# **Electronic Supplementary Information**

# Dynamics of the copper(I)-alkyne interaction and its use in a heteroleptic four-component catalytic rotor

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### 1. Synthesis

### **1.1 General information**

All solvents were dried by distillation prior to use while commercial reagents were used without any further purification. Bruker Avance (400 MHz), Jeol ECZ 500 (500 MHz) and Varian VNMR-S 600 (600 MHz) spectrometers were used to measure <sup>1</sup>H and <sup>13</sup>C NMR spectra applying the deuterated solvent as the lock and residual protiated solvent as internal reference (CDCl<sub>3</sub>:  $\delta_H$ 7.26 ppm,  $\delta_C$  77.0 ppm; CD<sub>2</sub>Cl<sub>2</sub>:  $\delta_H$  5.32 ppm,  $\delta_C$  53.8 ppm, DMSO- $d_6$ :  $\delta_H$  2.50 ppm,  $\delta_C$  39.52 ppm). The following abbreviations were used to define NMR peak patterns: s = singlet, d =doublet, t = triplet, dd = doublet of doublets, bs = broad signal, m = multiplet. The coupling constant values are given in Hertz (Hz) and, wherever possible, assignments of protons are provided. The numbering of different carbons in different molecular skeletons does not necessarily follow the IUPAC nomenclature rules; it is exclusively provided for assigning NMR signals. All electrospray ionization (ESI-MS) spectra were recorded on a Thermo-Quest LCQ deca and the theoretical isotopic distributions of the mass signals were calculated (https://omics.pnl.gov/software/molecular-weight-calculator) using a molecular weight calculator software. Melting points of compounds were measured on a BÜCHI 510 instrument and are not corrected. Infrared spectra were recorded on a Perkin Elmer Spectrum-Two FT-IR spectrometer. Elemental analysis was performed using the EA-3000 CHNS analyzer. UV-vis spectra were recorded on a Varian Cary 100 BioUV/Vis spectrometer. Column chromatography was performed either on silica gel (60-400 mesh) or neutral alumina (Fluka, 0.05-0.15 mm, Brockmann Activity 1). Merck silica gel (60 F254) or neutral alumina (150 F254) sheets were used for thin layer chromatography (TLC). All complexations were performed directly in the NMR tube using  $CD_2Cl_2$  as solvent.

# 1.2 Synthesis and characterization of ligands



Scheme S1. Reaction scheme to synthesis stator 2. Synthesis of  $8^{1}, 9^{2}, 10^{2}, 11^{2}$  and  $12^{3}$  was done following literature-known procedures.



Scheme S2. Reaction scheme to synthesis rotator 3. Synthesis of 15,<sup>4</sup>  $16^4$  and  $17^3$  was done following literature-known procedures.

Synthesis of stator  $1^5$  and model compounds  $4^6$  and  $5^7$  was accomplished by a literature-known procedure.

Synthesis of  $2^8$ 



Under N<sub>2</sub> atmosphere, compounds 12 (100 mg, 107 µmol) and 11 (195 mg, 430 µmol) were dissolved in dry DMF (20 mL) and dry Et<sub>3</sub>N (20 mL) in a sealed tube. The mixture was degassed by two freeze-pump-thaw cycles. Then, Pd(PPh<sub>3</sub>)<sub>4</sub> (15.0 mg, 13.0 µmol) was added under N<sub>2</sub> atmosphere. After degassing again by freeze-pump-thaw cycles, the reaction mixture was stirred at 75 °C for 18 h. The solvent was evaporated under reduced pressure and the residue worked up with ice-cold water to remove DMF. The organic part was extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated. The compound was first purified by column chromategraphy (SiO<sub>2</sub>) using CH<sub>2</sub>Cl<sub>2</sub> in as eluent ( $R_f = 0.3$ ). It was finally purified by size exclusion chromategraphy using SX-3 biobead using THF as eluent to afford a violet solid (94.5 mg, 70%). **mp:** > 250 °C. <sup>1</sup>**H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 400 MHz):**  $\delta$  = 1.84 (s, 12H, 15'-H), 1.85 (s, 6H, 18'-H), 2.01 (s, 6H, 1'-H), 2.07 (s, 6H, 10'-H), 2.36 (s, 3H, 11'-H), 2.61 (s, 3H, 19'-H), 2.62 (s, 6H, 16'-H), 2.69 (s, 6H, 2'-H), 6.98 (s, 2H, 9'-H), 7.29 (s, 2H, 17'-H), 7.30 (s, 4H, 14'-H), 7.59 (d,  ${}^{3}J = 8.2$ Hz, 1H, 3'/8'-H), 7.60 (d,  ${}^{3}J = 8.2$  Hz, 1H, 8'/3'-H), 7.93 (s, 2H, 5'+6'-H), 7.98 (d,  ${}^{3}J = 8.4$  Hz, 2H, 12'-H), 8.25 (d,  ${}^{3}J = 8.4$  Hz, 2H, 13'-H), 8.36 (d,  ${}^{3}J = 8.2$  Hz, 1H, 4'/7'-H), 8.39 (d,  ${}^{3}J = 8.2$ Hz, 1H, 7<sup>'</sup>/4<sup>'</sup>-H), 8.70 (s, 4H, β-H), 8.76 (d,  ${}^{3}J$  = 4.6 Hz, 2H, β-H), 8.93 (d,  ${}^{3}J$  = 4.6 Hz, 2H, β-H) ppm.

### Synthesis of 18<sup>9</sup>



In a dry sealed tube, compounds **17** (250 mg, 247 µmol) and **16** (233 mg, 1.98 mmol) were taken and dissolved in 20 mL of DMF and 20 mL of triethylamine. The solution was deaerated by the freeze-pump-thaw method twice. To this mixture, Pd(PPh<sub>3</sub>)<sub>4</sub> (57.0 mg, 49.4 µmol) was added. The solution was once more deaerated using the freeze-pump-thaw method and it was allowed to stir at 80 °C for 20 h. All the solvents were then evaporated under vacuum. The residue was worked up using CH<sub>2</sub>Cl<sub>2</sub> and ice-cold water. The organic part was separated, dried over Na<sub>2</sub>SO<sub>4</sub> and filtered to get rid of solids. The filtrate was evaporated under vacuum and subjected to column chromatography on silica (SiO<sub>2</sub>) using 50% ethyl acetate in hexane ( $R_f = 0.4$ ). Finally, the compound was purified over SX-3 bio-bead using THF as eluent to obtain the violet compound (159 mg, 65%). **Mp**: > 250 °C. <sup>1</sup>**H NMR (CDCl<sub>3</sub>, 500 MHz)**:  $\delta = 1.83$  (s, 12H, h-H), 2.63 (s, 6H, i-H), 5.00 (s, 2H, e-H), 6.89 (d, <sup>3</sup>*J* = 8.4 Hz, 4H, d-H), 7.29 (s, 4H, g-H), 7.58 (d, <sup>3</sup>*J* = 8.4 Hz, 4H, c-H), 7.89 (d, <sup>3</sup>*J* = 8.4 Hz, 4H, b-H), 8.22 (d, <sup>3</sup>*J* = 8.4 Hz, 4H, a-H), 8.79 (d, <sup>3</sup>*J* = 4.4 Hz, 4H, β-H), 8.90 (d, <sup>3</sup>*J* = 4.4 Hz, 4H, β-H) ppm. Synthesis of **3** 



Compound **18** (100 mg, 100 µmol) and K<sub>2</sub>CO<sub>3</sub> (110 mg, 800 µmol) were stirred in THF (30 mL) at room temp. for 10 min, then, propargyl bromide (19, 158 mg, 1.32 mmol) was added. After heating to reflux for 6 h, the reaction was complete as confirmed by TLC. The solvents were evaporated, and the resultant mixture was worked up with water and  $CH_2Cl_2$  (150 mL). The organic part was evaporated under reduced pressure and dried over Na<sub>2</sub>SO<sub>4</sub>. The compound was purified via column chromatography on silica (SiO<sub>2</sub>) using 30% CH<sub>2</sub>Cl<sub>2</sub> in hexane ( $R_f = 0.3$ ) yielding the violet colour product (91.5 mg, 85%). mp: >250 °C. IR (KBr): v = 675, 727, 799, 810, 831, 852, 872, 927, 999, 1027, 1065, 1107, 1138, 1175, 1205, 1224, 1258, 1286, 1304, 1337, 1376, 1400, 1451, 1512, 1570, 1600, 2214, 2916, 2968, 3118, 3294 cm<sup>-1</sup>. <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 400 MHz):  $\delta = 1.82$  (s, 12H, h-H), 2.63 (s, 6H, i-H), 2.64 (t,  ${}^{3}J = 2.4$  Hz, 2H, f-H), 4.79 (d,  ${}^{3}J = 2.4$  Hz, 4H, e-H), 7.05 (d,  ${}^{3}J = 8.8$  Hz, 4H, d-H), 7.31 (s, 4H, g-H), 7.64 (d,  ${}^{3}J = 8.8$ Hz, 4H, c-H), 7.91 (d,  ${}^{3}J = 8.4$  Hz, 4H, b-H), 8.22 (d,  ${}^{3}J = 8.4$  Hz, 4H, a-H), 8.78 (d,  ${}^{3}J = 4.4$  Hz, 4H,  $\beta$ -H), 8.92 (d,  ${}^{3}J$  = 4.4 Hz, 4H,  $\beta$ -H) ppm.  ${}^{13}C$  NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  = 21.5, 21.6, 55.9, 75.9, 78.2, 88.5, 90.2, 115.1, 116.5, 119.5, 119.6, 122.6, 127.7, 129.7, 130.9, 132.2, 133.2, 134.5, 137.5, 138.9, 139.2, 142.7, 149.8, 150.0, 157.7 ppm. Elemental analysis: Anal. Calcd for C<sub>72</sub>H<sub>52</sub>N<sub>4</sub>O<sub>2</sub>Zn•CH<sub>2</sub>Cl<sub>2</sub>: C, 75.88; H, 4.71; N, 4.85. Found: C, 75.59; H, 4.52; N, 4.66.

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### 1.3 Synthesis and characterization of complexes

Synthesis of model complex C1



In an NMR tube, compound **4** (565 µg, 1.36 µmol), [Cu(CH<sub>3</sub>CN)<sub>4</sub>]PF<sub>6</sub> (506 µg, 1.36 µmol) and **5** (350 µg, 1.36 µmol) were dissolved in 550 µL of CD<sub>2</sub>Cl<sub>2</sub> to quantitatively furnish complex **C1** = [Cu(**4**)(**5**)]<sup>+</sup>. **IR** (**KBr**): v = 507, 559, 625, 733, 838, 1017, 1084, 1144, 1177, 1221, 1285, 1401, 1484, 1510, 1586, 1611, 1970, 2855, 2918, 2969, 3189, 3238 cm<sup>-1</sup>. <sup>1</sup>H NMR (**CD<sub>2</sub>Cl<sub>2</sub>**, **400 MHz**):  $\delta = 2.05$  (s, 12H, 7"-H), 2.34 (s, 6H, 8"-H), 2.68 (t, <sup>3</sup>*J* = 2.2 Hz, 1H, f'-H), 4.09 (d, <sup>3</sup>*J* = 2.2 Hz, 2H, e'-H), 6.60 (d, <sup>3</sup>*J* = 9.0 Hz, 2H, d'-H), 7.03 (s, 4H, 6"-H), 7.58 (d, <sup>3</sup>*J* = 9.0 Hz, 2H, c'-H), 7.92 (d, <sup>3</sup>*J* = 8.2 Hz, 2H, 3"-H), 8.20 (s, 2H, 5"-H), 8.74 (d, <sup>3</sup>*J* = 8.2 Hz, 2H, 4"-H) ppm. **ESI-MS**: m/z (%) 738.4 (100) [Cu(**4**)(**5**)]<sup>+</sup>.

#### Synthesis of ASB-1



All the spacer protons 12-H, 13-H, a-H and b-H are assigned as s-H. After complexation 14-H, 15-H, g-H and h-H split into 1:1 ratio and they are assigned as 14-H, 14'-H, 15-H, 15'-H, g-H, g'-H, h-H and h'-H, respectively. We see a loss of symmetry in the <sup>1</sup>H NMR due to the restricted rotation of the phenyl group in **ASB1**.

In an NMR tube, compound **1** (1.15 mg, 690 nmol), rotator **3** (738 µg, 690 nmol), DABCO (77.4 µg, 690 nmol) and [Cu(CH<sub>3</sub>CN)<sub>4</sub>]PF<sub>6</sub> (514 µg, 1.38 µmol) were dissolved in 550 µL of CD<sub>2</sub>Cl<sub>2</sub> to furnish the complex **ASB-1** in quantitative yield. **mp:** > 250 °C. **IR** (**KBr**): v = 560, 641, 727, 797, 809, 846, 972, 996, 1064, 1081, 1103, 1179, 1204, 1226, 1286, 1306, 1337, 1378, 1458, 1492, 1511, 1551, 1587, 1608, 1694, 2213, 2729, 2842, 2870, 2916, 2956 cm<sup>-1</sup>. <sup>1</sup>H **NMR** (**CD<sub>2</sub>Cl<sub>2</sub>, 400 MHz**):  $\delta = -4.57 - 4.53$  (m, 6H, DABCO-H), -4.49 - -4.45 (m, 6H, DABCO-H), 1.11 (s, 6H, h/15-H), 1.13 (s, 6H, 15/h-H), 1.67 (s, 6H, h'/15'-H), 1.77 (s, 6H, 15'/h'-H), 2.02 (s, 12H, 1-H), 2.18 (s, 12H, 10-H), 2.47 (s, 6H, 11-H), 2.58 (s, 6H, 16-H), 2.59 (s, 6H, i-H), 2.64 (s, 12H, 2-H), 3.10 (t, <sup>3</sup>J = 1.4 Hz, 2H, f-H), 3.60 (d, <sup>3</sup>J = 1.4 Hz, 4H, e-H), 6.77 (d, <sup>3</sup>J = 8.8 Hz, 4H, d-H), 7.07 (s, 2H, g/14-H), 7.08 (s, 2H, 14/g-H), 7.10 (d, <sup>3</sup>J = 8.0 Hz, 2H, s-H), 7.21 (s, 4H, 9-H), 7.31 (d, <sup>3</sup>J = 8.0 Hz, 2H, s-H), 7.33 (s, 2H, g'/14'-H), 7.34 (s, 2H, 14/g'-H), 7.75 (d, <sup>3</sup>J = 8.8

Hz, 4H, c-H), 7.84 (d,  ${}^{3}J = 8.0$  Hz, 2H, s-H), 7.85 (d,  ${}^{3}J = 8.2$  Hz, 2H, 8/3-H), 7.90 (d,  ${}^{3}J = 8.0$  Hz, 2H, s-H), 7.93-7.99 (m, 4H, s-H), 8.02 (d,  ${}^{3}J = 8.2$  Hz, 2H, 3/8-H), 8.07 (d,  ${}^{3}J = 8.0$  Hz, 2H, s-H), 8.21 (bs, 2H, s-H), 8.29 (d,  ${}^{3}J = 8.8$  Hz, 2H, 6/5-H), 8.32 (d,  ${}^{3}J = 8.8$  Hz, 2H, 5/6-H), 8.35 (d,  ${}^{3}J = 4.6$  Hz, 4H,  $\beta$ (1)-H), 8.36 (d,  ${}^{3}J = 4.6$  Hz, 4H,  $\beta$ (3)-H), 8.54 (d,  ${}^{3}J = 4.6$  Hz, 4H,  $\beta$ (1)-H), 8.59 (d,  ${}^{3}J = 4.6$  Hz, 4H,  $\beta$ (3)-H), 8.83 (d,  ${}^{3}J = 8.2$  Hz, 2H, 7/4-H), 8.89 (d,  ${}^{3}J = 8.2$  Hz, 2H, 4/7-H) ppm. **ESI-MS:** *m*/*z* (%) 1489.1 (100) [Cu<sub>2</sub>(1)(3)(DABCO)]<sup>2+</sup>. **Elemental analysis:** Anal. Calcd for C<sub>194</sub>H<sub>160</sub>N<sub>14</sub>O<sub>2</sub>Zn<sub>2</sub>Cu<sub>2</sub>P<sub>2</sub>F<sub>12</sub>•5CH<sub>2</sub>Cl<sub>2</sub>•2CH<sub>3</sub>CN: C, 64.61; H, 4.70; N, 5.94. Found: C, 64.64; H, 4.91; N, 6.10.

Synthesis of ROT-1



All the spacer protons 12'-H, 13'-H, a-H and b-H are assigned as s-H.

In an NMR tube, compound **2** (976 µg, 776 nmol), rotator **3** (831 µg, 776 nmol), DABCO (87.1 µg, 776 nmol) and [Cu(CH<sub>3</sub>CN)<sub>4</sub>]PF<sub>6</sub> (289 µg, 776 nmol) were dissolved in 550 µL of CD<sub>2</sub>Cl<sub>2</sub> to furnish the complex **ROT-1** in quantitative yield. **mp:** > 250 °C. **IR** (**KBr**): v = 640, 668, 722, 798, 809, 831, 851, 996, 1062, 1103, 1139, 1175, 1204, 1225, 1286, 1304, 1336, 1378, 1400, 140

1456, 1479, 1511, 1608, 1696, 1809, 2213, 2730, 2868, 2917, 2954, 3116, 3307 cm<sup>-1</sup>. <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 600 MHz):  $\delta$  = – 4.51 (bs, 6H, DABCO-H), –4.40 (bs, 6H, DABCO-H), 1.20 (bs, 12H, h-H), 1.47 (bs, 18H, 15'-H+18'-H), 2.03 (s, 6H, 1'-H), 2.16 (s, 6H, 10'-H), 2.46 (s, 3H, 11'-H), 2.58 (s, 9H, 16'-H+19'-H), 2.60 (s, 6H, i-H), 2.65 (s, 6H, 2'-H), 2.88 (bs, 2H, f-H), 4.19 (bs, 4H, e-H), 6.93 (d, <sup>3</sup>*J* = 8.0 Hz, 4H, d-H), 6.98 (bs, 1H, s-H), 7.15 (d, <sup>3</sup>*J* = 8.4 Hz, 1H, s-H), 7.19-7.34 (m, 12H, g-H+14'-H+17'-H+9-H), 7.63 (d, <sup>3</sup>*J* = 8.4 Hz, 1H, s-H), 7.66 (d, <sup>3</sup>*J* = 8.4 Hz, 1H, s-H), 7.72 (d, <sup>3</sup>*J* = 8.0 Hz, 4H, c-H), 7.84-7.90 (m, 5H, 8'/3'-H+s-H), 7.94-7.97 (m, 2H, s-H), 8.00 (d, <sup>3</sup>*J* = 8.4 Hz, 1H, 3'/8'-H), 8.27-8.28 (bs, 2H, s-H), 8.28 (d, <sup>3</sup>*J* = 8.8 Hz, 2H, 6'/5'-H), 8.31 (d, <sup>3</sup>*J* = 8.8 Hz, 2H, 5'/6'-H), 8.33-8.37 (m, 8H, β(2)-H+β(3)-H), 8.53-8.55 (m, 8H, β(2)-H+β(3)-H), 8.84 (d, <sup>3</sup>*J* = 8.0 Hz, 1H, 7'/4'-H), 8.88 (d, <sup>3</sup>*J* = 8.4 Hz, 1H, 4'/7'-H) ppm. **ESI-MS:** *m*/*z* (%) 2388.8 (100) [Cu(2)(3)]<sup>+</sup>. **Elemental analysis:** Anal. Calcd for C<sub>164</sub>H<sub>138</sub>N<sub>12</sub>O<sub>2</sub>Zn<sub>2</sub>CuPF<sub>6</sub>•CH<sub>2</sub>Cl<sub>2</sub>: C, 72.51; H, 5.16; N, 6.15. Found: C, 72.19; H, 4.97; N, 6.48.

#### Synthesis of ROT-2



All the spacer protons 12'-H, 13'-H, a-H and b-H are assigned as s-H.

In an NMR tube, compound **2** (946 µg, 753 nmol), rotator **3** (806 µg, 753 nmol), DABCO (84.5 µg, 753 nmol) and [Cu(CH<sub>3</sub>CN)<sub>4</sub>]PF<sub>6</sub> (281 µg, 753 nmol) were dissolved in 100 µL of CD<sub>2</sub>Cl<sub>2</sub> to furnish the complex **ROT-1** in quantitative yield. Now, benzyl azide (200 µg, 1.50 µmol) and 1 µL of Et<sub>3</sub>N were added. Further 300 µL of CD<sub>2</sub>Cl<sub>2</sub> was added. The NMR tube was tightly closed and heated at 40 °C for 24 h. Then all the solvent was evaporated to remove residual Et<sub>3</sub>N. The residue was redissolved in 550 µL of CD<sub>2</sub>Cl<sub>2</sub> to furnish **ROT-2** in quantitative yield. **mp:** > 250 °C. <sup>1</sup>**H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 400 MHz):**  $\delta = -4.39$  (bs, 12H, DABCO-H), 1.26 (bs, 12H, h-H), 1.47 (bs, 18H, 15'-H+18'-H), 2.03 (s, 6H, 1'-H), 2.08 (s, 6H, 10'-H), 2.15 (s, 3H, 11'-H), 2.58 (s, 15H, 16'-H+19'+2'-H), 2.59 (s, 6H, i-H), 4.81 (bs, 4H, e-H), 5.52 (s, 4H, j-H), 6.61 (bs, 2H, 9'-H), 6.78 (bs, 4H, d-H), 7.21 (bs, 12H, g-H+14'-H+17'-H+s-H), 7.28-7.30 (m, 6H, k-H+s-H), 7.41-7.48 (m, 8H, 1-H+m-H+s-H), 7.61 (d, <sup>3</sup>*J* = 8.0 Hz, 4H, c-H), 7.85 (d, <sup>3</sup>*J* = 7.8 Hz, 4H, s-H), 7.92-7.94 (m, 4H, 3'-H+8'-H+s-H), 8.21 (s, 2H, 5'-H+6'-H), 8.28 (bs, 2H, β(2)-H), 8.30 (bs, 2H, β(2)-H), 8.35-8.36 (m, 6H, β(2)-H+β(3')-H), 8.53-8.54 (m, 6H, β(2)-H+β(3')-H), 8.73 (d, <sup>3</sup>*J* = 8.0 Hz, 2H.

4'-H+7'-H) ppm. **ESI-MS:** *m/z* (%) 2655.9 (100) [Cu(**2**)(**3**')]<sup>+</sup>. **Elemental analysis:** Anal. Calcd for C<sub>178</sub>H<sub>152</sub>N<sub>18</sub>O<sub>2</sub>Zn<sub>2</sub>CuPF<sub>6</sub>•5CH<sub>2</sub>Cl<sub>2</sub>: C, 65.82; H, 4.89; N, 7.55. Found: C, 65.55; H, 4.64; N, 7.32.

# 2. NMR spectra: <sup>1</sup>H, <sup>13</sup>C, <sup>1</sup>H-<sup>1</sup>H COSY



Figure S1. <sup>1</sup>H NMR of compound 2 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).



Figure S2. <sup>1</sup>H NMR of compound 18 in CDCl<sub>3</sub> (500 MHz, 298 K).



Figure S3. <sup>1</sup>H NMR of compound 3 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).



Figure S4. <sup>1</sup>H-<sup>1</sup>H COSY NMR of compound 3 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).



Figure S5. <sup>13</sup>C NMR of compound 3 in CDCl<sub>3</sub> (100 MHz, 298 K).



Figure S6. <sup>1</sup>H NMR of complex C1 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K, 2.5 mM).



Figure S7. <sup>1</sup>H NMR of [Cu(4)]<sup>+</sup>, 5 and C1 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K, 2.5 mM).



Figure S8. <sup>1</sup>H NMR of ASB-1 in  $CD_2Cl_2$  (400 MHz, 298 K).



**Figure S9.** <sup>1</sup>H-<sup>1</sup>H COSY NMR of **ASB-1** in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).



Figure S10. <sup>1</sup>H NMR of 1,  $[Cu_2(1)]^{2+}$ , 3 and ASB-1 in  $CD_2Cl_2$  (400 MHz, 298 K).



Figure S11. <sup>1</sup>H NMR of ROT-1 in CD<sub>2</sub>Cl<sub>2</sub> (600 MHz, 298 K).

![](_page_18_Figure_2.jpeg)

**Figure S12.** <sup>1</sup>H-<sup>1</sup>H COSY NMR of **ROT-1** in CD<sub>2</sub>Cl<sub>2</sub> (600 MHz, 298 K).

![](_page_19_Figure_0.jpeg)

Figure S13. <sup>1</sup>H NMR of 2, [Cu(2)]<sup>+</sup>, 3 and ROT-1 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).

![](_page_20_Figure_0.jpeg)

Figure S14. <sup>1</sup>H NMR of 3, ROT-1 and ASB-1 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).

![](_page_21_Figure_0.jpeg)

Figure S15. <sup>1</sup>H NMR of ROT-2 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).

![](_page_21_Figure_2.jpeg)

Figure S16. <sup>1</sup>H-<sup>1</sup>H COSY NMR of ROT-2 in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K).

![](_page_22_Figure_0.jpeg)

Figure S17. <sup>1</sup>H NMR of ROT-1 and ROT-2 in  $CD_2Cl_2$  (400 MHz, 298 K).

### 3. Variable temperature <sup>1</sup>H NMR spectra

The kinetics of rotational exchange at various temperatures was analyzed using the program WinDNMR through simulation of the experimental <sup>1</sup>H NMR spectra.<sup>10</sup> The spectra simulation was performed using the model of a 2-spin system undergoing mutual exchange and provided the rate constants. Activation parameters were determined from an Eyring plot.

![](_page_23_Figure_2.jpeg)

**Figure S18.** (a) VT <sup>1</sup>H-NMR (600 MHz) of **ROT-1** in  $CD_2Cl_2$  shows the splitting of proton signal e-H in 1:1 ratio. The corresponding rate constant at different temperatures was calculated from the simulation. (b) Eyring plot for exchange frequency in nanorotor **ROT-1**.

![](_page_23_Figure_4.jpeg)

**Figure S19.** (a) VT <sup>1</sup>H-NMR (600 MHz) of **ROT-2** in  $CD_2Cl_2$  shows the splitting of proton signal j-H in 1:1 ratio. The corresponding rate constant at different temperatures was calculated from the simulation. (b) Eyring plot for exchange frequency in nanorotor **ROT-2**.

### 4. DOSY NMR spectra

Calculation of hydrodynamic radius from DOSY

The diffusion coefficient D of each assembly was obtained from the DOSY spectrum, and the corresponding hydrodynamic radius was calculated by using the Stokes-Einstein equation:

 $r = k_B T / 6 \pi \eta D$ 

![](_page_24_Figure_4.jpeg)

**Figure S20.** DOSY NMR of **ASB-1** in CD<sub>2</sub>Cl<sub>2</sub> (600 MHz, 298 K). Diffusion coefficient  $D = 4.8 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$ , hydrodynamic radius r = 11.4 Å.

![](_page_24_Figure_6.jpeg)

**Figure S21.** DOSY NMR of **ROT-1** in CD<sub>2</sub>Cl<sub>2</sub> (600 MHz, 298 K). Diffusion coefficient  $D = 4.8 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$ , hydrodynamic radius r = 11.4 Å.

![](_page_25_Figure_0.jpeg)

**Figure S22.** DOSY NMR of **ROT-2** in CD<sub>2</sub>Cl<sub>2</sub> (600 MHz, 298 K). Diffusion coefficient  $D = 5.0 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$ , hydrodynamic radius r = 10.9 Å.

# 5. ESI-MS spectra

![](_page_26_Figure_1.jpeg)

Figure S23. ESI-MS of  $[Cu_2(1)(3)(DABCO)]^{2+}$ .

![](_page_26_Figure_3.jpeg)

**Figure S24.** ESI-MS of [Cu(2)(3)]<sup>+</sup>.

![](_page_27_Figure_0.jpeg)

**Figure S25.** ESI-MS of [Cu(2)(3')]<sup>+</sup>.

### 6. Binding constant determination

### a. Measurement of binding constant of compound 5 to [Cu(4)]<sup>+</sup>

To determine the binding constant of alkyne **5** to  $[Cu(4)]^+$ , an NMR titration was performed. A 3.13 mM solution of  $[Cu(4)]^+$  was prepared in CD<sub>2</sub>Cl<sub>2</sub> directly in an NMR tube. In another vial, a 62.2 mM stock solution of **5** was prepared CDCl<sub>3</sub>. An aliquot of 3.0 µL of **5** was added. After each addition, the <sup>1</sup>H NMR was recorded and the peak at 8.67 ppm (for 4"-H) was monitored for data analysis. The binding constant was determined using a nonlinear curve-fitting applying the following equation<sup>11</sup>:

 $Y = Y0 + DY^{*}((K^{*}(P+x)+1) - SQRT(((K^{*}(P+x)+1)^{2}) - 4^{*}K^{*}K^{*}P^{*}x))/(2^{*}K^{*}P)$ 

Y = Measured Chemical shift; Y0 = Chemical shift of empty host solution; DY = Maximum change in chemical shift: the difference in chemical shift of a fully occupied host and an empty host; K = Binding constant; P = Total host concentration; x = Total guest concentration.

![](_page_28_Figure_5.jpeg)

**Figure S26.** <sup>1</sup>H NMR of  $[Cu(4)]^+$  upon successive addition of compound **5** in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K). (b) Curve-fitting for determination of the binding constant of **5** to  $[Cu(4)]^+$ .

#### b. Measurement of binding constant of compound 6 to [Cu(4)]<sup>+</sup>

To determine the binding constant of triazole **6** to  $[Cu(4)]^+$ , an NMR titration was performed. A 3.13 mM solution of  $[Cu(4)]^+$  was prepared in CD<sub>2</sub>Cl<sub>2</sub> directly in an NMR tube. In another vial, a 32.3 mM stock solution of **6** was prepared in CD<sub>2</sub>Cl<sub>2</sub>. An aliquot of 4.82 µL of **6** was added. After each addition, the <sup>1</sup>H NMR was recorded and the peak at 7.01 ppm (for 6"-H) was monitored for data analysis. As before in chapter 6a, the binding constant was determined using non-linear curve-fitting.

![](_page_29_Figure_2.jpeg)

**Figure S27.** <sup>1</sup>H NMR of  $[Cu(4)]^+$  upon successive addition of compound **6** in CD<sub>2</sub>Cl<sub>2</sub> (400 MHz, 298 K). (b) Curve-fitting for determination of the binding constant of **6** to  $[Cu(4)]^+$ .

# 7. X-ray crystallography

![](_page_30_Figure_1.jpeg)

**Figure S28.** X-Ray crystal structure of complex **C1**. Carbon are shown in light grey; H, light green; N, blue; O, red; Cu<sup>+</sup>, indigo and I, violet.

Data were collected on a STOE IPDS II two-circle diffractometer with a Genix Microfocus tube with mirror optics using Mo $K_{\alpha}$  radiation ( $\lambda = 0.71073$  Å). The data were scaled using the frame scaling procedure in the *X*-AREA program system.<sup>12</sup> The structure was solved by direct methods using the program *SHELXS*<sup>13</sup> and refined against  $F^2$  with full-matrix least-squares techniques using the program *SHELXL*.<sup>13</sup> The F ligands of the PF<sub>6</sub> anion are disordered over two positions with a site occupation factor of 0.510(17) for the major occupied orientation. The displacement ellipsoids of the disordered F atoms were restrained to an isotropic behaviour. The contribution of the unidentifiable solvent was suppressed using the *SQUEEZE* routine in *PLATON*.<sup>14</sup> CCDC deposition number: 2199510.

 Table S1. Crystal data and structure refinement for complex C1.

Identification code	C1	
Empirical formula	C39 H35 Cu F6 I N2 O P	
Formula weight	883.10	
Temperature	173(2) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	P -1	
Unit cell dimensions	a = 11.2316(6) Å	a= 91.434(4)°.
	b = 12.5683(7) Å	b= 103.968(4)°.
	c = 14.3333(7) Å	$g = 100.004(4)^{\circ}$ .
Volume	1928.89(18) Å <sup>3</sup>	
Z	2	
Density (calculated)	1.520 Mg/m <sup>3</sup>	
Absorption coefficient	1.469 mm <sup>-1</sup>	
F(000)	884	
Crystal size	0.120 x 0.110 x 0.020 mm	1 <sup>3</sup>
Theta range for data collection	3.266 to 25.027°.	
Index ranges	-13<=h<=13, -14<=k<=14	4, -17<=l<=17
Reflections collected	29992	
Independent reflections	6784 [R(int) = 0.0588]	
Completeness to theta = $25.000^{\circ}$	99.8 %	
Absorption correction	Semi-empirical from equi	valents
Max. and min. transmission	1.000 and 0.609	
Refinement method	Full-matrix least-squares	on $F^2$
Data / restraints / parameters	6784 / 93 / 525	
Goodness-of-fit on F <sup>2</sup>	1.036	
Final R indices [I>2sigma(I)]	$R1 = 0.0549, wR2 = 0.14^{\circ}$	78
R indices (all data)	R1 = 0.0646, wR2 = 0.159	94
Extinction coefficient	n/a	
Largest diff. peak and hole	0.630 and -1.254 e.Å <sup>-3</sup>	

	х	у	Z	U(eq)
I(1)	-388(1)	3709(1)	-994(1)	58(1)
Cu(1)	6284(1)	1796(1)	3159(1)	29(1)
O(1)	5252(3)	3574(3)	638(2)	40(1)
N(1)	7166(3)	2684(3)	4400(2)	27(1)
C(2)	7896(3)	2103(3)	4996(3)	29(1)
C(3)	8730(3)	2542(3)	5869(3)	30(1)
C(4)	8779(4)	3646(4)	6129(3)	34(1)
C(5)	8006(4)	4213(4)	5541(3)	34(1)
C(6)	7184(4)	3724(3)	4678(3)	28(1)
C(7)	9450(4)	1854(4)	6460(3)	35(1)
C(8)	9337(4)	796(4)	6202(3)	35(1)
N(11)	6912(3)	578(3)	3879(2)	27(1)
C(12)	7768(3)	973(3)	4716(3)	27(1)
C(13)	8478(4)	317(4)	5313(3)	32(1)
C(14)	8279(4)	-784(4)	5017(3)	36(1)
C(15)	7409(4)	-1177(4)	4173(3)	38(1)
C(16)	6729(4)	-482(3)	3612(3)	31(1)
C(21)	6280(4)	4321(3)	4083(3)	30(1)
C(22)	6700(4)	5284(3)	3683(3)	34(1)
C(23)	5824(4)	5821(3)	3137(3)	36(1)
C(24)	4544(4)	5441(4)	2972(3)	39(1)
C(25)	4146(4)	4515(4)	3393(3)	36(1)
C(26)	4983(4)	3941(3)	3957(3)	32(1)
C(27)	8078(4)	5709(4)	3802(4)	41(1)
C(28)	3611(5)	6020(5)	2335(4)	54(1)
C(29)	4486(4)	2976(4)	4433(4)	40(1)
C(31)	5764(4)	-869(3)	2702(3)	29(1)
C(32)	4498(4)	-973(3)	2698(3)	33(1)
C(33)	3601(4)	-1250(4)	1824(3)	37(1)
C(34)	3931(4)	-1432(4)	963(3)	38(1)
C(35)	5185(4)	-1338(4)	990(3)	38(1)

**Table S2**. Atomic coordinates (x 10<sup>4</sup>) and equivalent isotropic displacement parameters ( $Å^2x$  10<sup>3</sup>) for ms3. U(eq) is defined as one third of the trace of the orthogonalized U<sup>ij</sup> tensor.

C(36)	6114(4)	-1065(3)	1844(3)	34(1)
C(37)	4093(5)	-769(5)	3599(4)	46(1)
C(38)	2940(5)	-1707(5)	23(4)	52(1)
C(39)	7470(4)	-931(5)	1825(4)	48(1)
C(41)	5251(5)	1412(4)	1845(3)	40(1)
C(42)	5570(4)	2391(4)	1935(3)	34(1)
C(43)	5782(5)	3518(4)	1649(3)	39(1)
C(51)	3996(4)	3587(3)	346(3)	37(1)
C(52)	3586(5)	3849(4)	-606(3)	44(1)
C(53)	2339(5)	3892(4)	-972(4)	49(1)
C(54)	1514(5)	3657(4)	-410(4)	44(1)
C(55)	1907(5)	3405(4)	532(4)	46(1)
C(56)	3155(5)	3360(4)	904(3)	45(1)
P(1)	9862(1)	2850(1)	2910(1)	50(1)
F(1)	10362(17)	3001(11)	4029(6)	103(5)
F(2)	8783(17)	3428(18)	2829(15)	158(9)
F(3)	9508(16)	2565(15)	1799(7)	74(5)
F(4)	9063(19)	1753(11)	3058(11)	144(9)
F(5)	10754(12)	3887(10)	2741(9)	77(4)
F(6)	11065(12)	2261(15)	2994(13)	121(6)
F(1')	9770(10)	3417(10)	3932(7)	75(3)
F(2')	8420(9)	2856(15)	2600(9)	98(5)
F(3')	9828(18)	2459(16)	1896(8)	100(7)
F(4')	9518(12)	1695(8)	3238(11)	100(5)
F(5')	10170(20)	4056(10)	2660(14)	149(10)
F(6')	11255(7)	2958(16)	3401(12)	148(7)

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