

Supporting Information

Regiodivergent cascade cyclization/alkoxylation of allenamides via N-protecting group driven rearrangement to access indole and indoline derivatives

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General Information

Unless otherwise noted, all the reactions were performed using oven-dried Schlenk tubes under nitrogen. The reactions were monitored by Merck silica gel 60 F₂₅₄ precoated plates (0.25 mm) visualizing under UV light (254 nm) or I₂ staining. The temperature mentioned for any reaction is corresponding to the oil bath temperature. Column chromatography was performed using silica gel 60-120 Å or 100-200 Å mesh of Merck Company.

All the commercial reagents and anhydrous solvents were purchased from Sigma-Aldrich, Alfa Aesar, Merck, Spectrochem, Avra Synthesis Pvt. Ltd. and directly used as received without any further purification.

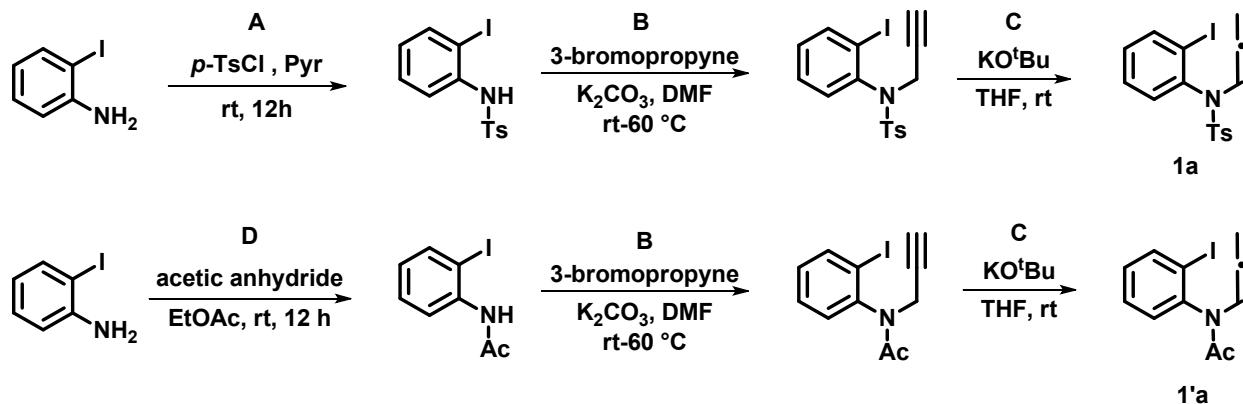
Analytical Methods

¹H, ¹³C and ¹⁹F nuclear magnetic resonance spectra were recorded on Bruker Advance III 400 MHz spectrometer at 25 °C. NMRs of the products were measured in CDCl₃. The chemical shifts in ¹H NMR and ¹³C{¹H} NMR spectra are reported in parts per million (ppm) and are referenced to the residual solvent signal as the internal standard; ¹H NMR spectra (CDCl₃ δ 7.26 ppm), ¹³C (CDCl₃ δ 77.16), ¹H (D₄-methanol δ 3.31 ppm), ¹³C (D₄-methanol δ 49.00) and ¹H (D₆-acetone δ 2.05 ppm), ¹³C (D₄-methanol δ 29.84, 206.26). The coupling constant (J) was reported in Hertz (Hz). Splitting patterns are denoted as "s" for singlet; "d" for doublet; "t" for triplet; "q" for quartet; "sext" for sextet; "sept" for septet; "m" for multiplet, "br" for broad; "dt" for doublet of triplets; "td" for triplet of doublets. ESI-HRMS were recorded on AGILENT 6520 Q-TOF spectrometer. IR spectra were recorded with Agilent Cary 630 FTIR Spectrometer.

2. Experimental procedures

The starting materials **1a-1x**^{1,2,3,4,5,6,7} and **1'a**⁵, **7a-7b**⁸ were prepared according to the previously reported methods.

2.1 Preparation of Allenamides:



General procedure A for tosylation:

To a solution of *o*-iodoaniline (1.0 equiv.) in pyridine was added *p*-TsCl (1.05 equiv.) at 0 °C. The reaction was stirred at rt over night before being quenched with H₂O. The quenched mixture was extracted three times with DCM. The combined organic layers were first washed with 1M HCl to remove excess pyridine, and then with saturated aq. NaHCO₃, H₂O, followed by saturated aq. NaCl, and dried over anhyd. Na₂SO₄. The filtrate was concentrated under reduced pressure and purified using silica gel flash column chromatography to give the desired product tosylamine.

General procedure B for propargylamides:

To a solution of the crude carbamate/tosylamine product (1.00 equiv.) in DMF, K₂CO₃ (1.5 equiv.) and 3-bromopropyne (1.5 equiv.) was added. The mixture was stirred in an oil bath at 60 °C for overnight. After the reaction was complete, the mixture was filtered through Celite. The filtrate was concentrated under reduced pressure and purified by flash column chromatography to give the propargylamide.

General procedure C for allenation reaction:

To a solution of propargylamide (1.0 equiv) in THF was added *t*-BuOK (0.5 equiv.) at 0 °C. The reaction was stirred at room temperature for 1 h before being concentrated under reduced pressure. Subsequently, the residue was suspended in DCM and then filtered through Celite. The filtrate was concentrated under reduced pressure and the crude residue was purified by flash column chromatography to give the allenamide.

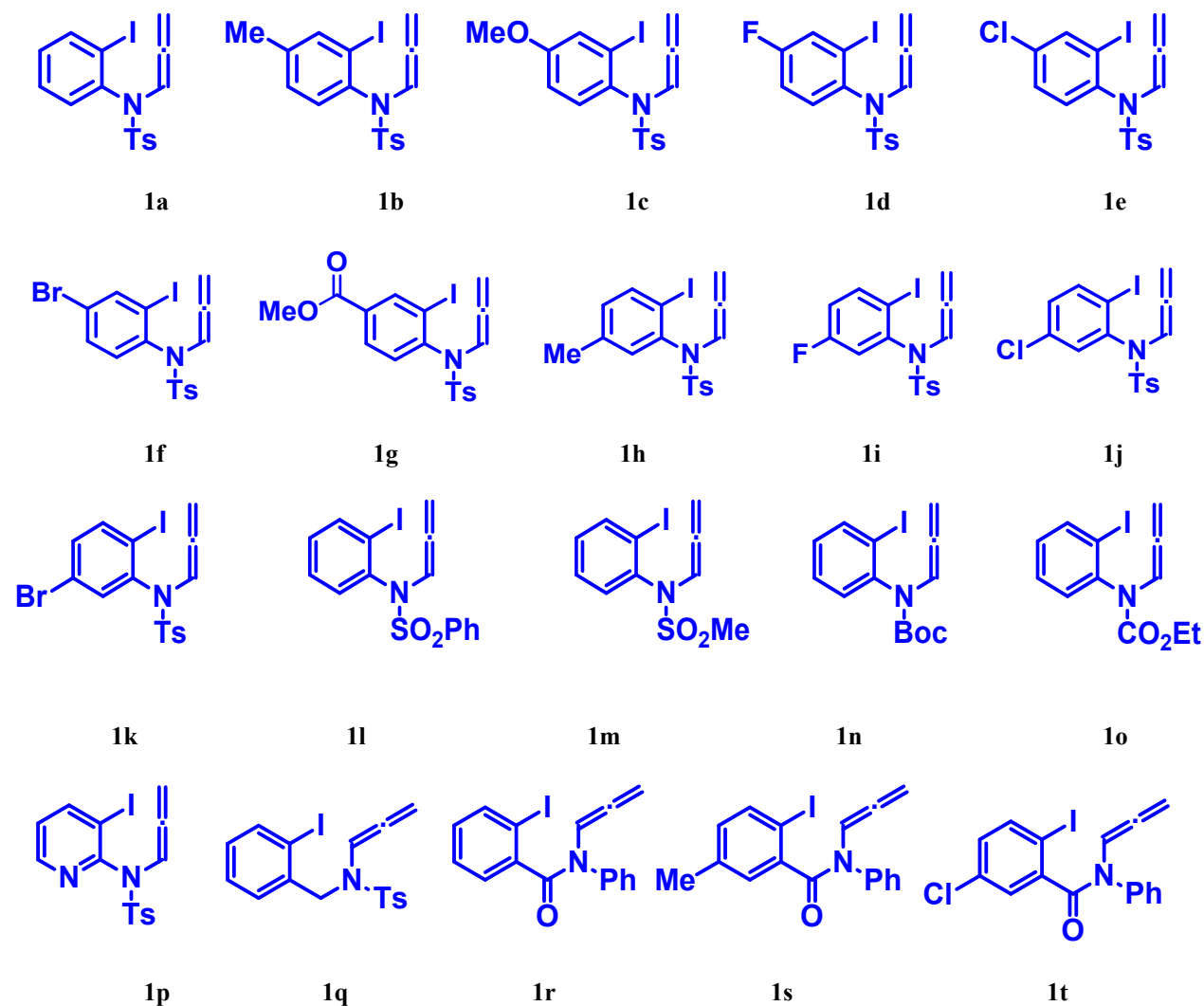
General procedure D for acetylation:^{11, 12}

In a 25 mL round bottom flask, *o*-iodoaniline (1.0 equiv) was dissolved in EtOAc, and acetic anhydride (1.4 equiv.) was added, and the reaction mixture was stirred at room temperature overnight. The solution was filtered and concentrate under reduced pressure to give the crude carbamate that was used in the next step without further purification.

General procedure D' for Benzoylation:¹³

To a solution of corresponding 2-iodo aniline (1.0 equiv.) in dichloromethane was added benzoyl chloride (1.2 mmol) followed by triethylamine (1.5 equiv.) at room temperature. After complete addition, the reaction was allowed to stir continuously until all the starting material was consumed completely (monitored by TLC, approx. 8-10 h after completion), the reaction mixture was quenched with water and extracted with dichloromethane. The combined organic layer was washed with brine solution, dried over Na₂SO₄, evaporated and washed with hexane to give pure benzamide product in quantitative yields.

Table S1. Substrates employed in the reaction:



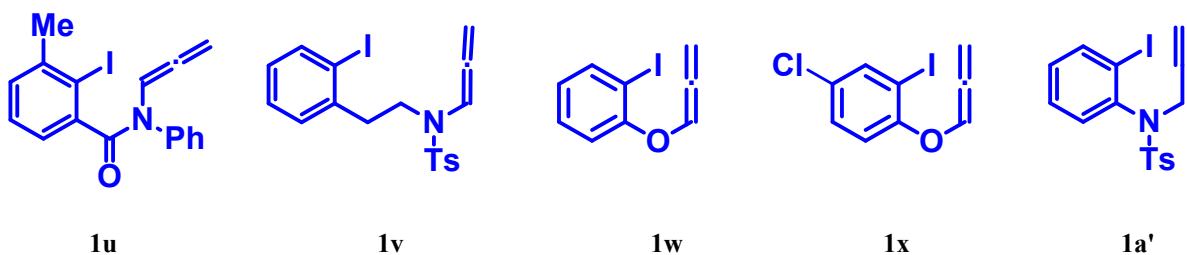
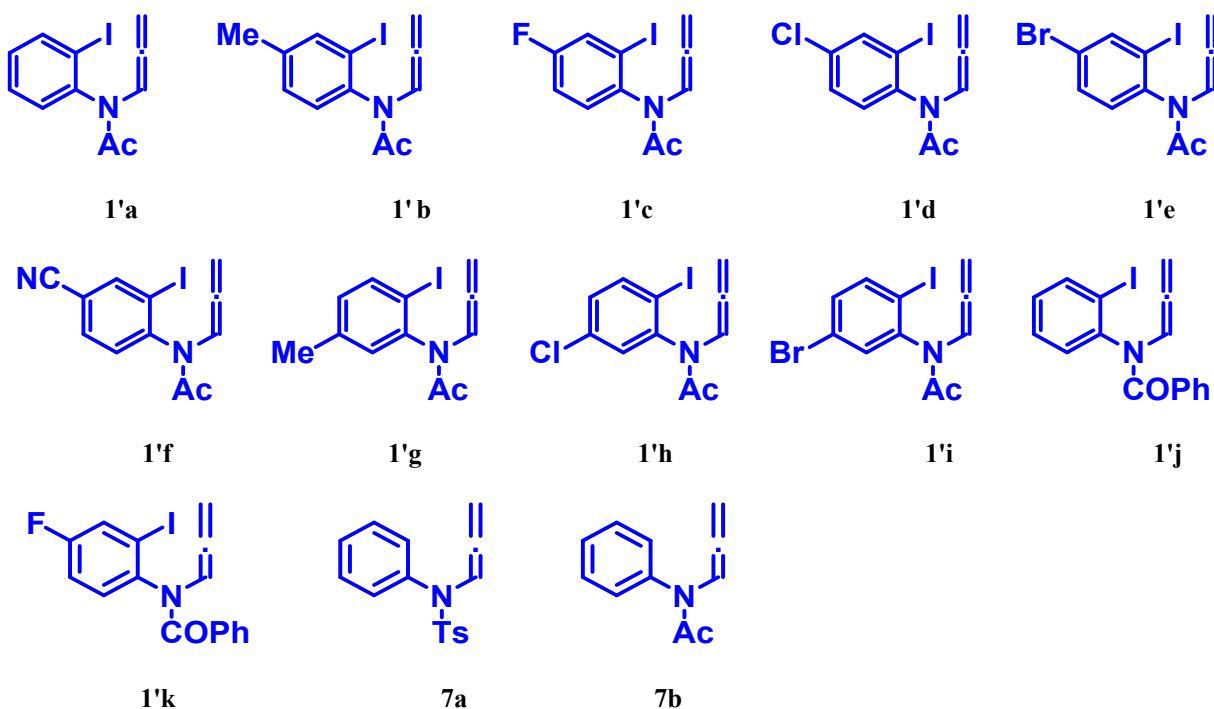


Table S2. Substrates employed in the reaction:



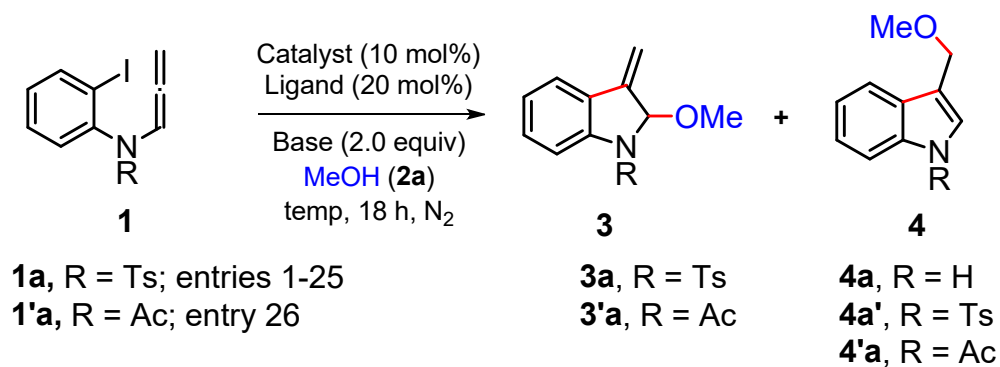
2.2 General Procedure E for the synthesis of 3a-4i:

To an oven dried Schlenk tube was charged with allenamide (1 equiv.), palladium chloride (10 mol%), tri(2-furyl)phosphine (20 mol%), and K_2CO_3 (2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N_2 gas (3X). Then, the solvent methanol/ethanol (**2a**) 3mL was added to the reaction mixture. The reaction was stirred at rt for (18-20 h), and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EtOAc) to yield the corresponding product.

2.3 General Procedure F for the Synthesis of 4ad:

To an oven dried Schlenk tube was charged with allenamide **1a** (1 equiv.), and piperidine (1.2 equiv.), palladium chloride (10 mol%), tri(2-furyl)phosphine(20 mol%), and Cs_2CO_3 (2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N_2 gas (3X). DCE (3mL) was added to the reaction mixture. The reaction was stirred at 80 °C for (18-20 h), the solvent was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (acetone in hexane) to afford the desired product **4ad**.

2.4 Optimization of the reaction conditions:^{a,b}



| Entry | Catalyst | Ligand | Base | Temp °C | 3a (%) | 4a (%) |
|-----------|--|-----------------------------|------------------------------------|-------------|-------------|--------------|
| 1 | Pd(OAc) ₂ | XPhos | Cs ₂ CO ₃ | 80 | 17 | 20 |
| 2 | " | DPPP | " | " | n.d. | 37 |
| 3 | " | PPh ₃ | " | " | (50) | (46) |
| 4 | " | 1,10 Phen | " | " | trace | nd |
| 5 | " | 2,2 bipyridyl | " | " | nd | nd |
| 6 | " | Tri(2-furyl)phosphine (TFP) | " | " | 62 | 9 |
| 7 | Pd(acac) ₂ | " | " | " | 67 | 15 |
| 8 | Pd(PPh ₃) ₂ Cl ₂ | " | " | " | 44 | n.d. |
| 9 | Pd(Ph ₃) ₄ | " | " | " | n.d. | n.d. |
| 10 | Pd ₂ (dba) ₃ | " | " | " | n.d. | n.d. |
| 11 | PdCl₂ | " | " | " | 78 | 11 |
| 12 | " | PPh ₃ | Cs ₂ CO ₃ | " | 11 | 56 |
| 13 | " | PCy ₃ | Cs ₂ CO ₃ | " | n.d. | n.d. |
| 14 | " | P(OPh) ₃ | Cs ₂ CO ₃ | " | 21 | 13 |
| 15 | " | JohnPhos | Cs ₂ CO ₃ | " | n.d. | n.d. |
| 16 | " | P(o-tol) ₃ | Cs ₂ CO ₃ | " | n.d. | n.d. |
| 17 | PdCl ₂ | TFP | K ₂ CO ₃ | " | (78) | n.d. |
| 18 | " | " | K ₃ PO ₄ | " | 73 | 8 |
| 19 | " | " | Na ₂ CO ₃ | " | 70 | n.d. |
| 20 | " | " | NaO ^t Bu | " | 61 | 40 |
| 21 | PdCl₂ | TFP | K₂CO₃ | r.t. | (88) | trace |
| 22 | PdCl ₂ (5 mol%) | TFP (10 mol%) | K ₂ CO ₃ | r.t. | (78) | trace |

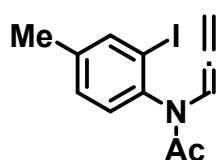
| | | | | | | |
|-----------------------|-----------------------------|---------------|------------------------------------|-------------|------------|-------------|
| 23 | PdCl ₂ (2 mol%) | TFP (4 mol%) | K ₂ CO ₃ | r.t. | (70) | ~5 |
| 24 | PdCl ₂ (10 mol%) | TFP (10 mol%) | K ₂ CO ₃ | r.t. | (65) | ~5 |
| 25 | PdCl ₂ | TFP | Ag ₂ CO ₃ | r.t. | 5 | trace |
| 26 | None | TFP | K ₂ CO ₃ | r.t. | n.d. | n.d. |
| 27 | PdCl ₂ | None | K ₂ CO ₃ | r.t. | 10 | n.d. |
| 28 | PdCl ₂ | TFP | None | r.t. | n.d. | n.d. |
| 29^c | PdCl₂ | TFP | K₂CO₃ | r.t. | (7) | (79) |

^aReaction conditions: **1a/1'a** (1.0 equiv.), catalyst (10 mol%), ligand (20 mol%), base (2.0 equiv.), MeOH (1 mL) at 80 °C/rt for 18 h under N₂. ¹HNMR yield using 1,3,5-trimethoxybenzene as internal standard. n.d. = not detected.

^bIsolated yields on parantheses. ^cR = Ac in **1**.

3. Characterization Data of Starting Materials:

N-(2-iodo-4-methylphenyl)-N-(propa-1,2-dien-1-yl)acetamide (**1'b**):



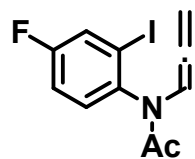
The representative general procedure **C** was followed, using propargylamide (470 mg, 1.5 mmol), in THF was added *t*-BuOK (0.5 equiv.) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 6% EtOAc in hexane) furnished **1'b** (320 mg, 68% yield) as a pale yellow solid. R_f 0.72 (20% EtOAc in hexane).

¹H NMR (400 MHz, CDCl₃): δ 7.73 (s, 1H), 7.59 (td, *J* = 6.3, 1.2 Hz, 1H), 7.20 (d, *J* = 7.9 Hz, 1H), 7.11 (d, *J* = 7.9 Hz, 1H), 5.05-4.94 (m, 2H), 2.34 (s, 3H), 1.84 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 202.4, 168.2, 140.6, 140.4, 140.0, 130.4, 129.2, 99.8, 99.6, 86.6, 23.0, 20.6.

HRMS (ESI) Calcd. for C₁₂H₁₂INO: [M+H]⁺, 314.0042. Found: *m/z* 314.0033.

N-(4-fluoro-2-iodophenyl)-N-(propa-1,2-dien-1-yl)acetamide (**1'c**):



The representative general procedure **C** was followed, using propargylamide (350 mg, 1.1 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 6% EtOAc in hexane) furnished **1'c** (195 mg, 56% yield) as a pale yellow solid. R_f 0.71 (20% EtOAc in hexane).

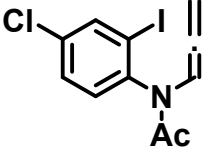
¹H NMR (500 MHz, CDCl₃): δ 7.63 (dd, *J* = 7.6, 2.8 Hz, 1H), 7.59 (t, *J* = 6.4, Hz, 1H), 7.23 (dd, *J* = 8.7, 5.3 Hz, 1H), 7.16-7.12 (m, 1H), 5.07-4.97 (m, 2H), 1.86 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 202.3, 167.9, 161.4 (d, *J* = 254.3 Hz), 139.0, 130.5 (d, *J* = 8.9 Hz), 127.0 (d, *J* = 24.6 Hz), 116.7 (d, *J* = 22.5 Hz), 99.9 (d, *J* = 8.8 Hz), 99.8, 86.9, 22.9.

^{19}F NMR (376 MHz): δ -110.5.

HRMS (ESI) Calcd. for $\text{C}_{11}\text{H}_9\text{FINO}$: $[\text{M}+\text{H}]^+$, 317.9791. Found: m/z 317.9785.

N-(4-chloro-2-iodophenyl)-N-(propa-1,2-dien-1-yl)acetamide (1'd):

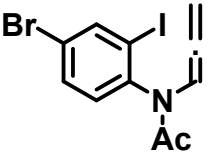
 The representative general procedure **C** was followed, using propargylamide (500 mg, 1.5 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 7% EtOAc in hexane) furnished **1'd** (300 mg, 60% yield) as a pale yellow solid. R_f 0.71 (20% EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 7.91 (d, $J = 2.3$ Hz, 1H), 7.59 (t, $J = 6.4$ Hz, 1H), 7.41 (dd, $J = 8.4, 2.3$ Hz, 1H), 7.18 (d, $J = 8.4$ Hz, 1H), 5.09-4.98 (m, 2H), 1.87 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 202.1, 167.6, 141.4, 139.5, 135.1, 130.4, 129.9, 100.4, 99.6, 87.1, 23.0.

HRMS (ESI) Calcd. for $\text{C}_{11}\text{H}_9\text{ClINO}$: $[\text{M}+\text{H}]^+$, 333.9496. Found: m/z 333.9488.

N-(4-bromo-2-iodophenyl)-N-(propa-1,2-dien-1-yl)acetamide (1'e):

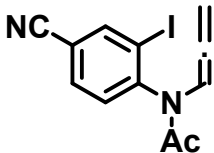
 The representative general procedure **C** was followed, using propargylamide (565 mg, 1.5 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **1'e** (342 mg, 60% yield) as a pale yellow solid. R_f 0.72 (20% EtOAc in hexane).

^1H NMR (500 MHz, CDCl_3): δ 8.07 (d, $J = 1.8$ Hz, 1H), 7.59-7.54 (m, 2H), 7.12 (d, $J = 8.3$ Hz, 1H), 5.08-4.99 (m, 2H), 1.86 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 202.1, 167.6, 142.2, 141.9, 132.9, 130.8, 123.1, 100.8, 99.6, 87.1, 23.0.

HRMS (ESI) Calcd. for $\text{C}_{11}\text{H}_9\text{BrINO}$: $[\text{M}+\text{H}]^+$, 377.8990. Found: m/z 377.8980.

N-(4-cyano-2-iodophenyl)-N-(propa-1,2-dien-1-yl)acetamide (1'f):

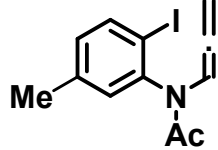
 The representative general procedure **C** was followed, using propargylamide (324 mg, 1.1 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **1'f** (220 mg, 62% yield) as a pale yellow solid. R_f 0.65 (20% EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 8.21 (d, $J = 1.2$ Hz, 1H), 7.73 (dd, $J = 8.0, 1.3$ Hz, 1H), 7.59 (t, $J = 6.3$ Hz, 1H), 7.37 (d, $J = 8.0$ Hz, 1H), 5.08-4.98 (m, 2H), 1.86 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 201.9, 166.8, 146.9, 143.4, 133.2, 130.6, 116.1, 114.3, 100.6, 99.4, 87.5, 23.1.

HRMS (ESI) Calcd. for $\text{C}_{12}\text{H}_9\text{IN}_2\text{O}$: $[\text{M}+\text{H}]^+$, 324.9838. Found: m/z 324.9828.

N-(2-iodo-5-methylphenyl)-N-(propa-1,2-dien-1-yl)acetamide (**1'g**):

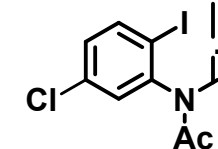
 The representative general procedure **C** was followed, using propargylamide (532 mg, 1.7 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 7% EtOAc in hexane) furnished **1'g** (350 mg, 66% yield) as a pale yellow solid. R_f 0.72 (20% EtOAc in hexane).

^1H NMR (500 MHz, CDCl_3): δ 7.76 (d, $J = 8.1$ Hz, 1H), 7.59 (t, $J = 6.4$ Hz, 1H), 7.06 (s, 1H), 6.90 (d, $J = 8.1$ Hz, 1H), 5.05-4.95 (m, 2H), 2.34 (s, 3H), 1.86 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 202.4, 168.0, 142.4, 140.0, 139.6, 131.2, 130.4, 99.7, 95.6, 86.6, 23.0, 20.9.

HRMS (ESI) Calcd. For $\text{C}_{12}\text{H}_{12}\text{INO}$: $[\text{M}+\text{H}]^+$, 314.0042. Found: m/z 314.0011.

N-(5-chloro-2-iodophenyl)-N-(propa-1,2-dien-1-yl)acetamide (**1'h**):

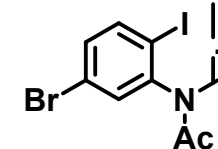
 The representative general procedure **C** was followed, using propargylamide (433 mg, 1.3 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 7% EtOAc in hexane) furnished **1'h** (250 mg, 58% yield) as a pale yellow solid. R_f 0.71 (20% EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 7.82 (d, $J = 8.5$ Hz, 1H), 7.57 (t, $J = 6.3$ Hz, 1H), 7.25 (s, 1H), 7.09 (dd, $J = 8.4, 2.1$ Hz, 1H), 5.08-5.00 (m, 2H), 1.87 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 202.1, 167.5, 143.8, 140.8, 135.3, 130.5, 130.0, 99.5, 97.5, 87.2, 23.1.

HRMS (ESI) Calcd. for $\text{C}_{11}\text{H}_9\text{ClINO}$: $[\text{M}+\text{H}]^+$, 333.9496. Found: m/z 333.9487.

N-(5-bromo-2-iodophenyl)-N-(propa-1,2-dien-1-yl)acetamide (**1'i**):

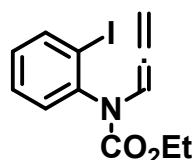
 The representative general procedure **C** was followed, using propargylamide (565 mg, 1.5 mmol), in THF was added *t*-BuOK (0.5 equiv) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 7% EtOAc in hexane) furnished **1'i** (350 mg, 62% yield) as a pale yellow solid. R_f 0.71 (20% EtOAc in hexane).

¹H NMR (500 MHz, CDCl₃): δ 7.76 (d, J = 8.4 Hz, 1H), 7.58 (t, J = 6.4 Hz, 1H), 7.41 (d, J = 2.2 Hz, 1H), 7.23 (dd, J = 8.4, 2.2 Hz, 1H), 5.08-5.01 (m, 2H), 1.88 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 202.1, 167.5, 143.9, 141.1, 133.4, 132.9, 122.8, 99.6, 98.4, 87.2, 23.1.

HRMS (ESI) Calcd. for C₁₁H₉BrINO: [M+H]⁺, 377.8990. Found: m/z 377.8988.

ethyl (2-iodophenyl(propa-1,2-dien-1-yl)carbamate (**1o**):



2-Iodoaniline (1.0 equiv), ethyl chloroformate (4.0 equiv.) and K₂CO₃ (6.0 equiv) were stirred in 40 mL of acetone at room temperature for 3 h. The reaction was monitored by TLC to establish completion. Then the reaction mixture was diluted with water. The organic phase was separated, and the aqueous phase was extracted with ether. Organic fractions were combined, washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. Column chromatography on silica gel using 8:1 hexanes/ethyl acetate to give the ethyl (2-iodophenyl)carbamate as off-white solid.

To a solution of the ethyl (2-iodophenyl)carbamate (1.0 equiv.) in DMF, potassium carbonate (1.5 equiv.) and 3-bromopropyne (1.5 equiv.) was added. The mixture stirred in an oil bath at 60 °C for overnight. After the reaction was complete, the mixture was filtered through Celite. The filtrate was concentrated under reduced pressure and purified by flash column chromatography to give the ethyl (2-iodophenyl)(prop-2-yn-1-yl)carbamate.

To a solution ethyl (2-iodophenyl)(prop-2-yn-1-yl)carbamate (324 mg, 1.1 mmol), in THF was added *t*-BuOK (0.5 equiv.) at 0 °C. The reaction was stirred at room temperature for 1 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **1o** (220 mg, 62% yield) as a sticky yellow solid. R_f 0.65 (20% EtOAc in hexane).

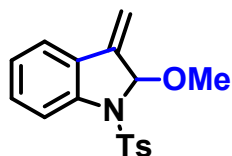
¹H NMR (400 MHz, CDCl₃): δ 7.86 (d, J = 7.9 Hz, 1H), 7.38 -7.19 (m, 3H), 7.02 (t, J = 7.6 Hz, 1H), 5.06-4.97 (m, 2H), 4.26-4.10 (m, 2H), 1.18 (t, J = 6.3 Hz, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 201.6, 152.6, 141.4, 139.5, 129.6, 129.5, 128.9, 101.4, 99.6, 87.2, 62.6, 14.6.

HRMS (ESI) Calcd. for C₁₂H₁₂INO₂: [M+Na]⁺, 351.9810. Found: m/z 351.9808.

4. Characterization Data of Isolated Products

2-methoxy-3-methylene-1-tosylindoline (**3a**):



The representative general procedure E was followed, using **1a** (82 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3a** (55 mg, 88% yield) as a brown solid. R_f 0.71 (20% EtOAc in hexane).

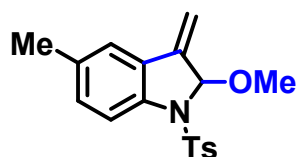
¹H NMR (400 MHz, CDCl₃): δ 7.65 (d, J = 8.3 Hz, 2H), 7.61 (d, J = 8.1 Hz, 1H), 7.35 (d, J = 7.6 Hz, 1H), 7.29-7.24 (m, 1H), 7.19 (d, J = 8.1 Hz, 2H), 7.04 (td, J = 7.5, 0.9 Hz, 1H), 5.89 (t, J = 1.6 Hz, 1H), 5.68 (d, J = 1.7 Hz, 1H), 5.35 (d, J = 1.3 Hz, 1H), 3.31 (s, 3H), 2.35 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 144.2, 142.7, 141.2, 135.4, 130.2, 129.6, 128.1, 127.2, 124.2, 120.9, 115.8, 109.3, 92.9, 52.2, 21.5.

Melting point: 125-127 °C.

HRMS (ESI) Calcd. for C₁₇H₁₇NO₃S: [M-OCH₃]⁺, 284.0740. Found: m/z 284.0751.

2-methoxy-5-methyl-3-methylene-1-tosylindoline (3b):



The representative general procedure **E** was followed, using **1b** (85 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3b** (56 mg, 85% yield) as a white solid. R_f 0.71 (20 % EtOAc in hexane).

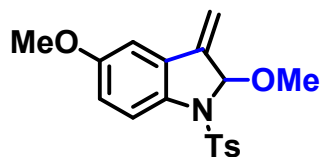
¹H NMR (500 MHz, CDCl₃): δ 7.63 (d, J = 8.3 Hz, 2H), 7.52 (d, J = 8.3 Hz, 1H), 7.18 (d, J = 8.1 Hz, 2H), 7.14 (s, 1H), 7.08 (d, J = 8.3 Hz, 1H), 5.85 (t, J = 1.5 Hz, 1H), 5.63 (d, J = 1.7 Hz, 1H), 5.30 (d, J = 1.3 Hz, 1H), 3.32 (s, 3H), 2.34 (s, 3H), 2.29 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 144.1, 141.4, 140.5, 135.3, 134.1, 131.0, 129.6, 128.2, 127.2, 121.3, 115.8, 108.9, 93.1, 52.3, 21.5, 20.9.

Melting point: 103-105 °C.

HRMS (ESI) Calcd. for C₁₈H₁₉NO₃S: [M-OCH₃]⁺, 298.0896. Found: m/z 298.0898.

2,5-dimethoxy-3-methylene-1-tosylindoline (3c):



The representative general procedure **E** was followed, using **1c** (88 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3c** (55 mg, 80% yield) as a brown solid. R_f 0.71 (20 % EtOAc in hexane).

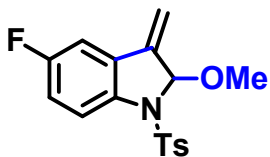
¹H NMR (400 MHz, CDCl₃): δ 7.56 (d, J = 8.4 Hz, 3H), 7.16 (d, J = 7.9 Hz, 2H), 6.86-6.82 (m, 2H), 5.77 (t, J = 1.5 Hz, 1H), 5.60 (d, J = 1.5 Hz, 1H), 5.31 (d, J = 1.0 Hz, 1H), 3.77 (s, 3H), 3.38 (s, 3H), 2.33 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 157.2, 144.1, 141.7, 136.4, 134.9, 129.7, 129.6, 127.2, 117.6, 116.6, 109.5, 105.5, 93.5, 55.7, 52.8, 21.5.

Melting point: 98-100 °C.

HRMS (ESI) Calcd. for C₁₈H₁₉NO₄S: [M-OCH₃]⁺, 314.0845. Found: m/z 314.0843.

5-fluoro-2-methoxy-3-methylene-1-tosylindoline (**3d**):



The representative general procedure **E** was followed, using **1d** (86 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 10% EtOAc in hexane) furnished **3d** (49 mg, 74% yield) as a sticky brown solid. R_f 0.71 (20 % EtOAc in hexane).

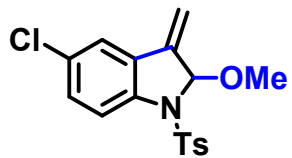
^1H NMR (400 MHz, CDCl_3): δ 7.60-7.57 (m, 3H), 7.19 (d, $J = 8.0$ Hz, 2H), 7.01-6.94 (m, 2H), 5.84 (t, $J = 1.6$ Hz, 1H), 5.64 (d, $J = 1.5$ Hz, 1H), 5.39 (s, 1H), 3.37 (s, 3H), 2.35 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 160.2 (d, $J = 243.5$ Hz), 144.4, 140.9 (d, $J = 2.8$ Hz), 138.7, 134.9, 130.1 (d, $J = 8.7$ Hz), 129.7, 127.2, 117.6 (d, $J = 8.5$ Hz), 117.0 (d, $J = 24.1$ Hz), 110.8, 107.8 (d, $J = 24.4$ Hz), 93.5, 52.9, 21.5.

^{19}F NMR (376 MHz): δ -118.2.

HRMS (ESI) Calcd. for $\text{C}_{17}\text{H}_{16}\text{FNO}_3\text{S}$: $[\text{M}-\text{OCH}_3]^+$, 302.0646. Found: m/z 302.0652.

5-chloro-2-methoxy-3-methylene-1-tosylindoline (**3e**):



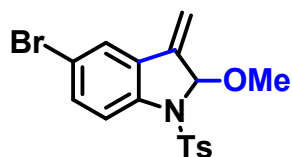
The representative general procedure **E** was followed, using **1e** (89 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3e** (44 mg, 63% yield) as a sticky yellow. R_f 0.71 (20 % EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 7.63 (d, $J = 8.4$ Hz, 2H), 7.55 (d, $J = 8.6$ Hz, 1H), 7.29 (d, $J = 2.1$ Hz, 1H), 7.24-7.18 (m, 3H), 5.88 (t, $J = 1.6$ Hz, 1H), 5.68 (d, $J = 1.2$ Hz, 1H), 5.39 (s, 1H), 3.31 (s, 3H), 2.35 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 144.5, 141.2, 140.2, 135.1, 130.1, 129.9, 129.8, 127.2, 121.1, 116.9, 110.8, 93.2, 52.4, 21.6.

HRMS (ESI) Calcd. for $\text{C}_{17}\text{H}_{16}\text{ClNO}_3\text{S}$: $[\text{M}-\text{OCH}_3]^+$, 318.0350. Found: m/z 318.0392.

5-bromo-2-methoxy-3-methylene-1-tosylindoline (**3f**):



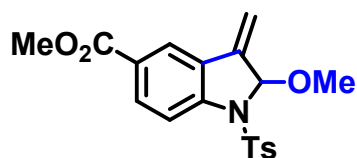
The representative general procedure **E** was followed, using **1f** (98 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3f** (62 mg, 79% yield) as a sticky yellow. R_f 0.71 (20 % EtOAc in hexane).

¹H NMR (400 MHz, CDCl₃): δ 7.63 (d, J = 8.3 Hz, 2H), 7.49 (d, J = 8.6 Hz, 1H), 7.44 (d, J = 1.9 Hz, 1H), 7.36 (dd, J = 8.6, 2.0 Hz, 1H), 7.20 (d, J = 8.0 Hz, 2H), 5.88 (t, J = 1.6 Hz, 1H), 5.68 (d, J = 1.2 Hz, 1H), 5.39 (s, 3H), 3.29 (s, 3H), 2.35 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 144.5, 141.7, 140.1, 135.1, 132.9, 130.1, 129.8, 127.2, 124.1, 117.3, 117.2, 110.8, 93.0, 52.4, 21.5.

HRMS (ESI) Calcd. for C₁₇H₁₆BrNO₃S: [M-OCH₃]⁺, 361.9845. Found: m/z 361.9841.

methyl 2-methoxy-3-methylene-1-tosylindoline-5-carboxylate (**3g**):



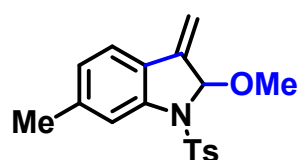
The representative general procedure **E** was followed, using **1g** (94 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 10% EtOAc in hexane) furnished **3g** (49 mg, 65% yield) as a sticky yellow. R_f 0.71 (20 % EtOAc in hexane).

¹H NMR (400 MHz, CDCl₃): δ 8.03 (d, J = 1.6 Hz, 1H), 7.96 (dd, J = 8.6, 1.7 Hz, 1H), 7.71 (d, J = 8.4 Hz, 2H), 7.62 (d, J = 8.5 Hz, 1H), 7.22 (d, J = 8.3 Hz, 2H), 6.02 (t, J = 1.5 Hz, 1H), 5.82 (d, J = 1.6 Hz, 1H), 5.44 (s, 1H), 3.89 (s, 3H), 3.23 (s, 3H), 2.36 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 166.4, 146.2, 144.6, 139.9, 135.4, 132.0, 129.8, 127.9, 127.2, 125.9, 122.6, 114.5, 110.7, 93.2, 52.2, 51.9, 21.5.

HRMS (ESI) Calcd. for C₁₉H₁₉NO₅S: [M-OCH₃]⁺, 342.0795. Found: m/z 342.0797.

2-methoxy-6-methyl-3-methylene-1-tosylindoline (**3h**):



The representative general procedure **E** was followed, using **1h** (85 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3h** (55 mg, 83% yield) as a brown solid. R_f 0.71 (20 % EtOAc in hexane).

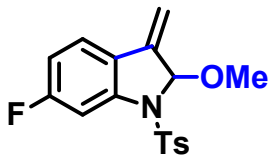
¹H NMR (500 MHz, CDCl₃): δ 7.66 (d, J = 8.3 Hz, 2H), 7.45 (s, 1H), 7.23 (d, J = 7.8 Hz, 1H), 7.19 (d, J = 8.2 Hz, 2H), 6.85 (d, J = 7.8 Hz, 1H), 5.88 (s, 1H), 5.59 (d, J = 1.5 Hz, 1H), 5.27 (s, 1H), 3.28 (s, 3H), 2.37 (s, 3H), 2.35 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 144.1, 142.9, 141.0, 140.8, 135.5, 129.6, 127.2, 125.5, 125.2, 120.7, 116.3, 108.0, 93.2, 52.0, 22.0, 21.5.

Melting point: 83-85 °C.

HRMS (ESI) Calcd. for C₁₈H₁₉NO₃S: [M-OCH₃]⁺, 298.0896. Found: m/z 298.0903.

6-fluoro-2-methoxy-3-methylene-1-tosylindoline (**3i**):



The representative general procedure **E** was followed, using **1i** (86 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 10% EtOAc in hexane) furnished **3i** (55 mg, 82% yield) as a brown solid. R_f 0.71 (20 % EtOAc in hexane).

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.62 (d, $J = 8.3$ Hz, 2H), 7.29-7.15 (m, 4H), 6.66 (td, $J = 8.6, 2.3$ Hz, 1H), 5.86 (t, $J = 1.6$ Hz, 1H), 5.54 (d, $J = 1.4$ Hz, 1H), 5.24 (s, 1H), 3.19 (s, 3H), 2.29 (s, 3H).

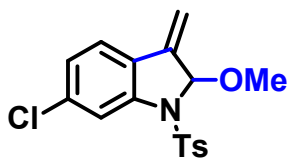
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 164.1 (d, $J = 248.4$ Hz), 144.5, 144.0 (d, $J = 11.9$ Hz), 140.0, 135.3, 129.8, 127.2, 123.9 (d, $J = 2.5$ Hz), 122.1 (d, $J = 10.1$ Hz), 111.3 (d, $J = 23.5$ Hz), 108.7 (d, $J = 2.7$ Hz), 103.6 (d, $J = 28.4$ Hz), 93.4, 52.1, 21.6.

$^{19}\text{F NMR}$ (376 MHz): δ -108.6.

Melting point: 88-90 °C.

HRMS (ESI) Calcd. for $\text{C}_{17}\text{H}_{16}\text{FNO}_3\text{S}$: $[\text{M}-\text{OCH}_3]^+$, 302.0646. Found: m/z 302.0646.

6-chloro-2-methoxy-3-methylene-1-tosylindoline (**3j**):



The representative general procedure **E** was followed, using **1j** (89 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3j** (51 mg, 73% yield) as a brown solid. R_f 0.71 (20 % EtOAc in hexane).

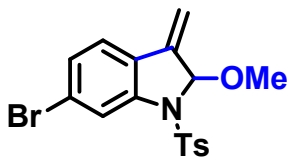
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.68 (d, $J = 8.4$ Hz, 2H), 7.62 (d, $J = 1.8$ Hz, 1H), 7.26-7.21 (m, 3H), 6.99 (dd, $J = 8.2, 1.8$ Hz, 1H), 5.92 (t, $J = 1.6$ Hz, 1H), 5.67 (d, $J = 1.3$ Hz, 1H), 5.36 (d, $J = 0.9$ Hz, 1H), 3.25 (s, 3H), 2.36 (s, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 144.5, 143.6, 140.1, 135.9, 135.2, 129.8, 127.2, 126.5, 124.4, 121.7, 115.8, 109.8, 93.2, 52.1, 21.6.

Melting point: 80-82 °C.

HRMS (ESI) Calcd. for $\text{C}_{17}\text{H}_{16}\text{ClNO}_3\text{S}$: $[\text{M}-\text{OCH}_3]^+$, 318.0350. Found: m/z 318.0386.

6-bromo-2-methoxy-3-methylene-1-tosylindoline (**3k**):



The representative general procedure **E** was followed, using **1k** (98 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3k** (67 mg, 85% yield) as a brown solid. R_f 0.71 (20 % EtOAc in hexane).

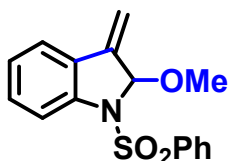
¹H NMR (400 MHz, CDCl₃): δ 7.79 (d, J = 1.5 Hz, 1H), 7.68 (d, J = 8.3 Hz, 2H), 7.26-7.14 (m, 4H), 5.92 (t, J = 1.6 Hz, 1H), 5.69 (d, J = 1.7 Hz, 1H), 5.38 (d, J = 0.9 Hz, 1H), 3.25 (s, 3H), 2.37 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 144.5, 143.7, 140.2, 135.2, 129.8, 127.3, 127.2, 126.9, 123.9, 122.0, 118.7, 110.0, 93.1, 52.1, 21.6.

Melting point: 98-100 °C.

HRMS (ESI) Calcd. for C₁₇H₁₆BrNO₃S: [M-OCH₃]⁺, 361.9845. Found: m/z 361.9844.

2-methoxy-3-methylene-1-(phenylsulfonyl)indoline (3l):



The representative general procedure E was followed, using **1l** (79 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3l** (49 mg, 82% yield) as a yellow solid. R_f 0.71 (20 % EtOAc in hexane).

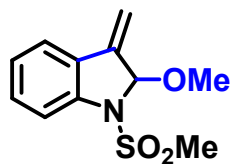
¹H NMR (400 MHz, CDCl₃): δ 7.75 (d, J = 7.6 Hz, 2H), 7.59 (d, J = 8.1 Hz, 1H), 7.48 (t, J = 7.4 Hz, 1H), 7.39-7.35 (m, 2H), 7.31 (d, J = 7.6 Hz, 1H), 7.23 (d, J = 7.1 Hz, 1H), 7.01 (t, J = 7.5 Hz, 1H), 5.88 (s, 1H), 5.65 (d, J = 1.3 Hz, 1H), 5.32 (s, 1H), 3.26 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 142.6, 141.1, 138.4, 133.3, 130.2, 129.0, 128.1, 127.2, 124.4, 121.0, 115.7, 109.4, 92.9, 52.3.

Melting point: 60-62°C.

HRMS (ESI) Calcd. for C₁₆H₁₅NO₃S: [M-OCH₃]⁺, 270.0583. Found: m/z 270.0583.

2-methoxy-3-methylene-1-(methylsulfonyl)indoline (3m):



The representative general procedure E was followed, using **1m** (67 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 15% EtOAc in hexane) furnished **3m** (40 mg, 84% yield) as a brown solid. R_f 0.51 (20 % EtOAc in hexane).

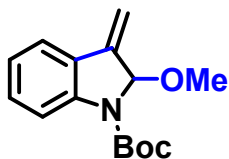
¹H NMR (400 MHz, CDCl₃): δ 7.49-7.45 (m, 2H), 7.32-7.28 (m, 1H), 7.10 (td, J = 7.5, 0.9 Hz, 1H), 5.85 (t, J = 1.5 Hz, 1H), 5.82 (d, J = 1.6 Hz, 1H), 5.46 (d, J = 1.1 Hz, 1H), 3.39 (s, 3H), 2.96 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 142.6, 141.0, 130.5, 127.5, 124.2, 121.4, 114.5, 109.6, 93.0, 52.8, 39.0.

Melting point: 78-80 °C.

HRMS (ESI) Calcd. for C₁₁H₁₃NO₃S: [M-OCH₃]⁺, 208.0427. Found: m/z 208.0425.

tert-butyl 2-methoxy-3-methyleneindoline-1-carboxylate (**3n**):



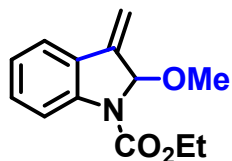
The representative general procedure **E** was followed, using **1n** (72 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3n** (44 mg, 83% yield) as a sticky yellow solid. R_f 0.71 (20 % EtOAc in hexane).

$^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.67 (br, 1H), 7.35 (d, $J = 7.5$ Hz, 1H), 7.17 (t, $J = 7.6$ Hz, 1H), 6.92 (t, $J = 7.5$ Hz, 1H), 5.86 (s, 1H), 5.67 (d, $J = 1.2$ Hz, 1H), 5.32 (s, 1H), 3.20 (s, 3H), 1.51 (s, 9H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 152.0, 143.6, 141.8, 130.0, 127.0, 122.8, 120.3, 115.5, 108.0, 90.1, 81.8, 52.0, 28.4.

HRMS (ESI) Calcd. for $\text{C}_{15}\text{H}_{19}\text{NO}_3$: $[\text{M}+\text{Na}]^+$, 284.1263. Found: m/z 284.1259.

ethyl 2-methoxy-3-methyleneindoline-1-carboxylate (**3o**):



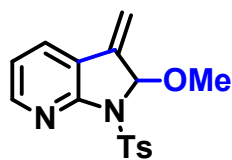
The representative general procedure **E** was followed, using **1o** (66 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3o** (36 mg, 77% yield) as a sticky yellow. R_f 0.71 (20 % EtOAc in hexane).

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.81 (br, 1H), 7.45 (dd, $J = 7.6, 0.6$ Hz, 1H), 7.30-7.25 (m, 1H), 7.03 (td, $J = 7.5, 0.9$ Hz, 1H), 6.02 (t, $J = 1.5$ Hz, 1H), 5.78 (d, $J = 1.8$ Hz, 1H), 5.42 (d, $J = 1.4$ Hz, 1H), 4.39-4.31 (m, 2H), 3.26 (s, 3H), 1.39 (t, $J = 7.1$ Hz, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 153.1, 141.5, 130.2, 127.0, 123.2, 123.1, 120.5, 115.5, 108.5, 108.4, 89.9, 53.0, 62.1, 14.6.

HRMS (ESI) Calcd. for $\text{C}_{13}\text{H}_{15}\text{NO}_3$: $[\text{M}+\text{Na}]^+$, 256.0950. Found: m/z 256.0941.

2-methoxy-3-methylene-1-tosyl-2,3-dihydro-1H-pyrrolo[2,3-b]pyridine (**3p**):



The representative general procedure **E** was followed, using **1p** (82 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3p** (52 mg, 82% yield) as a yellow solid. R_f 0.71 (20 % EtOAc in hexane).

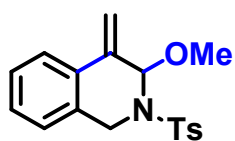
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.15 (dd, $J = 5.1, 1.6$ Hz, 1H), 7.97 (d, $J = 8.4$ Hz, 2H), 7.54 (dd, $J = 7.5, 1.6$ Hz, 1H), 7.19 (d, $J = 8.0$ Hz, 2H), 6.83 (dd, $J = 7.5, 5.1$ Hz, 1H), 6.13 (t, $J = 1.6$ Hz, 1H), 5.75 (d, $J = 1.5$ Hz, 1H), 5.47 (s, 1H), 3.10 (s, 3H), 2.31 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 155.9, 149.2, 144.2, 138.5, 136.8, 129.3, 128.7, 128.1, 120.7, 118.6, 111.9, 91.0, 51.8, 21.6.

Melting point: 95-96 °C.

HRMS (ESI) Calcd. for C₁₆H₁₆N₂O₃S: [M+H]⁺, 317.0960. Found: m/z 317.0958.

3-methoxy-4-methylene-2-tosyl-1,2,3,4-tetrahydroisoquinoline (3q)⁹:



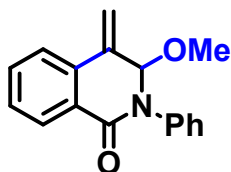
The representative general procedure **E** was followed, using **1q** (75 mg, 0.2 mmol), for 18 h. Purification by column chromatography (eluted with 7% EtOAc in hexane) furnished **3q** (58mg, 88% yield) as a yellow liquid. R_f 0.71 (20 % EtOAc in hexane).

¹H NMR (400 MHz, CDCl₃): δ 7.66 (d, *J* = 7.9 Hz, 2H), 7.52 (d, *J* = 6.1 Hz, 1H), 7.19 (d, *J* = 7.5 Hz, 4H), 7.02 (d, *J* = 6.0 Hz, 1H), 5.65 (d, *J* = 8.1 Hz, 2H), 5.25 (s, 1H), 4.56 (d, *J* = 16.1 Hz, 1H), 4.37 (d, *J* = 16.1 Hz, 1H), 3.36 (s, 3H), 2.35 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 143.5, 137.8, 136.2, 130.8, 129.8, 129.4, 128.4, 127.4, 127.2, 125.9, 124.8, 112.5, 87.3, 54.9, 43.2, 21.5.

HRMS (ESI) Calcd. for C₁₈H₁₉NO₃S: [M+Na]⁺, 352.0983. Found: m/z 352.0974.

3-methoxy-4-methylene-2-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3r)⁹:



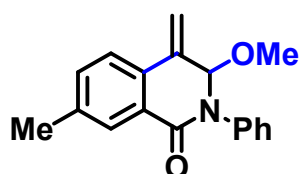
The representative general procedure **E** was followed, using **1r** (72 mg, 0.2 mmol), for 18 h. Purification by column chromatography (eluted with 10% EtOAc in hexane) furnished **3r** (44 mg, 84% yield) as a sticky red. R_f 0.71 (20 % EtOAc in hexane).

¹H NMR (400 MHz, CDCl₃): δ 8.22 (dd, *J* = 7.8, 0.9 Hz, 1H), 7.65-7.62 (m, 1H), 7.58 (td, *J* = 7.3, 1.4 Hz, 1H), 7.50-7.41 (m, 5H), 7.35-7.30 (m, 1H), 5.87 (s, 1H), 5.42 (s, 1H), 5.18 (s, 1H), 3.23 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 163.3, 141.8, 136.5, 133.8, 132.7, 129.1, 129.0, 128.8, 127.2, 126.8, 123.9, 115.9, 93.4, 54.1.

HRMS (ESI) Calcd. for C₁₇H₁₅NO₂: [M+H]⁺, 266.1181. Found: m/z 266.1179.

3-methoxy-7-methyl-4-methylene-2-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3s):



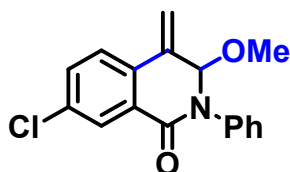
The representative general procedure **E** was followed, using **1s** (75 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3s** (49 mg, 88% yield) as a sticky yellow. R_f 0.71 (20 % EtOAc in hexane).

¹H NMR (400 MHz, CDCl₃): δ 8.02 (s, 1H), 7.53 (d, J = 7.9 Hz, 1H), 7.49-7.37 (m, 5H), 7.34-7.30 (m, 1H), 5.82 (s, 1H), 5.36 (s, 1H), 5.16 (s, 1H), 3.22 (s, 3H), 2.43 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 163.5, 141.9, 139.3, 136.4, 134.7, 133.6, 131.1, 129.0, 127.1, 126.9, 126.8, 123.9, 114.9, 93.4, 54.1, 21.3.

HRMS (ESI) Calcd. for C₁₈H₁₇NO₂: [M+H]⁺, 280.1338. Found: m/z 280.1341.

7-chloro-3-methoxy-4-methylene-2-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3t):



The representative general procedure **E** was followed, using **1t** (79 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3t** (50 mg, 83% yield) as a pale yellow solid. R_f 0.71 (20 % EtOAc in hexane).

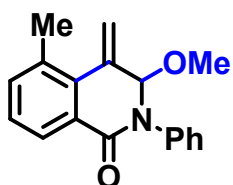
¹H NMR (400 MHz, CDCl₃): δ 8.15 (d, J = 8.4 Hz, 1H), 7.60 (d, J = 1.8 Hz, 1H), 7.45-7.41 (m, 5H), 7.36-7.31 (m, 1H), 5.87 (s, 1H), 5.46 (s, 1H), 5.16 (s, 1H), 3.22 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 162.5, 141.5, 138.9, 135.6, 135.3, 130.6, 129.3, 129.1, 127.4, 126.8, 125.7, 124.1, 116.9, 93.3, 54.3.

Melting point: 118-120 °C.

HRMS (ESI) Calcd. for C₁₇H₁₄ClNO₂: [M+H]⁺, 300.0791. Found: m/z 300.0788.

3-methoxy-5-methyl-4-methylene-2-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3u):



The representative general procedure **E** was followed, using **1u** (75 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3u** (42 mg, 75% yield) as a yellow solid. R_f 0.72 (20 % EtOAc in hexane).

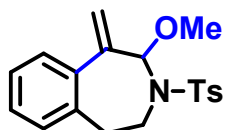
¹H NMR (400 MHz, CDCl₃): δ 8.09 (dd, J = 7.7, 0.8 Hz, 1H), 7.49-7.41 (m, 5H), 7.38-7.29 (m, 2H), 5.71 (s, 1H), 5.66 (s, 1H), 5.07 (s, 1H), 3.22 (s, 3H), 2.57 (s, 3H).

¹³C NMR (125 MHz, CDCl₃): δ 163.5, 141.8, 136.6, 135.4, 135.1, 132.4, 129.0, 128.7, 128.1, 127.1, 126.9, 126.6, 120.0, 95.1, 54.5, 21.2.

Melting point: 123-125 °C.

HRMS (ESI) Calcd. for C₁₈H₁₇NO₂: [M+H]⁺, 280.1338. Found: m/z 280.1326.

2-methoxy-1-methylene-3-tosyl-2,3,4,5-tetrahydro-1H-benzo[d]azepine (3v):



The representative general procedure **E** was followed, using **1v** (88 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 7%

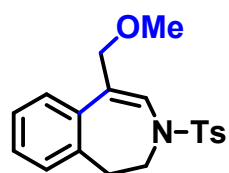
EtOAc in hexane) furnished **3v** (15 mg, 22% yield) as a yellow liquid. R_f 0.71 (20 % EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 7.63 (d, $J = 8.3$ Hz, 2H), 7.21-7.13 (m, 5H), 6.96-6.94 (m, 1H), 5.79 (s, 1H), 5.41 (s, 1H), 5.26 (d, $J = 0.9$ Hz, 1H), 3.65-3.59 (m, 1H), 3.48-3.40 (m, 1H), 3.29 (s, 3H), 2.89-2.74 (m, 2H), 2.38 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 144.1, 140.1, 135.3, 134.7, 129.9, 129.4, 128.7, 127.3, 127.1, 126.8, 126.5, 114.7, 77.8, 57.0, 49.4, 35.4, 21.5.

HRMS (ESI) Calcd. for $\text{C}_{19}\text{H}_{21}\text{NO}_3\text{S}$: $[\text{M}-\text{OCH}_3]^+$, 312.1053. Found: m/z 312.1050.

5-(methoxymethyl)-3-tosyl-2,3-dihydro-1H-benzo[d]azepine (**4v**):



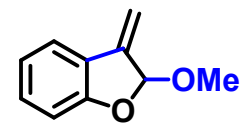
The representative general procedure **E** was followed, using **1v** (88 mg, 0.2 mmol) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **4v** (21 mg, 31% yield) as a yellow liquid. R_f 0.58 (20 % EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 7.68 (d, $J = 8.3$ Hz, 2H), 7.51 (dd, $J = 8.0, 0.7$ Hz, 1H), 7.29 (d, $J = 8.0$ Hz, 2H), 7.23-7.19 (m, 2H), 7.09 (td, $J = 7.4, 1.1$ Hz, 1H), 6.97 (dd, $J = 7.4, 0.8$ Hz, 1H), 4.31 (s, 2H), 3.78-3.76 (m, 2H), 3.34 (s, 3H), 2.80-2.78 (m, 2H), 2.40 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 144.1, 140.1, 135.3, 134.7, 129.9, 129.4, 128.7, 127.3, 127.1, 126.8, 126.5, 114.7, 77.8, 57.0, 49.4, 35.4, 21.5.

HRMS (ESI) Calcd. for $\text{C}_{19}\text{H}_{21}\text{NO}_3\text{S}$: $[\text{M}-\text{OCH}_3]^+$, 312.1053. Found: m/z 312.1057.

2-methoxy-3-methylene-2,3-dihydrobenzofuran (**3w**):



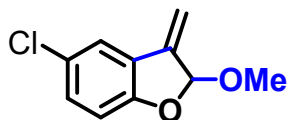
The representative general procedure **E** was followed, using **1w** (77 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 2% EtOAc in hexane) furnished **3w** (32 mg, 66% yield) as a yellow liquid. R_f 0.51 (5 % EtOAc in hexane).

^1H NMR (400 MHz, CDCl_3): δ 7.40 (dd, $J = 7.5, 0.8$ Hz, 1H), 7.25-7.20 (m, 1H), 6.92 (td, $J = 7.5, 0.9$ Hz, 1H), 6.87 (d, $J = 8.1$ Hz, 1H), 5.98 (t, $J = 1.9$ Hz, 1H), 5.70 (d, $J = 2.1$ Hz, 1H), 5.34 (d, $J = 1.7$ Hz, 1H) 3.52 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ 160.9, 143.2, 130.7, 124.3, 121.1, 121.0, 110.5, 107.1, 106.9, 54.5.

HRMS (ESI) Calcd. for $\text{C}_{10}\text{H}_{10}\text{O}_2$: $[\text{M}-2\text{H}]^+$, 161.0597. Found: m/z 161.0599.

5-chloro-2-methoxy-3-methylene-2,3-dihydrobenzofuran (**3x**):



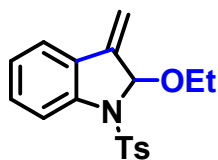
The representative general procedure **E** was followed, using **1x** (88 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 2% EtOAc in hexane) furnished **3x** (31 mg, 53% yield) as a yellow liquid. R_f 0.53 (5% EtOAc in hexane).

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.35 (d, $J = 2.2$ Hz, 1H), 7.17 (dd, $J = 8.6, 2.3$ Hz, 1H), 6.79 (d, $J = 8.5$ Hz, 1H), 5.98 (t, $J = 1.9$ Hz, 1H), 5.70 (d, $J = 2.0$ Hz, 1H), 5.38 (d, $J = 1.7$ Hz, 1H) 3.51 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 159.5, 142.2, 130.5, 126.2, 125.9, 121.1, 111.5, 108.5, 107.7, 54.7.

HRMS (ESI) Calcd. for $\text{C}_{10}\text{H}_9\text{ClO}_2$: $[\text{M}-2\text{H}]^+$, 195.0207. Found: m/z 195.0202.

2-ethoxy-3-methylene-1-tosylindoline (**3aa**):



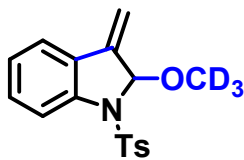
The representative general procedure **E** was followed, using **1a** (82 mg, 0.2 mmol), and **Ethanol** (3ml) at 50°C for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3aa** (35 mg, 53% yield) as a yellow liquid. R_f 0.71 (20 % EtOAc in hexane).

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.64 (d, $J = 8.3$ Hz, 2H), 7.60 (d, $J = 8.1$ Hz, 1H), 7.33 (dd, $J = 7.6, 0.6$ Hz, 1H), 7.27-7.23 (m, 1H), 7.18 (d, $J = 7.9$ Hz, 2H), 7.03 (td, $J = 7.5, 0.9$ Hz, 1H), 5.92 (t, $J = 1.6$ Hz, 1H), 5.64 (d, $J = 1.6$ Hz, 1H), 5.34 (d, $J = 1.3$ Hz, 1H), 3.77-3.69 (m, 1H), 3.61-3.54 (m, 1H), 2.34 (s, 3H), 1.19 (t, $J = 7.0$ Hz, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 144.1, 142.7, 141.9, 135.5, 130.1, 129.6, 128.2, 127.2, 124.3, 121.0, 116.0, 109.1, 92.3, 61.2, 21.5, 15.0.

HRMS (ESI) Calcd. for $\text{C}_{18}\text{H}_{19}\text{NO}_3\text{S}$: $[\text{M}+\text{Na}]^+$, 352.0983. Found: m/z 352.0982.

2-(methoxy- d_3)-3-methylene-1-tosylindoline (**3a-D**):



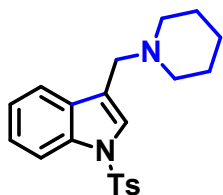
The representative general procedure **E** was followed, using **1a** (82 mg, 0.2 mmol), and CD_3OD (3mL) for 18 h. Purification by column chromatography (eluted with 8% EtOAc in hexane) furnished **3a-D** (31 mg, 49% yield) as a brown solid. R_f 0.71 (% EtOAc in hexane).

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.66 (d, $J = 8.3$ Hz, 2H), 7.61 (d, $J = 8.1$ Hz, 1H), 7.34 (d, $J = 7.6$ Hz, 1H), 7.28-7.24 (m, 1H), 7.18 (d, $J = 8.1$ Hz, 2H), 7.04 (td, $J = 7.5, 0.8$ Hz, 1H), 5.89 (t, $J = 1.5$ Hz, 1H), 5.68 (d, $J = 1.5$ Hz, 1H), 5.35 (s, 1H), 2.34 (s, 3H).

Melting point: 86-88 °C.

HRMS (ESI) Calcd. for $\text{C}_{17}\text{H}_{14}\text{D}_3\text{NO}_3\text{S}$: $[\text{M}+\text{Na}]^+$, 341.1015. Found: m/z 341.1001.

3-(piperidin-1-ylmethyl)-1-tosyl-1H-indole (4ad):



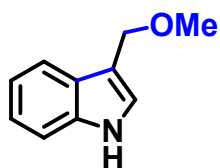
The representative general procedure **F** was followed, using **1a** (123 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 20% acetone in hexane) furnished **4ad** (62 mg, 56% yield) as a sticky yellow. R_f 0.52 (30 % acetone in hexane).

^1H NMR (500 MHz, CDCl_3): δ 7.96 (d, $J = 8.3$ Hz, 1H), 7.74 (d, $J = 8.3$ Hz, 2H), 7.65 (d, $J = 7.8$ Hz, 1H), 7.49 (s, 1H), 7.29 (t, $J = 7.7$ Hz, 1H), 7.22 (d, $J = 7.4$ Hz, 1H), 7.19 (d, $J = 8.3$ Hz, 2H), 3.61 (s, 2H), 2.41 (br, 4H), 2.32 (s, 3H), 1.59-1.54 (m, 4H), 1.41 (br, 2H).

^{13}C NMR (125 MHz, CDCl_3): δ 144.8, 135.4, 135.3, 131.2, 129.8, 126.8, 125.0, 124.6, 123.1, 120.4, 119.3, 113.6, 54.3, 53.6, 25.8, 24.2, 21.5.

HRMS (ESI) Calcd. for $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_2\text{S}$: $[\text{M}+\text{H}]^+$, 369.1637. Found: m/z 369.1629.

3-(methoxymethyl)-1H-indole (4a):¹⁰



The representative general procedure **E** was followed, using **1'a** (90 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4a** (38 mg, 79% yield) as a yellow solid. R_f 0.51 (20 % EtOAc in hexane).

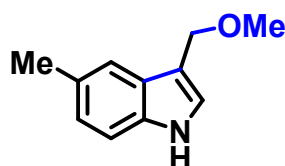
^1H NMR (400 MHz, CDCl_3): δ 8.17 (brs, 1H), 7.73-7.71 (m, 1H), 7.37-7.34 (m, 1H), 7.23-7.19 (m, 1H), 7.18-7.13 (m, 2H), 4.68 (d, $J = 0.6$ Hz, 2H), 3.40 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 135.4, 126.1, 122.7, 121.3, 118.8, 118.1, 112.2, 110.1, 65.5, 56.5.

Melting point: 98-99 °C.

HRMS (ESI) Calcd. for $\text{C}_{10}\text{H}_{11}\text{NO}$: $[\text{M}-\text{OCH}_3]^+$, 130.0651. Found: m/z 130.0650.

3-(methoxymethyl)-5-methyl-1H-indole (4b):¹⁰



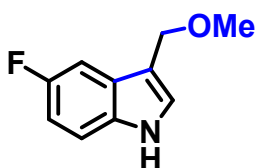
The representative general procedure **E** was followed, using **1'b** (94 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4b** (41 mg, 77% yield) as a sticky brown. R_f 0.51 (20 % EtOAc in hexane).

¹H NMR (400 MHz, D₆-acetone): δ 9.99 (brs, 1H), 7.44 (d, J = 0.7 Hz, 1H), 7.28 (d, J = 8.3 Hz, 1H), 7.24 (d, J = 2.3 Hz, 1H), 6.96 (dd, J = 8.3, 1.4 Hz, 1H), 4.60 (s, 2H), 3.29 (s, 3H), 2.41 (s, 3H).

¹³C NMR (100 MHz, D₆-acetone): δ 136.2, 128.8, 128.6, 125.4, 123.9, 119.5, 112.9, 111.9, 67.0, 57.1, 21.6.

HRMS (ESI) Calcd. for C₁₁H₁₃NO: [M-OCH₃]⁺, 144.0808. Found: m/z 144.0803.

5-fluoro-3-(methoxymethyl)-1H-indole (4c):¹⁰



The representative general procedure E was followed, using **1'c** (95 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4c** (37 mg, 68% yield) as a sticky brown. R_f 0.51 (20 % EtOAc in hexane).

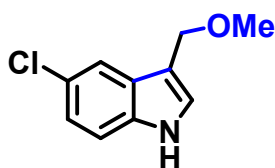
¹H NMR (400 MHz, D₆-acetone): δ 10.26 (brs, 1H), 7.41-7.37 (m, 2H), 7.31 (dd, J = 9.9, 2.5 Hz, 1H), 6.91 (td, J = 9.2, 2.5 Hz, 1H), 4.59 (s, 2H), 3.28 (s, 3H).

¹³C NMR (100 MHz, D₆-acetone): δ 158.5 (d, J = 232.1 Hz), 134.4, 128.7 (d, J = 9.9 Hz), 127.3, 113.8 (d, J = 4.7 Hz), 113.1 (d, J = 9.6 Hz), 110.4 (d, J = 26.4 Hz), 104.4 (d, J = 23.4 Hz), 66.8, 57.2.

¹⁹F NMR (376 MHz, D₆-acetone): δ -126.7.

HRMS (ESI) Calcd. for C₁₀H₁₀FNO: [M-OCH₃]⁺, 148.0557. Found: m/z 148.0551.

5-chloro-3-(methoxymethyl)-1H-indole (4d):¹⁰



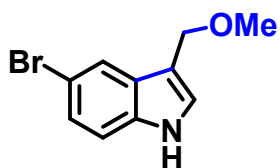
The representative general procedure E was followed, using **1'd** (100 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4d** (41 mg, 70% yield) as a sticky yellow. R_f 0.51 (20 % EtOAc in hexane).

¹H NMR (500 MHz, D₆-acetone): δ 10.35 (brs, 1H), 7.64 (d, J = 1.9 Hz, 1H), 7.41 (d, J = 8.6 Hz, 1H), 7.38 (d, J = 2.1 Hz, 1H), 7.11 (dd, J = 8.6, 1.7 Hz, 1H), 4.60 (s, 2H), 3.29 (s, 3H).

¹³C NMR (125 MHz, D₆-acetone): δ 136.2, 129.5, 126.9, 125.2, 122.4, 119.2, 113.6, 113.4, 66.8, 57.3.

HRMS (ESI) Calcd. for C₁₀H₁₀ClNO: [M-OCH₃]⁺, 164.0262. Found: m/z 164.0251.

5-bromo-3-(methoxymethyl)-1H-indole (4e):¹⁰



The representative general procedure **E** was followed, using **1'e** (113 mg, 0.3 mmol), for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4e** (49 mg, 68% yield) as a pale yellow solid. R_f 0.51 (20 % EtOAc in hexane).

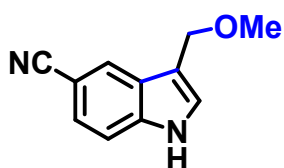
^1H NMR (400 MHz, D_4 -methanol): δ 7.73 (d, $J = 1.5$ Hz, 1H), 7.28-7.26 (m, 2H), 7.20 (dd, $J = 8.6, 1.9$ Hz, 1H), 4.60 (s, 2H), 3.33 (s, 3H).

^{13}C NMR (100 MHz, D_4 -methanol): δ 136.9, 130.2, 127.2, 125.4, 122.3, 113.9, 113.3, 112.7, 67.3, 57.4.

Melting point: 94-96 °C.

HRMS (ESI) Calcd. for $\text{C}_{10}\text{H}_{10}\text{BrNO}$: $[\text{M}-\text{OCH}_3]^+$, 207.9756. Found: m/z 207.9757.

3-(methoxymethyl)-1H-indole-5-carbonitrile (**4f**):



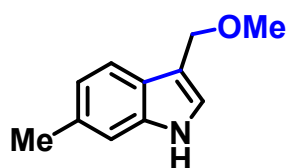
The representative general procedure **E** was followed, using **1'f** (97 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4f** (36 mg, 65% yield) as a sticky brown. R_f 0.51 (20 % EtOAc in hexane).

^1H NMR (400 MHz, D_4 -methanol): δ 8.02 (d, $J = 0.8$ Hz, 1H), 7.48 (dd, $J = 8.4, 0.6$ Hz, 1H), 7.41 (s, 1H), 7.38 (dd, $J = 8.5, 1.5$ Hz, 1H), 4.65 (s, 2H), 3.35 (s, 3H).

^{13}C NMR (100 MHz, D_4 -methanol): δ 140.0, 128.3, 128.2, 125.7, 125.4, 121.8, 114.3, 113.6, 102.8, 67.0, 57.7.

HRMS (ESI) Calcd. for $\text{C}_{11}\text{H}_{10}\text{N}_2\text{O}$: $[\text{M}+\text{Na}]^+$, 209.0691 Found: m/z 209.0688.

3-(methoxymethyl)-6-methyl-1H-indole (**4g**):



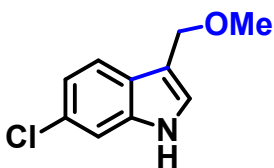
The representative general procedure **E** was followed, using **1'g** (94 mg, 0.3 mmol) for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4g** (40 mg, 76% yield) as a sticky brown. R_f 0.51 (20 % EtOAc in hexane).

^1H NMR (400 MHz, D_6 -acetone): δ 9.95 (brs, 1H), 7.53 (d, $J = 8.1$ Hz, 1H), 7.19 (d, $J = 8.3$ Hz, 2H), 6.89 (d, $J = 8.1$ Hz, 1H), 4.60 (s, 2H), 3.28 (s, 3H), 2.41 (s, 3H).

^{13}C NMR (100 MHz, D_6 -acetone): δ 138.3, 131.7, 126.3, 124.6, 121.6, 119.5, 113.3, 112.1, 67.1, 57.1, 21.8.

HRMS (ESI) Calcd. for $\text{C}_{11}\text{H}_{13}\text{NO}$: $[\text{M}-\text{OCH}_3]^+$, 144.0808. Found: m/z 144.0800.

6-chloro-3-(methoxymethyl)-1H-indole (**4h**):¹⁰



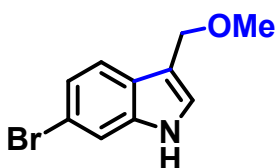
The representative general procedure **E** was followed, using **1h** (100 mg, 0.3 mmol), for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4h** (42 mg, 72% yield) as a sticky brown. R_f 0.51 (20 % EtOAc in hexane).

^1H NMR (400 MHz, D_6 -acetone): δ 10.30 (brs, 1H), 7.62 (d, $J = 8.4$ Hz, 1H), 7.44 (d, $J = 1.8$ Hz, 1H), 7.35 (d, $J = 2.4$ Hz, 1H), 7.04 (dd, $J = 8.4, 1.9$ Hz, 1H), 4.60 (s, 2H), 3.28 (s, 3H).

^{13}C NMR (100 MHz, D_6 -acetone): δ 138.2, 127.8, 127.1, 126.3, 121.1, 120.3, 113.8, 112.0, 66.8, 57.3.

HRMS (ESI) Calcd. for $\text{C}_{10}\text{H}_{10}\text{ClNO}$: $[\text{M}-\text{OCH}_3]^+$, 164.0262. Found: m/z 164.0260.

6-bromo-3-(methoxymethyl)-1H-indole (**4i**):¹⁰



The representative general procedure **E** was followed, using **1i** (113 mg, 0.3 mmol), for 18 h. Purification by column chromatography (eluted with 12% EtOAc in hexane) furnished **4i** (46 mg, 64% yield) as a dark yellow solid. R_f 0.51 (20 % EtOAc in hexane).

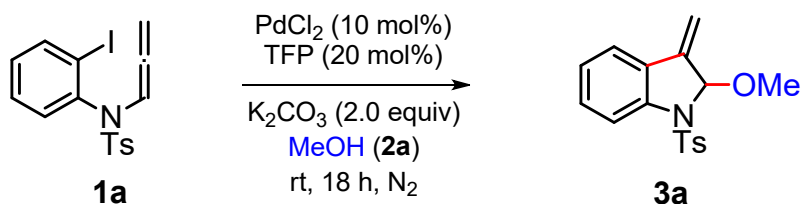
^1H NMR (400 MHz, D_4 -methanol): δ 7.52 (d, $J = 1.7$ Hz, 1H), 7.50 (d, $J = 8.5$ Hz, 1H), 7.22 (s, 1H), 7.14 (dd, $J = 8.4, 1.7$ Hz, 1H), 4.61 (s, 2H), 3.33 (s, 3H).

^{13}C NMR (100 MHz, D_4 -methanol): δ 139.0, 127.4, 126.6, 123.3, 121.2, 116.1, 115.2, 113.2, 67.2, 57.5.

Melting point: 78-80 °C.

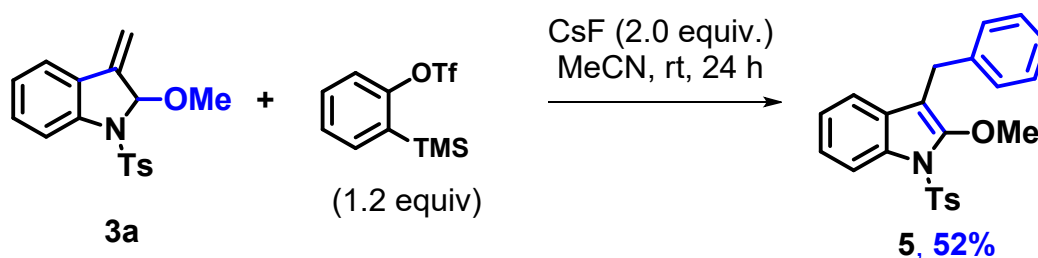
HRMS (ESI) Calcd. for $\text{C}_{10}\text{H}_{10}\text{BrNO}$: $[\text{M}-\text{OCH}_3]^+$, 207.9756. Found: m/z 207.9770.

5. Gram scale synthesis of **3a**:



To an oven dried Schlenk tube was charged with allenamide **1a** (1g, 2.43 mmol), palladium chloride (43 mg, 10 mol%), tri(2-furyl)phosphine (118 mg, 20 mol%), and K_2CO_3 (671 mg, 2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N_2 gas (3X). Methanol (**2a**) 20 mL was added to the reaction mixture. The reaction was stirred at rt for 18 h, and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography to give **3a** as a brown solid (550 mg, 72% yield).

Synthesis of 3-benzyl-2-methoxy-1-tosyl-1H-indole (5):



To an oven dried Schlenk tube was charged with **3a** (95 mg, 0.3 mmol, 1 equiv.), 2-(trimethylsilyl)phenyl trifluoromethanesulfonate (1.2 equiv.) and CsF (2 equiv.) in MeCN (3 mL) was added to the reaction mixture. The reaction was stirred at rt for 24 h, and the solvent was evaporated under reduced pressure. Purification by column chromatography (eluted with 5% EtOAc in hexane) furnished **5** (61 mg, 52% yield) as a white solid. R_f 0.72 (20 % EtOAc in hexane).

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.14 (d, $J = 8.3$ Hz, 1H), 7.70 (d, $J = 8.4$ Hz, 2H), 7.25-7.22 (m, 1H), 7.18-7.11 (m, 7H), 7.02-6.99 (m, 2H), 4.01 (s, 3H), 3.89 (s, 2H), 2.35 (s, 3H).

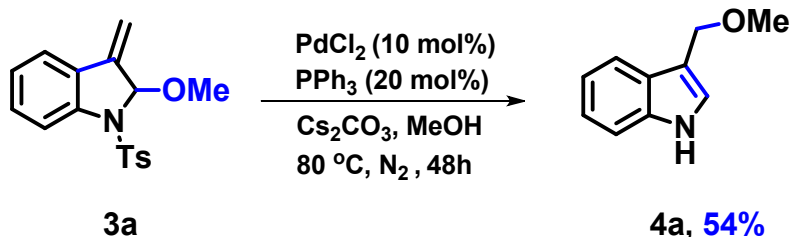
$^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 149.5, 144.7, 139.3, 135.2, 132.6, 129.5, 129.4, 128.3, 128.1, 126.9, 126.1, 123.9, 123.8, 118.9, 115.2, 105.8, 64.9, 28.4, 21.6.

Melting point: 120-122 °C.

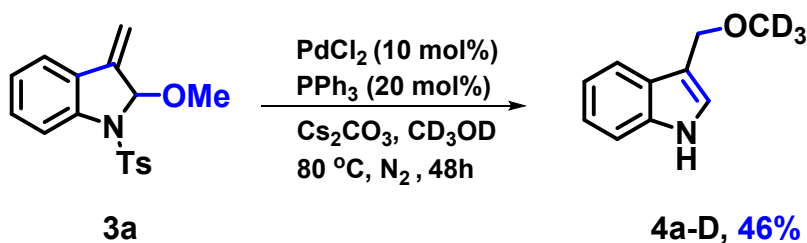
HRMS (ESI) Calcd. for $\text{C}_{23}\text{H}_{21}\text{NO}_3\text{S}$: $[\text{M}-\text{H}]^+$, 392.1320. Found: m/z 392.1312.

6. Mechanistic experiments:

a.

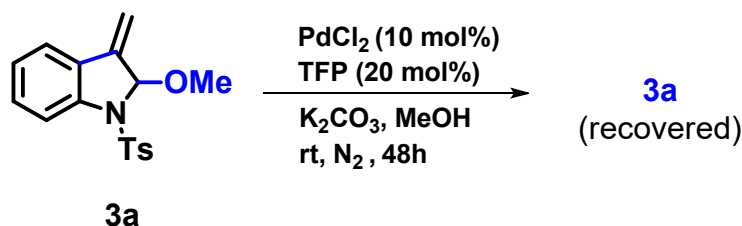


To an oven dried Schlenk tube was charged with **3a** (0.28 mmol, 1 equiv.), palladium chloride (10 mol%), triphenylphosphine (20 mol%), and Cs_2CO_3 (2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N_2 gas (3X). Then, the methanol (3 mL) was added to the reaction mixture. The reaction was stirred at rt for 48 h, and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography to give **4a** as a yellow solid (25 mg, 54% yield).



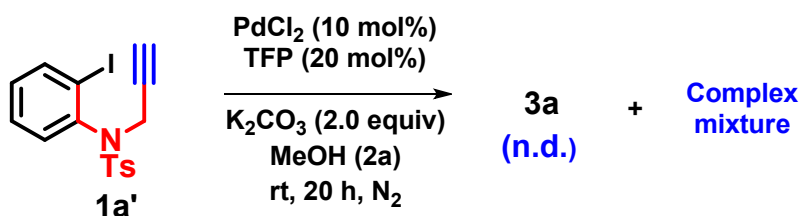
To an oven dried Schlenk tube was charged with **3a** (0.318 mmol, 1 equiv.), palladium chloride (10 mol%), triphenylphosphine (20 mol%), and Cs₂CO₃ (2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N₂ gas (3X). Then, the CD₃OD (3 mL) was added to the reaction mixture. The reaction was stirred at rt for 48 h, and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography to give **4a-D** as a yellow solid (24 mg, 46% yield).

¹H NMR (400 MHz, D₆-acetone): δ 10.14 (brs, 1H), 7.64 (d, *J* = 8.1 Hz, 1H), 7.39 (d, *J* = 8.1 Hz, 1H), 7.30 (d, *J* = 2.2 Hz, 1H), 7.13-7.09 (m, 1H), 7.05-7.01 (m, 1H), 4.61 (s, 2H).



To an oven dried Schlenk tube was charged with **3a** (0.28 mmol, 1 equiv.), palladium chloride (10 mol%), tri(2-furyl)phosphine (20 mol%), and K₂CO₃ (2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N₂ gas (3X). Methanol (**2a**) 3 mL was added to the reaction mixture. The reaction was stirred at rt for 18 h. No reaction was observed and **3a** was recovered.

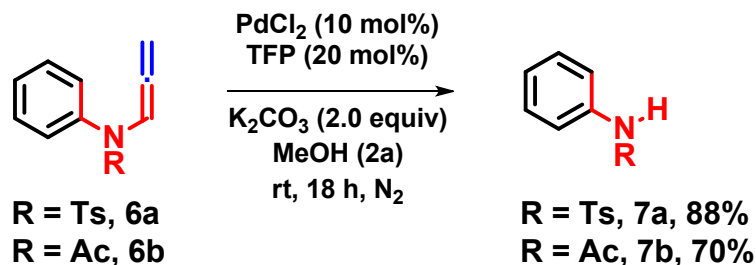
b.



To an oven dried Schlenk tube, was charged with allenamide **1a'** (1 equiv.), palladium chloride (10 mol%), tri(2-furyl)phosphine (20 mol%), and K₂CO₃ (2 equiv.), and the vial was sealed with a septum and put under vacuum, followed by flushing with N₂ gas (3X). Methanol (**2a**) 3

mL was added to the reaction mixture. The reaction was stirred at rt for 20 h. **3a** was not detected in this reaction, however, we observed a complex reaction mixture.

c.



To an oven dried Schlenk tube was charged with allenamide **6** (1 equiv.), palladium chloride (10 mol%), tri(2-furyl)phosphine (20 mol%), and K_2CO_3 (2 equiv.), the vial was sealed with a septum and put under vacuum, followed by flushing with N_2 gas (3X). Methanol (**2a**) 3 mL was added to the reaction mixture. The reaction was stirred at rt for 18 h, and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EtOAc).

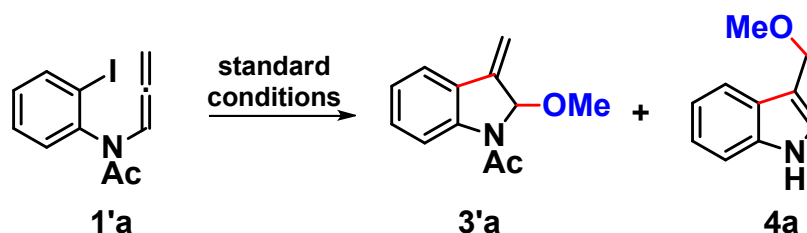
4-methyl-N-phenylbenzenesulfonamide (**7a**):

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.21 (d, $J = 7.8$ Hz, 2H), 7.24-7.20 (m, 4H), 7.11-7.04 (m, 4H), 2.36 (s, 3H).

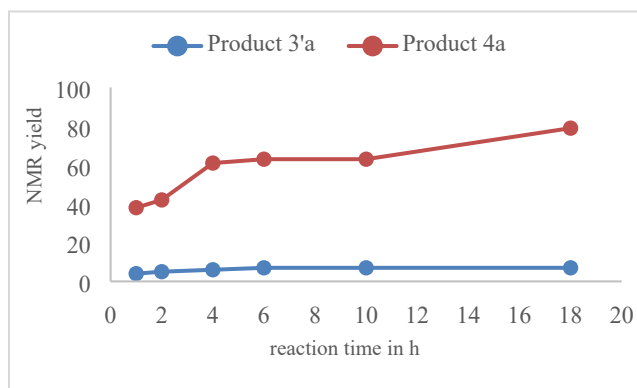
N-phenylacetamide (**7b**):

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.42 (d, $J = 7.9$ Hz, 2H), 7.23 (t, $J = 7.8$ Hz, 2H), 7.02 (t, $J = 7.4$ Hz, 1H), 2.08 (s, 3H).

d. Formation of **3'a** and **4a** versus time

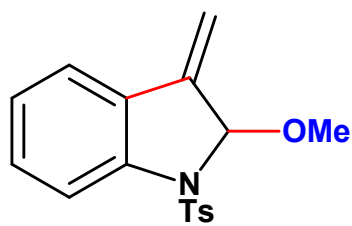


To an oven dried Schlenk tube was charged with allenamide **1'a** (0.034 mmol, 1 equiv.), palladium chloride (10 mol%), tri(2-furyl)phosphine (20 mol%), and K_2CO_3 (2 equiv.), the vial was sealed with a septum and put under vacuum, followed by flushing with N_2 gas (3X). methanol (2a) 1 mL was added to the reaction mixture. The reaction was stirred at rt. The same reaction was run for different time intervals (1-10h) and the $^1\text{H NMR}$ yield was calculated using 1,3,5-trimethoxybenzene as an internal standard.

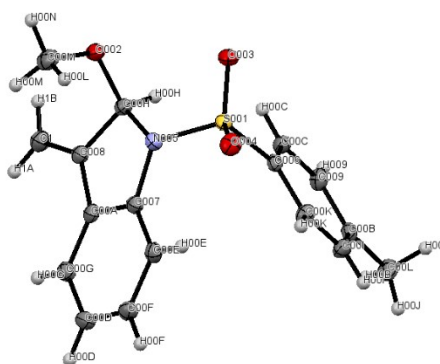


| Entry | Time | 3'a (NMR Yield %) | 4a (NMR Yield %) |
|-------|------|-------------------|------------------|
| 1. | 1h | 4 | 38 |
| 2. | 2h | 5 | 42 |
| 3. | 4h | 6 | 61 |
| 4. | 6h | 7 | 63 |
| 5. | 10h | 7 | 63 |
| 6. | 18h | 7 | 79 |

7. Crystal Structure



3a
CCDC 2168612



| | | | |
|----------------|------------------------------------|---------|---|
| Compound | 3a | | |
| CCDC | 2168612 | Formula | C ₁₇ H ₁₇ NO ₃ S |
| M _w | 315.38 | | |
| crystal system | Monoclinic | | |
| space group | <i>P</i> 2 ₁ / <i>c</i> | | |

| | | | |
|---|-----------------------|---------------------------|------------|
| T [K] | 296 | | |
| a [Å] | 9.6139(10) | α [°] | 90 |
| b [Å] | 14.7059(13) | β [°] | 100.759(3) |
| c [Å] | 10.8929(10) | γ [°] | 90 |
| Z | 4 | V [Å ³] | 1513.0(2) |
| D _{calc} [g cm ⁻³] | 1.385 | μ [mm ⁻¹] | 0.226 |
| total reflns | 3797 | unique reflns | 3789 |
| observed reflns | 3241 | | |
| $R_1[I > 2\sigma(I)]$ | 0.0362 | | |
| wR_2 [all] | 0.0927 | | |
| GOF | 0.994 | | |
| Diffractionmeter | Bruker APEX-II CCD | | |

8. References

1. Zhu, X.; Li, R.; Yao, H.; Lin, A. *Org. Lett.* **2021**, *23*, 4630–4634.
2. Wang, D-C.; Cheng, P-P.; Yang, T-T.; Wu, P-P.; Qu, G-R.; Guo, H-M. *Org. Lett.* **2021**, *23*, 7865–7872.
3. Deng, G.; Li, M.; Yu, K.; Liu, C.; Liu, Z.; Duan, S.; Chen, W.; Yang, X.; Zhang, H.; Walsh, P.J. *Angew. Chem. Int. Ed.* **2019**, *58*, 2826–2830.
4. Chen, X.; Qiu, G.; Liu, R.; Chen, D.; Chen, Z. *Org. Chem. Front.* **2020**, *7*, 890–895.
5. Guo, S.; Chen, J.; Yi, M.; Dong, L.; Lin, A.; Yao, H. *Org. Chem. Front.* **2021**, *8*, 1783–1788.
6. Grigg, R.; Sridharan, V.; Xu, L-H. *J. Chem. Soc., Chem. Commun.* **1995**, 1903-1904.
7. He, S.; Hsung, R. P.; Presser, W. R.; Ma, Z-X.; Haugen, B. J. *Org. Lett.* **2014**, *16*, 2180–2183.
8. Yang, X.; Toste, F. D. *Chem. Sci.* **2016**, *7*, 2653-2656.
9. Xie, Z.; Wu, P.; Cai, L.; Tong, X. *Tetrahedron Lett.* **2014**, *55*, 2160–2162.
10. Pi, C.; Yin, X.; Cui, X.; Ma, Y.; Wu, Y. *Org. Lett.* **2019**, *21*, 2081–2084.

11. Caiuby, C. A. D.; Ali, A.; Santana, V. T.; Lucas, F. W.; Santos, M. S.; Corrêa, A. G.; Nascimento, O. R.; Jianga, H.; Paixão, M. W. *RSC Adv.* **2018**, *8*, 12879–12886.
12. Tasker, S. Z.; Jamison, T. F. *J. Am. Chem. Soc.* **2015**, *137*, 9531–9534.
13. Garkhedkar, A. M.; Gore, B. S.; Hu, W-P.; Wang, J-J. *Org. Lett.* **2020**, *22*, 3531–3536.
14. Deposition number 2168612 contain the supplementary crystallographic data for this paper. These data are provided free of charge by the joint Cambridge Crystallographic Data Centre.

9.NMR Spectra

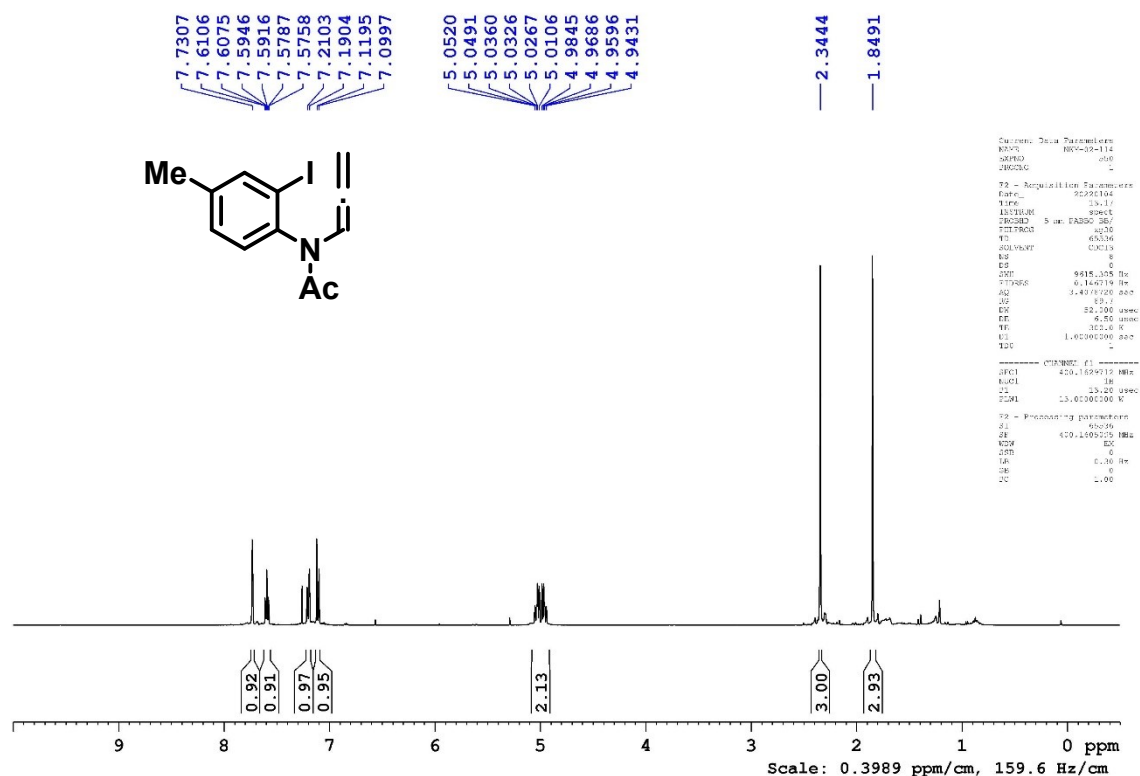


Figure S-1: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **1'b**

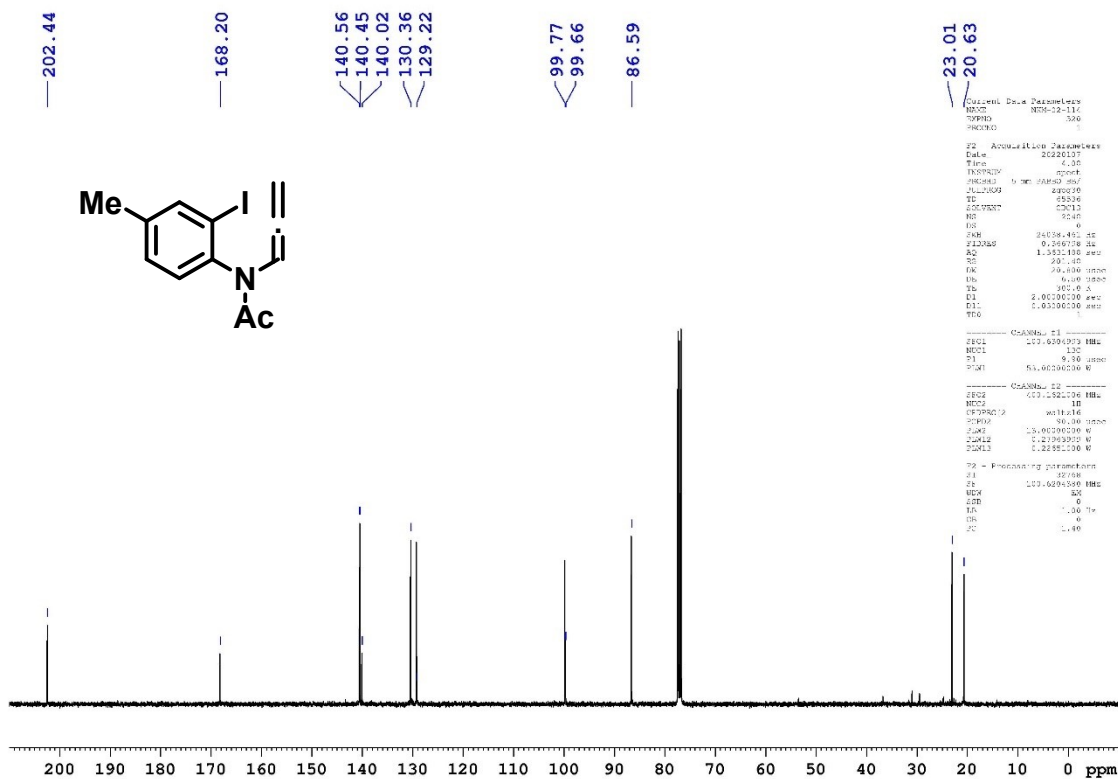


Figure S-2: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 1'b

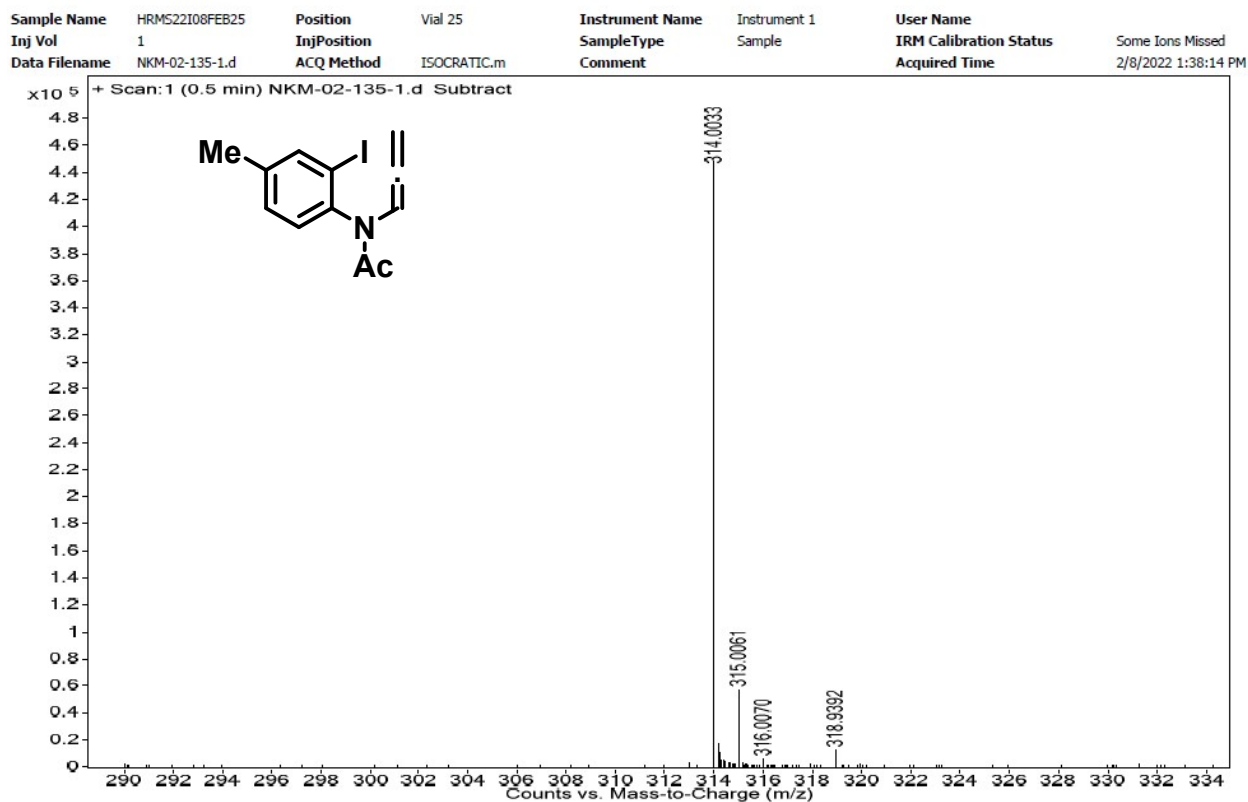


Figure S-3: HRMS spectrum of compound 1'b

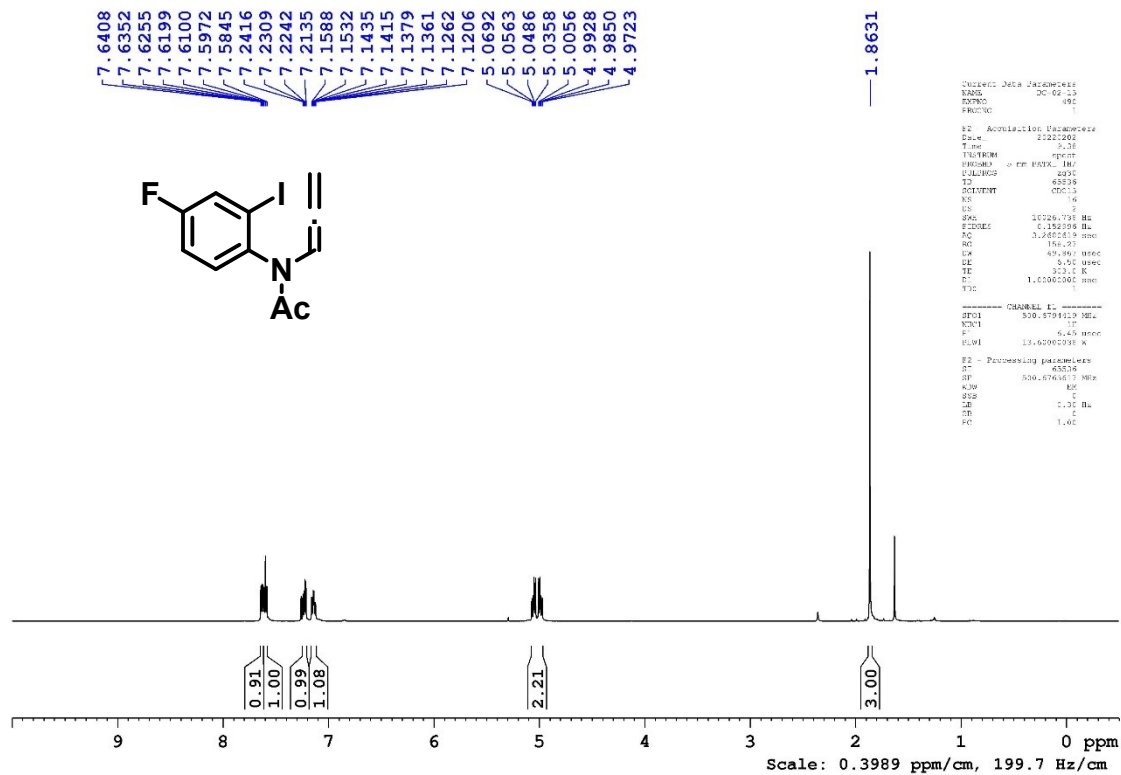


Figure S-4: ¹H NMR (500 MHz, CDCl₃) spectrum of compound 1c

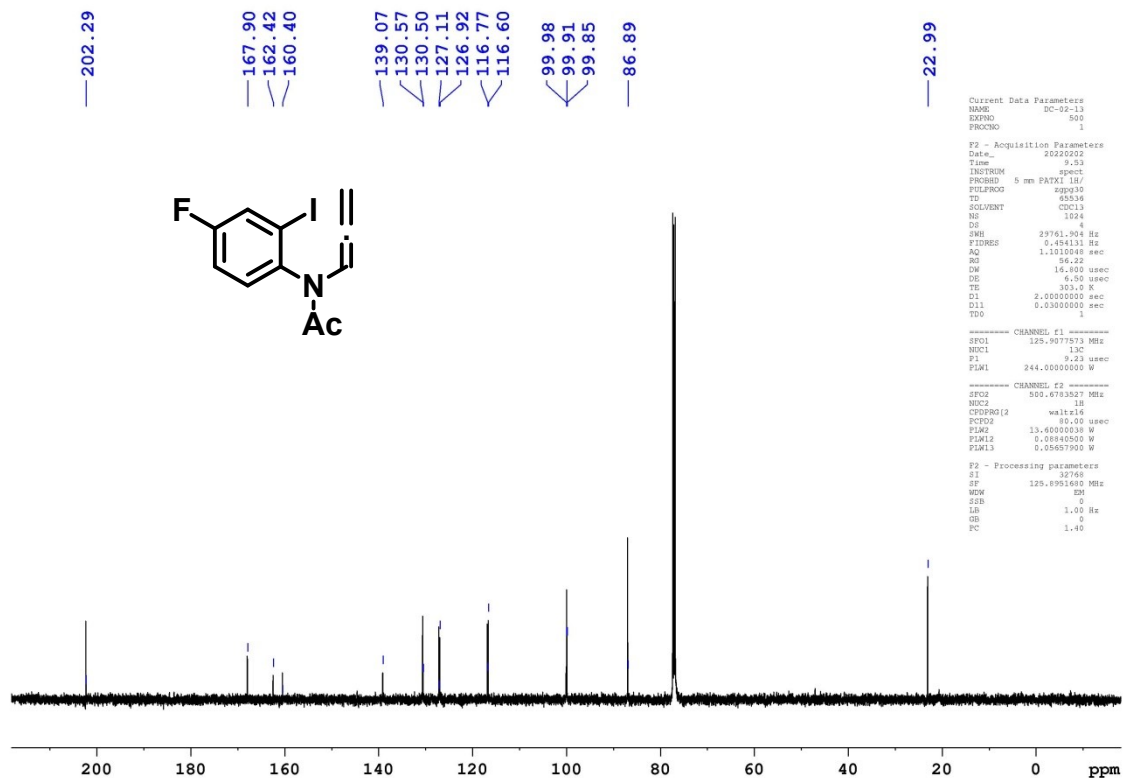


Figure S-5: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 1'c

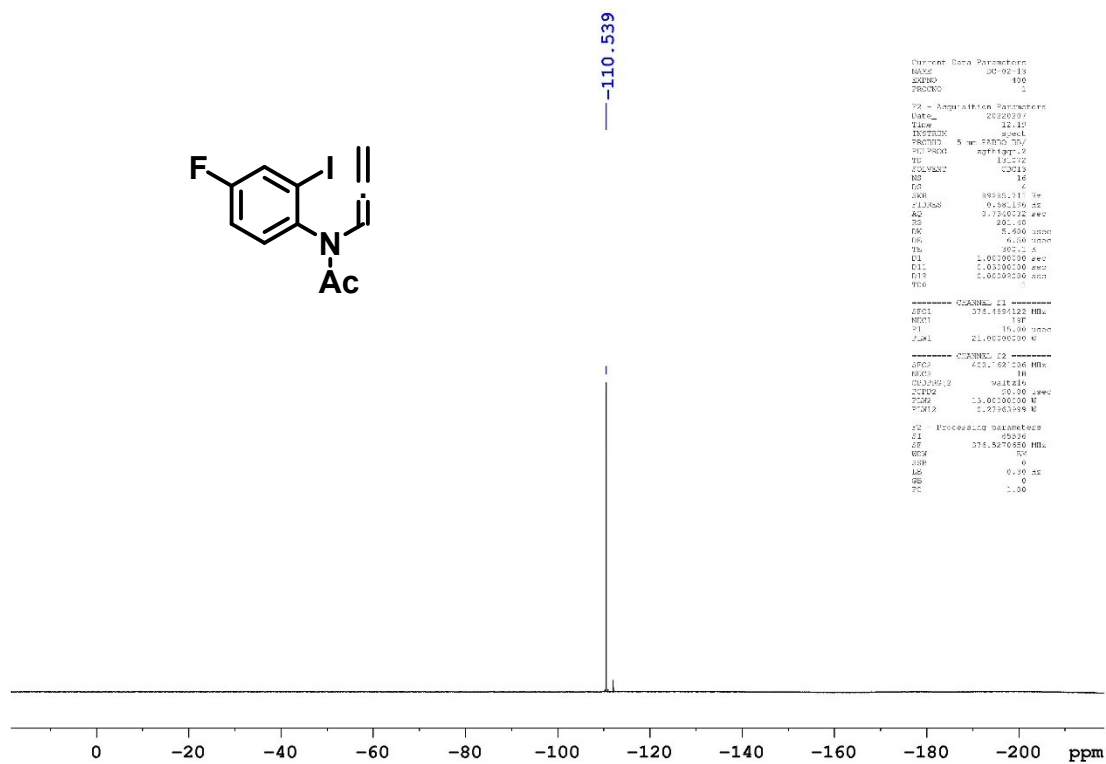


Figure S-6: ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound 1'c

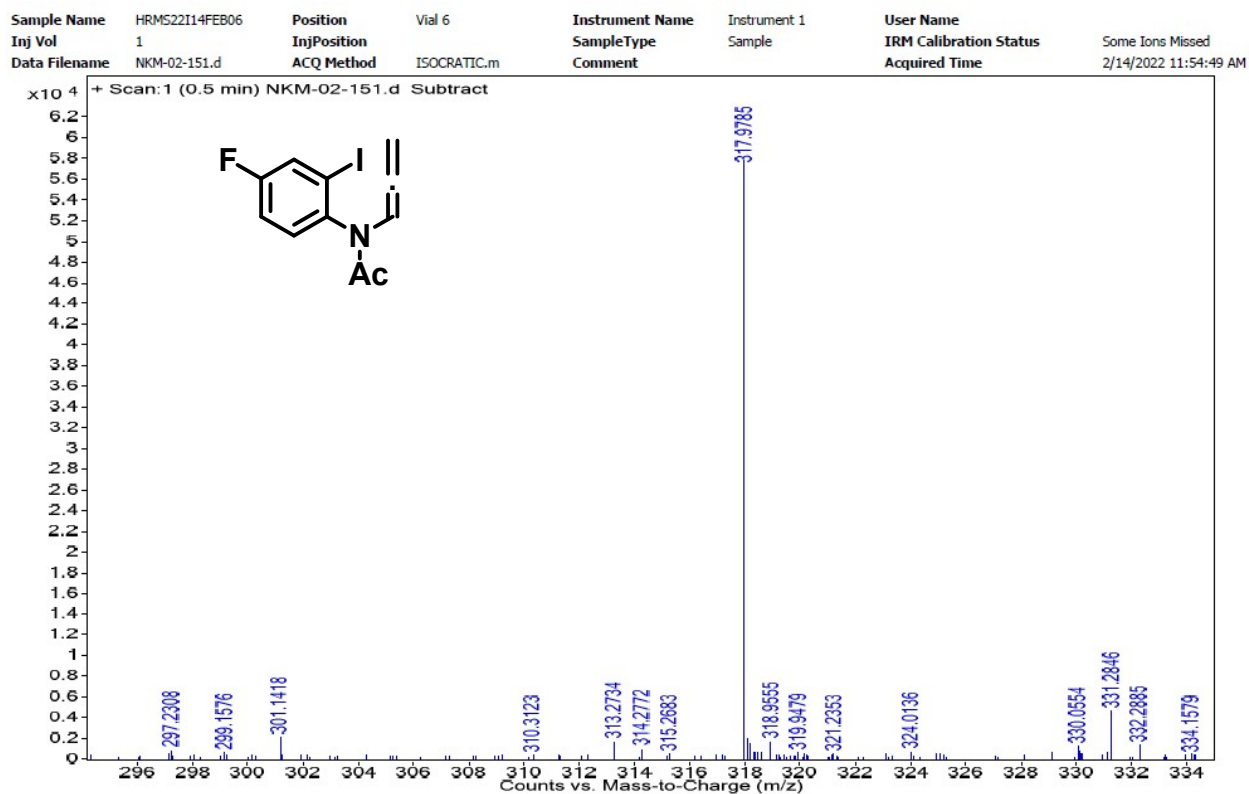


Figure S-7: HRMS spectrum of compound 1'c

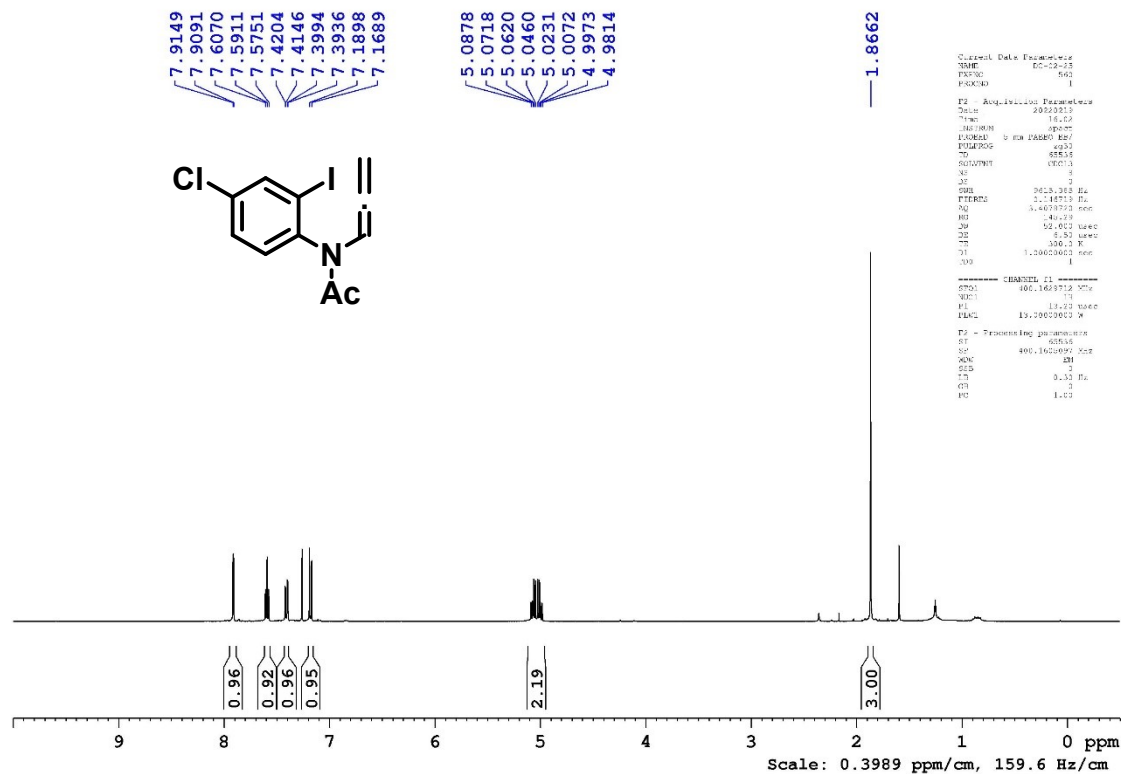


Figure S-8: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **1'd**

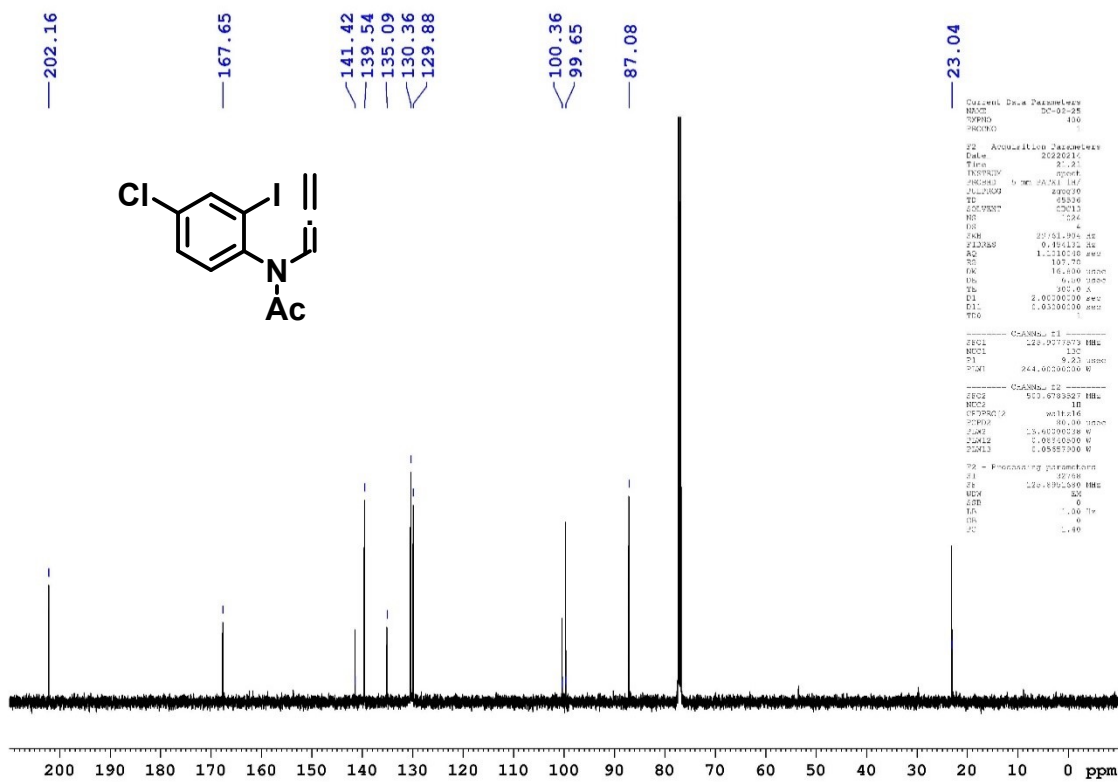


Figure S-9: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound **1'd**

| Sample Name | HRM522I15FEB23 | Position | Vial 23 | Instrument Name | Instrument 1 | User Name | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | DC-01-296.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/15/2022 12:26:29 PM |

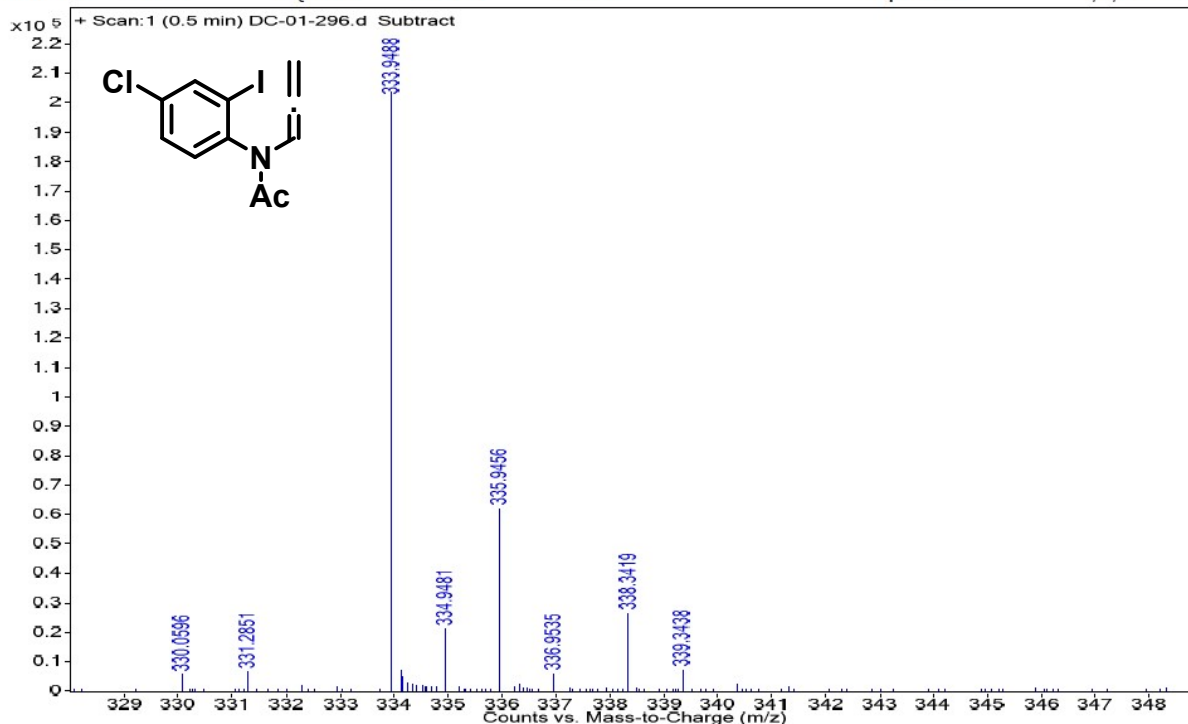


Figure S-10: HRMS spectrum of compound 1'd

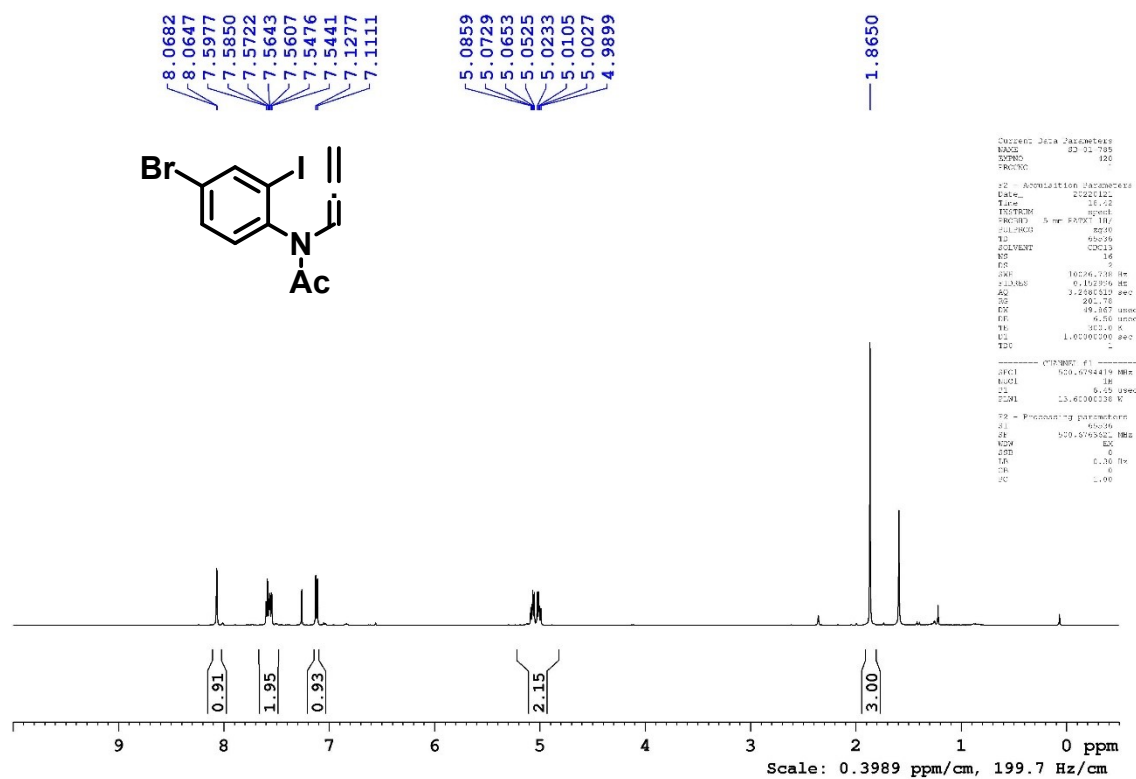


Figure S-11: ¹H NMR (500 MHz, CDCl₃) spectrum of compound 1'e

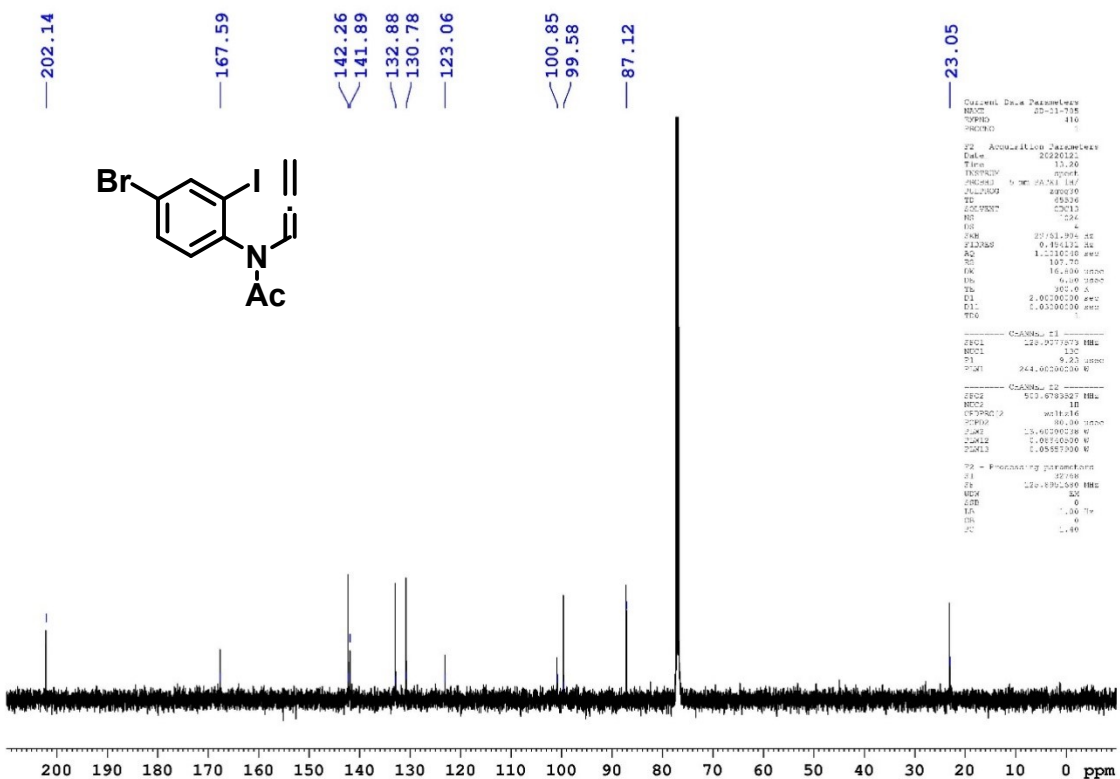


Figure S-12: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 1'e

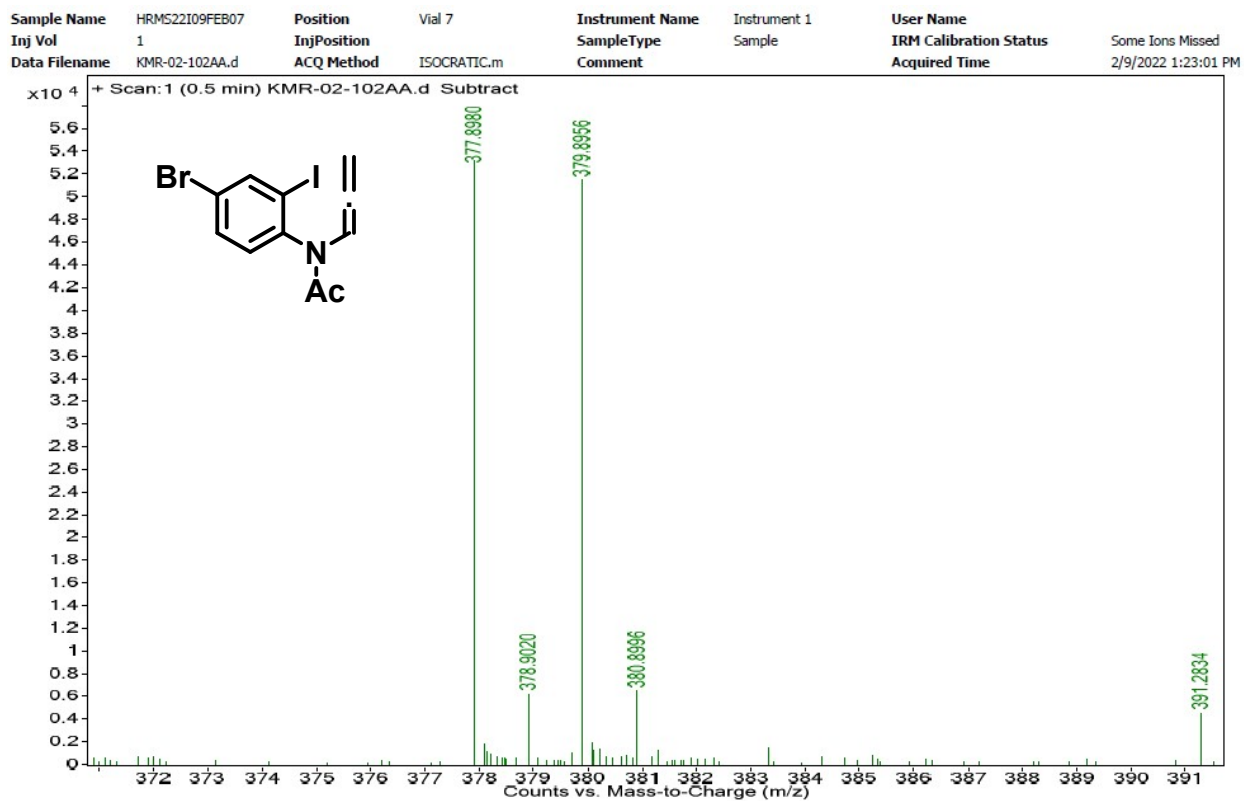


Figure S-13: HRMS spectrum of compound 1'e

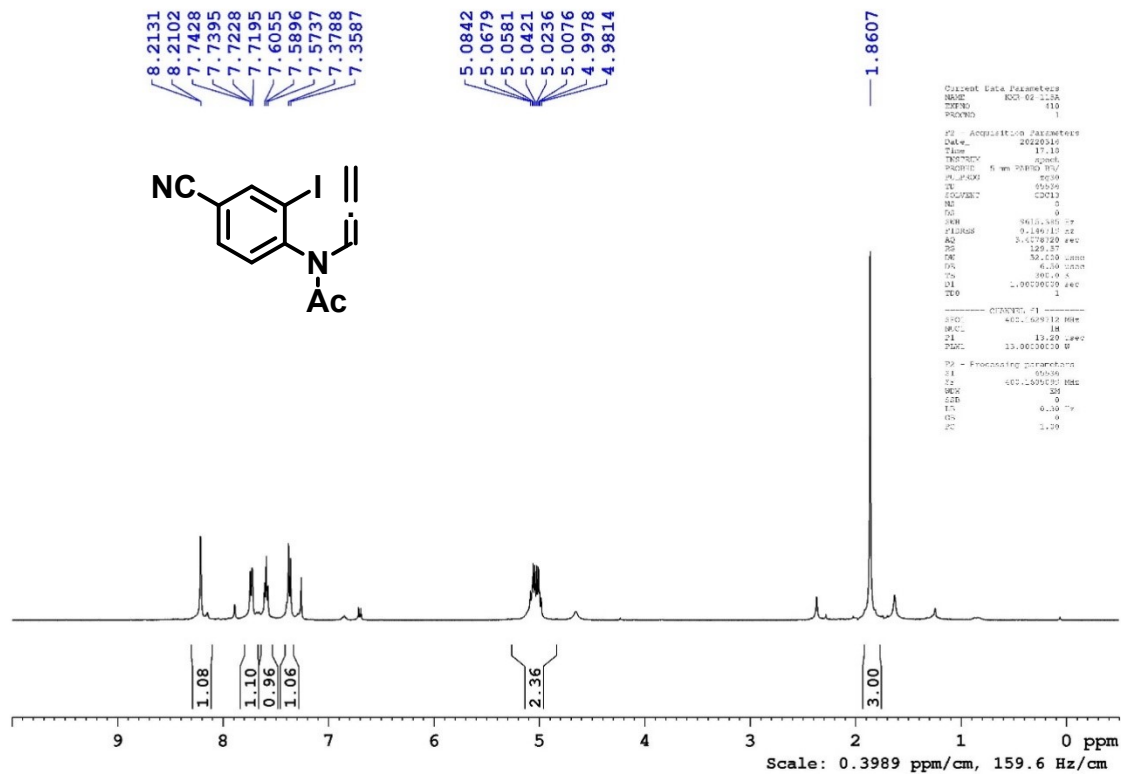


Figure S-14: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 1'f

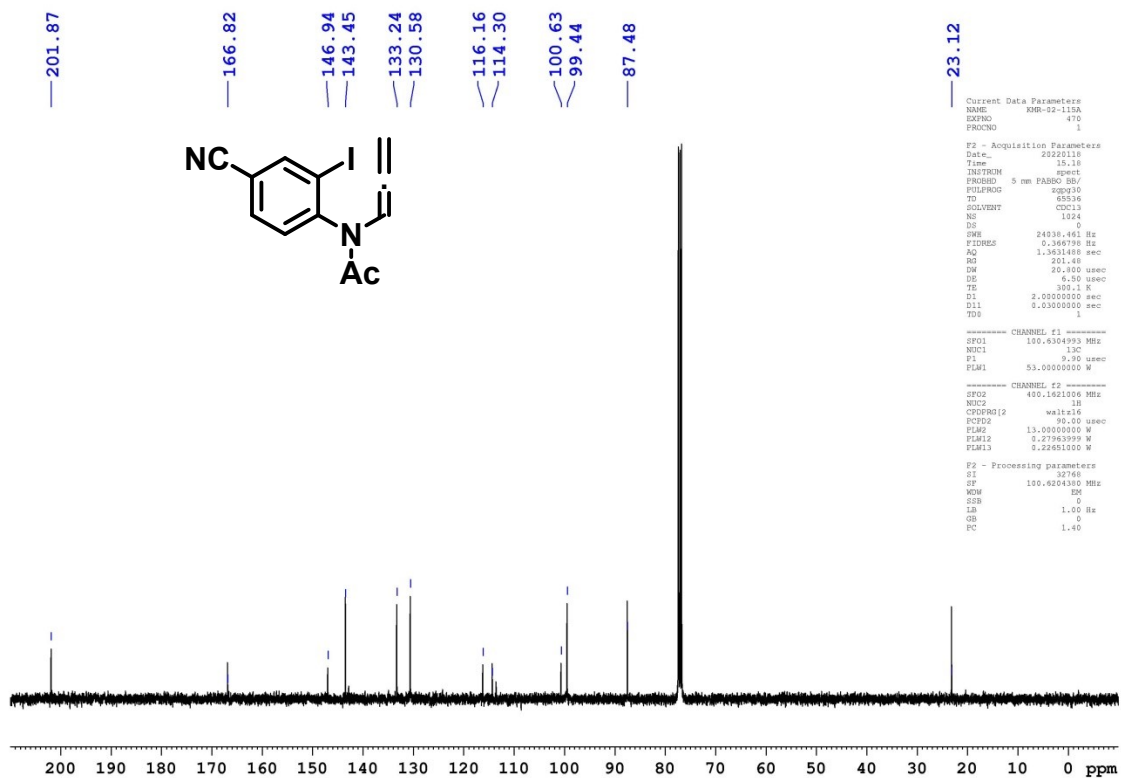


Figure S-15: ^{13}C NMR (100 MHz, CDCl_3) spectrum of compound 1'f

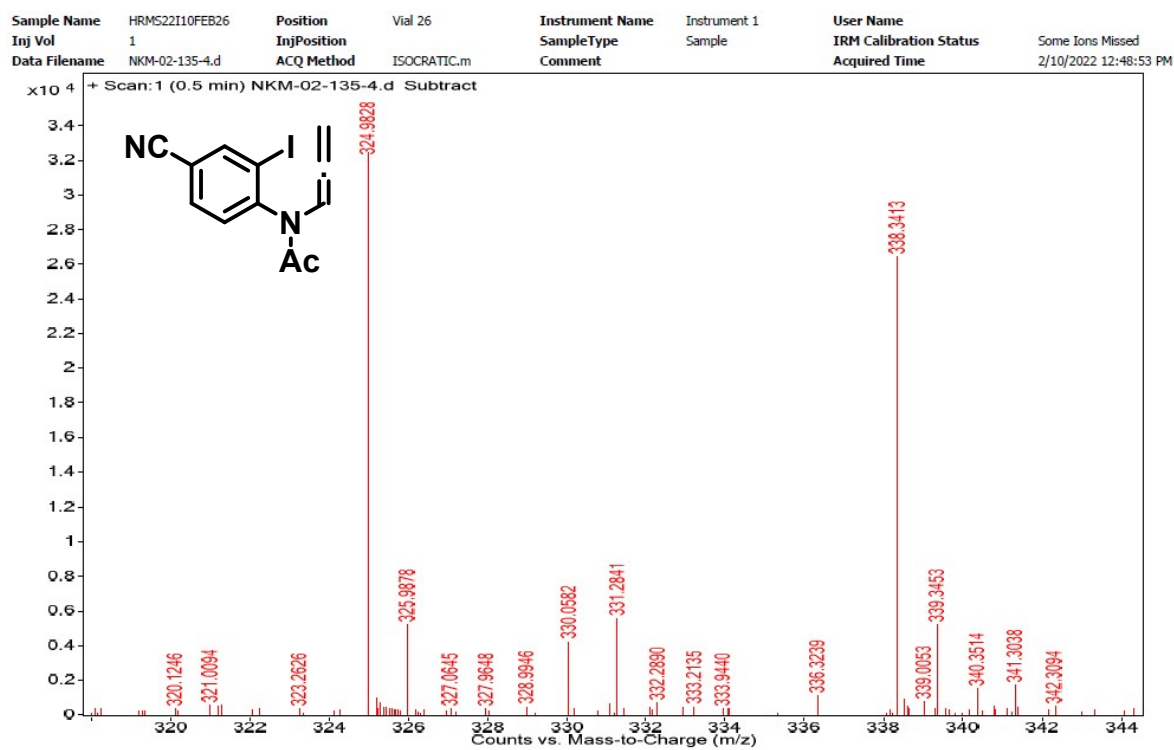


Figure S-16: HRMS spectrum of compound 1'f

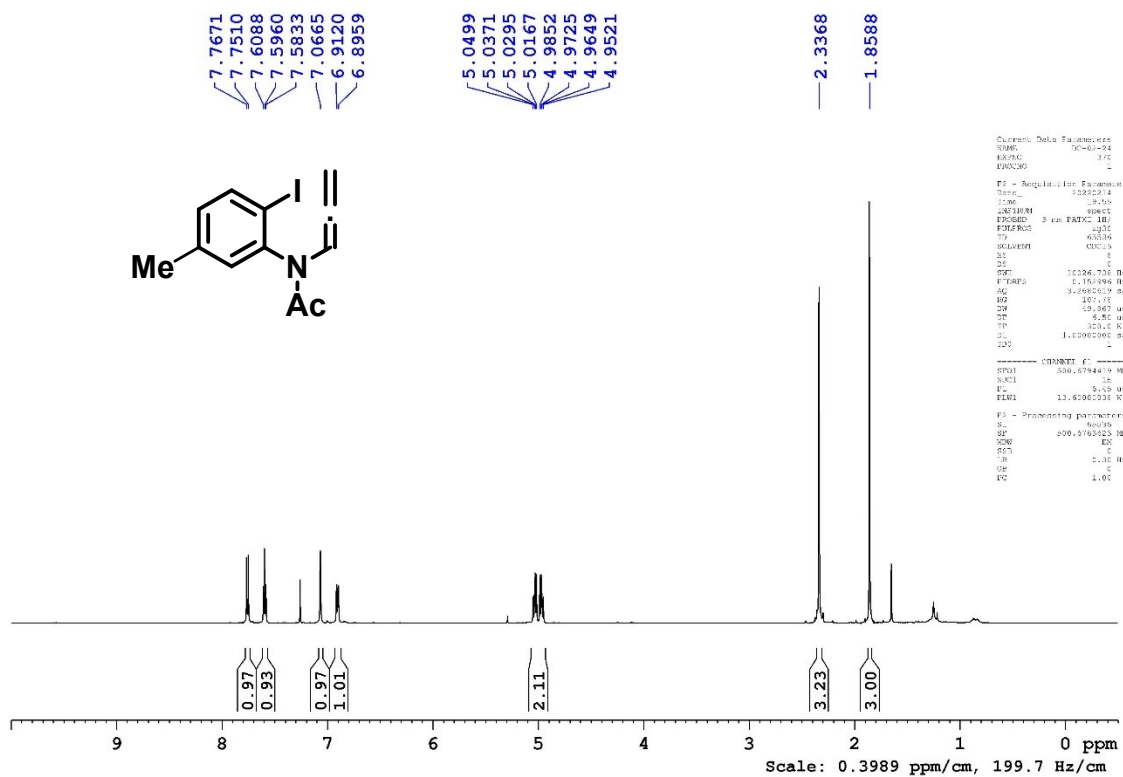


Figure S-17: ^1H NMR (500 MHz, CDCl_3) spectrum of compound 1'g

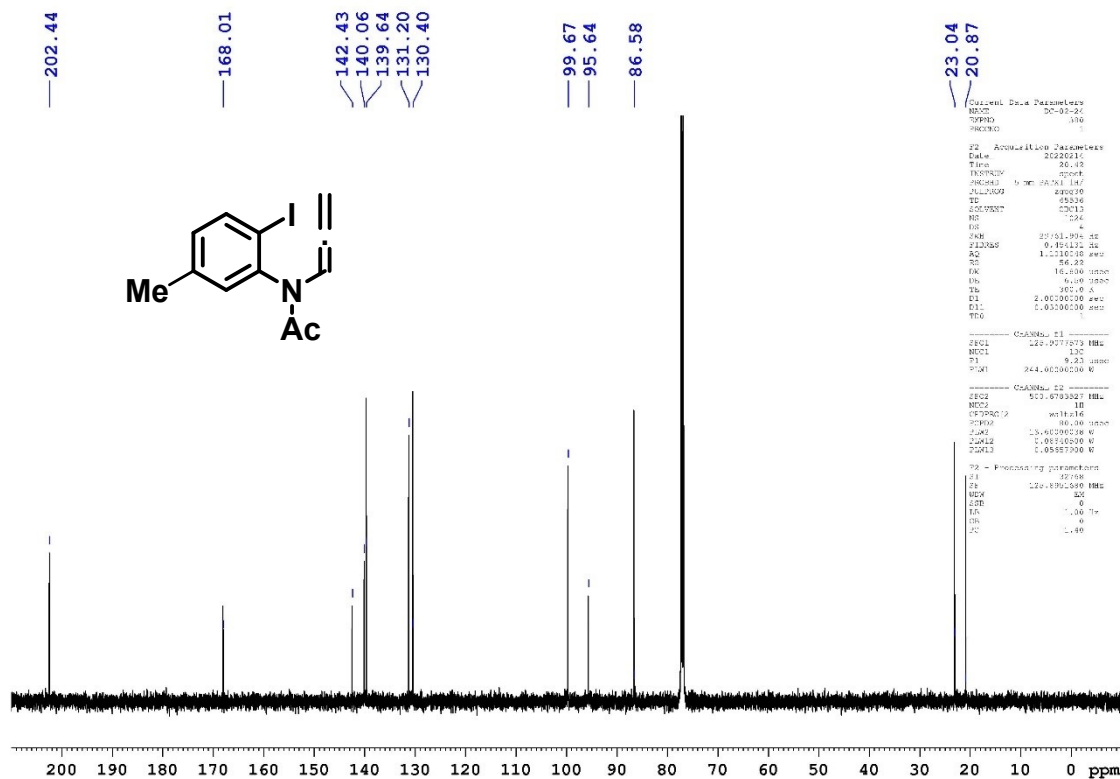


Figure S-18: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 1'g

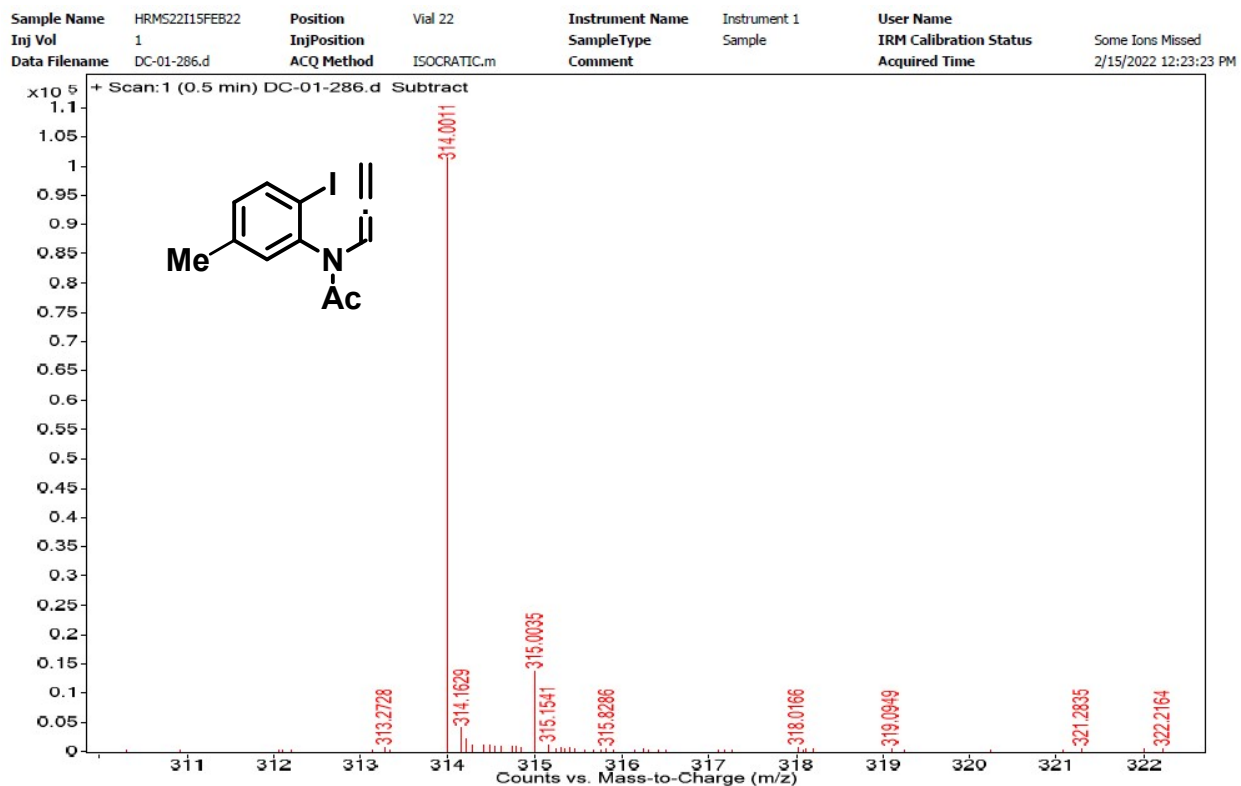


Figure S-19: HRMS spectrum of compound 1'g

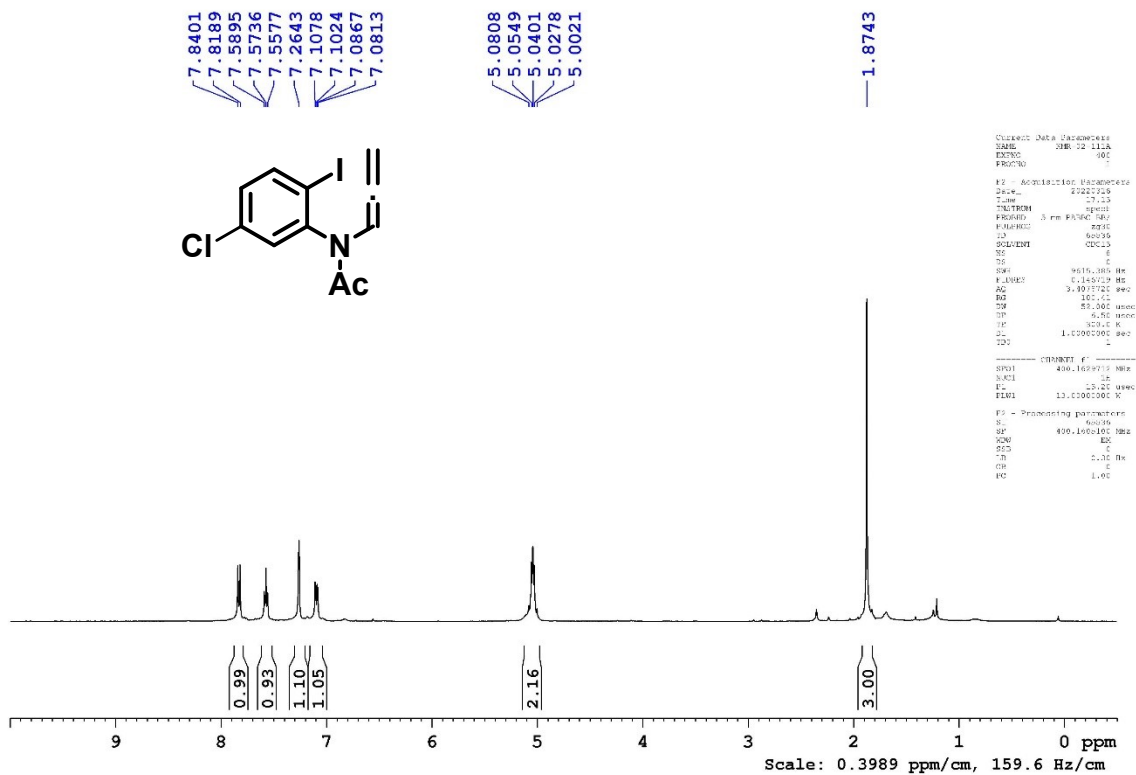


Figure S-20: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **1'h**

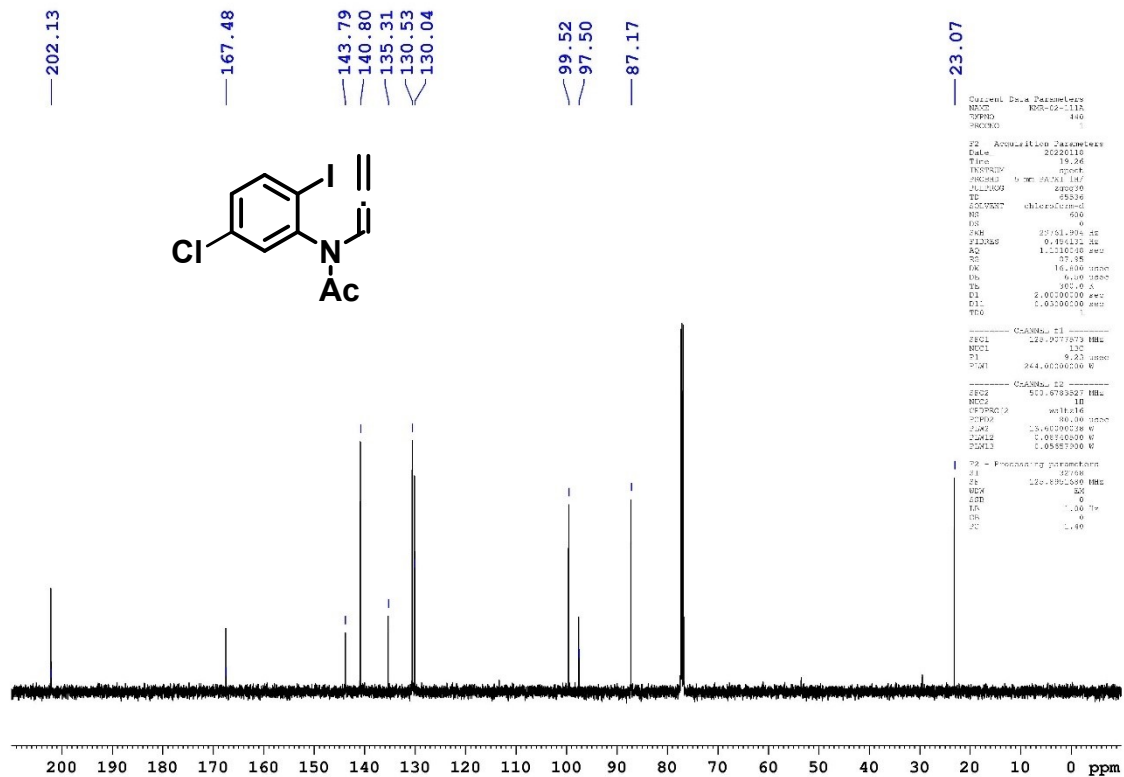


Figure S-21: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound **1'h**

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Sample Name | HRMS22I10FEB25 | Position | Vial 25 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | NKM-02-135-3.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/10/2022 12:45:42 PM |

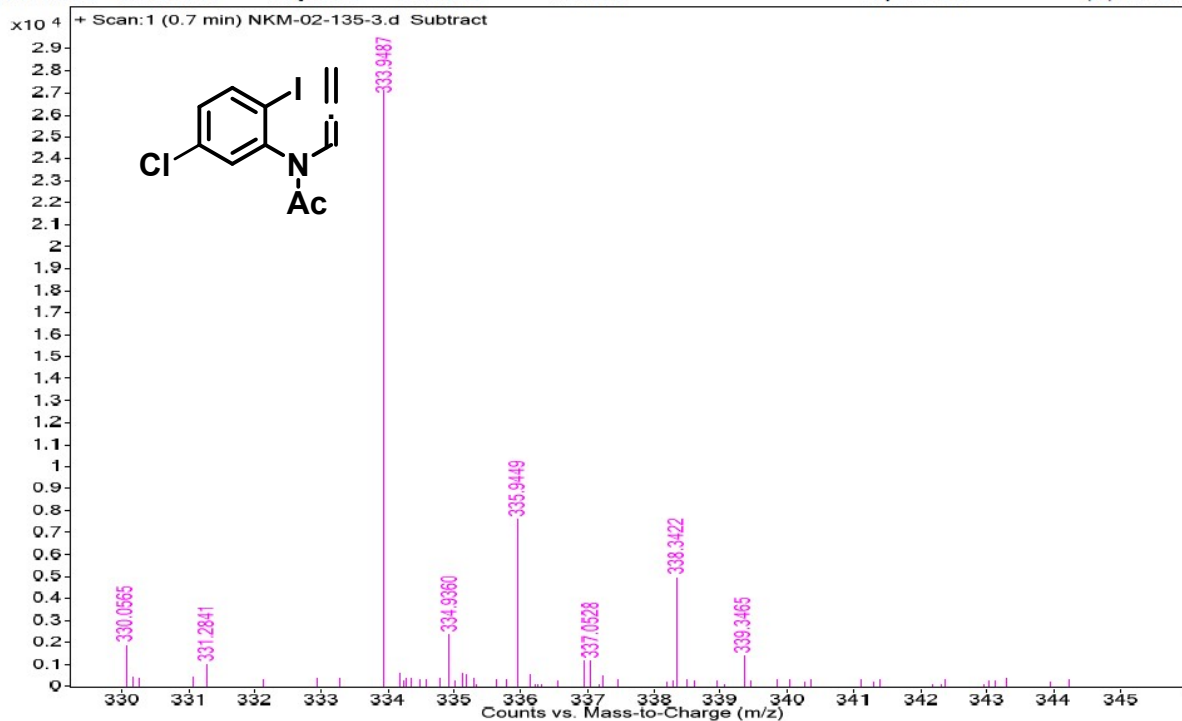


Figure S-22: HRMS spectrum of compound 1'h

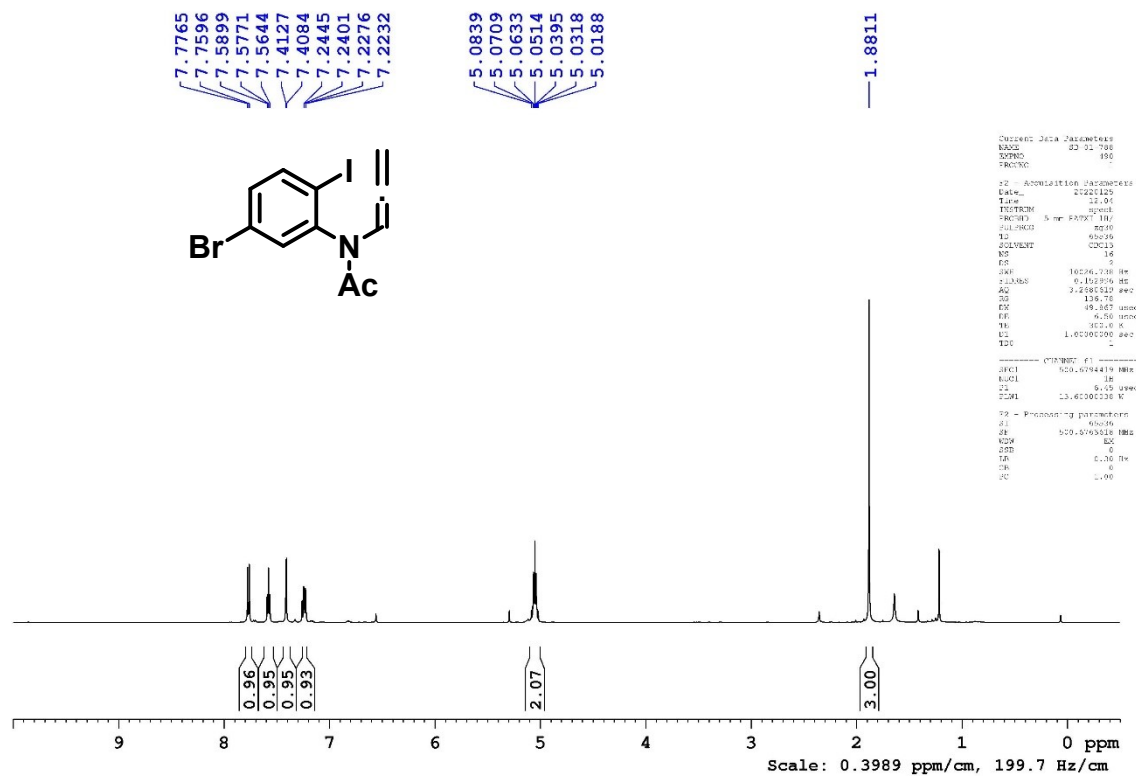


Figure S-23: ^1H NMR (500 MHz, CDCl_3) spectrum of compound 1'i

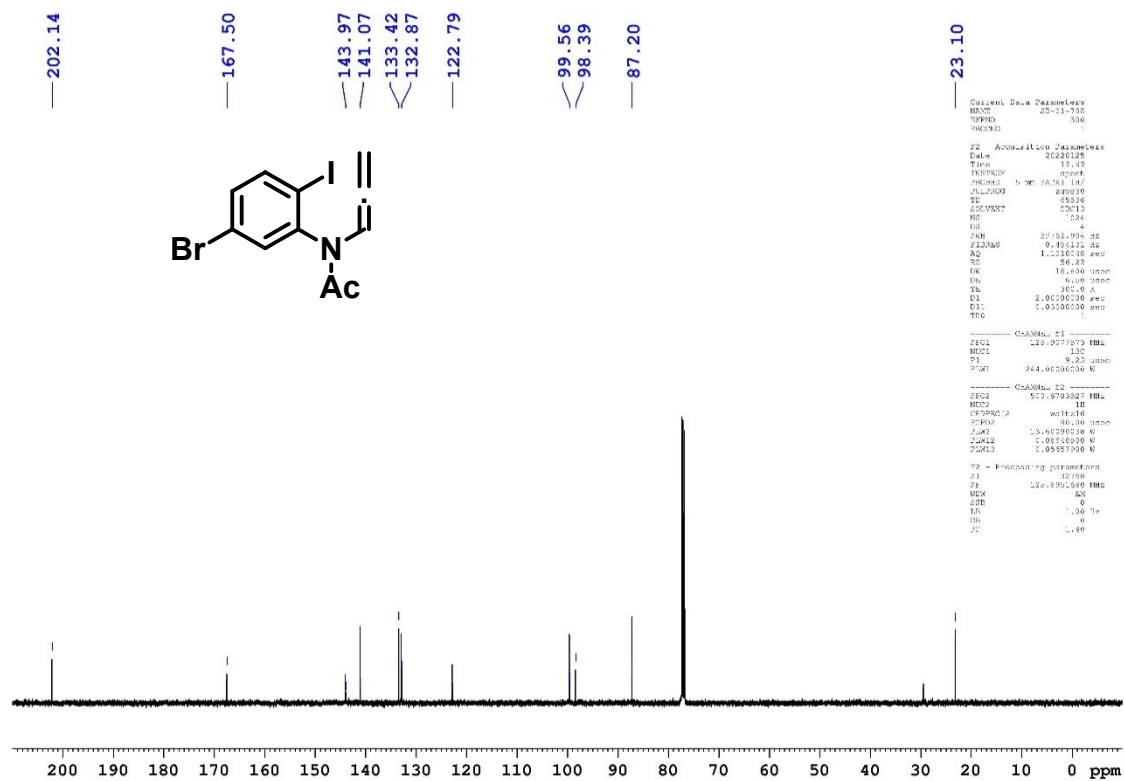


Figure S-24: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 1'i

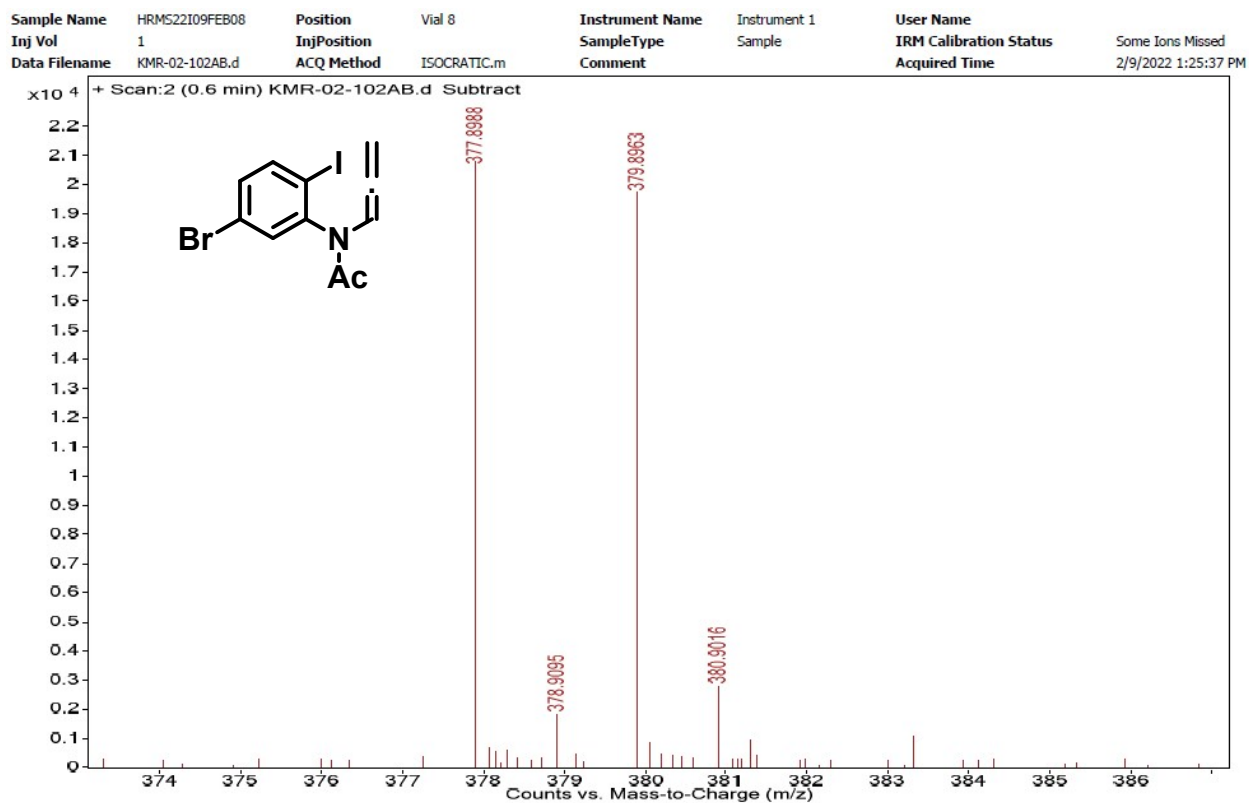


Figure S-25: HRMS spectrum of compound 1'i

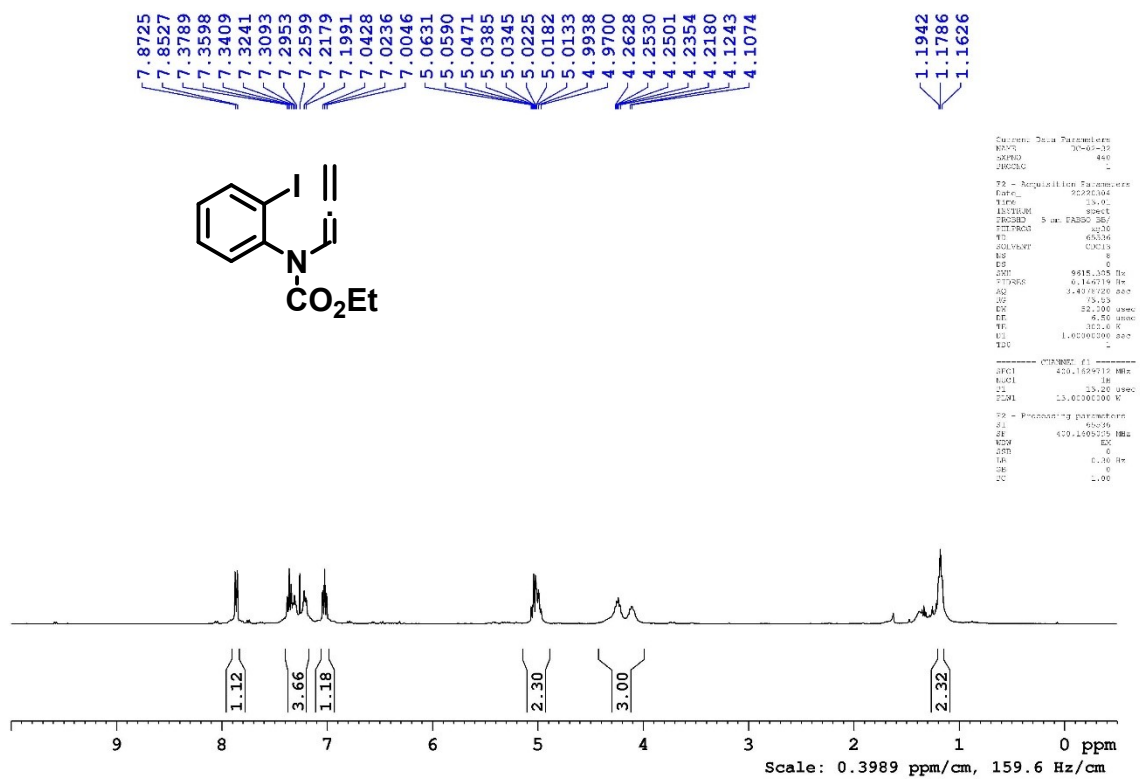


Figure S-26: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **10**

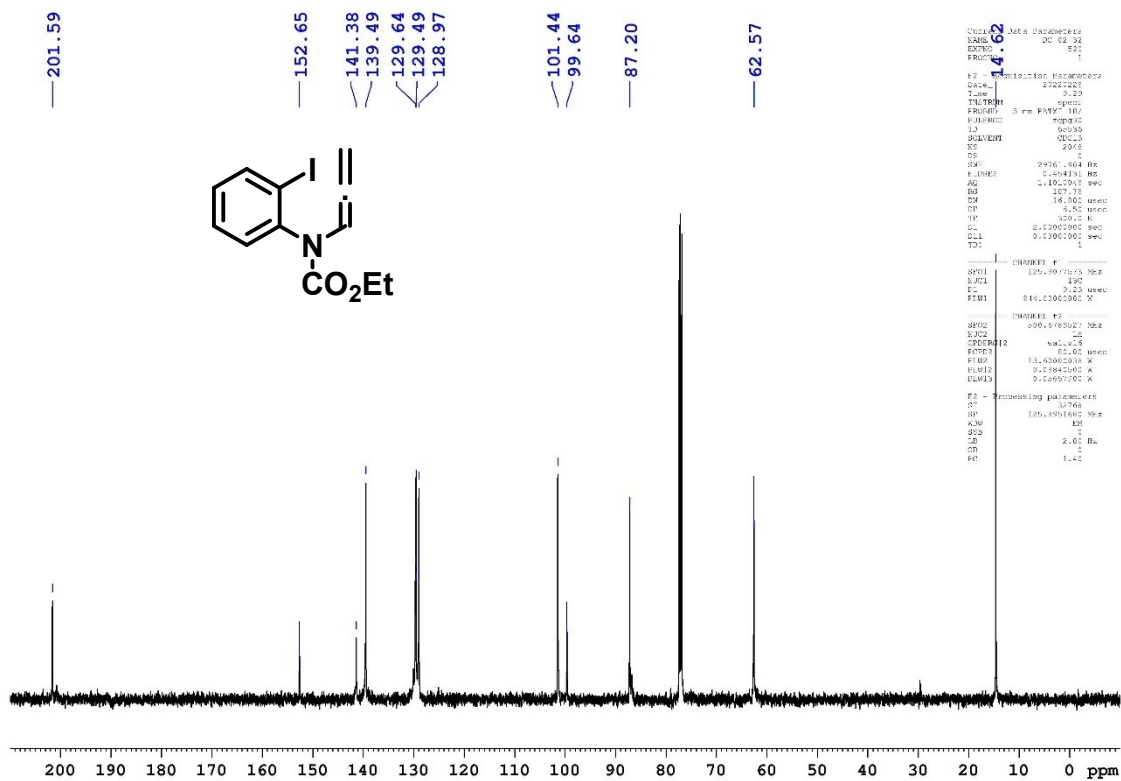


Figure S-27: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound **10**

| | | | | | | | |
|---------------|------------------|-------------|-------------|-----------------|--------------|------------------------|---------------------|
| Sample Name | HRMS22I02MARCH18 | Position | Vial 18 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | SD-01-822.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 3/2/2022 1:30:04 PM |

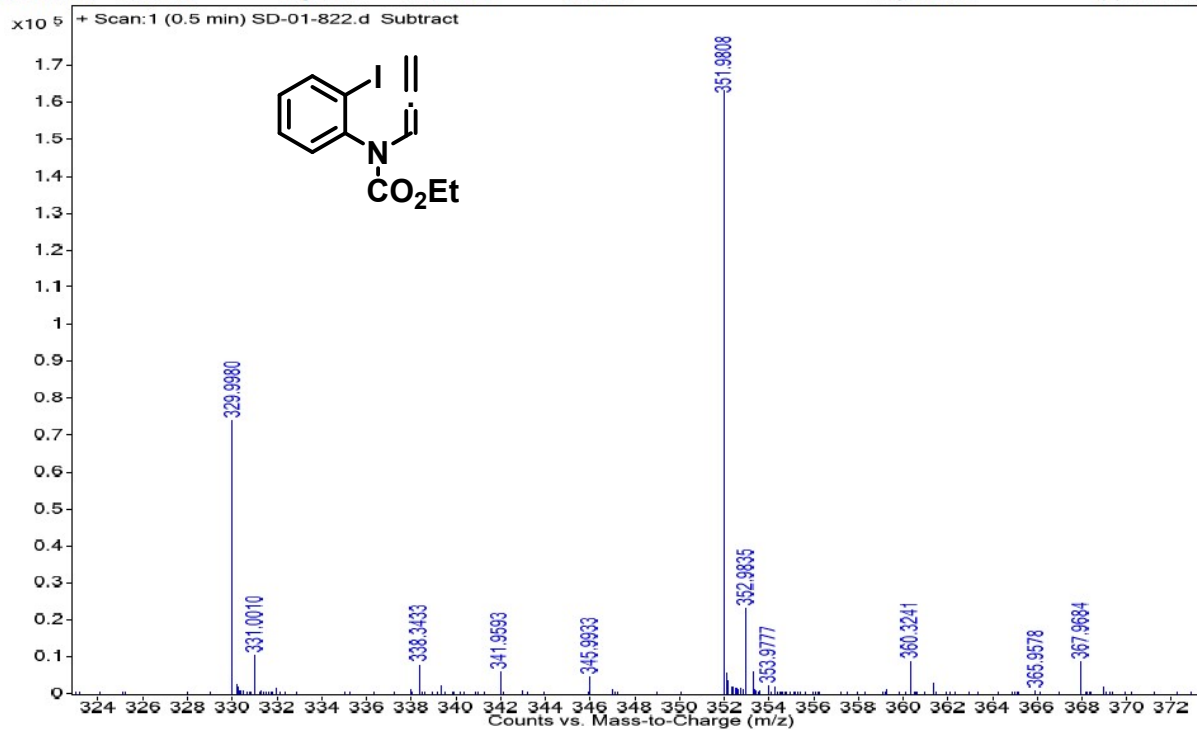


Figure S-28: HRMS spectrum of compound 1o

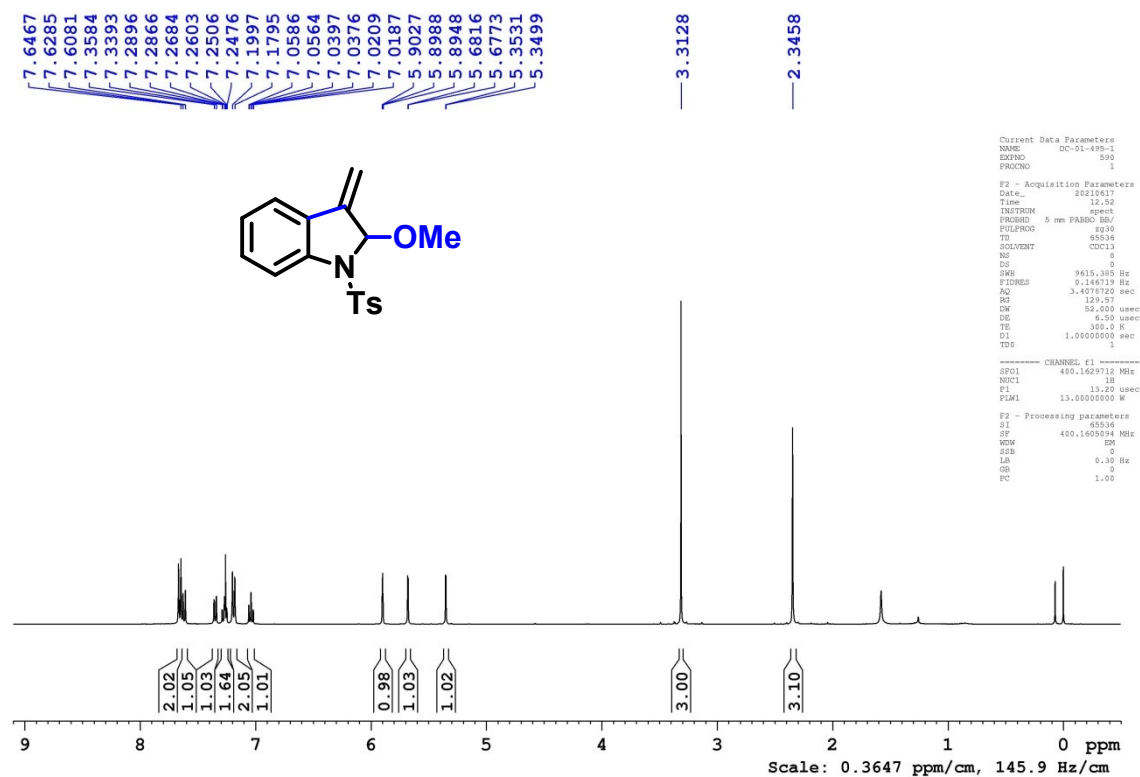


Figure S-29: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3a

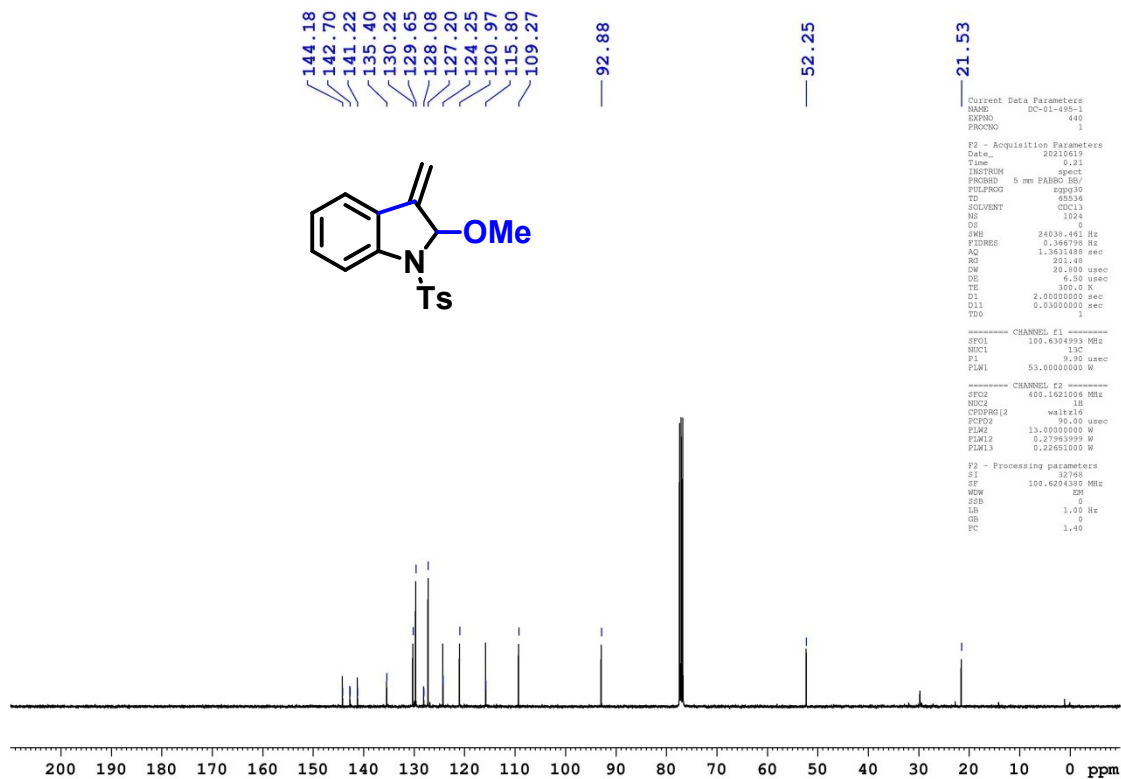


Figure S-30: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3a

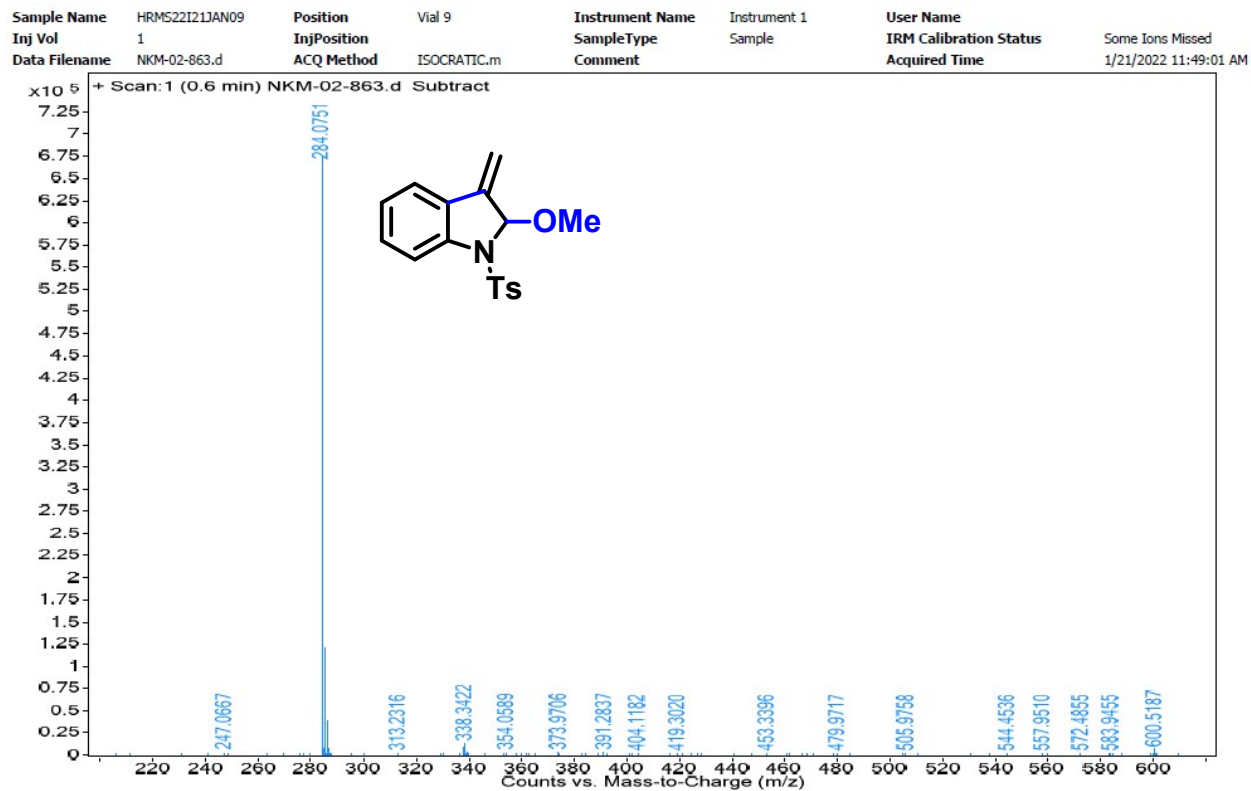


Figure S-31: HRMS spectrum of compound 3a

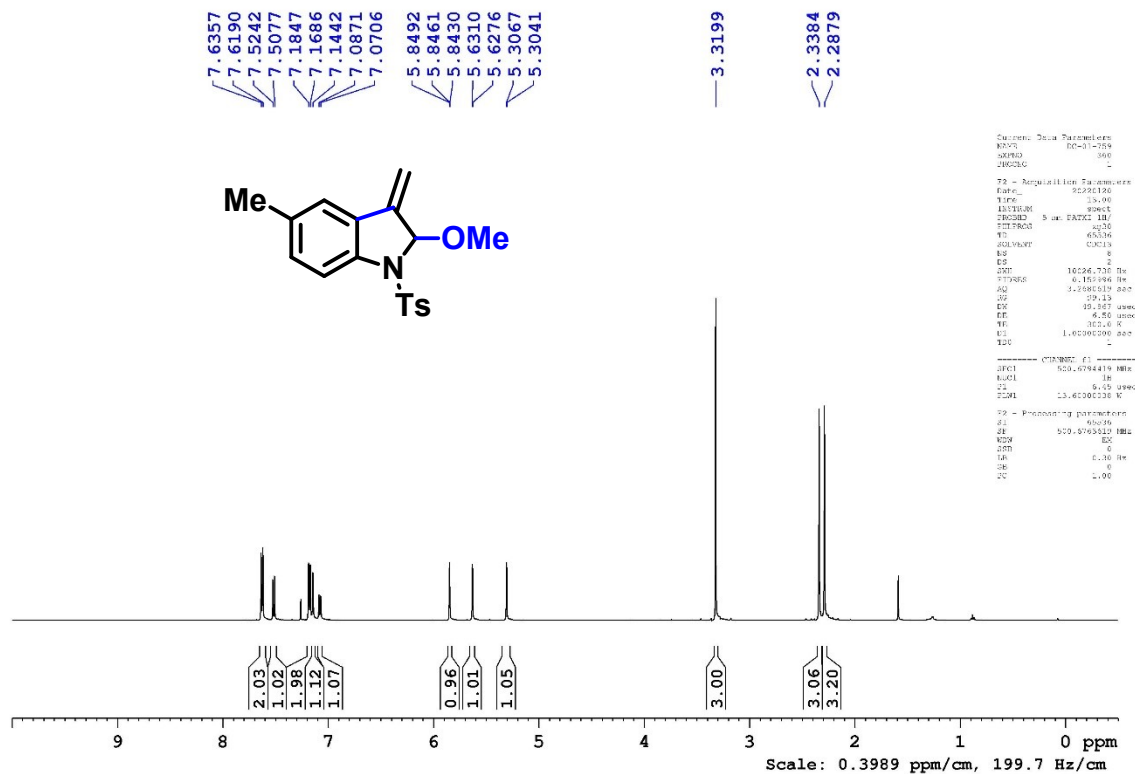


Figure S-32: ¹H NMR (500 MHz, CDCl₃) spectrum of compound **3b**

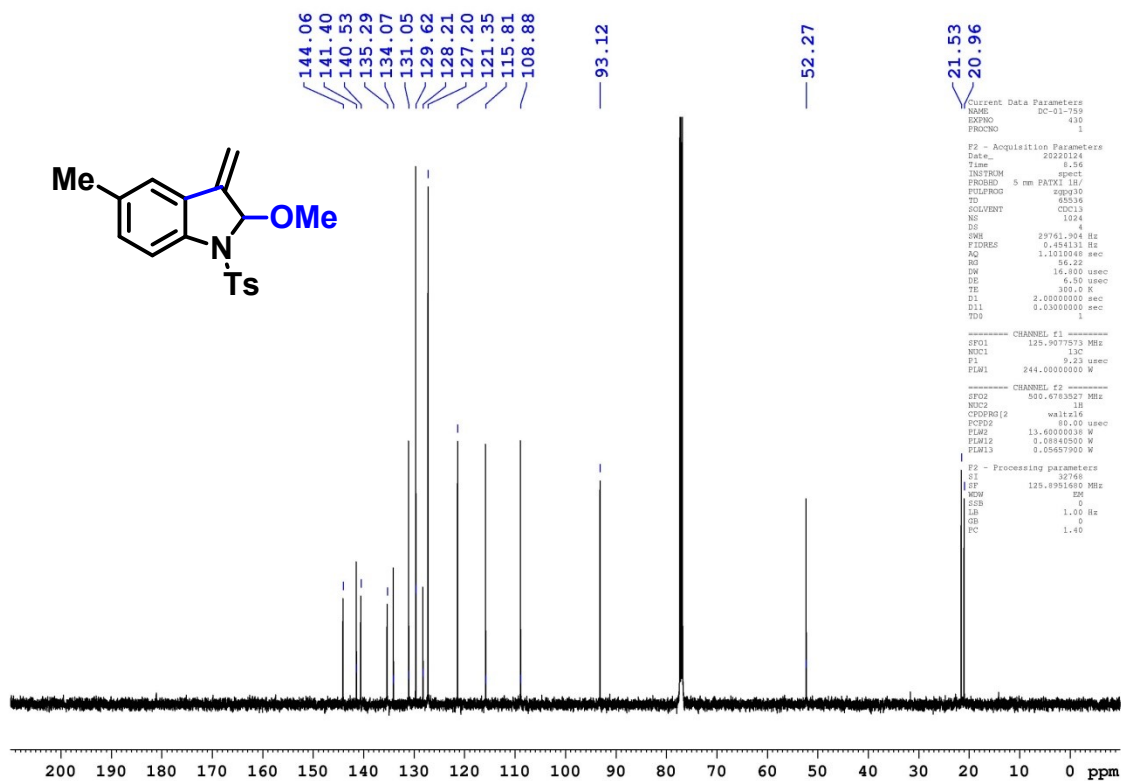


Figure S-33: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound **3b**

| Sample Name | HRMS22124JAN22 | Position | Vial 22 | Instrument Name | Instrument 1 | User Name | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | KM-CY-P2.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 1/24/2022 12:53:07 PM |

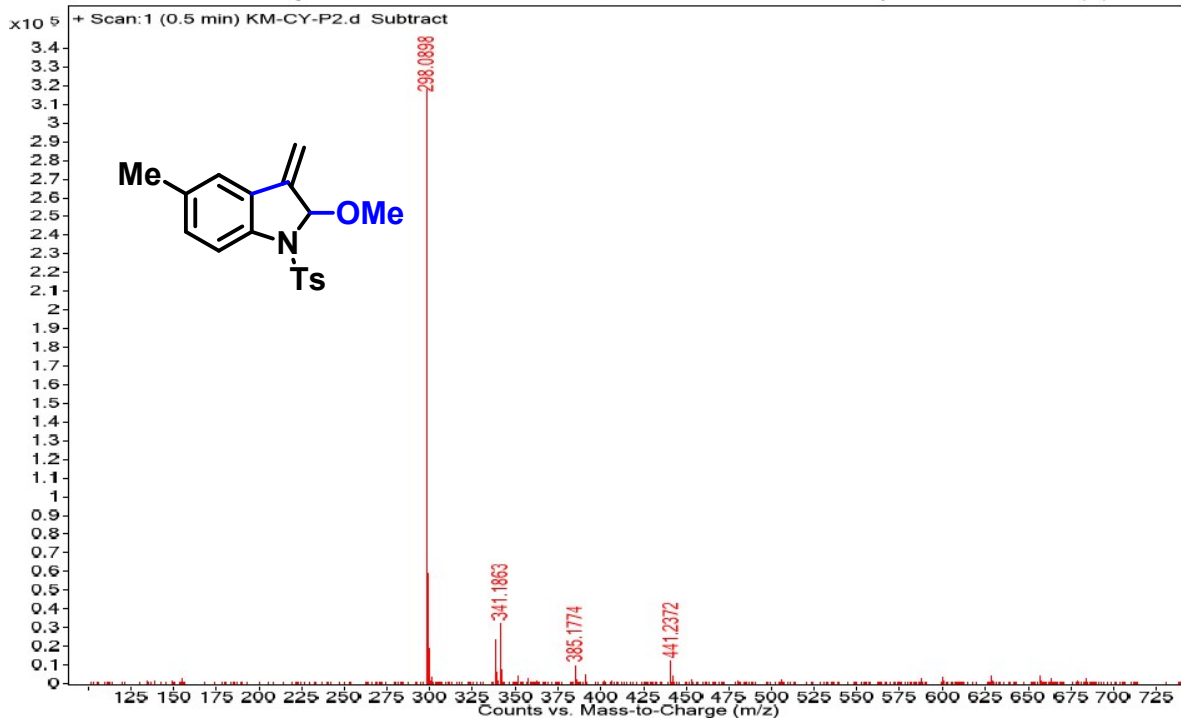


Figure S-34: HRMS spectrum of compound 3b

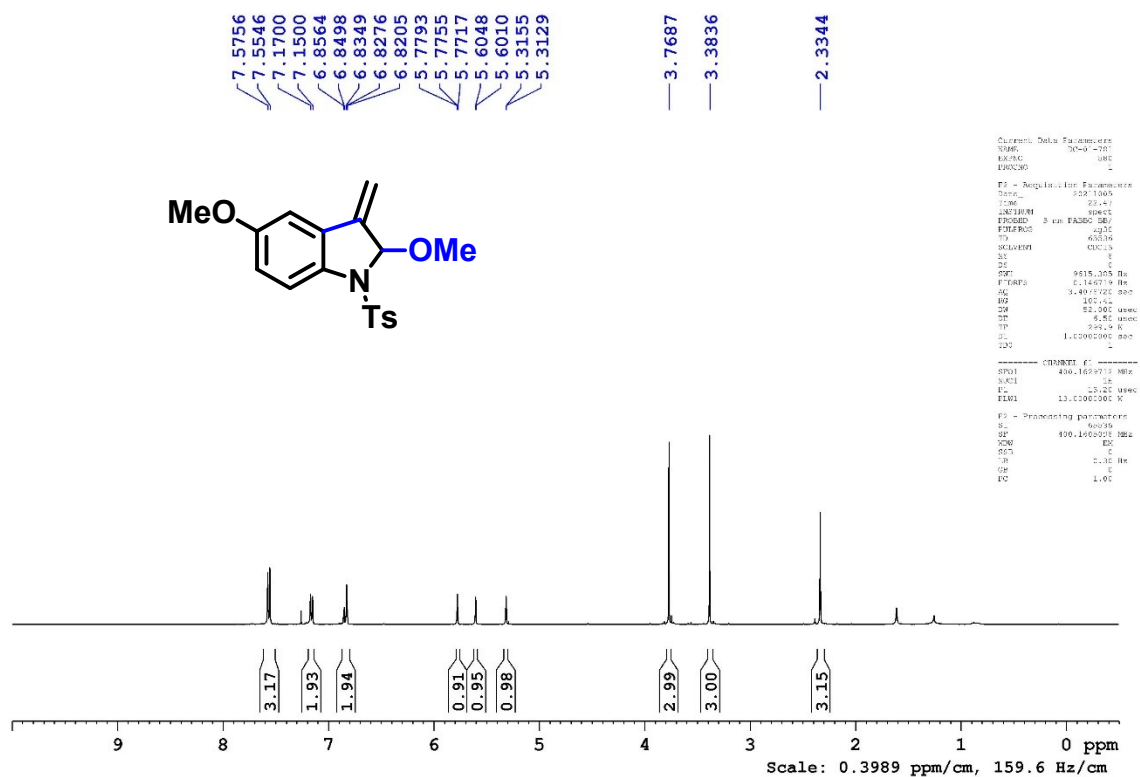


Figure S-35: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3c

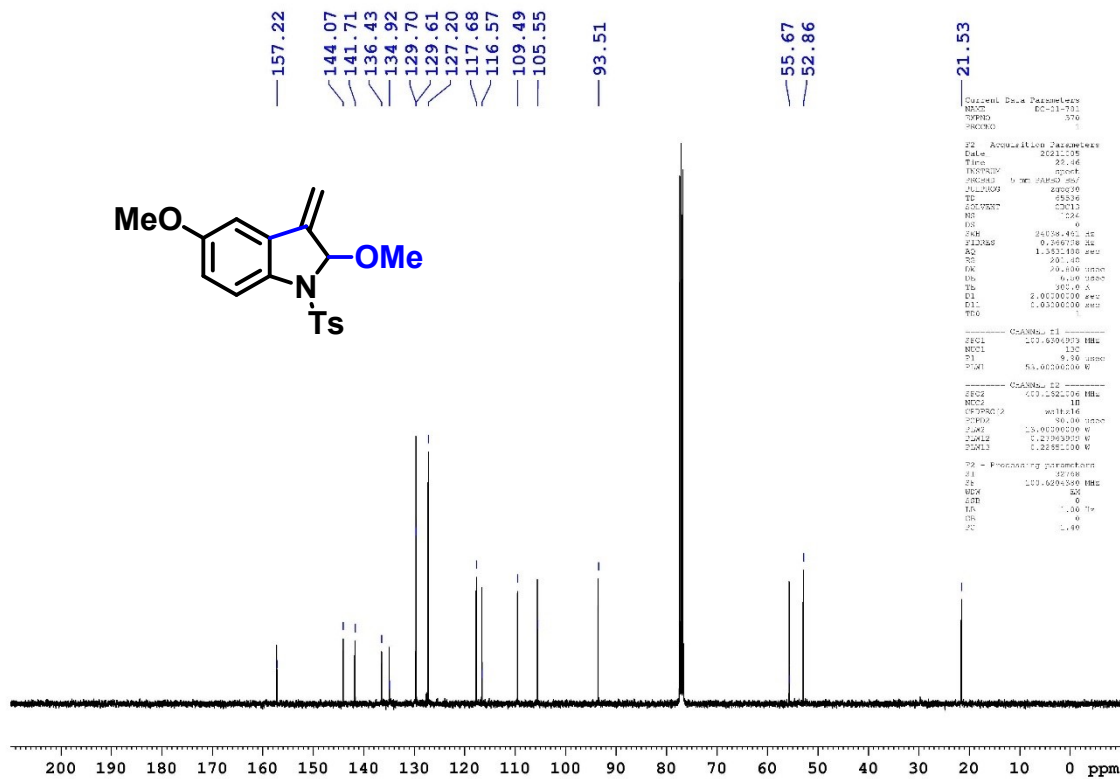


Figure S-36: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3c

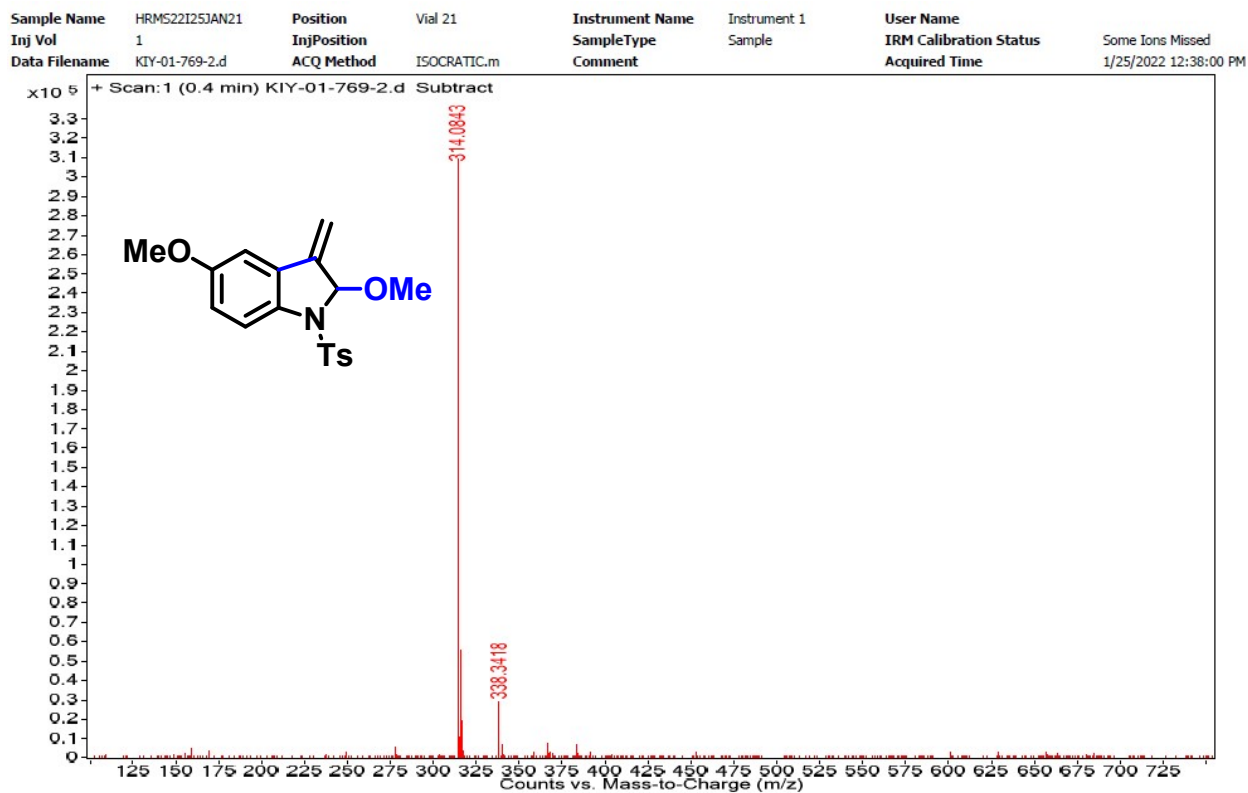


Figure S-37: HRMS spectrum of compound 3c

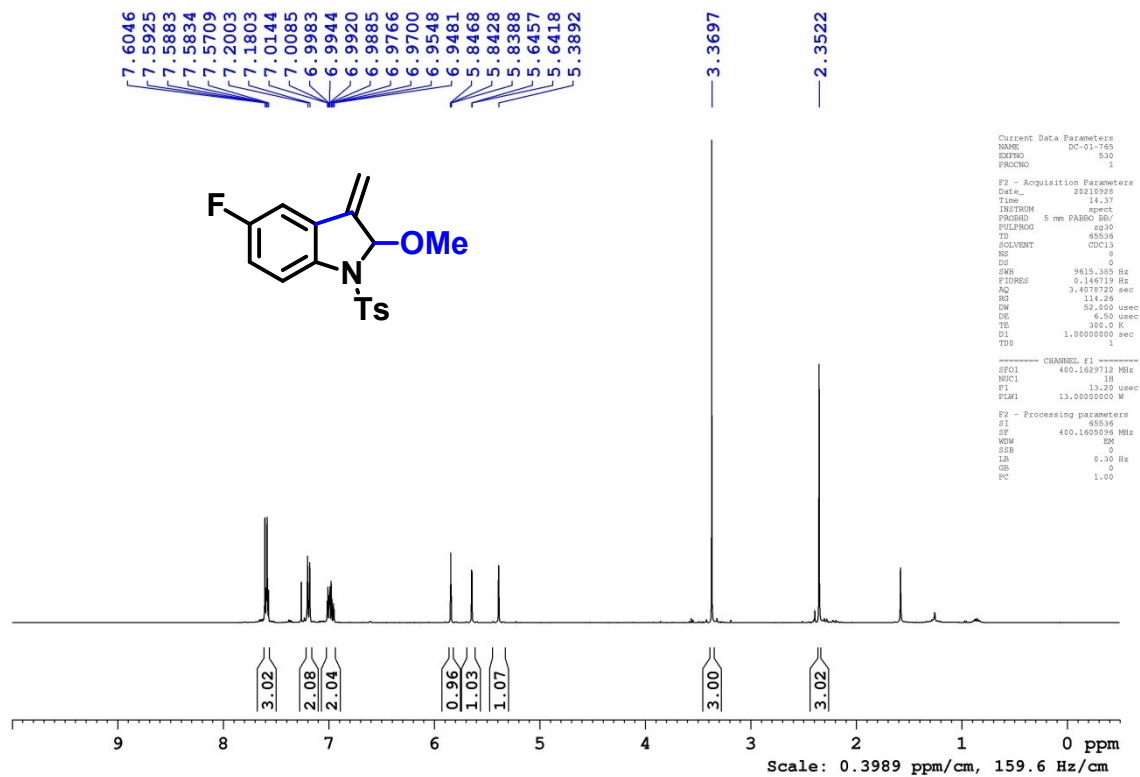


Figure S-38: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3d

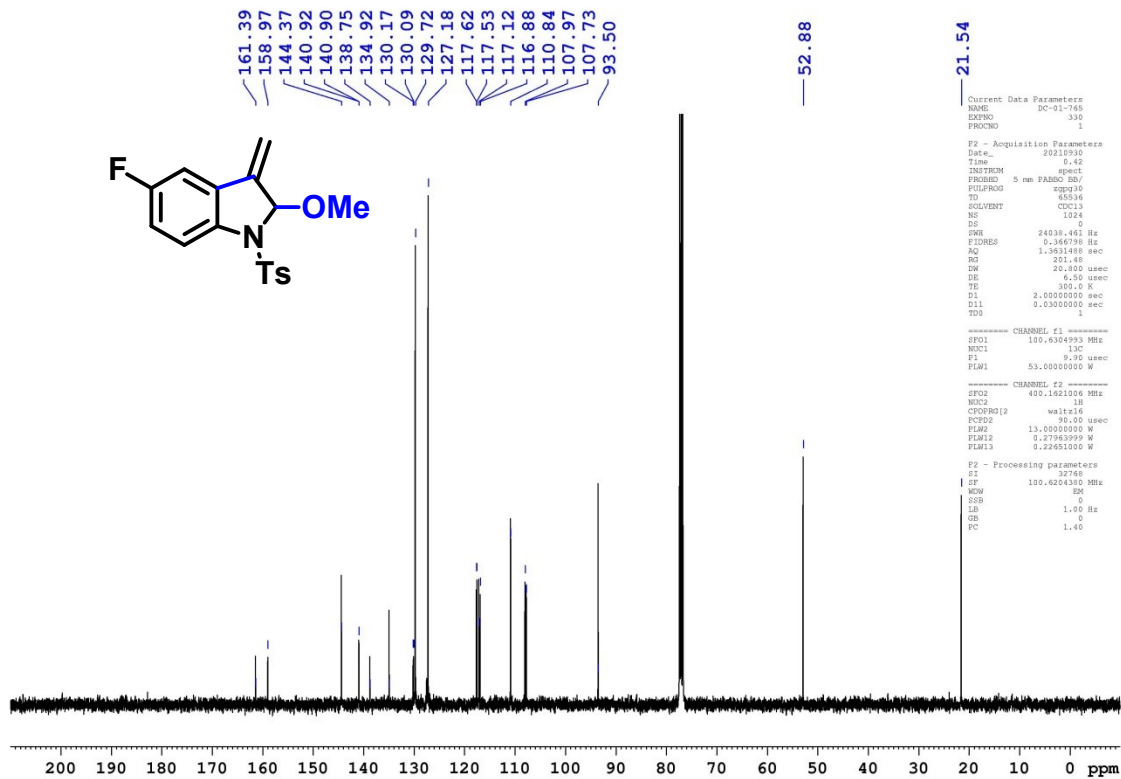


Figure S-39: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3d

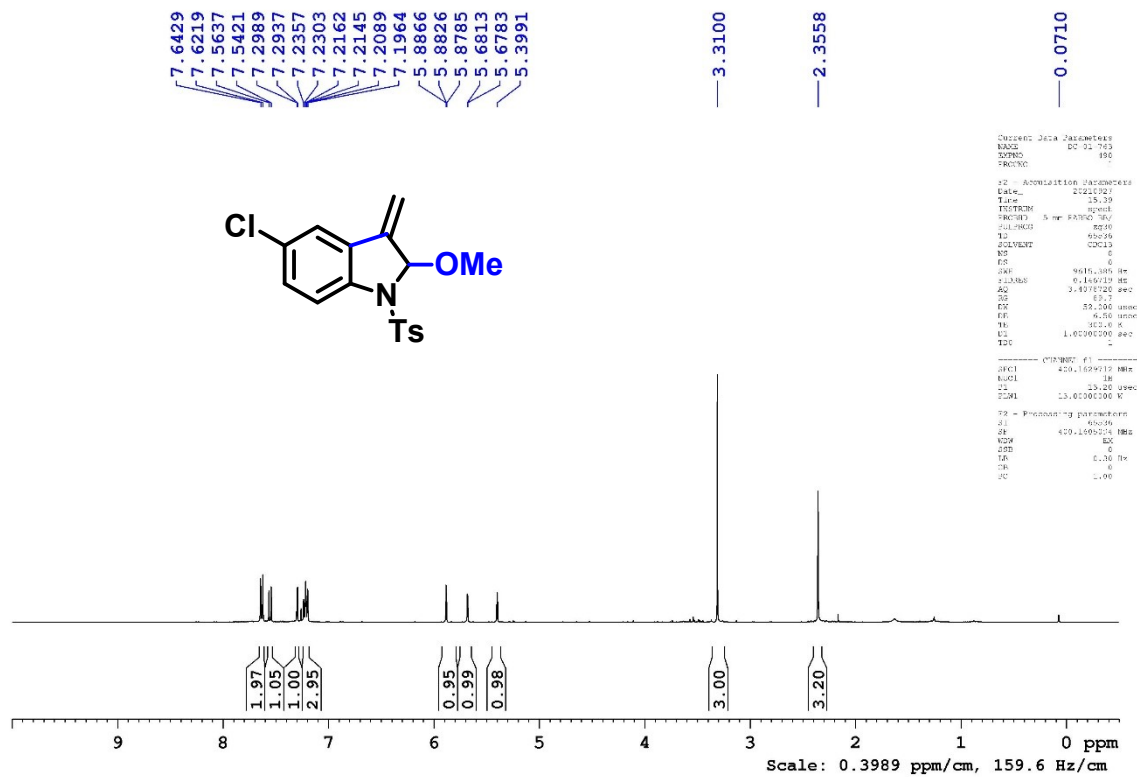


Figure S-42: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3e

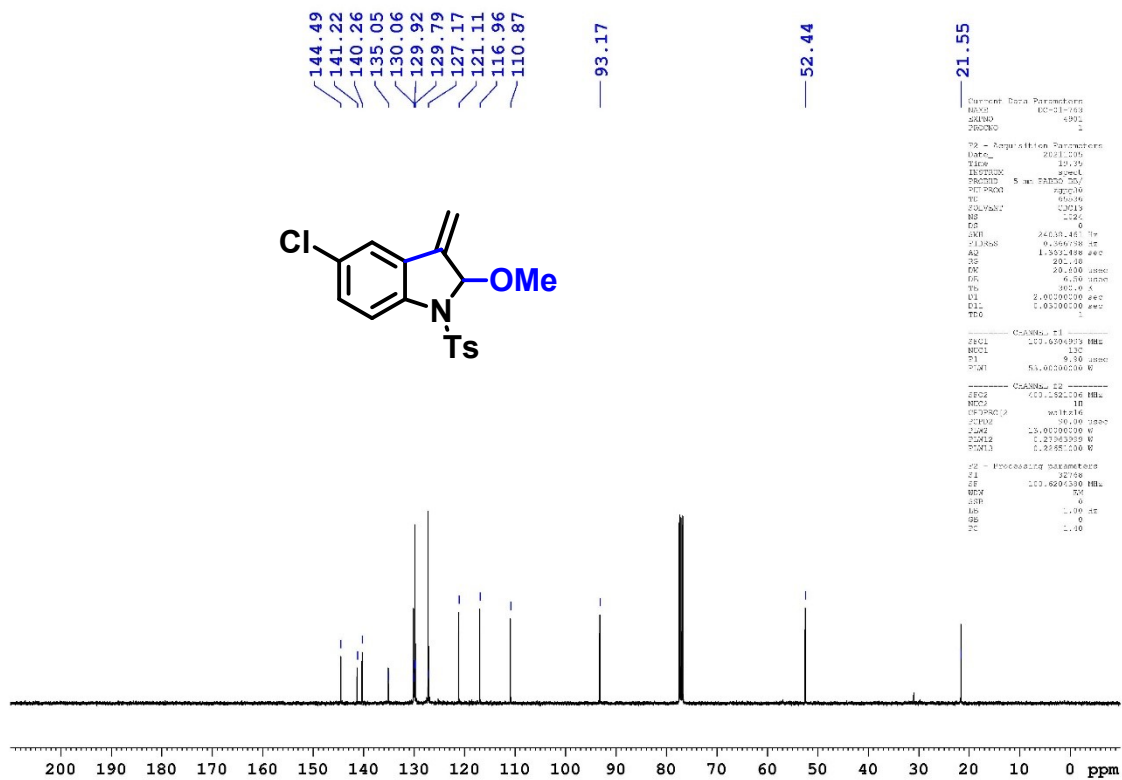


Figure S-43: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3e

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|---------------------|
| Sample Name | HRMS22101FEB20 | Position | Vial 20 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | KMR-02-79A3.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/1/2022 1:02:24 PM |

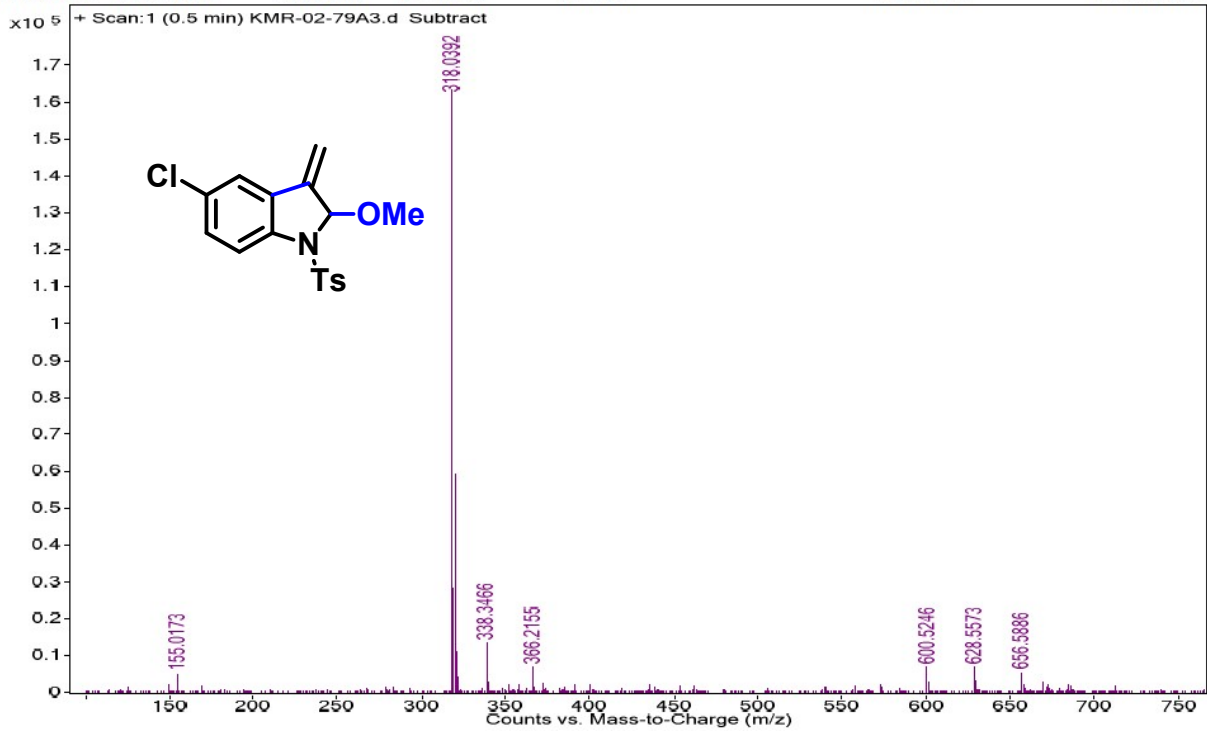


Figure S-44: HRMS spectrum of compound 3e

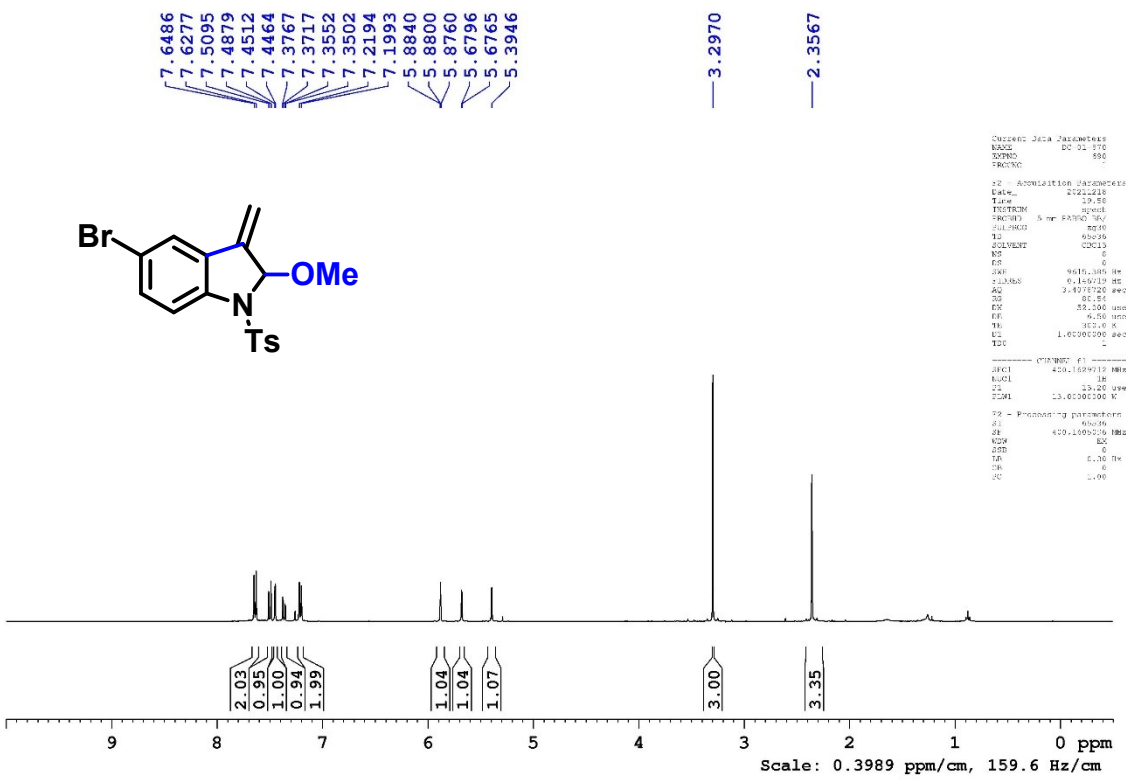


Figure S-45: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3f

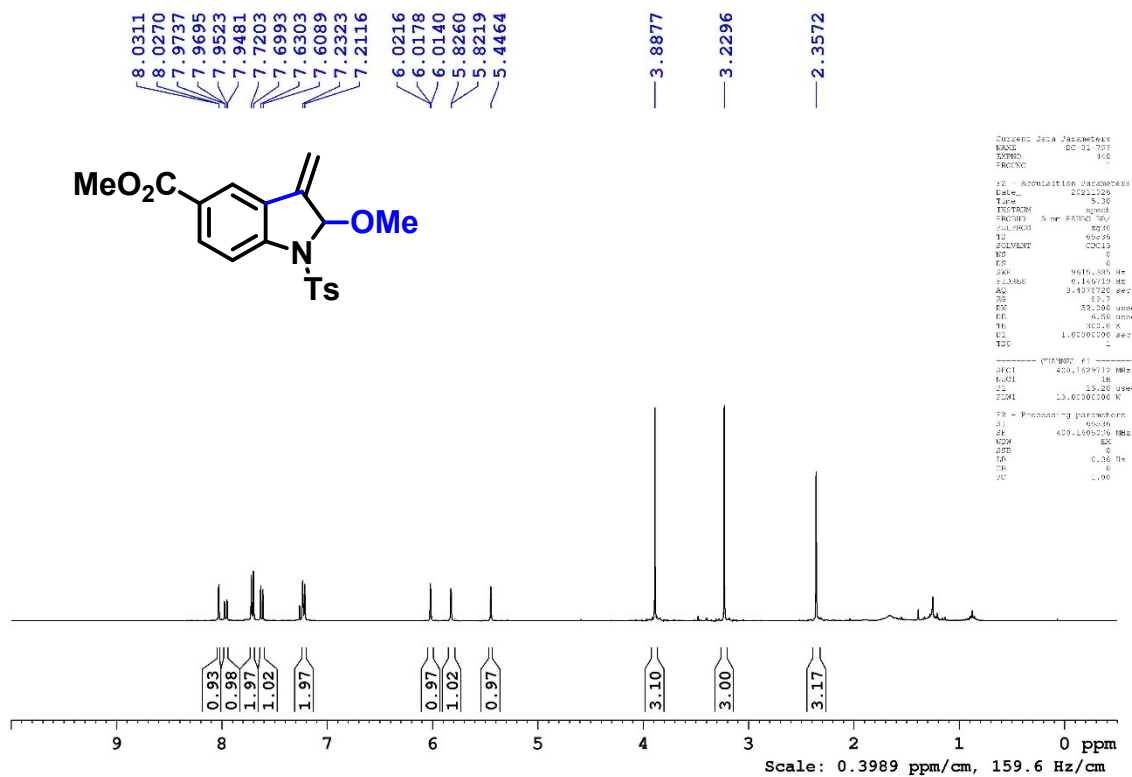


Figure S-48: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3g**

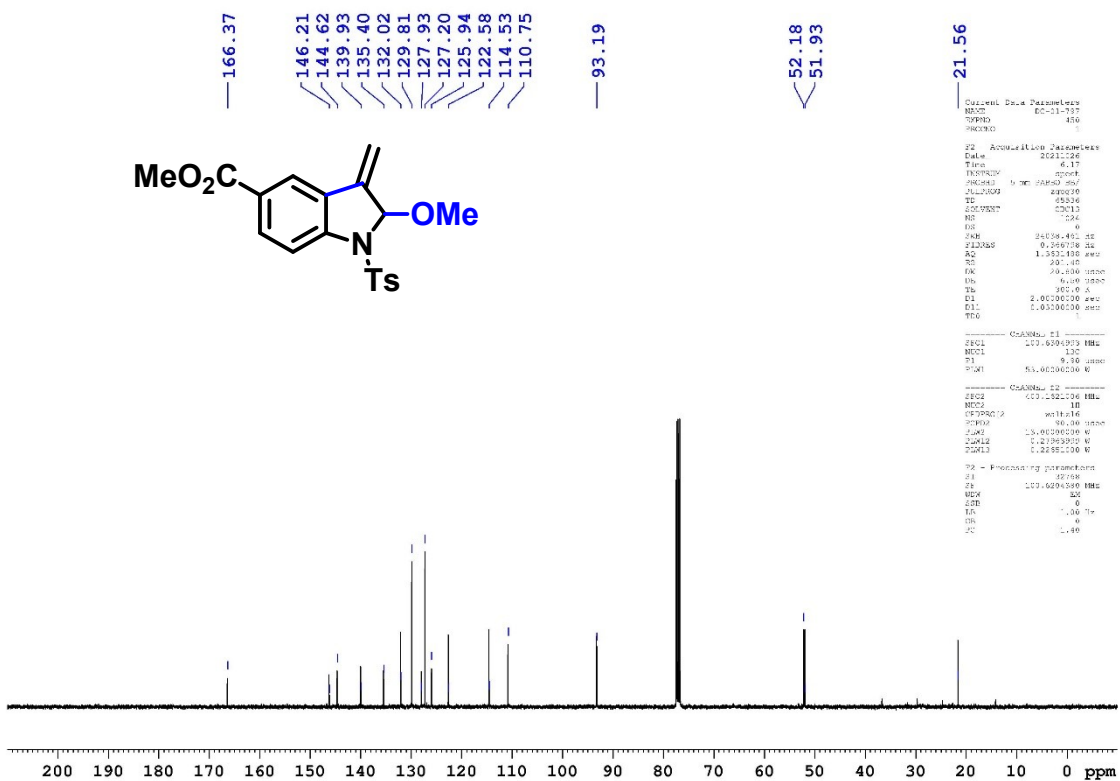


Figure S-49: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3g**

| Sample Name | HRMS22131JAN07 | Position | Vial 7 | Instrument Name | Instrument 1 | User Name | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | NKM-02-115.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 1/31/2022 11:56:34 AM |

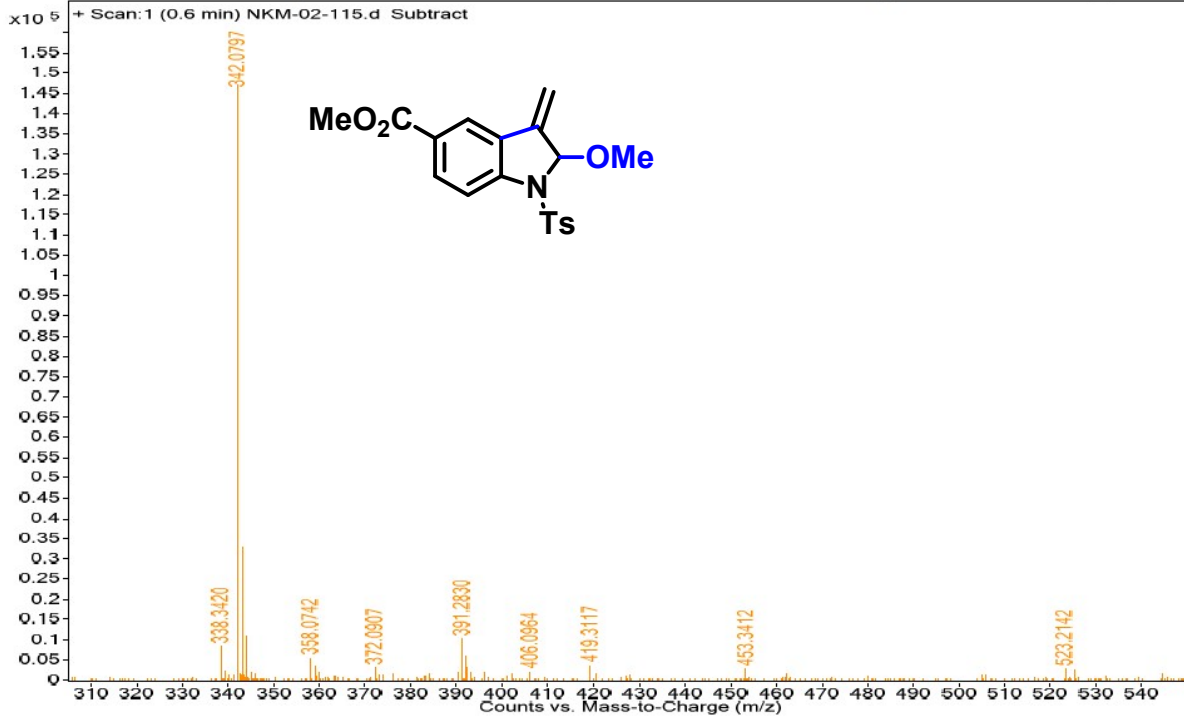


Figure S-50: HRMS spectrum of compound **3g**

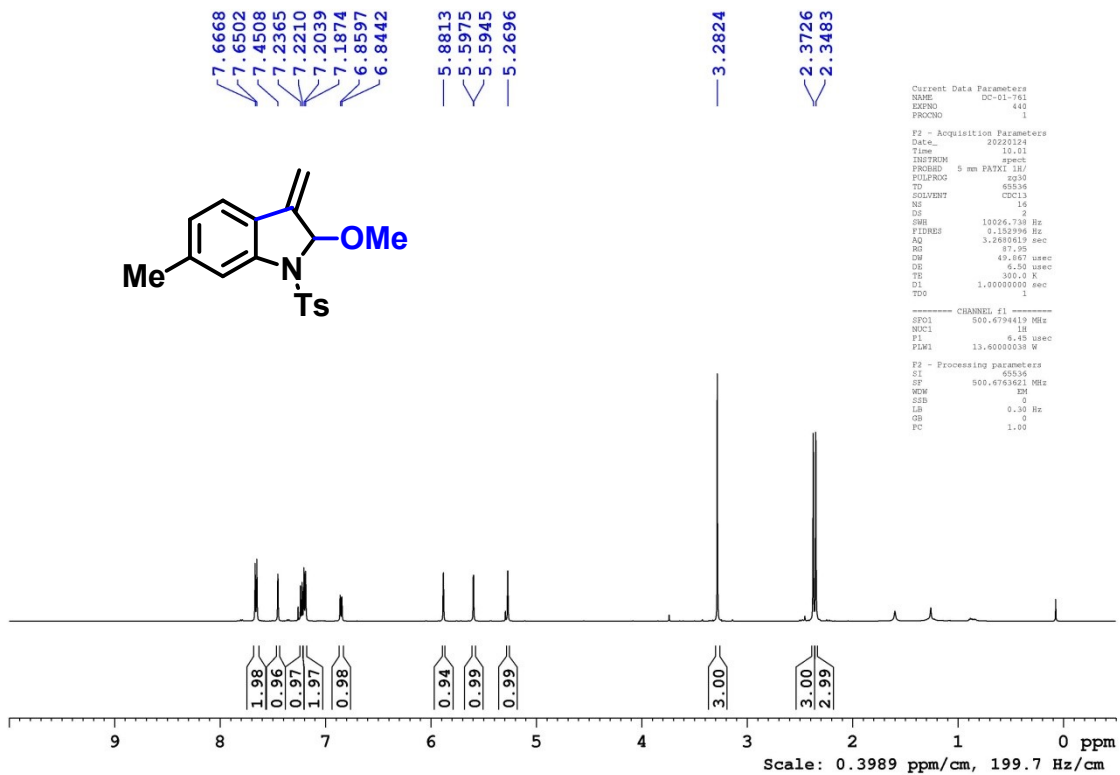


Figure S-51: ^1H NMR (500 MHz, CDCl_3) spectrum of compound **3h**

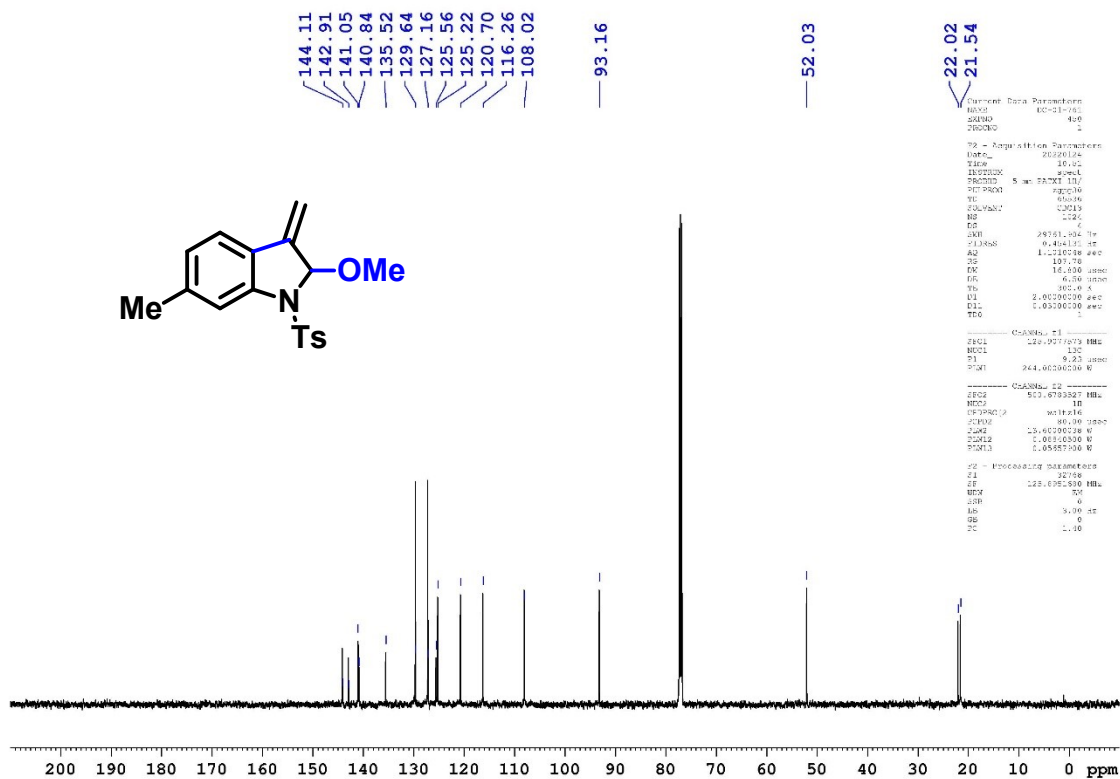


Figure S-52: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3h

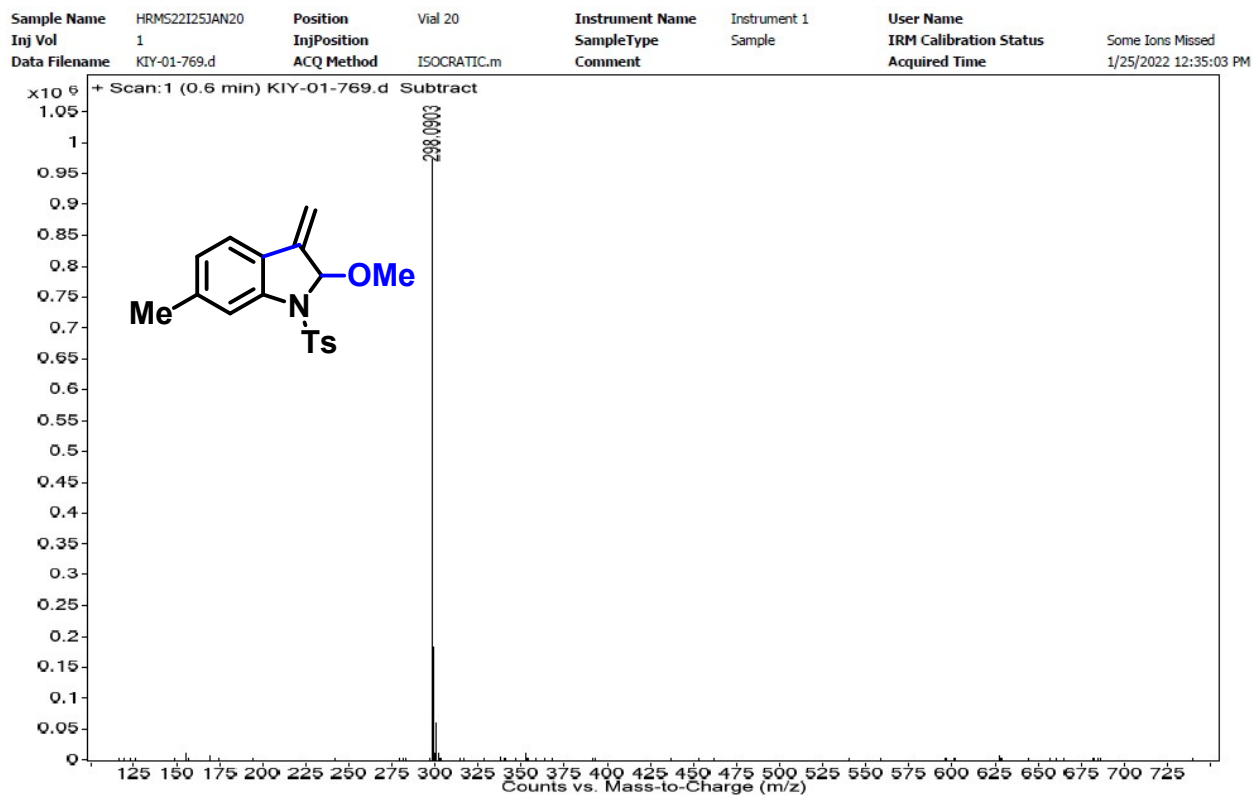


Figure S-53: HRMS spectrum of compound 3h

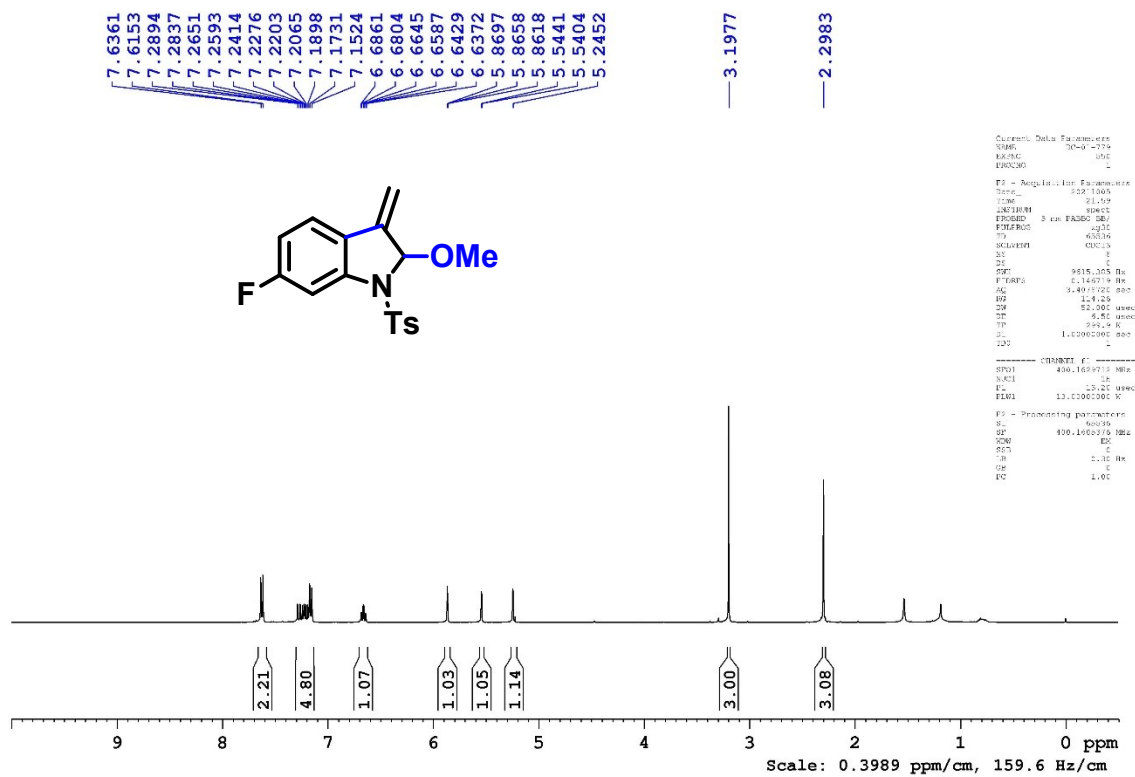


Figure S-54: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3i**

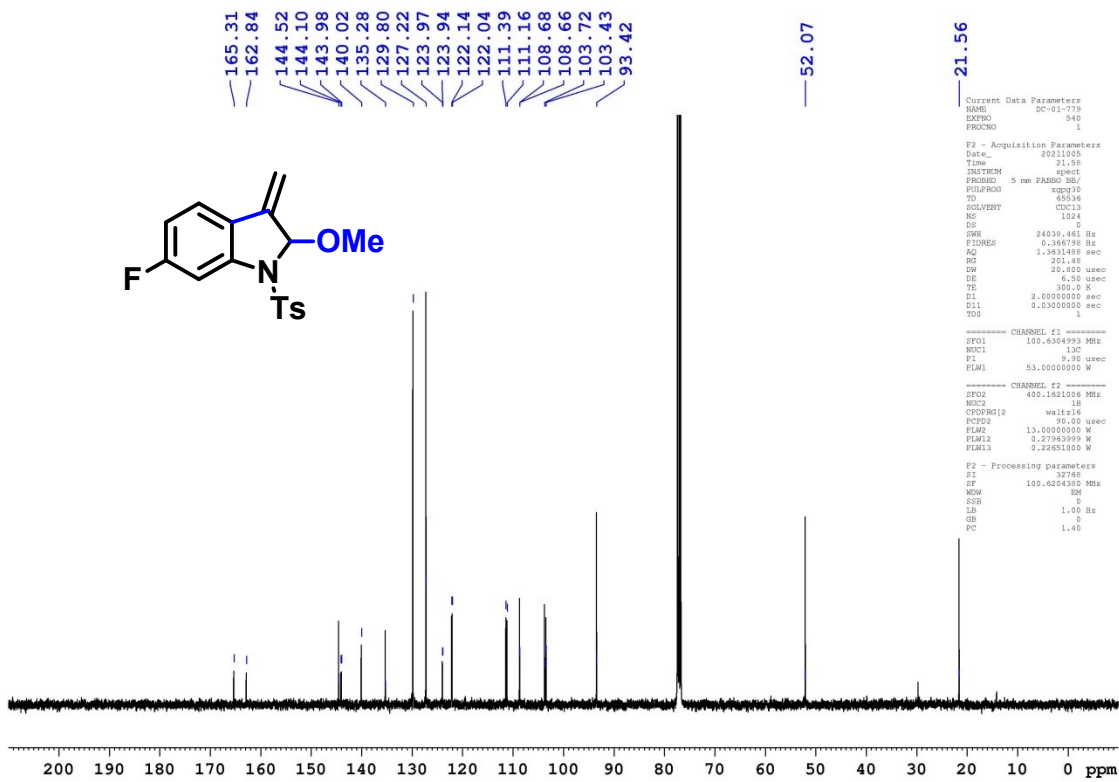


Figure S-55: ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3i**

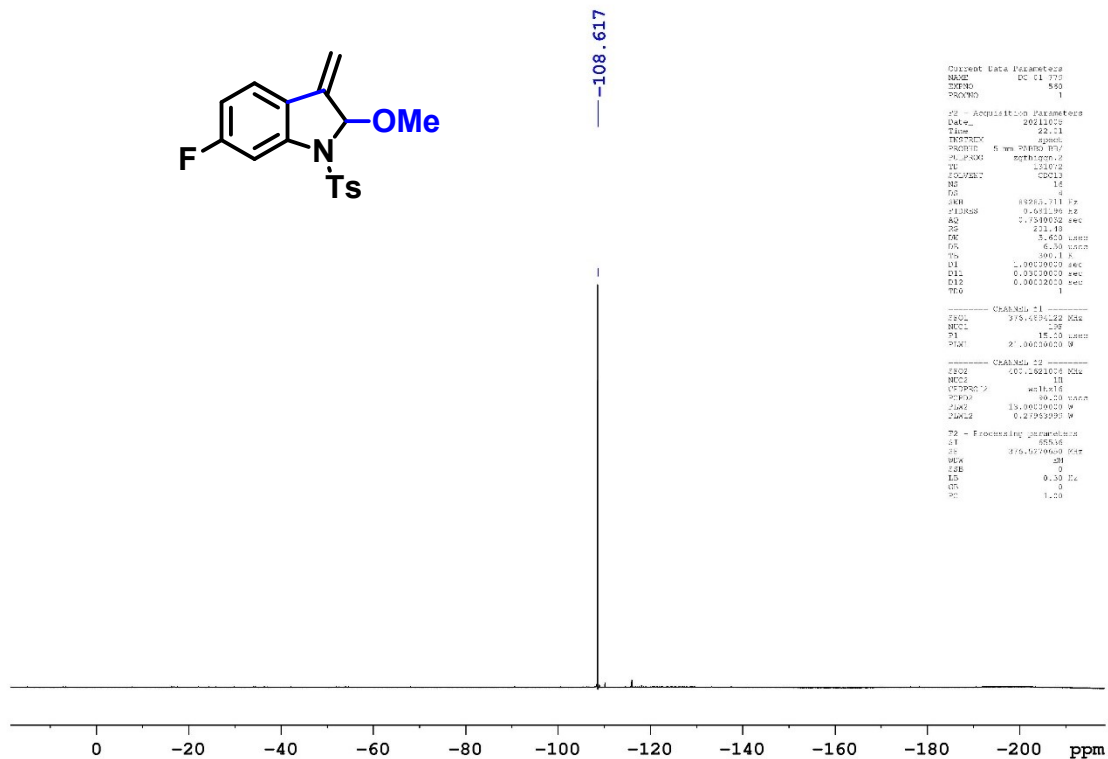


Figure S-56: ^{19}F NMR (376 MHz, CDCl_3) spectrum of compound 3i

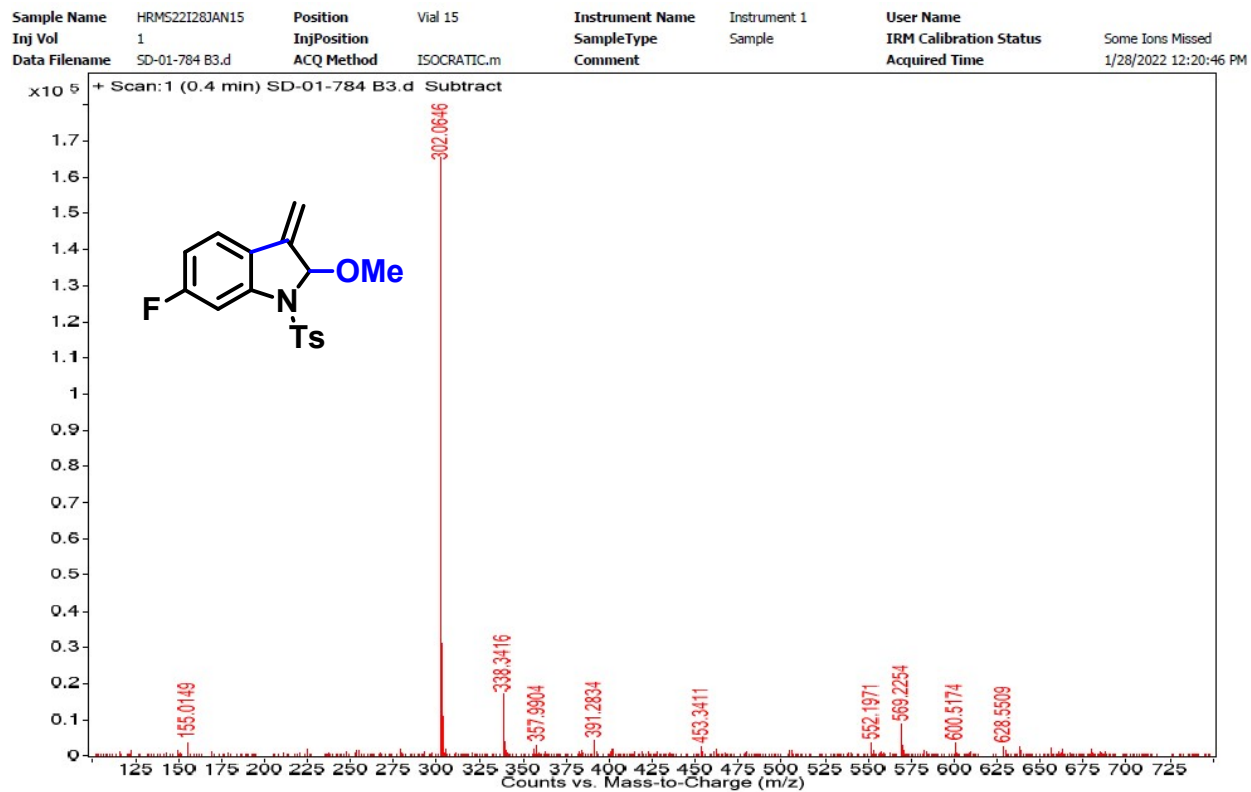


Figure S-57: HRMS spectrum of compound 3i

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|----------------------|
| Sample Name | HRM522101FEB19 | Position | Vial 19 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | KMR-02-79A2.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/1/2022 12:59:17 PM |

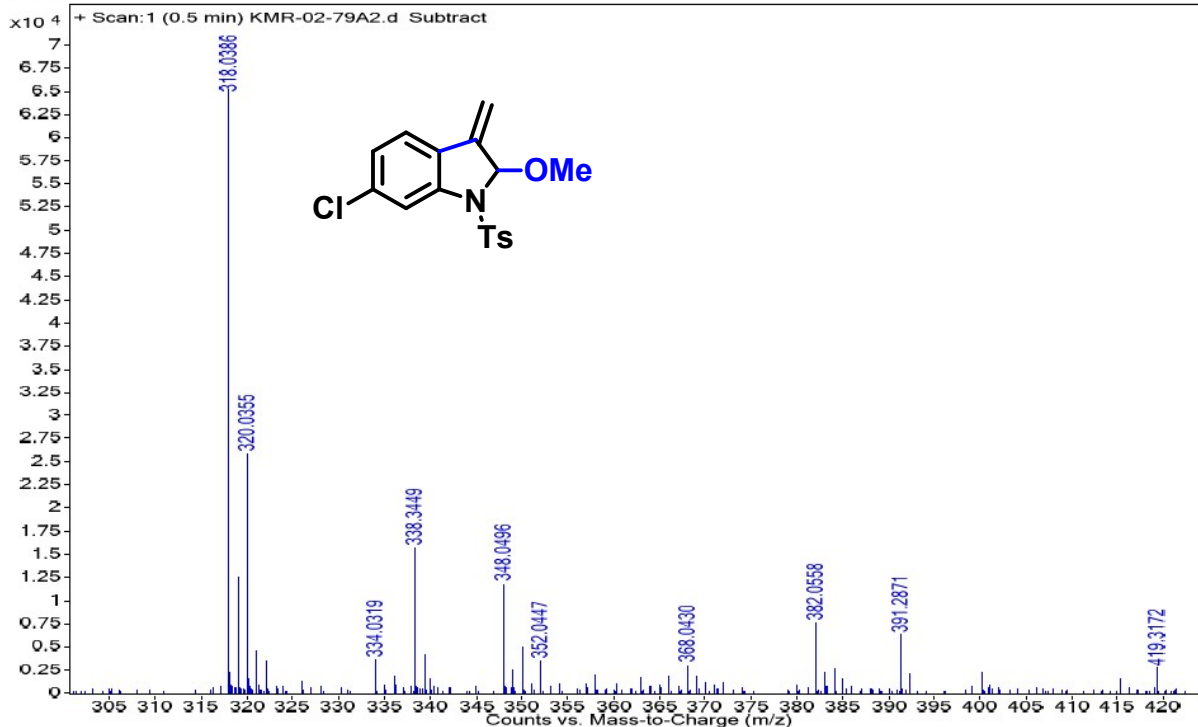


Figure S-60: HRMS spectrum of compound 3j

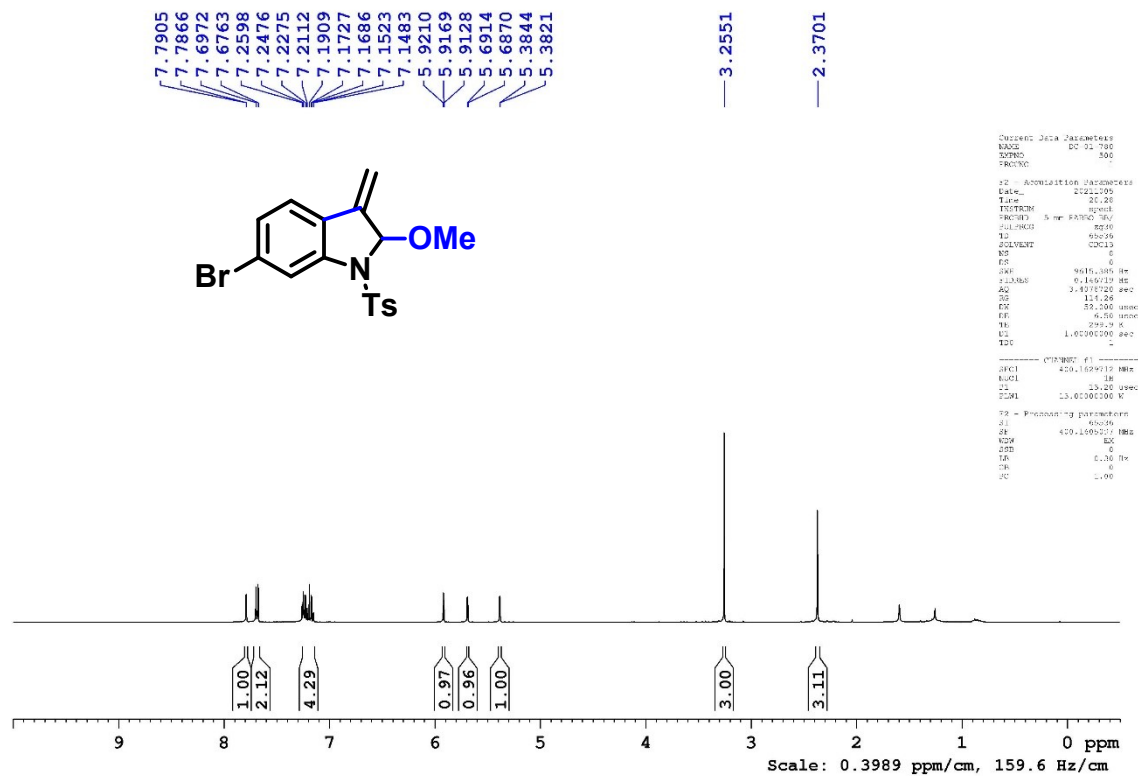


Figure S-61: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3k

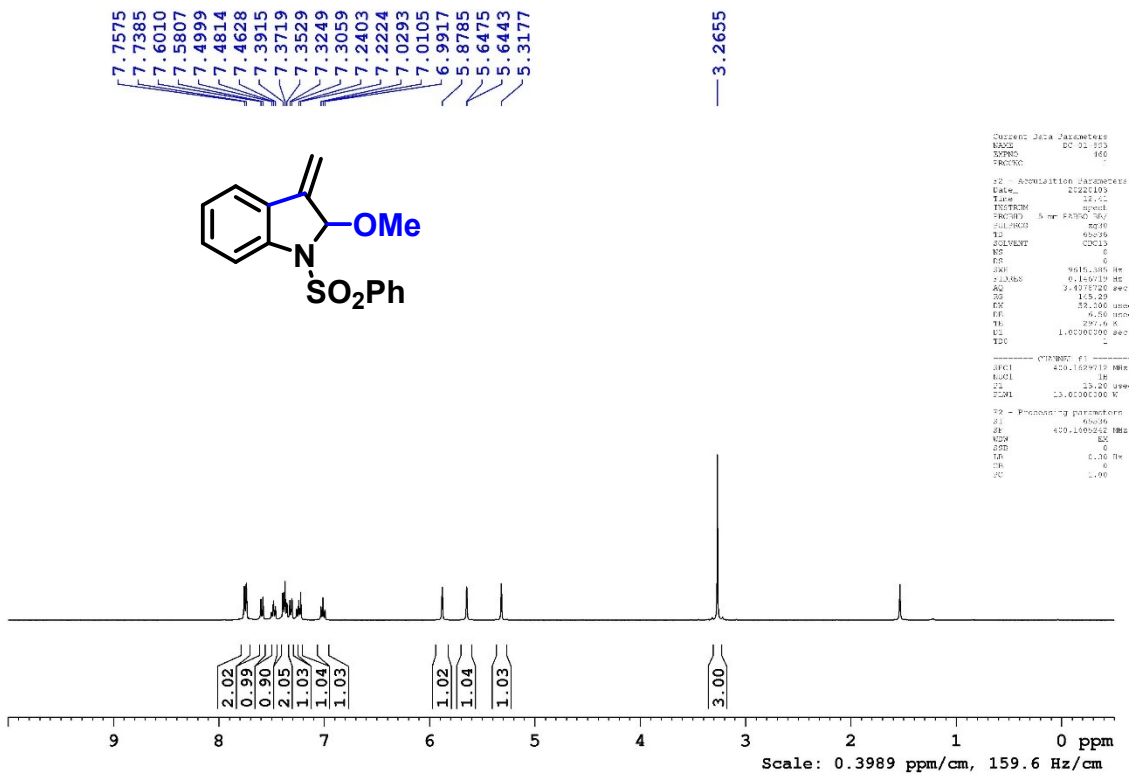


Figure S-64: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **31**

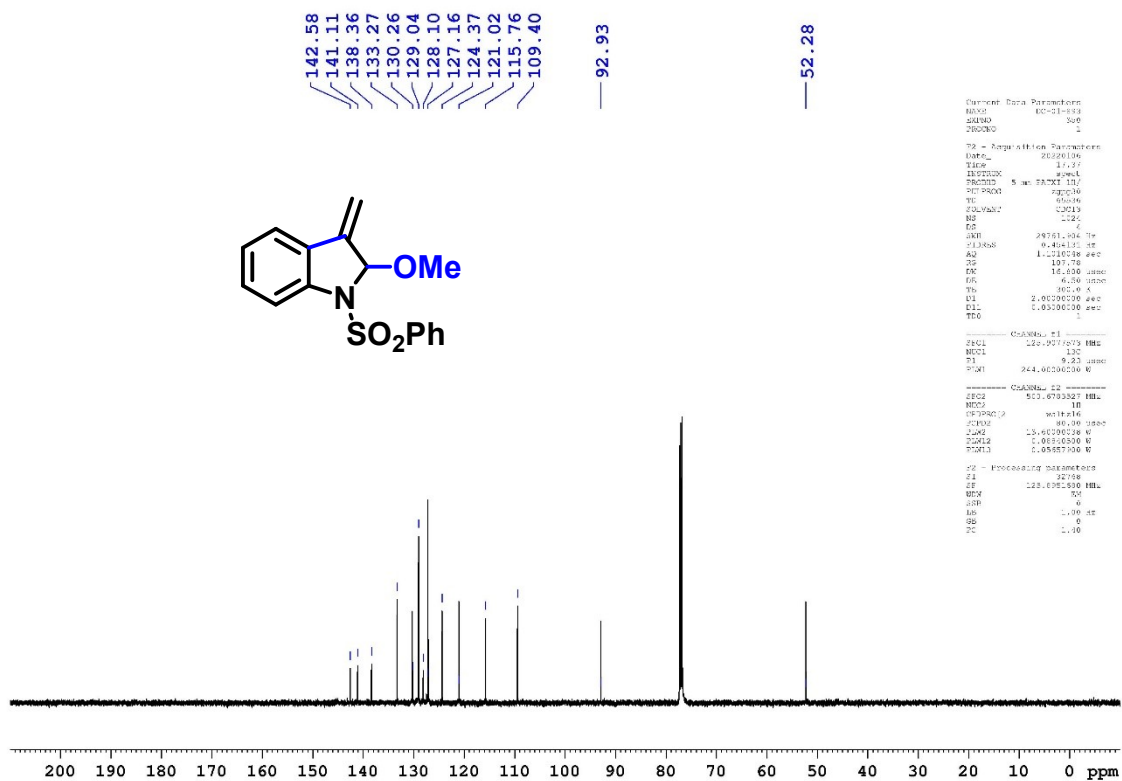


Figure S-65: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound **31**

Sample Name HRMS22124JAN06 Position Vial 6 Instrument Name Instrument 1 User Name
 Inj Vol 1 InjPosition Instrument 1 Sample IRM Calibration Status Some Ions Missed
 Data Filename KMR-02-94Aa.d ACQ Method ISOCRATIC.m Comment Acquired Time 1/24/2022 4:35:05 PM

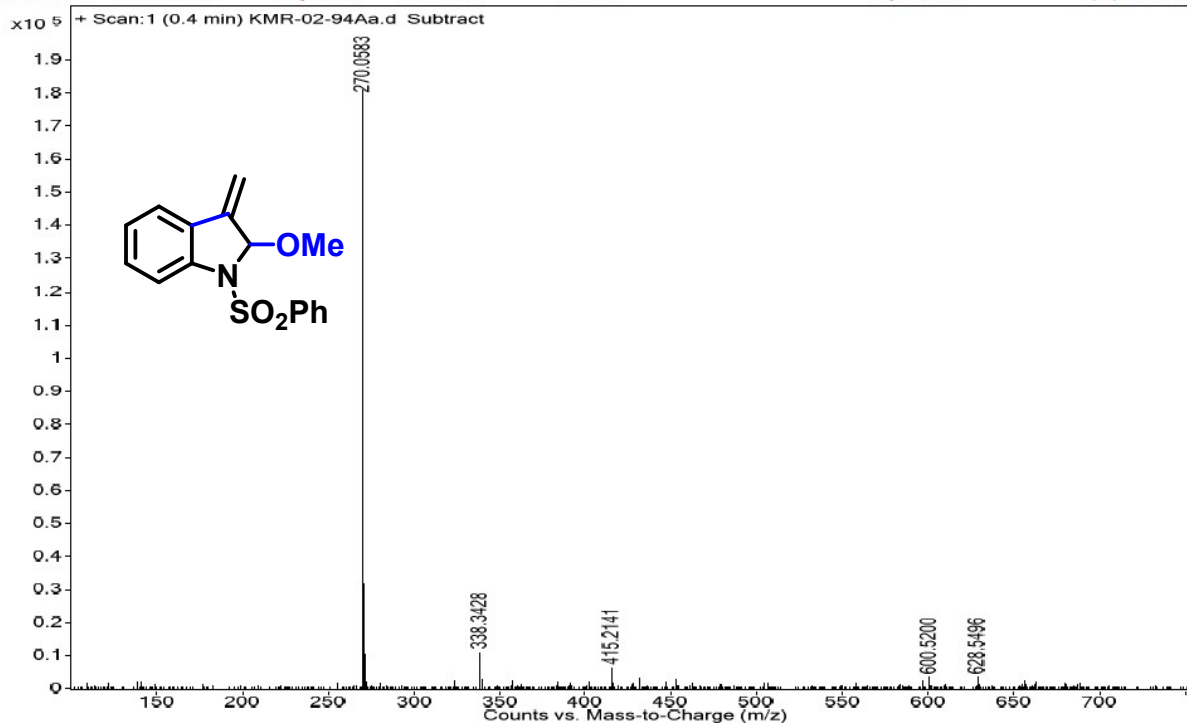


Figure S-66: HRMS spectrum of compound 31

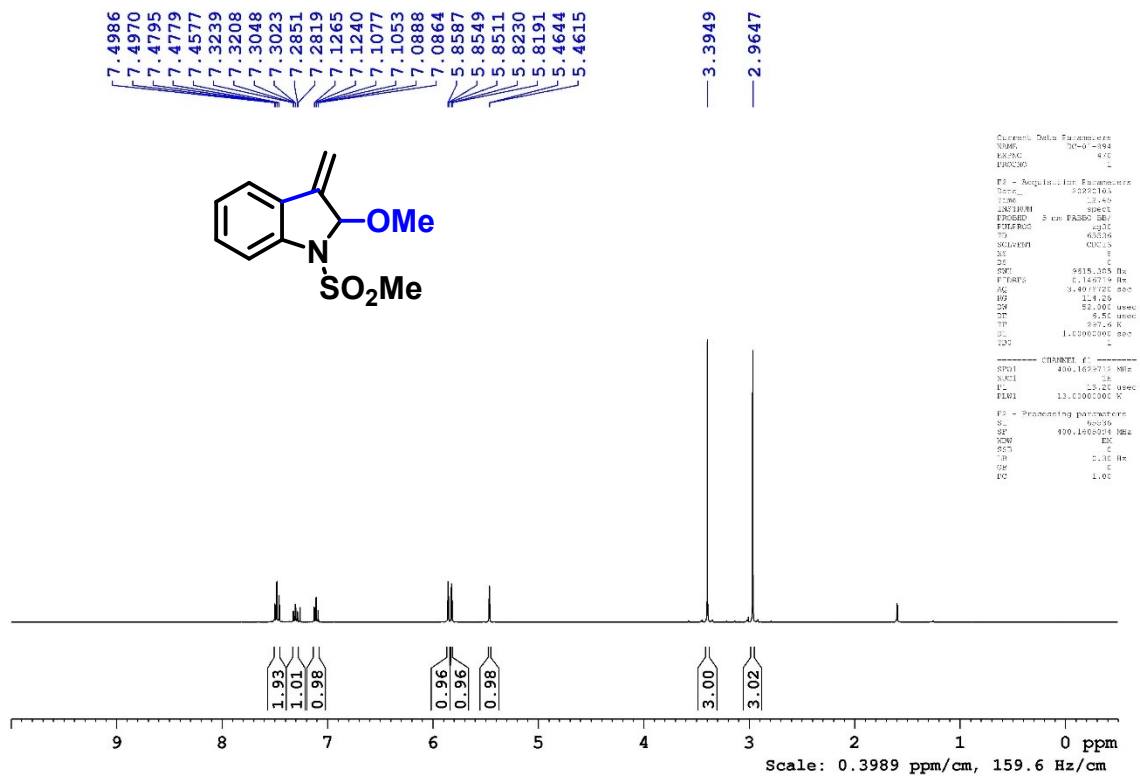


Figure S-67: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3m

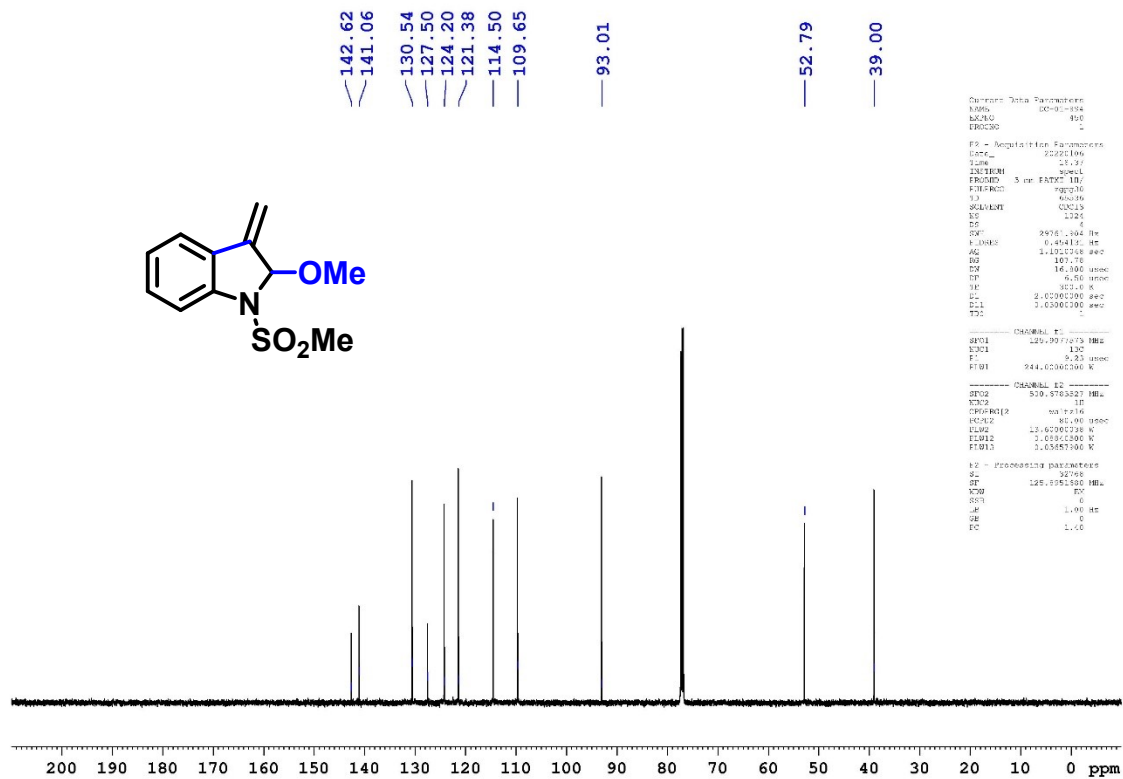


Figure S-68: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3m

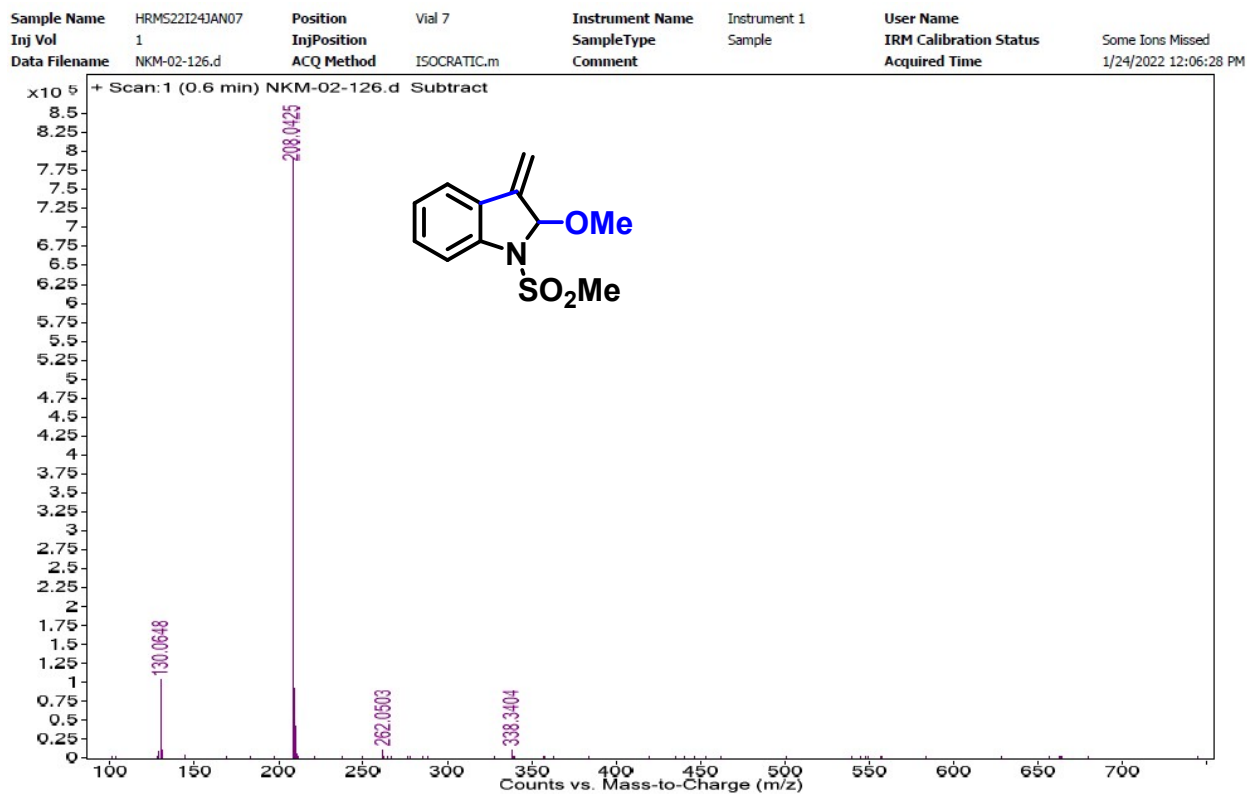


Figure S-69: HRMS spectrum of compound 3m

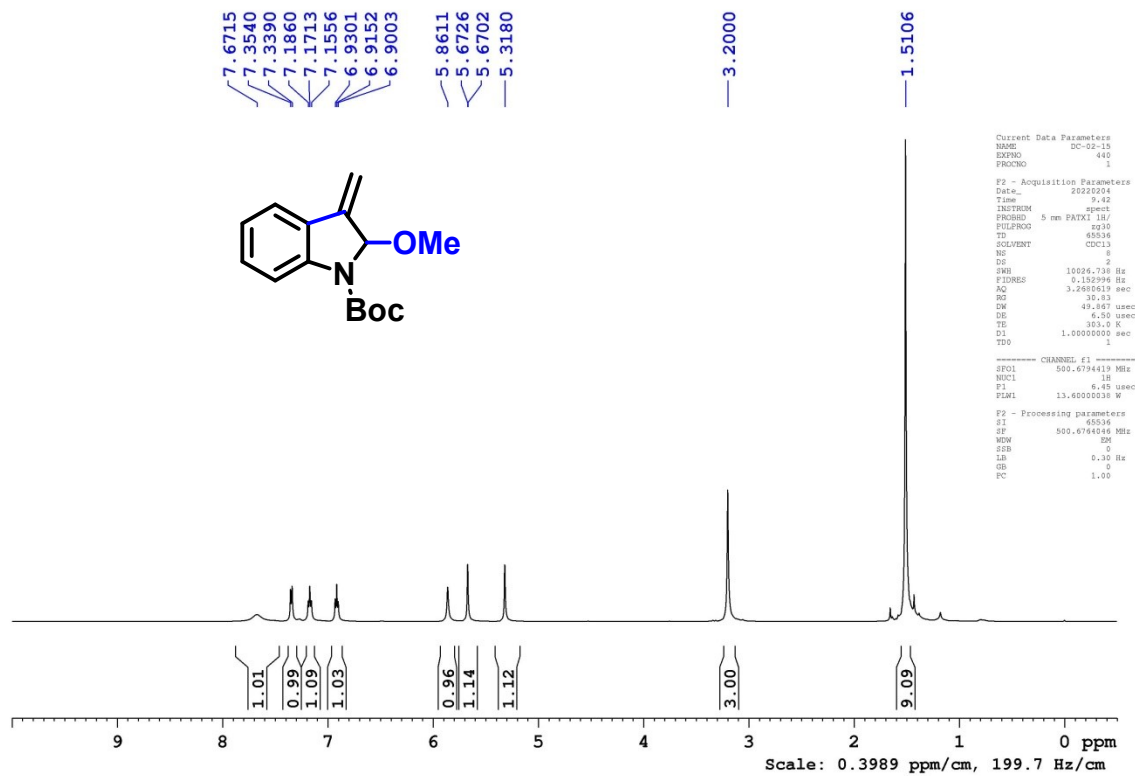


Figure S-70: ^1H NMR (500 MHz, CDCl_3) spectrum of compound **3n**

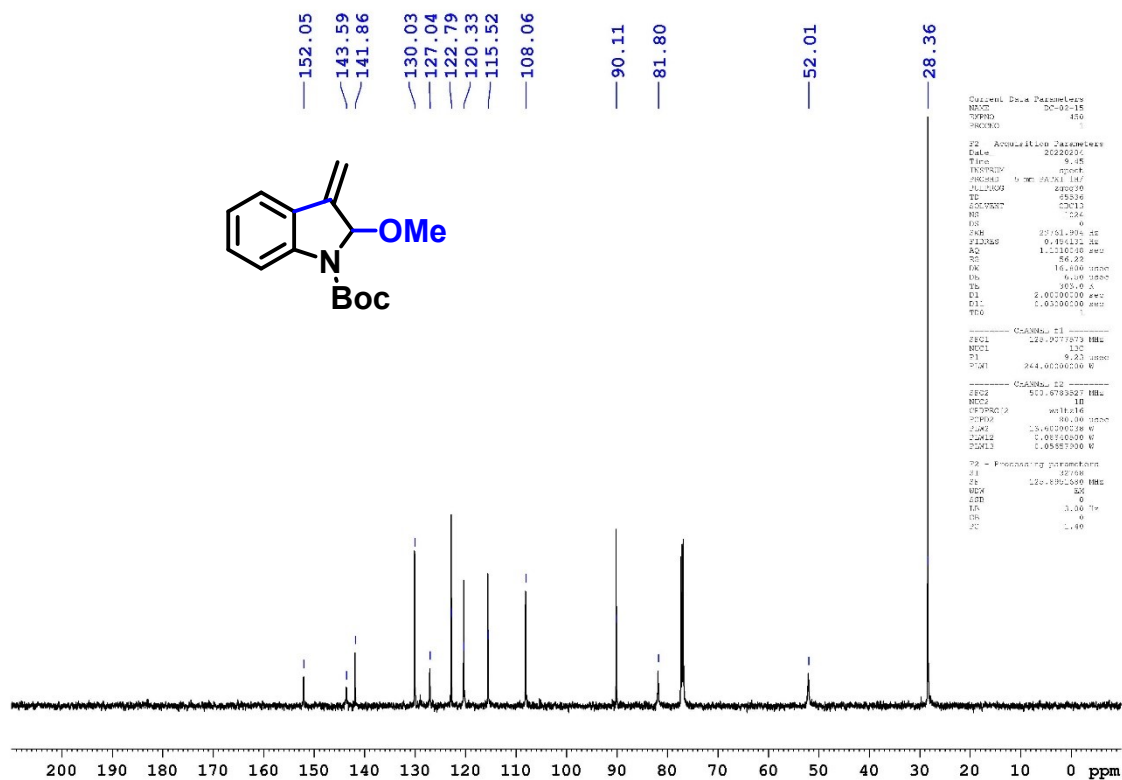


Figure S-71: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound **3n**

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|----------------------|
| Sample Name | HRMS22I07FEB18 | Position | Vial 18 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | KMR-02-97A.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/7/2022 12:30:37 PM |

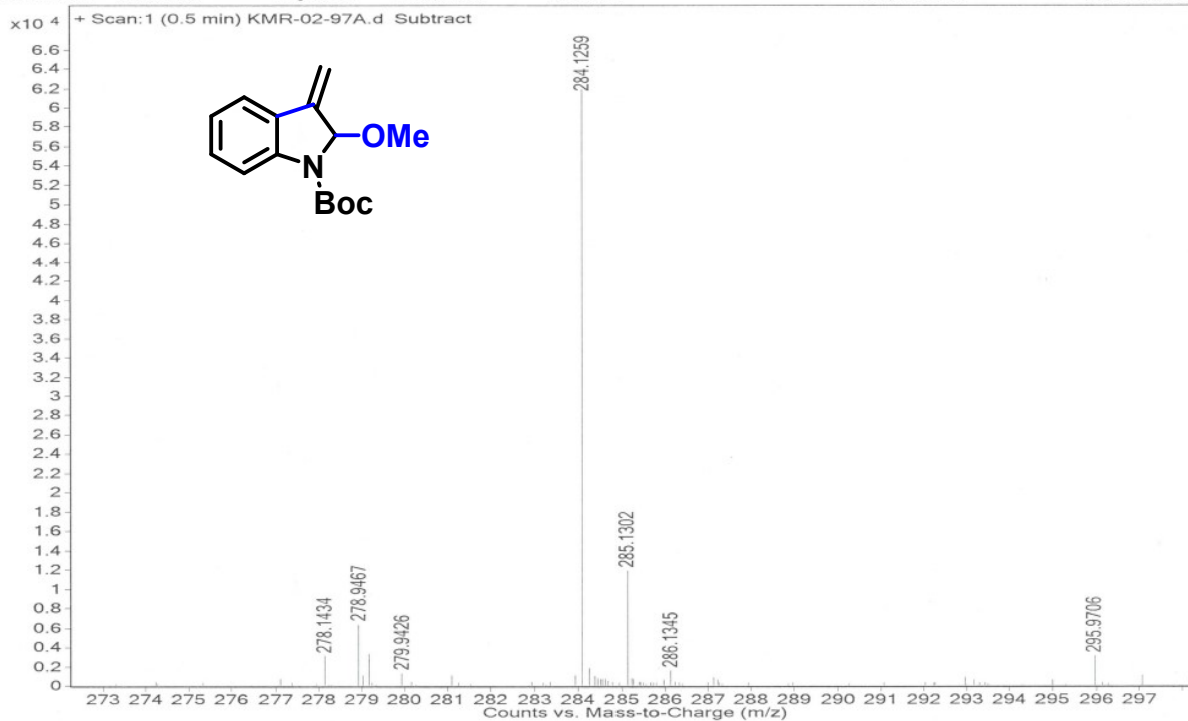


Figure S-72: HRMS spectrum of compound **3n**

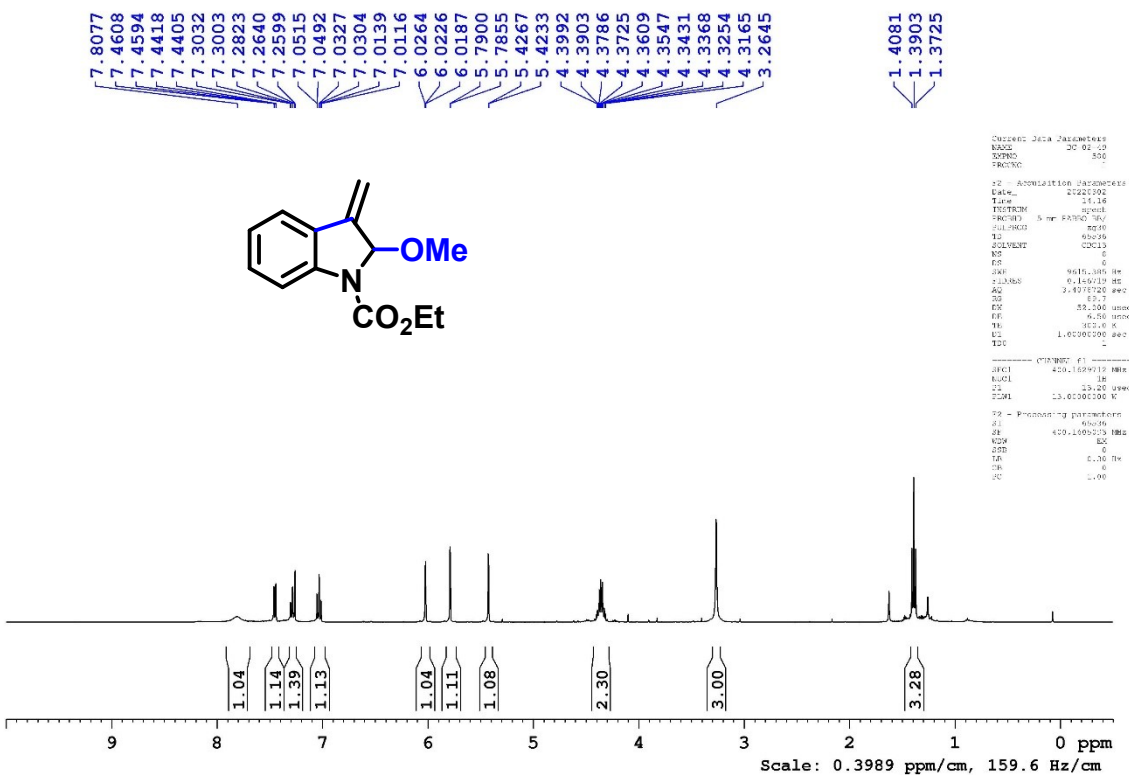


Figure S-73: ^1H NMR (400 MHz, CDCl_3) spectrum of compound **3o**

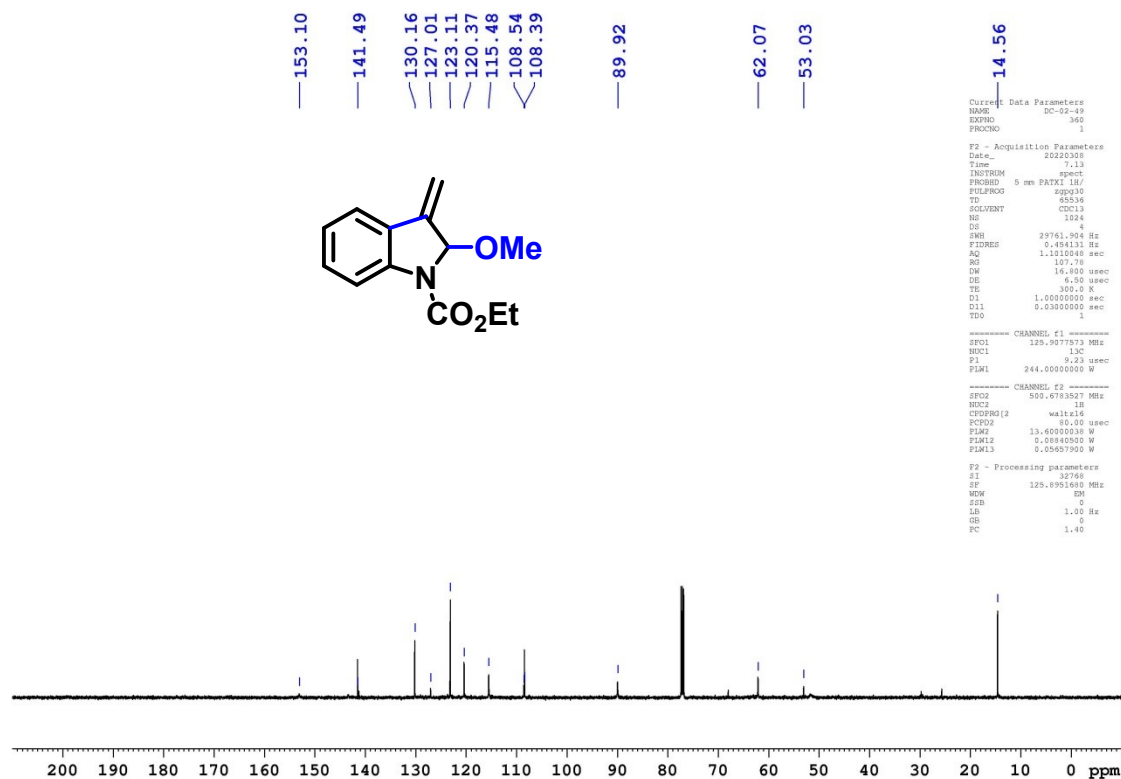


Figure S-74: ^{13}C NMR (100 MHz, CDCl_3) spectrum of compound 30

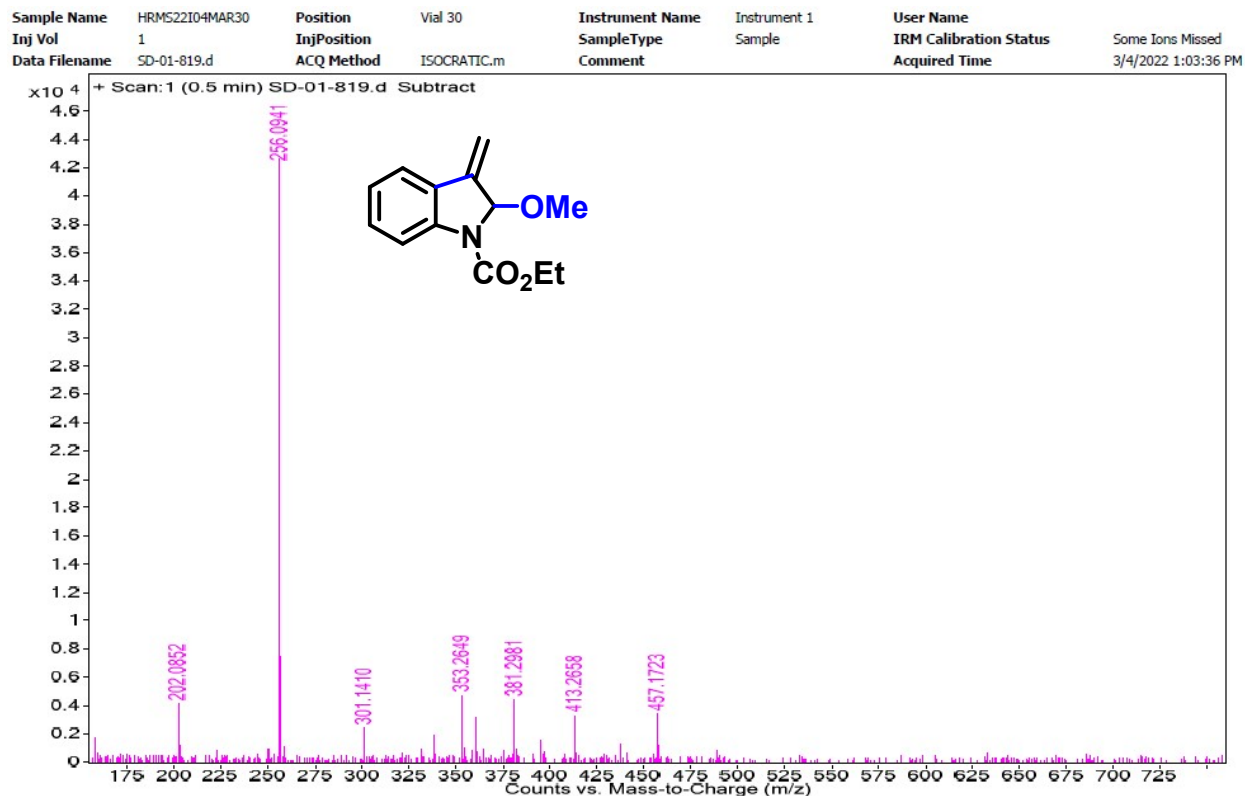


Figure S-75: HRMS spectrum of compound 30

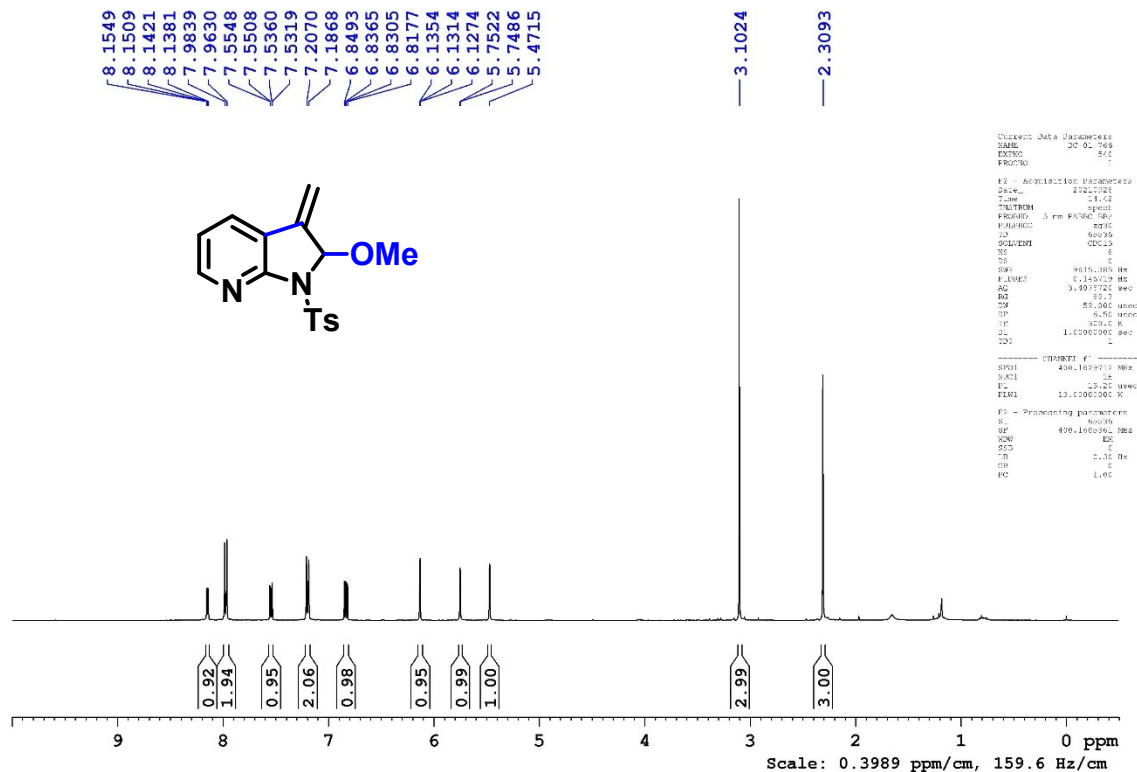


Figure S-76: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3p

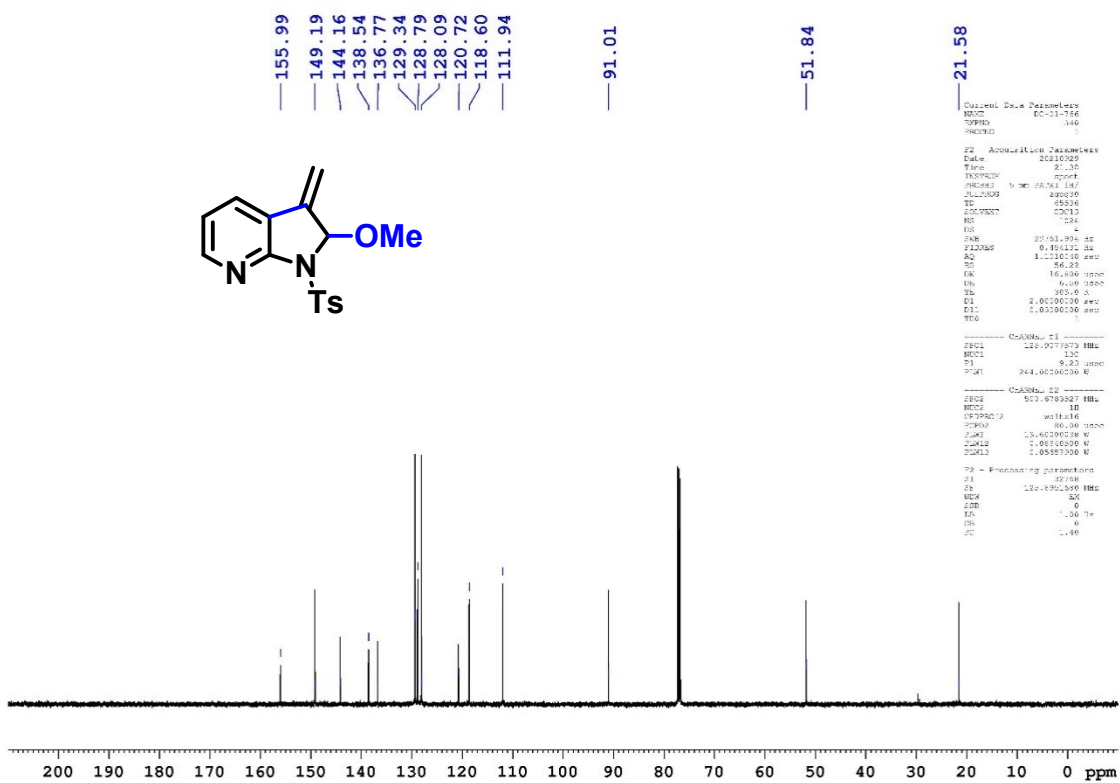


Figure S-77: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3p

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Sample Name | HRMS22I24JAN21 | Position | Vial 21 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | KM-CY-P.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 1/24/2022 12:50:03 PM |

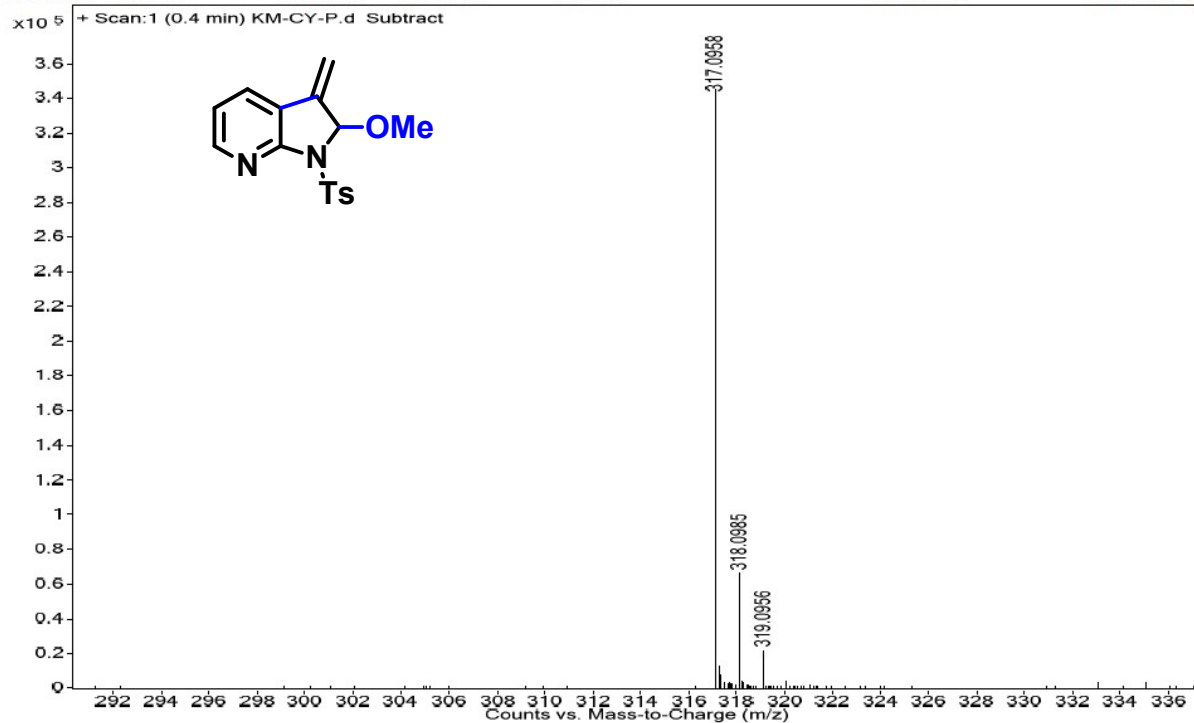


Figure S-78: HRMS spectrum of compound **3p**

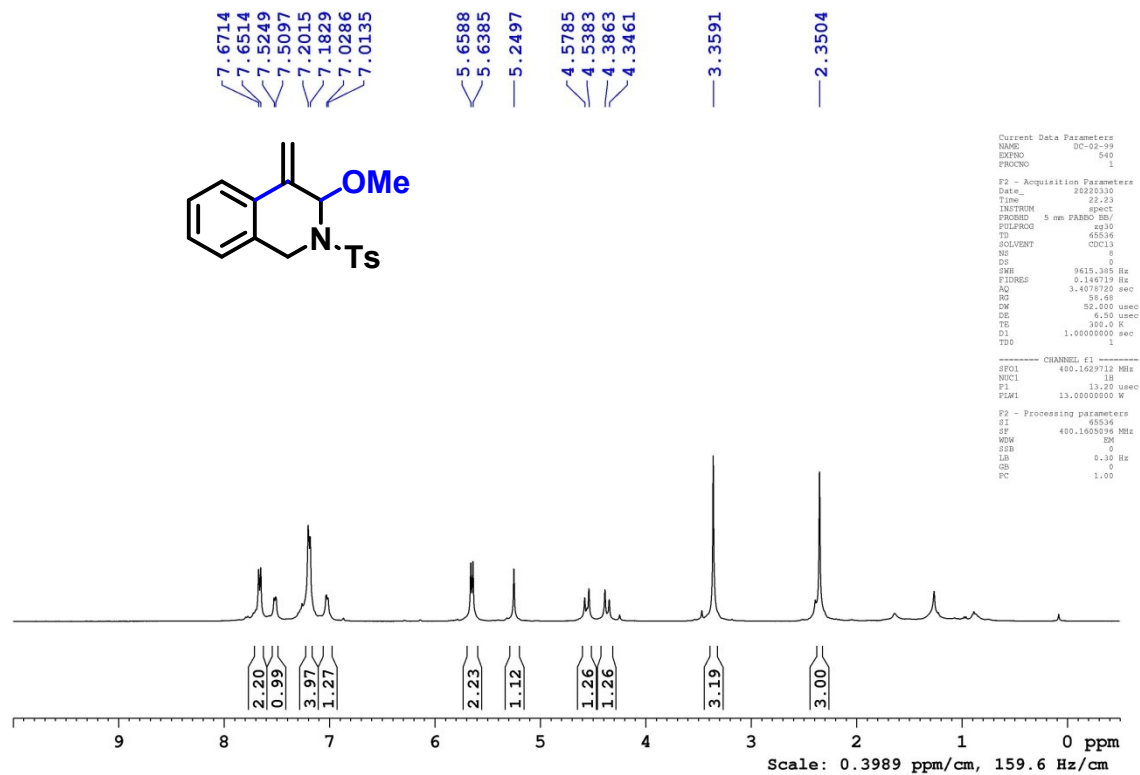


Figure S-79: ^1H NMR (400 MHz, CDCl_3) spectrum of compound **3q**

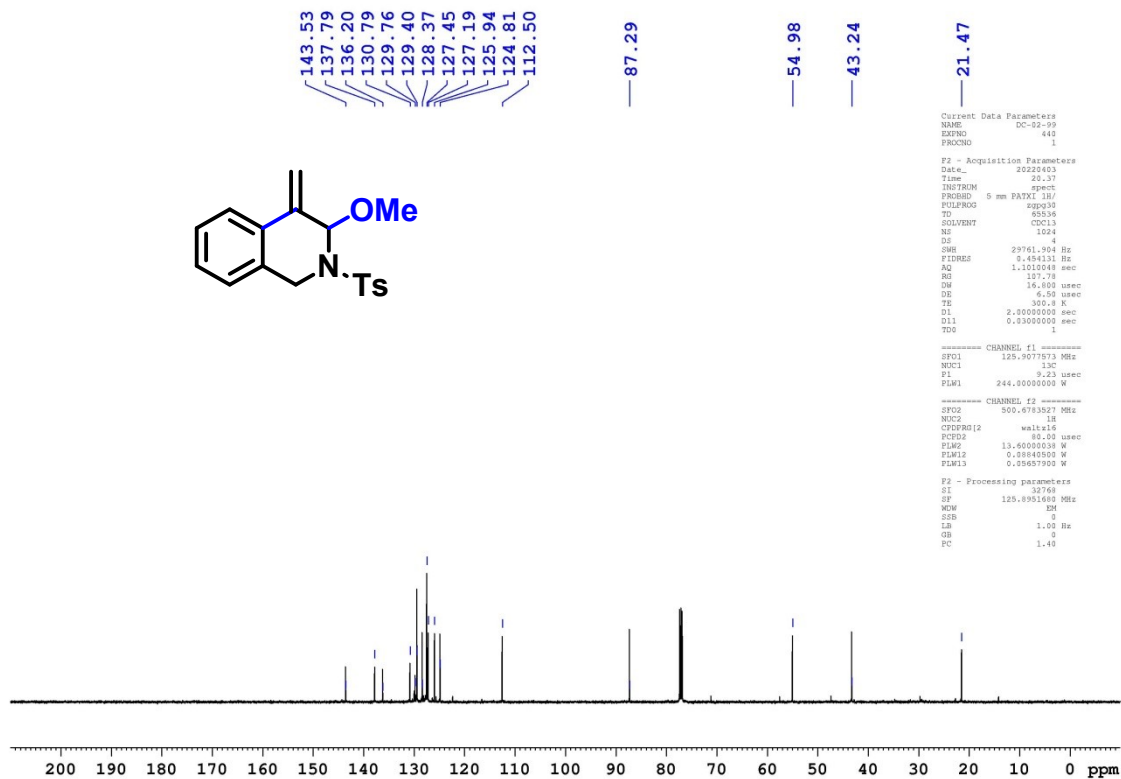


Figure S-80: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 3q

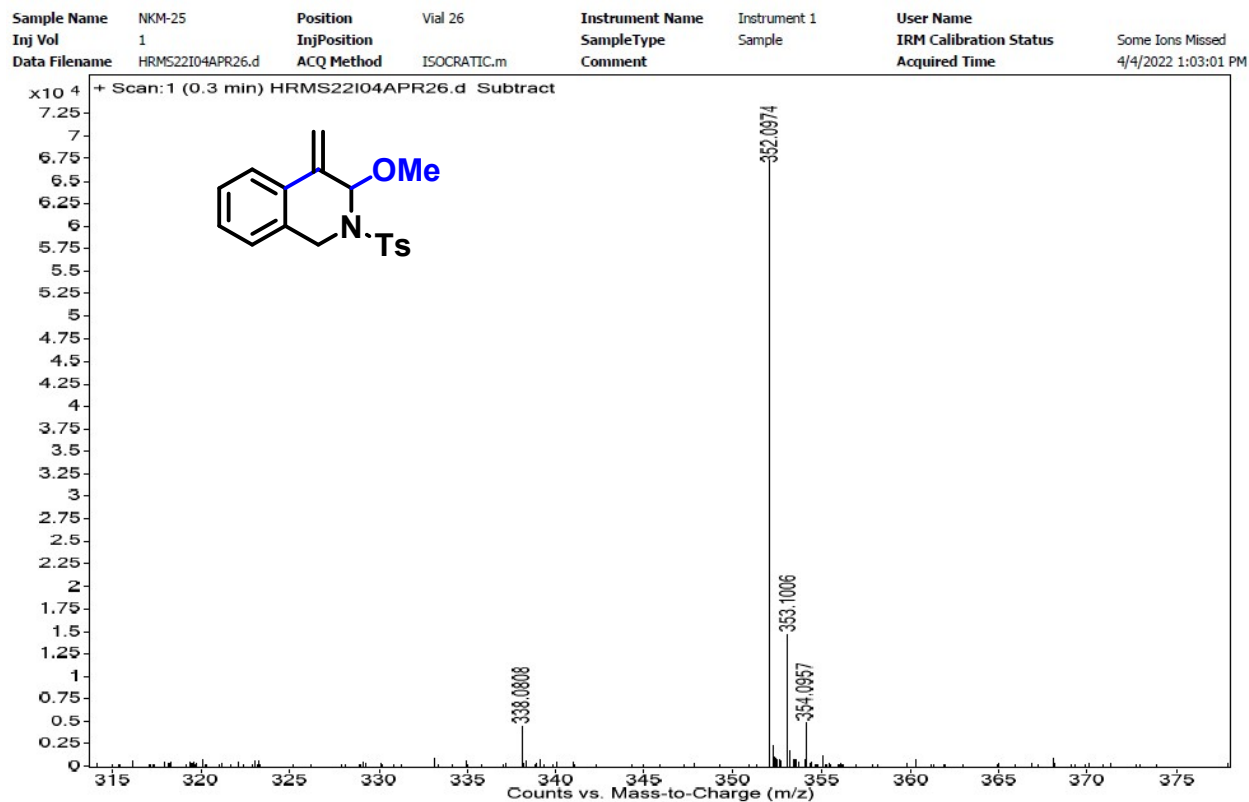


Figure S-81: HRMS spectrum of compound 3q

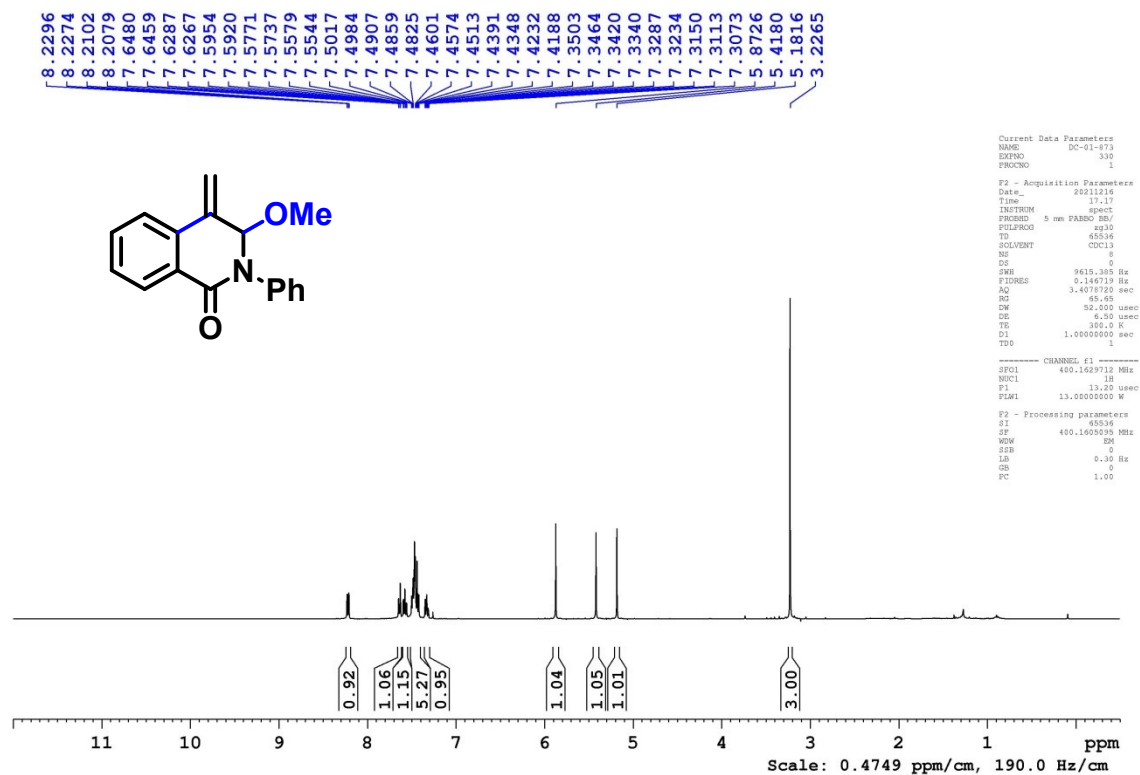


Figure S-82: ^1H NMR (400 MHz, CDCl_3) spectrum of compound **3r**

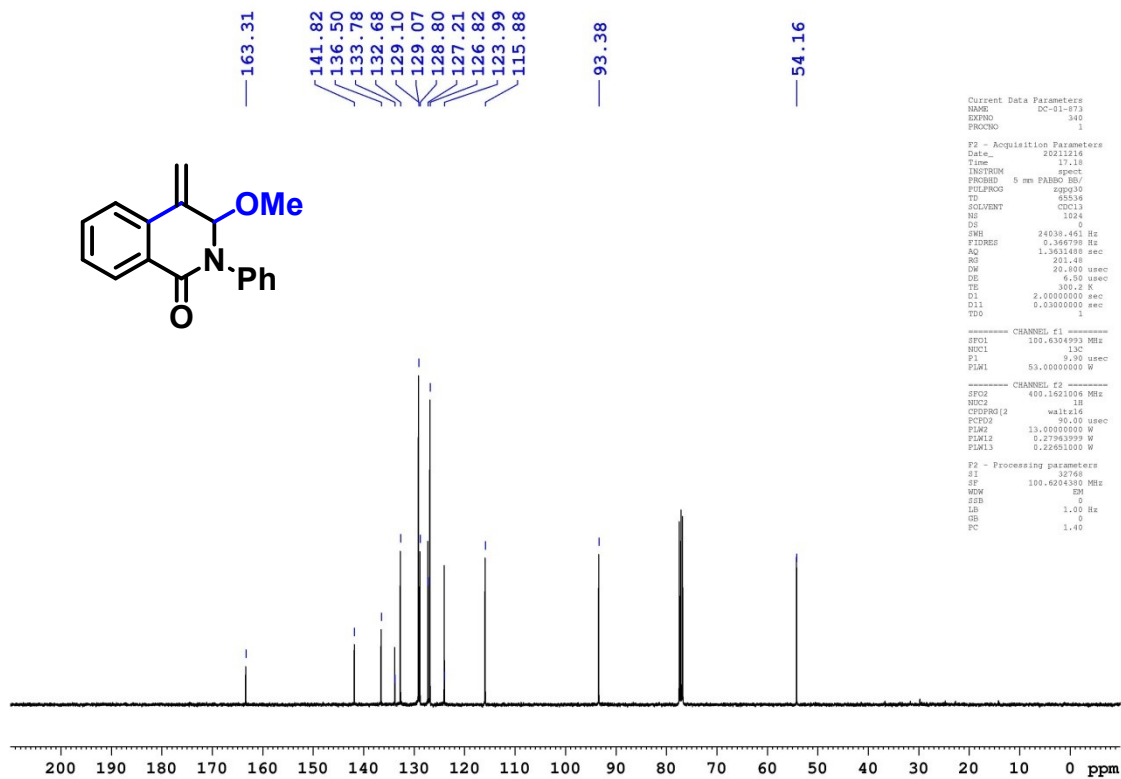


Figure S-83: ^{13}C NMR (100 MHz, CDCl_3) spectrum of compound **3r**

| Sample Name | HRMS22118JAN17 | Position | Vial 17 | Instrument Name | Instrument 1 | User Name | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | DC-01-903.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 1/18/2022 12:31:43 PM |

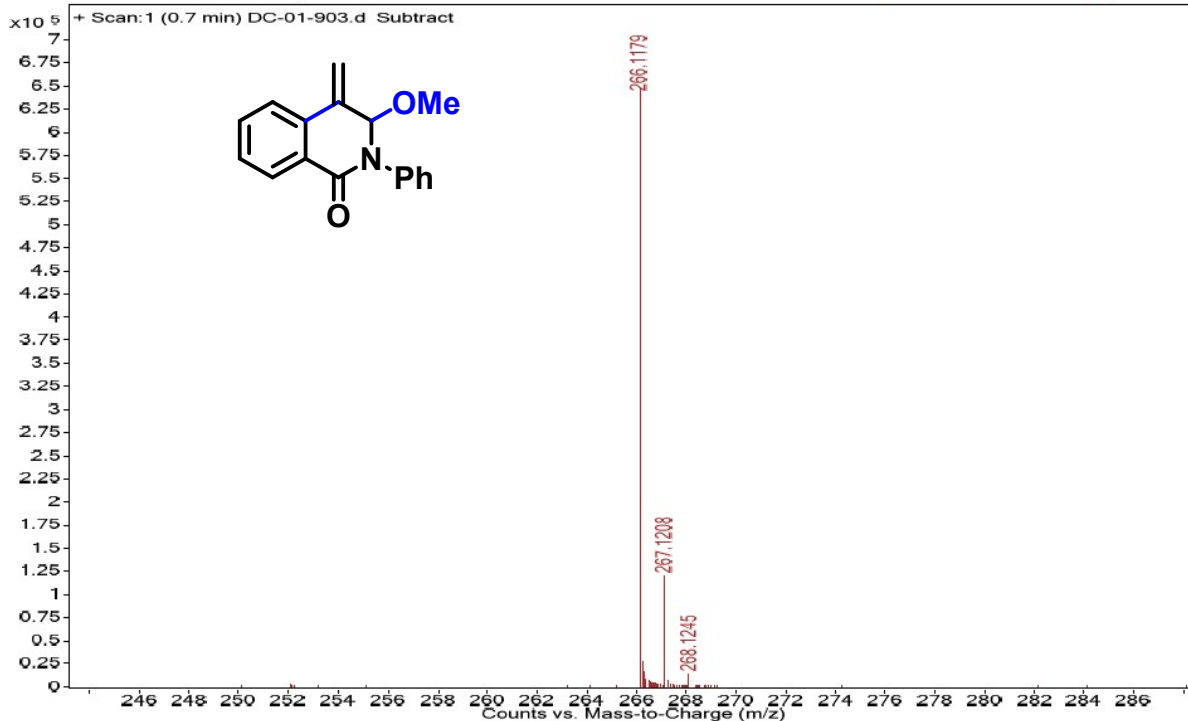


Figure S-84: HRMS spectrum of compound 3r

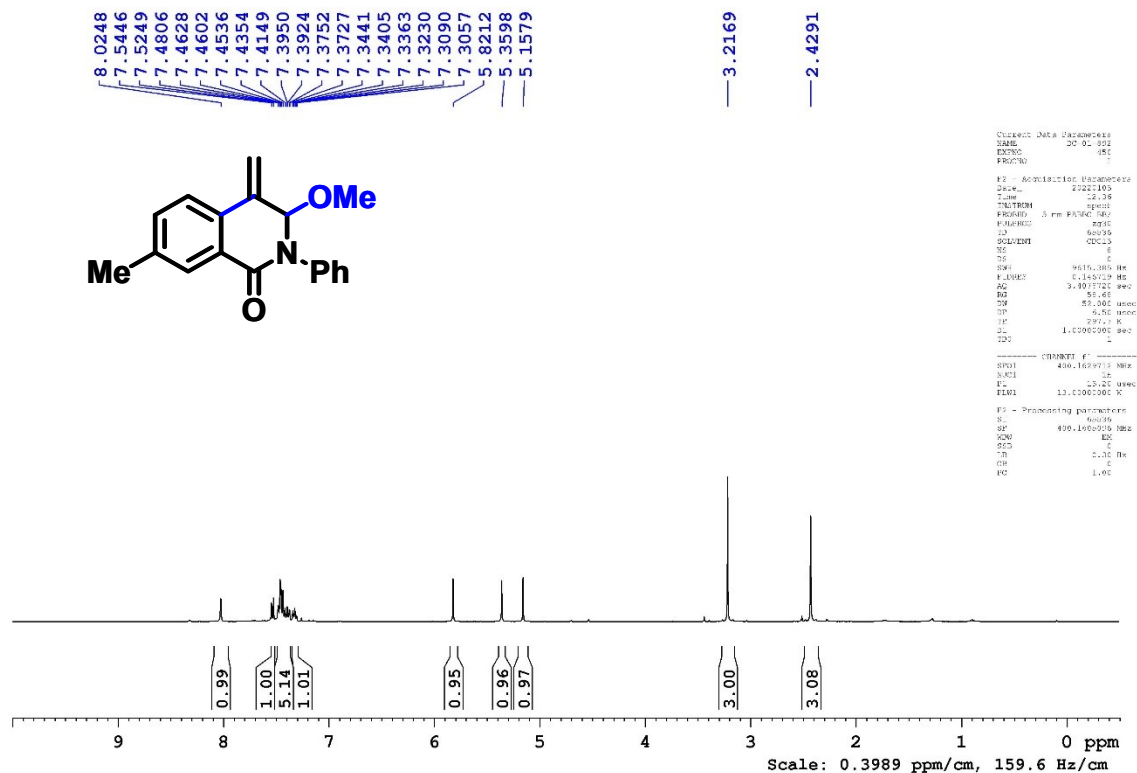


Figure S-85: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3s

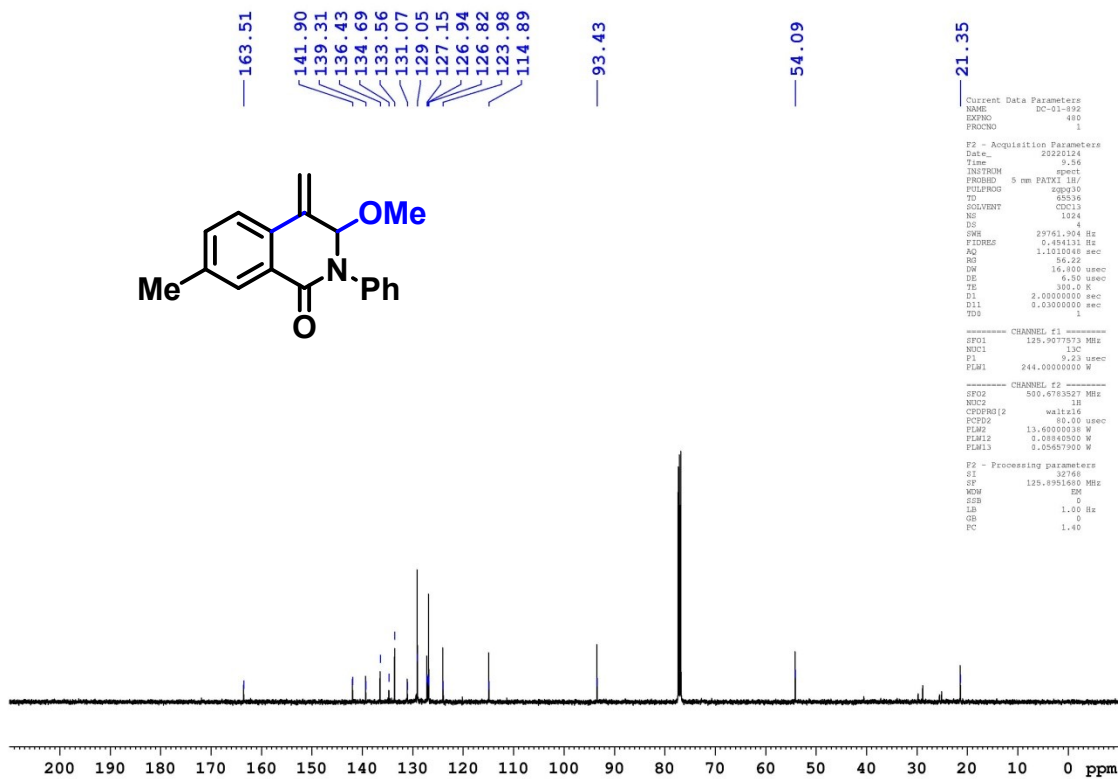


Figure S-86: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 3s

| Sample Name | HRMS22I18JAN18 | Position | Vial 18 | Instrument Name | Instrument 1 | User Name | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | DC-01-898.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 1/18/2022 12:34:51 PM |

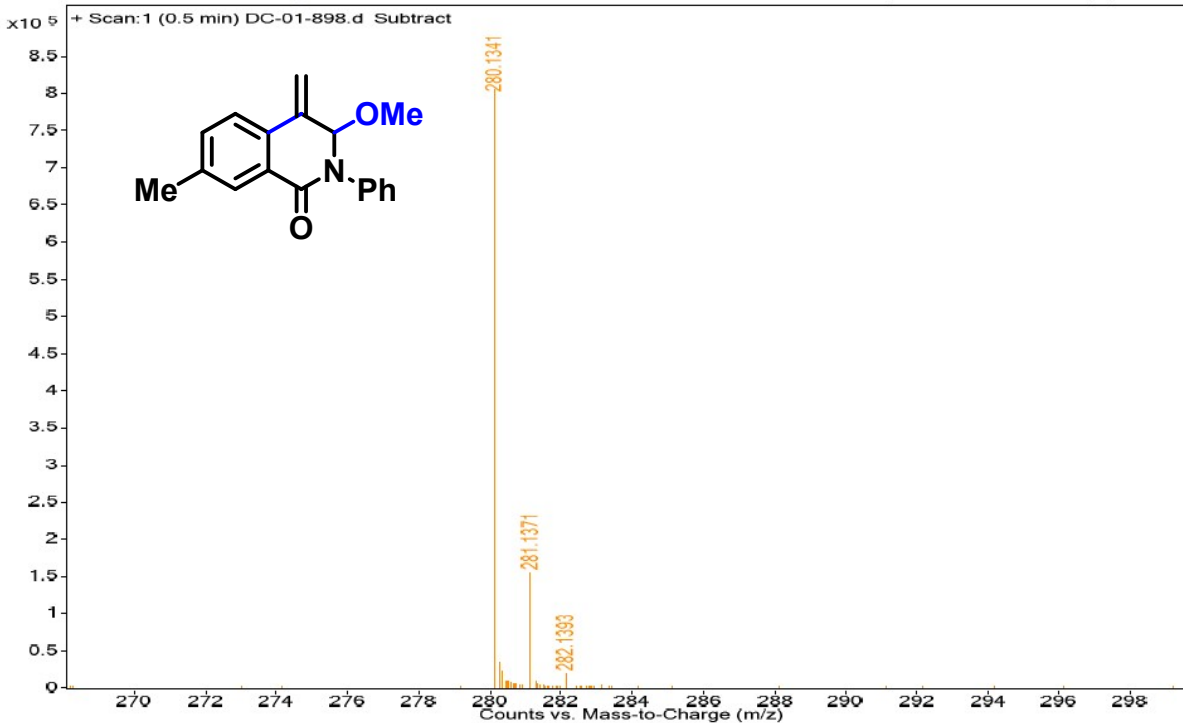


Figure S-87: HRMS spectrum of compound 3s

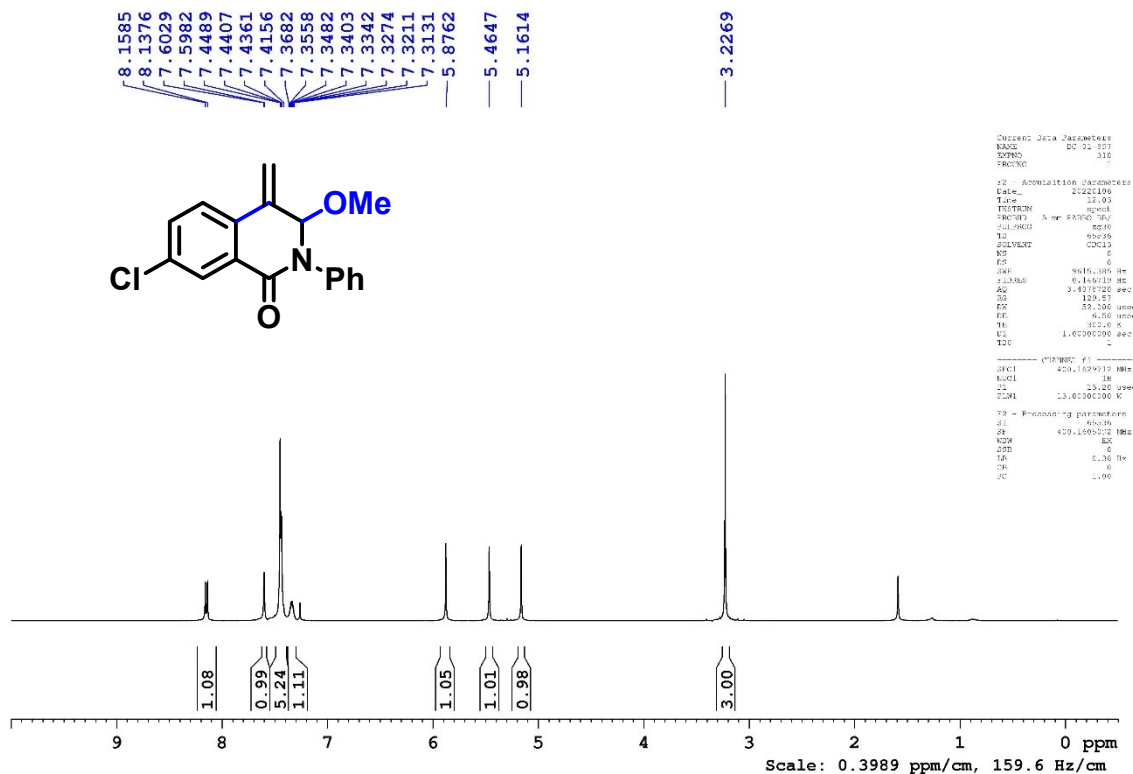


Figure S-88: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3t

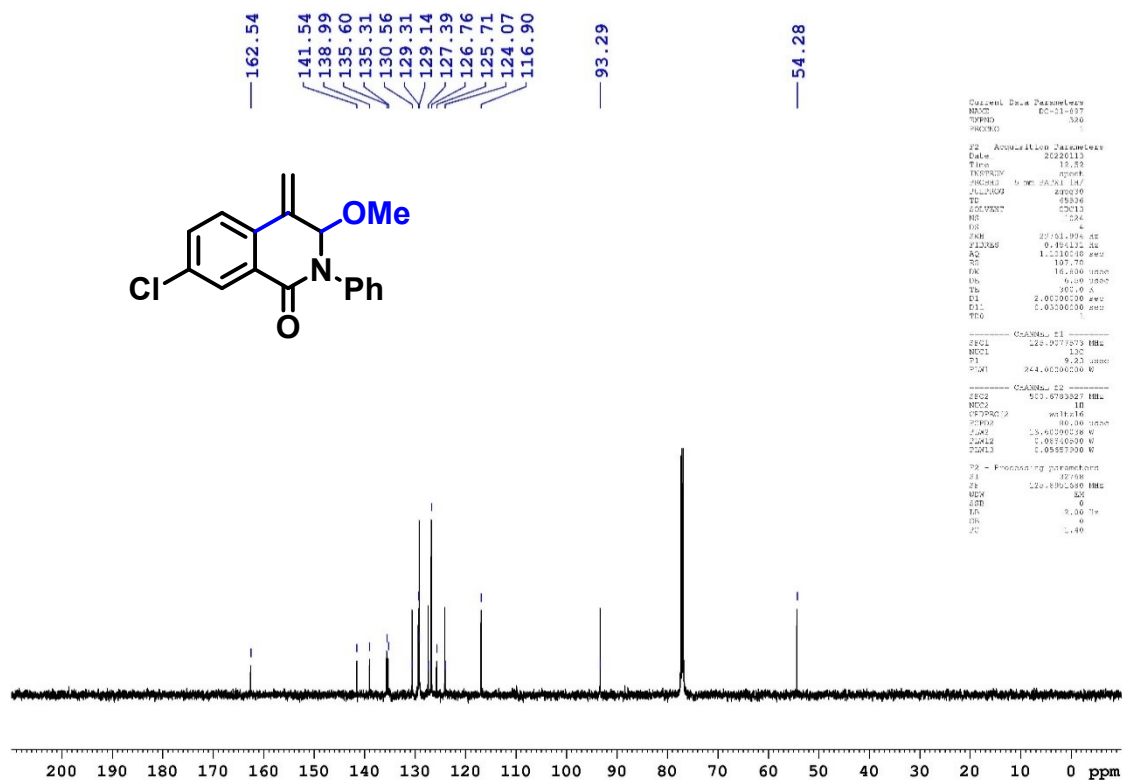


Figure S-89: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3t

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Sample Name | HRMS22119JAN11 | Position | Vial 11 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | SD-01-763.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 1/19/2022 12:03:46 PM |

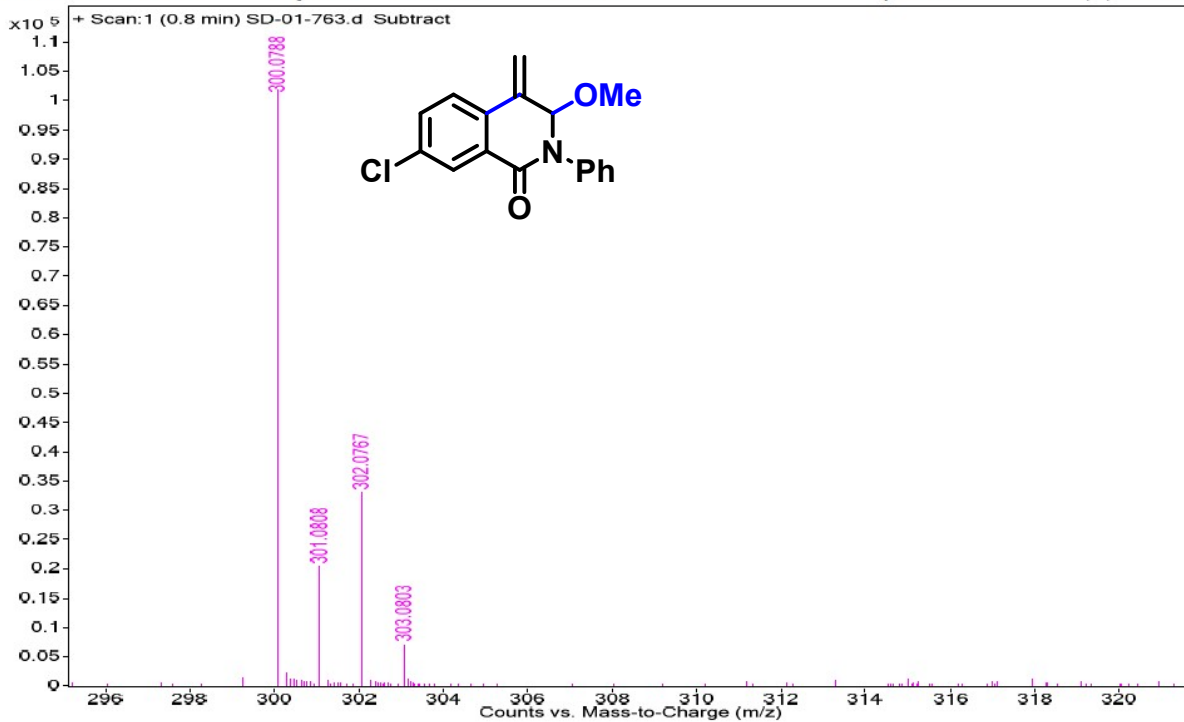


Figure S-90: HRMS spectrum of compound 3t

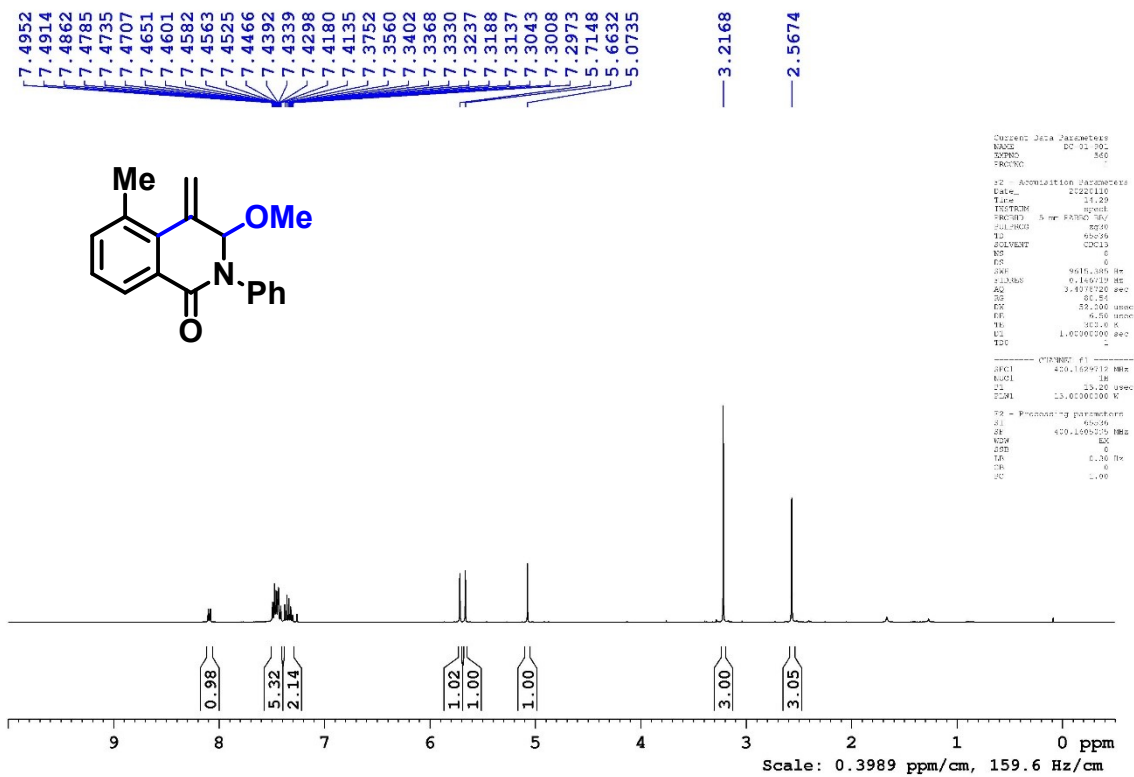


Figure S-91: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3u

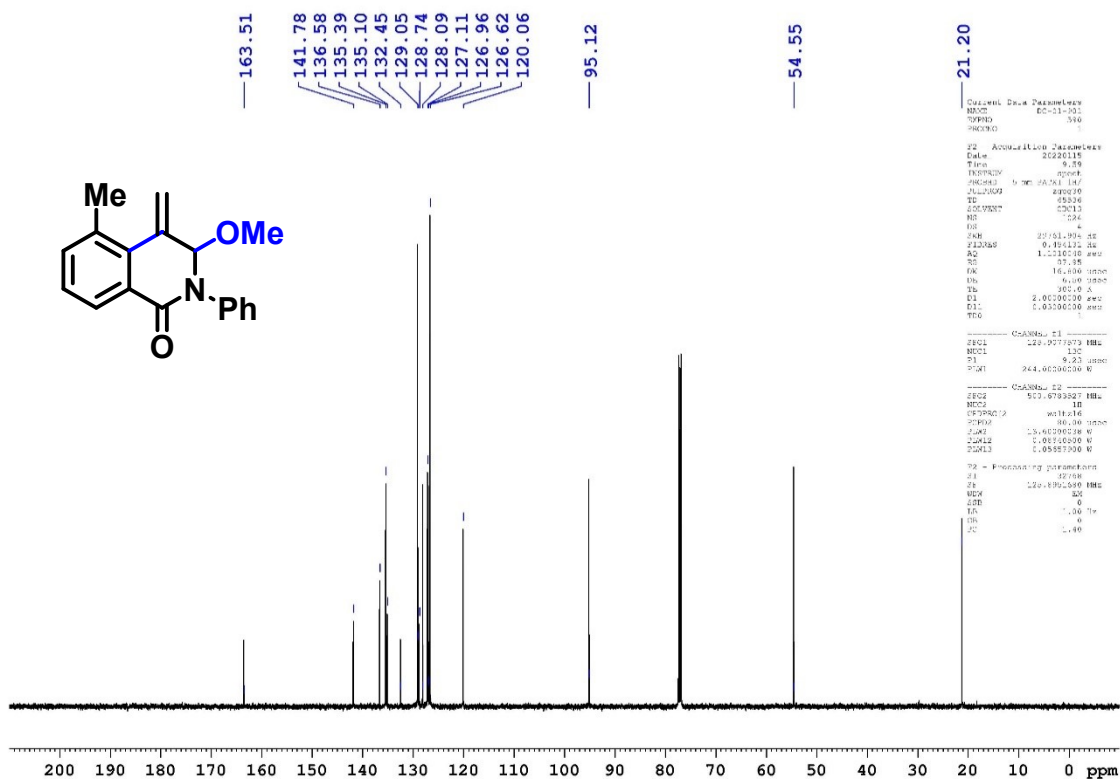


Figure S-92: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 3u

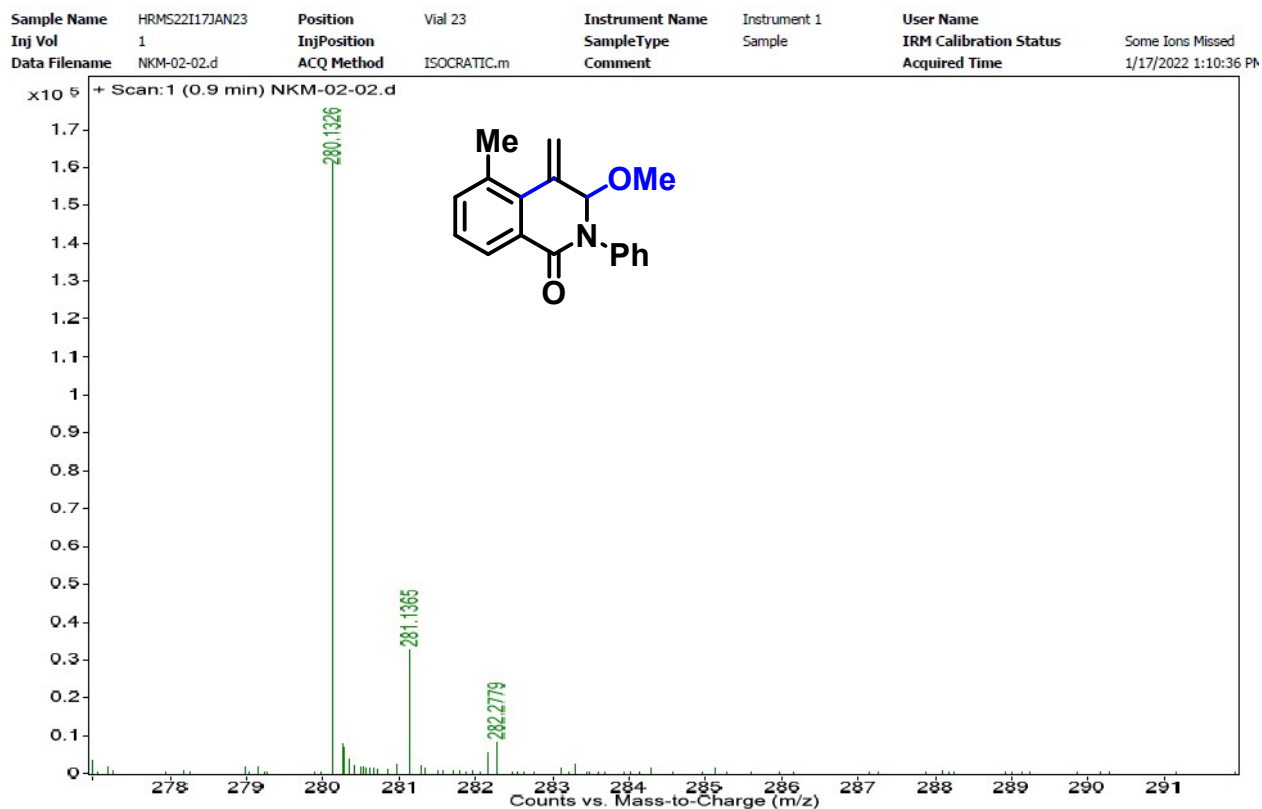


Figure S-93: HRMS spectrum of compound 3u

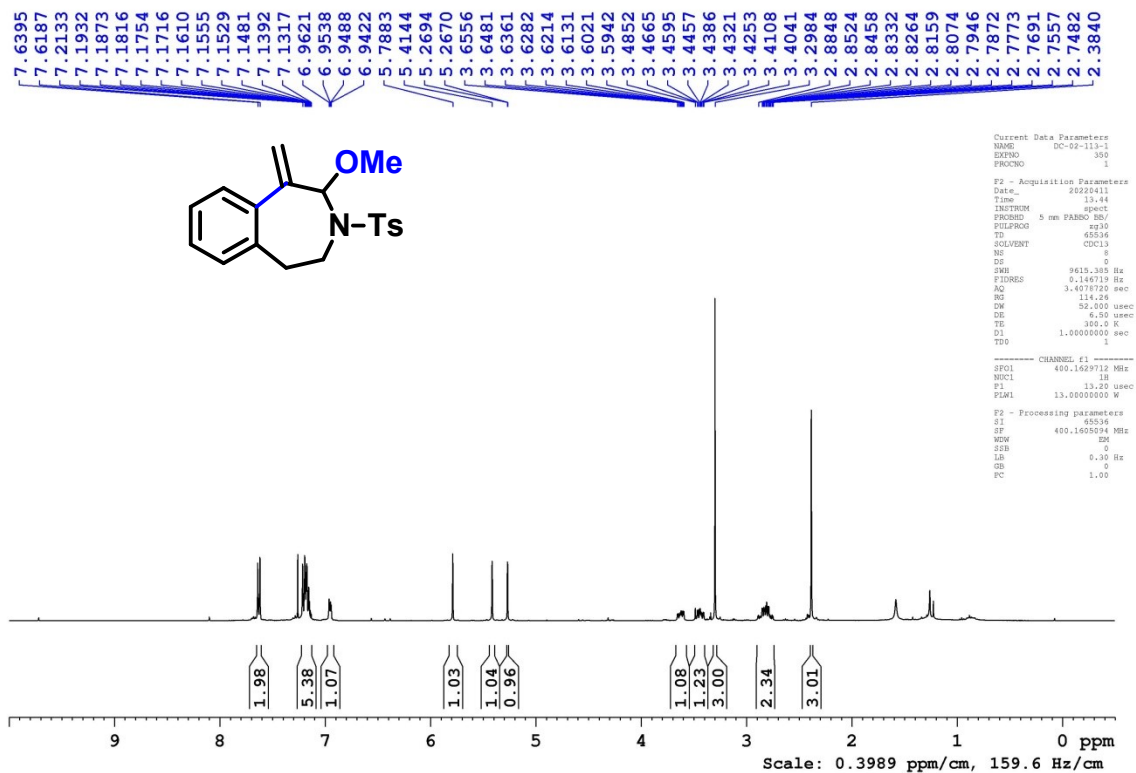


Figure S-94: ^1H NMR (400 MHz, CDCl_3) spectrum of compound 3v

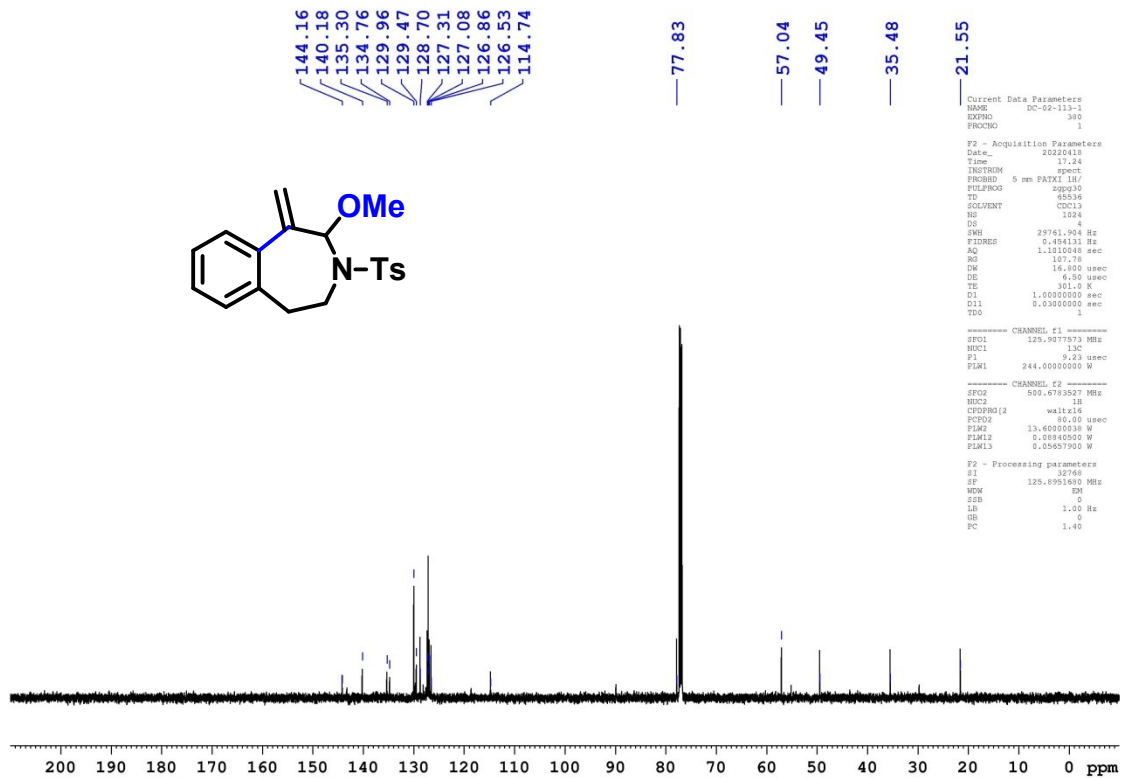


Figure S-95: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 3v

Sample Name NKM-02-95 Position Vial 7 Instrument Name Instrument 1 User Name
 Inj Vol 1 InjPosition SampleType Sample IRM Calibration Status Some Ions Missed
 Data Filename HRMS22112APR07.d ACQ Method ISOCRATIC.m Comment Acquired Time 4/12/2022 12:54:18 PM

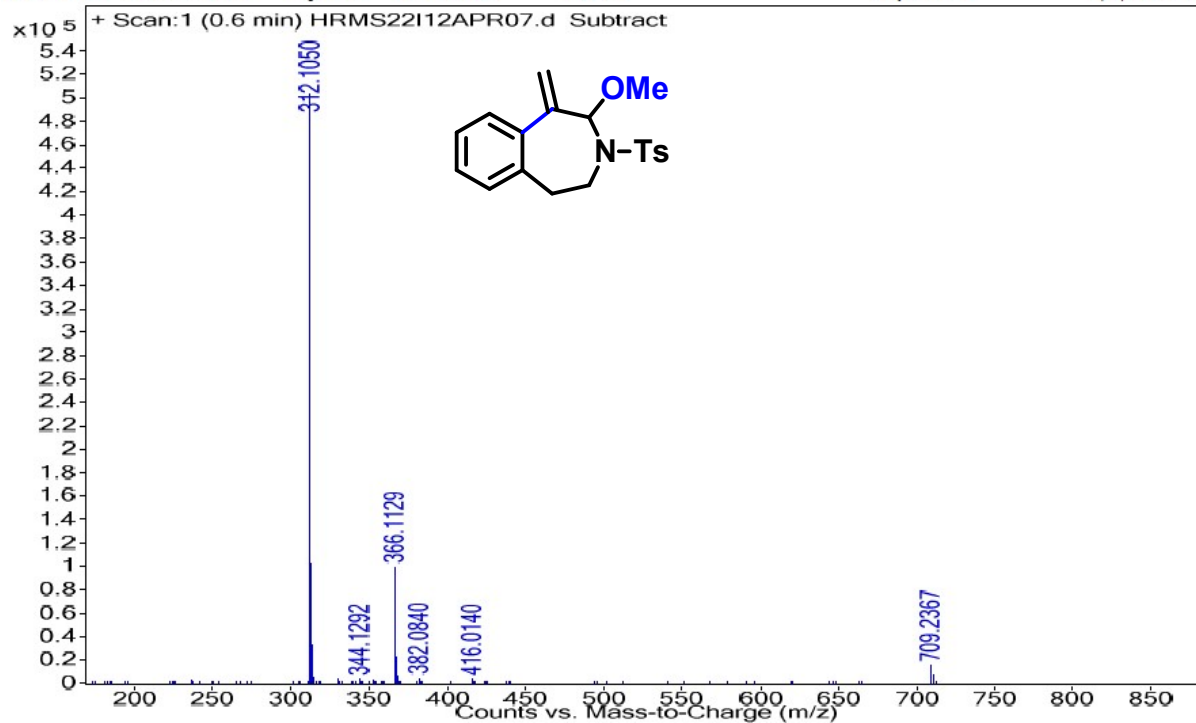


Figure S-96: HRMS spectrum of compound 3v

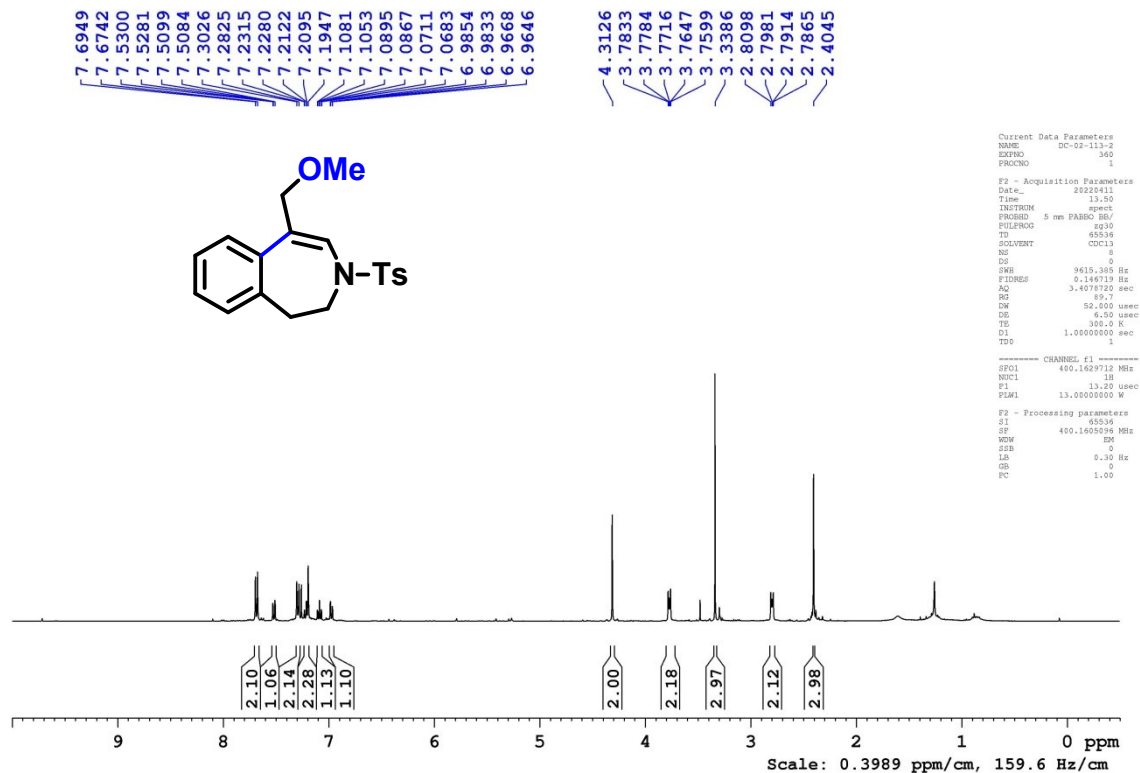


Figure S-97: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 4v

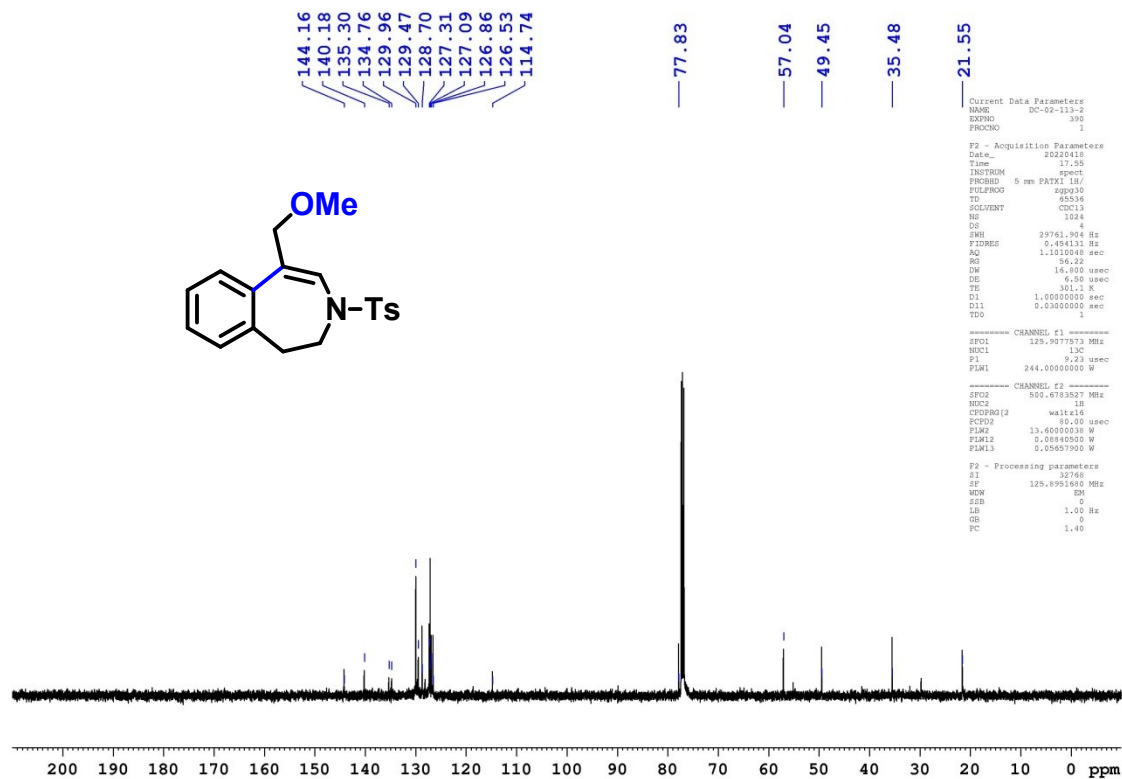


Figure S-98: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 4v

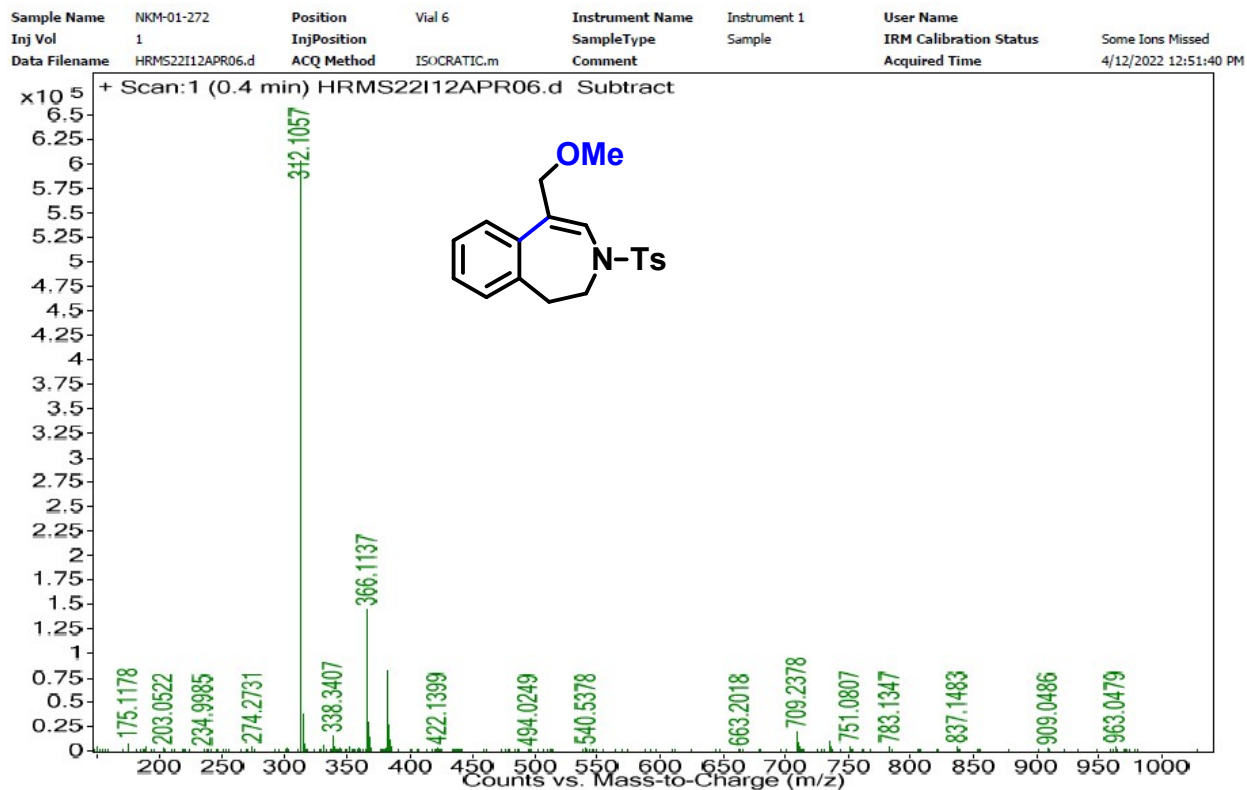


Figure S-99: HRMS spectrum of compound 4v

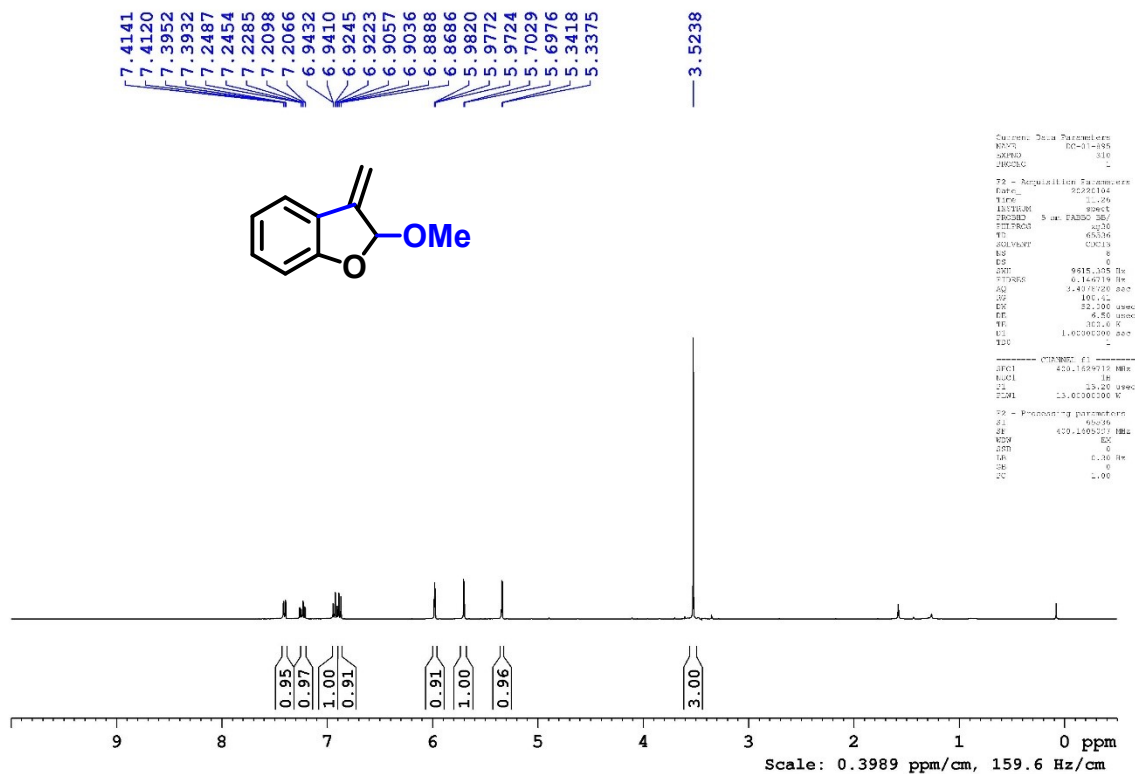


Figure S-100: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3w

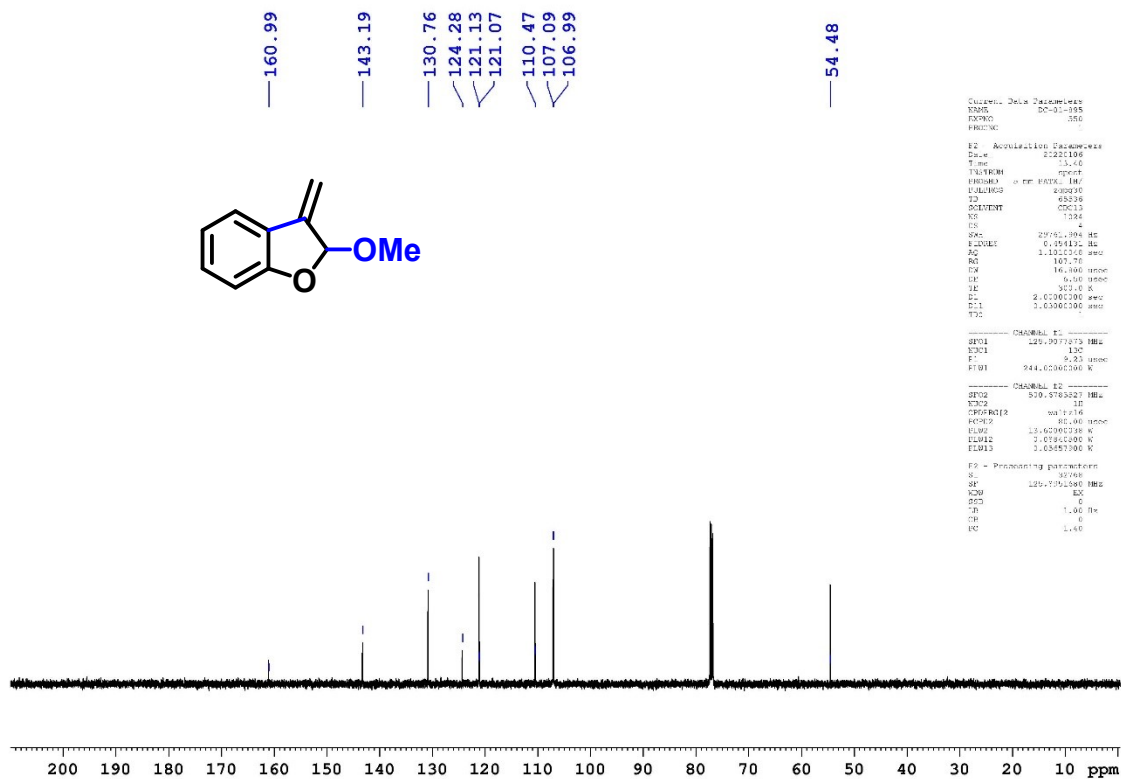


Figure S-101: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3w

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|---------------------|
| Sample Name | HRMS22103FEB15 | Position | Vial 15 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | NKM-01-257.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/3/2022 2:15:28 PM |

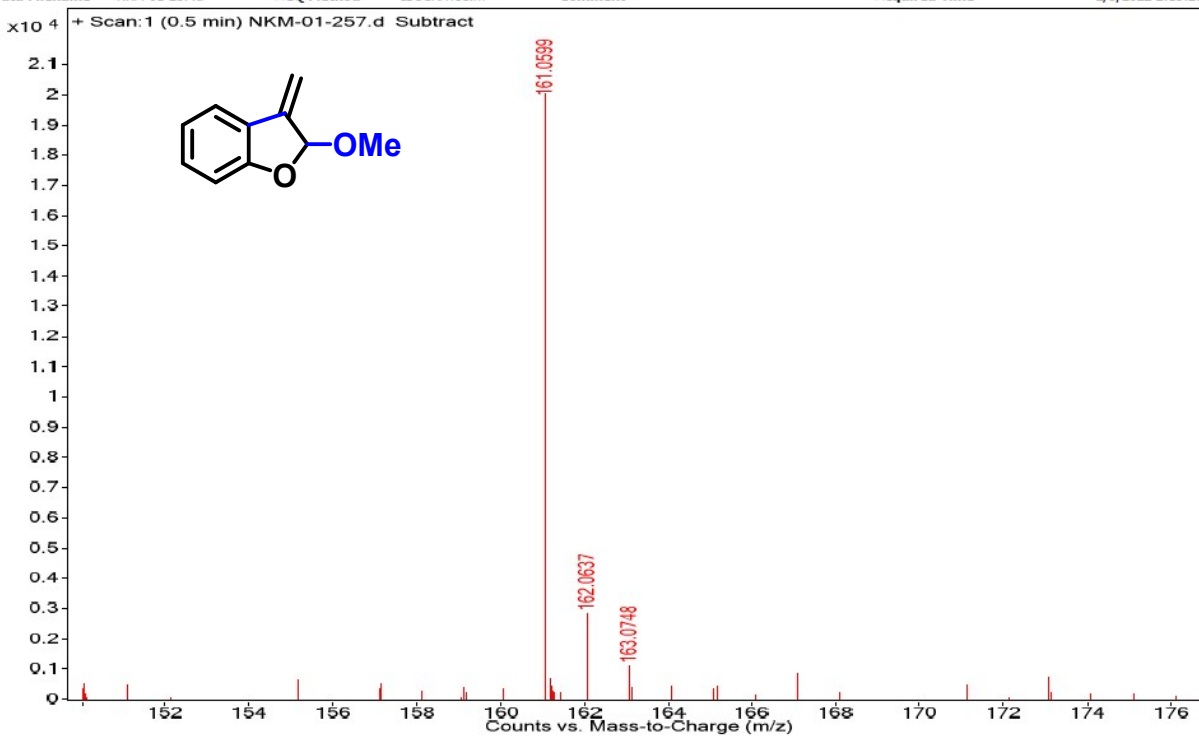


Figure S-102: HRMS spectrum of compound 3w

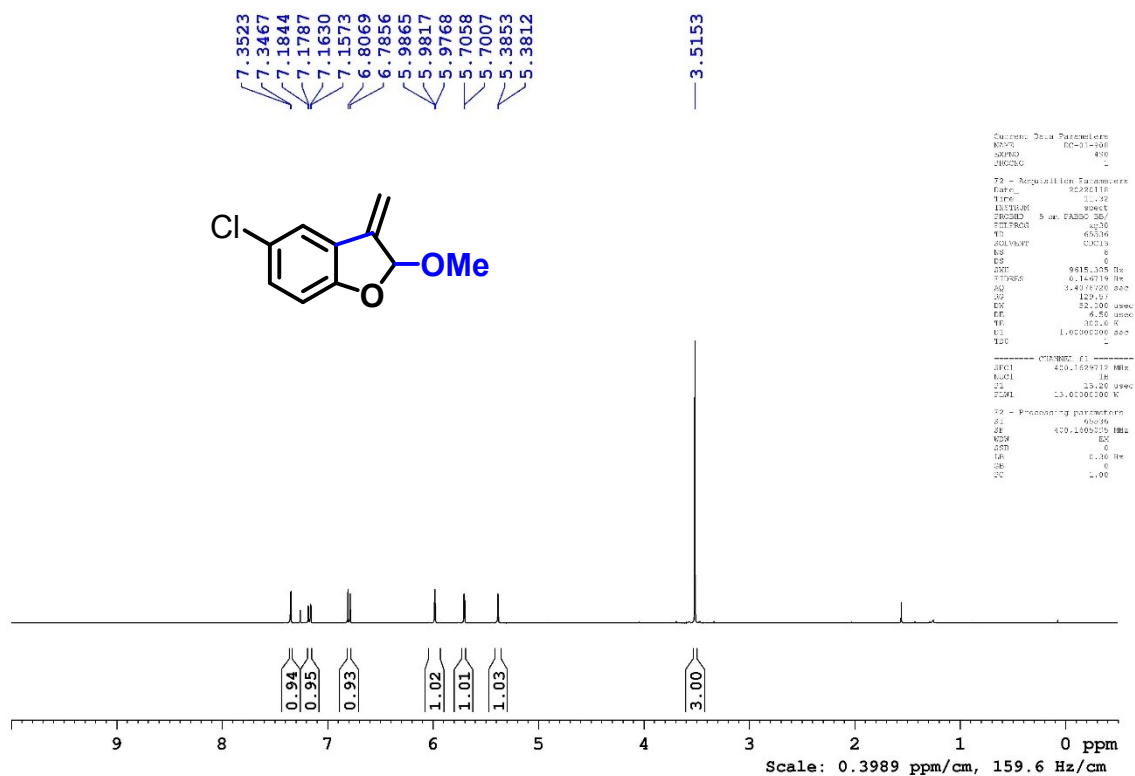


Figure S-103: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3x

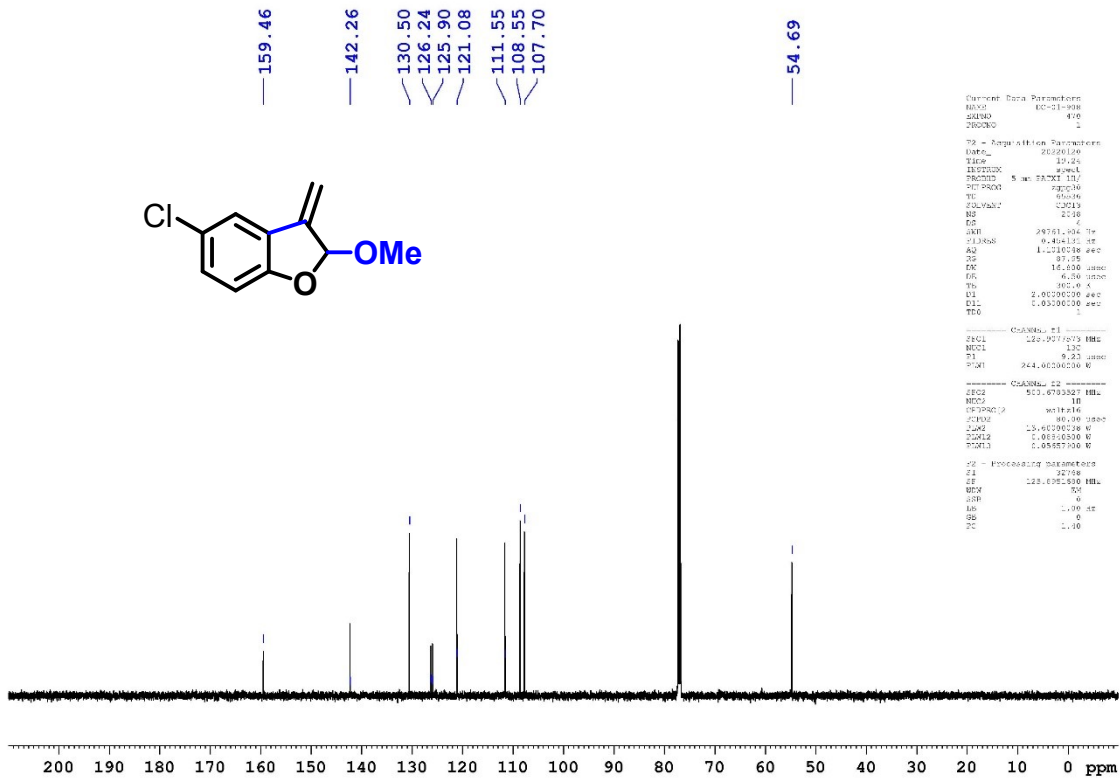


Figure S-104: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3x

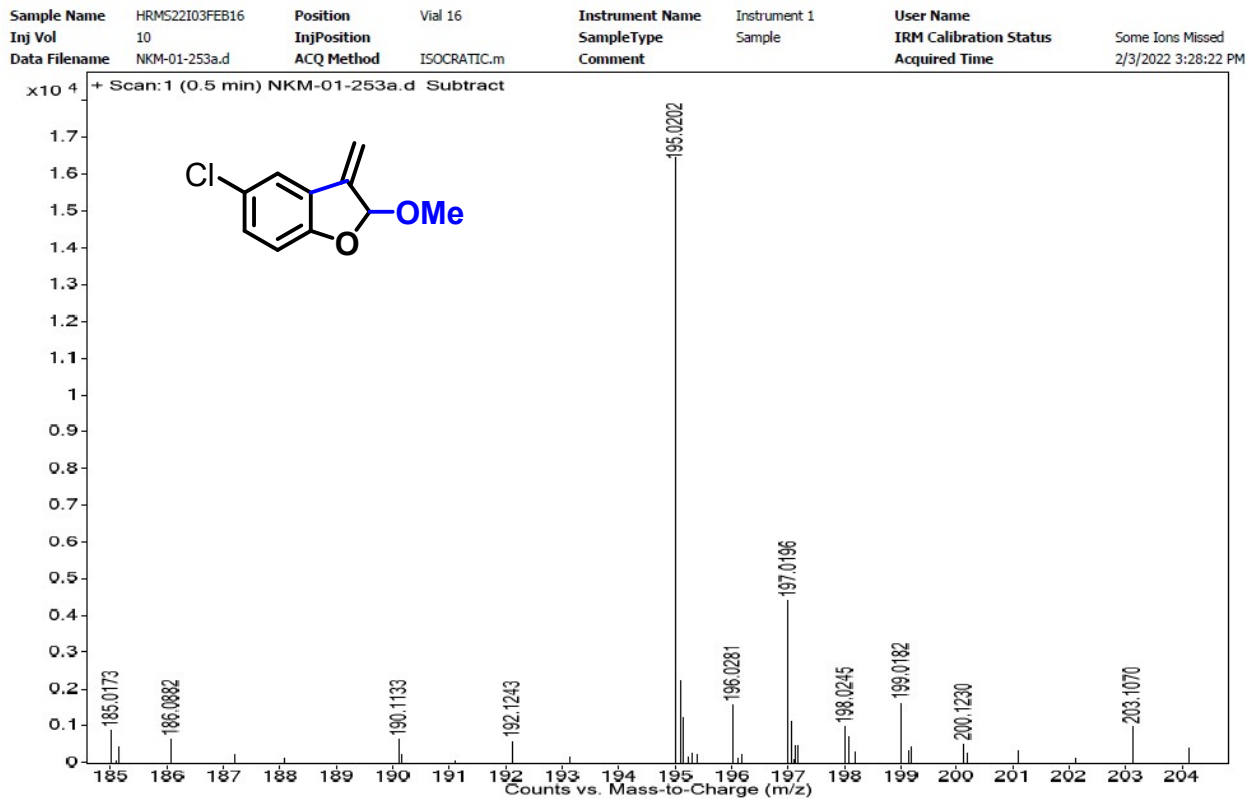


Figure S-105: HRMS spectrum of compound 3x

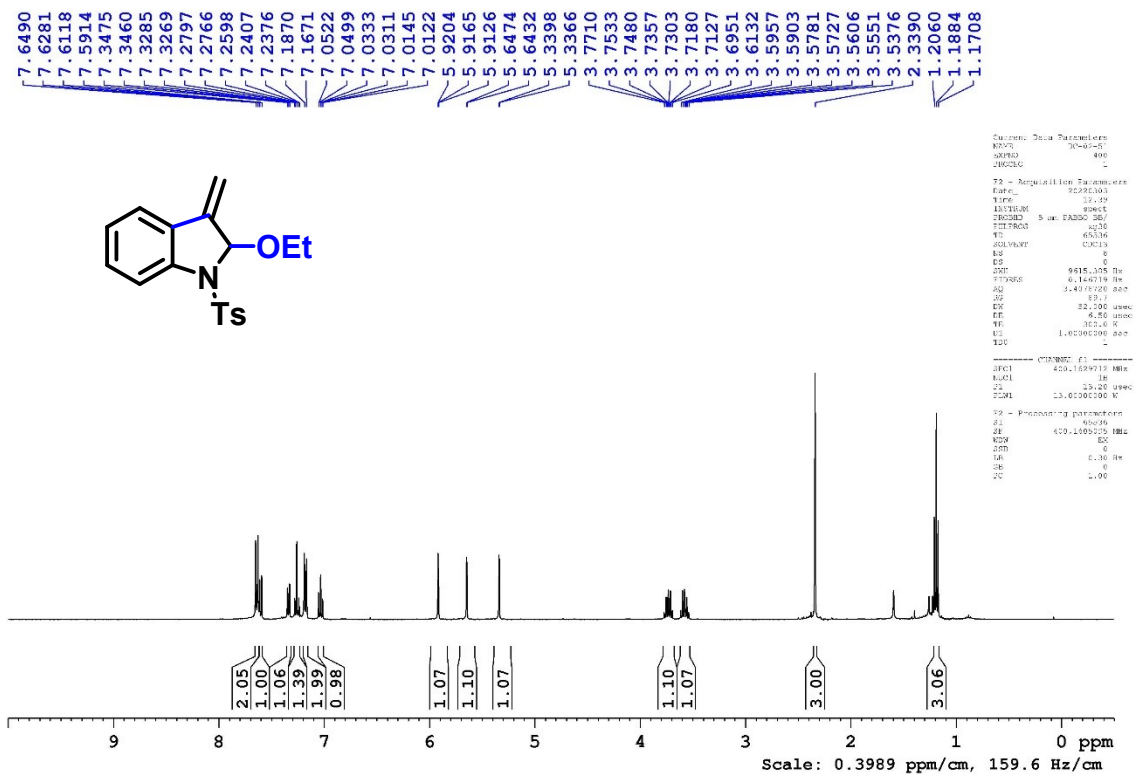


Figure S-106: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3aa

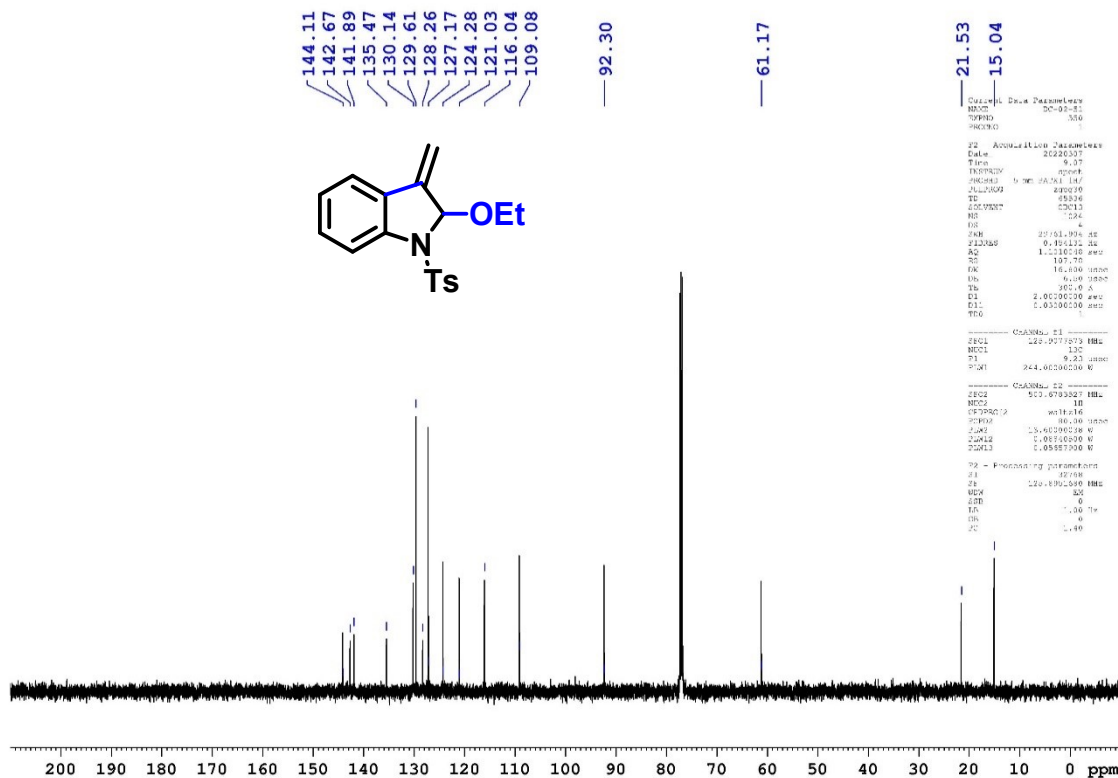


Figure S-107: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 3aa

| Sample Name | HRMS22107MAR28 | Position | Vial 28 | Instrument Name | Instrument 1 | User Name | IRM Calibration Status | Some Ions Missed |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|------------------------|---------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | | 3/7/2022 1:06:35 PM |
| Data Filename | DC-01-415.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | | |

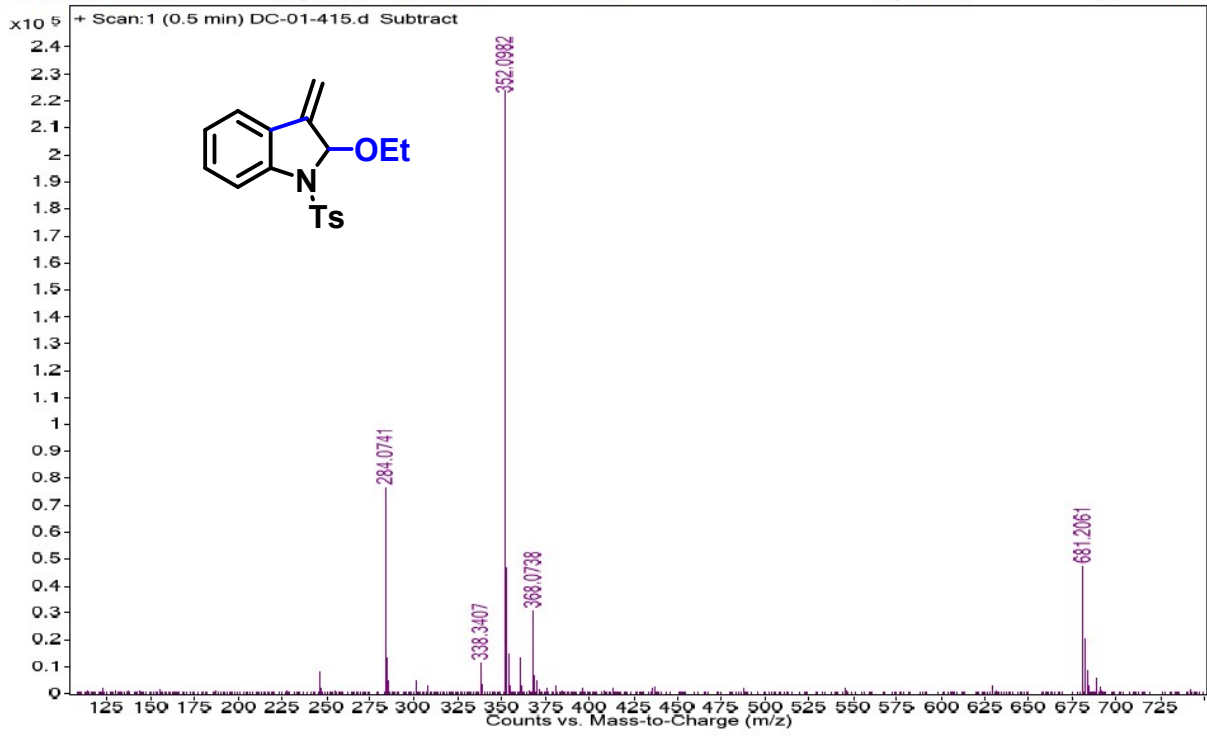


Figure S-108: HRMS spectrum of compound 3aa

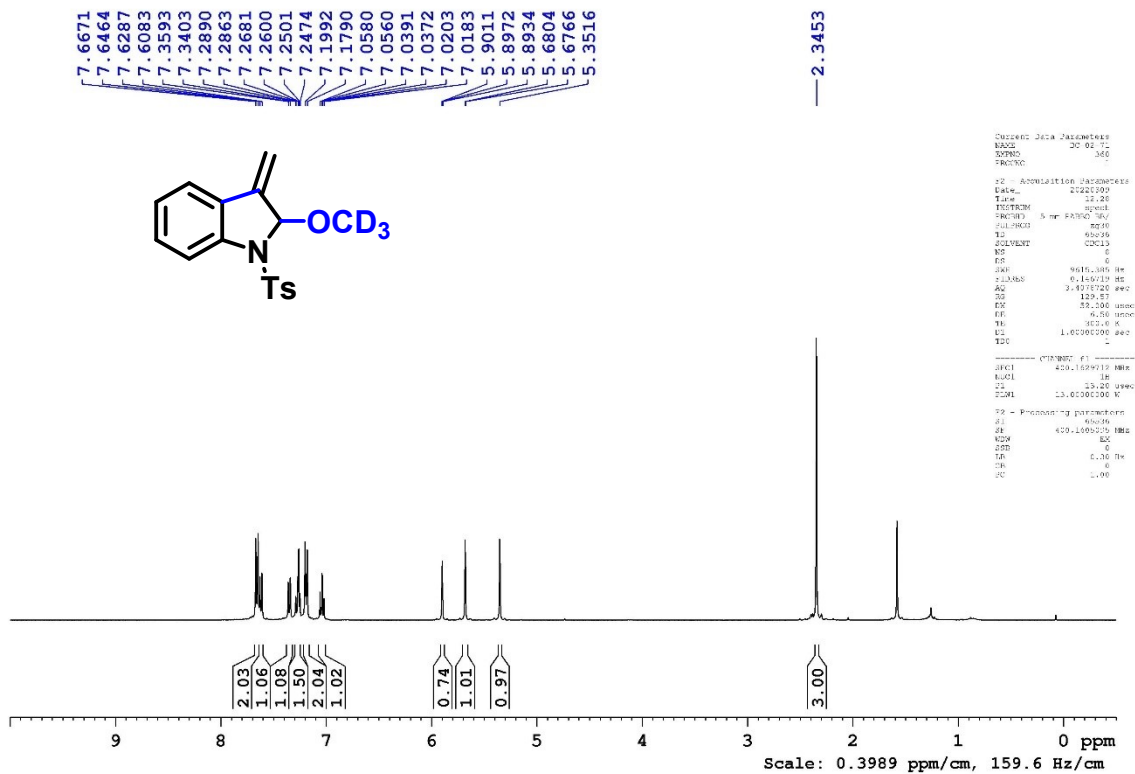


Figure S-109: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3a-D

| Sample Name | HRMS22110MAR25 | Position | Vial 25 | Instrument Name | Instrument 1 | User Name | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | DC-01-286.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 3/10/2022 12:42:40 PM |

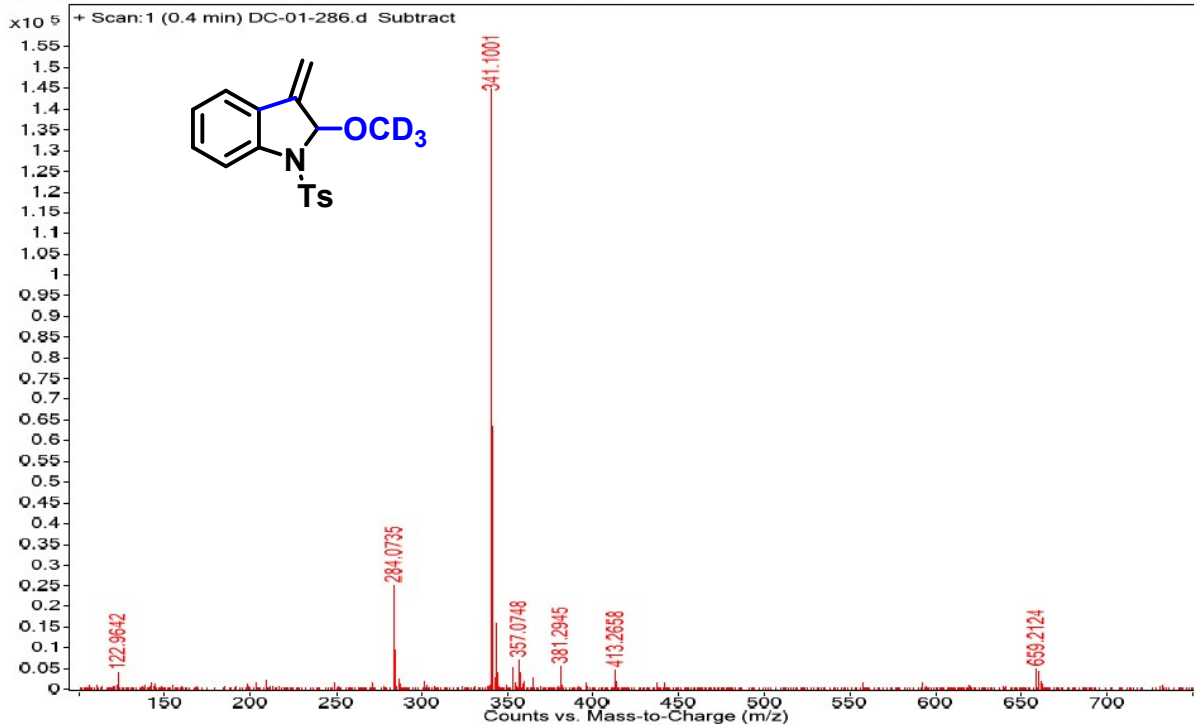


Figure S-110: HRMS spectrum of compound 3a-D

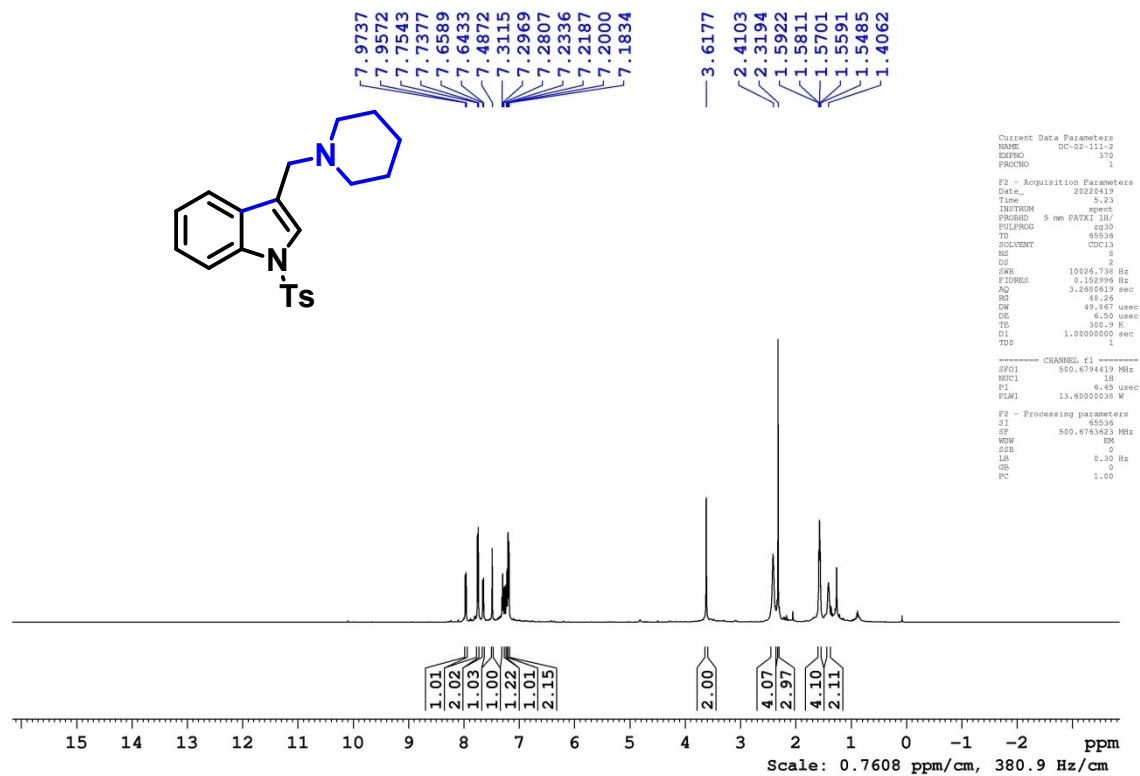


Figure S-111: ¹H NMR (500 MHz, CDCl₃) spectrum of compound 4ad

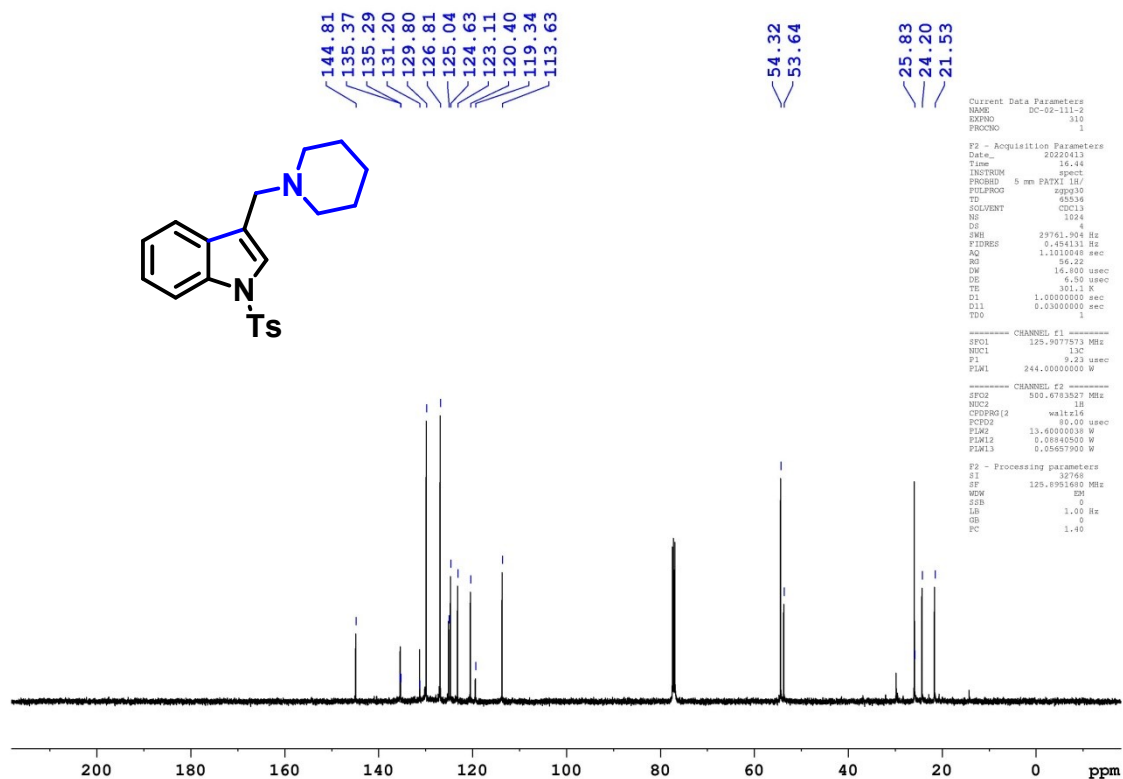


Figure S-112: ^{13}C NMR (125 MHz, CDCl_3) spectrum of compound 4ad

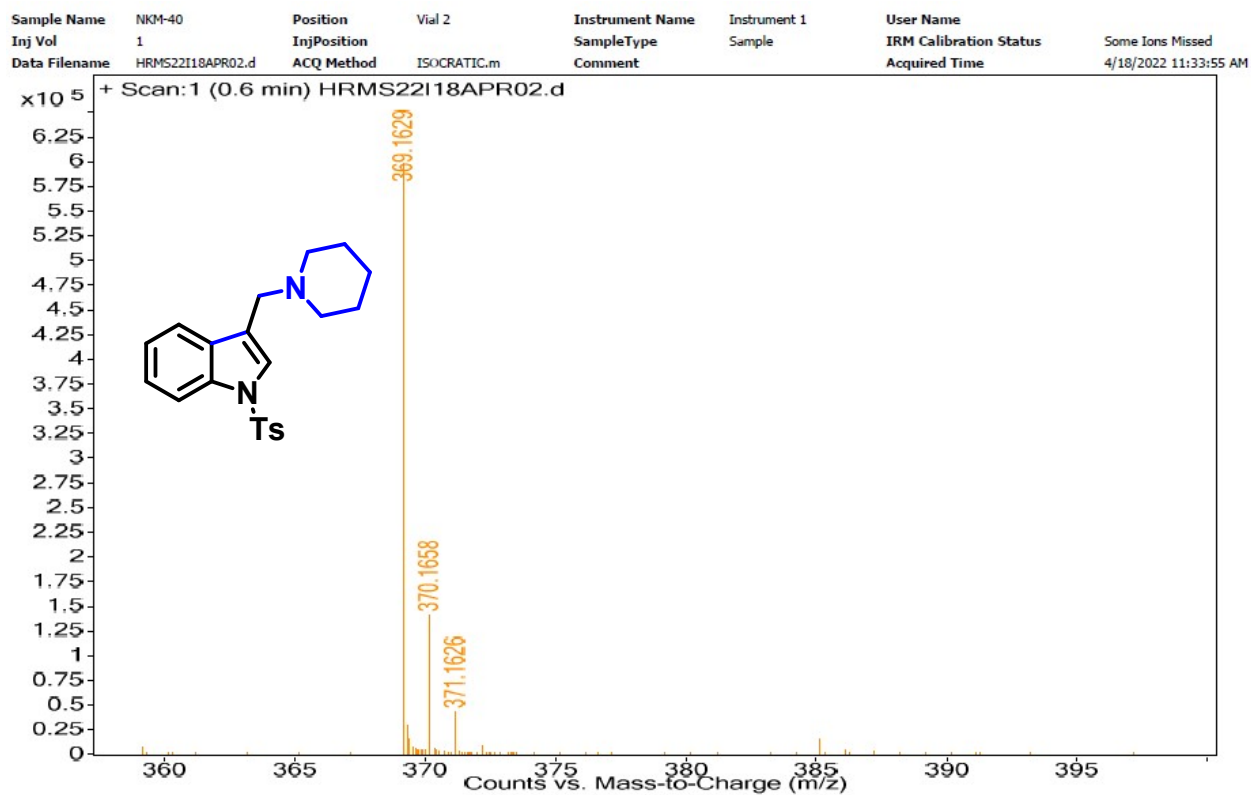


Figure S-113: HRMS spectrum of compound 4ad

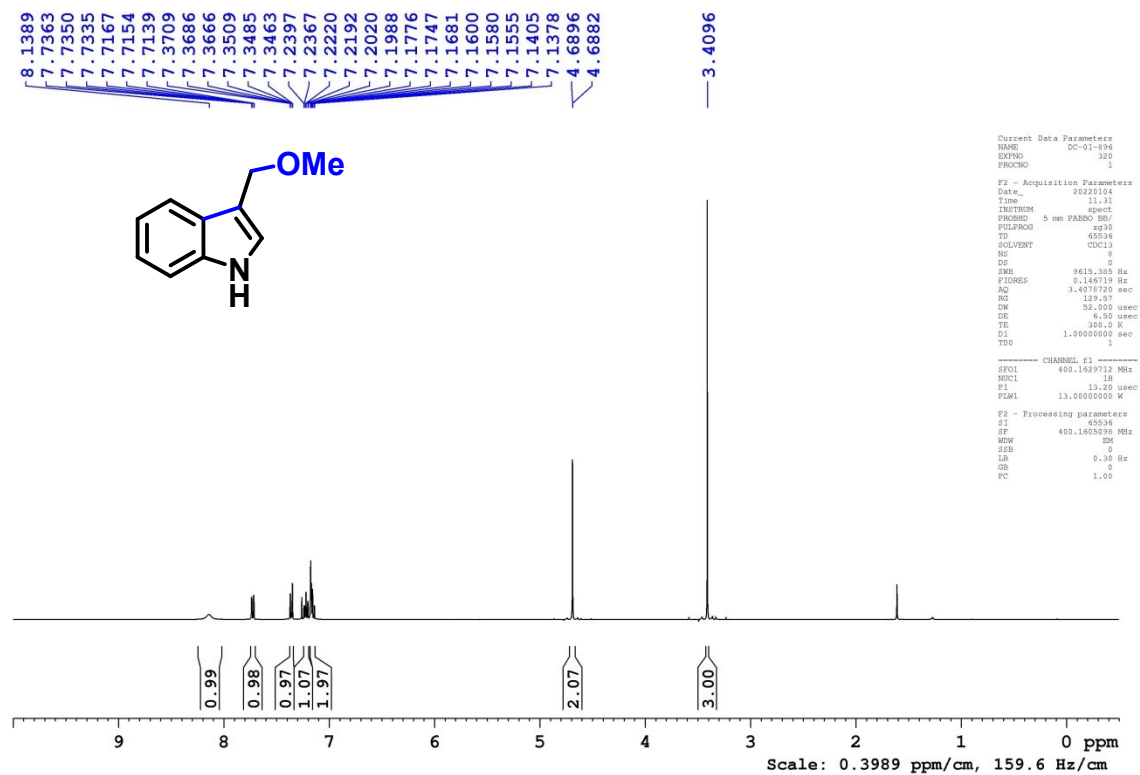


Figure S-114: ^1H NMR (400 MHz, CDCl_3) spectrum of compound **4a**

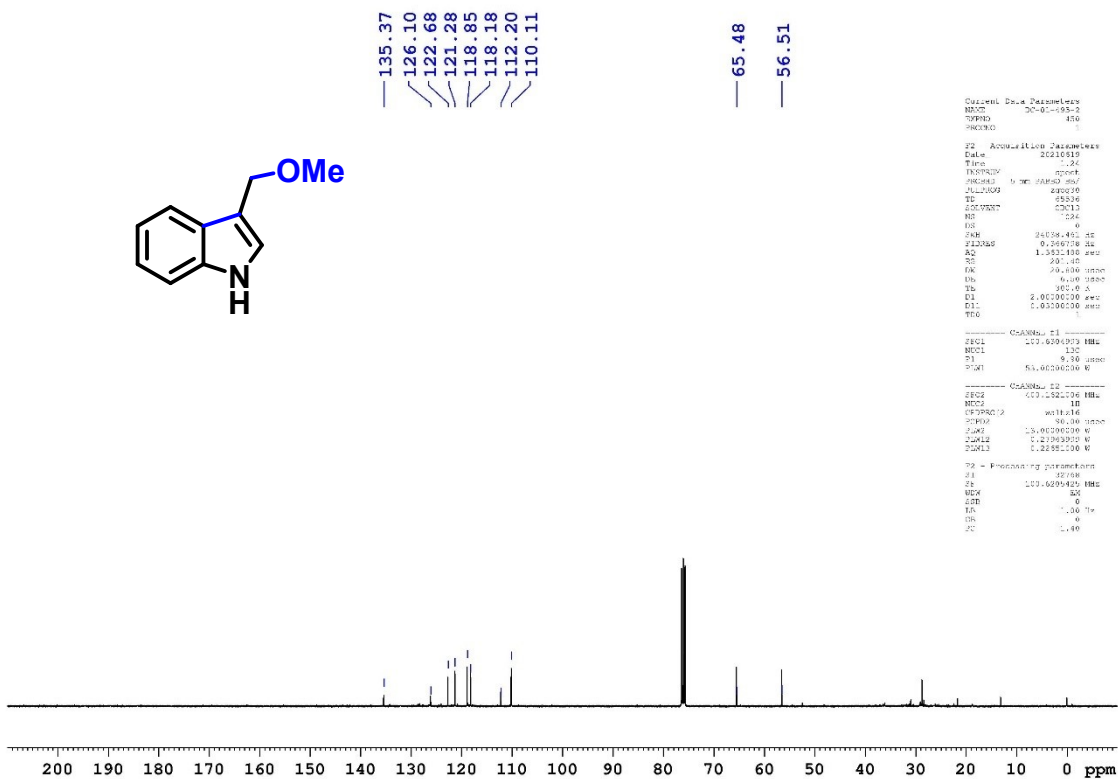


Figure S-115: ^{13}C NMR (100 MHz, CDCl_3) spectrum of compound **4a**

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|---------------------|
| Sample Name | HRMS22107FEB19 | Position | Vial 19 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 5 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | KMR-02-93A6a.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/7/2022 4:20:43 PM |

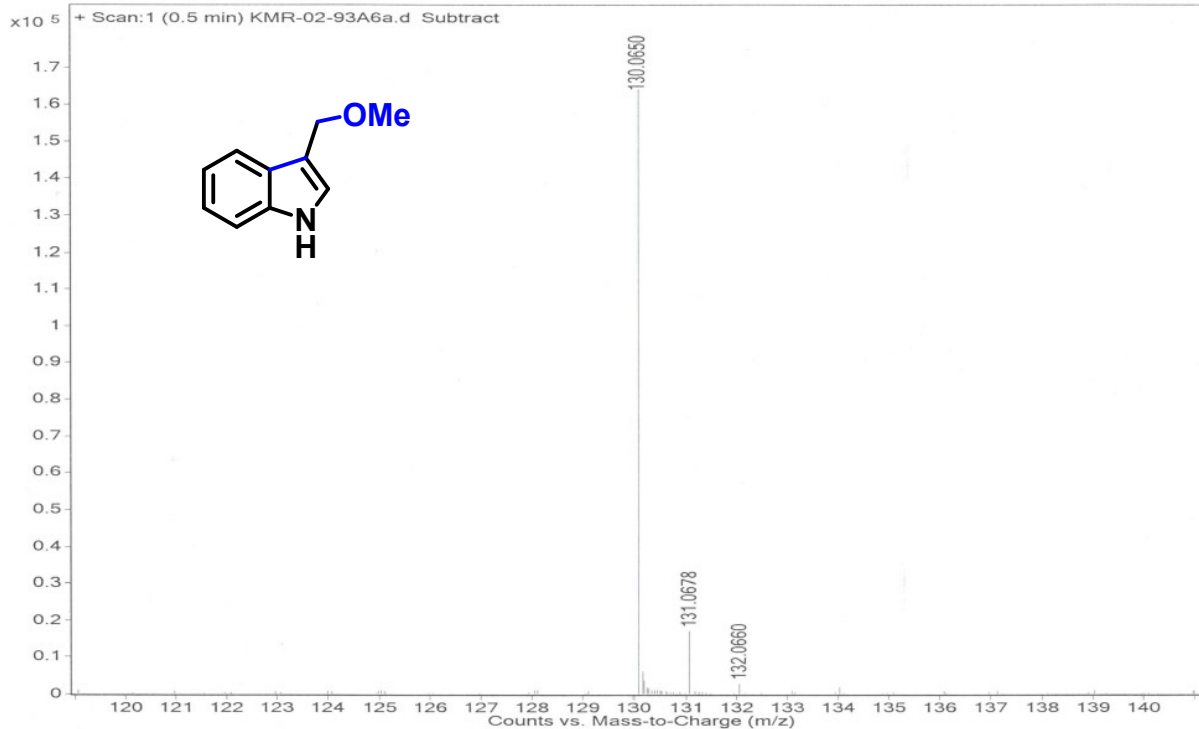


Figure S-116: HRMS spectrum of compound **4a**

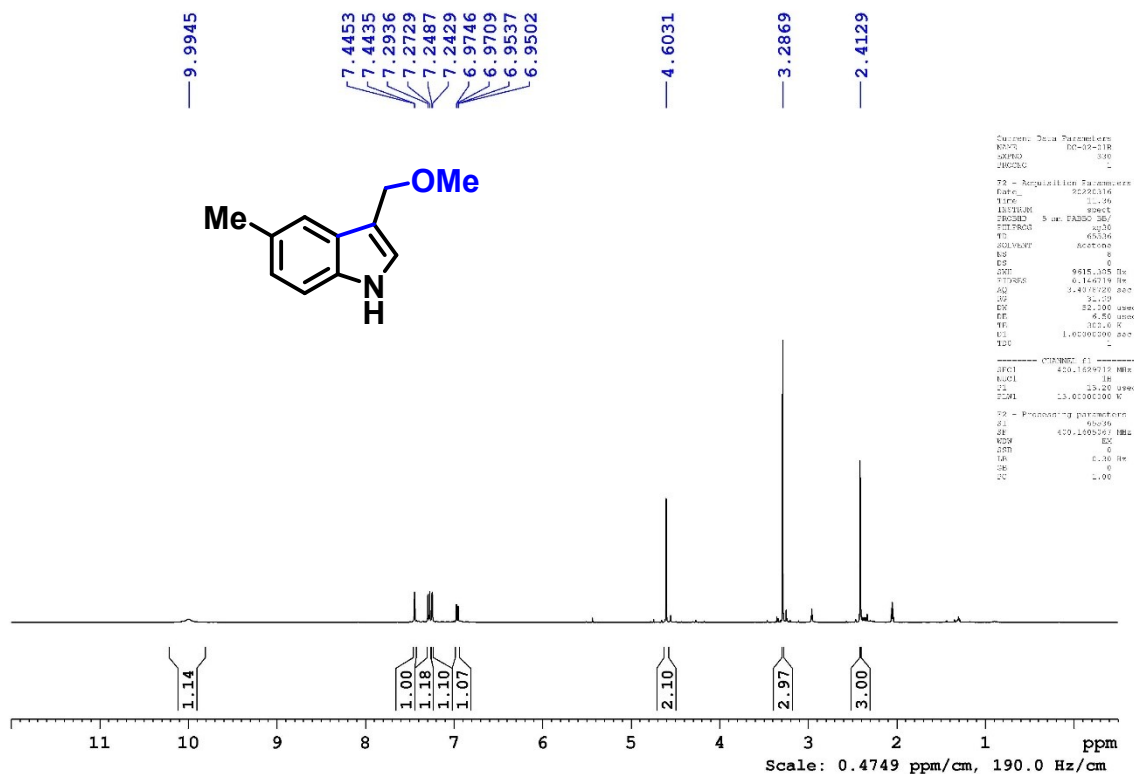


Figure S-117: ¹H NMR (400 MHz, D₆-acetone) spectrum of compound **4b**

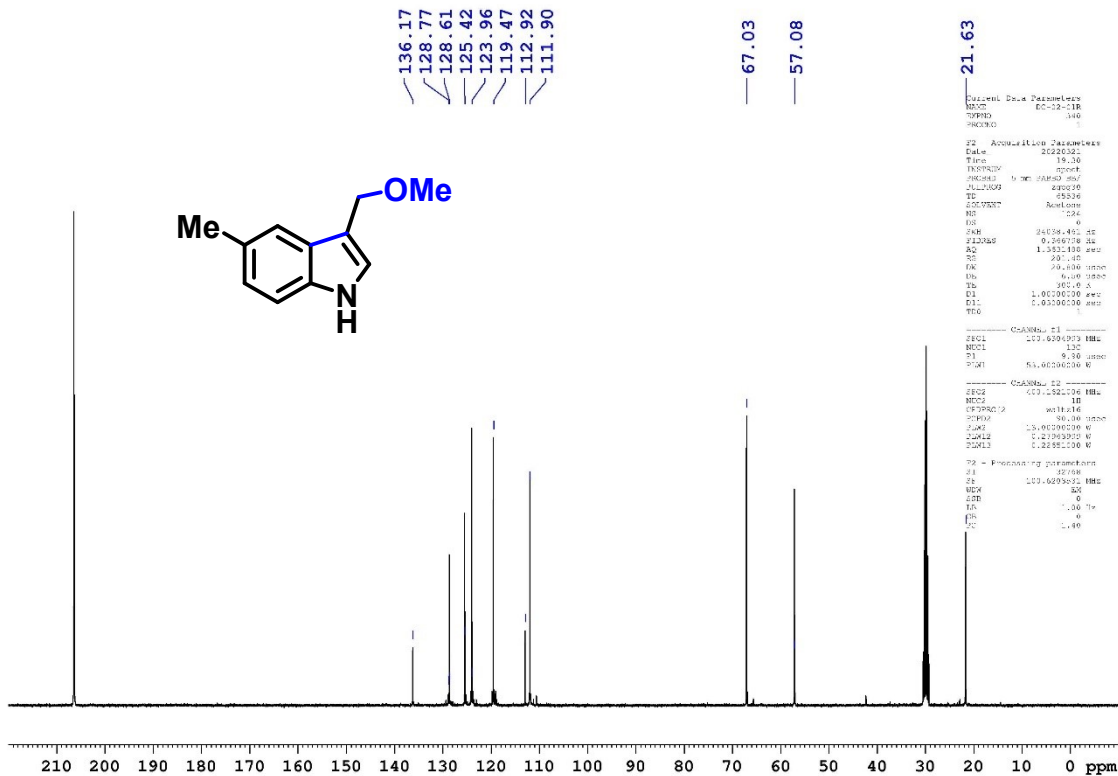


Figure S-118: ¹³C NMR (100 MHz, D₆-acetone) spectrum of compound 4b

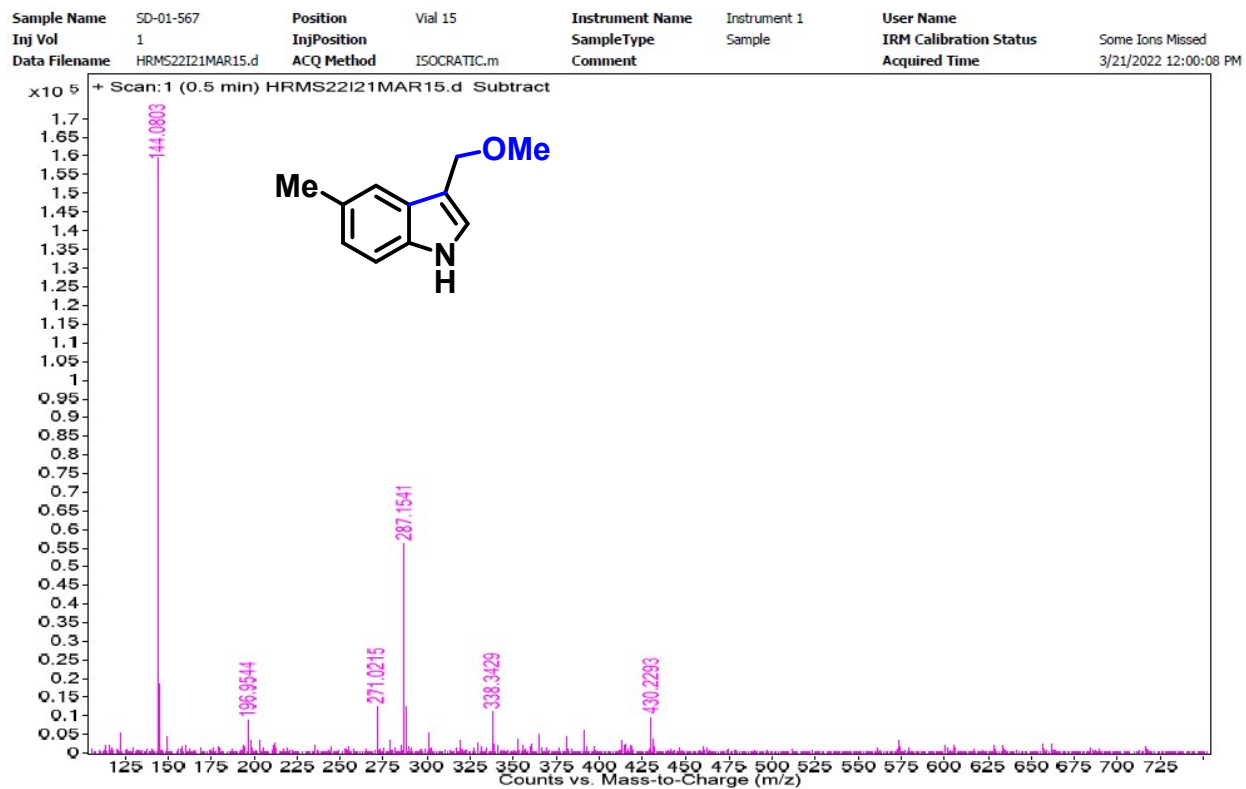


Figure S-119: HRMS spectrum of compound 4b

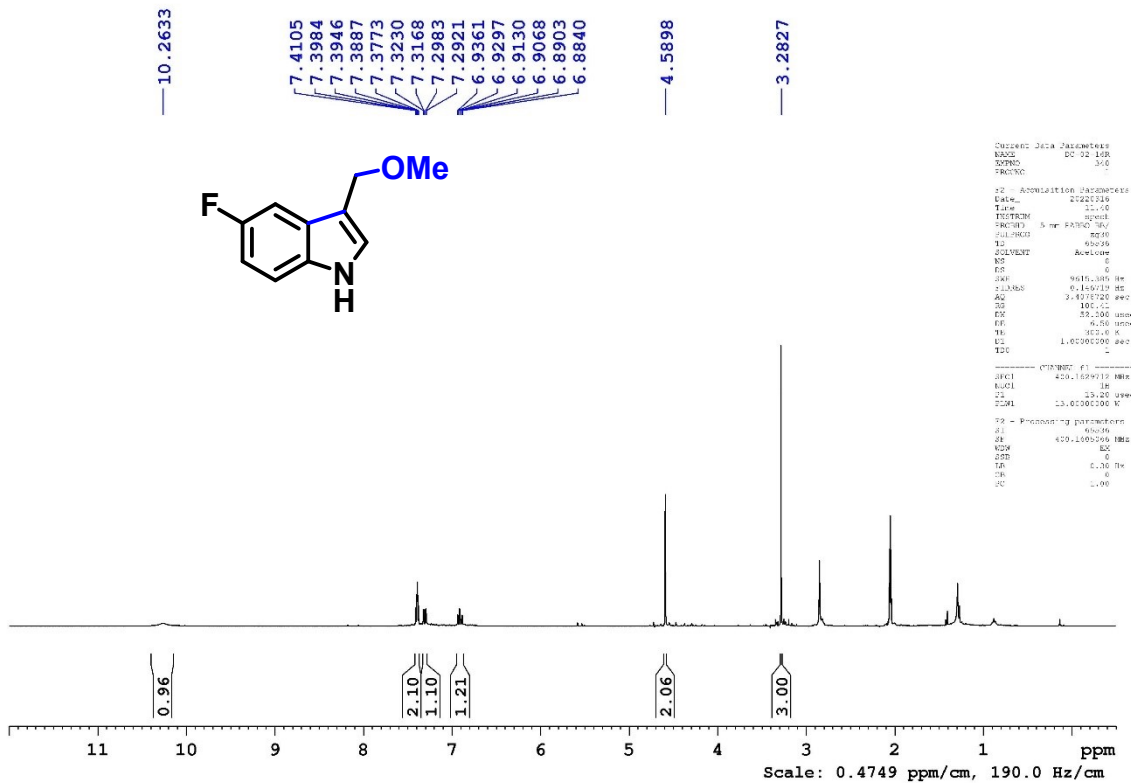


Figure S-120: ¹H NMR (400 MHz, D₆-acetone) spectrum of compound 4c

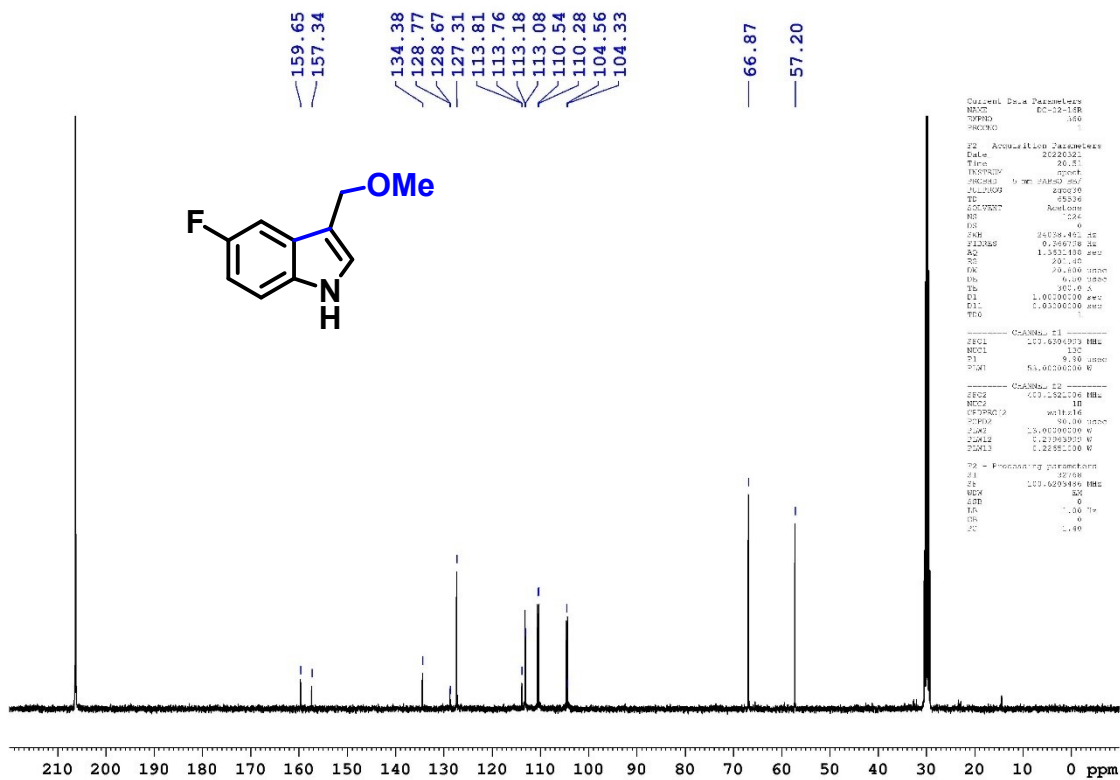


Figure S-121: ¹³C NMR (100 MHz, D₆-acetone) spectrum of compound 4c

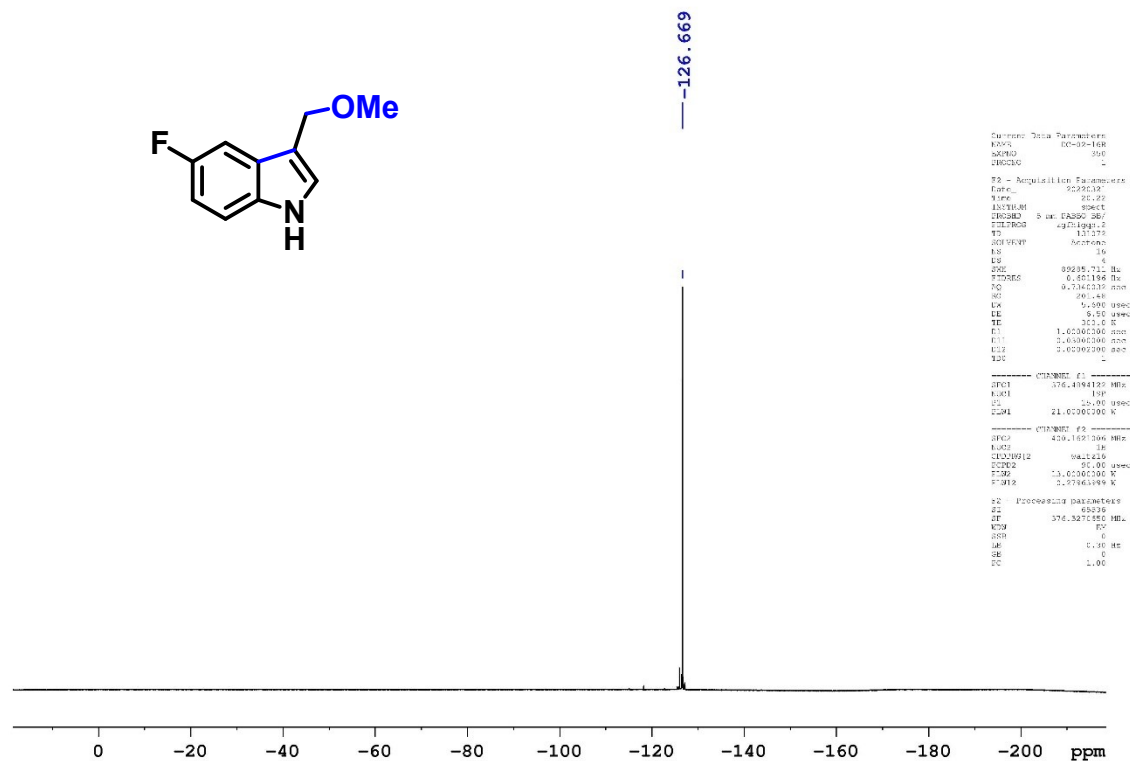


Figure S-122: ¹⁹F NMR (376 MHz, D₆-acetone) spectrum of compound 4c

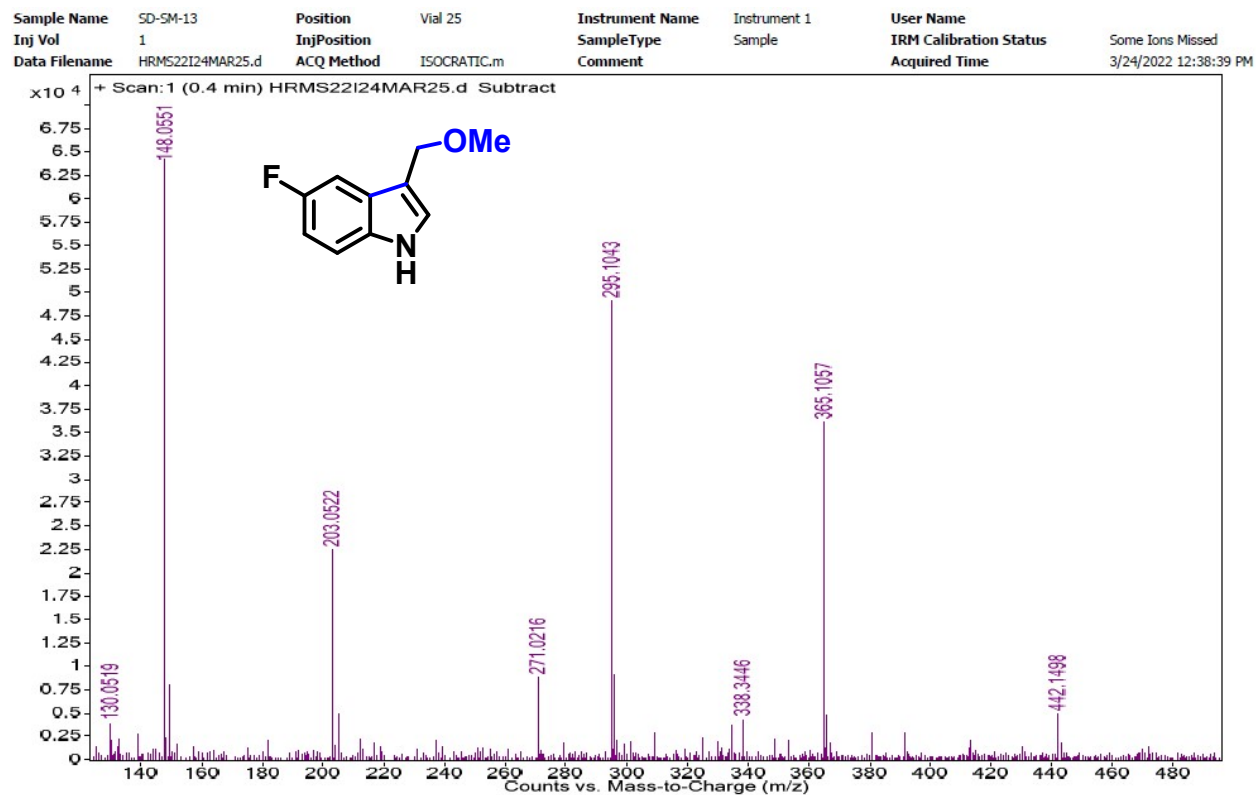


Figure S-123: HRMS spectrum of compound 4c

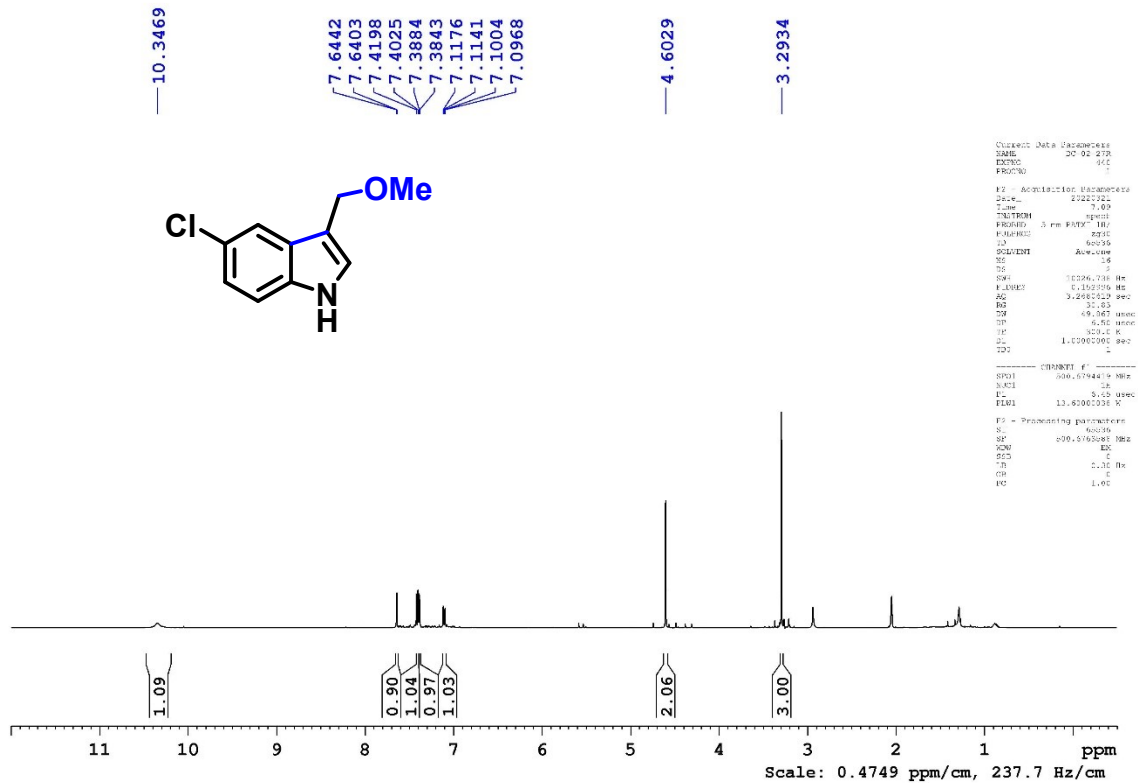


Figure S-124: ¹H NMR (500 MHz, D₆-acetone) spectrum of compound 4d

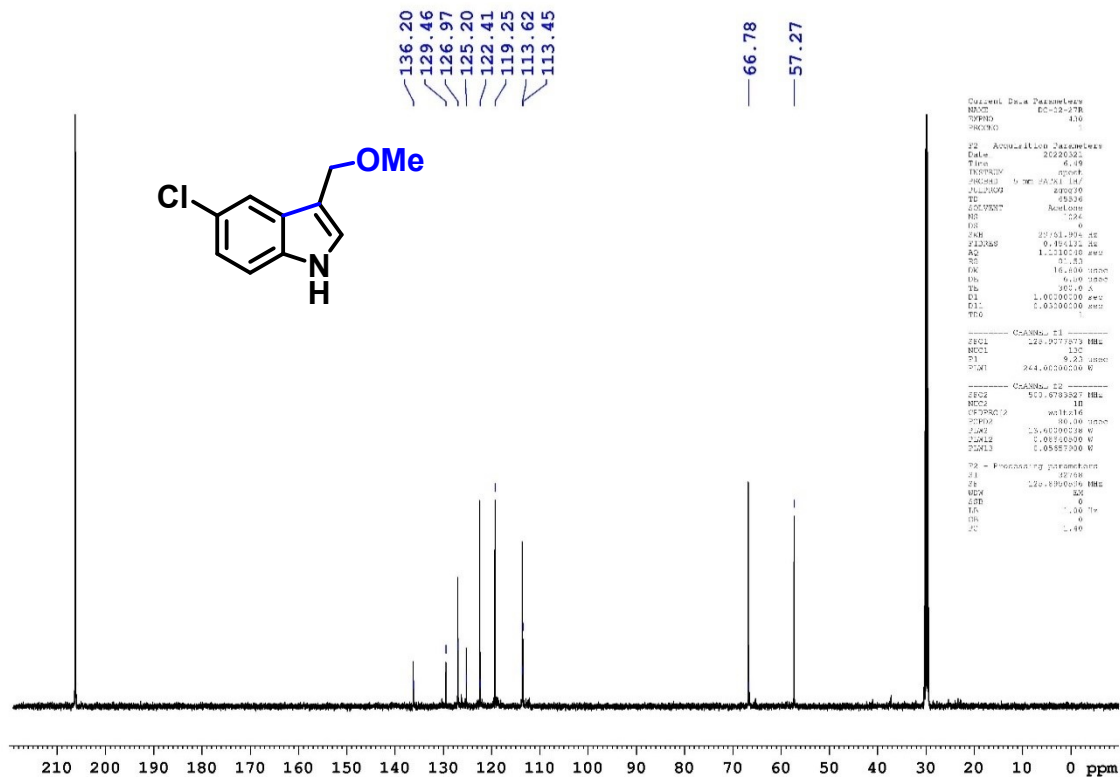


Figure S-125: ¹³C NMR (125 MHz, D₆-acetone) spectrum of compound 4d

| Sample Name | DC-01-913 | Position | Vial 18 | Instrument Name | Instrument 1 | User Name | |
|---------------|------------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | HRMS22122MAR18.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 3/22/2022 12:15:59 PM |

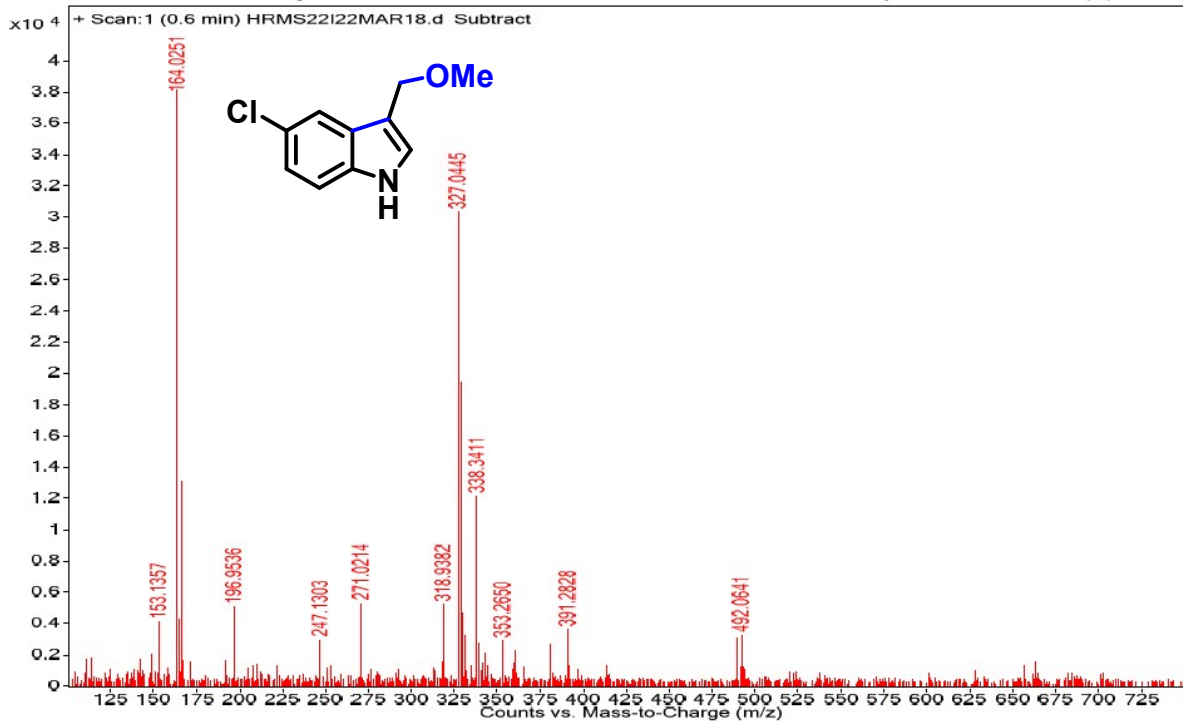


Figure S-126: HRMS spectrum of compound 4d

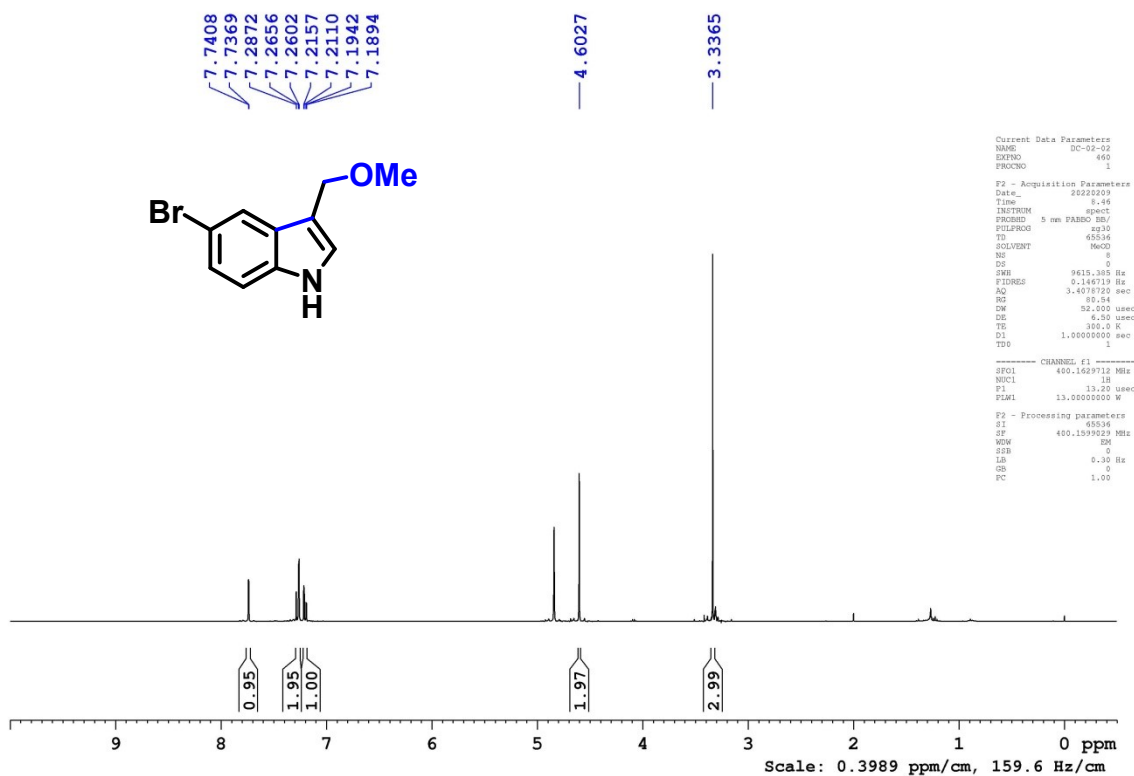


Figure S-127: ¹H NMR (400 MHz, D₄-methanol) spectrum of compound 4e

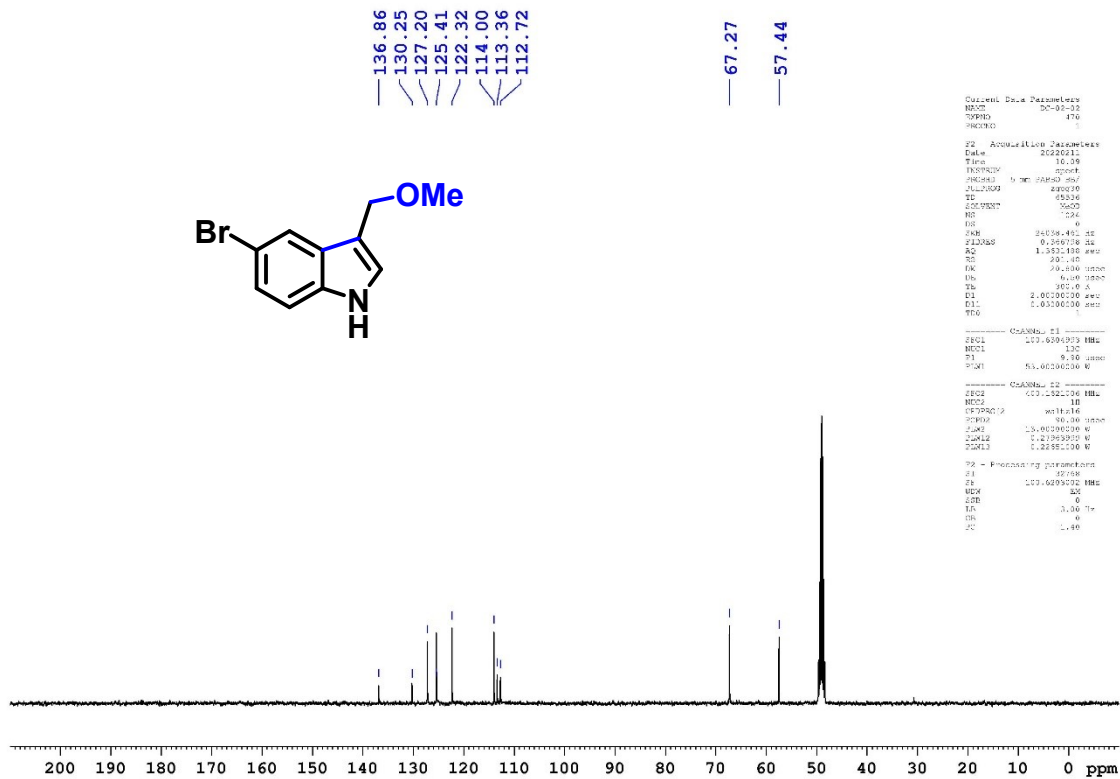


Figure S-128: ¹³C NMR (100 MHz, D₄-methanol) spectrum of compound 4e

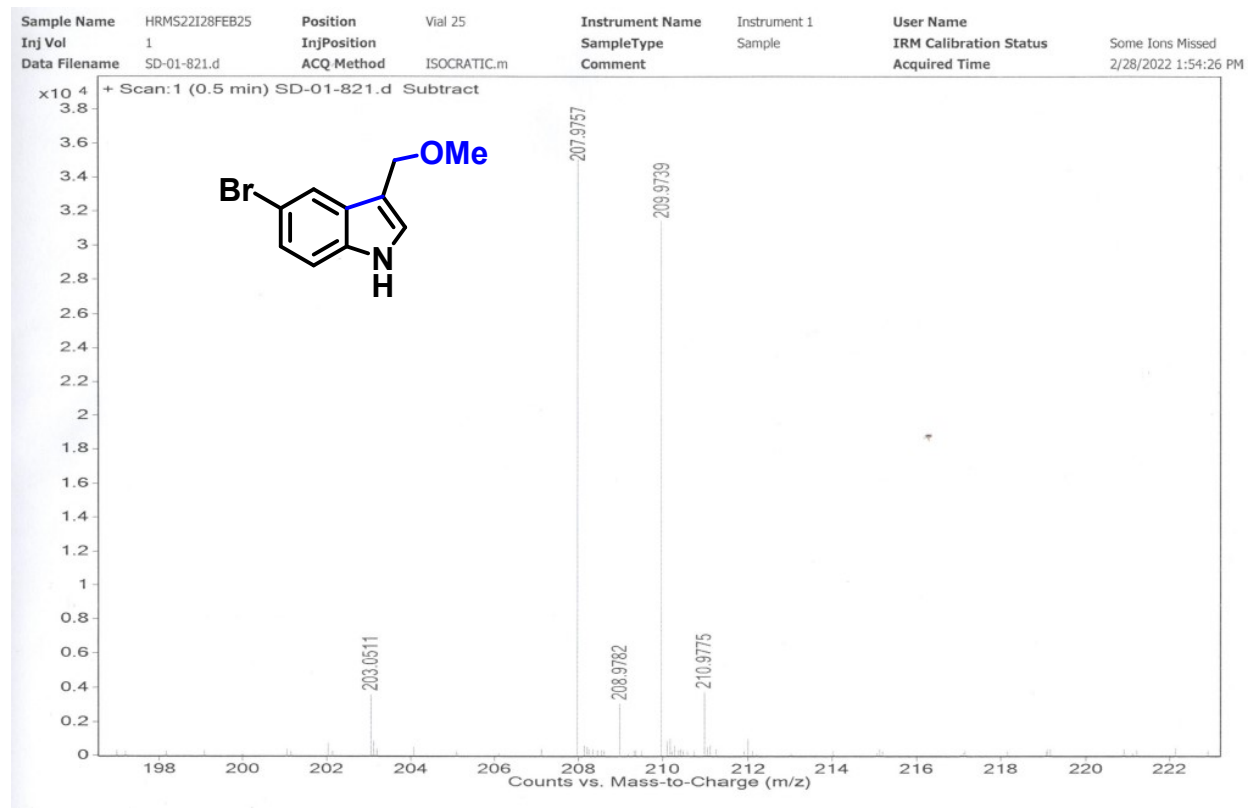


Figure S-129: HRMS spectrum of compound 4e

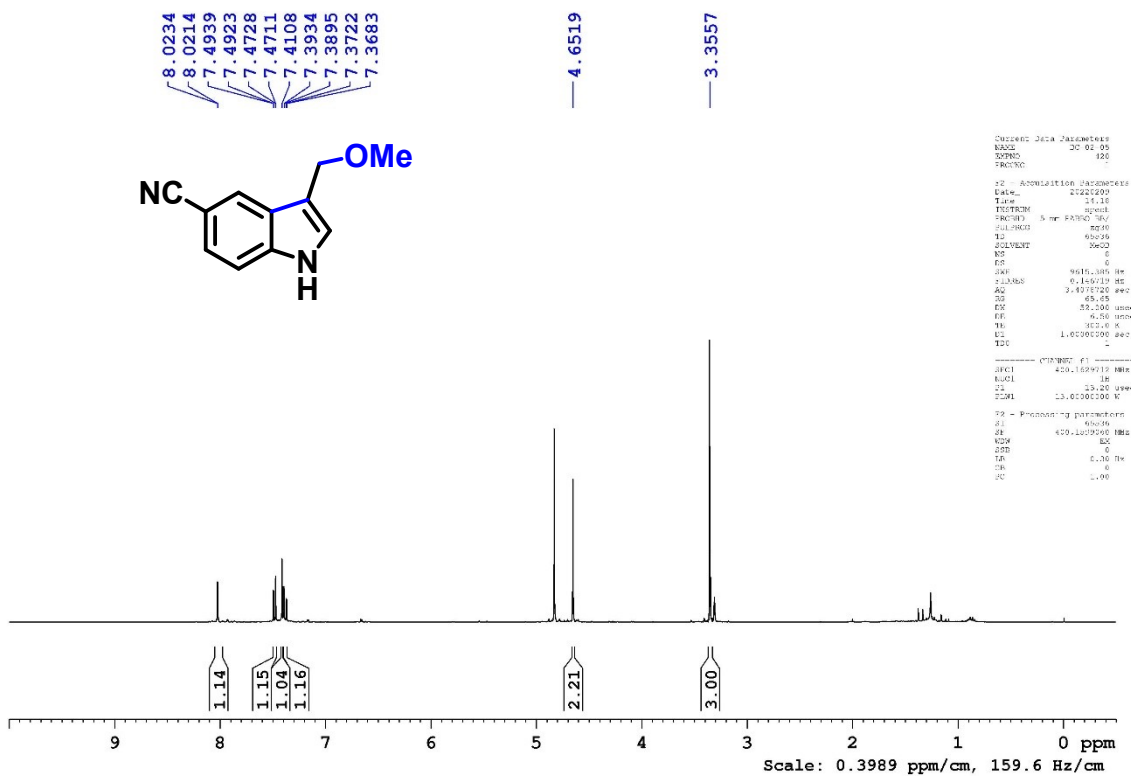


Figure S-130: ¹H NMR (400 MHz, D₄-methanol) spectrum of compound 4f

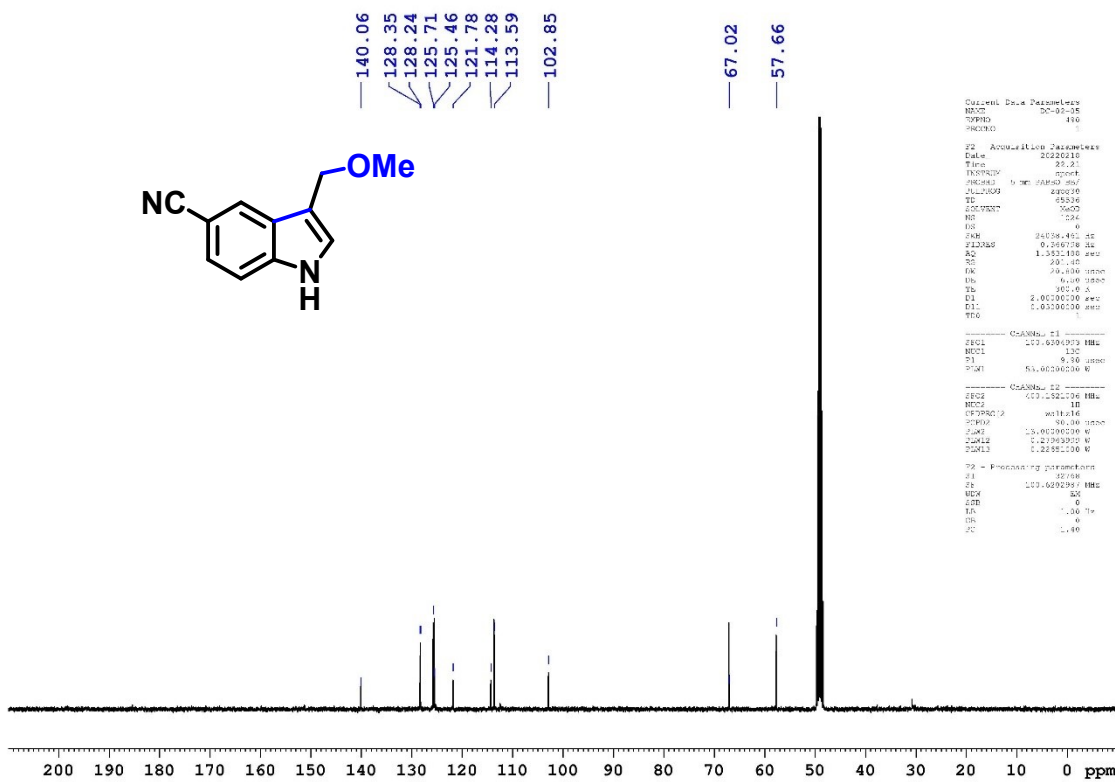


Figure S-131: ¹³C NMR (100 MHz, D₄-methanol) spectrum of compound 4f

| | | | | | | | |
|---------------|----------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Sample Name | HRMS22I18FEB10 | Position | Vial 10 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 5 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | DC-01-314_1.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 2/18/2022 12:50:23 PM |

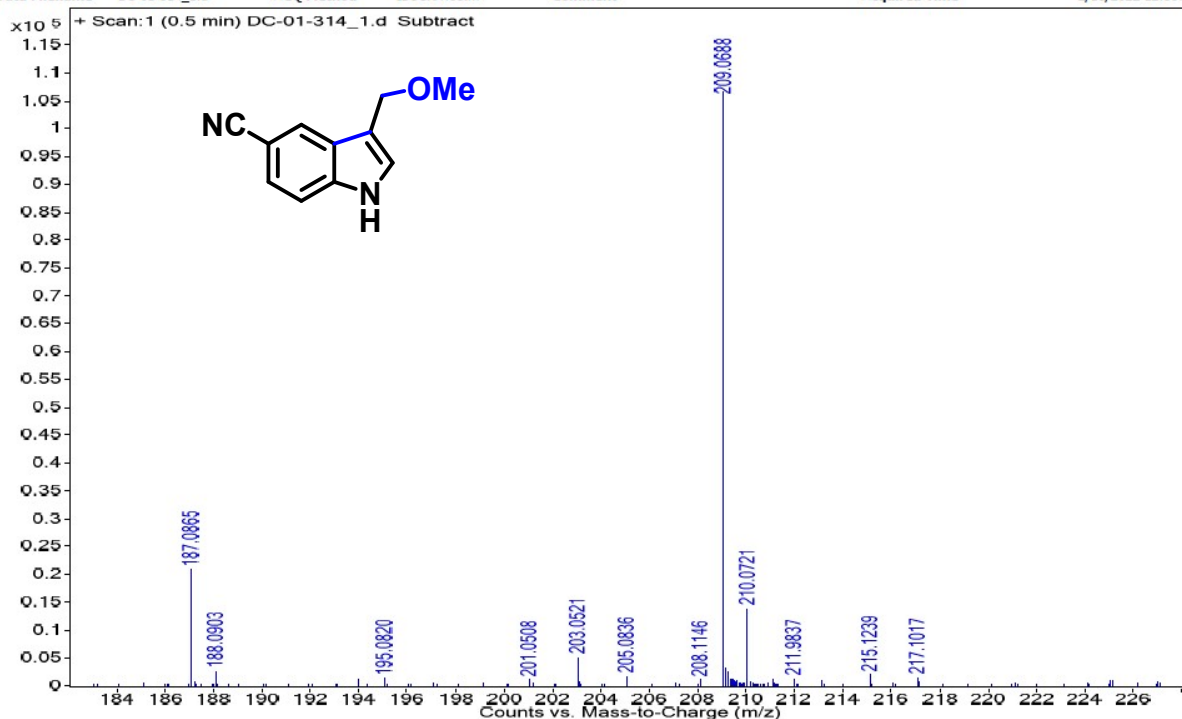


Figure S-132: HRMS spectrum of compound 4f

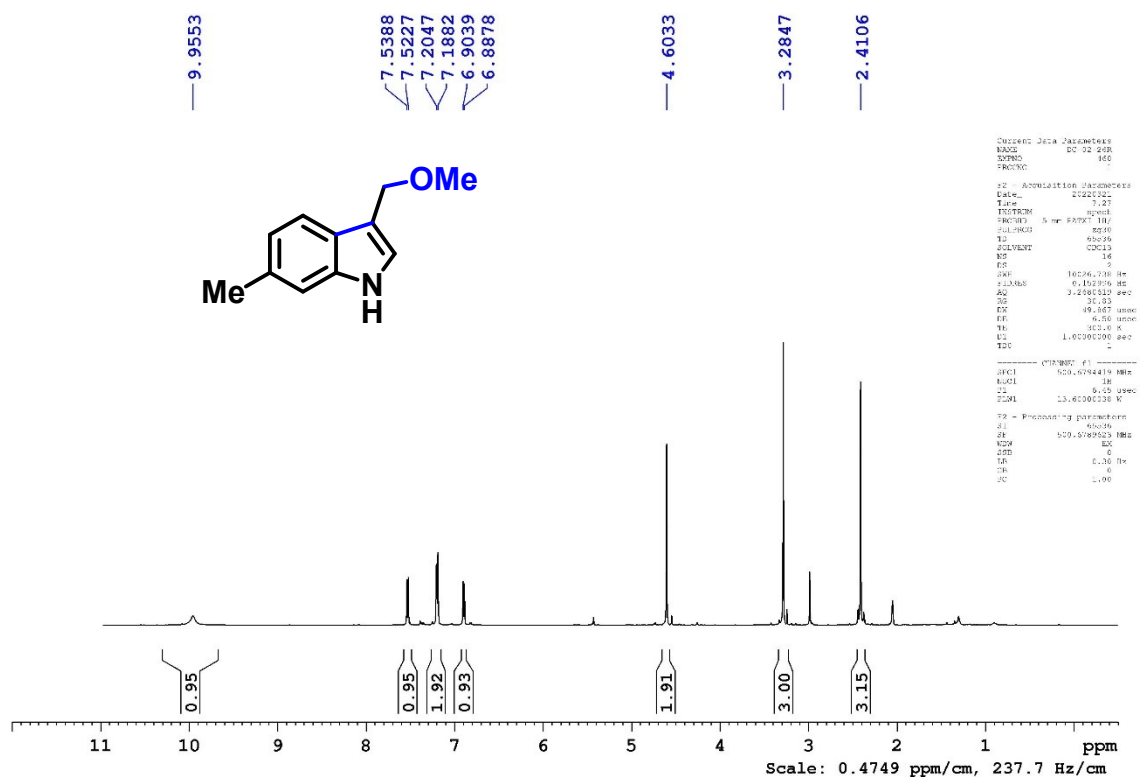


Figure S-133: ¹H NMR (500 MHz, CDCl₃) spectrum of compound 4g

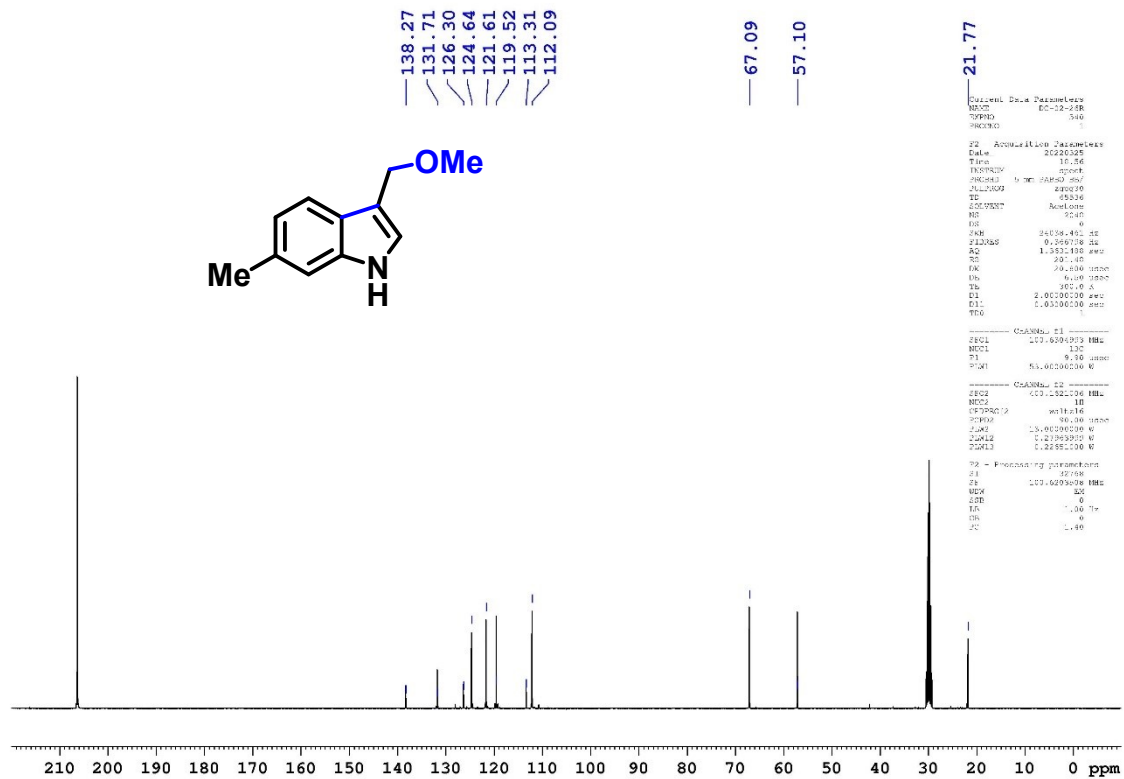


Figure S-134: ¹³C NMR (100 MHz, D₆-acetone) spectrum of compound 4g

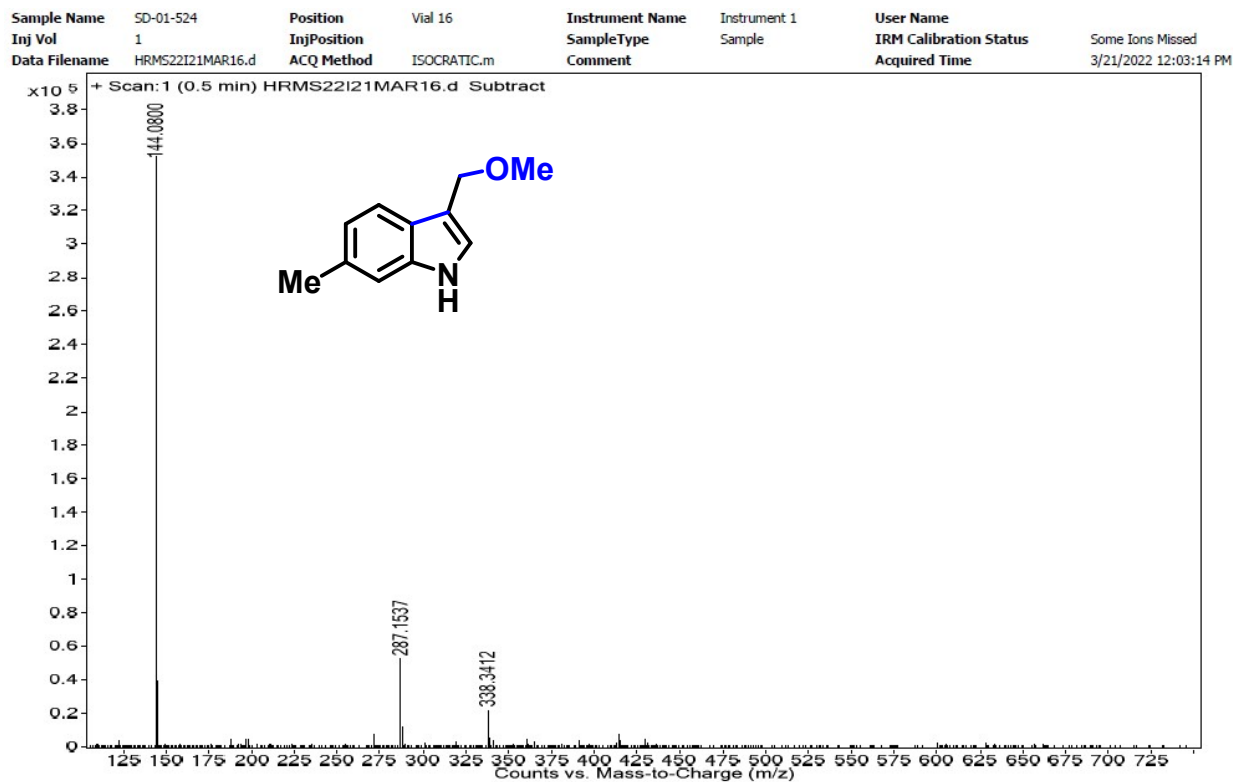


Figure S-135: HRMS spectrum of compound 4g

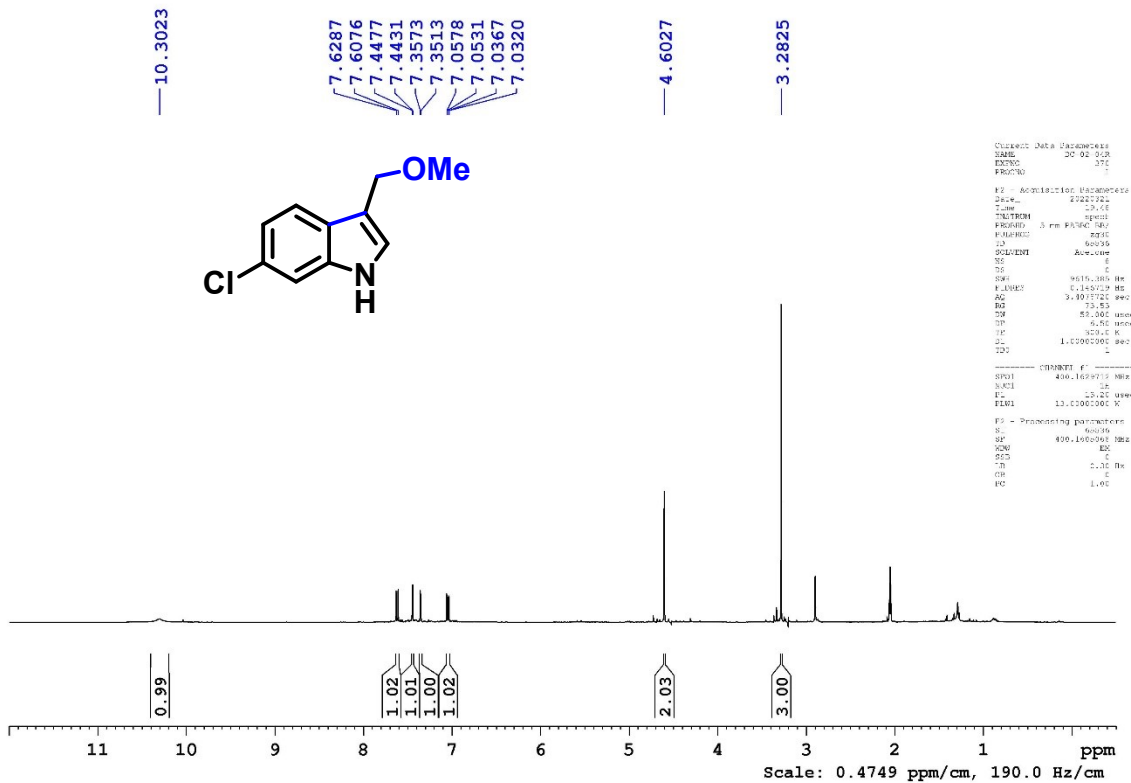


Figure S-136: ¹H NMR (400 MHz, D₆-acetone) spectrum of compound 4h

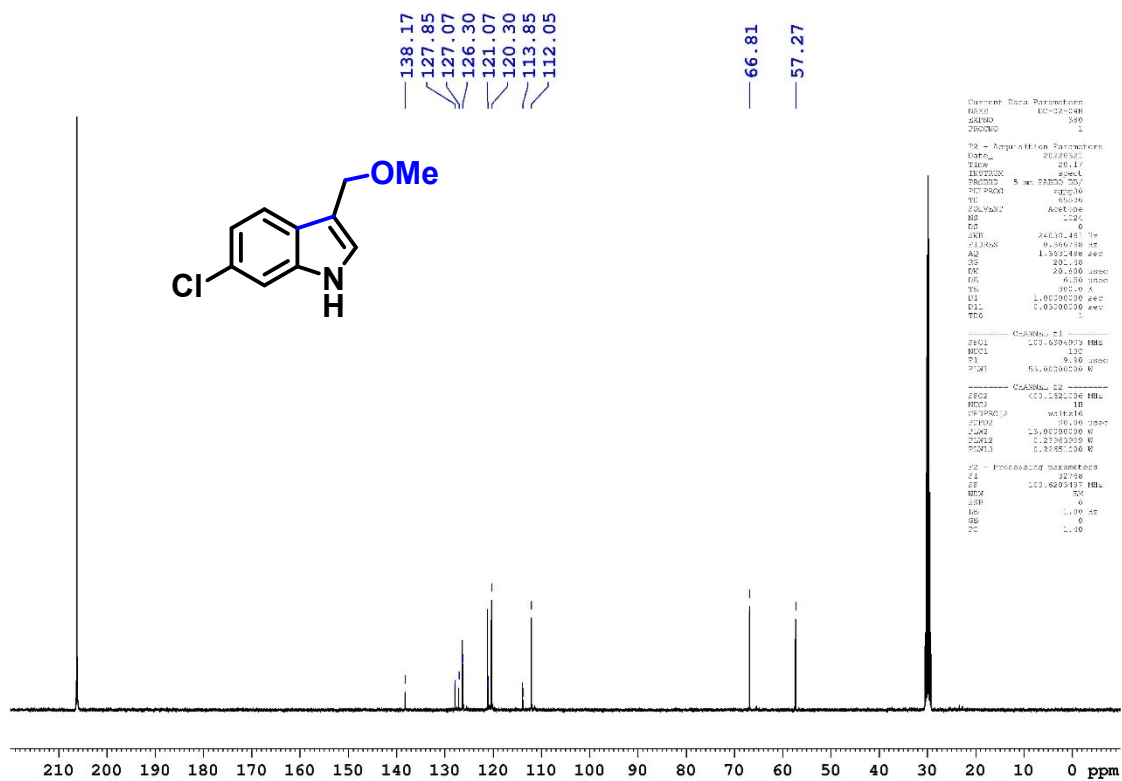


Figure S-137: ¹³C NMR (100 MHz, D₆-acetone) spectrum of compound 4h

| | | | | | | | |
|---------------|------------------|-------------|-------------|-----------------|--------------|------------------------|-----------------------|
| Sample Name | SD-01-772-3 | Position | Vial 26 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | HRMS22124MAR26.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 3/24/2022 12:41:47 PM |

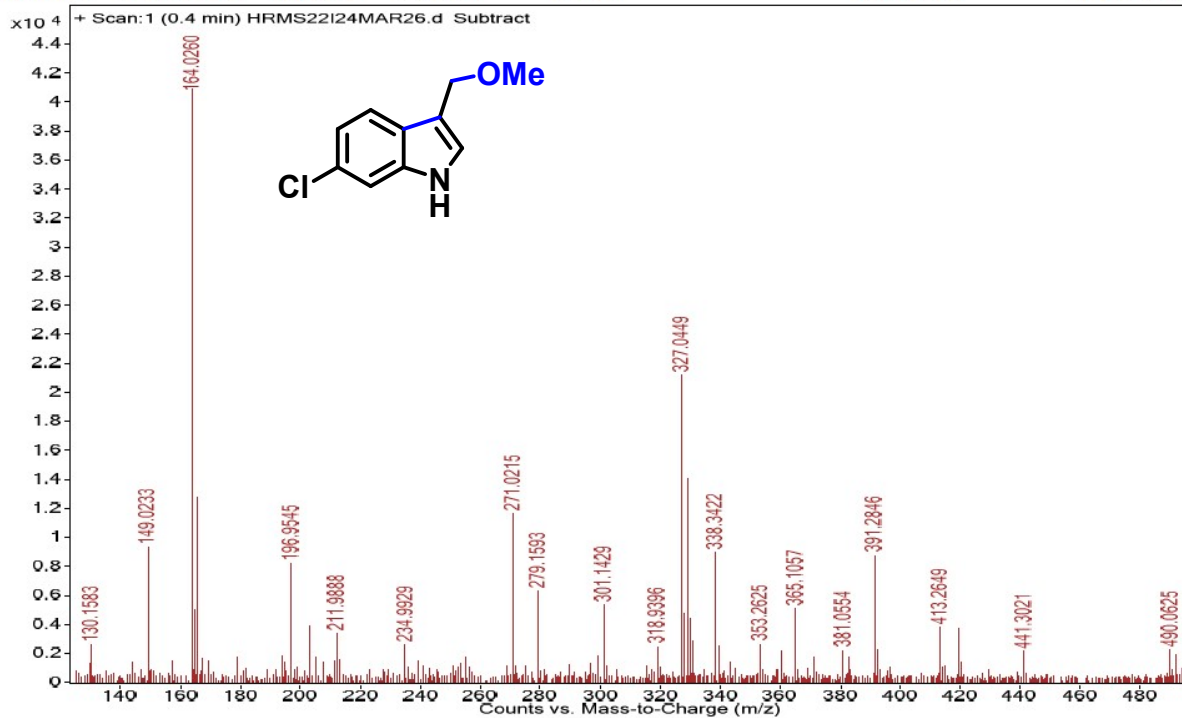


Figure S-138: HRMS spectrum of compound 4h

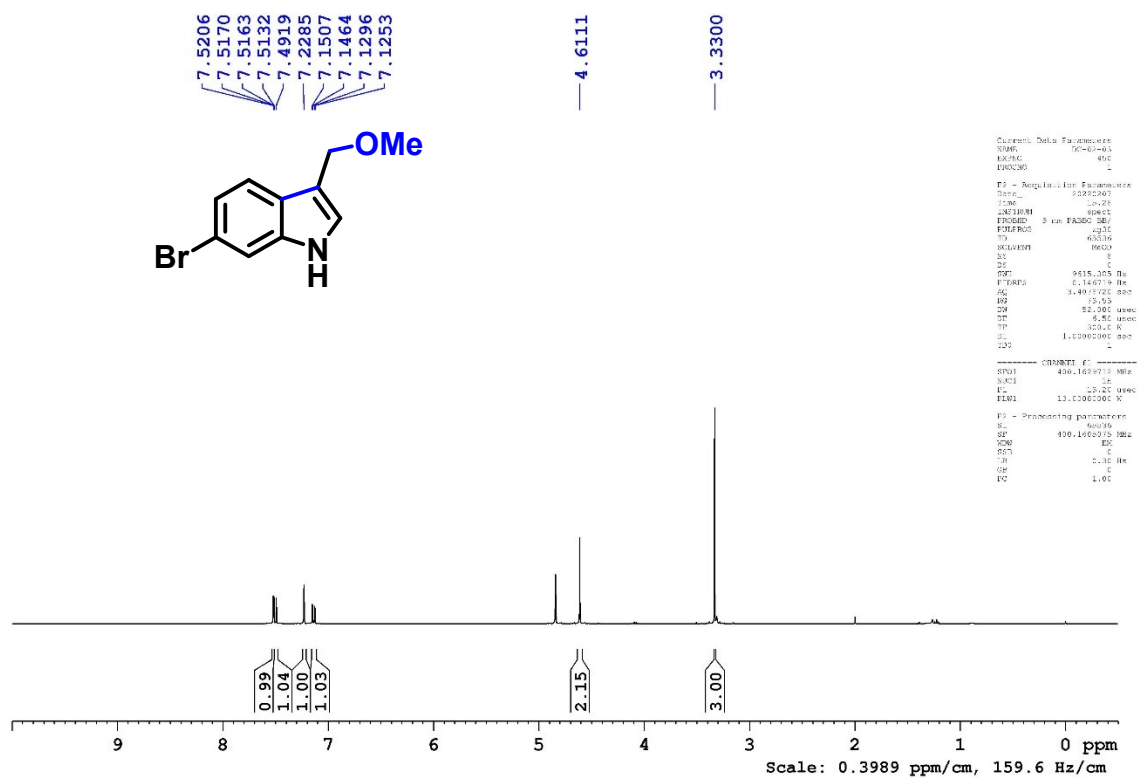


Figure S-139: ¹H NMR (400 MHz, D₄-methanol) spectrum of compound 4i

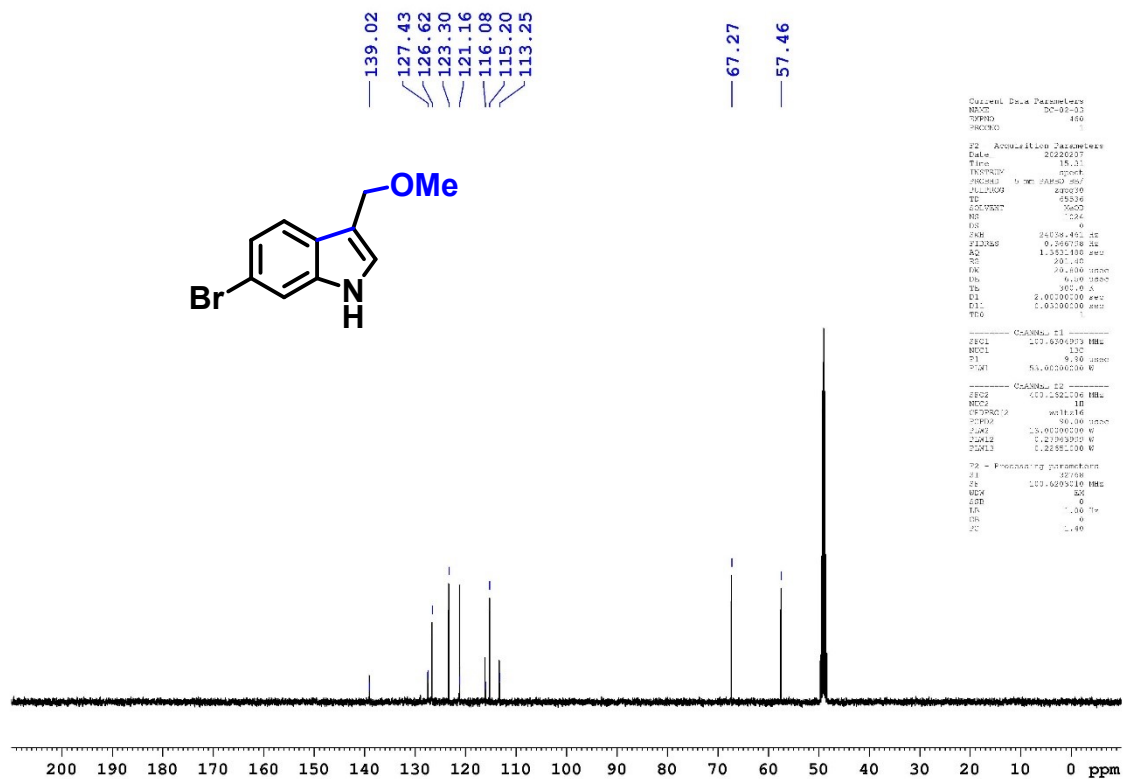


Figure S-140: ¹³C NMR (100 MHz, D₄-methanol) spectrum of compound 4i

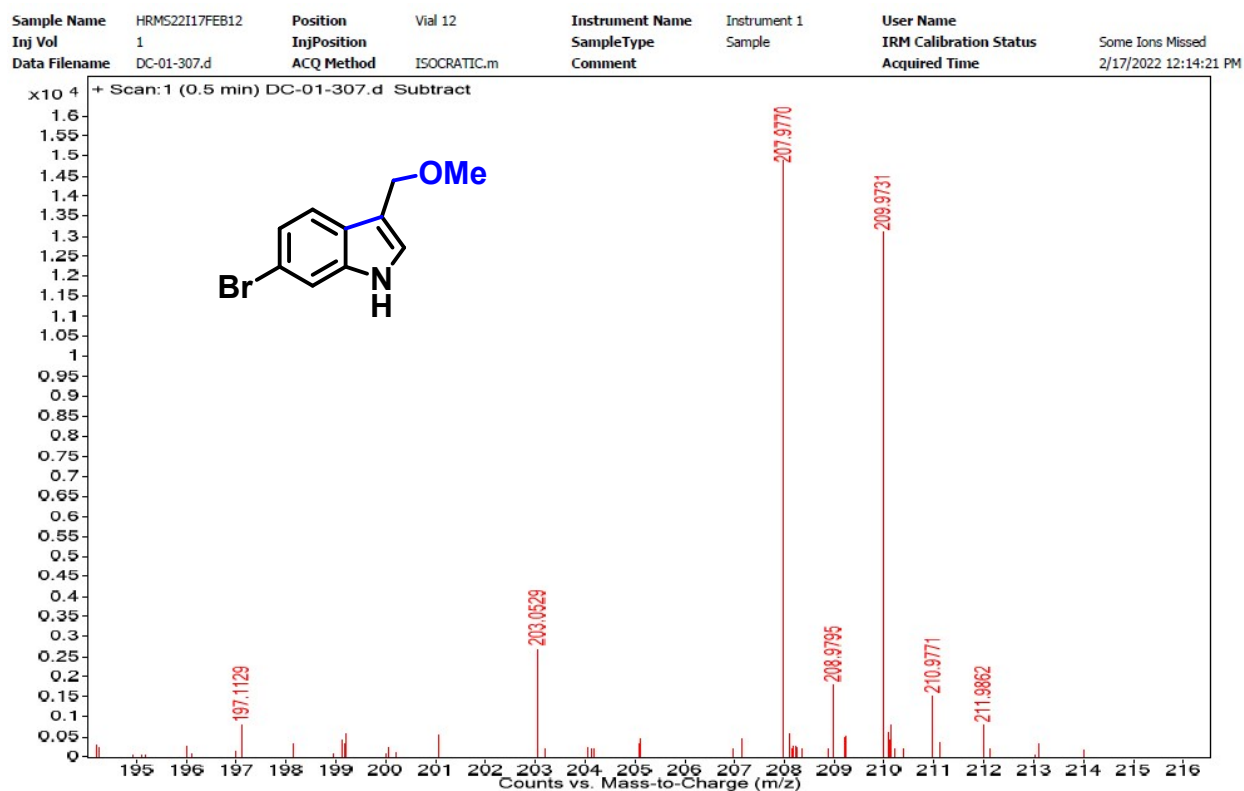


Figure S-141: HRMS spectrum of compound 4i

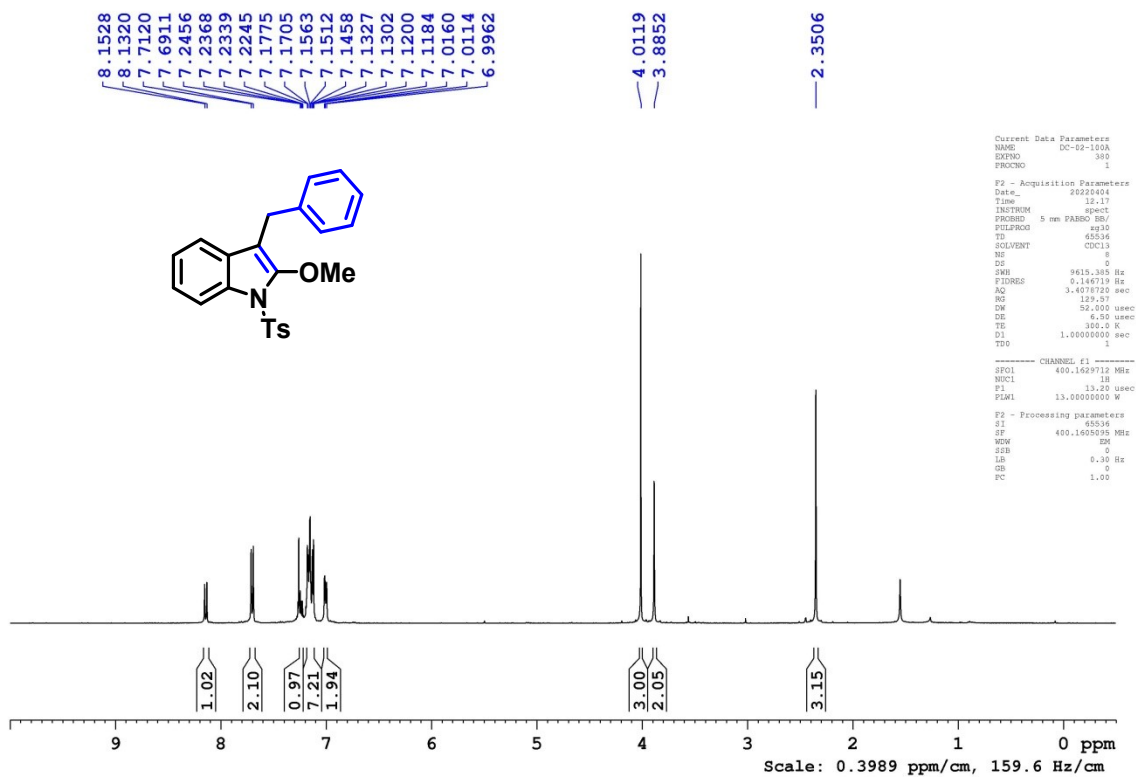


Figure S-142: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5

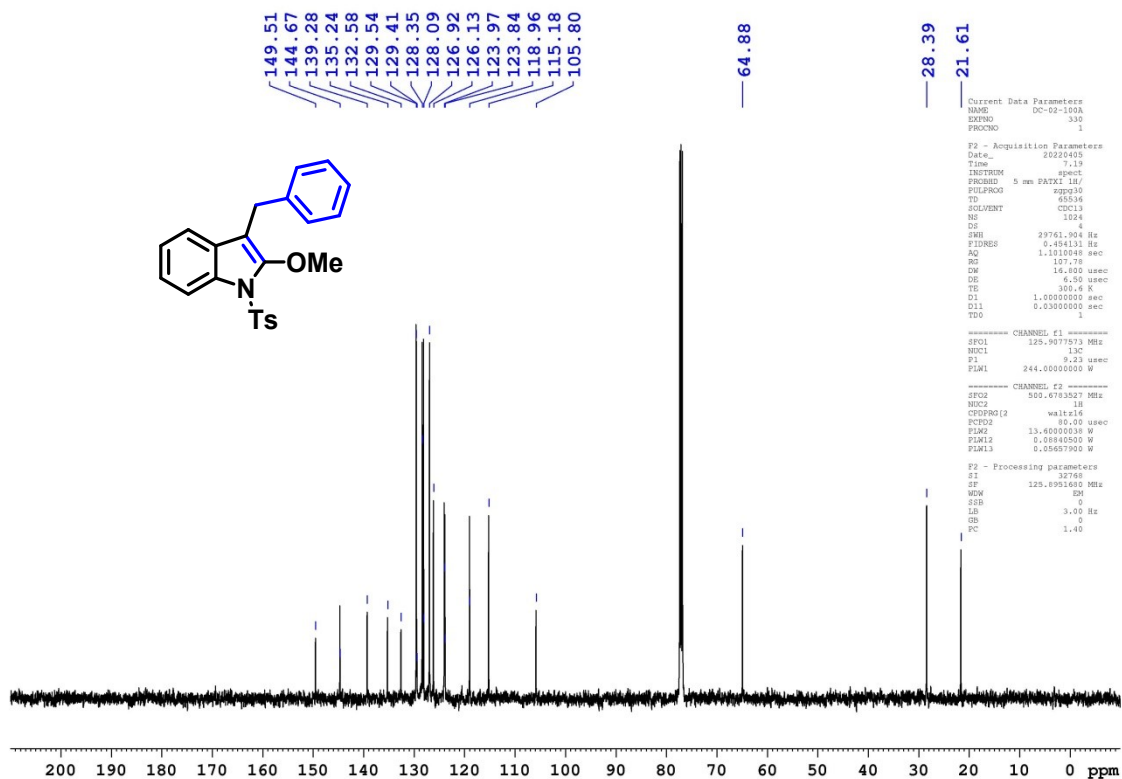


Figure S-143: ¹³C NMR (125 MHz, CDCl₃) spectrum of compound 5

| | | | | | | | |
|---------------|------------------|-------------|-------------|-----------------|--------------|------------------------|---------------------|
| Sample Name | KM-PHU | Position | Vial 26 | Instrument Name | Instrument 1 | User Name | |
| Inj Vol | 1 | InjPosition | | SampleType | Sample | IRM Calibration Status | Some Ions Missed |
| Data Filename | HRMS22105APR26.d | ACQ Method | ISOCRATIC.m | Comment | | Acquired Time | 4/5/2022 4:48:16 PM |

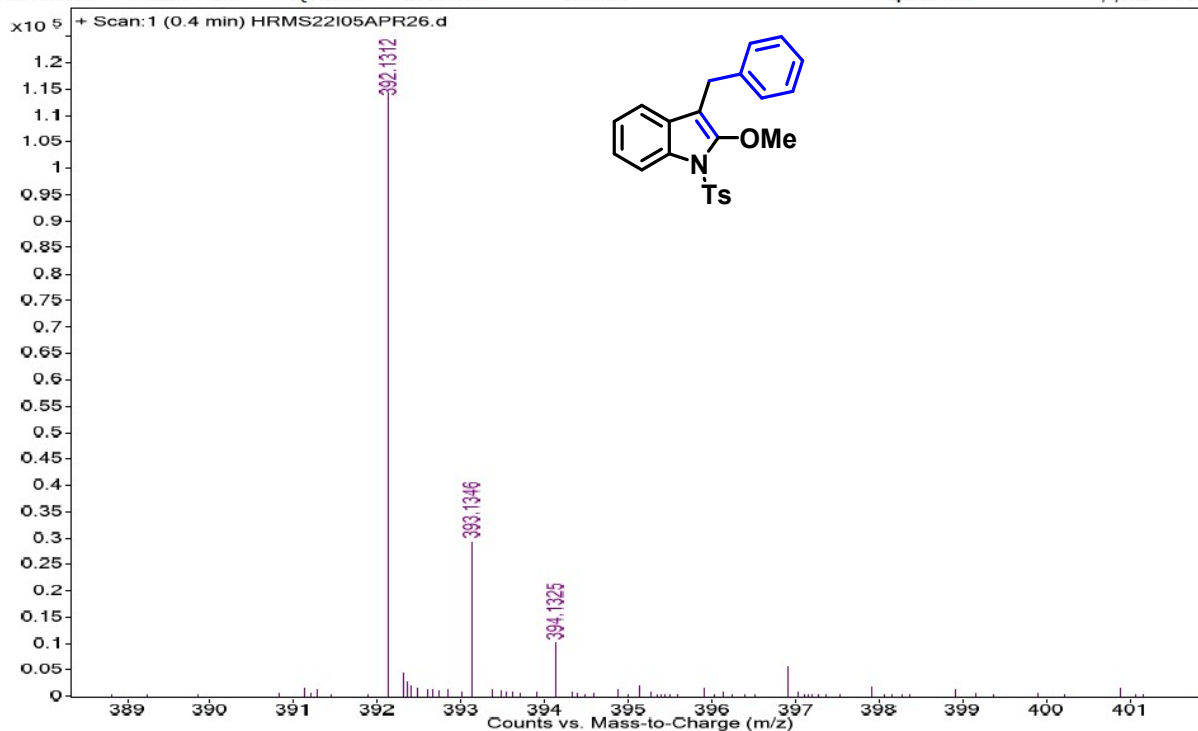


Figure S-144: HRMS spectrum of compound 5

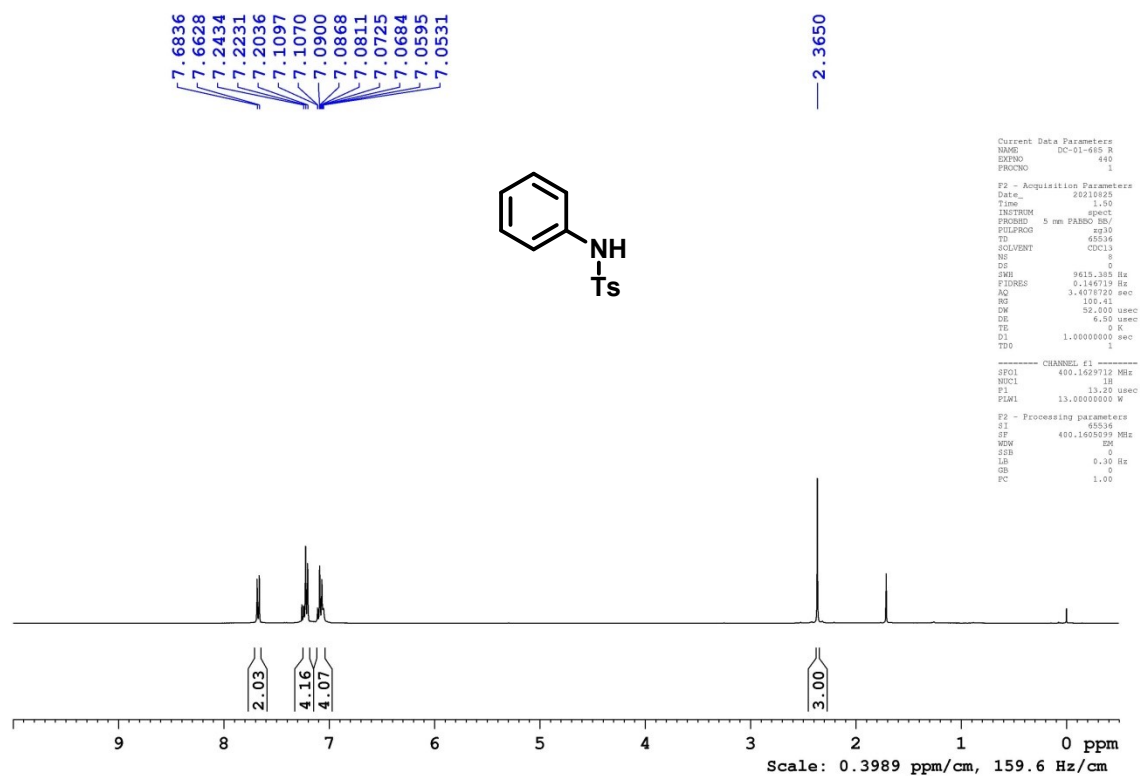


Figure S-145: ¹H NMR (400 MHz, CDCl₃) spectrum of compound 7a

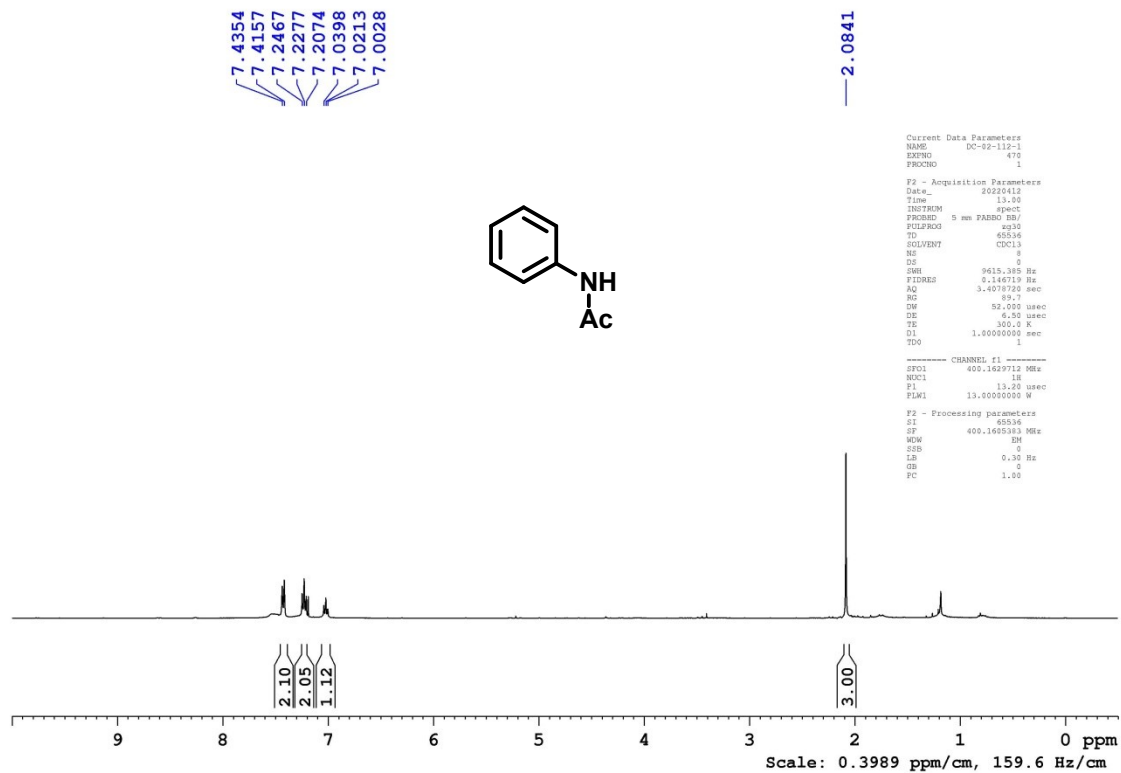


Figure S-146: ¹H NMR (400 MHz, CDCl₃) spectrum of compound **7b**

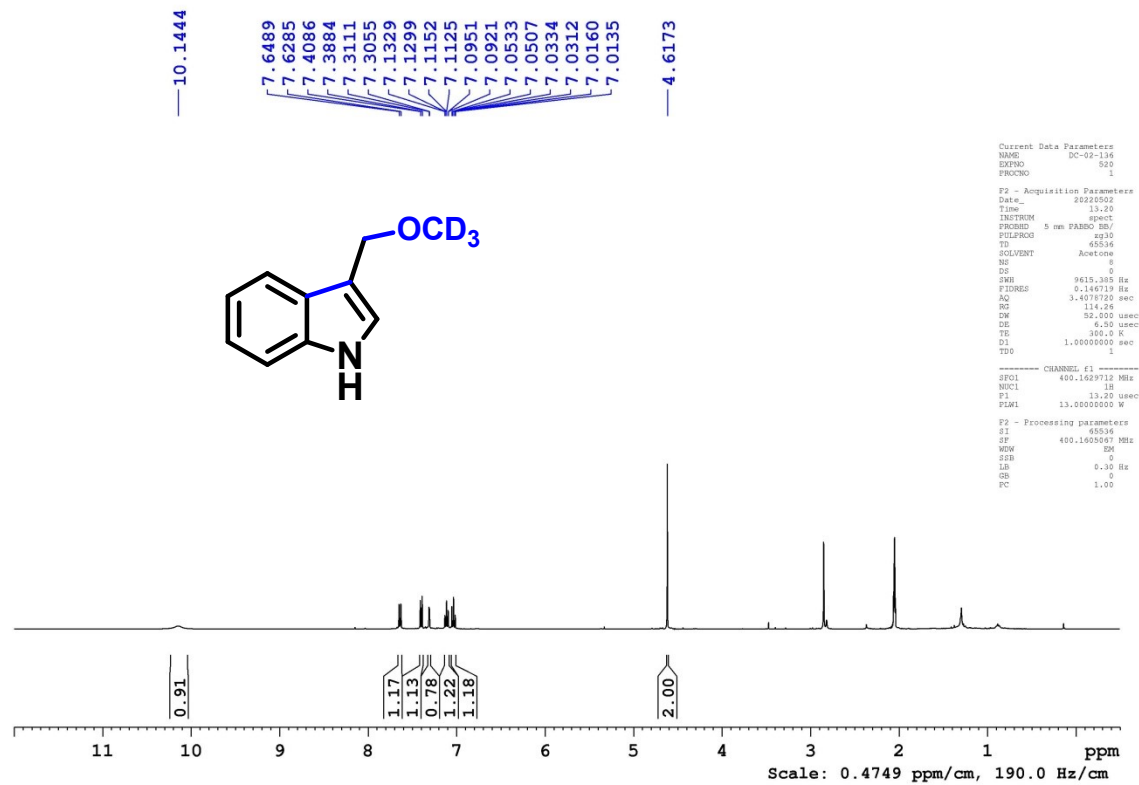


Figure S-147: ¹H NMR (400 MHz, D₆-acetone) spectrum of compound **4a-D**