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Supporting information

Ultrasonic interfacial crosslinking of TiO₂-based nanocomposite hydrogels through thiolnorbornene reactions for sonodynamic antibacterial treatment

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Note - ESI updated 08/08/2023 to correct error in Figure 10(a) as requested by the authors.



Scheme S1. Synthetic schemes of (a) Nor-Dex and (b) TiO₂@MS-SH NPs.





Figure S1. Representative SEM images of (a, b) pure TiO₂ and (c, d) TiO₂@MS-SH NPs. (e) Photographs of NPs dispersed in aqueous solution (left: TiO₂ NPs; right: TiO₂@MS-SH NPs).



Figure S2. (a) FT-IR spectra of TiO₂ and TiO₂@MS-SH NPs. (b) Quantification of the thiol group of the TiO₂@MS-SH NPs using Ellman's assay. Hydrodynamic diameter distribution of (c) TiO₂ and (d) TiO₂@MS-SH NPs.



Figure S3. (a) XPS survey data of TiO₂@MS-SH NPs. High-resolution XPS spectra for (b) C 1s, (c) O 1s, (d) Ti 2p, (e) Si 2p, and (f) S 2p of TiO₂@MS-SH NPs.



Figure S4. Stable gelation of $TiO_2@MS-SH/Nor-Dex NC$ hydrogels was formed in the presence of DTT with thiol/norbornene ratio N= 0.4.



Figure S5. Representative stress-strain curves of TiO₂@MS-SH/Nor-Dex NC hydrogels prepared with varied ultrasound frequencies.



Figure S6. Measurements of ROS generation in the solution. (a) Hydroxyl radical (·OH) reaction with terephthalic acid (TA) and radical production under ultrasound irradiation. (b) Fluorescent spectra of TA acid with different samples (i.e., Nor-Dex polymer, Nor-Dex hydrogel (crosslinked with DTT), TiO₂@MS-SH NPs, and TiO₂@MS-SH/Nor-Dex NC hydrogels) under 120 kHz ultrasound irradiation. (c) TiO₂@MS-SH/Nor-Dex NC hydrogels under varied frequencies of ultrasound irradiation. (d) The reaction of singlet oxygen (¹O₂) with DPBF. The decay UV-vis absorption curves of DPBF in aqueous solution as a function of the (e) varied frequencies and (f) ultrasound irradiation time.



Figure S7. Cell membrane permeability of (a) *S. aureus* and (b) *E. coli* in the absence and presence of TiO₂@MS-SH/Nor-Dex NC hydrogels under various ultrasound frequencies and sonication times. S: *S. aureus*; E: *E. coli*; SH: *S. aureus* incubated with hydrogel; EH: *E. coli* incubated with hydrogel. S-2 (or SH-2, E-2, and EH-2) and S-6 (or SH-6, E-6, and EH-6) indicate the absorbance of *S. aureus* (or *E. coli*) suspensions with ONPG obtained after the reaction time for 2 and 6 hr, respectively.



Figure S8. Reprehensive SEM images of *S. aureus* in the (a) absence and (b) presence of TiO₂@MS-SH/Nor-Dex NC hydrogels. Reprehensive SEM images of *E. coli* in the (c) absence and (d) presence of TiO₂@MS-SH/Nor-Dex NC hydrogels.



Figure S9. Stability test of TiO₂@MS-SH/Nor-Dex NC hydrogels in serum-containing DMEM and PBS solution.



Figure S10. Cytocompatibility tests of the extract solutions obtained from the $TiO_2@MS-SH/Nor-Dex NC$ hydrogels. (a) Fluorescence images of the live/dead staining of MEFs at different time points, where calcein-AM stained live cells green while ethidium homodimer-1 stained dead cells red. Scale bar: 100 µm. (b) Metabolic activity of MEFs cultured in the extract solutions obtained from the NC hydrogels.

 Table S1. Summary of the sample conditions of TiO2@MS-SH/Nor-Dex NC hydrogels used in the compression tests.

Tests ^{a,b}	TiO2@MS-SH	KPS	Frequency (kHz)
	concentration (wt%)	concentration (wt%)	
TiO2@MS-SH	0, 0.5, 1, 1.5	1	120
concentration			
KPS	1	0, 0.25, 0.5, 1.25, 1.5	120
concentration			
Frequency	1	1	40, 80, 120
^a The Nor-Dex polyme	er was fixed at 10 wt% in ea	ich test.	
^b The thiol/norborene 1	ratio was fixed at 0.4 in each	h test.	