

Supporting Information

Electrochemical Collective Synthesis of Labeled Pyrroloindoline

Alkaloids with Freon-type Methanes as Functional C1 Synthons

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1. General information

Reagents: All commercial materials were used as received from Energy Chemical or Adamas-beta, Alfa Aesar, Acros, Bidepharm and Leyan unless otherwise noted.

Reactions: All reactions were carried out in undivided electrochemical cells (10 mL) using pre-dried glassware, if not noted otherwise. The electrochemical cells were fitted with a rubber stopper with electrical feed-throughs. Electrocatalysis was conducted using an DC-power supplier HY3005ET in constant current mode, CV studies were performed using a CHI660E workstation.

Chromatography: Thin layer chromatography (TLC) was carried out on silica gel 60 F254 pre-coated glass plates. Visualization was detected by irradiation with UV light (254 nm), or by treatment with a solution of phosphomolybdic acid in ethanol followed by heating. Flash chromatography was carried out on 200 – 300 mesh silica gel, eluting with a mixture of petroleum ether (b.p. 60 – 90 °C) and ethyl acetate.

NMR Spectroscopy: ^1H NMR and ^{13}C NMR spectra were recorded on a Bruker AVANCE III HD 400, 500, 600 or 700 M spectrometer, operating at 400 (or 500 or 600 or 700) MHz and 100 (or 125 or 150 or 175) MHz respectively. Chemical shifts (δ) were given in parts per million (ppm), and referenced relative to residual solvent CHCl_3 (7.26 ppm) in CDCl_3 , or tetramethylsilane (0.00 ppm) as an internal standard for ^1H NMR spectra and deuterated solvent CDCl_3 (77.0 ppm) for ^{13}C NMR spectra. Coupling constants (J) were reported in hertz (Hz). The following abbreviations are used to indicate the multiplicity of the signals: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and associated combinations, e.g. dd = doublet of doublets.

Mass Spectrometry: High-resolution mass spectra (HRMS) were obtained on a Bruker Buker impact II using the electrospray ionization (ESI) technique.

X-Ray: X-ray diffraction data were collected on a Bruker APEX-II CCD diffractometer.

Preparation of substrates: The acrylamides were prepared according to the related references. [1-6]

2. Cyclic voltammetry studies

General information: Cyclic voltammetry (CV) experiments were conducted in a 20 mL glass vial fitted with a glassy carbon working electrode (3 mm in diameter, BASi), a saturated calomel electrode as reference electrode, and a platinum wire counter electrode. The solution of interest was sparged with nitrogen for 3-5 minutes before data collection. The diagrams were made using OriginLab 8.0.

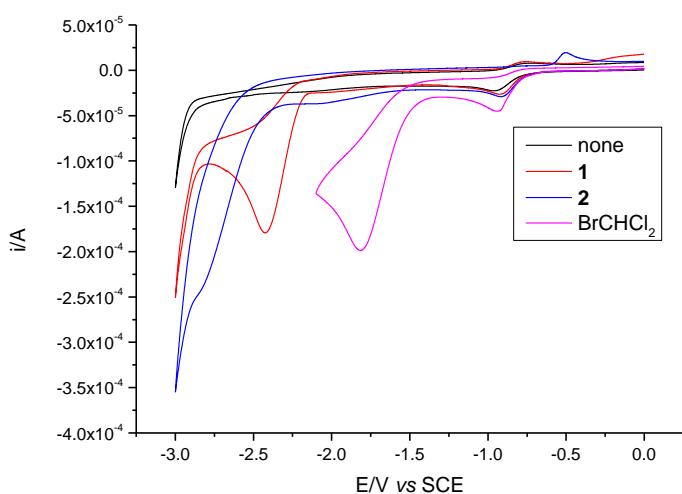


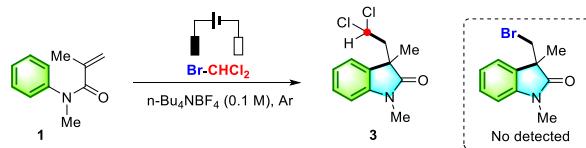
Fig. S1. Cyclic voltammogram of **1** (10 mM), **3** (10 mM), and BrCHCl₂ (10 mM) in DMF (10 mL) containing n-Bu₄NBF₄ (1 mmol). **Scan rate: 50 mV/s.**

3. Optimization table of reaction conditions

The desired chlorinated oxindoles could be obtained in 89% yield using a carbon felt as the anode and a foam Ni plate as the cathode, in an undivided cell containing DMF and n-Bu₄NBF₄ at 100 °C (**Table S1**, entry 1). Decrease of amounts of BrCHCl₂ to 5 equiv. resulted in 79% yield, while reaction temperature of 80 °C resulted in a sharp decrease in the yield (entry 2 and 3). A 66% yield was obtained when DMSO was used as the solvent (entry 4). Then various anodes and cathodes were screened, and Pt cathode, carbon felt cathode, Pt anode and even foam Ni anode can enable the reaction to proceed smoothly, although in lower yield (entries 5-8). A reaction process analysis was conducted to monitor the reaction (see SI), and we found that about 90% of acrylamide **1** was consumed and 81% of product was obtained when the reaction time

was 6 h (entry 9). However, when we conducted the reaction at a current of 10 mA, 3h instead of 5 mA, 6h to ensure the same quantity of electricity in the reaction, the yield decreased to 63%, which indicated that high current density may not be beneficial to the reaction probably because of overreduction of BrCHCl₂ and limited mass transfer rate to the electrode surface (entry 10). Exposure to air instead of Ar atmosphere resulted in a sharp decrease of yield (entry 11), while no products was obtained without electricity, confirming the indispensable role of electricity in the reaction (entry 12).

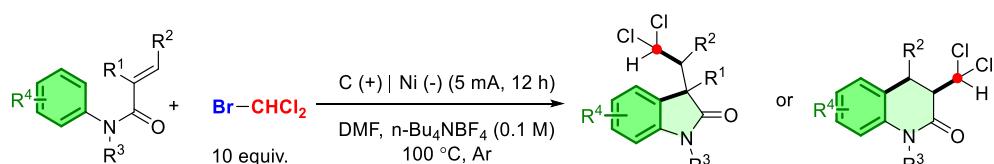
Table S1. Optimization.^a



entry	(+)/(-)	Reagents (equiv.)	yield
1	C/Ni	BrCHCl ₂ (10), 100 °C, DMF	89%
2	C/Ni	BrCHCl ₂ (5), 100 °C, DMF	79%
3	C/Ni	BrCHCl ₂ (10), 80 °C, DMF	13%
4	C/Ni	BrCHCl ₂ (10), 100 °C, DMSO	66%
5	C/Pt	BrCHCl ₂ (10), 100 °C, DMF	76%
6	C/C	BrCHCl ₂ (10), 100 °C, DMF	88%
7	Pt/Ni	BrCHCl ₂ (10), 100 °C, DMF	86%
8	Ni/Ni	BrCHCl ₂ (10), 100 °C, DMF	79%
9 ^b	C/Ni	BrCHCl ₂ (10), 100 °C, DMF	81%
10 ^c	C/Ni	BrCHCl ₂ (10), 100 °C, DMF	63%
11 ^d	C/Ni	BrCHCl ₂ (10), 100 °C, DMF	22%
12 ^e	C/Ni	BrCHCl ₂ (10), 100 °C, DMF	<5%

^aReaction conditions: acrylamide (0.2 mmol), BrCHCl₂, solvent (4 mL), n-Bu₄NBF₄ (0.1 M), Ar, anode area (0.5 cm × 0.5 cm), cathode area (0.5 cm × 0.5 cm), constant current (5 mA, 12h), undivided cell. Isolated yield. ^b5 mA, 6h. ^c10 mA, 3h. ^dAir. ^eNo current.

4. General procedure for electrochemical radical cyclization

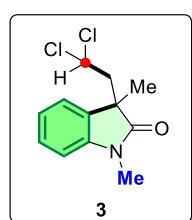


General procedure 1: To an oven-dried Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 0.2 mmol) and n-Bu₄NBF₄ (0.1 M, 0.4 mmol, 132 mg) were

added. The rubber plug equipped with a carbon felt anode (10 mm * 10 mm) and a foam Ni cathode (10 mm* mm) was used to keep the Schlenk tube sealed. The tube was evacuated and filled with Ar, repeating three times. BrCHCl₂ (10 equiv., 2 mmol, 165 µL) was dissolved in 4 mL DMF and then injected into the tube via syringe. The mixture was stirred for 5 min and put in the oil bath (100 °C) with the electrolysis of 5 mA, 12h. After the reaction, the tube was cooled to room temperature and ethyl acetate (10 mL) and water (10 mL) was added. The aqueous layer was separated and extracted with ethyl acetate (3×10 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield the desired product.



Fig. S2. The electrochemical reaction setup.

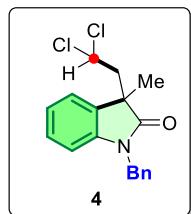


Purification of the crude product by flash column chromatography afforded the cyclization product **3** as colorless oil in 89% yield (46 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.31 (td, *J* = 7.7, 1.3 Hz, 1H), 7.20 – 7.18 (m, 1H), 7.11 – 7.07 (m, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 5.38 (dd, *J* = 9.2, 4.1 Hz, 1H), 3.20 (s, 3H), 3.03 (dd, *J* = 14.8, 9.2 Hz, 1H), 2.70 (dd, *J* = 14.8, 4.1 Hz, 1H), 1.39 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.0, 143.4, 131.0, 128.6, 122.6, 122.6, 108.6, 69.6, 50.1, 47.1, 26.4, 25.4.

HRMS (ESI) *m/z* calculated for C₁₂H₁₃Cl₂NNaO [M+Na⁺] 280.0266, found 280.0267.

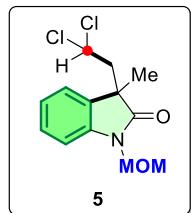


Purification of the crude product by flash column chromatography afforded the cyclization product **4** as pale yellow oil in 52% yield (35 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.25 (m, 5H), 7.23 – 7.19 (m, 2H), 7.09 – 7.05 (m, 1H), 6.80 – 6.78 (m, 1H), 5.45 (dd, *J* = 8.9, 4.4 Hz, 1H), 5.01 (d, *J* = 15.6 Hz, 1H), 4.81 (d, *J* = 15.6 Hz, 1H), 3.10 (dd, *J* = 14.8, 9.0 Hz, 1H), 2.76 (dd, *J* = 14.8, 4.4 Hz, 1H), 1.45 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.0, 142.5, 135.7, 131.1, 128.7, 128.5, 127.7, 127.5, 122.7, 122.7, 109.6, 69.6, 49.9, 47.2, 44.1, 26.1.

HRMS (ESI) *m/z* calculated for C₁₈H₁₇Cl₂NNaO [M+Na⁺] 356.0579, found 356.0581.

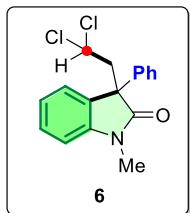


Purification of the crude product by flash column chromatography afforded the cyclization product **5** as colorless oil in 64% yield (37 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.32 (td, *J* = 7.7, 1.0 Hz, 1H), 7.22 (d, *J* = 6.9 Hz, 1H), 7.15 – 7.11 (m, 1H), 7.08 (d, *J* = 7.8 Hz, 1H), 5.44 (dd, *J* = 9.1, 4.3 Hz, 1H), 5.15 (d, *J* = 10.9 Hz, 1H), 5.08 (d, *J* = 10.9 Hz, 1H), 3.35 (s, 3H), 3.06 (dd, *J* = 14.9, 9.1 Hz, 1H), 2.74 (dd, *J* = 14.9, 4.3 Hz, 1H), 1.42 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.6, 141.7, 130.6, 128.7, 123.2, 122.7, 1101, 71.8, 69.6, 56.5, 49.9, 47.5, 26.3.

HRMS (ESI) *m/z* calculated for C₁₃H₁₅Cl₂NNaO₂ [M+Na⁺] 310.0372, found 310.0367.

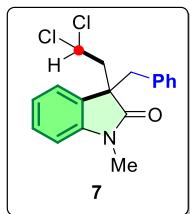


Purification of the crude product by flash column chromatography afforded the cyclization product **6** as white solid in 44% yield (28 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.34 (td, *J* = 7.7, 0.9 Hz, 1H), 7.27 – 7.18 (m, 6H), 7.13 – 7.09 (m, 1H), 6.87 (d, *J* = 7.9 Hz, 1H), 5.38 (dd, *J* = 9.5, 3.8 Hz, 1H), 3.52 (dd, *J* = 14.7, 9.5 Hz, 1H), 3.11 (s, 3H), 3.02 (dd, *J* = 14.7, 3.8 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 177.0, 144.4, 139.5, 129.2, 128.7, 128.6, 127.8, 126.4, 125.1, 122.7, 109.0, 69.8, 54.9, 50.1, 26.7.

HRMS (ESI) *m/z* calculated for C₁₇H₁₅Cl₂NNaO [M+Na⁺] 342.0423, found 342.0420.

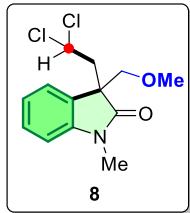


Purification of the crude product by flash column chromatography afforded the cyclization product **7** as white solid in 72% yield (48 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.25 – 7.22 (m, 1H), 7.14 – 7.02 (m, 5H), 6.79 – 6.76 (m, 2H), 6.61 (d, *J* = 7.8 Hz, 1H), 5.43 (dd, *J* = 9.3, 4.2 Hz, 1H), 3.18 (dd, *J* = 14.8, 9.3 Hz, 1H), 3.10 (d, *J* = 12.8 Hz, 1H), 2.99 (d, *J* = 12.8 Hz, 1H), 2.93 (s, 3H), 2.85 (dd, *J* = 14.8, 4.2 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 177.5, 144.0, 134.3, 130.0, 128.7, 128.1, 127.5, 126.8, 123.7, 122.1, 108.3, 69.7, 53.0, 48.6, 45.0, 26.0.

HRMS (ESI) *m/z* calculated for C₁₈H₁₇Cl₂NNaO [M+Na⁺] 356.0579, found 356.0580.

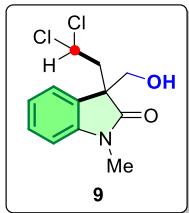


Purification of the crude product by flash column chromatography afforded the cyclization product **8** as pale yellow solid in 77% yield (44 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.30 (m, 2H), 7.11 – 7.07 (m, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 6.89 (d, *J* = 7.8 Hz, 1H), 5.40 (dd, *J* = 8.2, 5.3 Hz, 1H), 3.61 (d, *J* = 8.8 Hz, 1H), 3.40 (d, *J* = 8.8 Hz, 1H), 3.25 (s, 3H), 3.19 (s, 3H), 2.97 – 2.94 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 176.5, 144.1, 128.8, 128.1, 124.0, 122.5, 108.4, 76.5, 69.7, 59.5, 52.4, 45.6, 26.5.

HRMS (ESI) *m/z* calculated for C₁₃H₁₅Cl₂NNaO₂ [M+Na⁺] 310.0372, found 310.0375.

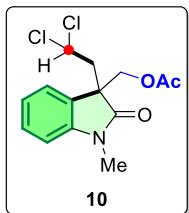


Purification of the crude product by flash column chromatography afforded the cyclization product **9** as pale gray solid in 83% yield (44 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.36 (td, *J* = 7.7, 1.2 Hz, 1H), 7.25 – 7.23 (m, 1H), 7.12 (td, *J* = 7.5, 0.8 Hz, 1H), 6.90 (d, *J* = 7.8 Hz, 1H), 5.43 (dd, *J* = 9.4, 4.3 Hz, 1H), 3.77 – 3.67 (m, 2H), 3.21 (s, 3H), 3.15 (dd, *J* = 14.9, 9.4 Hz, 1H), 2.85 (dd, *J* = 14.9, 4.2 Hz, 1H), 2.67 (br, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 177.7, 144.2, 129.2, 127.1, 123.3, 122.9, 108.8, 69.7, 67.7, 53.0, 45.5, 26.5.

HRMS (ESI) *m/z* calculated for C₁₂H₁₃Cl₂NNaO₂ [M+Na⁺] 296.0216, found 296.0214.

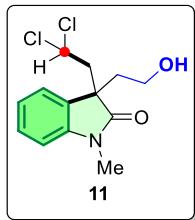


Purification of the crude product by flash column chromatography afforded the cyclization product **10** as colorless oil in 84% yield (53 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.36 (td, *J* = 7.8, 1.2 Hz, 1H), 7.25 – 7.23 (m, 1H), 7.10 (td, *J* = 7.5, 0.7 Hz, 1H), 6.89 (d, *J* = 7.8 Hz, 1H), 5.44 (dd, *J* = 9.2, 4.2 Hz, 1H), 4.41 (d, *J* = 10.8 Hz, 1H), 4.09 (d, *J* = 10.8 Hz, 1H), 3.22 (s, 3H), 3.03 (dd, *J* = 14.7, 9.3 Hz, 1H), 2.85 (dd, *J* = 14.7, 4.2 Hz, 1H), 1.94 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 175.7, 170.1, 144.2, 129.4, 126.7, 123.8, 122.7, 108.7, 69.1, 67.2, 51.0, 45.5, 26.6, 20.5.

HRMS (ESI) *m/z* calculated for C₁₄H₁₅Cl₂NNaO₃ [M+Na⁺] 338.0321, found 338.0320.

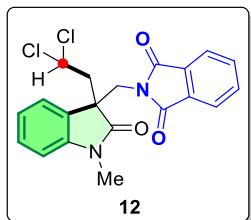


Purification of the crude product by flash column chromatography afforded the cyclization product **11** as colorless oil in 59% yield (34 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.34 (td, J = 7.7, 1.2 Hz, 1H), 7.20 – 7.18 (m, 1H), 7.14 – 7.10 (m, 1H), 6.89 (d, J = 7.8 Hz, 1H), 5.36 (dd, J = 9.1, 4.1 Hz, 1H), 3.56 – 3.49 (m, 1H), 3.45 – 3.39 (m, 1H), 3.20 (s, 3H), 3.13 – 3.06 (m, 1H), 2.76 (dd, J = 14.8, 4.1 Hz, 1H), 2.24 – 2.17 (m, 1H), 2.04 – 1.97 (m, 1H), 1.90 (br, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 178.9, 143.9, 129.0, 123.0, 122.9, 108.8, 69.3, 58.4, 49.9, 49.7, 41.2, 26.5.

HRMS (ESI) *m/z* calculated for C₁₃H₁₅Cl₂NNaO₂ [M+Na⁺] 310.0372, found 310.0372.

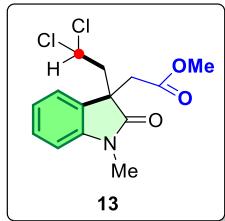


Purification of the crude product by flash column chromatography afforded the cyclization product **12** as white solid in 72% yield (58 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.80 (dd, J = 5.5, 3.0 Hz, 2H), 7.70 (dd, J = 5.5, 3.1 Hz, 2H), 7.29 (td, J = 7.8, 1.2 Hz, 1H), 7.20 – 7.18 (m, 1H), 7.03 (td, J = 7.5, 0.8 Hz, 1H), 6.83 (d, J = 7.8 Hz, 1H), 5.41 (dd, J = 8.6, 4.9 Hz, 1H), 4.01 (d, J = 14.1 Hz, 1H), 3.94 (d, J = 14.1 Hz, 1H), 3.22 – 3.16 (m, 4H), 3.04 (dd, J = 14.9, 4.9 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 176.1, 167.9, 144.0, 134.1, 131.6, 129.3, 126.8, 123.7, 123.5, 122.5, 108.7, 69.3, 51.4, 47.0, 44.1, 26.6.

HRMS (ESI) *m/z* calculated for C₂₀H₁₆Cl₂N₂NaO₃ [M+Na⁺] 425.0430, found 425.0429.



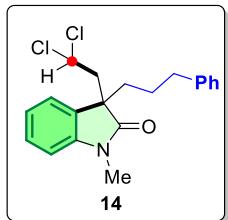
Purification of the crude product by flash column chromatography afforded the

cyclization product **13** as colorless oil in 87% yield (55 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.34 (td, *J* = 7.7, 1.2 Hz, 1H), 7.28 – 7.26 (m, 1H), 7.08 (td, *J* = 7.5, 0.8 Hz, 1H), 6.89 (d, *J* = 7.8 Hz, 1H), 5.42 (dd, *J* = 8.4, 4.6 Hz, 1H), 3.50 (s, 3H), 3.23 (s, 3H), 3.00 – 2.88 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 177.4, 169.5, 144.3, 129.2, 128.3, 123.3, 122.6, 108.6, 68.9, 51.7, 49.0, 48.5, 41.4, 26.6.

HRMS (ESI) *m/z* calculated for C₁₄H₁₅Cl₂NNaO₃ [M+Na⁺] 338.0321, found 338.0318.

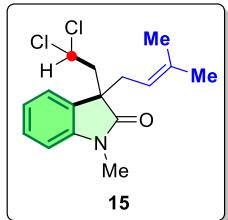


Purification of the crude product by flash column chromatography afforded the cyclization product **14** as colorless cream in 84% yield (61 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.28 (m, 1H), 7.23 – 7.19 (m, 2H), 7.15 – 7.06 (m, 3H), 7.01 (d, *J* = 7.0 Hz, 2H), 6.84 (d, *J* = 7.8 Hz, 1H), 5.34 (dd, *J* = 9.3, 4.0 Hz, 1H), 3.17 (s, 3H), 3.00 (dd, *J* = 14.8, 9.4 Hz, 1H), 2.68 (dd, *J* = 14.8, 4.0 Hz, 1H), 2.55 – 2.38 (m, 2H), 2.00 – 1.92 (m, 1H), 1.78 – 1.71 (m, 1H), 1.39 – 1.25 (m, 1H), 1.15 – 1.04 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 178.2, 144.2, 141.4, 129.1, 128.6, 128.3, 128.2, 125.8, 122.8, 122.6, 108.4, 69.5, 51.4, 49.9, 38.7, 35.6, 26.3, 25.0.

HRMS (ESI) *m/z* calculated for C₂₀H₂₁Cl₂NNaO [M+Na⁺] 384.0892, found 384.0894.

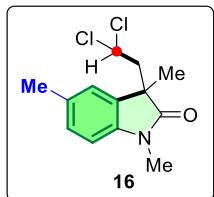


Purification of the crude product by flash column chromatography afforded the cyclization product **15** as colorless oil in 58% yield (36 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.31 (td, *J* = 7.7, 1.1 Hz, 1H), 7.17 (d, *J* = 7.2 Hz, 1H), 7.09 – 7.05 (m, 1H), 6.85 (d, *J* = 7.7 Hz, 1H), 5.37 (dd, *J* = 9.4, 4.2 Hz, 1H), 4.87 – 4.83 (m, 1H), 3.19 (s, 3H), 3.01 (dd, *J* = 14.8, 9.4 Hz, 1H), 2.76 (dd, *J* = 14.8, 4.2 Hz, 1H), 2.50 – 2.37 (m, 2H), 1.60 (s, 3H), 1.48 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.4, 144.0, 136.7, 129.2, 128.5, 123.4, 122.3, 116.5, 108.3, 69.9, 51.6, 48.3, 37.3, 26.4, 25.9, 18.0.

HRMS (ESI) m/z calculated for C₁₆H₁₉Cl₂NNaO [M+Na⁺] 334.0736, found 334.0735.

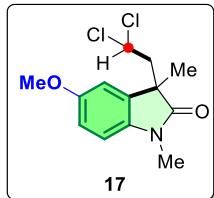


Purification of the crude product by flash column chromatography afforded the cyclization product **16** as colorless oil in 62% yield (34 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.12 – 7.10 (m, 1H), 7.00 – 6.99 (m, 1H), 6.76 (d, *J* = 7.9 Hz, 1H), 5.40 (dd, *J* = 9.4, 4.0 Hz, 1H), 3.18 (s, 3H), 3.02 (dd, *J* = 14.8, 9.3 Hz, 1H), 2.68 (dd, *J* = 14.8, 4.0 Hz, 1H), 2.36 (s, 3H), 1.38 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.9, 141.0, 132.2, 131.1, 128.9, 123.4, 108.3, 69.7, 50.2, 47.2, 26.4, 25.4, 21.1.

HRMS (ESI) m/z calculated for $C_{13}H_{15}Cl_2NNaO$ [M+Na $^+$] 294.0423, found 294.0424.

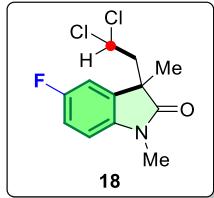


Purification of the crude product by flash column chromatography afforded the cyclization product **17** as pale yellow solid in 75% yield (43 mg).

¹H NMR (400 MHz, CDCl₃) δ 6.84 – 6.75 (m, 3H), 5.41 (dd, *J* = 9.1, 4.2 Hz, 1H), 3.80 (s, 3H), 3.17 (s, 3H), 3.01 (dd, *J* = 14.8, 9.2 Hz, 1H), 2.66 (dd, *J* = 14.8, 4.2 Hz, 1H), 1.37 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.6, 156.1, 136.8, 132.4, 112.3, 110.5, 108.8, 69.6, 55.8, 50.1, 47.5, 26.5, 25.4.

HRMS (ESI) m/z calculated for $C_{13}H_{15}Cl_2NNaO_2 [M+Na^+]$ 310.0372, found 310.0371.



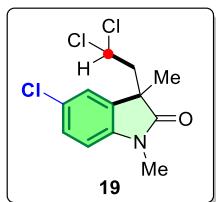
Purification of the crude product by flash column chromatography afforded the

cyclization product **18** as pale yellow solid in 89% yield (49 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.01 (td, *J* = 8.7, 2.5 Hz, 1H), 6.95 (dd, *J* = 7.7, 2.5 Hz, 1H), 6.79 (dd, *J* = 8.5, 4.0 Hz, 1H), 5.41 (dd, *J* = 8.8, 4.6 Hz, 1H), 3.18 (s, 3H), 3.01 (dd, *J* = 14.8, 8.8 Hz, 1H), 2.68 (dd, *J* = 14.9, 4.6 Hz, 1H), 1.38 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.5, 159.2 (d, *J* = 241.4 Hz), 139.3 (d, *J* = 1.6 Hz), 132.8 (d, *J* = 7.8 Hz), 114.8 (d, *J* = 23.4 Hz), 110.9 (d, *J* = 24.7 Hz), 109.1 (d, *J* = 8.1 Hz), 69.3, 49.9, 47.6 (d, *J* = 1.7 Hz), 26.5, 25.4.

HRMS (ESI) *m/z* calculated for C₁₂H₁₂Cl₂FNNaO [M+Na⁺] 298.0172, found 298.0172.

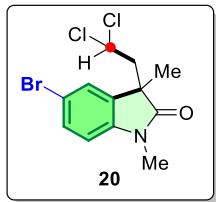


Purification of the crude product by flash column chromatography afforded the cyclization product **19** as white solid in 85% yield (50 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.29 (dd, *J* = 8.3, 2.1 Hz, 1H), 7.18 (d, *J* = 2.0 Hz, 1H), 6.79 (d, *J* = 8.3 Hz, 1H), 5.41 (dd, *J* = 9.0, 4.5 Hz, 1H), 3.80 (s, 3H), 3.19 (s, 3H), 3.03 (dd, *J* = 14.9, 9.0 Hz, 1H), 2.68 (dd, *J* = 14.9, 4.5 Hz, 1H), 1.39 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.4, 142.0, 132.9, 128.6, 128.1, 123.2, 109.5, 69.3, 49.9, 47.4, 26.6, 25.4.

HRMS (ESI) *m/z* calculated for C₁₂H₁₂Cl₃NNaO [M+Na⁺] 313.9877, found 313.9877.

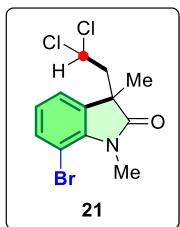


Purification of the crude product by flash column chromatography afforded the cyclization product **20** as pale yellow solid in 87% yield (58 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.43 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.31 (d, *J* = 1.7 Hz, 1H), 6.75 (d, *J* = 8.4 Hz, 1H), 5.40 (dd, *J* = 9.2, 4.8 Hz, 1H), 3.18 (s, 3H), 3.01 (dd, *J* = 14.8, 8.8 Hz, 1H), 2.68 (dd, *J* = 14.8, 4.4 Hz, 1H), 1.38 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.3, 142.2, 133.2, 131.5, 125.9, 115.3, 110.0, 69.3, 49.8, 47.3, 26.5, 25.4.

HRMS (ESI) m/z calculated for $C_{12}H_{12}BrCl_2NNaO$ [M+Na⁺] 357.9372, found 357.9370.

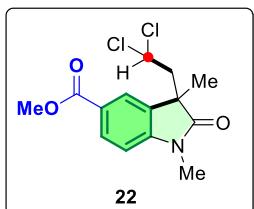


Purification of the crude product by flash column chromatography afforded the cyclization product **21** as colorless oil in 77% yield (52 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (dd, $J = 8.2, 1.0$ Hz, 1H), 7.10 (dd, $J = 7.3, 1.0$ Hz, 1H), 6.95 – 6.92 (m, 1H), 5.38 (dd, $J = 9.0, 4.4$ Hz, 1H), 3.58 (s, 3H), 3.02 (dd, $J = 14.8, 9.0$ Hz, 1H), 2.67 (dd, $J = 14.8, 4.4$ Hz, 1H), 1.38 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.3, 140.8, 134.3, 134.2, 123.8, 121.7, 103.0, 69.3, 50.2, 46.9, 30.0, 25.8.

HRMS (ESI) m/z calculated for $C_{12}H_{12}BrCl_2NNaO$ [M+Na⁺] 357.9372, found 357.9371.

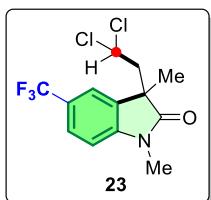


Purification of the crude product by flash column chromatography afforded the cyclization product **22** as pale yellow solid in 57% yield (36 mg).

¹H NMR (400 MHz, CDCl₃) δ 8.06 (dd, $J = 8.2, 1.7$ Hz, 1H), 7.87 (d, $J = 1.5$ Hz, 1H), 6.90 (d, $J = 8.2$ Hz, 1H), 5.37 (dd, $J = 9.2, 4.3$ Hz, 1H), 3.92 (s, 3H), 3.23 (s, 3H), 3.05 (dd, $J = 14.9, 9.3$ Hz, 1H), 2.75 (dd, $J = 14.9, 4.3$ Hz, 1H), 1.41 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.3, 166.6, 147.5, 131.3, 131.1, 124.6, 123.9, 108.1, 69.4, 52.1, 49.8, 47.0, 26.6, 25.4.

HRMS (ESI) m/z calculated for $C_{14}H_{15}Cl_2NNaO_3$ [M+Na⁺] 338.0321, found 338.0318.

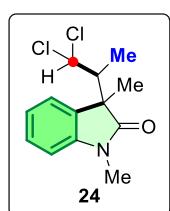


Purification of the crude product by flash column chromatography afforded the cyclization product **23** as colorless oil in 25% yield (16 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.61 (d, *J* = 8.1 Hz, 1H), 7.43 (s, 1H), 6.95 (d, *J* = 8.2 Hz, 1H), 5.39 (dd, *J* = 8.9, 4.6 Hz, 1H), 3.24 (s, 3H), 3.06 (dd, *J* = 14.9, 8.9 Hz, 1H), 2.75 (dd, *J* = 14.9, 4.6 Hz, 1H), 1.42 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.9, 146.3, 131.8, 126.5 (q, *J* = 3.9 Hz, 1H), 125.0 (q, *J* = 32.8 Hz, 1H), 124.2 (q, *J* = 271.6 Hz, 1H), 119.7 (d, *J* = 3.5 Hz, 1H), 108.4, 69.2 (d, *J* = 5.6 Hz, 1H), 49.8, 47.1, 26.7, 25.4.

HRMS (ESI) m/z calculated for C₁₃H₁₂Cl₂F₃NNaO [M+Na⁺] 348.0140, found 348.0141.

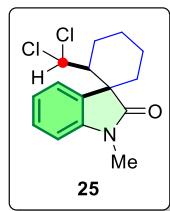


Purification of the crude product by flash column chromatography afforded the cyclization product **24** as colorless oil in 68% yield (37 mg, a mixture of diastereoisomers, d.r. ~ 4.1:1).

¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.26 (m, 1H), 7.21 (d, *J* = 7.1 Hz, 1H), 7.11 – 7.03 (m, 1H), 6.88 – 6.84 (m, 1H), 5.77 (d, *J* = 2.8 Hz, 0.19H), 5.65 (d, *J* = 1.6 Hz, 0.77H), 3.24 (s, 0.6H), 3.17 (s, 2.4H), 2.82 – 2.76 (m, 0.2H), 2.71 – 2.66 (m, 0.8H), 1.54 – 1.51 (m, 3H), 1.45 – 1.42 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.3, 178.0, 143.2, 131.9, 131.0, 128.5, 128.2, 124.3, 122.8, 122.6, 122.5, 108.4, 108.3, 75.3, 75.0, 51.6, 50.6, 50.0, 49.5, 26.3, 26.2, 23.5, 22.8, 9.0, 7.5.

HRMS (ESI) m/z calculated for $C_{13}H_{15}Cl_2NNaO [M+Na^+]$ 294.0423, found 294.0421.



Purification of the crude product by flash column chromatography afforded the cyclization product **25** in 41% yield (25 mg, diastereoisomers can be isolated with d.r.

~ 1.2:1).

25-up (pale yellow solid):

¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.28 (m, 1H), 7.13 – 7.07 (m, 2H), 6.84 (d, *J* = 7.8 Hz, 1H), 5.35 (s, 1H), 3.15 (s, 3H), 2.54 – 2.46 (m, 2H), 2.43 – 2.31 (m, 1H), 2.14 – 2.11 (m, 2H), 1.82 – 1.73 (m, 2H), 1.60 – 1.47 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 177.1, 143.3, 131.8, 128.5, 122.7, 121.6, 108.1, 73.9, 55.8, 49.3, 36.6, 26.1, 25.5, 20.1, 19.3.

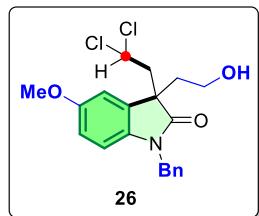
HRMS (ESI) *m/z* calculated for C₁₅H₁₇Cl₂NNaO [M+Na⁺] 320.0579, found 320.0580.

25-down (pale yellow solid):

¹H NMR (400 MHz, CDCl₃) δ 7.51 (d, *J* = 7.5 Hz, 1H), 7.33 (t, *J* = 7.8 Hz, 1H), 7.04 (t, *J* = 7.4 Hz, 1H), 6.90 (d, *J* = 7.8 Hz, 1H), 5.24 (d, *J* = 6.2 Hz, 1H), 3.23 (s, 3H), 2.83 – 2.77 (m, 1H), 2.41 – 2.36 (m, 1H), 2.11 – 2.08 (m, 1H), 1.95 – 1.88 (m, 1H), 1.85 – 1.65 (m, 4H), 1.48 – 1.42 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 179.6, 143.4, 130.0, 128.2, 125.7, 121.8, 108.6, 75.1, 51.5, 51.5, 36.6, 26.6, 24.9, 24.6, 20.9.

HRMS (ESI) *m/z* calculated for C₁₅H₁₇Cl₂NNaO [M+Na⁺] 320.0579, found 320.0581.

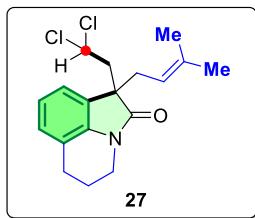


Purification of the crude product by flash column chromatography afforded the cyclization product **26** as pale yellow oil in 49% yield (38 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.27 (m, 5H), 6.79 (d, *J* = 2.4 Hz, 1H), 6.75 – 6.72 (m, 1H), 6.69 – 6.67 (m, 1H), 5.44 (dd, *J* = 8.8, 4.4 Hz, 1H), 5.01 (d, *J* = 15.6 Hz, 1H), 4.72 (d, *J* = 15.6 Hz, 1H), 3.60 – 3.54 (m, 1H), 3.51 – 3.45 (m, 1H), 3.13 (dd, *J* = 14.8, 8.8 Hz, 1H), 2.76 (dd, *J* = 14.8, 4.4 Hz, 1H), 2.28 – 2.21 (m, 1H), 2.06 – 1.99 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 178.6, 156.2, 136.6, 135.6, 130.6, 128.8, 127.7, 127.6, 112.6, 110.8, 110.2, 69.2, 58.5, 55.8, 50.3, 49.8, 44.5, 41.6

HRMS (ESI) *m/z* calculated for C₂₀H₂₁Cl₂NNaO₃ [M+Na⁺] 416.0791, found 416.0785.

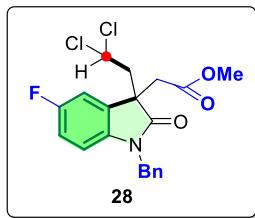


Purification of the crude product by flash column chromatography afforded the cyclization product **27** as colorless oil in 74% yield (50 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.06 – 7.04 (m, 1H), 7.01 – 7.00 (m, 1H), 6.97 – 6.93 (m, 1H), 5.37 (dd, *J* = 9.6, 3.9 Hz, 1H), 4.93 – 4.89 (m, 1H), 3.73 – 3.62 (m, 2H), 2.99 (dd, *J* = 14.8, 9.6 Hz, 1H), 2.79 – 2.73 (m, 3H), 2.48 (dd, *J* = 14.0, 7.7 Hz, 1H), 2.36 (dd, *J* = 14.0, 7.5 Hz, 1H), 1.99 – 1.93 (m, 2H), 1.62 (s, 3H), 1.50 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 177.3, 139.8, 136.6, 127.6, 127.3, 121.7, 121.3, 120.4, 116.8, 70.2, 52.8, 48.0, 39.0, 36.9, 25.9, 24.7, 21.1, 18.1.

HRMS (ESI) *m/z* calculated for C₁₈H₂₁Cl₂NNaO [M+Na⁺] 360.0892, found 360.0890.

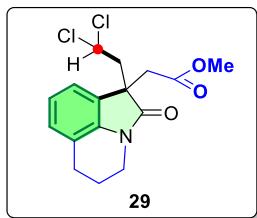


Purification of the crude product by flash column chromatography afforded the cyclization product **28** as pale yellow solid in 59% yield (48 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.28 (m, 5H), 7.07 (dd, *J* = 7.8, 2.6 Hz, 1H), 6.91 (td, *J* = 8.8, 2.6 Hz, 1H), 6.68 (dd, *J* = 8.6, 4.2 Hz, 1H), 5.51 (dd, *J* = 7.8, 5.2 Hz, 1H), 5.01 (d, *J* = 15.6 Hz, 1H), 4.81 (d, *J* = 15.7 Hz, 1H), 3.51 (s, 3H), 3.03 – 2.90 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 177.2, 169.3, 159.1 (d, *J* = 241.9 Hz), 139.5 (d, *J* = 2.1 Hz), 135.2, 130.1 (d, *J* = 8.0 Hz), 128.8, 127.8, 127.5, 115.3 (d, *J* = 23.2 Hz), 111.8 (d, *J* = 25.0 Hz), 110.3 (d, *J* = 8.1 Hz), 68.6, 51.9, 49.0, 48.9 (d, *J* = 1.8 Hz), 44.6, 41.4.

HRMS (ESI) *m/z* calculated for C₂₀H₁₈Cl₂FNNaO₃ [M+Na⁺] 432.0540, found 432.0539.

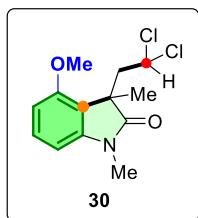


Purification of the crude product by flash column chromatography afforded the cyclization product **29** as pale yellow oil in 67% yield (46 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.10 – 7.06 (m, 2H), 6.97 – 6.93 (m, 1H), 5.42 (t, *J* = 6.5 Hz, 1H), 3.75 – 3.65 (m, 2H), 3.51 (s, 3H), 2.94 (d, *J* = 6.5 Hz, 2H), 2.91 – 2.81 (m, 2H), 2.77 (t, *J* = 6.1 Hz, 2H), 2.04 – 1.93 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 176.2, 169.7, 140.0, 127.9, 126.6, 122.1, 121.3, 120.7, 69.3, 51.7, 49.6, 48.6, 41.1, 39.1, 24.6, 20.9.

HRMS (ESI) *m/z* calculated for C₁₆H₁₇Cl₂NNaO₃ [M+Na⁺] 364.0478, found 364.0478.

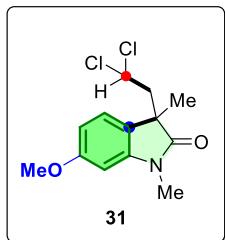


Purification of the crude product by flash column chromatography afforded the cyclization product **30** as pale yellow solid in 56% yield (32 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.31 – 7.27 (m, 1H), 6.64 (d, *J* = 8.5 Hz, 1H), 6.54 (d, *J* = 7.8 Hz, 1H), 5.33 (dd, *J* = 8.3, 5.3 Hz, 1H), 3.88 (s, 3H), 3.18 (s, 3H), 2.99 – 2.97 (m, 2H), 1.41 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.6, 155.8, 144.7, 129.9, 116.3, 105.6, 101.9, 70.6, 55.5, 48.4, 47.5, 26.6, 23.1.

HRMS (ESI) *m/z* calculated for C₁₃H₁₅Cl₂NNaO₂ [M+Na⁺] 310.0372, found 310.0373.

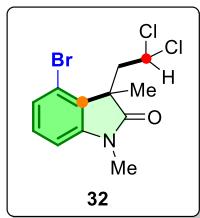


Purification of the crude product by flash column chromatography afforded the cyclization product **31** as colorless oil in 29% yield (17 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, *J* = 8.2 Hz, 1H), 6.59 (dd, *J* = 8.1, 2.3 Hz, 1H), 6.45 (d, *J* = 2.2 Hz, 1H), 5.37 (dd, *J* = 9.4, 4.0 Hz, 1H), 3.84 (s, 3H), 3.18 (s, 3H), 3.00 (dd, *J* = 14.8, 9.4 Hz, 1H), 2.66 (dd, *J* = 14.8, 4.0 Hz, 1H), 1.36 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 179.6, 160.5, 144.7, 123.3, 122.8, 106.5, 96.6, 69.8, 55.5, 50.3, 46.7, 26.5, 25.5.

HRMS (ESI) *m/z* calculated for C₁₃H₁₅Cl₂NNaO₂ [M+Na⁺] 310.0372, found 310.0374.

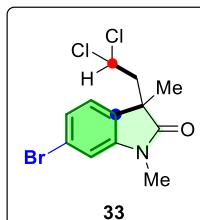


Purification of the crude product by flash column chromatography afforded the cyclization product **32** as white solid in 59% yield (40 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.22 – 7.17 (m, 2H), 6.82 (dd, *J* = 5.9, 2.9 Hz, 1H), 5.38 (dd, *J* = 10.1, 3.6 Hz, 1H), 3.25 – 3.20 (m, 4H), 2.98 (dd, *J* = 14.8, 10.1 Hz, 1H), 1.50 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.6, 145.6, 130.2, 128.9, 126.5, 118.9, 107.6, 70.0, 49.2, 47.6, 26.7, 22.5.

HRMS (ESI) *m/z* calculated for C₁₂H₁₂BrCl₂NNaO [M+Na⁺] 357.9372, found 357.9370.

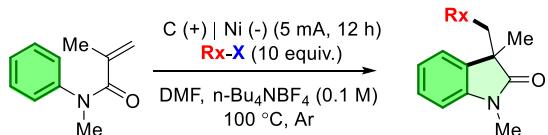


Purification of the crude product by flash column chromatography afforded the cyclization product **33** as colorless oil in 22% yield (15 mg).

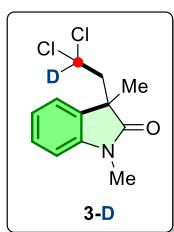
¹H NMR (400 MHz, CDCl₃) δ 7.24 (dd, *J* = 7.8, 1.7 Hz, 1H), 7.06 (d, *J* = 7.8 Hz, 1H), 7.02 (d, *J* = 1.7 Hz, 1H), 5.38 (dd, *J* = 9.0, 4.4 Hz, 1H), 3.19 (s, 3H), 3.02 (dd, *J* = 14.9, 9.0 Hz, 1H), 2.69 (dd, *J* = 14.9, 4.4 Hz, 1H), 1.38 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.7, 144.8, 130.0, 125.4, 124.0, 122.2, 112.2, 69.4, 49.9, 47.0, 26.6, 25.4.

HRMS (ESI) m/z calculated for $\text{C}_{12}\text{H}_{12}\text{BrCl}_2\text{NNaO}$ [$\text{M}+\text{Na}^+$] 357.9372, found 357.9367.



General procedure 2: To an oven-dried Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 0.2 mmol, 35 mg) and $n\text{-Bu}_4\text{NBF}_4$ (0.1 M, 0.4 mmol, 132 mg) were added. The rubber plug equipped with a carbon felt anode (10 mm * 10 mm) and a foam Ni cathode (10 mm* mm) was used to keep the Schlenk tube sealed. The tube was evacuated and filled with Ar, repeating three times. halomethanes (10 equiv., 2 mmol) was dissolved in 4 mL DMF and then injected into the tube via syringe. The mixture was stirred for 5 min and put in the oil bath (100 °C) with the electrolysis of 5 mA, 12h. After the reaction, the tube was cooled to room temperature and ethyl acetate (10 mL) and water (10 mL) was added. The aqueous layer was separated and extracted with ethyl acetate (3×10 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield the desired product.

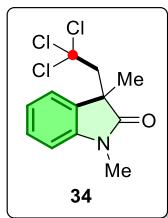


Purification of the crude product by flash column chromatography afforded the cyclization product **3-D** as colorless oil in 83% yield (43 mg).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.31 (td, $J = 7.7, 1.2$ Hz, 1H), 7.20 – 7.18 (m, 1H), 7.11 – 7.07 (m, 1H), 6.87 (d, $J = 7.8$ Hz, 1H), 3.20 (s, 3H), 3.02 (d, $J = 14.8$ Hz, 1H), 2.69 (d, $J = 14.8$ Hz, 1H), 1.39 (s, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 178.9, 143.3, 131.0, 128.6, 122.6, 122.6, 108.6, 69.4 ($J = 27.3$ Hz), 49.9, 47.1, 26.4, 25.4.

HRMS (ESI) m/z calculated for $\text{C}_{12}\text{H}_{12}\text{Cl}_2\text{DNNaO}$ [$\text{M}+\text{Na}^+$] 281.0329, found 281.0327.

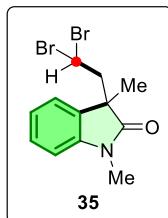


Purification of the crude product by flash column chromatography afforded the cyclization product **34** as pale yellow solid in 17% yield (10 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.35 (d, *J* = 7.4 Hz, 1H), 7.33 – 7.29 (m, 1H), 7.06 (t, *J* = 7.5 Hz, 1H), 6.88 (d, *J* = 7.8 Hz, 1H), 3.69 (d, *J* = 15.2 Hz, 1H), 3.34 (d, *J* = 15.3 Hz, 1H), 3.23 (s, 3H), 1.39 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.6, 143.2, 129.5, 128.5, 125.6, 122.0, 108.4, 96.1, 59.8, 48.0, 26.8, 26.6.

HRMS (ESI) *m/z* calculated for C₁₂H₁₂Cl₃NNaO [M+Na⁺] 313.9877, found 313.9875.

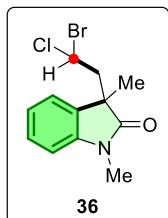


Purification of the crude product by flash column chromatography afforded the cyclization product **35** as colorless oil in 64% yield (44 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.32 (td, *J* = 7.7, 1.3 Hz, 1H), 7.19 – 7.17 (m, 1H), 7.12 – 7.08 (m, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 5.30 (dd, *J* = 9.6, 4.3 Hz, 1H), 3.27 (dd, *J* = 15.1, 9.6 Hz, 1H), 3.20 (s, 3H), 2.99 (dd, *J* = 15.2, 4.3 Hz, 1H), 1.37 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.8, 143.5, 130.7, 128.6, 122.6, 108.6, 51.5, 48.5, 39.6, 26.5, 25.6.

HRMS (ESI) *m/z* calculated for C₁₂H₁₃Br₂NNaO [M+Na⁺] 367.9256, found 367.9254.

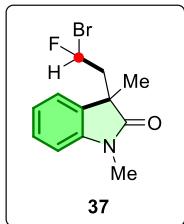


Purification of the crude product by flash column chromatography afforded the cyclization product **36** as colorless oil in 66% yield (40 mg, a mixture of diastereoisomers, d.r. ~ 1.2:1).

¹H NMR (400 MHz, CDCl₃) δ 7.32 (tdd, *J* = 7.7, 3.8, 1.1 Hz, 1H), 7.20 – 7.18 (m, 1H), 7.12 – 7.08 (m, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 5.43 – 5.36 (m, 1H), 3.21 – 3.09 (m, 4H), 2.87 – 2.81 (m, 1H), 1.39 (s, 1.3H), 1.38 (s, 1.5H).

¹³C NMR (101 MHz, CDCl₃) δ 179.0, 178.7, 143.5, 143.4, 131.1, 130.7, 128.6, 128.5, 122.7, 122.6, 122.6, 108.6, 108.6, 55.7, 55.6, 51.1, 50.8, 48.0, 47.8, 26.4, 25.6, 25.3.

HRMS (ESI) *m/z* calculated for C₁₂H₁₃BrClNNaO [M+Na⁺] 323.9761, found 323.9762.



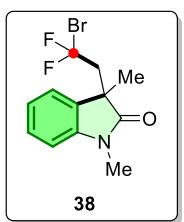
Purification of the crude product by flash column chromatography afforded the cyclization product **37** as colorless oil in 13% yield (8 mg, a mixture of diastereoisomers, d.r. ~ 1.5:1).

¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.28 (m, 1H), 7.23 – 7.19 (m, 1H), 7.11 – 7.08 (m, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 6.28 (dd, *J* = 7.4, 4.6 Hz, 0.2H), 6.25 (dd, *J* = 9.9, 2.0 Hz, 0.3H), 6.16 (dd, *J* = 7.4, 4.6 Hz, 0.2H), 6.12 (dd, *J* = 9.9, 2.0 Hz, 0.3H), 3.23 (s, 1.2H), 3.20 (s, 1.8H), 3.14 – 3.05 (m, 0.66H), 2.92 – 2.70 (m, 0.86H), 2.56 (dd, *J* = 14.9, 2.0 Hz, 0.32H), 2.46 (dd, *J* = 14.9, 2.0 Hz, 0.32H), 1.42 (s, 1.2H), 1.41 (s, 1.8H).

¹³C NMR (101 MHz, CDCl₃) δ 179.0, 178.7, 143.3, 142.9, 131.8, 131.3, 128.6, 128.4, 122.9, 122.8, 122.7, 122.7, 108.5, 108.5, 92.5 (d, *J* = 43.2 Hz), 90.0 (d, *J* = 43.2 Hz), 48.5 (d, *J* = 19.7 Hz), 47.6 (d, *J* = 19.4 Hz), 47.0, 46.9, 46.9, 26.4, 26.3, 24.7, 24.0.

¹⁹F NMR (376 MHz, CDCl₃) -130.1 (s, 1F), -133.8 (s, 1.5F).

HRMS (ESI) *m/z* calculated for C₁₂H₁₃BrFNNaO [M+Na⁺] 308.0057, found 308.0055.



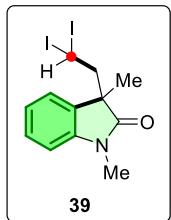
Purification of the crude product by flash column chromatography afforded the cyclization product **38** as colorless oil in 38% yield (23 mg).

¹H NMR (400 MHz, CDCl₃) δ 7.31 (td, *J* = 7.7, 1.1 Hz, 1H), 7.27 – 7.25 (m, 1H), 7.08 (td, *J* = 7.5, 0.7 Hz, 1H), 6.88 (d, *J* = 7.8 Hz, 1H), 3.34 – 3.22 (m, 4H), 3.10 – 2.98 (m, 1H), 1.38 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.3, 142.8, 130.8, 128.4, 123.8 (t, *J* = 1.7 Hz), 122.5, 119.5 (t, *J* = 307.5 Hz), 108.4, 50.9 (t, *J* = 21.0 Hz), 45.9, 26.4, 25.3.

¹⁹F NMR (376 MHz, CDCl₃) δ ppm –39.9 (d, *J* = 155.0 Hz, 1F), -43.9 (d, *J* = 154.9 Hz, 1F).

HRMS (ESI) *m/z* calculated for C₁₂H₁₂BrF₂NNaO [M+Na⁺] 325.9963, found 325.9963.



Purification of the crude product by flash column chromatography afforded the cyclization product **39** as colorless cream in 29% yield (26 mg).

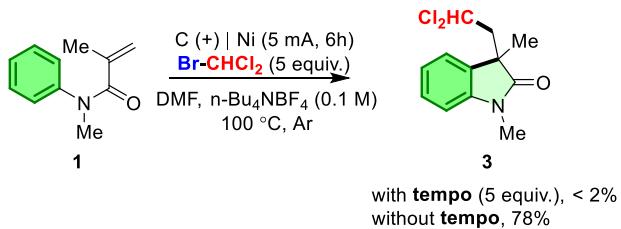
¹H NMR (400 MHz, CDCl₃) δ 7.33 (td, *J* = 7.7, 1.4 Hz, 1H), 7.17 – 7.15 (m, 1H), 7.12 – 7.08 (m, 1H), 6.87 (d, *J* = 7.8 Hz, 1H), 4.68 (dd, *J* = 9.7, 4.9 Hz, 1H), 3.35 (dd, *J* = 15.3, 9.7 Hz, 1H), 3.21 – 3.15 (m, 4H), 1.31 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 178.5, 143.9, 130.3, 128.6, 122.7, 122.7, 108.7, 54.0, 50.8, 26.6, 25.9, -38.6.

HRMS (ESI) *m/z* calculated for C₁₂H₁₃I₂NNaO [M+Na⁺] 463.8979, found 463.8977.

5. Mechanistic studies

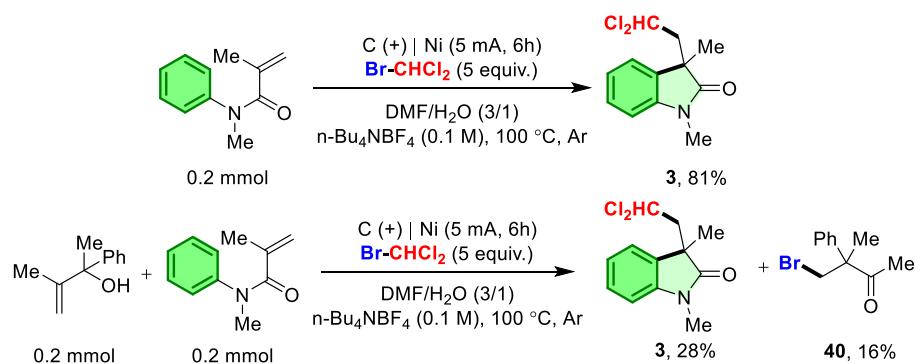
a) Radical scavenging experiments



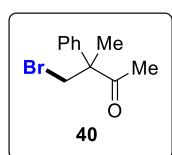
To an oven-dried Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 0.2 mmol, 35 mg), n-Bu₄NBF₄ (0.1 M, 0.4 mmol, 132 mg) and radical scavenger (tempo, 5 equiv., 156 mg) were added. The rubber plug equipped with a carbon felt

anode (5 mm * 5 mm) and a foam Ni cathode (5 mm* 5 mm) was used to keep the Schlenk tube sealed. The tube was evacuated and filled with Ar, repeating three times. BrCHCl₂ (5 equiv., 2 mmol, 83 µL) was dissolved in 4 mL DMF and then injected into the tube via syringe. The mixture was stirred for 5 min and put in the oil bath (100 °C) with the electrolysis of 5 mA, 6h. After the reaction, the tube was cooled to room temperature and trace **3** was detected by thin layer chromatography.

b) Detection of generated bromide ion



To an oven-dried Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 0.2 mmol, 35 mg), n-Bu₄NBF₄ (0.1 M, 0.4 mmol, 132 mg) and allyl alcohol (1 equiv., 0.2 mmol, 33 mg) were added. The rubber plug equipped with a carbon felt anode (5 mm * 5 mm) and a foam Ni cathode (5 mm* 5 mm) was used to keep the Schlenk tube sealed. The tube was evacuated and filled with Ar, repeating three times. BrCHCl₂ (5 equiv., 2 mmol, 83 µL) was dissolved in DMF/H₂O (3/1 mL) and then injected into the tube via syringe. The mixture was stirred for 5 min and put in the oil bath (100 °C) with the electrolysis of 5 mA, 6h. After the reaction, the tube was cooled to room temperature and ethyl acetate (10 mL) and water (10 mL) was added. The aqueous layer was separated and extracted with ethyl acetate (3×10 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield **3** in 28% yield and **40** in 16% yield.

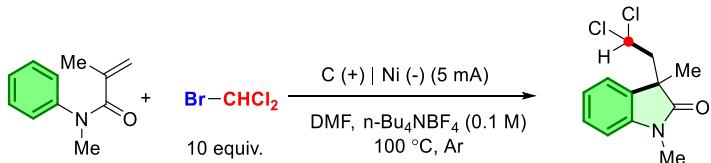


¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.36 (m, 2H), 7.33 – 7.30 (m, 1H), 7.26 – 7.24

(m, 2H), 3.95 (d, $J = 10.4$ Hz, 1H), 3.71 (d, $J = 10.5$ Hz, 1H), 2.00 (s, 3H), 1.72 (s, 3H).

The ^1H NMR spectra is consistent with the literature.

c) Reaction process analysis



To an oven-dried Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 0.2 mmol, 35 mg) and n-Bu₄NBF₄ (0.1 M, 0.4 mmol, 132 mg) were added. The rubber plug equipped with a carbon felt anode (5 mm * 5 mm) and a foam Ni cathode (5 mm * 5 mm) was used to keep the Schlenk tube sealed. The tube was evacuated and filled with Ar, repeating three times. BrCHCl₂ (10 equiv., 2 mmol, 165 μL) was dissolved in 4 mL DMF and then injected into the tube via syringe. The mixture was stirred for 5 min and put in the oil bath (100 °C) with the electrolysis of 5 mA for different reaction time (1h, 2h, 3h, 6h, 12h). After the reaction, the tube was cooled to room temperature and ethyl acetate (10 mL) and water (10 mL) was added. The aqueous layer was separated and extracted with ethyl acetate (3×10 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield the desired product and the acrylamide was recycled.

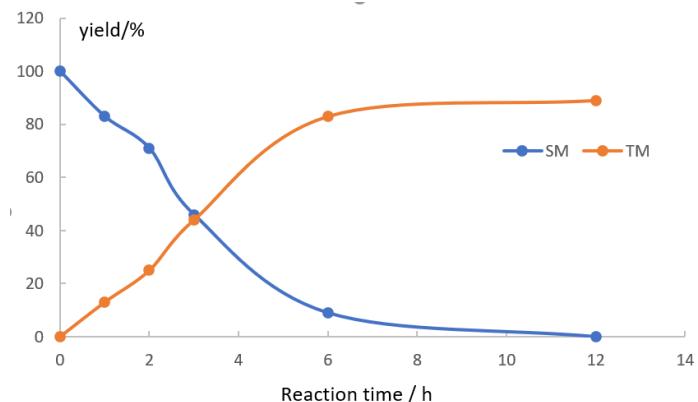
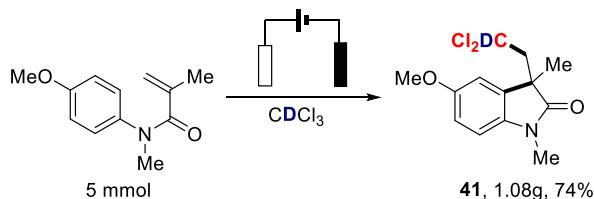


Fig. S3. Reaction process analysis.

6. Synthetic applications

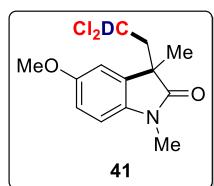
(a) Gram-scale synthesis



To an oven-dried 100 mL Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 5 mmol, 1.025 g), CDCl_3 (10 equiv., 50 mmol, 4.05 mL), $n\text{-Bu}_4\text{NBF}_4$ (0.1 M, 5 mmol, 1.65 g) and dry DMF (50 mL) were added. The tube was flushed with Ar for 3 min to exclude the air. The rubber plug equipped with a carbon felt anode (25 mm * 20 mm) and a foam Ni cathode (25 mm * 20 mm) was used to keep the Schlenk tube sealed. The mixture was stirred for 5 min and put in the oil bath (100°C) with the electrolysis of 35 mA, 72 h. After the reaction, the tube was cooled to room temperature and ethyl acetate (50 mL) and water (200 mL) was added. The aqueous layer was separated and extracted with ethyl acetate (3×50 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield the desired product **41** as white solid (1.08 g, 74%).



Fig. S4. The electrochemical reaction setup for gram-scale synthesis.

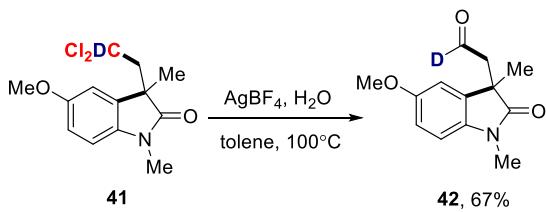


¹H NMR (400 MHz, CDCl₃) δ 6.83 – 6.75 (m, 3H), 3.79 (s, 3H), 3.16 (s, 3H), 2.99 (d, *J* = 14.8 Hz, 1H), 2.65 (d, *J* = 14.8 Hz, 1H), 1.36 (s, 3H).

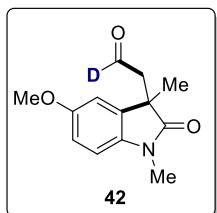
¹³C NMR (101 MHz, CDCl₃) δ 178.5, 156.0, 136.8, 132.4, 112.3, 110.4, 108.8, 69.3 (t, *J* = 27.1 Hz), 55.7, 49.9, 47.5, 26.4, 25.4.

HRMS (ESI) m/z calculated for $C_{13}H_{14}Cl_2DNNaO_2$ [M+Na $^+$] 311.0435, found 311.0432.

(b) Synthesis of deuterated (\pm) -Physostigmine



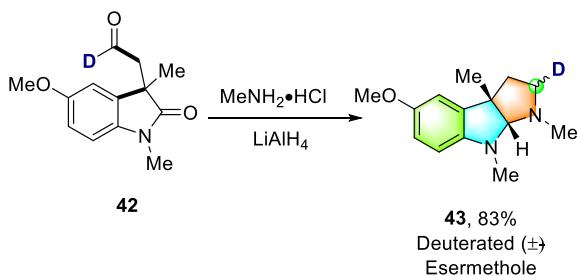
To an oven-dried 10 mL reaction tube equipped with a magnetic stir bar, **41** (1 equiv., 0.2 mmol, 58 mg), AgBF₄ (4 equiv., 0.8 mmol, 160 mg), Celite (100 mg), H₂O (100 μL) and toluene (1 mL) were added. The tube was flushed with Ar for 1 min to exclude the air. The tube was sealed and put in the oil bath (100 °C) with vigorous stirring. After the reaction, the tube was cooled to room temperature and the reaction mixture was subjected to flash column chromatography on silica gel directly to yield the desired product **42** as white solid (31 mg, 67%).



¹H NMR (400 MHz, CDCl₃) δ 6.81 – 6.76 (m, 3H), 3.78 (s, 3H), 3.24 (s, 3H), 2.97 (d, *J* = 17.2 Hz, 1H), 2.90 (d, *J* = 17.2 Hz, 1H), 1.41 (s, 3H).

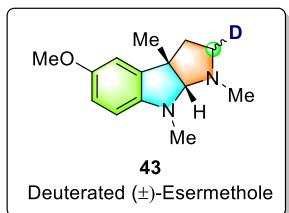
¹³C NMR (101 MHz, CDCl₃) δ 198.3 (t, *J* = 26.8 Hz), 178.9, 155.8, 136.4, 133.9, 111.9, 110.1, 108.4, 55.5, 50.1 (t, *J* = 3.6 Hz), 45.1, 26.2, 23.7.

HRMS (ESI) *m/z* calculated for C₁₃H₁₄DNNaO₃ [M+Na⁺] 257.1007, found 257.1006.



To an oven-dried 100 mL flask equipped with a magnetic stir bar, **42** (1 equiv., 0.8 mmol, 180 mg), MeNH₂ · HCl (10 equiv., 8 mmol, 540 mg), Et₃N (10 equiv., 8 mmol, 1.115 mL), MgSO₄ (730 mg) and dry THF (45 mL) were added. The tube was flushed with Ar for 3 min to exclude the air. The tube was sealed and reacted for 16 h at room temperature. LiAlH₄ (10 equiv., 8 mmol, 306 mg) was then added and the mixture was refluxed at 80 °C for 1.5 h. After the reaction, the tube was cooled to room temperature

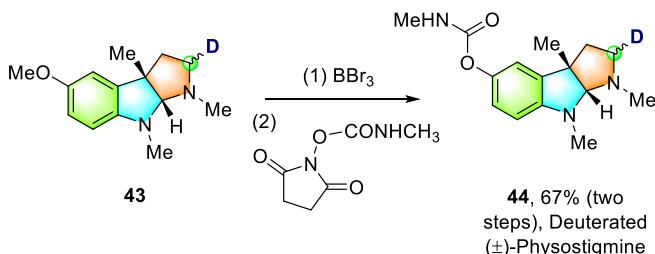
and ethyl acetate (30 mL) and saturated NaHCO₃ aq. solution (30 mL) were added. The aqueous layer was separated and extracted with ethyl acetate (3×20 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield the desired product **43** as colorless oil (155 mg, 83%, a mixture of diastereoisomers, d.r. ~ 1.1:1).



¹H NMR (400 MHz, CDCl₃) δ 6.66 – 6.63 (m, 2H), 6.35 (d, *J* = 8.1 Hz, 1H), 4.06 (s, 0.52H), 4.05 (s, 0.43H), 3.74 (s, 3H), 2.89 (s, 3H), 2.70 (t, *J* = 5.2 Hz, 0.51H), 2.61 (t, *J* = 7.5 Hz, 0.58H), 2.53 (s, 3H), 1.94 (d, *J* = 7.1 Hz, 2H), 1.43 (s, 3H).

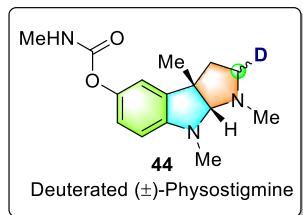
¹³C NMR (101 MHz, CDCl₃; values in brackets were corresponding to the same carbon in diastereoisomers) δ 152.9, 146.5 (146.4), 138.2, 112.1, 109.7, 107.4, 98.3 (98.2), 55.9, 52.8 (td, *J* = 21.4, 3.1 Hz), 52.7, 40.7 (40.6), 38.1 (38.0), 37.9 (37.8), 27.4 (27.3).

HRMS (ESI) *m/z* calculated for C₁₄H₂₀DN₂O [M+H⁺] 234.1711, found 234.1711.



To an oven-dried 50 mL flask equipped with a magnetic stir bar, **43** (1 equiv., 0.5 mmol, 116 mg) was dissolved in dry DCM (10 mL). BBr₃ solution (1 M in DCM, 3 equiv., 1.5 mL) was slowly added at 0 °C and then reacted at rt for 5 h. After the reaction, the tube was cooled to 0 °C and quenched with H₂O (10 mL). The aqueous layer was separated and extracted with DCM (3×10 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was directly subjected to the next step of reaction. The resulting phenol was dissolved in dry THF (10 mL) and was added NaH (60% in mineral oil, 2.2 equiv., 1.1 mmol, 44 mg) slowly at 0 °C. After 10 min, N-succinimidyl-N-

methylcarbamate (1.1 equiv., 0.55 mmol, 95 mg) was added and reacted at room temperature for 2 h. Then the reaction was quenched with water and extracted with ethyl acetate. The organic layers were then combined, washed with brine, dried by Na₂SO₄, and concentrated. The residue was purified by using flashcolumn chromatography to give **44** as pale yellow oil (92 mg, 67% for two steps, a mixture of diastereoisomers, d.r. ~ 1.1:1).

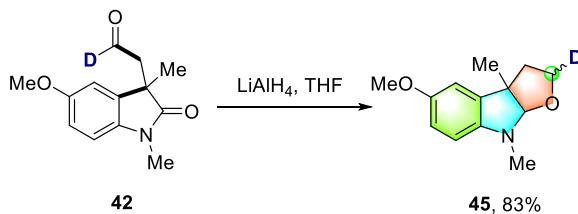


¹H NMR (400 MHz, CDCl₃) δ 6.79 (dd, *J* = 8.4, 2.2 Hz, 1H), 6.75 (d, *J* = 2.2 Hz, 1H), 6.33 (d, *J* = 8.4 Hz, 1H), 4.97 (d, *J* = 3.4 Hz, 1H), 4.16 (s, 0.50H), 4.15 (s, 0.49H), 2.91 (s, 3H), 2.86 (s, 1.29H), 2.85 (s, 1.50H), 2.73 (t, *J* = 4.9 Hz, 0.65H), 2.61 (t, *J* = 7.6 Hz, 0.63H), 2.53 (s, 3H), 1.95 (d, *J* = 7.2 Hz, 2H), 1.42 (s, 3H).

¹³C NMR (101 MHz, CDCl₃; values in brackets were corresponding to the same carbon in diastereoisomers) δ 156.1, 149.4 (149.3), 143.2, 137.3, 120.5, 116.1, 106.6, 97.8 (97.7), 52.7 (t, *J* = 20.5 Hz), 52.6, 40.5 (40.4), 38.0 (37.9), 37.1 (37.0), 27.7, 27.2 (27.1).

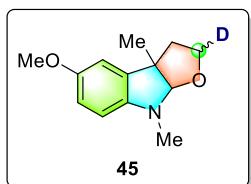
HRMS (ESI) *m/z* calculated for C₁₅H₂₁DN₃O₂ [M+H⁺] 277.1769, found 277.1767.

(c) Synthesis of furoindoline



To an oven-dried 10 mL reaction tube equipped with a magnetic stir bar, **42** (1 equiv., 0.3 mmol, 70 mg) was dissolved in dry THF (2 mL). LiAlH₄ solution (2.5 M in THF, 4 equiv., 0.48 mL) was slowly added at 0 °C with the atmosphere of Ar. After the reaction for 5 min, ethyl acetate and H₂O were added to quench the reaction. The aqueous layer was separated and extracted with ethyl acetate (3×10 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was purified by using flashcolumn chromatography to give **45** as colorless oil (55 mg, 83%, a mixture of diastereoisomers,

d.r. ~ 1.5:1).

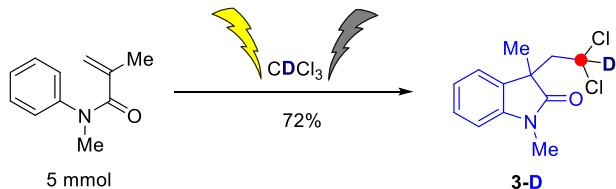


¹H NMR (400 MHz, CDCl₃) δ 6.69 (d, *J* = 2.2 Hz, 1H), 6.66 (d, *J* = 8.4, 2.3 Hz, 1H), 6.29 (d, *J* = 8.3 Hz, 1H), 5.03 (s, 1H), 3.93 (d, *J* = 7.1 Hz, 0.61H), 3.75 (s, 3H), 3.46 (dd, *J* = 10.9, 5.3 Hz, 0.43H), 2.88 (s, 3H), 2.14 – 2.01 (m, 2H), 1.45 (s, 3H).

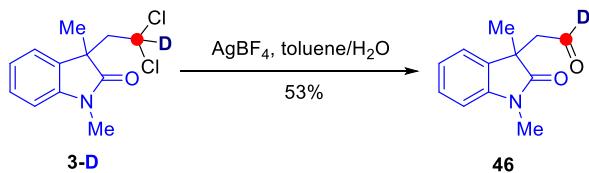
¹³C NMR (101 MHz, CDCl₃) δ 152.7, 144.9, 136.0, 112.1, 110.4, 105.6, 105.2, 67.0 (t, *J* = 22.1 Hz), 56.0, 52.4, 41.3, 31.6, 24.4.

HRMS (ESI) *m/z* calculated for C₁₃H₁₇DNO₂ [M+H⁺] 221.1395, found 221.1392.

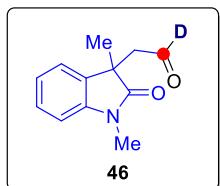
(d) Synthesis of deuterated (\pm)-Lansai B



To an oven-dried 100 mL Schlenk tube equipped with a magnetic stir bar, acrylamide (1 equiv., 5 mmol, 857 mg), CDCl₃ (10 equiv., 50 mmol, 4.05 mL), n-Bu₄NBF₄ (0.1 M, 5 mmol, 1.65 g) and dry DMF (50 mL) were added. The tube was flushed with Ar for 3 min to exclude the air. The rubber plug equipped with a carbon felt anode (25 mm * 20 mm) and a foam Ni cathode (25 mm * 20 mm) was used to keep the Schlenk tube sealed. The mixture was stirred for 5 min and put in the oil bath (100 °C) with the electrolysis of 35 mA, 72 h. After the reaction, the tube was cooled to room temperature and ethyl acetate (50 mL) and water (200 mL) was added. The aqueous layer was separated and extracted with ethyl acetate (3×50 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to flash column chromatography on silica gel to yield the desired product **3-D**. Six parallel experiments (5 mmol×6) provided **3-D** in 72% yield (5.6g, 21.6 mmol).



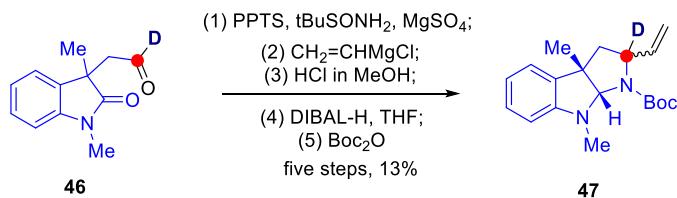
To an oven-dried 25 mL reaction tube equipped with a magnetic stir bar, **3-D** (1 equiv., 1 mmol, 259 mg), AgBF₄ (5 equiv., 5 mmol, 1 g), Celite (500 mg), H₂O (500 μ L) and toluene (5 mL) were added. The tube was flushed with Ar for 3 min to exclude the air. The tube was sealed and put in the oil bath (100 °C) with vigorous stirring for 2 h. After the reaction, the tube was cooled to room temperature and the reaction mixture was subjected to flash column chromatography on silica gel directly to yield the desired product **46** as colorless oil (109 mg, 53%).



¹H NMR (400 MHz, CDCl₃) δ 7.29 – 7.25 (m, 1H), 7.18 (dd, *J* = 7.3, 0.7 Hz, 1H), 7.04 (td, *J* = 7.6, 0.8 Hz, 1H), 6.88 (d, *J* = 7.8 Hz, 1H), 3.25 (s, 3H), 2.96 (d, *J* = 1.7 Hz, 2H), 1.41 (s, 3H).

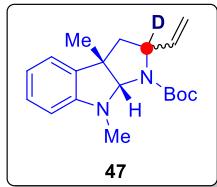
¹³C NMR (101 MHz, CDCl₃) δ 198.4 (t, *J* = 26.9 Hz), 179.3, 143.0, 132.6, 128.1, 122.5, 122.2, 108.2, 50.2 (t, *J* = 3.7 Hz), 44.7, 26.2, 23.8.

HRMS (ESI) *m/z* calculated for C₁₂H₁₂DNNaO₂ [M+Na⁺] 227.0901, found 227.0903.



To an oven-dried 50 mL reaction tube equipped with a magnetic stir bar, **46** (1 equiv., 7.5 mmol, 1.53 g), PPTS (5 mol%, 0.38 mmol, 96 mg), MgSO₄ (5 equiv., 37.5 mmol, 4.52 g), ^tBuSONH₂ (2 equiv., 15 mmol, 1.82 g) and 20 mL dry DCM were added. The tube was sealed and reacted at room temperature overnight. After the reaction, the mixture was vacuum filtrated to remove salts firstly, and then filtrated through a short silica gel column quickly to remove PPTS and ^tBuSONH₂. The filtrate was concentrated, dried and used for the next step of reaction without further purification. The resulting

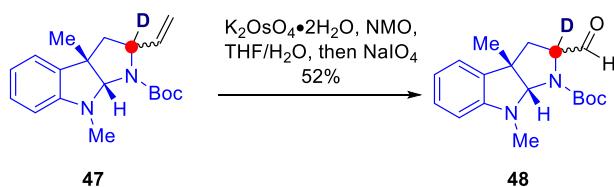
imine was dissolved in dry THF (15 mL) and cooled to -65°C . Vinylmagnesium chloride (1 M in THF, 2 equiv., 15 mL) was added slowly to the reaction and the reaction proceeded at -65°C for 2 h. After the reaction, saturated aq. NH₄Cl solution was added to quench the reaction and ethyl acetate was added for extraction. The aqueous layer was separated and extracted with ethyl acetate (3×20 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to the further transformation. The resulting sulfinamide was dissolved in MeOH (20 mL) and then added aq. HCl (4 M, 2 equiv., 3.8 mL). The reaction proceeded at room temperature and monitored by TLC. After the reaction, the mixture was concentrated to remove solvent and then alkalized with aq. NaHCO₃ solution followed by extraction with DCM. The combined organic layer was washed with 1N HCl aqueous solution until there was not amine in organic layer through TLC monitoring. Then aqueous layer was alkalized again with saturated aq. NaHCO₃ solution and extracted with DCM for three times. The combined organic phase was washed with brine, dried by anhydrous Na₂SO₄ and concentrated for the next step of reaction. The obtained amine was dissolved in dry THF (20 mL) and cooled to 0°C . DIBAL-H (1 M in hexane, 3 equiv., 22.5 mL) was added to the reaction and reacted at 0°C for 1h, then at room temperature for 2h and at 80°C for 15 h. After cooling to room temperature, a saturated aqueous solution of potassium sodium tartrate was added and ethyl acetate was used for extraction. The aqueous layer was separated and extracted with ethyl acetate (3×20 mL), and the combined organic layers were washed with brine and dried over sodium sulfate. Following concentration in vacuo, the crude residue was subjected to the further transformation. The resulting amine was dissolved in dry THF (10 mL) and Boc₂O (2 equiv., 3.5 mL) was added. The reaction proceeded at room temperature overnight and was then concentrated. The crude residue was subjected to flash column chromatography on silica gel to yield the **47** as colorless oil (307 mg, 13% for five steps, a mixture of diastereomers, d.r. $\sim 4:1$).



¹H NMR (400 MHz, CDCl₃; compound exists as a mixture of diastereomers, the major is denoted by *, minor denoted by §) δ 7.13 – 7.09 (m, 1H*§), 6.70 – 6.98 (m, 1H*§), 6.69 (t, J = 7.2 Hz, 0.87H*), 6.66 – 6.64 (m, 0.15H§), 6.43 (d, J = 7.8 Hz, 0.82H*), 6.38 (d, J = 7.8 Hz, 0.18H§), 5.88 (dd, J = 15.8, 10.5 Hz, 0.78H*), 5.48 (dd, J = 17.0, 10.0 Hz, 0.26H§), 5.23 (s, 1H*§), 5.07 – 5.01 (m, 1.91H*§), 4.84 (d, J = 10.0 Hz, 0.20H§), 3.01 (s, 2.55H*), 2.95 (s, 0.62H§), 2.30 (d, J = 12.7 Hz, 0.90H*), 2.18 (d, J = 12.8 Hz, 0.22H§), 2.01 (d, J = 12.8 Hz, 0.25H§), 1.85 (d, J = 12.8 Hz, 0.92H*), 1.53 (s, 8.84H*§), 1.45 (s, 2.72H*§), 1.39 (s, 0.55H*§).

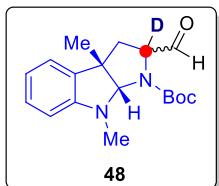
¹³C NMR (101 MHz, CDCl₃; compound exists as a mixture of diastereomers, the major is denoted by *, minor denoted by §) δ 155.2§, 150.0*, 149.6§, 146.7*, 140.8*, 139.3§, 135.0*, 134.9§, 128.1*, 127.8§, 121.6§, 121.4*, 117.7*, 117.3§, 114.3§, 113.4*, 106.7*, 106.1§, 90.7*, 90.3§, 85.0*, 80.0§, 60.7* (t, J = 22.0 Hz), 50.0*, 46.3*, 45.1§, 35.2*, 32.7§, 28.2§, 27.3*, 24.5*, 24.1§.

HRMS (ESI) *m/z* calculated for C₁₉H₂₆DN₂O₂ [M+H⁺] 316.2130, found 316.2128.



To an oven-dried 50 mL reaction tube equipped with a magnetic stir bar, **47** (1 equiv., 1.9 mmol, 595 mg), K₂OsO₄ · 2H₂O (0.2 equiv., 0.38 mmol, 140 mg), NMO (2 equiv., 3.8 mmol, 445 mg) and THF/H₂O (15/5 mL) were added. The tube was sealed and reacted at room temperature with stirring for 15 h. Then NaIO₄ (2 equiv., 3.8 mmol, 813 mg) was added and reacted for 2 h. NaIO₄ (2 equiv., 3.8 mmol, 813 mg) was added and reacted for 2 h again. After the reaction, a saturated aqueous solution of Na₂S₂O₃ (10 mL) was introduced. The mixture was then extracted with ethyl acetate (3 × 20 mL), and the combined organic phases were dried over anhydrous sodium sulfate. The filtrate was concentrated, and the residue was chromatographed on silica gel column to yield

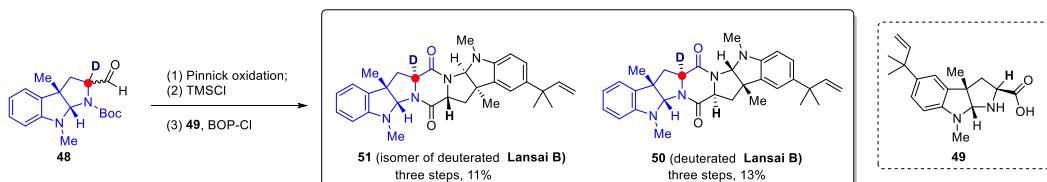
48 as colorless oil (312 mg, 52%, a mixture of diastereomers, d.r. ~ 5:1, both diastereomers exist as a ~ 2:1 mixture of rotamers).



¹H NMR (400 MHz, CDCl₃; compound exists as a mixture of diastereomers, the major is denoted by *, minor denoted by §) δ 9.54 (s, 0.26H*), 9.51 (s, 0.50H*), 9.22 (s, 0.14H§), 7.16 – 7.10 (m, 1H*§), 7.01 – 6.94 (m, 1H*§), 6.73 – 6.67 (m, 1H*§), 6.46 – 6.43 (m, 1H*§), 5.30 (s, 0.51H*), 5.27 (s, 0.11H§), 5.14 (s, 0.06H§), 5.11 (s, 0.27H*), 3.05 (s, 1.51H*), 3.02 (s, 0.12H§), 3.01 (s, 0.29H§), 2.98 (s, 0.89H*), 2.36 (d, *J* = 13.2 Hz, 0.58H*), 2.31 – 2.25 (m, 0.44H*§), 2.18 (d, *J* = 12.8 Hz, 0.19H§), 2.03 (d, *J* = 13.5 Hz, 0.59H*), 1.98 (d, *J* = 13.3 Hz, 0.31H*), 1.55 (s, 3H*§), 1.45 (s, 5.45H*§), 1.42 (s, 1.59H*§), 1.40 (s, 1.81H*§).

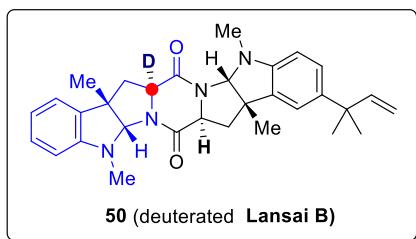
¹³C NMR (101 MHz, CDCl₃; compound exists as a mixture of diastereomers, the major is denoted by *, minor denoted by §) δ 200.8§, 199.3*, 198.9*, 155.0§, 154.4*, 154.2*, 149.8*, 149.7*, 133.7*, 133.6*, 133.2§, 128.8*, 128.6*, 128.4§, 122.2§, 122.0§, 121.7*, 121.6*, 118.2*, 118.1*, 107.1*, 107.0*, 90.8*, 90.5§, 90.3*, 90.0§, 81.7*, 81.6*, 81.4§, 81.3§, 65.4* (t, *J* = 23.0 Hz), 65.2* (t, *J* = 23.0 Hz), 52.1*, 51.0§, 50.6*, 41.0*, 40.7§, 40.1*, 35.1*, 34.7*, 33.5§, 29.7§, 28.3*, 28.2§, 28.1*, 24.4*, 24.1*, 23.0§.

HRMS (ESI) *m/z* calculated for C₁₈H₂₃DN₂NaO₃ [M+Na⁺] 340.1742, found 340.1741.



To an oven-dried flask equipped with a magnetic stir bar, **48** (1 equiv., 0.41 mmol, 130 mg) was dissolved in THF/H₂O/tBuOH (5/5/1.3 mL). 2-Methyl-2-butene (2.5 mL), NaClO₂ (2 equiv., 0.82 mmol, 80wt%, 93 mg), KH₂PO₄ (5 equiv., 2.05 mmol, 279 mg) were added and the reaction proceeded with stirring at room temperature for 3 h. After the reaction, a saturated aqueous solution of NH₄Cl (10 mL) was introduced. The mixture was then extracted with ethyl acetate (3 × 10 mL), and the combined organic

phases were dried over anhydrous sodium sulfate. The filtrate was concentrated, and the residue was subjected to the next step of the reaction without further purification. The resulting crude acid was dissolved in dry MeCN (5 mL) and TMSI (2 equiv., 0.82 mmol, 117 μ L) was added at 0 °C. The reaction proceeded at 0 °C for 1 h and then TMSI (2 equiv., 0.82 mmol, 117 μ L) was added and reacted for one more hour. After the reaction, a saturated aqueous solution of Na₂S₂O₃ (2 mL) and H₂O (10 mL) was introduced. The mixture was then extracted with ethyl acetate (3×10 mL) and it is ensured that there was product in aqueous phase instead of organic phase. Organic phase was discarded and 1 M HCl aq. solution was added slowly to adjust the pH of aqueous phase to ~ 6. Then the aqueous phase was concentrated and dried on the rotary evaporator to remove moisture. The resulting mixture was then dissolved in DCM and turbid liquid was observed. The mixture was vacuum filtrated though filter paper and the filtrate was collected, concentrated and dried for the further transformation. The obtained amino acid and **49** (**49** was prepared according to the reference^[7], 0.21 mmol, 63 mg) were dissolved in dry DCM (6 mL), followed by the addition of DIPEA (0.97 mmol, 167 μ L) and BOP-Cl (0.84 mmol, 214 mg) at 0 °C. The reaction was then conducted at ambient temperature for 13 h before it was quenched with a saturated aqueous solution of Na₂CO₃ (5 mL). The resulting mixture was extracted with ethyl acetate (3×5 mL). The organic phases were combined, dried over anhydrous sodium sulfate, and concentrated. The residue was purified by silica gel column chromatography (two times for silica gel column chromatography, eluent: PE/EA = 20:1; PE/acetone = 20:1) to afford **50** (target molecule, deuterated Lansai B, 13 mg, 13%) as pale yellow solid and **51** (isomers of deuterated Lansai B, 11 mg, 11%) as pale yellow solid.



¹H NMR (400 MHz, CDCl₃) δ 7.11 – 7.06 (m, 2H), 7.04 (d, *J* = 7.0 Hz, 1H), 7.01 (d, *J* = 1.8 Hz, 1H), 6.70 (t, *J* = 7.4 Hz, 1H), 6.34 (d, *J* = 7.8 Hz, 1H), 6.28 (d, *J* = 8.2 Hz,

1H), 5.97 (dd, $J = 17.4$, 10.6 Hz, 1H), 5.44 (s, 1H), 5.42 (s, 1H), 5.02 (dd, $J = 11.9$, 1.3 Hz, 1H), 4.98 (dd, $J = 5.1$, 1.3 Hz, 1H), 4.17 (dd, $J = 11.2$, 6.1 Hz, 1H), 2.98 (s, 3H), 2.95 (s, 3H), 2.73 – 2.68 (m, 2H), 2.19 – 2.13 (m, 2H), 1.47 (s, 3H), 1.46 (s, 3H), 1.34 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.8, 165.5, 150.1, 148.6, 148.2, 138.7, 132.9, 132.8, 128.7, 126.3, 122.3, 120.2, 118.1, 110.1, 105.8, 105.4, 86.9, 86.5, 60.1, 50.5, 50.3, 42.7, 42.6, 40.7, 33.1, 32.9, 28.5, 25.5, 25.4.

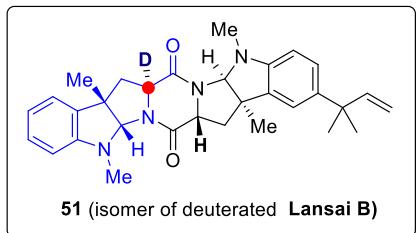
^{13}C NMR (176 MHz, CDCl_3) δ 165.8, 165.7, 149.9, 148.4, 147.5, 139.6, 133.2, 133.0, 128.8, 126.5, 122.4, 120.3, 118.3, 110.3, 106.3, 106.1, 87.1, 86.7, 60.2, 59.8 (t, $J = 21.9$ Hz), 50.5, 50.3, 42.7, 42.6, 40.7, 33.8, 33.1, 28.5, 25.6, 25.5.

HRMS (ESI) m/z calculated for $\text{C}_{31}\text{H}_{35}\text{DN}_4\text{NaO}_2$ [$\text{M}+\text{Na}^+$] 520.2793, found 520.2793.

Tuntiwachwuttikul et al. Report, ¹ Natural (–)-lansai B ¹ H NMR, 300 MHz, CDCl ₃	Reisman et al. Report ² Synthetic (–)-lansai B ¹ H NMR, 500 MHz, CDCl ₃	This Work, Synthetic Deuterated (±)-lansai B ¹ H NMR, 400 MHz, CDCl ₃
δ 7.11 (dt, <i>J</i> = 7.8, 1.5 Hz, 1H)	δ 7.13 – 7.05 (m, 1H)	δ 7.11 – 7.06 (m, 2H)
7.08 (dd, <i>J</i> = 8.1, 1.5 Hz, 1H)	7.07 (dd, <i>J</i> = 8.1, 1.8 Hz, 1H)	
7.06 (dd, <i>J</i> = 7.8, 1.5 Hz, 1H)	7.04 (dd, <i>J</i> = 7.3, 1.2 Hz, 1H)	7.04 (d, <i>J</i> = 7.0 Hz, 1H)
7.02 (d, <i>J</i> = 1.5 Hz, 1H)	7.01 (d, <i>J</i> = 1.9 Hz, 1H)	7.01 (d, <i>J</i> = 1.8 Hz, 1H)
6.71 (dt, <i>J</i> = 7.8, 1.5 Hz, 1H)	6.70 (td, <i>J</i> = 7.4, 0.9 Hz, 1H)	6.70 (t, <i>J</i> = 7.4 Hz, 1H)
6.36 (dd, <i>J</i> = 7.8, 1.5 Hz, 1H)	6.34 (d, <i>J</i> = 7.8 Hz, 1H)	6.34 (d, <i>J</i> = 7.8 Hz, 1H)
6.29 (d, <i>J</i> = 8.1 Hz, 1H)	6.28 (d, <i>J</i> = 8.1 Hz, 1H)	6.28 (d, <i>J</i> = 8.2 Hz, 1H)
5.99 (dd, <i>J</i> = 17.5, 10.6 Hz, 1H)	5.97 (dd, <i>J</i> = 17.4, 10.6 Hz, 1H)	5.97 (dd, <i>J</i> = 17.4, 10.6 Hz, 1H)
5.46 (s, 1H)	5.44 (s, 1H)	5.44 (s, 1H)
5.44 (s, 1H)	5.42 (s, 1H)	5.42 (s, 1H)
5.03 (d, <i>J</i> = 10.6 Hz, 1H)	5.01 (dd, <i>J</i> = 14.0, 1.4 Hz, 1H)	5.02 (dd, <i>J</i> = 11.9, 1.3 Hz, 1H)
5.01 (d, <i>J</i> = 17.5 Hz, 1H)	4.99 (dd, <i>J</i> = 7.2, 1.4 Hz, 1H)	4.98 (dd, <i>J</i> = 5.1, 1.3 Hz, 1H)
4.16 (m, 1H)	4.15 (app dddd, <i>J</i> = 13.5, 11.1, 6.1, 2.1 Hz, 2H)	4.17 (dd, <i>J</i> = 11.2, 6.1 Hz, 1H)
4.16 (m, 1H)	–	–
2.99 (s, 3H)	2.97 (s, 3H)	2.98 (s, 3H)
2.97 (s, 3H)	2.95 (s, 3H)	2.95 (s, 3H)
2.72 (dd, <i>J</i> = 12.3, 5.9 Hz, 1H)	2.71 (dd, <i>J</i> = 12.7, 3.0 Hz, 1H)	2.73 – 2.68 (m, 2H)
2.72 (dd, <i>J</i> = 12.3, 5.9 Hz, 1H)	2.69 (dd, <i>J</i> = 12.7, 3.0 Hz, 1H)	
2.18 (dd, <i>J</i> = 12.3, 11.3 Hz, 1H)	2.18 (dd, <i>J</i> = 12.6, 6.3 Hz, 1H)	2.19 – 2.13 (m, 2H)
2.17 (dd, <i>J</i> = 12.3, 11.3 Hz, 1H)	2.15 (dd, <i>J</i> = 12.7, 6.4 Hz, 1H)	
1.49 (s, 3H)	1.47 (s, 3H)	1.47 (s, 3H)
1.48 (s, 3H)	1.46 (s, 3H)	1.46 (s, 3H)
1.36 (s, 6H)	1.34 (s, 6H)	1.34 (s, 6H)

Tuntiwachwuttikul et al. Report, ¹ Natural (-)-lansai B ¹³ C NMR, 75 MHz, CDCl ₃	Reisman et al. Report ² Synthetic (-)-lansai B ¹³ C NMR, 126 MHz, CDCl ₃	This Work, Synthetic Deuterated (±)-lansai B ¹³ C NMR, 101 MHz, CDCl ₃	Chemical Shift Difference, Δδ
δ 165.8	δ 165.7	165.8	0.1
165.6	165.5	165.5	0
150.1	150.1	150.1	0
148.6	148.6	148.6	0
148.3	148.2	148.2	0
138.7	138.7	138.7	0
132.9	132.9	132.9	0
132.8	132.8	132.8	0
128.8	128.7	128.7	0
126.3	126.3	126.3	0
122.3	122.3	122.3	0
120.2	120.2	120.2	0
118.1	118.1	118.1	0
110.2	110.1	110.1	0
105.8	105.8	105.8	0
105.4	105.4	105.4	0
86.9	86.9	86.9	0
86.6	86.5	86.5	0
60.1	60.1	60.1	0
60.1	60.1	\	\
50.5	50.5	50.5	0
50.3	50.3	50.3	0
42.7	42.7	42.7	0
42.7	42.7	42.6	0.1
40.7	40.7	40.7	0
33.1	33.1	33.1	0
32.9	32.9	32.9	0
28.5	28.5	28.5	0
28.5	28.5	–	–
25.5	25.5	25.5	0
25.4	25.4	25.4	0

Tuntiwachwuttikul et al. Report, ⁸ Natural (–)-lansai B ¹³ C NMR, 75 MHz, CDCl ₃	Reisman et al. Report ⁷ Synthetic (–)-lansai B ¹³ C NMR, 126 MHz, CDCl ₃	This Work, Synthetic Deuterated (±)-lansai B ¹³ C NMR, 175 MHz, CDCl ₃	Chemical Shift Difference, Δδ
δ 165.8	δ 165.7	165.8	0.1
165.6	165.5	165.7	0.2
150.1	150.1	149.9	0.2
148.6	148.6	148.4	0.2
148.3	148.2	147.5	0.7
138.7	138.7	139.6	0.9
132.9	132.9	133.2	0.3
132.8	132.8	133.0	0.2
128.8	128.7	128.8	0.1
126.3	126.3	126.5	0.2
122.3	122.3	122.4	0.1
120.2	120.2	120.3	0.1
118.1	118.1	118.3	0.2
110.2	110.1	110.3	0.2
105.8	105.8	106.3	0.5
105.4	105.4	106.1	0.7
86.9	86.9	87.1	0.2
86.6	86.5	86.7	0.2
60.1	60.1	60.2	0.1
60.1	60.1	59.8 (t, <i>J</i> = 21.9 Hz)	0.3
50.5	50.5	50.5	0
50.3	50.3	50.3	0
42.7	42.7	42.7	0
42.7	42.7	42.6	0.1
40.7	40.7	40.7	0
33.1	33.1	33.8	0.7
32.9	32.9	33.1	0.2
28.5	28.5	28.5	0
28.5	28.5	—	—
25.5	25.5	25.6	0.1
25.4	25.4	25.5	0.1

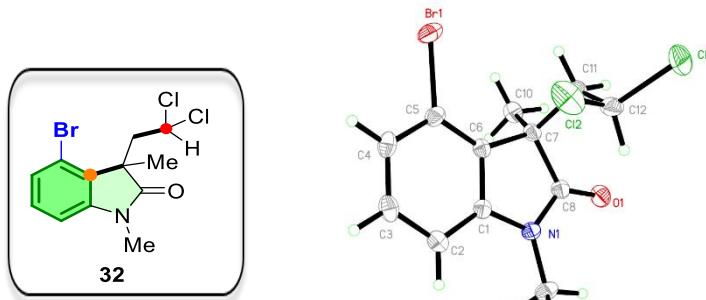


¹H NMR (400 MHz, CDCl₃) δ 7.14 – 7.08 (m, 2H), 7.05 (d, *J* = 7.0 Hz, 1H), 7.02 (d, *J* = 1.8 Hz, 1H), 6.71 (t, *J* = 7.3 Hz, 1H), 6.38 (d, *J* = 7.8 Hz, 1H), 6.31 (d, *J* = 8.2 Hz, 1H), 5.99 (dd, *J* = 17.4, 10.6 Hz, 1H), 5.48 (s, 1H), 5.47 (s, 1H), 5.03 (dd, *J* = 14.4, 1.2 Hz, 1H), 4.99 (dd, *J* = 7.6, 1.2 Hz, 1H), 4.04 (dd, *J* = 12.3, 5.2 Hz, 1H), 2.98 (s, 3H), 2.96 (s, 3H), 2.69 – 2.65 (m, 2H), 2.03 – 1.97 (m, 2H), 1.47 (s, 3H), 1.46 (s, 3H), 1.36 (s, 6H).

¹³C NMR (176 MHz, CDCl₃) δ 164.8, 164.5, 150.7, 148.7, 148.6, 138.7, 132.4, 132.3, 128.8, 126.4, 122.9, 120.7, 118.0, 110.1, 105.4, 105.1, 87.4, 87.0, 59.7, 59.4 (t, *J* = 22.0 Hz), 50.9, 50.7, 45.2, 45.0, 40.7, 32.3, 31.9, 28.5, 26.0, 25.8.

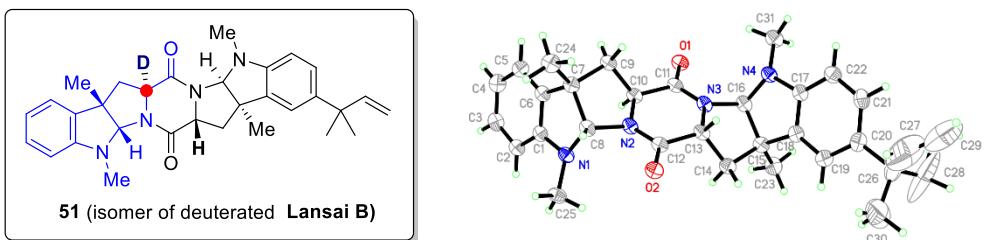
HRMS (ESI) *m/z* calculated for C₃₁H₃₆DN₄O₂ [M+H⁺] 498.2974, found 498.2969.

7. X-ray structure of compound 32, 51



A colorless block shaped crystal of **32** (C₁₂H₁₂BrCl₂NO) was used for the X-ray crystallographic analysis. The X-ray intensity data were measured at 173(2) K, on a Bruker D8 VENTURE CMOS photon 100 diffractometer with helios mx multilayer monochrmator Cu-Kα radiation (λ = 1.54178 Å). The X-ray crystallographic files, in CIF format, are available from the Cambridge Crystallographic Data Centre on quoting the deposition numbers **CCDC 2128341** for **32**. Copies of the data can be obtained free of charge from the Director, CCDC, 12 Union Road, Cambridge CB2 IEZ, UK (Fax:

+44-1223-336033; E-mail: deposit@ccdc.cam.ac.uk or www: <http://www.ccdc.cam.ac.uk>).

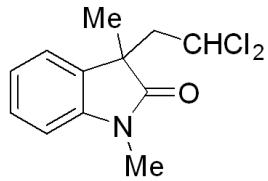


A colorless block shaped crystal of **51** ($C_{31}H_{35}DN_4O_2$) was used for the X-ray crystallographic analysis. The X-ray intensity data were measured at 173(2) K, on a Bruker D8 VENTURE CMOS photon 100 diffractometer with helios mx multilayer monochrmator Cu-K α radiation ($\lambda = 1.54178 \text{ \AA}$). The X-ray crystallographic files, in CIF format, are available from the Cambridge Crystallographic Data Centre on quoting the deposition numbers **CCDC 2128343** for **51**. Copies of the data can be obtained free of charge from the Director, CCDC, 12 Union Road, Cambridge CB2 IEZ, UK (Fax: +44-1223-336033; E-mail: deposit@ccdc.cam.ac.uk or www: <http://www.ccdc.cam.ac.uk>).

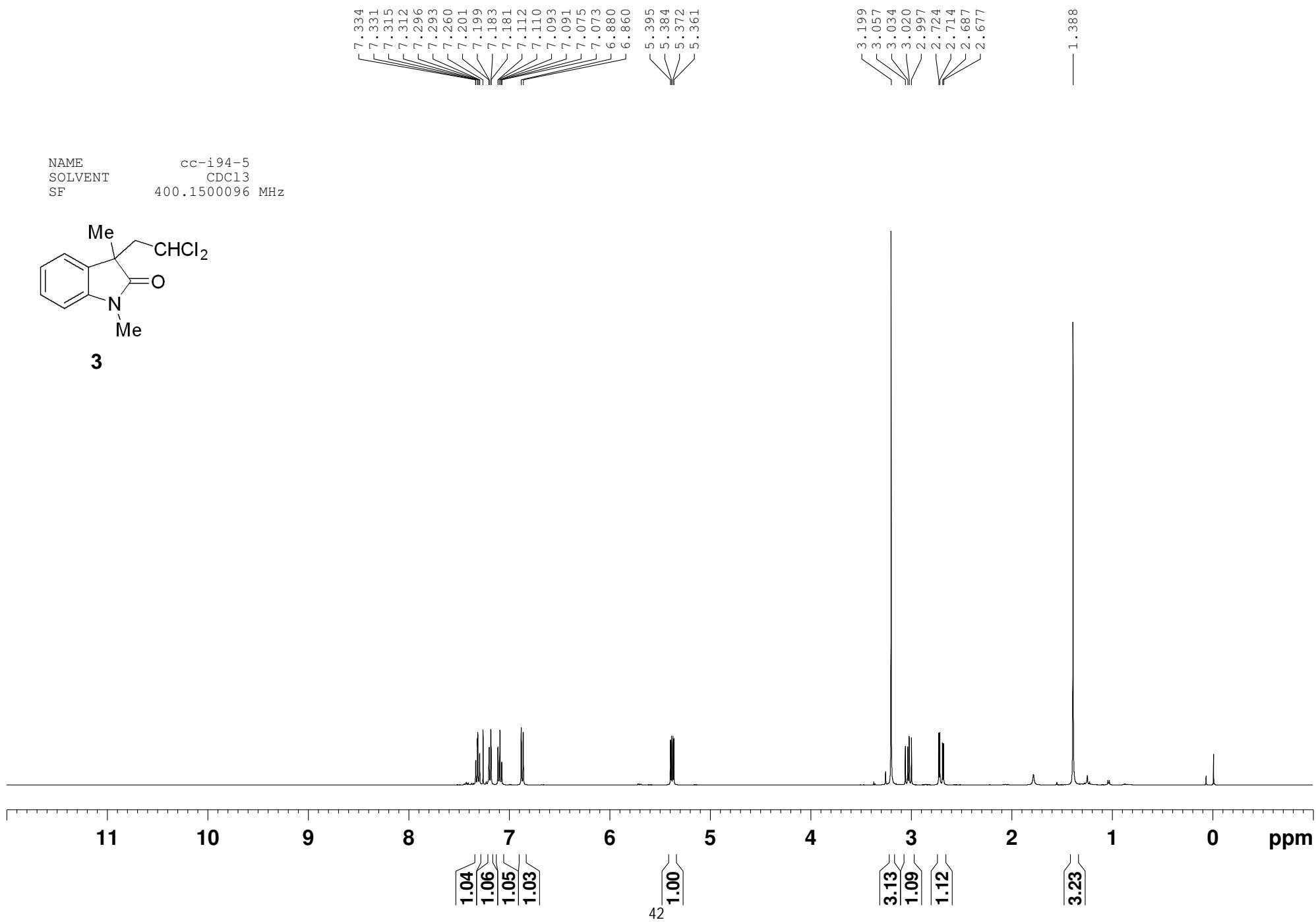
8. Reference

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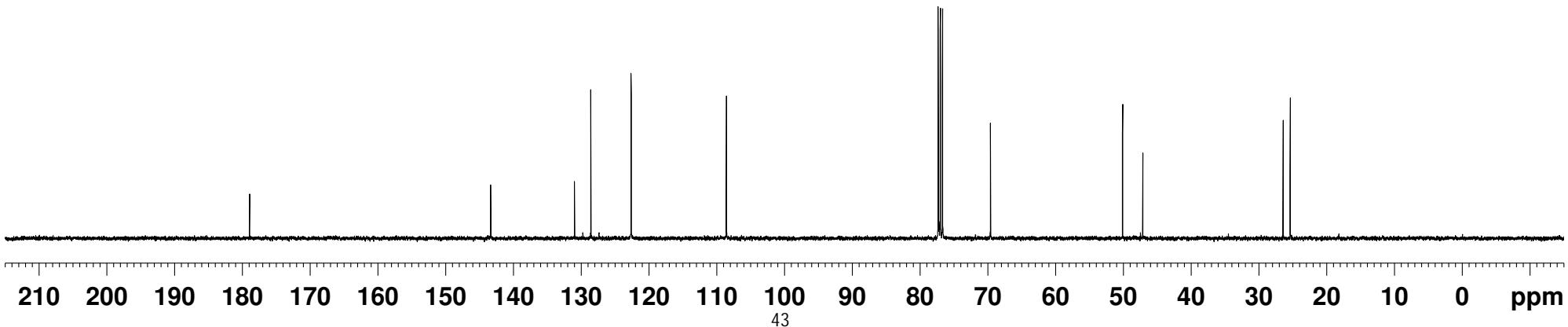
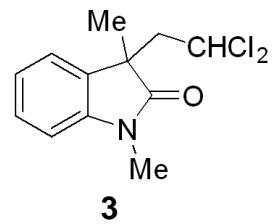
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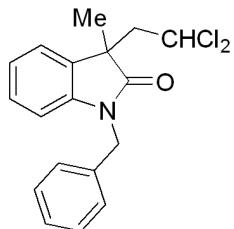
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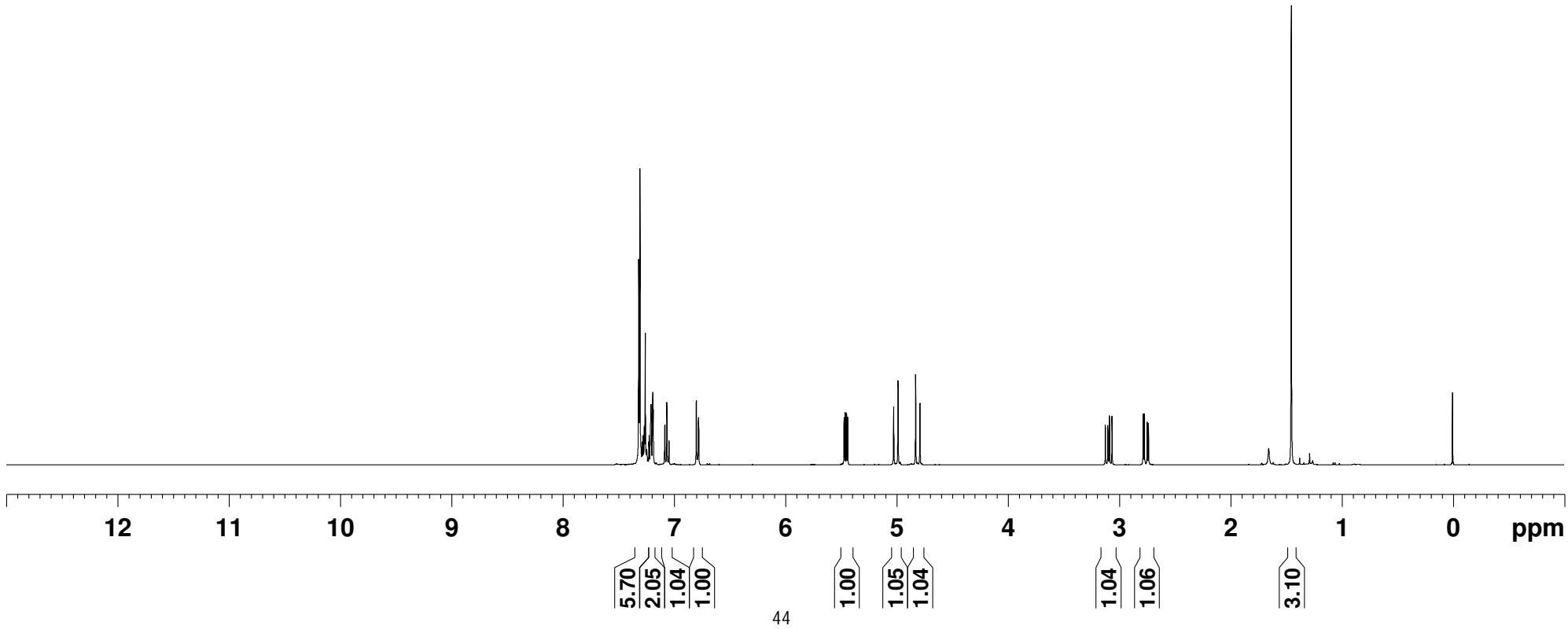
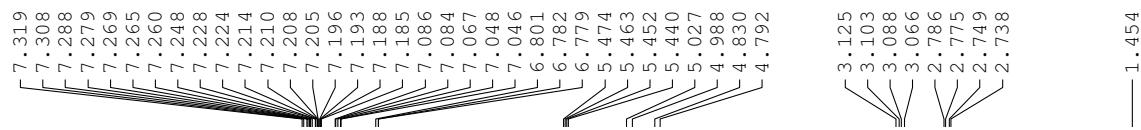
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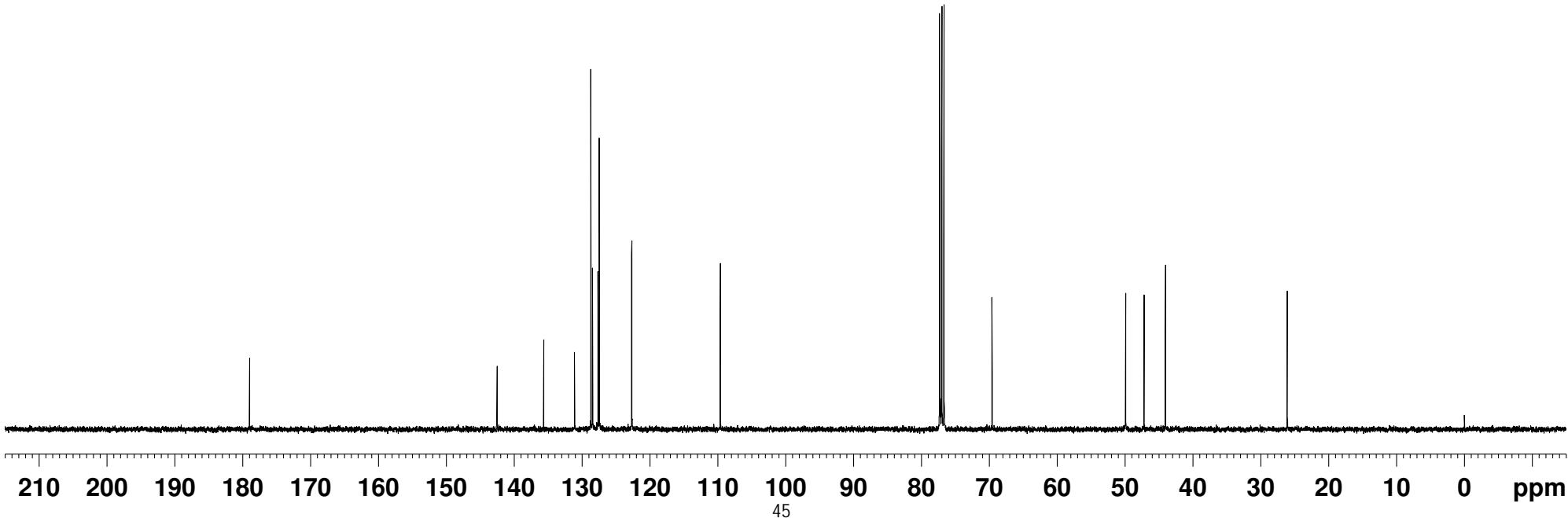
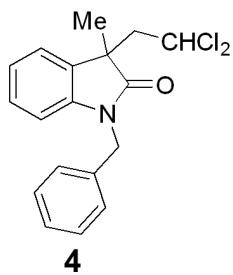
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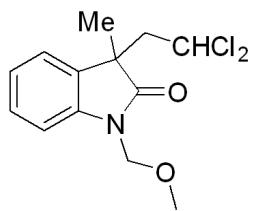
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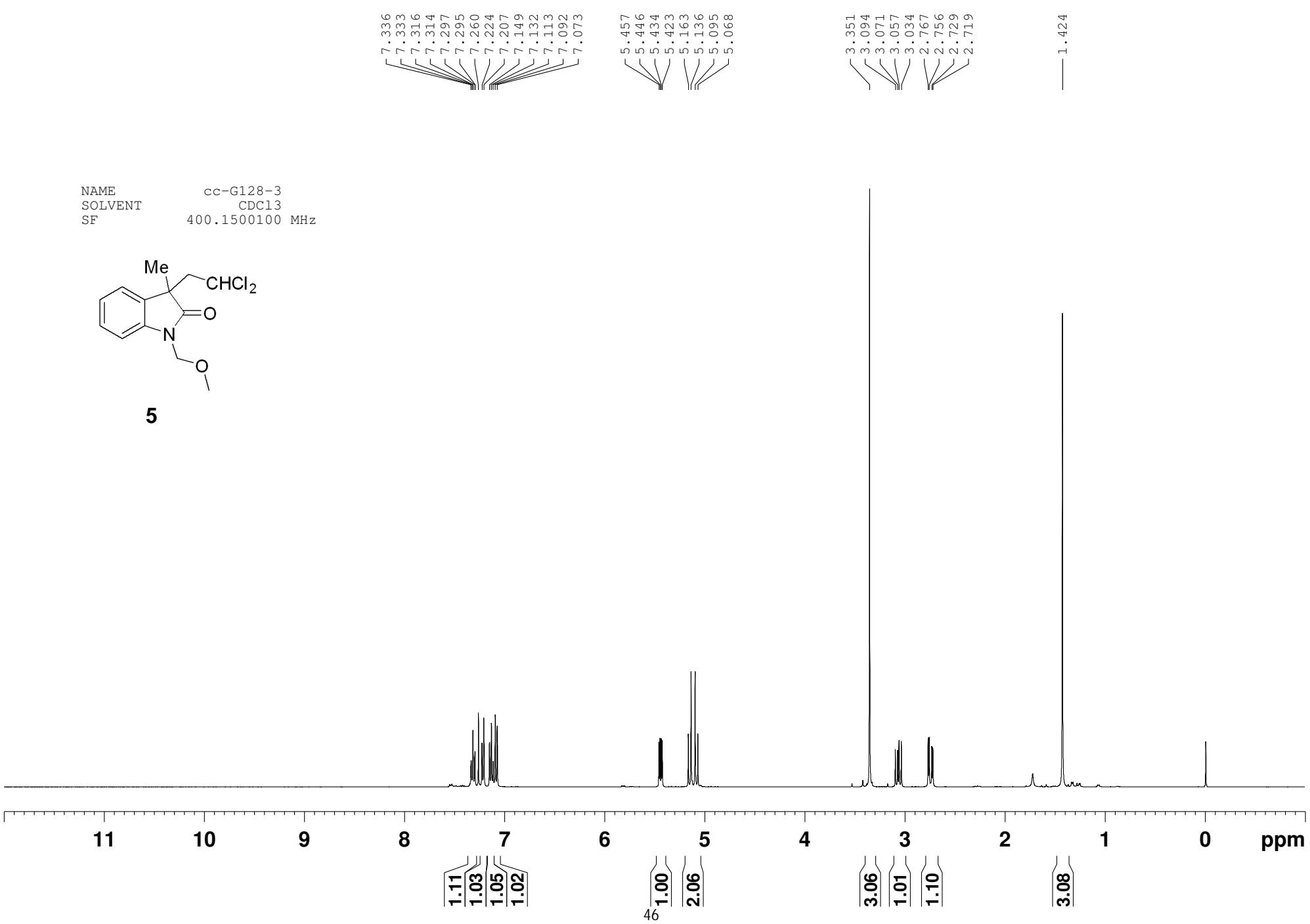
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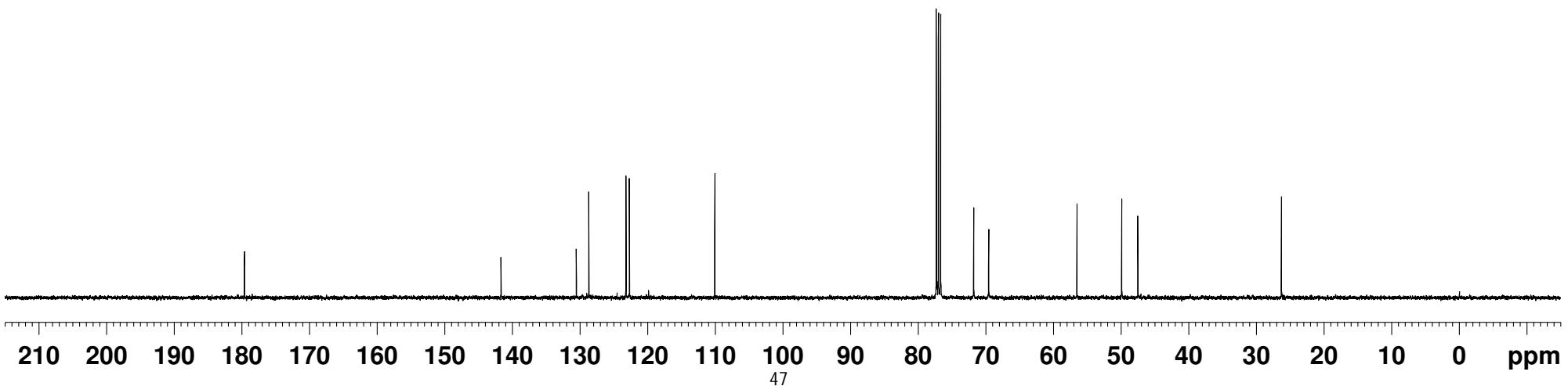
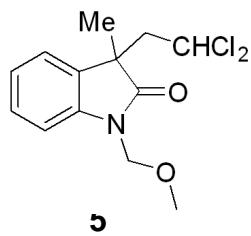
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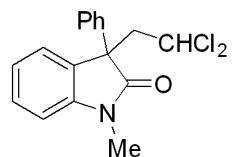
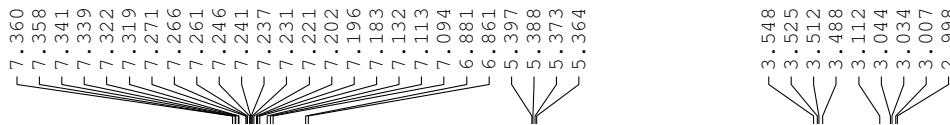
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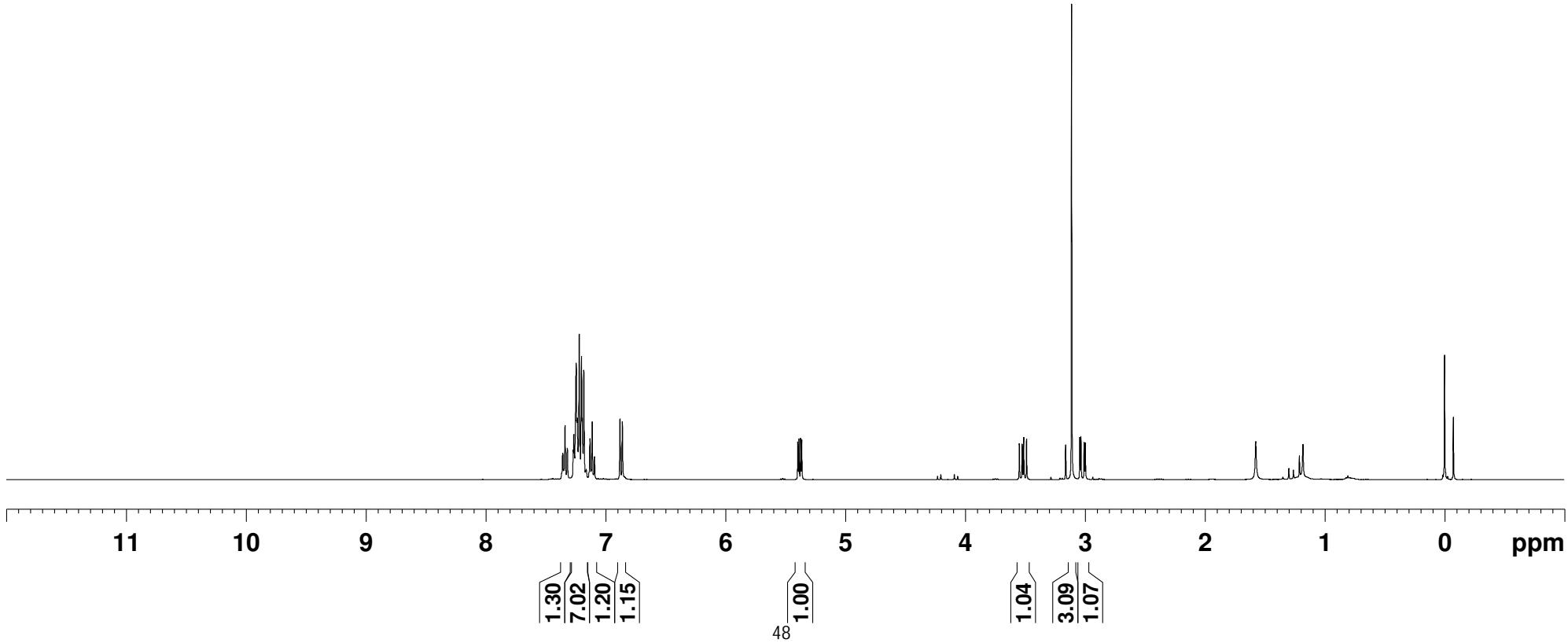
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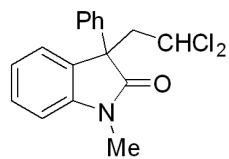
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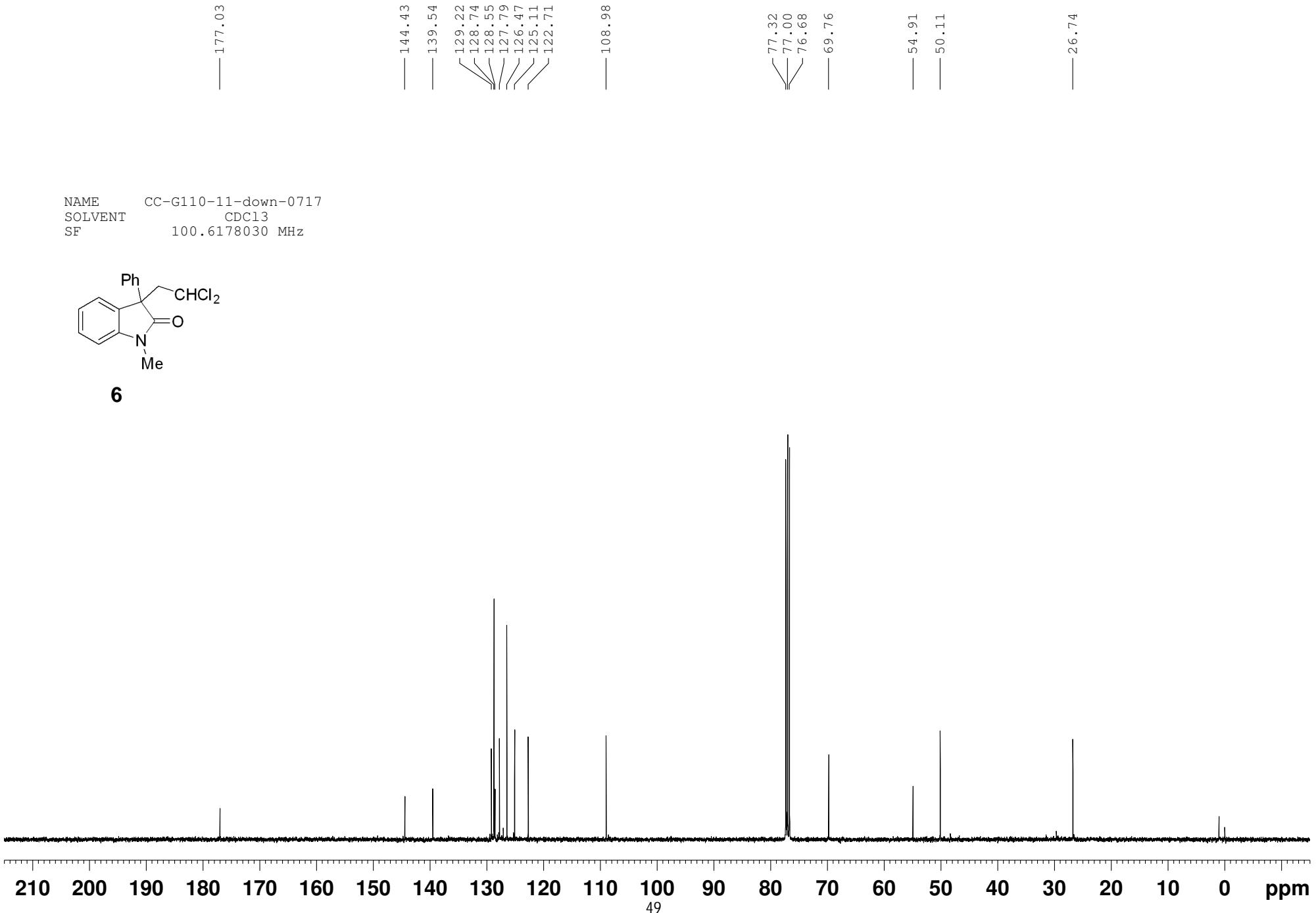
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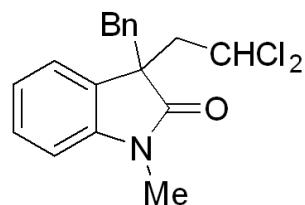
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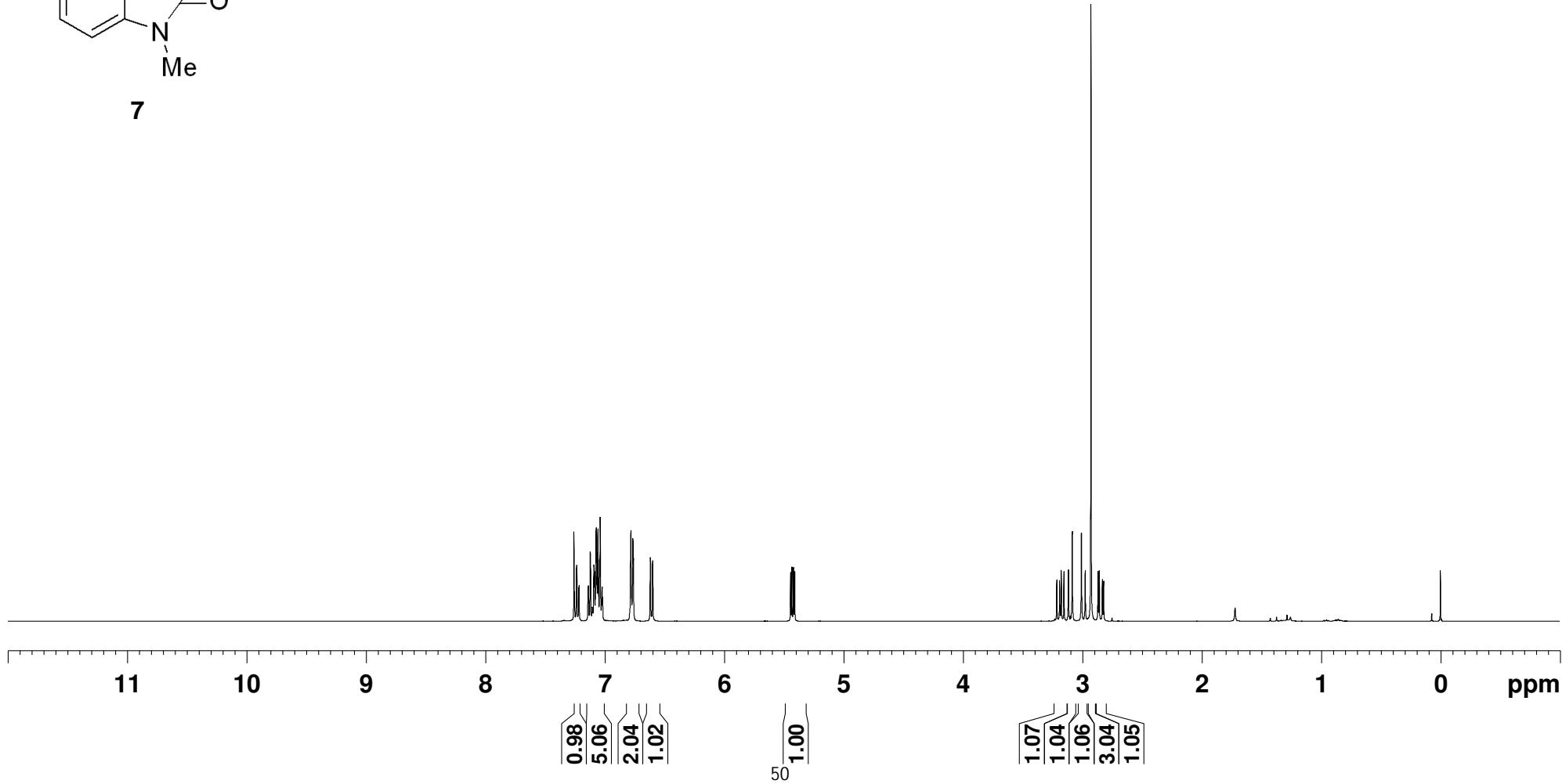
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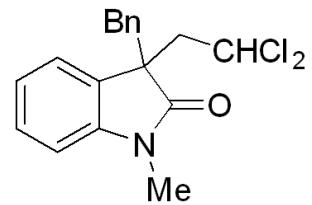
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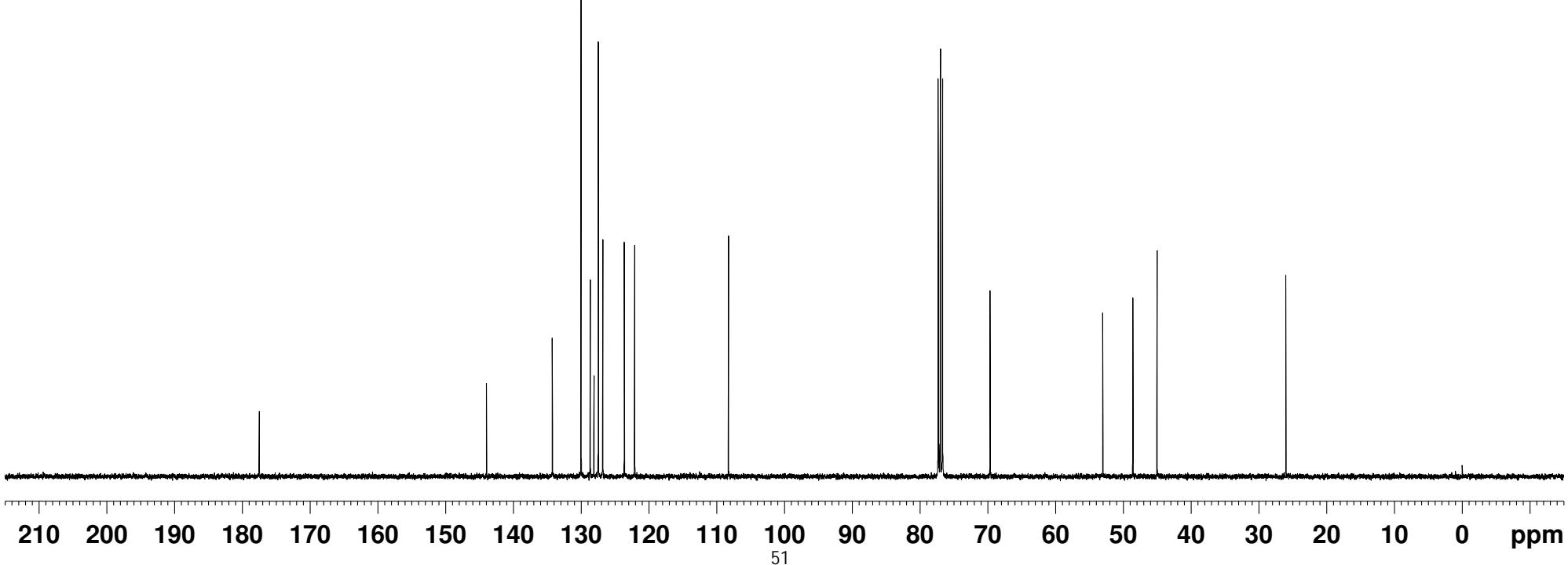
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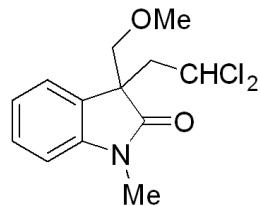
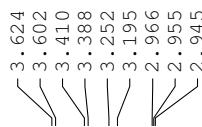
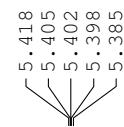
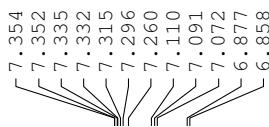
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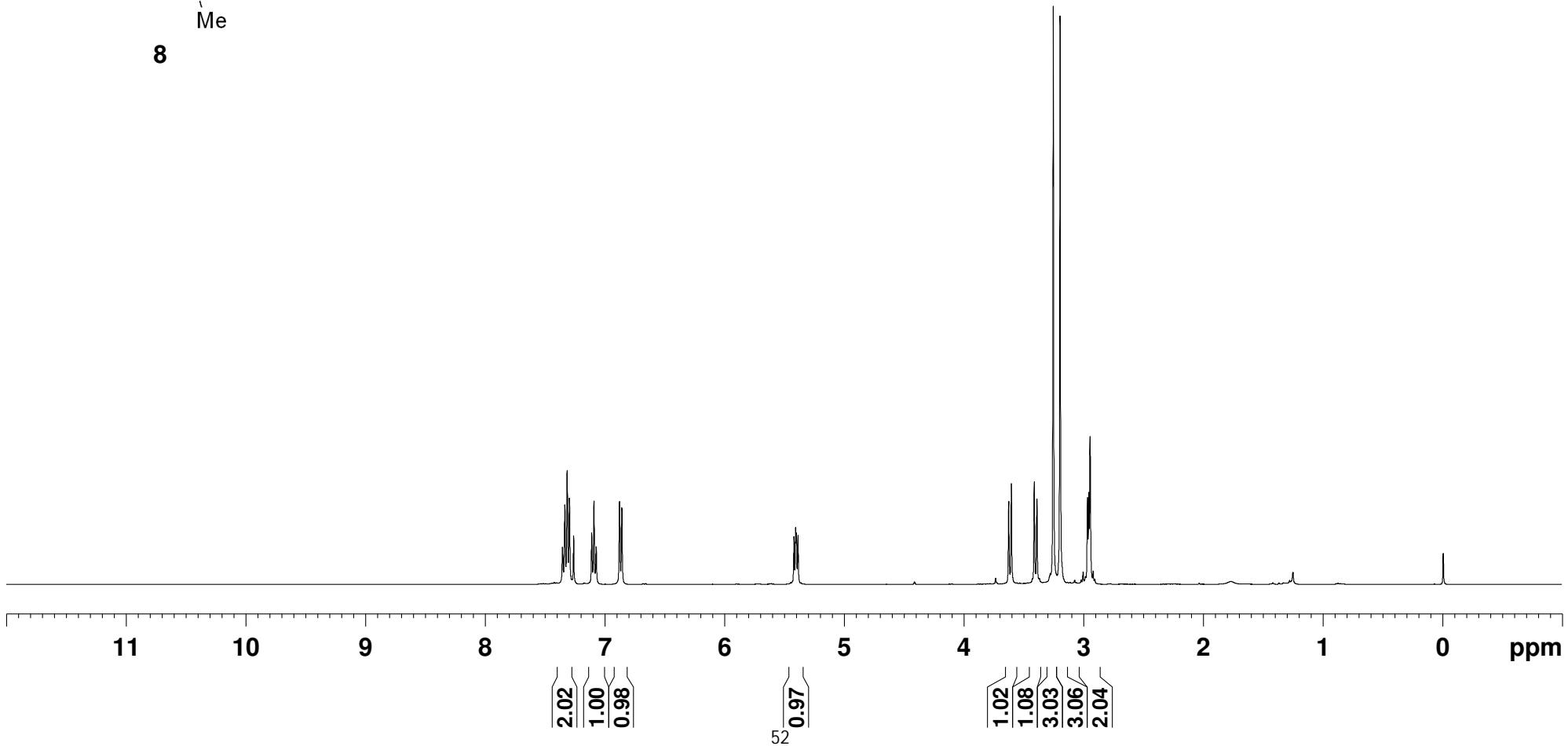
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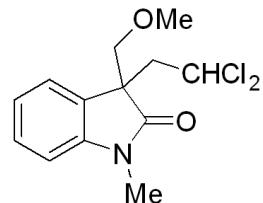
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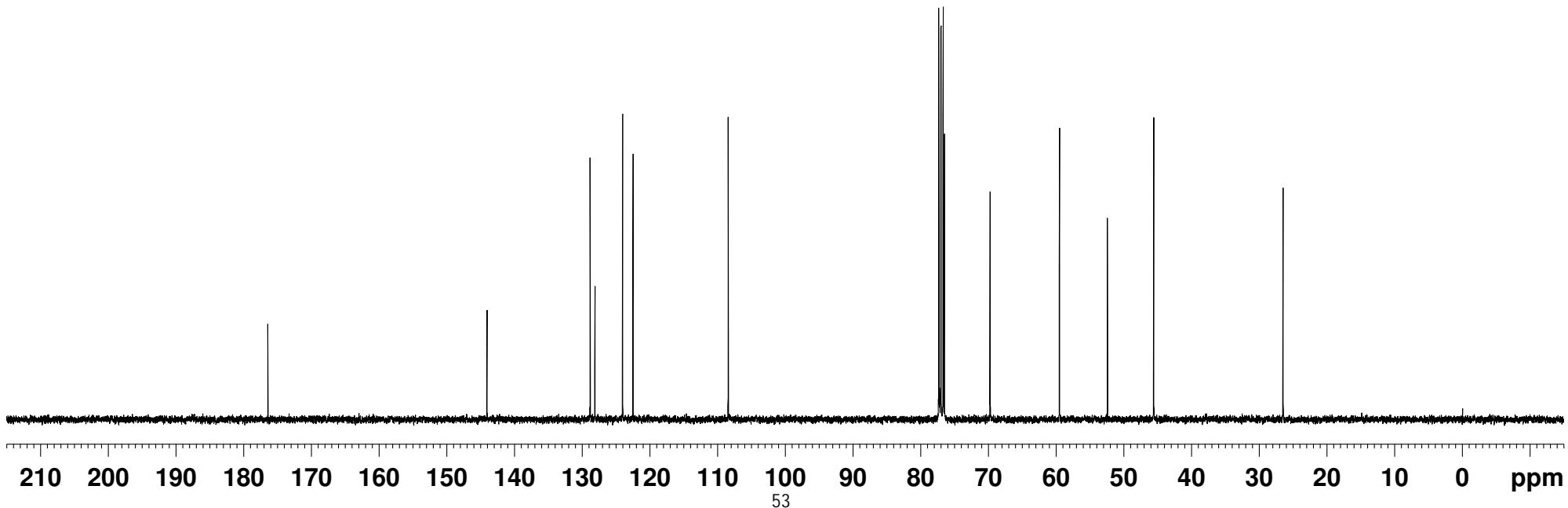
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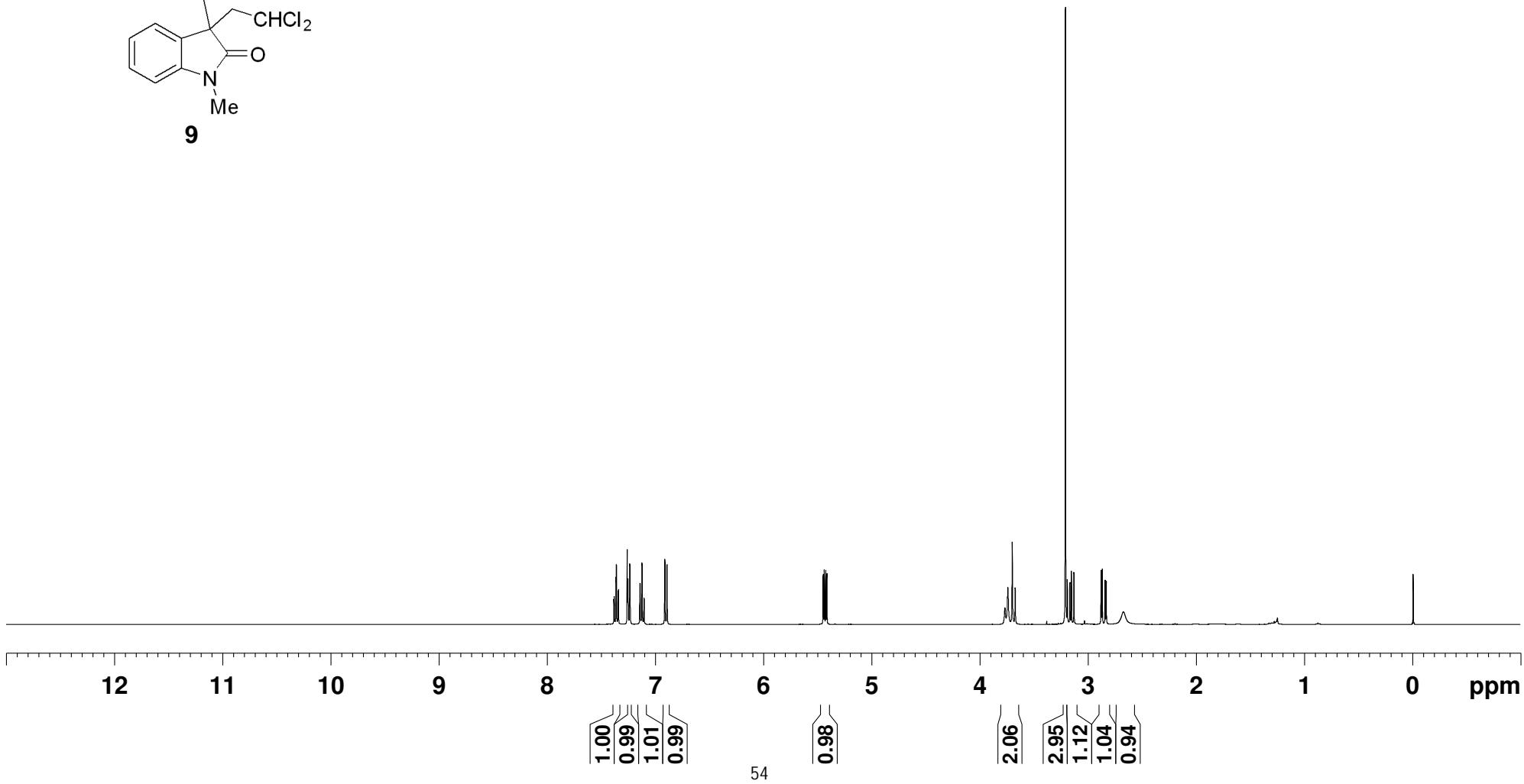
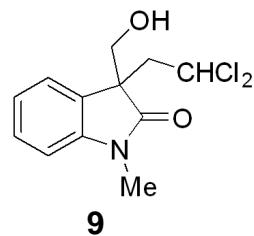
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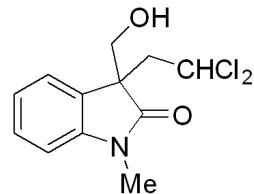
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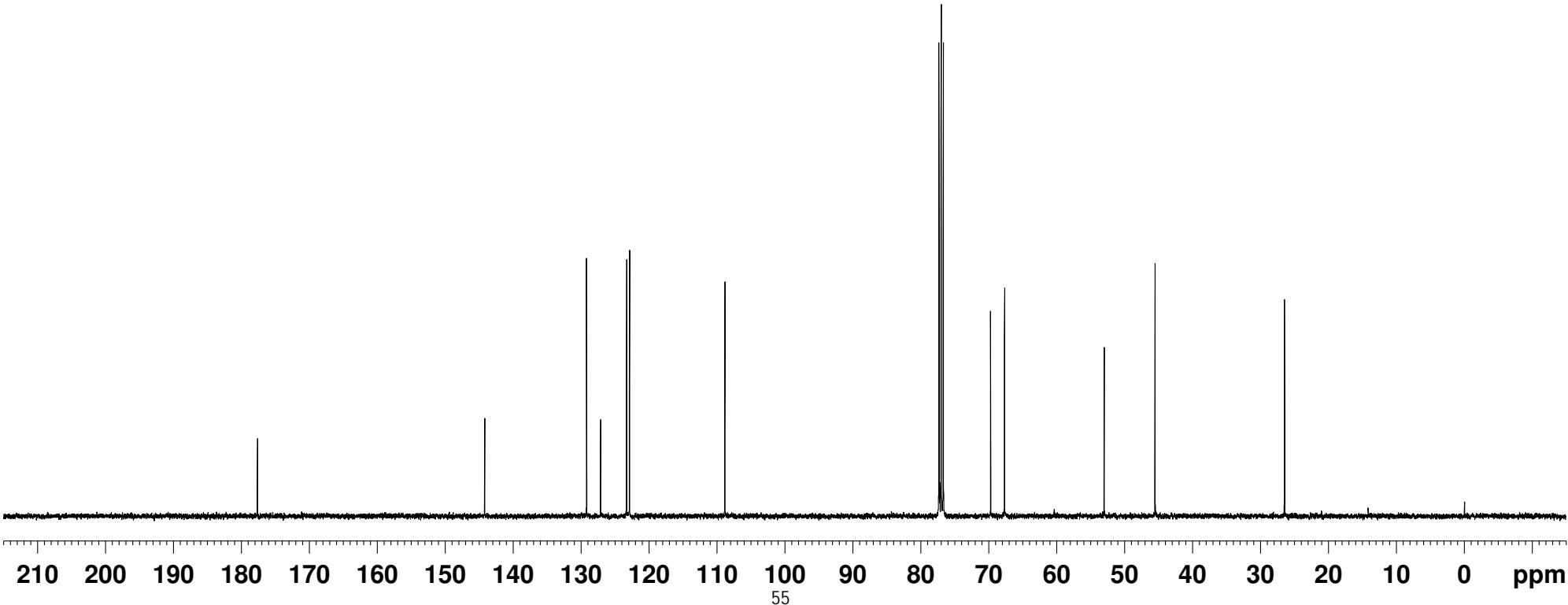
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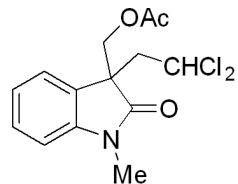
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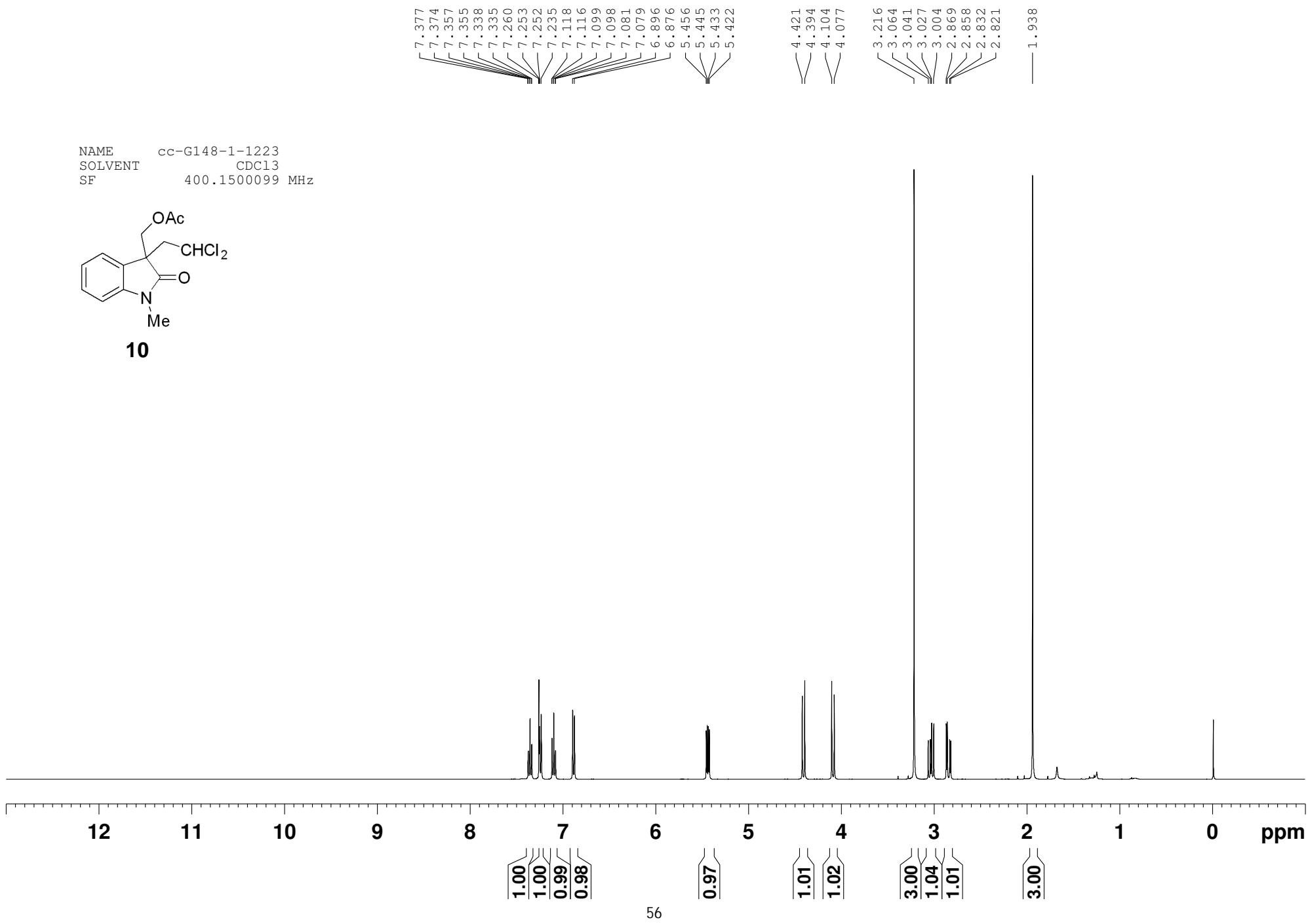
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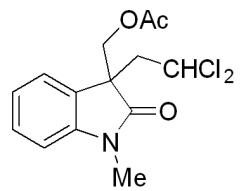
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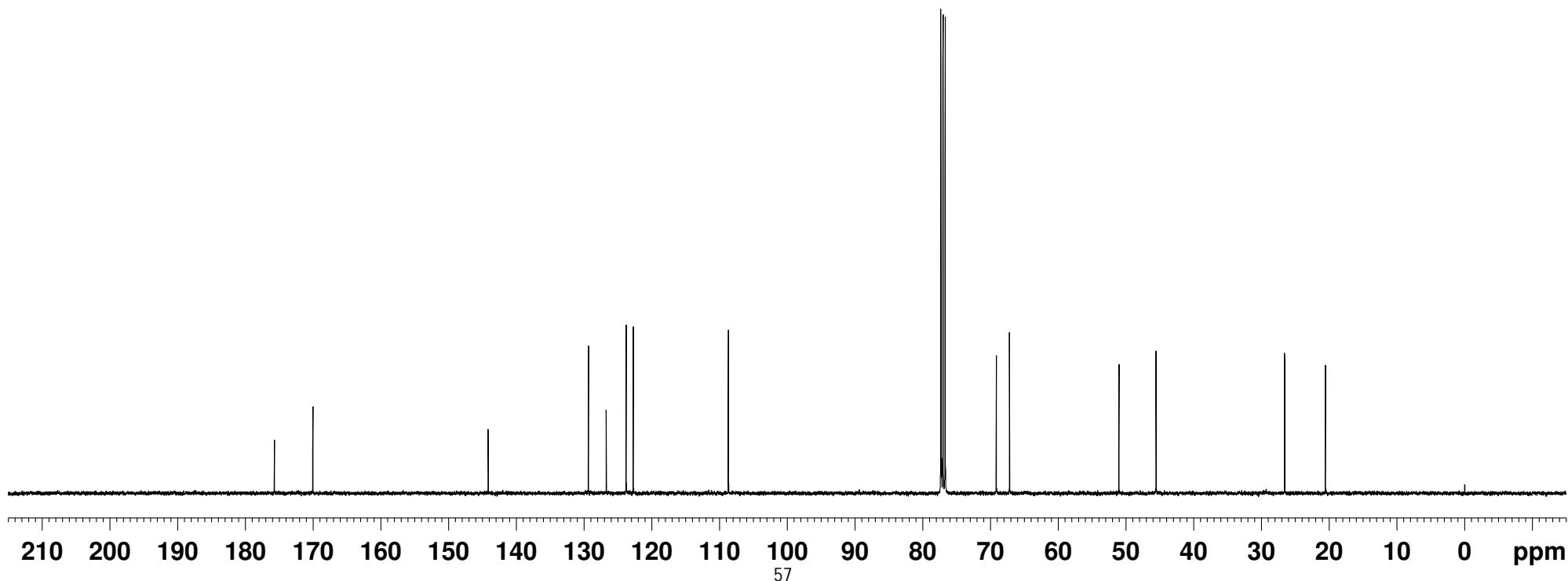
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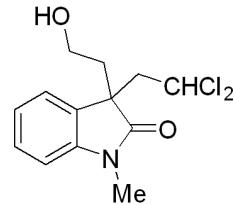
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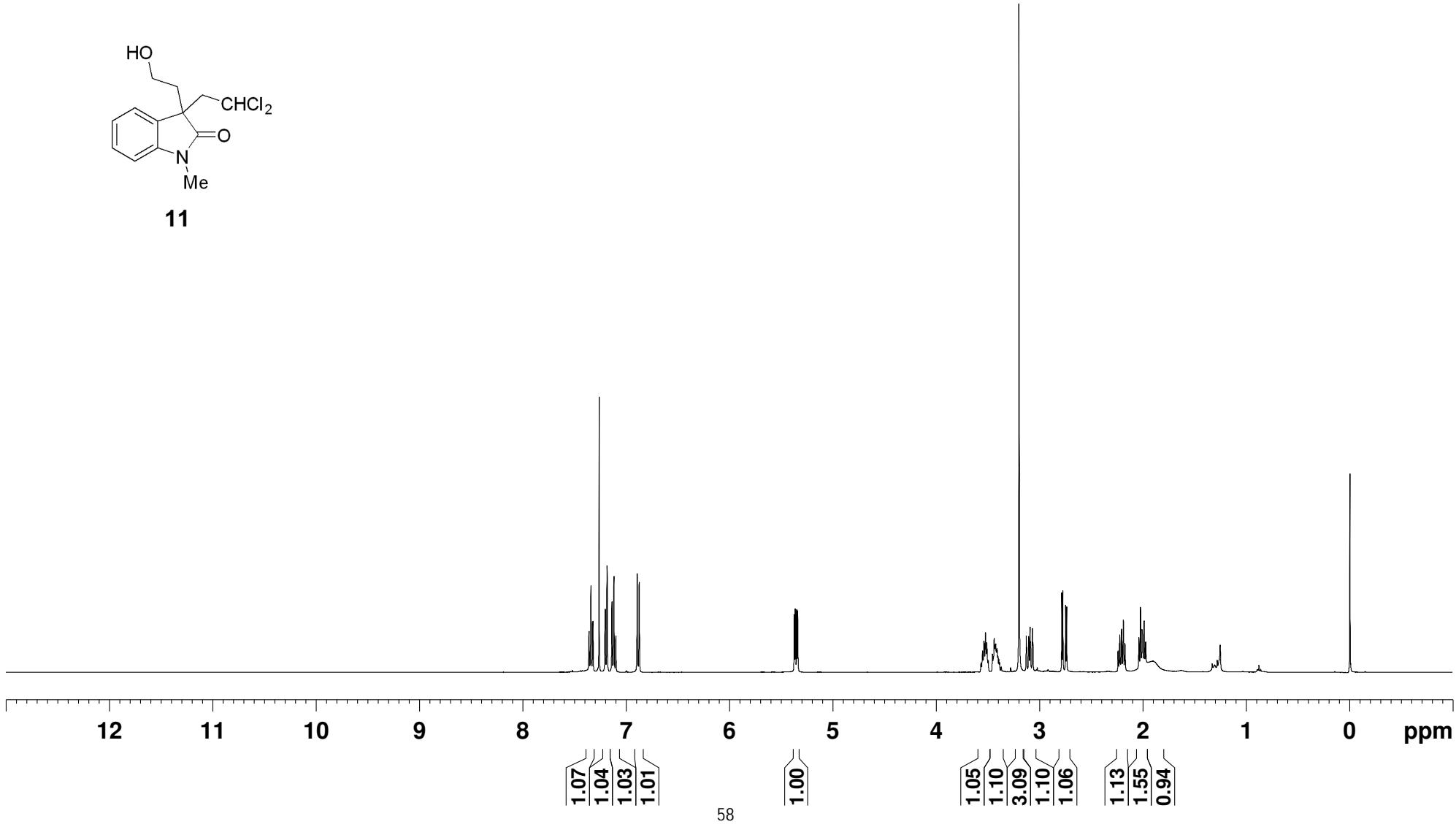
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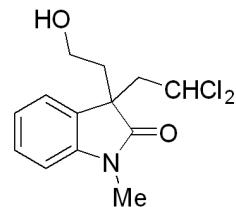
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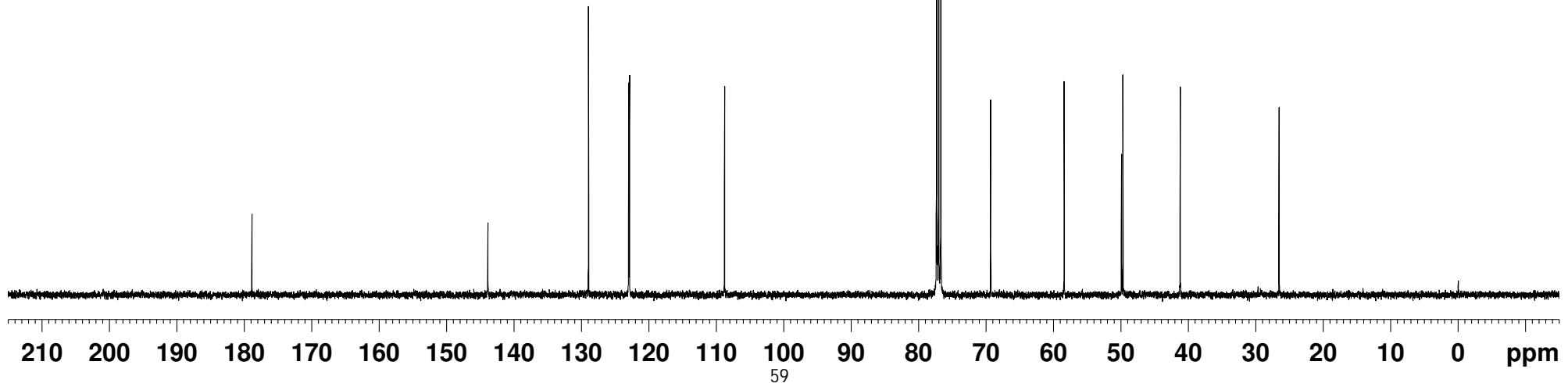
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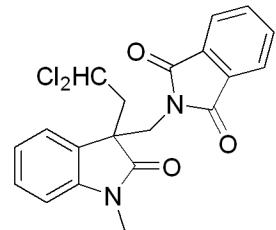
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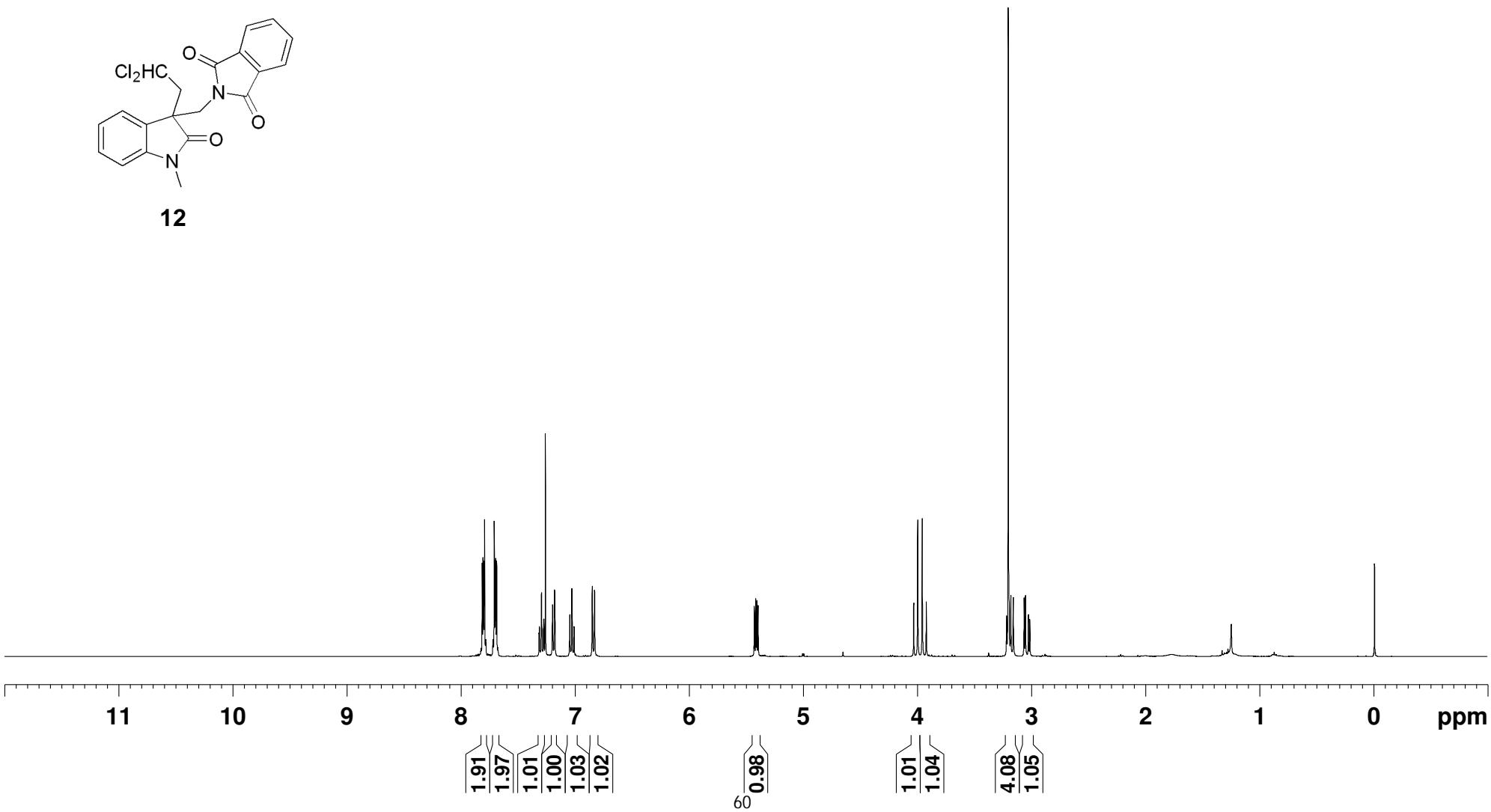
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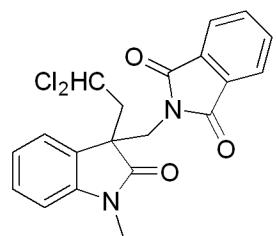
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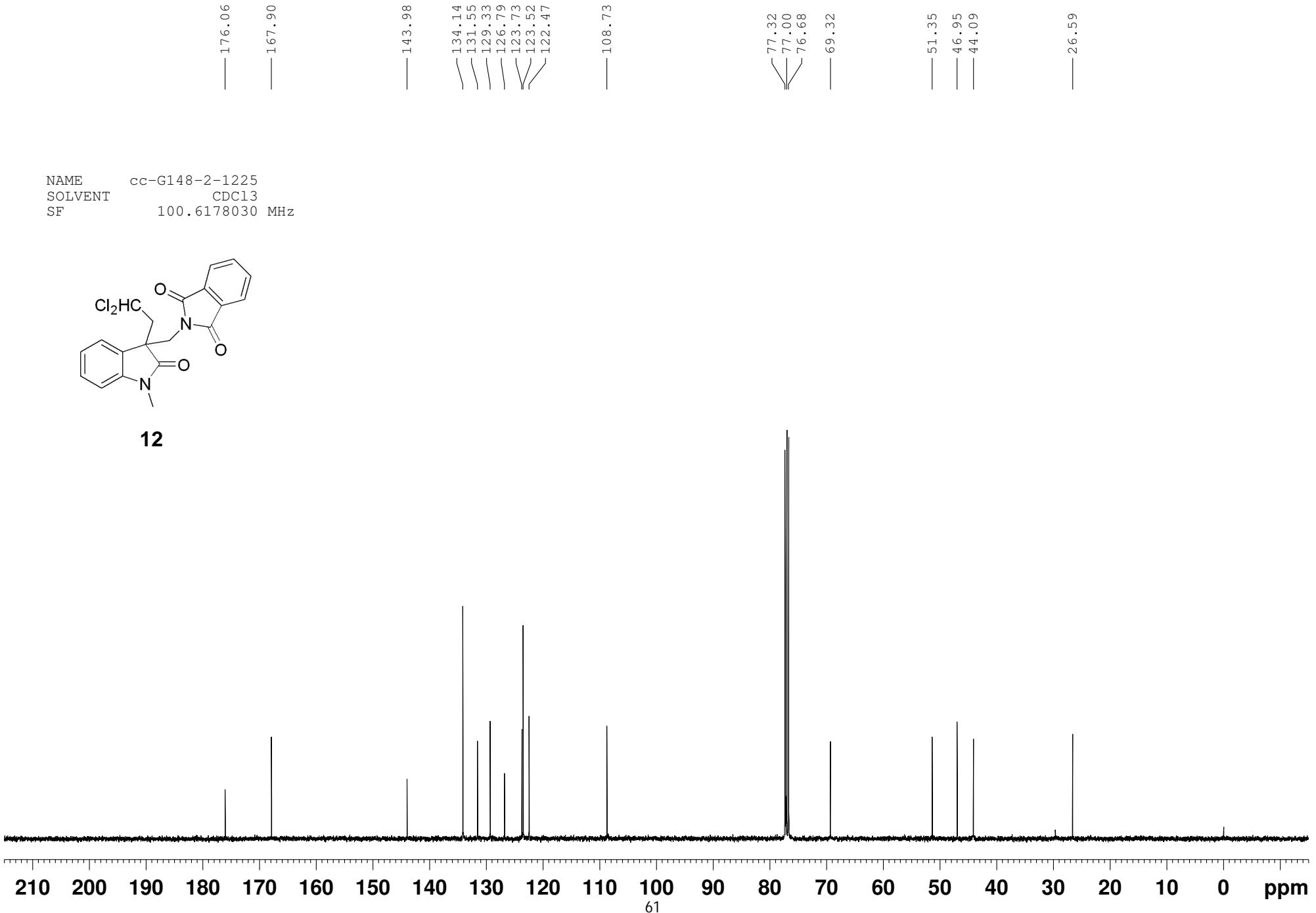
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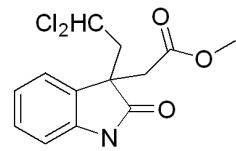
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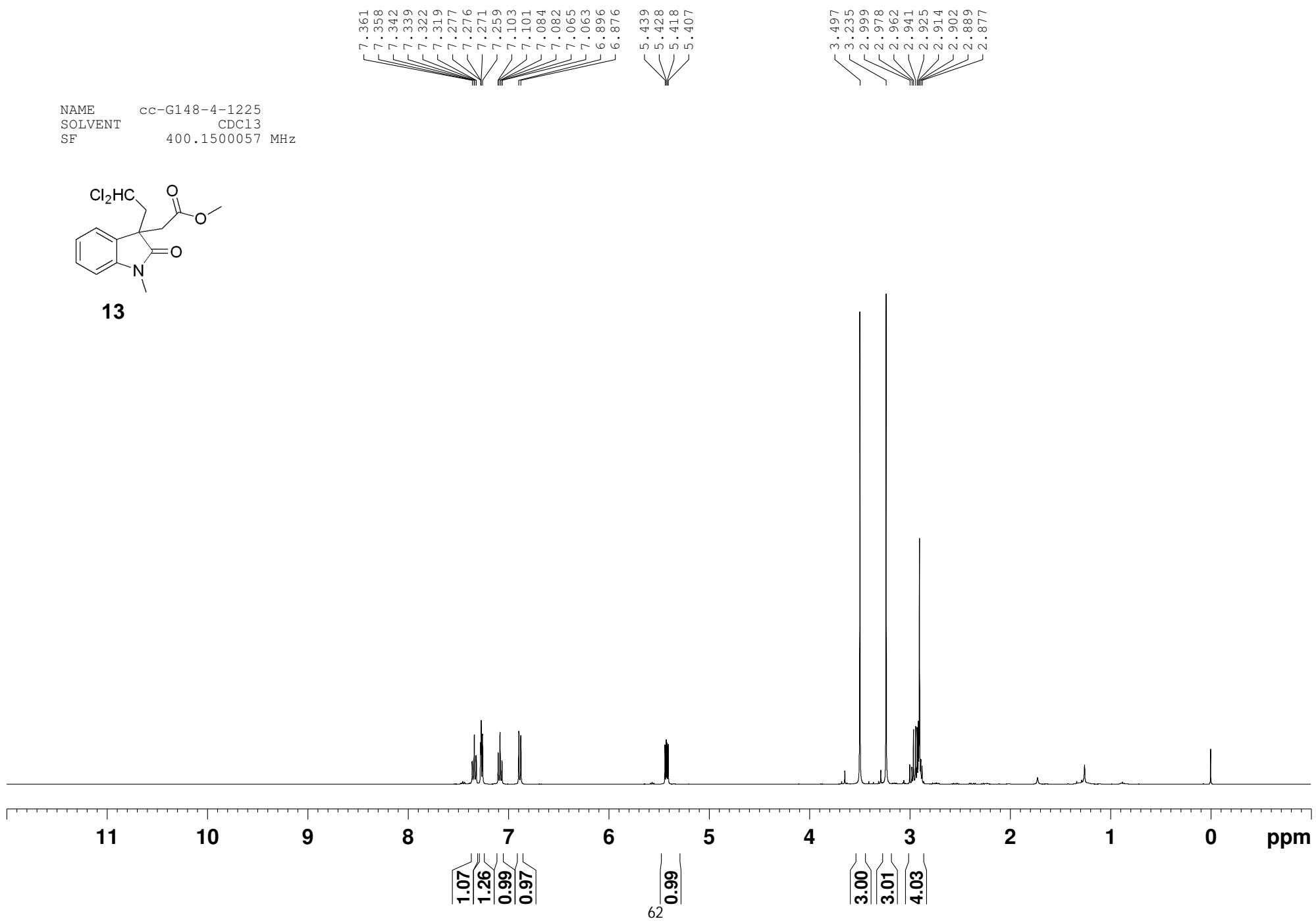
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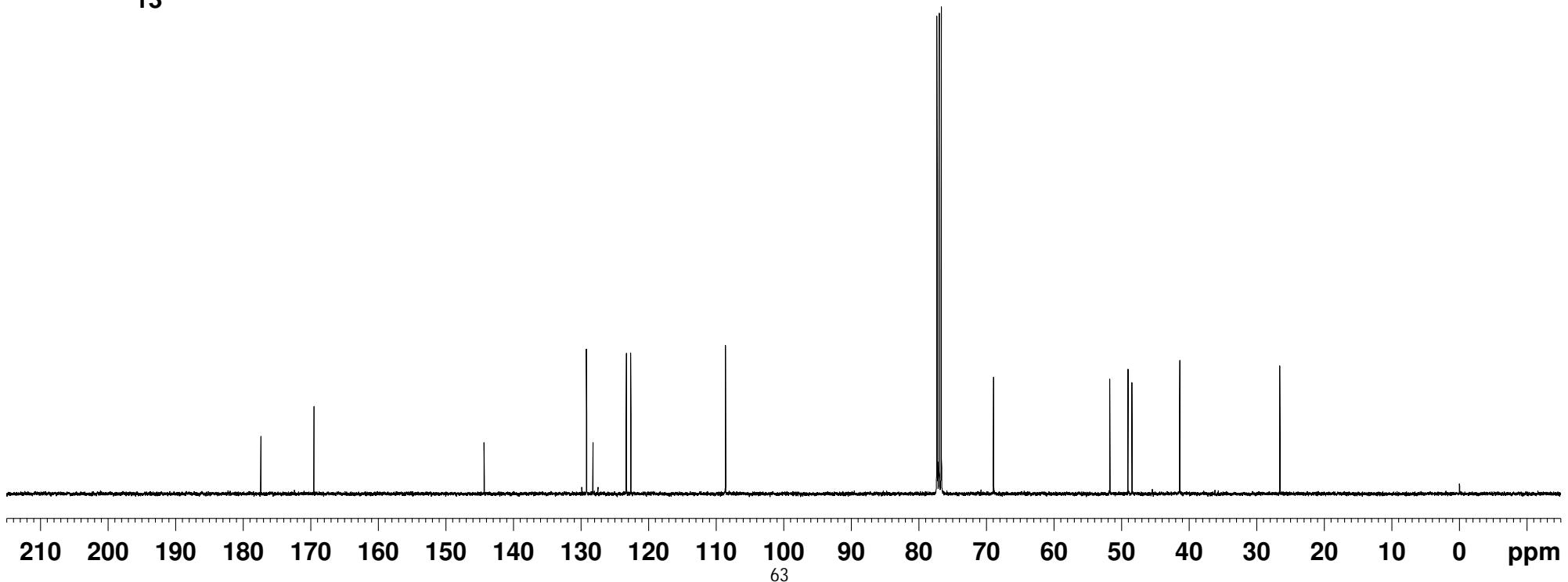
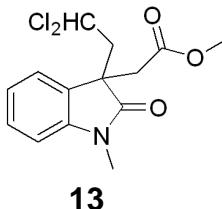
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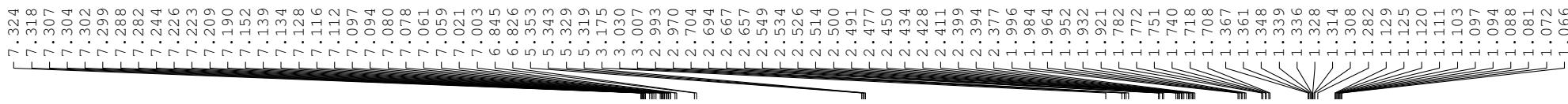


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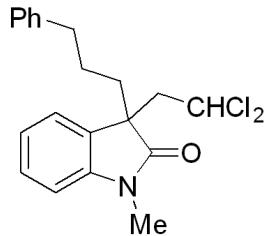


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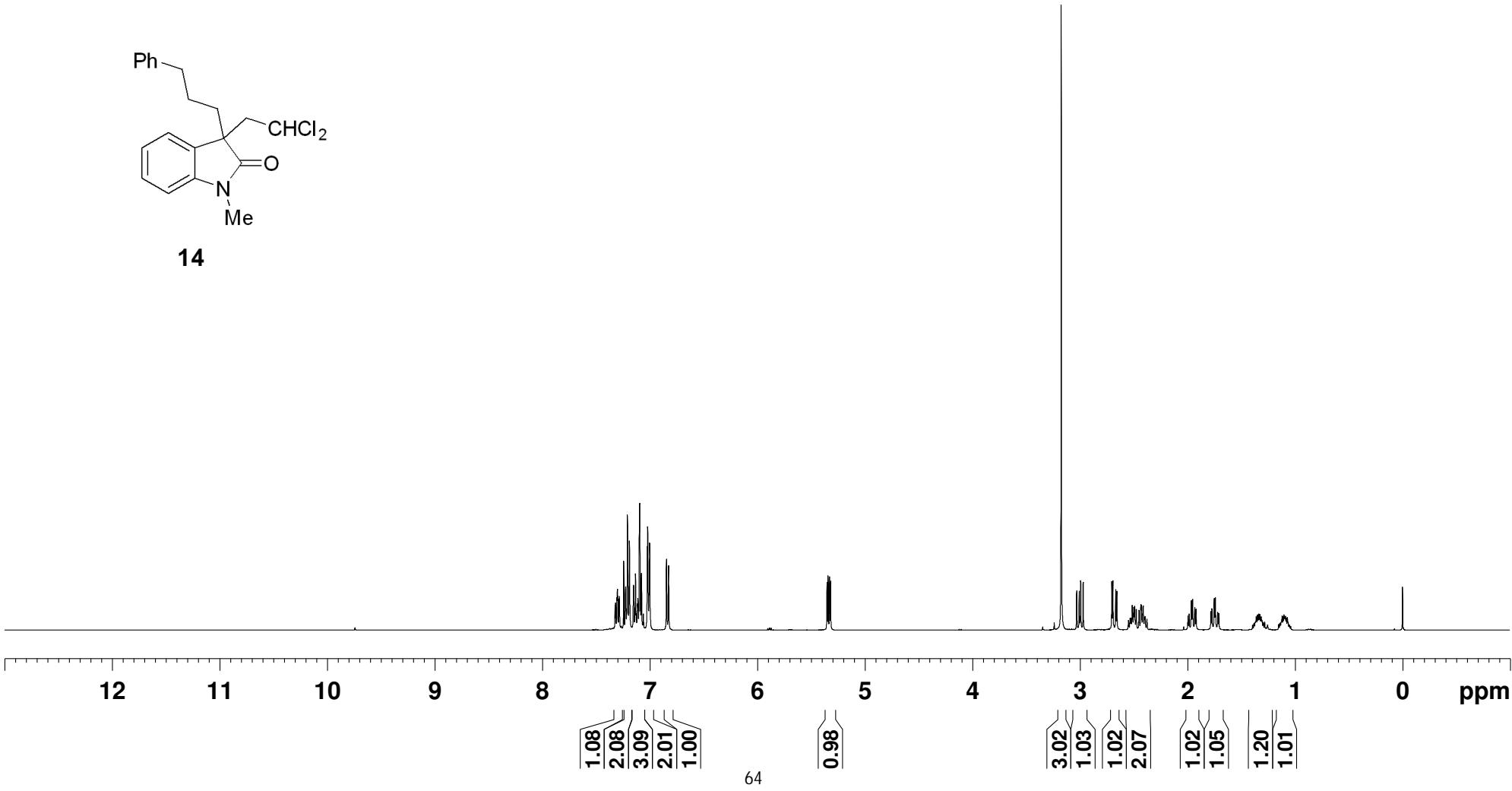




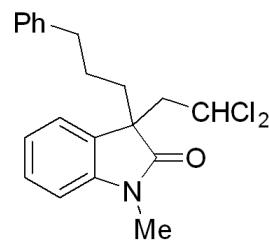
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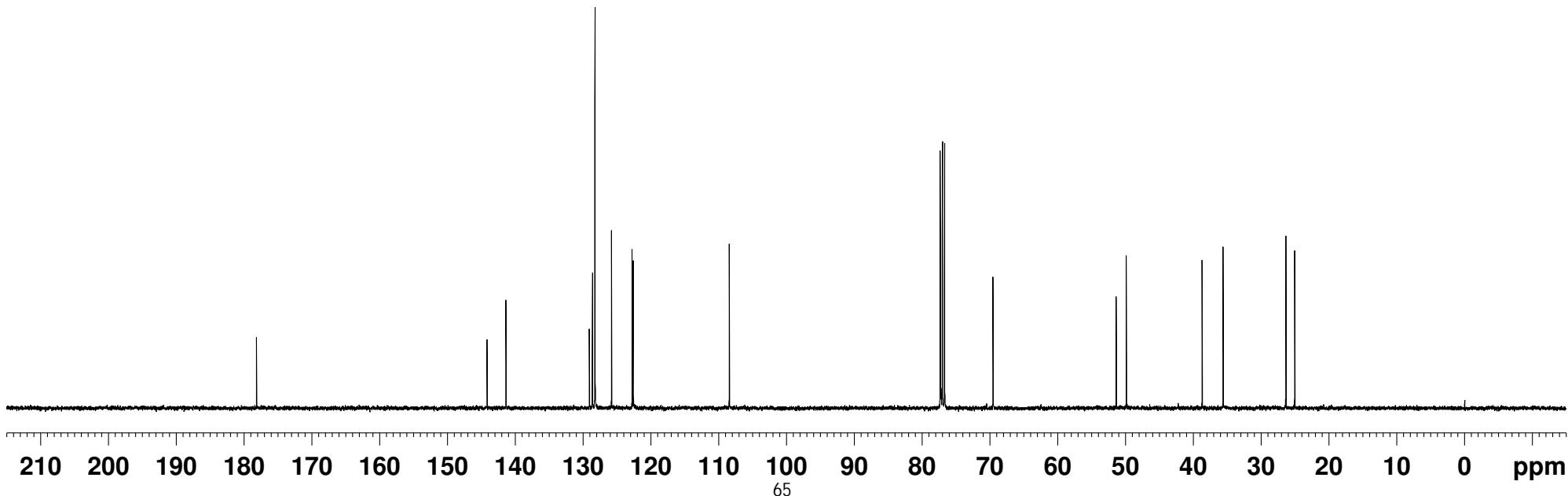
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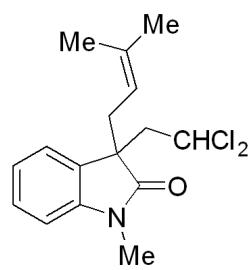
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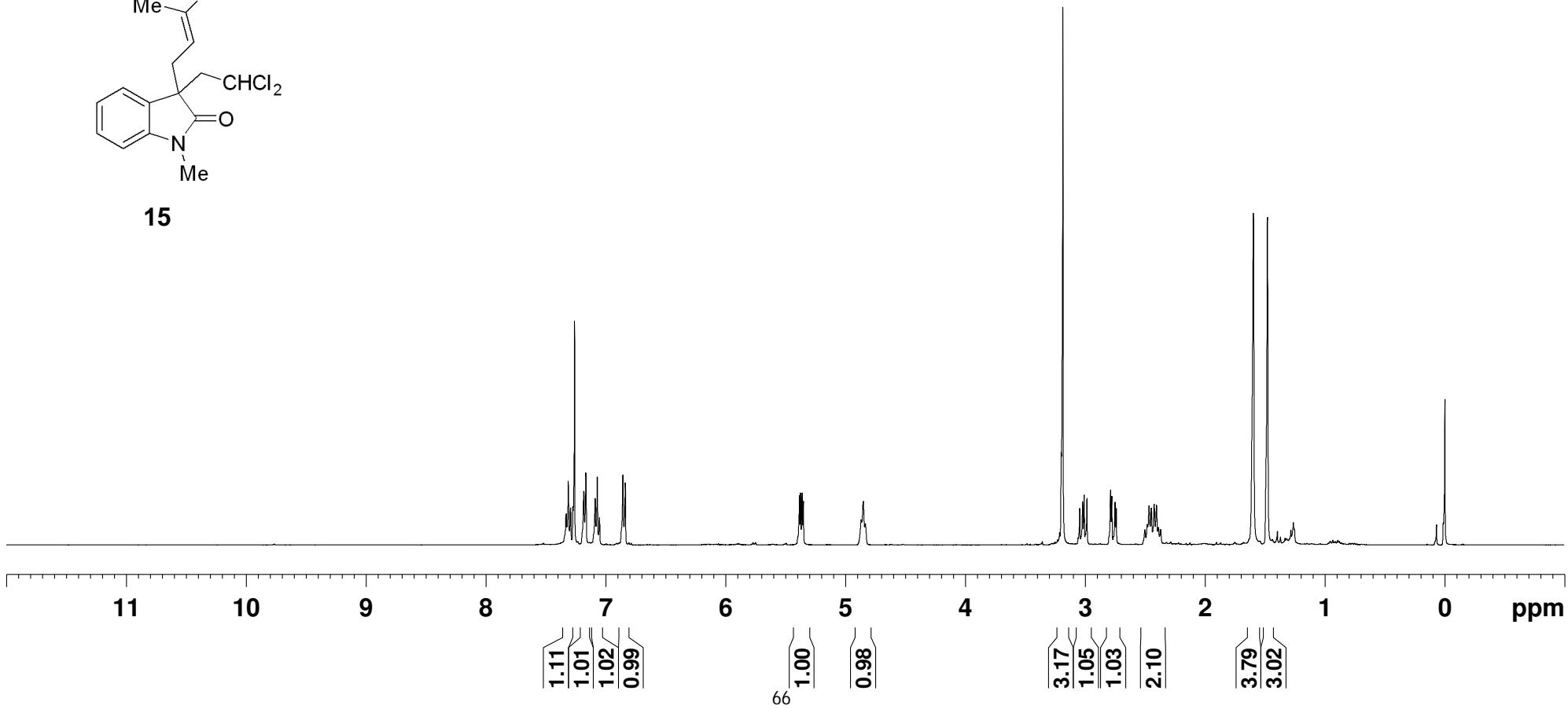
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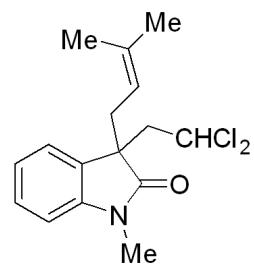
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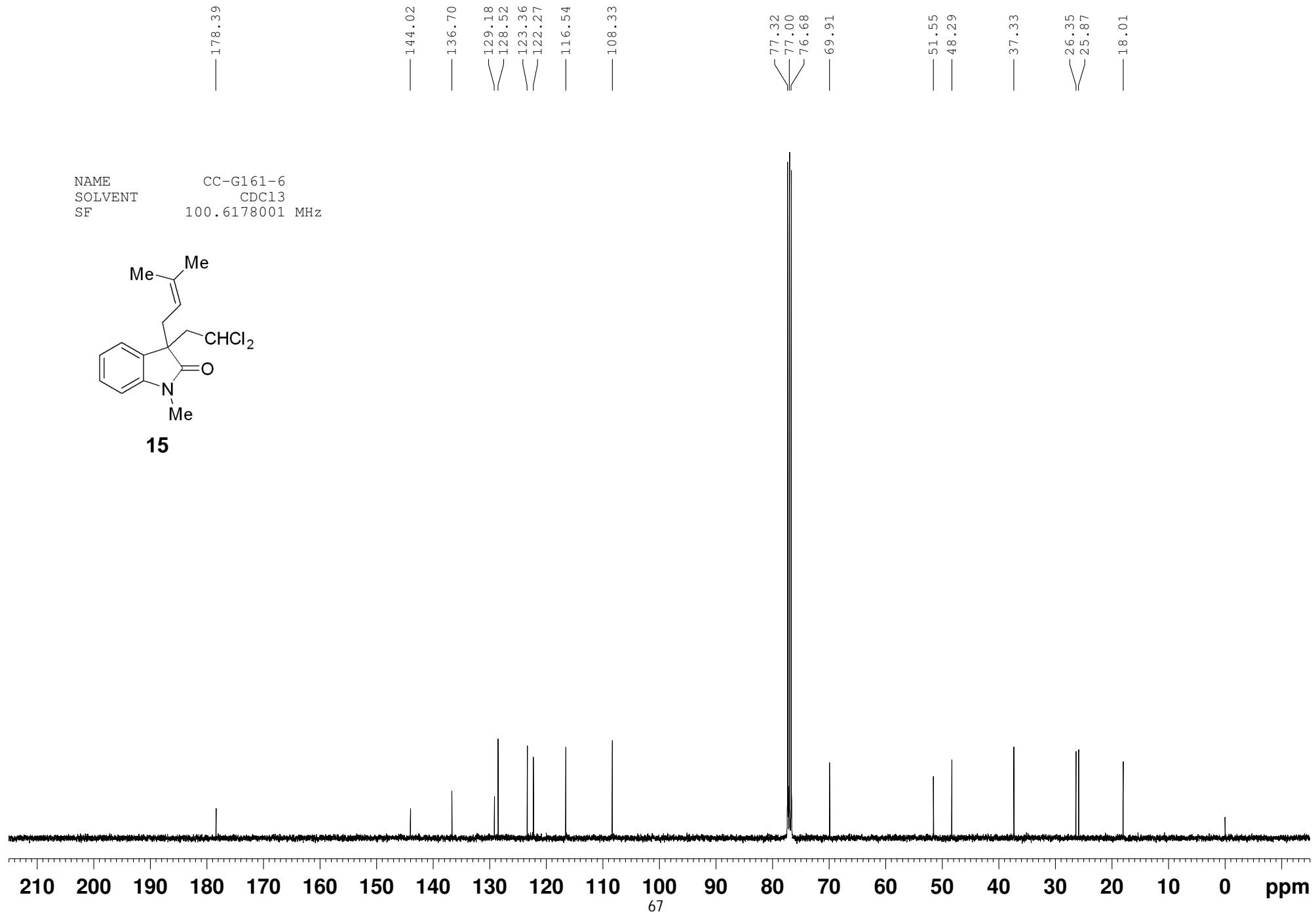
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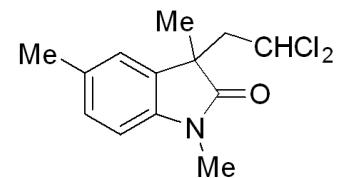
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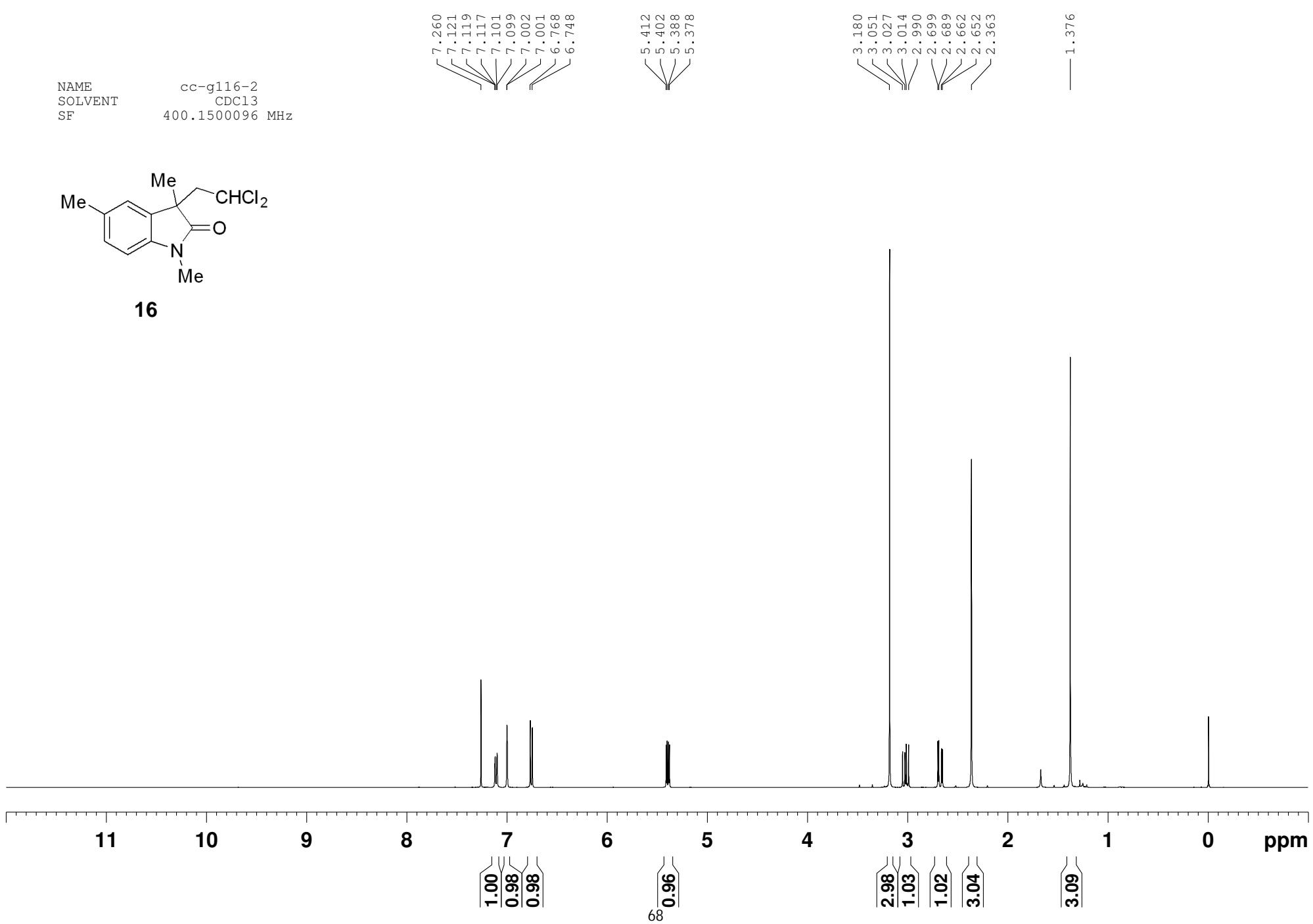
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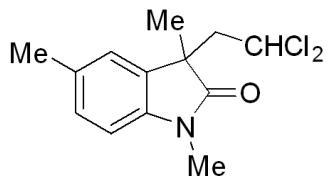
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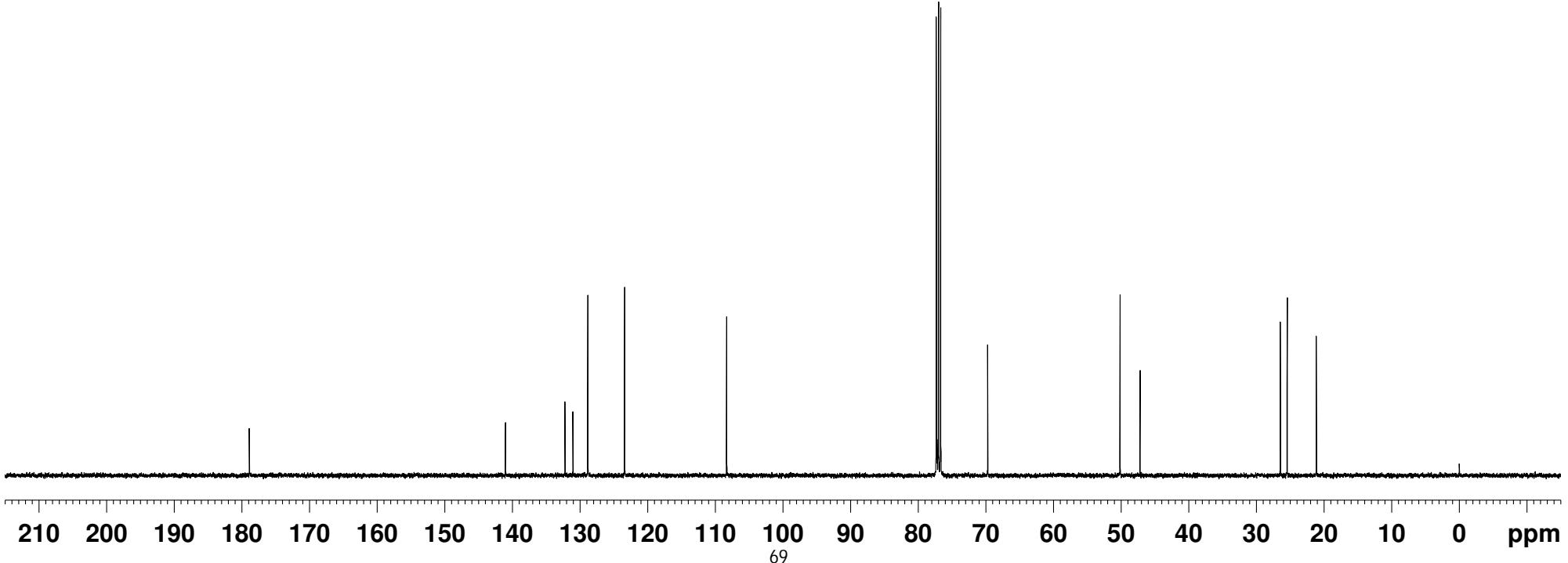
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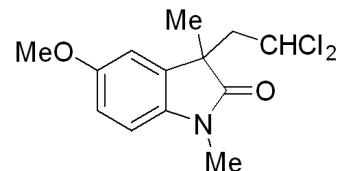
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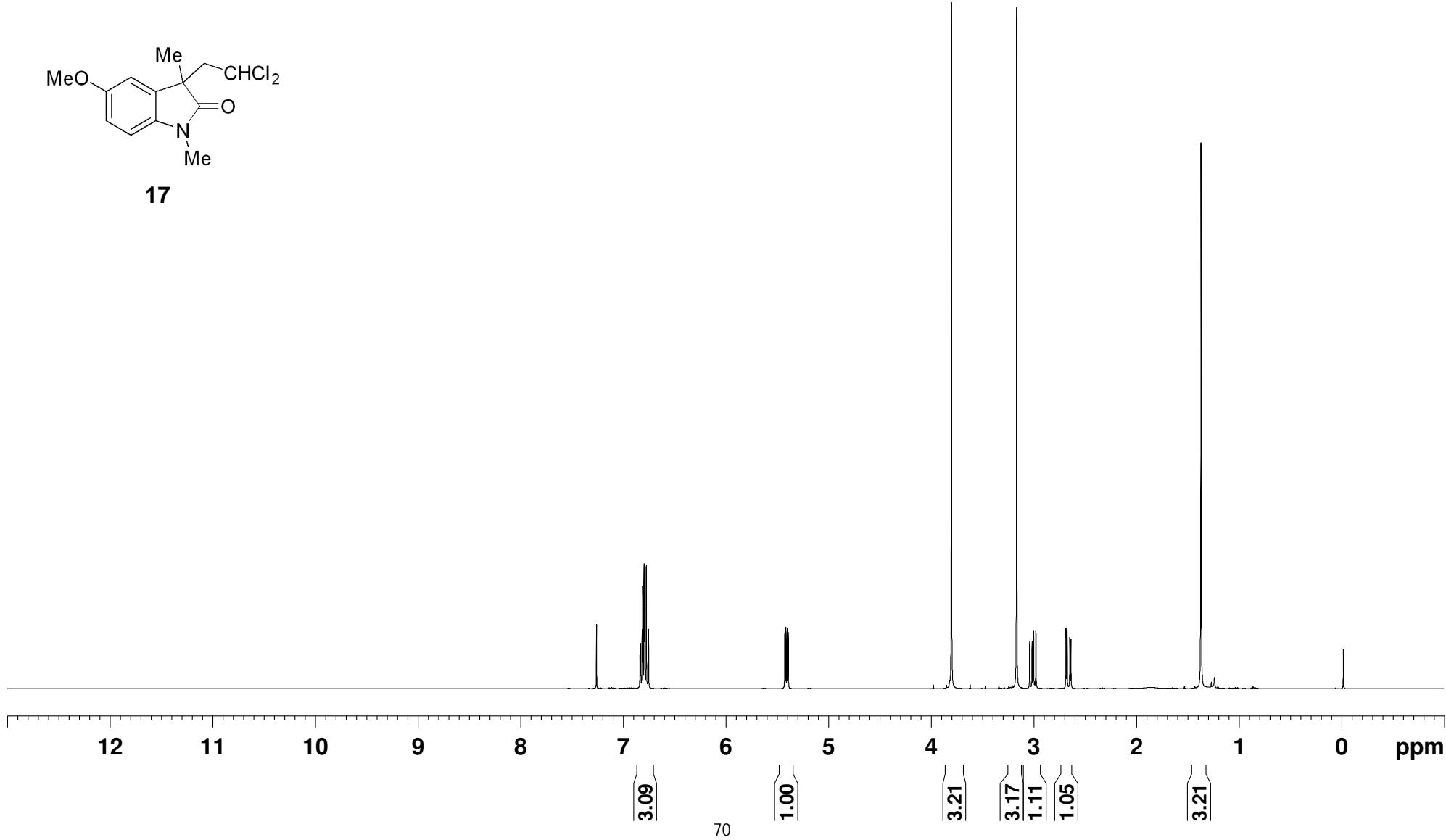
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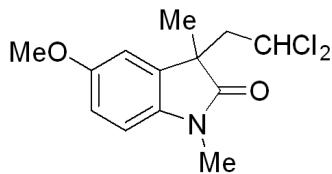
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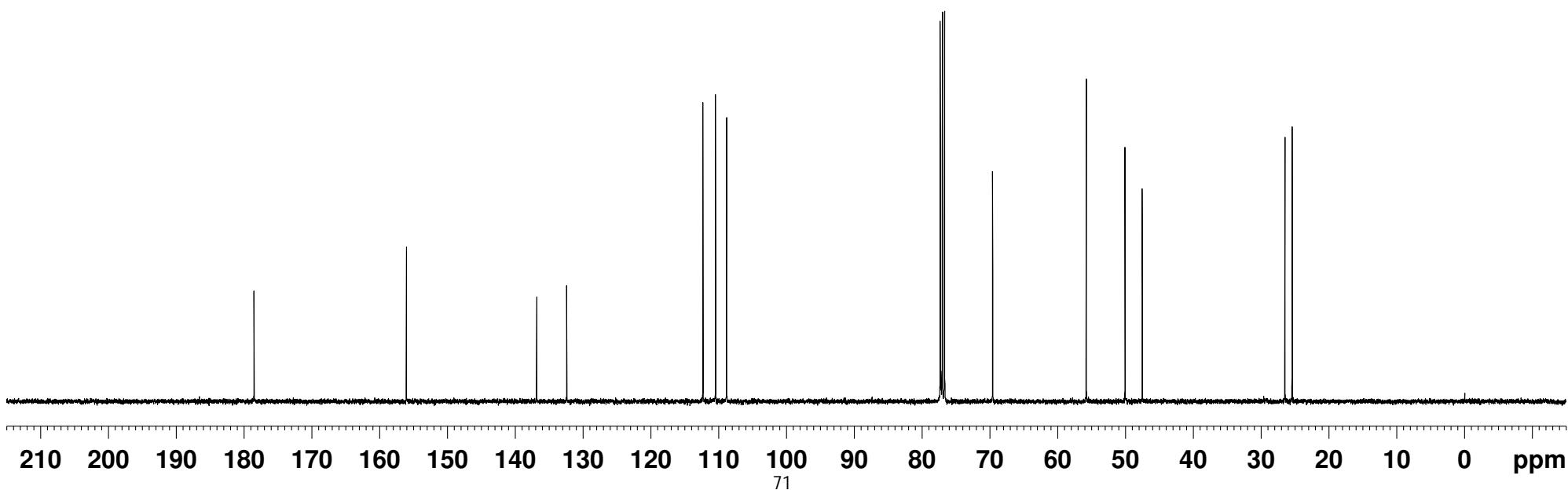
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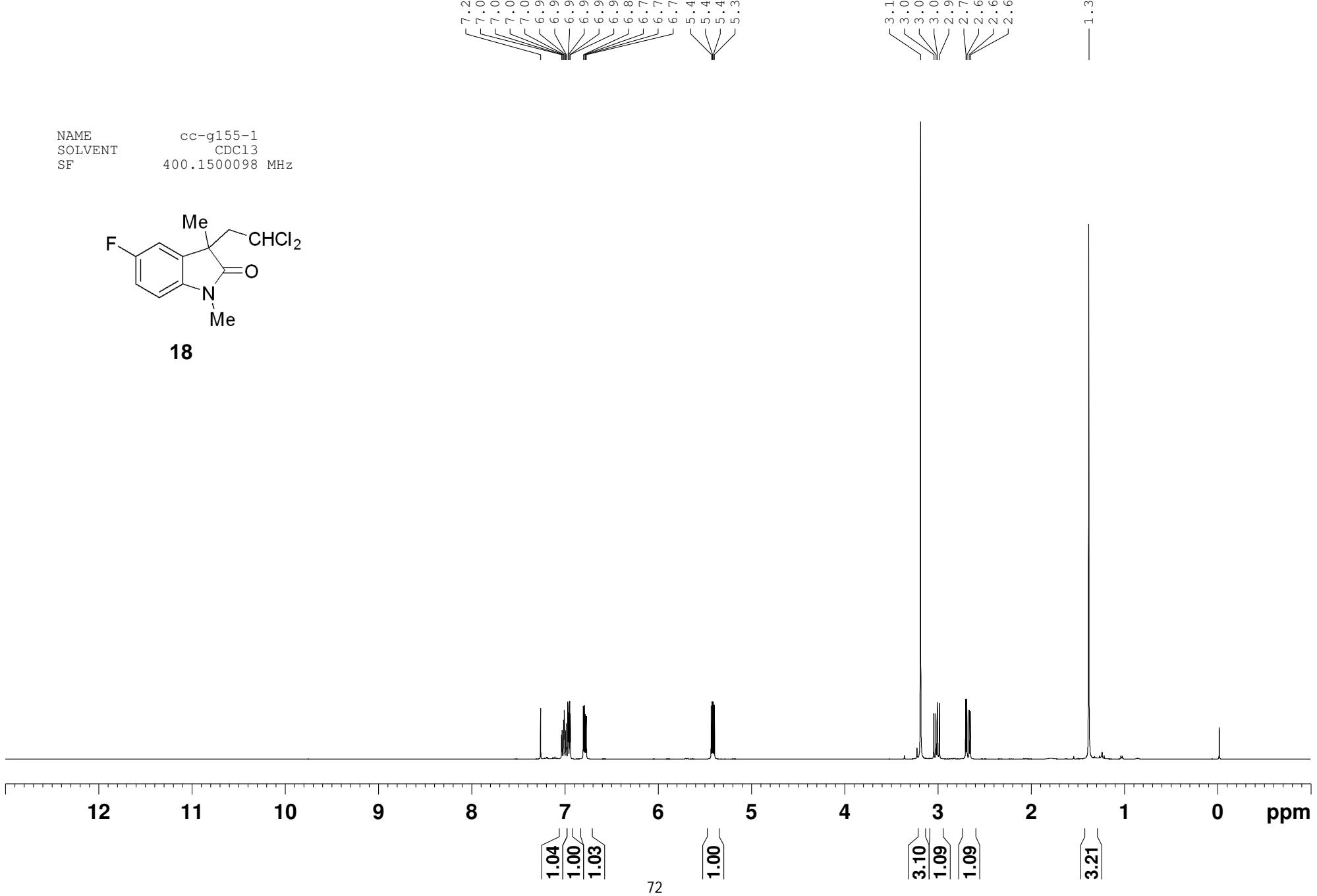
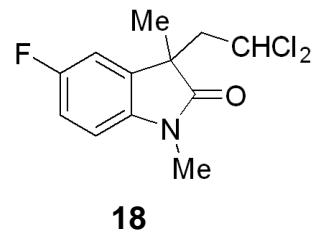
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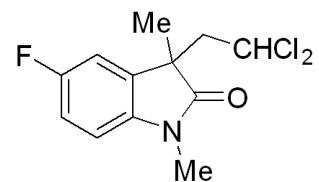


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SOLVENT CDCl₃
SF 400.1500098 MHz

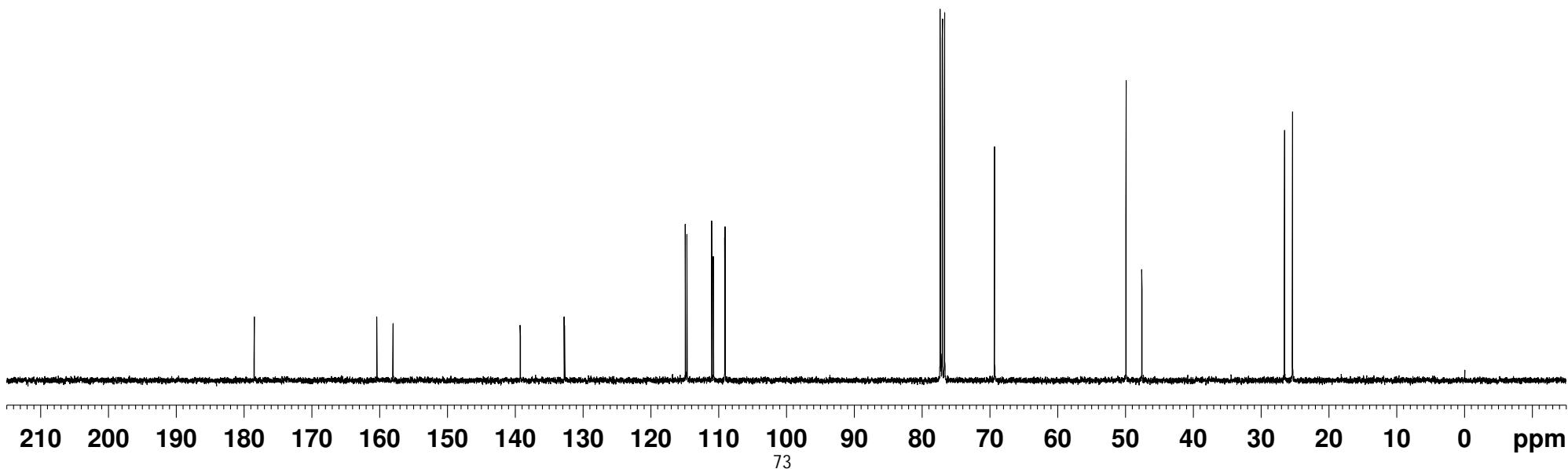


NAME
SOLVENT

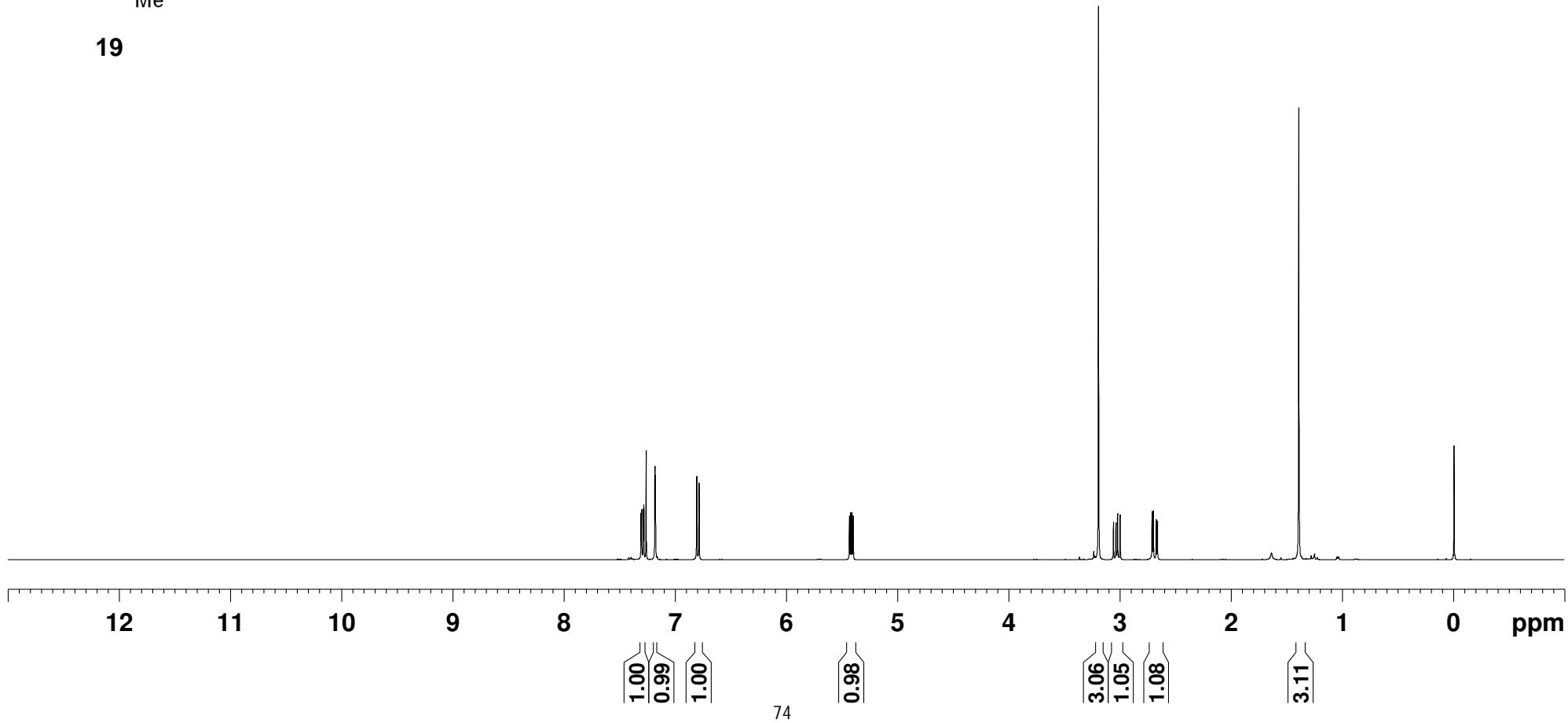
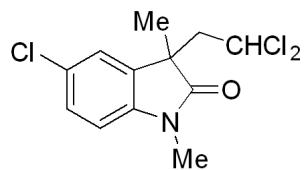
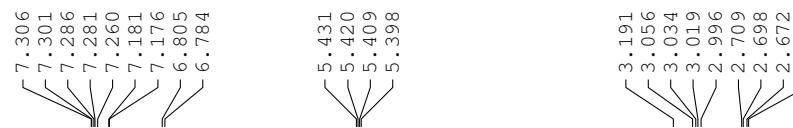
cc-g155-1
CDCl₃

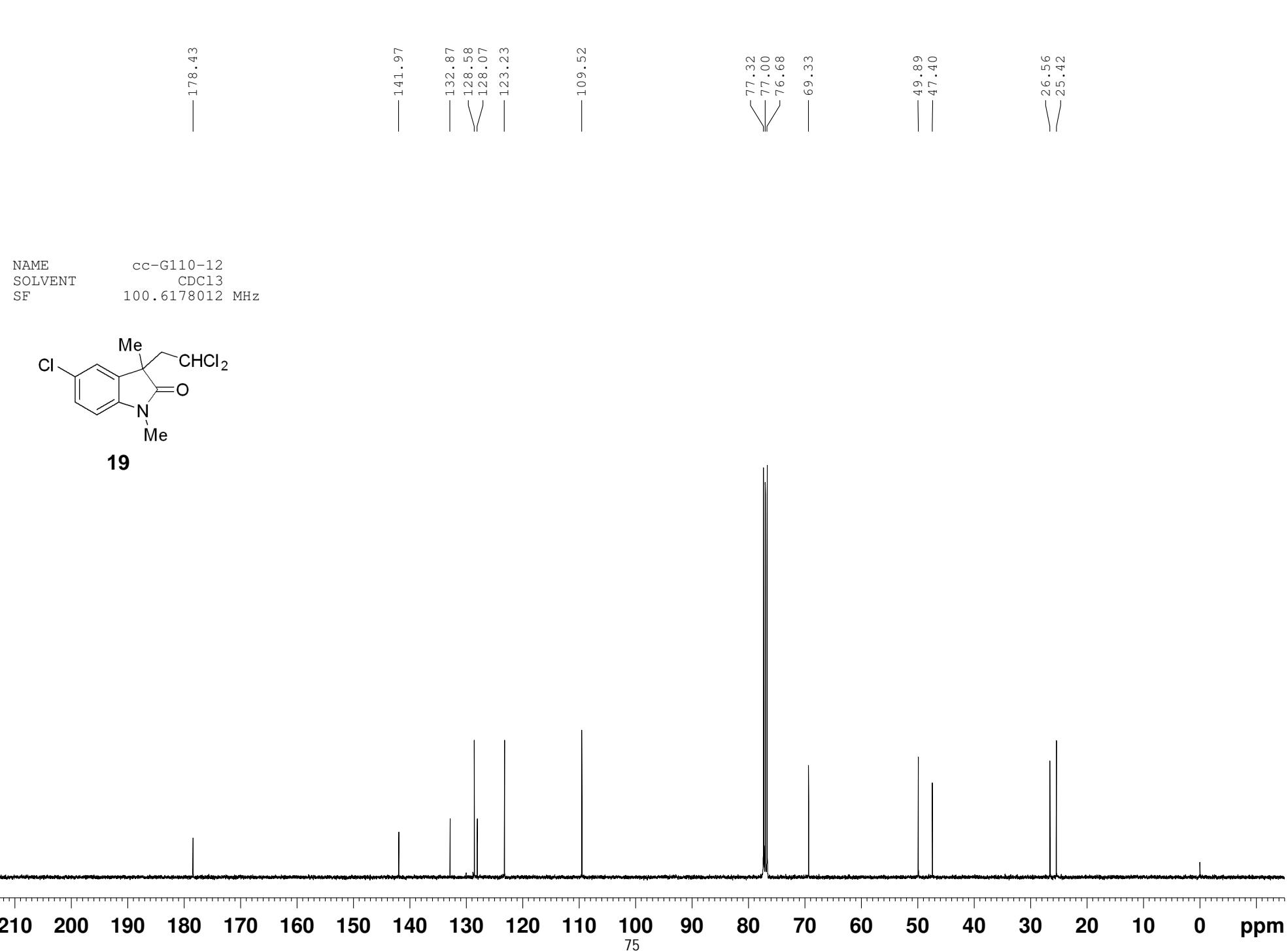


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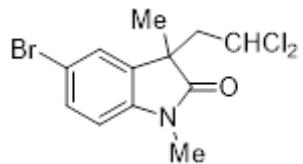


NAME cc-G110-12
SOLVENT CDCl₃
SF 400.1500096 MHz

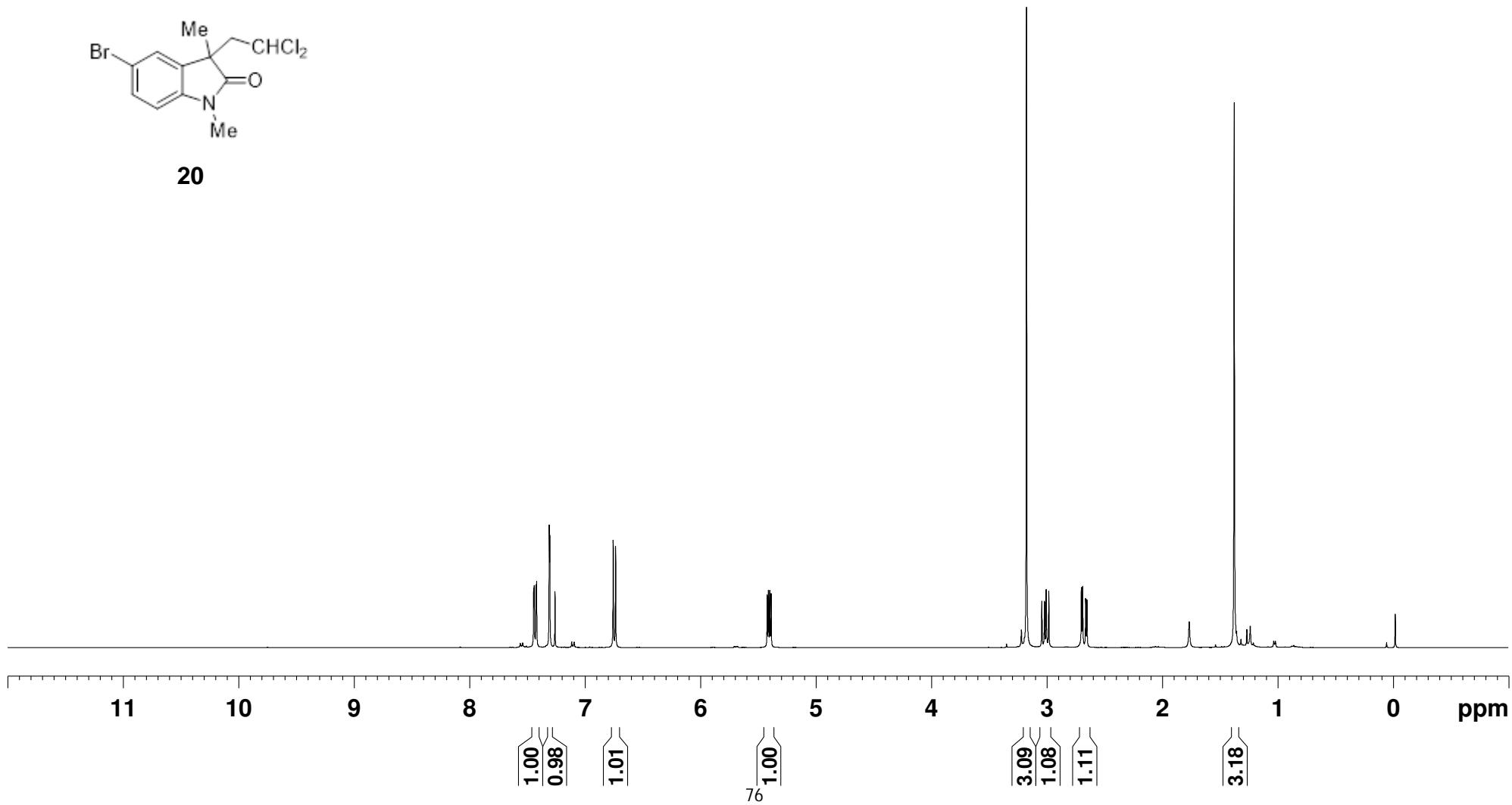




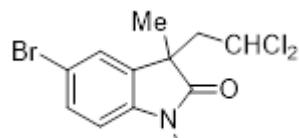
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SOLVENT CDCl₃
SF 400.1500092 MHz



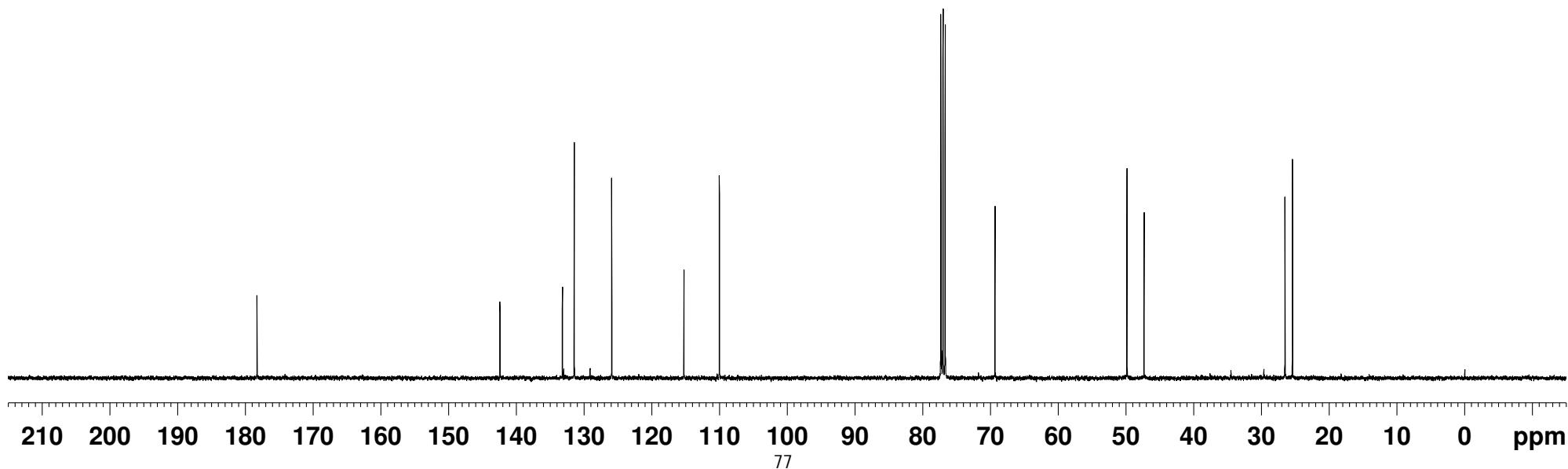
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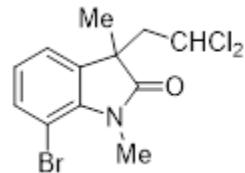
NAME cc-g155-2-0716
SOLVENT CDCl₃
SF 100.6178045 MHz



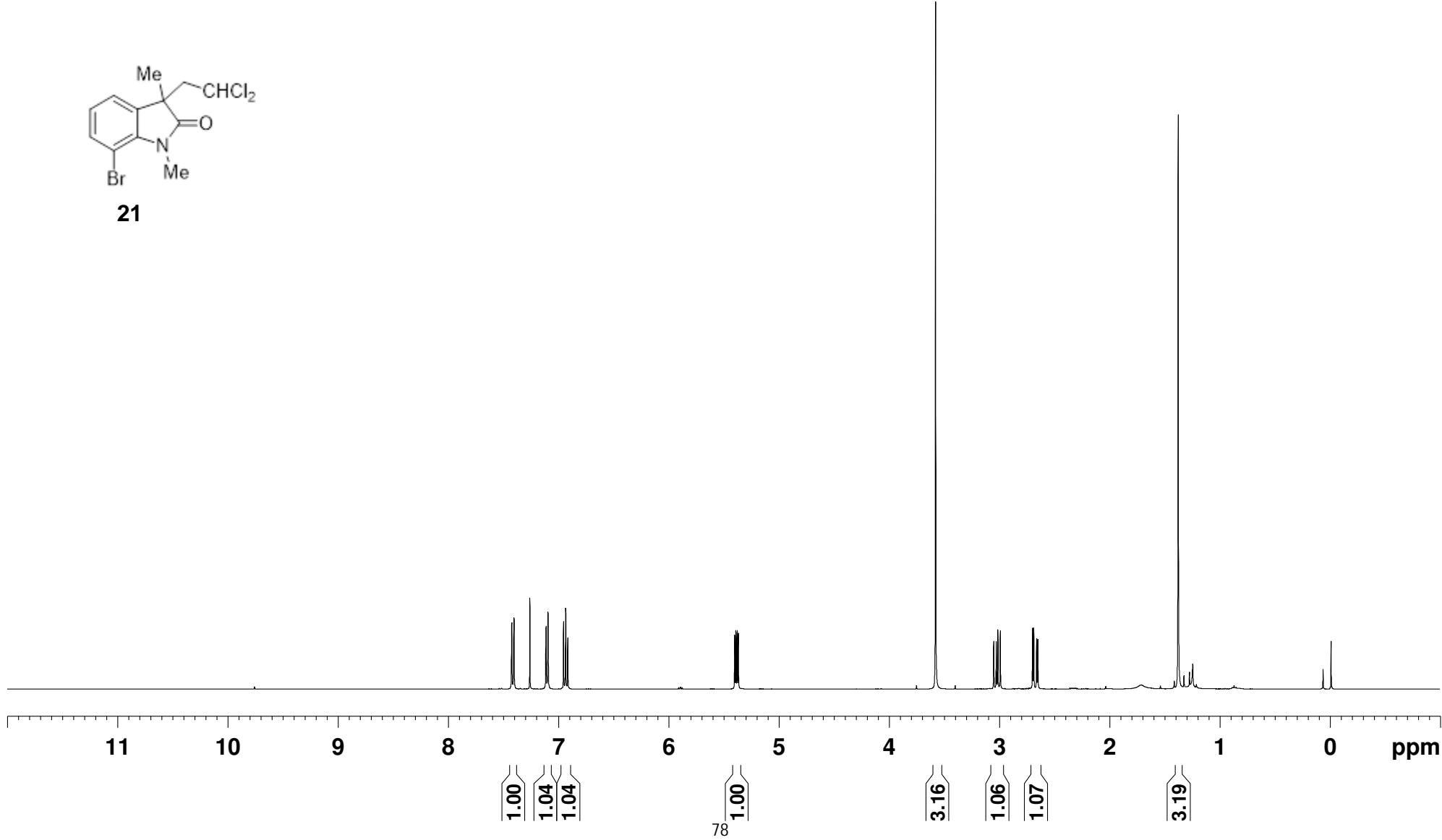
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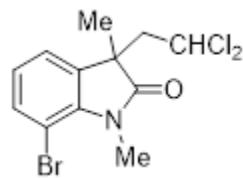
NAME cc-g155-4-1009
SOLVENT CDCl₃
SF 400.1500090 MHz



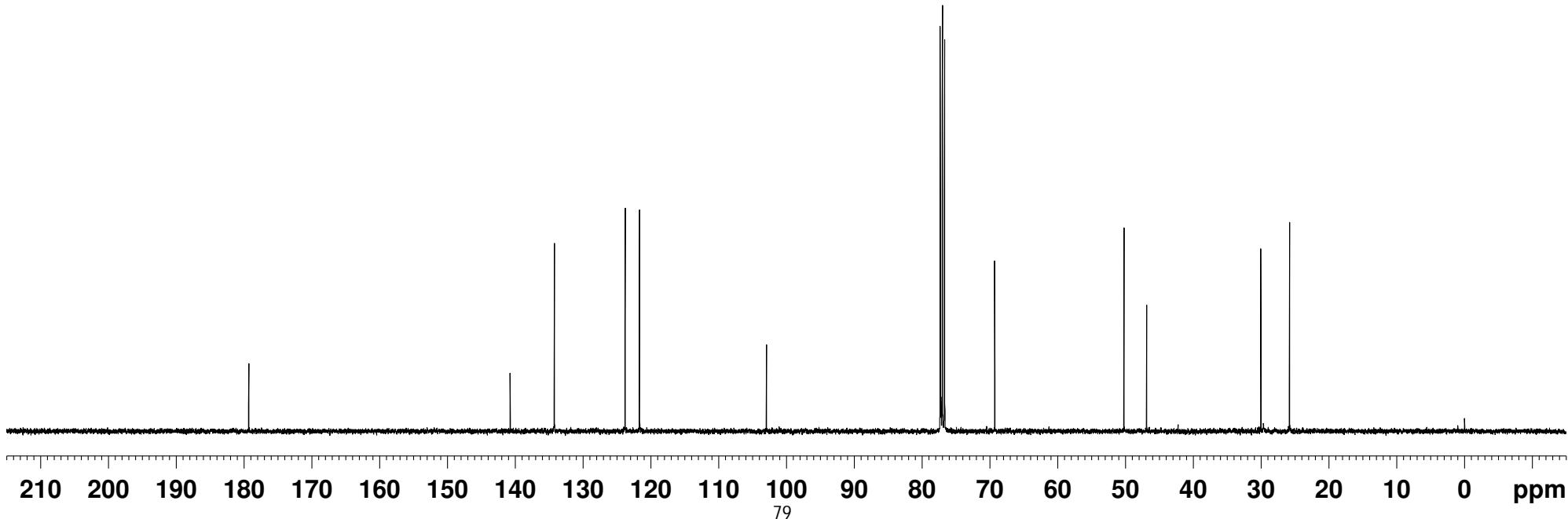
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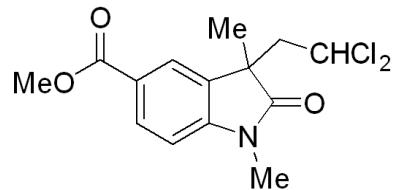
NAME cc-g155-4-1009
SOLVENT CDCl₃
SF 100.6178023 MHz



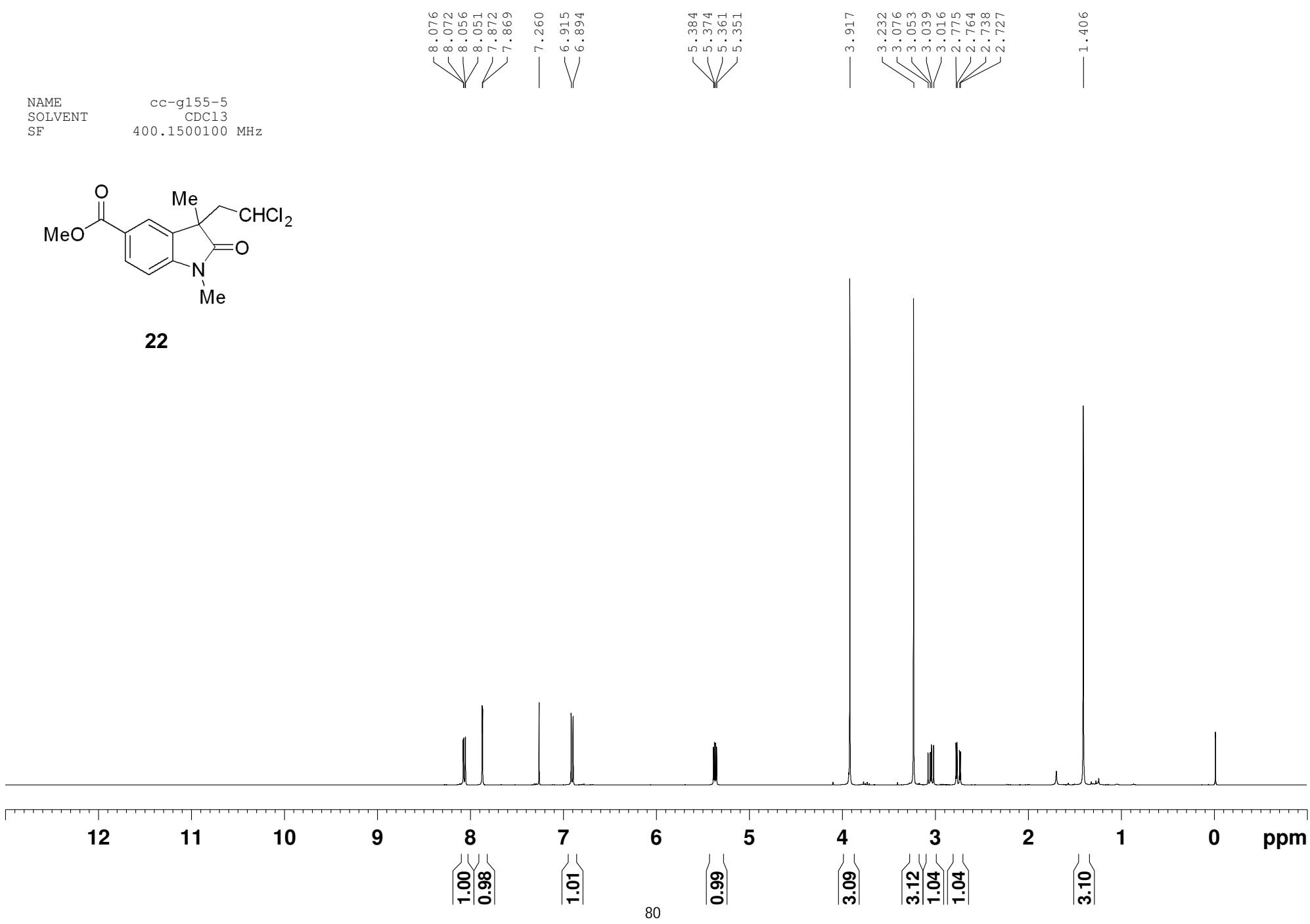
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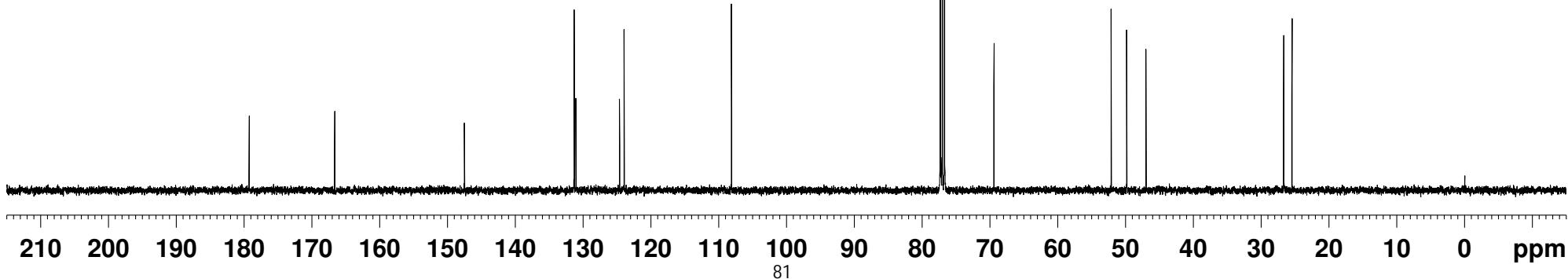
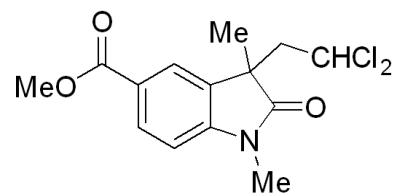
NAME cc-g155-5
SOLVENT CDCl₃
SF 400.1500100 MHz



22



NAME cc-g155-5
SOLVENT CDCl₃
SF 100.6178023 MHz



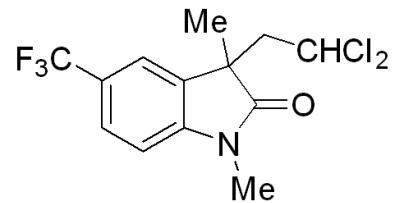
NAME cc-G116-3-1125
SOLVENT CDCl₃
SF 400.1500101 MHz

7.620
7.600
7.432
7.260
6.958
6.938

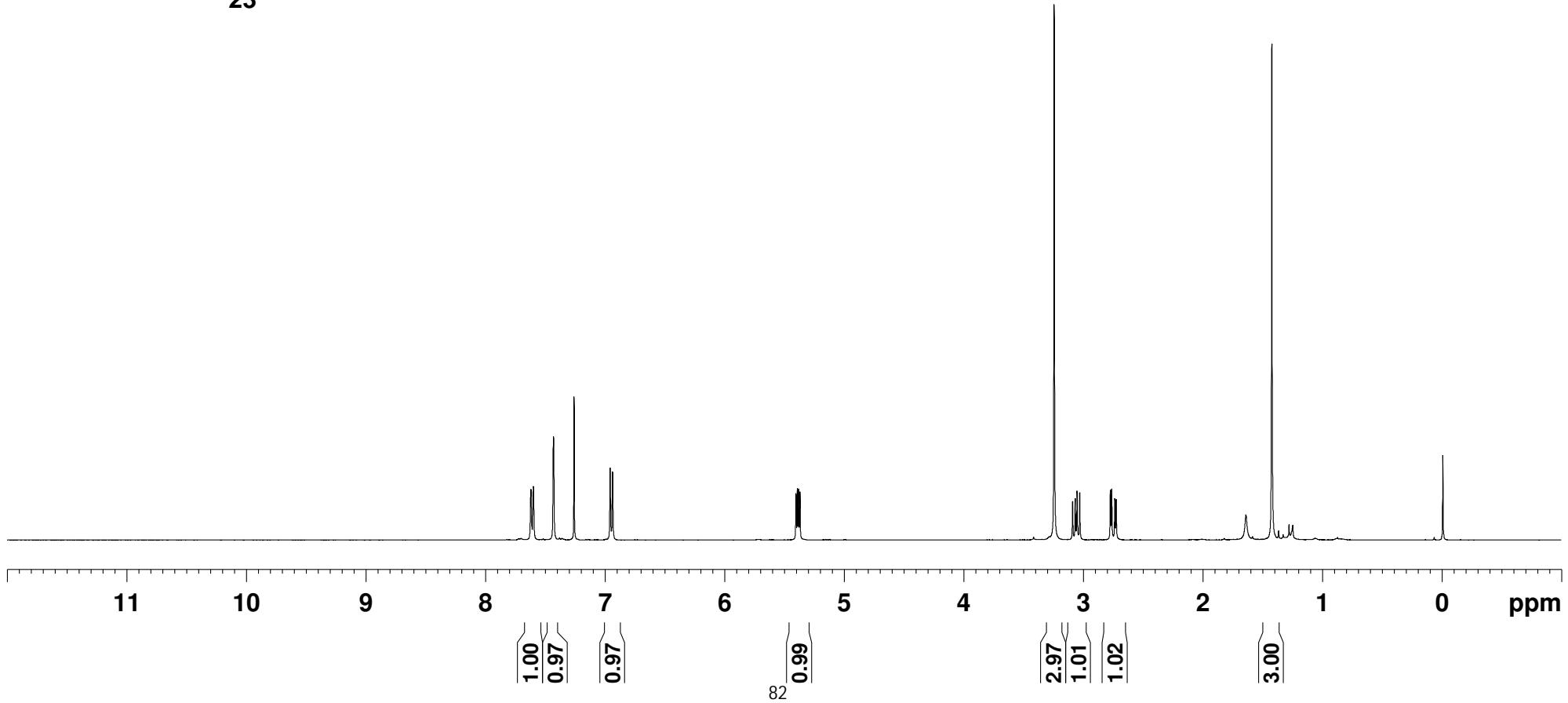
5.404
5.393
5.382
5.370

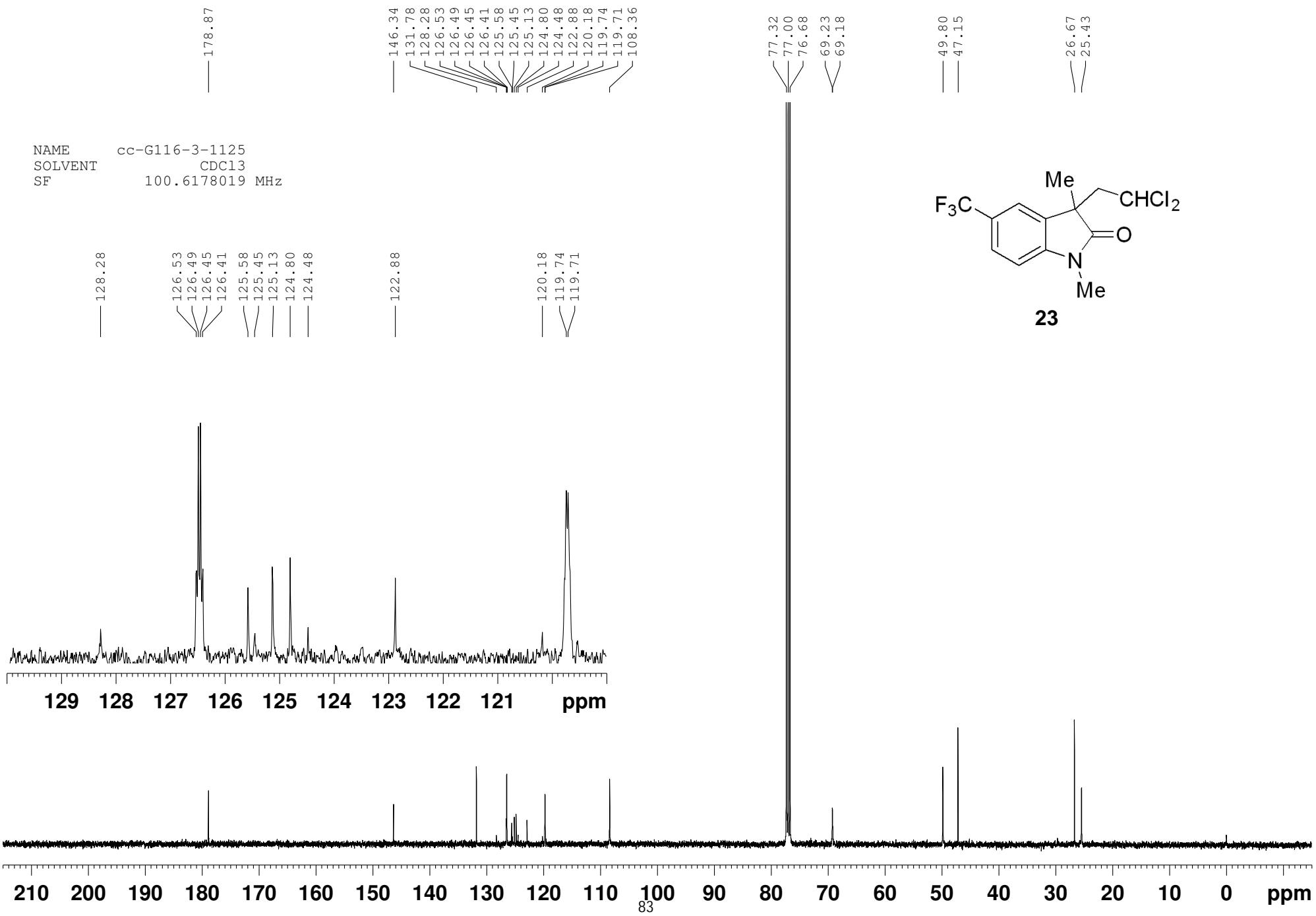
3.244
3.089
3.067
3.052
3.030
2.774
2.762
2.736
2.725

1.423



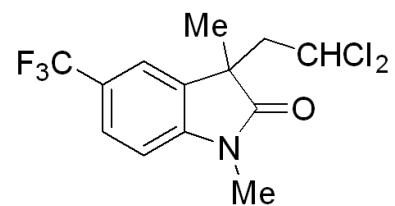
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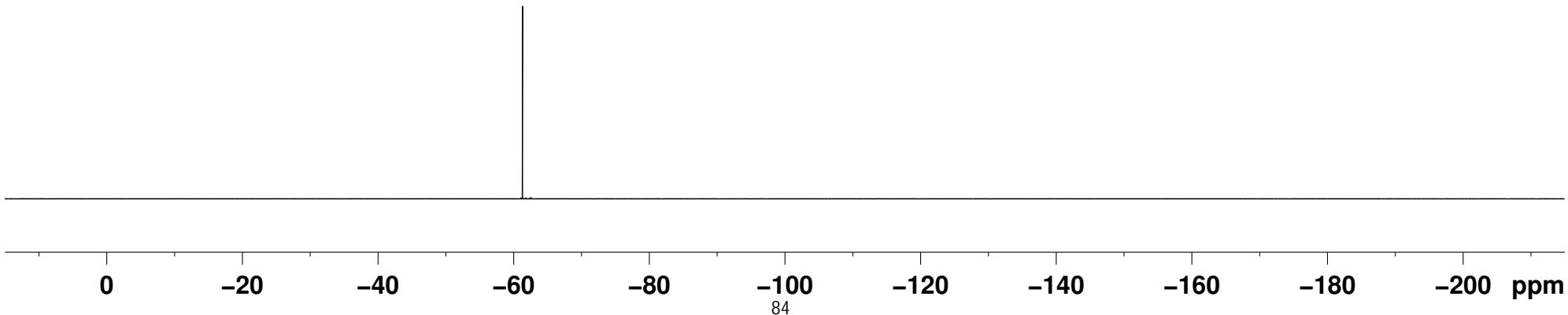


NAME cc-G116-3-1125
SOLVENT CDCl₃
SF 376.5171850 MHz

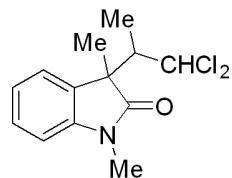
-61.324



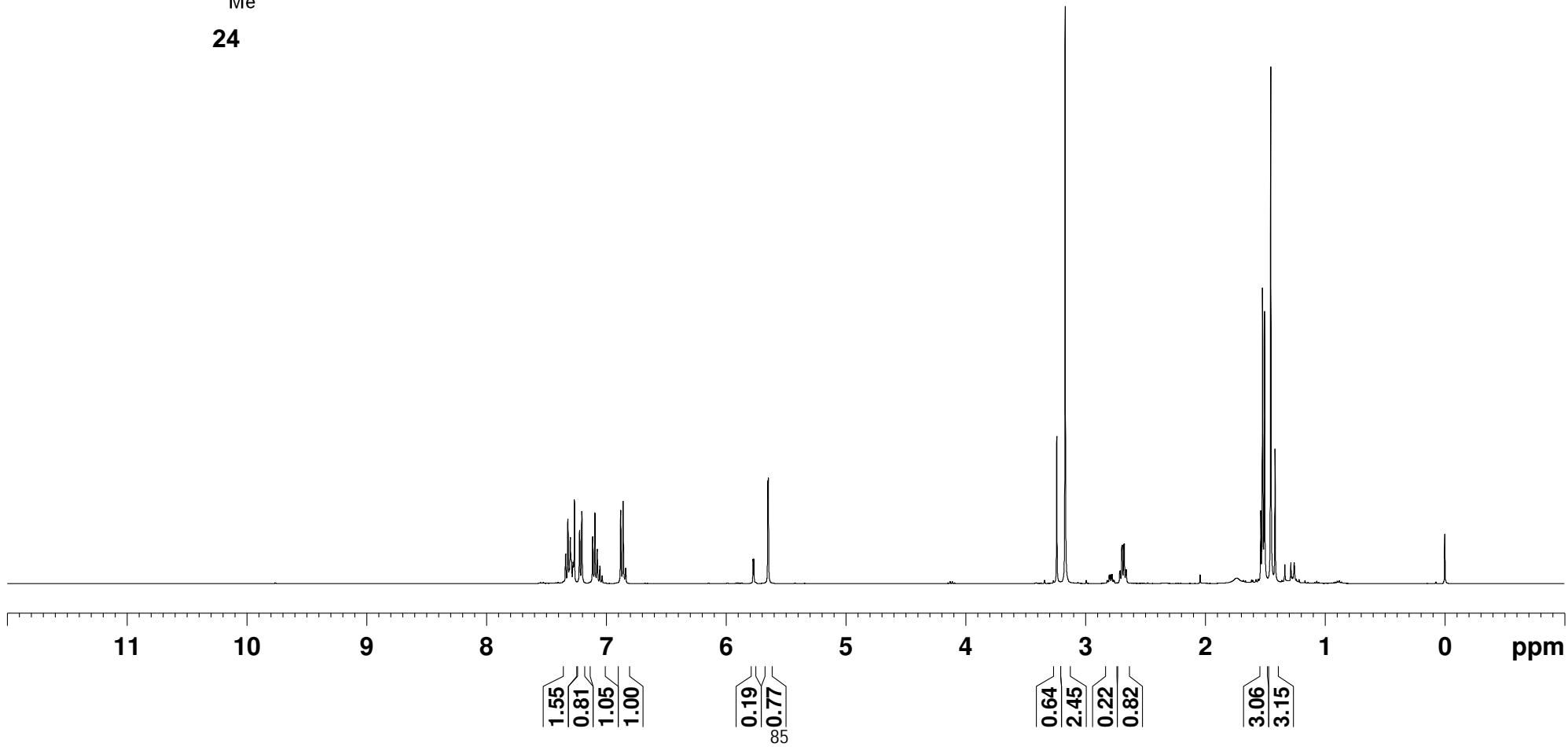
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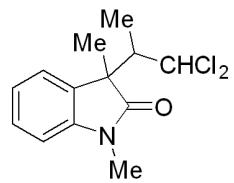
NAME cc-h129-3-0608
SOLVENT CDCl₃
SF 400.1500070 MHz



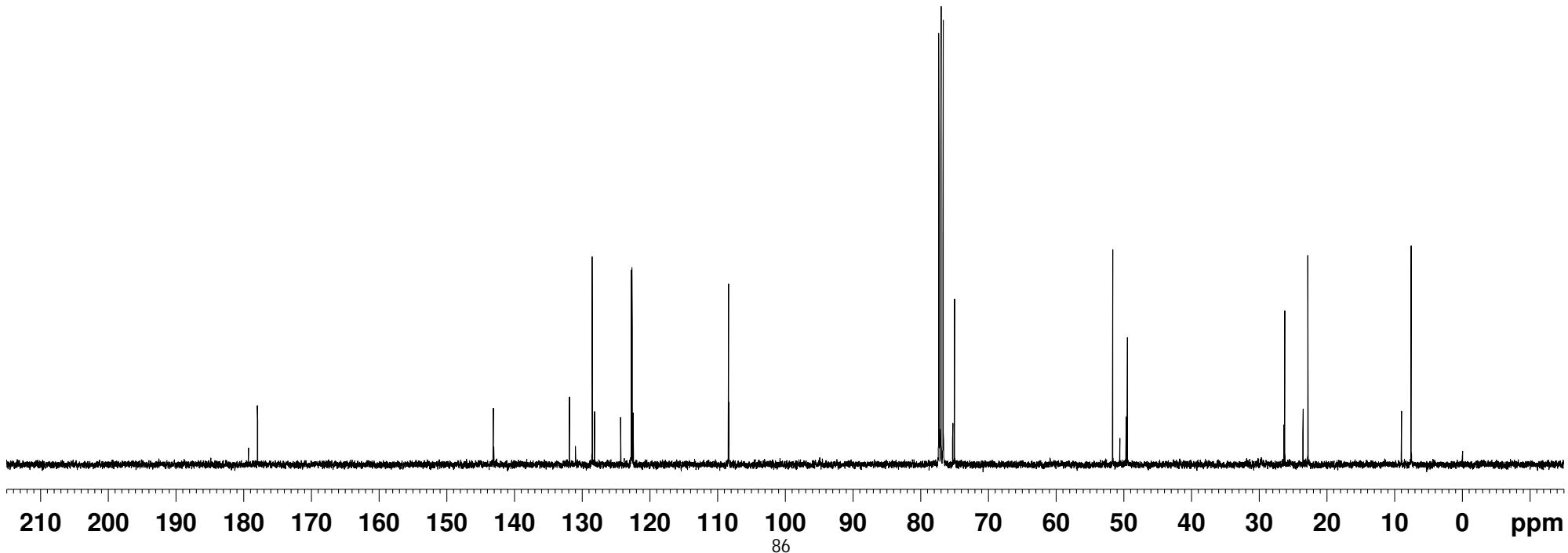
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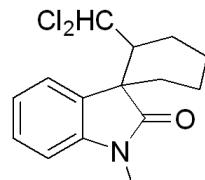
NAME cc-h129-3-0608
SOLVENT CDC13
SF 100.6178030 MHz



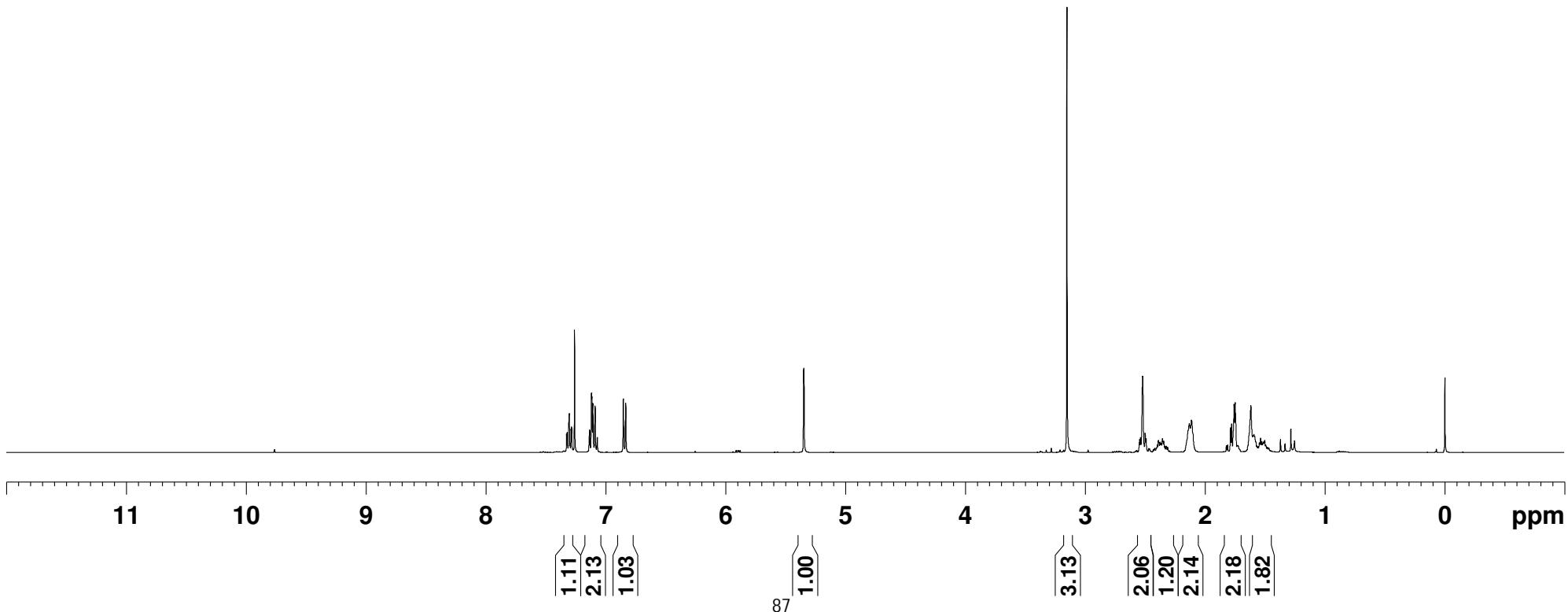
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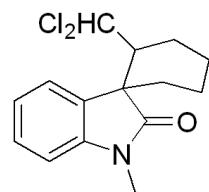
NAME cc-h163-up
SOLVENT CDCl₃
SF 400.1500096 MHz



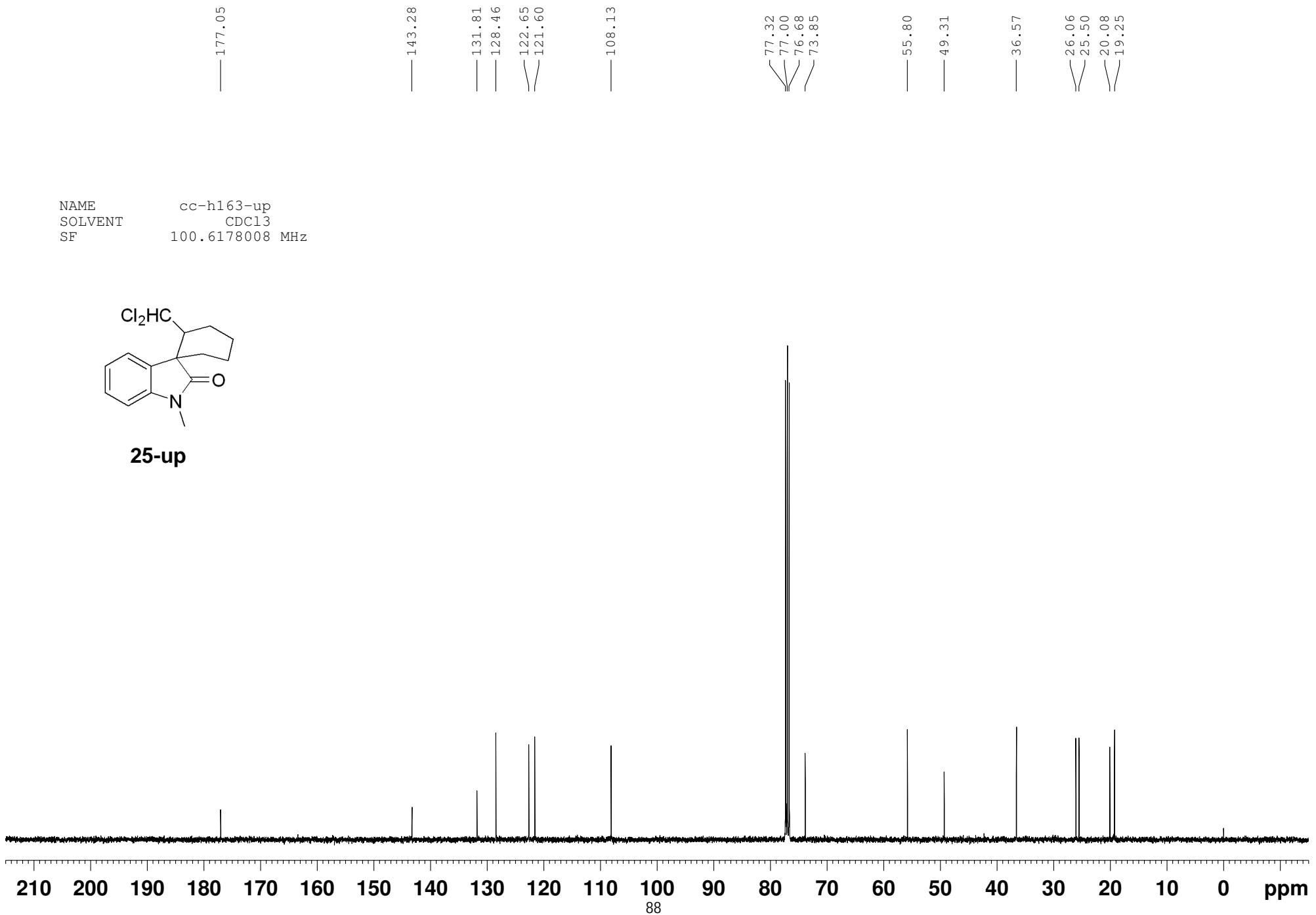
25-up



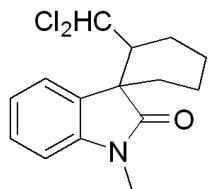
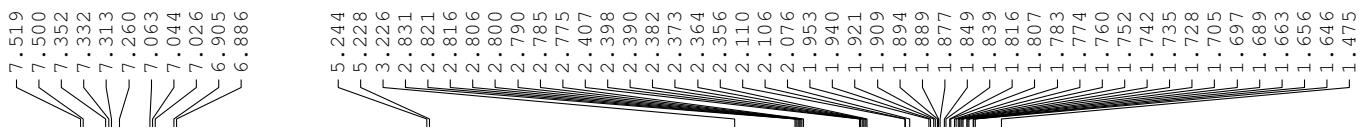
NAME cc-h163-up
SOLVENT CDC13
SF 100.6178008 MHz



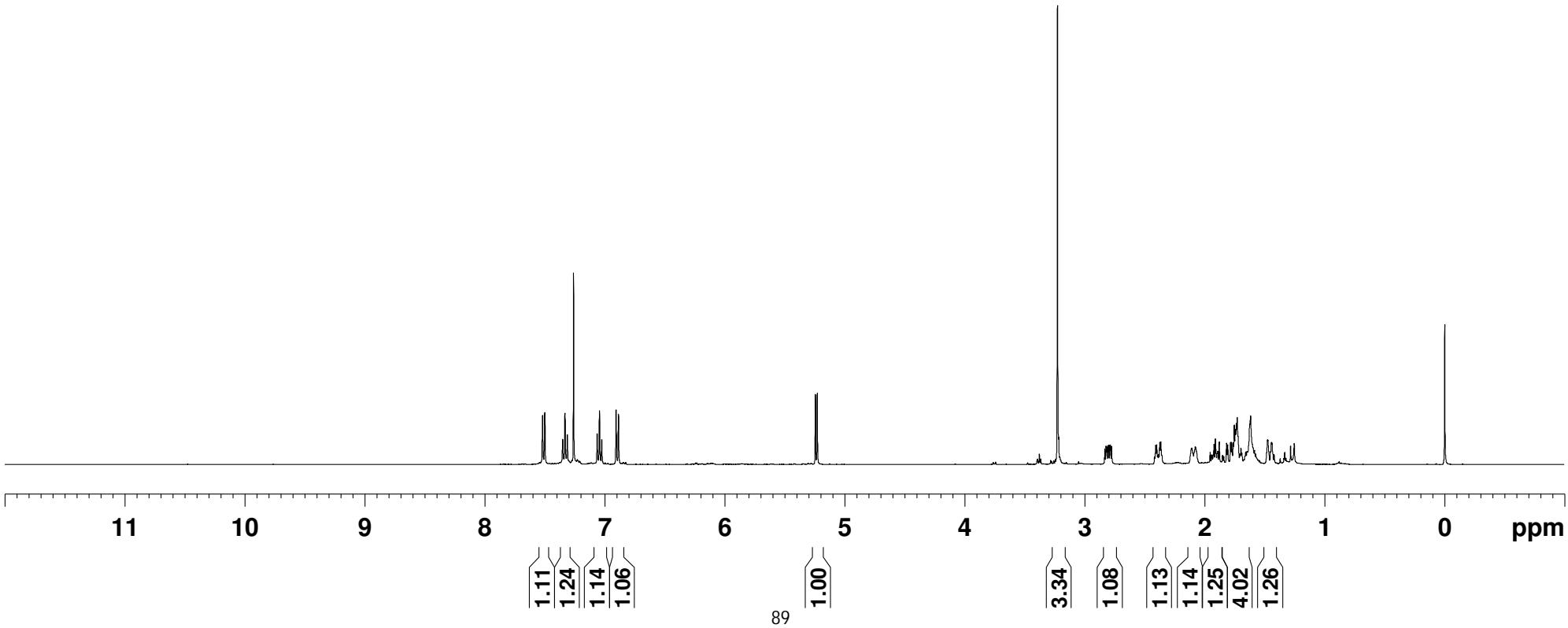
25-up

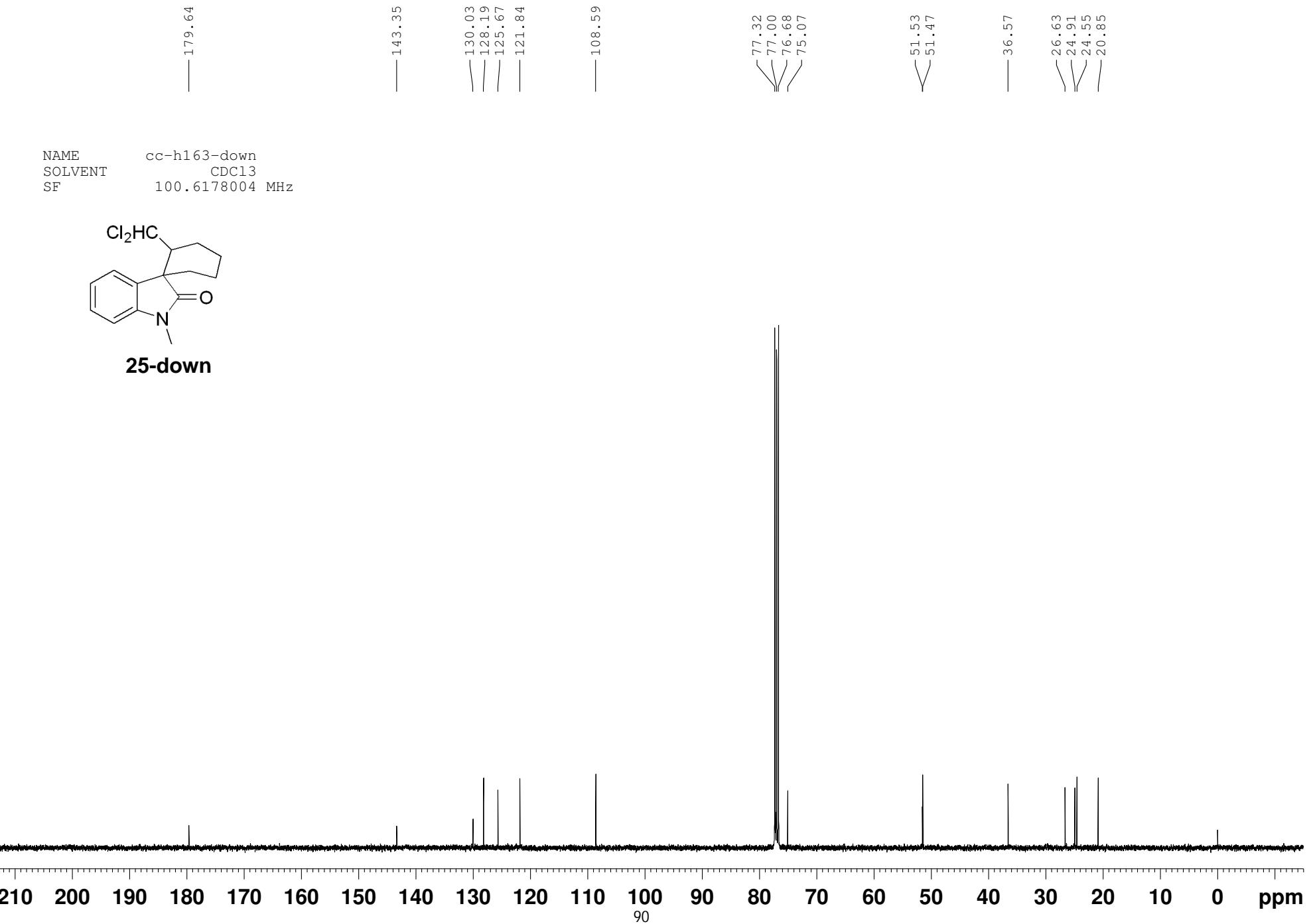


NAME cc-h163-down
SOLVENT CDCl₃
SF 400.1500096 MHz

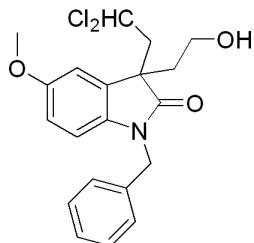


25-down

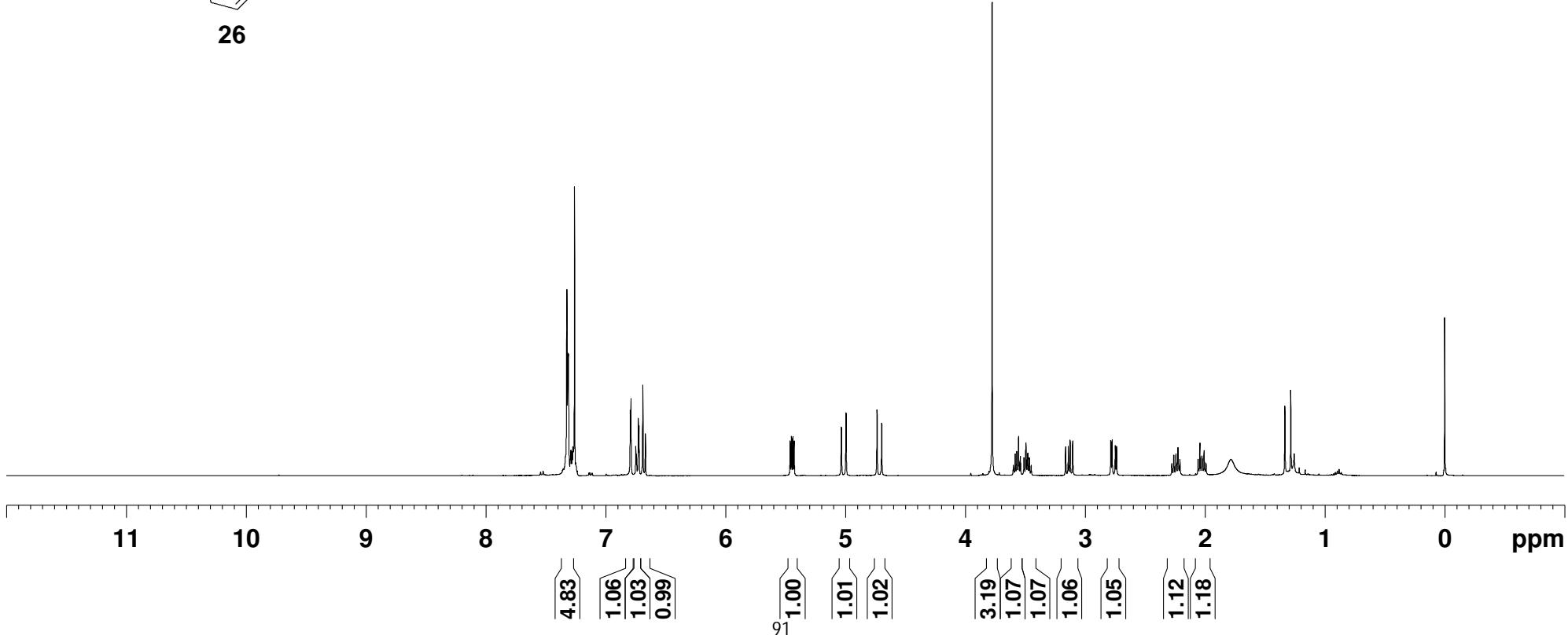




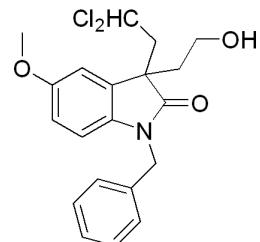
NAME CC-I120
SOLVENT CDC13
SF 400.1500095 MHz



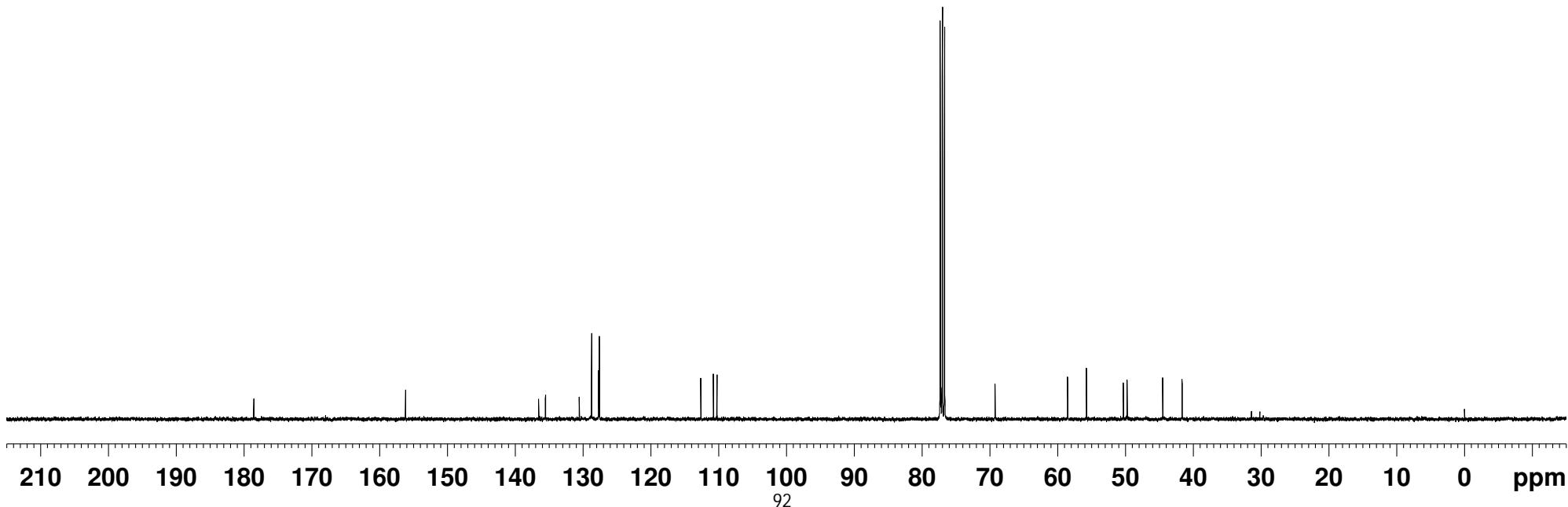
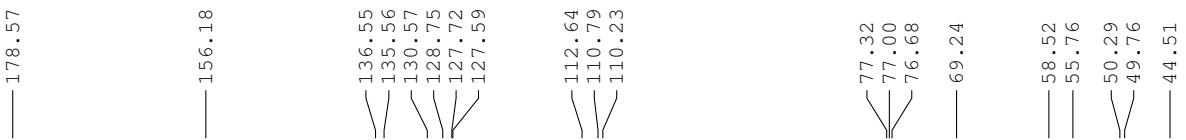
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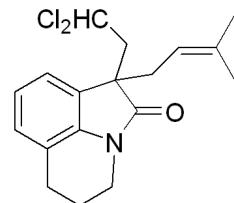
NAME CC-I120
SOLVENT CDCl₃
SF 100.6178001 MHz



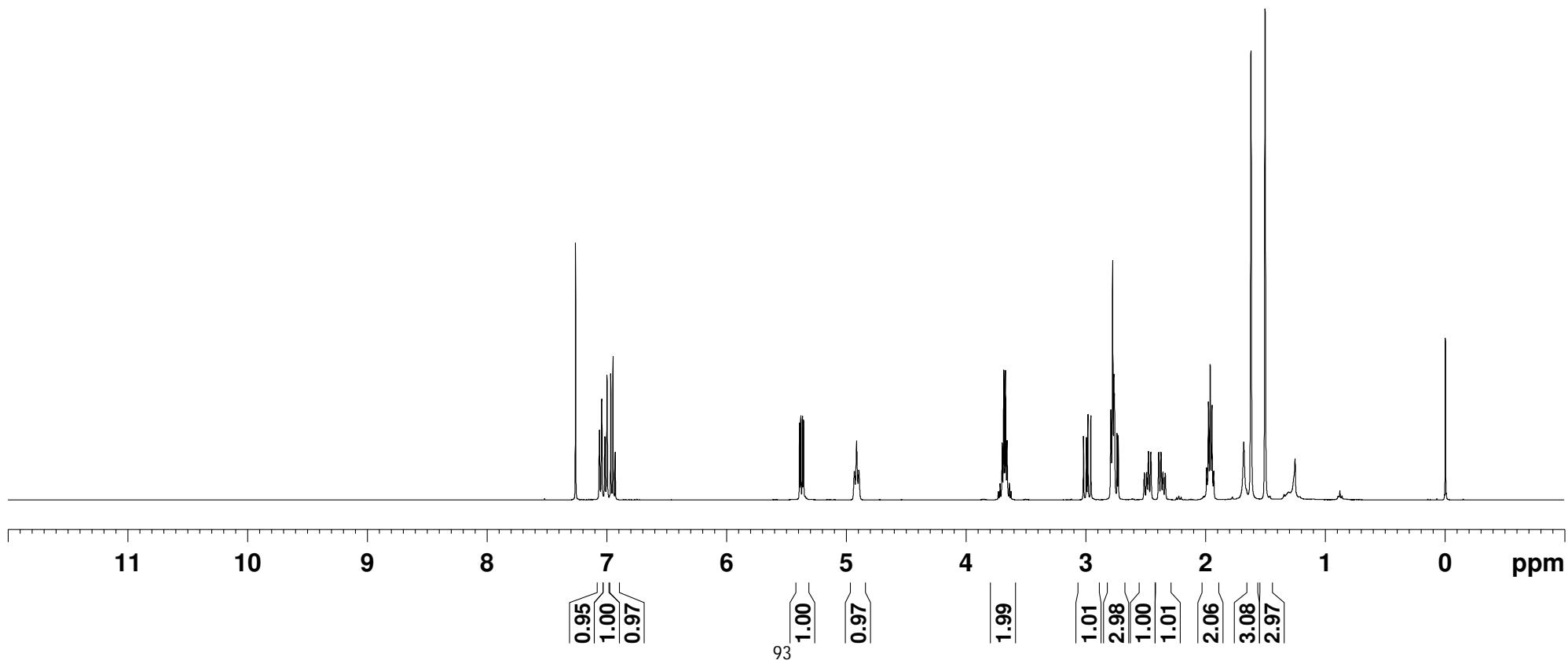
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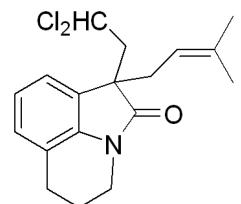
NAME cc-I27-0709
SOLVENT CDC13
SF 400.1500091 MHz



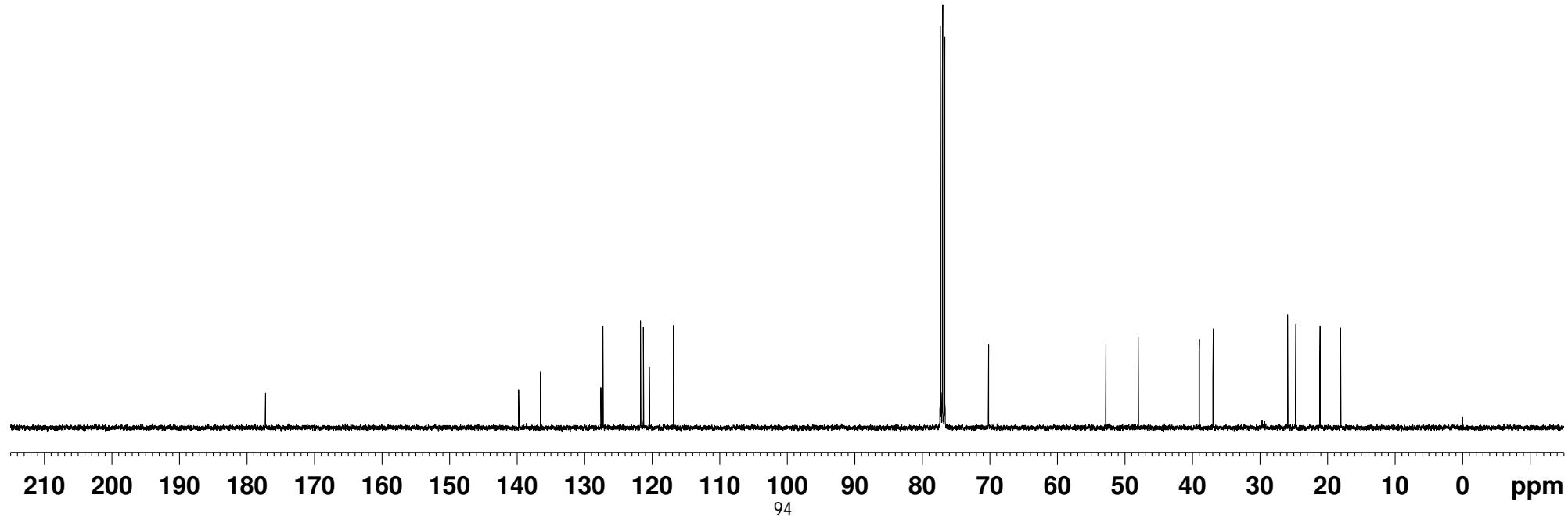
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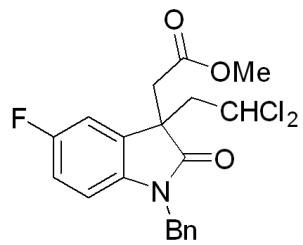
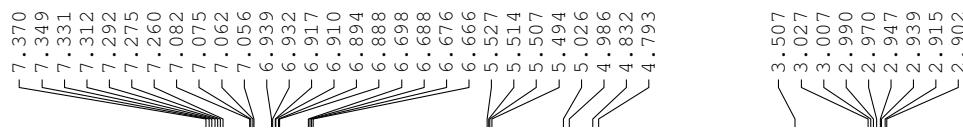
NAME cc-I27-0709
SOLVENT CDCl₃
SF 100.6178008 MHz



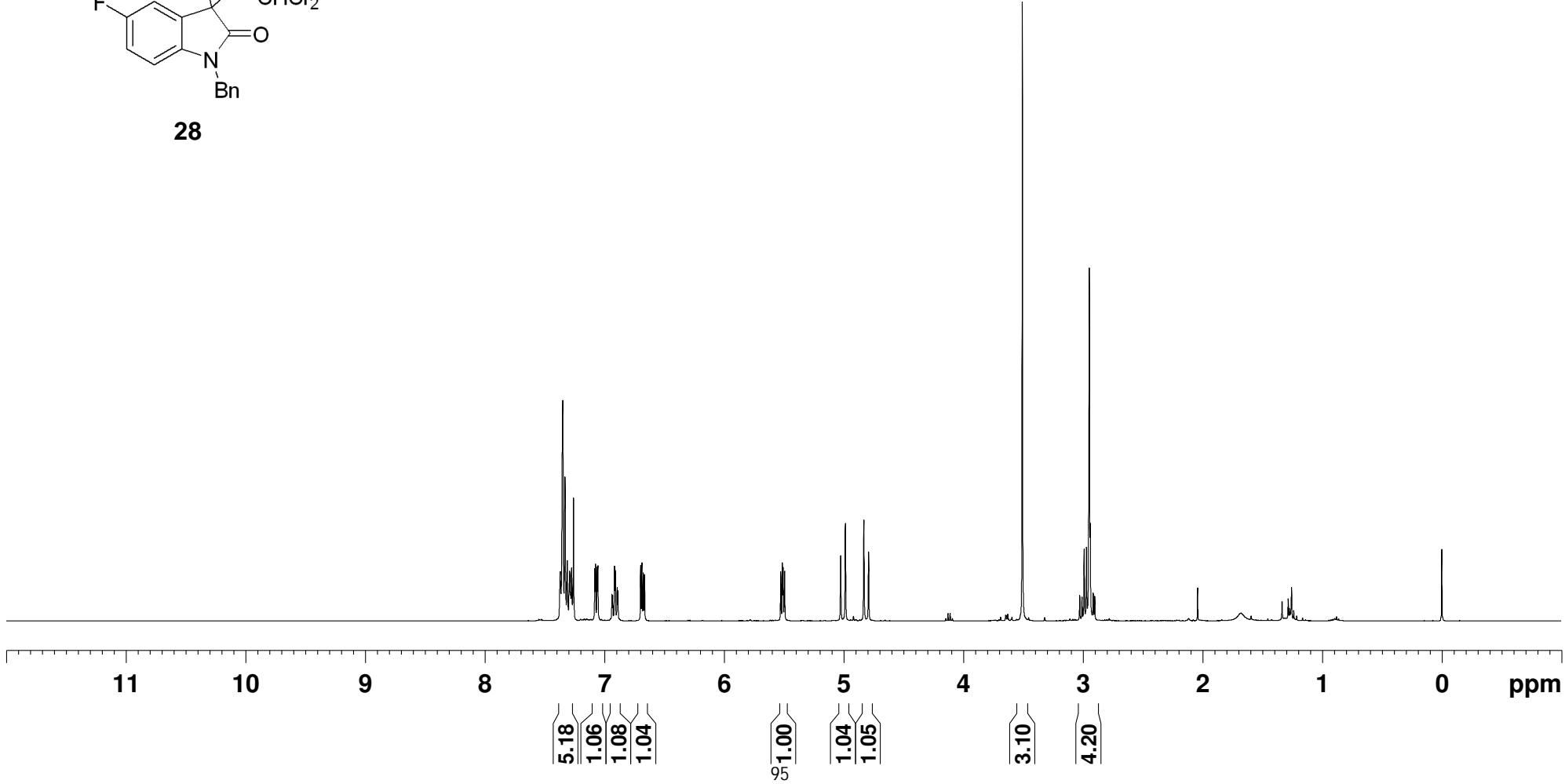
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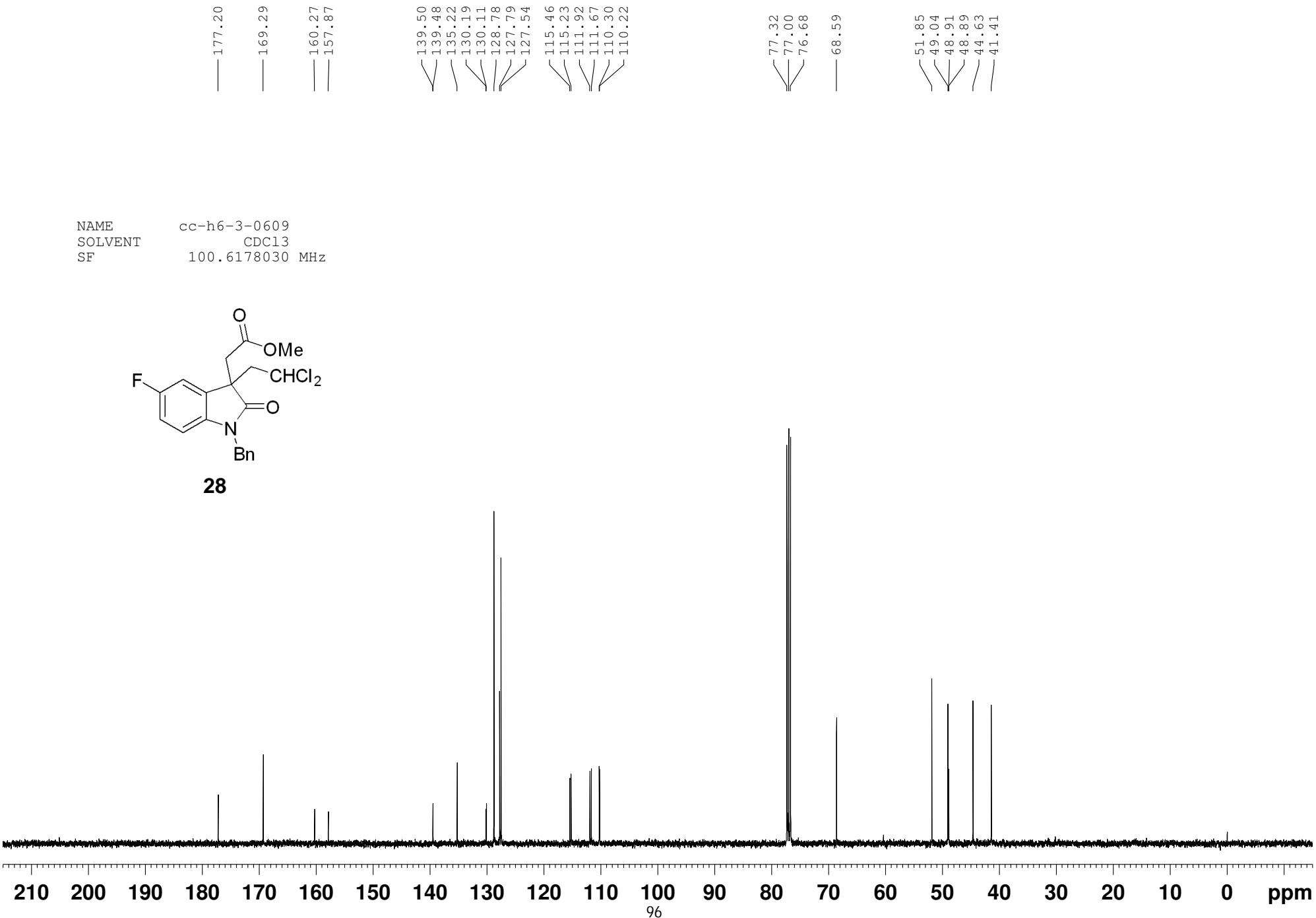


NAME cc-h6-3-0608
SOLVENT CDCl₃
SF 400.1500098 MHz

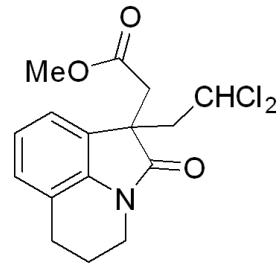


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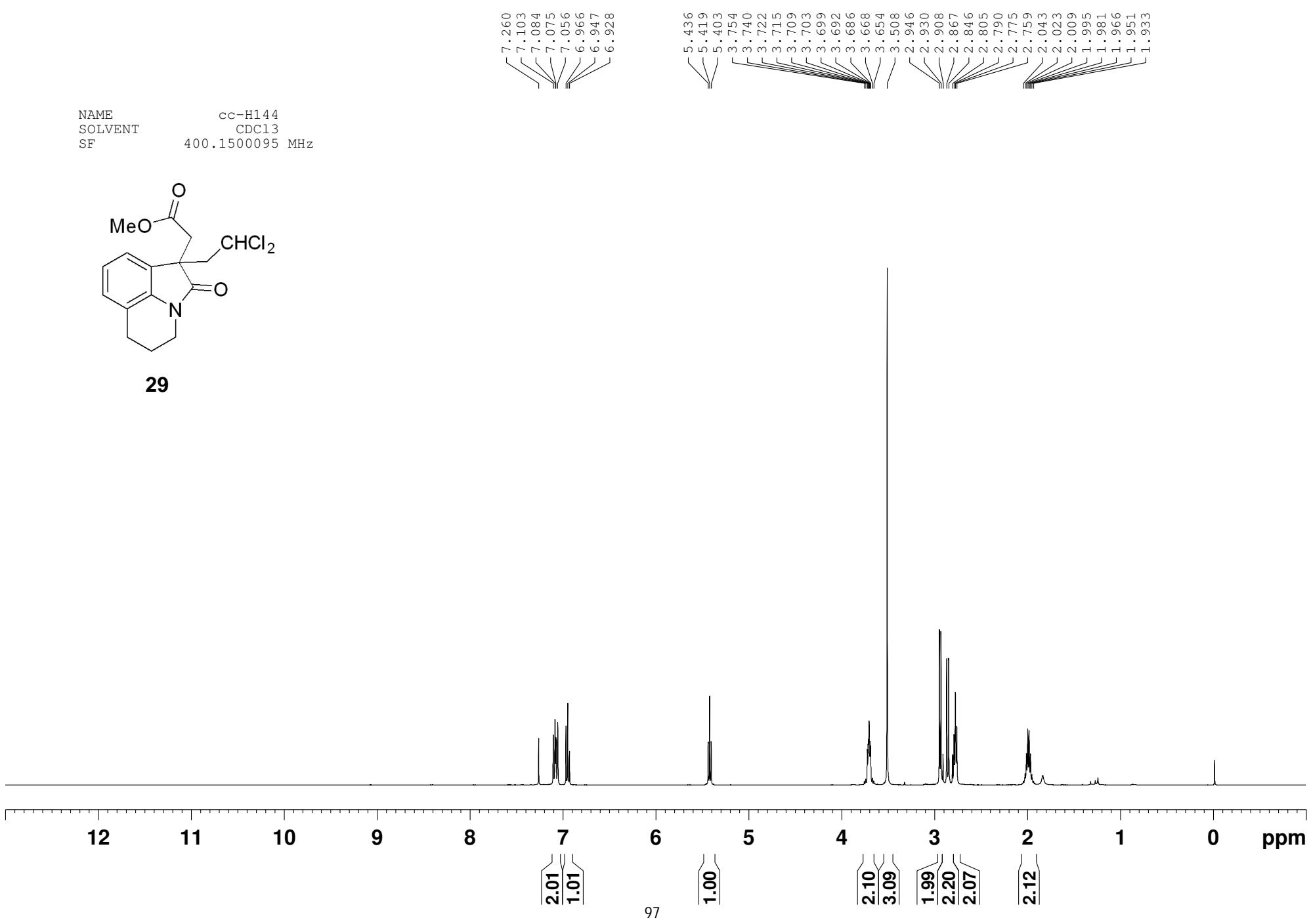




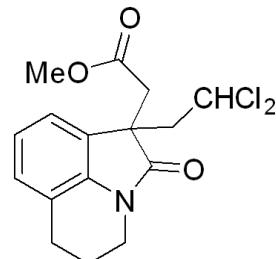
NAME cc-H144
SOLVENT CDCl₃
SF 400.1500095 MHz



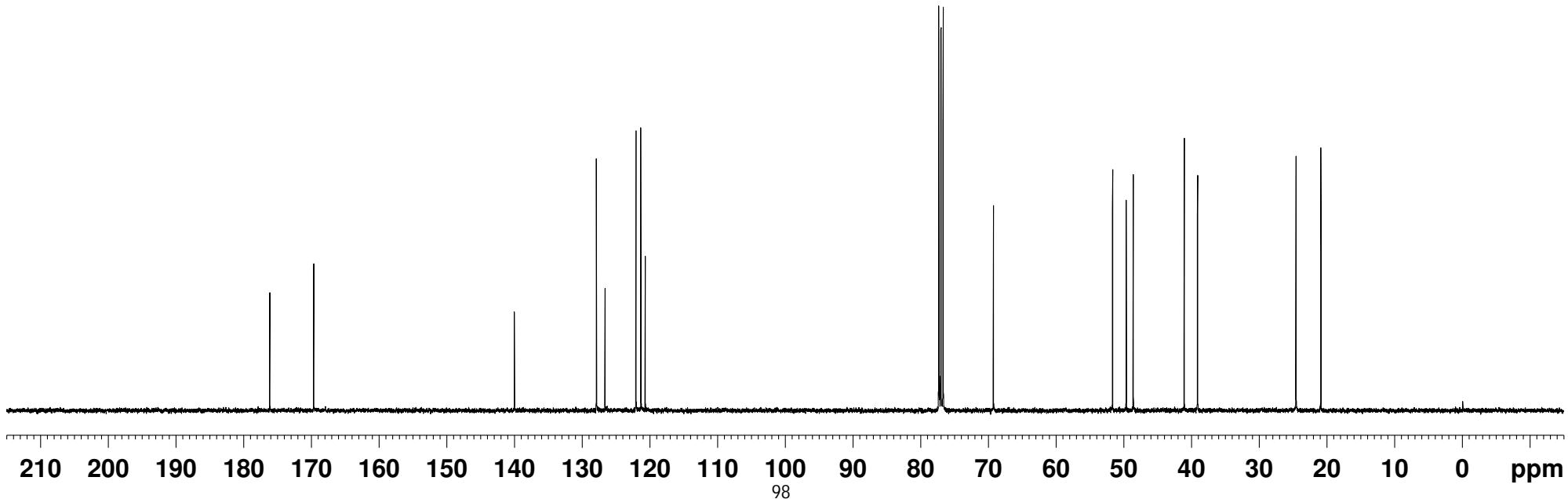
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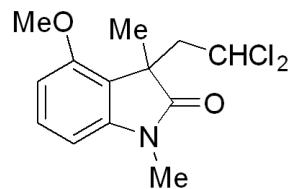
NAME cc-H144
SOLVENT CDCl₃
SF 100.6178063 MHz



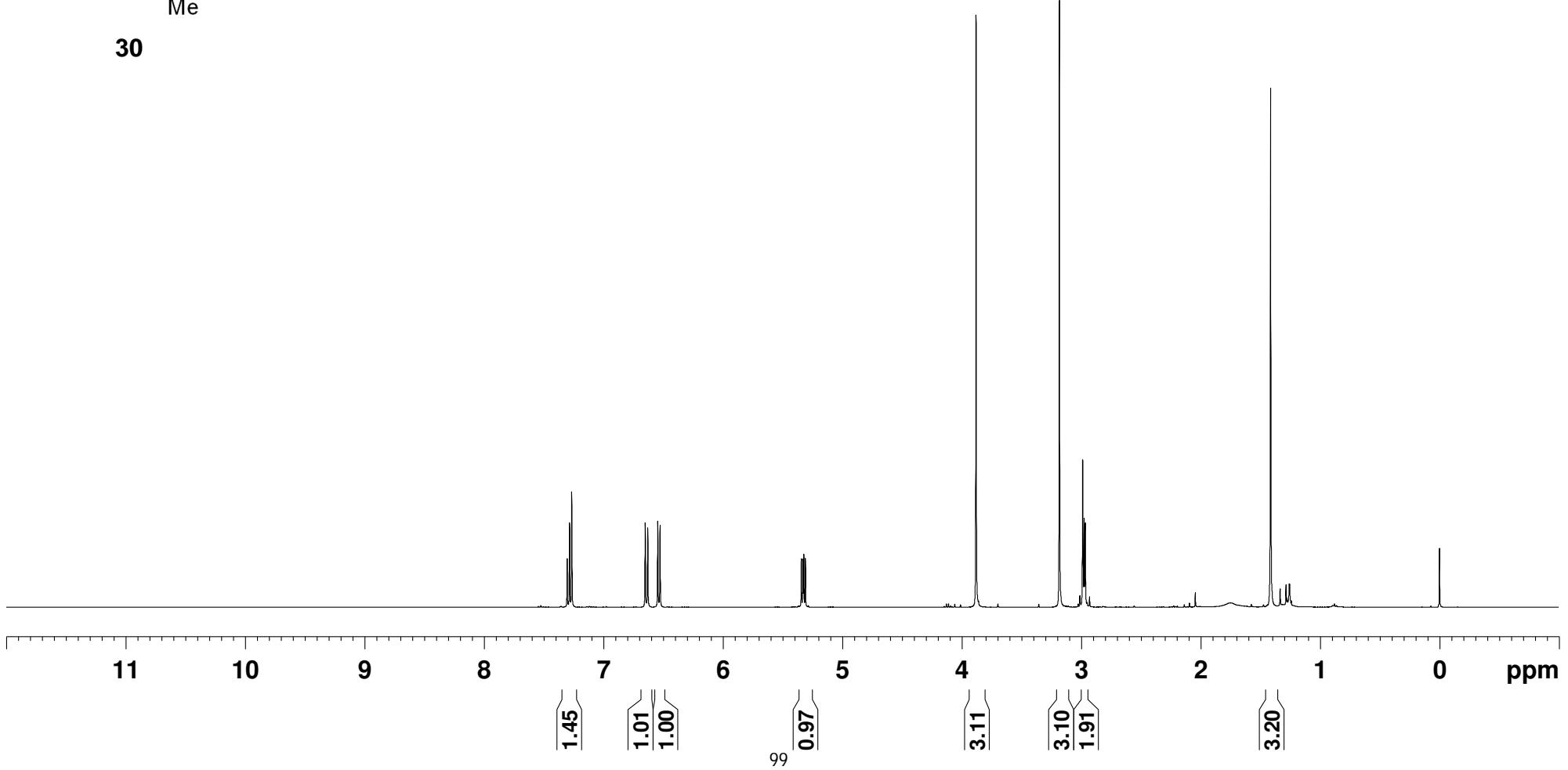
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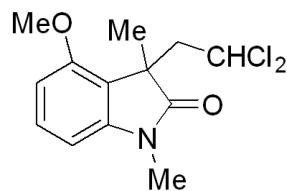
NAME cc-g161-1-up-0710
SOLVENT CDCl₃
SF 400.1500061 MHz



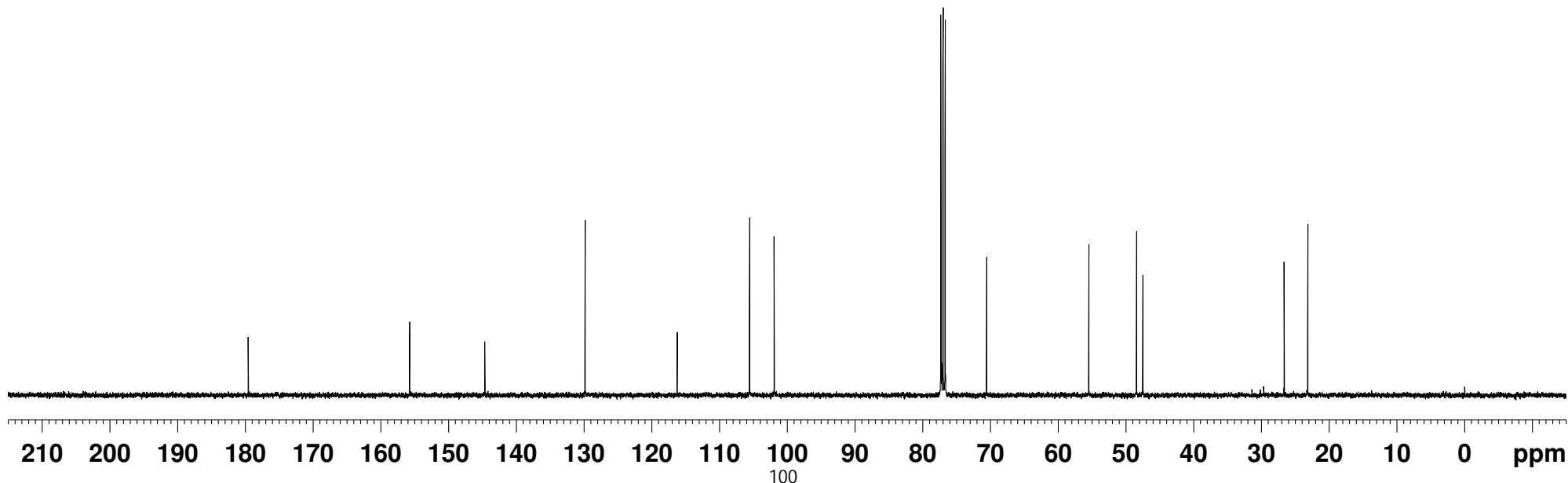
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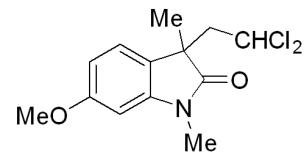
NAME cc-g161-up
SOLVENT CDCl₃
SF 100.6178023 MHz



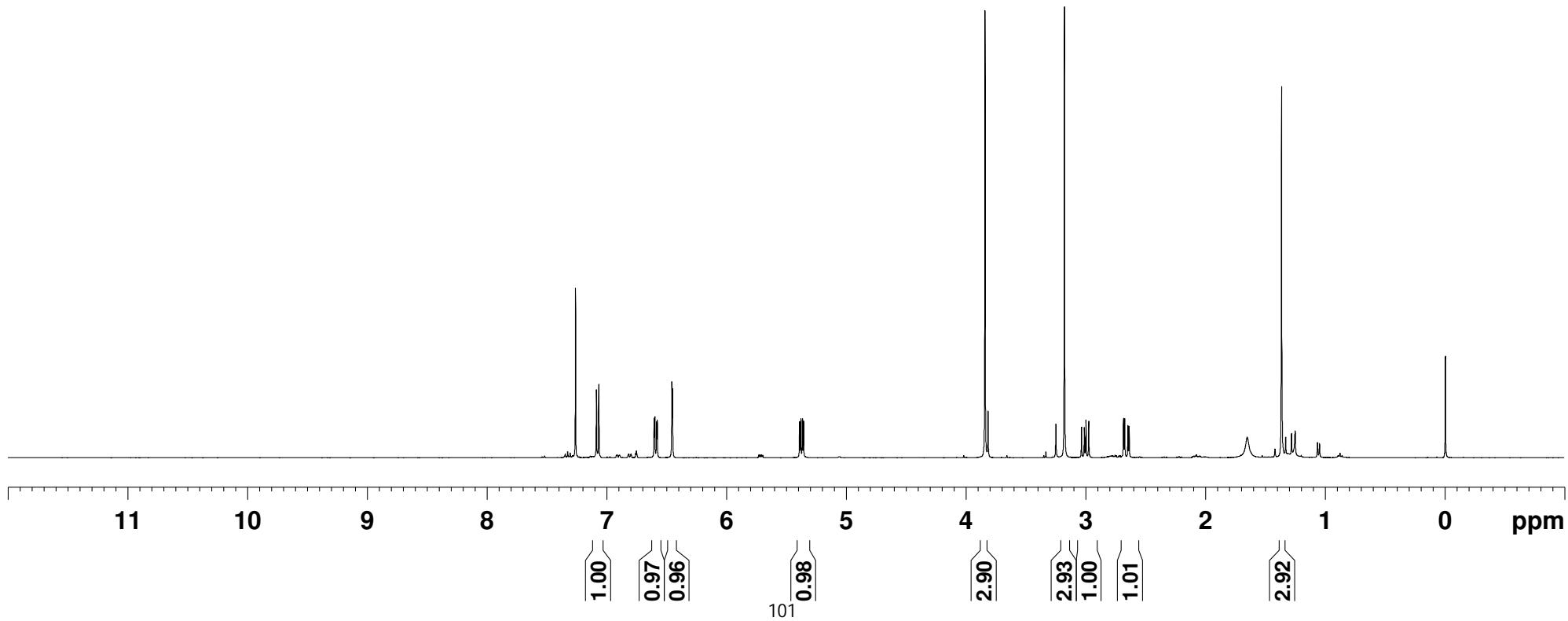
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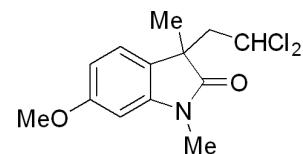
NAME cc-g161-1-down-0710
SOLVENT CDCl₃
SF 400.1500090 MHz



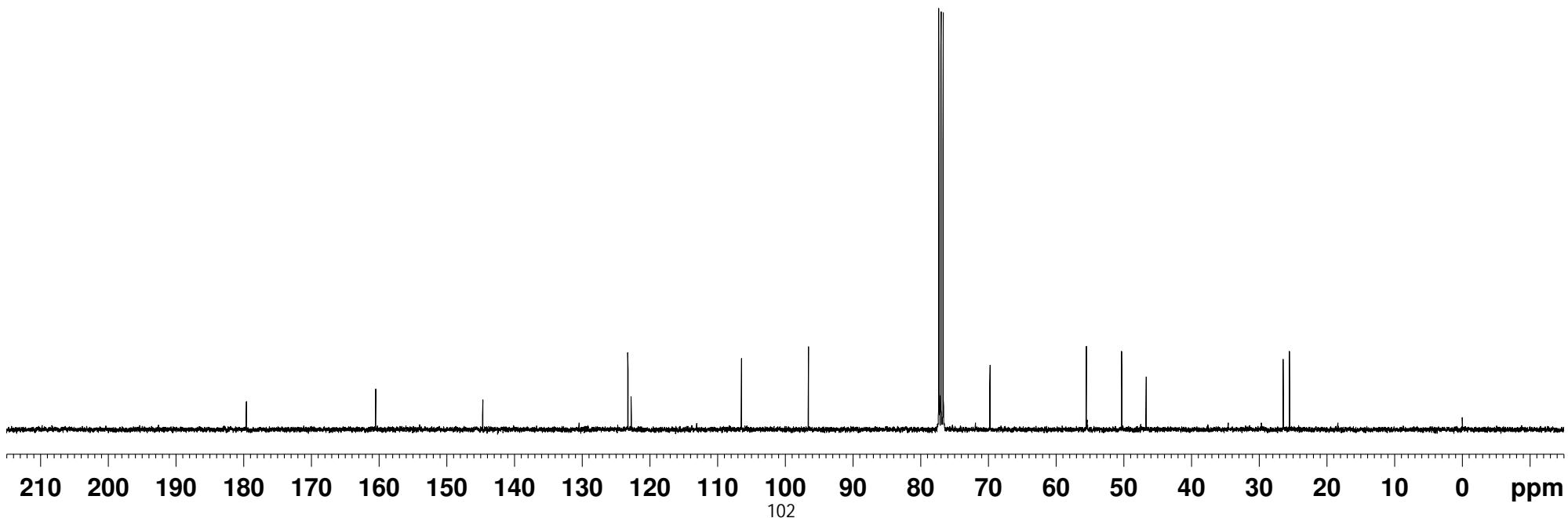
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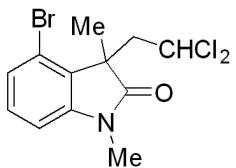
NAME cc-g161-1-down
SOLVENT CDCl₃
SF 100.6178008 MHz



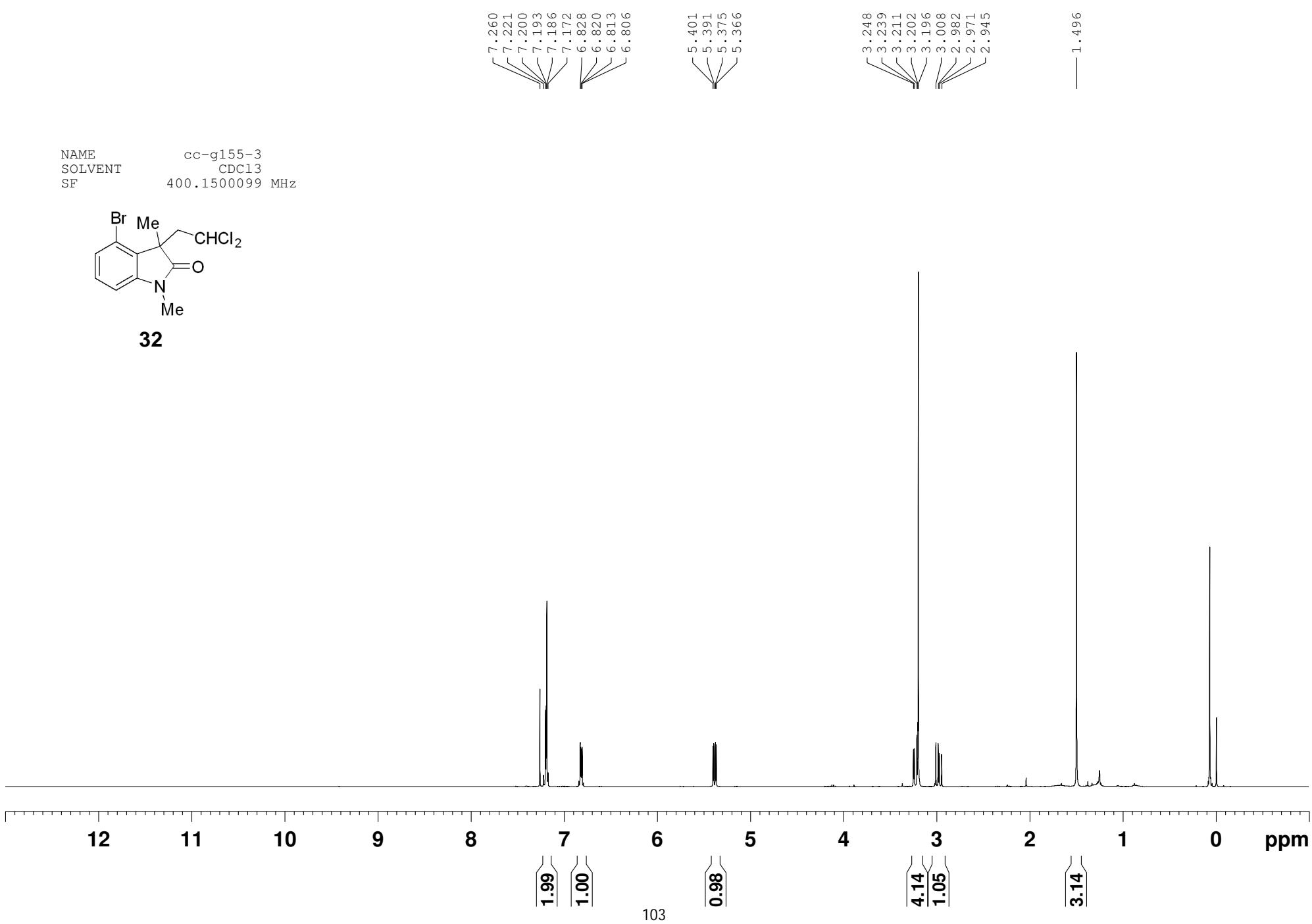
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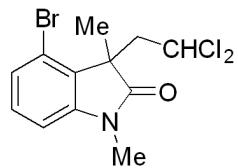
NAME cc-g155-3
SOLVENT CDCl₃
SF 400.1500099 MHz



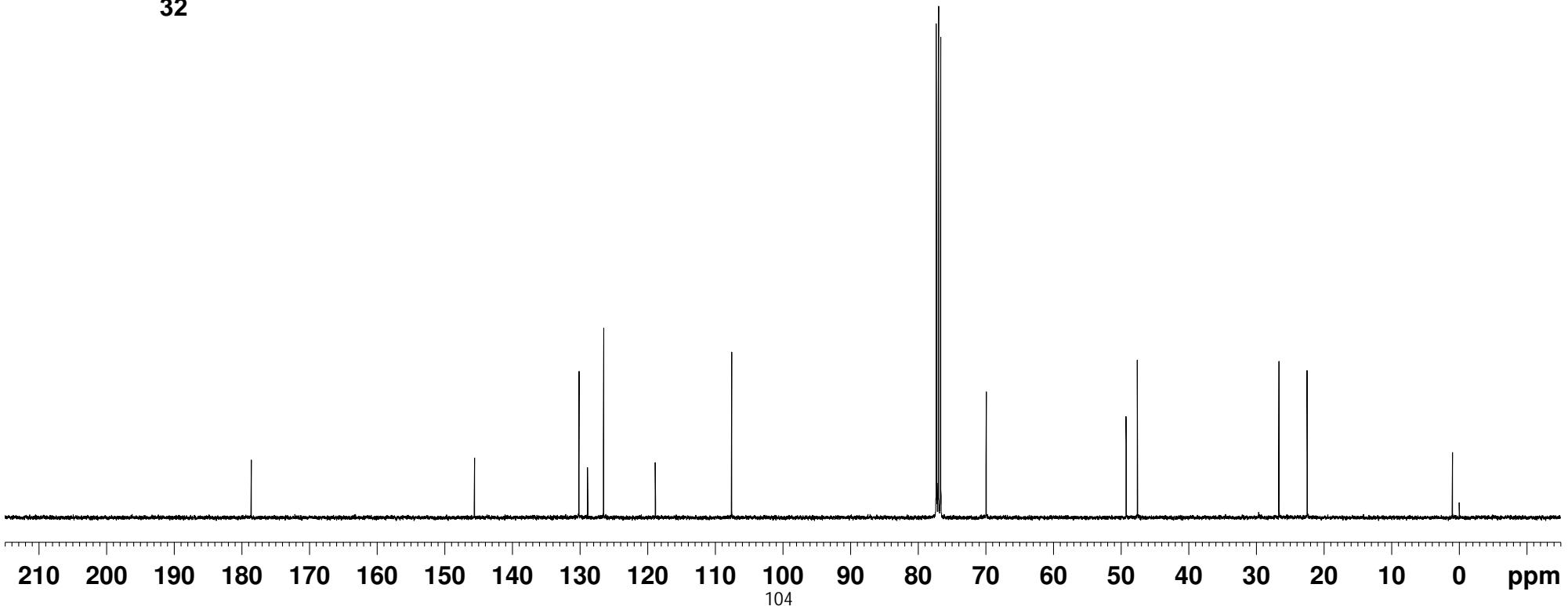
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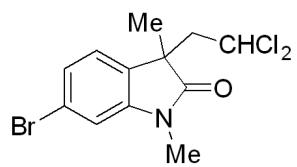
NAME cc-g155-3
SOLVENT CDCl₃
SF 100.6178016 MHz



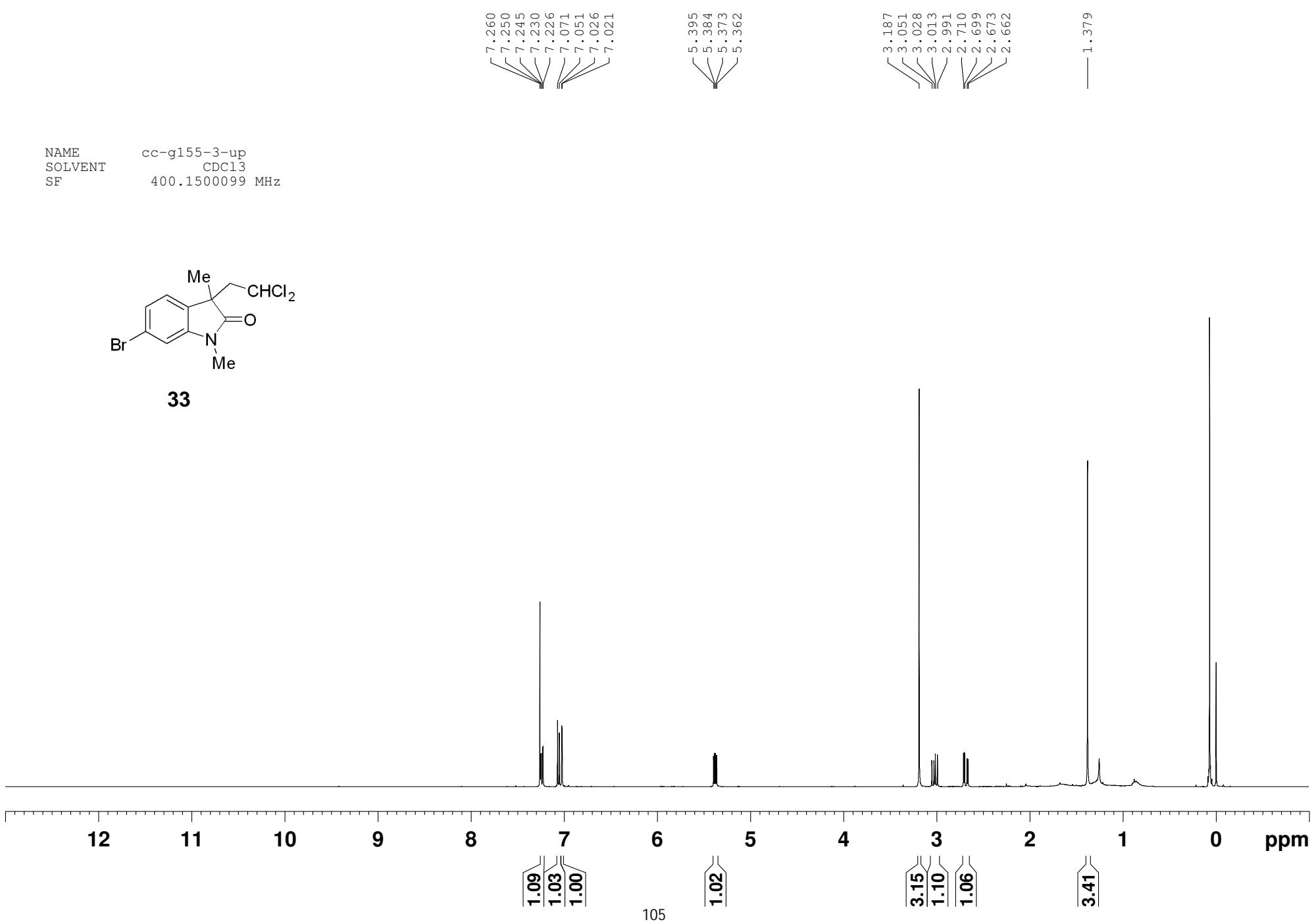
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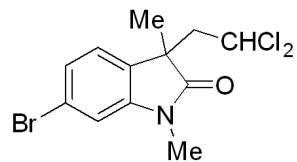
NAME cc-g155-3-up
SOLVENT CDCl₃
SF 400.1500099 MHz



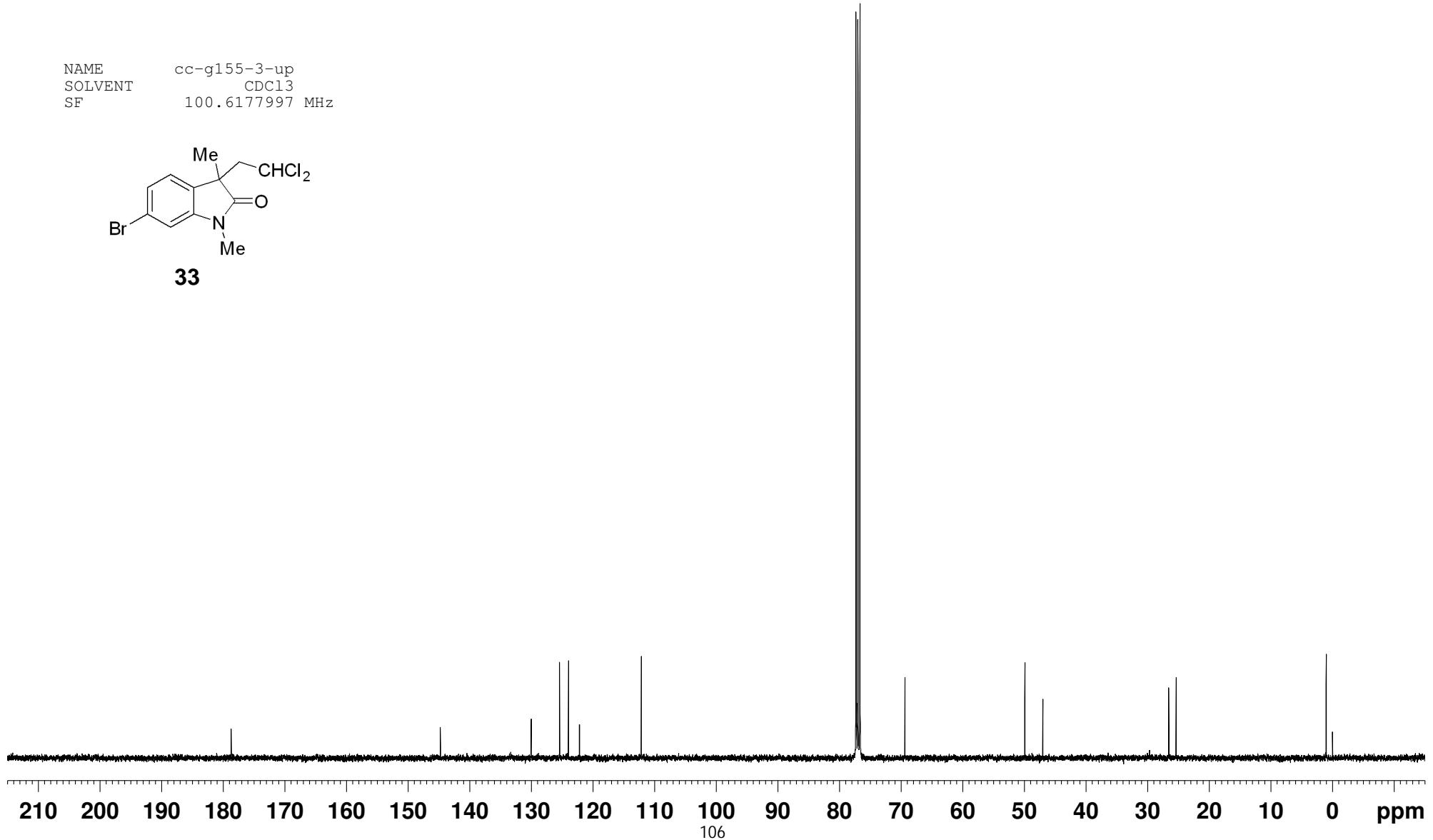
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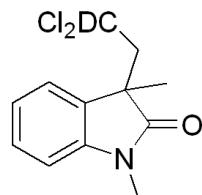
NAME cc-g155-3-up
SOLVENT CDCl₃
SF 100.6177997 MHz



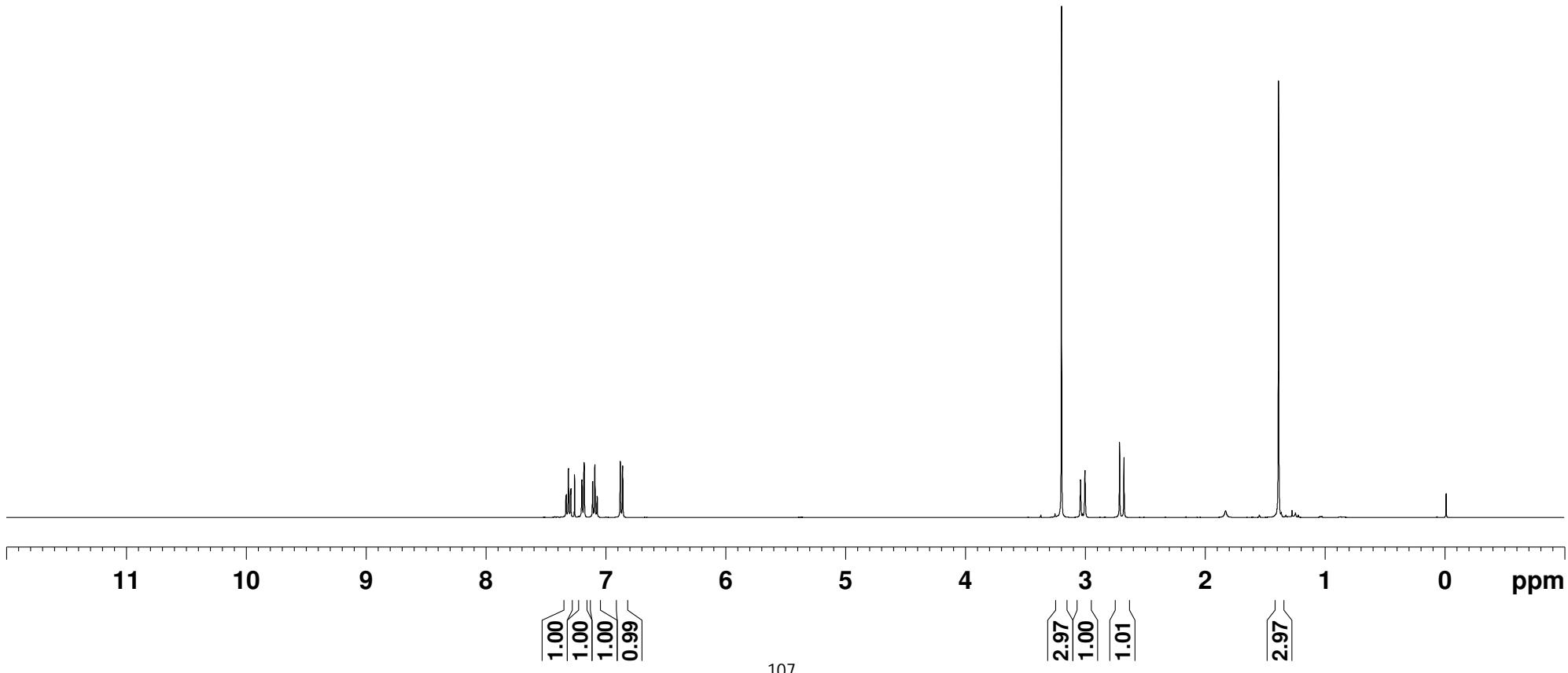
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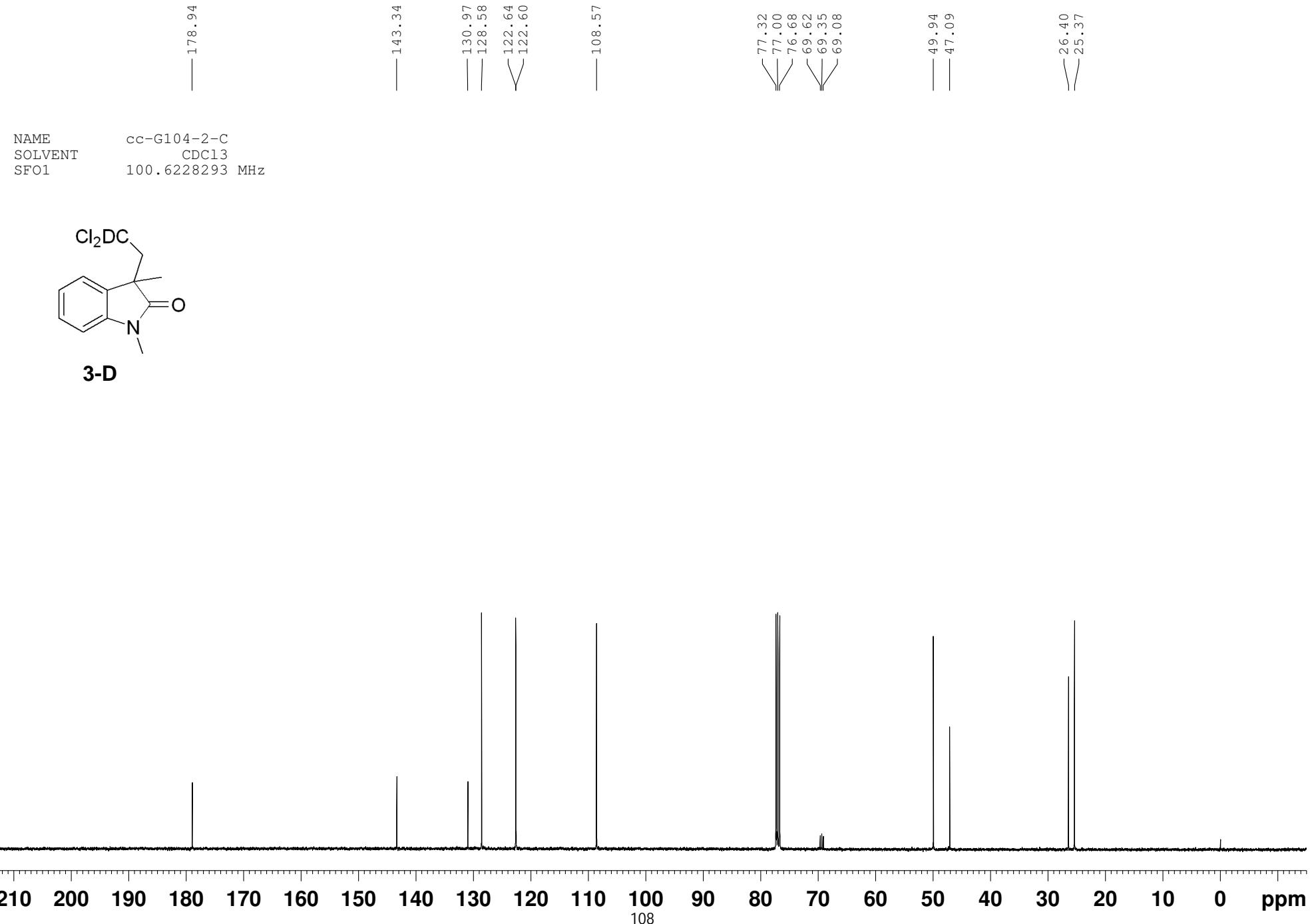


NAME cc-g104-2
SOLVENT CDCl₃
SF 400.1500096 MHz

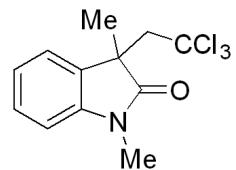


3-D

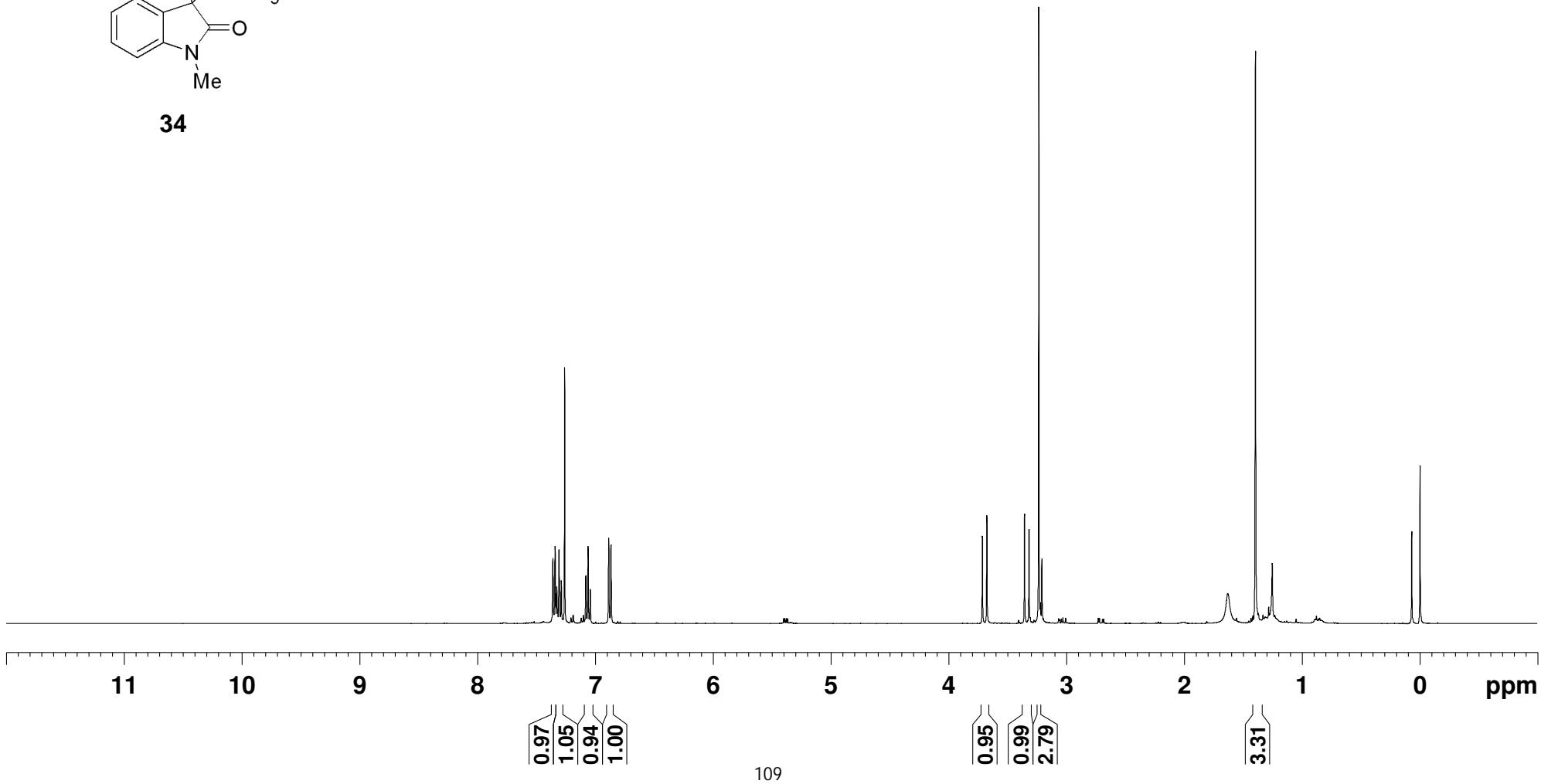




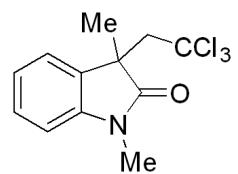
NAME cc-i94-2
SOLVENT CDCl₃
SF 400.1500096 MHz



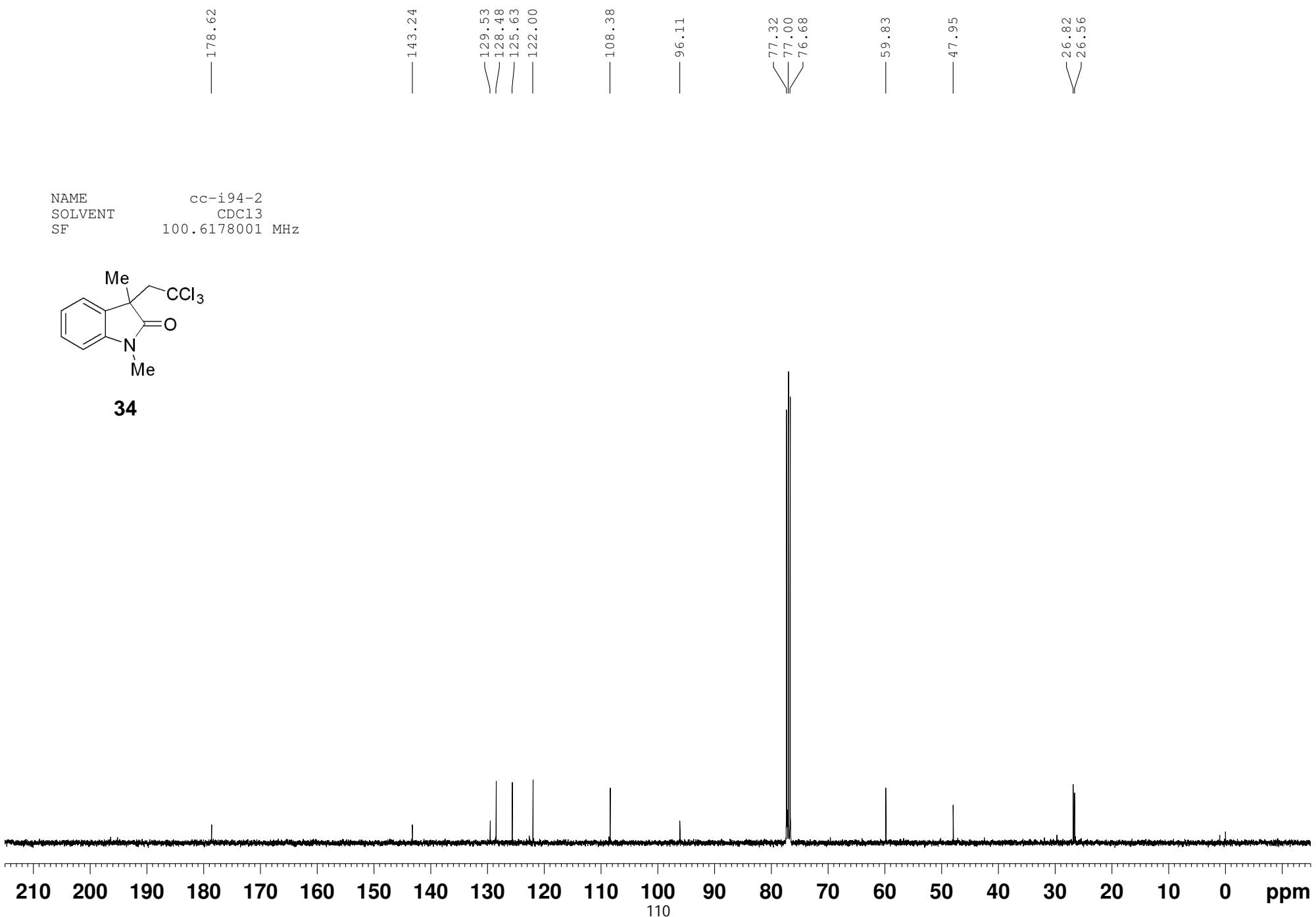
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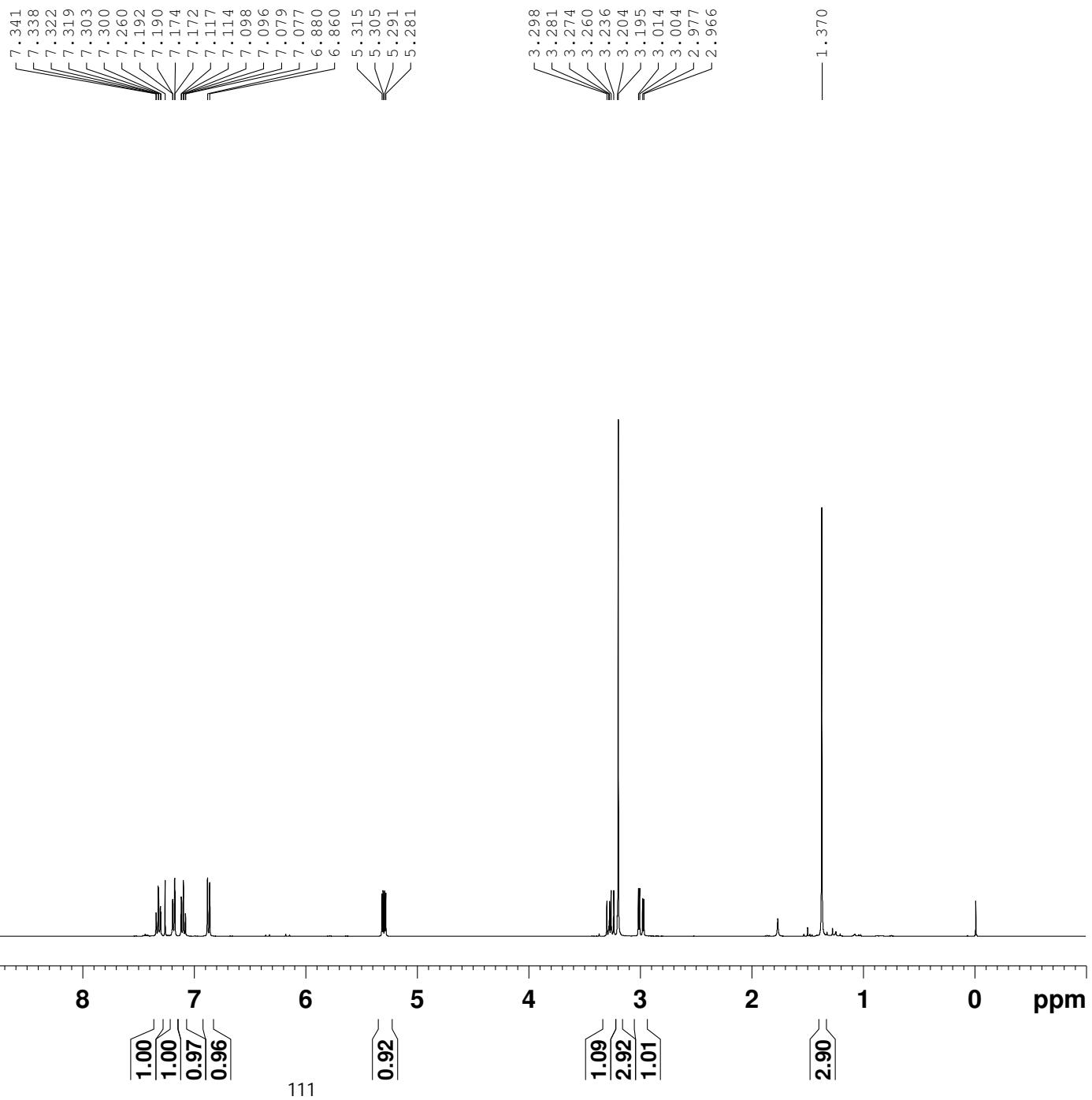
NAME cc-i94-2
SOLVENT CDCl₃
SF 100.6178001 MHz



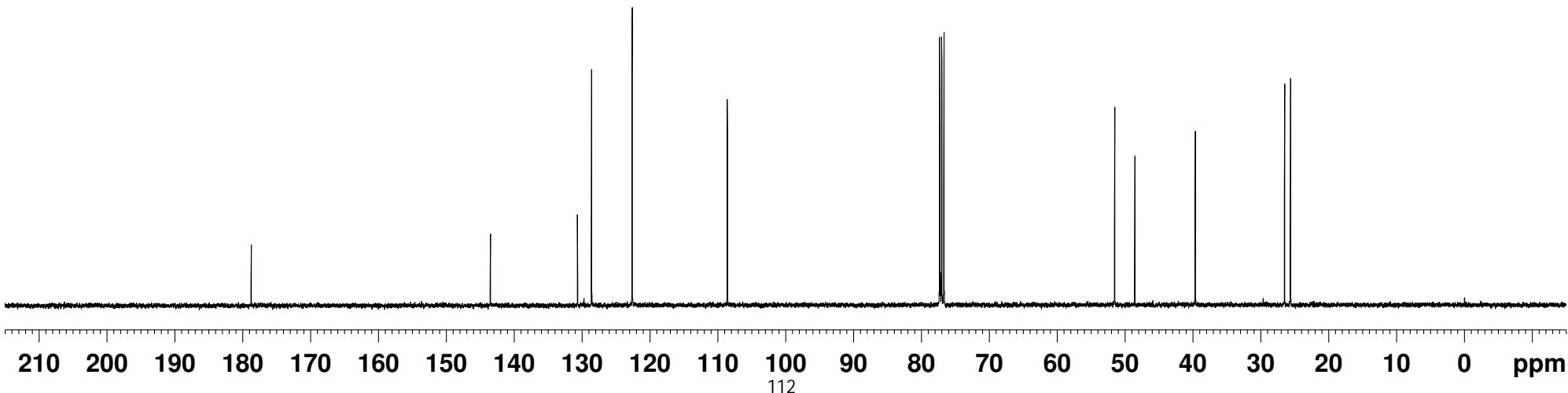
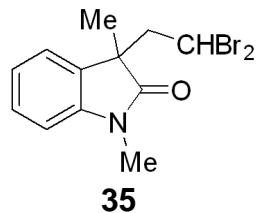
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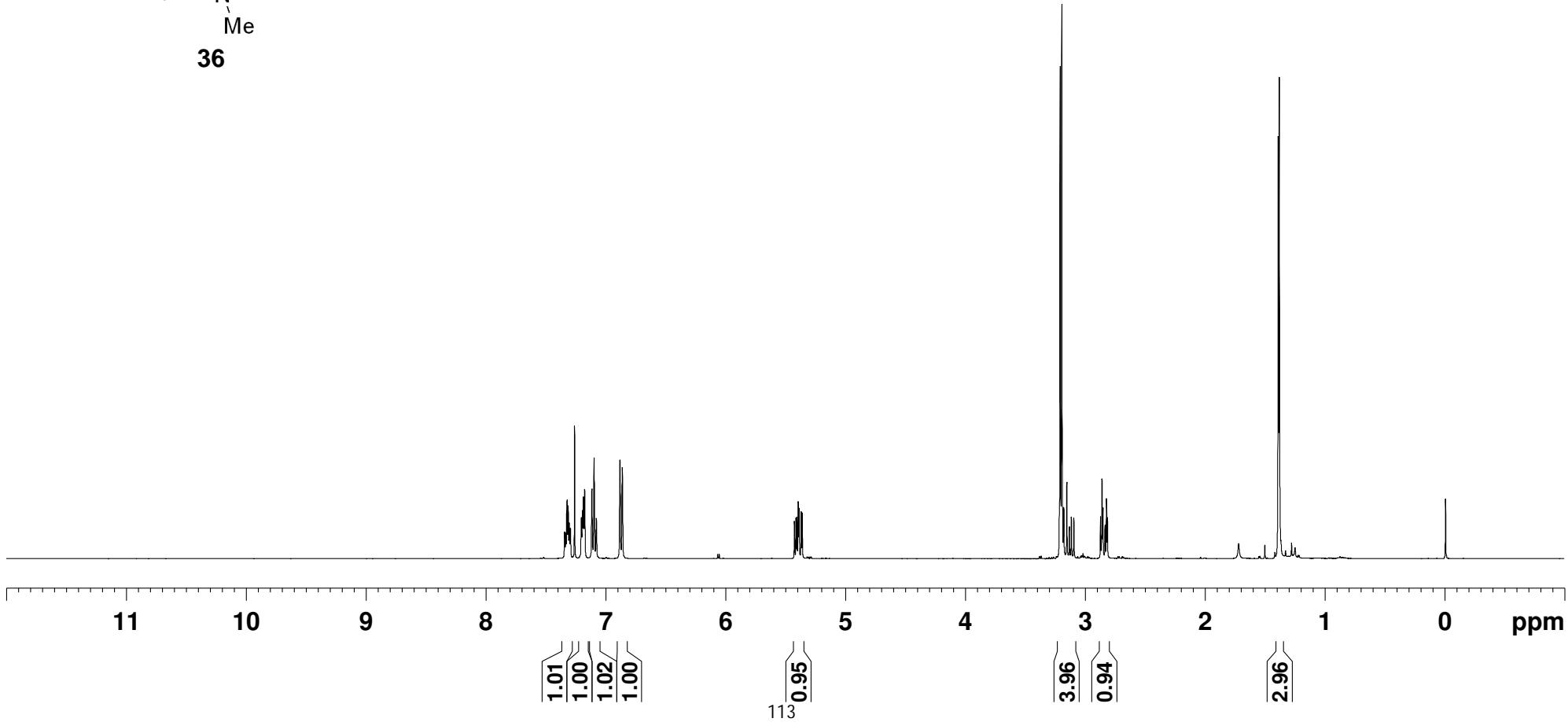
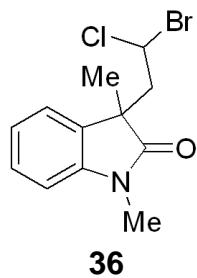
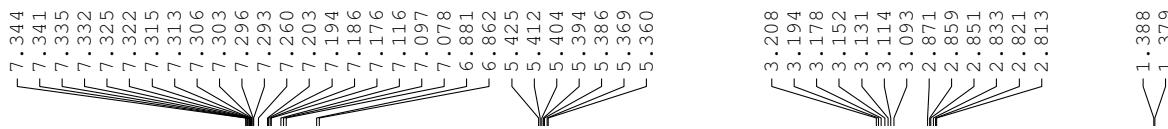
NAME cc-G104-5
SOLVENT CDCl₃
SF 400.1500096 MHz

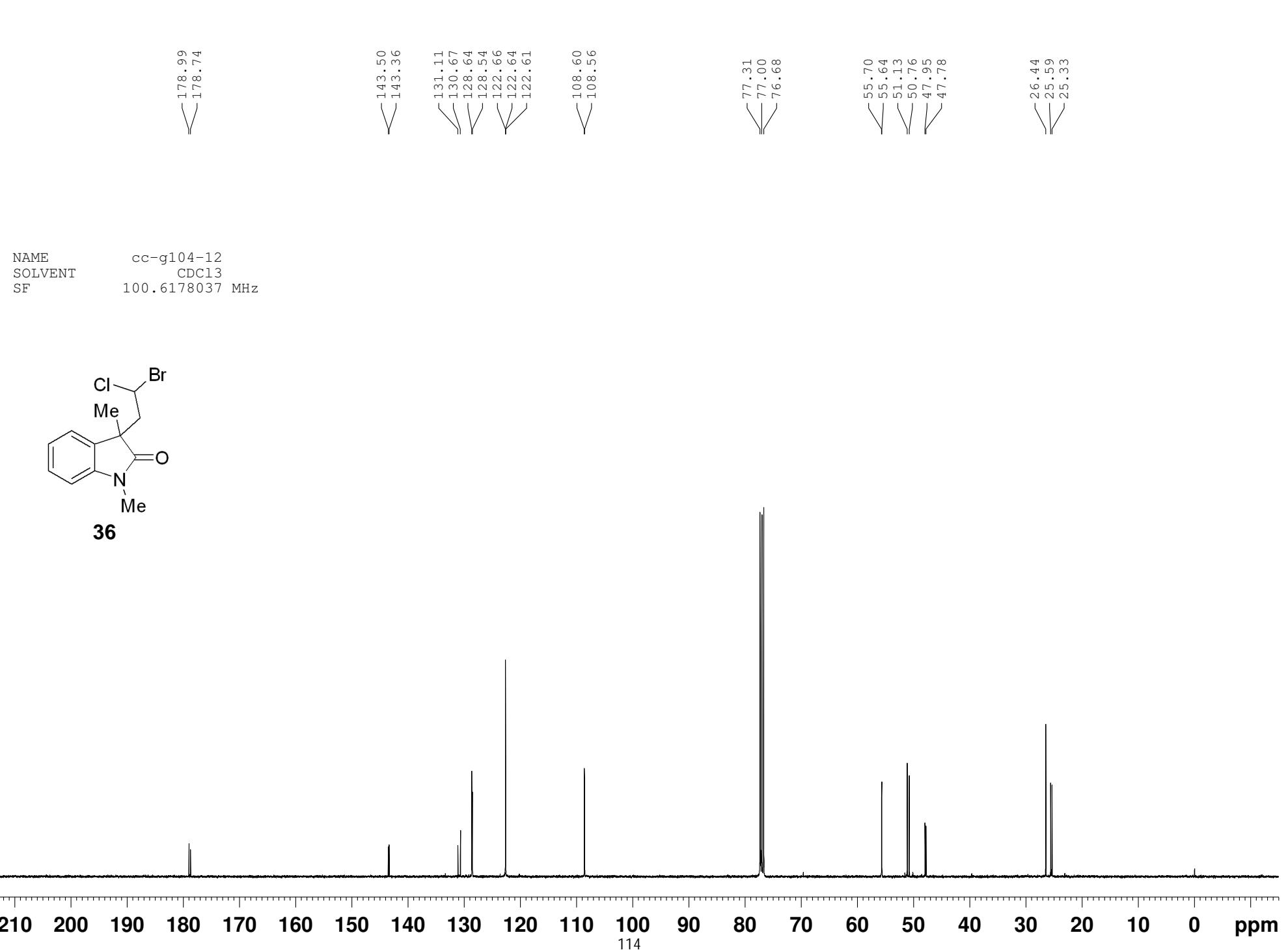


NAME cc-I94-1-C
SOLVENT CDCl₃
SF 100.6127761 MHz

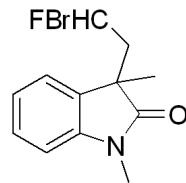


NAME cc-q104-12
SOLVENT CDCl₃
SFO1 400.1524711 MHz

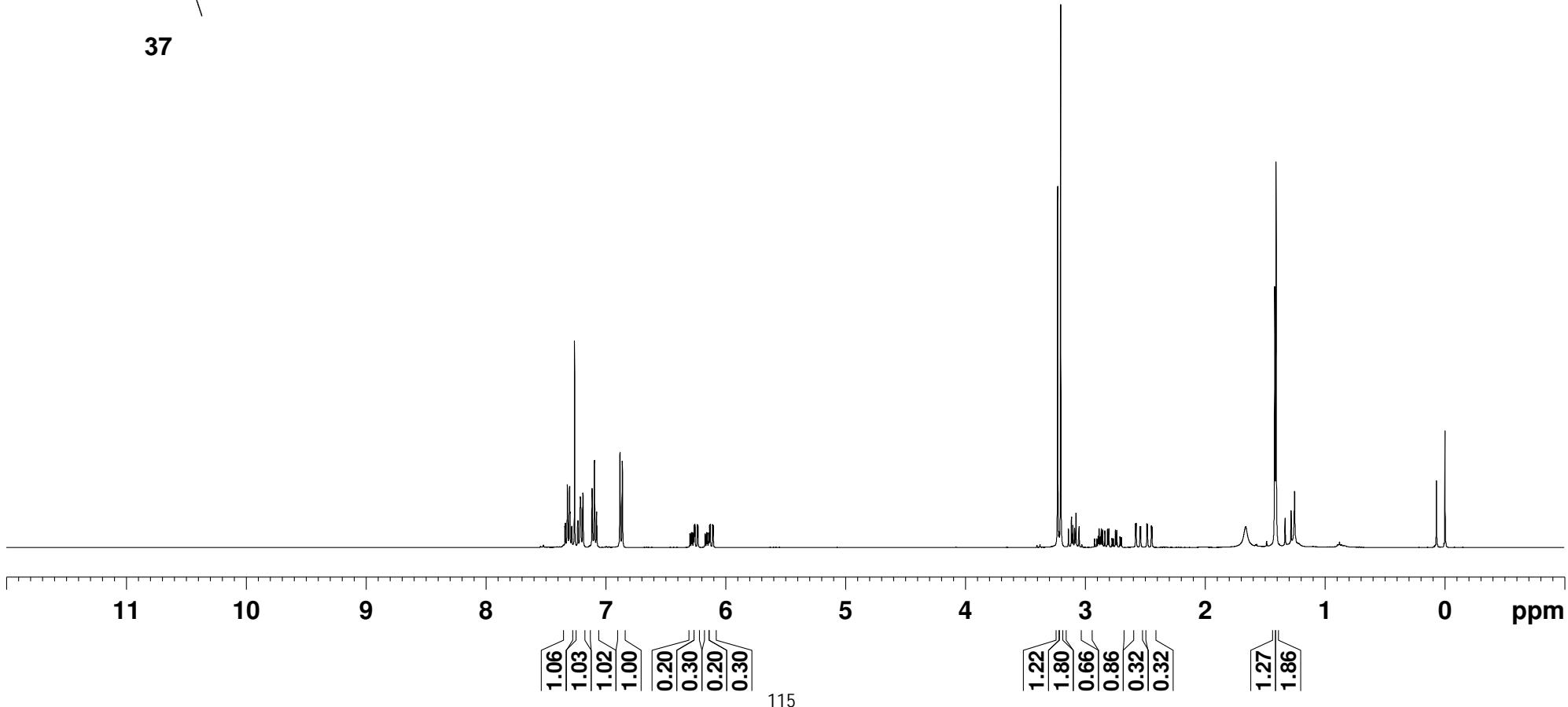




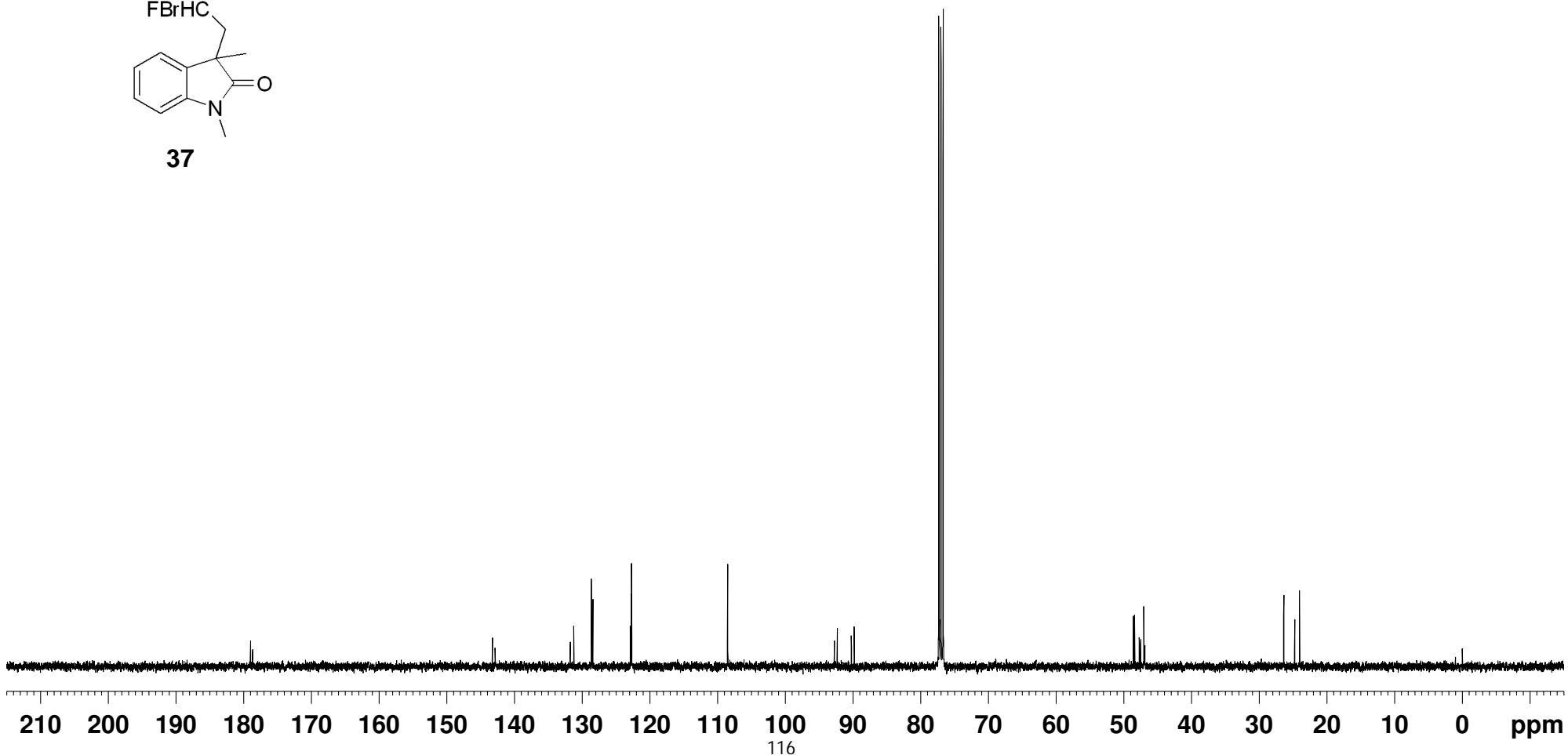
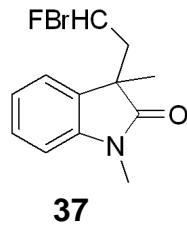
NAME cc-i94-4-H
SOLVENT CDCl₃
SF 400.1500090 MHz



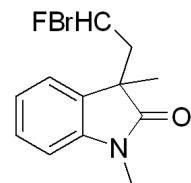
37



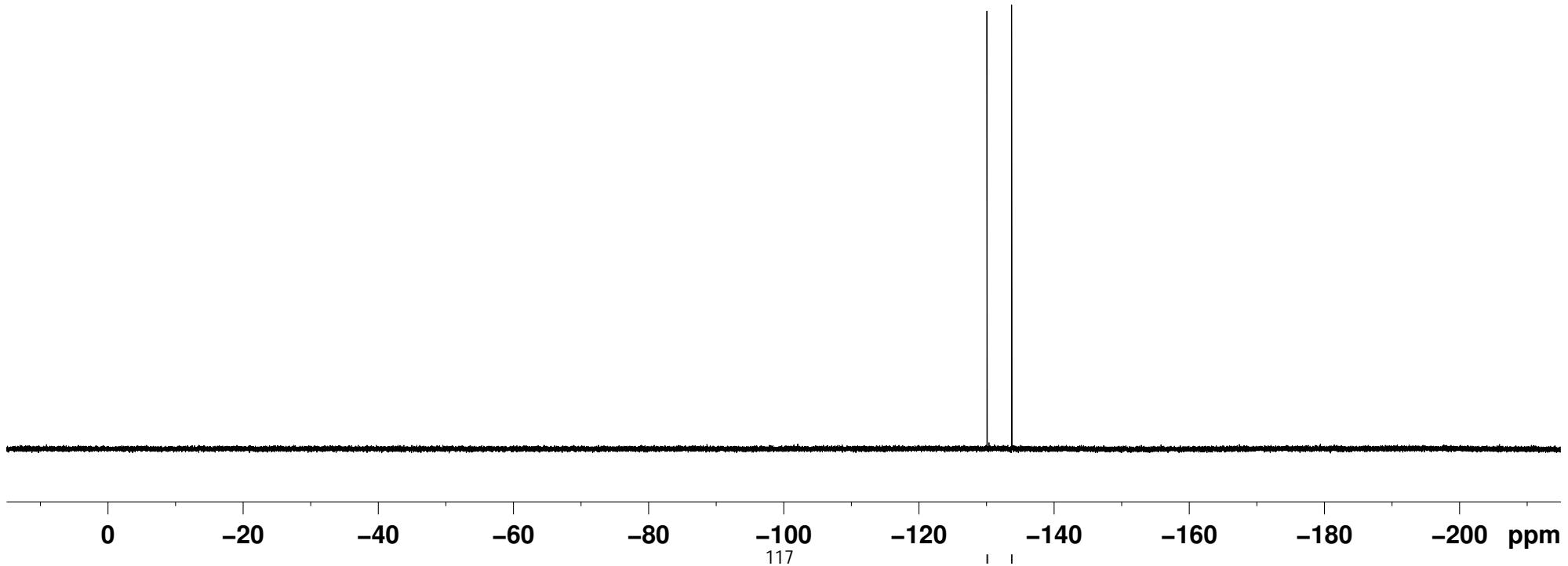
NAME cc-i94-4
SOLVENT CDCl₃
SF 100.6178006 MHz



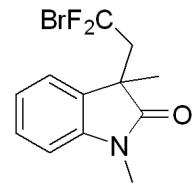
NAME cc-i94-4
SOLVENT CDCl₃
SF 376.5171850 MHz



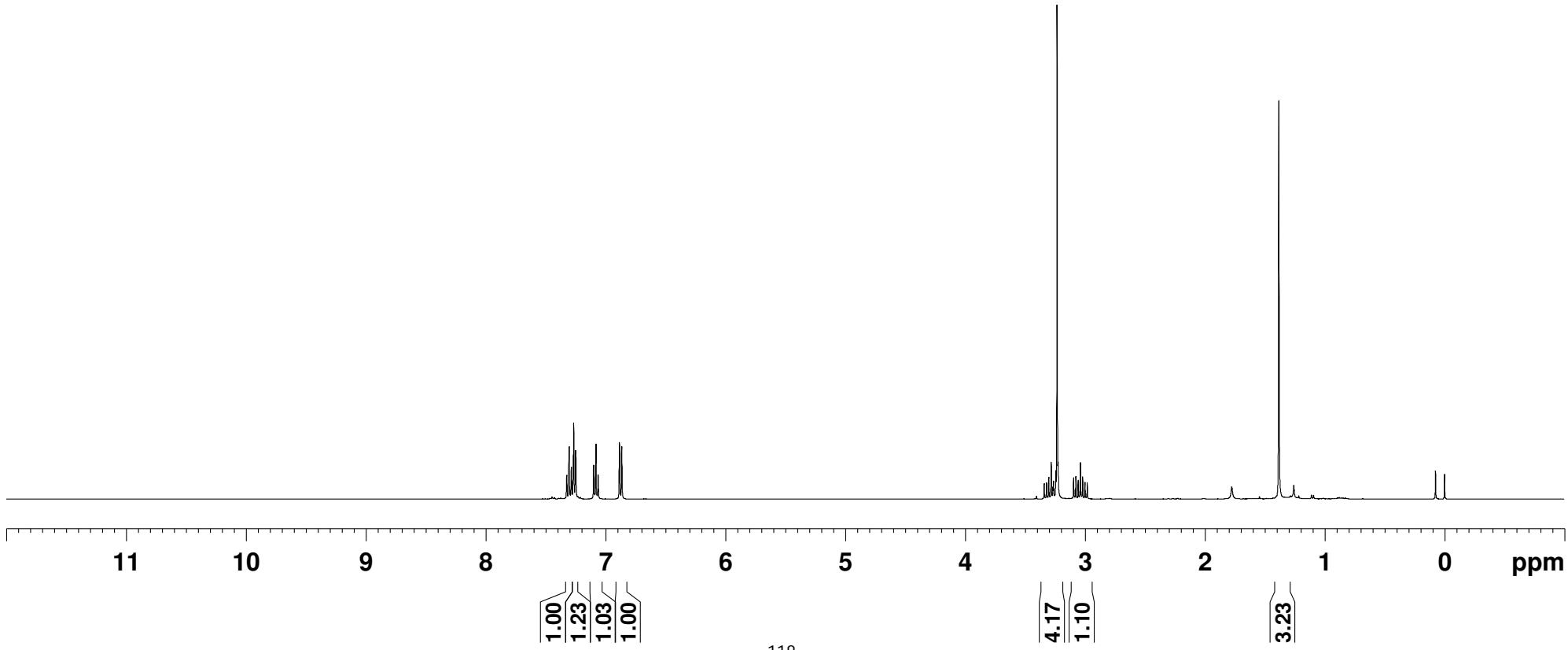
38

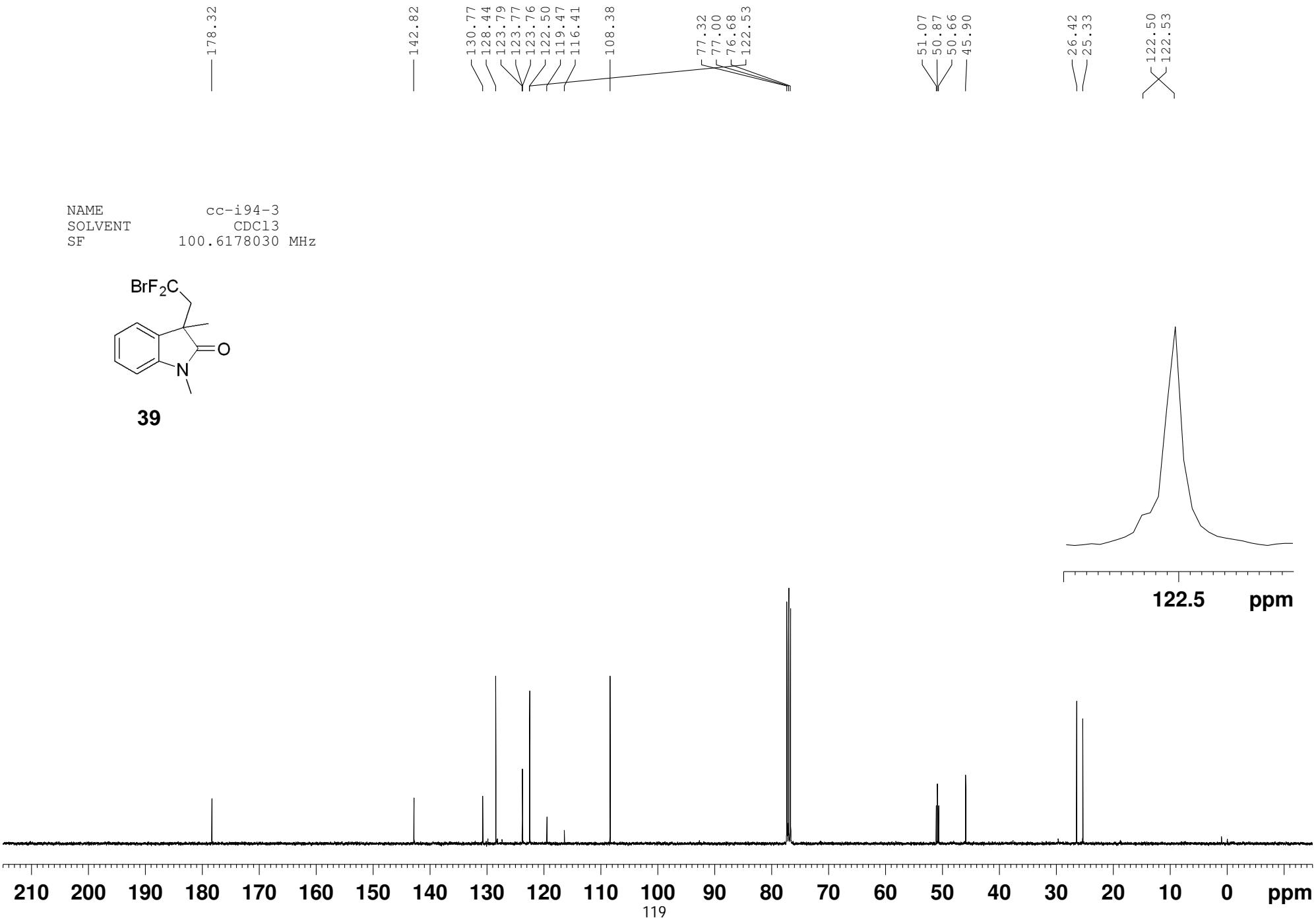


NAME cc-i94-3
SOLVENT CDCl₃
SF 400.1500069 MHz



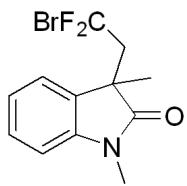
38



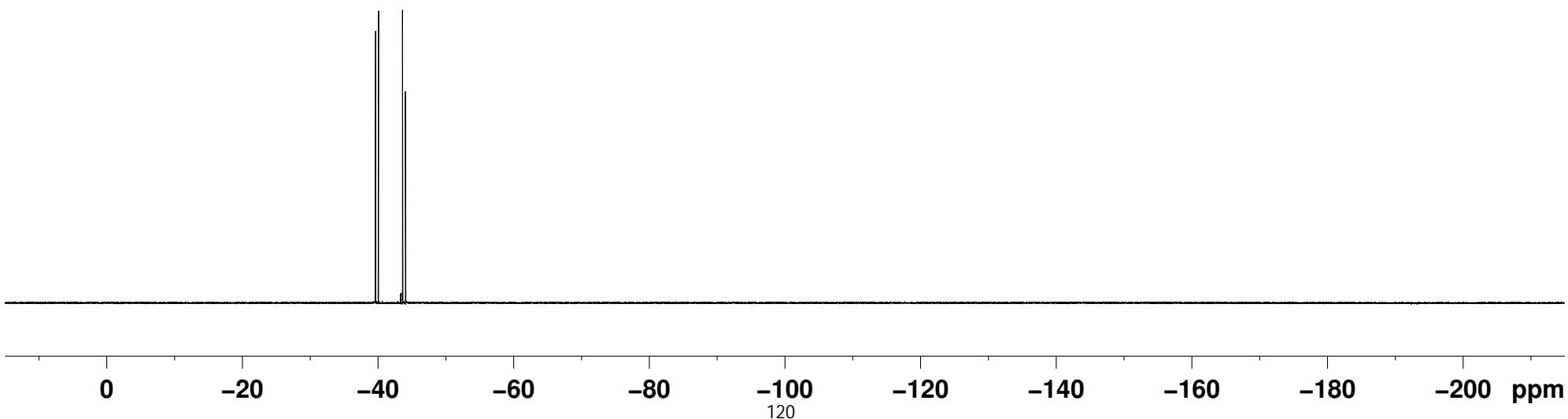


-39.669
-40.080
-43.649
-44.060

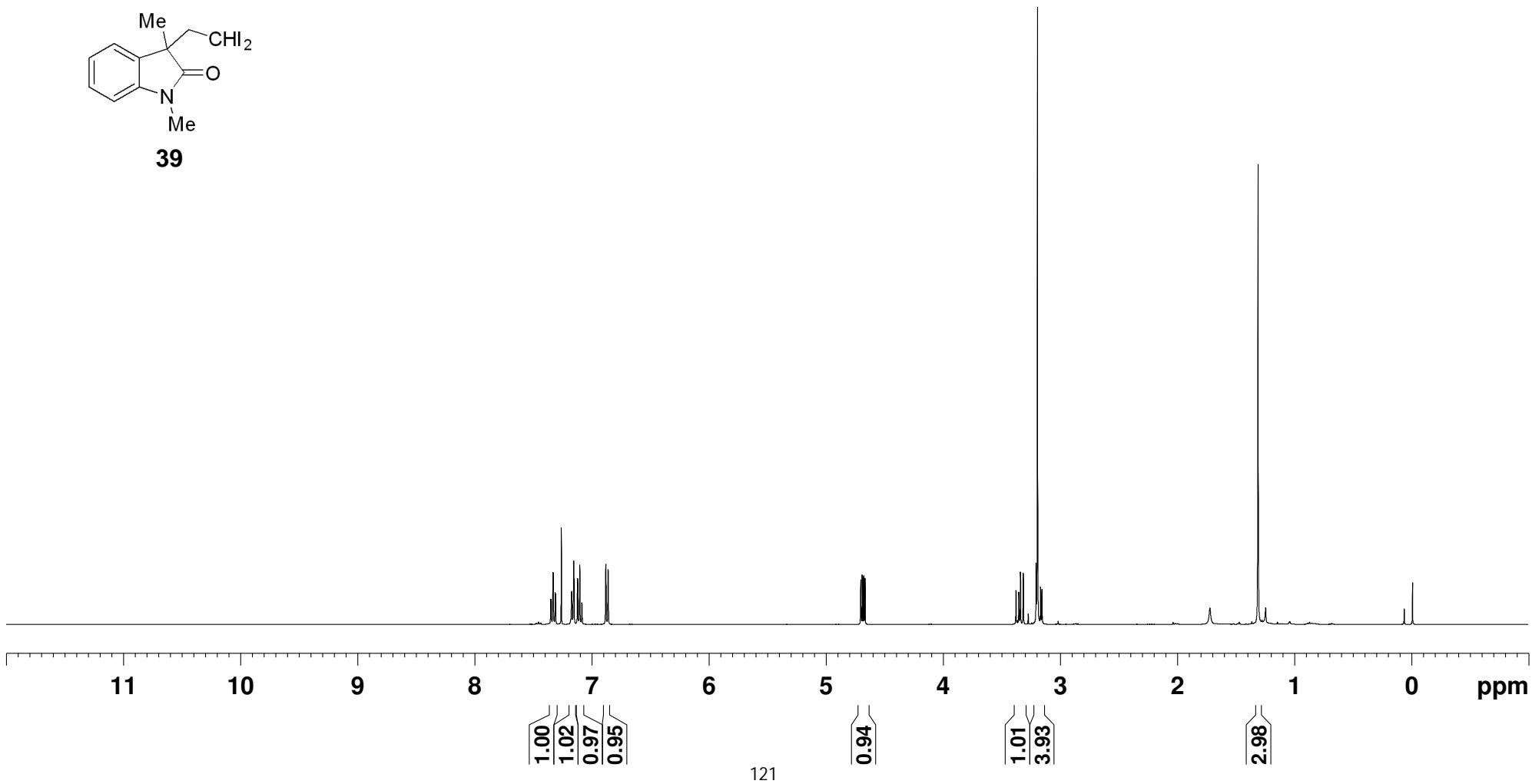
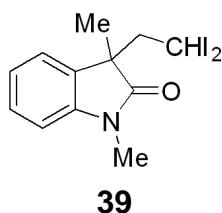
NAME cc-i94-3-0930
SOLVENT CDCl₃
SF 376.5171850 MHz



38



NAME cc-i94-6
SOLVENT CDCl₃
SF 400.1500095 MHz



— 178.49

— 143.87

— 130.31
— 128.61
— 122.69
— 122.66

— 108.67

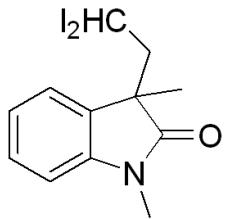
— 77.32
— 77.00
— 76.68

— 54.00
— 50.77

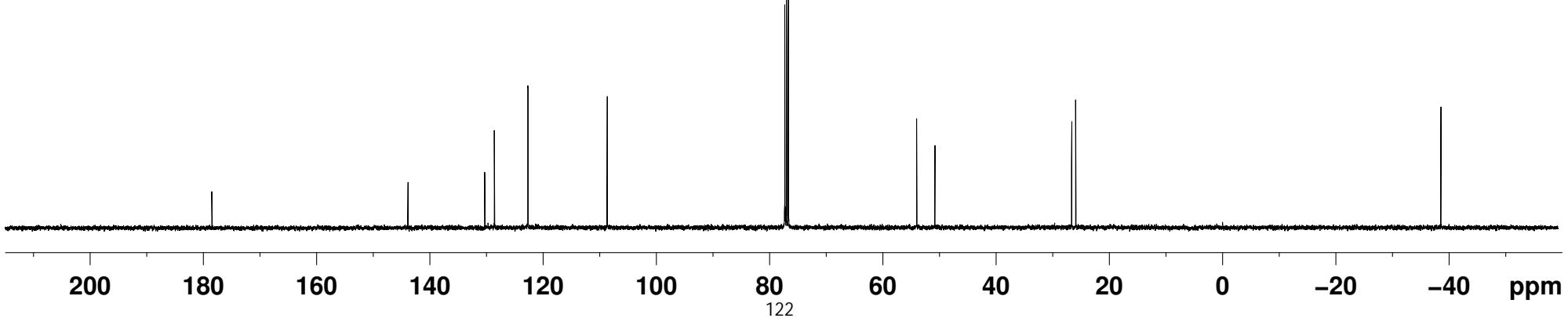
— 26.64
— 25.92

— -38.60

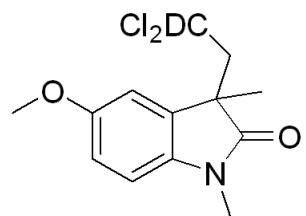
NAME cc-i94-6-0930
SOLVENT CDCl₃
SF 100.6178033 MHz



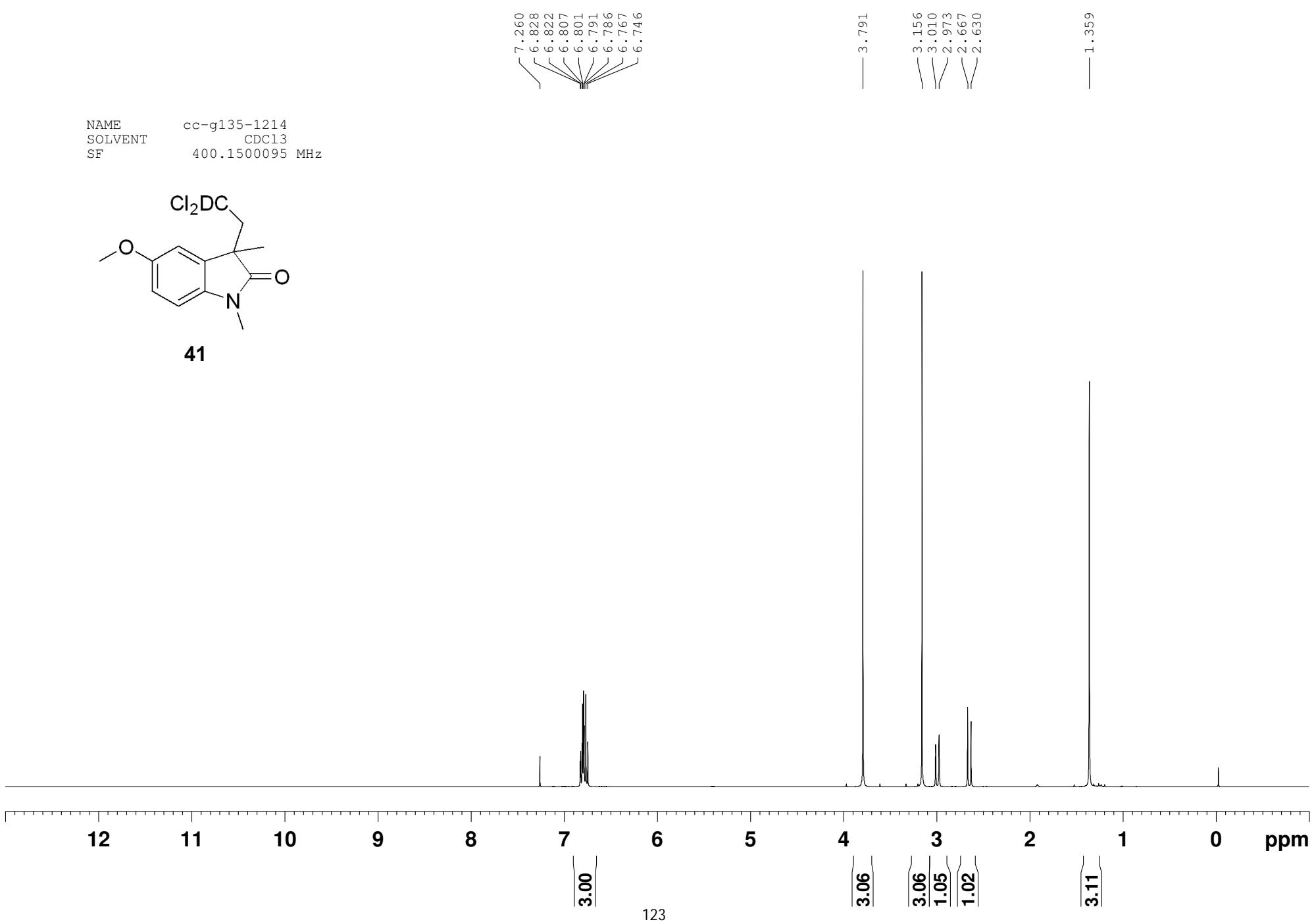
39



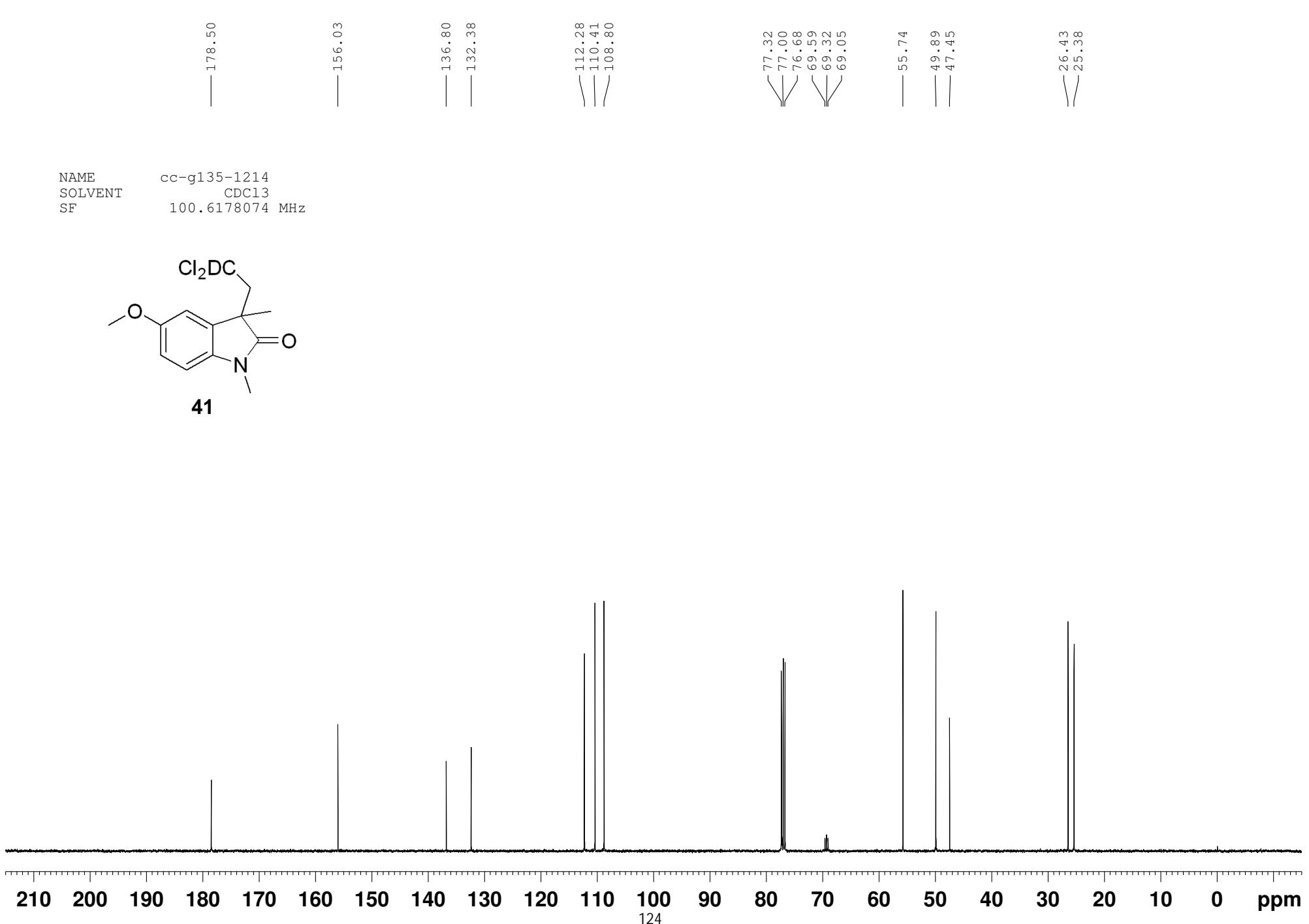
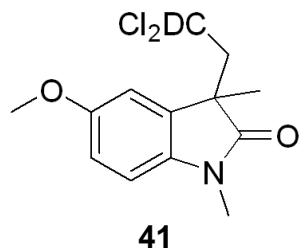
NAME cc-g135-1214
SOLVENT CDCl₃
SF 400.1500095 MHz



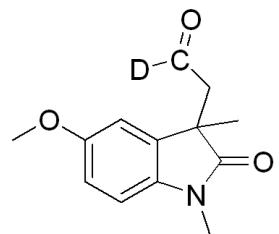
41



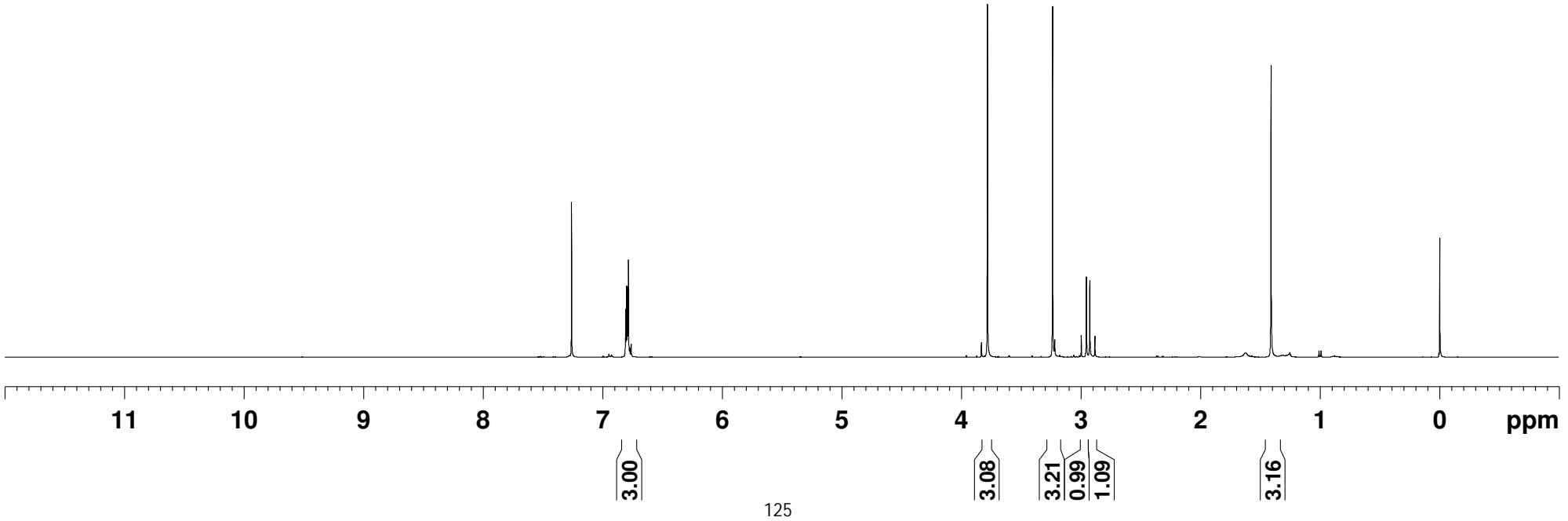
NAME cc-g135-1214
SOLVENT CDCl₃
SF 100.6178074 MHz



NAME cc-G156
SOLVENT CDCl₃
SF 400.1500099 MHz



42



— 198.52
— 198.26
— 197.99

— 178.89

— 155.83

— 136.41
— 133.91

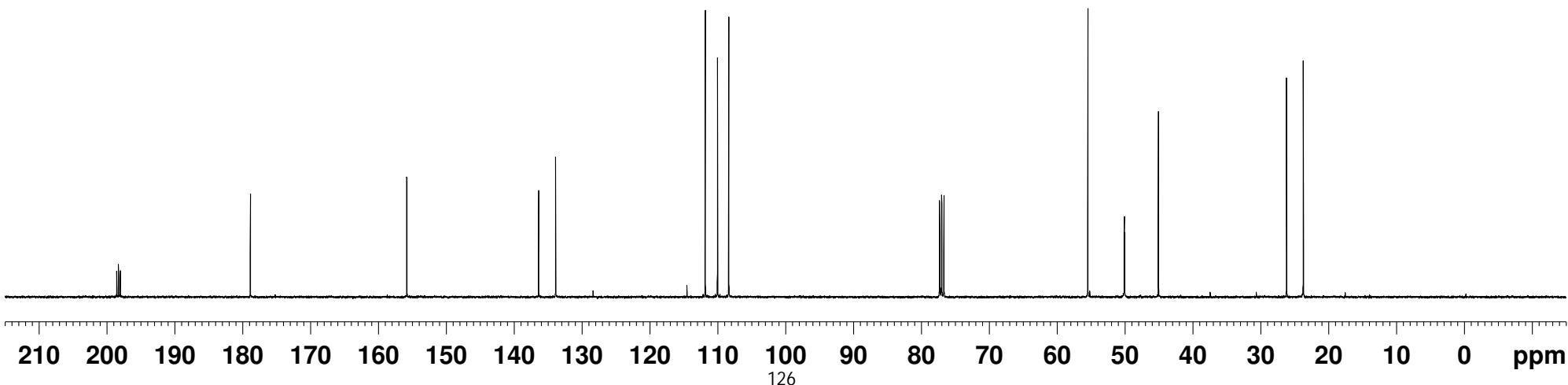
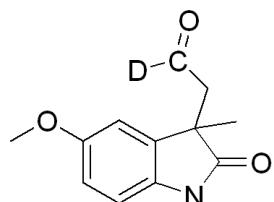
— 111.85
— 110.05
— 108.39

— 77.32
— 77.00
— 76.68

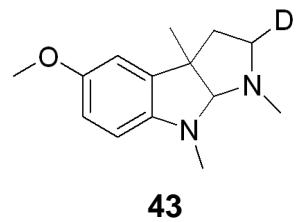
— 55.47
— 50.11
— 50.07
— 50.04
— 45.07

— 26.19
— 23.72

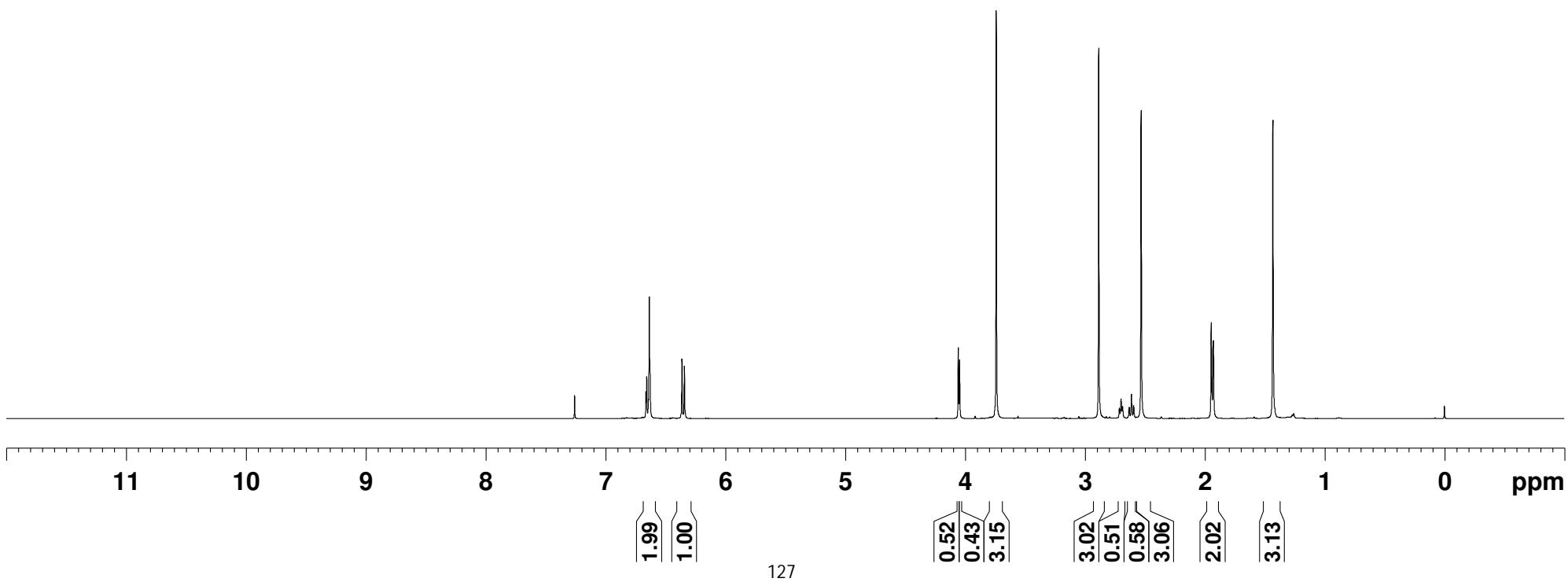
NAME CC-G156-0107
SOLVENT CDCl₃
SF 100.6178213 MHz



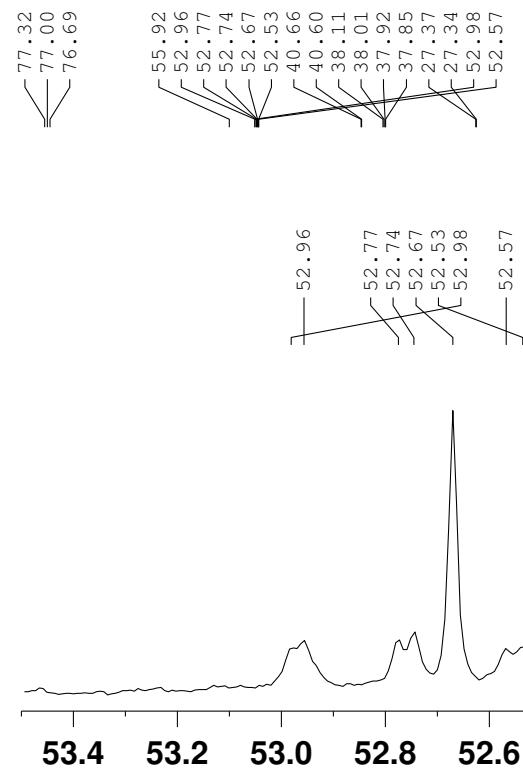
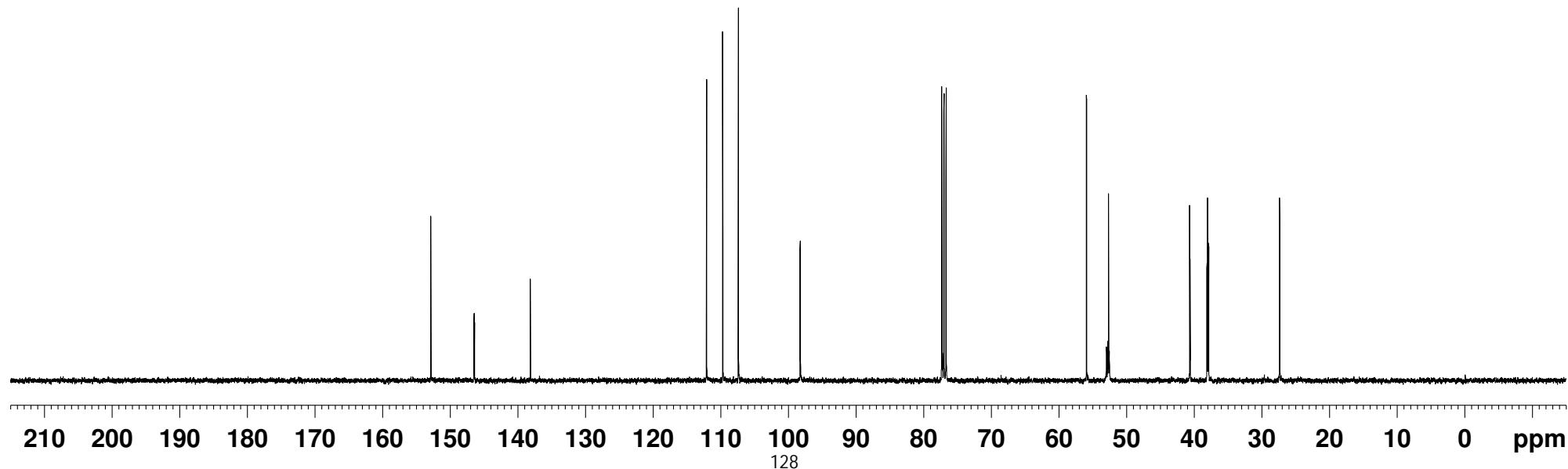
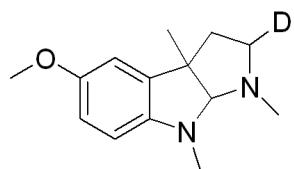
NAME cc-h9
SOLVENT CDCl₃
SF 400.1500104 MHz



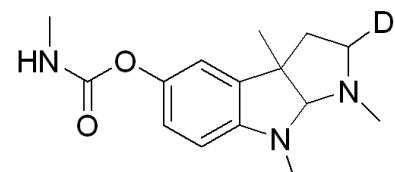
43



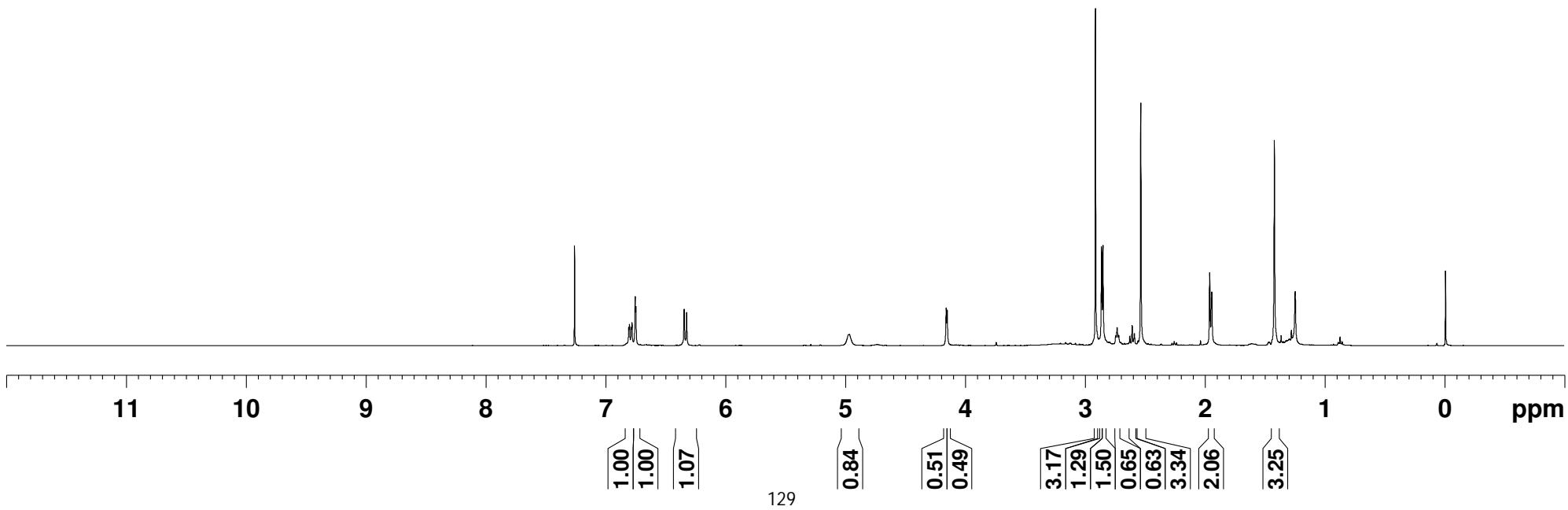
NAME cc-h9-C
SOLVENT CDC13
SF 100.6178081 MHz



NAME cc-I78-2
SOLVENT CDCl₃
SF 400.1500093 MHz



44



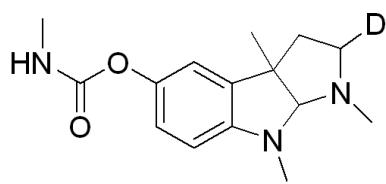
NAME cc-I78-2-c
SOLVENT CDCl₃
SF 100.6178030 MHz

— 156.14
— 149.40
— 149.33
— 143.19
— 137.30

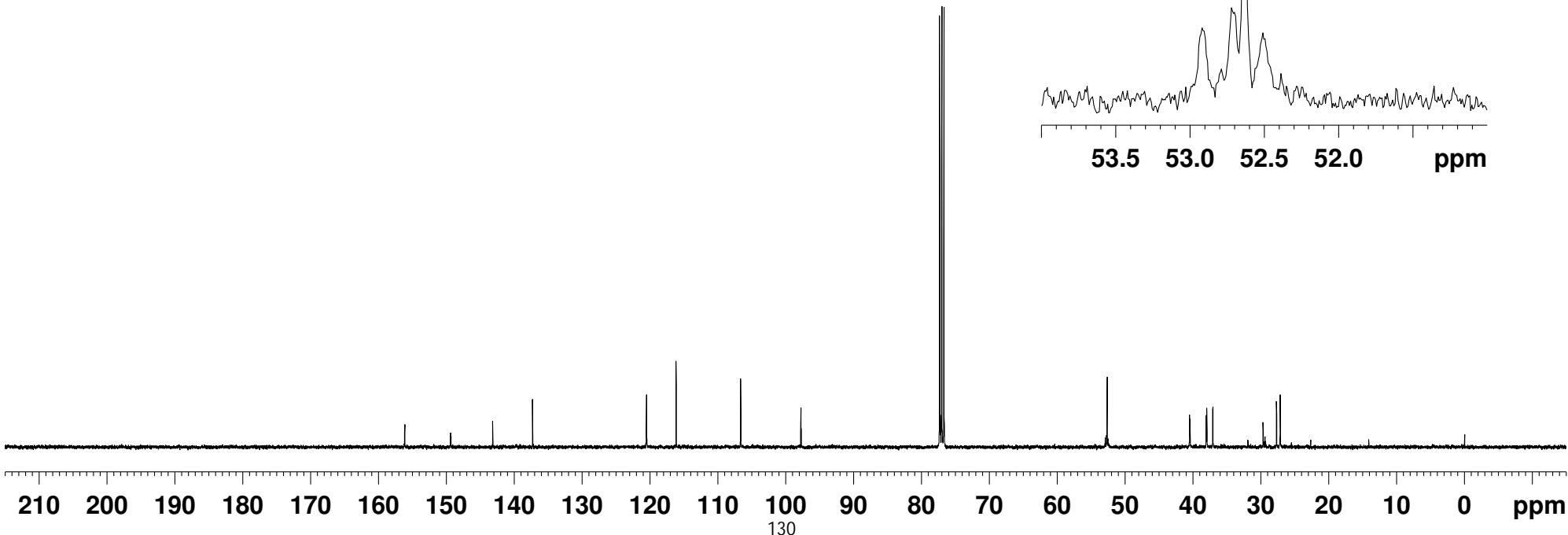
— 120.50
— 116.14
— 106.63

— 97.76
— 97.73

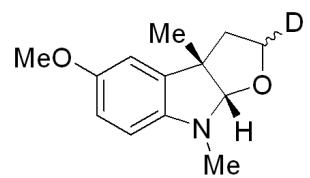
— 77.32
— 77.00
— 76.68
— 52.92
— 52.72
— 52.63
— 52.51
— 40.47
— 40.42
— 38.03
— 37.94
— 37.09
— 37.02
— 27.66
— 27.15
— 27.12



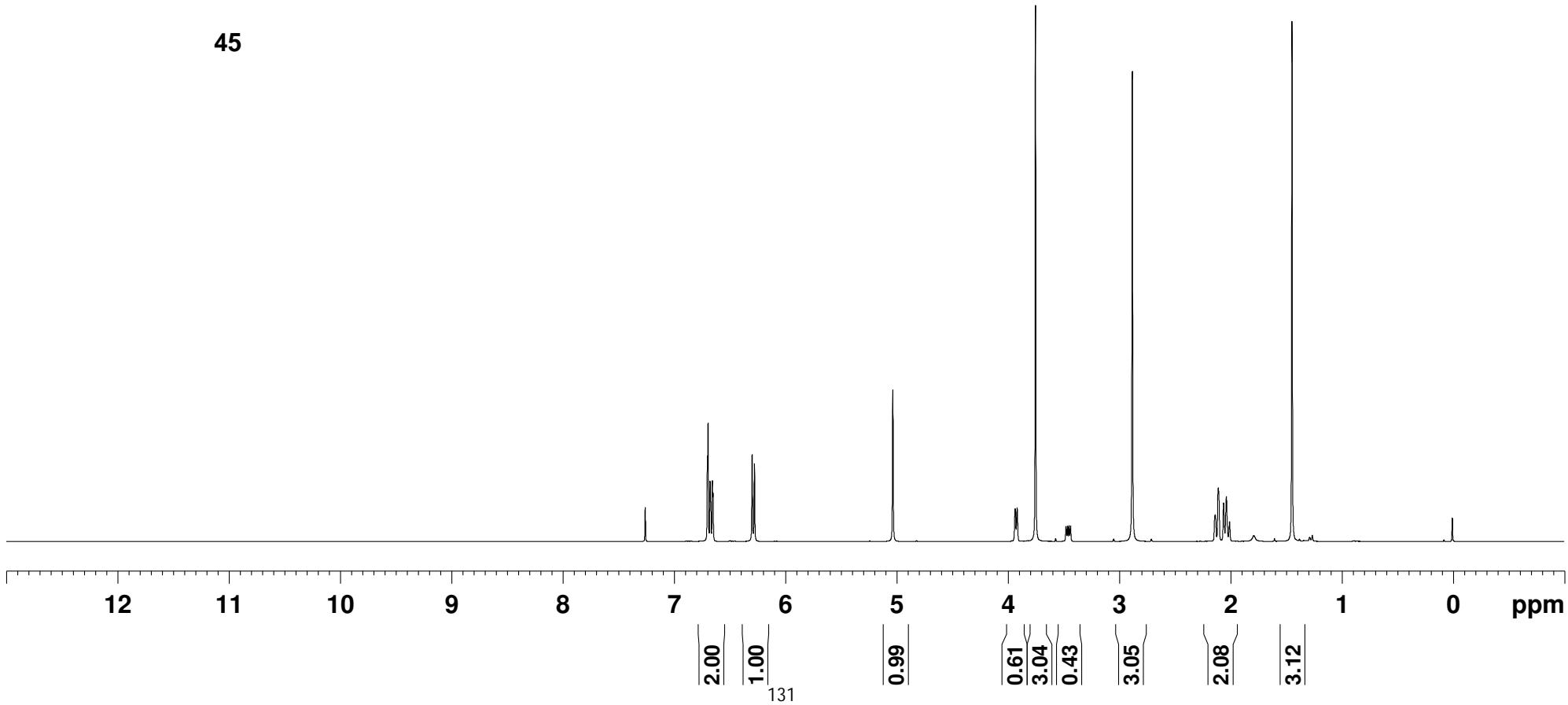
44



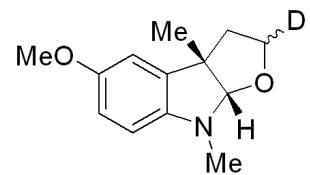
NAME CC-H47
SOLVENT CDC13
SF 400.1500102 MHz



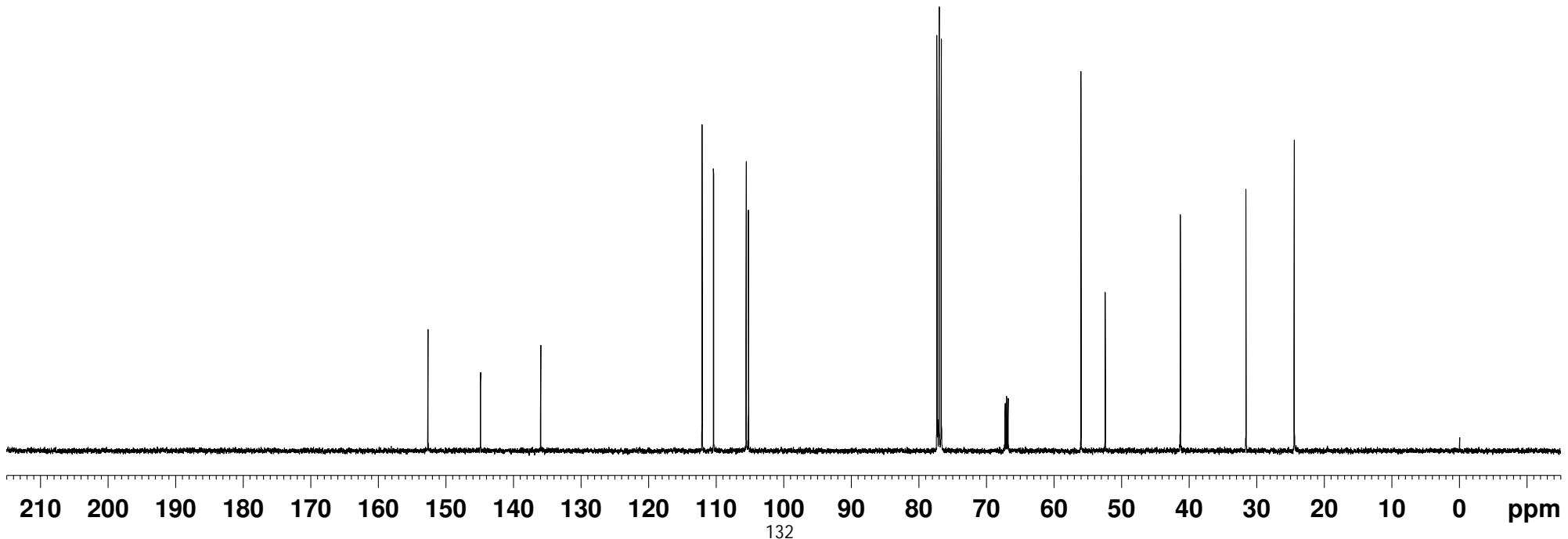
45



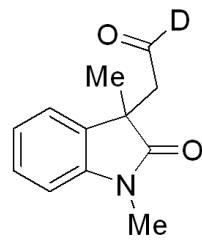
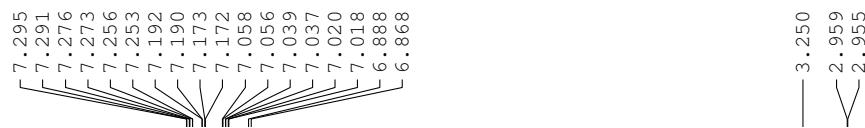
NAME CC-H47
SOLVENT CDCl₃
SF 100.6178052 MHz



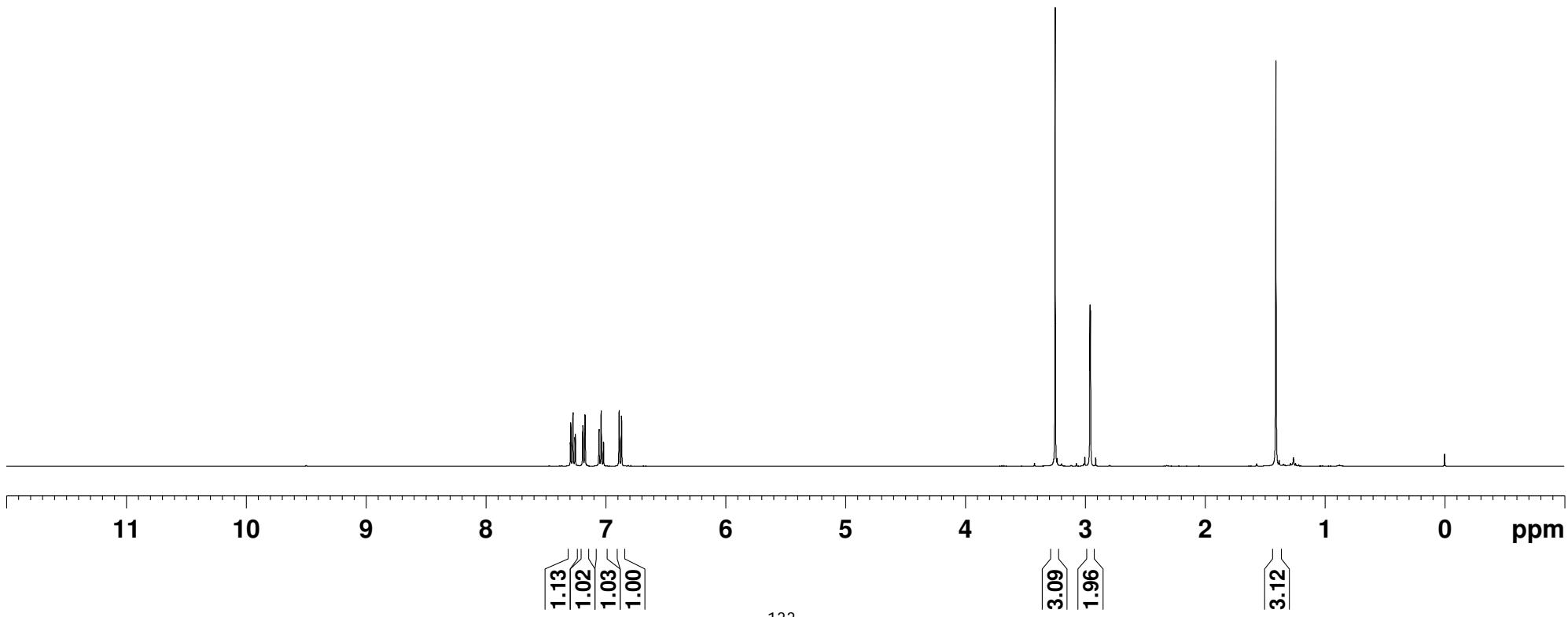
45



NAME CC-I119
SOLVENT CDCl₃
SF 400.1499969 MHz



46



— 198.56
— 198.30
— 198.03

— 179.34

— 143.00

— 132.59
— 128.09
— 122.46
— 122.24

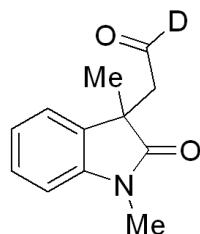
— 108.19

— 77.32
— 77.00
— 76.68

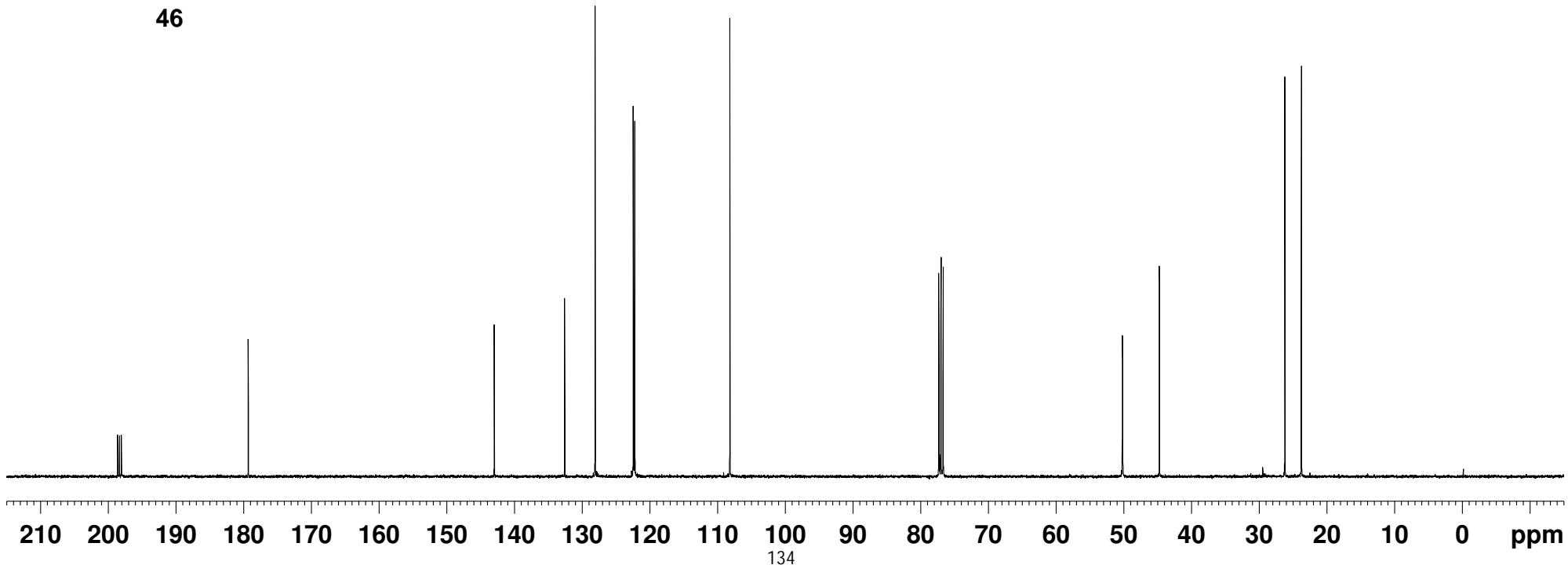
— 50.21
— 50.18
— 50.14
— 44.73

— 26.20
— 23.76

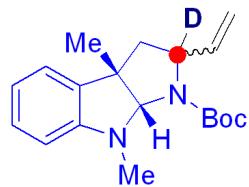
NAME CC-I119
SOLVENT CDCl₃
SF 100.6178169 MHz



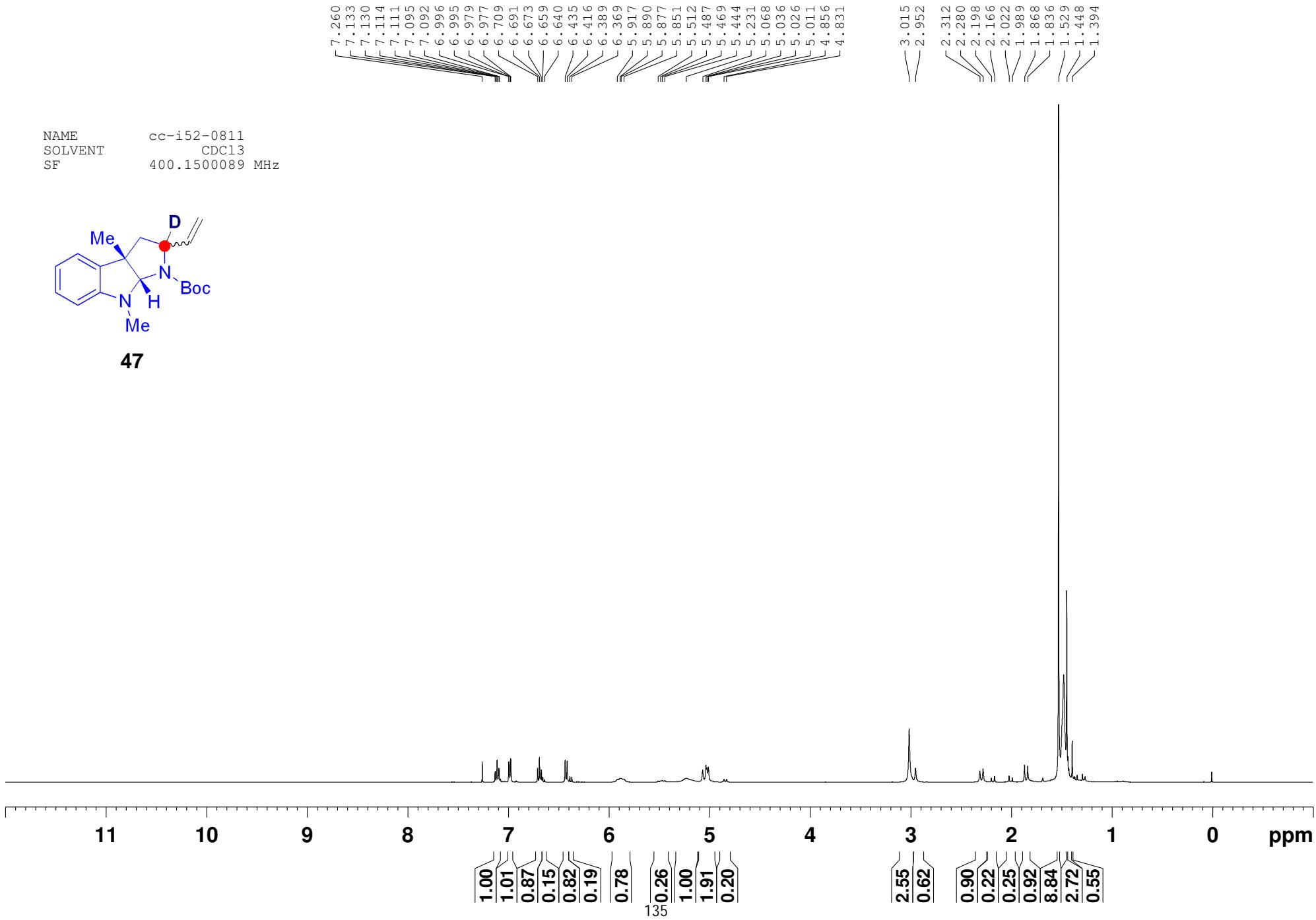
46

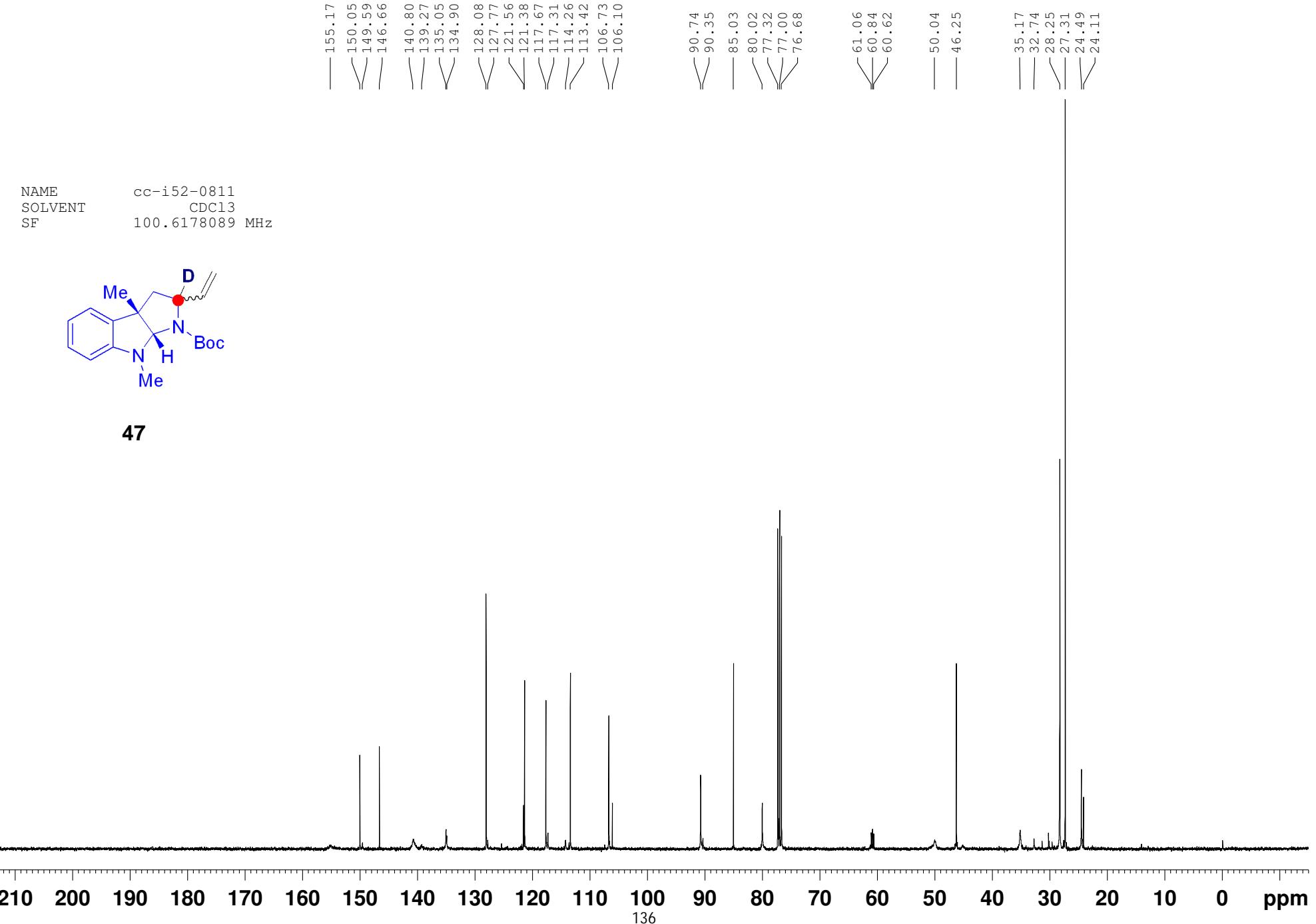


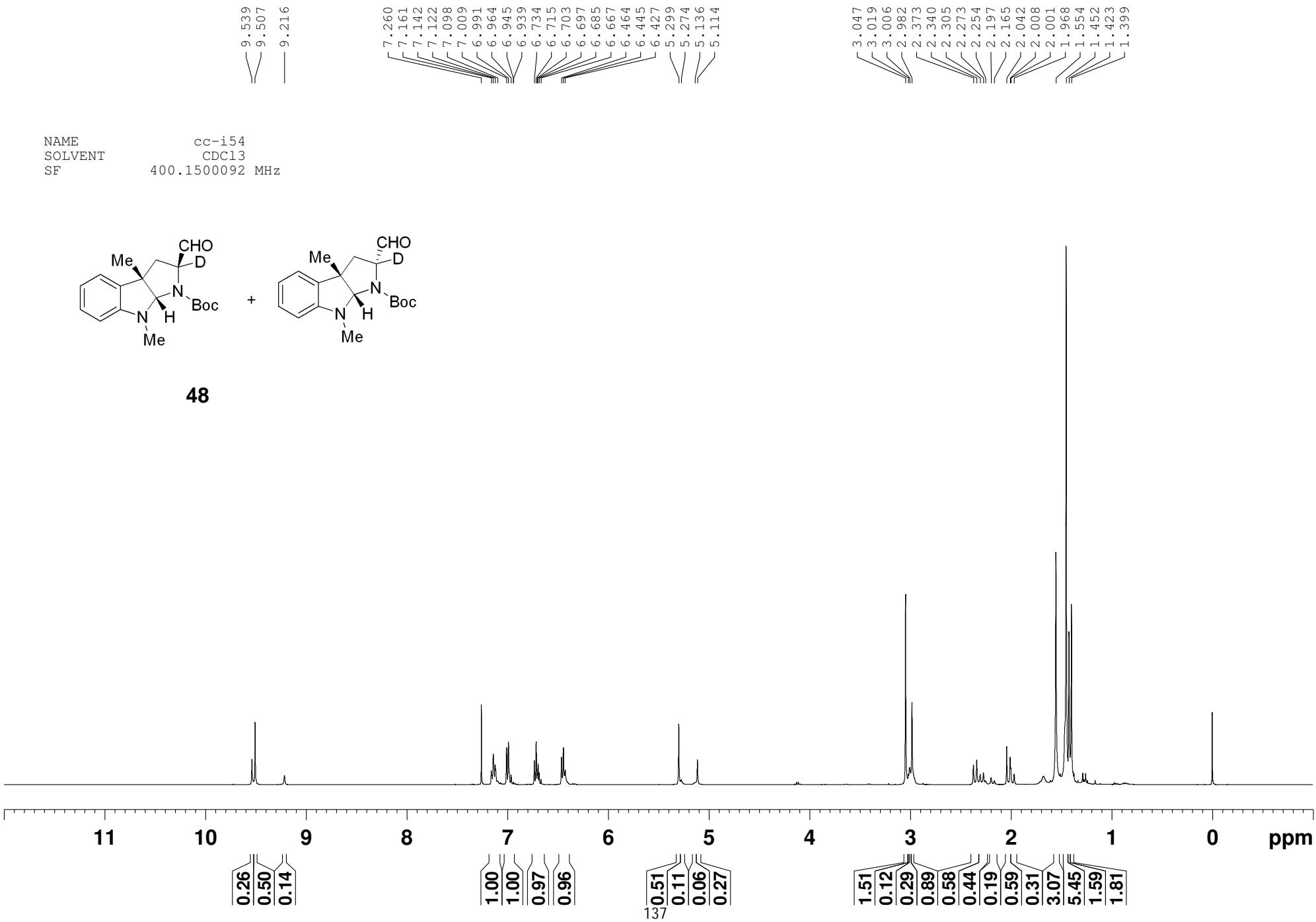
NAME cc-i52-0811
SOLVENT CDCl₃
SF 400.1500089 MHz

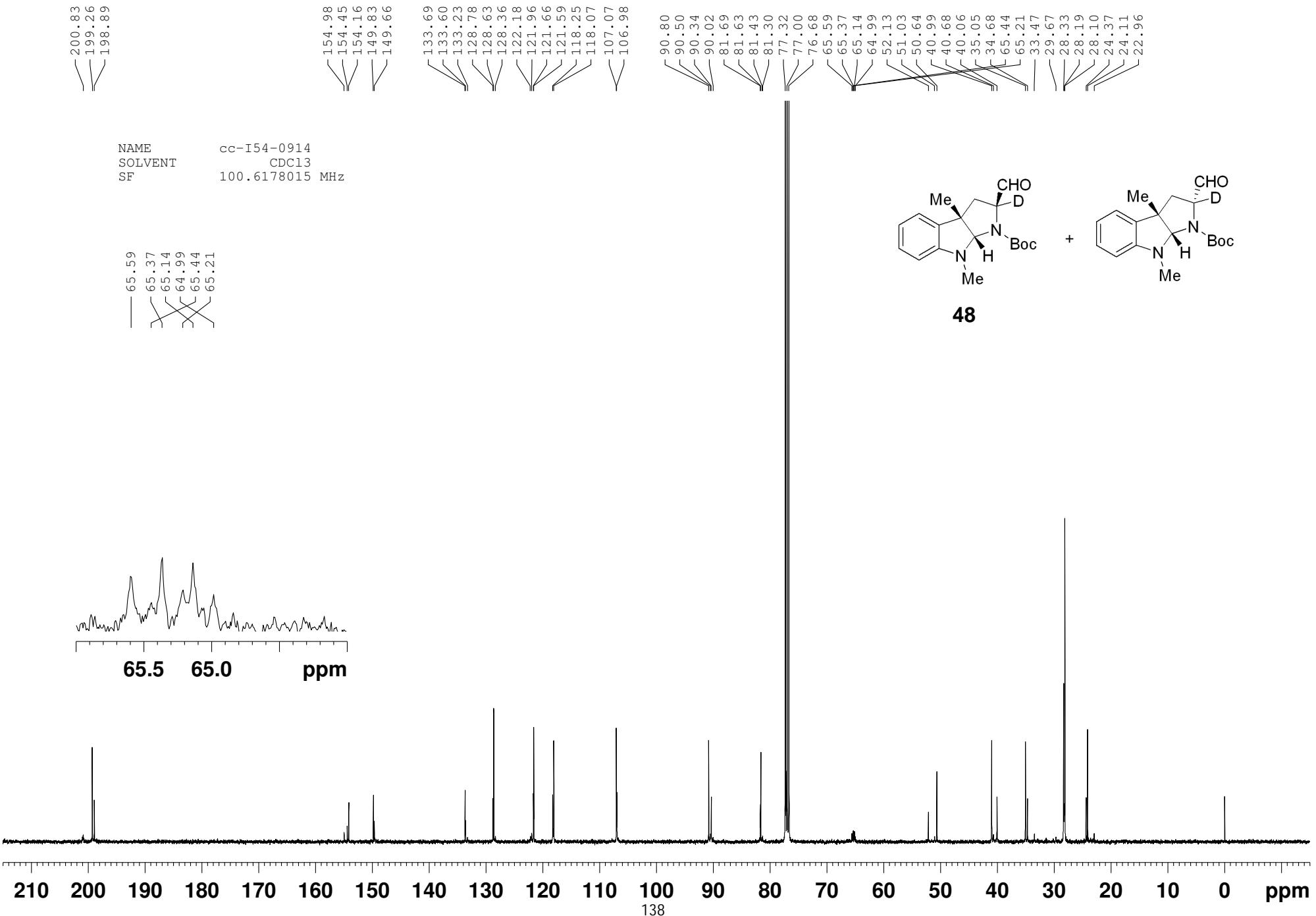


47

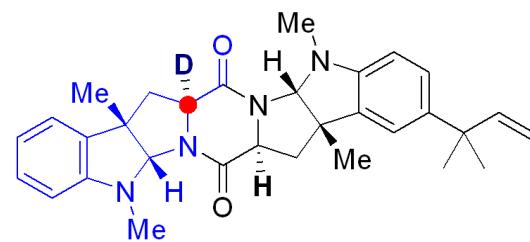




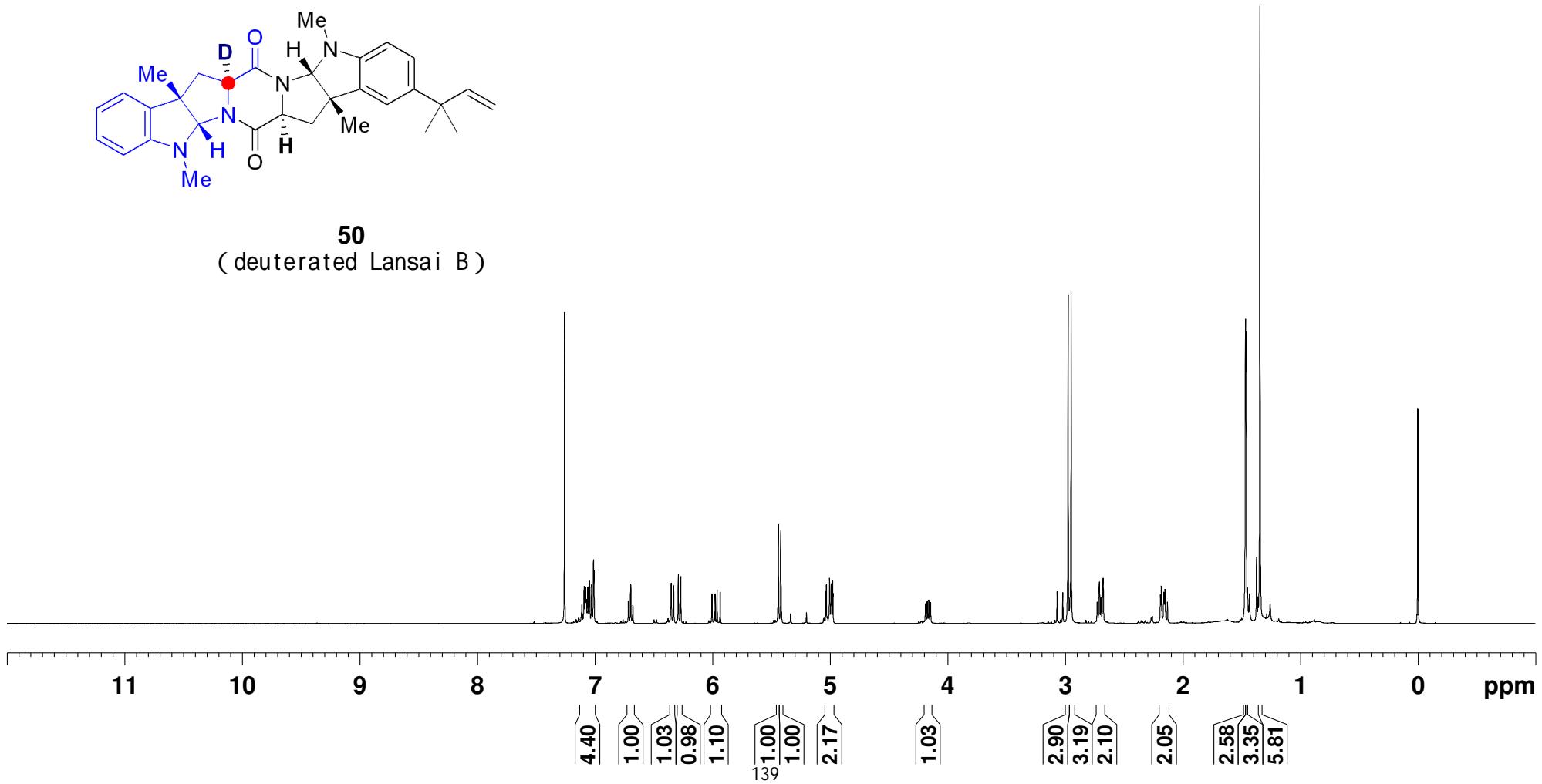




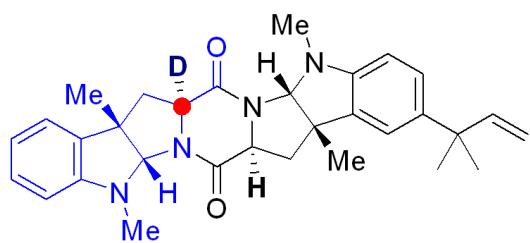
NAME cc-LansaiB-true
SOLVENT CDCl₃
SF 400.1300097 MHz



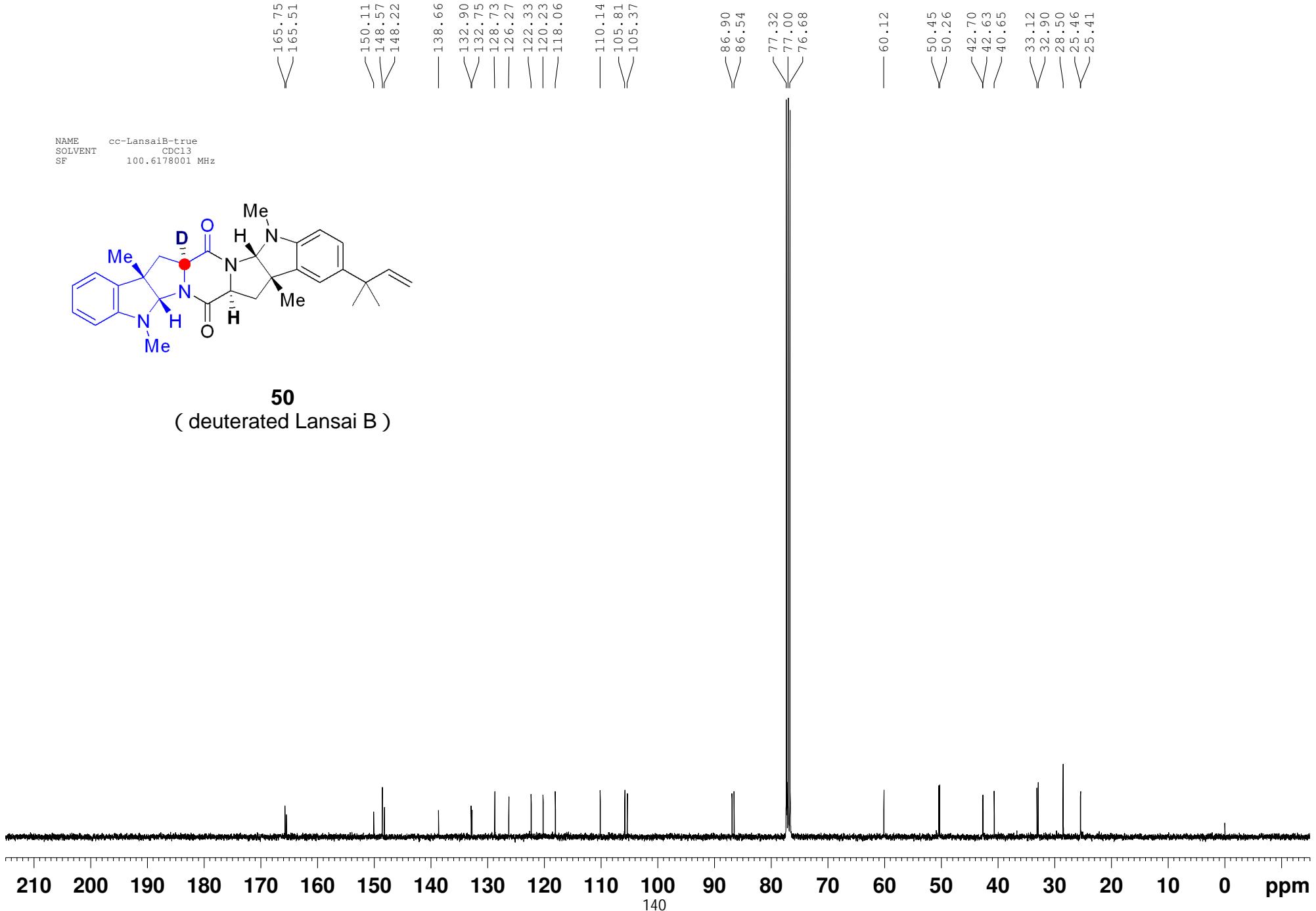
50
(deuterated Lansai B)

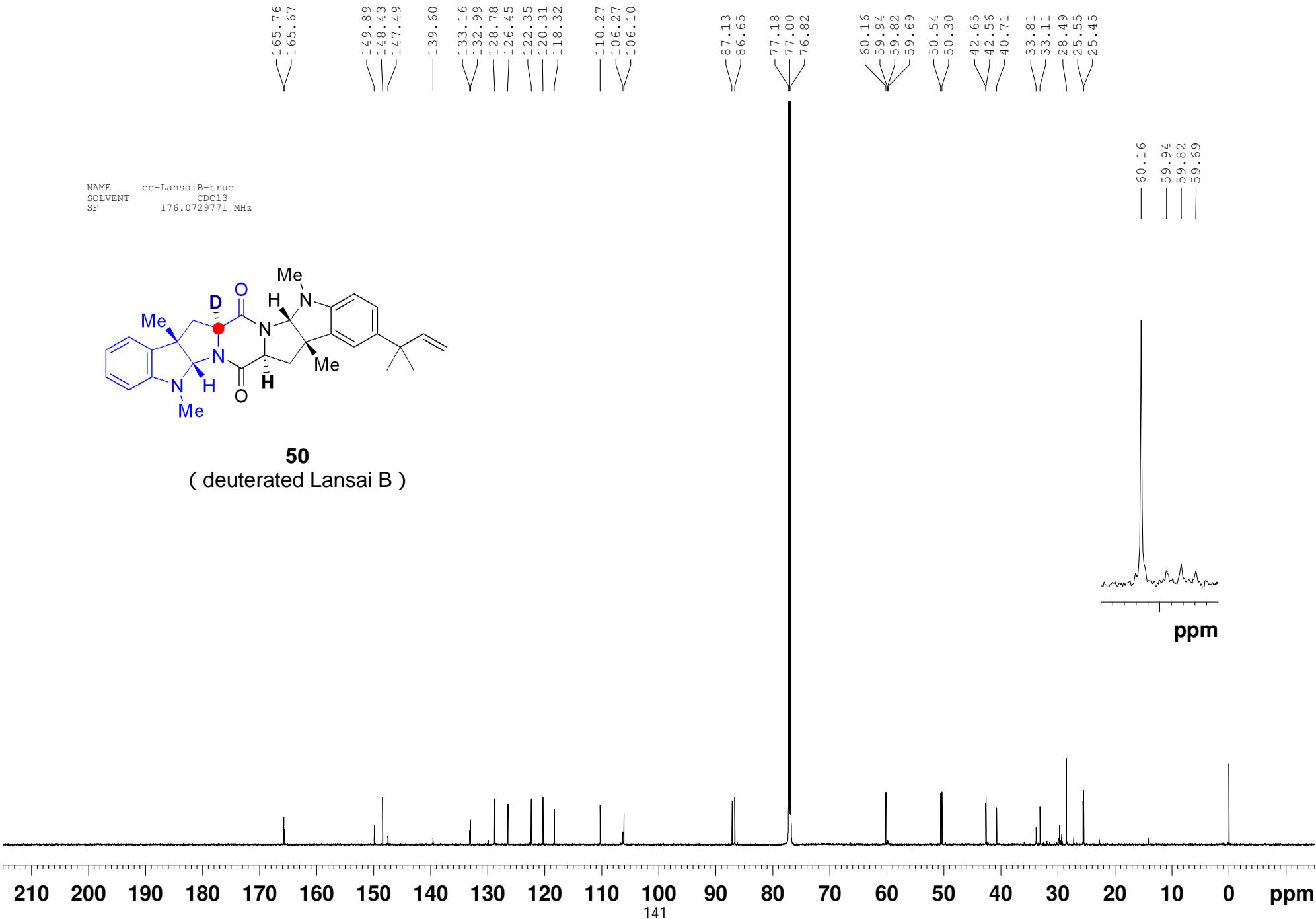


NAME cc-LansaiB-true
SOLVENT CDCl₃
SF 100.6178001 MHz



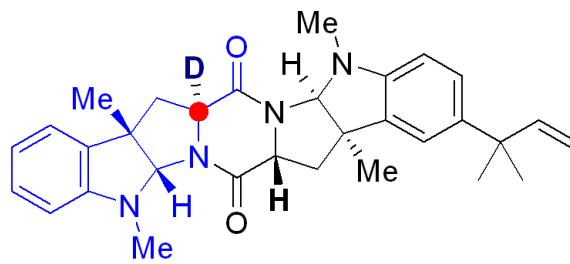
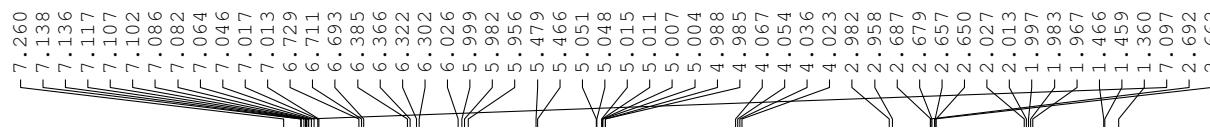
50
(deuterated Lansai B)





NAME
SOLVENT
SF

CC-I80-T
CDCl₃
400.1500092 MHz



51

