Electronic Supplementary Information

Experimental Section

Materials: Copper nitrate trihydrate (Cu(NO₃)₂·3H₂O), ferric nitrate nonahydrate (Fe(NO₃)₂·9H₂O), N,N-Dimethylformamide (DMF), ammonium chloride (NH₄Cl), sulfuric acid (H₂SO₄), hydrochloric acid (HCl) · Cupric oxide (CuO) were purchased from Kelong chemical Ltd (Chengdu, China). Polyvinylpyrrolidone (PVP Mw = 1300000), polyacrylonitrile (PAN Mw=150000), sodium nitrate (NaNO₃) · sodium nitrite (NaNO₂), sodium nitroferricyanide dihydrate (C₃FeN₆Na₂O·H₂O), sodium salicylate (C₇H₅NaO₃), salicylic acid (C₇H₆O₃), trisodium citrate dihydrate (C₆H₅Na₃O₇·2H₂O), p-dimethylaminobenzaldehyde (C₉H₁₁NO), 0.8 wt% sulfamic acid solution (H₃NO₃S), sodium hypochlorite solution (NaClO), sodium dihydrogen phosphate dihydrate (NaH₂PO₄·H₂O) · disodium phosphate (Na₂HPO₄) were bought from Aladdin Ltd (Shanghai, China). Nafion solution (5 wt%) came from Sigma-Aldrich Chemical Reagent Co., Ltd. Deionized water was purified through a Millipore system.

Preparation of CuFe₂O₄ and CuO@CuFe₂O₄: CuFe₂O₄ nanotubes was synthesized according to electrospinning. 0.5g PVP and 0.5g PAN were dissolved in 10 ml DMF with stirring for 1 h. Then 0.096 g Cu(NO₃)₂·3H₂O and 0.323g Fe(NO₃)₂·9H₂O were added into the above solution with stirring for 24 h at room temperature to form precursor solution.

During the spinning process, a high voltage of 18 kV was applied between the tip of a 10 mL syringe the grounding electrode and the aluminum foil collector placed at a distance of 10 cm, with the precursor fed at a rate of 0.8 mL h⁻¹. Annealing of the collected composite fibers. The fibers were firstly heated up to 200 °C at a 1 °C min⁻¹ heating rate and held for one hour to allow the decomposition of the organic fraction without altering the microstructure. Then, the samples were heated to 600 °C at a rate of 1°C min ⁻¹ and kept 600 °C for 2h. Finally, they are cooled to room temperature at a rate of 1°C min ⁻¹. As for CuO@CuFe₂O₄, 0.193 g Cu(NO₃)₂·3H₂O and 0.162 g Fe(NO₃)₂·9H₂O were added into the solution, and the fabrication process was the same as that for the CuFe₂O₄ nanotubes.

Here comes the reaction equation:

$$Fe^{2+} + 2OH^{-} \leftrightarrow Fe(OH)_{2(s)}$$
 $KSP = 4.87 \times 10^{-17}$ (1)

$$Cu^{2+} + 2OH^{-} \leftrightarrow Cu(OH)_{2(s)} \qquad KSP = 4.8 \times 10^{-20}$$
⁽²⁾

$$Cu(OH)_2 \rightarrow CuO + H_2O \tag{3}$$

$$Fe(OH)_2 \rightarrow FeO + H_2O \tag{4}$$

$$2FeO + \frac{1}{2}O_2 \rightarrow Fe_2O_3 \tag{5}$$

$$Fe_2O_3 + CuO \rightarrow CuFe_2O_4$$
 (6)

$$CuO + CuFe_2O_4 \rightarrow CuO@CuFe_2O_4$$
(7)

Characterization: X-ray diffraction (XRD) analysis of the samples was performed using a LabX XRD-6100 X-ray diffractometer with Cu K α radiation at 0.154 nm. Hitachi S-4800 scanning electron microscope (SEM) was used to observe the morphology and microstructure of the samples. The samples were observed using a JEOL JEM 2100F high resolution field emission transmission electron microscope (TEM) (the samples were dispersed over a copper grid). X-ray photoelectron spectroscopy (XPS) measurements were performed using an Al K ALPH source on Thermo Scientific K-Alpha X-ray Photoelectron spectrometer in a high-purity nitrogen atmosphere.

Electrochemical methods: Absorbance data are acquired from SHIMADZU UV-1800 uV-vis spectrophotometer. The concentrations of ammonium and nitrite are calculated from the standard concentration curve. Throughout all the measurement, a three-electrode system is utilized. Catalyst is deposited on the treated carbon paper and Nafion is used as the protective film. Carbon rod is employed as the counter electrode, and stable Ag/AgCl/KCl is used as the reference electrode. The neutral 0.1 M PBS

electrolyte is replaced when the voltage is changed. Ammonia production rate and Faraday efficiency are calculated using the following formulas:

where V means the electrolytic liquid volume, Q the amount of reaction charge, m the mass of the catalyst.

Determination of NH₃: Indophenol blue method is adopted to quantify the concentration of produced NH₃. The electrolytes after 1 h electrolysis is diluted 20 times to 2 mL and then added 2 mL of coloring solution (1 M NaOH containing 5 wt% salicylic acid and 5 wt% sodium citrate), 1 mL of the oxidizing solution (0.05 M sodium hypochlorite solution) and 200 uL catalyst solution (1 wt% sodium nitroferricyanide (III) dehydrate) one by one. After the mixed solution reacts for 2 h in the dark, the UV-Vis absorption spectra are performed at $\lambda = 665$ nm. We calibrate the standard NH₄Cl solutions with different concentration and then calculate the fitting curve (y=0.449x +0.0336, R²=0.999).

Determination of NO₂⁻: The electrolytes after 1 h electrolysis is diluted 20 times to 1 mL and 1 mL color reagent (1.0 g sulfonamide, 2.94 mL H₃PO₄, 0.1 g N-(1-naphthyl)-ethylenediamine dihydrochloride) and 2 mL H₂O are added one by one. After the mixed solution reacts for 10 min in the dark, the UV-Vis absorption spectra are performed at $\lambda = 540$ nm. We calibrate the standard NO₂⁻ solutions with different concentration and calculate the fitting curve (y=0.247x +0.0147, R²=0.999).

Determination of N₂H₄: The electrolytes after 1 h electrolysis is diluted 20 times to 1 mL \cdot 5.99 g C₉H₁₁NO , 30 ml HCl and 300mL C₂H₅OH was mixed to form a uniform solution used as a color reagent. Then, 1 mL color reagent was added into electrolyte after 2 h electrolysis. The absorbance was performed at a wavelength of 455 nm after the mixture stand 20 min in dark. The fitting curve (y = 0.5718x + 0.1543, R² = 0.9998) shows good linear relation of absorbance value with N₂H₄ concentration.

¹⁵N isotope labeling experiments: After electrolysis in 0.1 M PBS with 0.1 M $^{15}NO_3^{-1}$ or $^{14}NO_3^{-1}$ at -1.0 V for 24 h, the pH value of the obtained electrolyte is adjusted to be 3.0 by the addition of 0.5 M H₂SO₄ solution. Then, 1 mL of electrolyte, 0.2 mL of

DMSO and 0.2 mL of D₂O are added into the NMR tube for further NMR detection.

Computational Details:

First-principles calculations are performed by using the Vienna Ab initio Simulation Package (VASP)¹⁻⁴ to investigate the NO₃RR process on CuO and CuFe₂O₄ surface. The valence-core electrons interactions are treated by Projector Augmented Wave (PAW)⁵ potentials and the electron exchange correlation interactions are described by the generalized gradient approximation (GGA) with the Perdew-Burke-Ernzerhof (PBE)⁶ functional. Considered long-range interaction at the interface, Van der Waals interactions are considered using DFT-D3 correlation⁷. To avoid interaction come from other slabs, a vacuum of 20 Å is added along z direction. The convergence criterion of geometry relaxation is set to 0.03 eV•Å⁻¹ in force on each atom. The energy cutoff for plane wave-basis is set to 500 eV. The K points are sampled with $3\times3\times1$ by Monkhorst-Pack method⁸.

Gibbs free energy change (ΔG) is evaluated based on the computational hydrogen electrode (CHE) model, which takes one-half of the chemical potential of gaseous hydrogen under standard conditions as the free energy of the proton-electron pairs. ΔG is calculated by the following equation⁹:

$$\Delta G = \Delta E + \Delta E_{ZPE} - T\Delta S + neU$$

where ΔE , ΔE_{ZPE} , ΔS are the reaction energy from DFT calculation, the correction of zero-point energy and the change of simulated entropy, respectively. T is the temperature (T = 300 K). *n* and U are the number of transferred electrons and applied potential respectively.

Supplementary Figures



Figure S1. XRD pattern of pure CuFe₂O₄ fibers



Figure S2. SEM image of CO@CFO before calcination



Figure S3. SEM image of pure CFO



Figure S4. EPR spectra of CO@CFO and CFO.



Figure S5. LSV curve of CO@CFO in 0.1M PBS electrolyte with and without NO₃⁻.



Figure S6. Time-dependent current density curves at different potentials.



Figure S7. UV–vis absorption spectra at 5 various potentials with the indophenol indicator.



Figure S8. (a) UV-Vis absorption spectra of indophenol assays with NH_3 . (b) Calibration curve used for the determination of NH_3 concentration.



Figure S9. (a) UV-Vis absorption spectra of indophenol assays with NO_2^- . (b) Calibration curve used for the determination of NO_2^- concentration.



Figure S10. NO_2^- yields and FEs for CO@CFO in 0.1M PBS with 0.1M NO_3^- at different potentials



Figure S11. (a) UV-Vis absorption spectra of indophenol assays with N_2H_4 . (b) Calibration curve used for the determination of N_2H_4 concentration. (c) UV–Vis absorption spectra at 5 various potentials with the indophenol indicator.



Figure S12. Time-dependent current density curves of CO@CFO at a fixed potential of -1.0 V vs. RHE for 24 h.



Figure S13. (a)XRD pattern, (b)TEM image, XPS(c-f) spectra for CO@CFO after 24

h electrolysis



Figure S14. UV–Vis absorption spectra at identical potentials of -1.0 V vs. RHE with the indophenol indicator.



Figure S15. (a)Time-dependent current density curves at different potentials of CFO. (b) UV–Vis absorption spectra at 5 various potentials with the indophenol indicator of CFO. (c) NH_3 yields and FEs for CFO in 0.1M PBS with 0.1M NO_3^- at different potentials



Figure S16. (a-c) CVs of CO@CFO, CFO and CuO various scan rates (20-100 mV s⁻¹) from -0.0 V to -0.10 V vs. Ag/AgCl. (d-f) Capacitive current densities at -0.05 V vs. Ag/AgCl as a function of scan rates.



Figure S17. EIS Nyquist plots of different samples by applying an impedance amplitude of 5 mV with the frequency range from 10^4 to 10^{-1} Hz.



Figure S18. Comparison of amount of produced NH₃ under four different conditions.



Figure S19. 1H NMR spectra of the electrolyte after electrocatalytic reaction using $15NO_3^-$ and $14NO_3^-$ as the nitrogen source.



Figure S20. Ion chromatography test before and after 2 h electrolysis for CO@CFO.



Figure S21. Calculated free energy profiles of (a) CuO and (b) CFO surfaces. The charge density difference of NO_3^- adsorption configuration is plotted in the illustration. (c) Calculated density of states of pristine CuO and CFO.

Catalysts	Electrolyte	NH ₃ yield	FE	Ref
CuO@CuFe ₂ O ₄	0.1 M PBS with	9296.21 μg h ⁻¹ mg _{cat.} -	91.08%	This
	0.1 M KNO ₃	1		work
CuOx	0.1 M KOH	$449.41 \pm 12.18 \ \mu g \ h^{-1}$	74.18 ±	10
nanoparticles	(50 ppm KNO ₃)	mg _{cat.} ⁻¹	2.27%	
Cu/Cu ₂ O NWAs	$0.5 \text{ M} \text{ Na}_2 \text{SO}_4$	4148 µg h ⁻¹ mg _{cat.} ⁻¹	95.8 %	11
	(200 ppm NO ₃ ⁻)			
Cu nanosheets	0.1 M KOH	390.1 μ g h ⁻¹ mg _{cat} ⁻¹	99.7%	12
	(10 mM KNO ₃)			
NFP	0.5 M Na ₂ SO ₄	952 μg h ⁻¹ mg _{cat.} ⁻¹	99.2%	13
	(80 mg L ⁻¹ NaNO ₃)			
TiO ₂ -x	0.5 M Na ₂ SO ₄	765 µg h ⁻¹ mg _{cat.} ⁻¹	85%	14
	(50 ppm NO ₃ ⁻)			
TiO ₂	0.5 M Na ₂ SO ₄	408 µg h ⁻¹ mg _{cat.} ⁻¹	66.3%	14
	(50 ppm NO ₃ ⁻)			
Co3O4@NiO	0.5 M Na ₂ SO ₄	117.8µg h ⁻¹ mg _{cat.} ⁻¹	54.97%	15
HNTs	(200 ppm NO ₃ ⁻)			
In-S-G	-	3740 mmol $g_{cat}^{-1}h^{-1}$	75%	16
BCN@Cu	-	4609.1µg h ⁻¹ mg _{cat.} ⁻¹	91.15%	17
Co ₃ O ₄ /Cu	0.4 MNa ₂ SO ₄	$684 \ \mu g \ h^{-1} \ mg \ _{cat.}^{-1}$	94.6%	18
	$(50 \text{ mg } \text{L}^{-1} \text{NO}_3^{-})$			
PA-RhCu cNCs	0.1 м HClO ₄	2400 μg h ⁻¹ mg _{cat} ⁻¹	93.7%	19

Table S1. Comparison of electrocatalytic NO3RR performance for CuO@CuFe2O4with other Cu or Fe-related electrocatalysts under ambient conditions.

(0.05 м КNО ₃)			
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