## SUPPLEMENTARY INFORMATION for

A NanoCurvS platform for quantitative and multiplex analysis of curvature-sensing proteins


Supplementary Figure 1. Schematic illustrations of SLBs on nanostructures.
(A) Schematic illustrations of the NanoCurvS assay. (B) Illustrations of solvent-assisted lipid bilayer (SALB) formation method.


Supplementary Figure 2. SLBs on nanostructures show similar fluidities as surrounding flat areas. (A) Representative fluorescence images of lipid bilayers on a nanoX array at different time points after photobleaching. (B) Representative fluorescence images of lipid bilayers on a gradient nanoU array at different time points after photobleaching. For both (A) and (B), the bilayer consists of 70\% DOPC, 15\% DOPS, $15 \% \mathrm{PI}(4,5) \mathrm{P}_{2}$ and $\sim 1 \%$ Texas Red-DHPE. White-dashed circles indicate the bleached region. (C) A plot of the time-lapsed fluorescence recovery signals. Scale bar: $20 \mu \mathrm{~m}$.


Supplementary Figure 3. Purified protein and lysates of FBP17 BAR show similar curvature sensitivity on nanobar arrays.
(A) Illustration and a SEM image of a 200-nm nanobar array. All nanobars are 200 nm in width, $1 \mu \mathrm{~m}$ in height and $5 \mu \mathrm{~m}$ in spacing. Scale bar: $5 \mu \mathrm{~m}$. (B) Constructs of the GFP-FBP17 and GFP-FBP17(BAR)-6XHis variants used in this study. (C) Representative fluorescence images of GFP-FBP17(BAR)-6XHis purified protein and lysates on SLB-coated nanobar arrays. The lipid bilayer is made of $90 \%$ DOPC, 10\% DGS-Ni-NTA and doped with ~1\% Texas Red-DHPE for bilayer visualization. Scale bar: $5 \mu \mathrm{~m}$. (D) Averaged images of GFP-FBP17(BAR)-6XHis purified protein and lysates on SLB-coated nanobar arrays. (E) Quantification of fluorescence intensity of GFP-FBP17(BAR)-6xHis purified protein and lysates on SLB-coated nanobar arrays. Error bars represent standard deviation. Welch's t tests (unpaired, two-tailed, not assuming equal variance) are applied for all statistical analyses.


Supplementary Figure 4. Representative fluorescence images of U2OS cells expressing IRSp53(BAR)-GFP, IRSp53(FL)-GFP, and GFP-FBP17(FL). Scale bar: $20 \mu \mathrm{~m}$.


Supplementary Figure 5. Western blots of protein lysates confirming the presence of transfected proteins.
(A) IRSp53-GFP variants. (B) Full-length GFP-FBP17. Both GFP-FBP17 and IRSp53-GFP variants were probed by rabbit anti-GFP antibodies. GAPDH was stained with mouse anti-GAPDH antibodies as a control.


Supplementary Figure 6. Quantification of TXR-lipid bilayer signals on gradient nanoU and nanoX arrays.
(A-B) $70 \%$ DOPC, $15 \%$ DOPS, $15 \% \mathrm{PIP}_{2}$ and $\sim 1 \%$ Texas Red-DHPE on (A) gradient nanoX arrays or (B) gradient nanoU arrays. (C) 55\% DOPC, 15\% DOPS, 15\% PIP ${ }_{2}$, 15\% DGS-Ni-NTA, and $\sim 1 \%$ Texas Red-DHPE on gradient nanoU arrays. Error bars represent standard error of mean.


Supplementary Figure 7. Representative whole-field fluorescence images of BAR protein lysates on SLB-coated nanoX arrays. (A) IRSp53(BAR)-GFP; (B) GFP-FBP17(FL). Scale bar: $20 \mu \mathrm{~m}$.


Supplementary Figure 8. Representative whole-field and zoom-in fluorescence images of BAR protein lysates on SLB-coated nanoU arrays. (A) IRSp53(BAR)-GFP; (B) GFP-FBP17(FL); (C) IRSp53(FL)-6XHis-GFP; (D) IRSp53(FL)-GFP; (E) Empty vector. Scale bar: $20 \mu \mathrm{~m}$.


Supplementary Figure 9. Representative fluorescence images from the c-Abl kinase experiments on SLB-coated nanoU arrays. (A) c-Abl-catalyzed phosphorylation upon ATP addition significantly reduces the intensity of GFP-FBP17 on the lipid bilayer. (B) GFP-FBP17 fluorescence intensity does not change upon the addition of a control buffer. Scale bar: $20 \mu \mathrm{~m}$.

| Designed values |  |  | Measured values (Diameter of curvature) |  |  | Number of <br> nanostructures <br> measured |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Arm <br> width <br> $(\mathrm{nm})$ | Crossing <br> angle $(\boldsymbol{\theta})$ | Complementary <br> crossing angle $(\boldsymbol{\theta})$ | Arm end (nm) | Crossing angle (nm) | Complementary <br> crossing angle (nm) | 10 |
| 400 | 15 | 165 | $447 \pm 43.2$ | $354 \pm 15.8$ | $5749 \pm 1449$ | 10 |
|  | 30 | 150 | $397 \pm 26.3$ | $436 \pm 51.8$ | $3797 \pm 840$ | 10 |
|  | 45 | 135 | $438 \pm 36.2$ | $491 \pm 45.4$ | $2680 \pm 293$ | 10 |
|  | 60 | 120 | $480 \pm 46.1$ | $648 \pm 45.2$ | $1050 \pm 270$ | 10 |



Supplementary Table 1. Characterization of gradient nanoX arrays.
Error bars represent standard deviation.

| Desired <br> Arm width (nm) | Measured values (Diameter of curvature) |  |  | Number of nanostructures <br> measured |
| :---: | :---: | :---: | :---: | :---: |
|  | Arm end (nm) | Inner groove (nm) | Outer face (nm) |  |
| 400 | $382 \pm 39.4$ | $388 \pm 39.9$ | $1148 \pm 75.5$ | $1384 \pm 92.0$ |
| 450 | $443 \pm 28.0$ | $516 \pm 62.2$ | $1487 \pm 31.9$ | 10 |
| 500 | $489 \pm 21.1$ | $593 \pm 32.8$ | $1672 \pm 141$ | 10 |
| 550 | $562 \pm 23.0$ | $623 \pm 59.2$ | $1811 \pm 89.4$ | 10 |
| 600 | $599 \pm 38.6$ | $663 \pm 54.2$ | $1928 \pm 74.2$ | 10 |
| 650 | $644 \pm 36.0$ | $708 \pm 39.1$ | $2136 \pm 85.2$ | 10 |
| 700 | $711 \pm 29.5$ | $783 \pm 49.3$ | $2230 \pm 168$ | 10 |
| 750 | $757 \pm 33.5$ | $823 \pm 61.2$ | $2378 \pm 152$ | 10 |
| 800 | $795 \pm 44.1$ | $859 \pm 67.1$ |  | 10 |



Supplementary Table 2. Characterization of gradient nanoU arrays.
Error bars represent standard deviation.
A

| Target | Desired Crossing angle ( $\theta$ ) | Normalized intensity (A.U.) |  |  |  | Number of fields of view considered ( N ) | Number of nanostructures measured ( n ) | Corresponding figure |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | NanoX region | Mean | SD | SEM (SD/ $\sqrt{N}$ ) |  |  |  |
| IRSp53 (BAR)GFP | 15 | End | 1.56 | 0.80 | 0.23 | 12 | 168 | Fig. 1E |
|  |  | Inner face | 5.50 | 3.49 | 1.01 |  |  |  |
|  |  | Complementary inner face | 0.83 | 0.56 | 0.16 |  |  |  |
|  | 30 | End | 0.88 | 0.15 | 0.04 | 12 | 182 |  |
|  |  | Inner face | 4.29 | 2.71 | 0.78 |  |  |  |
|  |  | Complementary inner face | 0.71 | 0.30 | 0.09 |  |  |  |
|  | 45 | End | 0.83 | 0.12 | 0.03 |  |  |  |
|  |  | Inner face | 3.91 | 2.16 | 0.62 | 12 | 196 |  |
|  |  | Complementary inner face | 0.92 | 0.25 | 0.07 |  |  |  |
|  |  | End | 0.86 | 0.10 | 0.03 |  |  |  |
|  | 60 | Inner face | 3.36 | 1.62 | 0.47 | 12 | 186 |  |
|  |  | Complementary inner face | 1.31 | 0.19 | 0.05 |  |  |  |
|  |  | End | 0.73 | 0.13 | 0.04 |  |  |  |
|  | 75 | Inner face | 2.53 | 0.95 | 0.29 | 11 | 171 |  |
|  |  | Complementary inner face | 1.55 | 0.19 | 0.06 |  |  |  |
|  | 90 | End | 0.73 | 0.14 | 0.04 | 11 | 167 |  |
|  |  | Inner face | 1.98 | 0.55 | 0.17 |  |  |  |
|  |  | Complementary inner face | 1.96 | 0.54 | 0.16 |  |  |  |

B

| Target | Desired Crossing angle ( $\theta$ ) | Normalized intensity (A.U.) |  |  |  | Number of fields of view considered ( N ) | Number of nanostructures measured ( n ) | Corresponding figure |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | NanoX region | Mean | SD | SEM (SD/ $\sqrt{N}$ ) |  |  |  |
| $\begin{aligned} & \text { GFP- } \\ & \text { FBP17 } \\ & \text { (FL) } \end{aligned}$ | 15 | End | 2.22 | 0.32 | 0.16 |  |  | Fig. 1H |
|  |  | Inner face | 1.18 | 0.15 | 0.07 | 4 | 72 |  |
|  |  | Complementary inner face | 1.08 | 0.04 | 0.02 |  |  |  |
|  | 30 | End | 2.20 | 0.56 | 0.25 |  |  |  |
|  |  | Inner face | 0.99 | 0.24 | 0.11 | 5 | 90 |  |
|  |  | Complementary inner face | 0.97 | 0.12 | 0.05 |  |  |  |
|  |  | End | 2.21 | 0.47 | 0.21 |  |  |  |
|  | 45 | Inner face | 0.94 | 0.09 | 0.04 | 5 | 90 |  |
|  |  | Complementary inner face | 0.85 | 0.13 | 0.06 |  |  |  |
|  |  | End | 2.64 | 0.38 | 0.17 |  |  |  |
|  | 60 | Inner face | 0.93 | 0.10 | 0.04 | 5 | 90 |  |
|  |  | Complementary inner face | 0.92 | 0.09 | 0.04 |  |  |  |
|  | 75 | End | 2.44 | 0.22 | 0.10 | 5 | 90 |  |
|  |  | Inner face | 0.90 | 0.17 | 0.07 |  |  |  |
|  |  | Complementary inner face | 1.16 | 0.18 | 0.08 |  |  |  |
|  | 90 | End | 2.36 | 0.46 | 0.21 | 5 | 90 |  |
|  |  | Inner face | 0.99 | 0.13 | 0.06 |  |  |  |
|  |  | Complementary inner face | 0.98 | 0.14 | 0.06 |  |  |  |

Supplementary Table 3. Statistical analysis for Figure 1.


Supplementary Table 4. Statistical analysis for Figure 2.


Supplementary Table 5. Statistical analysis for Figure 3.

| Target | Desired Arm width ( nm ) | Normalized intensity (A.U.) |  |  |  | Number of fields of view considered ( N ) | Number of nanostructures measured ( n ) | Corresponding figure |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | NanoU region | Mean | SD | SEM (SD/ $\sqrt{N}$ ) |  |  |  |
| $\begin{gathered} \text { GFP- } \\ \text { FBP17 } \\ \text { (FL) } \end{gathered}$ | 400 | End | 1.38 | 0.21 | 0.07 | 9 | 162 | Fig. 4E |
|  | 450 |  | 1.30 | 0.22 | 0.07 | 9 | 162 |  |
|  | 500 |  | 1.21 | 0.18 | 0.06 | 9 | 162 |  |
|  | 550 |  | 1.16 | 0.16 | 0.05 | 9 | 162 |  |
| Before adding ATP | 600 |  | 1.23 | 0.13 | 0.04 | 9 | 162 |  |
|  | 650 |  | 1.17 | 0.20 | 0.07 | 9 | 162 |  |
|  | 700 |  | 1.12 | 0.13 | 0.04 | 9 | 162 |  |
|  | 750 |  | 1.13 | 0.14 | 0.05 | 9 | 162 |  |
|  | 800 |  | 1.11 | 0.21 | 0.07 | 9 | 162 |  |
| $\qquad$ <br> Target $\qquad$ <br> GFPFBP17 (FL) <br> 1-hr After adding ATP |  |  |  |  |  |  |  |  |
|  | Desired Arm width (nm) | Normalized intensity (A.U.) |  |  |  | Number of fields of view considered ( N ) | Number of nanostructures measured ( n ) | Corresponding figure |
|  |  | NanoU region | Mean | SD | SEM (SD/ $\sqrt{N}$ ) |  |  |  |
|  | 400 | End | 1.34 | 0.24 | 0.09 | 7 | 124 | Fig. 4F |
|  | 450 |  | 1.27 | 0.23 | 0.09 | 7 | 124 |  |
|  | 500 |  | 1.23 | 0.27 | 0.10 | 7 | 124 |  |
|  | 550 |  | 1.16 | 0.18 | 0.07 | 7 | 124 |  |
|  | 600 |  | 1.01 | 0.18 | 0.07 | 7 | 124 |  |
|  | 650 |  | 1.09 | 0.18 | 0.07 | 7 | 124 |  |
|  | 700 |  | 1.06 | 0.16 | 0.06 | 7 | 124 |  |
|  | 750 |  | 1.02 | 0.17 | 0.06 | 7 | 124 |  |
|  | 800 |  | 1.04 | 0.19 | 0.07 | 7 | 124 |  |

Supplementary Table 6. Statistical analysis for Figure 4.

