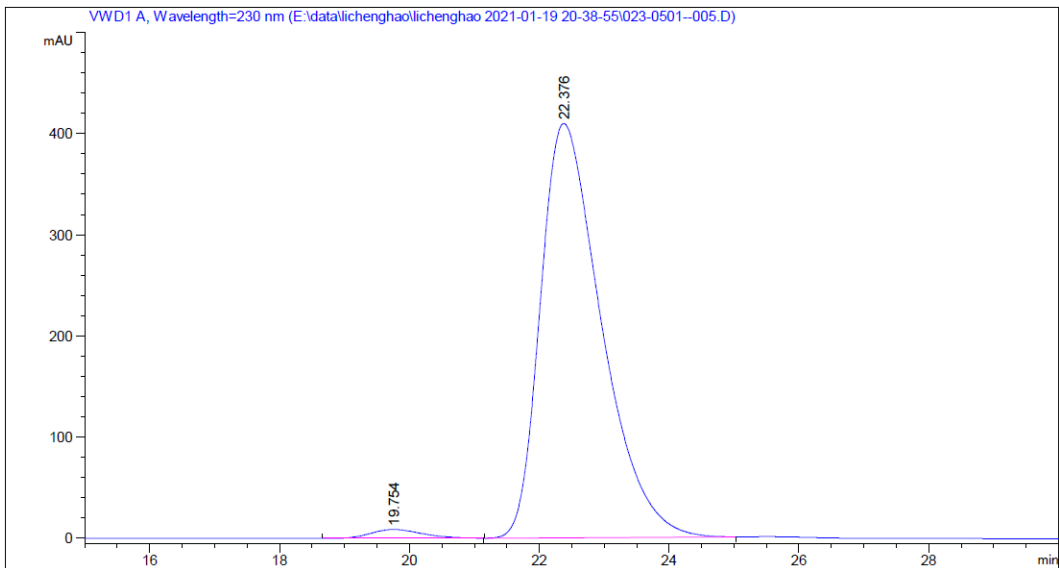
总量 : 6.80870e4 771.00158

Figure S67. Copy of HPLC spectra of 21



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	19.351	BV	0.8755	7.72724e4	1342.56787	49.8178
2	22.109	VB	1.0277	7.78378e4	1147.04639	50.1822

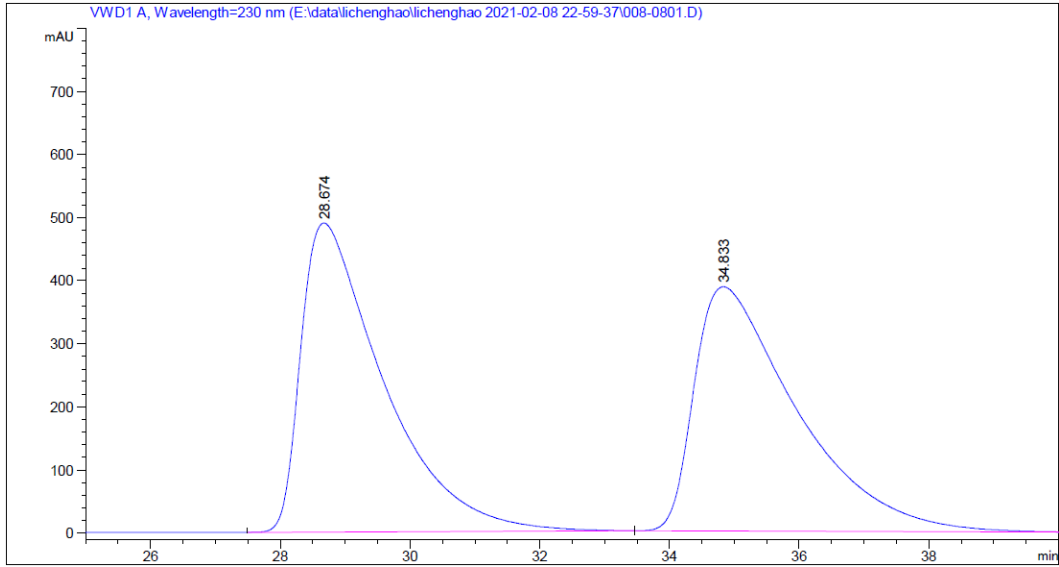
总量 : 1.55110e5 2489.61426



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	19.754	BB	0.8310	474.83063	8.72411	1.7481
2	22.376	BB	0.9947	2.66886e4	409.33804	98.2519

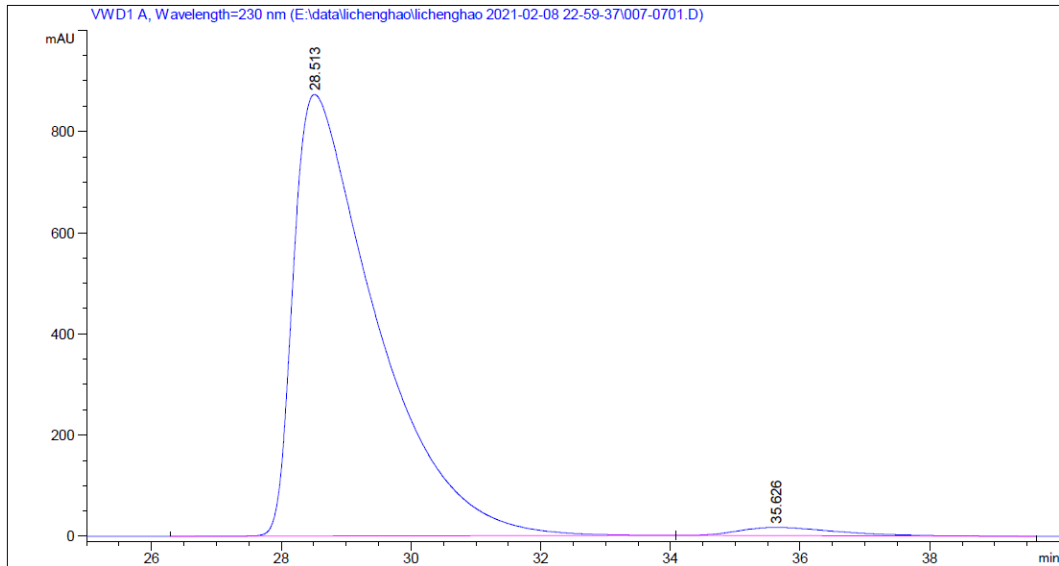
总量 : 2.71634e4 418.06216

Figure S68. Copy of HPLC spectra of 2m



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	28.674	BB	1.2903	4.31724e4	489.82043	50.4465
2	34.833	BBA	1.6079	4.24082e4	387.18066	49.5535

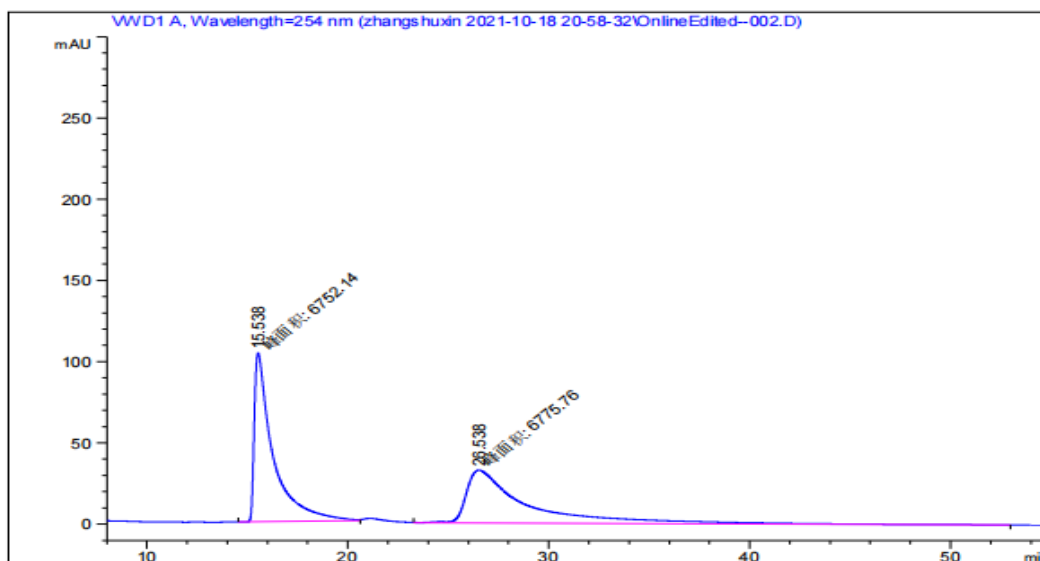
总量 : 8.55806e4 877.00110



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	28.513	BB	1.2949	7.73398e4	872.73737	97.7836
2	35.626	BBA	1.5515	1753.00134	16.51332	2.2164

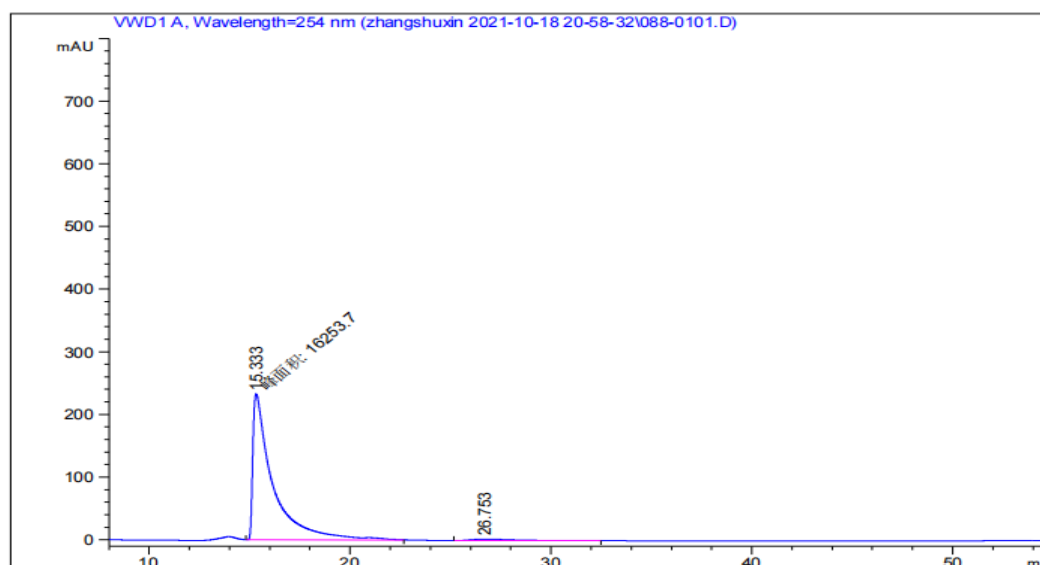
总量 : 7.90928e4 889.25069

Figure S69. Copy of HPLC spectra of 2n



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.538	MM	1.0839	6752.13721	103.82909	49.9127
2	26.538	MM	3.4808	6775.76123	32.44366	50.0873

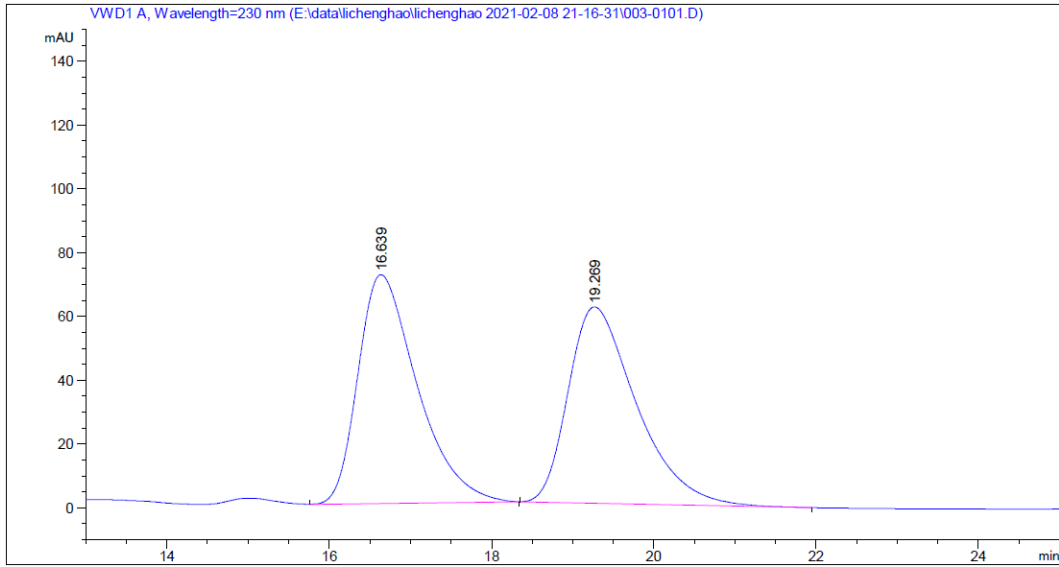
总量 : 1.35279e4 136.27275



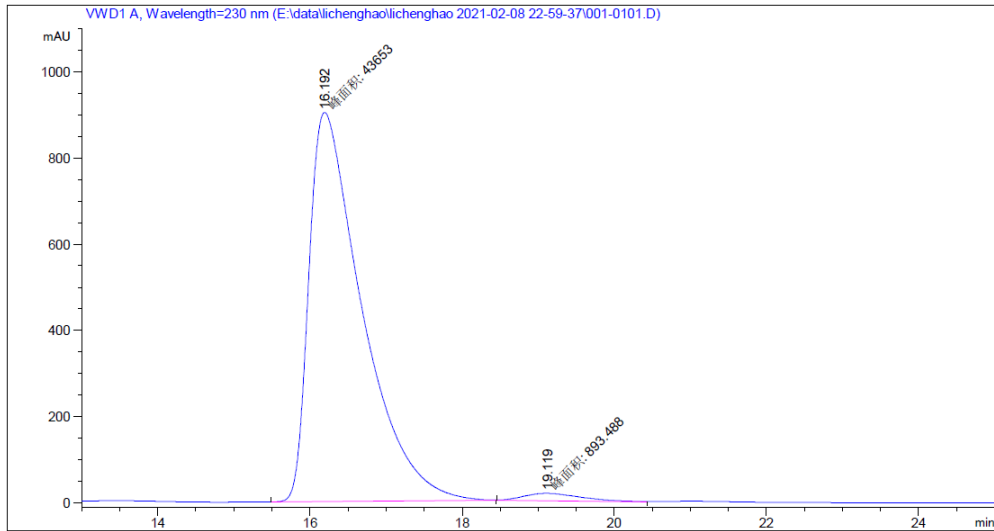
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.333	MM	1.1643	1.62537e4	232.66092	97.8706
2	26.753	BB	1.6851	353.63116	2.46596	2.1294

总量 : 1.66073e4 235.12688

Figure S70. Copy of HPLC spectra of 2o



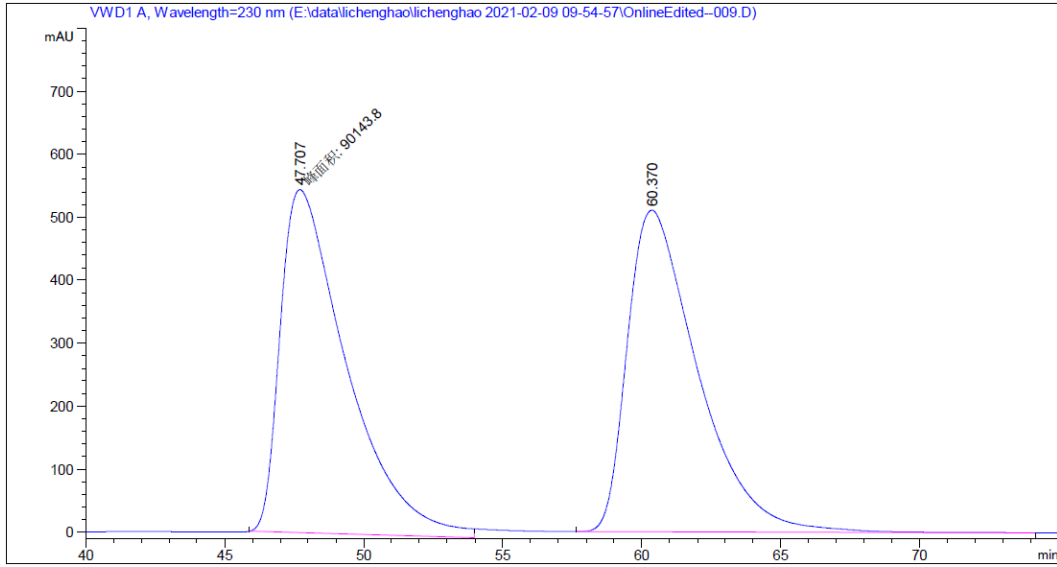
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	16.639	BB	0.7678	3633.31470	71.74956	50.2501
2	19.269	BB	0.8852	3597.14575	61.61126	49.7499



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	16.192	MM	0.8057	4.36530e4	902.95105	97.9943
2	19.119	MM	0.8744	893.48846	17.03008	2.0057

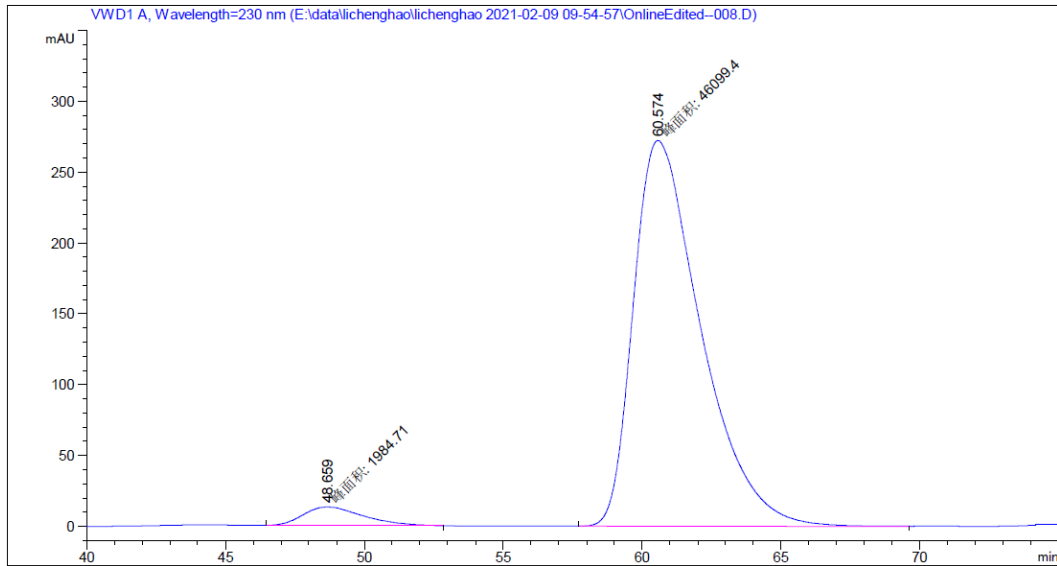
总量 : 4.45465e4 919.98113

Figure S71. Copy of HPLC spectra of 2p



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	47.707	MM	2.7590	9.01438e4	544.54565	50.0476
2	60.370	BB	2.4921	8.99722e4	511.00781	49.9524

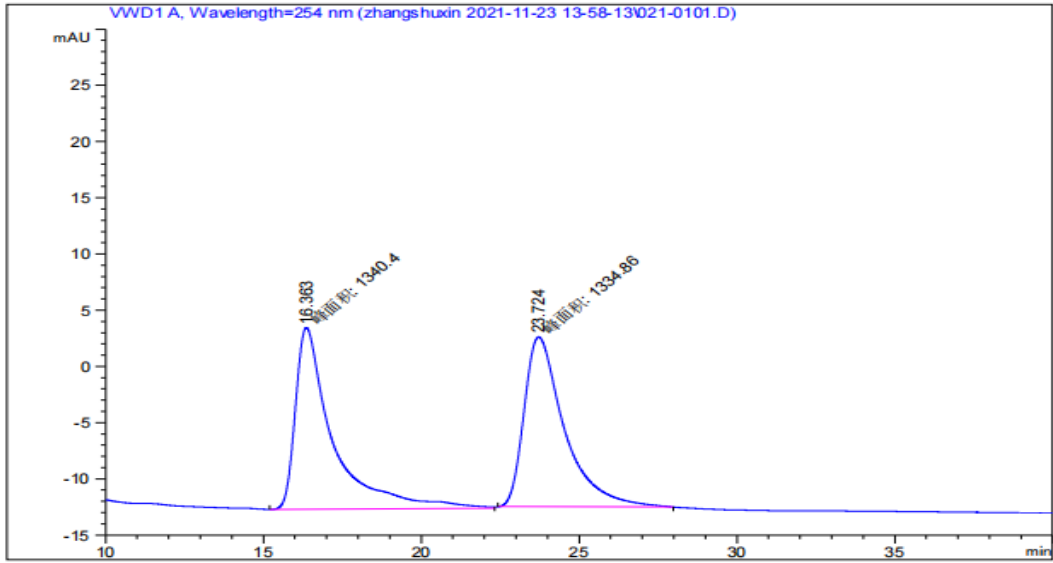
总量 : 1.80116e5 1055.55347



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	48.659	MM	2.5096	1984.71082	13.18088	4.1276
2	60.574	MM	2.8209	4.60994e4	272.36975	95.8724

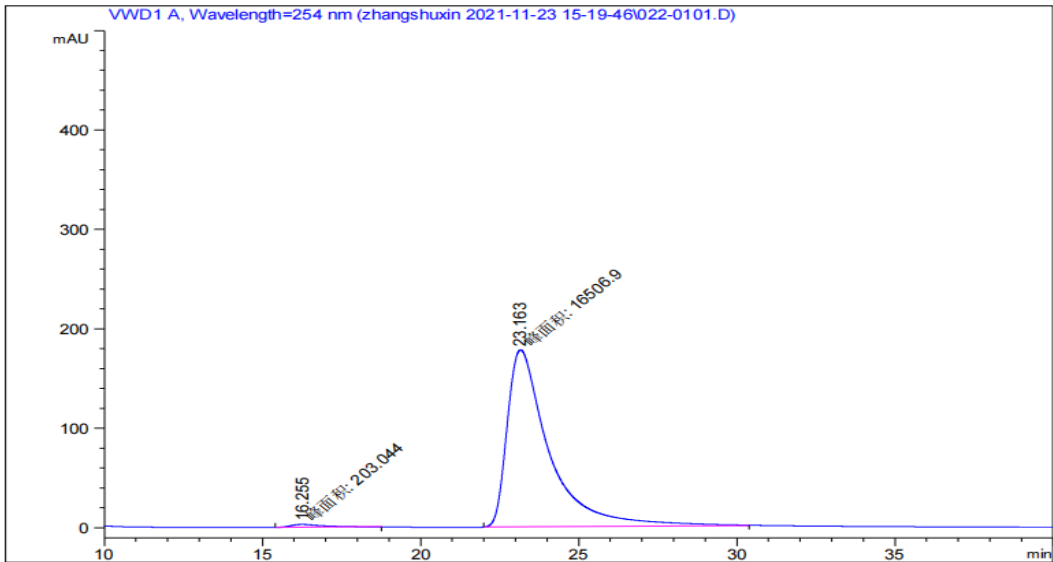
总量 : 4.80841e4 285.55063

Figure S72. Copy of HPLC spectra of 2q



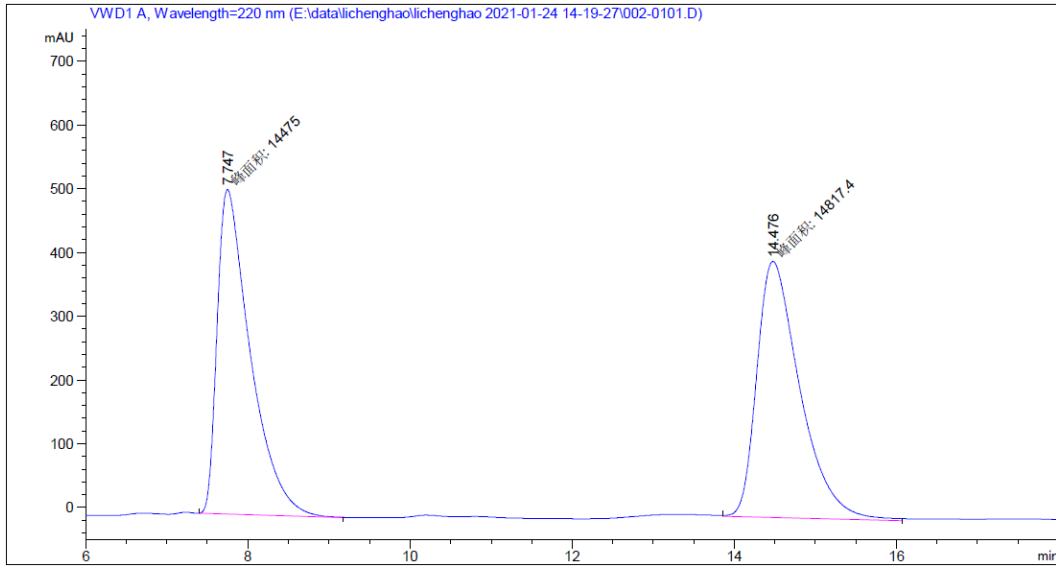
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	16.363	MM	1.3820	1340.39758	16.16502	50.1035
2	23.724	MM	1.4746	1334.86047	15.08761	49.8965

总量 : 2675.25806 31.25263



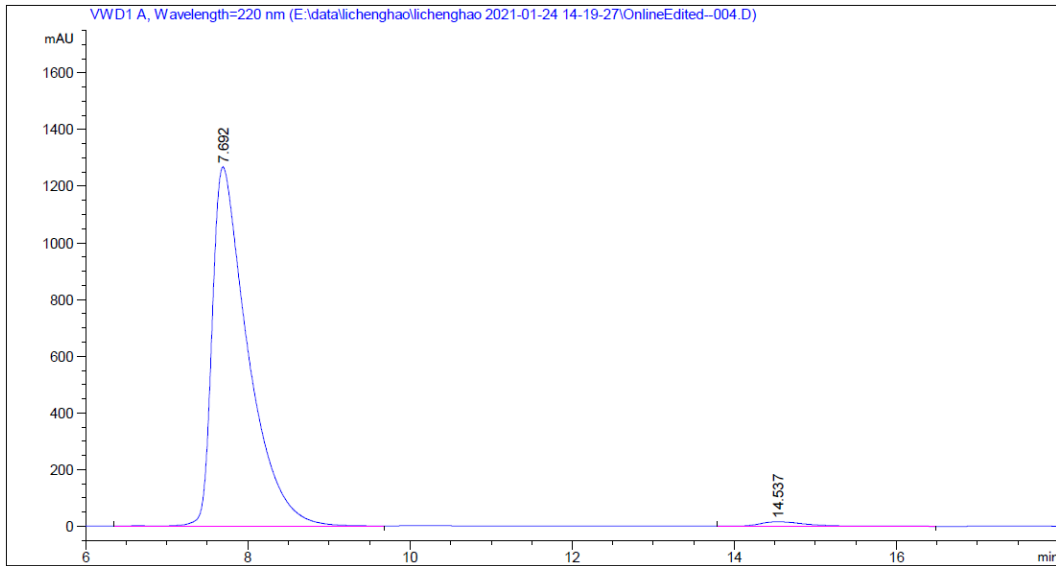
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	16.255	MM	1.1742	203.04422	2.88195	1.2151
2	23.163	MM	1.5445	1.65069e4	178.12419	98.7849

Figure S73. Copy of HPLC spectra of 2r



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.747	MM	0.4739	1.44750e4	509.12399	49.4155
2	14.476	MM	0.6151	1.48174e4	401.51736	50.5845

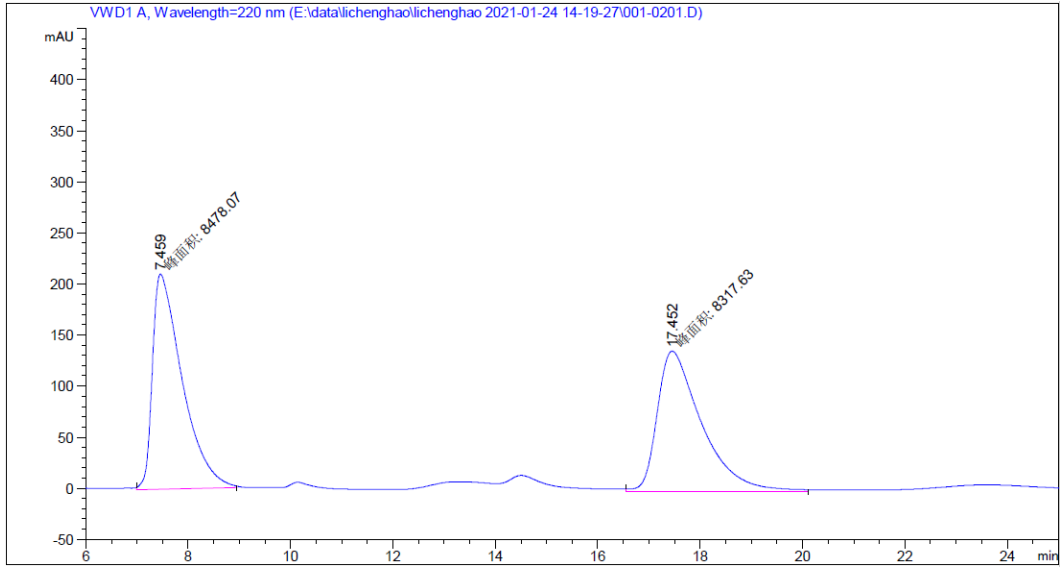
总量 : 2.92924e4 910.64136



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.692	VB R	0.4447	3.87396e4	1267.04321	98.4327
2	14.537	BB	0.5716	616.83783	16.29574	1.5673

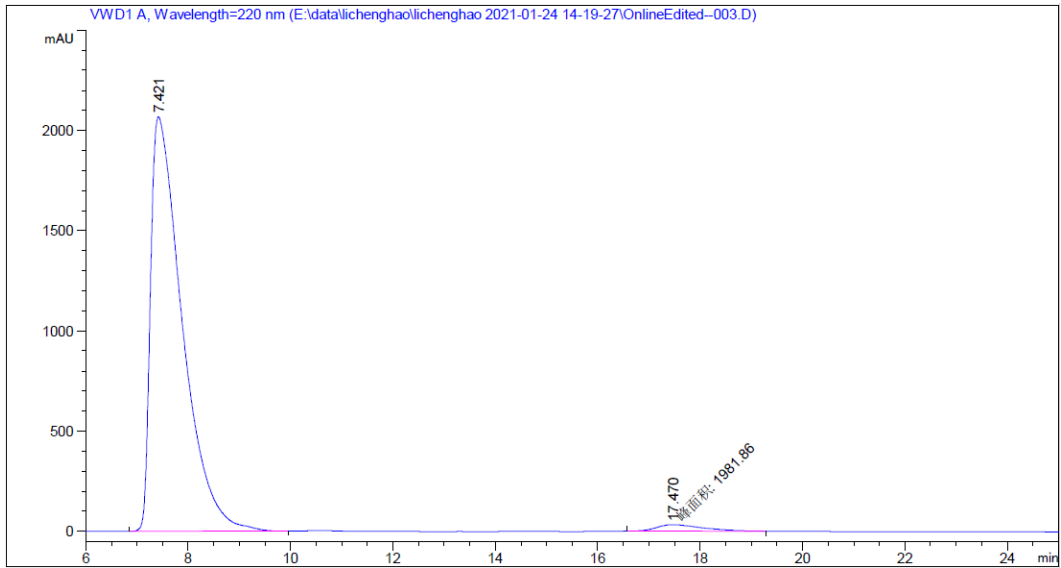
总量 : 3.93565e4 1283.33895

Figure S74. Copy of HPLC spectra of 2s



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.459	MM	0.6711	8478.07031	210.56204	50.4776
2	17.452	MM	1.0084	8317.62793	137.47853	49.5224

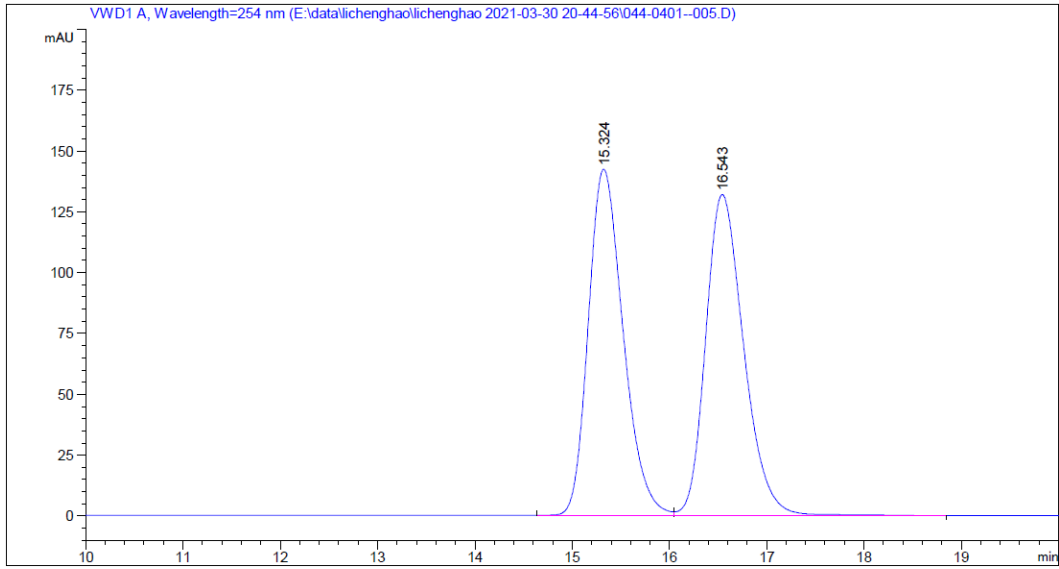
总量 : 1.67957e4 348.04057



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.421	BB	0.6482	8.85527e4	2069.71875	97.8109
2	17.470	MM	1.0188	1981.86365	32.42062	2.1891

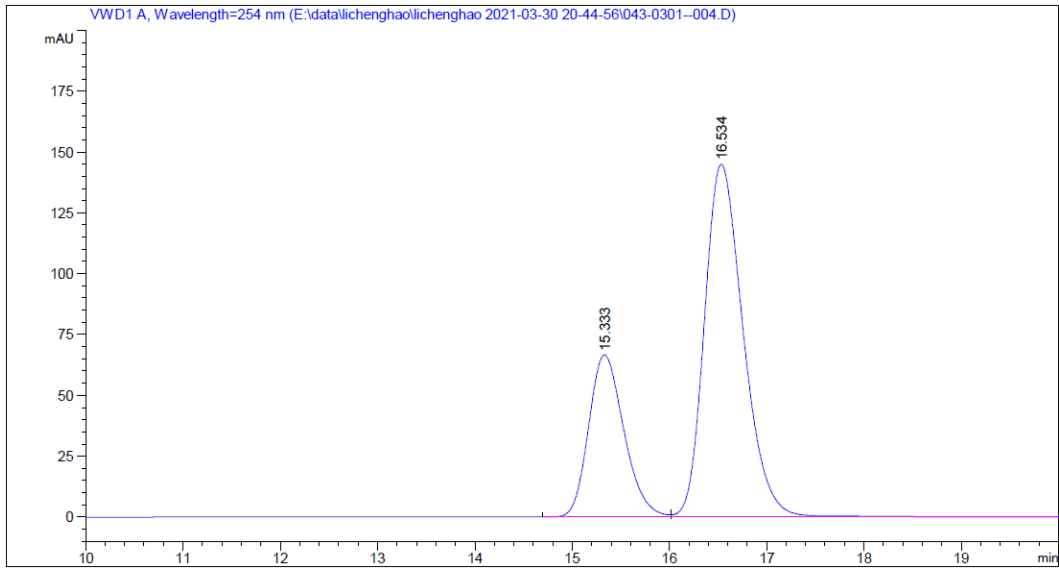
总量 : 9.05346e4 2102.13937

Figure S75. Copy of HPLC spectra of 4a



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.324	BV	0.3837	3531.19019	142.21953	49.7363
2	16.543	VB	0.4178	3568.62793	131.84190	50.2637

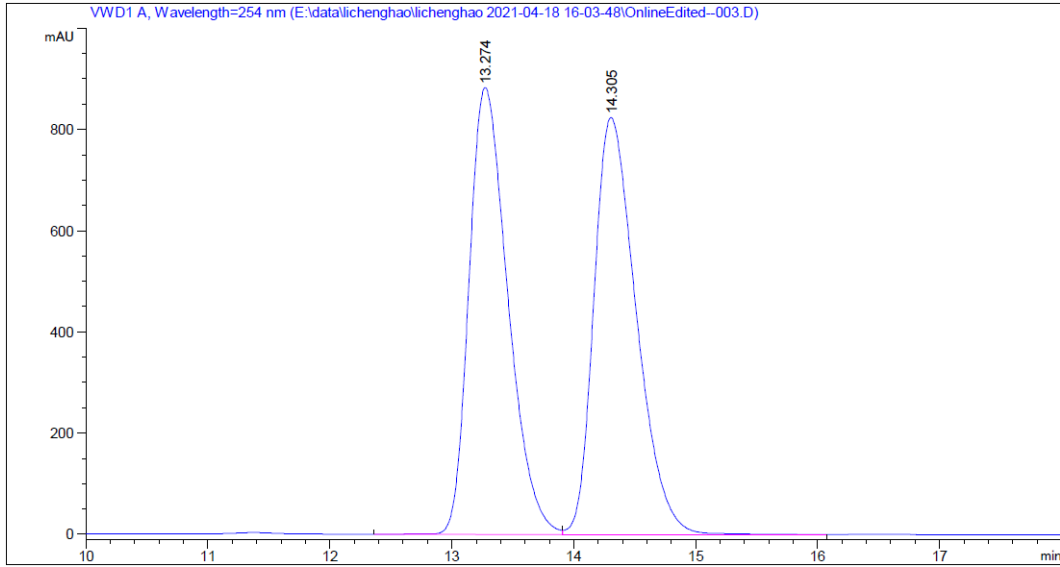
总量 : 7099.81812 274.06143



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.333	BV	0.3896	1676.02954	66.59358	29.4559
2	16.534	VBA	0.4270	4013.92847	144.99921	70.5441

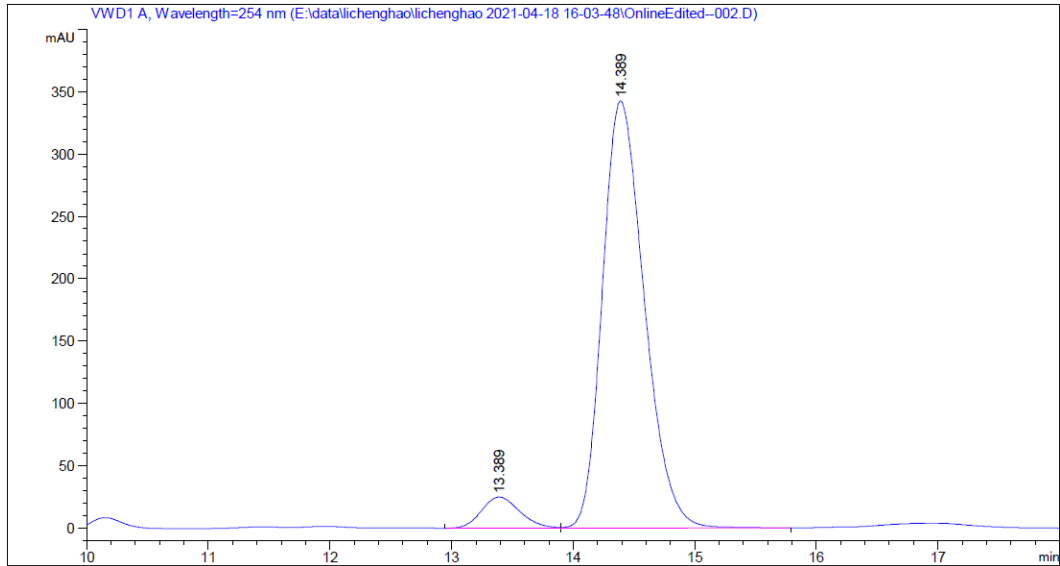
总量 : 5689.95801 211.59279

Figure S76. Copy of HPLC spectra of 4b



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	13.274	VV R	0.3420	1.95312e4	883.24719	49.5563
2	14.305	VB	0.3735	1.98810e4	824.10339	50.4437

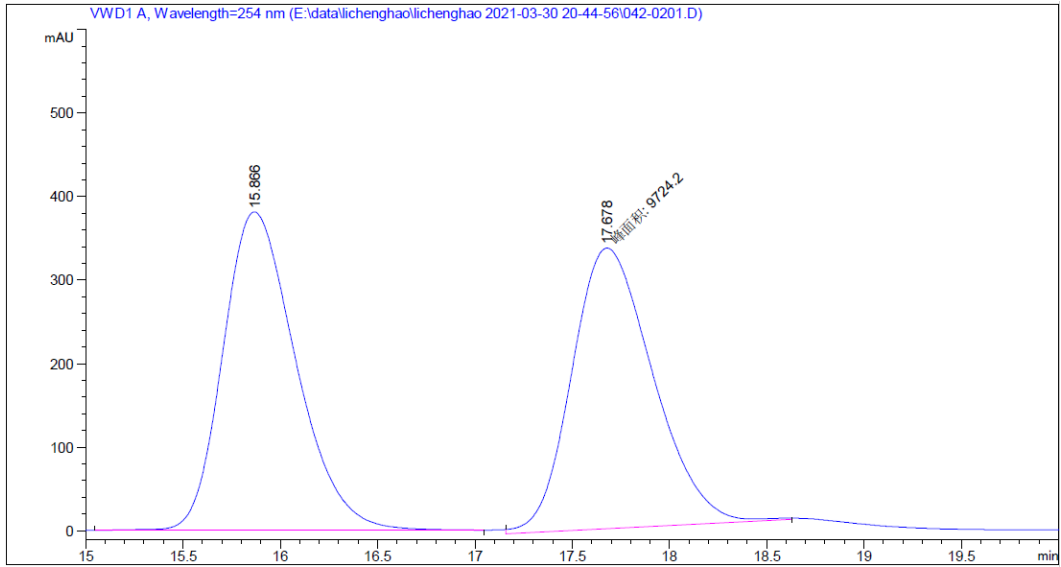
总量 : 3.94121e4 1707.35059



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	13.389	BV	0.3404	545.08173	24.89834	6.2406
2	14.389	VB	0.3709	8189.32959	342.58258	93.7594

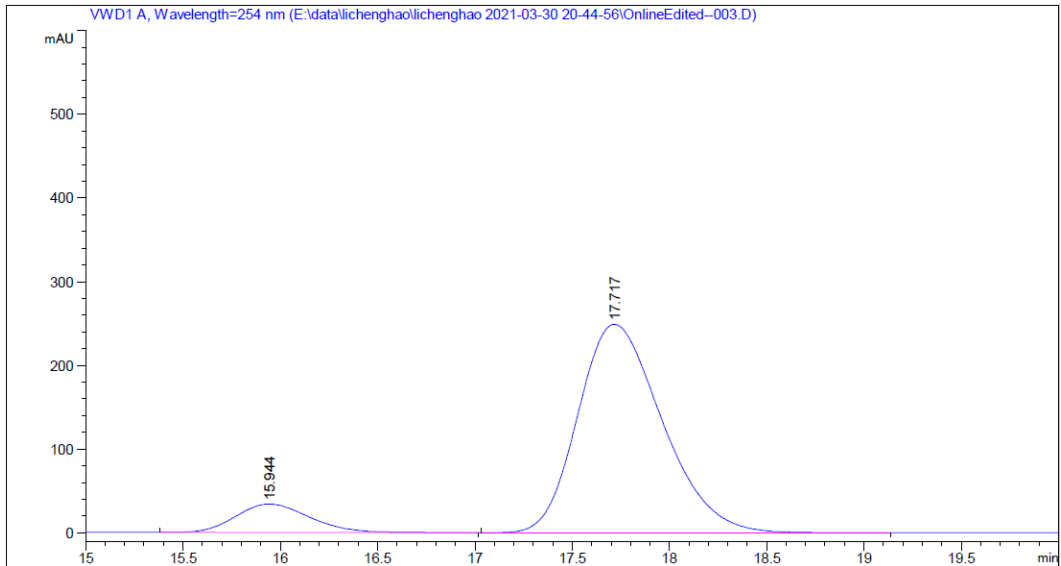
总量 : 8734.41132 367.48092

Figure S77. Copy of HPLC spectra of 4c



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.866	BB	0.4032	9924.75000	380.65872	50.5103
2	17.678	MM	0.4831	9724.20410	335.44534	49.4897

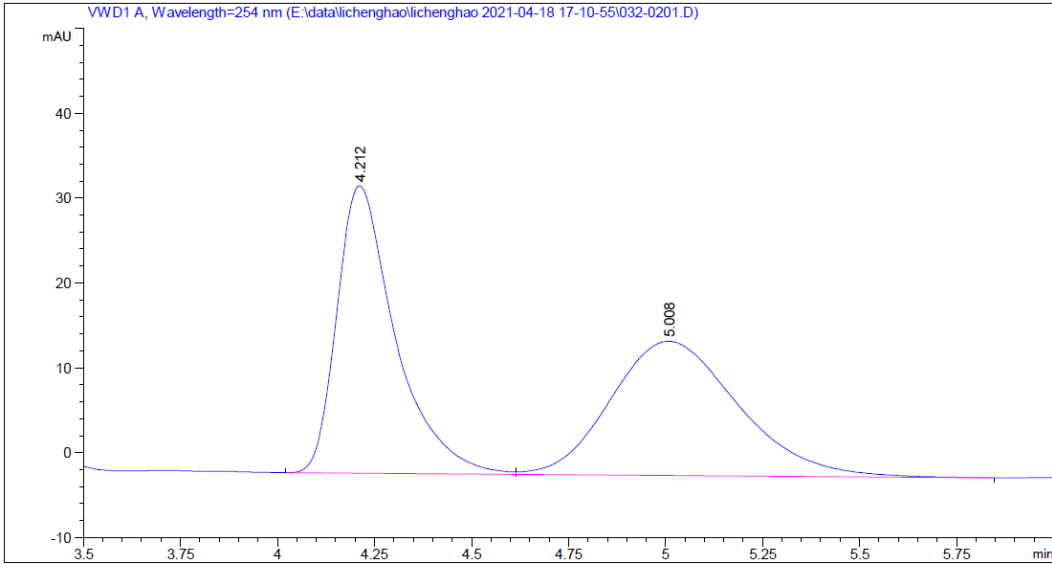
总量 : 1.96490e4 716.10406



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	15.944	BB	0.4128	907.71216	34.07307	10.7943
2	17.717	BB	0.4676	7501.45605	248.93977	89.2057

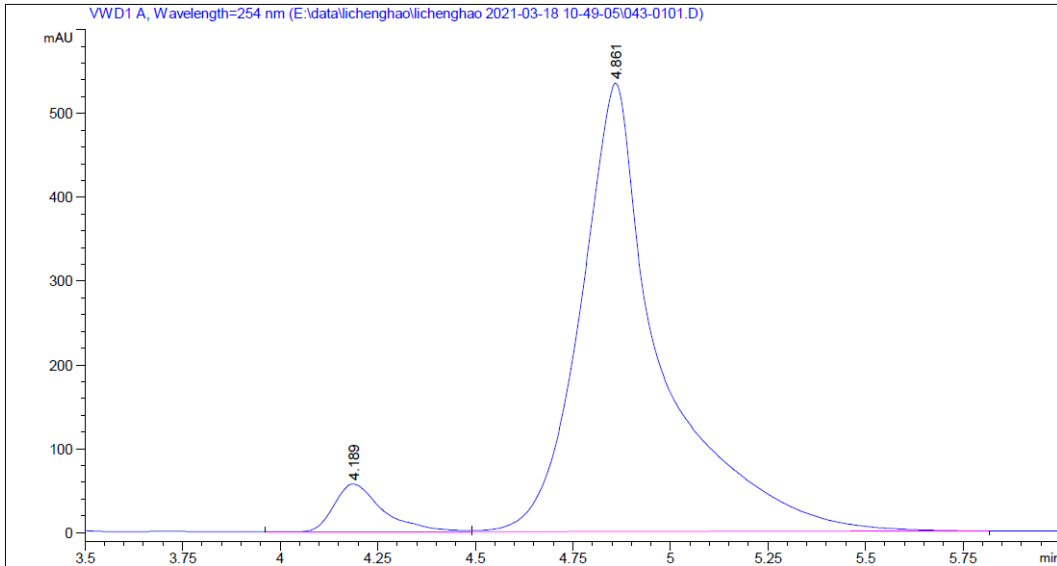
总量 : 8409.16821 283.01284

Figure S78. Copy of HPLC spectra of 4d



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	4.212	BV	0.1571	354.45859	33.87715	49.5439
2	5.008	VB	0.3535	360.98492	15.80097	50.4561

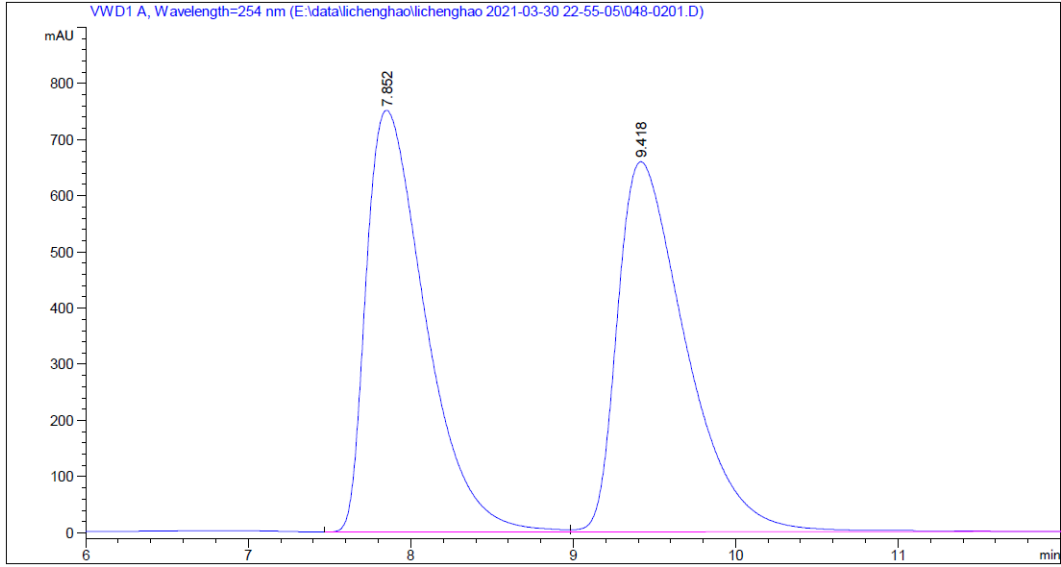
总量 : 715.44351 49.67812



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	4.189	BV	0.1298	496.66385	56.92661	6.0831
2	4.861	VB	0.1965	7667.97656	534.12988	93.9169

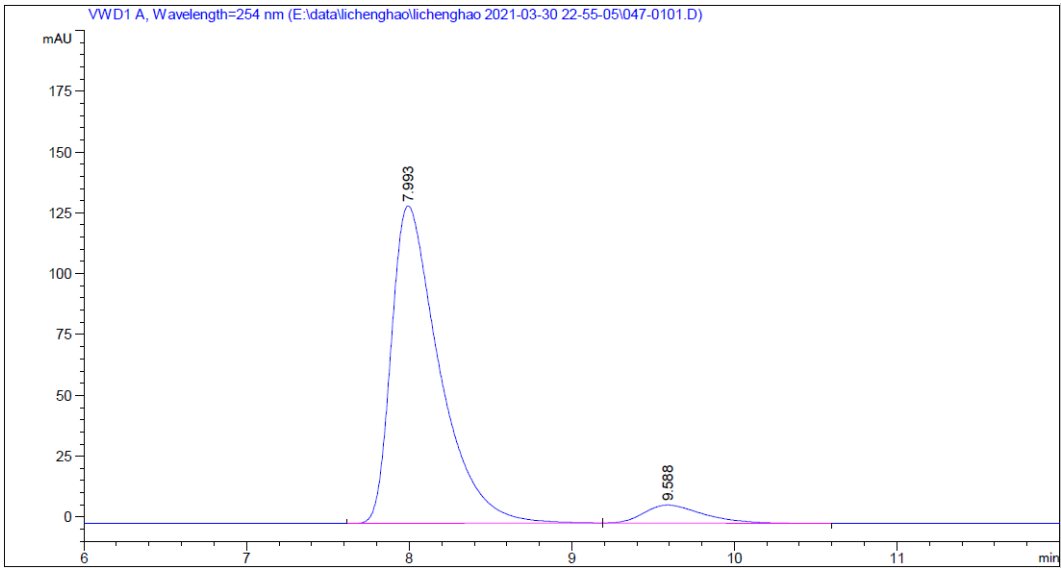
总量 : 8164.64041 591.05649

Figure S79. Copy of HPLC spectra of 4e



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.852	BV	0.3895	1.89609e4	751.26050	49.5380
2	9.418	VB	0.4448	1.93145e4	659.29108	50.4620

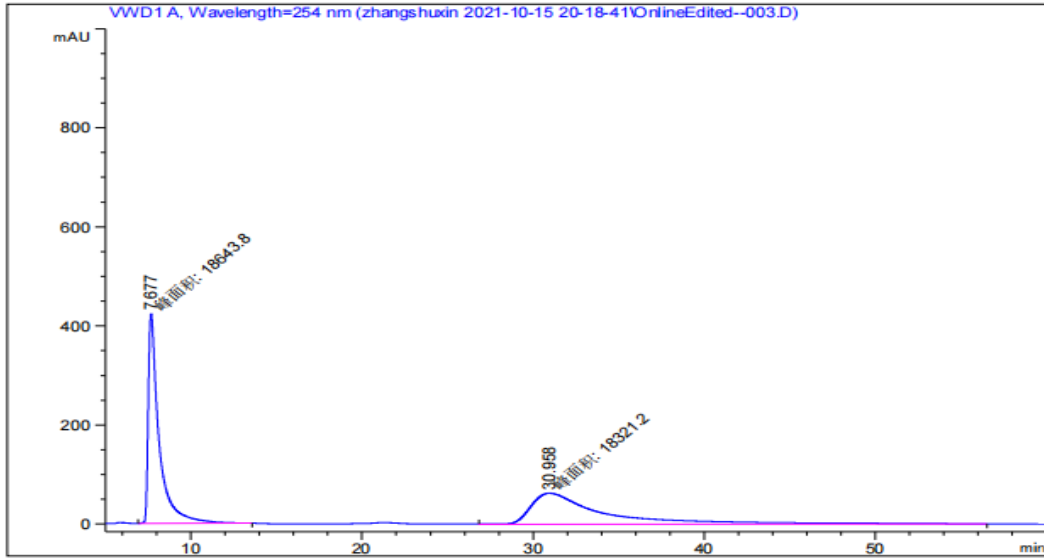
总量 : 3.82754e4 1410.55157



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.993	BB	0.3048	2673.29053	130.58397	93.4889
2	9.588	BB	0.3836	186.18167	7.37292	6.5111

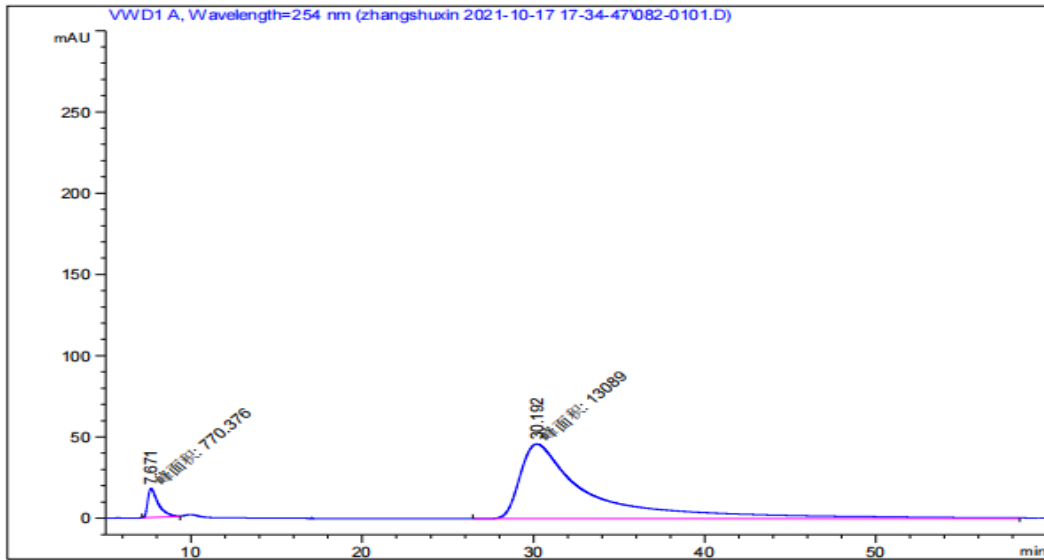
总量 : 2859.47220 137.95688

Figure S80. Copy of HPLC spectra of 4f



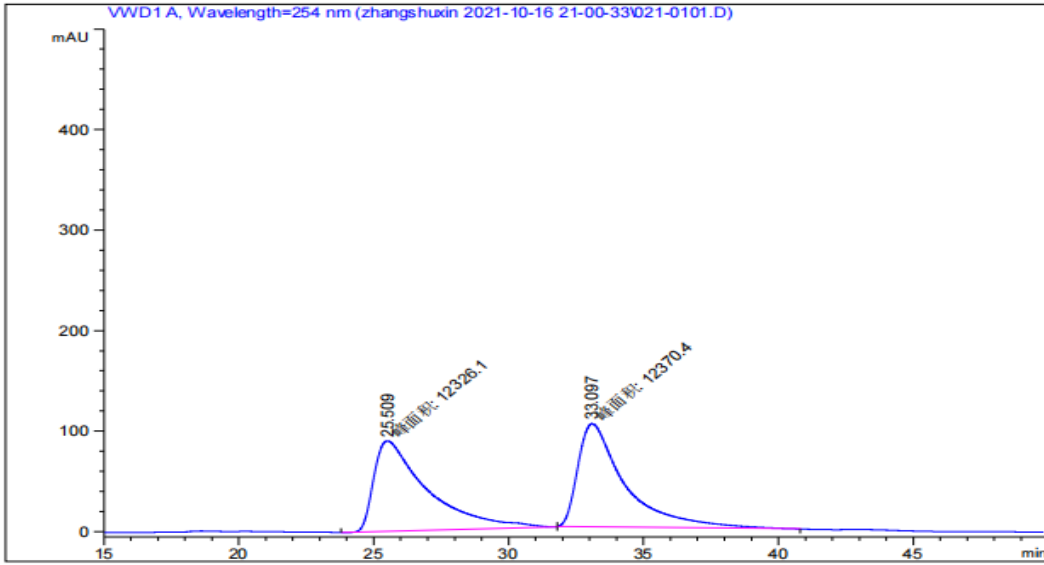
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.677	MM	0.7351	1.86438e4	422.72028	50.4364
2	30.958	MM	4.8984	1.83212e4	62.33804	49.5636

总量 : 3.69651e4 485.05831



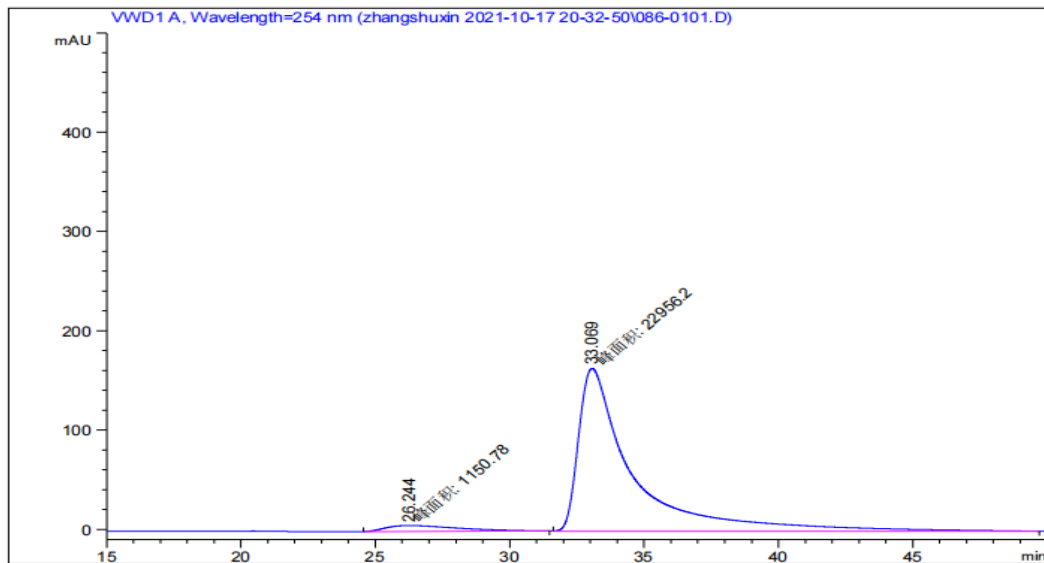
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.671	MM	0.7237	770.37561	17.74267	5.5585
2	30.192	MM	4.7499	1.30890e4	45.92723	94.4415

Figure S81. Copy of HPLC spectra of 4g



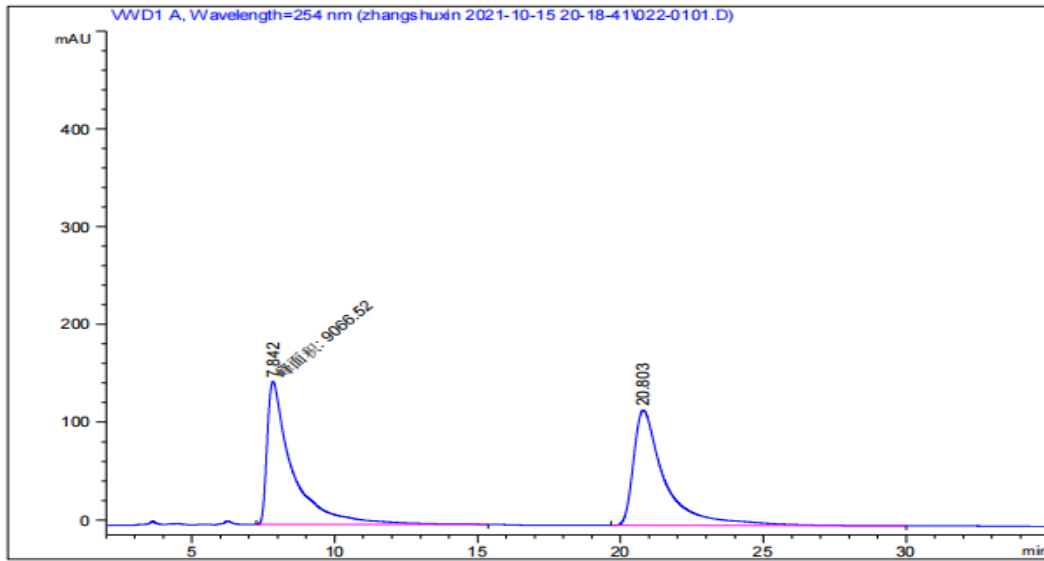
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	25.509	MM	2.2859	1.23261e4	89.86923	49.9103
2	33.097	MM	2.0124	1.23704e4	102.45327	50.0897

总量 : 2.46965e4 192.32250

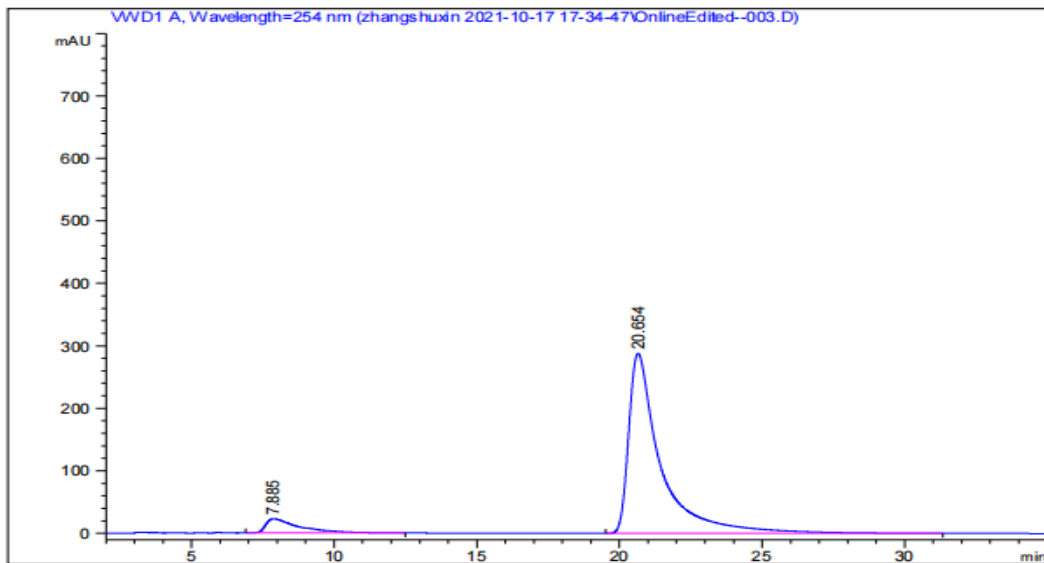


峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	26.244	MM	3.1600	1150.78040	6.06959	4.7736
2	33.069	MM	2.3370	2.29562e4	163.71204	95.2264

Figure S82. Copy of HPLC spectra of 4h



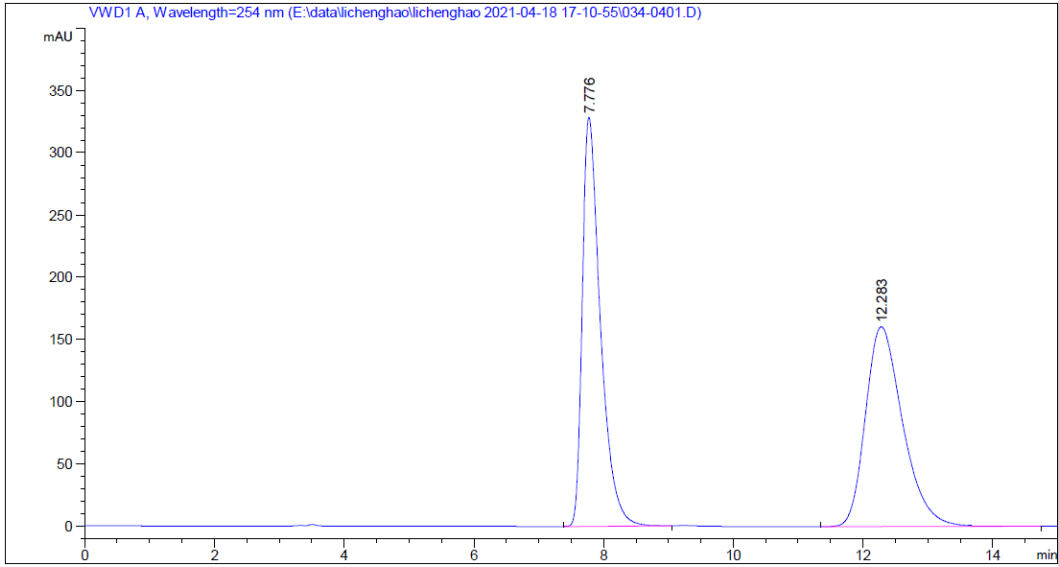
峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.842	MM	1.0341	9066.51563	146.12718	50.0503
2	20.803	BB	1.0952	9048.27832	117.91988	49.9497



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.885	BB	1.1402	1850.11279	22.64442	7.6067
2	20.654	BB	1.1053	2.24720e4	287.99750	92.3933

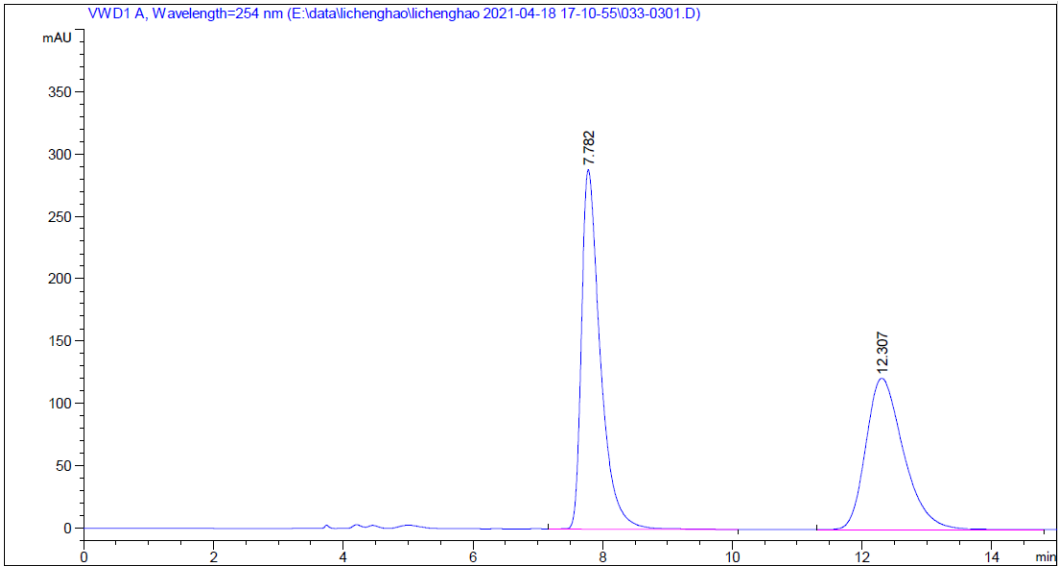
总量 : 2.43221e4 310.64191

Figure S83. Copy of HPLC spectra of 4i



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.776	BB	0.2893	6379.14795	328.89249	49.8845
2	12.283	BB	0.6096	6408.68457	160.57660	50.1155

总量 : 1.27878e4 489.46909



峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	7.782	BB	0.2984	5795.67773	288.47263	54.2645
2	12.307	BB	0.6154	4884.74316	121.41721	45.7355

总量 : 1.06804e4 409.88984