

## Supplementary material

# MoS<sub>2</sub>/PDA@Cu Peroxidase-Mimicking Enzyme with High Effect Antibacterial and Anticancer Activity

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# **1 Materials Synthetic**

## **1.1 Preparation of PDA**

In a typical experiment, 50mg DA were dissolved in 100ml Tris buffer (10mM pH=8.5) to prepare the PDA. The reaction was carried out at room temperature for 6 h under dark conditions. After the reaction, the product was separated by centrifuging at 7000 rpm for 5 min and washed with water and ethanol three times, and PDA were prepared.

## **1.2 Preparation of PDA@Cu**

40mg PDA and 0.5 g Copper acetate were added into 250ml anhydrous ethanol stirring for 2 h. After, the above solution was set aside for 12 h, the precipitate separated and was then dispersed into 200 mL ethanol. Next, 0.445 g  $\text{NaH}_2\text{PO}_2 \cdot \text{H}_2\text{O}$  was dissolved in a 50 mL ethanol solution, which acted as a reducing agent. Then, the re-prepared PDA mixed in the  $\text{NaH}_2\text{PO}_2 \cdot \text{H}_2\text{O}$  solution was added into a 250 mL round-bottom flask refluxed in an oil bath at a temperature of 80 °C for 30 min. The following treatment method was the same as for the preparation of PDA, and finally PDA@Cu nano material was prepared.

## **1.3 Preparation of MoS<sub>2</sub> Modified By Copper Acetate**

40mg MoS<sub>2</sub> and 0.5 g Copper acetate were added into 250ml anhydrous ethanol stirring for 2 h. After, the above solution was set aside for 12 h, the precipitate separated and was then dispersed into 200 mL ethanol. Next, 0.445 g  $\text{NaH}_2\text{PO}_2 \cdot \text{H}_2\text{O}$  was dissolved in a 50 mL ethanol solution, which acted as a reducing agent. Then, the re-prepared MoS<sub>2</sub> mixed in the  $\text{NaH}_2\text{PO}_2 \cdot \text{H}_2\text{O}$  solution was added into a 250 mL round-

bottom flask refluxed in an oil bath at a temperature of 80 °C for 30 min. The following treatment method was the same as for the preparation of PDA, and finally MoS<sub>2</sub>@Cu nano material was prepared.

## **2 The Preparation for Antibacterial Experiments**

### **2.1 Preparation of Samples**

The samples were dispersed in sterilized PBS buffer, diluted to an appropriate concentration with sterilized PBS buffer, and irradiated under uv lamp for 30 min for backup use.

### **2.2 Dilution of Inoculation Liquid**

1 mL of bacterial suspension was taken after 18 h of oscillation and diluted with sterilized PBS buffer. The absorbance (OD) of the diluted bacterial suspension was measured by UV-vis spectrophotometer until the OD value was 0.03.

### **2.3 Constant Temperature Water Bath Oscillation**

2 mL of diluted bacterial suspension was placed in a 10 mL centrifuge tube, and 1 mL of the sample to be tested was added. The centrifuge tube was placed in a constant temperature water bath oscillator at a rotational speed of 150 r/min at 37 °C for 18 h, and the plate antibacterial experimental bacterial suspension was obtained.

### **2.4 Preparation of Culture Medium**

Pour 30 mL of sterilized medium into sterilized petri dish and cool naturally in ultra-clean workbench.

### **2.5 Petri Dish Inoculation**

After shaking for 18 h, 50 μL of bacterial suspension was absorbed with a pipette

gun, then inoculated on the AGAR medium after cooling and solidification, and evenly coated with sterile coating stick for 3 times.

### **2.6 Constant Temperature Incubator Culture**

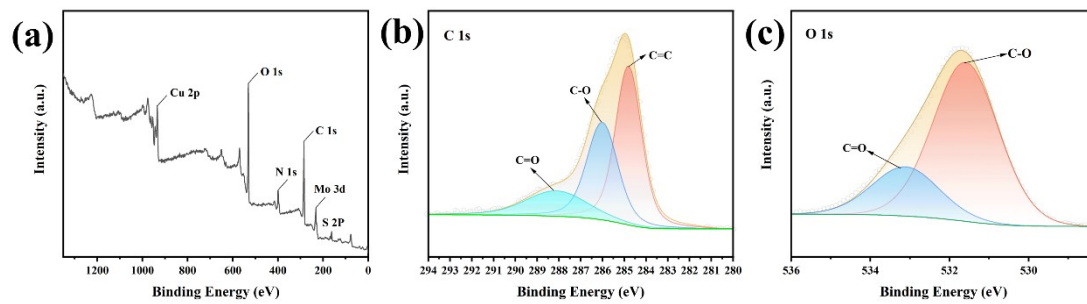
The coated petri dishes and the blank control petri dishes without samples were placed in a 37 °C constant temperature and humidity box at the same time for 18-24 h culture (there are changes according to different strains).

### **2.7 Record The Results**

The petri dishes were taken out and the colony photos of each petri dish were taken. The photos were imported into Image J software to calculate the colony number.

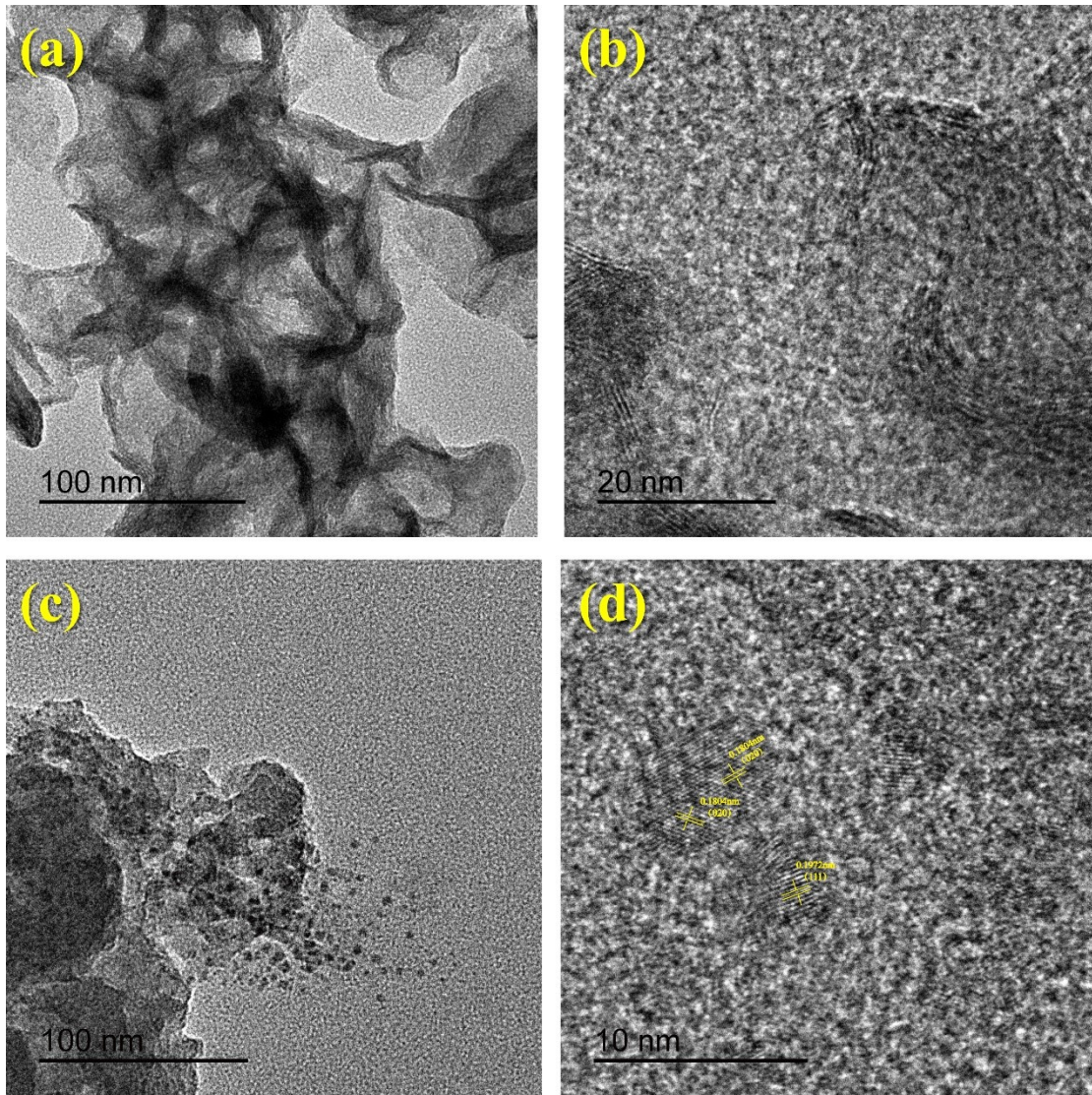
## **3. Cu Ion dissolution test**

The 0.05 mg/ml sample of MoS<sub>2</sub> modified by cupper acetate and MoS<sub>2</sub>/PDA@Cu were dispersed with DI water. After quiescence for one day, they were filtered using a filter with 0.22 μm pore size and detected using Inductive Coupled Plasma Emission Spectrometer (ICP).

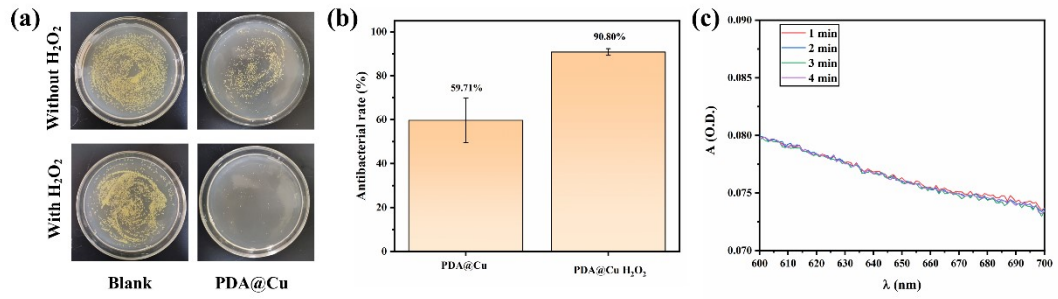


**Figure. S1** XPS survey patterns of (a) MoS<sub>2</sub>/PDA@Cu, (b) C 1s spectra, (c) O 1s spectra of

MoS<sub>2</sub>/PDA@Cu



**Figure. S2** TEM images (a-b) MoS<sub>2</sub> modified by copper acetate, (c-d) PDA@Cu



**Figure. S3** (a) Growth situation of *S. aureus* treated by PDA@Cu, (b) (c) The absorbance changes of PDA@Cu (100  $\mu$ g/ml, 0.5 ml) in the solution, which contained TMB (0.5 mM, 0.5 ml), H<sub>2</sub>O<sub>2</sub>(100 mM, 1 ml) and citrate buffer, were measured by UV-vis spectrophotometer (600 nm to 700 nm)

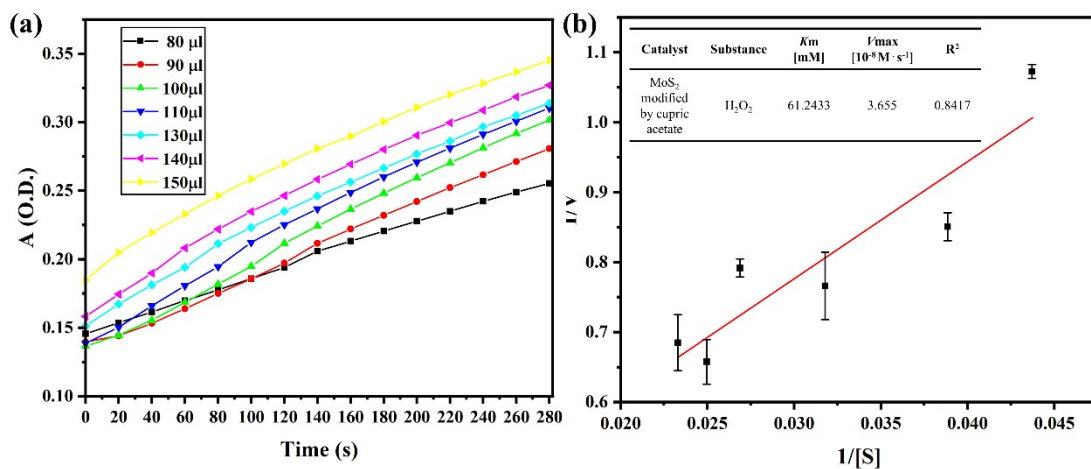


Figure. S4 (a) Time-dependent absorbance changes of MoS<sub>2</sub> modified by cupper acetate (60 μg/ml) at 652 nm of TMB (1.5 mM) with different volume of H<sub>2</sub>O<sub>2</sub> (100 mM) in a pH 4.0 Citrate buffer (0.2 M) at 25°C; (b) Double-reciprocal plot of the calculation of initial reaction rate and substrate concentration of MoS<sub>2</sub> modified by cupper acetate.



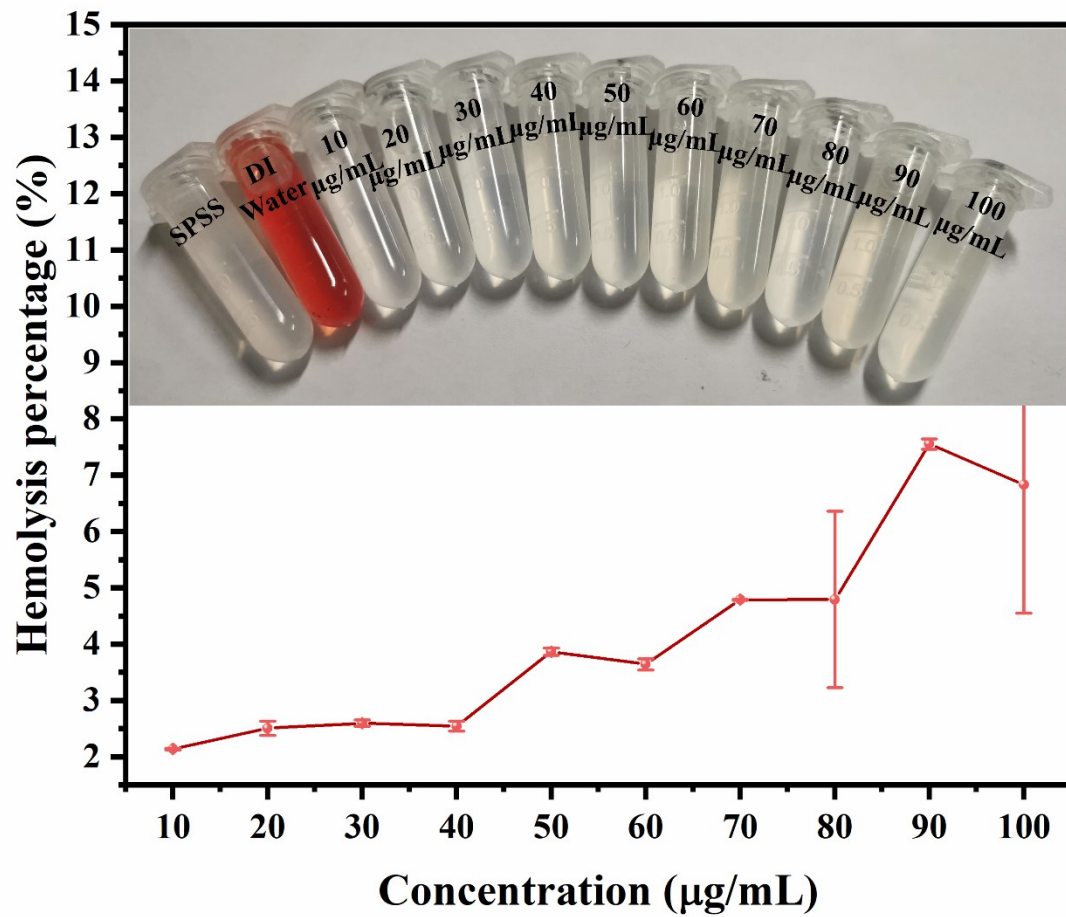


Figure S5 Hemolytic percentage of RBCs after incubated with MoS<sub>2</sub> modified by copper acetate at different concentrations (10 ~ 100 µg/mL) for 3 h; inset image is results of hemolysis assay.

Table S1 Cu ion dissolution concentration

Sample	Cu ion concentration (mg/L)	Standard deviation (mg/L)
MoS <sub>2</sub> modified by copper acetate	12.1825	0.0816
MoS <sub>2</sub> /PDA@Cu	0.6114	0.0113