Supporting Information

Targeted perfusion adsorption for hyperphosphatemia using Mixed Matrix Microspheres (MMMs) encapsulated NH₂-MIL-101(Fe)

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The supplementary file includes:

Fig. S1. Digital pictures of MMMs.

Fig. S2. Dynamic contact angle of MMMs and PSF microspheres.

Fig. S3. Effect of S/L ratio on the phosphate adsorption to NH₂-MIL-101(Fe) in water

 $(C_0 = 50 \text{ mg } \text{L}^{-1})$ at 25°C.

Fig. S4. SEM images and element distribution after surface (a and a') and crosssection (b and b') adsorption.

Fig. S5. Zeta potential of NH₂-MIL-101(Fe).

Fig. S6. BSA adsorption onto the MMMs.

Text. S1. The pseudo-first-order and pseudo-second-order equation.

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Table S1. Phosphate adsorption capacities of various adsorbents.

Table S2. Molecular weight and molecular size of urea, creatinine, lysozyme,

phosphate.

Table S3. The Lorentzian peak shape fitting parameters for O 1s XPS peak of NH₂-

MIL-101(Fe) before and after phosphate adsorption.



Fig. S1. Digital pictures of MMMs.



Fig. S2. Dynamic contact angle of MMMs and PSF microspheres.



Fig. S3. Effect of S/L ratio on the phosphate adsorption to NH₂-MIL-101(Fe) in water $(C_0 = 50 \text{ mg L}^{-1})$ at 25°C.



Fig. S4. SEM images and element distribution after surface (a and a') and cross-section (b and b') adsorption.







Fig. S6. BSA adsorption onto the MMMs.

Text. S1

The pseudo-first-order equation: $\ln (Q_e - Q_t) = \ln Q_e - k_1 t$ The pseudo-second-order equation:

 $\frac{t}{Q_t} = \frac{1}{K_2 Q_e^2} + \frac{t}{Q_e}$

Where k_1 and k_2 are the pseudo-first-order and pseudo-second-order rate constant, Q_e (mg/g) is the adsorption capacity coefficient at equilibrium time; Q_t (mg/g) is the amount adsorbed per unit weight of sorbent at t time.

Text. S2

The Langmuir model:

$$Q_e = Q_m K_L \frac{C_e}{1 + K_L C_e}$$

The Freundlich model:

$$Q_e = K_F C_e^{1/n}$$

 $Q_e (mg/g)$ is the adsorption capacity coefficient at equilibrium time; $Q_m (mg/g)$ is the maximal adsorption capacity in Langmuir model; $C_e (mg/L)$ is the equilibrium concentration; $K_F [(mg/g) (L/mg)^{1/n}]$ is the Freundlich capacity coefficient, and 1/n is isotherm curvature.

Adsorbent	Adsorption capacity	Ref	
	(mg/g)		
Fe-Cu binary oxides	35.2	1	
La-doped ordered	47.89	2	
mesoporous hollow silica			
spheres			
activated carbon nanofiber	26.3	3	
(ACF-ZrFe)			
graphene nanosheets (GNS)	41.96	4	
supported lanthanum			
hydroxide (LaOH)			
lanthanum oxide decorated	82.6	5	
graphene composite			
β-FeOOH/GO	45.2	6	
MWCNTs-COOH-La	48.02	7	
Weak-base resin	61.38	8	
UiO-66	85	9	
UiO-66-NH ₂	92	9	
NH ₂ -MIL-101(Fe)	62.38	This Study	

Table S1. Phosphate adsorption capacities of various adsorbents.

Table S2. Molecular weight and molecular size of urea, creatinine, lysozyme and phosphate.

Uremic Toxis	MW (g/mol)	Molecular size (nm)
Urea	60.056	0.36
Creatinine	113.12	0.54
Lysozyme	895.9	3 x 4.5
Phosphate	94.971	0.23

Table S3. The Lorentzian peak shape fitting parameters for O 1s XPS peak of NH ₂ -
MIL-101(Fe) before and after phosphate adsorption.

with total of before and after phosphate adsorption.					
Sample	Peak	Position	Area	percent	
		BE (eV)	(cps eV)	(%)	
	Fe-O-Fe	529.4	1767.4	22.5	
NH ₂ -MIL-101(Fe)	Fe-OH	530.3	4526.5	57.5	
	O-C=O	531.6	1574.7	20	
	Fe-O-Fe	529.3	3243.8	23.1	
NH ₂ -MIL-101(Fe)/Phosphate	Fe-OH/Fe-O-P/P=O	530.2	5977.8	42.5	

P-OH	531.2	2716.4	19.3
 O-C=O	531.9	2134	15.2

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