

1 **Supporting Information for**

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3 **Destruction of Per/poly-fluorinated Alkyl Substances by Magnetite**
4 **Nanoparticle-Catalyzed UV-Fenton reaction**

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24 **ST1.** Percent destruction of PFOA and PFOS with variable UV exposure time, hydrogen peroxide
 25 (H₂O₂) concentration, and Fe₃O₄ concentration.

UV time (min)	H ₂ O ₂ conc. (M)	Fe ₃ O ₄ conc. (ppm)	% Destruction PFOA ^a	% Destruction PFOS ^a
5	0.10	100	13.3	20.0
5	2.55	100	26.7	38.7
5	2.55	1050	68.0	88.0
5	2.55	2000	66.7	84.7
5	5.00	1050	56.0	75.3
5	5.00	2000	68.7	83.3
32.5	0.10	1050	6.7	36.0
32.5	0.10	2000	0.0	33.3
32.5	2.55	100	67.3	76.0
32.5	2.55	1050	61.3	86.0
32.5	2.55	2000	66.0	80.7
32.5	5.00	100	74.7	74.0
32.5	5.00	2000	69.3	82.7
60	0.10	1050	0.0	33.3
60	0.10	2000	6.7	34.0
60	2.55	100	65.3	78.7
60	2.55	2000	63.3	74.0
60	5.00	100	39.3	64.7
60	5.00	1050	13.3	47.3
60	5.00	2000	54.0	72.7

26 ^a Percent error for samples was calculated to be ±7.0% for PFOA and ±4.9% for PFOS.

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28 **S1. High Resolution Mass Spectrometry (HRMS) Experimental Details.**

29 HRMS analysis was conducted on a Thermo Scientific Orbitrap Exploris 240 instrument. Control
 30 PFOA and PFOS samples (100 ppm) and samples after reaction at specific conditions (pH 7 or pH
 31 9, 2.5M or 5M) were analyzed under the same instrumental conditions. Samples were prepared by
 32 filtration through 0.2 μm PTFE syringe filters followed by dilution in methanol in a 1:10 ratio of
 33 sample to methanol. Sample solutions were introduced by continuous direct infusion (10 μl/min)

34 into the electrospray ion source of the instrument and were analyzed in both negative and positive
35 ion modes. Optimized source conditions were 3600V and 2600V spray voltage for positive and
36 negative ion generation, respectively. The instrument was autotuned (rf lens rel. value 70%) with
37 automatic gain control and custom injection time functions switched on. The system was mass
38 calibrated in both polarities each day before each analysis by following the manufacturer's
39 protocols with calibration compound mixtures purchased from the manufacturer. The resolving
40 power was set at 240,000 FWHM. For each analyzed sample, 150 individual ion spectra were
41 collected in a range from 50 to 500 m/z (50 to 700 m/z for PFOS) in MS or tandem MS mode for
42 either polarity, respectively. The estimated experimental mass accuracy was better than 5 ppm in
43 the entire range. Tandem mass spectrometry was performed by isolating monoisotopic precursor
44 ions (window 1 m/z) and subsequent manual optimization of the relative collision energy value for
45 efficient collision-induced dissociation (CID). Elemental composition assignments from accurate
46 mass determination of select ions in the averaged negative ion spectra were performed using the
47 Web-based software Chem Calc (<https://www.chemcalc.org/mf-finder>). The mass of a single
48 electron (0.00054 Da) was subtracted from each experimentally determined anion mass, i.e., the
49 mass of the neutral species was input into the program. The calculated masses of the species with
50 tentative elemental composition assignment were restricted to be less than 5 ppm than the
51 measured neutral species mass.

52 **S2. XANES Experimental Details.**

53 Fe₃O₄ nanoparticles were filtered from solutions following Fenton reactions in the UV oven and
54 collected on P5 grade filter paper. A section of filter paper approximately 1 cm × 2 cm with a
55 uniform layer of magnetite was cut from the whole filter paper. The filtered Fe₃O₄ section was
56 mounted between Kapton tape and X-ray clean polyfilm. Fe XANES were collected for each

57 sample from -100 to -20 eV below the Fe k-edge (5.0 eV step size), from -20 eV below to 50 eV
58 above the Fe k-edge (0.2 eV step size), and 50 eV to 150 eV above the Fe k-edge (5.0 eV step size)
59 with a 0.5 sec point⁻¹ acquisition time. Spectra were collected in both transmission and
60 fluorescence modes, and a standard ionization chamber and a four element Silicon drift detector
61 were used for each of these measurement types, respectively. Fe XANES standard spectra of
62 Fe₃O₄, Fe foil, hematite (Fe₂O₃), and wustite (FeO) were collected and used to calibrate all samples
63 to the same Fe K-edge energy and to conduct linear combination fitting of each spectrum.

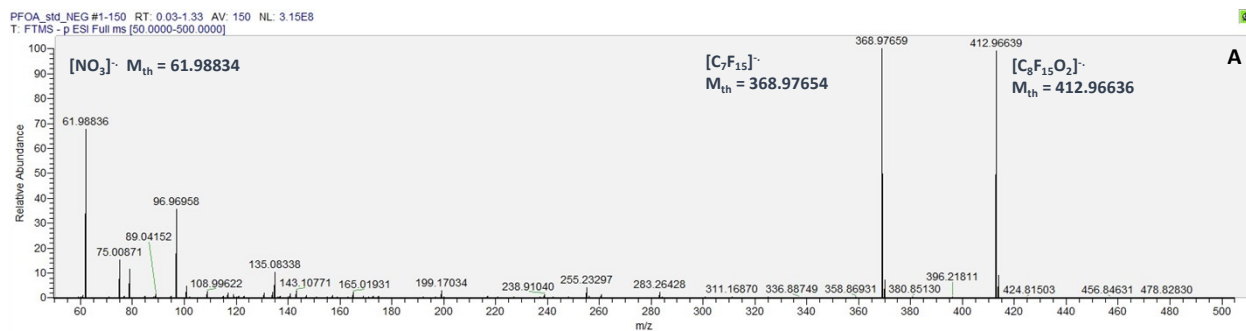
64 **S3. Results of HRMS and XANES Analyses.**

65 **S3.1. HRMS.**

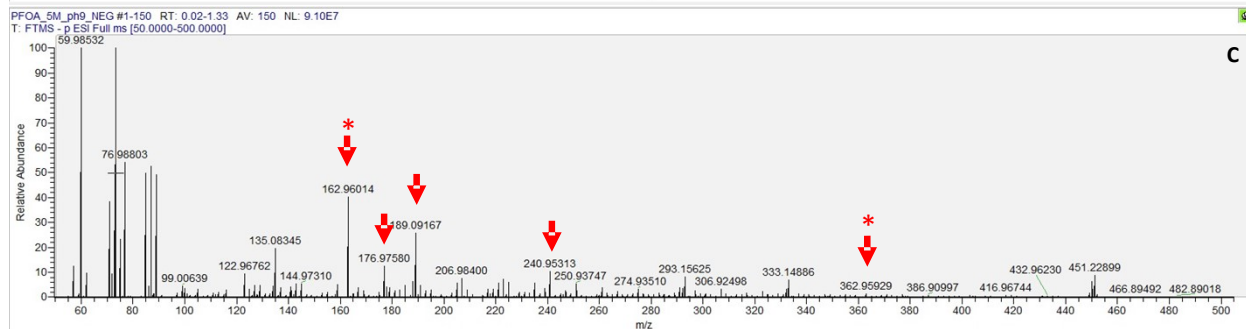
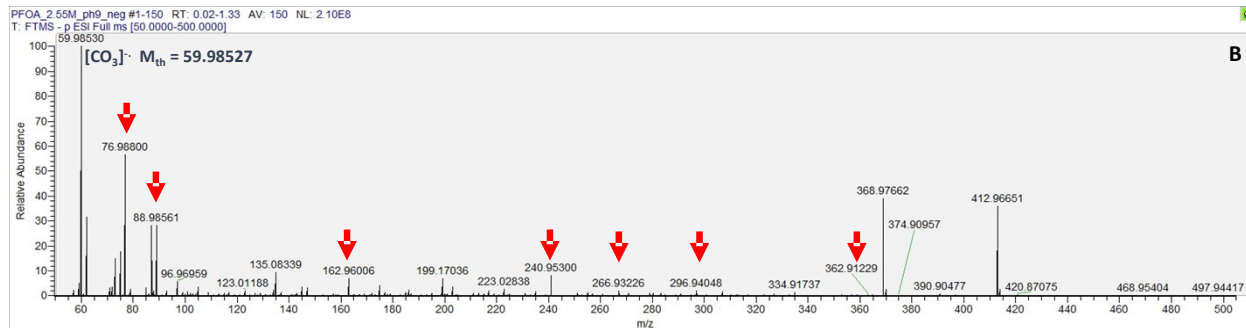
66 High resolution negative ion mass spectra of PFOS control and PFOS reacted under varying
67 conditions are plotted on Figure SF1. For the control, the most intense ion peak corresponds to
68 intact PFOS anion [C₈F₁₇O₃S]⁻ at m/z 498.93015 (molecular ion). The absolute intensity of the
69 intact PFOS peak in spectra from reacted samples is decreasing considerably. Similar to PFOA,
70 the initial PFOS sample has been completely degraded at 5M H₂O₂ and pH9, and no molecular ion
71 is observed in the spectrum (Figure SF2.C). Additional peaks observed in spectra from reacted
72 PFOA and PFOS are interpreted as the respective degradation products (Supplementary Tables ST2
73 and ST3).

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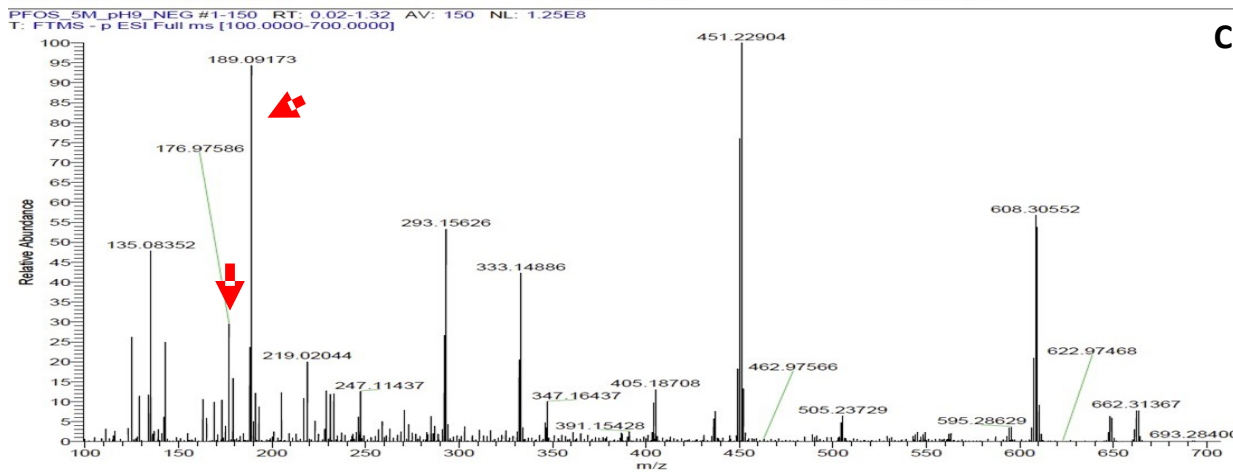
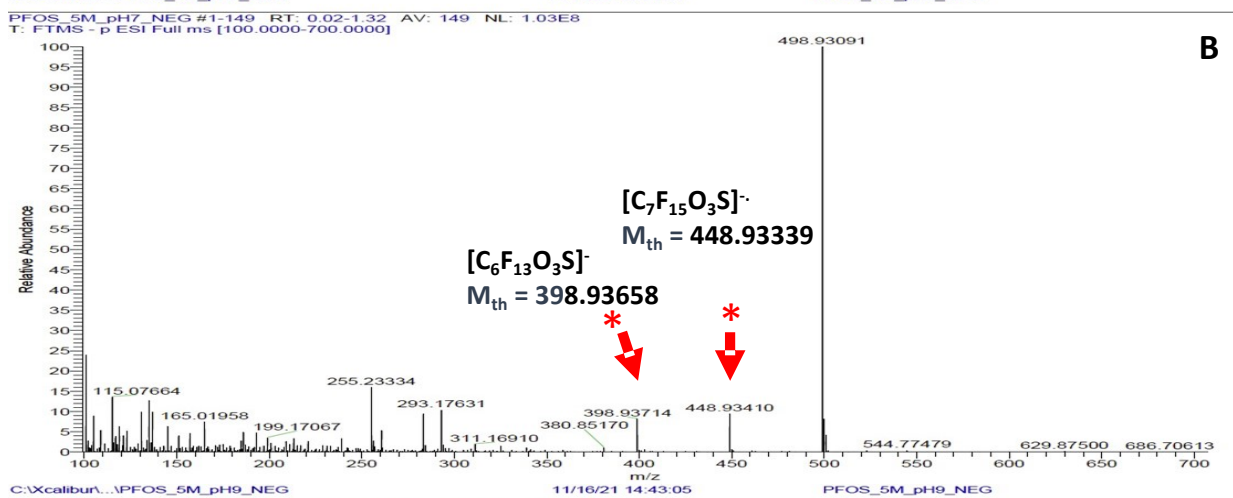
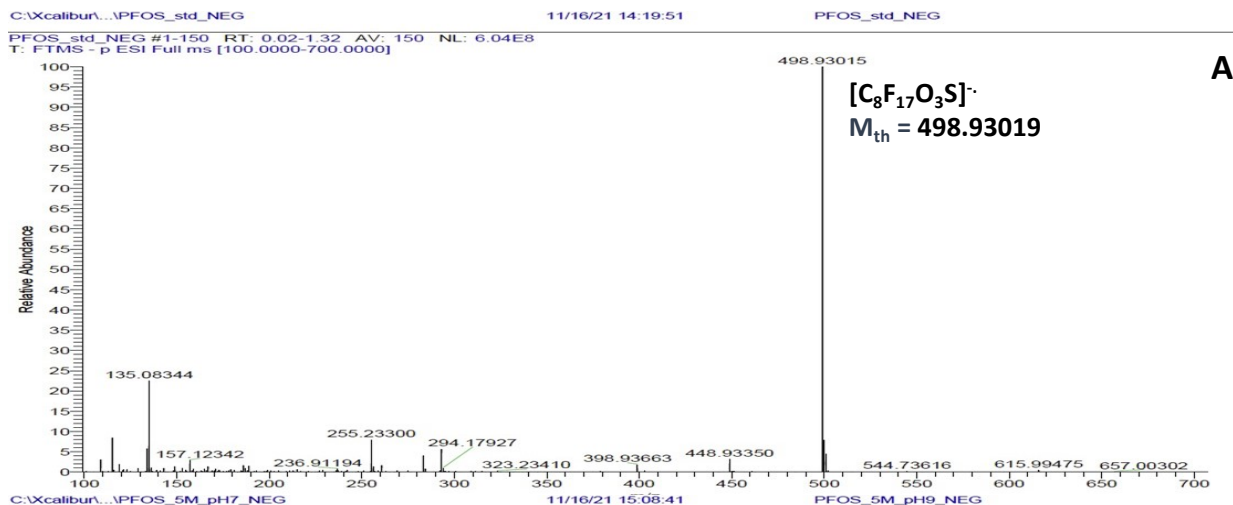


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78 **Figure SF1.** Comparison between negative ion spectra of PFOA control (A) and PFOA reacted at
 79 different conditions: 2.55M H₂O₂ at pH9 (B), and 5M H₂O₂ at pH9 (C). Arrows indicate peaks
 80 corresponding to degradation products, asterisks indicate peaks reported in the literature.
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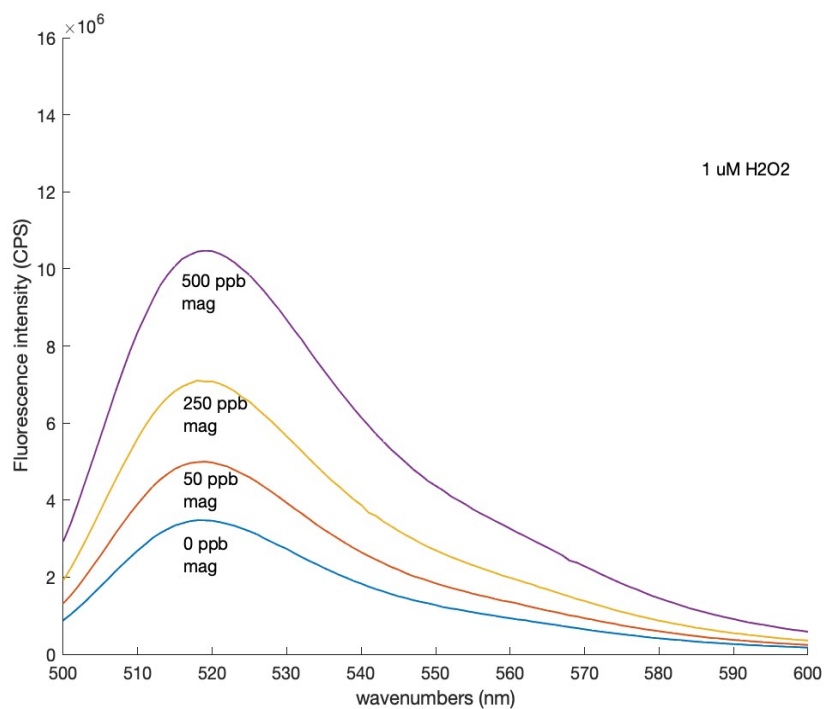
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85 **Figure SF2.** Comparison between negative ion spectra of PFOS control (A), and PFOS reacted at
 86 different conditions - 5M H₂O₂ at pH7 (B), and 5M H₂O₂ at pH9 (C). Arrows indicate peaks
 87 corresponding to degradation products, asterisks indicate peaks reported in the literature.² Strong
 88 peaks in spectrum C (at m/z 293, 333, 405, 451, 608) are attributed to metalloorganic cluster
 89 anions, that are also observed in spectra of blank (non-PFOS-containing) samples at the same
 90 experimental conditions.

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92 S3.2. FLUORESCENCE PROBE MEASUREMENTS.



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94 **Figure SF3.** Fluorescence intensity of detected ROS with increasing concentration of Fe₃O₄
95 nanoparticles.

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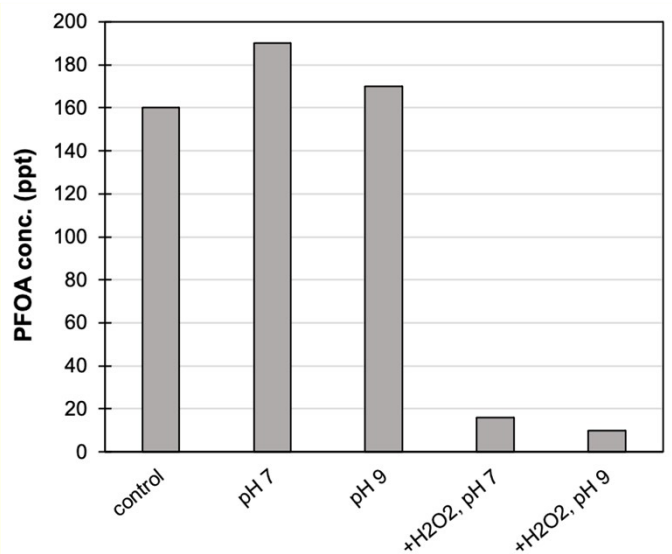
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113 S3.3. MAGNETITE ADSORPTION CONTROLS

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116 **Figure SF4.** PFOA concentrations in solutions with magnetite with and without added H₂O₂ and
117 exposed to UV radiation. There is no significant change in PFOA concentration with magnetite
118 particles suspended in the solution, compared to the control. Therefore, adsorption to magnetite is
119 not a contributing factor to PFAS removal. In contrast, ROS generated when H₂O₂ is present in
120 the solution result in a significant reduction in PFAS concentration.

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123 S3.3. XANES.

124 All samples were prepared with 1000 ppm Fe₃O₄ and underwent 30 min of UV-C exposure before

125 analysis. Linear combination fitting results for these spectra are given in Table ST4. Analysis

126 indicates no change in the peak energy of the K-edge (generally associated with oxidation state),

127 disappearance or appearance of pre-edge or K-edge features, or intensity of these features. In all

128 cases, the collected spectrum matches well with the Fe₃O₄ standard spectrum collected and shown

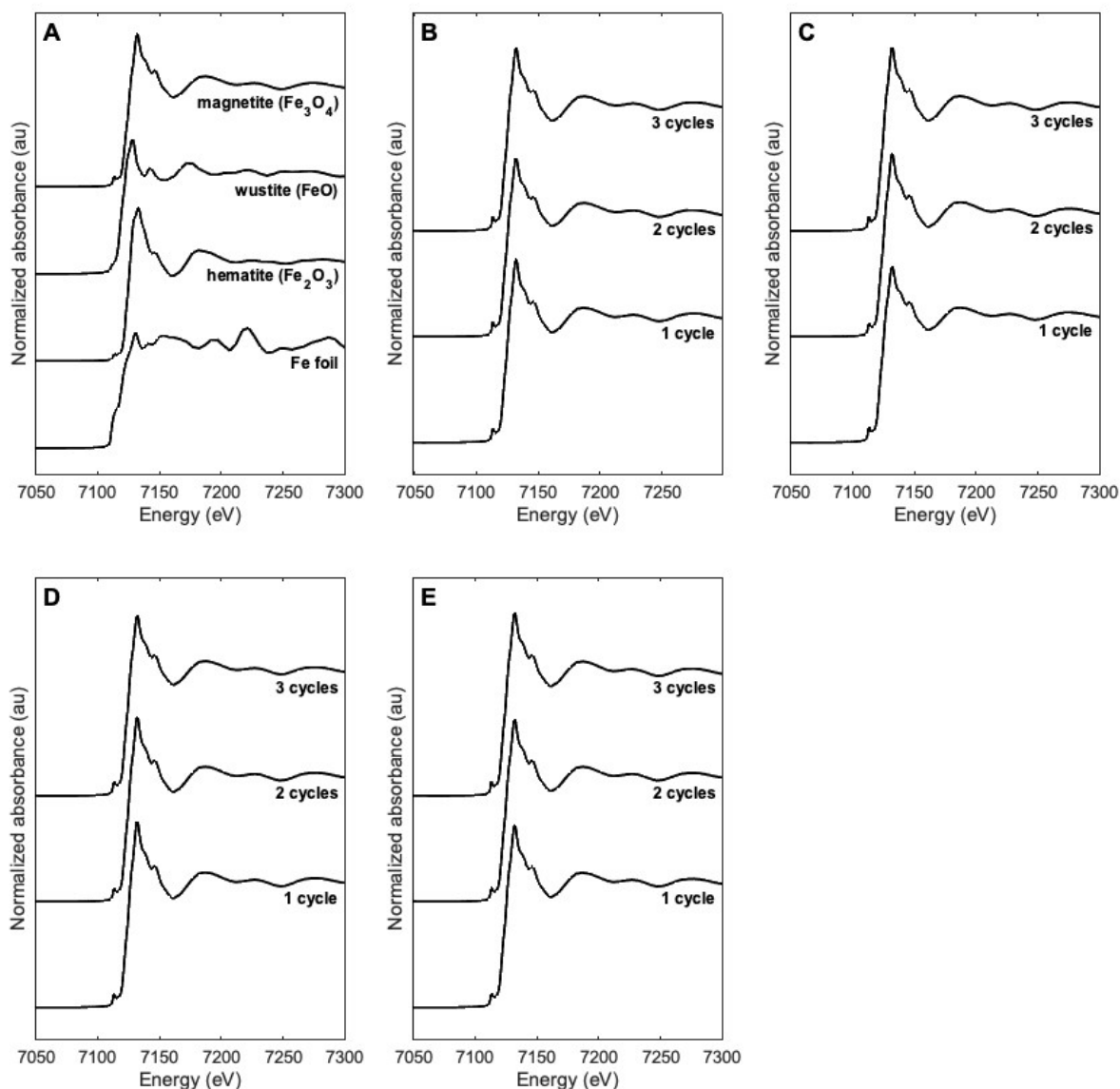
129 in Figure SF3A. Further analysis by linear combination fitting (Table ST4) of 4 common Fe

130 mineral spectra containing varied oxidation states of Fe emphasizes there is no change in the Fe₃O₄

131 after multiple reaction cycles. All samples are shown to be between 94-100% (estimated fit error

132 ± 3%) Fe₃O₄ in their Fe bonding characteristics. There is one outlier in the 2.55M, pH9 sample

133 after one cycle, which shows only about 89% Fe₃O₄, and this outlier is likely due to difficulty in
134 normalizing this specific spectrum, which may cause the fit to have a larger error.



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136 **Figure SF5.** Fe K-edge XANES spectra for Fe standards (A) and Fe₃O₄ NPs after variable number
137 of UV-Fenton cycles and solution conditions: (B) 2.55 M H₂O₂, pH 7, (C) 2.55 M H₂O₂, pH 9, (D)
138 5 M H₂O₂, pH 7, (E) 5 M H₂O₂, pH 9.

139

140 **Table ST2.** Linear combination fitting of Fe K-edge XANES using four common Fe standards
 141 with varied Fe speciation and local bonding states. The reported R-factor represents the goodness-
 142 of-fit.

Samples	Fe foil	Fe₂O₃	FeO	Fe₃O₄	R-factor
2.55 M H ₂ O ₂ , pH 7					
1 cycle	0.016	0	0.014	0.970	0.000055
2 cycles	0.049	0	0.015	0.930	0.000325
3 cycles	0.022	0	0.004	0.974	0.000068
2.55 M H ₂ O ₂ , pH 9					
1 cycle	0.075	0	0.032	0.889	0.000633
2 cycles	0.020	0	0.007	0.972	0.000054
3 cycles	0.017	0	0.010	0.974	0.000044
5 M H ₂ O ₂ , pH 7					
1 cycle	0.000	0	0.005	0.997	0.000007
2 cycles	0.010	0	0.004	0.987	0.000025
3 cycles	0.045	0	0.013	0.941	0.000242
5 M H ₂ O ₂ , pH 9					
1 cycle	0.028	0	0.017	0.955	0.000094
2 cycles	0.028	0	0.016	0.956	0.000112
3 cycles	0.022	0	0.006	0.971	0.000059

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146 **Supplementary References**

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