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Supporting Information

Synthesis of Tetraphenylethene-based D-A Conjugated Molecules with Near-

Infrared AIE Features, and Application in Photodynamic Therapy

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Scheme S1. The structures and synthetic routes of TPE-T-Py, MTPE-T-Py, MTPE-T-Qy, MTPE-DT-Py and MTPE-DT-Qy.

Synthesis of Compound 1. 1-(4-bromophenyl)-1,2,2-triphenylethylene (1.025 g, 2.5 mmol) and Pd(dppf)Cl₂ (183 mg, 0.25 mmol) was dissolved in 24 mL anhydrous toluene under nitrogen, then 5-formyl-2-thiopheneboronic acid (780 mg, 5 mmol), K₂CO₃ (1.38 g, 7.5 mmol) in anhydrous methanol was added. The mixture was heated to reflux for 16 h under nitrogen. After cooling to room temperature, the reaction is quenched by water. The organic layer was extracted by dichloromethane, washed with brine, dried over anhydrous Na₂SO₄ and evaporated to dryness under reduced pressure. The crude product was purified by silica gel with column to give compound **1** as yellow powder (770 mg, yield: 70%). ¹HNMR (500 MHz, Chloroform-d) δ 9.90 (s, 1H), 7.74 (d, J = 4.0 Hz, 1H), 7.46 (d, J = 8.1 Hz, 2H), 7.38 (d, J = 4.0 Hz, 1H), 7.21 – 7.02 (m, 17H). HRMS (ESI⁺): calcd. for C₃₁H₂₂OS⁺[M+H]⁺ 443.1464, found 443.1459.



Figure S1. ¹H NMR spectrum of compound 1



Figure S2. HRMS spectrum of compound 1

Synthesis of Compound 2. 4-pyridineacetonitrile (56.68 mg, 0.48 mmol) and *t*-BuOK (67 mg, 0.6 mmol) were successively added to anhydrous EtOH (15 mL) in a round bottomed flask, which was stirred at room temperature for 10 min. Then compound 1 (176 mg, 0.4 mmol) was added to the solution, and the mixture was further stirred at room temperature for 36 h. Then the solid was filtered off, washed with 30 mL EtOH and dried under vacuum. The obtained solid was purified by silica gel chromatography using petroleum ether/CH₂Cl₂ mixture as an orange yellow powder (195 mg, yield: 90%). ¹HNMR (400 MHz, Chloroform-*d*) δ 8.66 (d, *J* = 6.1 Hz, 2H), 7.80 (s, 1H), 7.63 (d, *J* = 4.0 Hz, 1H), 7.55 – 7.49 (m, 2H), 7.43 (d, *J* = 8.3 Hz, 2H), 7.32 (d, *J* = 4.0 Hz, 1H), 7.16 – 7.03 (m, 17H). ¹³C NMR (100 MHz, Chloroform-d) δ 151.47, 150.21, 145.05, 143.45, 141.98, 140.00, 137.01, 136.24, 136.00, 132.16, 131.38, 131.32, 127.93, 127.84, 127.70, 126.84, 126.71, 126.66, 125.53, 123.83, 119.56, 117.26, 104.20. HRMS (ESI⁺): calcd. for C₃₈H₂₇N₂S⁺[M+H]⁺ 543.1889, found 543.1879.







210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

Figure S4. ¹³C NMR spectrum of compound 2



Figure S5. HRMS spectrum of compound 2

Synthesis of 1,1'-[1-(4-bromophenyl)-1,2-ethenediylidene] bis[methoxybenzene] (3). 4-Bromobenzophenone (2.6 g, 10 mmol), 4, 4'dimethoxybenzophenone (3.1 g, 12 mmol) and zinc powder (6.5 g, 100 mmol) were mixed in anhydrous THF (150 mL) and stirred at ice water bath. Titanium tetrachloride (6.7 mL, 60 mmol) was then injected dropwise for 30 min. After recovering to room temperature, the mixture was heated at 80 °C for 24 h. The reaction was quenched with 10% K₂CO₃ aqueous solution and filtrated with silica gel powder. The residue was washed with ethyl acetate (50 mL × 3). The organic phase was dried with Na₂SO₄ and the solvent was removed under reduced pressure. The obtained residue was purified with chromatography (hexane/dichloromethane = 4/1, v/v) to give compound **3** as a white powder (2.05 g, yield: 43%). ¹HNMR (400 MHz, Chloroform-d) δ 7.25–6.83 (m, 13H), 6.70–6.57 (m, 4H), 3.84–3.69 (m, 6H). HRMS (ESI+): calcd. For C₂₈H₂₃BrO₂Na⁺[M+Na]⁺493.0774, found 493.0761.







Figure S7. HRMS spectrum of compound 3

2-[4-[1,2-bis(4-methoxyphenyl)-2-phenylethenyl]phenyl]-**Synthesis** of 4,4,5,5-tetramethyl-1,3,2-dioxaborolane (4). A mixture of Compound 3 (705 mg, 1.5 mmol), 4,4,4',4',5,5,5',5'-octamethyl-2,2'-bi(1,3,2-dioxaborolane) (457 mg, 1.8 mmol), potassium acetate (588 mg, 6 mmol), and Pd(dppf)Cl₂ (54 mg, 0.074 mmol) in anhydrous dimethyl sulfoxide (15 mL) was stirred at 100 °C under Nitrogen for 24 h and then water added. The crude product was extracted into ethyl acetate, washed with brine, and dried over anhydrous sodium sulfate. After removing solvent under reduced pressure, the residue was purified by column chromatography (CH₂Cl₂/petroleum ether, 1/2, v/v) on silica gel to yield a white powder (650 mg, yield: 83.6%). ¹HNMR (400 MHz, DMSO-d6) δ 7.43 (d, J = 8.2 Hz, 2H), 7.19 – 7.04 (m, 3H), 7.00 - 6.90 (m, 4H), 6.86 (dd, J = 8.8, 5.6 Hz, 4H), 6.74 - 6.66 (m, 4H), 3.68(d, J = 2.7 Hz, 6H), 1.26 (s, 12H). ¹³CNMR (100 MHz, DMSO-d₆) δ 158.27, 147.54, 144.09, 140.90, 138.97, 135.93, 134.44, 132.52, 131.22, 130.74, 128.34, 126.80, 113.65, 81.82, 55.39, 25.30



ure S8. ¹H NMR spectrum of compound 4



igure S9. ¹³C NMR spectrum of compound 4

Synthesis of Compound 5. The same synthetic procedure as described compound **1** for using compound **3** (345 mg, 0.8 mmol), affording the product as an orange yellow powder (321 mg, yield: 64%). ¹HNMR (400 MHz, Chloroform-d) δ 9.86 (s, 1H), 7.70 (d, J = 3.9 Hz, 1H), 7.42 (d, J = 8.3 Hz, 2H), 7.34 (d, J = 4.0 Hz, 1H), 7.18 – 6.88 (m, 11H), 6.65 (dd, J = 12.4, 8.8 Hz, 4H), 3.75 (d, J = 3.7 Hz, 6H). HRMS (ESI+): calcd. for C₃₃H₂₆O₃S⁺[M+H]⁺ 503.1675, found 503.1665. HRMS (ESI+): calcd. for C₃₃H₂₆O₃S⁺[M+Na]⁺ 525.1495, found 525.1489.







Figure S11. HRMS spectrum of compound 5

Synthesis of Compound 6. The same synthetic procedure as described compound 2 for using compound 5 (345 mg, 0.8 mmol), affording the product as an orange yellow powder (90 mg, yield: 75%).¹HNMR (400 MHz, Chloroform-d) δ 8.66 (d, J = 6.3 Hz, 2H), 7.83 – 7.60 (m, 2H), 7.52 (dd, J = 4.7, 1.5 Hz, 2H), 7.44 (d, J = 8.4 Hz, 2H), 7.32 (d, J = 4.0 Hz, 1H), 7.13 (d, J = 7.2 Hz, 3H), 7.08 – 7.03 (m, 4H), 7.00 – 6.92 (m, 4H), 6.66 (dd, J = 14.6, 8.8 Hz, 4H), 3.75 (d, J = 6.8 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-d) δ 158.39, 158.24, 151.52, 150.39, 145.69, 143.87, 141.68, 141.19, 138.24, 136.91, 136.18, 136.05, 135.89, 132.61, 132.18, 131.43, 130.44, 127.86, 126.36, 125.53, 123.68, 119.49, 117.31, 113.25, 113.03, 104.15, 55.12. HRMS (ESI+): calcd. for C₄₀H₃₂N₂O₂S⁺[M+H]⁺603.2101, found 603.2098. ¹⁷C⁻²³⁻¹¹¹⁸



Figure S12. ¹H NMR spectrum of compound 6



Figure S13. ¹³C NMR spectrum of compound 6



Figure S14. HRMS spectrum of compound 6

Synthesis of Compound 7. The same synthetic procedure as described compound **2** for using compound **5** (200 mg, 0.4 mmol) and 2-(quinolin-4-yl)acetonitrile (84 mg, 0.5 mmol), affording the product as an orange red powder (90 mg, yield: 35%).¹H NMR (400 MHz, Chloroform-*d*) δ 8.94 (d, *J* = 4.5 Hz, 1H), 8.22 (dd, *J* = 21.1, 8.2 Hz, 2H), 7.78 (dd, *J* = 16.7, 8.3 Hz, 1H), 7.70 – 7.62 (m, 1H), 7.60 (d, *J* = 4.4 Hz, 1H), 7.52 (d, *J* = 6.8 Hz, 1H), 7.49 – 7.43 (m, 2H), 7.33 (d, *J* = 4.0 Hz, 1H), 7.17 – 7.04 (m, 6H), 7.01 – 6.84 (m, 6H), 6.72 – 6.57 (m, 4H), 3.76 (s, 6H). ¹³C NMR (100MHz, Chloroform-d) δ 158.38, 158.20, 150.85, 149.99, 148.79, 143.89, 141.60, 141.43, 141.15, 138.26, 136.06, 135.79, 135.56, 132.63, 132.55, 131.43, 131.32, 130.54, 130.36, 130.04, 127.86, 127.55, 126.35, 125.53, 123.51,120.80, 113.24, 113.03, 55.10. HRMS (ESI+): calcd. For C₄₄H₃₃N₂O₂S⁺ [M+H]⁺ 653.2257, found 653.2222.







Figure S16. ¹³C NMR spectrum of compound 7





Synthesis of Compound 8. The same synthetic procedure as described compound **1** for using compound **4** (414 mg, 0.8 mmol) and 7-bromo-2,3-dihydrothieno[3,4-b][1,4]dioxine-5-carbaldehyde (98 mg, 0.4 mmol) affording the product as an orange powder (190 mg, yield: 85%). ¹H NMR (400 MHz, Chloroform-d) δ 9.90 (s, 1H), 7.55 (d, J = 8.5 Hz, 2H), 7.15 – 6.89 (m, 11H), 6.64 (dd, J = 12.2, 8.8 Hz, 4H), 4.40 – 4.32 (m, 4H), 3.74 (d, J = 4.8 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-d) δ 179.59, 158.30, 158.19, 145.14, 143.95, 140.98, 138.40, 137.69, 136.12, 132.63, 131.82, 131.43, 129.31, 127.80, 126.28, 126.08, 115.38, 113.21, 113.00, 65.09, 64.49, 55.12.



Figure S18. ¹H NMR spectrum of compound 8



Figure S19. ¹³C NMR spectrum of compound 8

Synthesis of Compound 9. The same synthetic procedure as described compound 2 for using compound 8 (200 mg, 0.36 mmol), affording the product as an orange red powder (163 mg, yield: 69%).¹H NMR (400 MHz, Chloroform-d) δ 8.61 (d, J = 6.2 Hz, 2H), 7.95 (s, 1H), 7.59 (s, 2H), 7.49 (d, J = 6.3 Hz, 2H), 7.11 (d, J = 9.1 Hz, 3H), 7.06 – 7.02 (m, 4H), 6.96 (dd, J = 15.8, 8.8 Hz, 4H), 6.70 – 6.59 (m, 4H), 4.36 (dd, J = 15.0, 5.2 Hz, 4H), 3.75 (d, J = 8.3 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-d) δ 158.33, 158.18, 150.19, 146.60, 144.87, 143.94, 142.22, 141.00, 138.45, 137.65, 136.16, 136.11, 132.62, 132.00, 131.85, 131.46, 129.30, 127.80, 126.28, 126.08, 119.19, 117.90, 113.23, 113.00, 111.17, 100.26, 65.20, 64.59, 55.12. HRMS (ESI⁺): calcd. for C₄₂H₃₃N₂O₄S⁺[M+H]⁺661.2156, found 661.2147.



Figure S20. ¹H NMR spectrum of compound 9



Figure S21. ¹³C NMR spectrum of compound 9



Figure S22. HRMS spectrum of compound 9

Synthesis of Compound 10. The same synthetic procedure as described compound **2** for using compound **8** (200 mg, 0.36 mmol) and 2-(quinolin-4-yl)acetonitrile (84 mg, 0.5 mmol) affording the product as an orange red powder (165 mg, yield: 66%).¹H NMR (400 MHz, Chloroform-d) δ 8.91 (d, J = 4.5 Hz, 1H), 8.29 (d, J = 8.3 Hz, 1H), 8.19 (d, J = 8.3 Hz, 1H), 7.78 (t, J = 8.1 Hz, 1H), 7.72 – 7.57 (m, 4H), 7.46 (d, J = 4.5 Hz, 1H), 7.11 (t, J = 6.7 Hz, 3H), 7.05 (s, 4H), 6.97 (dd, J = 17.4, 8.7 Hz, 4H), 6.66 (dd, J = 16.3, 8.7 Hz, 4H), 4.33 (s, 4H), 3.75 (d, J = 10.8 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-d) δ 158.36 , 158.20 , 149.73 , 146.27, 140.98 , 138.50 , 137.60, 137.05 , 136.20 , 136.14 , 132.64 , 131.87 , 131.49 , 130.02 , 129.40 , 127.82 , 127.43 , 126.29 , 126.08 , 125.61 , 124.57 , 120.69 , 113.25 , 113.03 , 110.91 , 98.79 , 65.14, 64.60, 55.11. HRMS (ESI⁺): calcd. for C₄₆H₃₅N₂O₄S⁺[M+H]⁺711.2312, found 711.2300.



Figure S23. ¹H NMR spectrum of compound 10



Figure S24. ¹³C NMR spectrum of compound 10



Figure S25. HRMS spectrum of compound 10

Synthesis of TPE-T-Py. To a stirred solution of compound **2** (60 mg, 0.12 mmol) in acetonitrile, iodomethane (1.92 mmol, 16 equiv) was added and the mixture was heated to reflux for 24 h under nitrogen. After cooling to room temperature, the mixture was poured into diethyl ether. The precipitates were filtered and re-dissolved in 20 mL methanol, followed by adding saturated KPF₆ solution (15 mL). After stirring for 1 h, the solvent was evaporated and the residue was filtered again, washed with water and dried under reduced pressure to give target compound as an orange red powder (60mg, yield: 71%).¹H NMR (400 MHz, DMSO-d₆) δ 8.96 (d, J = 7.1 Hz, 3H), 8.30 (d, J = 7.1 Hz, 2H), 7.97 (d, J = 4.2 Hz, 1H), 7.80 (d, J = 4.1 Hz, 1H), 7.63 (d, J = 8.4 Hz, 2H), 7.21 – 6.96 (m, 17H), 4.30 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 153.54, 149.39, 145.98, 145.49, 144.18, 143.40, 143.20, 142.07, 141.86, 140.15, 136.11, 132.30, 131.22, 131.17, 131.10, 130.64, 128.52, 128.47, 128.33, 127.42, 127.28, 127.23, 126.21, 126.12, 122.74, 116.95, 100.61, 47.59. HRMS (ESI+): calcd. for C₃₉H₂₉N₂S⁺ [M-PF₆]+557.2046, found 557.2025.







Figure S27. ¹³C NMR spectrum of TPE-T-Py



Figure S28. HRMS spectrum of TPE-T-Py

Synthesis of MTPE-T-Py. The same synthetic procedure as described TPE-T-Py for using compound **6** (90 mg, 0.15 mmol), affording the product as a red powder (80 mg, yield: 70%).¹H NMR (400 MHz, DMSO-d₆) δ 8.96 (d, J = 7.2 Hz, 3H), 8.30 (d, J = 7.0 Hz, 2H), 7.97 (d, J = 4.2 Hz, 1H), 7.80 (d, J = 4.1 Hz, 1H), 7.63 (d, J = 8.4 Hz, 2H), 7.24–7.04 (m, 5H), 7.03–6.97 (m, 2H), 6.93 (d, J = 8.7 Hz, 2H), 6.87 (d, J = 8.7 Hz, 2H), 6.72 (dd, J = 18.5, 8.8 Hz, 4H), 4.30 (s, 3H), 3.69 (d, J = 2.4 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆) δ 158.50, 158.35, 153.77, 149.36, 146.27, 145.98, 144.17, 143.89, 141.89, 141.46, 138.29, 136.00, 135.87, 132.63, 132.56, 132.38, 131.33, 130.28, 128.50, 126.99, 126.24, 126.01, 122.71, 113.87, 113.67, 100.49, 55.41, 47.58. HRMS (ESI+): calcd. for C₄₁H₃₃N₂O₂S⁺ [M-PF₆]⁺ 617.2257, found 617.2231.







Figure S30. ¹³C NMR spectrum of MTPE-T-Py



Figure S31. HRMS spectrum of MTPE-T-Py

Synthesis of MTPE-T-Qy. The same synthetic procedure as described **TPE-T-Py** for using compound **7** (90 mg, 0.15 mmol), affording the product as a black red powder (40 mg, yield: 55%).¹H NMR (400 MHz, DMSO-d₆) δ 9.58 (d, J = 6.4 Hz, 1H), 8.69 (d, J = 8.4 Hz, 1H), 8.59 (d, J = 9.0 Hz, 1H), 8.42 (s, 1H), 8.39 – 8.32 (m, 2H), 8.20 – 8.11 (m, 1H), 7.99 (d, J = 4.1 Hz, 1H), 7.78 (d, J = 4.0 Hz, 1H), 7.64 (d, J = 8.4 Hz, 2H), 7.16 (dd, J = 13.3, 7.2 Hz, 3H), 7.09 (d, J = 8.4 Hz, 2H), 7.01 (d, J = 8.1 Hz, 2H), 6.95 (d, J = 6.9 Hz, 2H), 6.89 (d, J = 8.7 Hz, 2H), 6.76 (d, J = 8.8 Hz, 2H), 6.72 – 6.69 (m, 2H), 4.66 (s, 3H), 3.69 (d, J = 5.3 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆) δ 158.51 , 158.36 , 152.16 , 150.98 , 149.91 , 146.93 , 145.97 , 143.94 , 141.39 , 140.90 , 139.28 , 138.35 , 135.98 , 132.62 , 132.36 , 131.33 , 130.91 , 130.45 , 128.50 , 127.35 , 126.97 , 126.47 , 126.14 , 125.56 , 118.16 , 113.89 , 99.43 , 55.42 , 45.94 . HRMS (ESI+): calcd. for C₄₅H₃₅N₂O₂S⁺[M-PF₆]⁺ 667.2414, found 667.2409.







Figure S33. ¹³C NMR spectrum of MTPE-T-Qy



Figure S34. HRMS spectrum of MTPE-T-Qy

Synthesis of MTPE-DT-Py. The same synthetic procedure as described TPE-T-Py for using compound 9 (120 mg, 0.18 mmol), affording the product as an orange red powder (110 mg, yield: 75%).¹H NMR (400 MHz, DMSO-d₆) δ 8.84 (d, J = 7.0 Hz, 2H), 8.47 (s, 1H), 8.27 (d, J = 7.0 Hz, 2H), 7.61 (d, J = 8.5 Hz, 2H), 7.21 – 7.05 (m, 5H), 7.02 – 6.97 (m, 2H), 6.90 (dd, J = 20.4, 8.7 Hz, 4H), 6.72 (dd, J = 17.4, 8.8 Hz, 4H), 4.63 – 4.40 (m, 4H), 4.27 (s, 3H), 3.69 (d, J = 3.6 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆) δ 158.48, 158.38, 150.75, 149.56, 145.75, 145.61, 143.85, 141.48 ,139.06, 138.37, 137.72, 135.88, 132.63, 132.55, 132.15, 131.32, 129.11, 128.48, 127.95, 126.98, 126.58, 122.31, 113.86, 113.68, 111.04, 96.68, 66.18, 65.18, 55.42, 47.34. HRMS (ESI+): calcd. for C₄₃H₃₅N₂O₄S⁺[M-PF₆]⁺ 675.2312, found 675.2307.



Figure S35. ¹H NMR spectrum of MTPE-DT-Py



Figure S36. ¹³C NMR spectrum of MTPE-DT-Py



Figure S37. HRMS spectrum of MTPE-DT-Py

Synthesis of MTPE-DT-Qy. The same synthetic procedure as described TPE-T-Py for using compound 10 (110 mg, 0.15 mmol), affording the product as an orange red powder (70 mg, yield: 54%).¹H NMR (400 MHz, DMSO-d₆) δ 9.47 (d, J = 6.5 Hz, 1H), 8.64 (d, J = 8.5 Hz, 1H), 8.56 (d, J = 9.0 Hz, 1H), 8.33 (dd, J = 10.4, 6.8 Hz, 2H), 8.17–8.09 (m, 1H), 8.04 (s, 1H), 7.63 (d, J = 8.5 Hz, 2H), 7.16 (dt, J = 12.0, 6.8 Hz, 3H), 7.09 (d, J = 8.4 Hz, 2H), 7.03 – 6.98 (m, 2H), 6.94 (d, J = 8.7 Hz, 2H), 6.88 (d, J = 8.7 Hz, 2H), 6.76 (d, J = 8.8 Hz, 2H), 6.70 (d, J = 8.8 Hz, 2H), 4.63 (s, 3H), 4.46 (dd, J = 19.3, 4.5 Hz, 4H), 3.69 (d, J = 7.5 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆) δ 158.47, 158.36, 151.44, 149.67, 149.62, 145.42, 143.90, 141.38, 139.25, 138.89, 138.41, 135.91, 132.63, 132.55, 132.14, 131.32, 130.70, 130.12, 129.25, 128.48, 126.46, 126.04, 121.89, 113.87, 113.68, 110.68, 95.78, 66.08, 65.24, 55.44, 55.41, 45.71. HRMS (ESI+): calcd. for C₄₇H₃₇N₂O₄S⁺[M-PF₆]⁺ 725.2469, found 725.2460



Figure S38. ¹H NMR spectrum of MTPE-DT-Qy



Figure S39. ¹³C NMR spectrum of MTPE-DT-Qy



Figure S40. ¹H NMR spectrum of MTPE-DT-Qy



Figure S41. PL spectra of (A) TPE-T-Py (B) MTPE-T-Py (C) MTPE-T-Qy (D) MTPE- DT-Qy in DMSO/toluene mixtures with different toluene fractions. [AIE-PSs]=10⁻⁵ M.



Figure S42. Absorption spectra of ABDA in the presence of Rose Bengal under (A) 20 mW cm⁻², (B) 50 mW cm⁻², and (C) 100 mW cm⁻² white light irradiation. (D) Decomposition rates of ABDA in the presence of Rose Bengal under different light intensities, where A_0 and A are the absorbance of ABDA at 378 nm.



Figure S43. Absorption spectra of ABDA (50×10^{-6} M) in (A) TPE-T-Py and (B) MTPE-T-Py (C) MTPE-T-Qy (D) MTPE-DT-Qy solutions (5×10^{-6} M) irradiated for different durations with white light irradiation (20 mW cm^{-2})



Figure S44. Particle size distributions of (A) TPE-T-Py. (B) MTPE-T-Py. (C) MTPE-T-Qy. (D) MTPE-DT-Py. (E) MTPE-DT-Qy aggregates in DMSO/H₂O mixture with 99% H₂O fraction



Figure S45. Confocal images of MCF-7 cells incubated with (A) TPE -T-Py and (B) MitoTracker Green, and (C) their merged images. (D, E) Intensity correlation plots (ICP) of red and green signals in the (A) and (B), respectively. (F) PDM image for visualizing the extent of colocalization. Scale bar: 50 μ m. The value of tMr (tMg) indicates the percentage of the red (green) signals colocalized with green (red) signals in the corresponding thresholded image. ICQ value indicated the correlation intensity of two channels and ranged from -0.5 to 0.5 (+0.1 to +0.5 implies a strong covariance).



Figure S46. Confocal images of MCF-7 cells incubated with (A) MTPE-T-Py and (B) MitoTracker Green, and (C) their merged images. (D, E) Intensity correlation plots (ICP) of red and green signals in the (A) and (B), respectively. (F) PDM image for visualizing the extent of colocalization. Scale bar: 50 μ m. The value of tMr (tMg) indicates the percentage of the red (green) signals colocalized with green (red) signals in the corresponding thresholded image. ICQ value indicated the correlation intensity of two channels and ranged from -0.5 to 0.5 (+0.1 to +0.5 implies a strong covariance).



Figure S47. Confocal images of MCF-7 cells incubated with (A) MTPE-T-Qy and (B) MitoTracker Green, and (C) their merged images. (D, E) Intensity correlation plots (ICP) of red and green signals in the (A) and (B), respectively. (F) PDM image for visualizing the extent of colocalization. Scale bar: 50 μ m. The value of tMr (tMg) indicates the percentage of the red (green) signals colocalized with green (red) signals in the corresponding thresholded image. ICQ value indicated the correlation intensity of two channels and ranged from -0.5 to 0.5 (+0.1 to +0.5 implies a strong covariance).



Figure S48. Confocal images of MCF-7 cells incubated with (A) MTPE-DT-Qy and (B) MitoTracker Green, and (C) their merged images. (D, E) Intensity correlation plots (ICP) of red and green signals in the (A) and (B), respectively. (F) PDM image for visualizing the extent of colocalization. Scale bar: 50 μ m. The value of tMr (tMg) indicates the percentage of the red (green) signals colocalized with green (red) signals in the corresponding thresholded image. ICQ value indicated the correlation intensity of two channels and ranged from -0.5 to 0.5 (+0.1 to +0.5 implies a strong covariance).



Fig S49. Confocal images of cancer cells (top: MDA-MB-231 cells, bottom: A549 cells) incubated with MTPE-DT-Py (red, 10 μ M) and MitoTracker Green (green), and their merged images. Scale bar: 50 μ m.



Fig S50. Confocal images of normal cells (top: RAW 264.7 cells, bottom: DC 2.4 cells) incubated with MTPE-DT-Py (red, 10 μ M) and MitoTracker Green (green), and their merged images. Scale bar: 50 μ m.



Figure S51. Photostability of MTPE-DT-Py and MitoTracker Green inside MCF-7 cells. The MCF-7 cells were excited sequentially by 488 nm laser and the emission fluorescence intensity was recorded at different time. Emission fluorescence signal intensity was normalized to the maximum intensity at the beginning of irradiation (n=10).



Figure S52. Confocal images and bright-field of *E. coli* and *S. aureus* incubated with 2×10^{-6} M of AIE-PSs for 20 min. λ_{ex} : 488 nm, λ_{em} : 600–740 nm.