New Bifunctional Monomers from Methyl Vinyl Glycolate.

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Supporting Information

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1. General information

Reagents: Methyl vinylglycolate can be obtained from Sigma-Aldrich. For larger quantities it can be convenient to use a high yielding synthesis from (E)-4-iodobut-2- enoate,¹ which itself is easily prepared from the commercially available bromide. All the other reagents and solvents were obtained from commercial sources and used as received unless noted otherwise. Dry solvents were obtained from a solvent purification system or purchased water-free in a bottle with septum. All the reagents and solvents were handled in oven-dried glassware using standard Schlenk techniques, unless otherwise stated.

<u>MMR-Spectroscopy</u>¹H-NMR and ¹³C-NMR were recorded at ambient temperature on 300 MHz spectrometers (Avance 300 or Fourier 300) or a 400 MHz spectrometer (Avance 400) from Bruker. The chemical shifts δ are given in ppm and referenced to the residual proton signal of the deuterated solvent used.

Gas Chromatography (GC): GC analysis was carried out on an Agilent 7890B GC system with a HP-5 normal-phase silica column, using He as a carrier gas and dodecane as internal standard.

Gel permeation chromatography (GPC): Gel permeation chromatograms were recorded with 1260 Infinity GPC/SEC System from Agilent Technologies. The setup consisted of a SECcurity Isocratic Pump, SECcurity 2-Canal-Inline-Degaser, SECcurity GPC-Column thermostat TCC6000, SECcurity Fraction Collector and SECcurity Differential Refractometer detector. The measurements were performed at a constant temperature of 50 °C using three columns with a polyester co-polymer network as the stationary phase (PSS GRAM 30 Å, 10 µm particle size, 8.0 × 300 mm; PSS GRAM 30 Å, 10 µm particle size, 8.0 × 300 mm). THF was applied as the mobile phase with a flow rate of 1 mL·min⁻¹. Polystyrene standards from ReadyCal (PSS-pskitr1I-10, $M_p = 370-2520000$ g·mol⁻¹) were used for calibration purposes.

Differential scanning calorimetry (DSC): Melting points and glass transition temperatures of polyesters were measured with a *Star-SW DSC* from *Mettler Toledo* using the following temperature program: -90.00 °C isothermal 5.00 min; Ramp 10.00 °C min⁻¹ to 150.00 °C; Ramp 10.00 °C/min to -90.00 °C; -90.00 °C isothermal 5.00 min; Ramp 10.00 °C min to 150.00 °C; Ramp 10.00 °C/min to -90.00 °C.

Infrared Spectroscopy: ATR-IR measurements were recorded on a Nicolet iS5 FT-IR (ThermoFisher) device calibrated on 1.5 mil polystyrene and equipped with a GladiATR 210 accessory from PIKE technologies.

<u>Mass spectroscopy (ESI-MS)</u>: measurements were recorded on an Agilent 6210 time-of-flight LC/MS (ESI) or on a Thermo Electron MAT 95-XP (EI, 70 eV). Peaks as listed correspond to the highest abundant peak and are of the expected isotope pattern.

2. Experimental procedures

Hydroformylation of MVG: In a glovebox, dicarbonyl(acetylacetonato)rhodium(I) (1.2 mg, 0.005 mmol, 0.5 mol%) and the desired ligand (0.095 mmol, 0.1 eq) were weighted into a vial. The vial was sealed, equipped with a magnetic stirrer and transferred out of the glovebox. The desired solvent (toluene or THF, 1.35 mL, 0.7 mol/L with respect to MVG) and MVG (110 mg, 0.95 mmol, 1 eq) were added under argon atmosphere. The vials were placed into a 300 mL Parr stainless steel autoclave and pierced with a needle. The autoclave was flushed three times with nitrogen, then pressurized with 10 bar of syngas and heated to 80 °C. After stirring overnight, the reaction was cooled down to room temperature, the crude mixtures filtered over celite, and volatiles evaporated under reduced pressure. The crude residue was dissolved in CDCl₃ and analysed by ¹H NMR and GC-MS. Linear product (1) ¹H NMR (300 MHz, CDCl₃) δ 9.77 (t, J = 1.2 Hz, 1H), 4.23 (dd, J = 7.8, 4.2 Hz, 1H), 3.77 (s, 3H), 2.65 – 2.55 (m, 2H), 2.48 – 2.36 (m, 2H). Branched product (2), mixture of 2 diasteroisomers: ¹H NMR (300 MHz, CDCl₃) δ 9.73 (d, J = 0.7 Hz, 1H), 9.65 (s, 1H), 4.76 (d, J = 3.0 Hz, 1H), 4.42 (d, J = 3.9 Hz, 1H), 3.77 (s, 3H), 3.75 (s, 3H), 2.54 – 2.37 (m, 1H), 2.37 – 2.12 (m, 1H), 1.26 (d, J = 7.3 Hz, 3H), 1.12 (d, J = 7.2 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 201.9, 201.7, 174.9, 70.9, 69.6, 53.1, 53.0, 49.8, 49.6, 9.9, 7.6. Hydroxyacetal (3), mixture of 2 diasteroisomers: ¹H NMR (300 MHz, CDCl₃) δ 5.75 – 5.71 (m, 1H), 5.60 (t, J = 3.2 Hz, 1H), 4.71 (dd, J = 8.6, 4.2 Hz, 1H), 4.55 (dd, J = 8.2, 7.1 Hz, 1H), 3.75 (s, 3H), 3.72 (s, 3H), 2.11 – 1.82 (m, 6H). ¹³C NMR (75 MHz, CDCl₃) δ 174.8, 173.2, 100.2, 99.7, 77.2, 76.3, 52.5, 52.3, 33.7, 32.2, 28.1, 28.0.

Methylation of MVG: In a 250 mL Schlenk flask Ag₂O (19.6 g, 84.4 mmol, 2 eq) was suspended in 60 mL of diethyl ether (0.7 mol/L with respect to MVG). MVG (4.90 g, 42.2 mmol, 1 eq) was added under argon atmosphere. To the stirred suspension, methyl iodide (13.0 g, 84.4 mmol, 2 eq) was slowly added via syringe. The reaction was stirred at room temperature while monitoring the conversion by GC. After 60 hours, MVG was fully converted. The reaction was filtered to remove the solids, then the solvent and the excess of methylating agent removed by carefully distilling under vacuum, affording a colourless liquid (5.51 g, quantitative yield). The analytical data corresponds to the known literature.²

¹H NMR (300 MHz, CDCl₃) δ 5.85 (ddd, *J* = 17.2, 10.4, 6.4 Hz, 1H), 5.46 (dt, *J* = 17.2, 1.3 Hz, 1H), 5.34 (dt, *J* = 10.4, 1.3 Hz, 1H), 4.26 (dt, *J* = 6.4, 1.3 Hz, 1H), 3.76 (s, 3H), 3.40 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 171.1, 132.7, 119.6, 81.7, 57.5, 52.4.

<u>Acetylation of MVG</u>: In a 50 mL Schlenk flask, MVG (550 mg, 4.74 mmol, 1 eq) and pyridine (750 mg, 9.47 mmol, 2 eq) were dissolved in dichloromethane (16 mL, 0.3 mol/L with respect to MVG). To the stirred mixture, acetic anhydride (967 mg, 9.47 mmol, 2 eq) was slowly added via syringe. The reaction was stirred at room temperature overnight, then poured into ice-cold water and extracted three times with DCM. The organic phase was washed with 1M HCl, water and brine, then dried over Na₂SO₄ and concentrated in vacuum, affording 746 mg of colourless liquid (quantitative yield). The analytical data corresponds to the known literature.³

¹H NMR (400 MHz, CDCl₃) δ 5.87 – 5.77 (m, 1H), 5.41 – 5.31 (m, 2H), 5.24 (ddd, *J* = 10.5, 1.4, 0.9 Hz, 1H), 3.63 (s, 3H), 2.04 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.7, 168.7, 129.9, 119.5, 72.8, 52.3, 20.3.

Methoxycarbonylation reactions: In a glovebox, palladium(II) diacetate (1.7 mg, 0.01 mmol, 1.0 mol%) and the desired ligand (0.02 mmol, 2.0 ml%) were weighted into a vial. The vial was sealed, equipped with a magnetic stirrer, and transferred out of the glovebox. Methanol (1.5 mL, 0.5 mol/L with respect to the substrate), methanesulfonic acid (2.2 mg, 0.02 mmol, 2 mol%) and the desired MVG derivative (see Table 2 main text; 0.80 mmol, 1 eq) were added under argon atmosphere. The vials were placed into a 300 mL Parr stainless steel autoclave and pierced with a needle. The autoclave was flushed three times with nitrogen, then pressurized with the desired pressure of carbon monoxide and heated to the required temperature. After stirring for the desired time, the reaction was cooled down to room temperature, the crude mixtures filtered over celite, and volatiles evaporated under reduced pressure. The crude was purified by flash column chromatography (gradient elution, from 100% *n*-hexane to 100% ethyl acetate), affording the diester as a yellowish liquid.

¹H NMR (300 MHz, CDCl₃) δ 3.82 (dd, *J* = 7.8, 4.7 Hz, 1H), 3.75 (s, 3H), 3.67 (s, 3H), 3.38 (s, 3H), 2.44 (ddd, *J* = 7.7, 7.0, 3.6 Hz, 2H), 2.18 – 1.92 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 173.4, 172.7, 79.3, 58.4, 52.1, 51.8, 29.6, 27.9. ESI-MS (ES⁺): calculated for C₈H₁₄O₅: 190.1932; found: 213.0737 [M-Na]⁺.

Polycondensation reactions: A 5 mL vial equipped with stirring bar was charged with the desired diol (2.0 mmol, 1 eq) and **8** (380 mg, 2.0 mmol, 1 eq). The starting materials were extensively dried via vacuum-argon cycles, then titanium(IV) isopropoxide (3.2 mg, 0.01 mmol, 1 mol%) was added via syringe and the reaction heated up to 150 °C. After stirring under argon

atmosphere for 6 hours, vacuum was applied for 1 hour. Viscosity visibly increased up to the point that the mixture wasn't stirred. Temperature was then increased to 190 °C and the reaction kept in vacuum another hour. After cooling down to room temperature, the solid products were analysed by NMR and GPC.

3. Screening of reaction conditions for the protection of the OH group of MVG

Methylation



| Entry | Conditions |
|-------|--|
| 1 | NaH, THF, 0 °C – r.t., 16 h |
| 2 | Ag ₂ O, Et ₂ O, r.t., 24 h |
| 3 | K ₂ CO ₃ , THF, 80 °C, 1 h |
| 4 | K ₂ CO ₃ , MeOH, 80 °C, 1 h |
| 5 | K ₂ CO ₃ , neat, 80 °C, 1 h |
| 6 | K ₂ CO ₃ , acetone, 80 °C, 1 h |
| 7 | NaOMe, MeOH, 80 °C, 1 h |
| 8 | DIPEA, Et ₂ O, r.t., 48 h |

Outcome

C=C isomerization 6 (49 % after FCC)

MeOMVG dimerization

C=C isomerization C=C isomerization

Acetylation



| Entry | Conditions |
|-------|--------------------------|
| 1 | Et₃N, DCM, r.t., 24 h |
| 2 | iPr2EtN, DCM, r.t., 24 h |
| 3 | Pyridine, r.t., 16 h |

Outcome C=C isomerization C=C isomerization 7 (>99%)



4. Mechanisms involved in the Pd-catalysed methoxycarbonylation

5. Characterization of products

NMR Spectra

- Representative spectra in CDCl₃ of the crude mixture after hydroformylation of MVG using PPh₃ as ligand (Table 1 main text, entry 1 and 2):





- Representative spectra in CDCl₃ of the high linear containing mixture after hydroformylation of MVG using Xantphos as ligand (Table 1 main text, entry 3 and 4):







220720.403.12.fid Andrea Dell'Acqua ADE 783-1-KR Au13C-dept CDCl3 {C:\Bruker\TopSpin3.5pl6} 2207 3





- NMR spectra of isolated products:













200325.f351.10.fid Dell'Acqua / ADE345 C13CPD CDCl3 {C:\Bruker\TopSpin3.6.0} 2003 51



200331.423.10.fid Dell'Acqua/ ADE 353 Au13C CDCl3 {C:\Bruker\TopSpin3.5pl6} 2003 23







220624.f344.11.fid Dell'Acqua/ ADE 776 C13CPD CDCl3 {C:\Bruker\TopSpin3.6.2} 2206 44



220506.f364.11.fid Andrea Dell'Acqua ADE 773 C13CPD CDCl3 {C:\Bruker\TopSpin3.6.2} 2205 4





GC-MS







ESI-MS Chromatograms



<u>ATR-IR</u>







GPC Chromatograms













<u>TGA</u>









6. References

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