

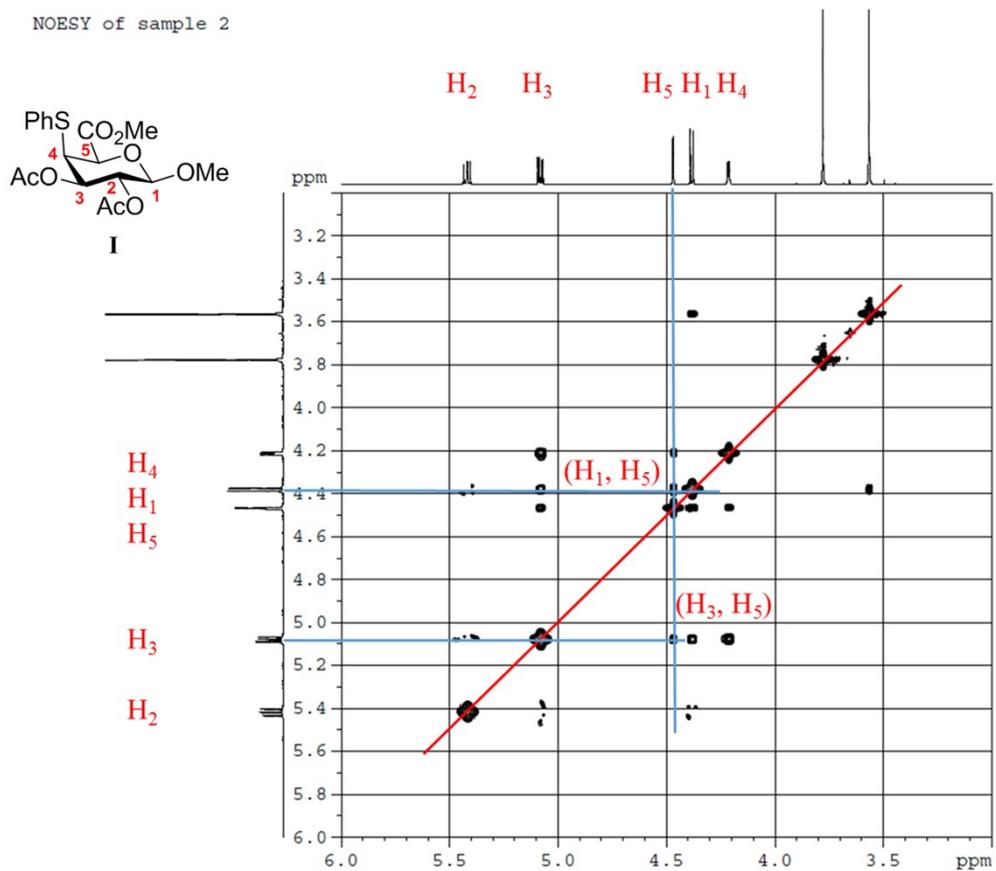
Supporting Information

Synthesis of 4-thiol-furanosidic uronate *via* hydrothiolation reaction

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New Taipei City 24205, Taiwan

(A)



(B)

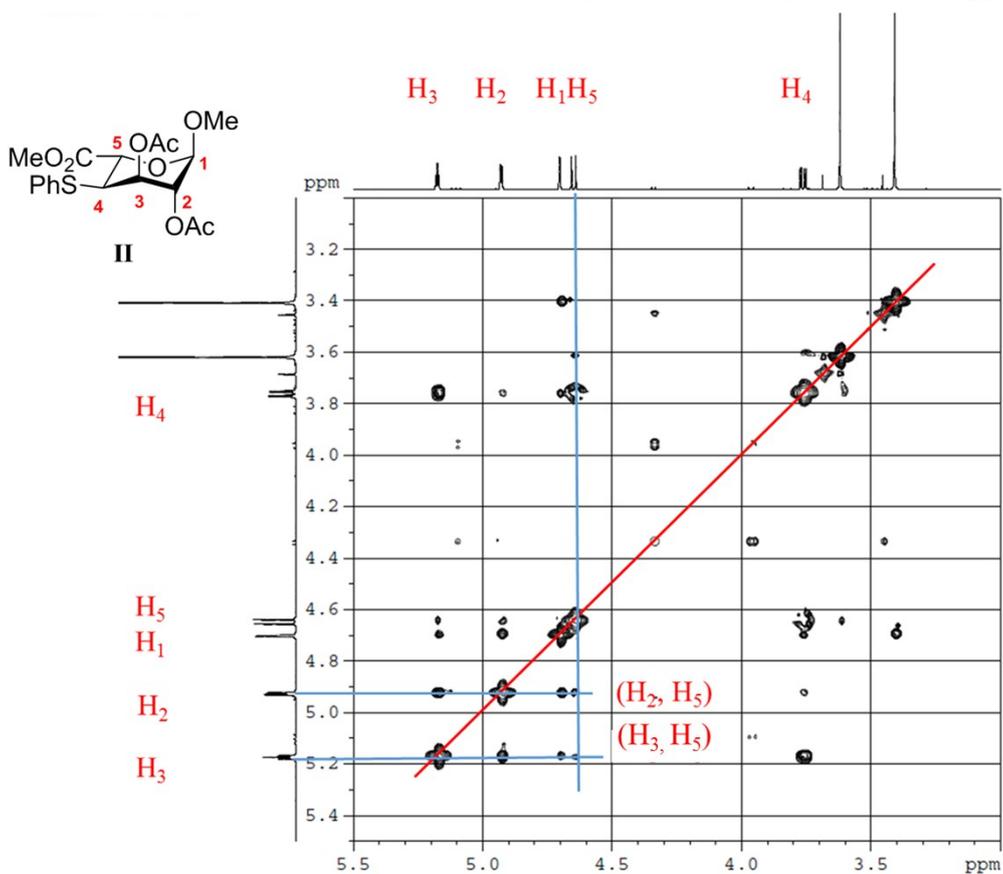


Figure S1. The NOESY spectra of compound I (A) and II (B).

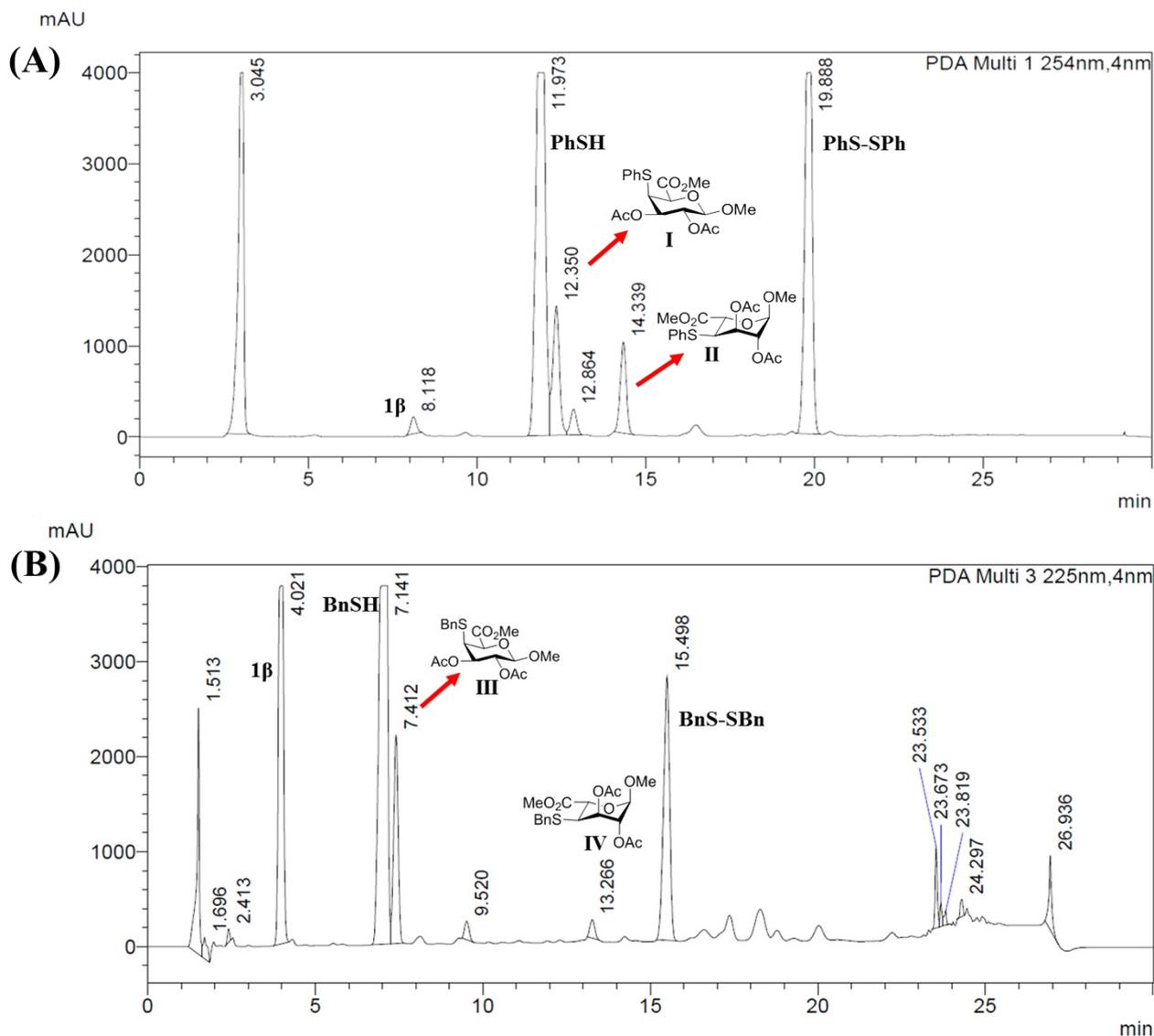
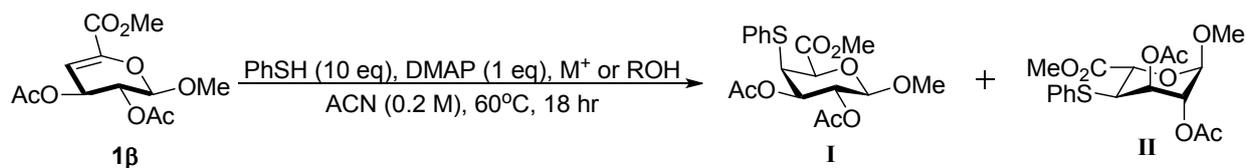


Figure S2. The ratio of desired products was determined by HPLC analysis. Using PhS (A) and BnSH (B), respectively, as nucleophiles for base-mediated thiol-Michael addition, the reaction crude mixture was diluted with acetonitrile and that was immediately analyzed by HPLC.

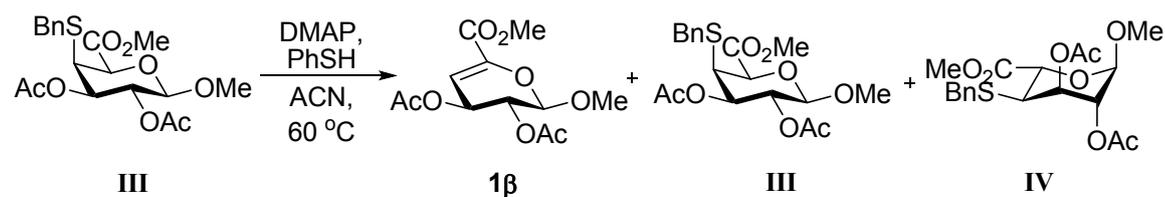
Table S1. **1 β** with metallic salts and alcohols containing bulky alkyl group.



Entry	Additives (equiv.)	Yield ^[a]	I:II ^[a]
1	none	72 %	1.6:1
2	MgBr ₂ .OEt ₂ (0.1 eq)	73 %	1.1:1
3	Mg(ClO ₄) ₂ (0.1 eq)	78 %	1.1:1
4	Mg(OTf) ₂ (0.1 eq)	76 %	1.1:1
5	NaBr (0.1 eq)	73 %	1.2:1
6	ZnCl ₂ (0.1 eq)	65 %	1.3:1
7	IPA (1 eq)	75 %	1.3:1
8	t-BuOH (1 eq)	76 %	1.3:1

[a] yield percentage and the ratio of products were determined by high performance liquid chromatography (HPLC) analysis.

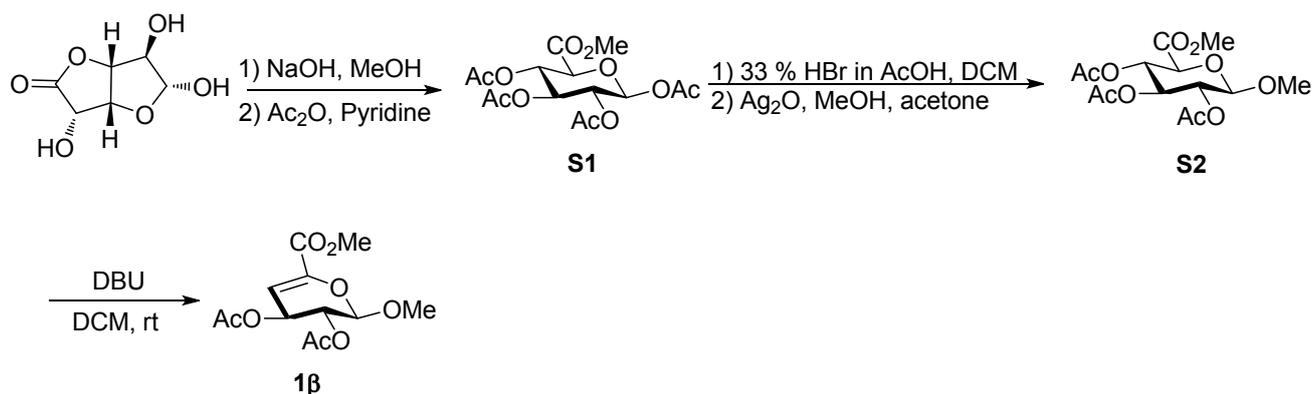
Table S2. Retro thiol-Michael addition.



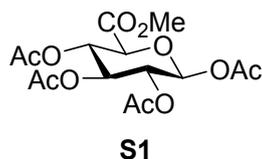
Entry	Thiol (Eq.)	Time	Result ^[a]
1	-	1 hr	III (100)
2	-	3 hr	III (100)
3	-	12 hr	1 β + III + IV (1 : 97 : 2)
4	BnSH (10)	1 hr	III (100)
5	BnSH (10)	3 hr	1 β + III (1: 99)
6	BnSH (10)	12 hr	1 β + III + IV (2 : 96 : 2)

[a] The ratio of products was determined by HPLC analysis.

Synthesis of compound 1 β :



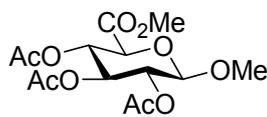
Scheme S1. Synthesis of methyl (methyl 2,3-di-O-acetyl-4-deoxy- α -L-threo-hex-4-enopyranosid)uronate (1 β).



β -D-Glucopyranuronic acid, methyl ester, 1,2,3,4-tetraacetate (S1)

To a solution of D-Glucurono-6,3-lactone (10.0 g, 56.8 mmol) in MeOH (30.0 mL) was added NaOH/MeOH (0.01 wt%, 90.0 mL) at 0 °C for 20 min, then the reaction mixture was allowed to warm up to room temperature. After another 16 hr stirring at room temperature, the solvent of this reaction mixture was removed under vacuum and acetic anhydride (47.2 mL) and pyridine (113.6 mL) was added to this crude residue at 0 °C. The reaction was stirred at 0 °C for overnight, then quenched in ice/water. Thereafter, the reaction mixture was diluted with EtOAc and the organic layer was washed with 1 N HCl_(aq), and brine to remove pyridine. The organic layer was dried over MgSO₄, and the organic layer was concentrated under reduced pressure. The crude product was recrystallized from methanol to give a slightly pale yellow powder (45.0 g, 0.1 mol, 52%). R_f = 0.33 (EtOAc : *n*-Hex = 1 : 3 v/v). ¹H NMR (300 MHz, CDCl₃, 298 K): δ = 5.76 (d, J = 7.8 Hz, 1H), 5.30 (dd, J_1 = J_2 = 9.2 Hz, 1H), 5.23 (dd, J_1 = J_2 = 9.3 Hz, 1H), 5.13 (dd, J_1 = J_2 = 8.3 Hz, 1H), 4.17 (d, J = 9.3 Hz, 1H), 3.74 (s, 3H), 2.11 (s, 3H), 2.03 (s, 6H), 2.02 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃, 298 K): δ = 170.03,

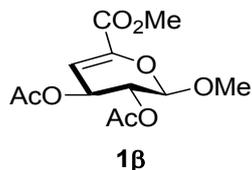
169.54, 169.31, 168.96, 166.93, 91.48, 73.11, 71.94, 70.27, 69.04, 53.15, 20.89, 20.68, 20.66, 20.59 ppm. ^1H and ^{13}C NMR spectra were agreed with reported literature^{S1}.



S2

β -D-Glucopyranosiduronic acid, methyl, methyl ester, 2,3,4-triacetate (S2)

To a solution of **S1** (2.0 g, 5.3 mmol) in dry dichloromethane (24.4 mL) was added 33% HBr in acetic acid (12.4 mL). The reaction was stirred at 0 °C for 20 min then left at 4 °C until the starting material was finished. After which, the reaction was quenched in cool sat. NaHCO_3 and the aqueous layer was extracted with dichloromethane. The combined organic layer was quickly washed with cold water, cold sat. NaHCO_3 and brine. After drying over MgSO_4 , the organic layer was concentrated *via* reduce pressure and the concentrated residue was directly used for next step without purification. To a solution of crude product in dry acetone (43.7 mL) was added Ag_2O (3.7 g, 16.0 mmol) and dry MeOH (19.6 mL, 483.3 mmole) under nitrogen. The reaction mixture was vigorously stirred at room temperature in the dark. After an overnight stirring, the reaction mixture was filtered through a plug of celite and the filtrate was concentrated under reduced pressure. The concentrated residue was diluted with EtOAc, and then it was washed with sat. NaHCO_3 , water and brine, sequentially. Thereafter, the organic layer was dried over MgSO_4 , concentrated *via* reduce pressure, and purified by flash column chromatography to yield desired product (1.4 g, 4.0 mmol, 76 %). R_f = 0.20 (EtOAc : *n*-Hex = 1 : 1.5 v/v). ^1H NMR (300 MHz, CDCl_3): δ = 5.29-5.19 (m, 2H), 5.00 (td, J_1 = 8.2 Hz, J_2 = 3.1 Hz, 1H), 4.48 (d, J = 7.7 Hz, 1H), 4.04 (dd, J_1 = 6.6 Hz, J_2 = 3.0 Hz, 1H), 3.77 (s, 3H), 3.52 (s, 3H), 2.05 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H) ppm. ^{13}C NMR (75 MHz, CDCl_3 , 298 K): δ = 170.28, 169.53, 169.47, 167.41, 101.82, 72.77, 72.23, 71.29, 69.61, 57.46, 53.05, 20.82, 20.75, 20.65 ppm. ^1H and ^{13}C NMR spectra were agreed with reported literature^{S1}.



methyl (methyl 2,3-di-O-acetyl-4-deoxy- α -L-threo-hex-4-enopyranosid)uronate (1 β)

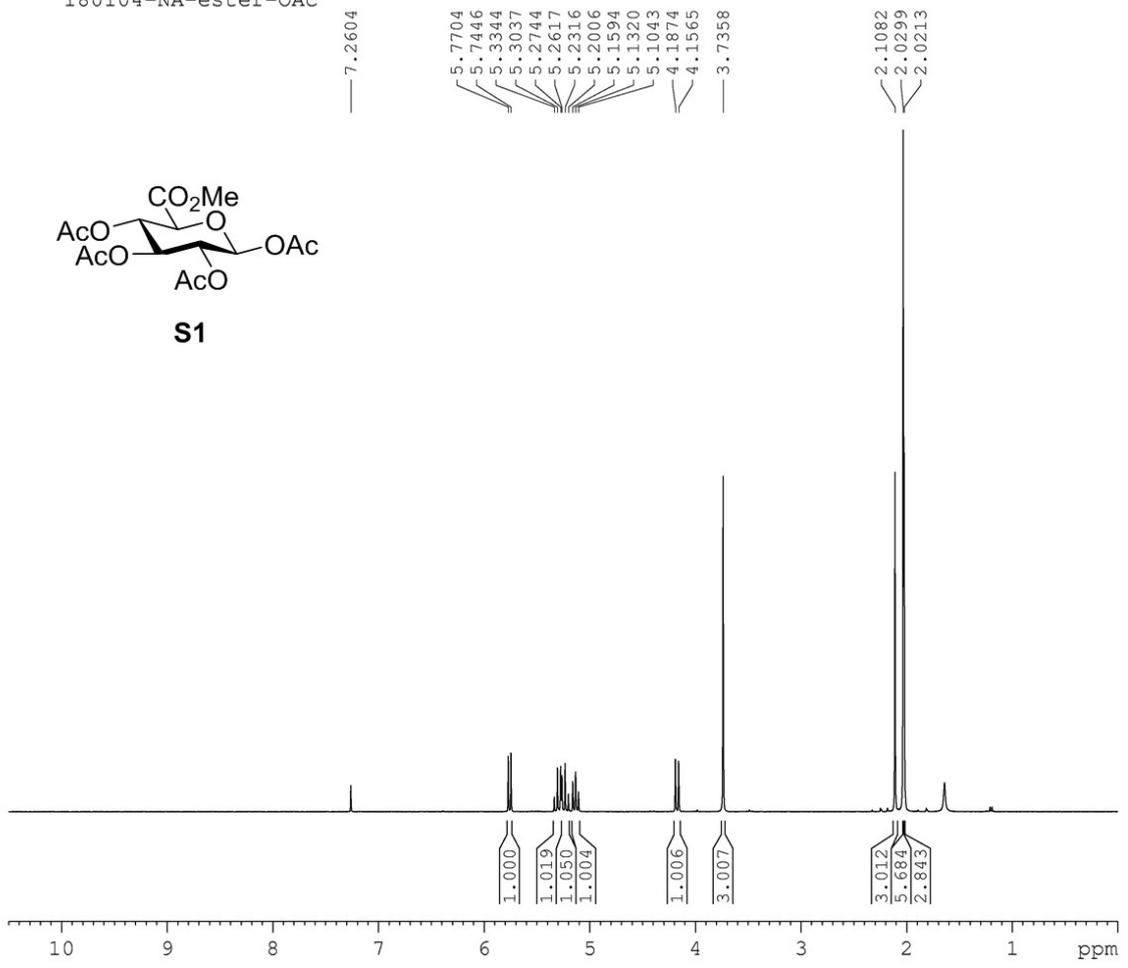
To a solution of compound **S3** (30.0 mg, 0.1 mmol) in dry dichloromethane (0.8 mL) was slowly added DBU (66.0 μ L, 0.7 mmol) under Ar_(g). After 4 hr stirring at room temperature, the reaction was quenched by sat. NH₄Cl_(aq) and diluted with dichloromethane. The organic layer was washed with water and brine, sequentially. Thereafter, the organic layer was dried over MgSO₄, concentrated under reduced pressure, then purification by flash column chromatography giving a white powder (21.6 mg, 0.07 mmol, 89 %). R_f = 0.30 (EtOAc : *n*-Hex = 1 : 2 v/v). ¹H NMR (300 MHz, CDCl₃): δ = 6.22 (dd, J_1 = 4.5 Hz, J_2 = 1.2 Hz, 1H), 5.19 (ddd, J_1 = 0.7 Hz, J_2 = 2.0 Hz, J_3 = 4.5 Hz, 1H), 5.13 (dd, J_1 = 2.8 Hz, J_2 = 0.6 Hz, 1H), 5.09-5.07 (m, 1H), 3.84 (s, 3H), 3.52 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃, 298 K): δ = 170.08, 169.50, 142.31, 107.34, 97.74, 68.74, 64.34, 57.06, 52.68, 20.99, 20.81 ppm. ¹H and ¹³C spectra were agreed with reported literature^{S1}.

References:

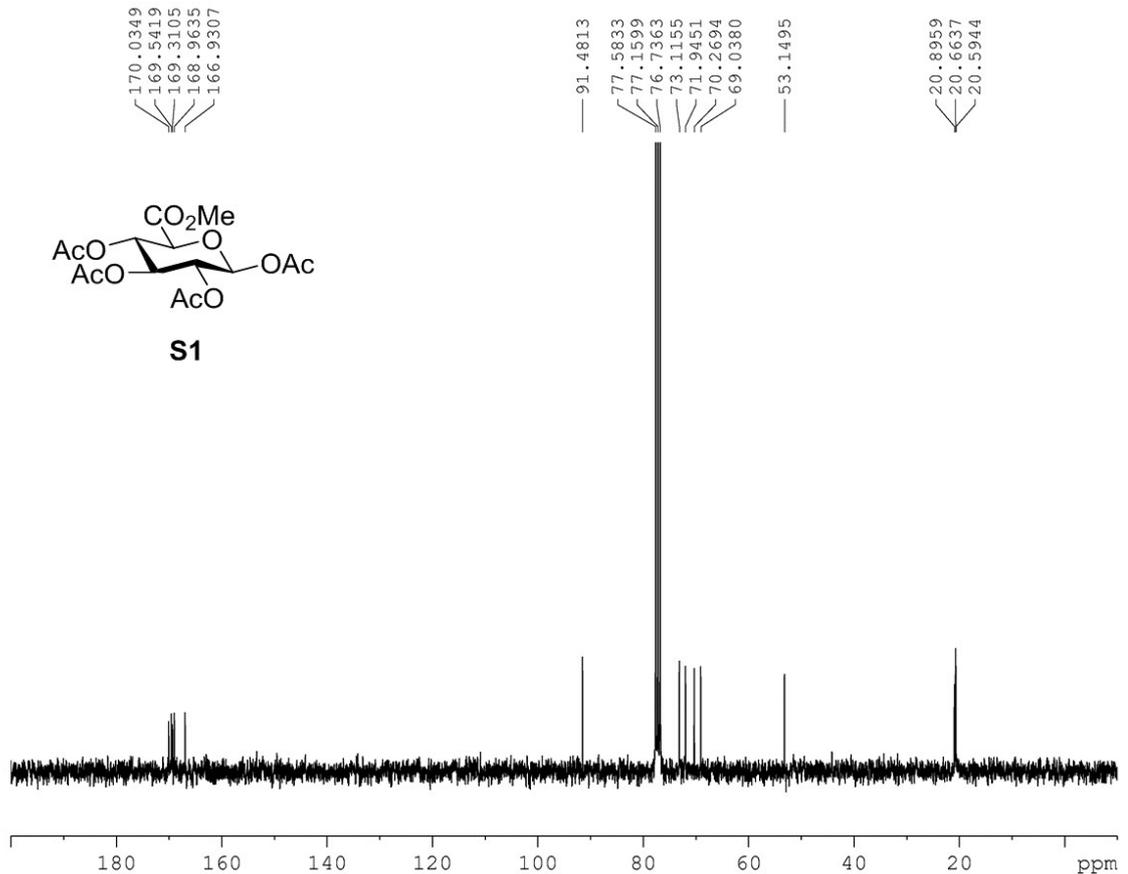
- S1. Jongkees, S. A. K.; Withers, S. G., Glycoside Cleavage by a New Mechanism in Unsaturated Glucuronyl Hydrolases. *J. Am. Chem. Soc.* **2011**, *133*, 19334-19337.

Appendix ^1H and ^{13}C NMR spectra

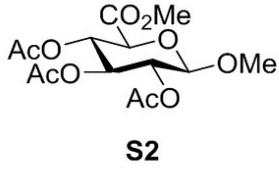
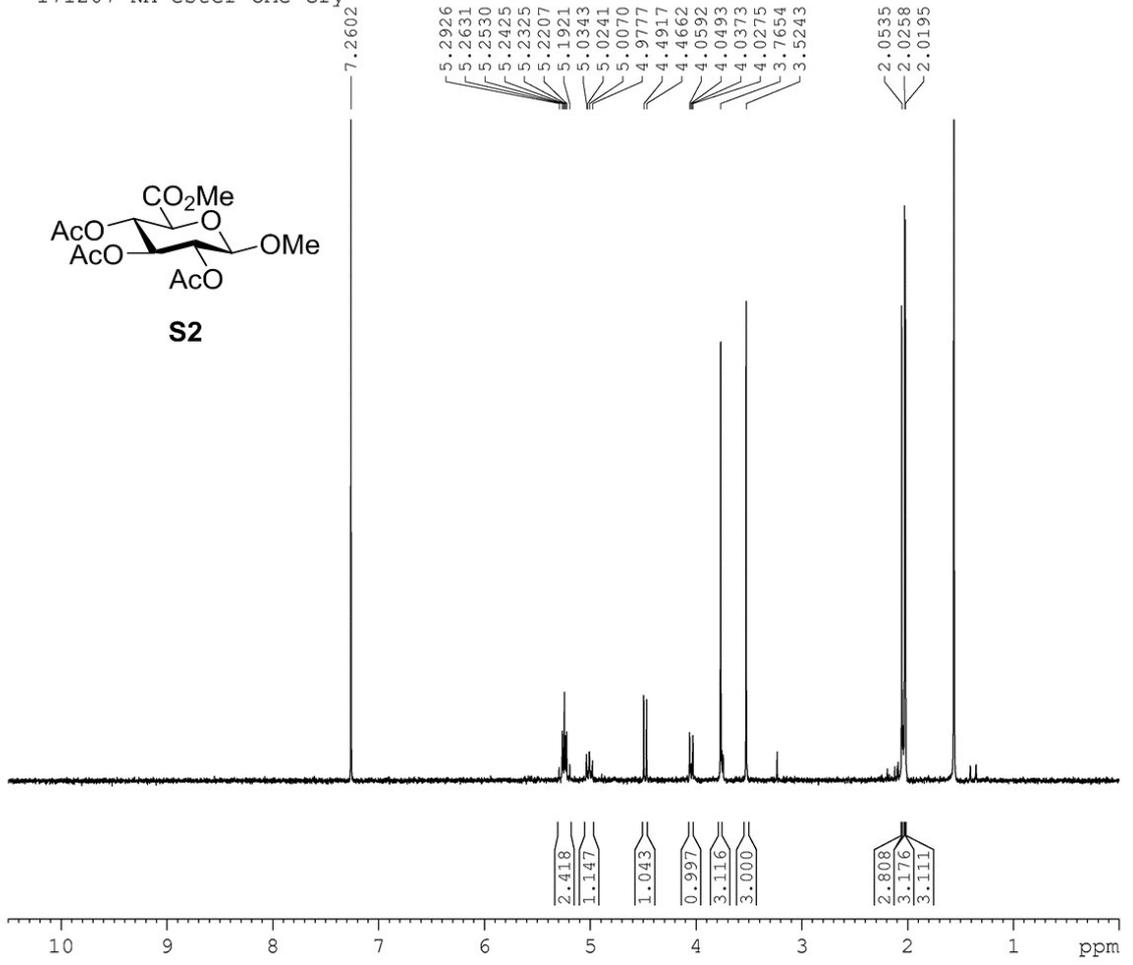
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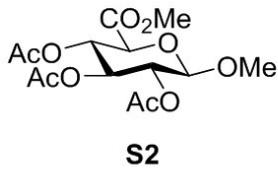
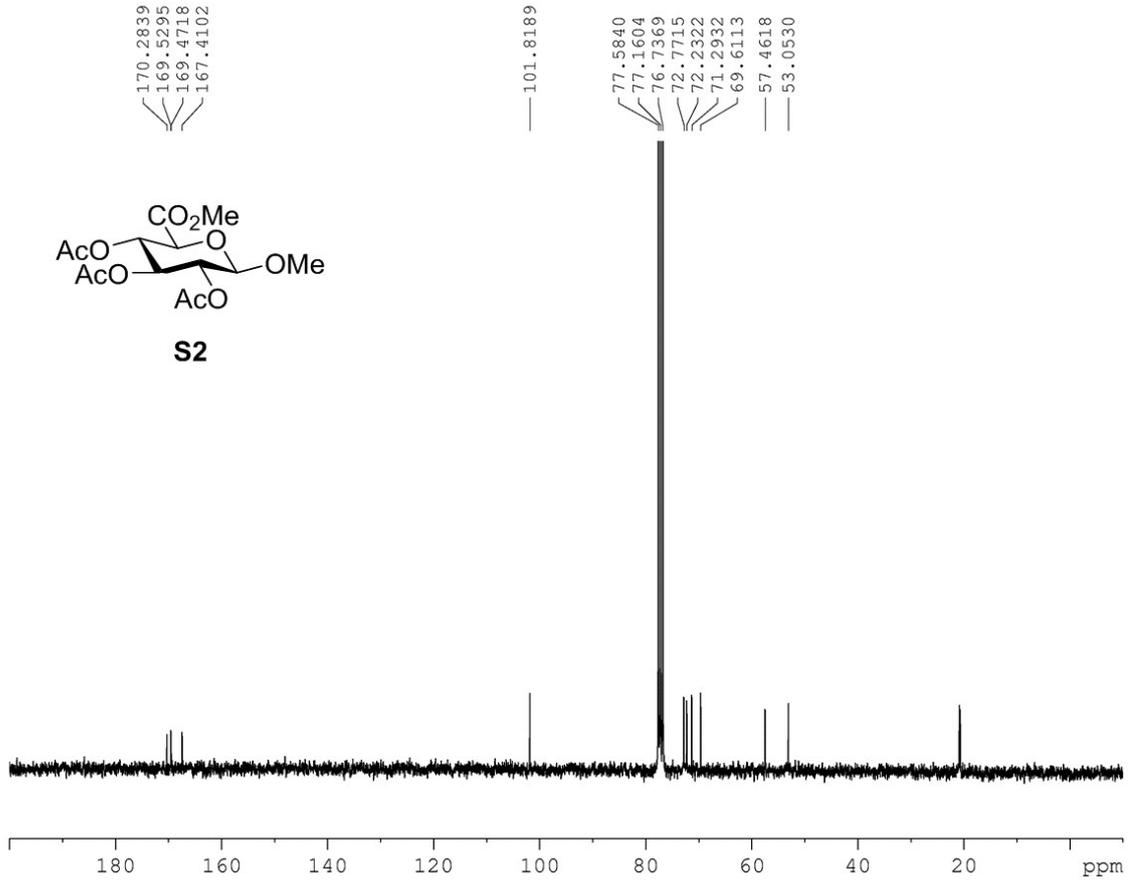
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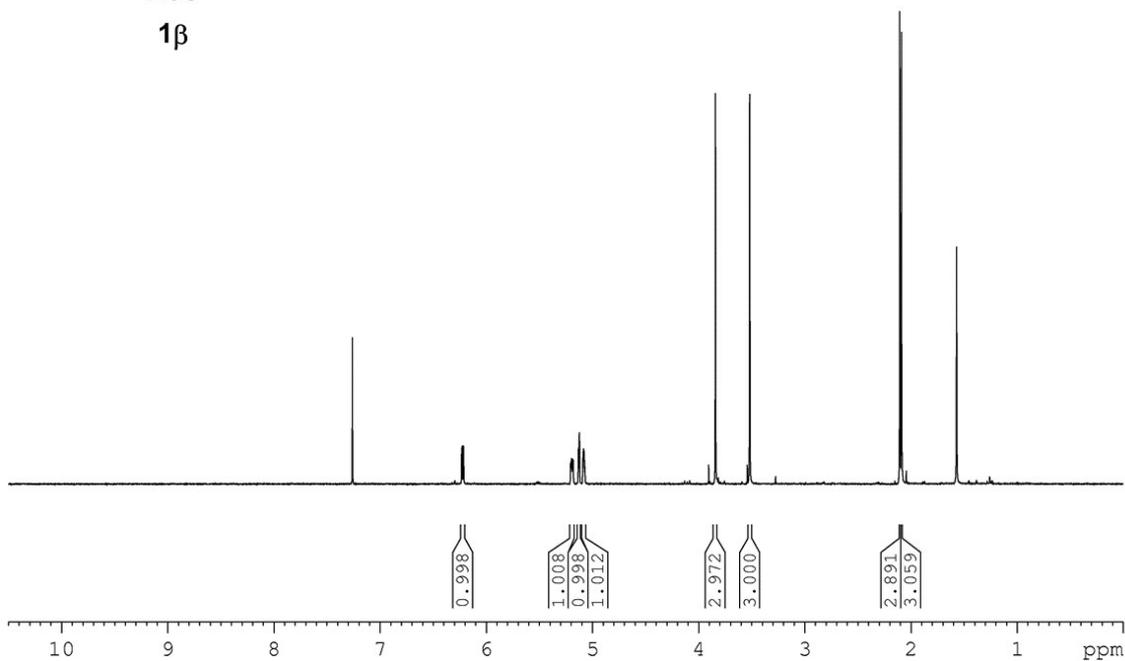
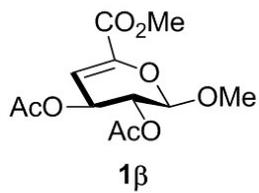


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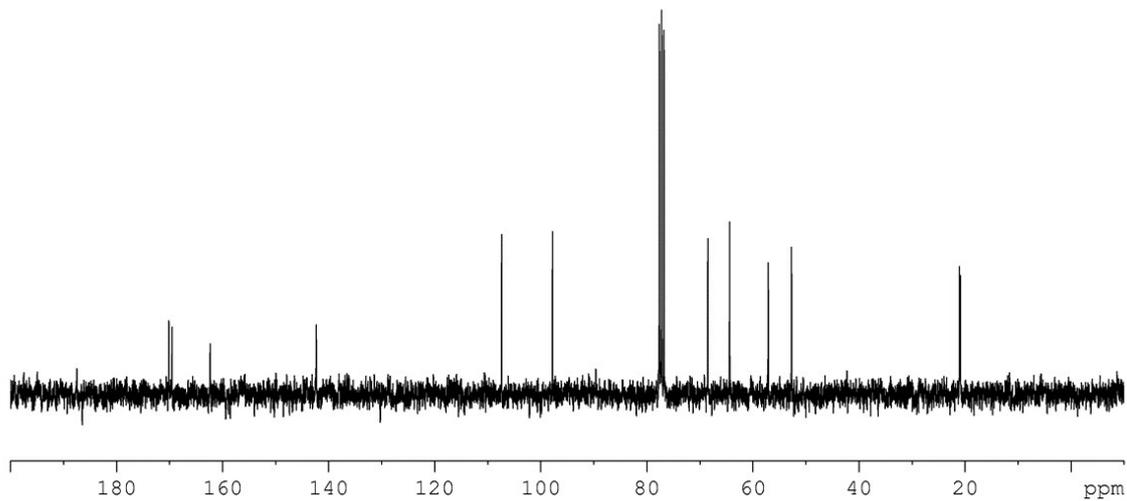
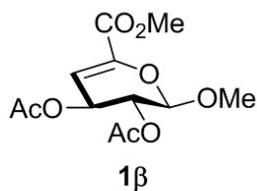
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6.2118
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5.2011
5.1969
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5.1862
5.1793
5.1319
5.1224
5.0893
5.0843
5.0689
3.8398
3.5172
2.1038
2.0877



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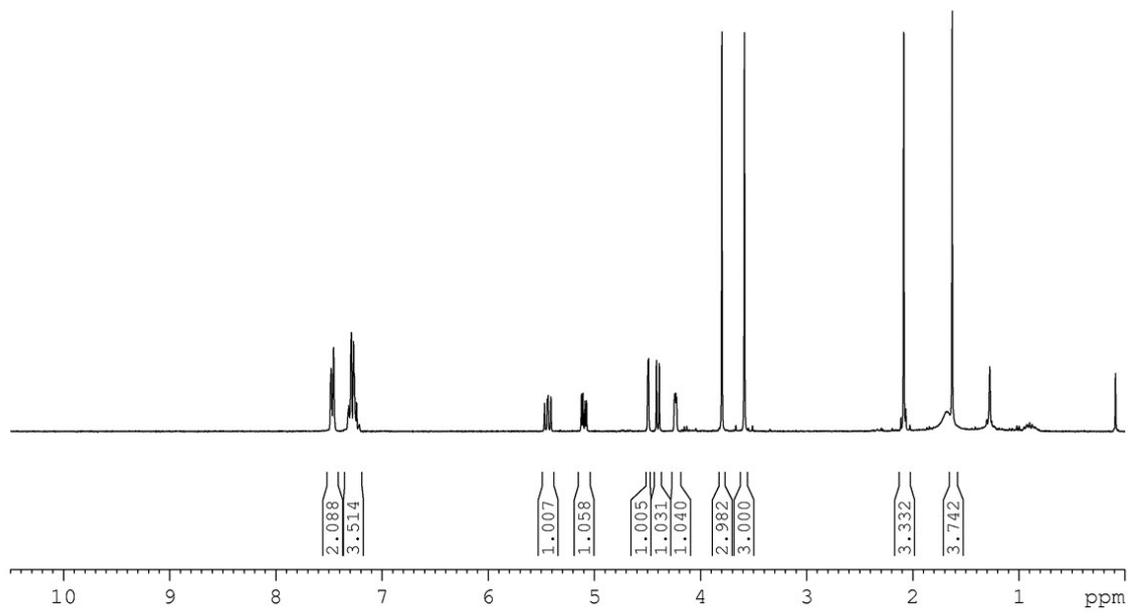
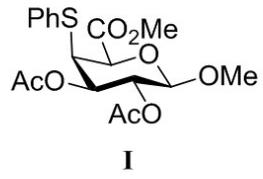


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7.2345

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5.4338
5.4073
5.1198
5.1052
5.0863
5.0713
4.4936
4.4876
4.4129
4.3865
4.2435
4.2370
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— 2.0820
— 1.6257



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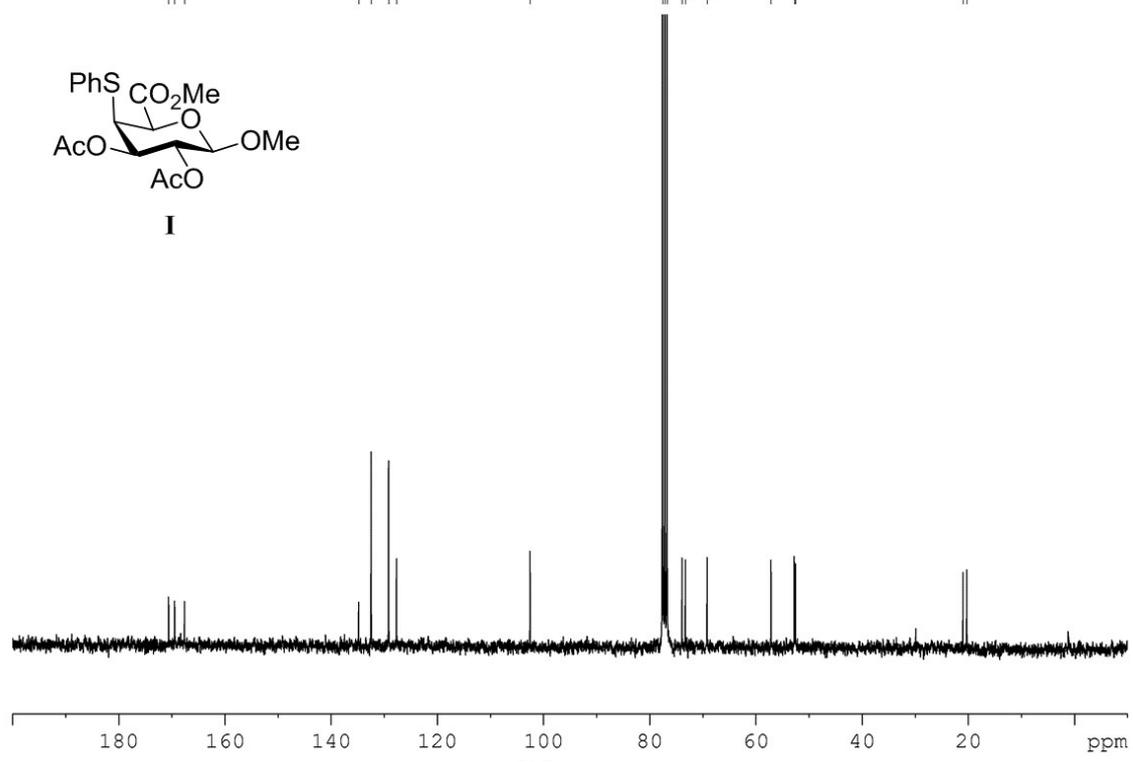
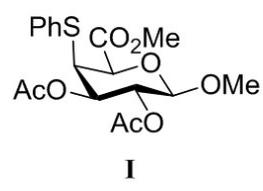
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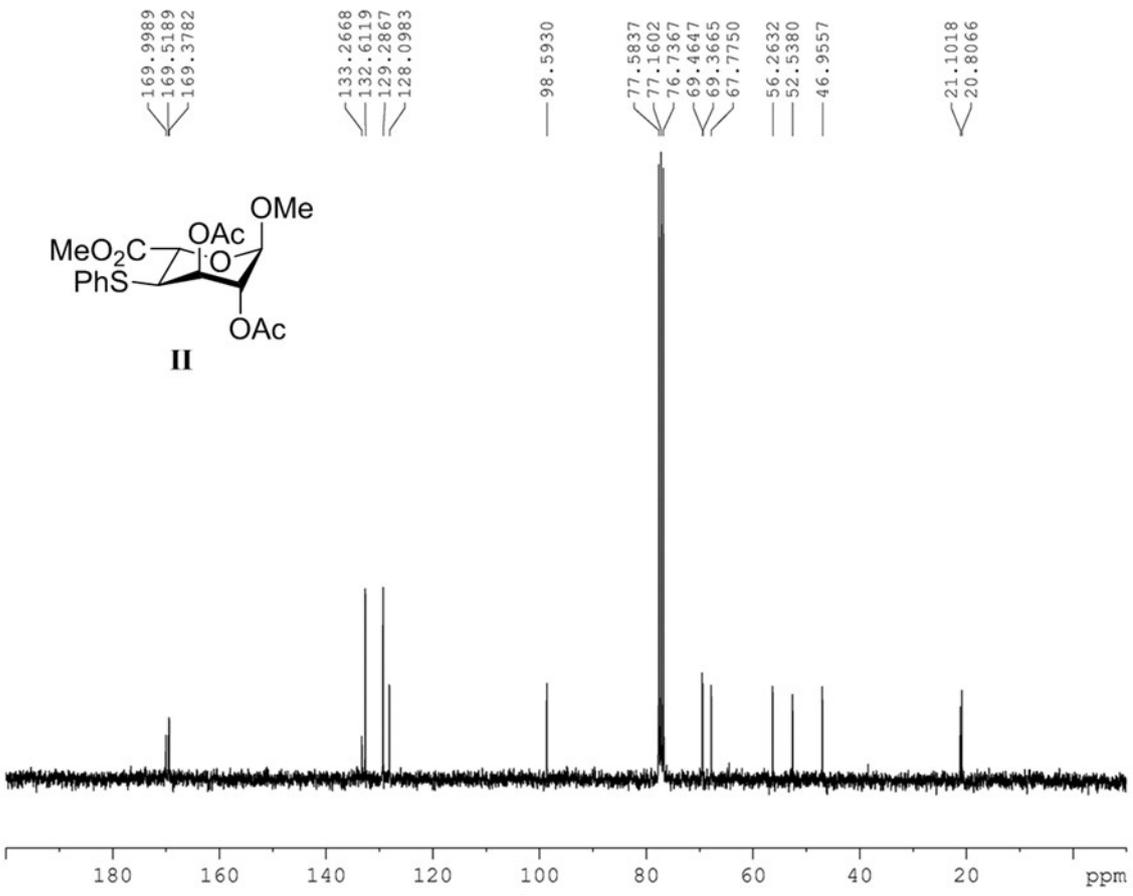
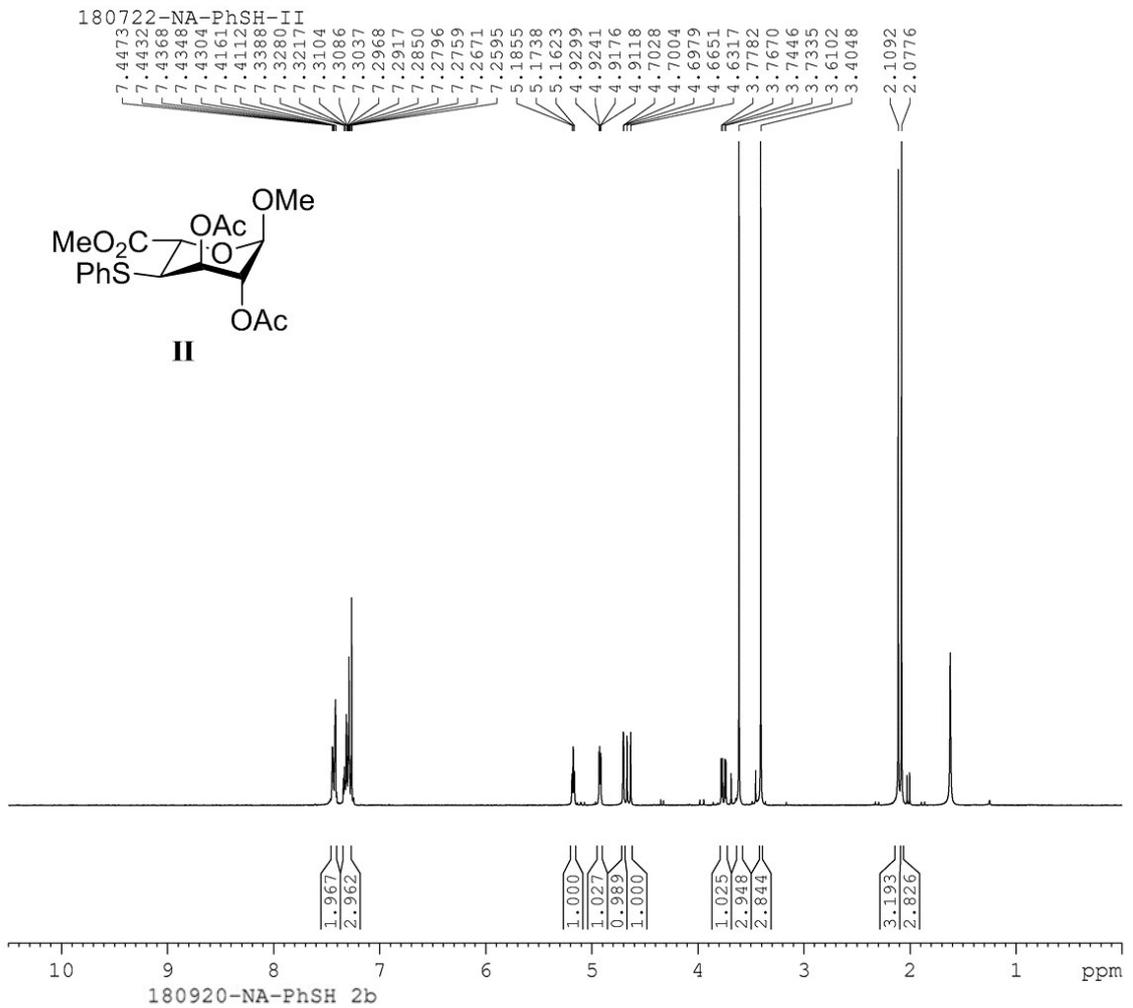
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129.1522
127.6656

— 102.5115

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52.7390
52.5144

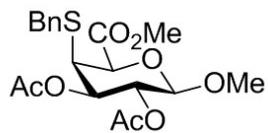
— 20.9438
— 20.2411



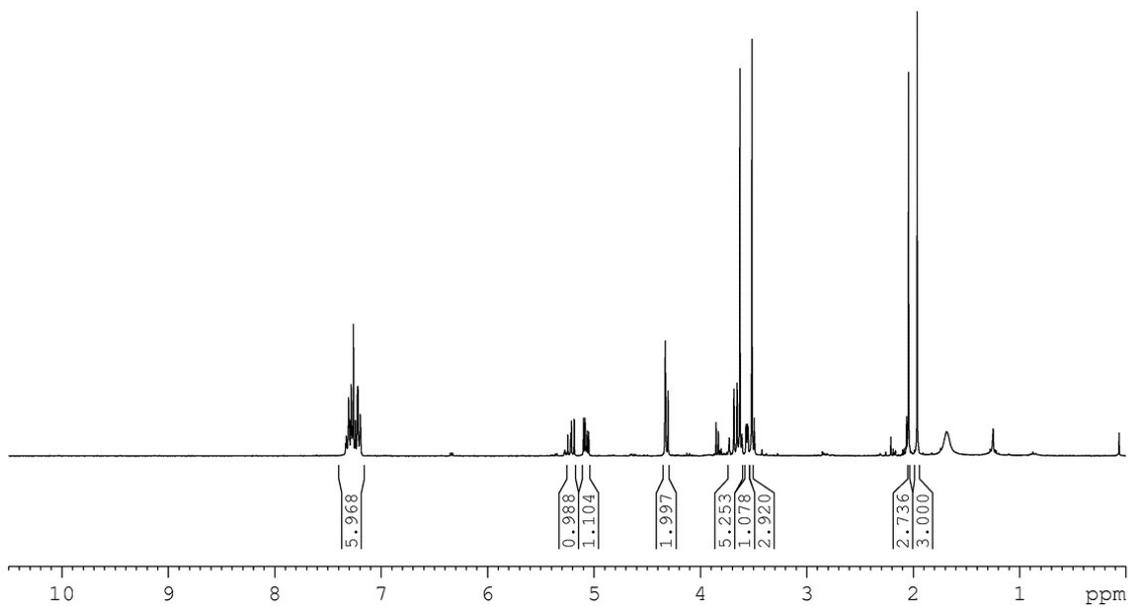


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7.2216
7.2158
7.1948
5.2428
5.2169
5.2089
5.1830
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5.0814
5.0623
5.0476
4.3278
4.3234
4.3015
3.7271
3.6819
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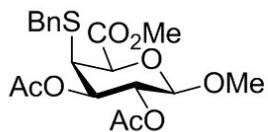


III

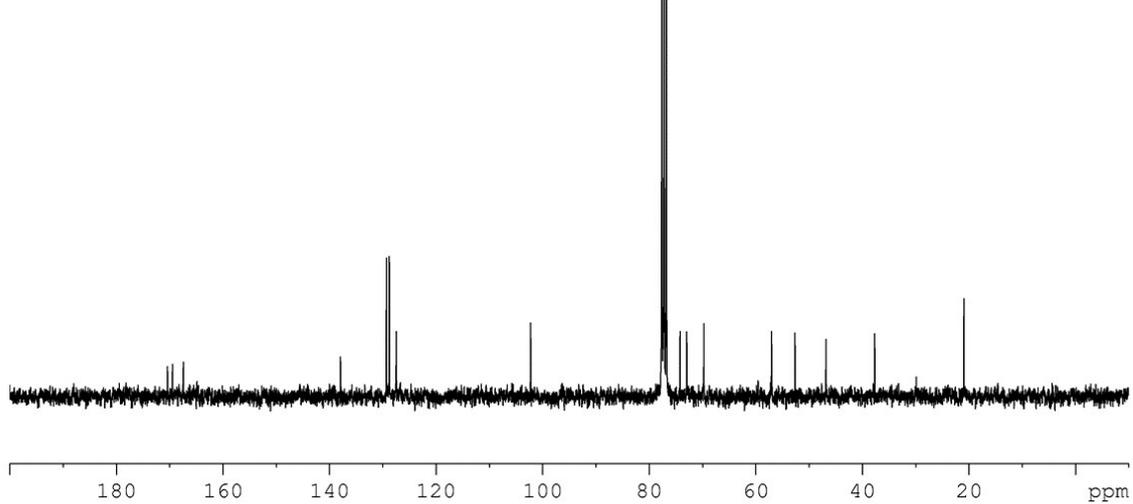


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III

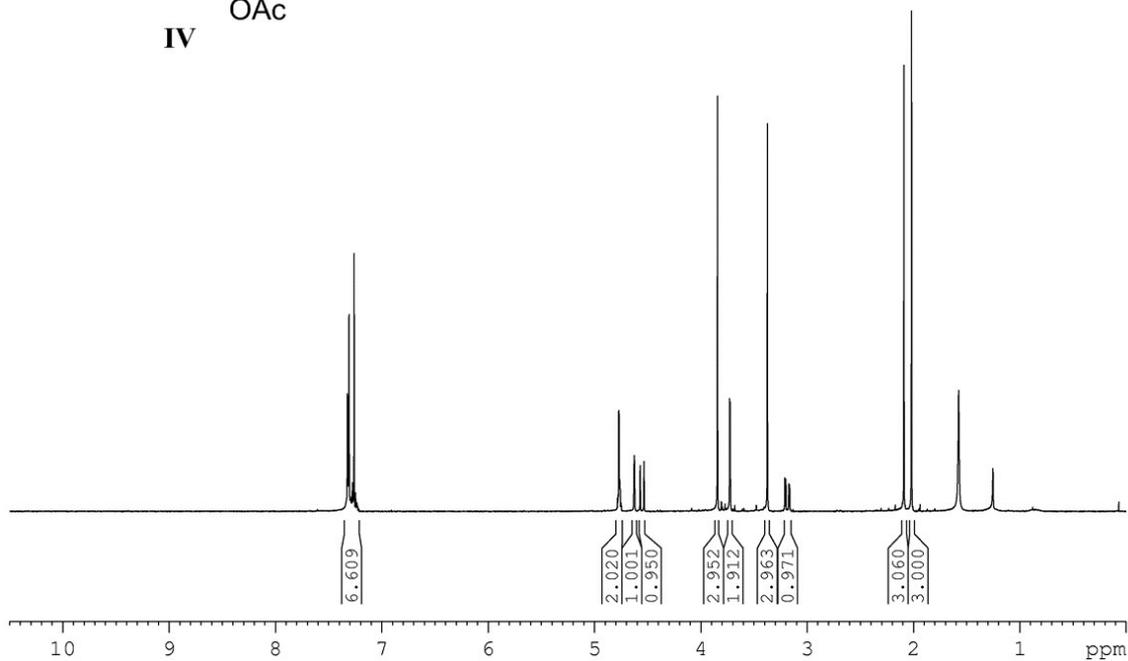
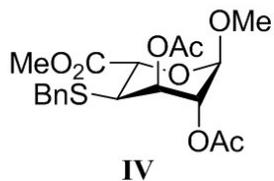


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4.7501
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4.6229
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4.5323
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3.7265
3.7220
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3.1710
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2.0869
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190518-NA-314475-tube 10 13C

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98.4726

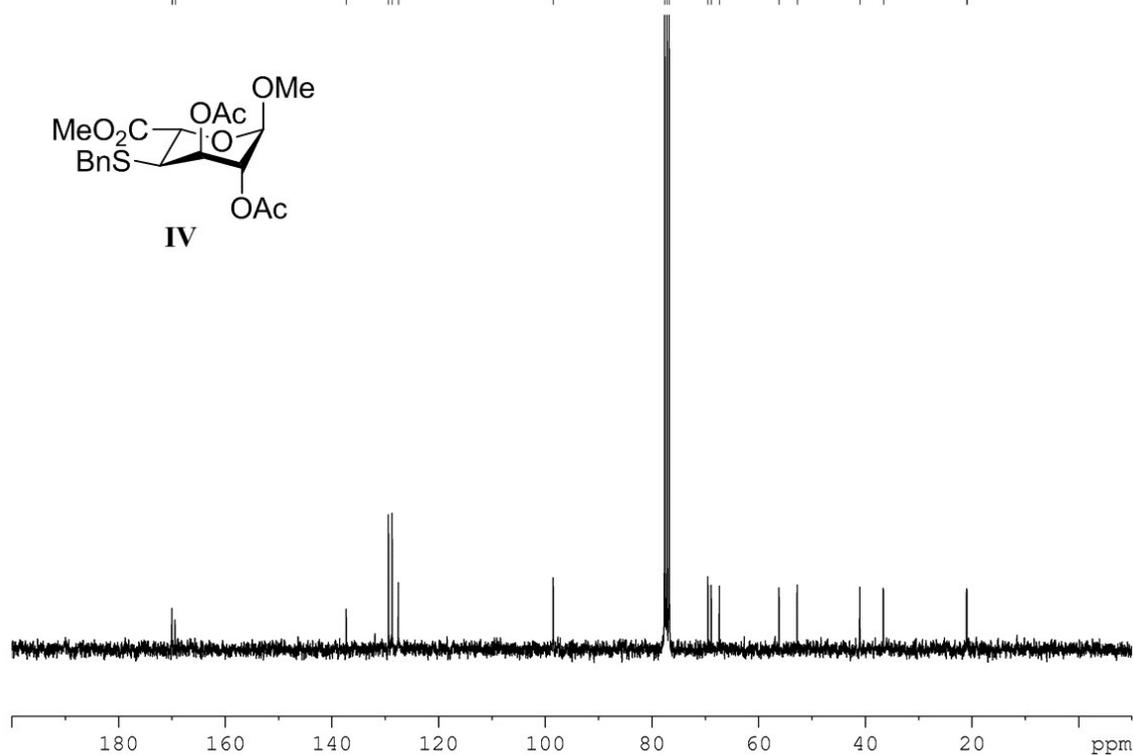
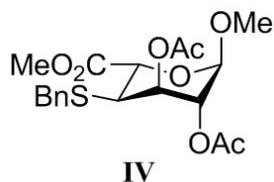
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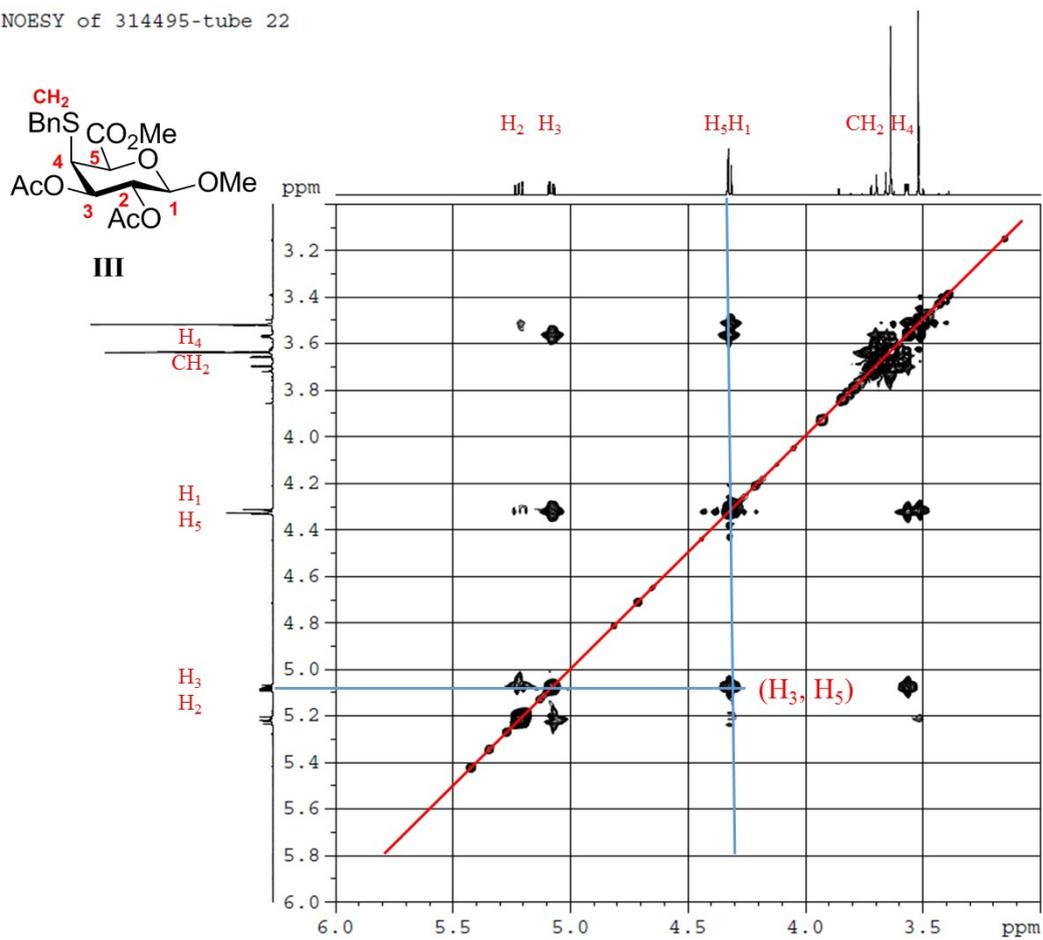
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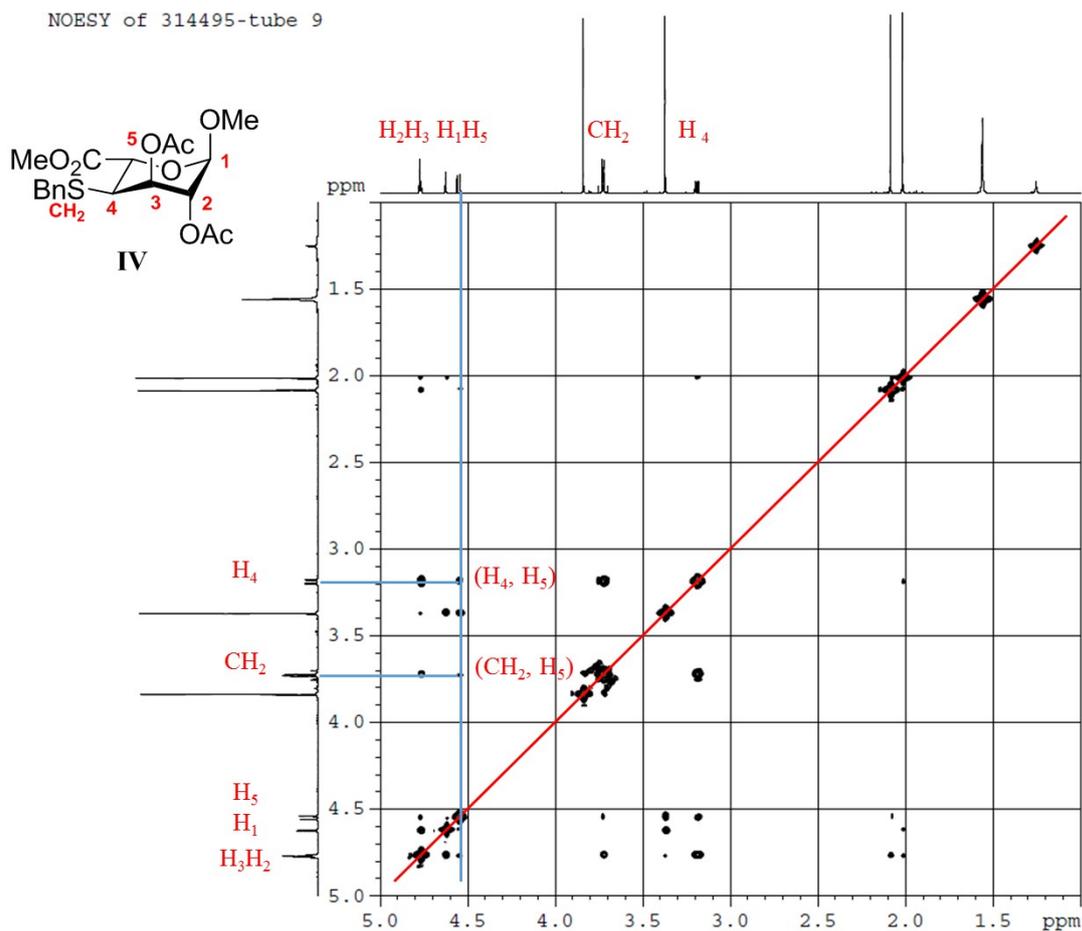
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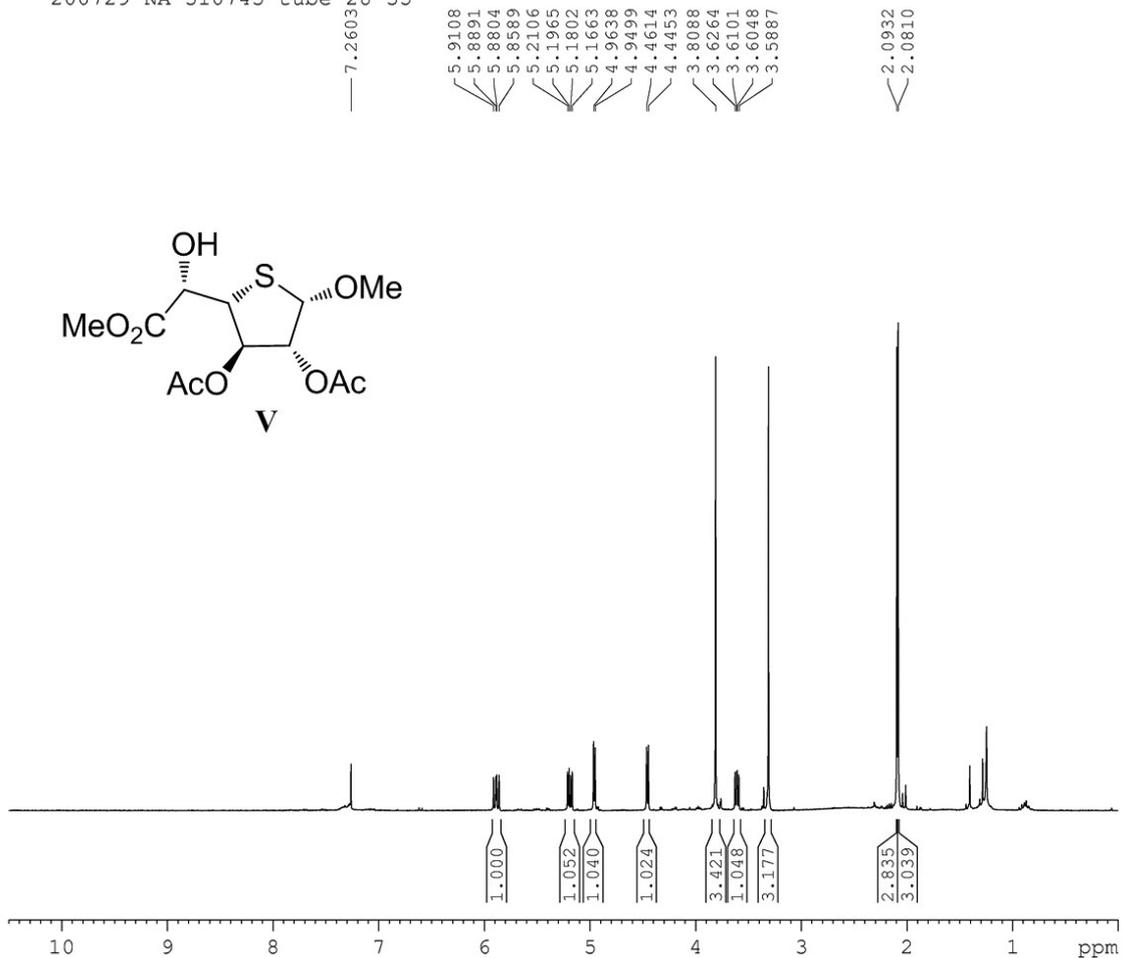
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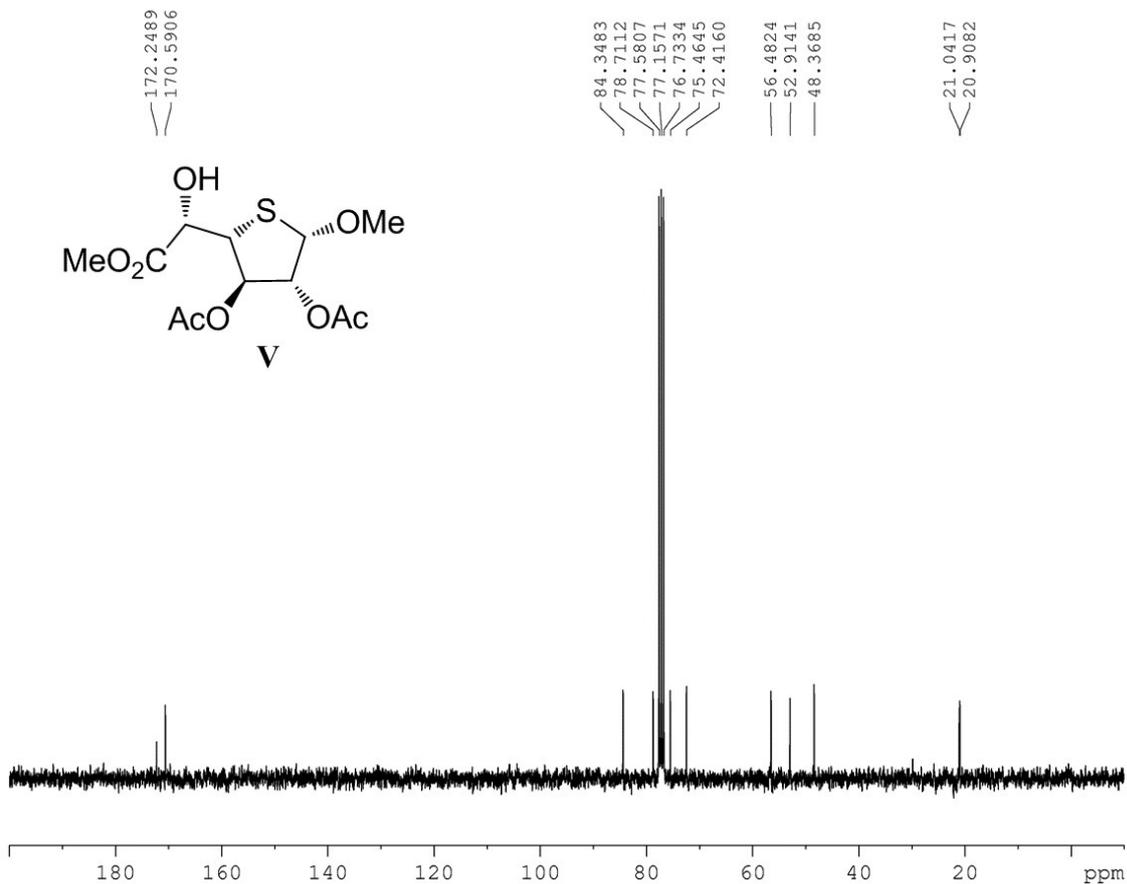
NOESY of 314495-tube 9



200729-NA-310743-tube 28-35



200729-NA-310743-tube 28-35 13C



NOESY of 310743

