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Electronic Supplementary Information

New carvacrol and thymol derivatives as potential insecticides: synthesis, biological activity, computational studies and nanoencapsulation

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1. ¹H and ¹³C NMR spectra of compounds 3a-c, 4a-c, 5, 6 and 8

¹H NMR spectra of compounds **3a-c**, **4a-c**, **5**, **6** and **8** are shown. These spectra confirm the corresponding structure and purity of each compound. In addition, ¹³C spectra are also shown. This information serves as the statement for confirming the purity (\geq 95%) of the compounds extracted/synthesized in the reported work.



































2. RMSD Analysis

A graphical representation of the RMSD values is presented in Figure S1. Low RMSD values indicate that the protein-ligand systems are well equilibrated and that compounds **6** and **8** maintained the binding pose anticipated in the inverted virtual screening stage. Compounds **6** and **8**, when bound to OBP, show a low RMSD contrary to AChE. This target displays higher values, which may suggest that there is an induced-fit adjustment to the AChE-binding pocket during the MD simulation. When looking at the RMSD of the ligands, compound **6** is the molecule that possesses lower RMSD values. This is mainly due to its own chemical structure, which is less flexible.



Figure S1. Protein and ligand RMSD (Å) of the AChE and OBP – ligand complexes.



Figure S2. Variation of the percentage of potential ligand SASA buried by protein (%) when in complex with

AChE and OBP.





Figure S3. Hydrogen bonds formed throughout the simulation between AChE-compound **6**; compound **8** and OBP-compound **6**; compound **8**.

3. Creation of a homology model for 1QON

The SWISS-MODEL homology model created for the AChE molecular structure is shown in Figure S4. There are two metrics used to evaluate the quality of the model: GMQE and QMEAN. GMQE - Global Model Quality Estimation, which is expressed between 0 and 1 with a higher number meaning higher reliability. QMEAN - provides an estimate of the "degree of nativeness" of the structural features observed in the model. A value of QMEAN around zero indicate a good agreement between the model and experimental structure.



Figure S4. Homology model created for 1QON. Presented in green is the original structure. The only area that was built was the loop represented in red. An example of one of the ligands studied represented in yellow to illustrate that the area that was modelled is far from the active site.

Table S1. Average scores of compounds 6 and 8 against all the protein targets evaluated with the five different scoring functions. Overall ranking of the most likely protein targets for interaction.

Target	PDB	PLP	ASP	ChemScore	GoldScore	Vina	Overall ranking	
Ecdysone receptor	1R20	53.60	24.36	27.34	42.29	-7.65	7	
	1R1K	61.76	23.85	29.22	48.89	-8.30	1	
Chitinase	3WL1	67.02	34.59	28.88	47.86	-7.90	2	
	3WQV	65.04	41.06	30.99	56.28	-7.80	5	
beta-N-acetyl-D-hexosaminidase OfHex1	30ZP	60.18	45.46	29.46	57.33	-7.30	4	
	3NSN	65.70	38.69	28.23	54.04	-6.60	4	
N-Acetylglucosamine-1-phosphate uridyltransferase (GlmU)	2V0K	50.25	16.83	22.49	40.16	-7.10	12	
	2VD4	43.77	18.73	20.24	38.70	-6.10	12	
Acetylcholinesterase	1QON	80.74	42.82	34.98	53.35	-9.80		
	4EY6	63.53	32.14	31.85	52.20	-8.70	2	
	1DX4	68.13	33.73	29.94	48.72	-8.20		
Prophenoloxidase (PPO)	3HHS	45.59	16.01	19.11	46.26	-5.40	13	
<i>p</i> -Hydroxyphenylpyruvate dioxygenase	6ISD	57.29	25.10	24.51	24.87	-7.20	9	
Voltage-gated sodium channel	6A95	53.48	18.32	22.42	46.25	-6.60	11	
Octopamine receptor	4N7C	45.21	33.21	37.59	41.31	-5.65	8	
Sterol carrier protein-2 (HaSCP-2)	4UEI	56.94	31.26	32.57	48.87	-8.10	6	
Peptide deformylase	5CY8	55.95	26.70	24.46	52.73	-5.30	10	
α-Esterase-7	5TYJ	56.04	33.93	30.58	52.01	-7.10	5	
	5TYP	56.29	35.23	32.03	54.43	-7.50		
Odorant binding protein	5V13	70.27	34.43	34.79	52.32	-8.50	1	
	3K1E	73.06	36.41	34.38	55.92	-8.80		
	2GTE	63.90	34.64	35.77	56.02	-8.35		
	3N7H	65.42	34.01	30.95	57.93	-7.25		

Table S2. Docking scores for Human and Insect AChE when in complex with compounds 6 and 8.

	Compound	PLP	ASP	ChemScore	GoldScore	Vina
Human AChE	8	69.42	41.22	35.15	63.06	-7.9
Insect AChE	8	92.19	60.29	42.82	70.73	-9.7
Human AChE	6	65.43	33.64	33.54	50.44	-8.6
Insect AChE	6	71.4	42.82	34.95	53.35	-9.8

Table S3. Targets selected for the inverted virtual screening assay

Target	Organism	PDB target	Resolution (Å)	Description	Ref	
Ecdysone receptor	Heliothis	1R20	3.00	VS based on 1R20 bound to an agonist as a model for the development of a receptor-based pharmacophore model.		
	virescens	1R1K	2.90 VS of 2 million compounds against 1R1K, an ecd receptor structure bound to its known ligand ponasterc		2	
Chitinase beta-N-acetyl-D- hexosaminidase OfHex1		3WL1	1.77	Pharmacophore-based screening using two crystal structures	_	
	Ostrinia furnacalis	3WQV	2.04	of chitinases: 3WL1 bound to its reaction product and 3WQV bound to an inhibitor.		
		3NSN	2.10	VS of the ZINC database to identify OfHex1 inhibitors using 3NSN crystal structure bound to a known inhibitor.	4	
		30ZP	2.00	VS of the ZINC data-base targeting 3OZP, a crystal structure of OfHex1 bound to an inhibitor.	5	
N-		2V0K	2.30			
Acetylglucosami- ne-1-phosphate uridyltransferase (GImU)	Xanthomonas oryzae	2VD4	1.90	Homology model built for docking using 2V0K and 2VD4 as templates. 2V0K crystal structure is bound to its known ligand and 2VD4 is bound to a possible inhibitor.	6	
Acetylcholines-	Aedes aegypti	1QON	2.72	Search for new molecules with insecticidal activity against Ae.		
		4EY6	2.40	Aegypti using acetylcholinesterase crystal structures 1QON and 4EY6 as targets, both bound to possible inhibitors.		
Drosophila melanogaster		1DX4	2.70	Homology 3D model built for VS using 1DX4 as template. 1DX4 crystal structure is bound to a potent inhibitor.		
Prophenoloxidas e (PPO)	Manduca sexta	3HSS	1.97	Crystal structure of a prophenoloxidase from Manduca sexta.	9	
<i>p</i> - Hydroxyphenyl- pyruvate dioxygenase	Arabidopsis thaliana	6ISD	2.40	Development of a receptor-ligand pharmacophore model based on the crystal structure 6ISD bound to a commonly used pesticide. The best model created was then used for VS studies.	10	
Voltage-gated sodium channel	Periplaneta americana	6A95	2.60	Crystallographic structure of a Voltage-gated sodium channel NavPaS bound to a pore blocker, tetrodotoxin (TTX)	11	
Octopamine receptor	Blattella germanica	4N7C	1.75	Crystal structure of Bla g 4, an octopamine receptor, bound to tyramine.	12	
Sterol carrier protein-2 (HaSCP-2)	Helicoverpa armigera	4UEI	Solution NMR	Structure-based VS of a database of commercially available compounds to find potential inhibitors of HaSCP-2. The residues Phe53, Thr128, and Gln131 were selected for the binding cavity.	13	
Peptide deformylase	Xanthomonas oryzae	5CY8	2.38	Docking and VS of a library of 318 phytochemicals. 5CY8 crystal structure is bound to a possible inhibitor.	14	
Alpha-Esterase-7 (αΕ7)	Lucilia cuprina	5TYJ	1.75	Computational design of potent and selective covalent		
		5TYP	1.88	inhibitors of α E7. 5TYJ and 5TYP crystal structures are bound to inhibitors: (3-bromo-5-phenoxylphenyl)boronic acid and (3-bromo-4-methylphenyl)boronic acid, respectively.	15	
Odorant Binding Protein	Aedes aegypti	5V13	1.84	Search for new molecules with insecticidal activity against <i>Ae. Aegypti</i> using a crystal structure of a mosquito juvenile hormone-binding protein, 5V13 bound to its natural hormone.	7	
	Drosophila melanogaster	2GTE	1.40	2GTE crystal structure is bound to its natural ligand.	16	
	Anopheles gambiae	3N7H	1.60	QSAR and docking studies for the rational design of mosquito repellents using the crystal structure 3K1E bound to a	17	
	Aedes aegypti	3K1E	1.85	polyethylene glycol molecule. 3N7H crystal structure is bound to a commonly used repellent.		

References

- X. Hu, B. Yin, K. Cappelle, L. Swevers, G. Smagghe, X. Yang and L. Zhang, J. Mol. Graph Model., 2018, 81, 77-85.
- 2. T. Harada, Y. Nakagawa, T. Ogura, Y. Yamada, T. Ohe and H. Miyagawa, *J. Chem. Inf. Model.*, 2011, **51**(2), 296-305.
- Y. Dong, X. Jiang, T. Liu, Y. Ling, Q. Yang, L. Zhang and X. He, J. Agric. Food Chem., 2018, 66, 3351-3357.
- 4. J. Liu, M. Liu, Y. Yao, J. Wang, Y. Li, G. Li and Y. Wang, Int. J. Mol. Sci., 2012, 13, 4545-4563.
- 5. L. Dong, S. Shen, Y. Xu, et al., J. Biomol. Struct. Dyn., 2021, 39, 1735-1743.
- 6. J. Min, D. Lin, Q. Zhang, J. Zhang and Z. Yu, Eur. J. Med. Chem., 2012, 53, 150-158.
- 7. R. Ramos, J. Costa, R. Silva, G. Costa, A. Rodrigues, E. Rebelo, R. Souto, C. Taft, C. Silva, J. Rosa, C. Santos and W. Macêdo, *Pharmaceuticals*, 2019, **12**, 20.
- 8. C. Riva, P. Suzanne, G. Charpentier, F. Dulin, M. Halm-Lemeille and J. Sopkova-de Oliveira Santos, *Pestic. Biochem. Physiol.*, 2019, **160**, 11-19.
- 9. Y. Li, Y. Wang, H. Jiang and J. Deng, Proc. Natl. Acad. Sci., 2009, 106, 17002-17006.
- 10. Y. Fu, Y. Liu, T. Kang, Y. Sun, J. Li and F. Ye, J. Taiwan Inst. Chem. Eng., 2019, 103, 33-43.
- 11. H. Shen, Z. Li, Y. Jiang, X. Pan, J. Wu, B. Cristofori-Armstrong, J. Smith, Y. Chin, J. Lei, Q. Zhou, G. King and N. Yan, *Science*, 2018, **362**(6412), 1-8.
- 12. L. Offermann, S. Chan, T. Osinski, Y. Tan, F. Chew, J. Sivaraman, Y. Mok, W. Minor and M. Chruszcz, *Mol. Immunol.*, 2014, **60**, 86-94.
- 13. J. Cai, X. Du, C. Wang, J. Lin and X. Du, J. Econ. Entomol., 2017, 110, 1779-1784.
- 14. T. Joshi, T. Joshi, P. Sharma, S. Chandra and V. Pande, J. Biomol. Struct. Dyn., 2021, **39**, 823-840.
- 15. G. Correy, D. Zaidman, A. Harmelin, S. Carvalho, P. Mabbitt, V. Calaora, P. James, A. Kotze, C. Jackson and N. London, *Proc. Natl. Acad. Sci.*, 2019, **116**, 21012-21021.
- 16. J. Laughlin, T. Ha, D. Jones and D. Smith, *Cell*, 2008, **13**, 1255-1265.
- 17. P. Oliferenko, A. Oliferenko, G. Poda, D. Osolodkin, G. Pillai, U. Bernier, M. Tsikolia, N. Agramonte, G. Clark, K. Linthicum and A. Katritzky, *PLoS One*, 2013, **8**, e64547.