

## Supporting Information For:

### **Diamino group-functionalized Zr-based metal-organic framework for fluorescence sensing of free chlorine in aqueous phase and Knoevenagel condensation**

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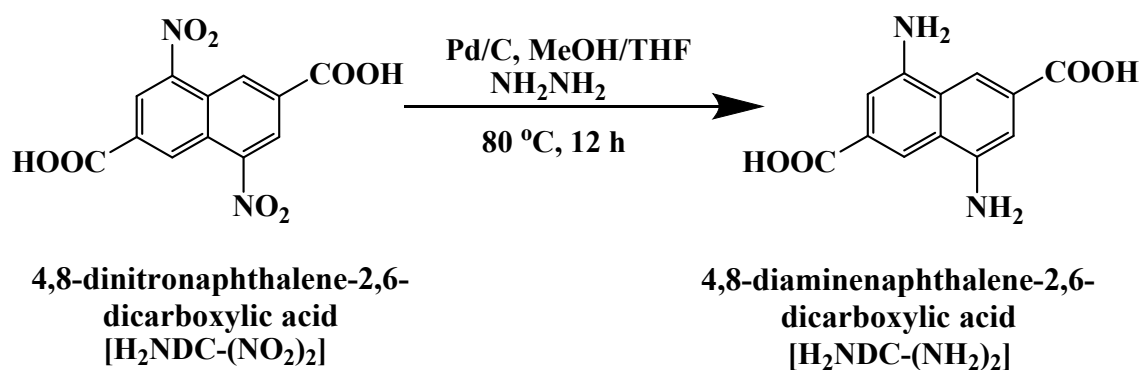
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## Materials and General Methods:

All the reagents and solvents were procured from commercial sources and used without purification, except the  $\text{H}_2\text{NDC}-(\text{NH}_2)_2$  ligand. The notations used for characterization of the bands are broad (br), strong (s), very strong (vs), medium (m), weak (w) and shoulder (sh). 9KW Powder X-Ray Diffraction System, Make: Rigaku Technologies, JAPAN, Model: Smartlab was employed for X-ray powder diffraction (XRPD) measurements. The Attenuated Total Reflectance Infrared (ATR-IR) spectra were recorded using PerkinElmer UATR Two at ambient condition in the region  $400\text{-}4000\text{ cm}^{-1}$  (USA). Thermogravimetric analysis (TGA) was carried out with a TG 209 F1 Libra; Make: M/s Netzsch, Germany thermogravimetric analyzer in the temperature range of  $25\text{-}700\text{ }^\circ\text{C}$  in an air atmosphere at the rate of  $10\text{ }^\circ\text{C min}^{-1}$ .  $\text{N}_2$  sorption isotherms were recorded by using Quantachrome AutosorbIQ-MP (USA) volumetric gas adsorption equipment at  $-196\text{ }^\circ\text{C}$ . Before the sorption analysis, the degassing of the compound was carried out at  $100\text{ }^\circ\text{C}$  under a high vacuum for 12 h. Fluorescence sensing studies were performed with a HORIBA JOBIN YVON Fluoromax-4 (Japan) spectrofluorometer. A BrukerAvance III 600 NMR (Germany) spectrometer was used for recording  $^1\text{H}$  NMR spectra at 600 MHz. Mass spectra were recorded with an Agilent 6520 Q-TOF (USA) high-resolution mass spectrometer (HR-MS). Picosecond Time-resolved and Steady State Luminescence Spectrometer, Make: Edinburg Instruments, Model: Lifespec II & FSP 920 was used for the TRPL study. Pawley refinement was carried out using Materials Studio software. The DICVOL program<sup>1</sup> incorporated within STOE's WinXPow software package<sup>2</sup> was used to determine the lattice parameters.

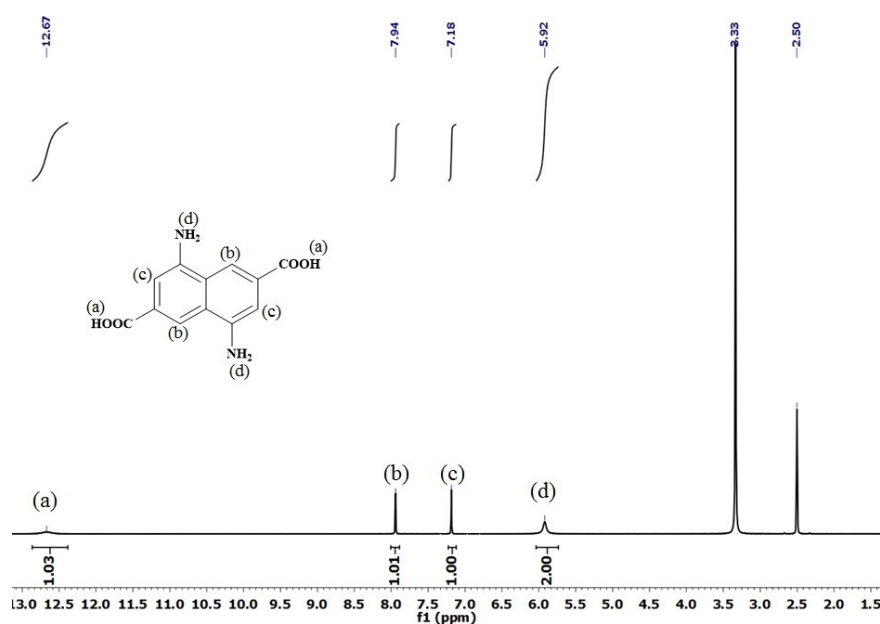
## Synthesis of $\text{H}_2\text{NDC}-(\text{NH}_2)_2$ linker:

4,8-Dinitronaphthalene-2,6-dicarboxylic acid ( $\text{H}_2\text{NDC}-(\text{NO}_2)_2$ ) linker was prepared from naphthalene-2,6-dicarboxylic acid ( $\text{H}_2\text{NDC}$ ) by following the protocol reported in the literature.<sup>3</sup> In a 250 mL round-bottom flask, 1 g (3.3 mmol) of  $\text{H}_2\text{NDC}-(\text{NO}_2)_2$  was taken and 100 mL of methanol and 50 mL of tetrahydrofuran (THF) were added at room temperature. The reaction mixture was stirred to completely dissolve the reactant. Then, 280 mg of Pd/C was added very slowly in stirring conditions at room temperature. The flask was fitted with a condenser and a nitrogen filled balloon. Then, the solution was heated at  $60\text{ }^\circ\text{C}$ . After 10 min, 4 mL of hydrazine hydrate was added very slowly and after complete addition, the mixture was heated at  $80\text{ }^\circ\text{C}$  for overnight. A dark greenish precipitate was obtained and the precipitate was filtered through a filter paper. The precipitate was dissolved in 1M NaOH (100 mL). Then, the precipitate was again filtered off and conc. HCl was slowly added to the filtrate to acidify the solution, resulting in dark yellowish precipitation. The obtained solid product, i.e. 4,8-diaminenaphthalene-2,6-dicarboxylic acid ( $\text{H}_2\text{NDC}-(\text{NH}_2)_2$ ) linker was filtered off, washed with large amount of water and dried in an oven at  $60\text{ }^\circ\text{C}$ . The reaction scheme for the preparation of  $\text{H}_2\text{NDC}-(\text{NH}_2)_2$  linker is shown in Scheme S1.

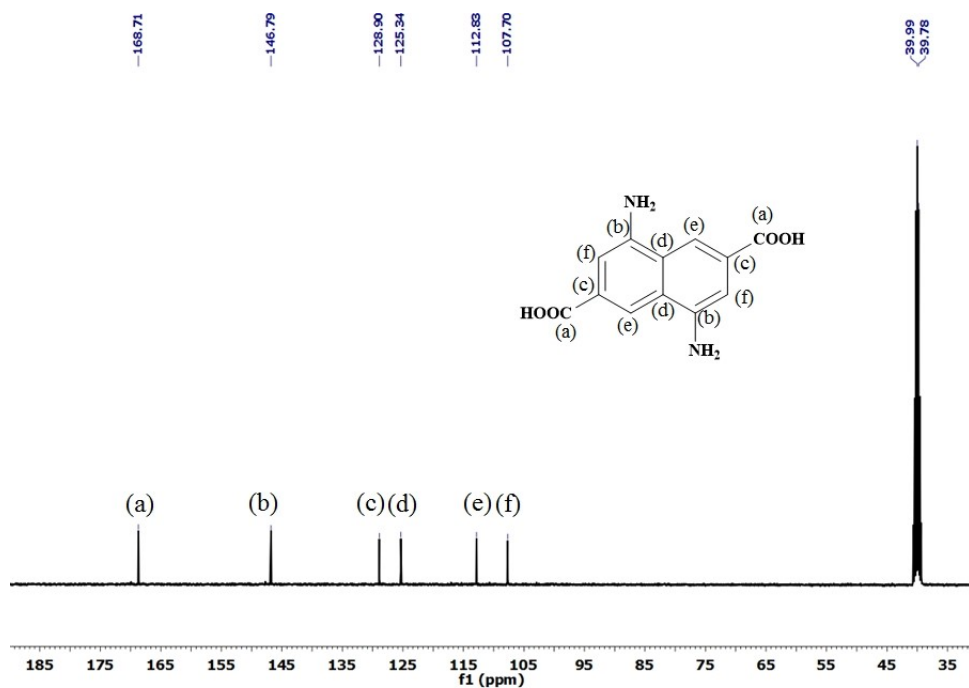


**Scheme S1.** Synthesis of H<sub>2</sub>NDC-(NH<sub>2</sub>)<sub>2</sub> linker.

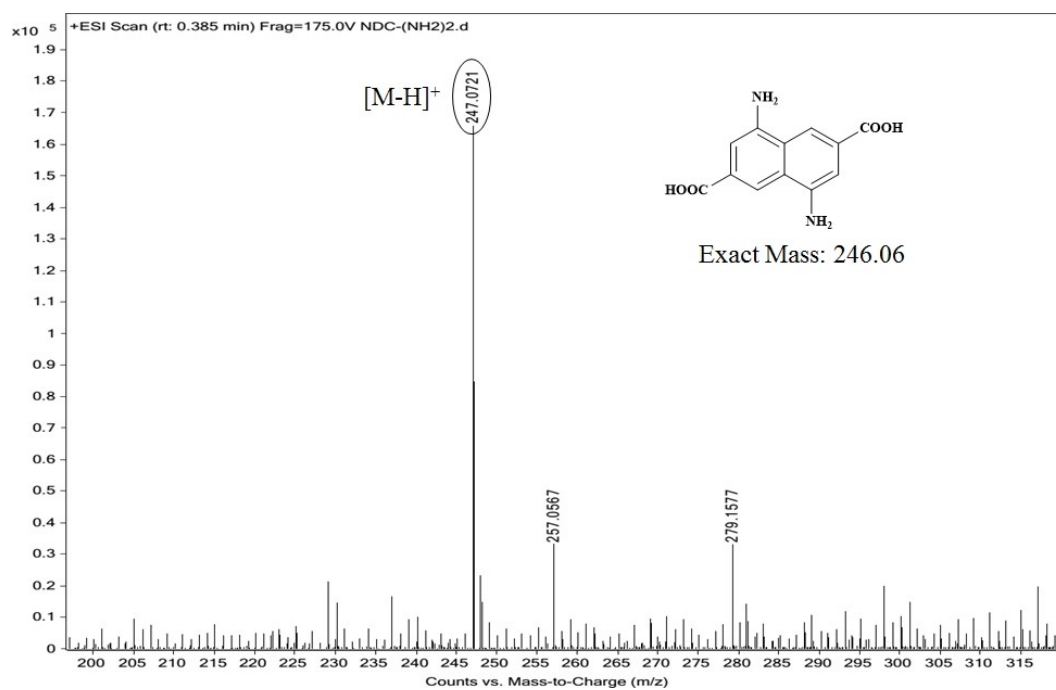
The characterization of the linker was carried out by mass spectrometry, <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy. The <sup>1</sup>H NMR spectrum (Figure S1) confirmed the presence of four types of protons in the H<sub>2</sub>NDC-(NH<sub>2</sub>)<sub>2</sub> linker. The naphthalene moiety contains two types of protons and one type of proton is present at the amine group and one type of proton is present in the carboxylic acid group. The different types of protons are marked by (a), (b), (c) and (d). The chemical shift values are observed at 12.67 (s, 2H, -COOH), 7.94 (s, 2H, Ar-H), 7.18 (s, 2H, Ar-H) and 5.92 (s, 4H, -NH<sub>2</sub>) ppm, respectively. Since there are no additional protons in the <sup>1</sup>H NMR spectrum, the formation of by-products is ruled out. The <sup>13</sup>C NMR spectrum (Figure S2) revealed that there are six types of carbon atoms present in the ligand. The observed chemical shift values are 107.70, 112.83, 125.34, 128.90, 146.79 and 168.71 ppm. In the mass spectrum of the linker (Figure S3), the most intense peak was found at m/z = 247.072 (measured in positive ion mode), which corresponds to (M-H)<sup>+</sup> ion (M is the mass of H<sub>2</sub>NDC-(NH<sub>2</sub>)<sub>2</sub> ligand). The theoretically calculated mass of the linker is 246.06 g mol<sup>-1</sup>. Therefore, it can be concluded that the desired H<sub>2</sub>NDC-(NH<sub>2</sub>)<sub>2</sub> linker has been synthesized in pure form.



**Figure S1.** <sup>1</sup>H NMR spectrum of H<sub>2</sub>NDC-(NH<sub>2</sub>)<sub>2</sub> linker measured in DMSO-d<sub>6</sub>.



**Figure S2.**  $^{13}\text{C}$  NMR spectrum of  $\text{H}_2\text{NDC}-(\text{NH}_2)_2$  linker measured in  $\text{DMSO}-d_6$ .



**Figure S3.** Mass spectrum of  $\text{H}_2\text{NDC}-(\text{NH}_2)_2$  linker.



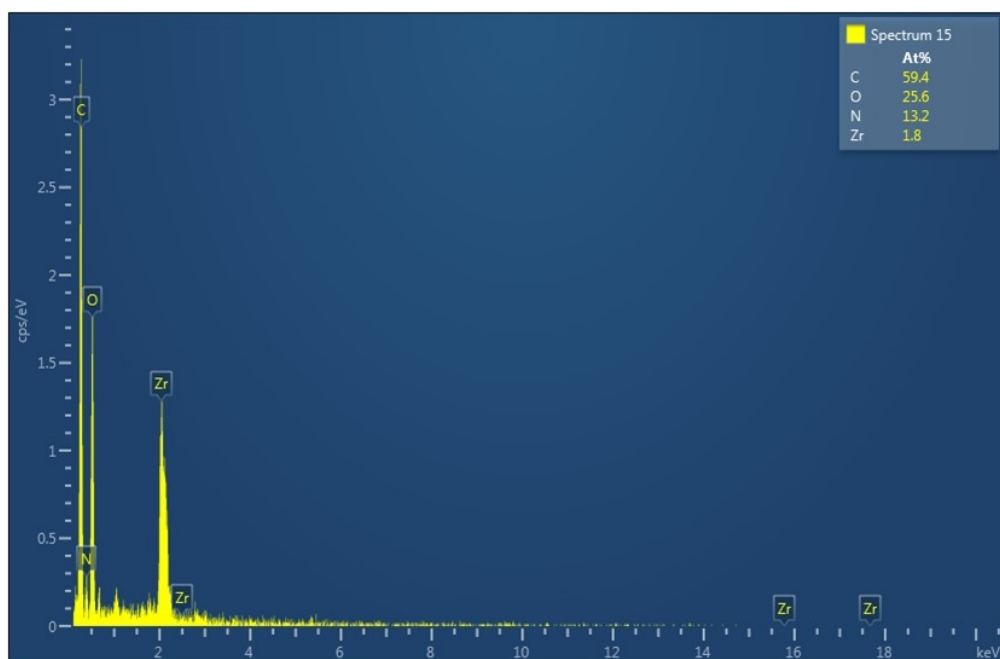


Figure S4. EDX spectrum of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1.

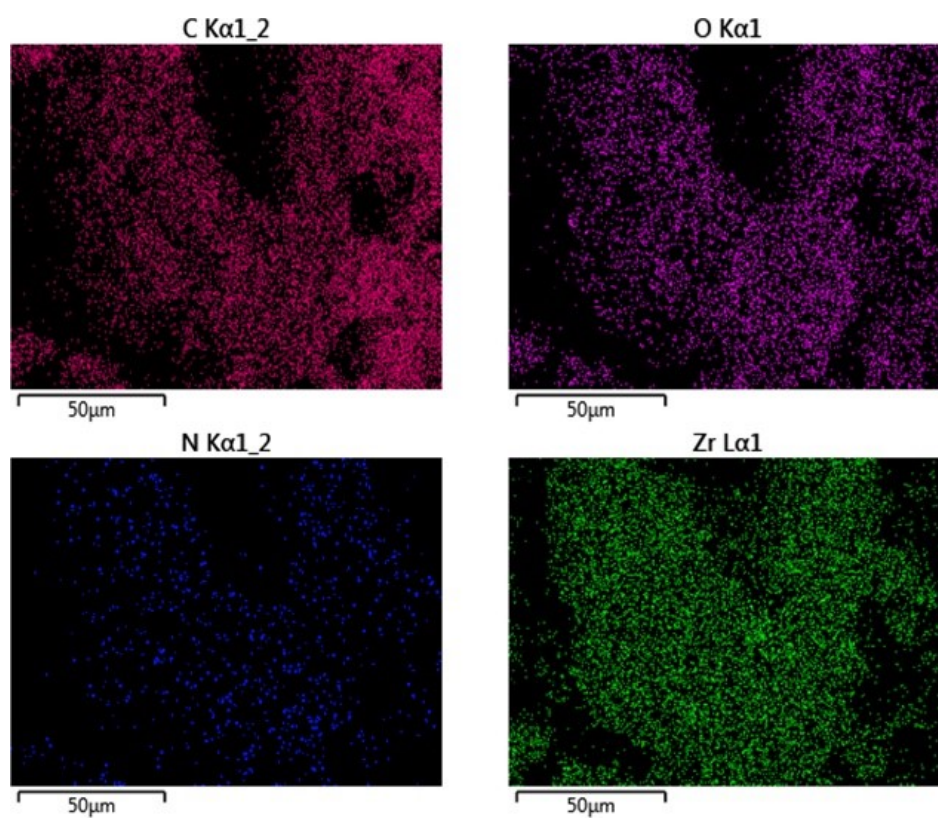
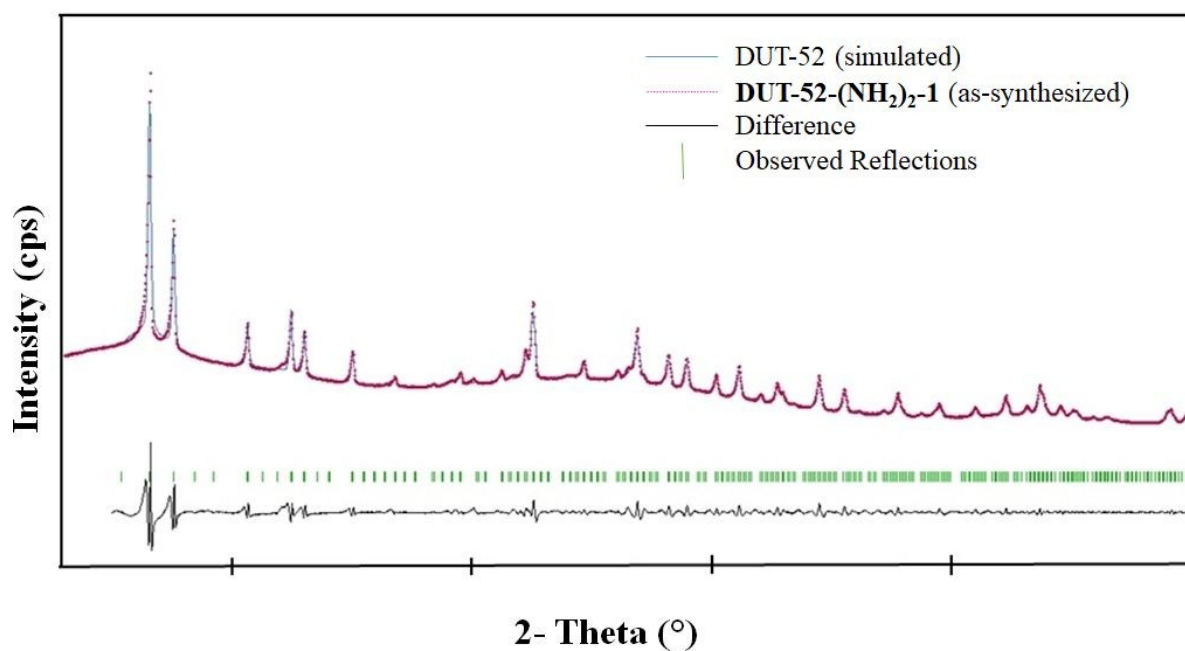


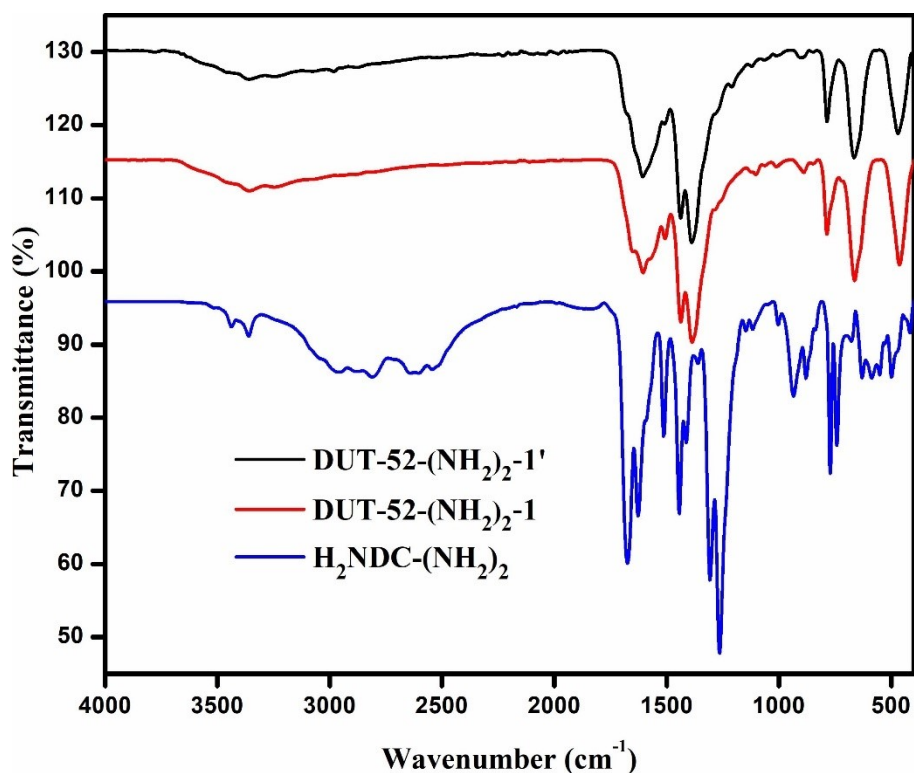
Figure S5. EDX elemental mapping of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1.



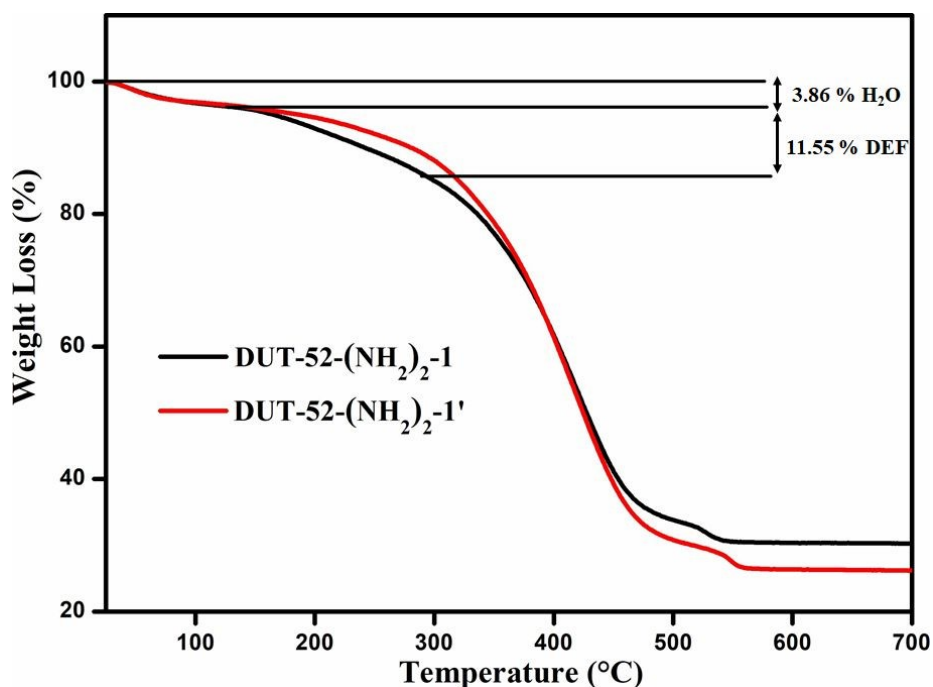
**Figure S6.** Pawley fit for the XRPD pattern of as-synthesized **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1**. Blue lines and red dots denote simulated and observed patterns, respectively. The peak positions and difference plot are displayed at the bottom ( $R_p = 2.13$ ,  $R_{wp} = 3.54$ ).

**Table S1.** Unit cell parameters of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1** and parent Zr-DUT-52 MOF.

Compound name	<b>DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1</b> (This work)	Zr-DUT-52 (Reported)
Space Group	Fm $\bar{3}$ m	Fm $\bar{3}$ m
Crystal System	Cubic	Cubic
a = b = c (Å)	23.723 (4)	23.910 (3)
$\alpha = \beta = \gamma$ (°)	90	90
V (Å <sup>3</sup> )	13350 (38)	13669 (3)
Figure of Merit (FOM)	30.6	-



**Figure S7.** FT-IR spectra of  $\text{H}_2\text{NDC}-(\text{NH}_2)_2$  linker, as-synthesized **DUT-52- $(\text{NH}_2)_2-1$**  (red) and activated **DUT-52- $(\text{NH}_2)_2-1'$**  (black).



**Figure S8.** TG curves of as-synthesized **DUT-52- $(\text{NH}_2)_2-1$**  (black) and activated **DUT-52- $(\text{NH}_2)_2-1'$**  (red) recorded in an air atmosphere in the temperature range of 25-700 °C at a heating rate of 5 °C min<sup>-1</sup>.

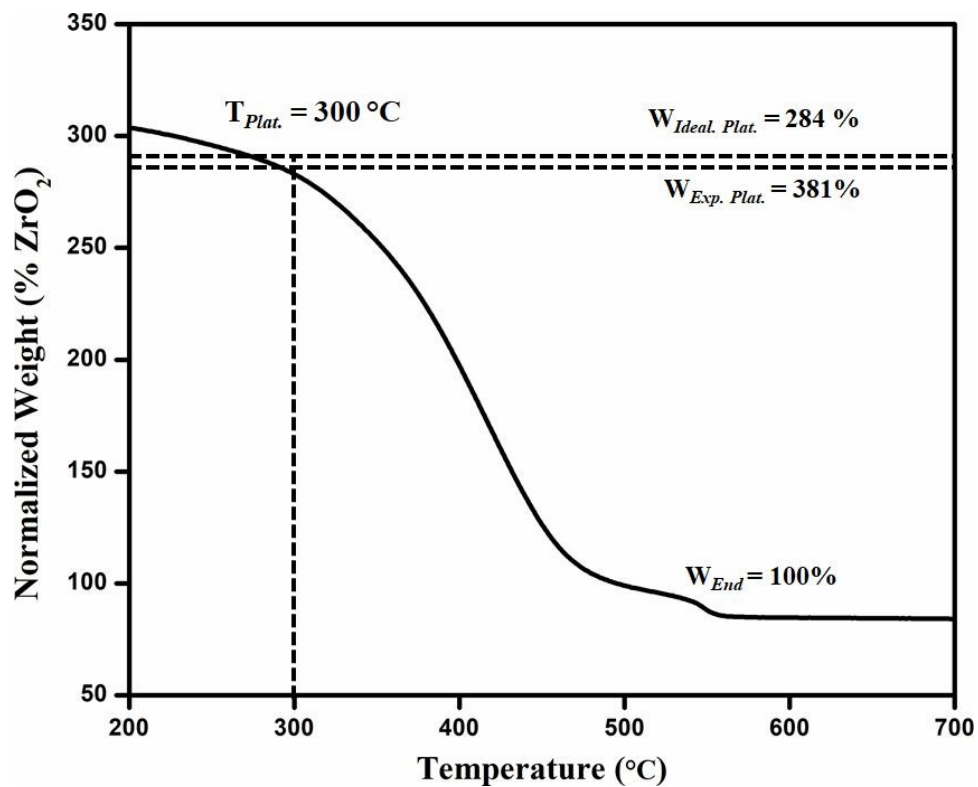


Figure S9. Calculation of missing linker defects from the TG curve of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'**.

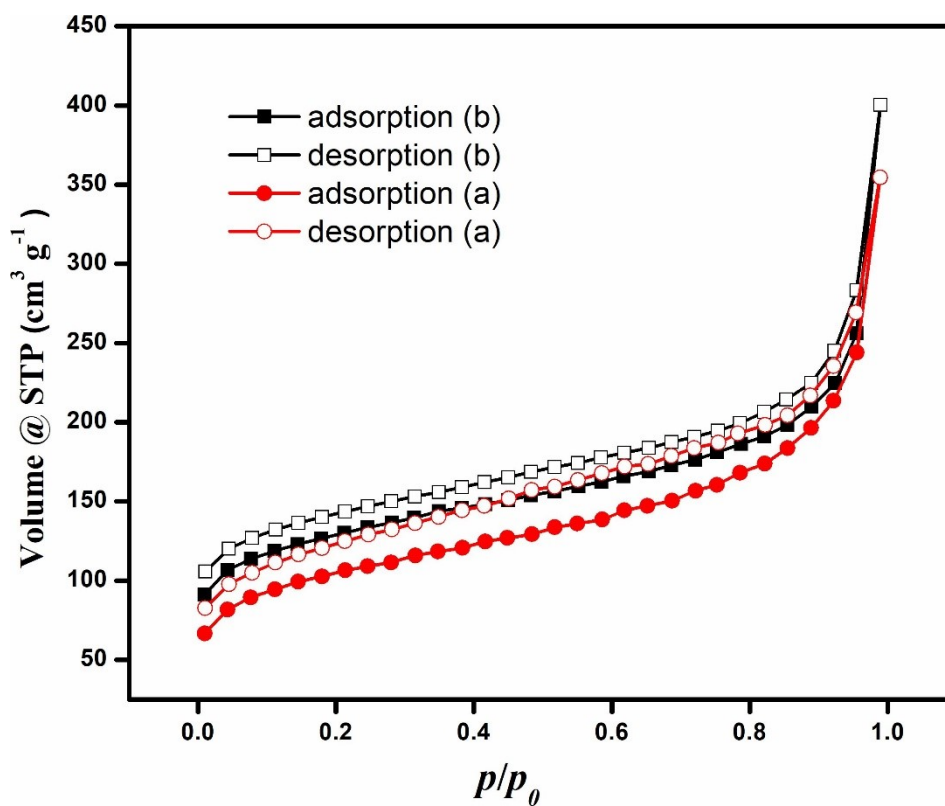


Figure S10. N<sub>2</sub> adsorption and desorption isotherms of (a) **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1** (b) **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** recorded at -196 °C.

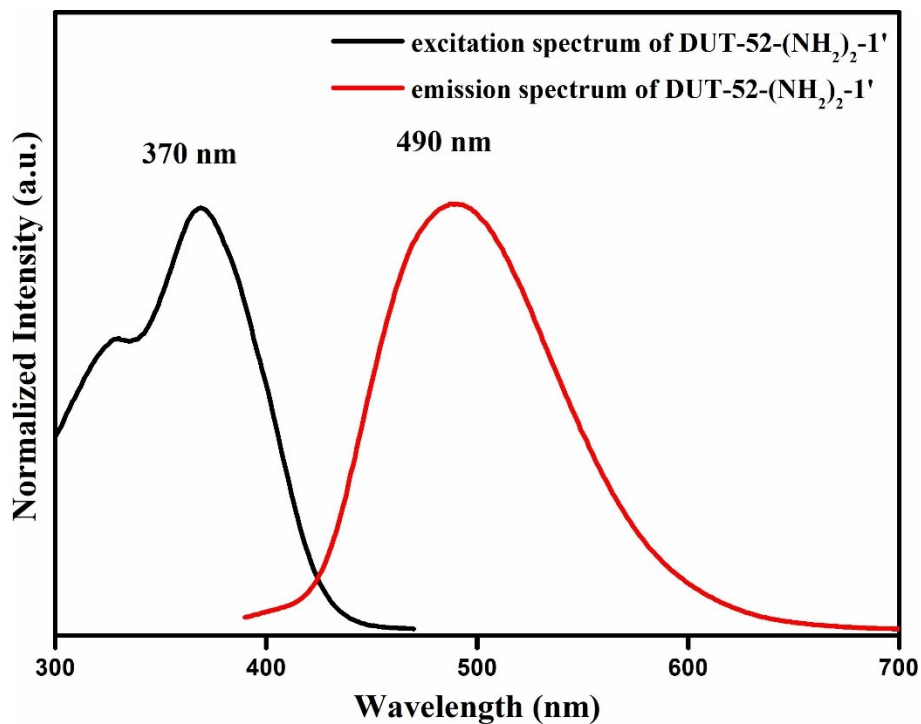


Figure S11. Fluorescence emission spectra and excitation DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'.

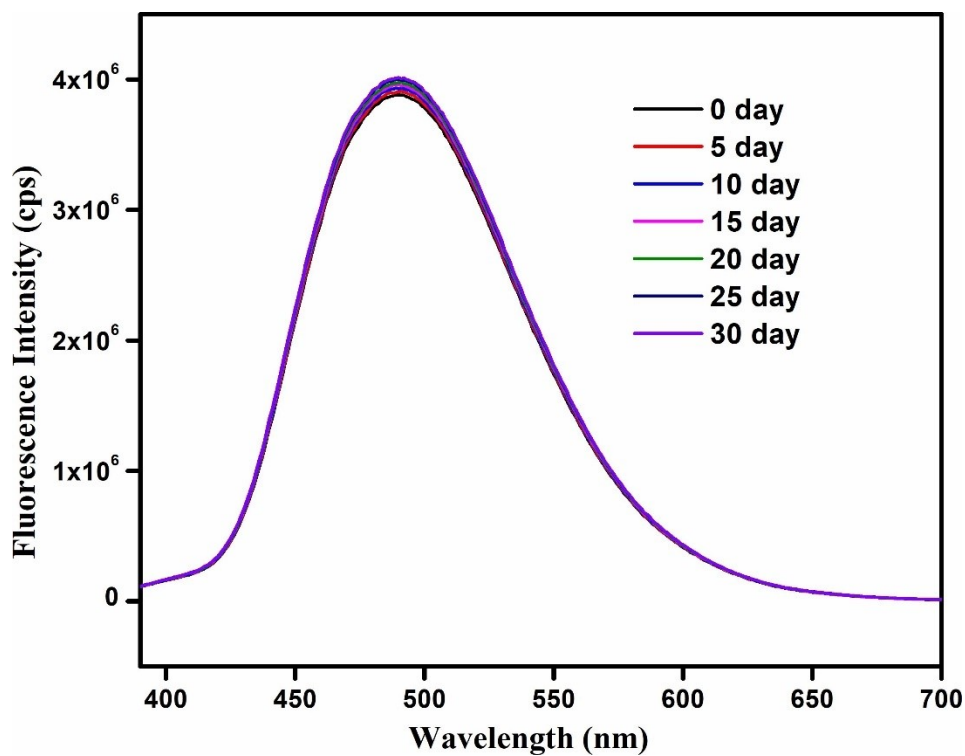
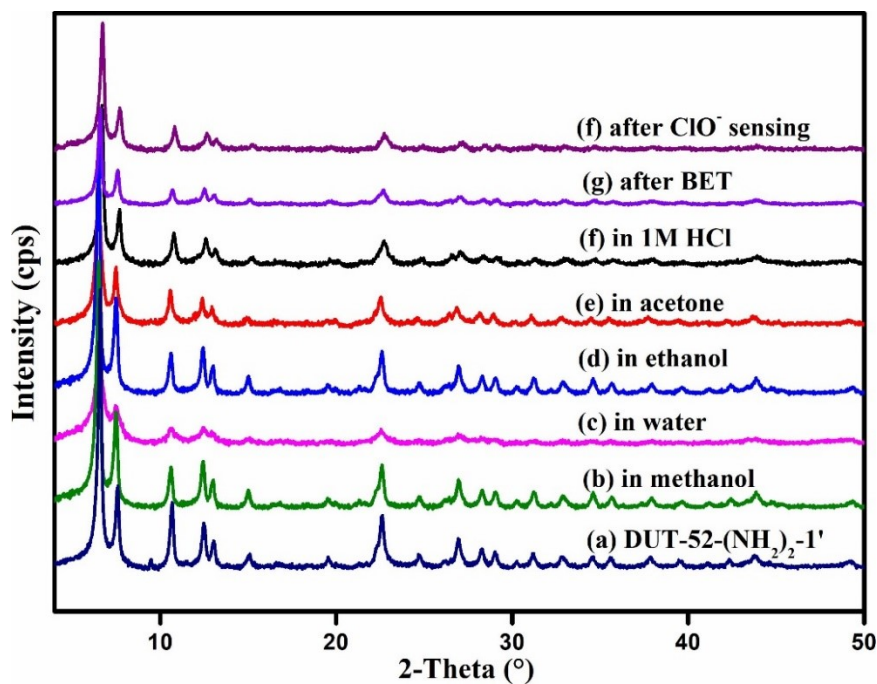
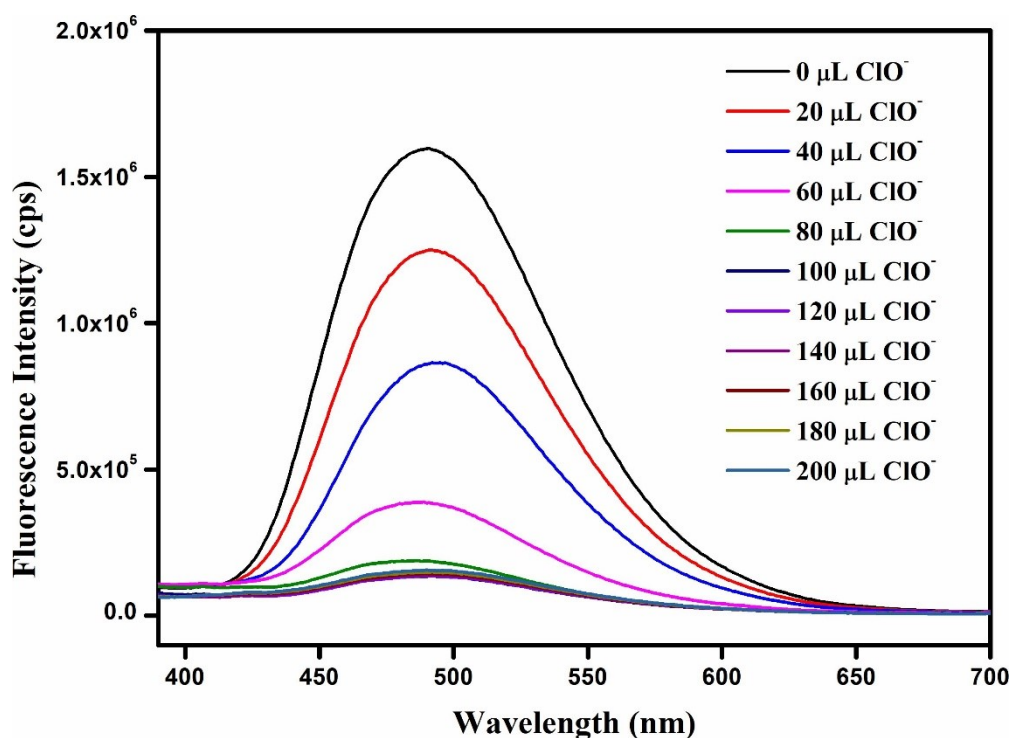


Figure S12. Day-to-day fluorescence stability of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' in PBS (pH = 7.4).

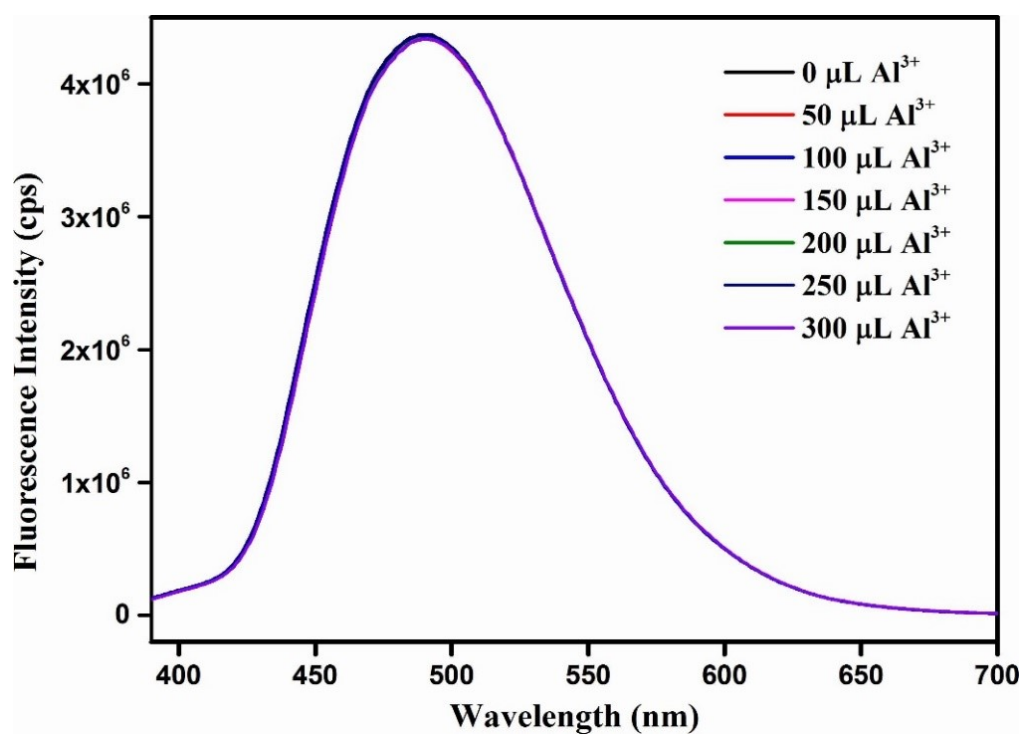


**Figure S13.** XRPD patterns of (a) **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** showing its chemical stability in (b) methanol, (c) water, (d) ethanol, (e) acetone and (f) 1M HCl. The XRPD pattern of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** after (g) BET and (f) ClO<sup>-</sup> sensing experiments.

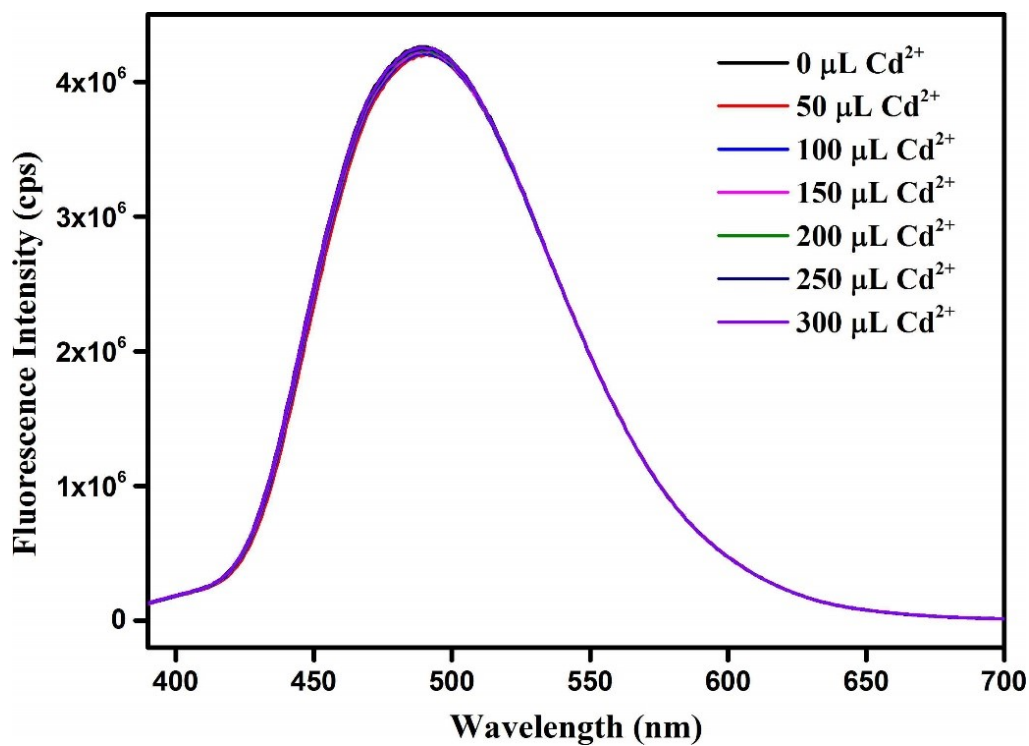


**Figure S14.** Change in fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** after incremental addition of 5 mM ClO<sup>-</sup> in water.

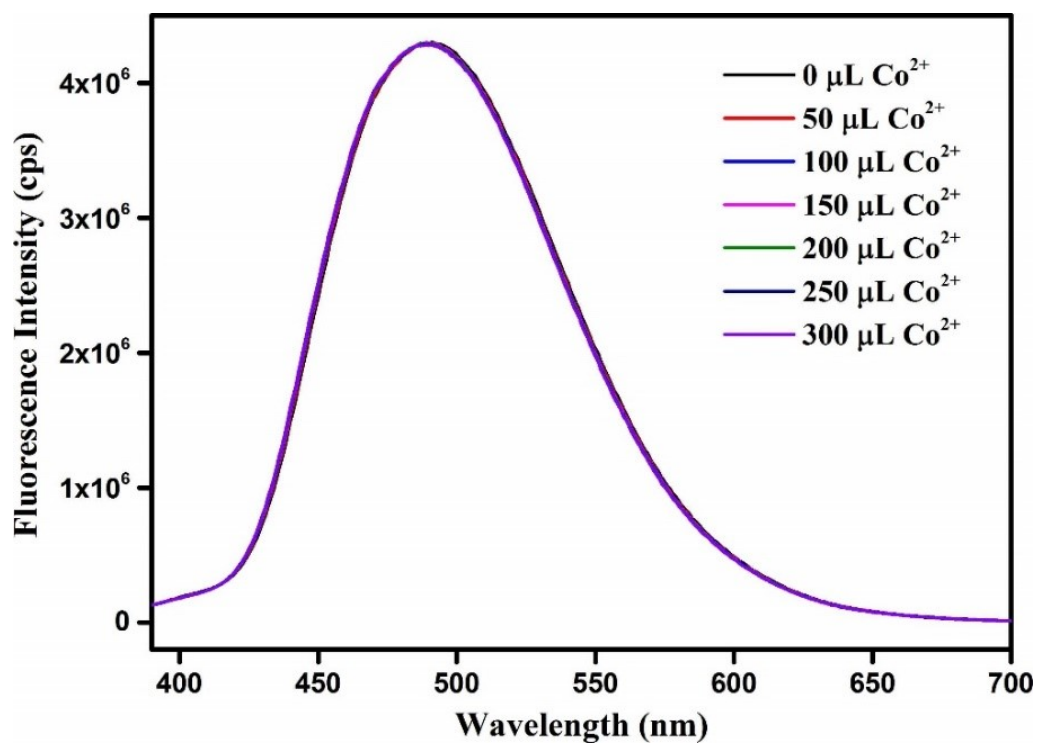




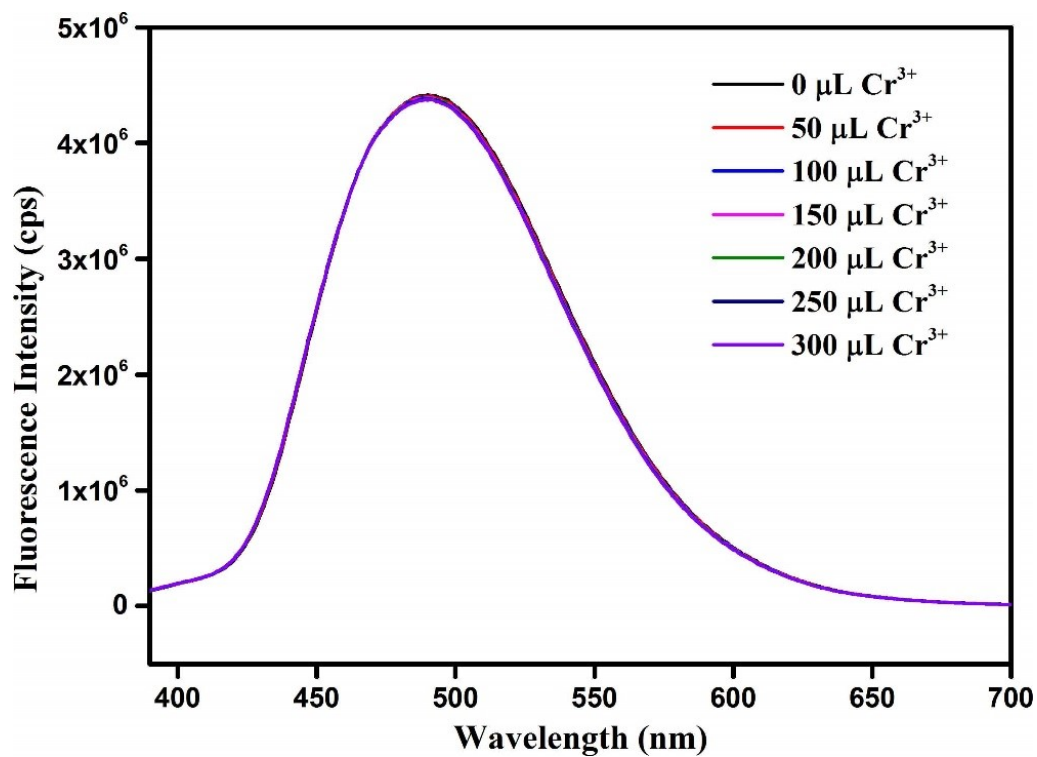
**Figure S15.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Al<sup>3+</sup> (300 μL).



**Figure S16.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Cd<sup>2+</sup> (300 μL).

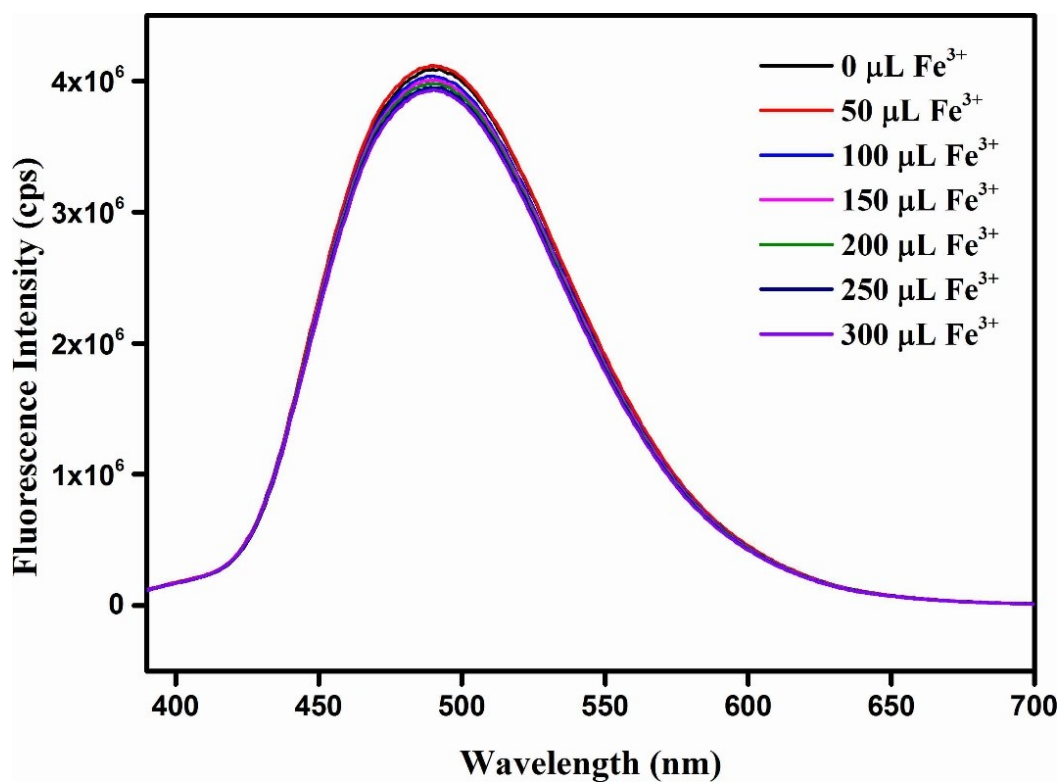


**Figure S17.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Co<sup>2+</sup> (300 μL).

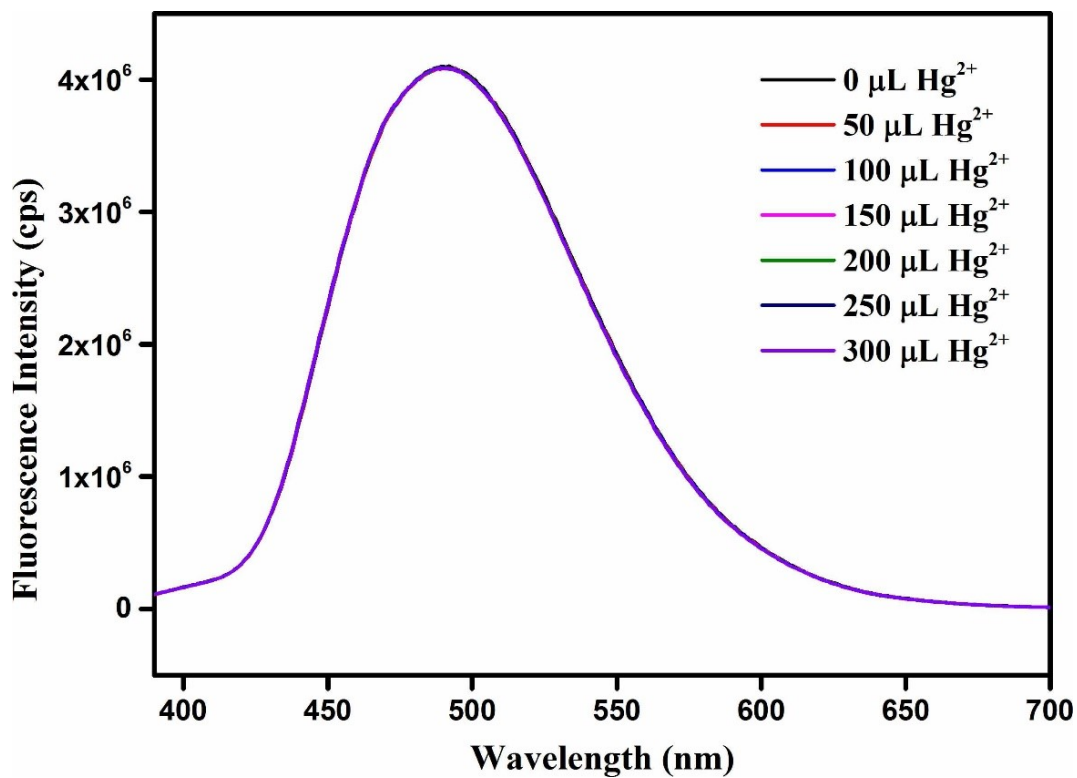


**Figure S18.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Cr<sup>3+</sup> (300 μL).

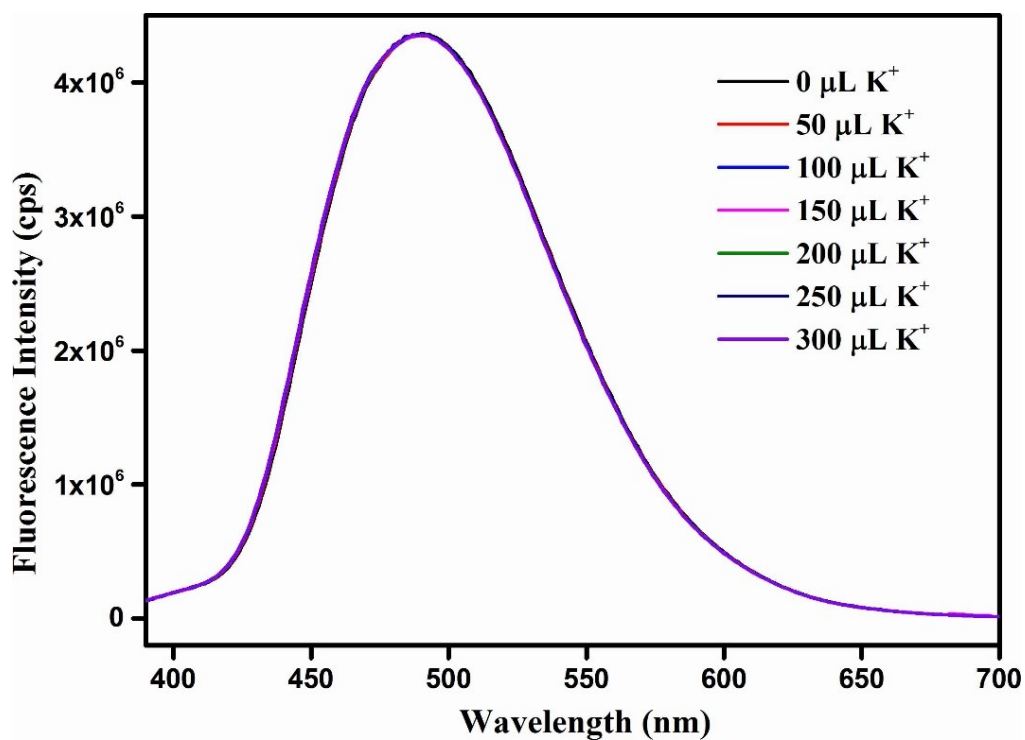




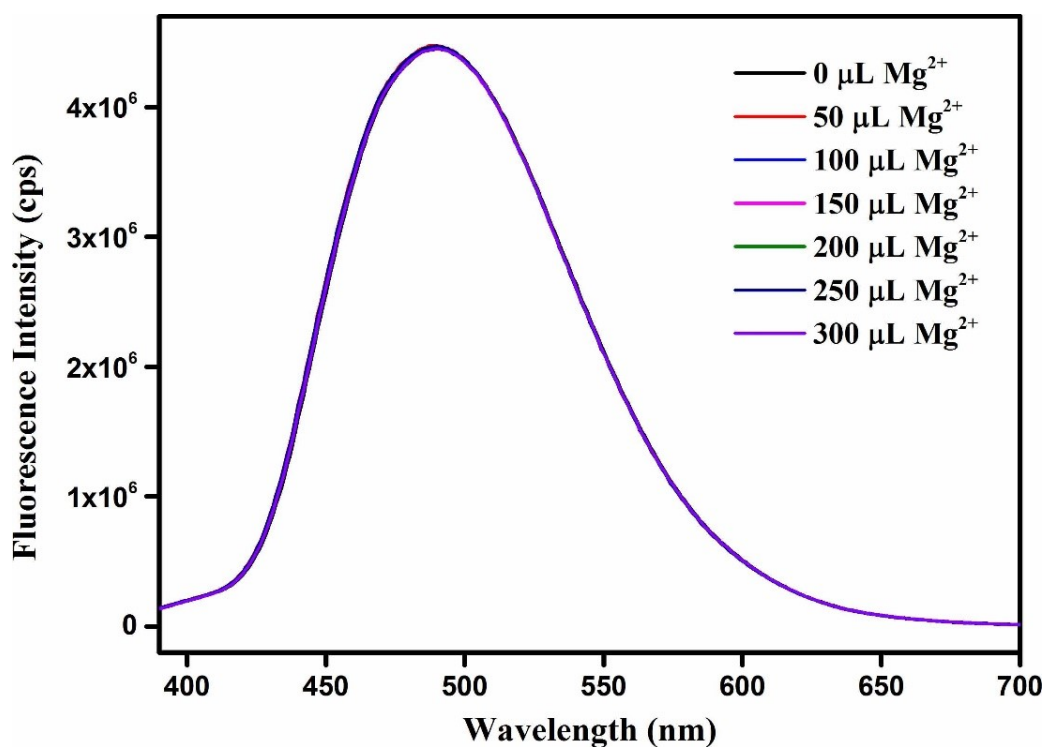
**Figure S19.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Fe<sup>3+</sup> (300 μL).



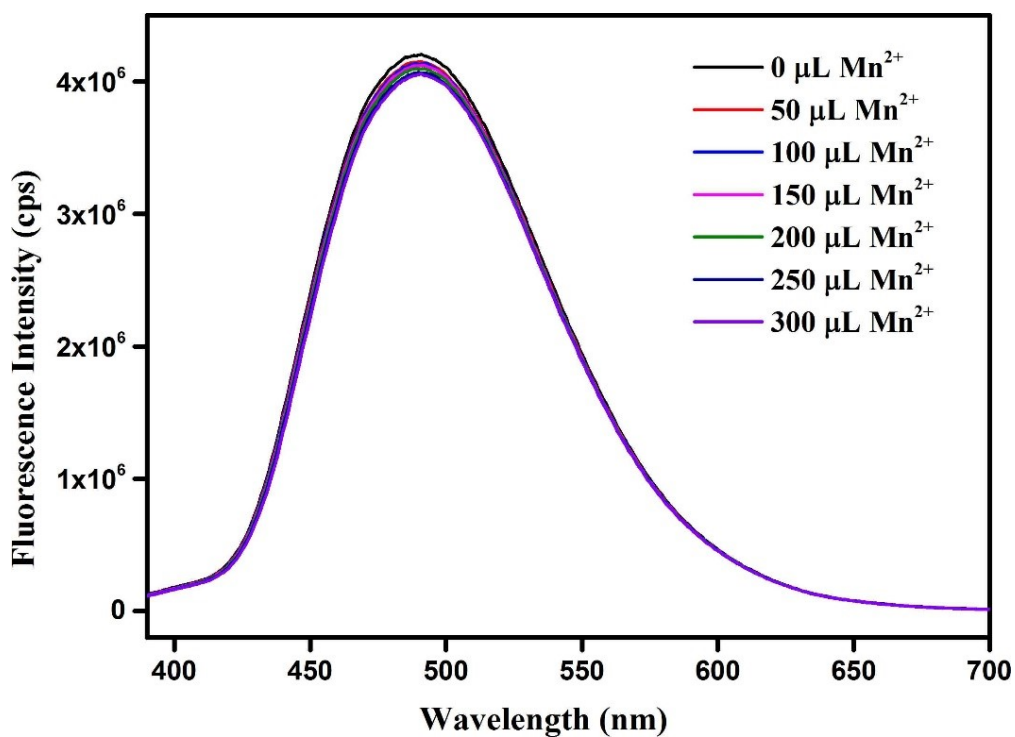
**Figure S20.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Hg<sup>2+</sup> (300 μL).



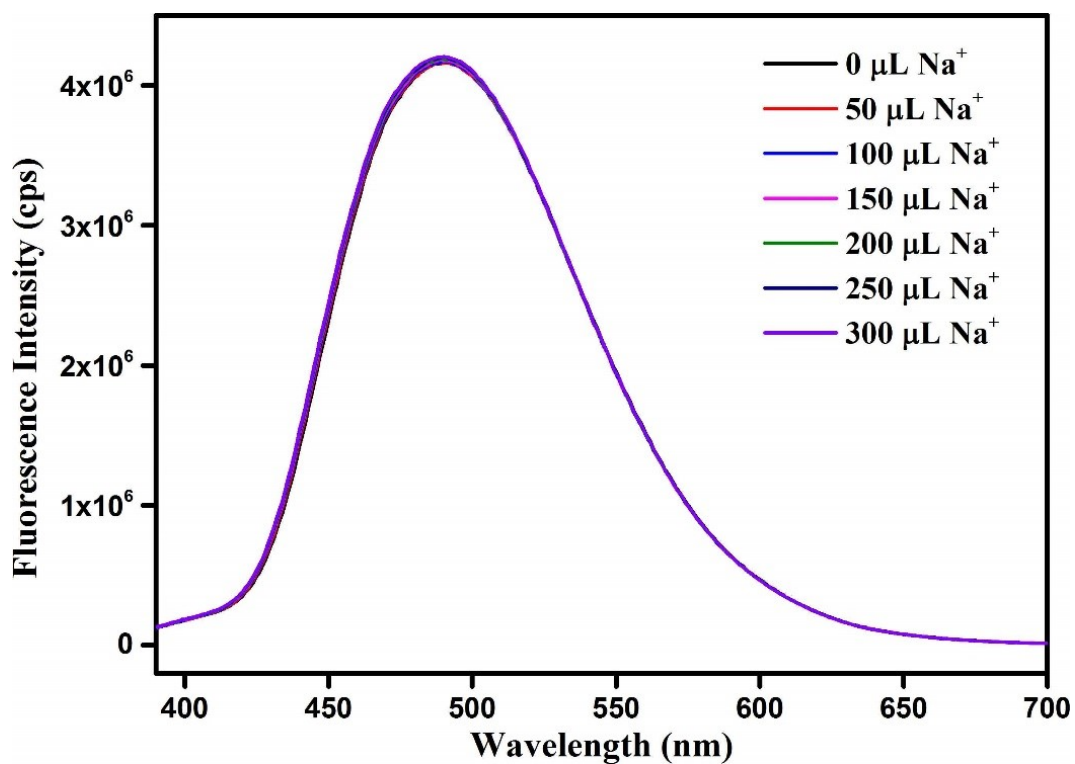
**Figure S21.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of K<sup>+</sup> (300 μL).



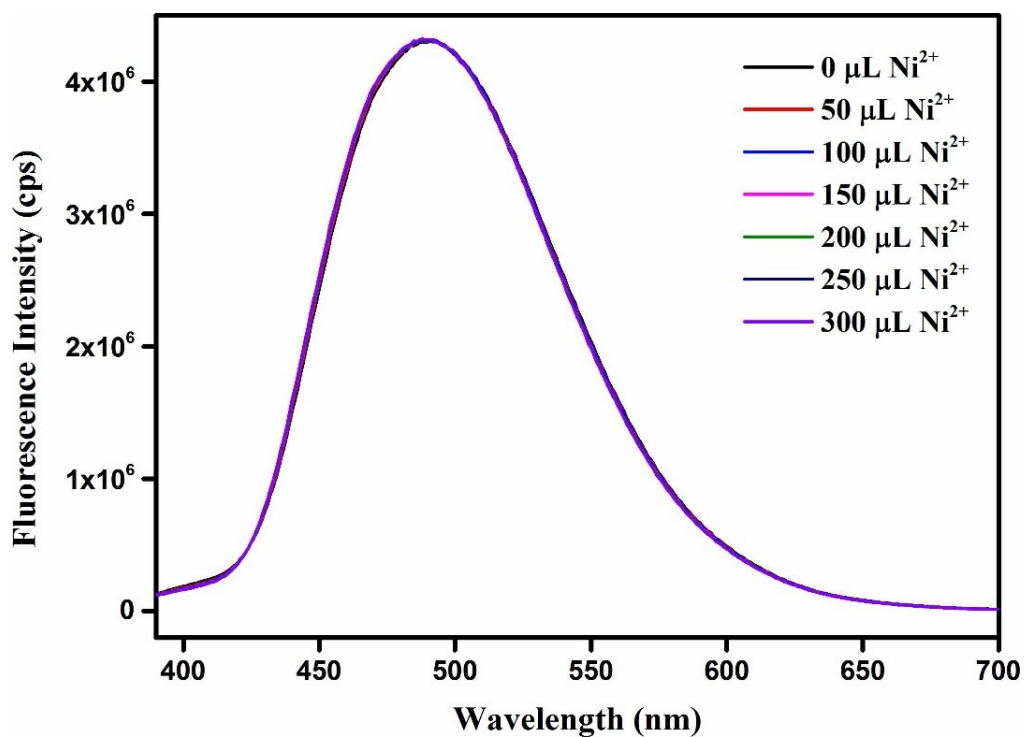
**Figure S22.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Mg<sup>2+</sup> (300 μL).



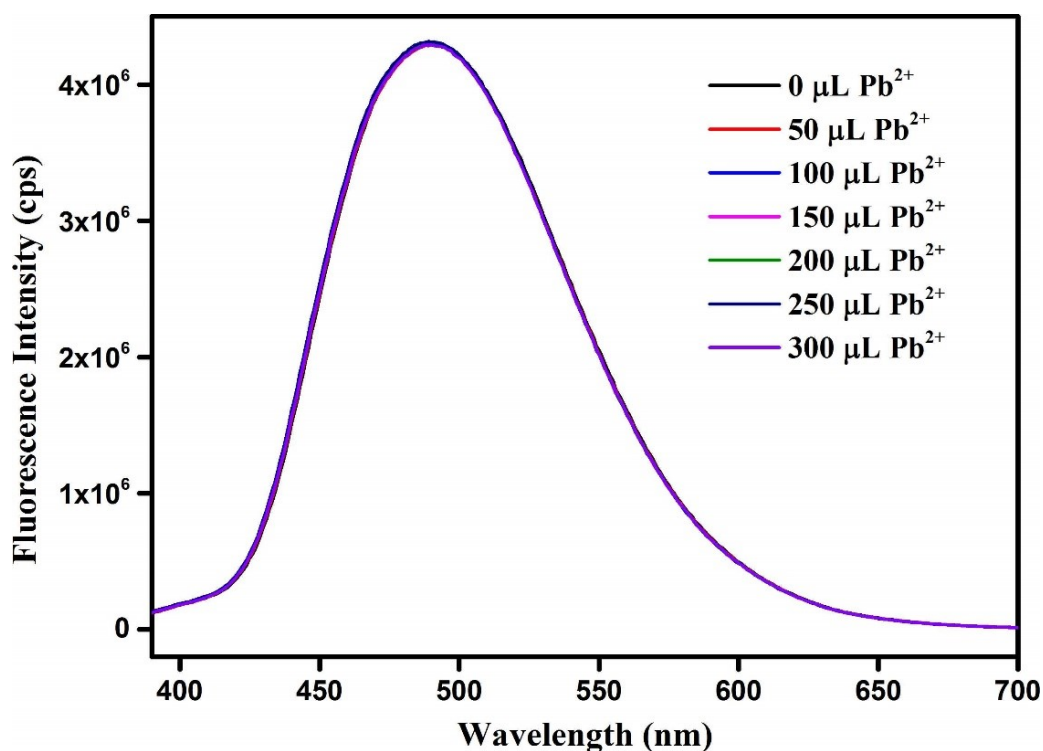
**Figure S23.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Mn<sup>2+</sup> (300  $\mu\text{L}$ ).



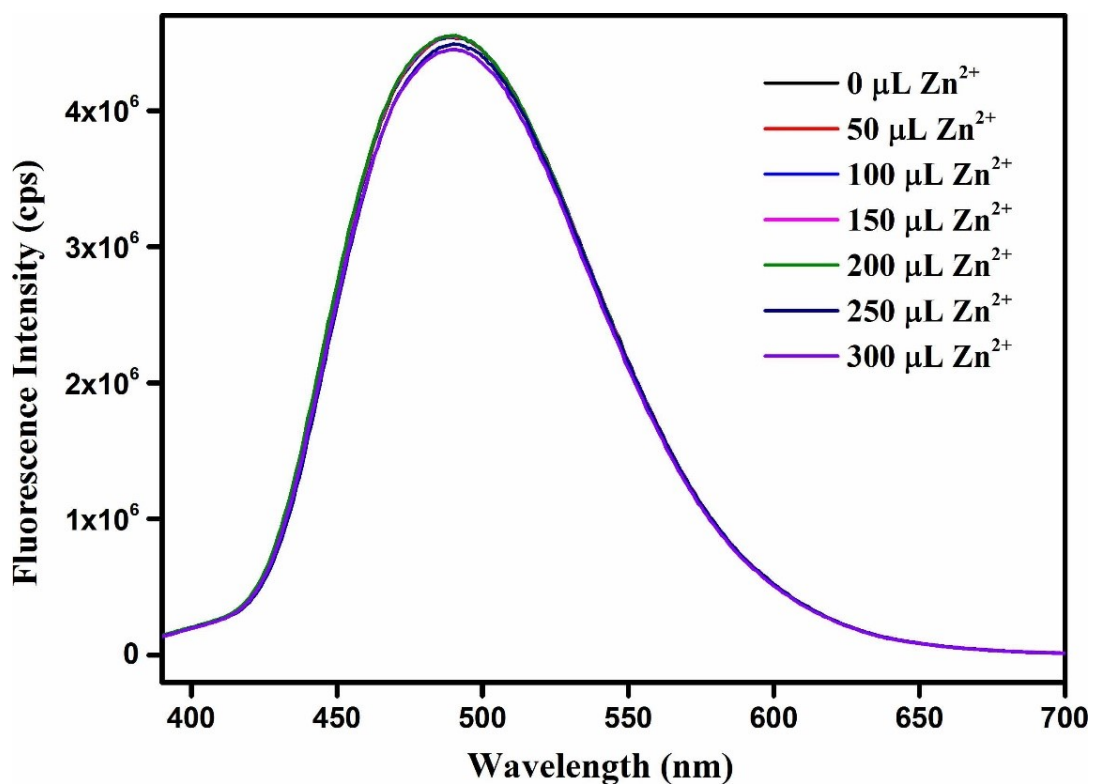
**Figure S24.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Na<sup>+</sup> (300  $\mu\text{L}$ ).



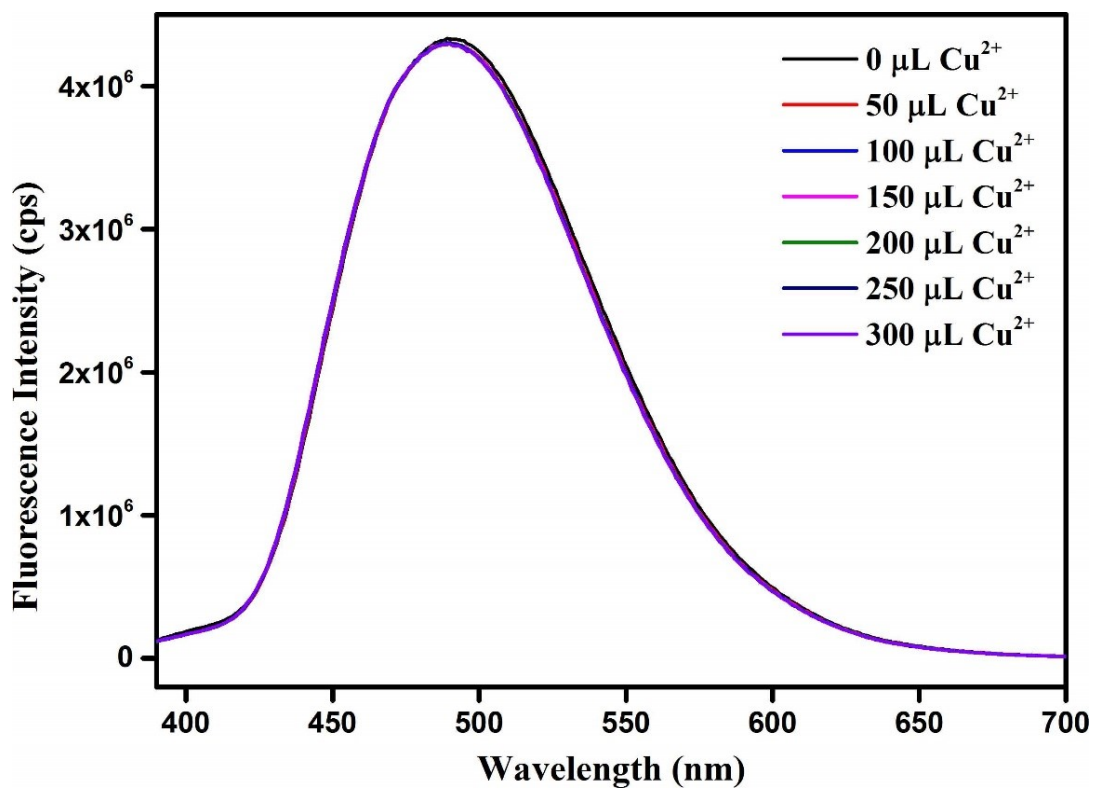
**Figure S25.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Ni<sup>+</sup> (300 μL).



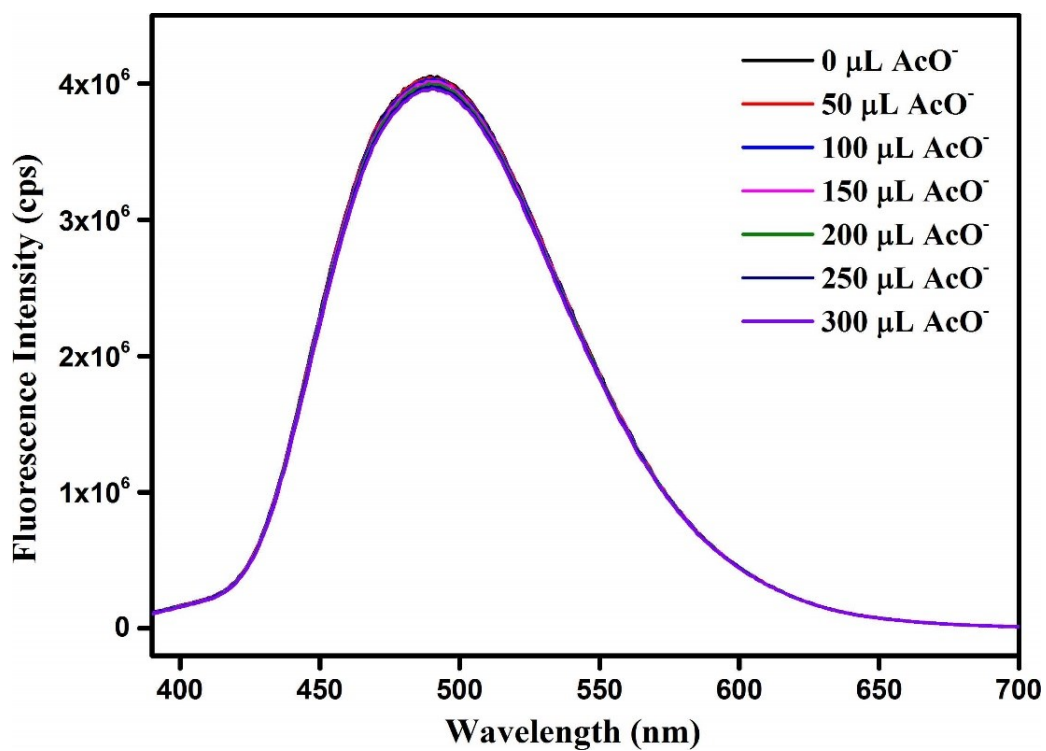
**Figure S26.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Pb<sup>2+</sup> (300 μL).



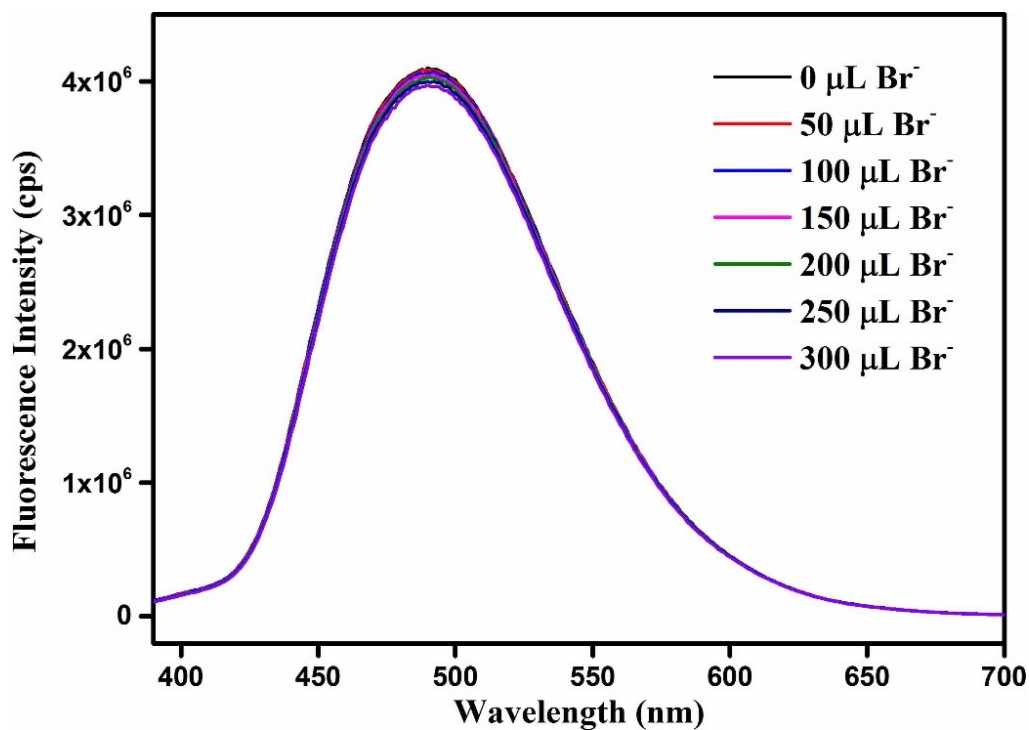
**Figure S27.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Zn<sup>2+</sup> (300 µL).



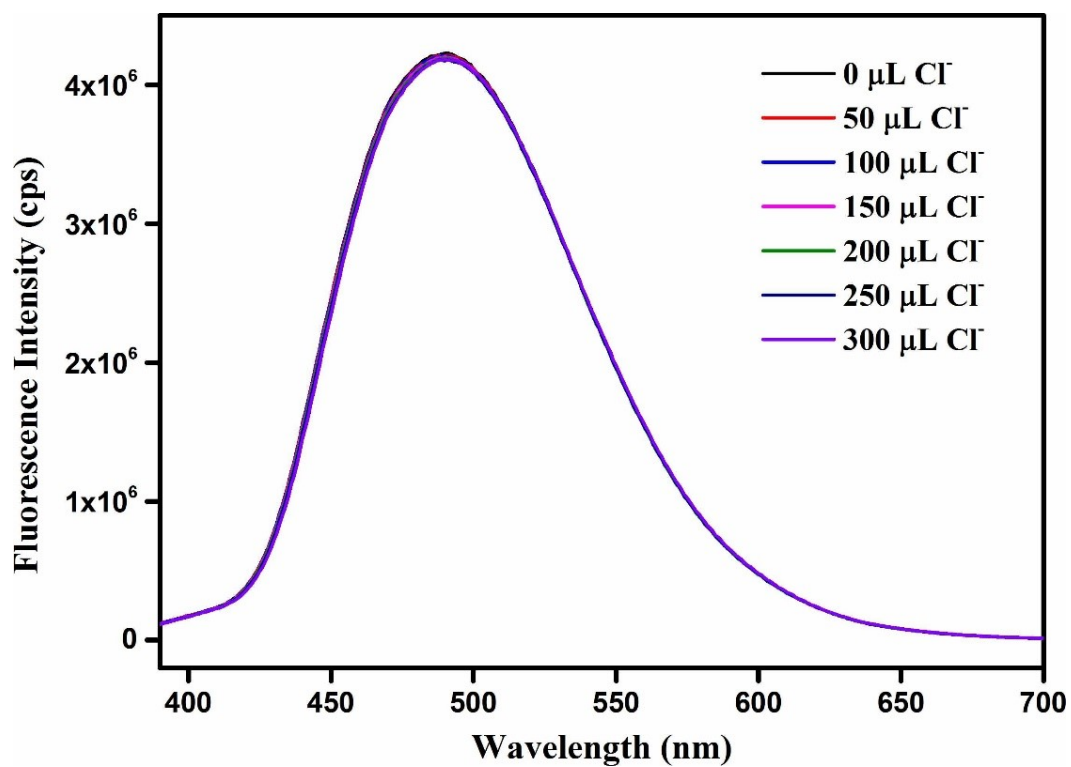
**Figure S28.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Cu<sup>2+</sup> (300 µL).



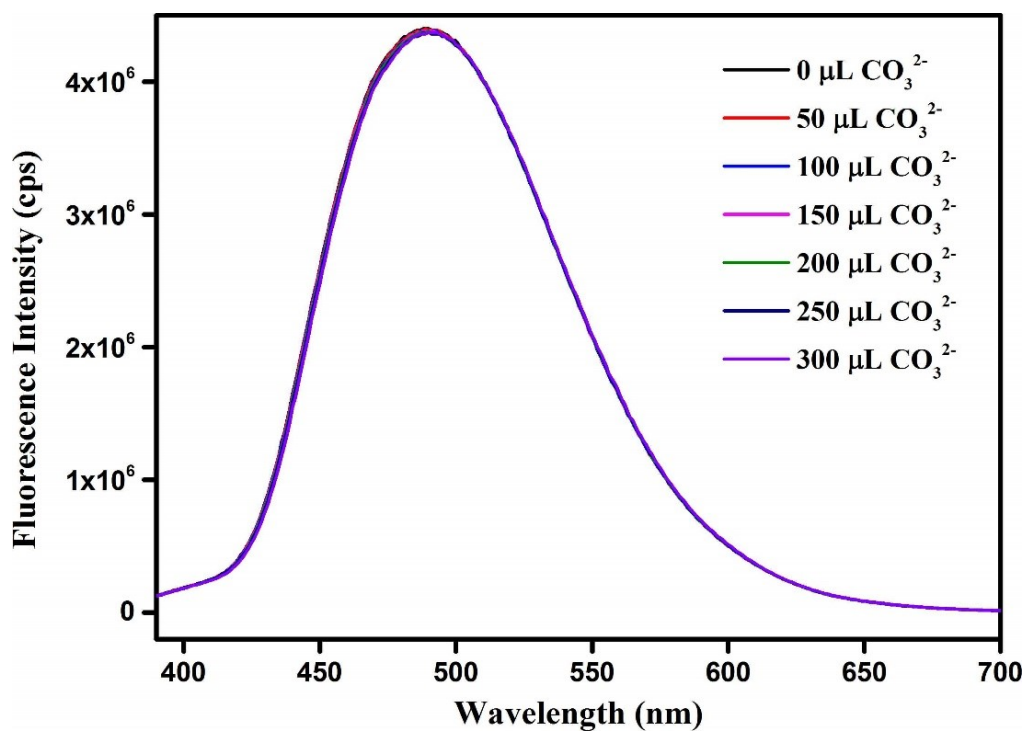
**Figure S29.** Change of fluorescence intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** dispersed in PBS after addition of 5 mM solution of AcO<sup>-</sup> (300 μL).



**Figure S30.** Change of fluorescence intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** dispersed in PBS after addition of 5 mM solution of Br<sup>-</sup> (300 μL).

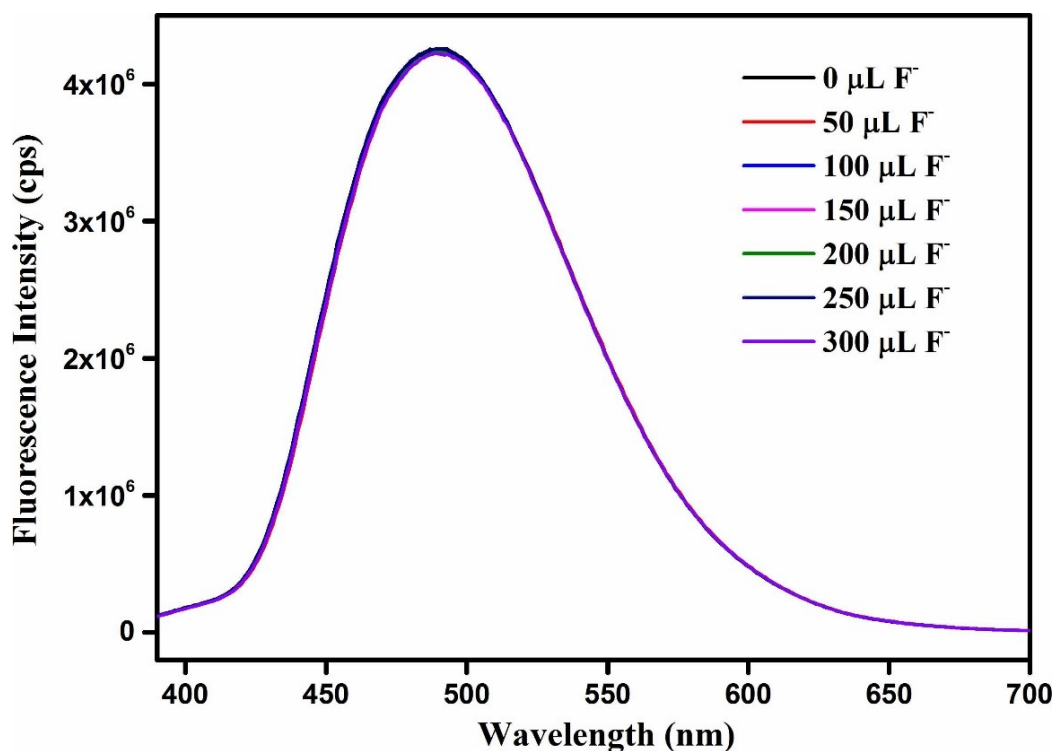


**Figure S31.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of Cl<sup>-</sup> (300 μL).

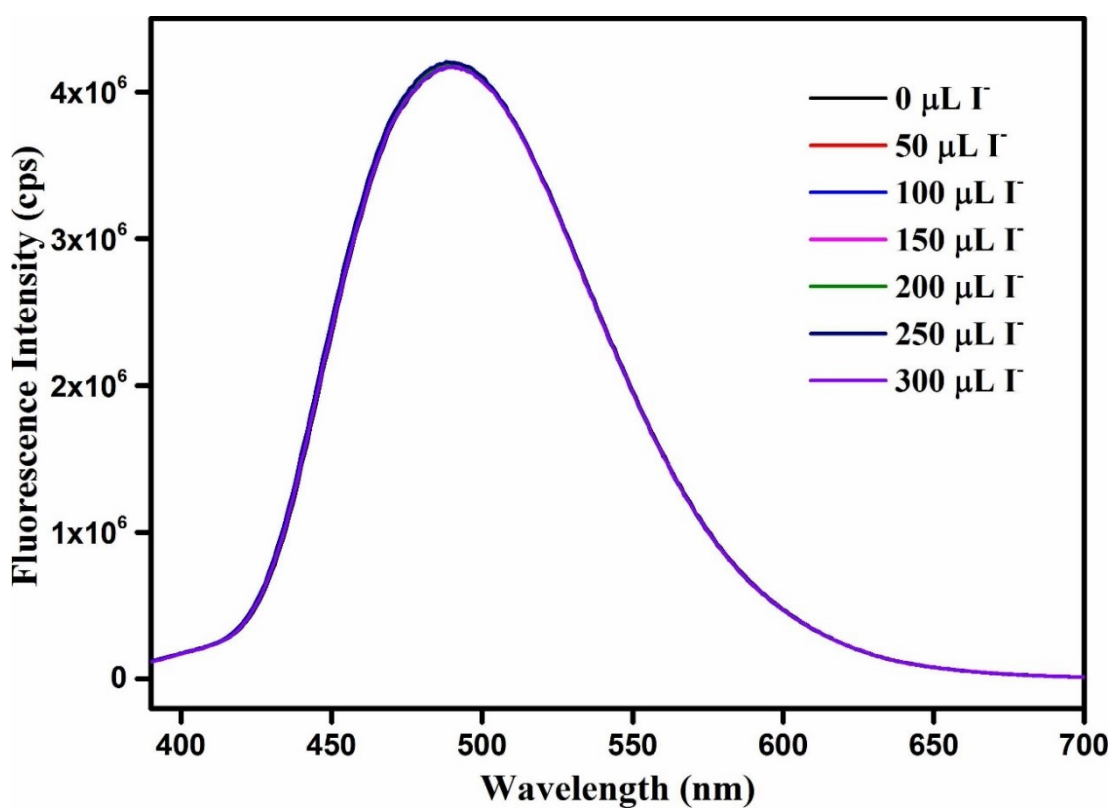


**Figure S32.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of CO<sub>3</sub><sup>2-</sup> (300 μL).



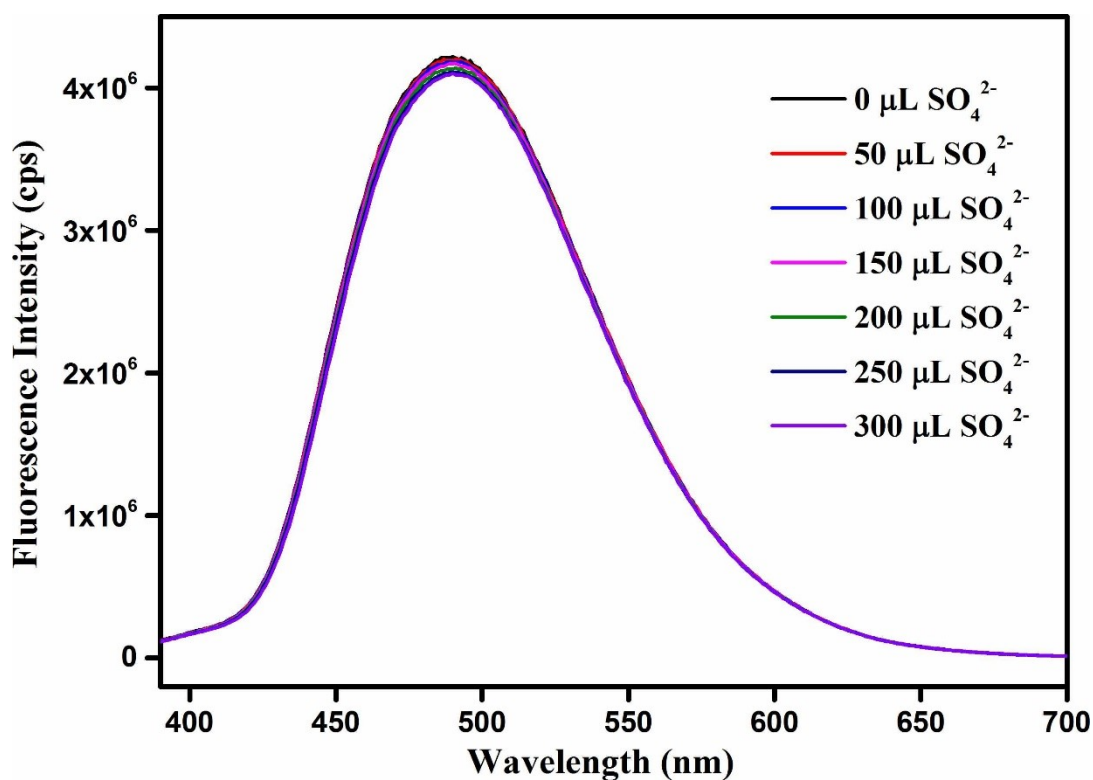


**Figure S33.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of F<sup>-</sup> (300 μL).

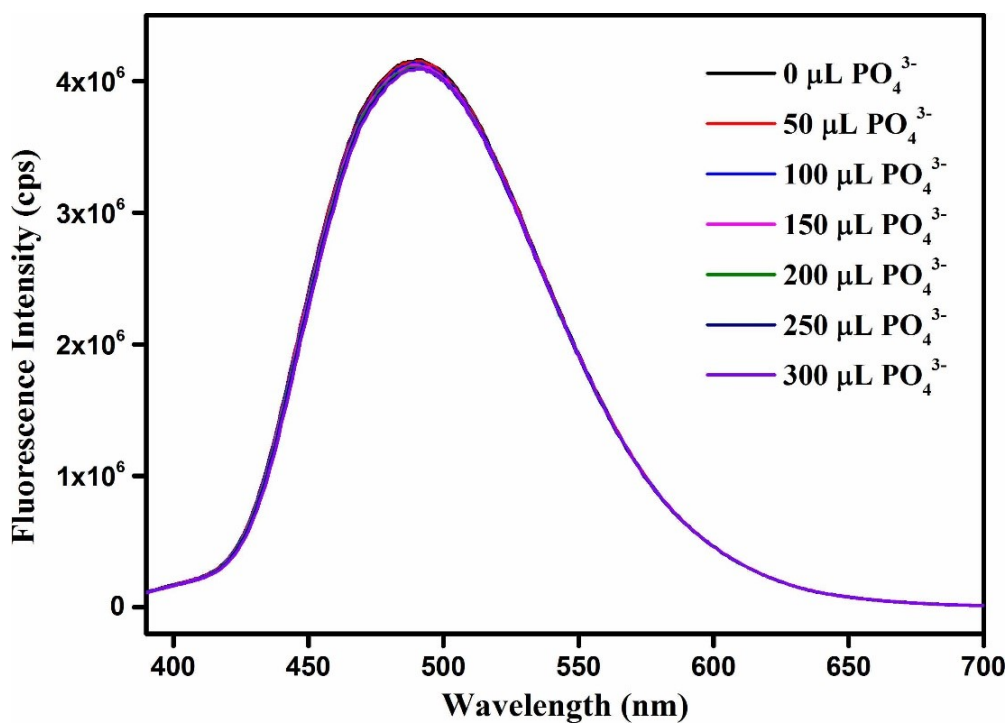


**Figure S34.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of I<sup>-</sup> (300 μL).

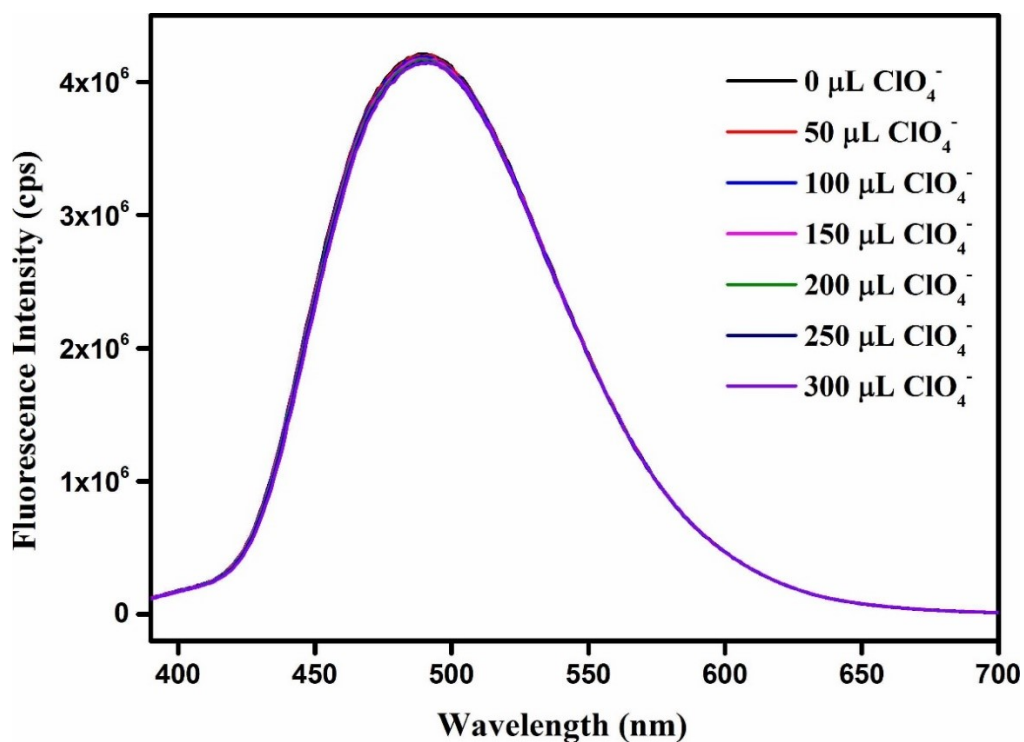




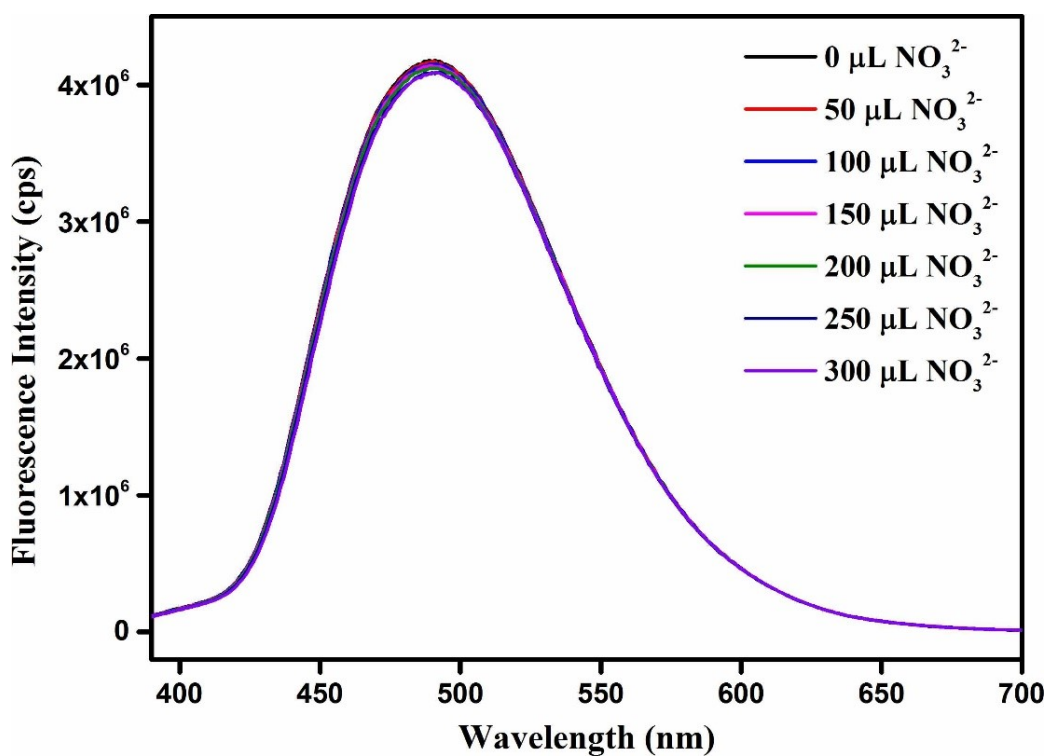
**Figure S35.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of SO<sub>4</sub><sup>2-</sup> (300 μL).



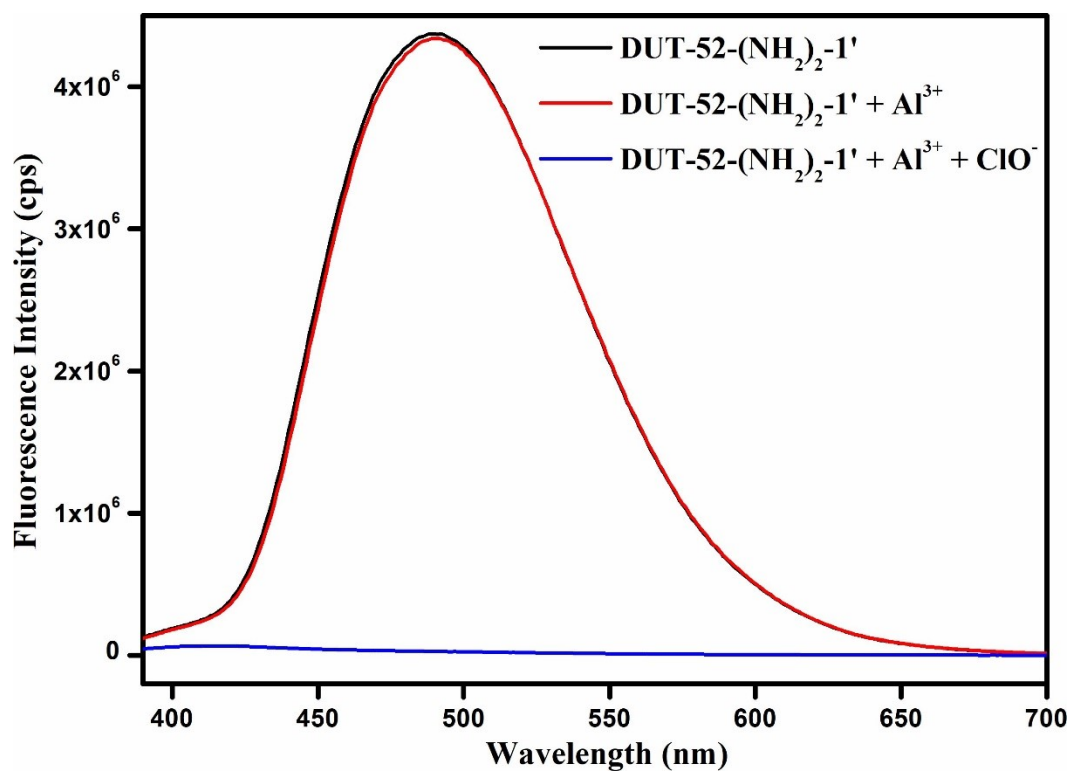
**Figure S36.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of PO<sub>4</sub><sup>3-</sup> (300 μL).



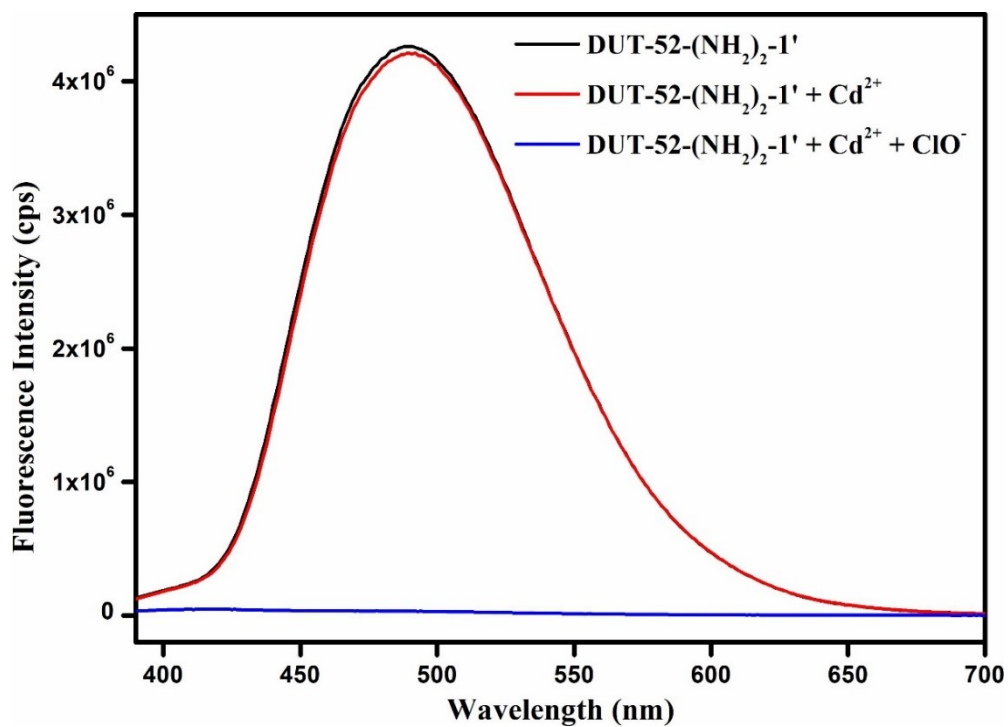
**Figure S37.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of ClO<sub>4</sub><sup>-</sup> (300 μL).



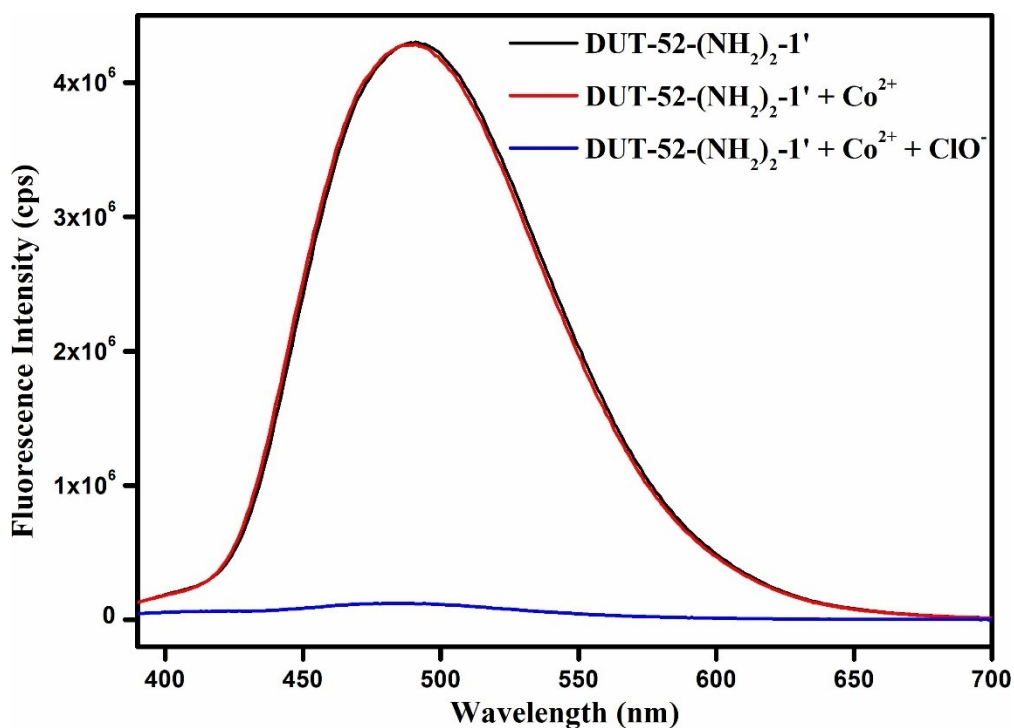
**Figure S38.** Change of fluorescence intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' dispersed in PBS after addition of 5 mM solution of NO<sub>3</sub><sup>2-</sup> (300 μL).



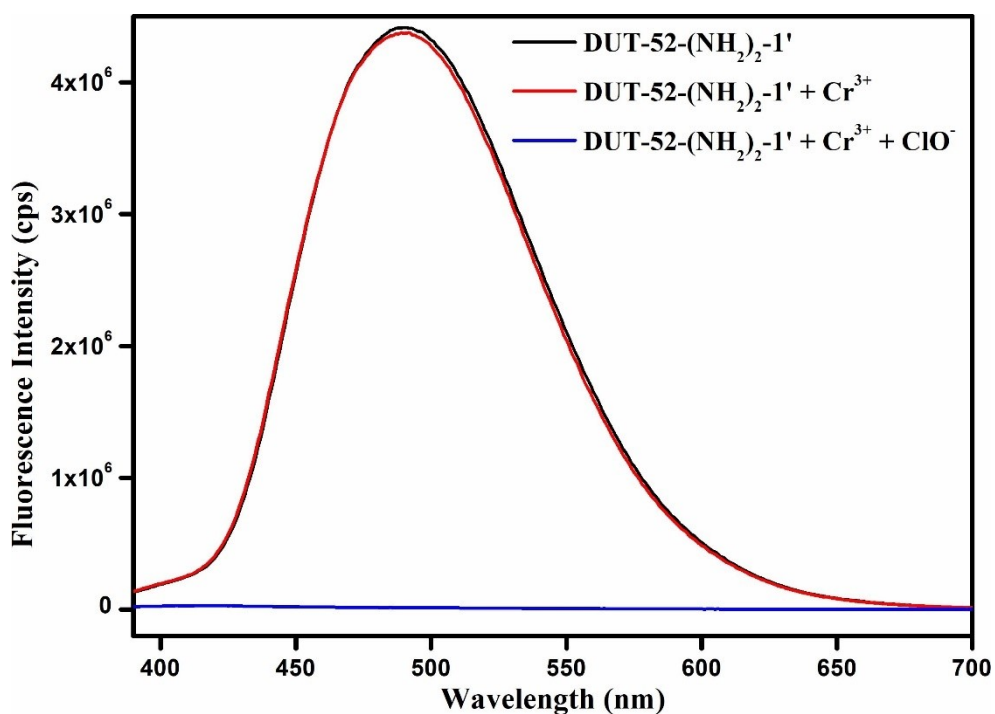
**Figure S39.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Al<sup>3+</sup> (300 μL) solution.



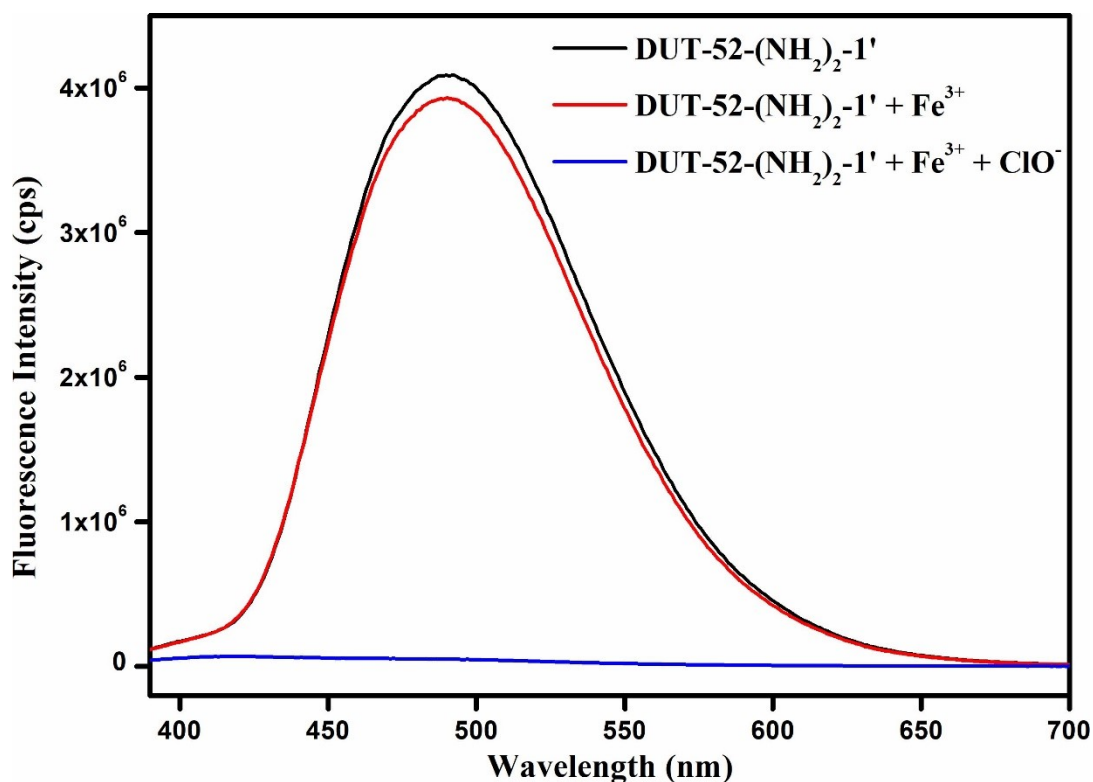
**Figure S40.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Cd<sup>2+</sup> (300 μL) solution.



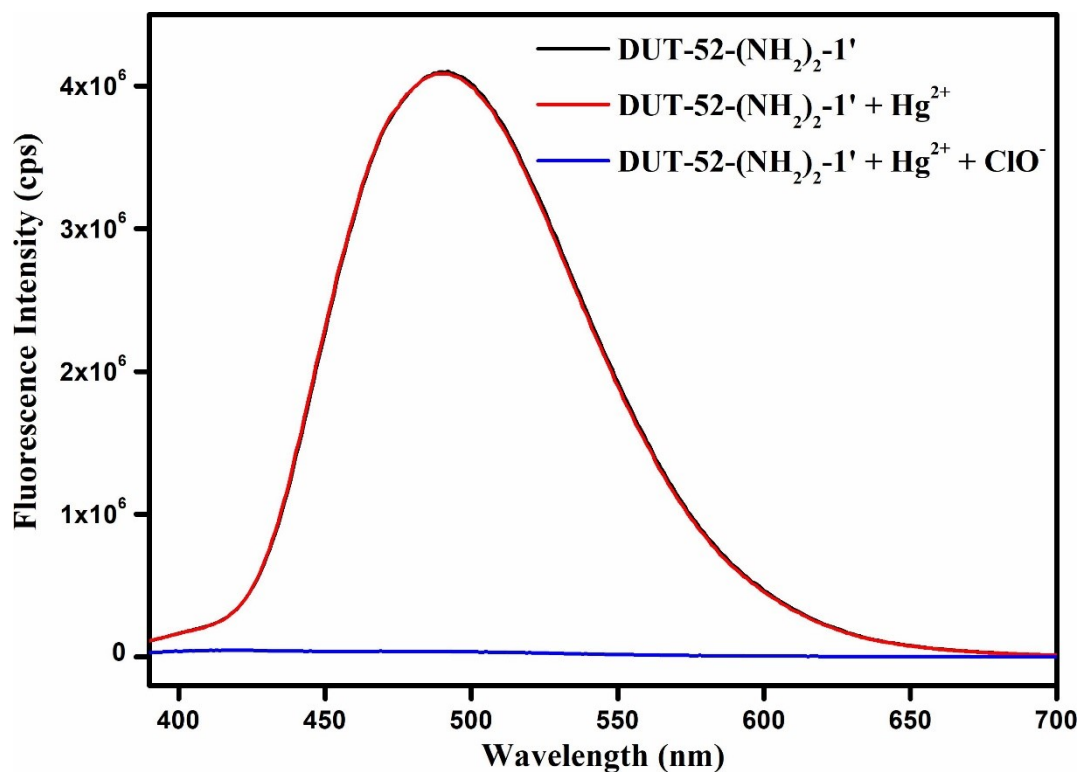
**Figure S41.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Co<sup>2+</sup> (300 μL) solution.



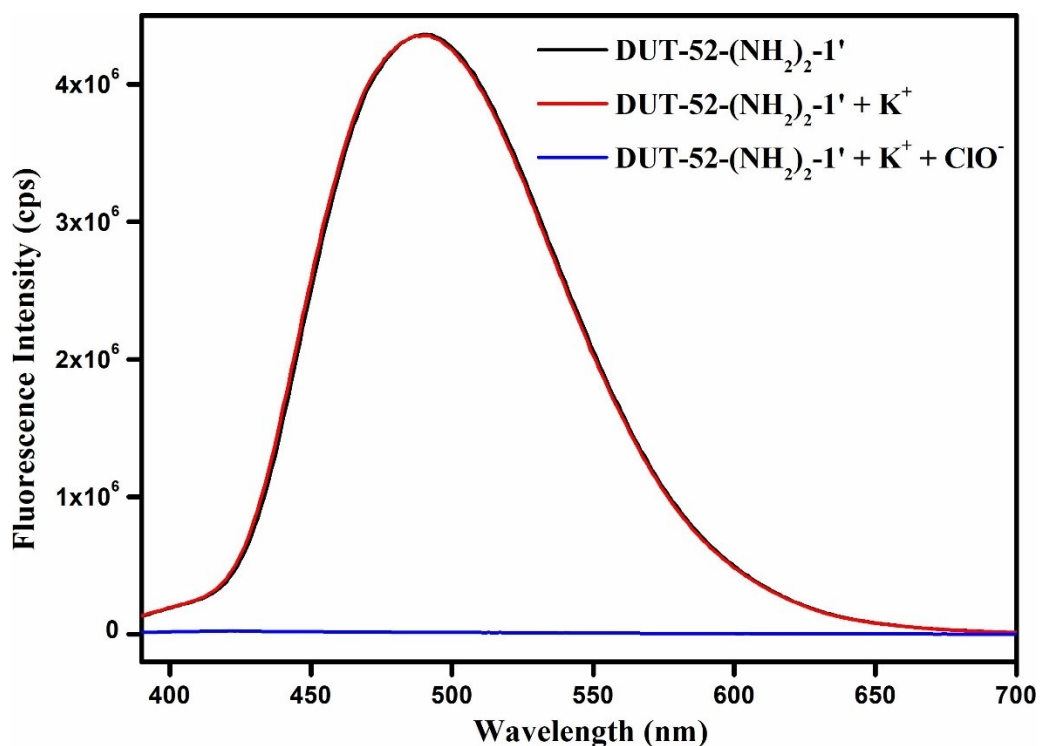
**Figure S42.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Cr<sup>3+</sup> (300 μL) solution.



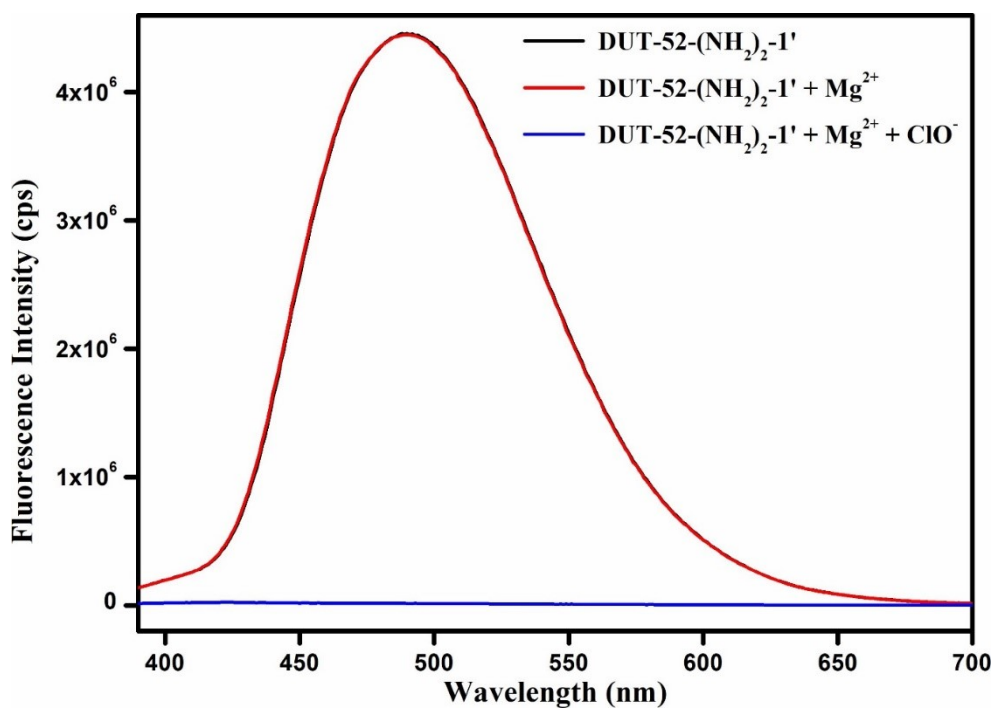
**Figure S43.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Fe<sup>3+</sup> (300 μL) solution.



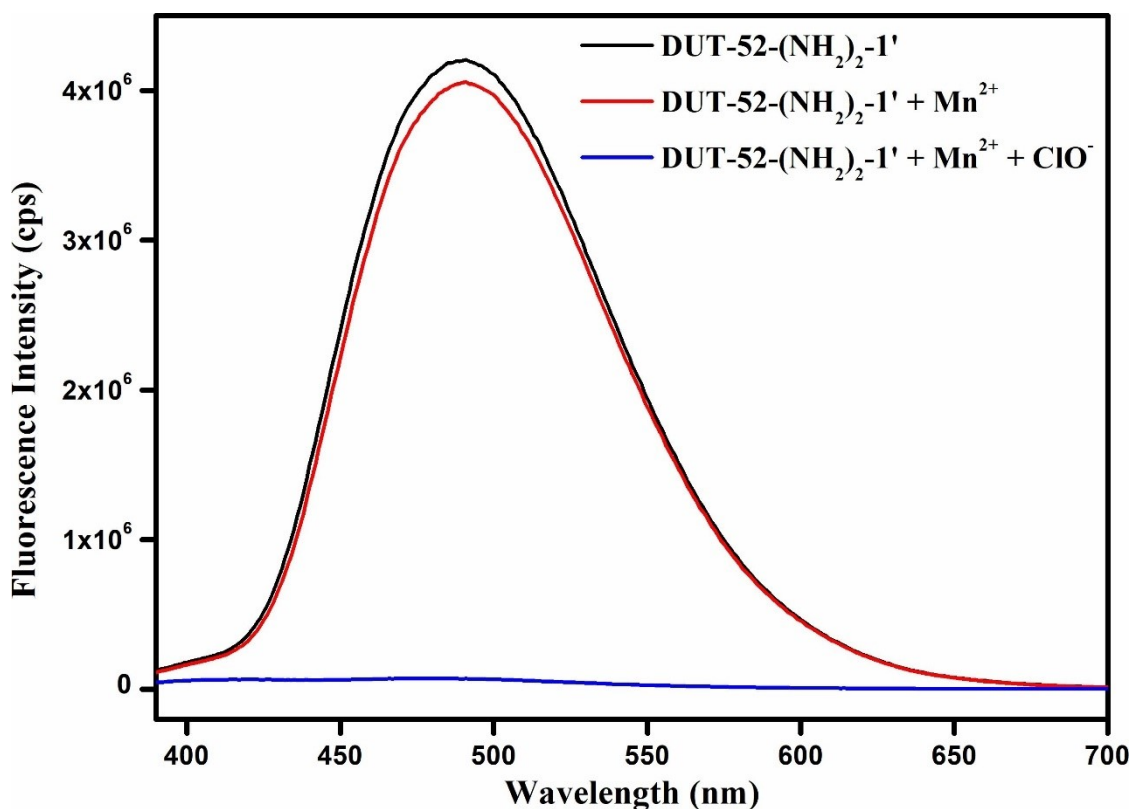
**Figure S44.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Hg<sup>2+</sup> (300 μL) solution.



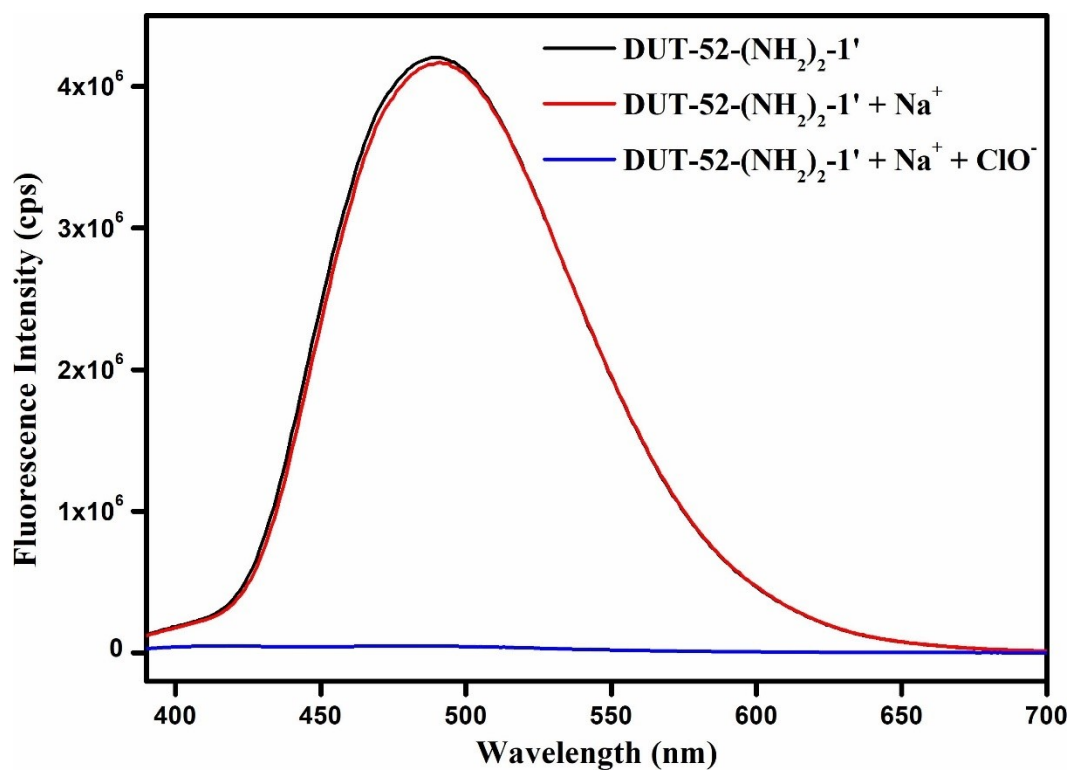
**Figure S45.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM K<sup>+</sup> (300 μL) solution.



**Figure S46.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Mg<sup>2+</sup> (300 μL) solution.

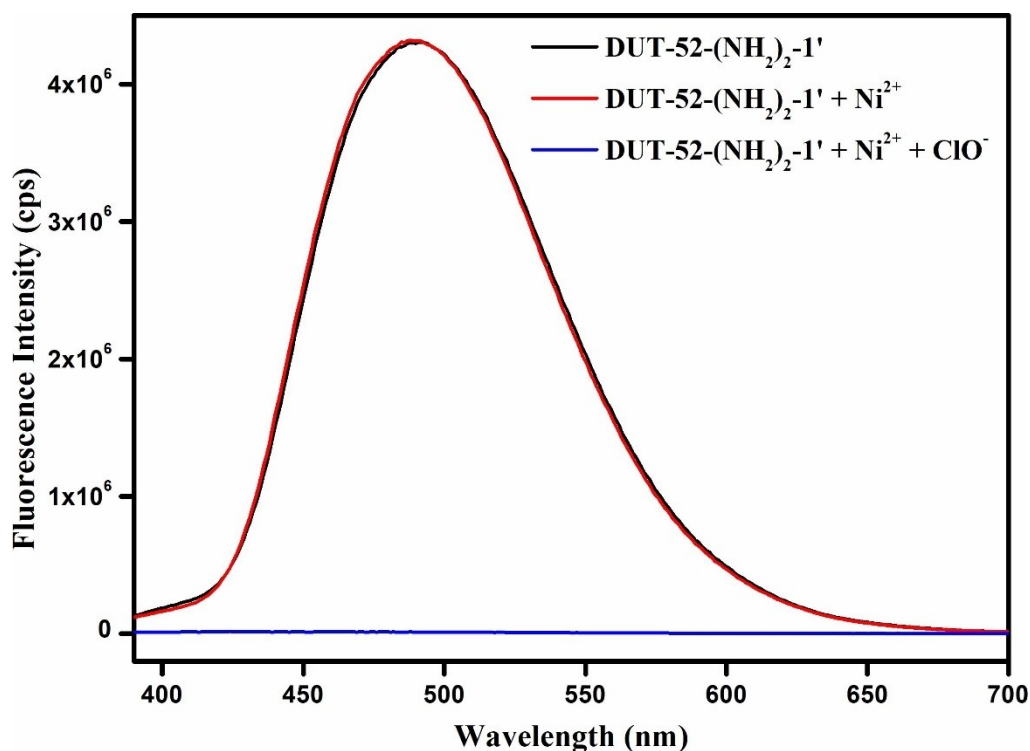


**Figure S47.** Change in the fluorescence emission intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Mn<sup>2+</sup> (300 μL) solution.

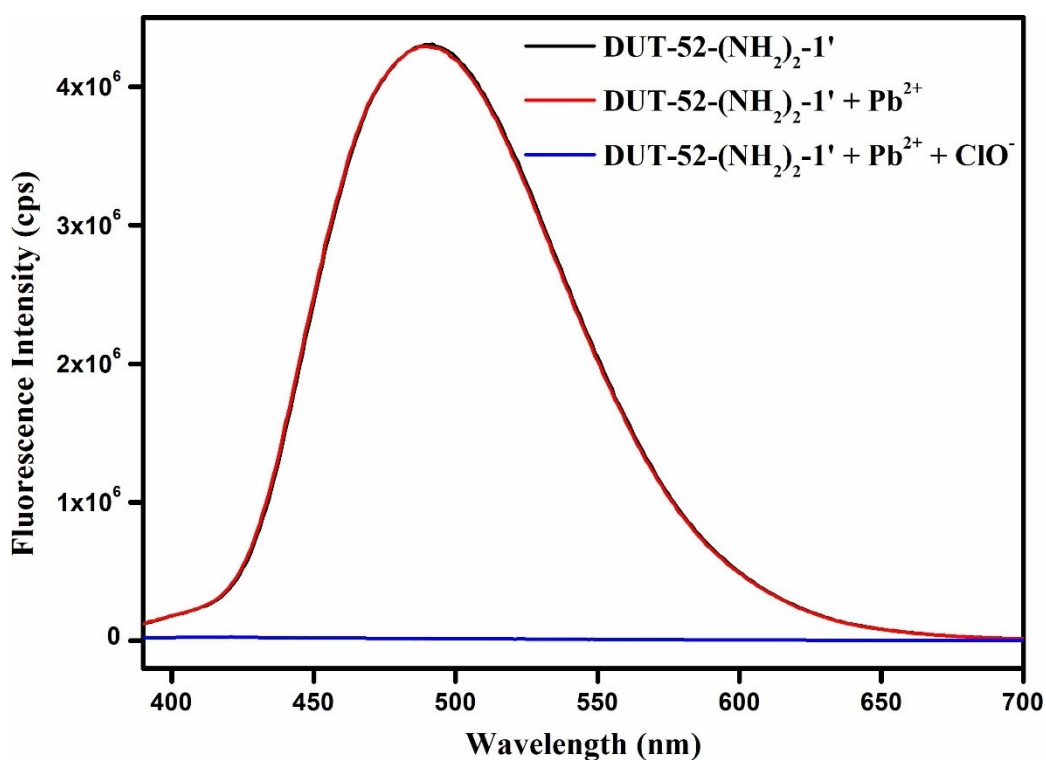


**Figure S48.** Change in the fluorescence emission intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Na<sup>+</sup> (300 μL) solution.



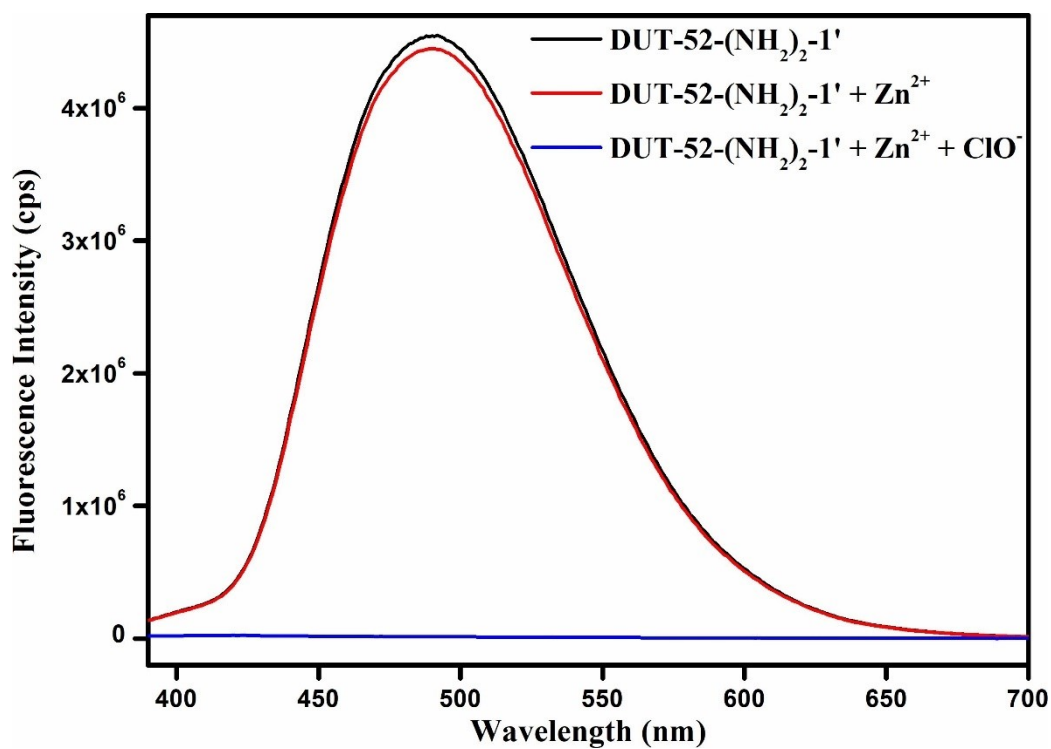


**Figure S49.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Ni<sup>2+</sup> (300 μL) solution.

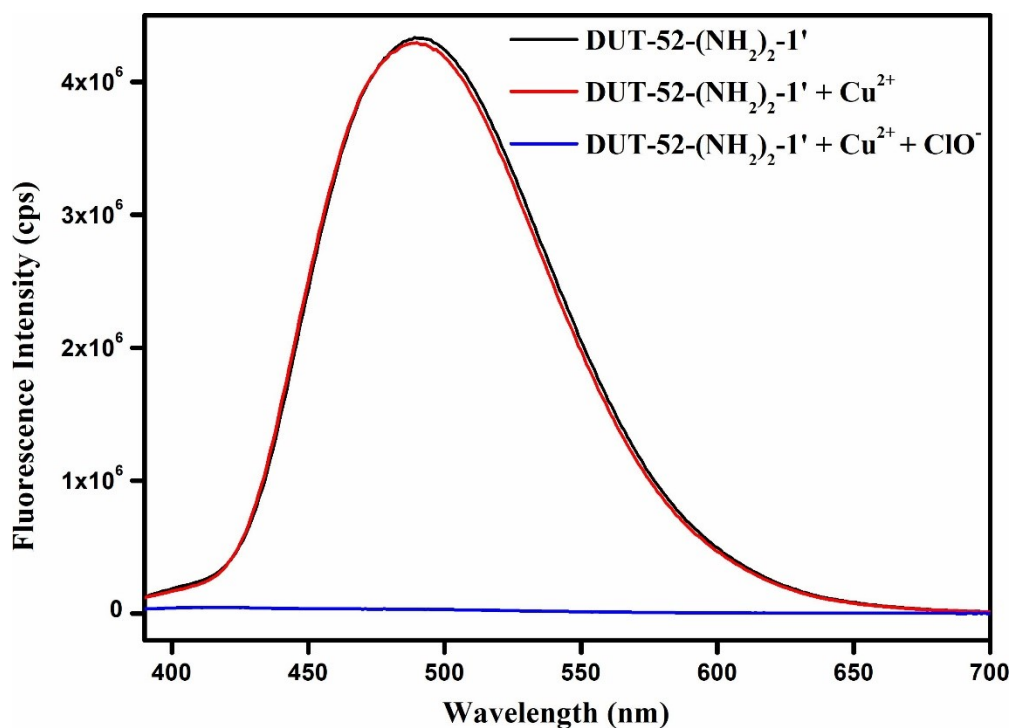


**Figure S50.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Pb<sup>2+</sup> (300 μL) solution.

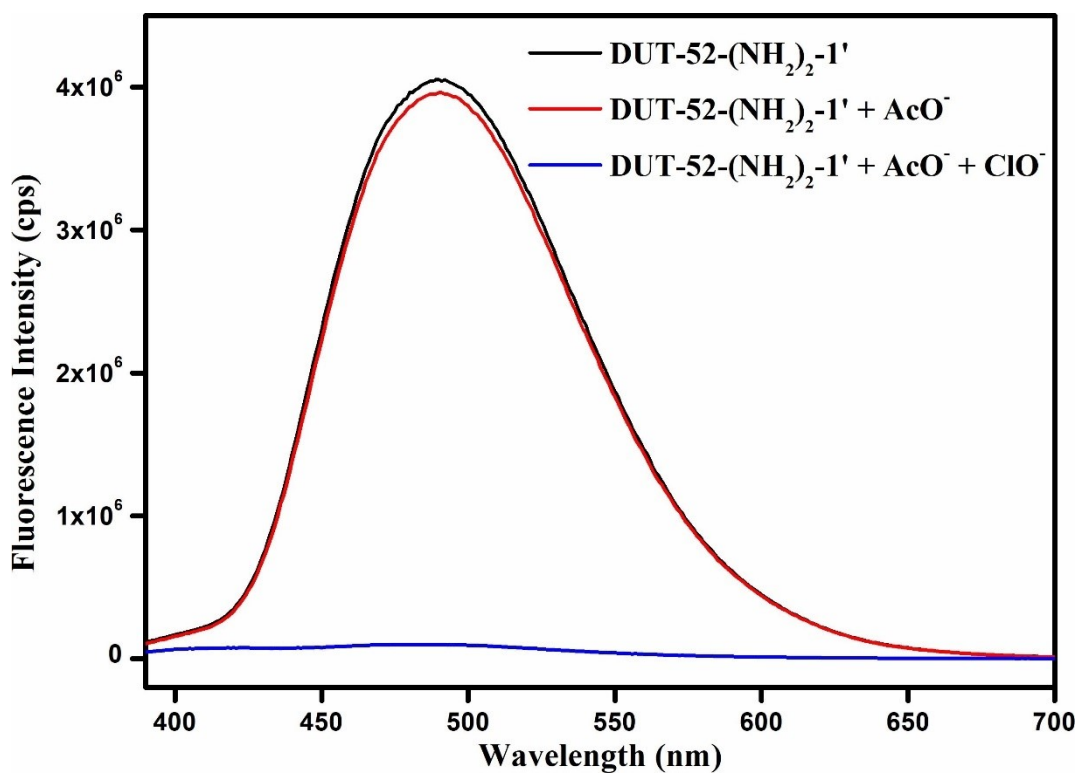




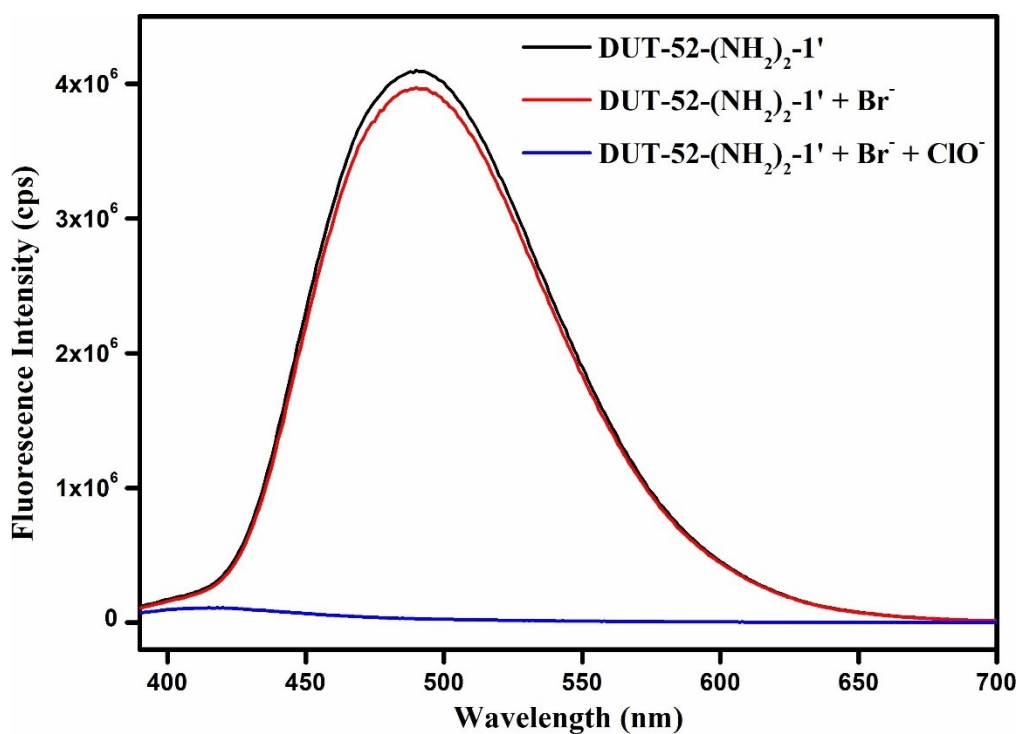
**Figure S51.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Zn<sup>2+</sup> (300 μL) solution.



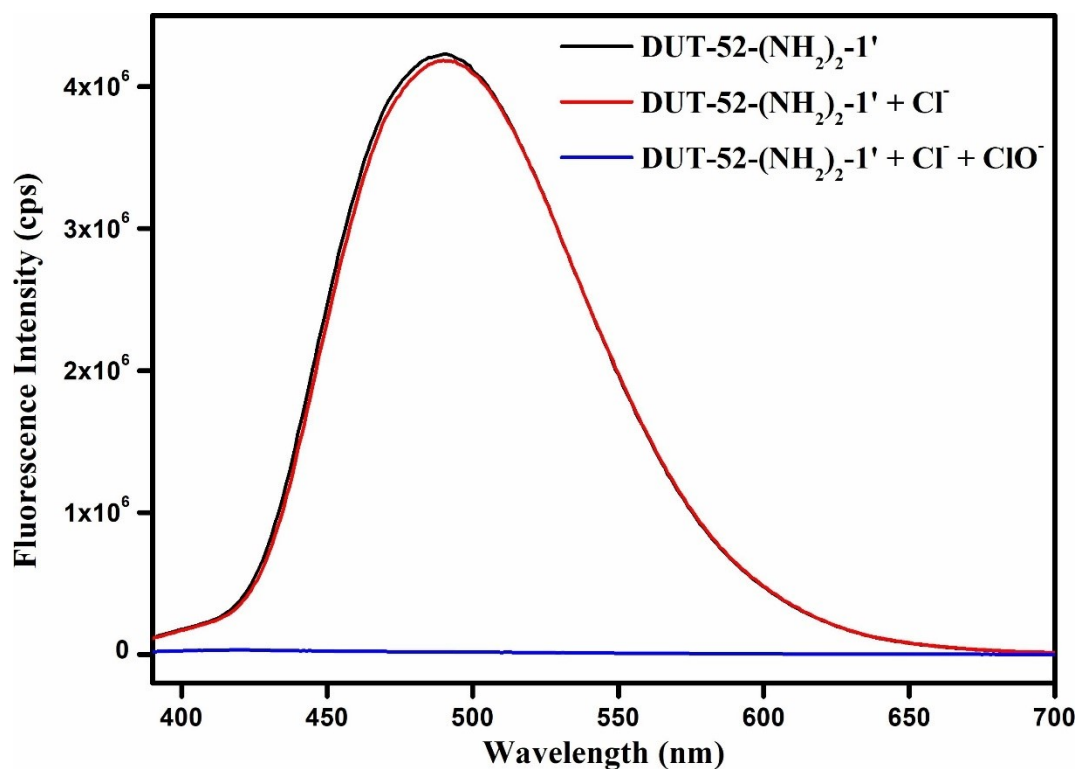
**Figure S52.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Cu<sup>2+</sup> (300 μL) solution.



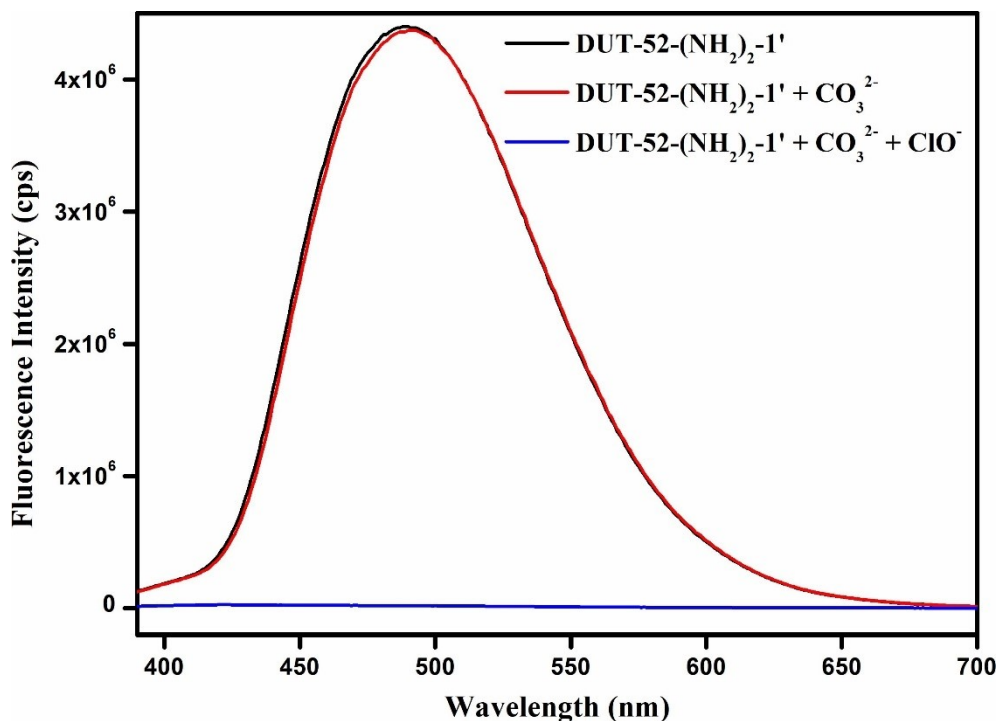
**Figure S53.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM AcO<sup>-</sup> (300 μL) solution.



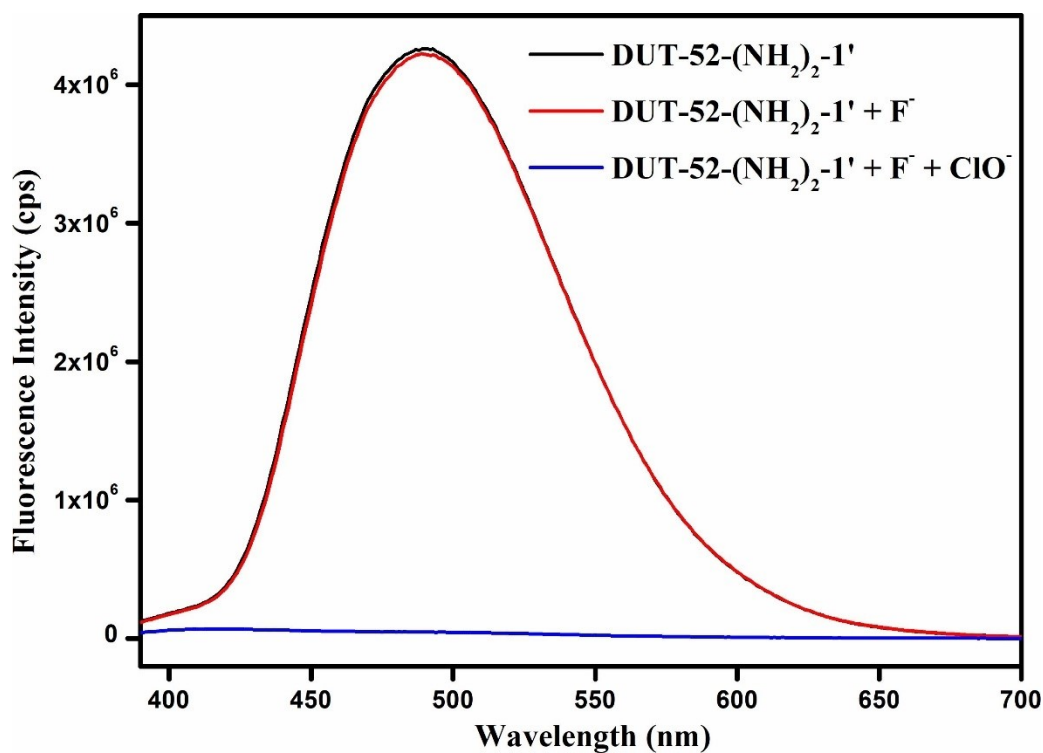
**Figure S54.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Br<sup>-</sup> (300 μL) solution.



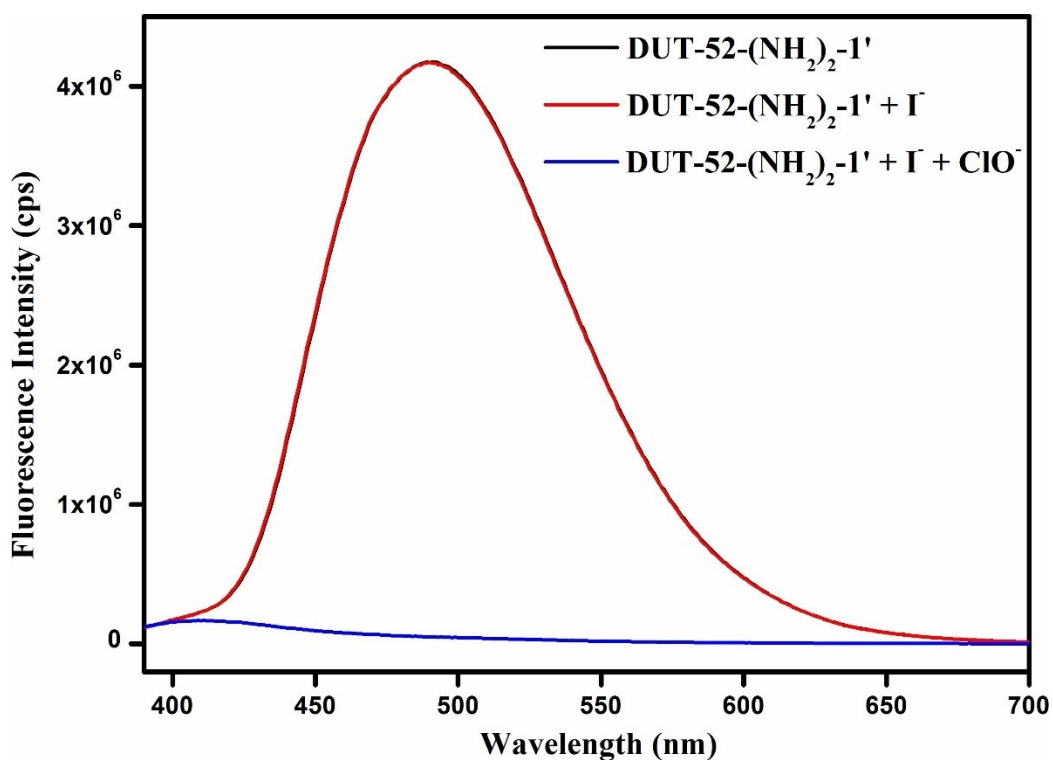
**Figure S55.** Change in the fluorescence emission intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM Cl<sup>-</sup> (300 μL) solution.



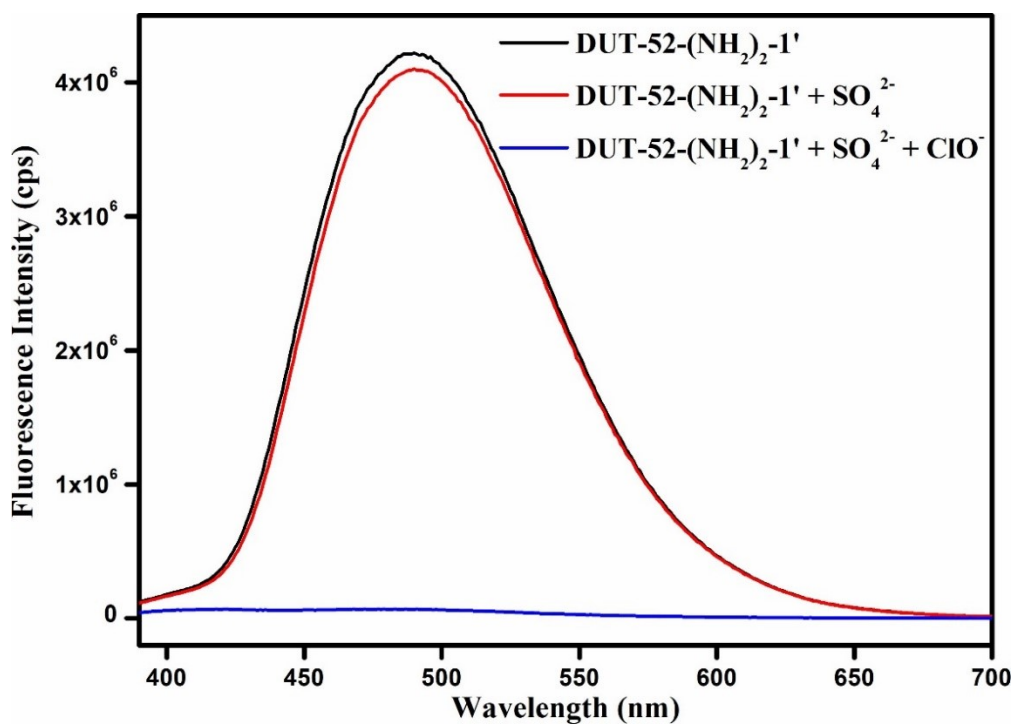
**Figure S56.** Change in the fluorescence emission intensity of DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM CO<sub>3</sub><sup>2-</sup> (300 μL) solution.



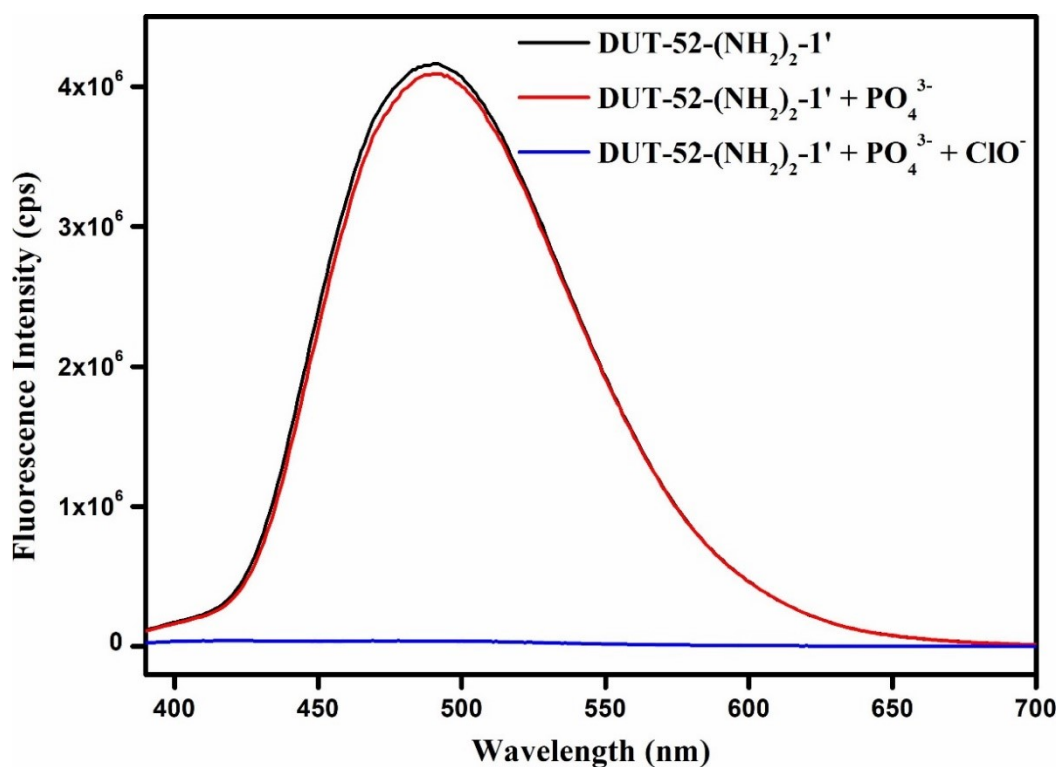
**Figure S57.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM F<sup>-</sup> (300 μL) solution.



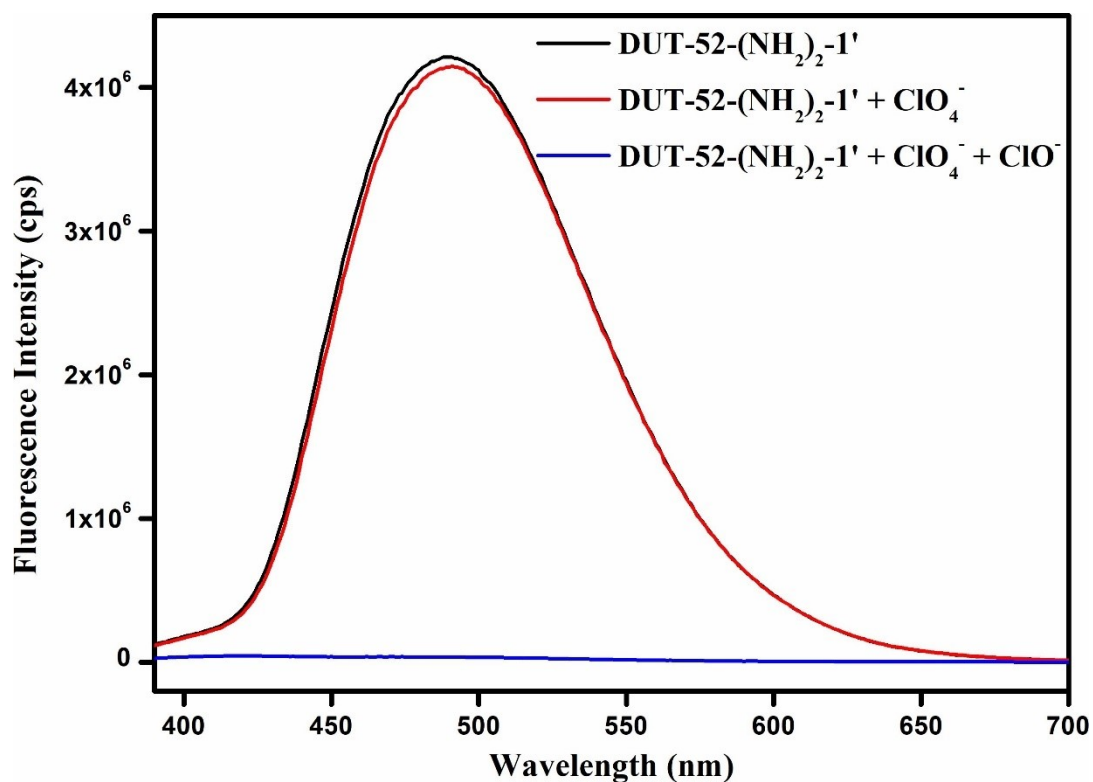
**Figure S58.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM I<sup>-</sup> (300 μL) solution.



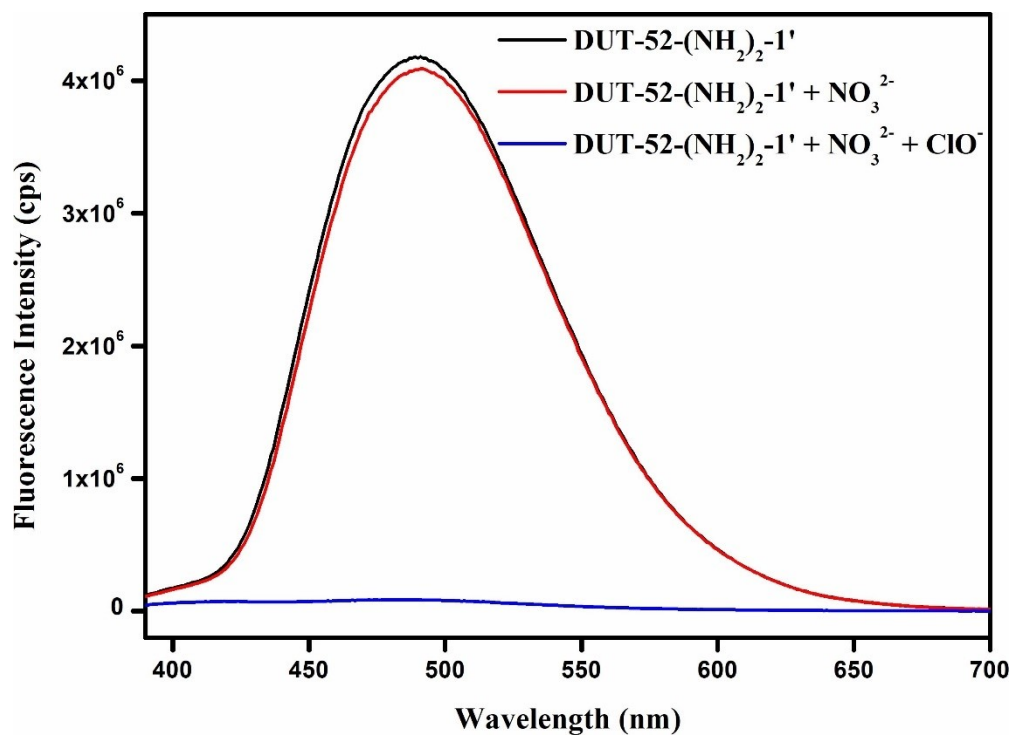
**Figure S59.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM SO<sub>4</sub><sup>2-</sup> (300 μL) solution.



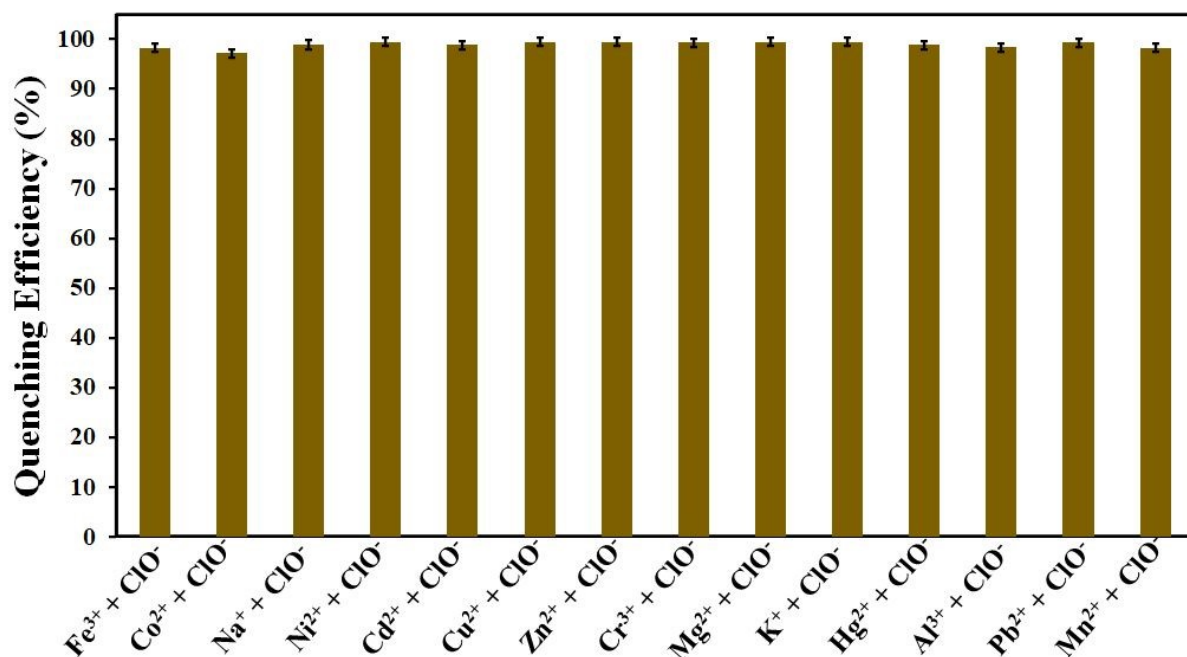
**Figure S60.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM PO<sub>4</sub><sup>3-</sup> (300 μL) solution.



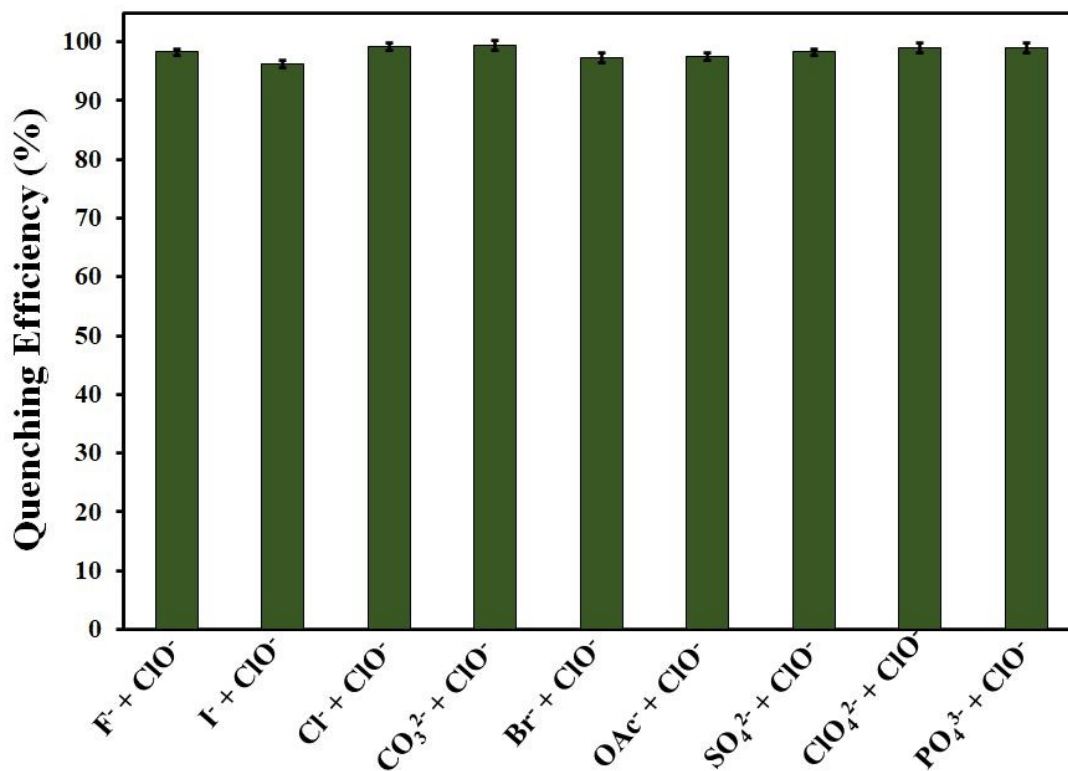
**Figure S61.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM ClO<sub>4</sub><sup>-</sup> (300 μL) solution.



**Figure S62.** Change in the fluorescence emission intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** upon addition of 5 mM ClO<sup>-</sup> solution (300 μL) in presence of 5 mM NO<sub>3</sub><sup>2-</sup> (300 μL) solution.

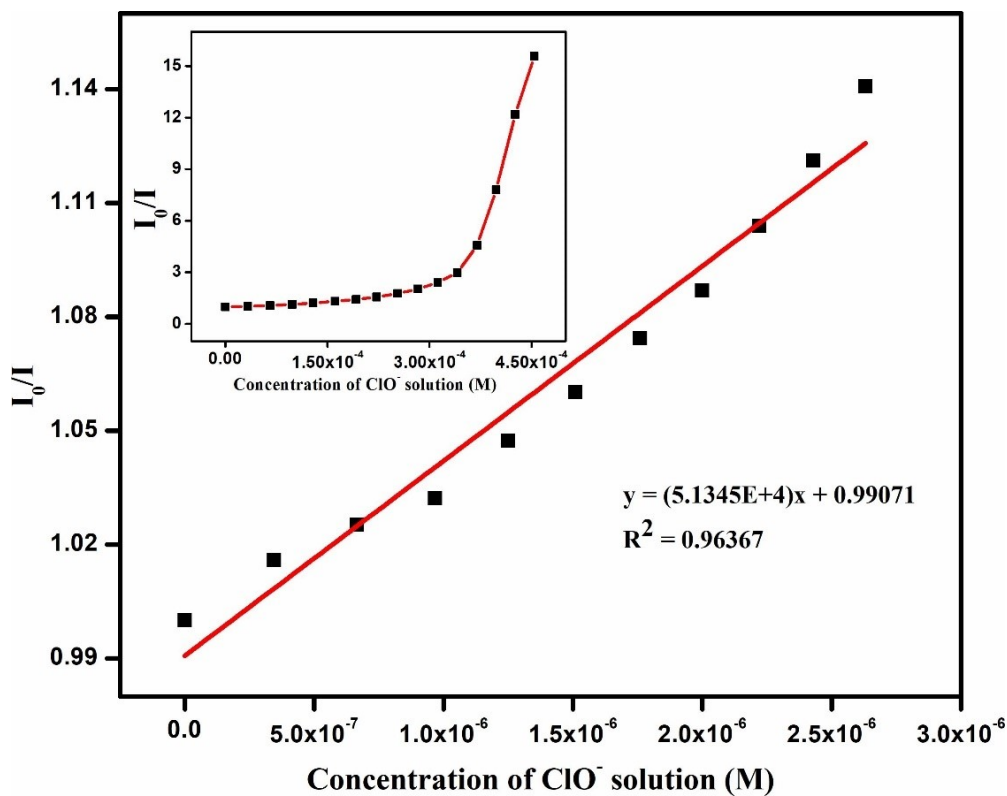


**Figure S63.** The effect of other metal anions on the quenching efficiency of ClO<sup>-</sup>. The error bars indicate the standard errors of three measurements.

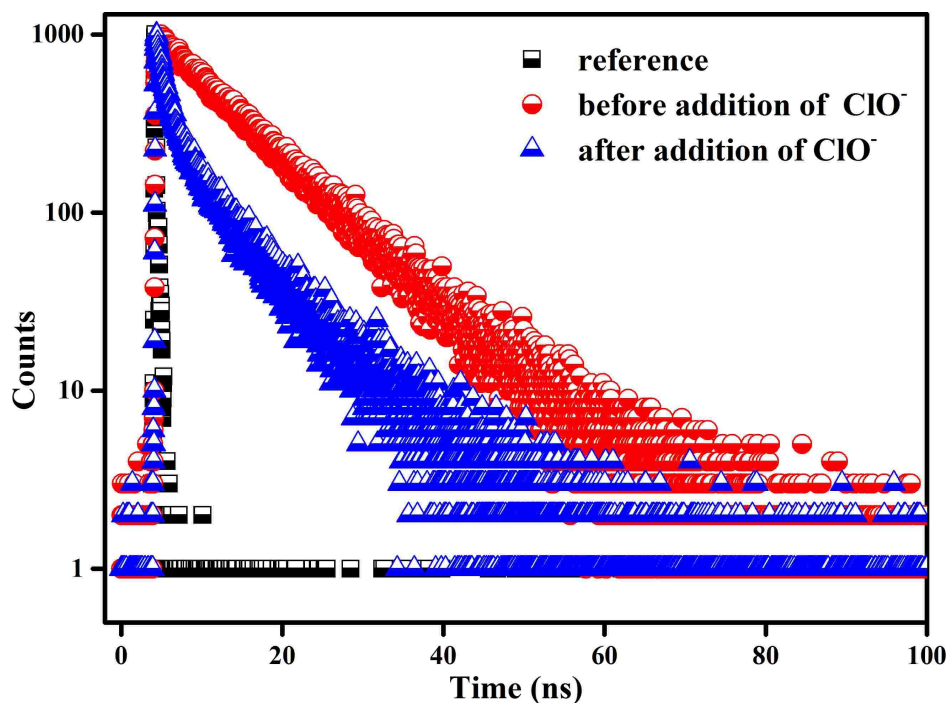


**Figure S64.** The effect of other metal anions on the quenching efficiency of ClO<sup>-</sup>. The error bars indicate the standard errors of three measurements.





**Figure S65.** Stern-Volmer plot for the fluorescence emission quenching of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** in presence of  $\text{ClO}^-$  solution.



**Figure S66.** Lifetime decay profiles of PBS buffer suspension of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** in absence and presence of  $\text{ClO}^-$  solution.



**Table S2.** Average excited state lifetime ( $\langle\tau\rangle$ ) values of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** before and after addition of 300  $\mu\text{L}$  of 5 mM  $\text{ClO}^-$  solution ( $\lambda_{\text{ex}} = 375 \text{ nm}$ ).

Volume Added ( $\mu\text{L}$ )	$B_1$	$B_2$	$a_1$	$a_2$	$\tau_1$ (ns)	$\tau_2$ (ns)	$\langle\tau\rangle^*$ (ns)	$\chi^2$
0	0.005	0.121	0.007	0.993	1.80	10.07	10.01	1.05
300	0.271	0.023	0.633	0.367	0.79	5.32	2.45	1.05

$$* \langle\tau\rangle = a_1\tau_1 + a_2\tau_2$$

Where,  $a_1$  = amplitude of the first component of the bi-exponential curve.

$a_2$  = amplitude of the second component of the bi-exponential curve.

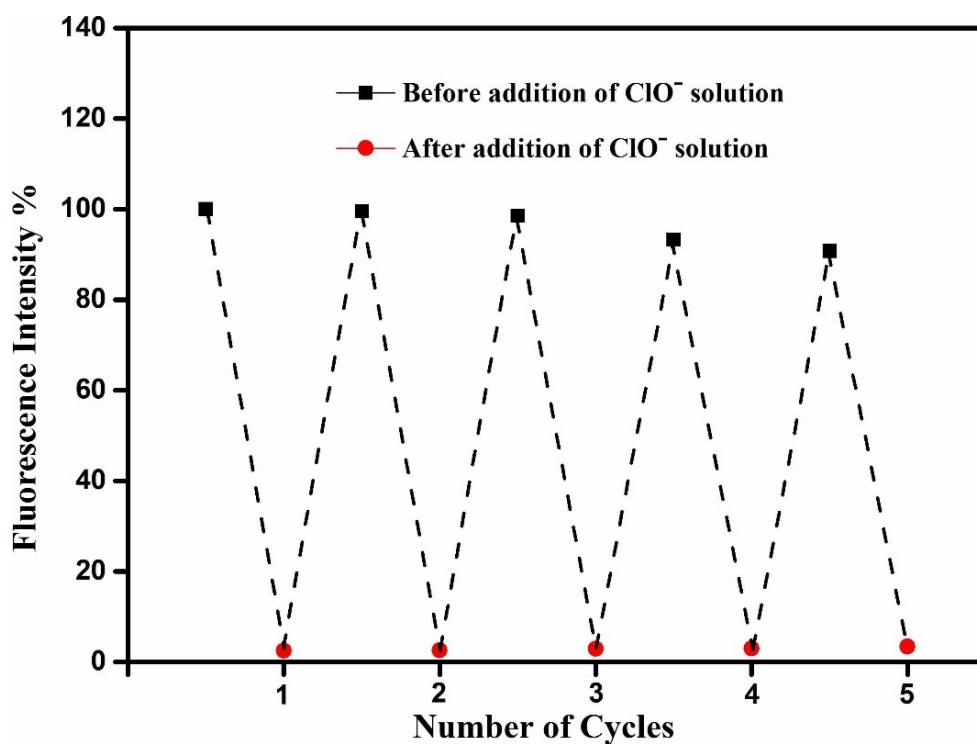
$\tau_1$  = life time of the first component of the bi-exponential curve.

$\tau_2$  = life time of the second component of the bi-exponential curve.

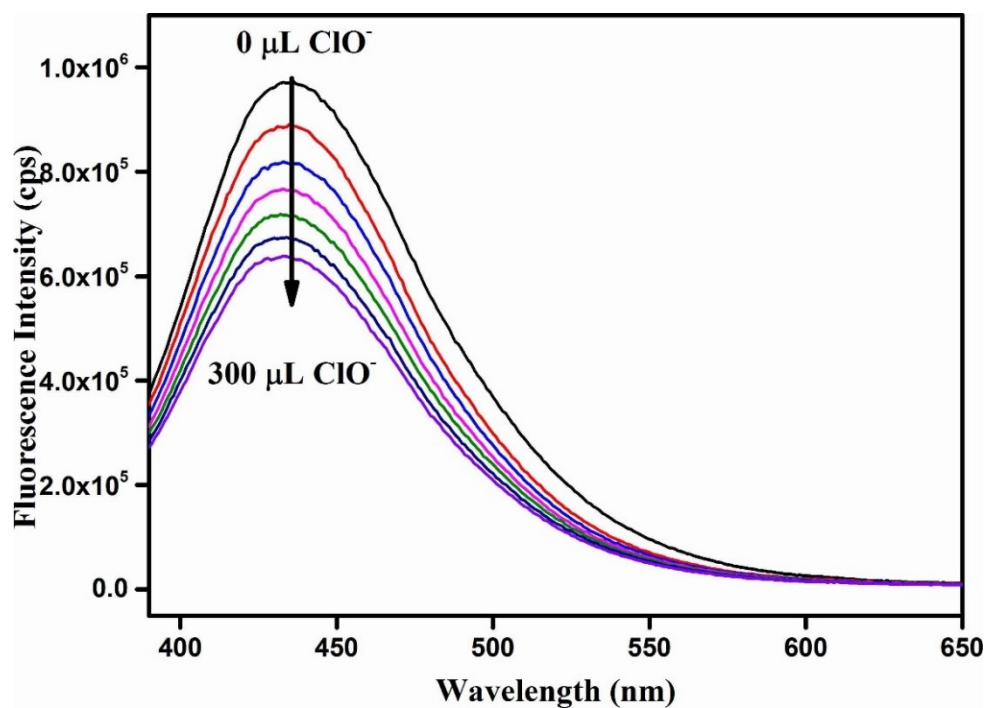
**Table S3.** Comparison of performance of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** with other reported MOF based probes.

Sl. No.	Name of the Compound	Response Time	Medium Used	Detection Limit ( $\mu\text{M}$ )	Ref.
1	<b>DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'</b>	< 30 s	PBS buffer	0.08	this work
2	NH <sub>3</sub> -MIL-53(Al) MOF	180 s	PBS buffer	0.04	4
3	UiO-68-PT	< 10 s	water	0.28	5
4	Hf-UiO-66-(NH <sub>2</sub> ) <sub>2</sub> MOF	seconds	HEPES buffer	0.02	6
5	Hf-UiO-66-(NH <sub>2</sub> ) <sub>2</sub> MOF	seconds	PBS buffer	0.009	6
6	Al-MIL-53-NH <sub>2</sub> MOF	seconds	HEPES buffer	0.54	6
7	NH <sub>2</sub> -HPU-17	-	water	0.026	7
8	Eu-BDC-NH <sub>2</sub> /DPA	20 s	water	0.037	8
9	PDA/Eu/PDA-UiO-66-NH <sub>2</sub>	< 15 s	Tris-HCl	0.10	9
10	AF@MOF-801	30 s	HEPES buffer	0.075	10
11	UiO-68-ol	5 s	PBS	0.10	11

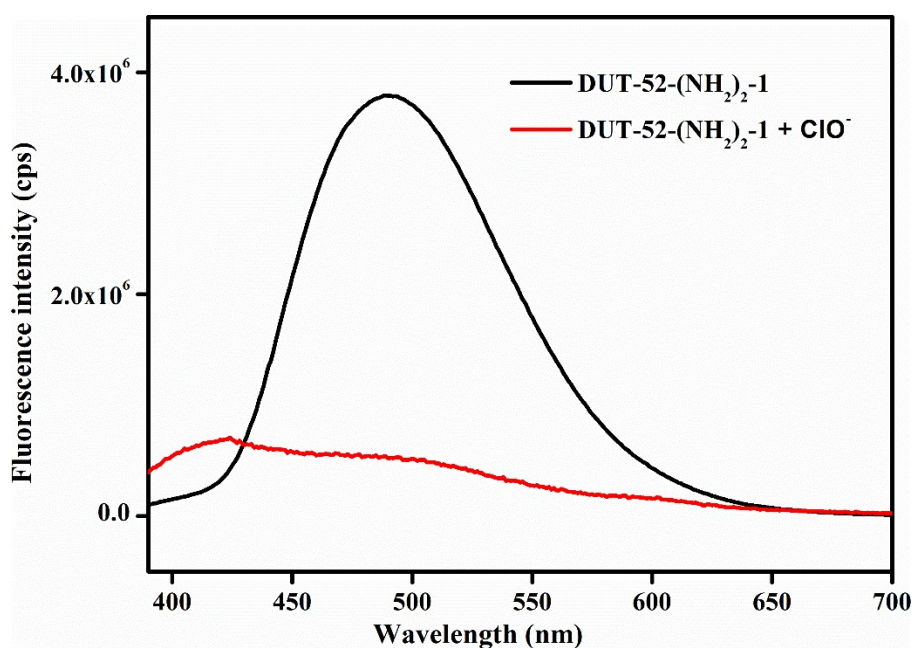
12	UiO-Eu-L1	5 s	water	0.016	12
13	CD/CCM@ZIF-8	-	ethanol	0.067	13
14	NH <sub>2</sub> -Cu-MOFs	10 s	water	0.036	14
15	UiO-66-NH <sub>2</sub>	180 s	water	0.3	15



**Figure S67.** Recyclability of compound DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1' up to five cycles of sensing experiments.



**Figure S68.** Change in the fluorescence intensity of DUT-52 MOF dispersed in PBS after the addition of 5 mM  $\text{ClO}^-$  solution.



**Figure S69.** Change in the fluorescence intensity of DUT-52-( $\text{NH}_2$ )<sub>2</sub>-1 MOF dispersed in PBS after the addition of 5 mM  $\text{ClO}^-$  solution.

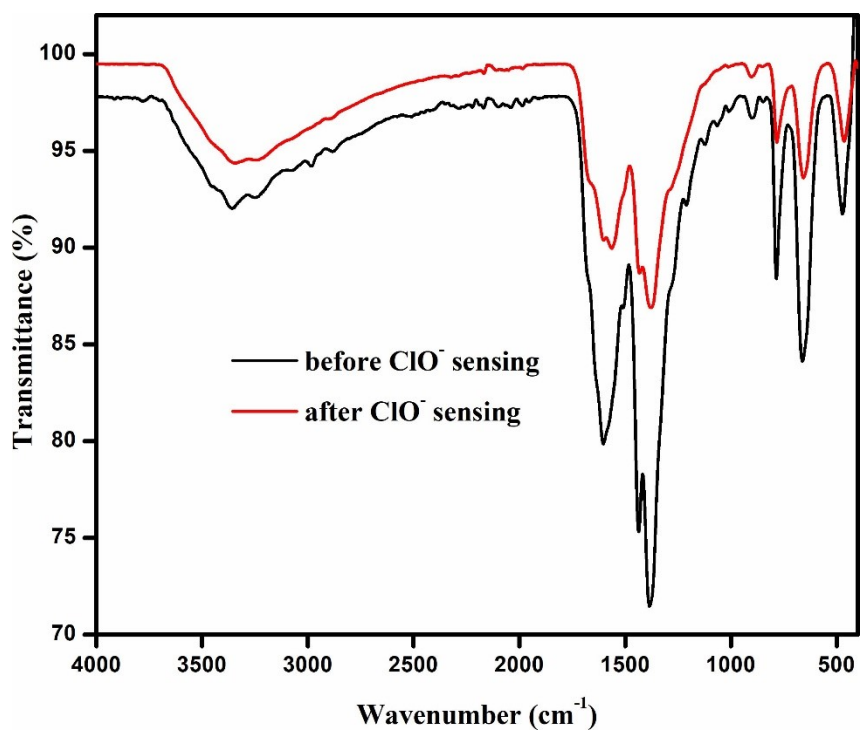


Figure S70. FT-IR spectra of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** before and after  $\text{ClO}^-$  sensing.

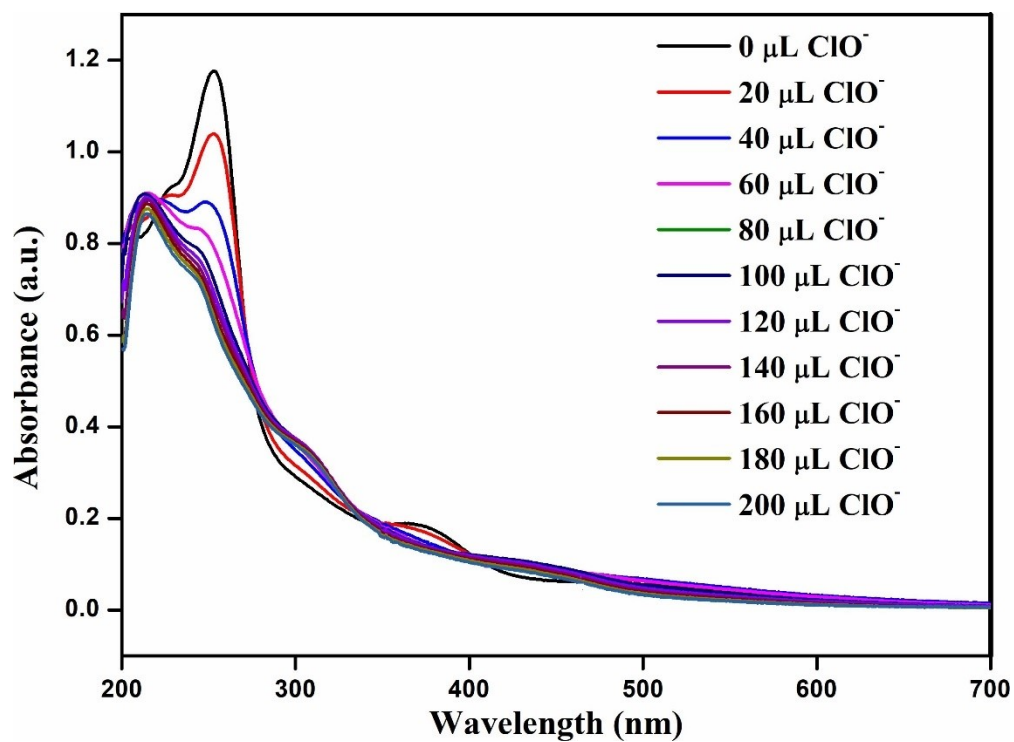
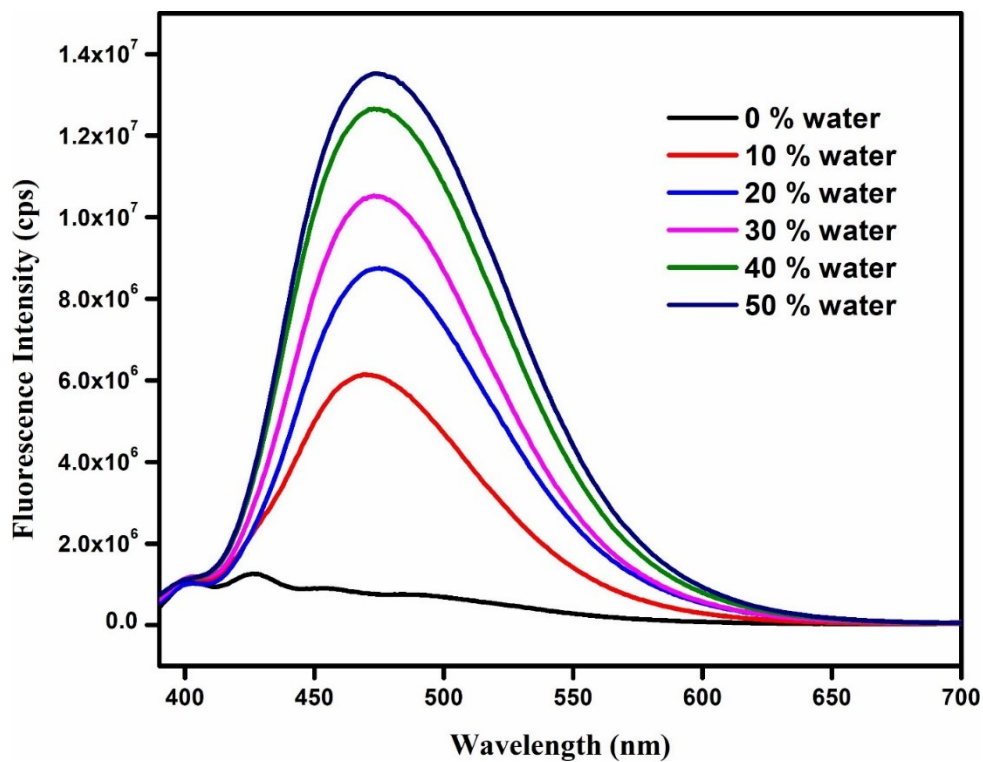
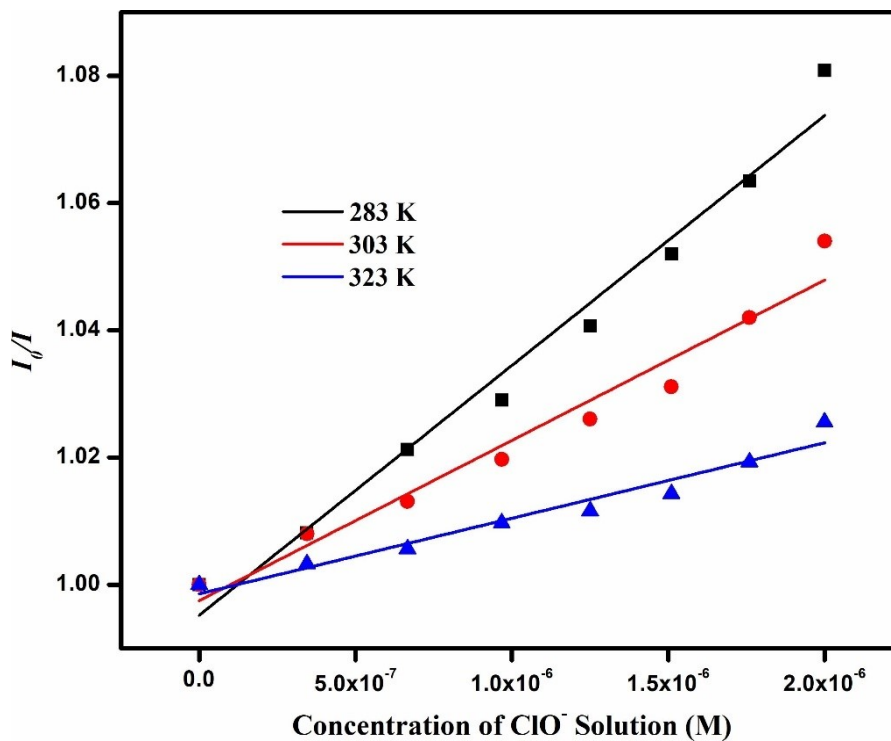


Figure S71. UV-vis spectra of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** in presence of  $\text{ClO}^-$  solutions (5 mM).



**Figure S72.** Fluorescence emission spectra of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** response to different water content in ethanol under excitation of 370 nm.



**Figure S73.** Stern-Volmer plots at different temperatures for quenching of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** by  $\text{ClO}^-$  solution.

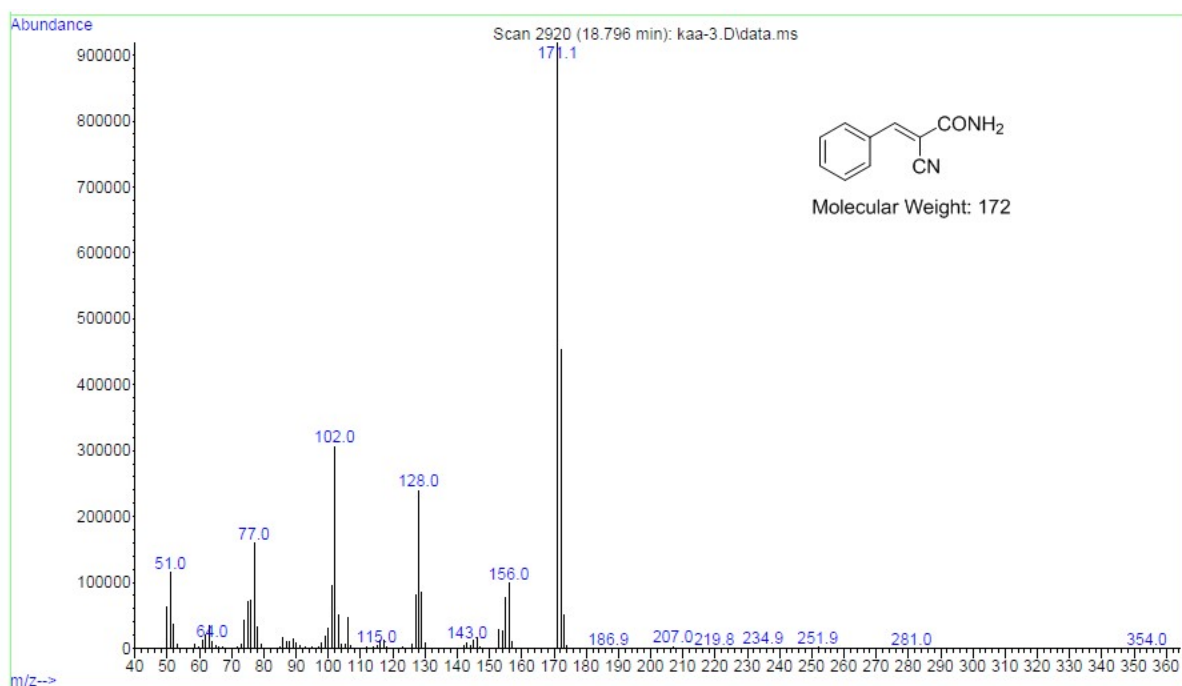
**Table S3.** Liner regression analysis of the Stern-Volmer plots (Figure S73) at different temperatures for the quenching of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** by ClO<sup>-</sup> solution.

T (K)	K <sub>sv</sub> (M <sup>-1</sup> ) x 10 <sup>4</sup>	R <sup>2</sup>
283	3.9270	0.97590
303	2.5197	0.95755
323	1.1876	0.95850

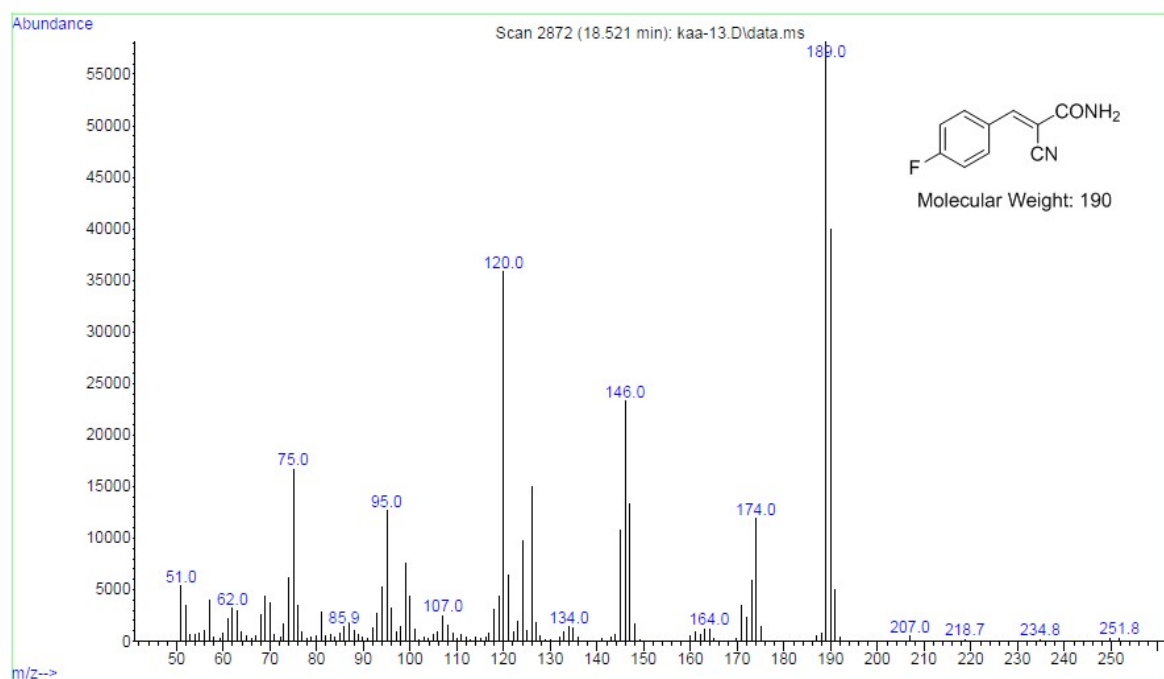
**Table S4.** Evaluation of intra-day, inter-day accuracy and precision study of change in fluorescence intensity of **DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1'** after incremental addition of 5 mM ClO<sup>-</sup>.

Parameter	Amount of ClO <sup>-</sup> added (μL)	Fluorescence Intensity (cps) at λ <sub>max</sub> = 490 nm			X (cps)	SD	RE%
Reproducibility Inter-day precision	0	4108760	4113620	4110620	4111000	2452	0.419
	50	2663950	2681820	2717140	2687637	27067	1.736
	100	1607700	1618730	1581910	1602780	18896	0.087
	150	8608421	869331	865643	865272	4256	0.773
	200	416166	420735	420143	419014	2484	1.014
	250	149736	148069	149952	149252	1030	1.947
	300	17790	18883	18475	18382	552	0.311
Repeatability Intra-day precision	0	4109640	4107200	4103300	4106713	3197	0.314
	50	2727430	2709920	2710650	2716000	9905	0.700
	100	1587070	1603810	1595640	1595507	8370	0.541
	150	863062	874866	876324	871417	7272	1.445
	200	421506	423536	421613	422218	1142	1.513
	250	150020	148770	153707	150832	2566	0.908
	300	18067	19073	19006	18715	562	1.493

Where: X, mean values; SD, standard deviation; %RE, percentage of relative error.

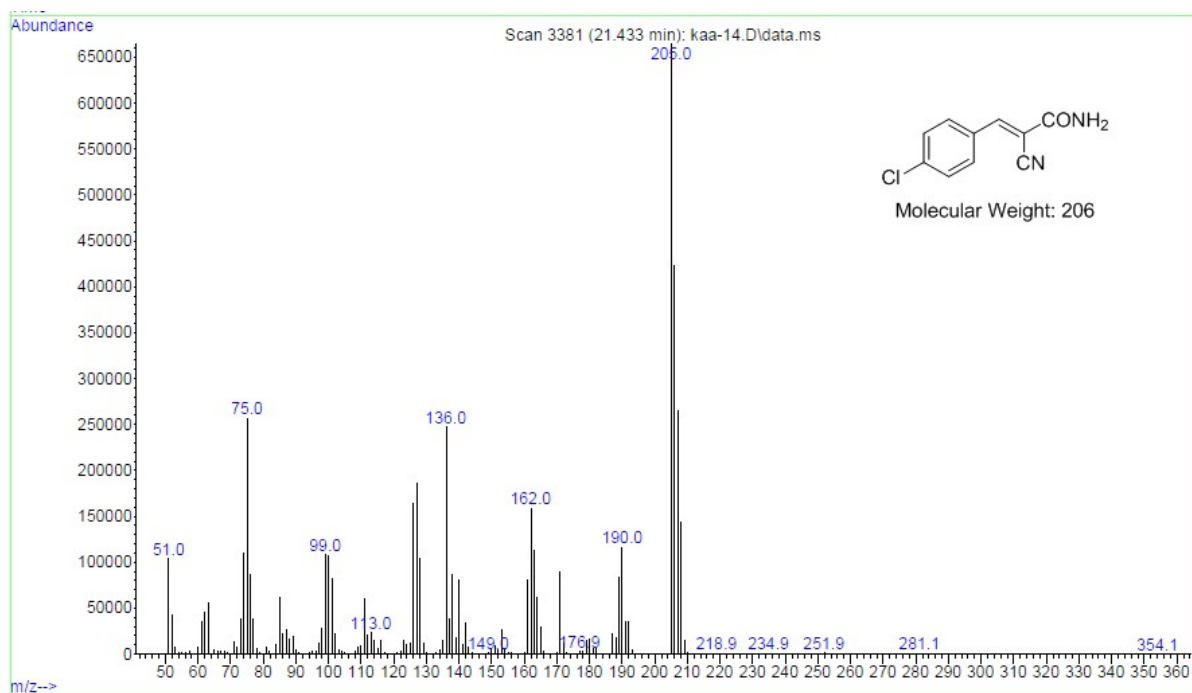


**Figure S74.** GC-MS trace of 2-cyano-3-phenylacrylamide.

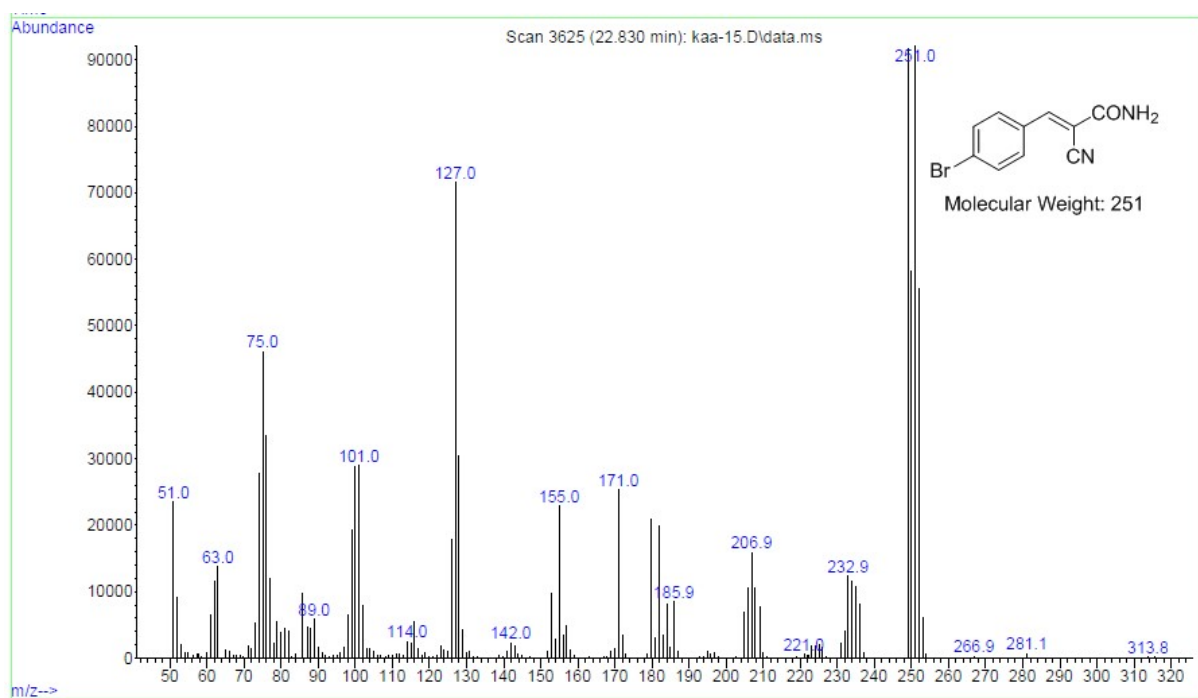


**Figure S75.** GC-MS trace of 2-cyano-3-(4-fluorophenyl)acrylamide.

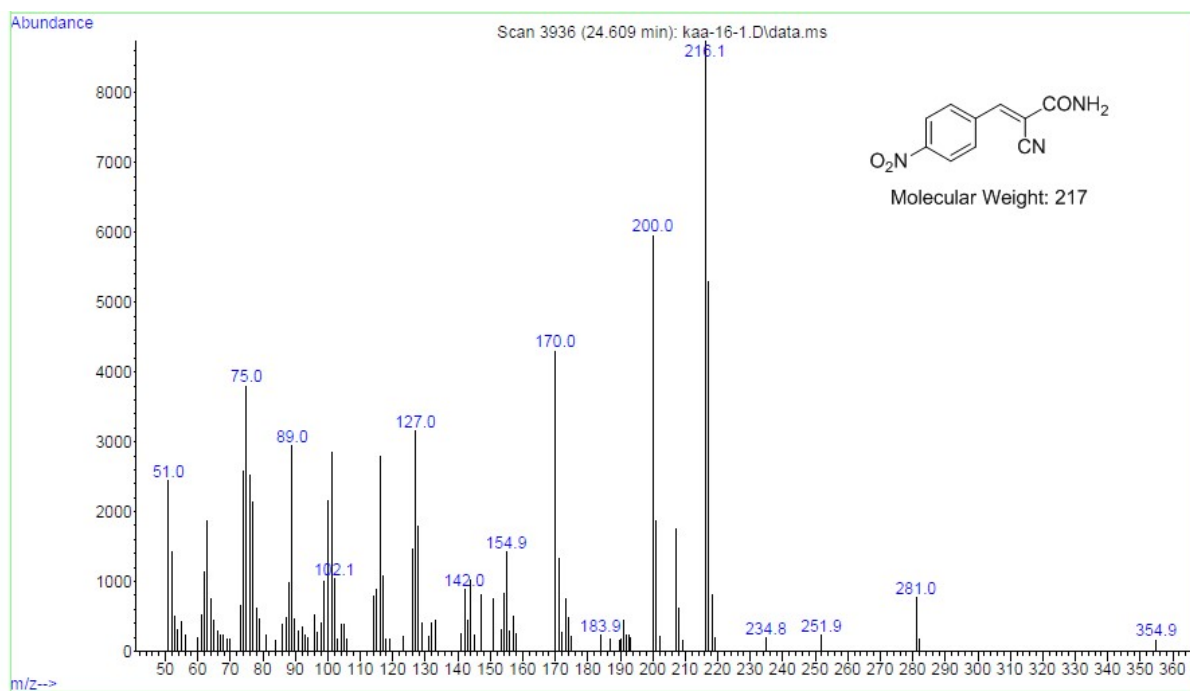




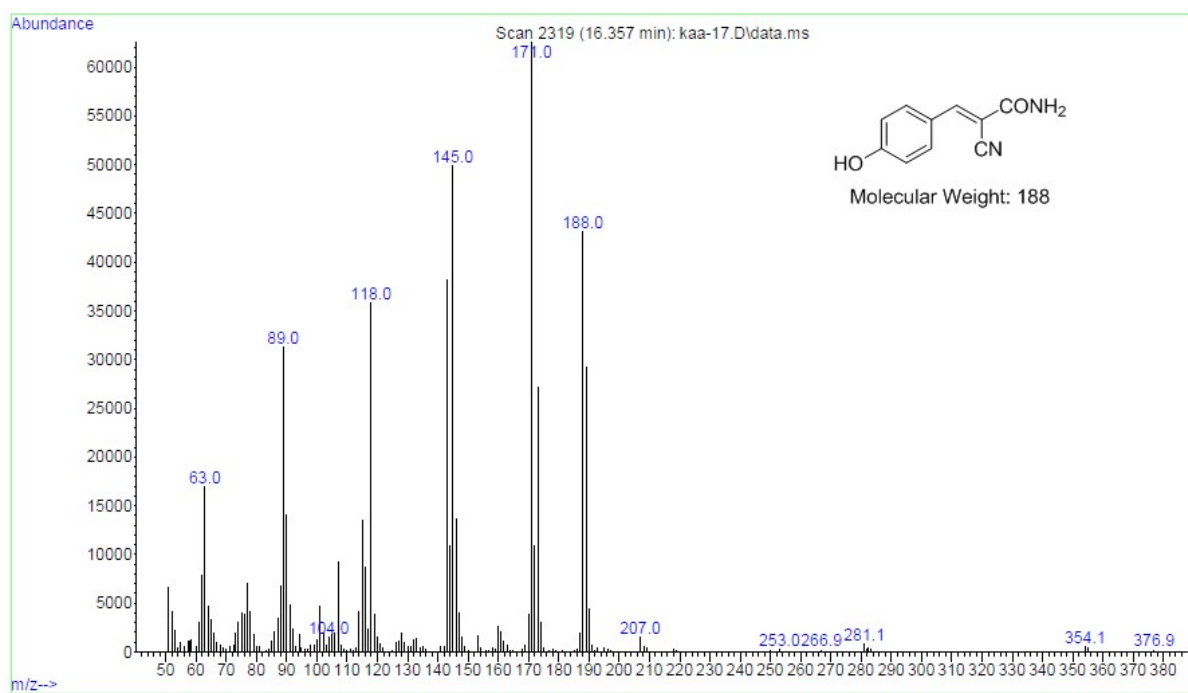
**Figure S76.** GC-MS trace of 3-(4-chlorophenyl)-2-cyanoacrylamide.



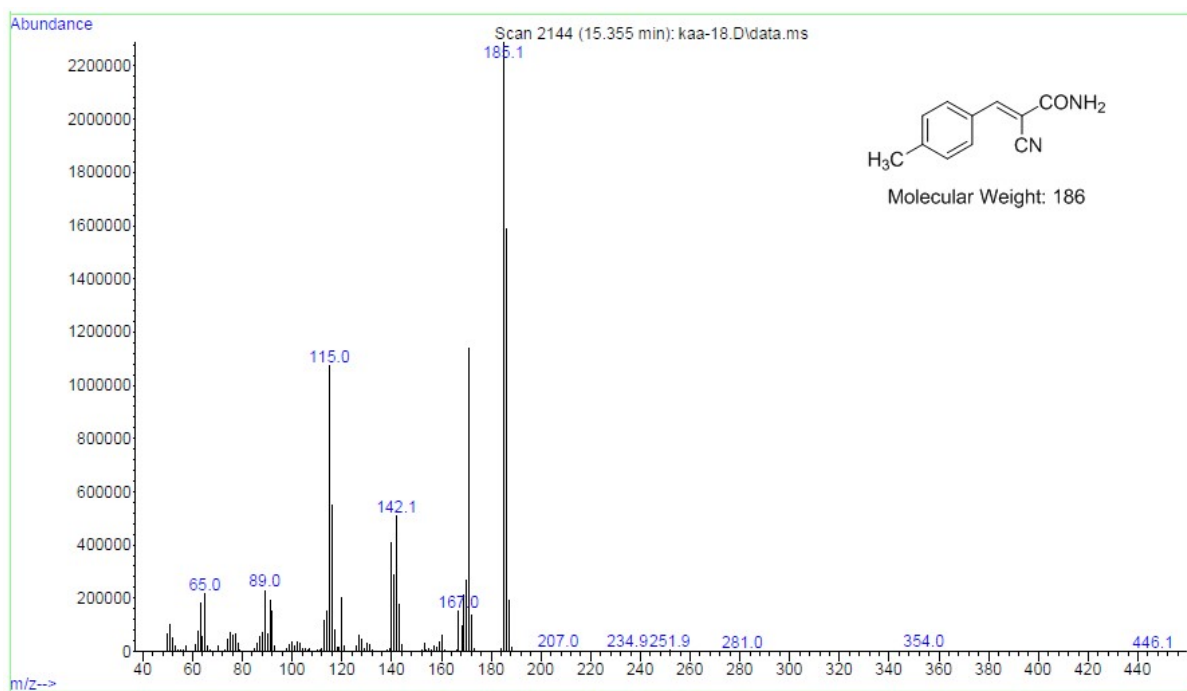
**Figure S77.** GC-MS trace of 3-(4-bromophenyl)-2-cyanoacrylamide.



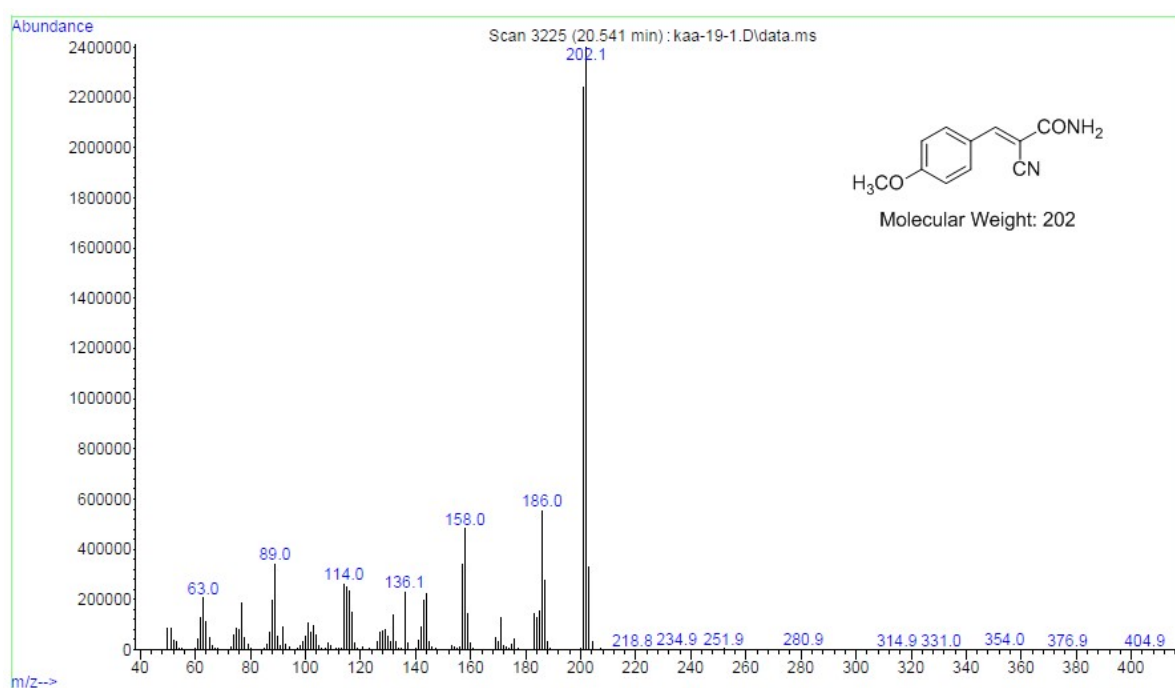
**Figure S78.** GC-MS trace of 2-cyano-3-(4-nitrophenyl)acrylamide.



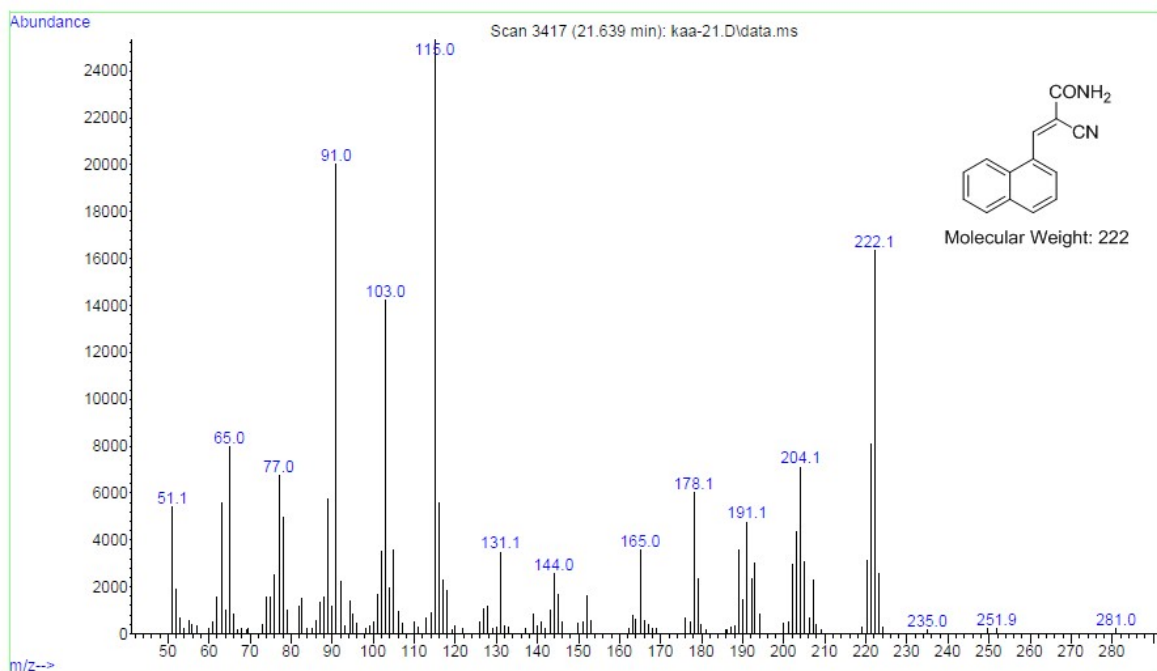
**Figure S79.** GC-MS trace of 2-cyano-3-(4-hydroxyphenyl)acrylamide.



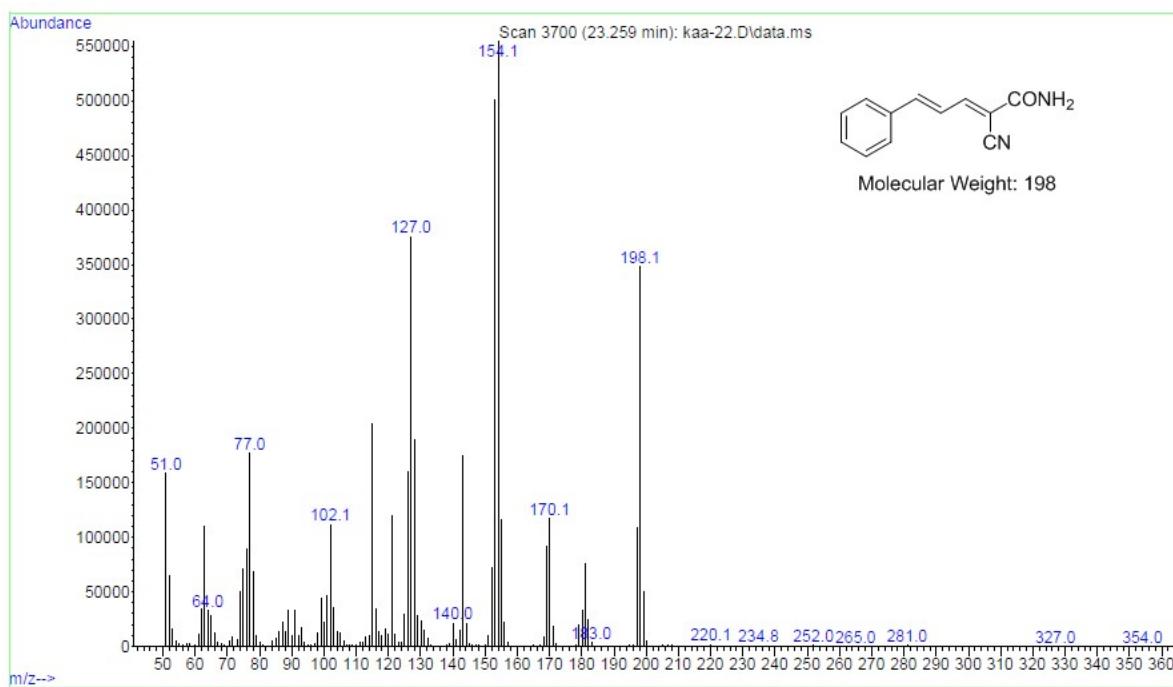
**Figure S80.** GC-MS trace of 2-cyano-3-(p-tolyl)acrylamide.



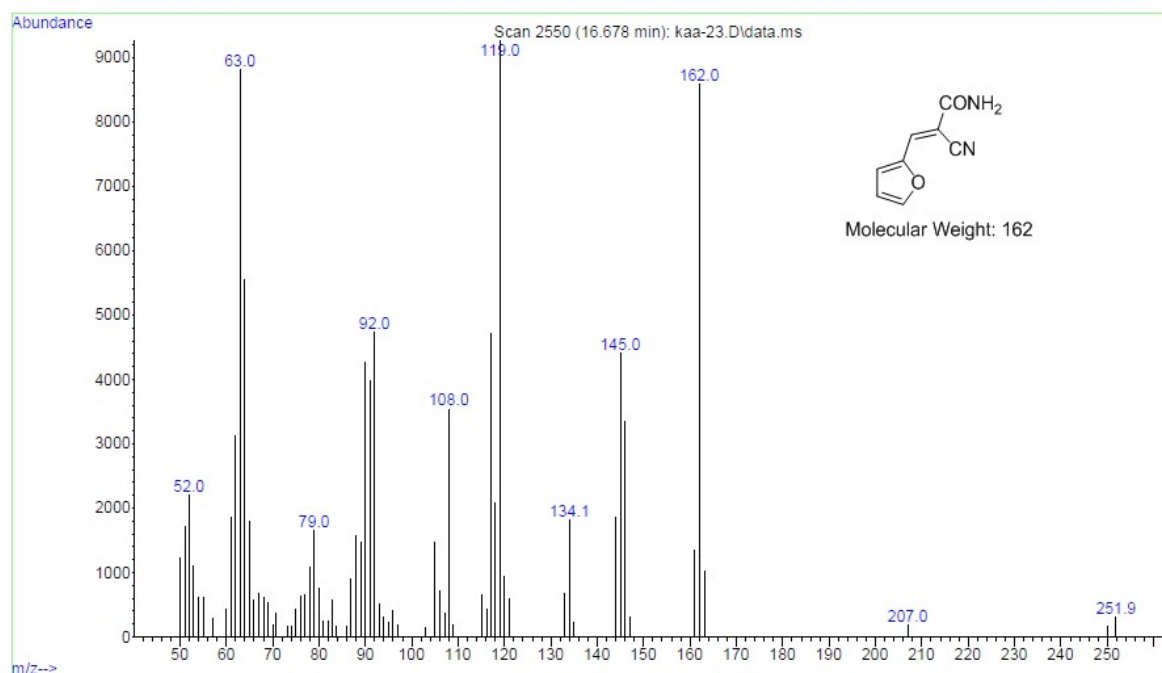
**Figure S81.** GC-MS trace of 2-cyano-3-(4-methoxyphenyl)acrylamide.



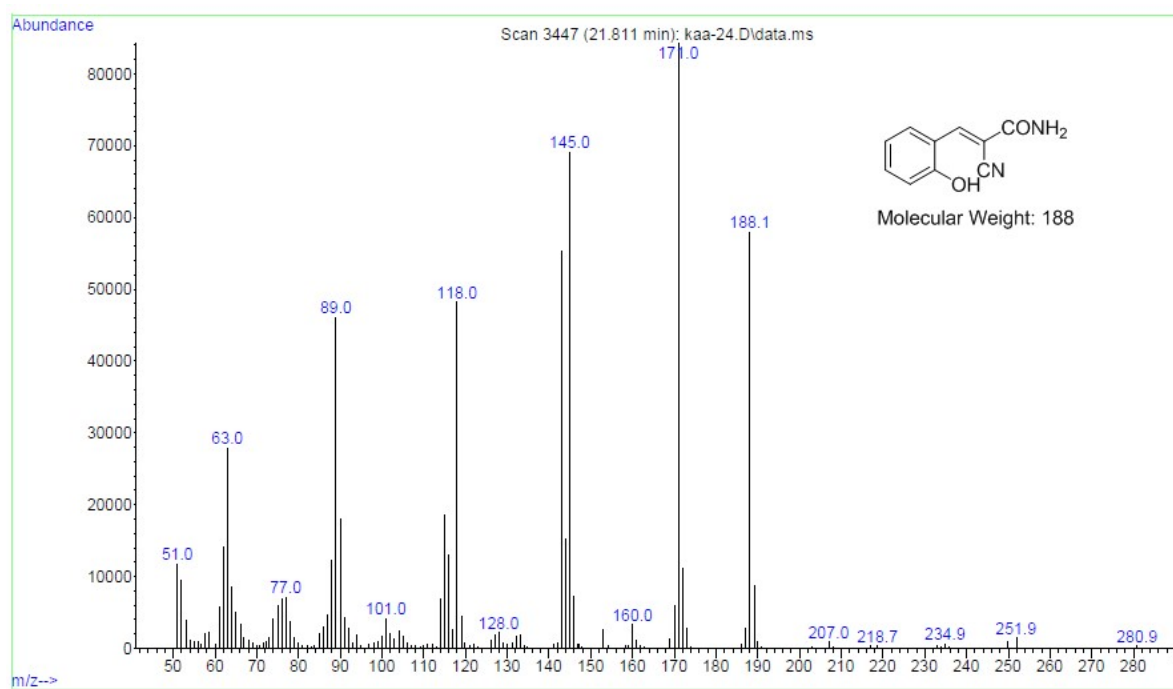
**Figure S82.** GC-MS trace of 2-cyano-3-(naphthalen-1-yl)acrylamide



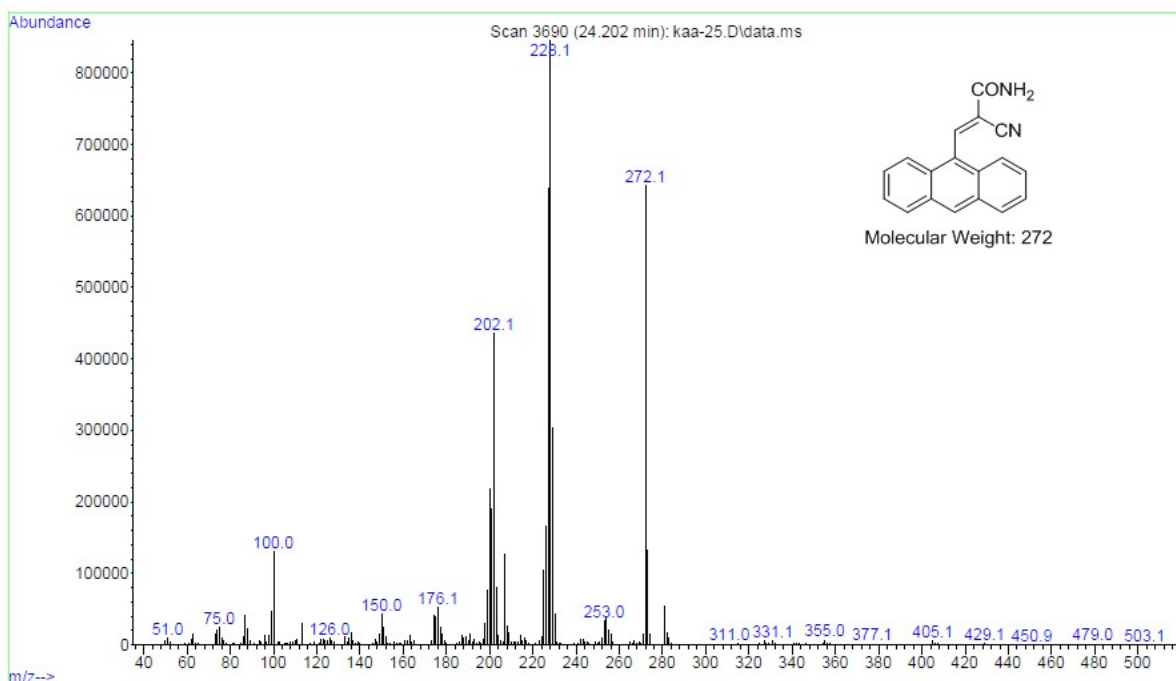
**Figure S83.** GC-MS trace of 2-cyano-5-phenylpenta-2,4-dienamide.



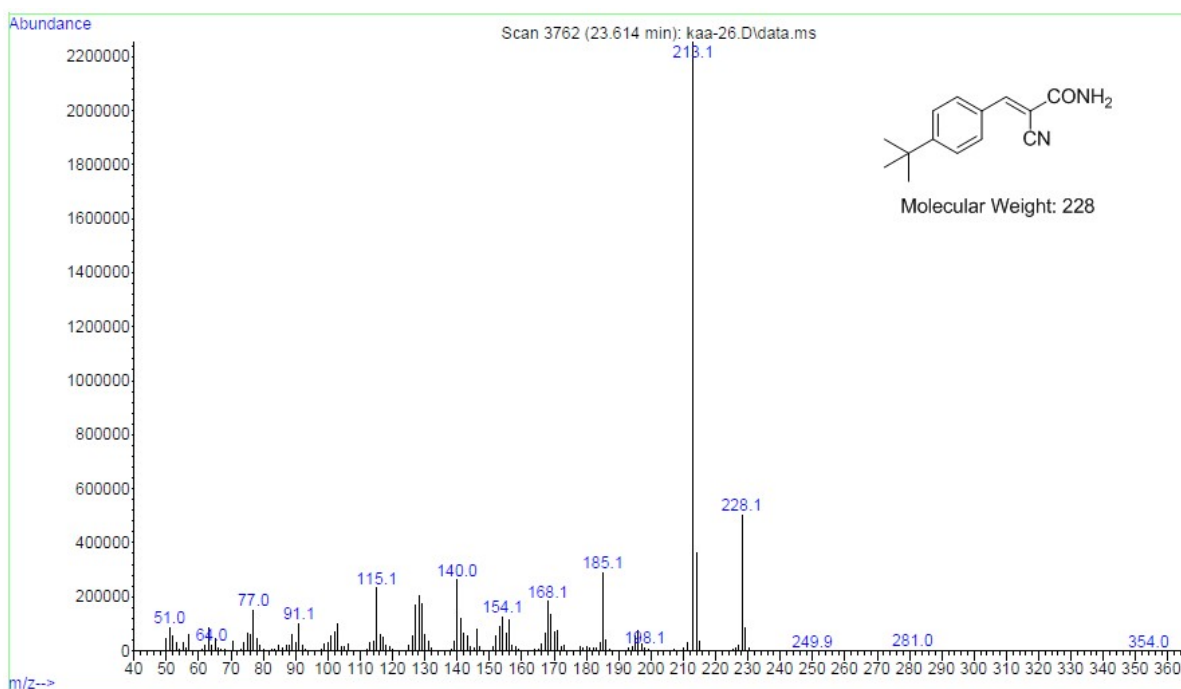
**Figure S84.** GC-MS trace of 2-cyano-3-(furan-2-yl)acrylamide.



**Figure S85.** GC-MS trace of 2-cyano-3-(2-hydroxyphenyl)acrylamide



**Figure S86.** GC-MS trace of 3-(anthracen-9-yl)-2-cyanoacrylamide.



**Figure S87.** GC-MS trace of 3-(4-(tert-butyl)phenyl)-2-cyanoacrylamide.

**Simulated CIF file of compound DUT-52-(NH<sub>2</sub>)<sub>2</sub>-1:**

data\_Zr-DUT-52-(NH<sub>2</sub>)<sub>2</sub>

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x,-y,-z

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-z,-x,y

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-y,-z,x

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\_atom\_site\_occupancy

C1	C	0.47818	0.31882	0.28228	0.00000	Uiso	1.00
O2	O	0.50215	0.14521	0.42073	0.00000	Uiso	1.00
C3	C	0.49278	0.26113	0.27721	0.00000	Uiso	1.00
C4	C	0.50330	0.12706	0.36803	0.00000	Uiso	1.00
O5	O	0.00030	0.07398	0.85274	0.00000	Uiso	1.00
C6	C	1.00139	0.23814	0.72211	0.00000	Uiso	1.00
O7	O	0.44795	0.05206	0.55204	0.00000	Uiso	1.00
O8	O	0.44809	0.05204	0.44790	0.00000	Uiso	1.00
O9	O	-0.07433	-0.35269	0.50219	0.00000	Uiso	1.00
C10	C	0.76112	-0.22273	0.50767	0.00000	Uiso	1.00
C11	C	0.18010	0.49404	0.33241	0.00000	Uiso	1.00
C12	C	0.13192	0.49706	0.37299	0.00000	Uiso	1.00
C13	C	0.31925	-0.00171	0.66692	0.00000	Uiso	1.00
C14	C	0.36788	-0.00083	0.62677	0.00000	Uiso	1.00
C15	C	0.26188	-0.22210	-0.00199	0.00000	Uiso	1.00
C16	C	0.12673	-0.13215	0.00044	0.00000	Uiso	1.00
C17	C	0.32603	0.99852	0.27399	0.00000	Uiso	1.00
O18	O	0.42054	1.00122	0.35488	0.00000	Uiso	1.00
C19	C	0.32111	-0.21567	-0.00530	0.00000	Uiso	1.00
O20	O	0.14501	-0.07947	0.00011	0.00000	Uiso	1.00
C21	C	0.35584	-0.26381	-0.00507	0.00000	Uiso	1.00
O22	O	0.42599	-0.35283	0.00021	0.00000	Uiso	1.00
O23	O	-0.07924	0.49893	-0.35489	0.00000	Uiso	1.00
C24	C	0.82540	0.50811	-0.27476	0.00000	Uiso	1.00
C25	C	0.23419	0.47919	0.35368	0.00000	Uiso	1.00
O26	O	0.14724	0.49731	0.42573	0.00000	Uiso	1.00
C27	C	0.50763	0.77474	0.32535	0.00000	Uiso	1.00
C28	C	0.49356	0.83239	0.31985	0.00000	Uiso	1.00

C29	C	1.00114	0.16693	0.81924	0.00000	Uiso	1.00
C30	C	0.99793	0.22602	0.82603	0.00000	Uiso	1.00
C31	C	0.14419	0.76380	1.00454	0.00000	Uiso	1.00
C32	C	0.17891	0.71567	1.00476	0.00000	Uiso	1.00
C33	C	0.85369	0.73424	0.52121	0.00000	Uiso	1.00
C34	C	0.81883	0.78234	0.52221	0.00000	Uiso	1.00
N35	N	0.84278	0.83607	0.53767	0.00000	Uiso	1.00
N36	N	0.15336	0.66058	1.00835	0.00000	Uiso	1.00
N37	N	0.34274	0.66400	0.53740	0.00000	Uiso	1.00
N38	N	0.65332	1.16059	0.99122	0.00000	Uiso	1.00
Zr39	Zr	0.50000	0.09965	0.50000	0.00000	Uiso	1.00
O40	O	0.05199	0.05199	-0.05199	0.00000	Uiso	1.00
Zr41	Zr	0.50000	-0.40056	0.00000	0.00000	Uiso	1.00
Zr42	Zr	0.50000	0.00000	-0.40028	0.00000	Uiso	1.00
Zr43	Zr	0.09954	0.00000	0.00000	0.00000	Uiso	1.00
O44	O	0.05209	-0.05209	-0.05209	0.00000	Uiso	1.00

loop\_

\_geom\_bond\_atom\_site\_label\_1

\_geom\_bond\_atom\_site\_label\_2

\_geom\_bond\_distance

\_geom\_bond\_site\_symmetry\_2

\_ccdc\_geom\_bond\_type

C1 C3 1.400 . A

C1 C25 1.392 9 A

C1 N37 1.426 8\_656 S

O2 C4 1.307 . A

O2 Zr39 2.144 . S

C3 C10 1.423 6\_565 A

C3 C27 1.407 2\_665 A

C4 O9 1.288 6 A

C4	C28	1.478	2_665	S
O5	C16	1.286	5_556	A
O5	Zr43	2.063	10_556	S
C6	C15	1.423	5_656	A
C6	C17	1.407	10_756	A
C6	C32	1.399	5	A
O7	Zr39	2.054	.	S
O7	Zr42	2.056	1_556	S
O7	Zr41	2.052	10_556	S
O8	Zr39	2.054	.	S
O8	Zr42	2.053	3_655	S
O8	Zr41	2.054	10_556	S
O9	C4	1.288	12	A
O9	Zr42	2.069	9	S
C10	C3	1.423	12_655	A
C10	C24	1.407	5_645	A
C10	C34	1.400	1_545	A
C11	C25	1.406	.	A
C11	C12	1.478	.	S
C11	C24	1.397	3_655	A
C12	O26	1.287	.	A
C12	O23	1.306	3	A
C13	C14	1.478	.	S
C13	C17	1.396	4_566	A
C13	C21	1.407	10_556	A
C14	O18	1.307	4_566	A
C14	O22	1.286	10_556	A
C15	C19	1.399	.	A
C15	C6	1.423	9_544	A
C15	C30	1.407	9_544	A

C16	O20	1.307	.	A
C16	O5	1.286	9_545	A
C16	C29	1.478	9_544	S
C17	C6	1.407	7_675	A
C17	C13	1.396	4_566	A
O18	C14	1.307	4_566	A
O18	Zr42	2.145	3_665	S
C19	C21	1.391	.	A
C19	N38	1.426	2_664	S
O20	Zr43	2.146	.	S
C21	C13	1.407	7_655	A
O22	Zr41	2.064	.	S
O22	C14	1.286	7_655	A
O23	C12	1.306	3	A
O23	Zr41	2.144	5	S
C24	C10	1.407	9_654	A
C24	C11	1.397	3_655	A
C25	C1	1.392	5	A
O26	Zr39	2.069	9	S
C27	C28	1.397	.	A
C27	C3	1.407	2_665	A
C28	C4	1.478	2_665	S
C28	C33	1.406	8_656	A
C29	C30	1.396	.	A
C29	C16	1.478	5_656	S
C29	C31	1.407	5	A
C30	C15	1.407	5_656	A
C31	C32	1.391	.	A
C31	C29	1.407	9	A
C32	N36	1.426	.	S

C32	C6	1.399	9	A
C33	C34	1.392	.	A
C33	C28	1.406	11_566	A
C34	N35	1.426	.	S
C34	C10	1.400	1_565	A
N37	C1	1.426	11_566	S
N38	C19	1.426	2_666	S
Zr39	O2	2.144	3_656	S
Zr39	O7	2.054	3_656	S
Zr39	O8	2.054	3_656	S
Zr39	O26	2.069	5	S
Zr39	O26	2.069	8_656	S
O40	Zr43	2.052	.	S
O40	Zr43	2.052	5	S
O40	Zr43	2.052	10	S
Zr41	O22	2.064	3_655	S
Zr41	O7	2.052	7_655	S
Zr41	O7	2.052	6	S
Zr41	O8	2.054	7_655	S
Zr41	O8	2.054	6	S
Zr41	O23	2.144	9	S
Zr41	O23	2.144	10_655	S
Zr42	O7	2.056	1_554	S
Zr42	O7	2.056	2_654	S
Zr42	O8	2.053	3_655	S
Zr42	O8	2.053	4	S
Zr42	O9	2.069	5	S
Zr42	O9	2.069	7_655	S
Zr42	O18	2.145	3_645	S
Zr42	O18	2.145	4_565	S

Zr43 O20 2.146 4 S  
 Zr43 O40 2.052 4 S  
 Zr43 O44 2.054 . S  
 Zr43 O44 2.054 4 S  
 Zr43 O5 2.063 7\_655 S  
 Zr43 O5 2.063 8\_655 S  
 O44 Zr43 2.054 6 S  
 O44 Zr43 2.054 10 S

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