

Supporting information:

# Nucleoside Analogs in ADAR Guide Strands Targeting 5'-UA sites

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**Table S1.** Average  $\chi$  angle of the -1 position across ADAR2 crystal structures (5ED1, 5ED2, 5HP2, 5HP3, 6D06, 6VFF, 7KFN), A-form -1 position, and the average of other adenosines in ADAR2d WT Bdf2-U structure (5HP2).

Measured Adenosine	$\chi$ angle
Average -1 position	$-90 \pm 6$
A-Form -1 position	-157
Average of other adenosines	$-160 \pm 7$

**Table S2.** The effect of -1 gRNA modifications on fitted end points and observed rate constants for ADAR2 deamination reactions with *MECP2*.<sup>a</sup>

-1 modification <sup>a</sup>	Fitted end point (%) <sup>b</sup>	$k_{\text{obs}}$ (min <sup>-1</sup> ) <sup>b</sup>	$k_{\text{rel}}$ <sup>c</sup>
Adenosine	$96.2 \pm 0.6$	$0.3 \pm 0.1$	1
dA	$95.2 \pm 0.1$	$0.7 \pm 0.3$	2.3
2'-O-Me A	$93.8 \pm 1.0$	$0.4 \pm 0.2$	1.3
LNA A	no reaction	no reaction	no reaction
UNA A	$87.2 \pm 0.7$	$0.061 \pm 0.006$	0.2
2'-F A	$94 \pm 1$	$0.18 \pm 0.02$	0.6
FANA A	$94.6 \pm 0.3$	$1.0 \pm 0.1$	3.3
$\beta$ -L-dA	$78.6 \pm 0.9$	$0.031 \pm 0.003$	0.1
C3 spacer	$55.1 \pm 0.5$	$0.17 \pm 0.02$	0.6
2-aminopurine	$90 \pm 2$	$0.071 \pm 0.008$	0.2
2,6-diaminopurine	$90 \pm 1$	$0.076 \pm 0.003$	0.3
dI	$93.0 \pm 0.6$	$0.42 \pm 0.04$	1.4
dN	$94.7 \pm 0.6$	$0.59 \pm 0.09$	2.0
7-deaza-dA	$94.5 \pm 0.2$	$0.68 \pm 0.07$	2.3
3-deaza-dA	$90.2 \pm 0.4$	$0.5 \pm 0.1$	1.7
dC	$76 \pm 1$	$0.0317 \pm 0.0008$	0.1
Abasic	$53.9 \pm 1.0$	$0.051 \pm 0.001$	0.2
dN -1, LNA -6	$92.8 \pm 0.2$	$1.72 \pm 0.07$	5.7
Orphan dZ	$69 \pm 2$	$0.41 \pm 0.09$	1.4
Orphan dZ, dN -1	$74.7 \pm 0.6$	$1.4 \pm 0.2$	4.7

<sup>a</sup> Reactions were carried out with 5 nM of target RNA and 15 nM ADAR2 WT

<sup>b</sup> Reactions were fitted to the equation  $[P]_t = \alpha[1 - \exp(-k_{\text{obs}} \cdot t)]$

<sup>c</sup>  $k_{\text{rel}} = k_{\text{obs}}$  for modification/ $k_{\text{obs}}$  for adenosine

**Table S3.** The effect of -1 gRNA modifications on observed rate constants for ADAR1 deamination reactions with *MECP2*.<sup>a</sup>

-1 modification <sup>a</sup>	$k_{\text{obs}} \times 10^{-2} (\text{min}^{-1})^{\text{b}}$	$k_{\text{rel}}^{\text{c}}$
Adenosine	0.22 ± 0.02	1
dA	1.2 ± 0.2	5.5
2'-O-Me A	0.17 ± 0.02	0.77
LNA A	no reaction	no reaction
UNA A	no reaction	no reaction
2'-F A	0.11 ± 0.01	0.50
FANA A	0.79 ± 0.04	3.6
dN	4.3 ± 0.7	20
7-deaza-dA	1.05 ± 0.06	4.8
Orphan dZ	0.36 ± 0.01	1.6
Orphan dZ, dN -1	1.89 ± 0.07	8.6

<sup>a</sup> Reactions were carried out with 15 nM of target RNA and 150 nM ADAR1 p110

<sup>b</sup> Reactions were fitted to the equation  $[P]_t = \alpha[1 - \exp(-k_{\text{obs}} \cdot t)]$ . End point was constrained to 65%

<sup>c</sup>  $k_{\text{rel}} = k_{\text{obs}}$  for modification/ $k_{\text{obs}}$  for adenosine

**Table S4.** The effect of -1 gRNA modifications on fitted end points and observed rate constants for ADAR2 deamination reactions with *IDUA*.<sup>a</sup>

-1 modification <sup>a</sup>	Fitted end point <sup>b</sup>	$k_{\text{obs}} (\text{min}^{-1})^{\text{b}}$	$k_{\text{rel}}^{\text{c}}$
Adenosine	94.6 ± 0.4	0.59 ± 0.01	1
dA	95.6 ± 0.1	0.6 ± 0.2	1
dN	92.3 ± 0.7	1.2 ± 0.2	2.0
7-deaza-dA	94.7 ± 0.4	0.59 ± 0.03	1.0

<sup>a</sup> Reactions were carried out with 5 nM of target RNA and 15 nM ADAR2 WT

<sup>b</sup> Reactions were fitted to the equation  $[P]_t = \alpha[1 - \exp(-k_{\text{obs}} \cdot t)]$

<sup>c</sup>  $k_{\text{rel}} = k_{\text{obs}}$  for modification/ $k_{\text{obs}}$  for adenosine

**Table S5.** The effect of -1 gRNA modifications on observed rate constants and fitted end points for ADAR1 deamination reactions with *IDUA*.<sup>a</sup>

-1 modification <sup>a</sup>	Fitted end point <sup>b</sup>	$k_{\text{obs}} \times 10^{-2} (\text{min}^{-1})^{\text{b}}$	$k_{\text{rel}}^{\text{c}}$
Adenosine	66 ± 2	4.1 ± 0.2	1
dA	66 ± 2	4.2 ± 0.3	1.0
dN	79 ± 1	17.5 ± 0.4	4.3
7-deaza-dA	73 ± 1	8.0 ± 0.8	2.0

<sup>a</sup> Reactions were carried out with 15 nM of target RNA and 150 nM ADAR1 p110

<sup>b</sup> Reactions were fitted to the equation  $[P]_t = \alpha[1 - \exp(-k_{\text{obs}} \cdot t)]$

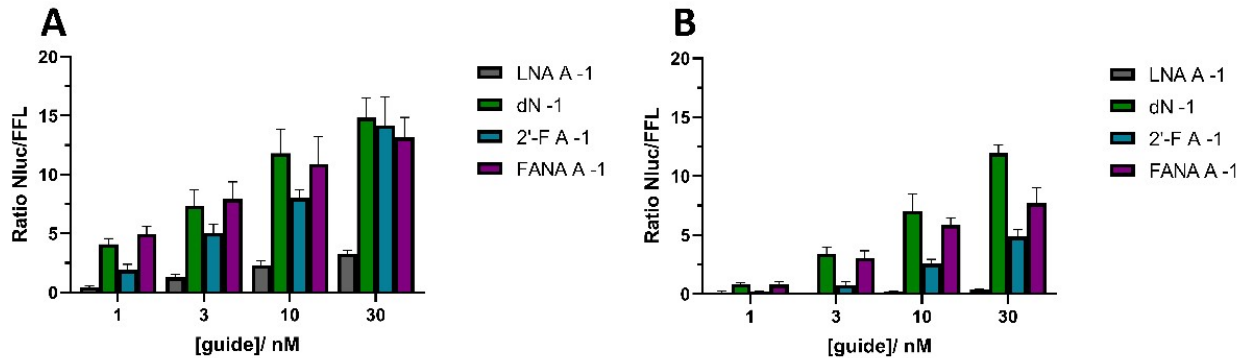
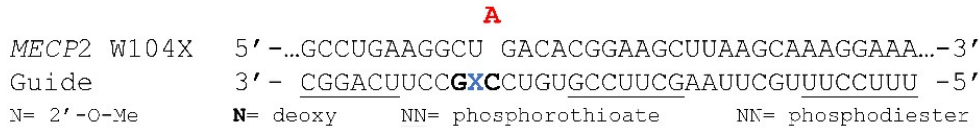
<sup>c</sup>  $k_{\text{rel}} = k_{\text{obs}}$  for modification/ $k_{\text{obs}}$  for adenosine



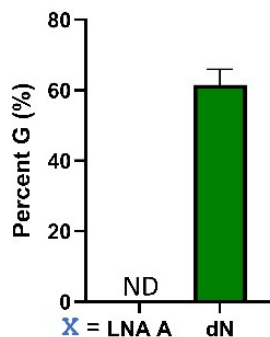
Mouse <i>IDUA</i> W392X gRNAs	5' -UUGAGACCUCUGCCC [A] GAGUUGUUCUCCA-3'
Human <i>ACTB</i> gRNAs	5' -GGGUGUAACGCAACC [A] AGUCAUAGUCCGC-3'
Human <i>MECP2</i> W104X cellular guides	5' - <u>UUUCCUUUGCUUAAGCUUCCGUGUCC</u> [A] <u>GCCUUCAGGC</u> -3'
Standard DNA oligo sequence for MALDI	5' -TTACGCCAGAATGCGTTCGCACAGCCGCCA-3'
<i>MECP2</i> RT-PCR and sequencing primer forward	5' -AGAGGCAGGCAAAGCAGA-3'
<i>MECP2</i> amplification and RT-PCR primer reverse	5' -TCATTAGGGTCCAGGGAT-3'
<i>MECP2</i> T7 DNA amplification forward	5' -TAATACGACTCACTATAGGGAGC-3'
<i>IDUA</i> RT-PCR primer forward	5' -CTCCCATCCTGTGGGCT-3'
<i>IDUA</i> amplification, RT-PCR and sequencing primer reverse	5' -GGATCCGGGTGTGC-3'
<i>IDUA</i> T7 DNA amplification forward	5' -TAATACGACTCACTATAGGGCTC-3'
<i>ACTB</i> RT-PCR and sequencing primer forward	5' -GTCACCTTCACCGTTCCA-3'
<i>ACTB</i> amplification and RT-PCR primer reverse	5' -TCCTCCTGAGCGCAAG-3'
<i>ACTB</i> T7 DNA amplification forward	5' -TAATACGACTCACTATAGGGTCCT-3'
<i>MECP2</i> W104X sequence for dual luciferase assay reporter plasmid (target A indicated in red)	5' -CGCTCCATCATCCGTGACCGGGACCCATGTATGATGACCCCACCCCTGCCTGAAGGCTAGACACGGAAGCTTAAGCAAAGGAAATCTGGC-3'
RT-PCR forward primer for sequencing of cellular editing	5' -CTGCTTACTGGCTTATCGAAATTAATACGACT-3'
RT-PCR reverse primer for sequencing of cellular editing	5' -TTACGCCAGAATGCGTTCGCACAGCCGCCA-3'
Nested PCR and sequencing forward primer for sequencing of cellular editing	5' -CACTATAGGGAGACCCAAGCTTGCCAC-3'
Nested PCR reverse primer for sequencing of cellular editing	5' -GCCGGTCACTCCGTTGATGGTTACT-3'

**Table S9.** Masses of gRNAs and guide oligonucleotides (cellular).

gRNA	Calculated Mass (Da)	Observed Mass (Da)
<i>MECP2</i> rA -1	9231	9240
<i>MECP2</i> dA -1	9215	9224
<i>MECP2</i> 2'-O-Me -1	9245	9248
<i>MECP2</i> LNA A -1	9243	9236
<i>MECP2</i> UNA A -1	9238	9238
<i>MECP2</i> 2'-F A -1	9233	9230
<i>MECP2</i> 2'-F-arabino A -1	9233	9224
<i>MECP2</i> $\beta$ -L-dA -1	9215	9221
<i>MECP2</i> C3 spacer -1	9040	9035
<i>MECP2</i> 2-aminopurine -1	9231	9229
<i>MECP2</i> 2,6-diaminopurine -1	9246	9240
<i>MECP2</i> dI -1	9216	9215
<i>MECP2</i> dN -1	9200	9205
<i>MECP2</i> 7-deaza-dA -1	9214	9225
<i>MECP2</i> 3-deaza-dA -1	9214	9225
<i>MECP2</i> dC -1	9191	9198
<i>MECP2</i> abasic -1	9082	9097
<i>MECP2</i> LNA -6, dN -1	9212	9228
<i>MECP2</i> orphan dZ	9259	9242
<i>MECP2</i> orphan dZ, dN -1	9228	9227
<i>IDUA</i> rA -1	9153	9144
<i>IDUA</i> dA -1	9137	9137
<i>IDUA</i> dN -1	9122	9123
<i>IDUA</i> 7-deaza-dA -1	9136	9141
<i>ACTB</i> rA -1	9301	9297
<i>ACTB</i> dA -1	9285	9280
<i>ACTB</i> dN -1	9270	9272
<i>MECP2</i> LNA A -1 (cellular)	12350	12350
<i>MECP2</i> dN -1 (cellular)	12307	12307
<i>MECP2</i> 2'-F A -1 (cellular)	12340	12340
<i>MECP2</i> FANA A -1 (cellular)	12340	12340



**Figure S1.** Concentration dependence of effect of -1 nucleoside analogs on cellular editing as measured by dual luciferase assay. Relative ratio of Nluc/FFL was measured in HEK293T cells with varying concentrations of guide oligonucleotides with four different -1 analogs with overexpressed (A) ADAR2 or (B) ADAR1. Error bars, s.d. ( $n \geq 5$  biological replicates). A Grubbs test for outliers was conducted at  $p = 0.05$  significance level.



**Figure S2.** Effect of -1 nucleoside analogs on cellular editing as measured by Sanger sequencing. The percent G at the *MECP2* W104X target site was measured in HEK293T cells after transfection of 30 nM of -1 nucleoside analog containing guide oligonucleotide with overexpressed ADAR1. Error bars, s.d. ( $n = 3$  biological replicates). ND = no detected editing.