

## **Fine-tuning of pharmacological potentials of novel thiazolium ionic liquids by anion alteration**

Mohammad Y. Alfaifi,<sup>a</sup> Ali A. Shati,<sup>a</sup> Serag Eldin I. Elbehairi,<sup>b</sup> Reda F. M. Elshaarawy,<sup>c,d,\*</sup>  
Emad M. Gad<sup>e</sup>

<sup>a</sup> Biology Department, Faculty of Science, King Khalid University, 9004 Abha, Saudi Arabia.

<sup>b</sup> Cell Culture Lab, Egyptian Organization for Biological Products and Vaccines (VACSERA Holding Company), Giza 12311, Egypt.

<sup>c</sup> Chemistry Department, Faculty of Science, Suez University, 43533 Suez, Egypt.

<sup>d</sup> Institut für Anorganische Chemie und Strukturchemie, Heinrich-Heine Universität Düsseldorf, Düsseldorf, Germany.

<sup>e</sup> Chemistry Department, Faculty of Science, Suez Canal University, Ismailia, Egypt.

### **Corresponding author:**

RFME; [reda.elshaarawy@suezuniv.edu.eg](mailto:reda.elshaarawy@suezuniv.edu.eg); [reel-001@hhu.de](mailto:reel-001@hhu.de)

### **Contents:**

- 1- Materials and instrumentation**
- 2- Synthesis of 1-alkyl-4-(chloromethyl)benzene (1a-c)**
- 3- Figures**

## 1. Materials and instrumentation

Chemicals were obtained from the following suppliers and used without further purification: toluene, cumene, tert-butylbenzene, 4-methylthiazole, sodium tetrafluoroborate, lithium bis(trifluoromethanesulfonimide), and anhydrous  $\text{MgCl}_2$  (Sigma–Aldrich); Zinc iodide (Acros).

Melting points were measured using a BÜCHI Melting point B-540 apparatus; all melting points were measured in open glass capillaries and are uncorrected. Elemental analyses for C, H, N, were performed with a Perkin–Elmer 263 elemental analyzer. FT-IR spectra were recorded on a BRUKER Tensor-37 FT-IR spectrophotometer in the range  $400\text{--}4000\text{ cm}^{-1}$  as KBr disc in the  $4000\text{--}550\text{ cm}^{-1}$  region with  $2\text{ cm}^{-1}$  resolution or with an ATR (attenuated total reflection) unit (Platinum ATR-QL, diamond). For signal intensities the following abbreviations were used: br (broad), sh (sharp), w (weak), m (medium), s (strong), vs (very strong). UV/Vis spectra were measured at  $25\text{ }^\circ\text{C}$  in ethanol ( $10^{-5}\text{ mol/L}$ ) on a Shimadzu UV-2450 spectrophotometer using quartz cuvettes (1 cm). NMR-spectra were obtained with a Bruker Avance DRX200 (200 MHz for  $^1\text{H}$ ) or Bruker Avance DRX500 (125, 97 and 470 MHz for  $^{13}\text{C}$ ,  $^{11}\text{B}$  and  $^{19}\text{F}$  respectively) spectrometer with calibration to the residual proton solvent signal in  $\text{DMSO-}d_6$  ( $^1\text{H}$  NMR: 2.52 ppm,  $^{13}\text{C}$  NMR: 39.5 ppm),  $\text{CDCl}_3$  ( $^1\text{H}$  NMR: 7.26 ppm,  $^{13}\text{C}$  NMR: 77.16 ppm) against TMS ( $\delta = 0.00\text{ ppm}$ ) for  $^1\text{H}$  and  $^{13}\text{C}$ , 85% phosphoric acid ( $\delta = 0.00\text{ ppm}$ ) for  $^{31}\text{P}$  and  $\text{CFC}_3$  ( $\delta = 0.00\text{ ppm}$ ) for  $^{19}\text{F}$  NMR. Multiplicities of the signals were specified s (singlet), d (doublet), t (triplet), q (quartet) or m (multiplet). The ESI-MS of the synthesized compounds were acquired in the linear mode for positive ions on a UHR-QTOF maXis 4G (Bruker Daltonics) and BRUKER Ultraflex MALDI-TOF instrument equipped with a 337 nm nitrogen laser pulsing at a repetition rate of 10 Hz. The 2+ charge assignment of ions in HR-ESI-MS was confirmed by the  $m/z = 0.5$  difference between the isotope peaks (x, x+1, x+2). Peaks with chlorine showed the isotope ratio  $^{35/37}\text{Cl} = 75.8:24.2$ .

## 2. Synthesis of 1-alkyl-4-(chloromethyl)benzene (1a-c)

General procedure: A flask was charged with 5 mol% of  $\text{ZnI}_2$  (1.3 mmol), chlorosulfonic acid (31 mmol) and  $\text{CH}_2\text{Cl}_2$  (30 mL), followed by dropwise addition of dimethoxymethane (31 mmol) at  $-10\text{ }^\circ\text{C}$ . After stirring the reaction mixture at  $-10\text{ }^\circ\text{C}$  for 30min, the aromatic compound (26 mmol) was slowly added. The resulting mixture was then stirred at  $5\text{--}10\text{ }^\circ\text{C}$  for the time 0.5 - 2 h. The reaction was monitored by TLC analysis. After

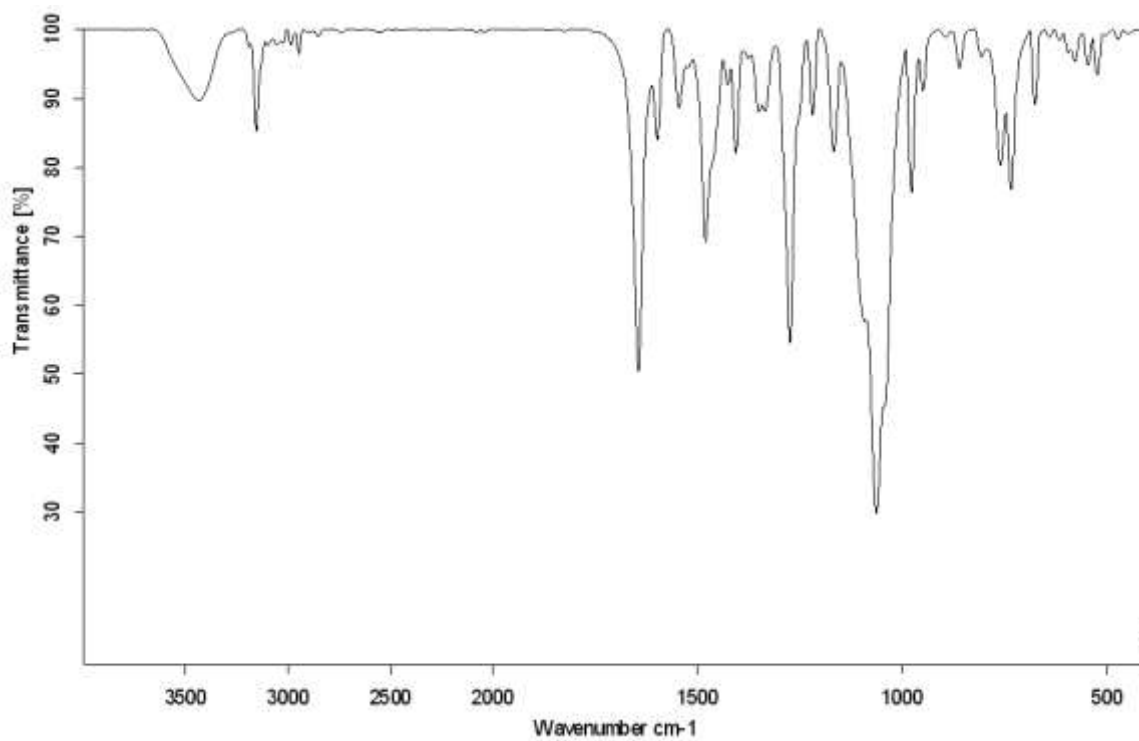
completion, the reaction was quenched by addition of water (10 mL) in an ice bath. After extraction with  $\text{CH}_2\text{Cl}_2$  (3 x 20 mL), the organic phase was washed with 5% sodium carbonate solution (2 x 10 mL), water (2 x 10 mL) and brine (2 x 20 mL), then evaporated to dryness under reduced pressure. The residue was purified by flash column chromatography on a silica gel using petroleum ether (boiling range: 60-90°C) and ethyl acetate as eluents to give the desired product.

*1-(Chloromethyl)-4-ethylbenzene (1a)* [24]: Colorless liquid.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm):  $\delta$ H 7.41 (d,  $J = 8.1$  Hz, 2H), 7.30 (d,  $J = 8.1$  Hz, 2H), 4.67 (s, 2H), 2.68 (s, 3H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 141.8, 131.5, 128.8, 127.9, 46.2, 28.4.

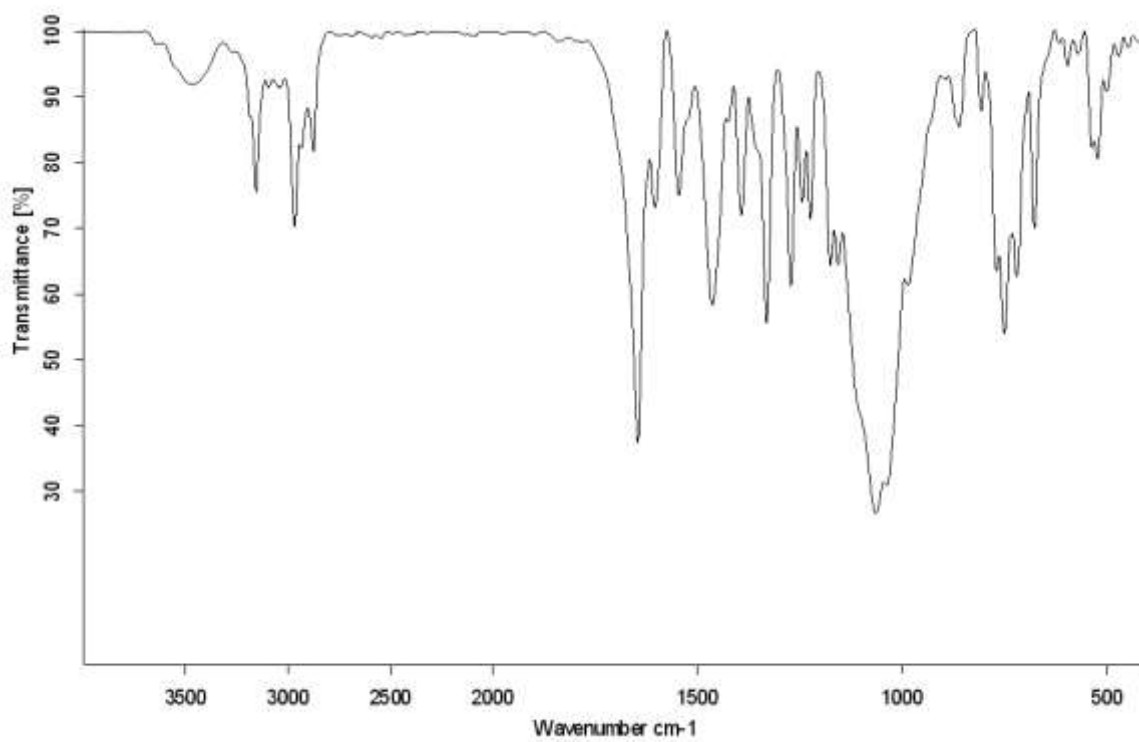
*1-(Chloromethyl)-4-isopropylbenzene (1b)* [24]: Colorless liquid.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 7.41 (d,  $J = 8.1$  Hz, 2H), 7.32 (d,  $J = 8.1$  Hz, 2H), 4.66 (s, 2H), 3.05–2.93 (m, 1H), 1.35 (d,  $J = 7.4$  Hz, 6H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 146.3, 138.7, 128.6, 126.7, 46.0, 33.9, 23.9.

*1-(tert-Butyl)-4-(chloromethyl)benzene (1c)* [24]: Colorless liquid.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm):  $\delta$ H 7.46 (d,  $J = 8.4$  Hz, 2H), 7.42–7.39 (m, 2H), 4.64 (s, 2H), 1.40 (s, 9H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 151.4, 134.5, 128.3, 125.6, 46.0, 34.5, 31.3.

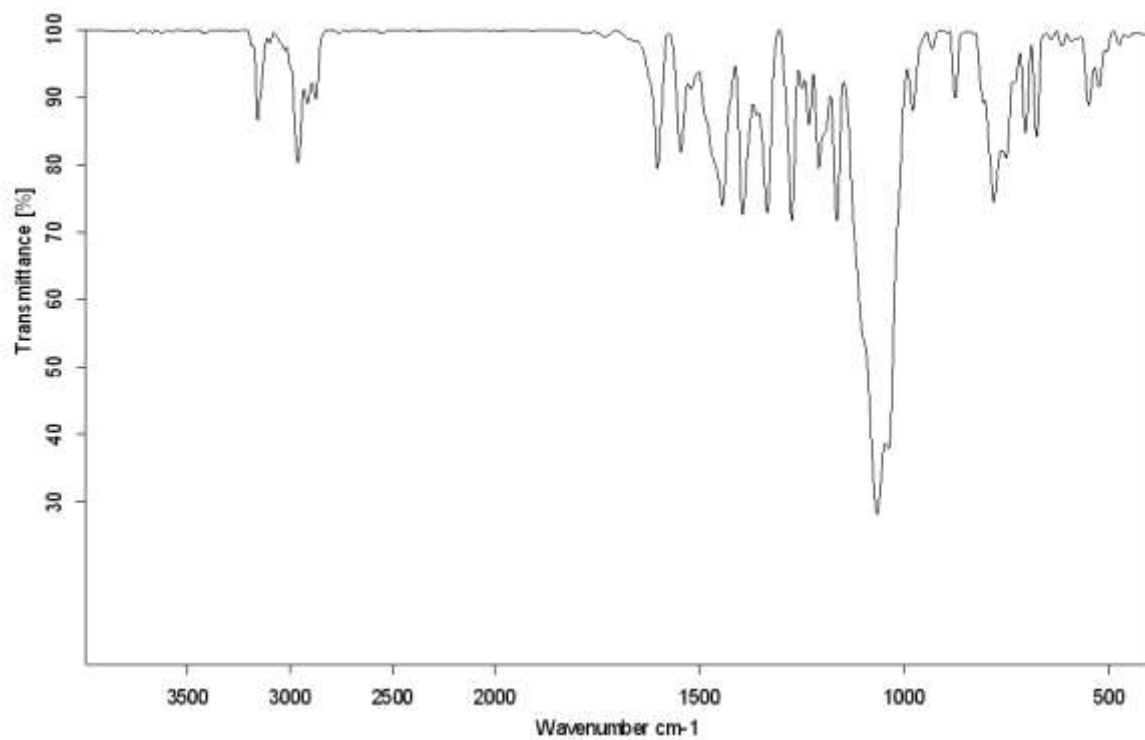
### 3. Figures



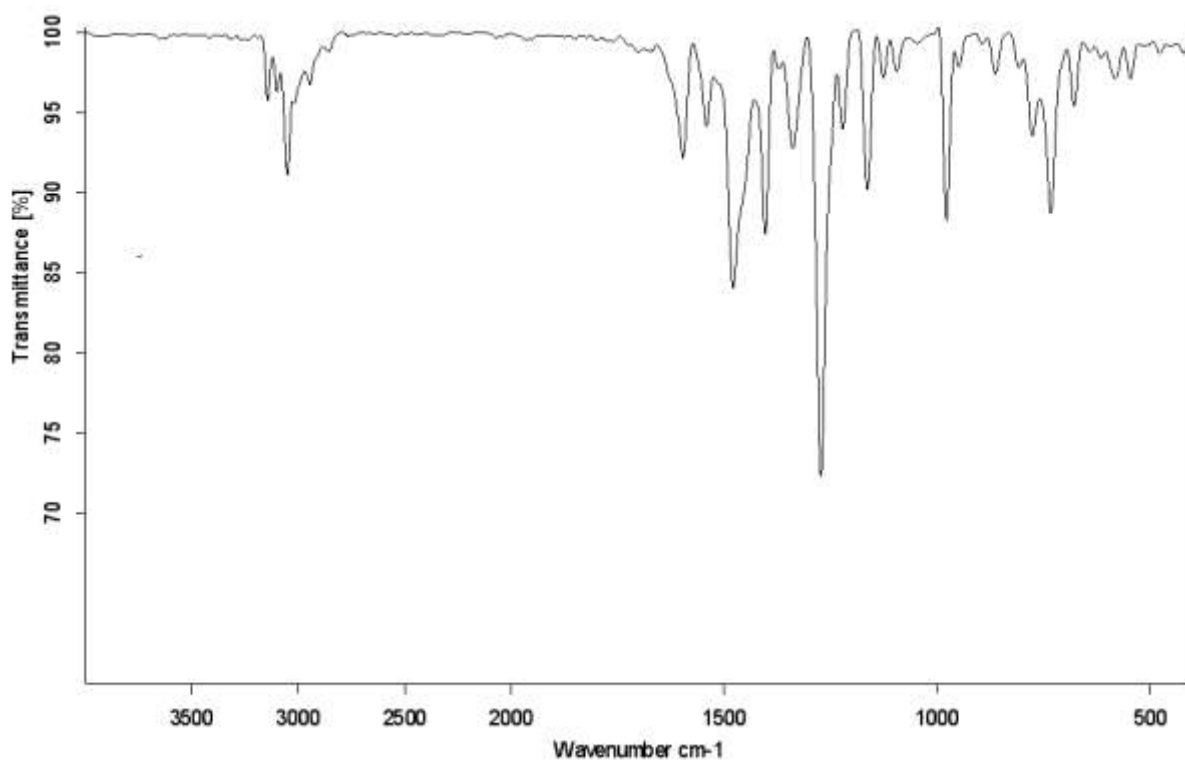
**Fig. S1:** FTIR spectrum of (3a)



**Fig. S2:** FTIR spectrum of (3b)



**Fig. S3:** FTIR spectrum of (3c)



**Fig. S4:** FTIR spectrum of (4a)

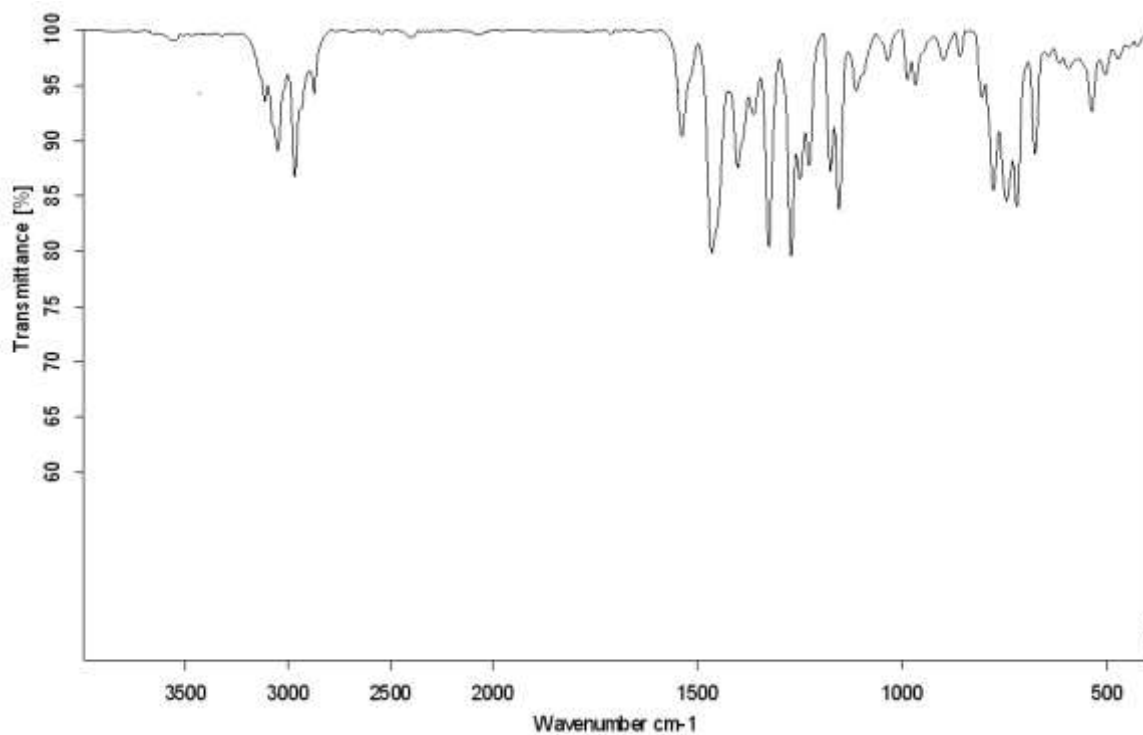


Fig. S5: FTIR spectrum of (4b)

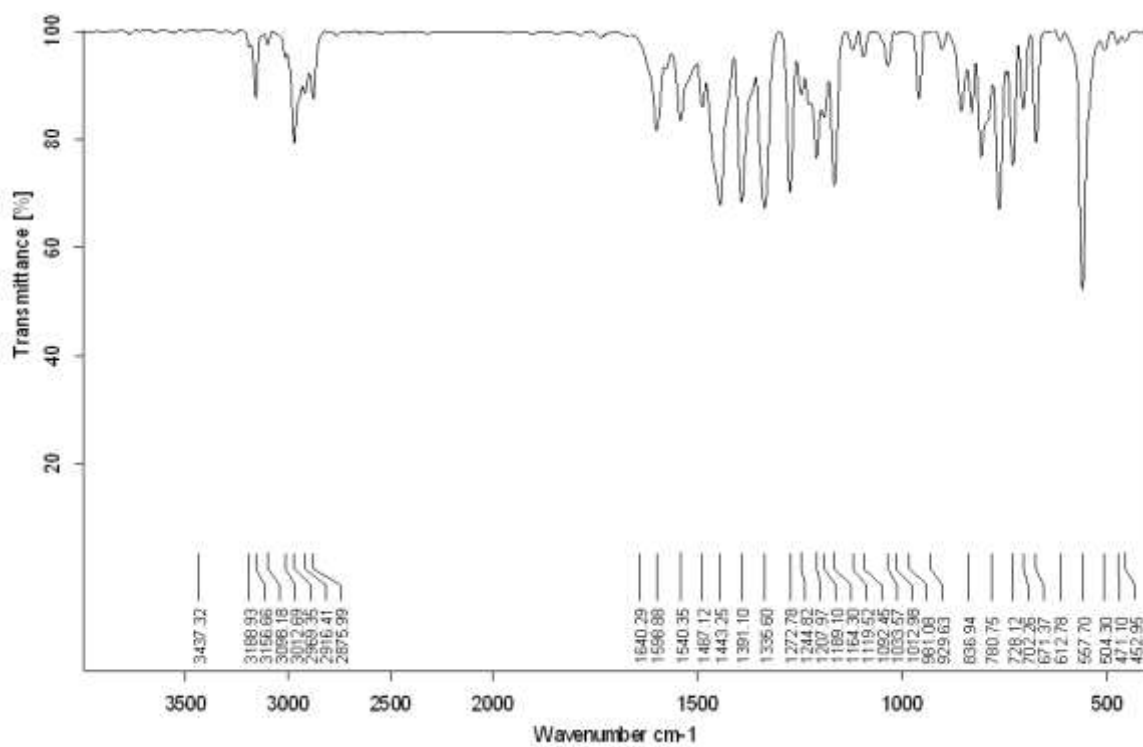


Fig. S6: FTIR spectrum of (4c)

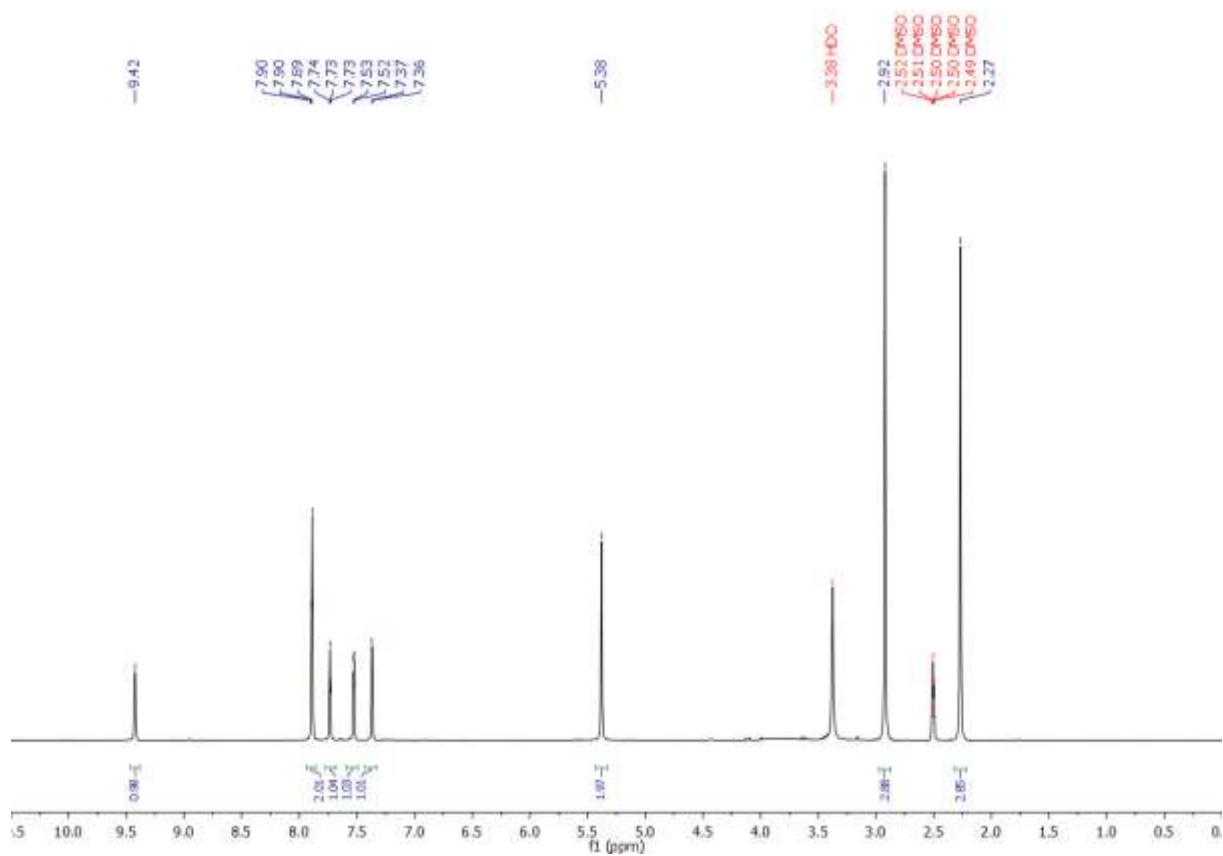


Fig. S7:  $^1\text{H}$  NMR of **2a** (200 MHz,  $\text{DMSO-}d_6$ )

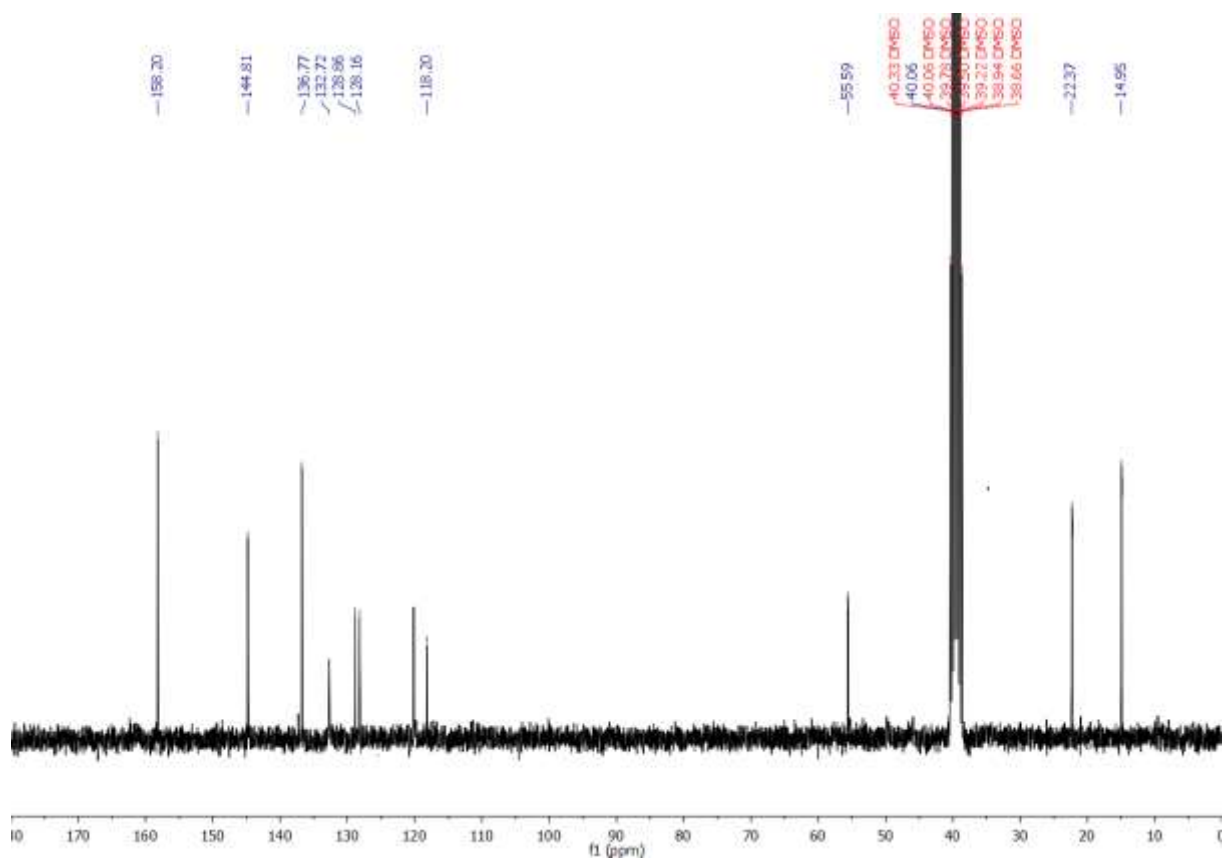


Fig. S8:  $^{13}\text{C}$  NMR of **2a** (125 MHz,  $\text{DMSO-}d_6$ )

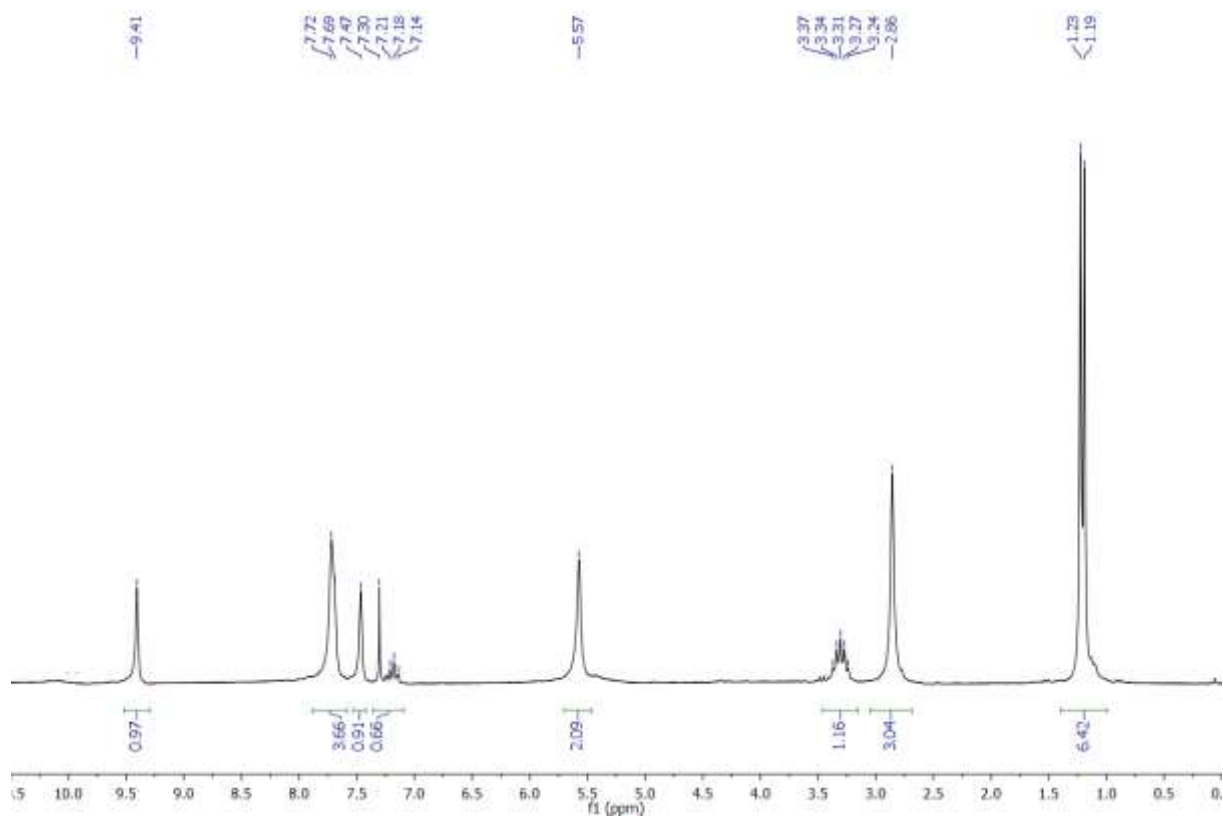


Fig. S9:  $^1\text{H}$  NMR of **2b** (200 MHz,  $\text{CDCl}_3$ )

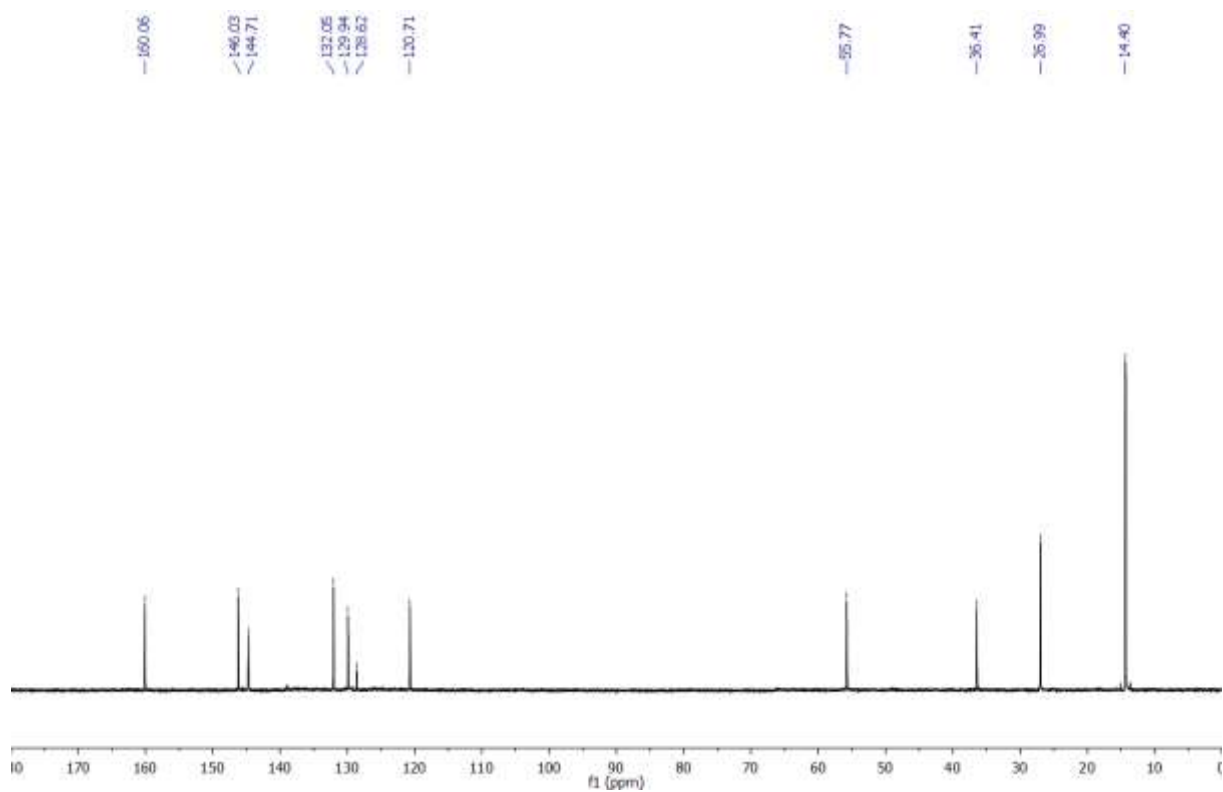


Fig. S10:  $^{13}\text{C}$  NMR of **2b** (125 MHz,  $\text{CDCl}_3$ )



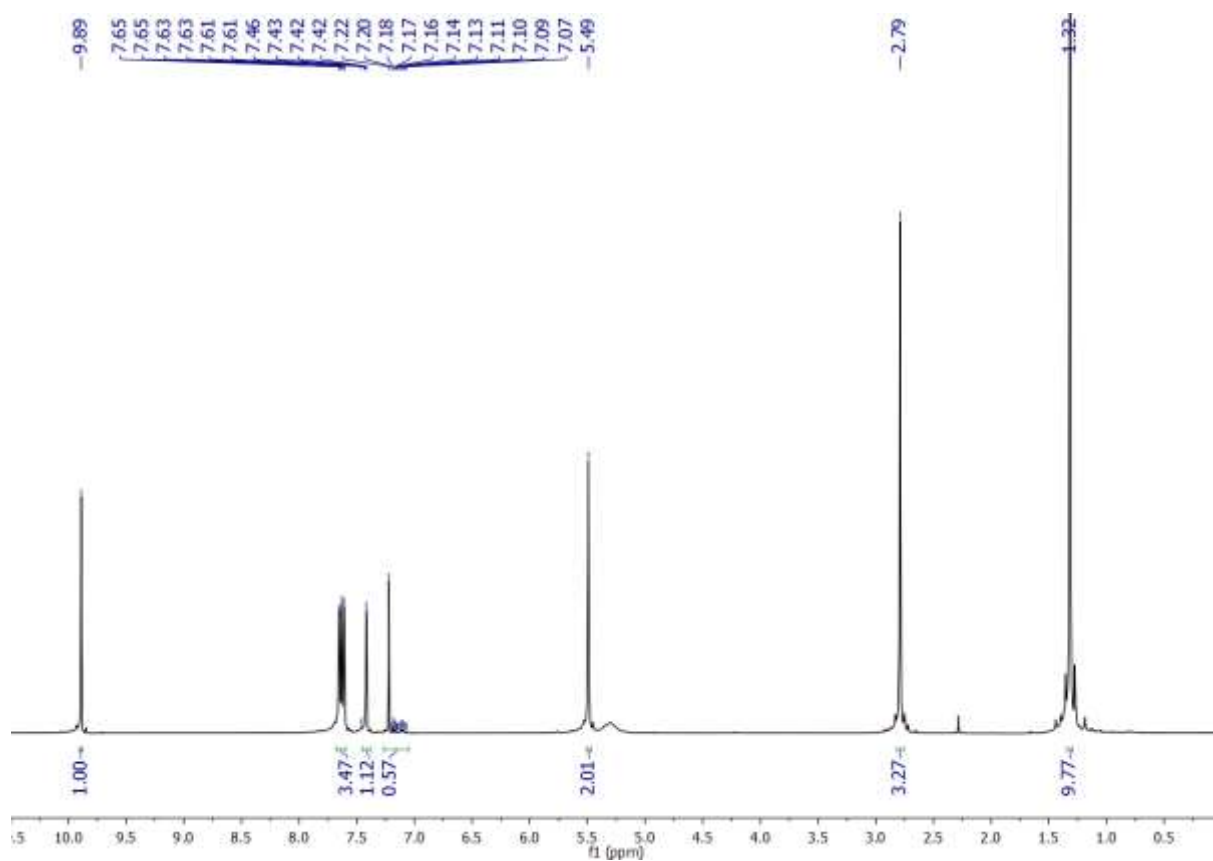


Fig. S11:  $^1\text{H}$  NMR of **2c** (200 MHz,  $\text{CDCl}_3$ )

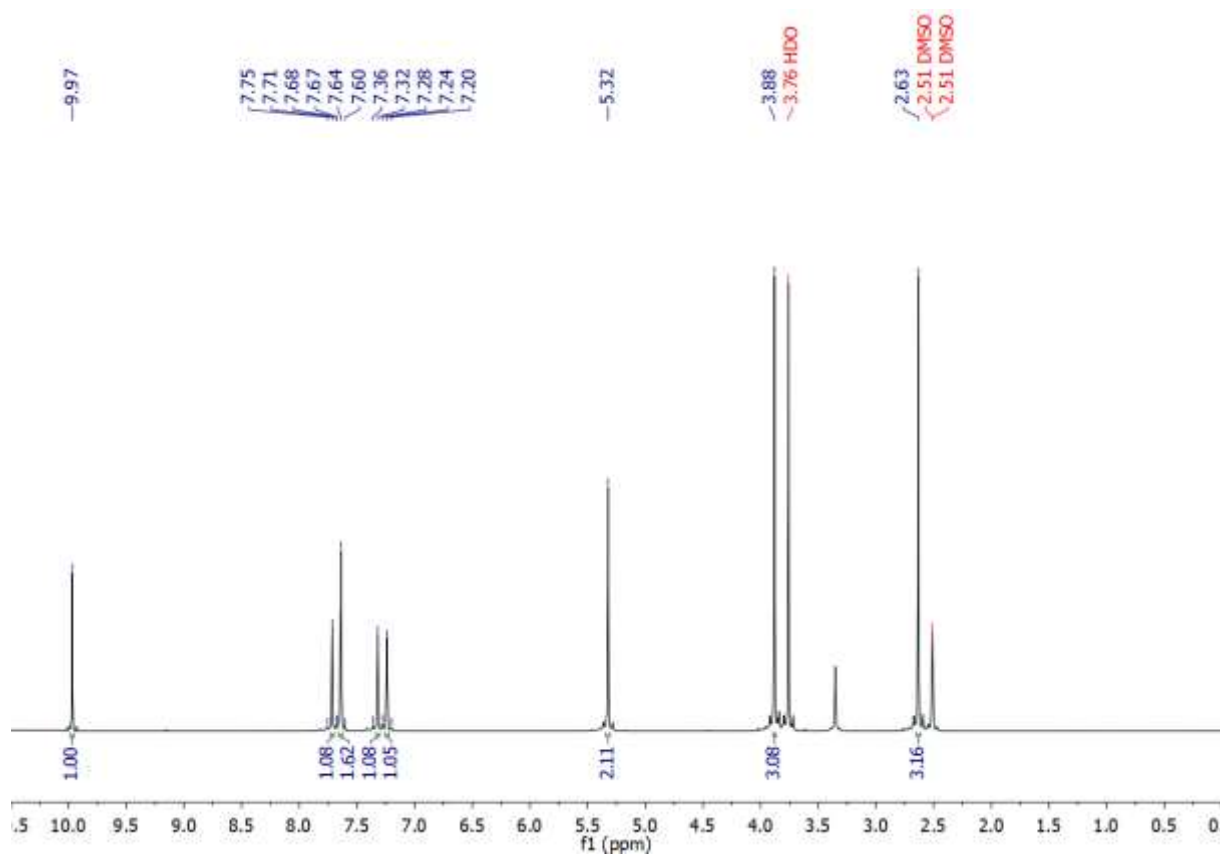


Fig. S12:  $^1\text{H}$  NMR of **3a** (200 MHz,  $\text{DMSO}-d_6$ )

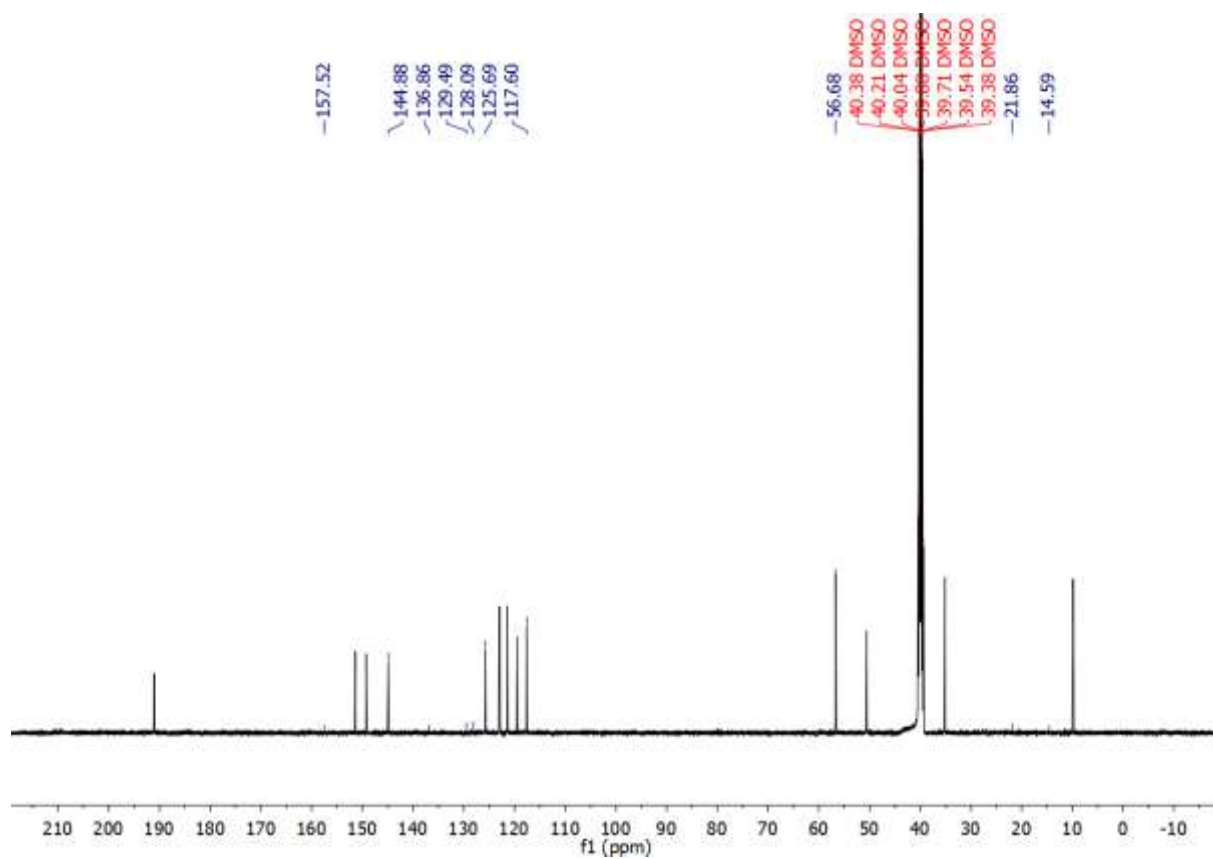


Fig. S13:  $^{13}\text{C}$  NMR of **3a** (125 MHz,  $\text{DMSO-}d_6$ )

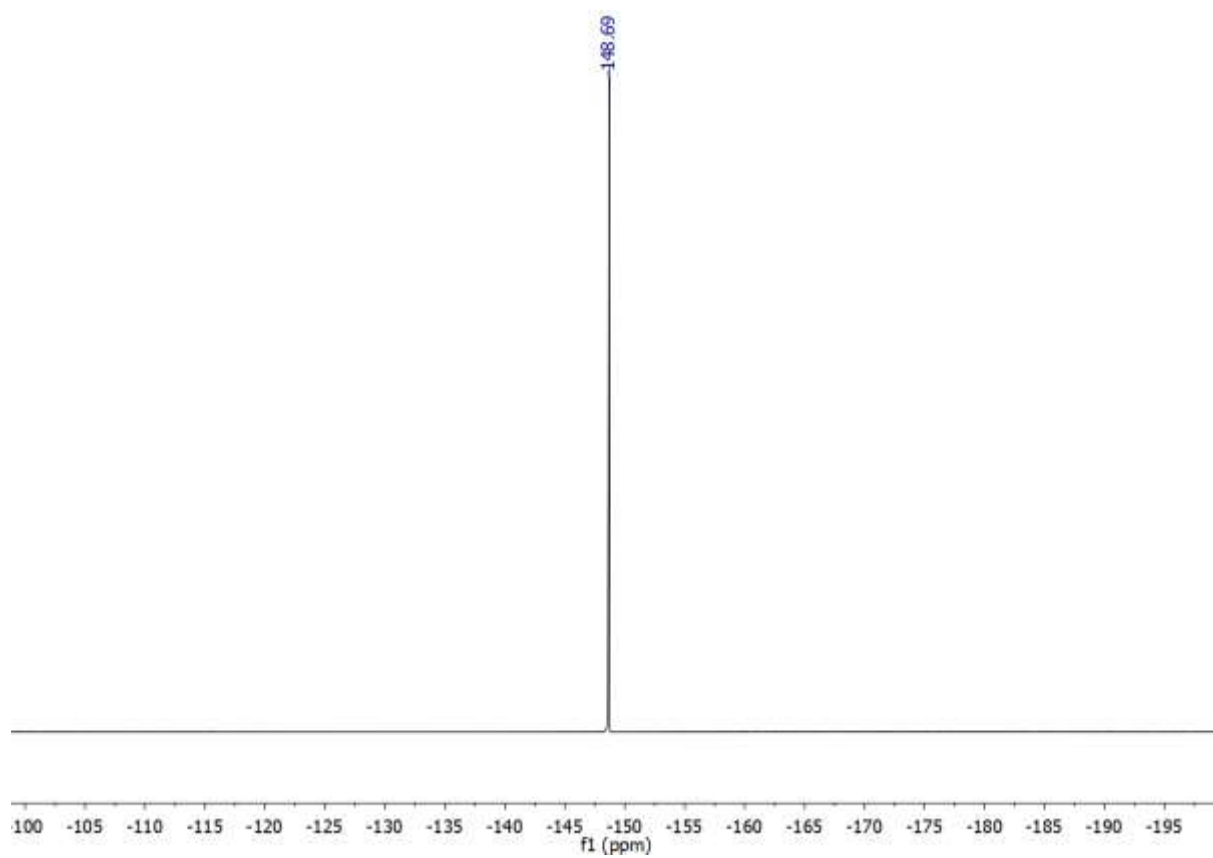


Fig. S14:  $^{19}\text{F}$  NMR of **3a** (471 MHz,  $\text{DMSO-}d_6$ )

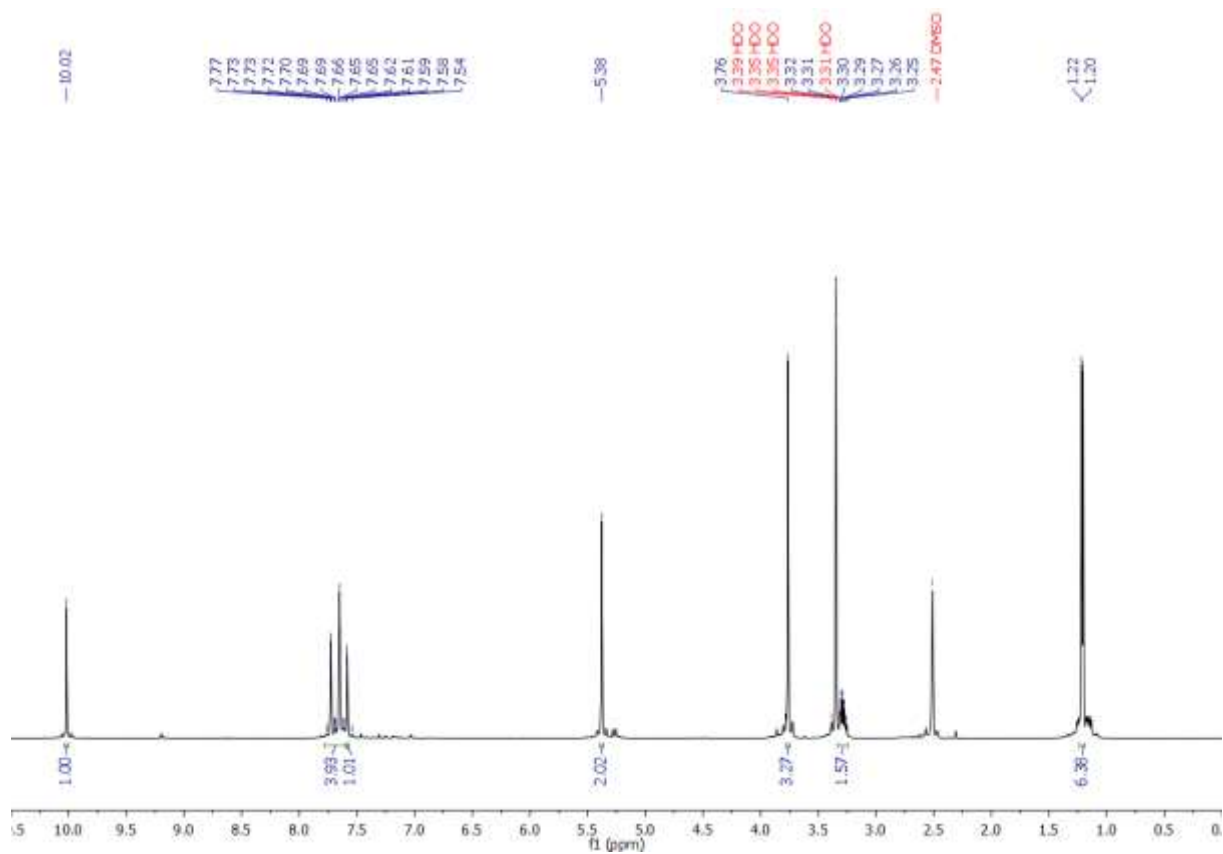


Fig. S15:  $^1\text{H}$  NMR of **3b** (200 MHz,  $\text{DMSO-}d_6$ )

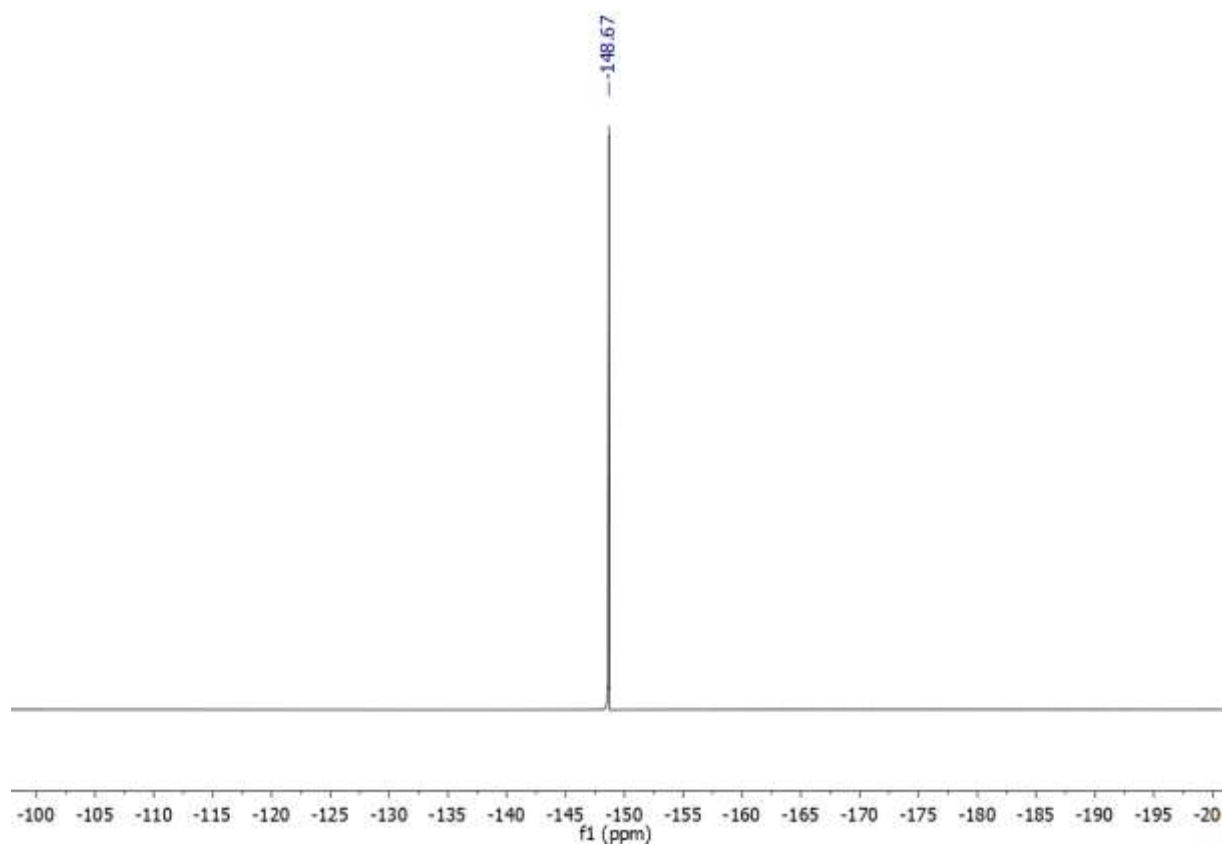


Fig. S16:  $^{19}\text{F}$  NMR of **3b** (471 MHz,  $\text{DMSO-}d_6$ )

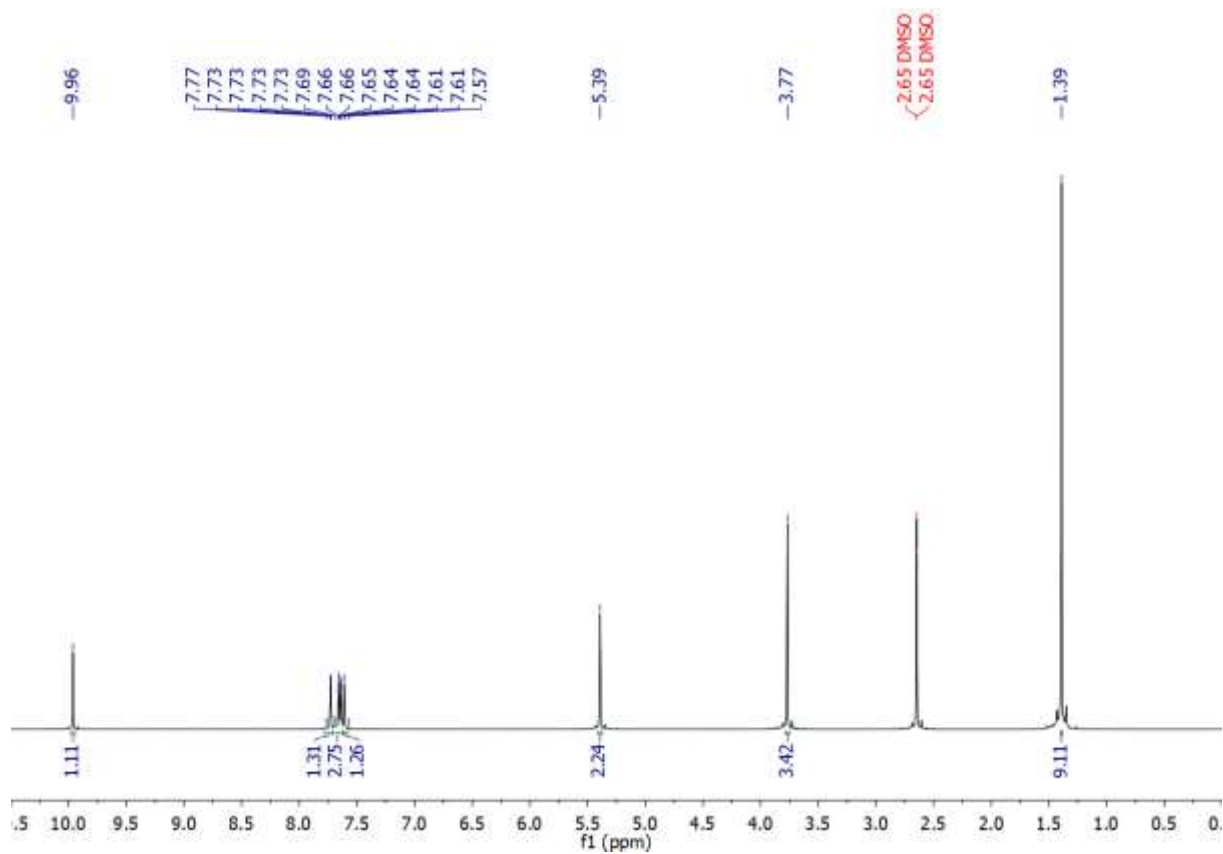


Fig. S17:  $^1\text{H}$  NMR of **3c** (200 MHz,  $\text{DMSO-}d_6$ )

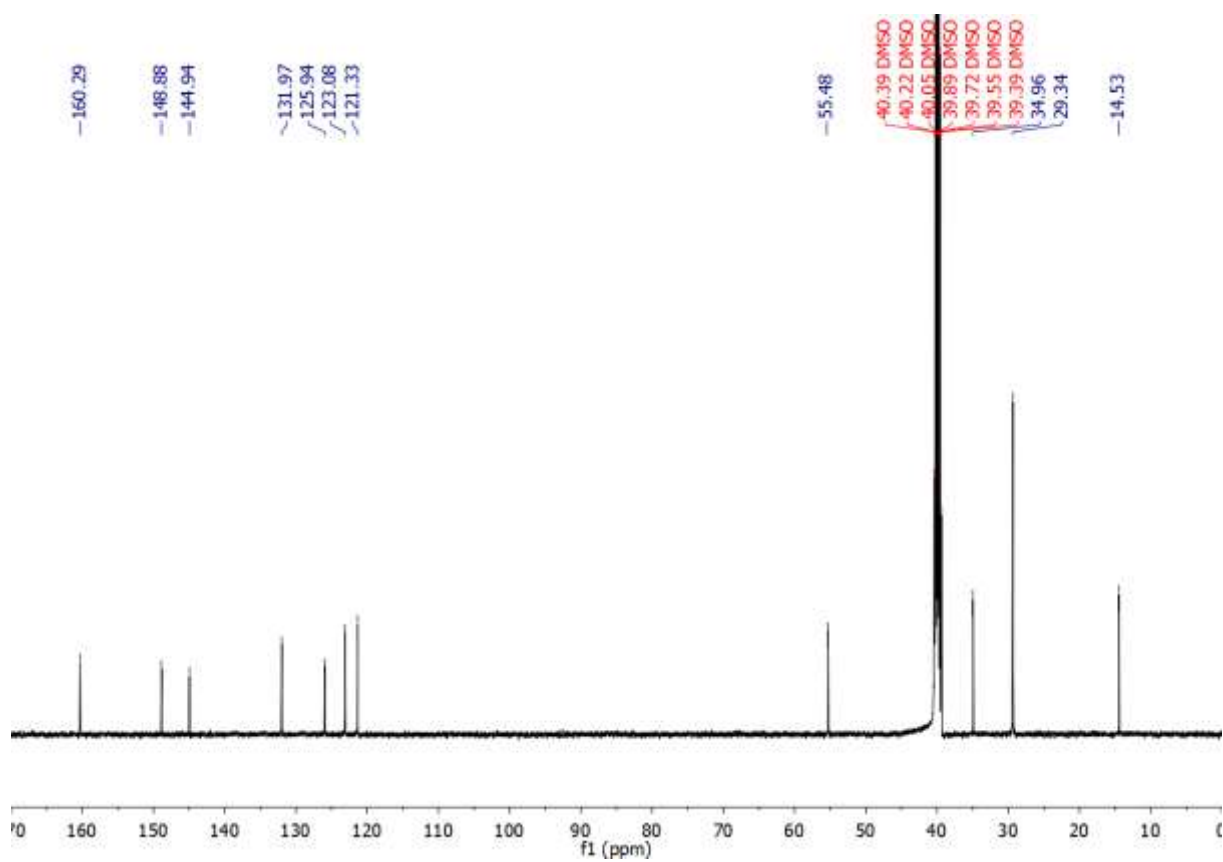


Fig. S18:  $^{13}\text{C}$  NMR of **3c** (125 MHz,  $\text{DMSO-}d_6$ )

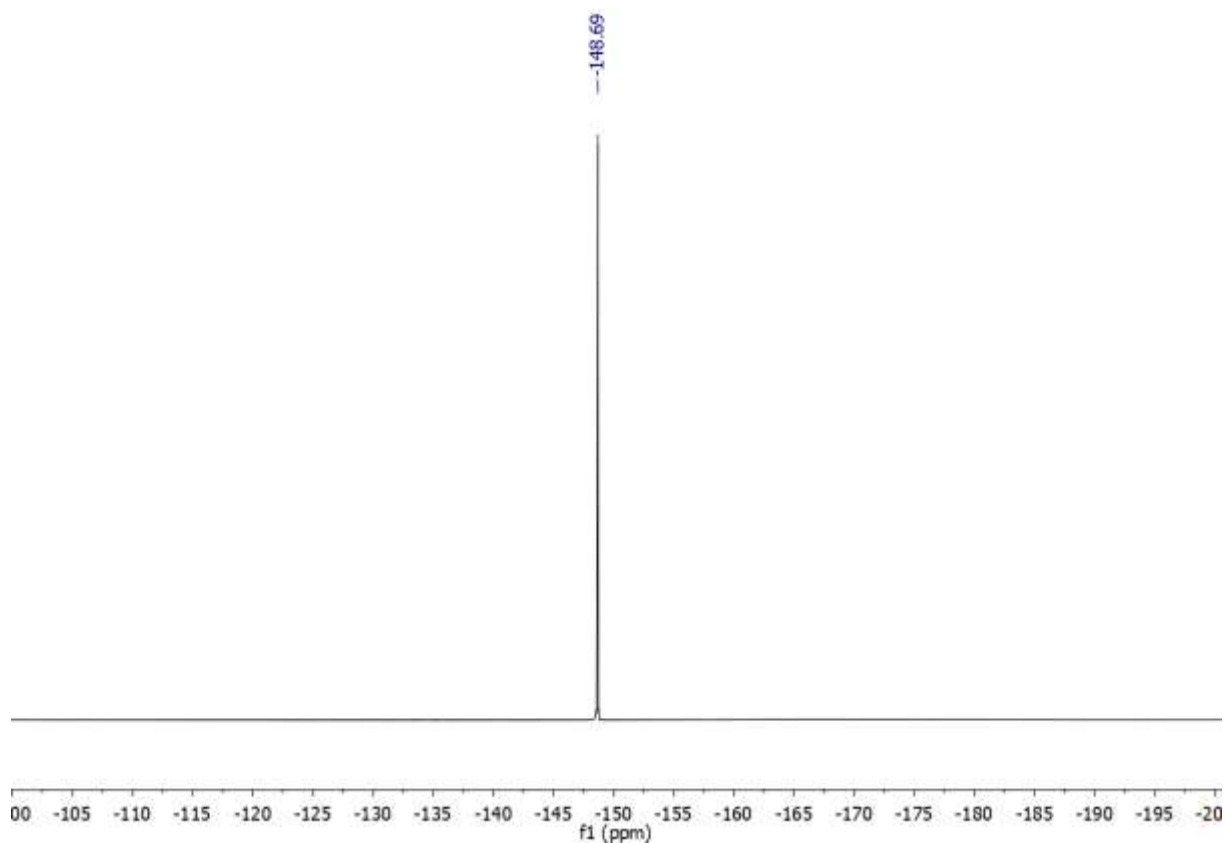


Fig. S19:  $^{19}\text{F}$  NMR of **3c** (471 MHz,  $\text{DMSO-}d_6$ )

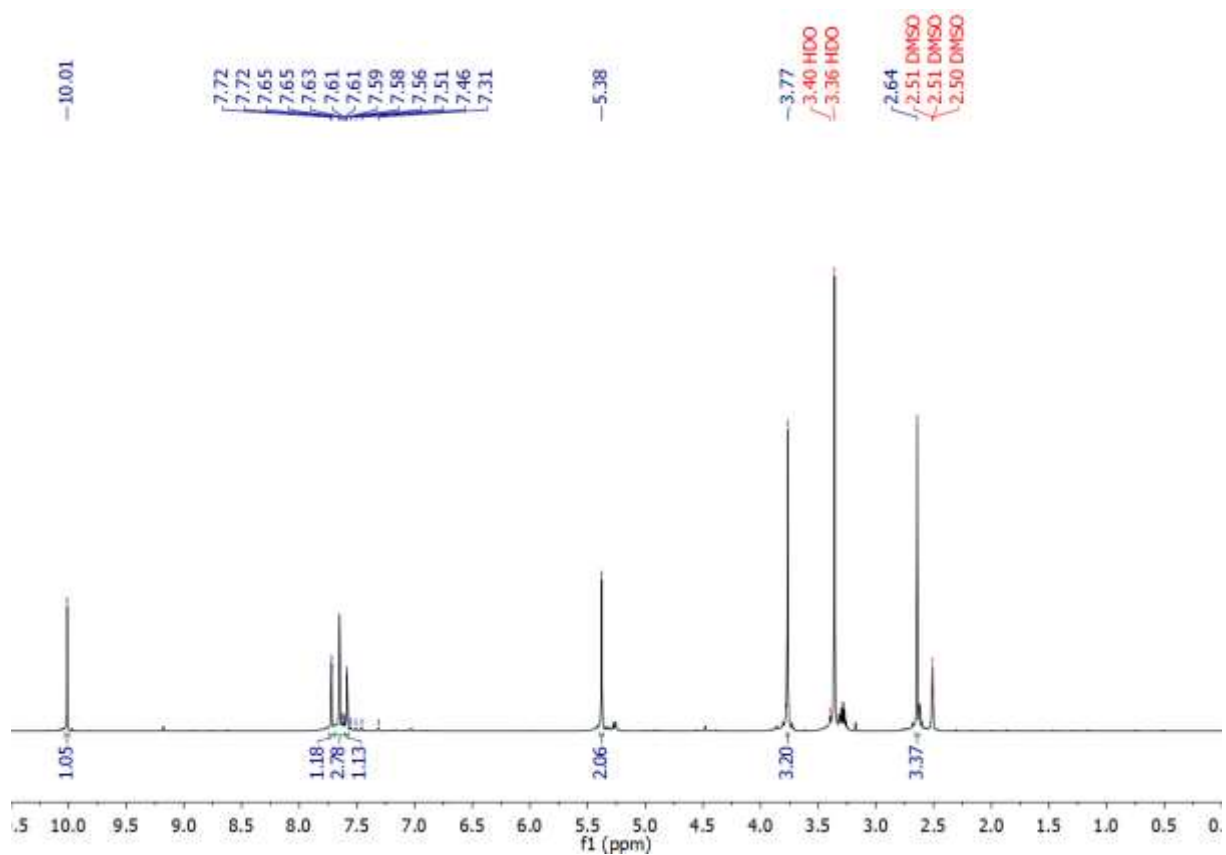


Fig. S20:  $^1\text{H}$  NMR of **4a** (200 MHz,  $\text{DMSO-}d_6$ )

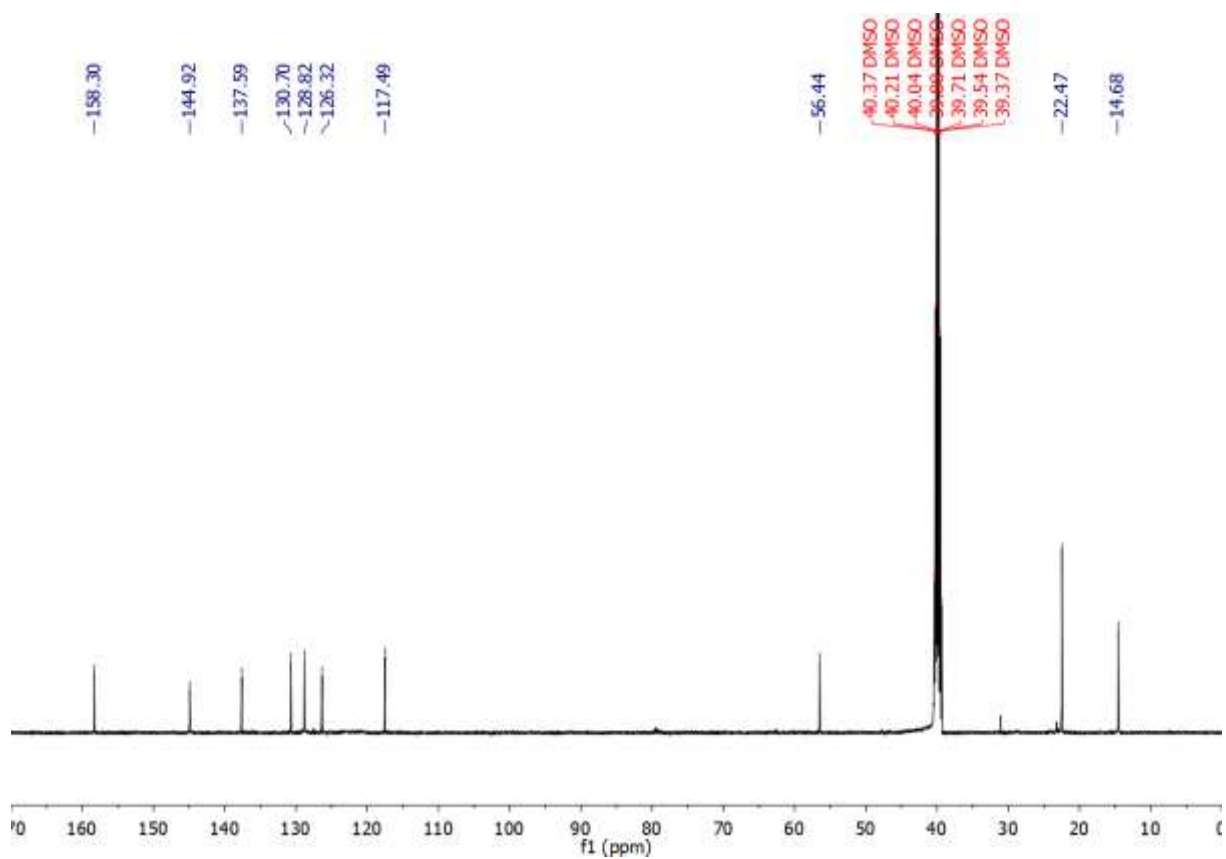


Fig. S21:  $^{13}\text{C}$  NMR of **4a** (125 MHz,  $\text{DMSO-}d_6$ )

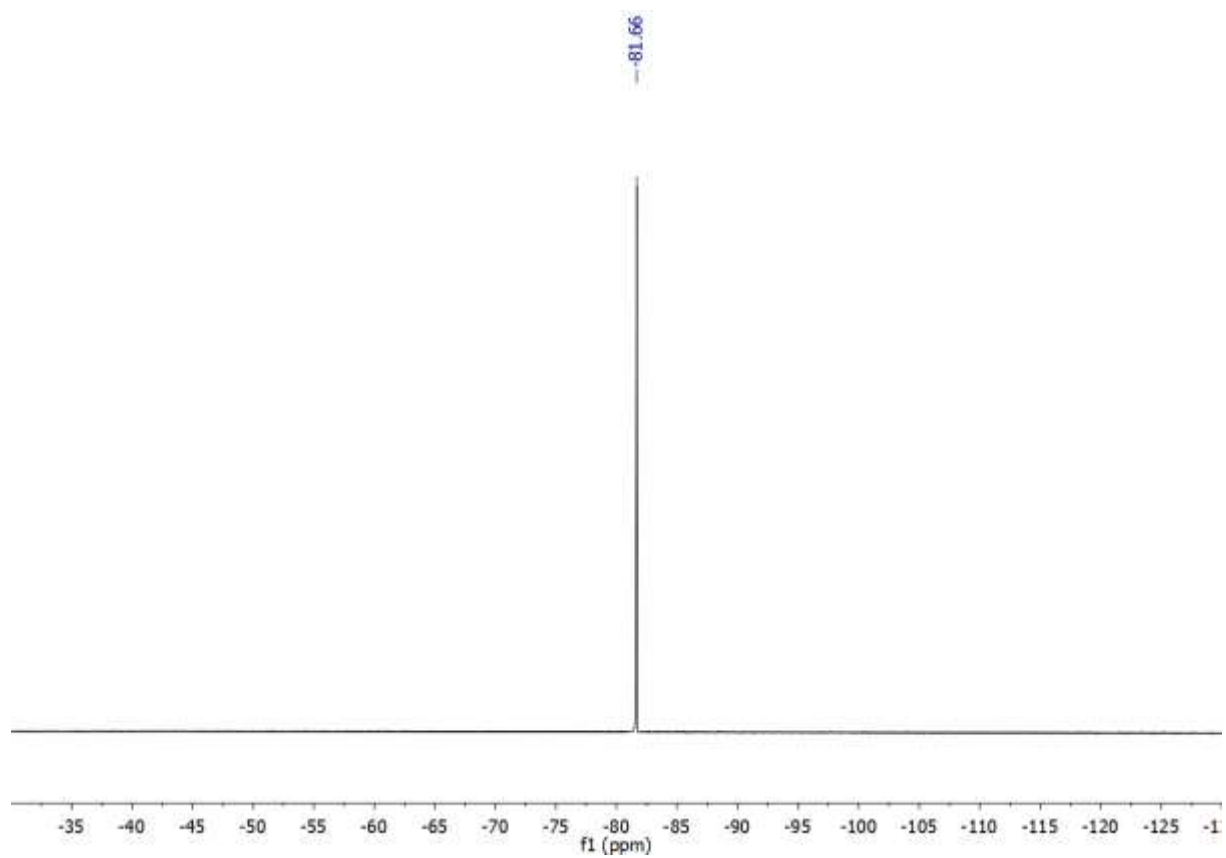


Fig. S22:  $^{19}\text{F}$  NMR of **4a** (565 MHz,  $\text{DMSO-}d_6$ )

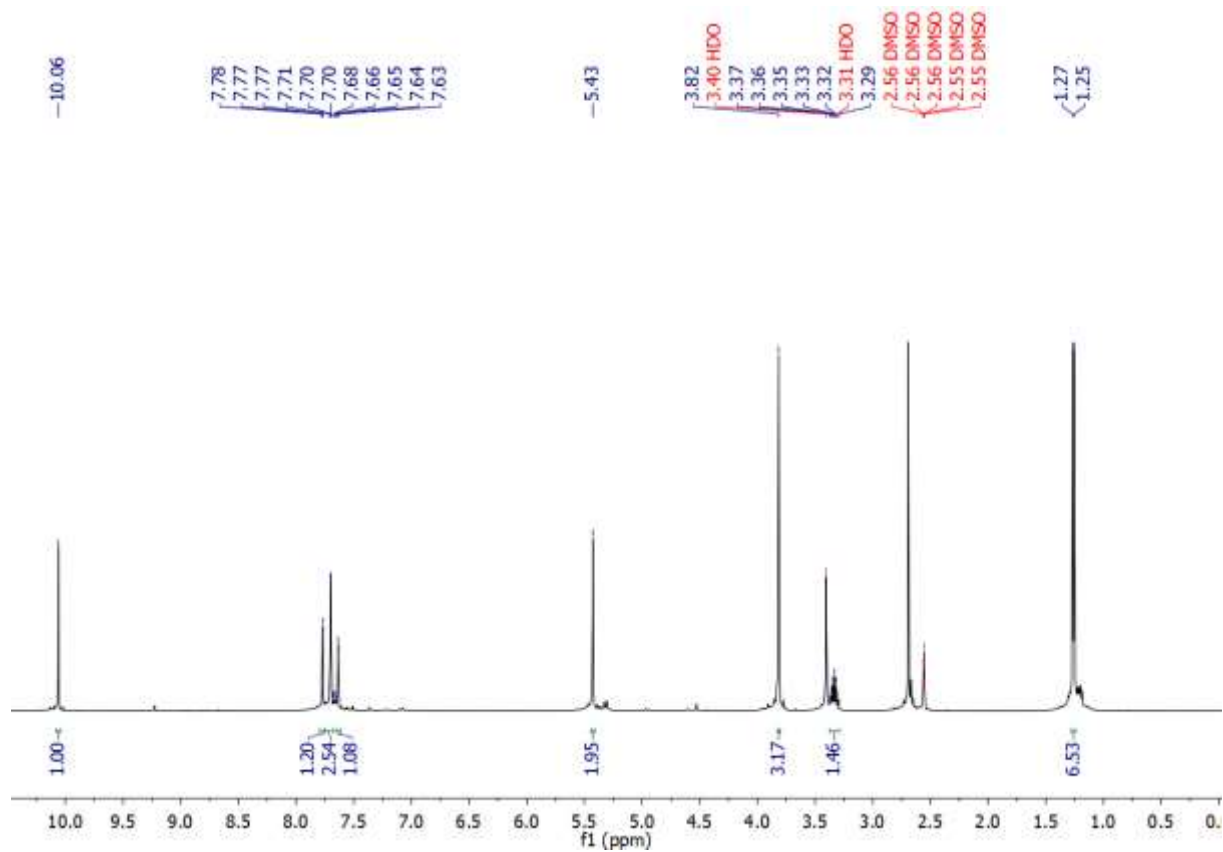


Fig. S23: <sup>1</sup>H NMR of **4b** (200 MHz, DMSO-*d*<sub>6</sub>)

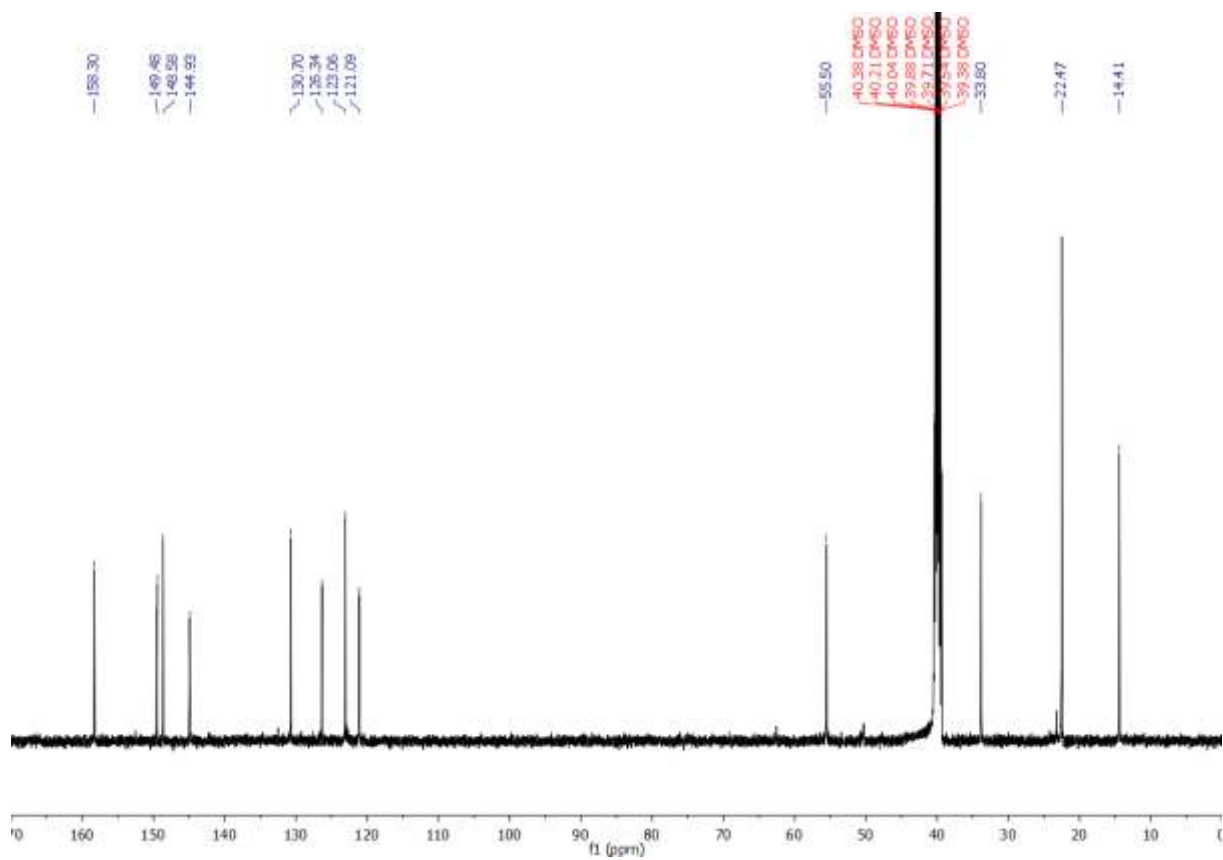


Fig. S24: <sup>13</sup>C NMR of **4b** (125 MHz, DMSO-*d*<sub>6</sub>)

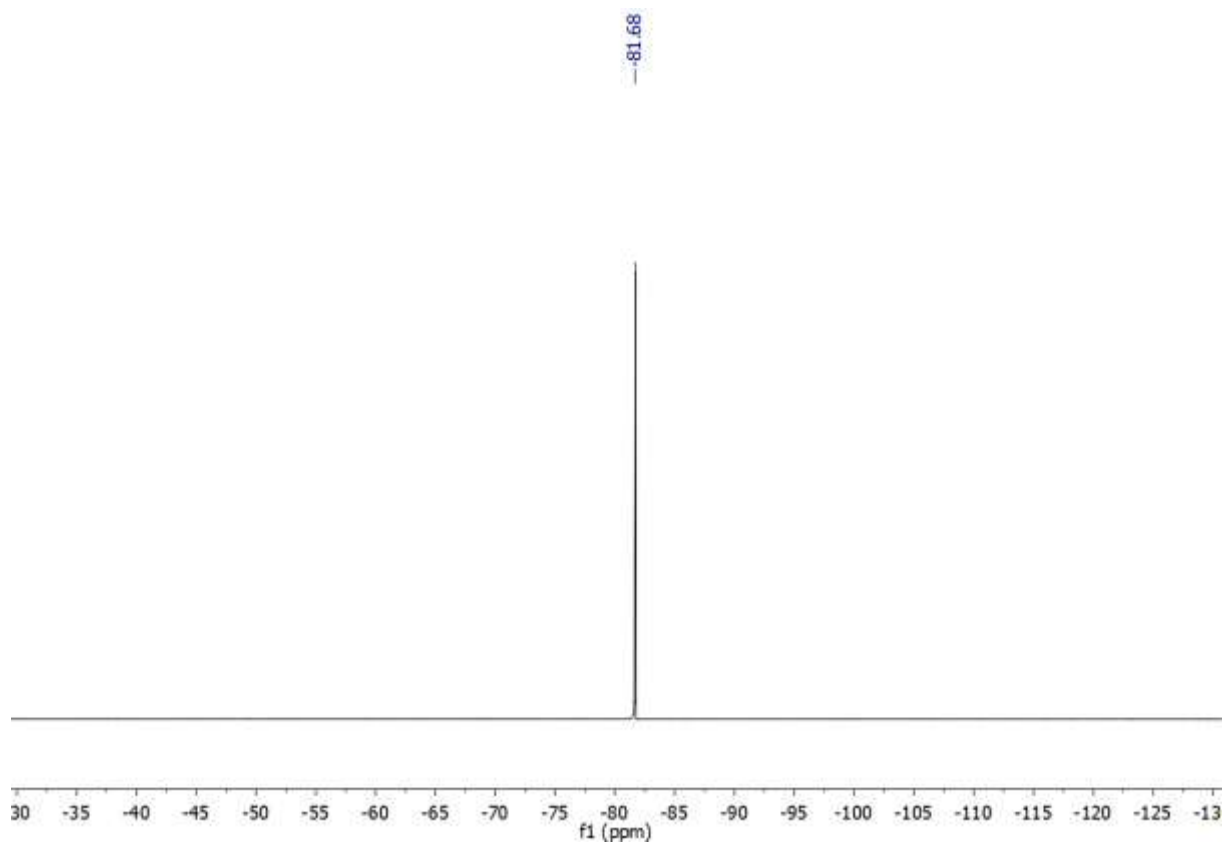


Fig. S25:  $^{19}\text{F}$  NMR of **4b** (565 MHz,  $\text{DMSO-}d_6$ )

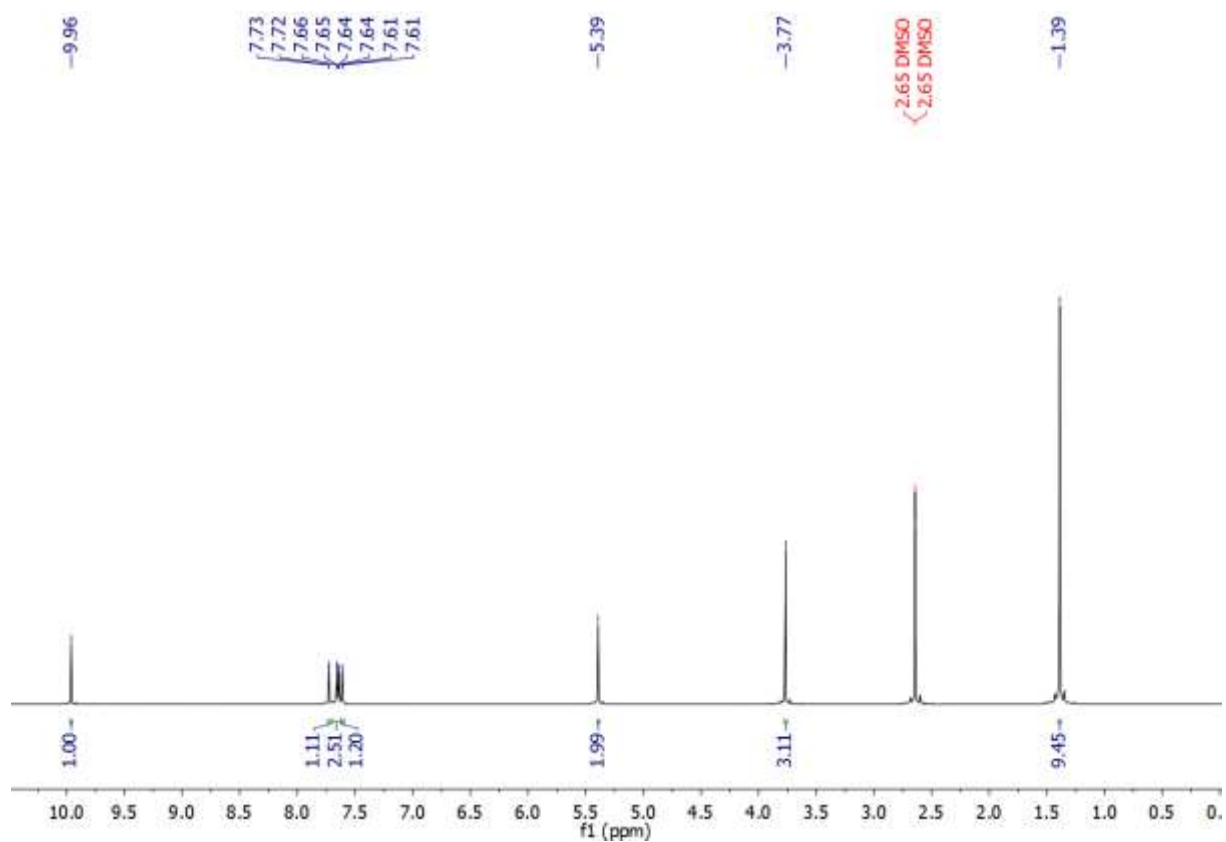


Fig. S26:  $^1\text{H}$  NMR of **4c** (200 MHz,  $\text{DMSO-}d_6$ )



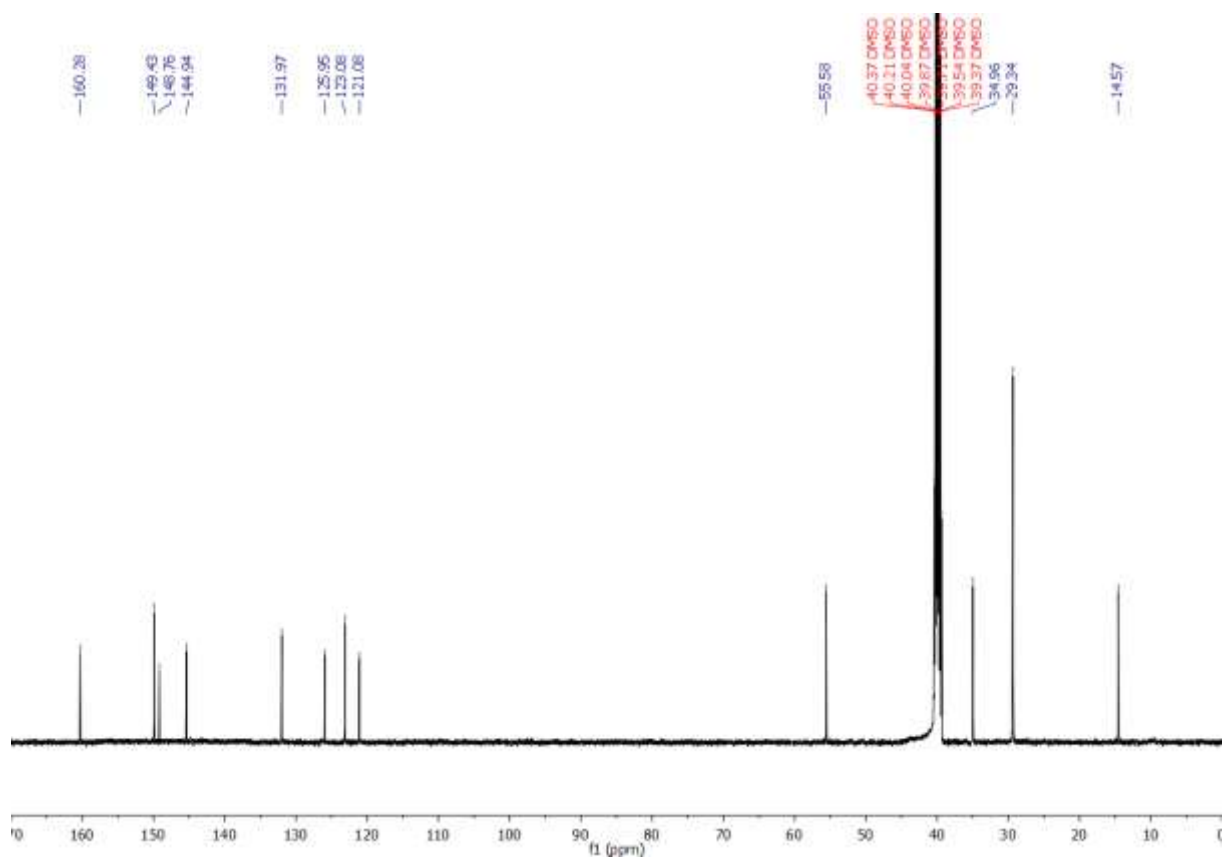


Fig. S27:  $^{13}\text{C}$  NMR of **4c** (125 MHz, DMSO- $d_6$ )

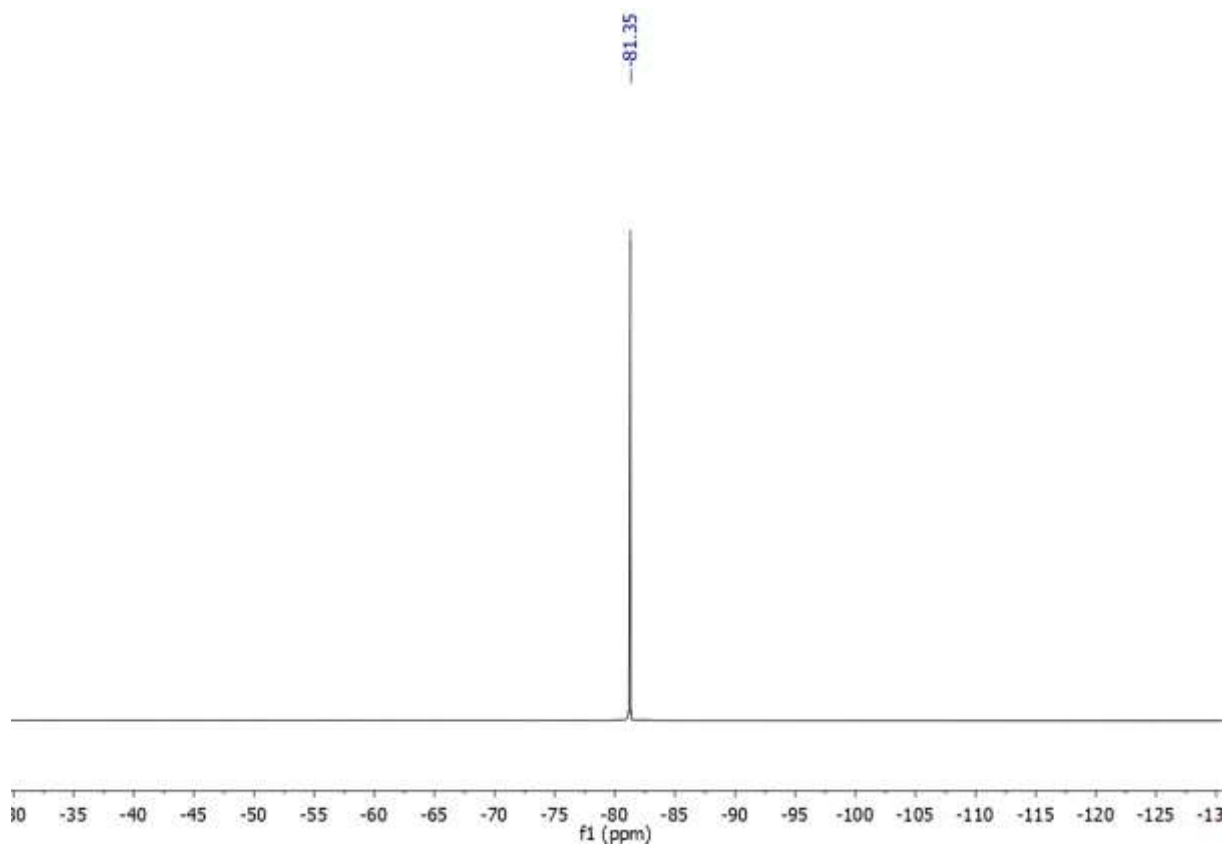
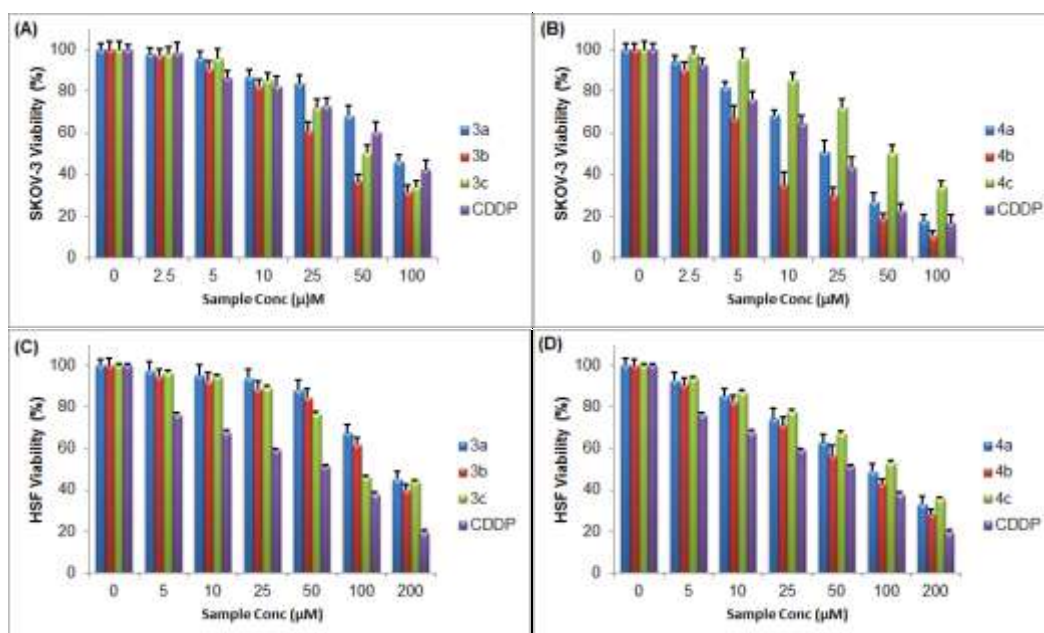


Fig. S28:  $^{19}\text{F}$  NMR of **4b** (565 MHz, DMSO- $d_6$ )



**Fig. S29:** The effect of different doses (0 – 200 μM) of TILs (**3a-c**) and (**4a-c**) on the viability of (**A,B**) ovarian carcinoma cell lines (SKOV-3) and (**C,D**) human skin fibroblast (HSF) cells after 24 h of treatment as compared to the clinical antitumor drug (cisplatin, CDDP).