

## Supporting Information

### **Unravelling the interactions of biodegradable dendritic nucleic acids carriers and neural cells**

*Ana Patrícia Spencer, Victoria Leiro\*, Ana Paula Pêgo\**

A.P. Spencer, Dr. V. Leiro, Dr. A.P. Pêgo

i3S – Instituto de Investigação e Inovação em Saúde, Universidade do Porto, Rua Alfredo Allen, 208, 4200-135 Porto, Portugal

INEB – Instituto de Engenharia Biomédica, Universidade do Porto, Rua Alfredo Allen, 208, 4200-135 Porto, Portugal

A.P. Spencer

FEUP – Faculdade de Engenharia, Universidade do Porto, Porto, R. Dr. Roberto Frias s/n, 4200-465 Porto, Portugal

Dr. A.P. Pêgo

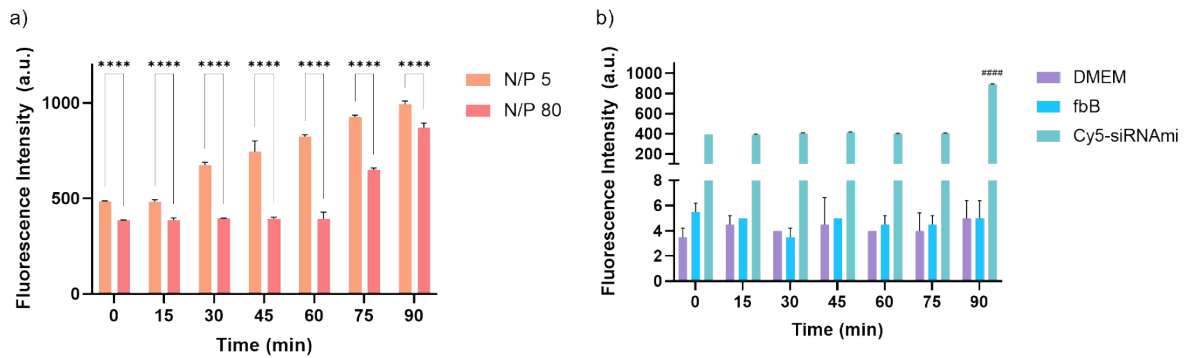
ICBAS – Instituto de Ciências Biomédicas Abel Salazar, Universidade do Porto, R. Jorge de Viterbo Ferreira 228, 4050-343 Porto, Portugal

\*Corresponding authors

## Table of Contents

	Page
1) Deposition assay of Cy5-siRNAmi drndriplexes	3
2) Internalization kinetics data in neuronal cell lines	4
3) Internalization kinetics data in primary cortical neurons	5
4) Endocytic mechanism in neuronal cell lines	7
5) Endocytic mechanism in primary cortical neurons	9
6) Intracellular path in neuronal cell lines (late endosomes)	10
7) Intracellular path in primary cortical neurons (late endosomes and autophagosomes)	11
8) Endocytosis study with inhibitors	13

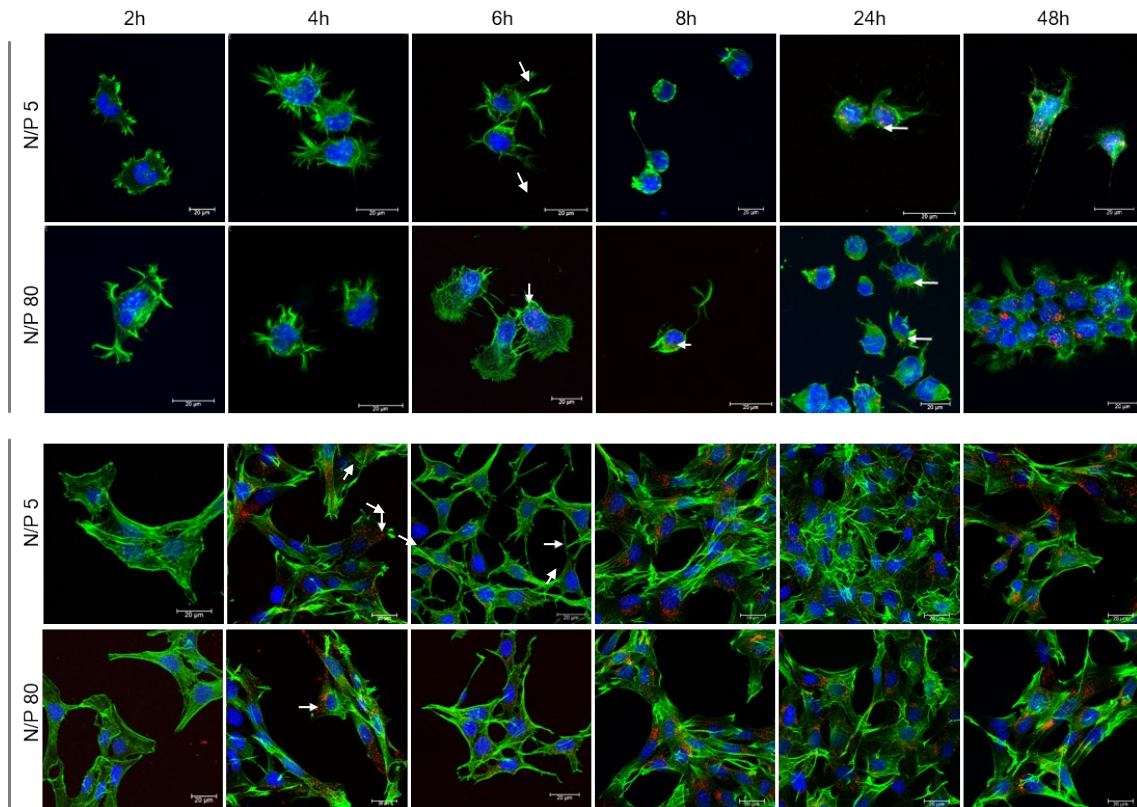
## 1) Deposition assay of Cy5-siRNAmi dendriplexes



**Figure S1.** Deposition assay of a) Cy5-siRNAmi dendriplexes at N/P 5 and 80. Dendriplexes were incubated in medium (DMEM with GlutaMAX™) at 37 °C and the fluorescence intensity (5 x 5 spots,  $\lambda_{exc} = 633$  nm,  $\lambda_{em} = 647$  nm) of the well bottom was measured every 15 minutes, up to 1.5 hours. b) DMEM with GlutaMAX™, free dendrimer and Cy5-siRNAmi were used as controls. Significant differences: ####, \*\*\*\* $p \leq 0.0001$ . For free Cy5-siRNA, the symbol # denotes significant differences between times of incubation.

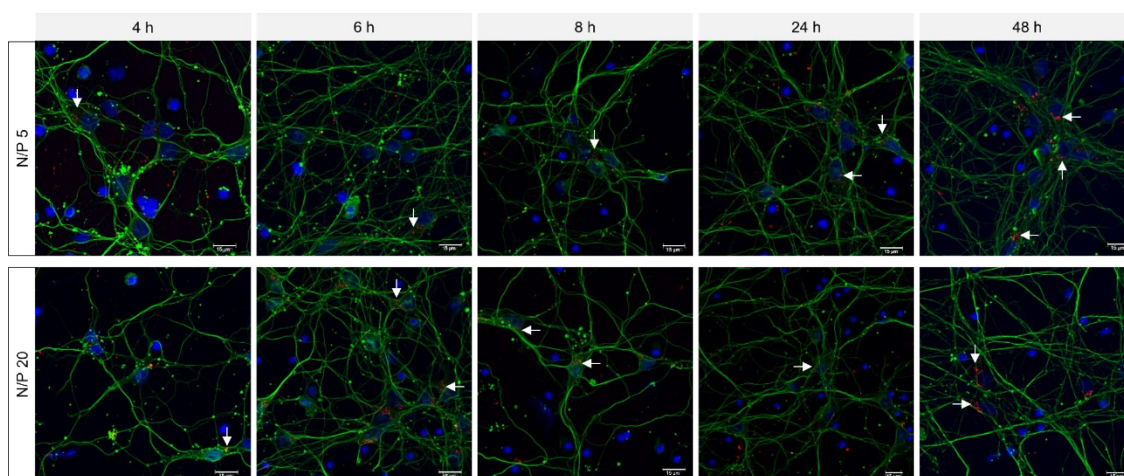


## 2) Internalization kinetics data in neuronal cell lines

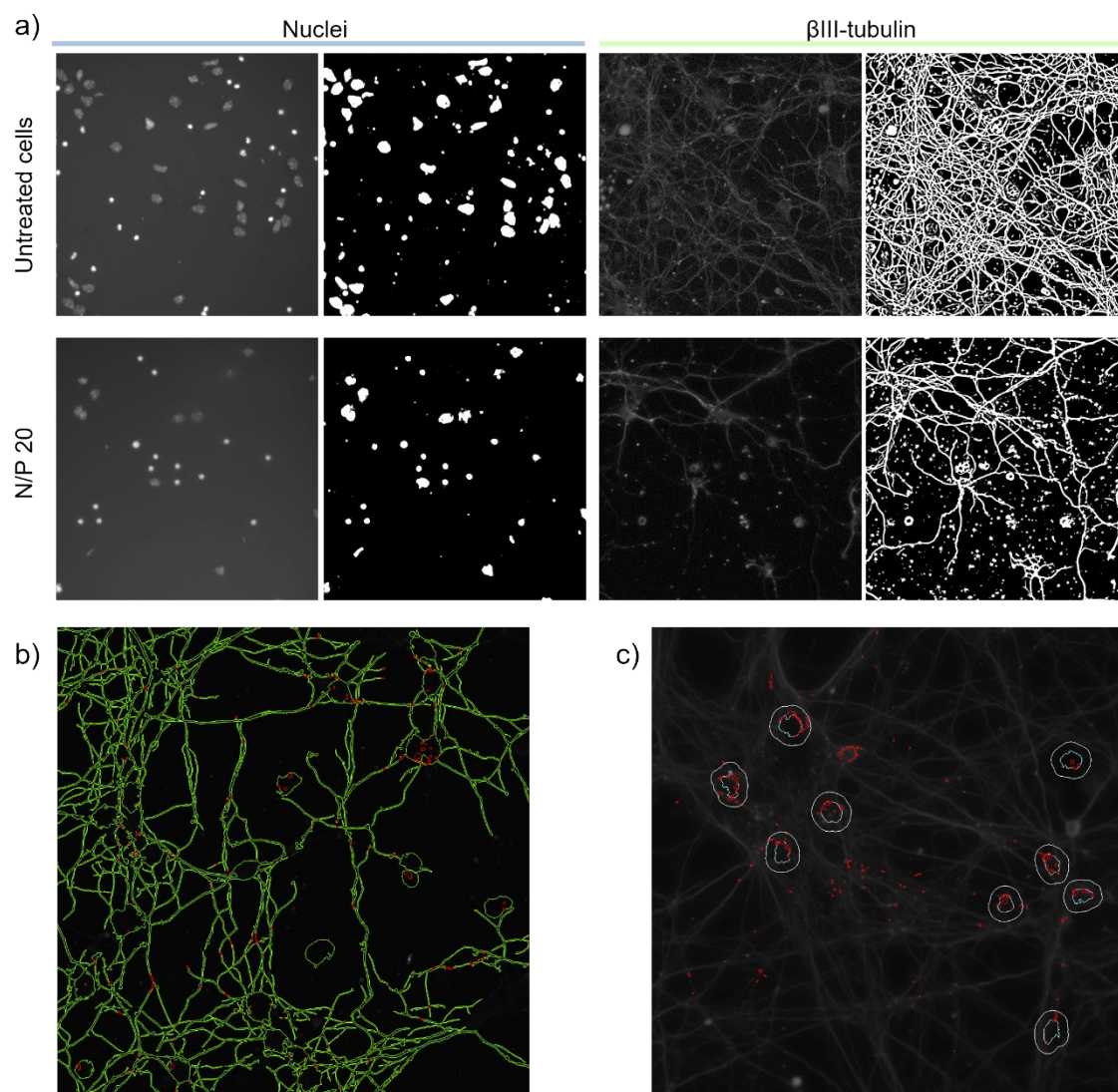


**Figure S2.** Confocal microscopy images of ND7/23 and HT22 cell lines after incubation with Cy5-siRNAmi-fbB dendriplexes at N/P 5 and 80 at different time points (2, 4, 6, 8, 24 and 48 hours). Nuclei stained with Hoechst 33342 (blue). Actin filaments stained with Alexa Fluor™ 488 Phalloidin (green). White arrows point to Cy5-siRNAmi dendriplexes (red). Scale bar: 20 μm.

### 3) Internalization kinetics data in primary cortical neurons

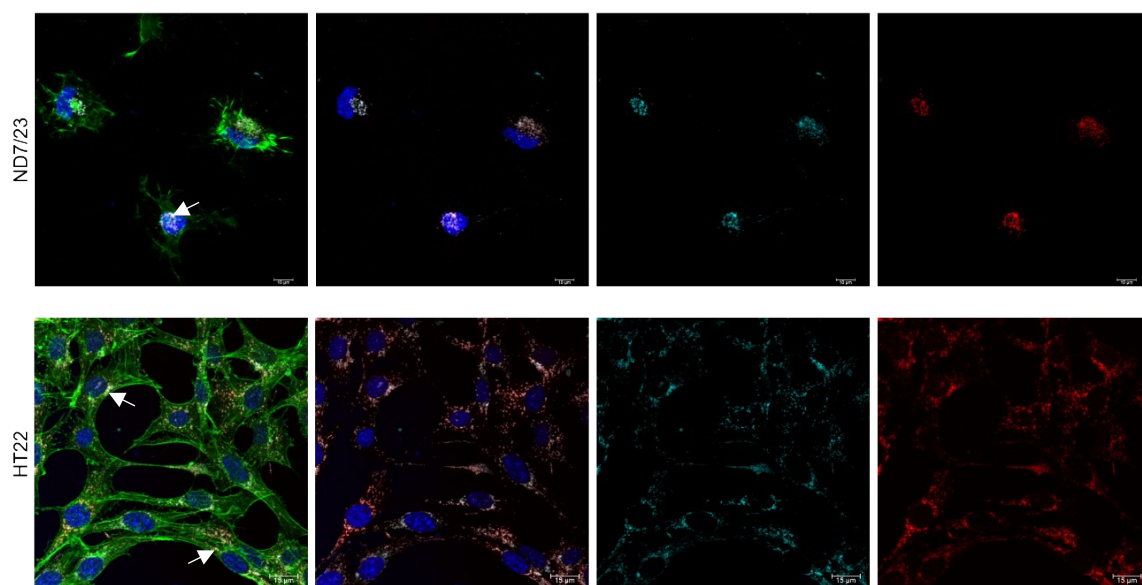


**Figure S3.** Confocal microscopy images of primary cortical neurons after incubation with Cy5-siRNAmi-fbB dendriplexes at N/P 5 and 80 at different time points (4, 6, 8, 24 and 48 hours). Staining: Nuclei with Hoechst 33342 (blue),  $\beta$ III-tubulin (green), and Cy5-siRNAmi dendriplexes (red). Scale bar: 15  $\mu$ m.



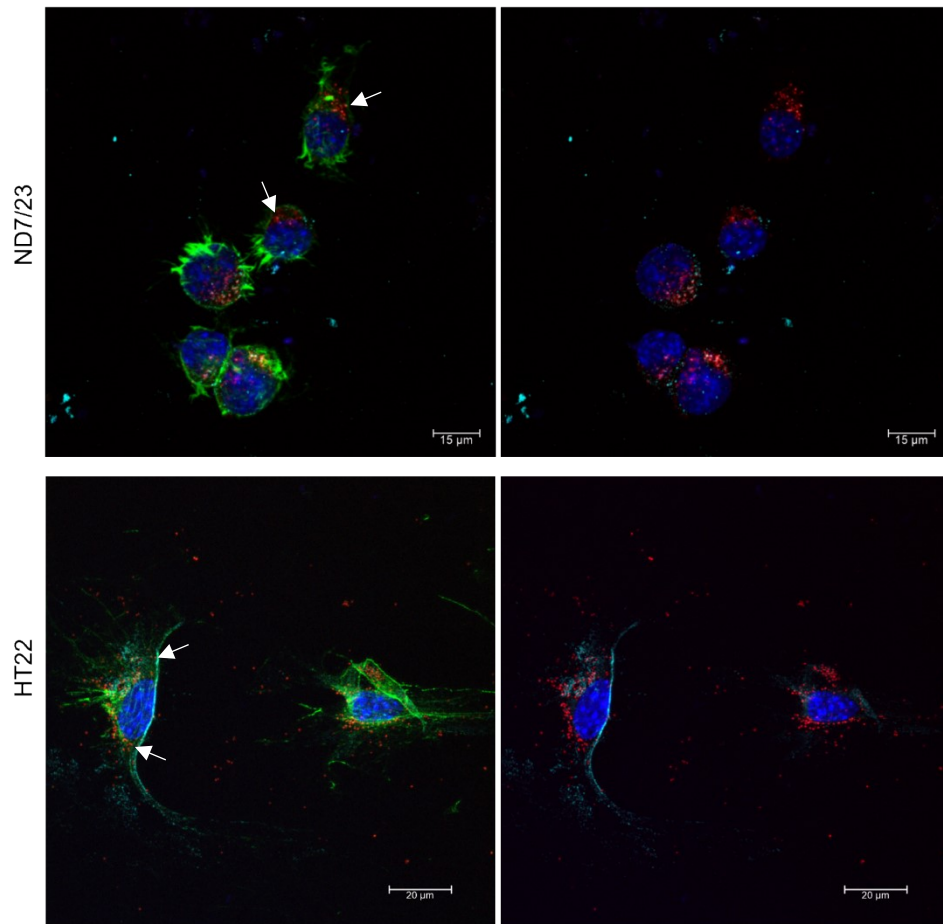
**Figure S4.** Representative images used in the analysis of internalization kinetics in primary cortical neurons. Cells were incubated with Cy5-siRNAmi-fbB dendriplexes at N/P 5 and 20 at different time points (4, 6, 8, 24 and 48 hours). Untreated cells were used as control. Staining: Nuclei with Hoechst 33342,  $\beta$ III-tubulin, and Cy5-siRNAmi dendriplexes. Acquired images were a) segmented and then Cy5 fluorescence intensity (red) was quantified in b) neuronal extension (green) and in c) perinuclear area (white).

#### 4) Internalization mechanism in neuronal cell lines



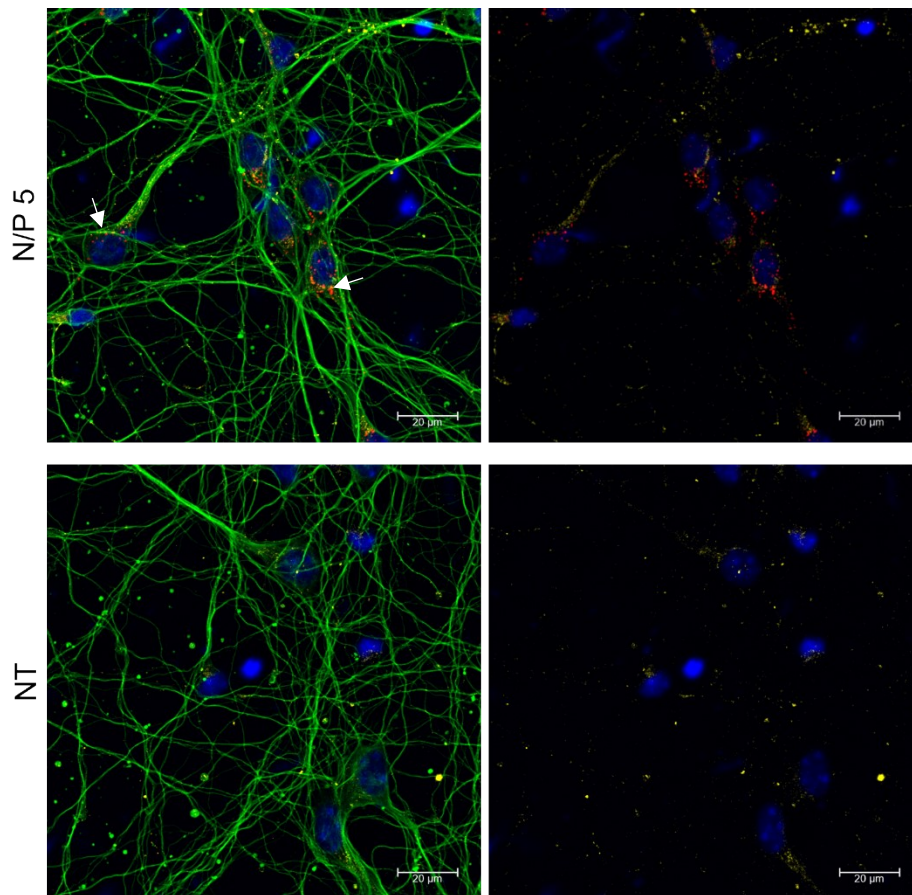
**Figure S5.** Confocal microscopy images of ND7/23 and HT22 cells after incubation with Cy5-siRNAmi-fbB dendriplexes at N/P 5 for 24 hours. Staining: nuclei with Hoechst 33342 (blue), actin filaments with Alexa Fluor™ 488 Phalloidin (green), Clathrin (cyan), and Cy5-siRNAmi dendriplexes (red). Scale bar: 15  $\mu$ m.





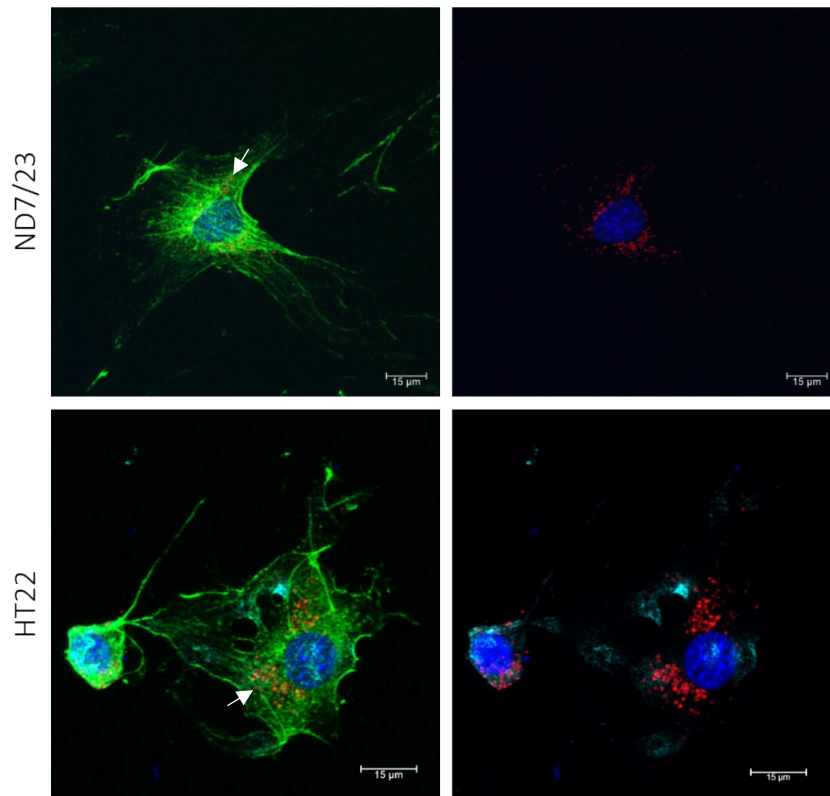
**Figure S6.** Confocal microscopy images of ND7/23 and HT22 cells after incubation with Cy5-siRNAmi-fbB dendriplexes at N/P 5 for 24 hours. Staining: nuclei with Hoechst 33342 (blue), actin filaments with Alexa Fluor™ 488 Phalloidin (green), Caveolin-1 (cyan), and Cy5-siRNAmi dendriplexes (red). Scale bar- ND7/23: 15 μm; HT22: 20 μm.

## 5) Internalization mechanism in primary cortical neurons



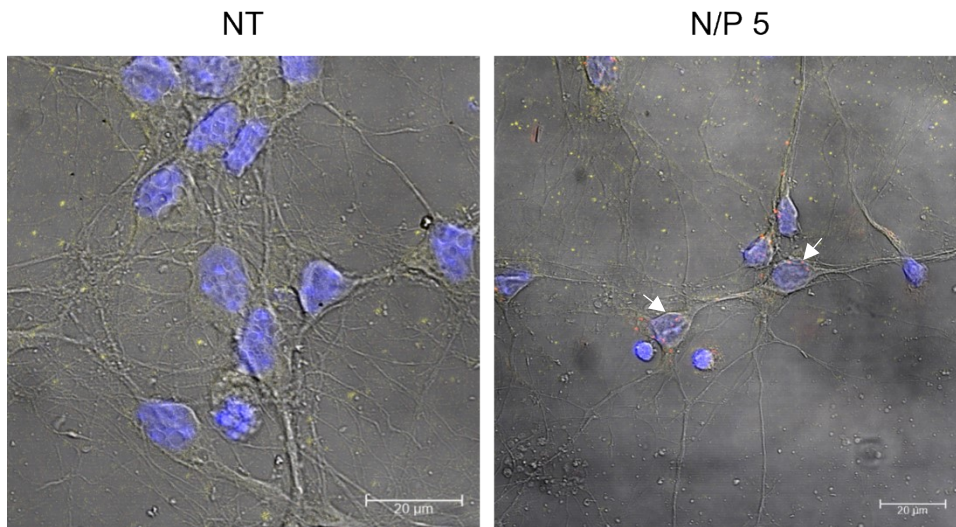
**Figure S7.** Confocal microscopy images of primary cortical neurons after incubation with Cy5-siRNAmi-fbB dendriplexes at N/P 5 for 24 h. Untreated cells (NT) were used as control. Staining: nuclei with Hoechst 33342 (in blue),  $\beta$ III-tubulin (in green), Clathrin (in yellow), and Cy5-siRNAmi dendriplexes (in red). Scale bar: 20  $\mu$ m.

6) Intracellular path in neuronal cell lines (late endosomes)

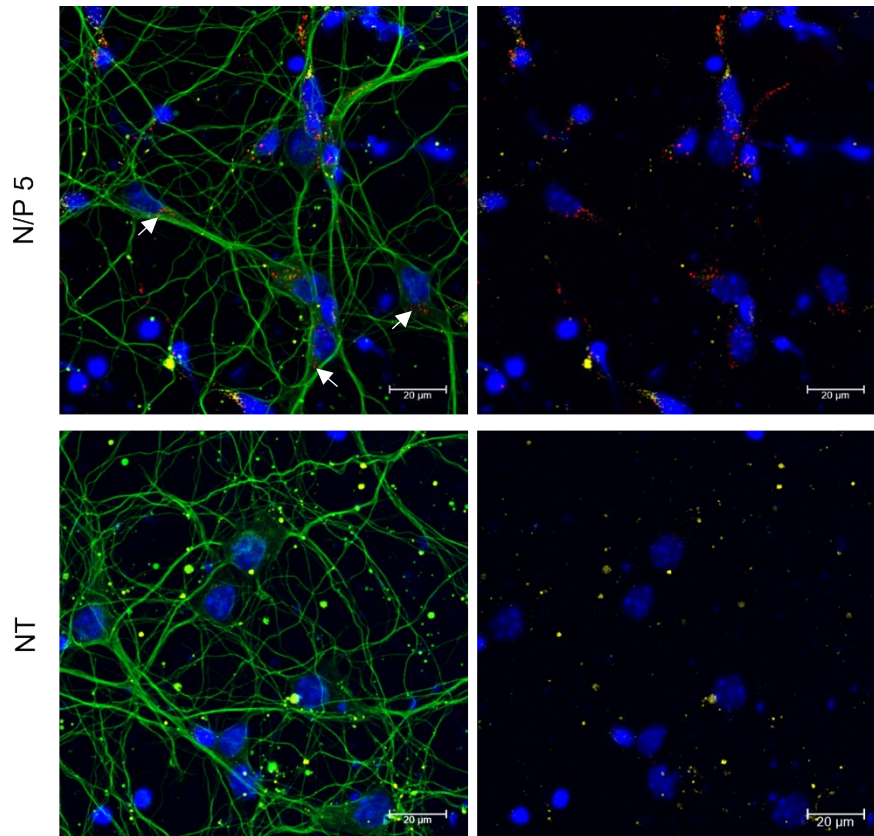


**Figure S8.** Confocal microscopy images of ND7/23 and HT22 cells after incubation with Cy5-siRNAmi/fbB dendriplexes at N/P 5 for 24 h. Staining: nuclei with Hoechst 33,342 (in blue), actin filaments with Alexa Fluor™ 488 Phalloidin (green), LAMP-1 (cyan), and Cy5-siRNAmi dendriplexes (red). Scale bar: 15 µm.

7) Intracellular path in primary cortical neurons (late endosomes and autophagosomes)



**Figure S9.** Confocal microscopy images of primary cortical neurons after incubation with Cy5-siRNAmi/fbB dendriplexes at N/P 5 for 24 hours. Untreated cells (NT) were used as control. Staining: nuclei with Hoechst 33342 (blue), late endosomes – LAMP-1 (yellow), and Cy5-siRNAmi dendriplexes (red). Scale bar: 20 µm.

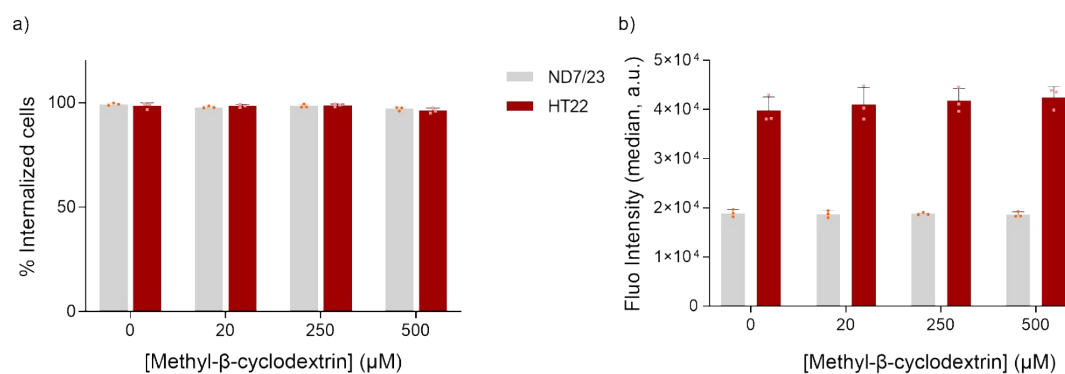


**Figure S10.** Confocal microscopy images of primary cortical neurons after incubation with Cy5-siRNAmi-fbB dendriplexes at N/P 5 for 24 hours. Untreated cells (NT) were used as control. Staining: nuclei with Hoechst 33342 (blue), autophagosomes - LC3 (yellow), and Cy5-siRNAmi dendriplexes (red). Scale bar: 20  $\mu\text{m}$ .

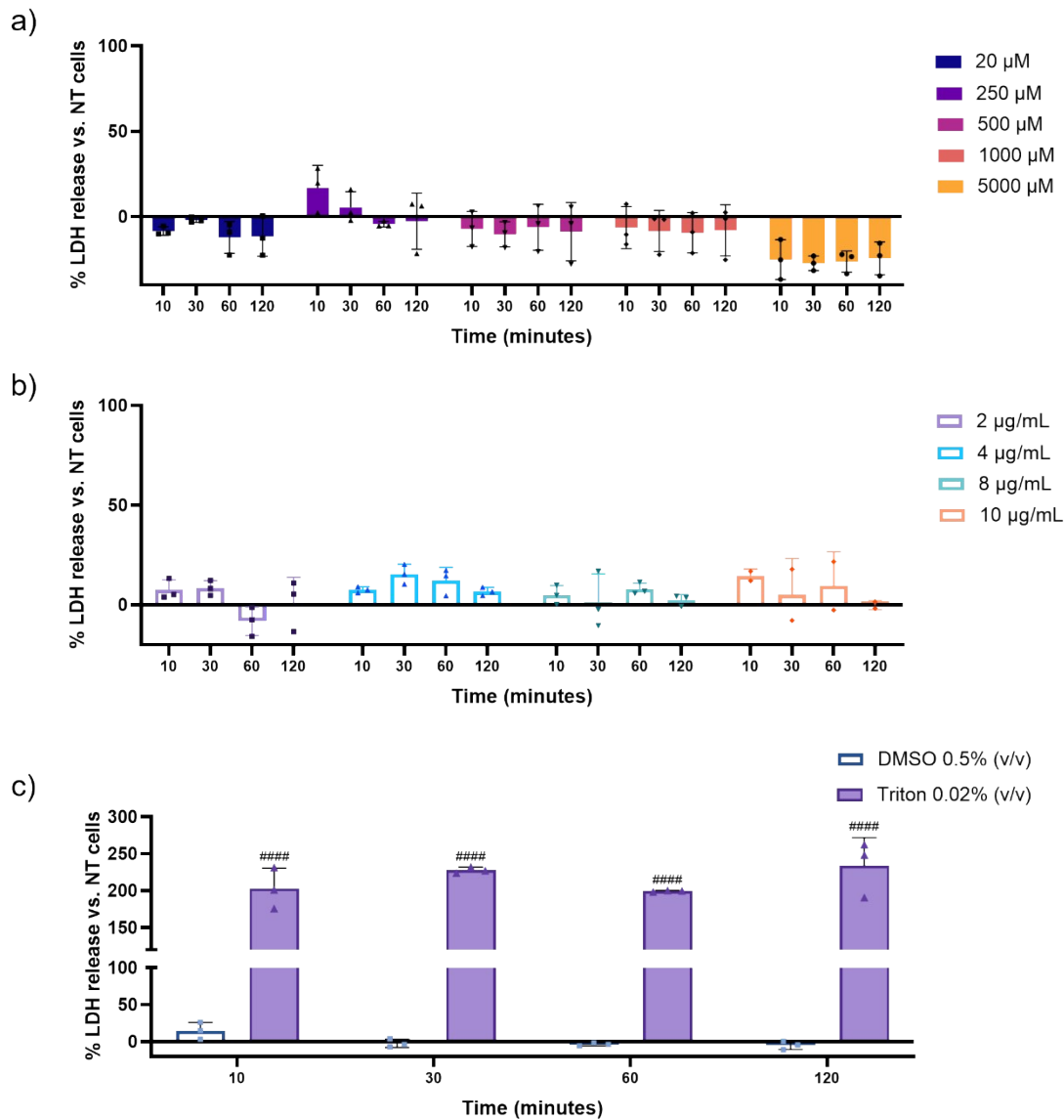
## 8) Endocytosis study with inhibitors

**Table S1.** Inhibitors of endocytosis (dependent on clathrin or caveolin) tested and the concentrations used by us in ND7/23 and HT22 cells. Despite the effects recorded on these cells, the use of these inhibitors is described in many other cells (mostly, non-neuronal cells).

Inhibitor	Endocytic mechanism	Concentrations tested	Effect in cells	Cells already tested	References
Chlorpromazine	Clathrin	2, 4, 6, 8 and 10 µg/mL	90-100% of cells died in less than 10 mins	HeLa, A549, 1321N1, COS-7, Vero, HuH-7, ARPE-19, D407, CIK, SK-Hep1, HEK293	dos Santos et al., 2011; Vercauteren et al., 2010; Rejman et al., 2005; Wang et al., 2016; Maddila et al., 2021; Itoh et al., 2008
Phenylarsine oxide	Clathrin	0.05, 0.1, 1 and 5 µM		H358, Calu-3, SNU-1327, H1703, adrenal chromaffin cells	Prichard et al., 2021; Kim et al., 2021
Genistein	Caveolin	50, 100, 200 and 400 µM	~90% of cells died in less than 30 mins	D407, Vero, COS-7, HuH-7, ARPE-19, A549, HCT116, COLO205, Sf1Ep	Vercauteren et al., 2010; Horibe et al., 2018; Singh et al., 2012
Filipin III	Caveolin	0.5, 1, 2, 5 and 10 µg/mL		A549, CIK, Sf1Ep, SK-Hep1	Rejman et al., 2005; Wang et al., 2016; Singh et al., 2012; Madilla et al., 2021
Methyl-β-Cyclodextrin	Caveolin	20, 250, 500, 1000 and 5000 µM	90-100% of cells died in less than 10 mins (concentrations above 1000 µM). At 20-500 µM, no uptake differences were observed	D407, Vero, COS-7, HuH-7, ARPE-19, A549, BY-2, HEK293, Sf1Ep	Vercauteren et al., 2010; Chen et al., 2018; Itoh et al., 2008; Singh et al., 2012; Rejman et al., 2005



**Figure S11.** Cellular interaction of N/P 5 Cy5-siRNAmi dendriplexes by cell lines pre-treated with a caveolin-mediated endocytosis inhibitor. ND7/23 and HT22 cells were incubated for 10 minutes with methyl-β-cyclodextrin (MBCD, 20-500 μM), and then incubated for 6 hours with dendriplexes carrying Cy5-siRNAmi ([Cy5-siRNAmi]<sub>f</sub> = 100 nM). Characterization by flow cytometry: a) Percentage of cells with fluorescence signal of Cy5-siRNAmi dendriplexes and b) Intensity of fluorescence in cells. Results are shown as mean ± SD of three independent experiments (n=3). Two-way ANOVA tests were used for statistical analysis. No significant differences between the non-treated cells (0 μM) and the MBCD-treated cells were observed.



**Figure S12.** Cytotoxic profile evaluation in primary cortical neuron cultures. Plasma membrane integrity assessed by LDH assay, after exposure of the cultures to: a) methyl- $\beta$ -cyclodextrin (MBCD) or b) chlorpromazine. Non-treated (NT) cells, cells treated with dimethylsulfoxide (DMSO, 0.5% (v/v)) and Triton™ X-100 (0.02% (v/v)) were used as controls. Results are represented as mean  $\pm$  SD of three independent experiments ( $n = 3$ ). Two-way ANOVA tests were used for statistical analysis. Significant differences: ####  $p \leq 0.0001$ . The symbol # indicates significant differences versus the NT cells.