Supporting information

Engineering non-covalently assembled protein nanoparticles for long-acting

gouty arthritis therapy

Jinrui Zhang^{1,2}#, Yao Sun³#, Qian Qu³#, Bo Li², Lili Zhang⁴, Rui Gu¹, Jianlin Zuo¹, Wei Wei⁴*, Chao Ma³, Lei Liu³, Kai Liu^{2,3}*, Jingjing Li²*, Hongjie Zhang^{2,3}

1. Department of Orthopedics, China-Japan Union Hospital of Jilin University, 130033, Changchun, China

2. State Key Laboratory of Rare Earth Resource Utilization, Changchun Institute of Applied Chemistry, Chinese Academy of Sciences, Changchun 130022, China

3. Department of Chemistry, Tsinghua University, 100084, Beijing, China

4. Key Laboratory of Organ Regeneration & Transplantation of the Ministry of Education, Institute of Translational Medicine, Institute of Virology and AIDS Research, The First Hospital of Jilin University, Changchun, Jilin Province 130061, China.

#These authors contributed equally to this work

Corresponding Authors:

*Jingjing Li, E-mail: jjingli@ciac.ac.cn; Wei Wei, E-mail: wwei6@jlu.edu.cn; Kai Liu. E-mail: kailiu@tsinghua.edu.cn.

Figures



Figure S1. SDS-PAGE analysis of (A) the IL-1Ra protein and (B) the IK-Pr. The protein purity was 97% and 91%, respectively.



Figure S2. Confocal laser scanning microscope (CLSM) was used for analysis of the biocompatibility and cytotoxicity of the IK-Np. (A) and (B) showed BMSCs in blank control group stained by calcein-AM and pripodium iodide (PI), respectively. (C) and (D) showed BMSCs stained by calcein-AM and PI after co-incubated with 400 μ g/mL of the IK-Np for 48 hours, respectively. Scale bar: 100 μ m. The results illustrated excellent biocompatibility of the IK-Np.



Figure S3. Cytokines in the plasm and joint fluid of rats after therapy were detected by ELISA. (A) and (B) showed the levels of IL-6 in serum and joint fluid of rats, respectively. (C) and (D) showed the levels of TNF- α in serum and joint fluid of rats, respectively. (E) and (F) showed the levels of IL-1 β in serum and joint fluid of rats, respectively.



Figure S4. Illustration of potential mechanism of the IK-Np involving the inhibition of IL-1 β signal pathway. MSU crystals stimulated innate immune cells including macrophages, monocytes and neutrophils, facilitating the production of active IL-1 β and TNF- α . IL-1 β further binds to IL-1 receptor (IL-1R) of surrounding synoviocytes and chondrocytes, resulting in inflammation process. Downstream cytokines (such as PGE2, MMP3 and IL-6) subsequently were secreted, promoting the inflammatory cascades and causes tissue damage through various signal pathways. The IK-Prs are slowly released from the nanoassembled IK-Np and reached to nidus during systemic circulation, where the IK-Pr can competitively bind to IL-1R and suppress the IL-1 β signal pathway, resulting in alleviation of inflammatory reaction.

Therapeutics based on IL- 1Ra	Modification strategy	Half-life (hour)	Formation	Mode of administration	Source
IK-Np	Fusing of IL-1Ra with ELP and PEGylation	27.16	Nanoparticle	Subcutaneous injection	This study
Anakinra	Human IL-1Ra expressed in <i>E. coli</i>	4-6	Protein	Subcutaneous injection	FDA-approved drug (https: //www.drugs.com/ kineret.html)
IL-1Ra-PF127	IL-1Ra loaded in Pluronic F-127 gels	12.53±2.48	Polymer-based gel	Subcutaneous injection	Akash M. S. H. et al., 2013 ¹
IL-1Ra-HSA	Fusing human serum albumin (HSA) to the carboxyl terminal of IL-1Ra	9.8 ± 1.7	Recombinant protein	Intravenous injection	Liu M. et al., 2012 ²
dAbm16-IL-1Ra	Fusing the Anti- serum albumin domain antibodies (dAbs) to the amino terminal of IL-1Ra	4.3	Recombinant protein	Intravenous injection	Holt L. J. et al., 2008 ³
IL-1Ra-MPs	IL-1Ra loaded by mineral coated microparticles (MPs)	detectable in serum within 24 hours	Microparticle	Local bolus injection	Clements A. E. B. et al., 2018 ⁴

Table S1. Half-lives of different therapeutics based on IL-IRa

References

- 1 M. S. H. Akash, K. Rehman, H. Sun and S. Chen, *PLoS One*, 2013, **8**, e55925.
- 2 M. Liu, Y. Huang, L. Hu, G. Liu, X. Hu, D. Liu and X. Yang, *BMC Biotechnol.*, 2012, **12**, 1–13.
- 3 L. J. Holt, A. Basran, K. Jones, J. Chorlton, L. S. Jespers, N. D. Brewis and I. M. Tomlinson, *Protein Eng. Des. Sel.*, 2008, **21**, 283–288.
- 4 A. E. B. Clements, E. R. Groves, C. S. Chamberlain, R. Vanderby and W. L. Murphy, *Adv. Healthc. Mater.*, 2018, **7**, 1800263.