Supporting Information

Self-assembly of monohydroxy Terpenoid Lupeol yielding nano-Fibers, Sheet and Gel: Environmental and drug Delivery application

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1. Energy minimized structure of lupeol



Figure S1: Energy minimized structure of Lupeol 1 using MMX force field as implemented in PCMODEL version 9.2 (Serena software)[®].



Figure S2: Energy minimized structure of Lupeol 1 obtained from DFT using (Gaussian 09 software)[®].

2. T_{gel} vs. Concentration Plots:



3. Calculation the thermodynamic parameter:

Calculation:

The thermo-reversibility of a gel melting can be expressed as:

 $\text{Gel} \rightarrow \text{sol}$

The equilibrium constant (K_{eqm}) for gel to sol transition:

 $K_{eqm} = [Gelator]/[Gel]$

Assuming unit activity of the gel, the equilibrium constant can be expressed as:

 $K_{eqm} = [Gelator]$

The Gibbs free energy (ΔG°) change during gel melting can be expressed as:

 $\Delta G^{\circ} = -RTlnK_{eqm} = \Delta H^{\circ} - T\Delta S^{\circ}$, Hence, $lnK_{eqm} = -\Delta^{\circ}/R$. $(1/T) + \Delta S^{\circ}/R$

The gel melting temperature (T_{gel}) increases with increasing concentration of the "solutes". A plot of lnK vs 1/T allowed us to calculate the thermodynamic parameters.





4. OPM images of the self-assemblies



Figure S5: OPM images of self-assemblies of 1: (a,b) THF (1.5% w/v, 35.2 mM), (c) cyclohexane (1.5 % w/v, 35.2 mM), (d) DMF-H₂O (1:1, 2.0 % w/v, 46.9 mM), (e,f) n-hexane (1.5 %w/v, 35.2 mM), (g,h) n-heptane (1.5 %w/v, 35.2 mM)

5. AFM images of the self-assemblies



Figure S6: AFM micrographs of dried self-assemblies of 1 : (a) n-heptane (0.12%w/v), (b) odichlorobenzene(0.15 %w/v), (c) DMF-H₂O (1:1, 0.12 % w/v)

6. FESEM images of the self-assemblies



Figure S7: FESEM images of self-assemblies of 1: (a,b) DMF (1.5 %w/v), (c,d) EtOH (1.25 %w/v).

7. HRTEM images of the self-assembled twisted fibers



Figure S8: HRTEM images of self-assemblies of 1: (a,b) n-heptane DMF (0.8 %w/v).

8. Epifluorescence microscopy of dye entraped gel



Figure S9: Epifluorescence microscopy images of the adsorbed fluorophores by the gels of **1** in DMSO-water (2:1): (a-c) adsorbed rhodamine-B (0.2 mM), (d-f) adsorbed CF (0.2 mM), (g-i) adsorbed doxorubicin (0.15 mM) after release respectively. Grey, green and red images are under normal, blue, green lights respectively.

9. Rheology of gels



Figure S10 : Rheological amplitude stress Sweep experiments of the gels of 1 (a-c) in DMSO-H₂O (2:1 v/v), (d-f) in DMF 2.5 %w/v, (g-i) in n -octane at concentration 2.0 % w/v, 2.5 %w/v, 3.5 % w/v, respectively.

10. FTIR spectrum of Lupeol



Figure S11: FTIR spectra of xerogel of 1 (a) butanol (5 % w/v) and o-dichlorobenze (4 % w/v), (b) n-hexane (1.7 % w/v) and n-octane (2 % w/v) respectively

11. Various possible modes of assembly of lupeol



Fig. S12: Schematic representation of two interacting lupeol molecules within van der Waals contact and facing opposite (α - β) to each other



Fig. S13: Schematic presentation of two interacting lupeol molecules within van der Waals contact and facing (α - α) to each other



Fig. S14: Schematic representation of two interacting lupeol molecules within van der Waals contact shown as terpenoid β -face facing each other



Fig. S15: Schematic representation of two interacting lupeol molecules within H-bonding





Fig. 16 Schematic representation of interacting lupeol molecules forming selfassembled architecture via within van der Waals interaction and contact and Hbonding

12. Experimental

12.1 Materials.

All solvents used for purification purposes were commercial grade and were distilled before use. The liquids used for gelation studies were laboratory-grade reagents and dried using standard literature methods and distilled before use. 12.2 Method of Sample Preparation and Characterization.

For self-assembly studies, 2–5 mg of compound 1 contained in a vial (1 cm id) was heated with a liquid with continuous magnetic stirring over a hot plate until a clear solution was obtained. The solution was then allowed to cool at room temperature (24-25)°C). When the material did not flow as observed by turning the vial upside down we called it a gel. The selfassemblies have been characterized by various spectroscopic and microscopic techniques. Scanning electron microscopy samples were prepared by placing a dilute solution of the sample on a aluminium foil and then allowing it to dry initially in air for 24 h and then under reduced pressure for 12 h and then sputter coated with Au before use for 120 s and studied using a Zeiss field-emission scanning electron microscope (FESEM). For optical microscopy, an aliquot of sample was taken on a glass plate and covered with a coverslip and observed both under normal and polarized light using a Nikon LV100 POL microscope. TEM images of the self-assemblies were recorded on dilute solution samples placed on 300 mesh carbon coated copper grids and dried at ambient temperatures in the air for 24 h and then under reduced pressure for 24 h and studied using JEOL transmission electron microscopy. For wide-angle X-ray scattering (WAXS) experiment, a thin layer of self-assemblies was placed on a glass plate and the volatiles were removed initially in air and then under reduced pressure and the diffractions were recorded in a Bruker X-ray diffractometer at 25 °C using Co-Ka filament ($\lambda = 1.789$ Å). For the measurement of diffraction pattern of a powder sample, it was taken directly in the diffractometer cell and measured. For FTIR spectra of the neat powder and self-assemblies were analyzed by using a PerkinElmer Spectrum.