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Supporting information for

Magnet-actuated microfluidic array chip for high-throughput pretreatment,

amplification and detection of multiple pathogens

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Bacteria	Target	GeneBna	Server (5, 2,)
	Gene	Acession	
Escherichia coli	mal B	J01648	GCCATCTCCTGATGACGCATAGTCAGC
			CCATCATGAATGTTGCTGTCGATGACA
			GGTTGTTACAAAGGGAGAAGGGCATG
			GCGAGCGTACAGCTGCAAAATGTAAC
			GAAAGCCTGGGGGCGAGGTCGTGGTAT
			CGAAAGATATCAATCTCGATATCCAT
			GAAGGTGAATTCGTGGTGTTTTGTCGG
			ACCGTCTGGCTGCGGTAAAT
Salmonella typhimurium	inv A	M90846	GGCGATATTGGTGTTTATGGGGTCGTT
			CTACATTGACAGAATCCTCAGTTTTTC
			AACGTTTCCTGCGGTACTGTTAATTAC
			CACGCTCTTTCGTCTGGCATTATCGAT
			CAGTACCAGTCGTCTTATCTTGATTGA
			AGCCGATGCCGGTGAAATTATCGCCA
			CGTTCGGGCAATTCGTTATTGGCGATA
			GCCTGGCGGTGGGTTTTGTTGTCTTCT
			CTATTGTCACCGTGGTCCAGTTTATCG
			TT

Table S1 Sequences of the specific genes used for pathogen detection in this paper



Figure S1. The Standard curve of bacteria. (a) Salmonella typhimurium. (b) Escherichia coli.



Figure S2. (a) Miniature peltier heater setup. (b)Chip imaging device.



Figure S3. LAMP test for PDMS compatibility. (a) Left to right (bright field): Amplification result with mixing layer, amplification result with mixed layer-free and blank control. (b) Dark field image under UV excitation. (c) Electrophoresis of LAMP reaction. Lane M: DNA marker, lane 1: Blank control, lane 2: Amplification result with mixing layer. lane3: Amplification result mixed without mixed layer.



Figure S4. Specificity of LAMP reaction in the amplification tube. lane M: DNA marker, lane 1-3: *invA* primer test results. Lane 1: blank control, lane 2: *malB* template, lane 3: *invA* template. Lane 4-6: *malB* primer test results. Lane 4: blank control, lane 5: *invA* template, lane 6: malB template.