Electrochemical Synthesis of α-Amino Amides via C(sp³)-H Bond Activation

Zhipeng Guan, ‡ Yanan Peng, ‡ Dongfeng Yang, Shuxiang Zhu, Heng Zhang, * Aiwen Lei*

 ^a Institute for Advanced Studies (IAS), College of Chemistry and Molecular Sciences, Wuhan University, Wuhan 430072, People's Republic of China
*Email: <u>aiwenlei@whu.edu.cn; hengzhang@whu.edu.cn</u>

Supporting Information

Table of Contents

General information					
Ex	xperimental section	S3			
1)	General procedure for electrochemical oxidative C(sp ³)-H function	alization of			
	<i>N</i> , <i>N</i> -dimethylanilines with of isocyanides	S3			
2)	Gram-scale experiments	S3			
3)	Screening of cathode materials	S4			
4)	Screening of other nucleophiles	S4			
5)	Control experiments	S5			
6)	General procedure for of cyclic voltammetry (CV) experiments	S7			
Detail descriptions for products					
Re	eference	S20			
¹ H	I NMR and ¹³ C NMR spectra	S21			

General information

All glassware was oven dried at 110 °C for hours and cooled down under vacuum. Unless otherwise noted, all reagents were purchased from commercial suppliers and used without further purification. The instrument for electrolysis was dual display potentiostat (DJS-292B) (made in China). Cyclic voltammograms were obtained on a CHI 605E potentiostat. The anodic electrode was graphite felt (1.5 cm×1.5 cm) and cathodic electrode was platinum sheet (1.5 cm×1.5 cm×0.3 mm). These electrodes were commercially available from GaossUnion and Huanqiujinxin, China. Thin layer chromatography (TLC) employed glass 0.25 mm silica gel plates. Flash chromatography columns were packed with 200-300 mesh silica gel in petroleum (bp. 60-90 °C). GC-MS spectra were recorded on a Varian GC-MS 3900-2100T. The known compounds were characterized by ¹H NMR, ¹³C NMR and ¹⁹F NMR. ¹H, ¹⁹F and ¹³C NMR data were recorded with ADVANCE III 400 MHz with tetramethylsilane as an internal standard. High resolution mass spectra (HRMS) were measured with a Waters Micromass GCT instrument. All chemical shifts (δ) were reported in ppm and coupling constants (*J*) in Hz. All chemical shifts were reported relative to tetramethylsilane (0 ppm for ¹H), Chloroform-*d* (77.16 ppm for ¹³C), respectively.

Experimental Section

1) General procedure for preparation electrochemical oxidative difunctionalization of isocyanides

$$R^{1} \xrightarrow{R^{2}} + CN - R^{3} + H_{2}O + HOAc \longrightarrow R^{1} \xrightarrow{R^{2}} O + HOAc$$

In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar and "Bu₄NOAc (0.5 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm) as the anode and platinum plate (1.5 cm × 1.5 cm × 1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (0.5 mmol), isocyanide (0.3 mmol), H₂O (50 μ L), HOAc (0.5 mL) and MeCN (5.5 mL) were injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 5 mA for 4 h at room temperature. When the reaction was finished, the pure product was purified by flash column chromatography (PE/EA/Et₃N) on silica gel.

2) Gram-scale experiments



In an oven-dried undivided three-necked bottle (100 mL) equipped with a stir bar and "Bu₄NBF₄ (10 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm) as the anode and platinum plate (1.5 cm × 1.5 cm × 1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (10 mmol), isocyanide (6 mmol), H₂O (1

mL), HOAc (7.5 mL) and MeCN (82.5 mL) were injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 20 mA for 22.5 h at room temperature. When the reaction was finished, the pure product was purified by flash column chromatography (PE/EA/Et₃N) on silica gel.

N N 1 0.6mmol	+	CN ^{CO} 2Et 2 0.3 mmol	H ₂ O (50 μL), HOAc (0.5 mL) C(+) Pt(-), ^{<i>n</i>} Bu ₄ NOAc (0.5 mmol) 5 mA, 4 h, N ₂ , rt, MeCN (5.5 mL)	N N CO ₂ Et 3 83% yield
Entry		va	riation from standard conditions	Yield/% ^a
1			none	83 ^b
2			C felt (+) C felt (-)	55
3			C felt (+) C rod (-)	48
4			C felt (+) Ni (-)	56
5			C felt (+) Stainless steel (-)	60
6			C felt (+) C cloth (-)	59
7			C felt (+) Ni foam (-)	36
8			C felt (+) Cu foam(-)	49
9			C felt (+) Cu (-)	79

3) Screening of other cathode materials

Conditions: ^a**1** (0.6 mmol), **2** (0.3 mmol), ^{*n*}Bu₄NOAc (0.5 mmol), H₂O (50 μ L), HOAc (0.5 mL), MeCN (5.5 mL), carbon felt anode, Pt cathode, undivided cell, constant current = 5 mA, 4 h, rt, N₂, GC yield. ^bisolated yield.

4) Screening of other nucleophiles



In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar and "Bu₄NOAc (0.5 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm) as the anode and platinum plate (1.5 cm × 1.5 cm × 1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (0.5 mmol), isocyanide (0.3 mmol), $H_2^{18}O$ (50 µL), HOAc (0.5 mL) and MeCN (5.5 mL) were injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 5 mA for 4 h at room temperature. When the reaction was finished, the pure product was purified by flash column chromatography (PE/EA/Et₃N) on silica gel.



In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar and "Bu₄NOAc (0.5 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm)

as the anode and platinum plate (1.5 cm \times 1.5 cm \times 1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (0.5 mmol), isocyanide (0.3 mmol), D₂O (50 µL), HOAc (0.5 mL) and MeCN (5.5 mL) were injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 5 mA for 4 h at room temperature. When the reaction was finished, the pure product was purified by flash column chromatography (PE/EA/Et₃N) on silica gel.



In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar and "Bu₄NOAc (0.5 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm) as the anode and platinum plate (1.5 cm × 1.5 cm × 1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (0.5 mmol), isocyanide (0.3 mmol), HOAc (0.5 mL) and MeCN (5.5 mL) were injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 5 mA for 4 h at room temperature.



In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar and TEMPO (0.6 mmol), ^{*n*}Bu₄NOAc (0.5 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm) as the anode and platinum plate (1.5 cm × 1.5 cm × 1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (0.5 mmol), isocyanide (0.3 mmol), H₂O (50 μ L), HOAc (0.5 mL) and MeCN (5.5 mL) were injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 5 mA for 4 h at room temperature.



In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar and BHT (0.6 mmol), "Bu₄NOAc (0.5 mmol) were combined and added. The bottle was equipped with carbon felt (1.5 cm×1.5 cm) as the anode and platinum plate (1.5 cm×1.5 cm×1 mm) as the cathode and was then charged with nitrogen. Under the protection of N₂, *N*,*N*-dimethylaniline (0.5 mmol), isocyanide (0.3 mmol), H₂O (50 μ L), HOAc (0.5 mL) and MeCN (5.5 mL) were injected respectively into the

tubes via syringes. The reaction mixture was stirred and electrolyzed at a constant current of 5 mA for 4 h at room temperature.

6) General procedure of cyclic voltammetry (CV) experiment

Cyclic voltammetry was performed in a three electrode cell connected to a schlenk line under nitrogen at room temperature. The working electrode was a glassy carbon electrode, the counter electrode was a platinum wire. The reference electrode was an Ag/AgCl electrode submerged in saturated aqueous KCl solution, and separated from reaction by a salt bridge. 0.2 mmol 1,10 mL of CH₃CN containing 1.0 mmol ^{*n*}Bu₄NOAc were poured into the electrochemical cell in all experiments. The scan rate is 0.1 V/s, ranging from 0 V to 2.0 V. The peak potentials vs. Ag/AgCl for used. The oxidation peak of 1 in acetonitrile was obtained at 1.15 V and 1.6 V. After 1 mL acetic acid was added to the reaction mixture (9 mL CH₃CN), the oxidation peak of 1 decreased to 1.0 V and 1.47 V.

Detail descriptions for products



ethyl *N*-methyl-*N*-phenylglycylglycinate (3). ¹ White solid was obtained in 83% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.34 – 7.24 (m, 2H), 7.14 – 6.98 (m, 1H), 6.85 (t, *J* = 7.4 Hz, 1H), 6.80 – 6.74 (m, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.06 (d, *J* = 5.7 Hz, 2H), 3.91 (s, 2H), 3.04 (s, 3H), 1.26 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.09, 169.65, 149.44, 129.49, 118.90, 113.45, 61.61, 58.89, 41.06, 39.82, 14.24.



ethyl *N*-(4-chlorophenyl)-*N*-methylglycylglycinate (4).¹ White solid was obtained in 76% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.24 – 7.17 (m, 2H), 7.02 (t, *J* = 5.7 Hz, 1H), 6.71 – 6.64 (m, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.04 (d, *J* = 5.7 Hz, 2H), 3.87 (s, 2H), 3.03 (s, 3H), 1.26 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.62, 169.59, 147.92, 129.20, 123.75, 114.49, 61.63, 58.69, 41.00, 39.98, 14.18.



ethyl *N*-(**4**-bromophenyl)-*N*-methylglycylglycinate (5). White solid was obtained in 75% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.39 – 7.32 (m, 2H), 7.01 (t, *J* = 5.8 Hz, 1H), 6.73 – 6.59 (m, 2H), 4.19 (q, *J* = 7.2 Hz, 2H), 4.05 (d, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.04 (s, 3H), 1.28 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.55, 169.58, 148.30, 132.10, 114.92, 110.94, 61.64, 58.56, 41.01, 39.92, 14.19. HRMS (ESI) calculated for C₁₃H₁₈BrN₂O₃⁺[M+H]⁺ 329.0495 found 329.0495.



ethyl *N*-(4-iodophenyl)-*N*-methylglycylglycinate (6). White solid was obtained in 68% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.52 (d, *J* = 8.9 Hz, 2H), 6.98 (t, *J* = 5.7 Hz, 2H), 6.54 (d, *J* = 9.0 Hz, 1H), 4.19 (q, *J* = 7.2 Hz, 2H), 4.04 (d, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.03 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 2H), 4.04 (z, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.03 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 2H), 4.04 (z, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.03 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 2H), 4.04 (z, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.03 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 2H), 4.04 (z, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.03 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 2H), 3.88 (s, 2H)

3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.45, 169.53, 148.83, 137.95, 115.43, 80.28, 61.60, 58.33, 41.00, 39.76, 14.19. HRMS (ESI) calculated for C₁₃H₁₈IN₂O₃⁺ [M+H]⁺ 377.0357 found377.0355.



ethyl *N*-methyl-*N*-(*p*-tolyl)glycylglycinate (7).² White solid was obtained in 76% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.17 (t, *J* = 5.8 Hz, 1H), 7.11 – 7.05 (m, 2H), 6.72 – 6.65 (m, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.04 (d, *J* = 5.8 Hz, 2H), 3.85 (s, 2H), 2.99 (s, 3H), 2.26 (s, 3H), 1.26 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.31, 169.63, 147.36, 129.90, 128.20, 113.66, 61.53, 59.16, 40.98, 40.00, 20.34, 14.19.



ethyl *N*-(4-methoxyphenyl)-*N*-methylglycylglycinate (8). White solid was obtained in 78% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.28 (t, *J* = 5.8 Hz, 1H), 6.89 – 6.83 (m, 2H), 6.80 – 6.72 (m, 2H), 4.19 (q, *J* = 7.1 Hz, 2H), 4.07 (d, *J* = 5.7 Hz, 2H), 3.81 (s, 2H), 3.77 (s, 3H), 2.96 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.37, 169.74, 153.29, 144.17, 115.55, 114.86, 61.61, 59.96, 55.80, 41.04, 40.70, 14.25. HRMS (ESI) calculated for C₁₄H₂₁N₂O₄+[M+H]+ 281.1496 found 281.1498.



ethyl *N*-(4-acetoxyphenyl)-*N*-methylglycylglycinate (9). White solid was obtained in 76% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.12 (t, *J* = 5.8 Hz, 1H), 6.98 (d, *J* = 9.0 Hz, 1H), 6.74 (d, *J* = 9.1 Hz, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.04 (d, *J* = 5.8 Hz, 2H), 3.88 (s, 2H), 3.02 (s, 3H), 2.27 (s, 3H), 1.26 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.85, 170.14, 169.54, 147.34, 143.01, 122.27, 113.94, 61.53, 58.98, 40.97, 40.05, 21.09, 14.16. HRMS (ESI) calculated for C₁₅H₂₁N₂O₅⁺ [M+H]⁺ 309.1445 found 309.1447.



ethyl *N*-methyl-*N*-(m-tolyl)glycylglycinate (10). White solid was obtained in 66% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.20 – 7.13 (m, 1H), 7.08 (d, *J* = 6.4 Hz, 1H), 6.71 – 6.65 (m, 1H),

6.59 (d, J = 6.6 Hz, 2H), 4.18 (q, J = 7.1 Hz, 2H), 4.06 (d, J = 5.7 Hz, 2H), 3.89 (s, 2H), 3.02 (s, 3H), 2.33 (s, 3H), 1.26 (t, J = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.22, 169.64, 149.52, 139.34, 129.34, 119.81, 114.22, 110.60, 61.61, 58.92, 41.05, 39.83, 21.94, 14.25. HRMS (ESI) calculated for C₁₄H₂₀N₂NaO₃⁺[M+Na]⁺ 287.1366 found 287.1368.



ethyl *N*-(2-bromophenyl)-*N*-methylglycylglycinate (11). White solid was obtained in 62% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.89 (t, *J* = 5.8 Hz, 1H), 7.59 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.31 (td, *J* = 7.7, 1.5 Hz, 1H), 7.18 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.00 (td, *J* = 7.7, 1.6 Hz, 1H), 4.23 (q, *J* = 7.1 Hz, 2H), 4.14 (d, *J* = 5.7 Hz, 2H), 3.66 (s, 2H), 2.81 (s, 3H), 1.30 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.39, 169.80, 150.26, 133.83, 128.64, 125.83, 122.64, 120.51, 61.55, 60.70, 42.65, 41.10, 14.29. HRMS (ESI) calculated for C₁₃H₁₇BrN₂NaO₃+[M+Na]+ 351.0315 found 351.0315.



ethyl *N*-(3-bromophenyl)-*N*-methylglycylglycinate (12). White solid was obtained in 70% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.12 (t, *J* = 8.1 Hz, 1H), 6.99 – 6.87 (m, 3H), 6.66 (m, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 4.06 (d, *J* = 5.6 Hz, 2H), 3.91 (s, 2H), 3.04 (s, 3H), 1.27 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.39, 169.59, 150.51, 130.72, 123.62, 121.63, 116.10, 111.89, 61.72, 58.37, 41.10, 39.78, 14.25. HRMS (ESI) calculated for C₁₃H₁₇BrN₂NaO₃⁺ [M+Na]⁺ 351.0315 found 351.0319.



ethyl *N*-methyl-*N*-(2-phenoxyphenyl)glycylglycinate (13). White solid was obtained in 72% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.31 – 7.23 (m, 2H), 7.18 – 7.08 (m, 2H), 7.06 – 6.98 (m, 3H), 6.91 – 6.86 (m, 2H), 6.82 (d, *J* = 6.9 Hz, 1H), 4.14 (q, *J* = 7.1 Hz, 2H), 3.73 (d, *J* = 6.0 Hz, 2H), 3.69 (s, 2H), 2.83 (s, 3H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.75, 169.59, 157.81, 147.95, 143.93, 129.79, 125.30, 123.71, 122.45, 122.13, 120.00, 116.62, 61.31, 59.92, 41.12, 40.89, 14.25. HRMS (ESI) calculated for C₁₉H₂₃N₂O₄+[M+H]⁺ 343.1652 found 343.1650.



ethyl *N*-(2,4-dimethylphenyl)-*N*-methylglycylglycinate (14). White solid was obtained in 78% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.94 (t, *J* = 5.5 Hz, 1H), 7.06 – 6.96 (m, 3H), 4.25 (q, *J* =

7.1 Hz, 2H), 4.16 (d, J = 5.3 Hz, 2H), 3.73 (s, 2H), 2.79 (s, 3H), 2.40 (s, 6H), 1.31 (t, J = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.79, 169.95, 148.92, 136.25, 129.60, 126.13, 61.60, 60.60, 41.47, 41.20, 19.76, 14.26. HRMS (ESI) calculated for C₁₅H₂₃N₂O₃⁺[M+H]⁺ 279.1703 found 279.1703.



ethyl *N*-(3,5-dimethylphenyl)-*N*-methylglycylglycinate (15). White solid was obtained in 78% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.10 (t, *J* = 5.8 Hz, 1H), 6.51 (s, 1H), 6.41 (s, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.05 (d, *J* = 5.8 Hz, 2H), 3.88 (s, 2H), 3.01 (s, 3H), 2.28 (s, 6H), 1.26 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.32, 169.61, 149.60, 139.15, 120.81, 111.38, 61.54, 58.90, 41.02, 39.79, 21.77, 14.22. HRMS (ESI) calculated for C₁₅H₂₃N₂O₃+[M+H]+ 279.1703 found 279.1705.



ethyl *N*-(**3,5-dichlorophenyl**)-*N*-methylglycylglycinate (**16**). White solid was obtained in 72% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.81 (m, 1H), 6.76 (d, *J* = 5.7 Hz, 1H), 6.60 (d, *J* = 1.7 Hz, 2H), 4.21 (q, *J* = 7.1 Hz, 2H), 4.07 (d, *J* = 5.6 Hz, 2H), 3.92 (s, 2H), 3.05 (s, 3H), 1.27 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.69, 169.56, 150.76, 135.93, 118.53, 111.51, 61.83, 57.93, 41.19, 39.83, 14.26. HRMS (ESI) calculated for C₁₃H₁₇Cl₂N₂O₃⁺[M+H]⁺ 319.0611 found 319.0618.



ethyl *N*-octyl-*N*-(*p*-tolyl)glycylglycinate (17). White solid was obtained in 70% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.09 – 7.02 (m, 3H), 6.68 – 6.61 (m, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 4.02 (d, *J* = 5.7 Hz, 2H), 3.87 (s, 2H), 2.25 (s, 3H), 1.61 (t, *J* = 7.7 Hz, 2H), 1.34 – 1.21 (m, 13H), 0.95 – 0.84 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.39, 169.55, 145.76, 130.00, 127.62, 113.53, 61.51, 56.59, 52.51, 41.04, 31.87, 29.49, 29.36, 27.18, 26.59, 22.73, 20.31, 14.20. HRMS (ESI) calculated for C₂₁H₃₅N₂O₃⁺[M+H]⁺ 363.2642 found 362.2645.



ethyl *N*-(4-fluorophenyl)-*N*-octylglycylglycinate (18). White solid was obtained in 67% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.04 (t, *J* = 5.6 Hz, 1H), 6.99 – 6.91 (m, 2H), 6.71 – 6.62 (m, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.03 (d, *J* = 5.6 Hz, 2H), 3.85 (s, 2H), 3.36 – 3.27 (m, 2H), 1.60 (t, *J* = 7.8 Hz, 2H), 1.33 – 1.24 (m, 13H), 0.92 – 0.85 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.04, 169.59, 156.44 (d, *J* = 237.7 Hz), 144.60, 115.96 (d, *J* = 22.1 Hz), 114.93 (d, *J* = 7.4 Hz), 77.48, 77.16, 76.84, 61.62, 57.00, 53.13, 41.10, 31.89, 29.50, 29.37, 27.19, 26.67, 22.75, 14.23, 14.21. ¹⁹F NMR (377 MHz, CDCl₃) δ -126.74. HRMS (ESI) calculated for C₂₀H₃₂FN₂O₃+[M+H]⁺ 367.2391 found 367.2392.



ethyl *N*-(4-phenylbutyl)-*N*-(*p*-tolyl)glycylglycinate (19). White solid was obtained in 66% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.31 – 7.23 (m, 2H), 7.18 (m, 3H), 7.04 (m, 2H), 6.62 (d, *J* = 8.6 Hz, 2H), 4.16 (q, *J* = 7.2 Hz, 2H), 4.00 (d, *J* = 5.7 Hz, 2H), 3.85 (s, 2H), 3.41 – 3.30 (m, 2H), 2.69 – 2.56 (m, 2H), 2.25 (s, 3H), 1.66 (p, *J* = 3.5 Hz, 4H), 1.24 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.27, 169.56, 145.72, 142.06, 130.04, 128.46, 128.44, 127.81, 125.97, 113.66, 61.53, 56.60, 52.40, 41.06, 35.73, 28.95, 26.29, 20.33, 14.22. HRMS (ESI) calculated for C₂₃H₃₁N₂O₃+[M+H]+ 383.2329 found 383.2331.



ethyl *N*-(cyclobutylmethyl)-*N*-(*p*-tolyl)glycylglycinate (20). White solid was obtained in 66% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.05 (m, 3H), 6.66 (d, *J* = 8.3 Hz, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 4.01 (d, *J* = 5.7 Hz, 2H), 3.87 (s, 2H), 3.36 (d, *J* = 7.0 Hz, 2H), 2.69 (p, *J* = 7.6 Hz, 1H), 2.26 (s, 3H), 2.14 – 2.02 (m, 2H), 1.97 – 1.82 (m, 2H), 1.77 (m, 2H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.38, 169.54, 146.37, 129.96, 128.09, 114.08, 61.54, 58.59, 57.15, 41.12, 34.13, 27.27, 20.36, 18.76, 14.22. HRMS (ESI) calculated for C₁₈H₂₇N₂O₃+[M+H]+ 319.2016 found 319.2017.



ethyl *N*-(2-methoxyethyl)-*N*-(*p*-tolyl)glycylglycinate (21). White solid was obtained in 47% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.80 (s, 1H), 7.11 – 7.02 (m, 2H), 6.64 (d, *J* = 8.6 Hz, 2H), 4.17 (q, *J* = 7.2 Hz, 2H), 4.01 (d, *J* = 5.8 Hz, 2H), 3.95 (s, 2H), 3.72 – 3.57 (m, 3H), 3.35 (s, 3H), 2.26 (s, 3H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 172.11, 169.69, 145.54, 130.01, 127.86, 113.52, 70.04, 61.41, 58.98, 58.63, 52.54, 41.29, 20.37, 14.26. HRMS (ESI) calculated for C₁₆H₂₄N₂NaO₄+[M+Na]+ 331.1628 found 331.1630.



ethyl *N*-(**2**-(**1,3-dioxolan-2-yl)ethyl)-***N***-(p-tolyl)glycylglycinate** (**22**). White solid was obtained in 72% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.12 (t, *J* = 5.6 Hz, 1H), 7.06 (d, *J* = 8.3 Hz, 2H), 6.72 – 6.59 (m, 2H), 4.63 (t, *J* = 4.7 Hz, 1H), 4.16 (q, *J* = 7.1 Hz, 2H), 4.13 – 4.06 (m, 2H), 4.00 (d, *J* = 5.7 Hz, 2H), 3.87 (s, 2H), 3.75 (m, 2H), 3.53 (t, *J* = 7.2 Hz, 2H), 2.25 (s, 3H), 1.92 (m, 2H), 1.24 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.35, 169.58, 145.52, 130.05, 127.76, 113.62, 100.47, 67.01, 61.53, 56.62, 47.22, 41.15, 32.38, 25.79, 20.36, 14.23. HRMS (ESI) calculated for C_{18H27}N₂O₅+[M+H]⁺ 351.1914 found 351.1912.



ethyl *N*-(4-acetoxybutyl)-*N*-(*p*-tolyl)glycylglycinate (23). White solid was obtained in 73% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.09 – 7.05 (m, 2H), 7.04 (t, *J* = 5.9 Hz, 1H), 6.67 – 6.63 (m, 2H), 4.17 (q, *J* = 7.2 Hz, 2H), 4.11 – 4.07 (m, 2H), 4.02 (d, *J* = 5.6 Hz, 2H), 3.88 (s, 2H), 3.42 – 3.35 (m, 2H), 2.26 (s, 3H), 2.05 (s, 3H), 1.71 – 1.66 (m, 4H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.27, 171.18, 169.60, 145.57, 130.11, 128.10, 113.78, 64.10, 61.58, 56.64, 52.14, 41.06, 26.22, 23.25, 21.10, 20.35, 14.22. HRMS (ESI) calculated for C₁₉H₂₉N₂O₅⁺[M+H]⁺ 365.2071 found 365.2075.



ethyl *N*-(3-cyanopropyl)-*N*-phenylglycylglycinate (24). White solid was obtained in 44% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.10 (d, *J* = 8.2 Hz, 2H), 6.98 (t, *J* = 5.6 Hz, 1H), 6.71 (d, *J* = 8.5 Hz, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.03 (d, *J* = 5.6 Hz, 2H), 3.88 (s, 2H), 3.49 (dd, *J* = 8.4, 6.4 Hz, 2H), 2.44 (t, *J* = 6.9 Hz, 2H), 2.27 (s, 3H), 1.98 (p, *J* = 7.1 Hz, 2H), 1.26 (t, *J* = 7.1 Hz, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 170.63, 169.70, 145.24, 130.33, 129.32, 119.17, 114.70, 61.70, 57.19, 51.36, 41.10, 22.81, 20.43, 15.02, 14.25. HRMS (ESI) calculated for C₁₇H₂₄N₃O₃⁺[M+H]⁺ 318.1812 found 318.1811.



ethyl *N*-(4-chlorobutyl)-*N*-(*p*-tolyl)glycylglycinate(25) White solid was obtained in 53% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) ¹H NMR (400 MHz, Chloroform-d) δ 7.32 – 7.23 (m, 2H), 7.18 (m, 2H), 7.04 (dd, *J* = 10.7, 8.3 Hz, 3H), 6.67 – 6.52 (m, 2H), 4.16 (q, *J* = 7.2 Hz, 2H), 4.00 (d, *J* = 5.7 Hz, 2H), 3.85 (s, 2H), 3.40 – 3.30 (m, 2H), 2.70 – 2.59 (m, 2H), 2.25 (s, 3H), 1.66 (p, *J* = 3.5 Hz, 4H), 1.24 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.27, 169.56, 145.72, 142.06, 130.04, 128.46, 128.44, 127.81, 125.97, 113.66, 61.53, 56.60, 52.40, 41.06, 35.73, 28.95, 26.29, 20.33, 14.22. HRMS (ESI) calculated for C₁₇H₂₆ClN₂O₃+[M+H]+ 341.1626 found 341.1628.



ethyl *N*-(4-(1,3-dioxoisoindolin-2-yl)butyl)-*N*-(*p*-tolyl)glycylglycinate (26). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.86 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.74 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.11 – 7.03 (m, 3H), 6.67 (d, *J* = 8.2 Hz, 2H), 4.15 (q, *J* = 7.2 Hz, 2H), 4.04 (d, *J* = 5.7 Hz, 2H), 3.88 (s, 2H), 3.73 (t, *J* = 6.9 Hz, 2H), 3.41 (t, *J* = 7.4 Hz, 2H), 2.26 (s, 3H), 1.83 – 1.63 (m, 4H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.14, 169.55, 168.46, 145.62, 134.04, 132.11, 130.04, 128.08, 123.32, 113.94, 61.45, 56.78, 52.05, 41.06, 37.58, 26.19, 23.98, 20.32, 14.19. HRMS (ESI) calculated for C₂₂H₂₄N₃O₅+[M+H]+ 407.1710 found 407.1709.



ethyl *N*-(*pent*-4-en-1-yl)-*N*-(*p*-tolyl)glycylglycinate (27). White solid was obtained in 75% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.13 – 6.96 (m, 3H), 6.70 – 6.60 (m, 2H), 5.82 (m, 1H), 5.09 – 4.95 (m, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 4.02 (d, *J* = 5.6 Hz, 2H), 3.87 (s, 2H), 3.41 – 3.31 (m, 2H), 2.26 (s, 3H), 2.17 – 2.07 (m, 2H), 1.81 – 1.66 (m, 2H), 1.25 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.30, 169.60, 145.69, 137.70, 130.09, 127.89, 115.58, 113.69, 61.58, 56.68, 51.99, 41.10, 31.22, 25.67, 20.36, 14.24. HRMS (ESI) calculated for C₁₈H₂₇N₂O₃+[M+H]+ 319.2016 found 319.2018.



ethyl *N*-(but-3-yn-1-yl)-*N*-(*p*-tolyl)glycylglycinate (28). White solid was obtained in 71% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.15 (t, *J* = 5.7 Hz, 1H), 7.08 (d, *J* = 8.4 Hz, 2H), 6.69 (d, *J* = 8.6 Hz, 2H), 4.17 (q, *J* = 7.2 Hz, 2H), 4.01 (d, *J* = 5.7 Hz, 2H), 3.94 (s, 2H), 3.58 (t, *J* = 7.0 Hz, 2H), 2.51 (td, *J* = 7.0, 2.7 Hz, 2H), 2.26 (s, 3H), 2.03 (t, *J* = 2.7 Hz, 1H), 1.25 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.95, 169.53, 145.07, 130.17, 128.76, 114.21, 81.85, 70.51, 61.54, 57.18, 51.57, 41.07, 20.38, 17.18, 14.22. HRMS (ESI) calculated for C₁₃H₁₇N₂O₃⁺[M+H]⁺ 303.1703 found 303.1708.



ethyl *N*-isopropyl-*N*-phenylglycylglycinate (29). White solid was obtained in 79% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.31 – 7.23 (m, 2H), 7.17 (t, *J* = 5.9 Hz, 1H), 6.87 – 6.75 (m, 3H), 4.17 (p, *J* = 6.8 Hz, 3H), 4.04 (d, *J* = 5.8 Hz, 2H), 3.78 (s, 2H), 1.32 – 1.18 (m, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 171.88, 169.59, 148.69, 129.50, 118.76, 114.22, 61.50, 49.77, 48.36, 40.98, 19.41, 14.19. HRMS (ESI) calculated for C₁₅H₂₃N₂O₃⁺[M+H]⁺ 279.1703 found 279.1701.



ethyl *N*-(*sec*-butyl)-*N*-(*p*-tolyl)glycylglycinate (30). White solid was obtained in 67% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.14 (t, *J* = 5.6 Hz, 1H), 7.09 – 7.03 (m, 2H), 6.77 – 6.69 (m, 2H), 4.16 (q, *J* = 7.1 Hz, 2H), 4.00 (d, *J* = 5.7 Hz, 2H), 3.80 (dt, *J* = 8.1, 6.2 Hz, 1H), 3.74 (s, 2H), 2.26 (s, 3H), 1.73 – 1.60 (m, 1H), 1.52 – 1.37 (m, 1H), 1.24 (t, *J* = 7.1 Hz, 3H), 1.16 (d, *J* = 6.6 Hz, 3H), 0.98 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.82, 169.57, 146.63, 129.99, 128.37, 115.04, 61.48, 56.07, 49.87, 41.09, 27.16, 20.34, 16.79, 14.20, 11.74. HRMS (ESI) calculated for C₁₇H₂₇N₂O₃+[M+H]+ 307.2016 found 307.2018.



ethyl *N*-cyclohexyl-*N*-phenylglycylglycinate (31). White solid was obtained in 79% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 – 7.22 (m, 2H), 7.14 (t, *J* = 5.8 Hz, 1H), 6.86 – 6.72 (m, 3H), 4.16 (q, *J* = 7.1 Hz, 2H), 4.03 (d, *J* = 5.7 Hz, 2H), 3.82 (s, 2H), 3.69 (m, 1H), 1.98 – 1.82 (m, 4H), 1.77 – 1.64 (m, 1H), 1.49 – 1.28 (m, 5H), 1.25 (t, *J* = 7.1 Hz, 3H), 1.20 – 1.07 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 171.95, 169.58, 148.59, 129.49, 118.61, 114.04, 61.50, 57.27, 50.90, 41.03, 29.92, 26.02, 25.77, 14.19. HRMS (ESI) calculated for C₁₈H₂₇N₂O₃⁺[M+H]⁺ 319.2016 found 319.2017.



ethyl diphenylglycylglycinate (32). White solid was obtained in 50% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.28 (m, 4H), 7.11 (t, *J* = 5.5 Hz, 1H), 7.07 – 7.01 (m, 6H), 4.37 (s, 2H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.05 (d, *J* = 5.4 Hz, 2H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.64, 169.55, 147.50, 129.78, 122.87, 120.83, 61.66, 57.52, 41.32, 14.23. HRMS (ESI) calculated for C₁₈H₂₁N₂O₃+[M+H]⁺ 313.1547 found 313.1545.



ethyl *N*-phenyl-*N*-(*p*-tolyl)glycylglycinate (33). White solid was obtained in 49% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.29 – 7.22 (m, 2H), 7.14 (t, *J* = 8.3 Hz, 3H), 7.05 – 6.99 (m, 2H), 6.98 – 6.89 (m, 3H), 4.33 (s, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 4.04 (d, *J* = 5.4 Hz, 2H), 2.33 (s, 3H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.75, 169.55, 147.97, 144.95, 133.57, 130.50, 129.56, 122.74, 121.58, 118.81, 61.62, 57.68, 41.33, 20.87, 14.22. HRMS (ESI) calculated for C₁₉H₂₃N₂O₃+[M+H]⁺ 327.1703 found 327.1709.



ethyl *N*-ethyl-*N*-phenylalanylglycinate (**34**). White solid was obtained in 43% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 – 7.23 (m, 2H), 7.16 (s, 1H), 6.89 – 6.80 (m, 3H), 4.27 – 4.07 (m, 4H), 3.97 (dd, *J* = 18.2, 5.2 Hz, 1H), 3.32 (dp, *J* = 21.8, 7.2 Hz, 2H), 1.42 (d, *J* = 7.1 Hz, 3H), 1.27 (t, *J* = 7.1 Hz, 3H), 1.20 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 174.05, 169.91, 147.52, 129.37, 119.50, 116.82, 61.56, 61.21, 42.51, 41.44, 14.28, 13.69, 13.43. HRMS (ESI) calculated for C₁₅H₂₃N₂O₃⁺[M+H]⁺ 279.1703 found 279.1707.



ethyl phenylprolylglycinate (35). White solid was obtained in 69% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 – 7.21 (m, 2H), 7.04 (t, *J* = 5.8 Hz, 1H), 6.81 (tt, *J* = 7.4, 1.1 Hz, 1H), 6.72 – 6.61 (m, 2H), 4.22 – 4.10 (m, 3H), 4.05 (dd, *J* = 8.4, 3.2 Hz, 1H), 3.82 (dd, *J* = 18.1, 4.9 Hz, 1H), 3.72 – 3.58 (m, 1H), 3.23 (q, *J* = 8.8 Hz, 1H), 2.34 – 2.19 (m, 2H), 2.12 – 1.99 (m, 2H), 1.24 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 174.42, 169.64, 147.49, 129.38, 118.24, 113.19, 64.33, 61.47, 49.72, 41.04, 31.57, 24.19, 14.18. HRMS (ESI) calculated for C₁₅H₂₁N₂O₃+[M+H]+ 277.1547 found 277.1547.



ethyl (1-phenylpiperidine-2-carbonyl)glycinate (36). White solid was obtained in 58% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.27 (t, *J* = 7.9 Hz, 2H), 7.01 (t, *J* = 5.8 Hz, 1H), 6.96 (d, *J* = 8.1 Hz, 2H), 6.87 (t, *J* = 7.3 Hz, 1H), 4.24 – 4.05 (m, 4H), 3.79 (dd, *J* = 18.2, 4.8 Hz, 1H), 3.46 – 3.25 (m, 2H), 2.19 (m, 1H), 1.84 (m, 1H), 1.64 (m, 4H), 1.25 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 173.35, 169.91, 150.66, 129.49, 120.11, 116.87, 77.48, 77.16, 76.84, 61.46, 47.92, 41.17, 26.09, 23.92, 21.32, 14.22. HRMS (ESI) calculated for C₁₆H₂₃N₂O₃⁺[M+H]⁺ 291.1703 found 291.1704.



methyl *N*-methyl-*N*-(*p*-tolyl)glycylglycinate (37). White solid was obtained in 91% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.19 (t, *J* = 5.8 Hz, 1H), 7.11 (d, *J* = 8.3 Hz, 2H), 6.72 (d, *J* = 8.7 Hz, 1H), 4.09 (d, *J* = 5.8 Hz, 2H), 3.88 (s, 2H), 3.76 (s, 3H), 3.03 (s, 3H), 2.30 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.41, 170.14, 147.39, 129.93, 128.30, 113.73, 59.19, 52.42, 40.86, 40.07, 20.36. HRMS (ESI) calculated for C₁₆H₂₃N₂O₃+[M+H]⁺ 251.1390 found 251.1392.



N-benzyl-2-(methyl(*p*-tolyl)amino)acetamide (38).³ White solid was obtained in 72% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.15 (m, 7H), 6.93 (s, 1H), 6.84 (td, *J* = 7.3, 1.0 Hz, 1H), 6.73 (dt, *J* = 7.8, 1.1 Hz, 2H), 4.48 (d, *J* = 6.0 Hz, 2H), 3.91 (s, 2H), 2.99 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.50, 149.36, 138.12, 129.49, 128.77, 127.60, 127.56, 118.86, 113.36, 59.10, 43.21, 40.01.



2-(methyl(phenyl)amino)-*N*-(tosylmethyl)acetamide (39).¹ White solid was obtained in 51% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.63 (d, *J* = 8.1 Hz, 2H), 7.33 (t, *J* = 6.9 Hz, 1H), 7.28 – 7.15 (m, 4H), 6.80 (t, *J* = 7.3 Hz, 1H), 6.58 (d, *J* = 8.1 Hz, 2H), 4.58 (d, *J* = 6.9 Hz, 2H), 3.66 (s, 2H), 2.90 (s, 3H), 2.37 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.43, 149.06, 145.60, 133.81, 130.00, 129.50, 128.91, 119.18, 113.43, 59.93, 58.51, 40.07, 21.87.



N-cyclohexyl-2-(methyl(phenyl)amino)acetamide (40). ¹ White solid was obtained in 81% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.33 – 7.22 (m, 2H), 6.84 (t, *J* = 7.3 Hz, 1H), 6.78 – 6.69 (m, 2H), 6.54 – 6.36 (m, 1H), 3.82 (m, 3H), 2.99 (s, 3H), 1.92 – 1.82 (m, 2H), 1.70 – 1.51 (m, 3H), 1.34 (m, 2H), 1.08 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.34, 149.57, 129.45, 118.78, 113.37, 59.30, 47.93, 39.82, 33.13, 25.53, 24.88.



N-(*tert*-butyl)-2-(methyl(*p*-tolyl)amino)acetamide (41).¹ White solid was obtained in 69% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.08 (d, *J* = 8.4 Hz, 2H), 6.72 – 6.62 (m, 2H), 6.49 (s, 1H), 3.67 (s, 2H), 2.94 (s, 3H), 2.27 (s, 3H), 1.33 (s, 8H). ¹³C NMR (101 MHz, CDCl₃) δ 169.84, 147.61, 129.88, 128.20, 113.75, 60.34, 50.95, 40.11, 28.81, 20.38.



2-(methyl(phenyl)amino)-*N*-(**2**,**4**,**4-trimethylpentan-2-yl)acetamide (42).** White solid was obtained in 75% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.32 – 7.23 (m, 2H), 6.84 (m, 1H), 6.77 – 6.71 (m, 2H), 6.56 (s, 1H), 3.72 (s, 2H), 2.97 (s, 3H), 1.66 (s, 2H), 1.40 (s, 6H), 0.94 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 169.30, 149.50, 129.37, 118.77, 113.41, 60.12, 54.98, 52.66, 39.89, 31.70, 31.52, 28.88. HRMS (ESI) calculated for C₁₇H₂₉N₂O⁺[M+H]⁺ 277.2274 found 277.2274.



N-(2,6-dimethylphenyl)-2-(methyl(*p*-tolyl)amino)acetamide (43). White solid was obtained in 33% isolated yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.21 (m, 3H), 7.14 (m, 2H), 6.76 – 6.64 (m, 3H),

 $4.44 (s, 2H), 3.01 (s, 3H), 2.16 (m, 9H). {}^{13}C NMR (101 MHz, CDCl_3) \\ \delta 173.10, 172.59, 149.12, 136.62, 135.67, 129.28, 129.24, 117.33, 112.36, 58.55, 39.52, 25.86, 18.09. HRMS (ESI) calculated for C_{18}H_{23}N_2O^+[M+H]^+ 283.1805 found 283.1804.$

Reference:

(1) Guerrero, I.; San Segundo, M.; Correa, A., Iron-catalyzed C(sp³)-H functionalization of N,N-dimethylanilines with isocyanides. *Chem. Commun.* **2018**, *54*, 1627-1630.

(2) Li, D.; Shen, X.; Lei, J., Metal-Free Iodine/TEMPO-Mediated Aerobic Oxidative Ugi-Type Multicomponent Reactions with Tertiary Amines. *J. Org. Chem.* **2020**, *85*, 2466-2475.

(3) Chanthamath, S.; Thongjareun, S.; Shibatomi, K.; Iwasa, S., Ru(II)-Pheox catalyzed N-H insertion reaction of diazoacetamides: synthesis of N-substituted α -aminoamides. *Tetrahedron Lett.* **2012**, *53*, 4862-4865.















210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)











120 110 100 90 80 fl (ppm) -10











10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)













S57

S61

