Supporting Information

Engineering Bacteria to Control Electron Transport Altering the Synthesis of Biopolymer

Mechelle R. Bennett, [a] Akhil Jain, [a, b] Katalin Kovacs, [b] Phil J. Hill, [c] Cameron Alexander, [d] Frankie J. Rawson*[a]

[a] Division of Regenerative Medicine and Cellular Therapies, Biodiscovery Institute, School of Pharmacy, University of Nottingham, University Park, Nottingham, NG7 2RD

*E-mail: Frankie.Rawson@nottingham.ac.uk

- [b] Synthetic Biology Research Centre, School of Life Sciences, University of Nottingham, University Park, Nottingham, NG7 2RD
- [c] Division of Microbiology, Brewing and Biotechnology. School of Bioscience, University

of Nottingham, Sutton Bonington Campus, Nottingham LE15 5RD

[d] Division of Molecular Therapeutics and Formulation, Boots Science Building, School of Pharmacy, University of Nottingham, University Park, Nottingham, NG7 2RD

Table S1. Bacteria and Plasmid Strains.

Strain/Plasmid	Description	Source/Reference
Escherichia coli wild type	Plasmid Storage Strain (K12 Top 10)	Invitrogen
pMTL_83153	Modular plasmid containing pCB102, catP, ColE1 + tra, P _{fdx} + MCS	http://www.clostron.com/p MTL80000.php

Table S2. Oligonucleotide Primers used for PCR of DNA regions. All Primers used for PCR with Q5 ® Polymerase. *Exceptions used for Colony PCR with Green DreamTaq.

Primers	Sequence (5'-3')	Tm (°C)	Function	
NapC_fwd_hifi	GAGCGAAATCATGGGAA	61.8	To amplify <i>napC</i> in cloning	
	ATTCTGACCGTAAG		with P_{BAD} promoter	
NapC_rev_hifi	ATCTCCATGGACGCGTG	59.3	To amplify <i>napC</i> in cloning	
	ACGTTAAAAACCTGGCTC		with P_{BAD} promoter	
	GAC			
	CAGGAAACAGCTATGAC	59.3		
P _{BAD} _araC-fwd	CGCTTATGACAACTTGAC		Amplify P _{BAD} promoter	
	GGC			
	AATTTCCCATTTTCTCCT	58.9		
P _{BAD} _araC_rev	CTTTAATCTAGAGAATTC		Amplify P_{BAD} promoter	
ColE + tra_F2*	CCATCAAGAAGA GCGAC	56.7	Colony PCR	
CD402 D4		EE A	Colony DCD	
pCB102_R1*	GATAGTCAAAAGGCATAA	55.4	Colony PCR	
	CAG			

Table S3. Arabinose induction concentrations for *E. coli* containing Inducible Promoter Vector.

Sample	Stock arabinose (w/v)	Final arabinose Concentration	

<i>E. coli</i> (IP_0%)	0	0	
<i>E. coli</i> (IP_0.000018%)	0.0018%	0.000018%	
<i>E. coli</i> (IP_0.0018%)	0.18%	0.0018%	
<i>E. coli</i> (IP_0.18%)	18%	0.18%	

Table S4. Protein standardisation calculations using BCA assay. *Calculated using BSA equation (y=0.0016x). **Note samples were diluted by 2; therefore, the concentrations were doubled to obtain original solution concentrations.

Vector	Abs _{562nm} (av)	Sample conc (µg/mL)*	Original conc (µg/mL)**	mg/ml
Empty Plasmid	1.57	982.7	1965	1.97
<i>E.coli</i> (IP_0%)	1.63	1019	2038	2.04
<i>E.coli</i> (IP_0.000018%)	1.69	1053	2106	2.11
<i>E.coli</i> (IP_0.0018%)	1.65	1029	2058	2.06
<i>E.coli</i> (IP_0.18%)	1.40	873	1746	1.75

Table S5. Reagent ratios for *E.coli_{IP}* initiated Fe ATRP.

Reagent	Ratio	Mmol	Mass (mg)	Vol (uL)
PEGMA	100	0.052	17	16.2
FeCl ₃ .6H ₂ 0	4.65	0.0024	0.65	-
Me ₆ TREN	13.95	0.0072	1.7	1.9
HEBIB	2	0.0010	0.22	0.15

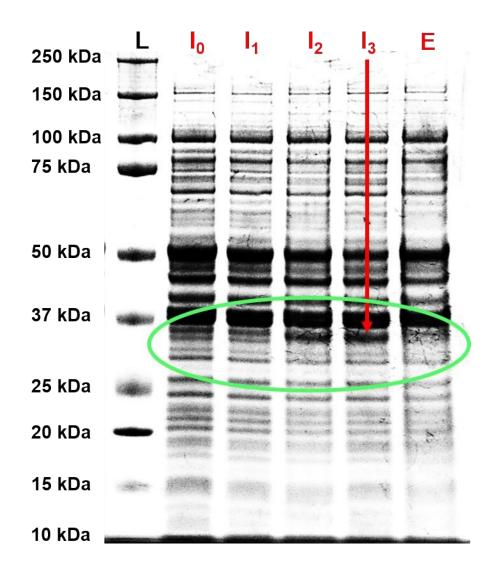


Figure S1. Protein expression analysis. SDS PAGE Gel for lysates of bacteria containing empty plasmid (E), Inducible promoter vector with 0% (I₀), 0.000018% (I₁), 0.0018% (I₂) and 0.18% (I₃) total arabinose concentration induction. Protein Gel against Precision Plus Protein TM Kaleidoscope ladder (L).

BCA Assay - Standard BSA curve

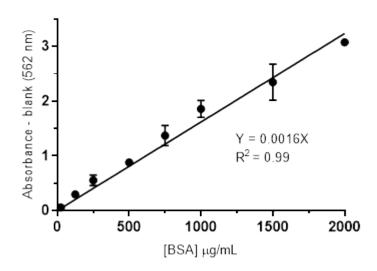


Figure S2. BCA Standard curve for BSA (Bovine Serum albumin).

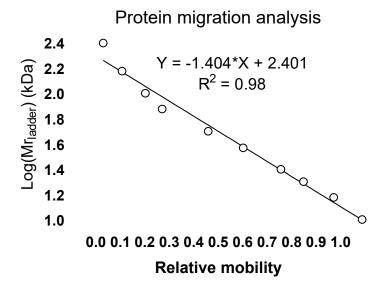


Figure S3. Protein migration analysis of protein ladder Log(Mr) against relative mobility on SDS PAGE Gel.

Calculation:

Migration (NapC) = 10.7 cm Dye front = 17.2 cm

Relative mobility (X) =
$$migration 10.7$$

 $dye front 17.2$ = 0.622
 $dye front 17.2$
 $log(Mr_{NapC}) = [-(1.404 * X) + 2.401]$
 $log(Mr_{NapC}) = (-1.404 * 0.622) + 2.401$
 $log(Mr_{NapC}) = (-1.404 * 0.622) + 2.401$

Fe ATRP with NapC Upregulation

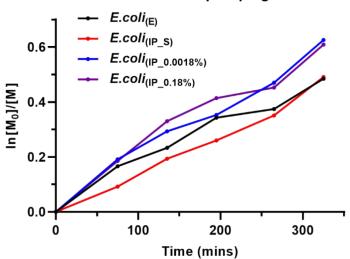


Figure S4. ¹H NMR kinetics of Fe ATRP activated by *E. coli* harbouring empty plasmids, *E. coli*_(E), or inducible promoter plasmids, *E. coli*_(IP) either i) suppressed by addition of glucose *E. coli*_(IP_S), ii) activated by 0.0018% total arabinose concentration *E. coli*_(IP_0.0018%) or ii) activated by 0.18% total arabinose concentration *E. coli*_(IP_0.18%).

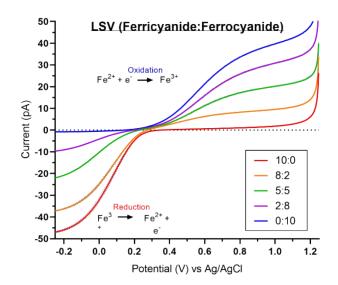


Figure S5. Linear sweep voltammogram of Current Vs Potential carried out using 3 electrode system with carbon fibre micro-disk electrode (33 μ m), Ag/AgCl reference electrode and Pt counter electrode in 1X PBS electrolyte. Scans were carried out at 100 mV/s from 1.25 V to -0.25 V. 1 mM potassium ferricyanide and ferrocyanide were made in PBS (1X) and mixed in ratios (10:0, 8:2, 5:5, 2:8, 0:10) and voltammograms observed (n=3, error = SD) for each sample, where electrode was polished between each scan.

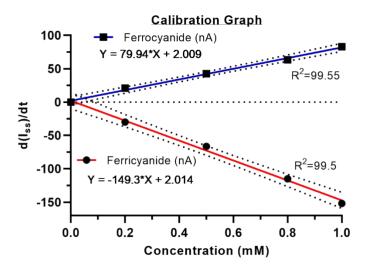
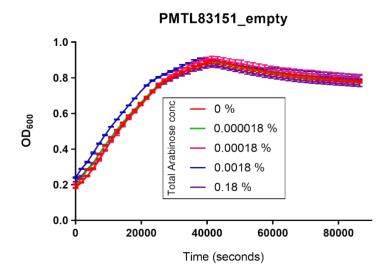


Figure S6. Calibration curve to determine Fe concentration. First derivative peaks from graphs in figures S5 and 3a against Ferricyanide or ferrocyanide concentration with line of best fit and 95% confidence limit.



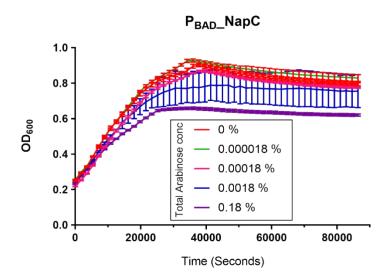


Figure S7. Arabinose toxicity study for, Top: *E. coli* containing empty plasmid and Bottom: *E. coli* containing Inducible vector Plasmid at different arabinose concentrations.