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Supporting Information

One-Bond 13C-13C Spin-Coupling Constants in Saccharides: A Comparison of Experimental and Calculated Values By Density Functional Theory Using Solid-State ¹³C NMR and X-Ray Crystallography

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Synthesis of 13C-Labeled Compounds 1–5

Starting materials and reagents were commercial grade and were used without further purification. 13C-Labeled D-aldohexoses were obtained from Omicron Biochemicals, Inc. (South Bend, IN) and used without further purification. All reactions were monitored by thin-layer

was measured in the two compounds ¹³C-labeled at C5 and C6.

chromatography (TLC) on aluminum sheets coated with Silica Gel 60 (F_{254} , 250 μ m thickness). Spots were detected with a cerium molybdate stain.¹ Column chromatography was conducted using Dowex 1 x 8 (200–400 mesh) anion-exchange resin (200–400 mesh) in the OH⁻ form.² High-resolution mass spectrometry (HRMS) data were obtained on a Bruker micrOTOF-QII quadrupole time-of-flight (QTOF) mass spectrometer operating in the ESI mode.

1.1. General Procedure To Prepare 13C-labeled 1–5 (Schemes S1 and S2). Dowex 50 x 8 (200-400 mesh) cation-exchange resin in the H⁺ form (250 mg; dried at >100 °C under high vacuum for 2 h), the ¹³C-labeled D-aldohexose (250 mg, 1.39 mmol) was added to MeOH (10 mL), and the mixture was refluxed overnight with stirring. After cooling to RT, the resin was removed by filtration and the filtrate was concentrated at 30° *in vacuo*. The crude residue was dissolved in a minimal volume of distilled water, and the solution was applied to a chromatography column (2.5 cm x 100 cm) containing Dowex 1 x 8 (200–400 mesh) anion-exchange resin in the OH⁻ form.² The column was eluted with decarbonated, distilled water and fractions (10 mL) were assayed by TLC analysis using a cerium molybdate stain1 and charring with a heat gun. For **1**–**4** (Glc and Gal), the α -anomer eluted first, followed by the β -anomer. For **5** (Man), the β -anomer eluted first, followed by the α -anomer. Yields: **1**^{1,2}, 45.6% as a white solid; **2**^{1,2}, 22.4% as a white solid; **1**5,6, 46.3% as a white solid; **2**5,6, 29.4% as a white solid; **3**1,2, 43.0% as a white solid; **4**1,2,

17.6% as a white solid; **5**1,2, 60.3% as a flaky white solid. 1H and 13C{1H} NMR spectra of isolated products matched those in the literature,^{3–7} except for the additional line-splitting from *J*_{CH} to the $13C$ -labeled carbons (¹H spectra), and relative line intensities (strong signals from the $13C$ -labeled carbons) and additional line-splitting from $J_{\rm CC}$ to the ¹³C-labeled carbons (¹³C spectra).

For 13C-labeled D-mannose: 100 mg (0**.**56 mmol), H+ resin (100 mg), MeOH (5 mL).

High-Resolution Mass Spectrometry (HRMS) Data for Compounds **1**–**5**

Crystallization and X-ray Crystallography of 1–5

All compounds **(1**1,2, **1**5,6, **2**1,2, **2**5,6, **3**1,2, **4**1,2, **5**1,2) were dissolved in a concentrated solution of **~**95:5 *v*/*v* methanol:water with heat and crystallized at RT by slow evaporation. Crystal structures were determined at 120 K (-173 °C) with monochromated copper K_{α} radiation to a resolution of 0.8 Å. All atoms were refined freely including hydrogen. The final structures were virtually identical to those reported previously;^{4,8–15} the presence of ¹³C atoms did not alter the structures to any appreciable extent.

The structures gave the following torsion angles involving hydroxyl hydrogens:

The values in parentheses are standard uncertainties (SU) of the average torsion angle. The values in brackets are the average torsion angle with range calculated as 3 x SU (99% confidence interval). Cartesian coordinates for the seven structures were deposited in the Cambridge Structural Database and can be accessed using the following deposition numbers:

- **1**1,2: Deposition Number 2252830
- **1**5,6: Deposition Number 2252831
- **2**1,2: Deposition Number 2252833
- **2**5,6: Deposition Number 2252828
- **3**1,2: Deposition Number 2252829
- **4**1,2: Deposition Number 2252827
- **5**1,2: Deposition Number 2252832

Solution 1H and 13C{1H} NMR Spectra of 11,2–51,2, 15,6 and 25,6

¹H NMR spectrum of methyl α -D-[1,2-¹³C₂]glucopyranoside (1^{1,2})

¹H NMR spectrum of methyl β -D-[1,2-¹³C₂]glucopyranoside (2^{1,2})

¹H NMR spectrum of methyl β -D-[5,6-¹³C₂]glucopyranoside (2^{5,6})

¹H NMR spectrum of methyl α -D-[1,2-¹³C₂]galactopyranoside (3^{1,2})

¹³C{¹H} NMR spectrum of methyl α -D-[1,2-¹³C₂]galactopyranoside (3^{1,2})

¹³C{¹H} NMR spectrum of methyl β -D-[1,2-¹³C₂]galactopyranoside (4^{1,2})

¹H NMR spectrum of methyl α -D-[1,2-¹³C₂]mannopyranoside (5^{1,2})

¹³C{¹H} NMR spectrum of methyl α -D-[1,2-¹³C₂]mannopyranoside (5^{1,2})

Figure S1. Partial 13C[1H} NMR spectrum of **1**1,2, showing the natural abundance carbons. Insets are expanded signals from C3, C4, C5, C6 and OCH $_3$ showing their multiplicities due to ¹³C-¹³C spin-coupling to the labeled carbons and the assigned $J_{\rm CC}$ values. Signs of the $^2J_{\rm CC}$ values were taken from references 16 and 17.

Figure S2. Partial 13C[1H} NMR spectrum of **1**5,6, showing the natural abundance carbons. Insets are expanded signals from C3, C4, and C5 showing their multiplicities due to 13C- ¹³C spin-coupling to the labeled carbons and the assigned $J_{\rm CC}$ values. Signs of the ² $J_{\rm CC}$ were taken from references 16 and 17. The OCH₃ signal was a sharp singlet, and the C2 signal (not shown) was a slightly broadened singlet.

Figure S3. Solid-state ¹³C NMR spectra of 1^{1,2} (A), $2^{1,2}$ (B), $1^{5,6}$ (C), $2^{5,6}$ (D), $3^{1,2}$ (E), $4^{1,2}$ (F) and $5^{1,2}$ (G) showing only the signals from the two ¹³C-labeled carbons. Figure S3. Solid-state 13C NMR spectra of **1**1,2 (A), **2**1,2 (B), **1**5,6 (C), **2**5,6 (D), **3**1,2 (E), **4**1,2 (F) and **5**1,2 (G) showing only the signals from the two 13C-labeled carbons.

Figure S4. (A) Plot of calculated *r*_{C1,C2} as a function of the C1–C2–O2–H torsion angle q2. Black filled, **1**c. Black open, **2**c. Blue filled, **3**c. Blue open, **4**c. Green filled, **5**c. (B) Plots of *r*C5,C6 as a function of the C4–C5–C6–O6 torsion angle ω . Black filled, 1^c. Blue filled, 2^c. The three staggered rotamers *gg*, *gt* and *tg* are shown associated with their corresponding values of ω .

	$1J_{\rm CC}$	detected spin	J-coupling	standard		average
compound			(Hz)	error	R^2	(STDEV)
				(Hz)		(Hz)
α -GlcOMe $(1^{1,2})$	$1J_{C1,C2}$	C1	48.07	0.10	0.986	
			47.97	0.10	0.988	
			47.97	0.11	0.985	48.00
		C ₂	48.10	0.11	0.985	(0.08)
			47.85	0.10	0.987	
			48.03	0.10	0.986	
β-GlcOMe $(2^{1,2})$	$1J_{C1,C2}$	C1	48.57	0.09	0.989	
			48.48	0.09	0.988	
			48.66	0.09	0.989	48.57
		C2	48.64	0.09	0.990	(0.09)
			48.42	0.10	0.987	
			48.63	0.10	0.987	
α -GalOMe $(3^{1,2})$	$1J_{C1,C2}$	C ₁	48.74	0.10	0.985	
			49.01	0.11	0.983	
			48.84	0.11	0.983	48.82
			48.80	0.10	0.986	(0.09)
		C2	48.76	0.10	0.988	
			48.74	0.10	0.988	
	$1J_{C1, C2}$	C1	49.12	0.09	0.988	
β-GalOMe (4 ^{1,2})			49.08	0.09	0.990	
			48.91	0.09	0.989	49.07
		C ₂	49.05	0.02	0.991	(0.09)
			49.20	0.09	0.988	
			49.04	0.08	0.992	
α -ManOMe $(5^{1,2})$	$1J_{C1, C2}$	C1	49.78	0.14	0.970	
			49.48	0.14	0.972	
			48.98	0.12	0.979	49.42
		C ₂	49.70	0.14	0.972	(0.27)
			49.35	0.14	0.974	
			49.24	0.12	0.981	
	$1J_{C5,C6}$	C5	43.36	0.11	0.993	
			43.10	0.10	0.994	
α -GlcOMe $(1^{5,6})$			43.18	0.10	0.994	43.16
		C6	43.21	0.11	0.992	(0.12)
			43.45	0.11	0.992	
			43.21	0.11	0.992	
β -GlcOMe $(2^{5,6})$	1J _{C5, C6}	C5	44.95	0.11	0.990	
			44.90	0.10	0.991	
			44.93	0.10	0.991	44.82
		C6	44.68	0.10	0.993	(0.11)
			44.73	0.11	0.990	
			44.73	0.10	0.992	

Table S1. Fitting Statistics from Solid-state ¹³C NMR Measurements of ¹*J*CC values in Crystalline **1**1,2**, 1**5,6**, 2**1,2**, 2**5,6**, 3**1,2**, 4**1,2**,** and **5**1,2.

Figure S5. Plots of S/S_o vs τ to determine ${}^1J_{C1,C2}$ or ${}^1J_{C5,C6}$ in $1{}^{1,2}$ (A and B), $2{}^{1,2}$ (C and D), **3**1,2 (E and F), **4**1,2 (G and H), **5**1,2 (I and J), **1**5,6 (K and L) and **2**5,6 (M and N). Normalized intensities of the *J*-modulated echo signals (*S*; red) and the reference echo signals (*So*; black) are plotted against the total echo interval, τ , in A, C, E, G, I, K and M. Intensity ratios, (S/S_o) , are plotted against τ in B, D, F, H, J, L and N. The solid black curves represent the best fits to the equation given in references 61 and 62 of the main text. All experiments were run in triplicate, and only one representative signal in the 13C NMR spectra (Figure S3) is shown for each compound.

Discussion of Equation [1] For *J***-Coupling Equation Parameterization**

The generalized form of the Karplus-like equation that was used to parameterize the dependence of *J*-couplings on a specific torsion angle was first described by Pachler (K. G. R. Pachler, Extended Hückel Theory MO Calculations of Proton-Proton Coupling Constants–II: The Effect of Substituents on Vicinal Couplings in Monosubstituted Ethanes, *Tetrahedron* 1971, *27*, 187–199). He proposed the use of this trigonometric function to account for asymmetry in the Karplus curve caused by substitution of a hydrogen atom in the coupling pathway. We adopted this trigonometric polynomial form because it provides the best parameterization to the DFT data with the smallest number of terms. This form of the equation is also amenable to simple integration which allowed us to develop the *MA'AT* equation for modeling torsional populations.

Additional DFT Calculations

In addition to the hydrogen atom optimization discussed in the main text, three additional sets of hydrogen atom optimizations were conducted. These calculations were performed to evaluate the effects of the initial geometry and hydrogen bonding on the optimization. In the first set of calculations (Set 3b), the optimizations were conducted as described in the main text except the input Cartesian coordinates were obtained from HAR structures, as described in the Experimental section and used for calculating 1_{OC} values in Set 2. The two remaining sets of calculations aimed to examine the effects of hydrogen bonding on optimization. In these calculations (Sets 3c and 3d), methanol molecules surrounding a single saccharide molecule were included to mimic the hydrogen bonding observed in the crystal (Set 1) and the refined (Set 2) structures, respectively. *J*-Couplings were calculated in each optimized structure as described in the main text. For Sets 3c (from X-ray) and 3d (from HAR), the hydrogen bonding was removed before calculating the ¹J_{CC} values. For Sets 3e (from X-ray) and 3f (from HAR) the hydrogen bonding was retained for the calculations of the ¹J_{CC} values. Sets 3c and 3e have identical geometries for the *J*-coupling calculations except that Set 3e includes the hydrogen bonding. The same applies to Sets 3d and 3f. The calculated 1_{OCC} values are shown in Table S2.

Input geometry		X-ray		HAR		
Packing included in geometry optimization	no	yes	yes	no	yes	yes
Packing included in J- coupling calculation	no	no	yes	no	no	yes
	Set 3	Set 3b	Set 3c	Set 3d	Set 3e	Set 3f
$1^{1,2}$ (α Glc)	49.3	49.2	49.4	48.6	48.9	49.0
$2^{1,2}$ (β Glc)	51.0	51.3	51.0	51.1	51.3	51.0
$3^{1,2}$ (α Gal)	50.5	50.4	50.4	50.4	50.3	50.3
$4^{1,2}$ (β Gal)	50.8	51.4	51.0	50.7	51.5	51.1
$5^{1,2}$ (α Man)	52.2	52.8	51.6	51.9	52.5	51.3
$1^{5,6}$ (α Glc)	45.7	45.6	46.1	45.9	46.0	46.5
$2^{5,6}$ (β Glc)	44.3	44.3	43.8	44.1	44.5	44.1

Table S2. Calculated ${}^{1}J_{\text{CC}}$ Values (In Hz) in $1^{1,2}$ -5^{1,2}, 1^{5,6} and 2^{5,6} Obtained from Different DFT Geometry Optimization and J-Coupling Calculations

In general, different starting structures and/or including hydrogen bonding during geometry optimizations and/or in the ¹J_{CC} calculations had little effect on the calculated ¹J_{CC} values. In all cases but **1**, including hydrogen bonding in the *J*-coupling calculations lowered the calculated ¹J_{CC} values by 0.0–1.2 Hz (Set 3b – Set 3c, and Set 3e – Set 3f), with an average difference of 0.24 Hz. Changing the input coordinates for geometry optimization resulted in minor changes in the calculated $1J_{\text{CC}}$ values (-0.2-0.7 Hz; Set 3 – Set 3d) with an average difference of 0.16 Hz. The average standard deviation of all methods was 0.28 Hz, indicating that neither the starting geometry nor hydrogen bonding has a significant impact on calculated ¹J_{CC} values.

Torsional Dependencies of 1*J***C5,C6 Values**

¹J_{C5,C6} values in aldohexopyranosyl rings depend strongly on both the ω (C4–C5–C6– O6) and θ_6 (C5–C6–O6–H) torsion angles. As discussed in prior work on $\frac{1}{\sqrt{C}}$ in vicinal diol fragments (*J. Am. Chem. Soc.* 2004, *126*, 15668–15685), it is possible to derive parameterized equations that include both angles and describe an hypersurface. However, due to the complexity of parameterizing a surface, the present work examined only the dependence on ω , since θ_6 is fixed at a known value in crystalline samples.

However, for the sake of completeness, the hypersurface was determined for **2** for comparison to results reported in the prior 2004 *JACS* study. In these DFT calculations, both ω and θ_6 in **2** were rotated in 15^o increments through 360^o, yielding 576 optimized structures. The geometry optimizations were conducted as described in the main text at the B3LYP/6-31G* level of theory, and ¹J_{C5}, C₆ values were calculated, again as described in the main text. The resulting hypersurface is shown in Figure S6.

Figure S6. Hypersurface showing the dual dependencies of ${}^1J_{C5,C6}$ in **2** on the ω (C4–C5–C6–O6) and θ_6 (C5–C6–O6– H) torsion angles. Maximal coupling is observed in the *tg* rotamer (ω = 300^o; O5 and O6 *anti*) when O6H is *anti* to C5 $(\theta_6=180^\circ)$.

X- and *Y*-axis 2D projections of the hypersurface are shown in Figure S7. These plots show the extent to which ¹J_{C5,C6} is affected by ω and θ_6 . Coupling is maximal when O5 and O6 are *anti* (ω = 300^o) (Plot A) and when C5 and O6<u>H</u> are *anti* (θ_6 = 180^o)(Plot B), a result consistent with those of prior studies of ¹J_{CC} in vicinal diols (see JACS reference above). The plane of symmetry of the hypersurface should allow an equation to be parameterized for ¹J_{C5,C6} containing two variables, ω and θ ₆.

Figure S7. Plots of *x*- and *y*axis projections of the data shown in Figure S6. (A) Plot of $1J_{\text{C5,C6}}$ *vs* ω , showing data scatter at discrete values of ω caused by θ_6 . (B) Plot of $1J_{\text{C5,C6}}$ *vs* θ_6 , showing data scatter at discrete values of θ_6 caused by ω .

Representative Cartesian Coordinates for DFT Structures 1c–5c

Structure **1**c

C 8.056 3.752 20.399

Structure **3**c

Structure **4**c

Structure **5**c

Complete Reference 50

M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, *Gaussian*16, Revision C.01, Gaussian, Inc., Wallingford CT, 2019.

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