## **Supplementary Information**

# Acetonitrile and Benzonitrile as Versatile Amino Sources in the Copper-Catalyzed Mild C-H Amidation Electrochemical Reactions

Sofia Strekalova\*,<sup>a</sup> Alexander Kononov,<sup>a,b</sup> Ildar Rizvanov<sup>a</sup> and Yulia Budnikova\*<sup>a,b</sup>

<sup>*a*</sup>Arbuzov Institute of Organic and Physical Chemistry, FRC Kazan Scientific Center of RAS, Kazan, 420088 Russian Federation

<sup>b</sup>Kazan National Research Technological University, Kazan, 420015 Russian Federation

E-mail: strekalova@iopc.ru, yulia@iopc.ru

#### **Contents of Supplementary Information:**

1. General Information	S-2
2. Materials	S-2
3. NMR and Mass measurements	S-2
4. Synthesis	S-3
4.1 Electrolysis setup	S-3
4.2 General electrolysis procedure	S-3
5. Characterization of the substrate and products	S-7
6. Electrochemical potentials	S-13
7. NMR Spectra	S-14

#### 1. General Information

Preparative electrolysis was performed in galvanostatic mode with simultaneous control of the working electrode potential in thermostatically controlled, cylindrical 40 mL cell with separate anodic and cathodic compartments using a B5-49 direct current source at a current of 40 mA. A ceramic plate with the pore size 900 nm was used as a membrane. The potential of the working electrode was detected by a V7-27 DC voltmeter. The platinum cylindrical cathode with a surface area of 20 cm<sup>2</sup> used as a working electrode and the platinum rod was used as the anode. The working electrode potential was determined using reference electrode Ag/AgNO<sub>3</sub> in CH<sub>3</sub>CN. The catholyte was a saturated solution of the PyHBF<sub>4</sub> in the CH<sub>3</sub>CN. During electrolysis, the electrolyte was stirred using a magnetic stirrer. Preparative chromatography BUCHI system including Pump Module C-601, ELS Detector C-605, Fraction Collector C-660, Control Unit C-620, Sepacore Control Package and software program were used for products isolation.

## 2. Materials

Toluene (99%), o-Xylene (99%), m-Xylene (99%), Naphthalene (99%), 2-Phenylpyridine (97%), Copper(II) acetate (anhydrous, 98%) were purchased from Alfa Aesar and used without preliminary purification.

Benzene (anhydrous, 99.8%), 2-Bromotoluene (99%), Mesitylene (98%), 2-Bromomesitylene (98%), 2,6-Lutidine (ReagentPlus<sup>®</sup>, 98%), 4-Bromoanisole ( $\geq$ 99.0%), Anisole (anhydrous, 99.7%), Benzonitrile (99+%) were purchased from Sigma Aldrich and used without preliminary purification.

Acetonitrile (CHROMASOLV<sup>®</sup> Plus,  $\geq$ 99.9%, «extra pure» by Acros Organics) was used without preliminary purification.

## 3. NMR and Mass measurements

NMR measurements were performed in the NMR department (A.E. Arbuzov Institute Organic and Physical Chemistry) of the Federal Collective Spectral Analysis Center for physical and chemical studies on the structure, properties, and composition of matter and materials. NMR experiments were conducted using Bruker spectrometers AVANCE-400 (399.93 MHz (<sup>1</sup>H), 100.6 MHz (<sup>13</sup>C) and AVANCE-600 (600.1 MHz (<sup>1</sup>H) 150.9 MHz (<sup>13</sup>C) equipped with a pulsed gradient unit capable of producing magnetic field pulse gradients in the z-direction of 53.5 G cm<sup>-1</sup>. All spectra were acquired in a 5 mm gradient inverse broad band probe head. As a result, chemical shifts were reported on the  $\delta$  (ppm) scale relative to the residual <sup>1</sup>H and <sup>13</sup>C C<sub>6</sub>D<sub>6</sub> signal resulting in external NMR spectra.

Mass spectrometric studies (electrospray ionization) of the samples was carried out on a Bruker Daltonik GmbH AmazonX (Germany). Nitrogen was used as a drying gas in the source with a temperature of 220°C. The source voltage was 4.5 kV. Solutions of the samples were diluted with acetonitrile to a concentration of ~10–3 mg mL<sup>-1</sup>. The samples were injected using an autosampler of an Agilent 1260 Infinity liquid chromatograph (Agilent Technologies, USA).

GC–MS analysis of the samples was carried out on an Agilent 6890N/5973 gas chromatograph (Agilent Technologies, USA) with a capillary column (30 mm×0.25 mm, film thickness 0.25  $\mu$ m) equipped with an Agilent 6890N-5973 mass selective detector in the electron impact mode. The GC was operated under the following conditions: manual injection 1  $\mu$ l; injector temperature, 300 °C; carrier gas (He) flow 1 ml/min, split – 1:5; oven temperature programmed 100–300 °C at a rate of 10 °C/min. The detector temperature was 230 °C. The MS was operated under 70 eV, scan range 29–800 amu.

#### 4. Synthesis

## 4.1 Electrolysis setup



**Figure S1.** Electrolysis setup: cylindrical cell with divided anodic and cathodic compartments. The ceramic plate with the pore size of 900 nm is a membrane, the cylindrical platinum with a surface area of 20 cm<sup>2</sup> is cathode and the platinum rod is anode. The cell can be thermostatted.

## 4.2 General electrolysis procedure

The substrate (1.0 mmol, 0.1 g), Copper acetate (0.1 mmol, 0.024 g), R-CN (40 mL) as solvent, were placed into electrochemical cell at 23°C. The electrolysis time is 2 to 4 hours (2F or 4F electricity). At the end of electrolysis the reaction mixture was evaporated on rotary evaporator. The residue was purified through passing through chromatographic column with silica gel (hexane-ethyl acetate). The product yields are given in Table S1.

Entry	Substrate	R-CN	Product	Yield <sup>a</sup> , %
la		MeCN		70
2a,b	Br	MeCN	Br H a b	68 (1:3 a:b)
3a	Br	MeCN		69
4a,b	CF <sub>3</sub>	MeCN	b CF <sub>3</sub> B b	76 (3:1 a:b)
5a		MeCN	<b>K</b>	78
6a		MeCN	<b>I</b>	69
7a		MeCN		68
8a		MeCN	T T	72
9a	Br	MeCN	Br	56

Table S1. Yield of the obtained acetamides. **a** Isolated. **b** by NMR

10a		MeCN	↓ N	62
11a	Br	MeCN	Br	51
12a		MeCN		trace <sup>b</sup>
13a		PhCN		60
14a,b	Br	PhCN	Br H a b	56 (1:3 a:b)
15a	Br	PhCN		59
16a,b	CF <sub>3</sub>	PhCN	CF <sub>3</sub> H b b	64 (3:1 a:b)
17a		PhCN		69
18a	Br	PhCN	Br N	48
19a		PhCN		59

20a		PhCN		58
21a		PhCN		61
22a		PhCN		54
23a	Br	PhCN	Br H	42
24a	CN	MeCN, PhCN		0

#### 5. Characterization of the products

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H} = 7.51-7.53$  (d, J = 7.9 Hz, 2H), 7.33–7.36 (t, J = 7.8 Hz, 2H), 7.11–7.15 (t, J = 7.3 Hz, 1H), 7.22 (s, NH), 2.20 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 168.18, 137.87, 129.01, 124.32, 119.85, 24.61$ . MS (ESI) m/z: [M+H]<sup>+</sup> 136.0.

**6** *N-(2-bromophenyl)acetamide* (**2a**). Light yellow solid (17 %, 0.048 g), mp 104-105 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H} = 8.44$  (d, J = 8.1 Hz, 1H), 7.71 (s, NH), 7.64 (d, J = 8.2 Hz, 1H), 7.48 (t, J = 7.8 Hz, 1H), 7.06 (t, J = 7.8 Hz, 1H), 2.26 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 168.31$ , 135.82, 132.18, 128.31, 122.32, 113.43, 125.30, 24.50.MS (ESI) m/z: [M+H]<sup>+</sup> 214.0.



**Br** *N*-(*4*-bromophenyl)acetamide (**2b**). Light yellow solid (51%, 0.144 g), mp 167-169°C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H} = 7.53$  (d, J = 8.5 Hz, 2H), 7. 41 (d, J = 8.5 Hz, 2H), 7.26 (s, NH), 2.17 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 168.82$ , 139.21, 132.07, 121.32, 114.85, 24.5.MS (ESI) m/z: [M+H]<sup>+</sup> 214.0.



Br *N-(2,5-dibromophenyl)acetamide* (**3a**). White solid (69%, 0.267 g), mp 172-174°C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H} = 8.36$ (s, 1H), 7.95 (s, 1H), 7.43 (d, *J* = 8.6 Hz, 1H), 7.16-7.09 (m, 1H), 2.28 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 168.18$ , 136.77, 133.08, 124.41, 122.09, 111,41, 24.87.MS (ESI) m/z: [M+H]<sup>+</sup> 294.0.

 $\underbrace{\text{N-}(2-(trifluoromethyl)phenyl)acetamide (4a). White solid (57\%, 0.153 g), mp 95-97 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): <math>\delta_{\text{H}} = 8.24$  (d, J = 7.9 Hz, 1H), 7.64 (d, J = 7.8 Hz, 1H), 7.59 (t, J = 7.8 Hz, 1H), 7.40 (s, NH), 7.19 (t, J = 7.6 Hz, 1H), 2.21 (s, 3H). <sup>13</sup>C NMR (100 MHz, 100 MHz, 100 MHz, 100 MHz) (t, J = 7.6 Hz, 1H), 2.21 (s, 3H). <sup>13</sup>C NMR (100 MHz) (t, J = 7.6 Hz, 1H), 2.21 (s, 3H). <sup>13</sup>C NMR (100 MHz) (t, J = 7.6 Hz) (t, J

CDCl<sub>3</sub>):  $\delta_C = 168.44$ , 135.11, 132.93, 126.14, 124.69, 124.53, 124.43, 24.87. MS (ESI) m/z: [M+H]<sup>+</sup> 204.0.



**F<sub>3</sub>C** *N-(4-(trifluoromethyl)phenyl)acetamide* (**4b**). White solid (19%, 0.051 g), mp 147-149 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta_{\rm H} = 7.68$  (d, J = 7.9 Hz, 2H), 7. 53 (d, J = 7.9 Hz, 2H), 7.37 (s, NH), 2.22 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 168.89$ , 142.65, 128.80, 123.40, 123.17, 123.00, 122.80, 119.20, 24.87. MS (ESI) m/z: [M+H]<sup>+</sup> 204.0.



**O** *N-benzyl-acetamide* (**5a**). Colorless crystals (78%, 0.126 g), mp 43–45 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.38 - 7.27$  (m, 5H), 5.86 (s, 1H), 4.45 (d, *J* = 5.5 Hz, 2H), 2.04 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 169.94$ , 138.23, 128.73, 127.86, 127.56, 43.8, 22.23. MS (ESI) m/z: [M+H]<sup>+</sup> 150.0.

**O** *N*-(2-methylbenzyl)acetamide (**6a**). White solid (69%, 0.106 g), mp 73–76 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.26 - 7.19$  (dt, 4H), 5.58 (s, 1H), 4.46 (d, *J* = 5.4 Hz, 2H), 2.35 (s, 3H), 2.04 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 169.67$ , 136.5, 135.81, 130.58, 128.66, 127.86, 126.25, 41.92, 23.18, 18.94. MS (ESI) m/z: [M+H]<sup>+</sup> 164.0.

 $\circ$  *N-(3-methylbenzyl)acetamide* (**7a**). White solid (67%, 0.103 g), mp 21–24 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.27$ -7.21 (m, 1H), 7.14 – 7.07 (m, 3H), 5.85 (s, 1H), 4.41 (d, *J* = 5.7 Hz, 2H), 2.37 (s, 3H), 2.04 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 169.76$ , 138.49, 138.14, 128.65, 128.32, 124.91, 43.8, 23.29, 21.33. MS (ESI) m/z: [M+H]<sup>+</sup> 164.0.



**O** *N-(4-methylbenzyl)acetamide* (**8a**). White solid (72 %, 0.11 g), mp 102–103 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H}$  = 7.20 (d, *J* = 8.2 Hz, 2H), 7.14 (d, *J* = 8.2 Hz, 2H), 5.59 (s,

1H), 4.45 (d, J = 5.4 Hz, 2H), 2.34 (s, 3H), 2.03 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 169.92, 137.31, 135.2, 129.41, 127.88, 43.53, 23.28, 21.09$ . MS (ESI) m/z: [M+H]<sup>+</sup> 164.0.



**O** *N*-(2-bromobenzyl)acetamide (**9a**). White solid (56%, 0.0746), mp. 75–76 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.61$  (d, *J* = 7.9 Hz, 1H), 7.49 (d, *J* = 7.7 Hz, 1H), 7.33 (t, *J* = 7.5 Hz, 1H), 7.23-7.17 (m, 1H), 6.31 (s, 1H), 4.64 (d, J = 5.7 Hz, 2H), 2.07 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 170.03$ , 138.17, 131.6, 129.73, 128.86, 127.56, 124.8, 43.7, 23.37. MS (ESI) m/z: [M+H]<sup>+</sup> 228.0.



**O** *N-(3,5-dimethylbenzyl)acetamide* (**10a**). White solid (62%, 0.091 g), mp. 80-81 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 6.92$  (d, J = 20.1 Hz, 3H), 6.24 (s, 1H), 4.37 (d, J = 4.5 Hz, 2H), 2.31 (s, 6H), 2.06 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 170.98$ , 138.46, 137.5, 129.35, 125.75, 44.11, 22.89, 21.1. MS (ESI) m/z: [M+H]<sup>+</sup> 178.0.

 $\circ$  *N-(4-bromo-3,5-dimethylbenzyl)acetamide* (**11a**). White solid (51%, 0.065 g), mp. 101-102 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.03$  (s, 2H), 5.97 (s, 1H), 4.45 (d, *J* = 5.6 Hz, 2H), 2.48 (s, 6H), 2.05 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 169.98$ , 139.92, 130.36, 127.48, 122.9, 44.25, 22.33, 20.3.MS (ESI) m/z: [M+H]<sup>+</sup> 256.0.

**O** *N-((6-methylpyridin-2-yl)methyl)acetamide* (**12a**). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.09$  (m, 2H), 7.86 (d, J = 7.8 Hz, 1H), 4.7 (d, J = 5.7 Hz, 2H), 2.5 (s, 3H), 2.07 (s, 3H). MS (ESI) m/z: [M+H]<sup>+</sup> 165.0.



*N-Phenylbenzamide* (**13a**). White solid (60%, 0.15 g), mp. 163-166 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.17$ -8.11 (m, 2H), 7.88-7.82 (m, 2H), 7.64-7.5 (m, 6H), 6.20 (s, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 166.82$ , 139.17, 135.44, 132.4, 128.77, 128.49, 128.01, 123.71, 120.28. MS (ESI) m/z: [M+H]<sup>+</sup> 198.



*N-(2-bromophenyl)benzamide* (**14a**). Light yellow solid (14%, 0.051 g), mp. 109-110 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.64$  (d, J = 7.8 Hz, 1H), 8.26 (s, NH), 8.04 (d, J = 7.2 Hz, 2H), 7.62 (d, J = 7.2 Hz, 2H), 7.48 (t, J = 7.0 Hz, 2H), 7.33 (t, J = 7.3 Hz, 1H), 6.97 (t, J = 7.3 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 165.31$ , 136.44, 134.11, 132.78, 131.87, 128.82, 128.47, 128.11, 127.92, 127.63, 120.56. MS (ESI) m/z: [M+H]<sup>+</sup> 276.



**Br** *N*-(*4*-bromophenyl)benzamide (**14b**). Light yellow solid (42%, 0.153 g), mp. 203-205 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.02$  (d, J = 7.6 Hz, 2H), 7.9 (s, NH), 7.81 (d, J = 7.6 Hz, 2H), 7.62-7.55 (m, 5H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 166.17$ , 137.09, 134.58, 131.6, 129.73, 128.86, 127.56, 117.23. MS (ESI) m/z: [M+H]<sup>+</sup> 276.



**Br** *N-(2,5-dibromophenyl)benzamide* (**15a**). White solid (59%, 0.277 g), mp. 146-148 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.11$  (d, J = 7.5 Hz, 2H), 7.85 (d, J = 7.5 Hz, 1H), 7.61-7.58 (m, 2H), 7.53 (s, NH), 7.6 (t, J = 7.0 Hz, 2H), 7.44 (t, J = 8.5 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 166.28$ , 137.21, 134.41, 132.78, 131.87, 128.82, 128.47, 128.11, 127.92, 127.63. MS (ESI) m/z: [M+H]<sup>+</sup> 354.



*N-(2-(Trifluoromethyl)phenyl)benzamide* (**16a**). White solid (48%, 0.168 g), mp. 144-145 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.41$  (d, J = 8.2 Hz, 1H), 8.25 (s, 1H), 7.89– 7.86 (m, 2H), 7.66–7.49 (m, 5H), 7.26 (t, J = 7.6 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} =$  166.73, 136.09, 134.57, 133.9, 132.66, 132.04, 129.67, 129.09, 127.4, 127.17, 127.0, 126.8. MS (ESI) m/z: [M+H]<sup>+</sup> 266.



**F**<sub>3</sub>**C** *N*-(2-(*Trifluoromethyl*)*phenyl*)*benzamide* (**16b**). White solid (16%, 0.056 g), mp. 200-203 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 8.01$  (s, 1H), 7.91 (d, *J* = 7.0 Hz, 2H), 7.79 (d, *J* = 8.2 Hz, 2H), 7.69 (d, *J* = 8.8 Hz, 2H), 7.62 (tt, *J* = 7.0, 1.8 Hz, 1H), 751 (t, *J* = 7.6 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 167.89$ , 144.15, 135.22, 132.73, 128.96, 125.84, 123.97, 121.07. MS (ESI) m/z: [M+H]<sup>+</sup> 266.



**O** *N-benzylbenzamide* (**17a**). White solid (69%, 0.158 g), mp. 94–96 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.81$  (m, 2H), 7.69-7.6 (m, 4H), 7.37 (m, 4H), 6.5 (s, 1H), 4.66 (d, J = 5.7 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 166.48$ , 138.16, 134.38, 133.11, 128.88, 128.61, 127.96, 127.64, 126.96, 44.17. MS (ESI) m/z: [M+H]<sup>+</sup> 212.



**O** *N-(2-bromobenzyl)benzamide* (**18a**). White solid (48%, 0.081 g), mp. 105-106 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.90-7.84$  (m, 2H), 7.75-7.32 (m, 6H), 7.25-7.16 (m, 1H), 6.73 (s, 1H), 4.66 (d, *J* = 5.7 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 168.43$ , 137.72, 135.19, 134.21, 132.77, 131.93, 129.44, 128.62, 128.06, 127.51, 123.87, 44.56. MS (ESI) m/z: [M+H]<sup>+</sup> 290.



**O** *N-(2-methylbenzyl)benzamide* (**19a**). White solid (59%, 0.125 g), mp. 107-109 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.82-7.76$  (m, 2H), 7.43–7.36 (m, 1H), 7.32–7.23 (m, 2H), 7.21–7.16 (m, 1H), 7.12–7.07 (m, 3H), 6.7 (s, 1H), 4.6 (d, *J* = 5.3 Hz, 2H), 2.26 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 167.91$ , 138.47, 137.89, 136.23, 131.58, 130.84, 128.70, 128.60, 128.12, 127.50, 125.79, 42.28, 19.80. MS (ESI) m/z: [M+H]<sup>+</sup> 226.



**O** *N-(3-methylbenzyl)benzamide* (**20a**). Light yellow solid (58%, 0.123 g) mp. 73-75 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.93-7.88$  (d, 2H), 7.49–7.41 (m, 1H), 7.36-7.29 (m, 2H), 7.27–7.19 (m, 1H), 7.15–7.06 (m, 3H), 6.68 (s, 1H), 4.58 (d, *J* = 5.6 Hz, 2H), 2.39 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 167.14$ , 139.62, 139.19, 134.81, 131.53, 128.78, 128.61, 128.44, 127.50, 125.83, 44.43, 20.88. MS (ESI) m/z: [M+H]<sup>+</sup> 226.



**O** *N-(4-methylbenzyl)benzamide* (**21a**). White solid (61%, 0.129 g) mp. 136-138 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.83-7.80$  (m, 2H), 7.67–7.65 (m, 1H), 7.46-7.42 (m, 2H), 7.25 (d, *J* = 7.5 Hz, 2H), 7.13 (d, *J* = 7.5 Hz, 2H), 6.50 (s, 1H), 4.62 (d, *J* = 5.6 Hz, 2H), 2.37 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 167.29$ , 137.11, 135.12, 134.28, 131.29, 129.32, 128.41, 127.82, 126.93, 43.74, 21.33. MS (ESI) m/z: [M+H]<sup>+</sup> 226.



**O** *N-(3,5-dimethylbenzyl)benzamide* (**22a**). White solid (54%, 0.107 g), mp. 68-70 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H} = 7.83-7.81$  (m, 2H), 7.57–7.52 (m, 1H), 7.49-7.46 (m, 2H), 7.03-6.94 (m, 3H), 6.45 (s, 1H), 4.6 (d, *J* = 4.5 Hz, 2H), 2.33 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C} = 167.77$ , 138.64, 136.24, 131.55, 129.42, 128.68, 127.20, 125.81, 42.32, 19.13. MS (ESI) m/z: [M+H]<sup>+</sup> 240.



**O** *N*-(4-bromo-3,5-dimethylbenzyl)benzamide (**23a**). White solid (42%, 0.067), mp. 96-98 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H}$  = 7.83-7.76 (m, 2H), 7.55–7.52 (m, 1H), 7.51- 7.40 (m, 2H), 6.99-6.87 (m, 2H), 6.59 (s, 1H), 4.59 (d, *J* = 4.5 Hz, 2H), 2.39 (s, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta_{\rm C}$  = 167.66, 138.85, 135.97, 131.44, 129.42, 128.81, 125.88, 42.56, 20.45. MS (ESI) m/z: [M+H]<sup>+</sup> 318.

#### 6. Electrochemical potentials

Table S2. Oxidation potentials (V) of compounds; (*a*) our results, conditions: WE- GC, RE – Ag/AgCl, Bu<sub>4</sub>NBF<sub>4</sub>, CH<sub>3</sub>CN; (*b*) literature data, conditions: WE- Pt, RE – Ag/AgNO<sub>3</sub>, perchlorate salts (LiClO<sub>4</sub>, NaClO<sub>4</sub>, (C<sub>2</sub>H<sub>5</sub>)<sub>4</sub>NClO<sub>4</sub>, (n-C<sub>8</sub>H<sub>7</sub>)<sub>4</sub>NClO<sub>4</sub>, (*n*-C<sub>4</sub>H<sub>9</sub>)<sub>4</sub>NClO<sub>4</sub>)), CH<sub>3</sub>CN

N⁰	Compound	$E_{ox}^{a}$ , V	$E_{ox}^{b}, V$	References
1	Benzene	2.53	2.08	1
2	Toluene	2.20	1.98	1
3	o-Xylene	2.15	1.58	1
4	m-Xylene	2.15	1.58	1, 2
5	p-Xylene	2.14	1.56	1
6	Bromobenzene	2.57	1.98	1
7	2-Bromtoluene	2.25		
8	Mesitylene	2.01	1.55	1
9	2-Bromomesitylene	2.05		
10	Naphthalene	1.70	1.34	1
11	2-Phenylpyridine	2.20, 2.65		
12	Cu(OAc) <sub>2</sub>	2.35		
13	Benzonitrile	2.35	1.8 vs SCE	3

#### References

- 1. N.L. Weinberg and H.R. Weinberg, Chem. Rev., 1968, 68, 449-523.
- 2. D.M. Heard and A.J.J. Lennox, Angew. Chem. Int. Ed., 2020, 59, 18866-18884.
- 3. K.M. Kadish and J.E. Anderson, Pure Appl. Chem., 1987, 59, 703-714.

# 7. NMR Spectra













































































Formation of a product with three substituted  $CH_3$  group of 2-Bromomesitylene was established by GC-MS analysis



Abundance





N,N',N''-((2-bromobenzene-1,3,5-triyl)tris(methylene))triacetamide,

M= 370

Abundance



Formation of 12 was determined GC-MC analysis

) M (12)

M=164

