

Supporting Information

In situ generated Fe₃C embedded Fe-N-doped carbon nanozymes with enhanced oxidase mimic activity for total antioxidant capacity assessment

Rui Li⁺, Xiaohong Qiao⁺, Huijun Ma, Hanmei Li, Cong Li and Lihua Jin*

Key Laboratory of Synthetic and Natural Functional Molecule Chemistry of the Ministry of Education, College of Chemistry & Materials Science, Northwest University, Xi'an 710069, People's Republic of China

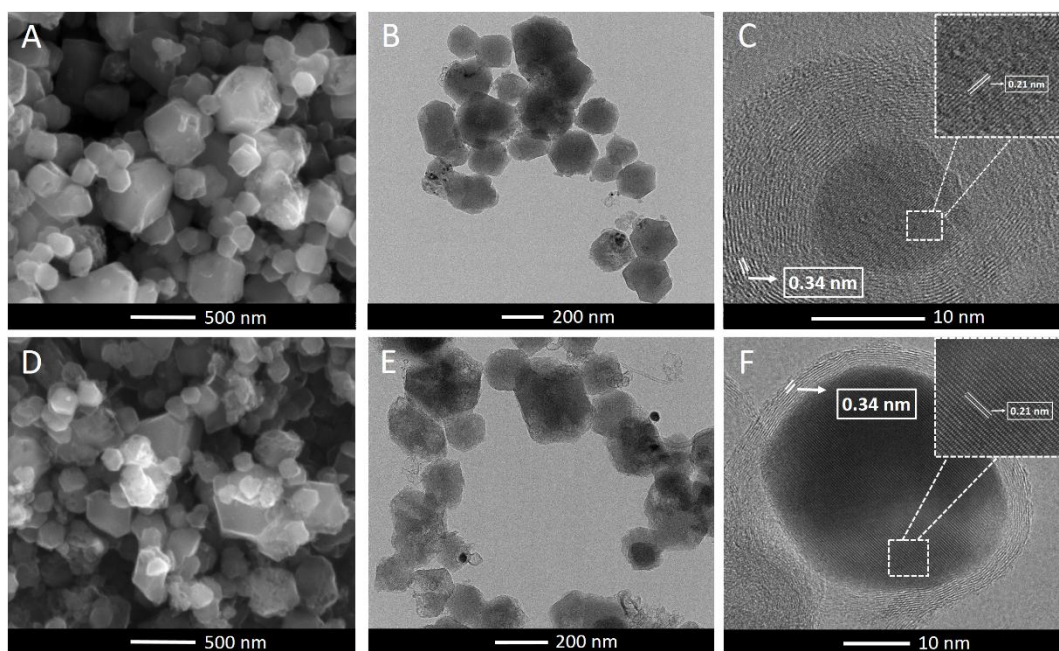


Fig. S1 SEM image, TEM image and HRTEM image of CMs-700 (A-C) and CMs-900(D-E).

* To whom correspondence should be addressed. Fax: +86-029-88302635. E-mail: jinlihua@nwu.edu.cn.

⁺ These authors contributed equally.

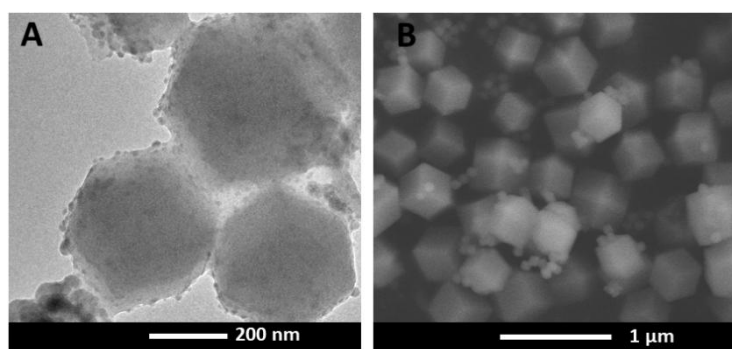


Fig. S2 SEM image (A) and TEM image (B) of ZIF framework

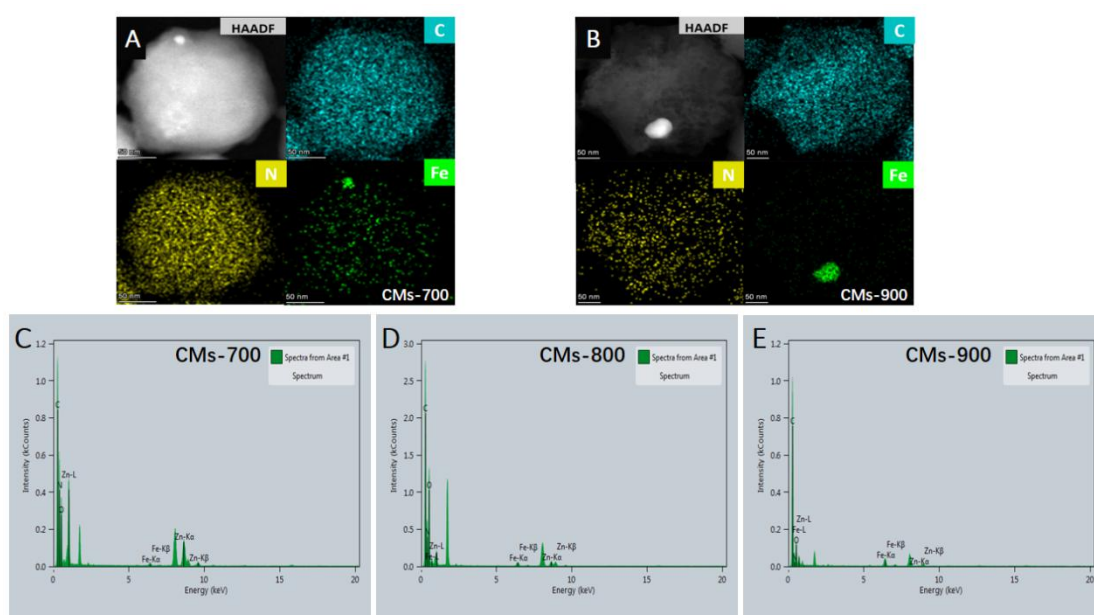


Fig. S3 Element mappings of CMS-700 (A) and CMS-800 (B); EDS images of CMS-700 (C), CMS-800 (D) and CMS-900(E).

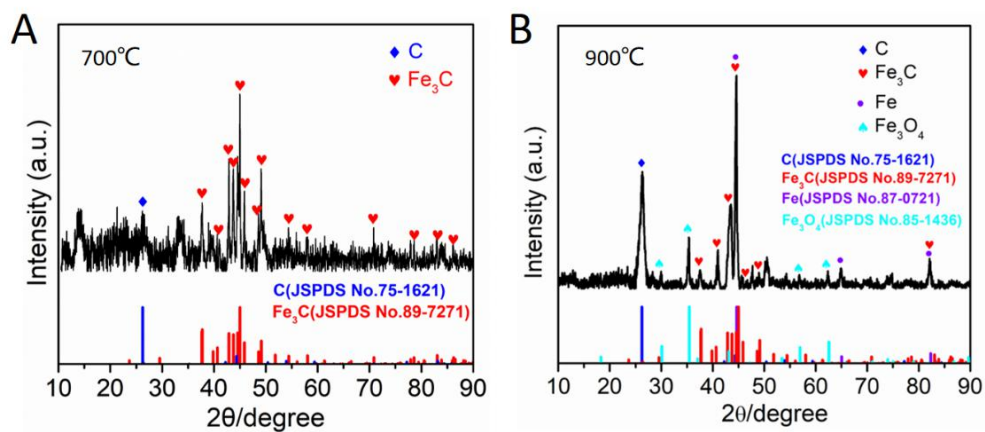


Fig. S4 XRD patterns of CMS-700 (A) and CMS-900 (B).

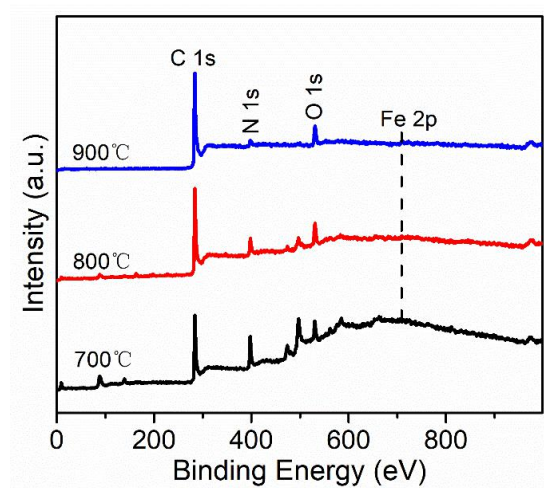


Fig. S5 The overview XPS spectra of the CMS-700, CMS-800 and CMS-900.

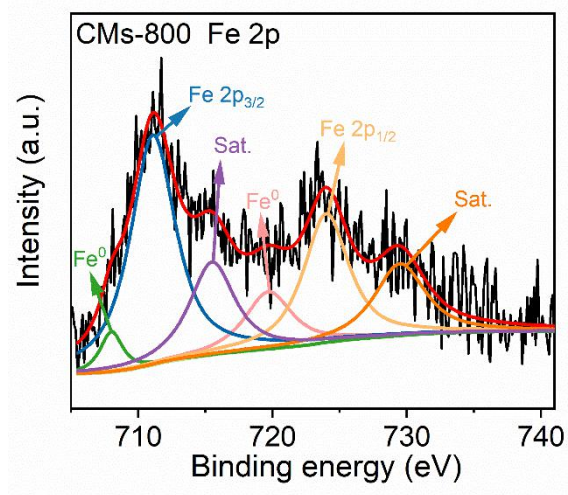


Fig. S6 High-resolution XPS Fe 2p spectra of CMs-800

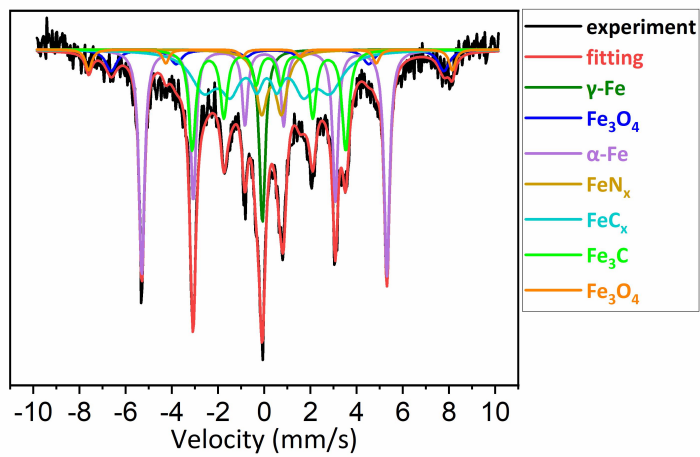


Fig. S7 ^{57}Fe Mössbauer fitted parameters of CMs-800

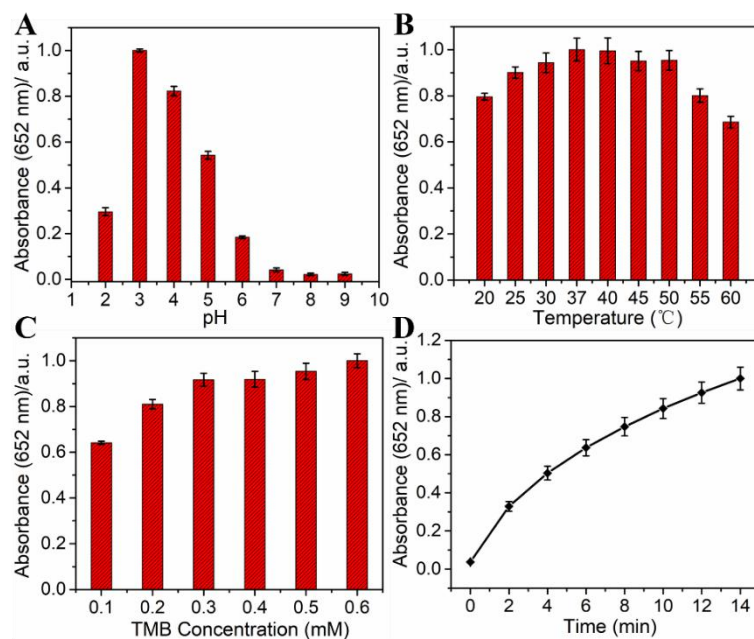


Fig. S8 The effects of reaction condition on the oxidase mimic activity of CMs-800 (A) pH; (B) temperature; (C) TMB concentration; (D) reaction time.

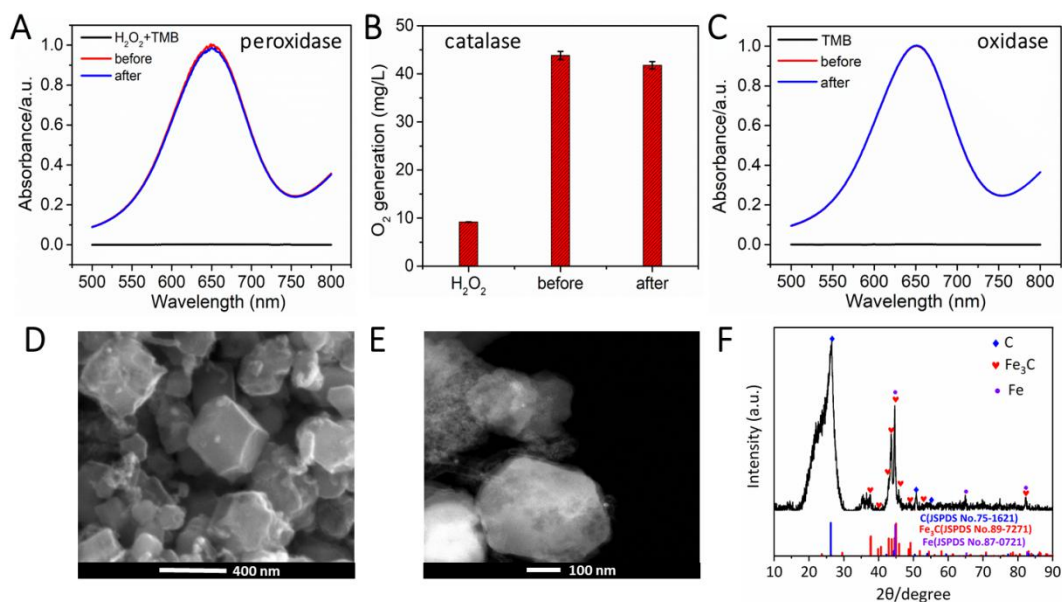


Fig. S9 (A-C) The comparison of peroxidase-like, catalase-like and oxidase-like activities of CMs-800 before and after acid leaching experiment; (D-F) the SEM image, TEM image and XRD data of CMs-800 after acid leaching experiment.

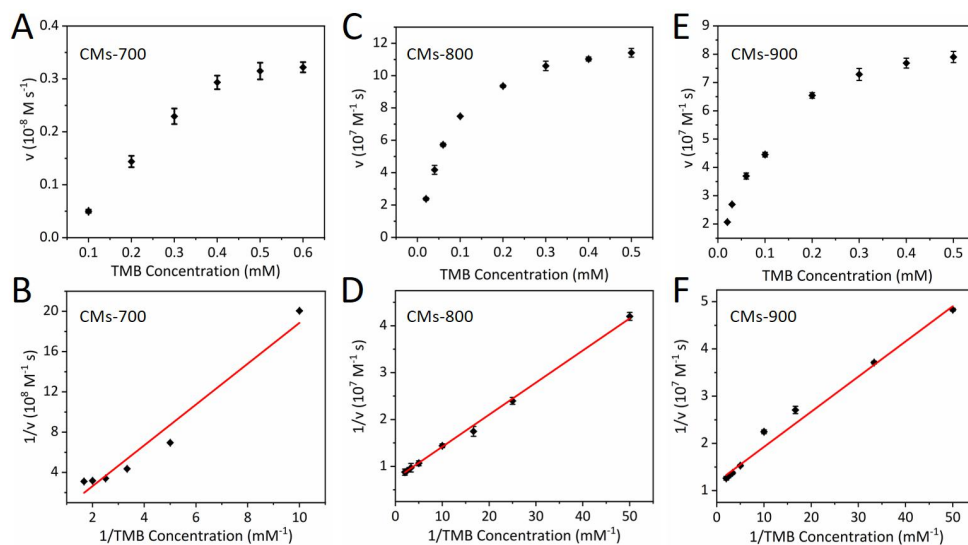


Fig. S10 Oxidase mimic steady-state kinetic assay of the CMs-700(A), CMs-800(C) and CMs-900(E) for TMB. Line weaver-Burk plot of CMs-700(B), CMs-800(D) and CMs-900(F) for the variation of TMB.

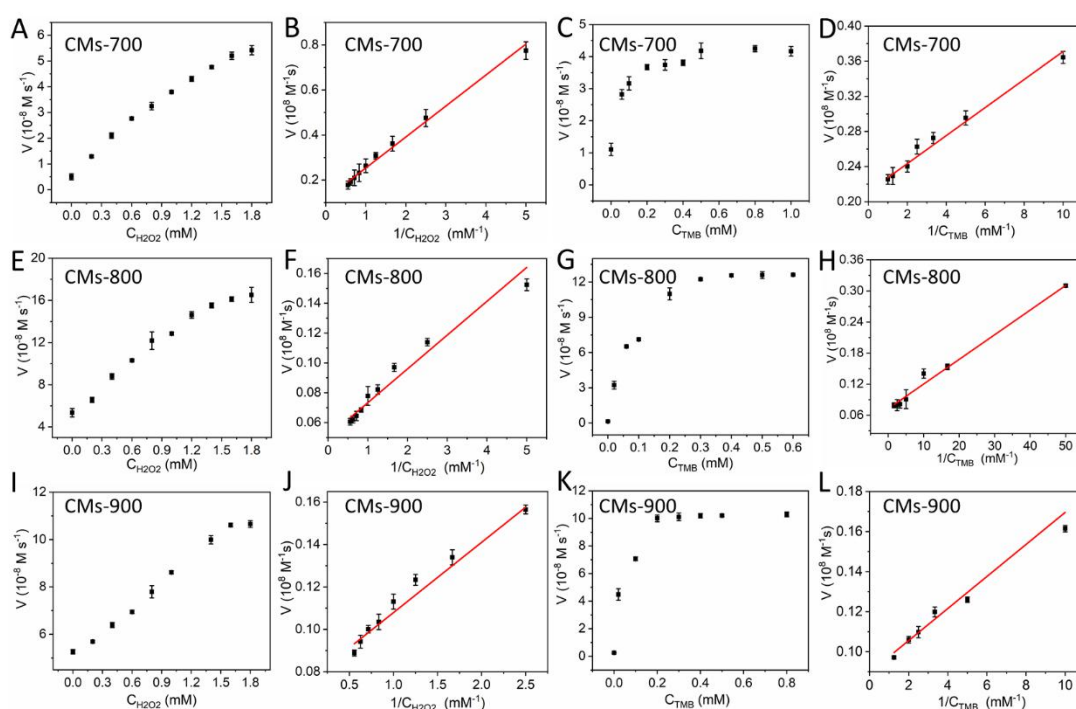


Fig. S11 Peroxidase mimic steady-state kinetic assay of the CMs-700(A), CMs-800(E) and CMs-900(I) for H_2O_2 ; Line weaver-Burk plot of CMs-700(B), CMs-800(F) and CMs-900(J) for the variation of H_2O_2 ; peroxidase mimic steady-state kinetic assay of the CMs-700(C), CMs-800(G) and CMs-900(K) for TMB; Line weaver-Burk plot of CMs-700(D), CMs-800(H) and CMs-900(L) for the variation of TMB.

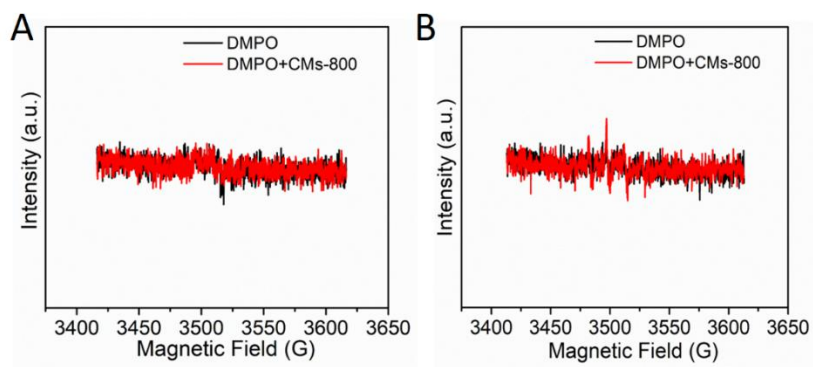


Fig. S12 The EPR spectra of the (A) DMPO+CMs-800 aqueous solution, and (B) DMPO+CMs-800 methanol solution.

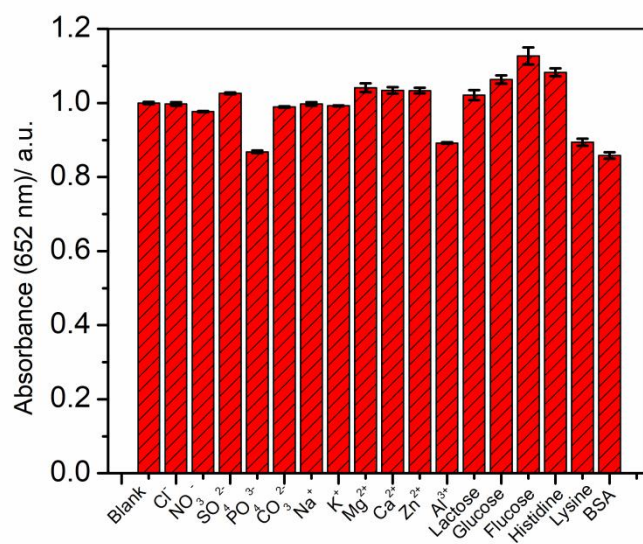


Fig. S13 Selectivity of AA sensing system for common anions, cations and small molecules. All common anions, cations and small molecules were 1mM except BSA was 0.1 mg/mL.

Table S1. ^{57}Fe Mössbauer fitted parameters of CMs-800.

Catalyst	IS(mm·s ⁻¹)	QS(mm·s ⁻¹)	H(T)	Phase
CMs-800	0.18785	0.02730	20.66	Fe ₃ C
	0.1114	~	16.96	FeC _x
	0.31537	0.864	~	FeN _x
	0.00099	~	33.00	α-Fe
	0.46536	0.22015	44.78	tetahed B-site of Fe ₃ O ₄
	0.27929	-0.02579	48.94	tetahed A-site of Fe ₃ O ₄

Table S2 Comparison of oxidase-like steady-state kinetic parameter for Fe₃C@Fe-N-CMs and other oxidase mimics.

Nanozyme	Substrate	K_m (mM)	v_{\max} (10 ⁻⁸ M s ⁻¹)	[E](μg/mL)	Reference
N-PCNSs-3		0.084	0.42	25	[1]
Fe SAEs		0.13	2.25	20	[2]
CeO ₂ NRs-MOF/Cr(VI)		0.108	0.977	1000	[3]
Iron alkoxide		0.12	2.11	—	[4]
Fe-Co-LDH		0.34	5.4	33	[5]
MOF(Co/2Fe)		0.199	0.39	20	[6]
Pt NCs		0.63	2.7	—	[7]
AWNRs-9	TMB	0.109	2.17	50	[8]
Nanoceria		0.42	10.04	5×10 ⁻⁵	[9]
Fe/NC-800		0.170	8.9	5.0	[10]
Commercial Pt/C		0.129	5.2	—	[11]
FeN ₅ SA/CNF		0.148	75.8	5.0	
CMs-700		1.4	0.7	2.0	This work
CMs-800		0.095	14.1	2.0	This work
CMs-900		0.063	8.5	2.0	This work

K_m is the Michaelis constant, v_{\max} is the maximal reaction velocity and k_{cat} is the catalytic constant, where $k_{\text{cat}} = v_{\max}/[E]$, [E] is the molar concentration of metal in nanozymes.

Table S3. The peroxidase-like enzyme kinetic parameters of Fe₃C@Fe-N-CMs

Catalyst	K_m (mM)		v_{max} ($10^{-8}M s^{-1}$)		[E]($\mu g/mL$)
	TMB	H ₂ O ₂	TMB	H ₂ O ₂	
CMS-700	1.183	1.183	4.73	8.66	2.0
CMS-800	0.0651	0.447	18.70	19.73	2.0
CMS-900	0.0893	0.442	11.13	13.37	2.0

K_m is the Michaelis constant, v_{max} is the maximal reaction velocity and k_{cat} is the catalytic constant, where $k_{cat} = v_{max}/[E]$, [E] is the molar concentration of metal in nanozymes.

Table S4. Recovery experiment of AA in diluted commercial vitamin drink

Sample	Original (μM)	Added (μM)	Found (U/L)	Recovery (%)	RSD(%, n=3)
1	67.92	10.00	10.16	101.6	3.5
2	67.92	20.00	20.54	102.7	2.8
3	67.92	30.00	30.38	101.3	4.3

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