

Supplementary Information

Fenton-like Chemistry Enables Catalytic Oxidative Desulfurization of Thioacetals and Thioketals with Hydrogen Peroxide

Guodong Zhao^{#*a,b}, Yaxin Wang^{#a}, Cheng Wang^{#a}, Haimin Lei^a, Bingqing Yi^a and Rongbiao Tong^{*b,c}

^aSchool of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing 102488, China. E-mail: 202001036@bucm.edu.cn; Fax: +(86) 010-64213817; Tel: +(86) 010-64286426

^bDepartment of Chemistry and ^cthe Southern Marine Science and Engineering Guangdong Laboratory (Guangzhou), The Hong Kong University of Science and Technology, Clearwater Bay, Kowloon, Hong Kong, China. E-mail: rtong@ust.hk; Fax: +(852) 23581594; Tel: +(852) 23587357

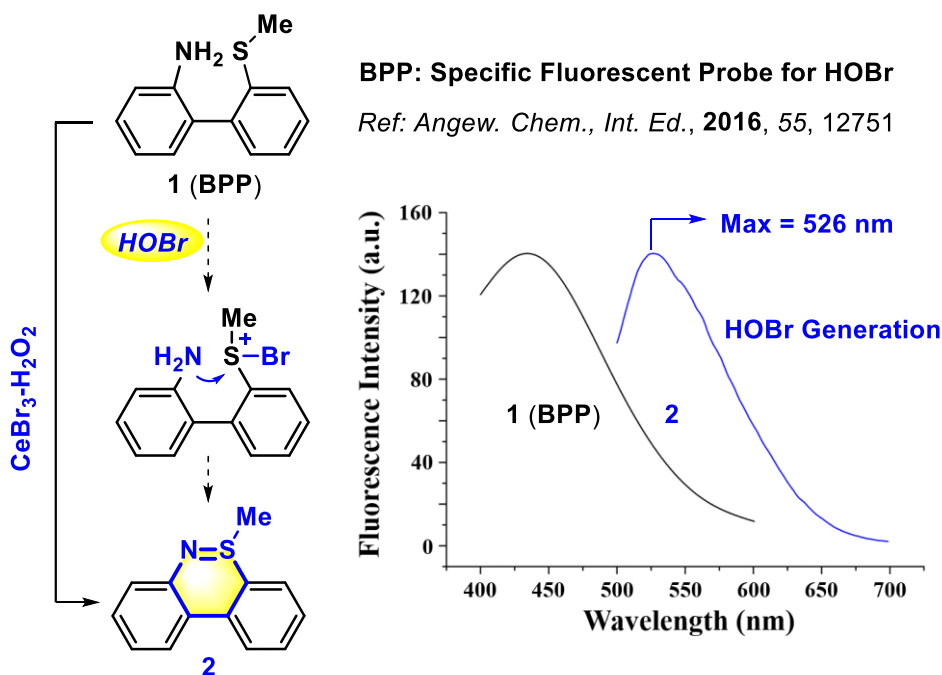
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1. General Information

Reactions were carried out in a round-bottom flask with vigorous stirring at room temperature with open-air condition, unless otherwise noted. Anhydrous ethanol (EtOH), acetonitrile (MeCN), tetrahydrofuran (THF), dichloromethane (DCM), dimethyl sulfoxide (DMSO), *N,N*-dimethylformamide (DMF), 1-butanol (*n*-BuOH) and pyridine were used as received without further purification unless otherwise noted. Anhydrous CeBr₃, FeBr₂ and FeBr₃ were purchased from Sigma-Aldrich and dried under vacuum before use. Solvents were used as received from commercial suppliers without prior purification for workup, extraction and column chromatography. Reactions were monitored by thin-layer chromatography (TLC, 0.25 mm) on pre-coated silica gel plates. Flash chromatography was performed with silica gel 60 (particle size 0.040-0.062 mm). ¹H- and ¹³C-NMR spectra were recorded on a 400 MHz spectrometer (400 MHz for ¹H, 100 MHz for ¹³C). Chemical shifts are reported in parts per million (ppm) as values relative to the internal chloroform (7.26 ppm for ¹H and 77.16 ppm for ¹³C) or DMSO (2.50 ppm for ¹H and 39.52 ppm for ¹³C). Abbreviations for signal coupling are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Mass spectra were detected by ESI-TOF (Agilent 6520 or G6125B) or GC-MS (Agilent 7890A/5975C). Infrared spectrometry was recorded on Nicolet iS10 FT-IR (Thermo). Fluorescence emission spectra were recorded on LS-45 fluorescence spectrophotometer (PerkinElmer). Optical rotations were measured on Autopol I polarimeter (Rudolph Research Analytical) with $[\alpha]_D$ values reported in degrees. Melting points were determined on MP70 instrument (Mettler Toledo) without correction.

2. [Br⁺] from CeBr₃-H₂O₂ system was confirmed to be HOBr.

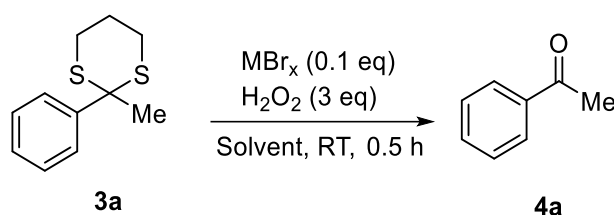


To a stirred solution of fluorescent probe **BPP**¹ (compound **1**, 108 mg, 0.5 mmol) in MeCN/H₂O (3/1, 10 mL) were added CeBr₃ (56.9 mg, 0.15 mmol) and H₂O₂ (30%, 150 μL, 1.5 mmol). After completion of the addition, the reaction mixture was stirred at room temperature (rt) for 1.5 h. The reaction was quenched by aqueous Na₂S₂O₃ solution (0.1 M, 50 mL) and ethyl acetate (50 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (2 × 20 mL). The combined organic fractions were washed with H₂O, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. Compound **2**¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:1 to 3:1) as a gray solid (85.3 mg, 80%). M.P. 142–144 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 7.95 (d, *J* = 8.1 Hz, 1H), 7.89 (d, *J* = 8.0 Hz, 1H), 7.63–7.57 (m, 1H), 7.42 (d, *J* = 4.6 Hz, 2H), 7.34–7.27 (m, 1H), 7.22 (d, *J* = 8.2 Hz, 1H), 6.97 (t, *J* = 7.5 Hz, 1H), 2.35 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 148.7, 132.6, 131.9, 130.5, 127.6, 125.8, 124.8, 124.6, 124.3, 123.6, 120.8, 119.8, 32.1. IR 2971.9, 2895.4, 1590.2, 1563.1, 1465.4, 1413.1, 1280.4, 1218.5, 1148.6, 1109.4, 1037.1, 738.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₃H₁₂NS [M+H]⁺ 214.0685; found 214.0683.

Notably, no product **2** was furnished after the combination of Br₂ (38.4 μL, 0.75 mmol) or H₂O₂ (30%, 150 μL, 1.5 mmol) with fluorescent probe **BPP** (108 mg, 0.5 mmol) following the above protocol (1.5 h at rt in MeCN/H₂O 3/1), further indicating the specificity of fluorescent probe **BPP** for HOBr¹ detection.

The emission spectra of fluorescent probe **BPP** **1** (0.9 mM in MeCN, black) and **2** (7.5 mM in MeCN, blue) were got under the excitation wavelength of 375 nm and 480 nm respectively, and the emission wavelength collected were 400–600 nm (for **BPP**) and 500–700 nm (for compound **2**), which were reported in the literature¹.

3. Optimization of the 1,3-dithiane deprotection.



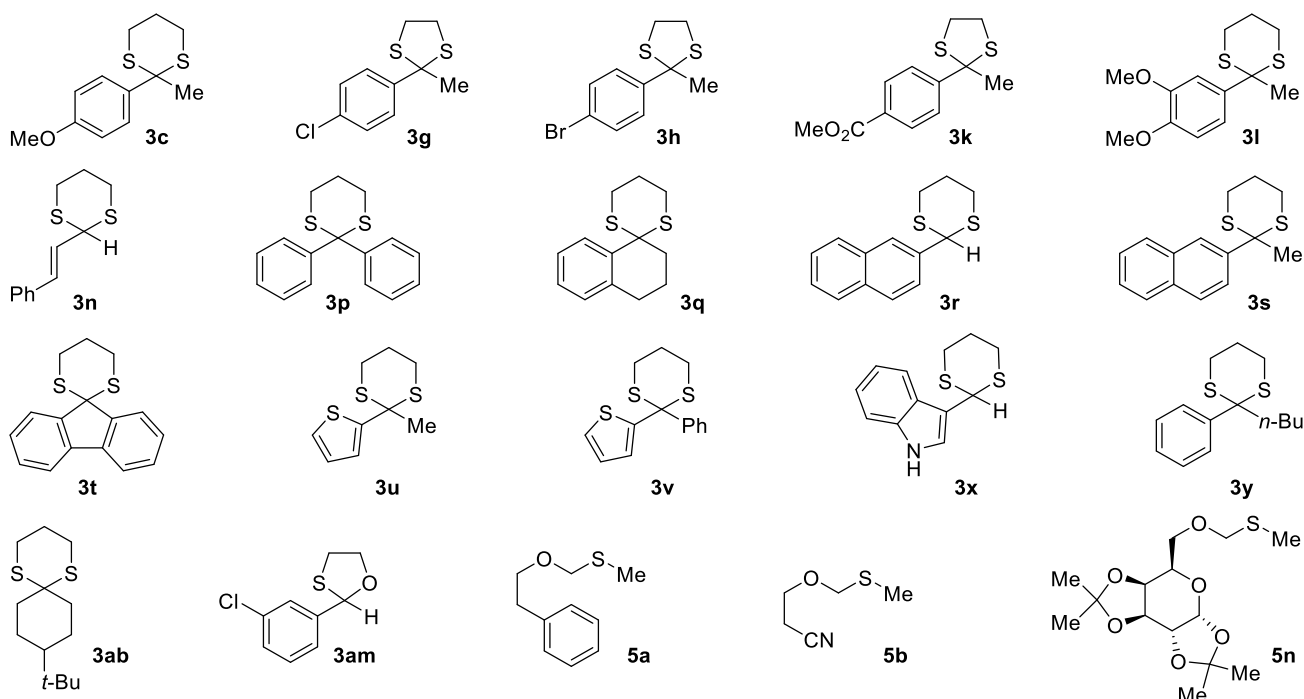
General Procedure: To a stirred solution of compound **3a** (21.0 mg, 0.1 mmol) in solvent (1 mL) were added MBr_x (0.01 mmol) and H_2O_2 (30 wt%, 30 μ L, 0.3 mmol). After completion of the addition, the reaction mixture was allowed to stir at room temperature (rt) for 1.5 h. The reaction was quenched by dilute aqueous $Na_2S_2O_3$ solution (0.1 M, 10 mL) and ethyl acetate (10 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (2×10 mL). The combined organic fractions were washed with H_2O , dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. Yield was determined by 1H -NMR of the crude reaction mixture using CH_2Br_2 as the internal reference.

entry ^a	MBr_x (0.1 eq)	Solvent	Yield ^c (%)
1	CeBr ₃	DCM/H ₂ O (10/1)	<5
2	CeBr ₃	DMSO/H ₂ O (10/1)	<5
3	CeBr ₃	DMF/H ₂ O (10/1)	<5
4	CeBr ₃	THF/H ₂ O (10/1)	13
5	CeBr ₃	<i>n</i> -BuOH/H ₂ O (10/1)	32
6	CeBr ₃	MeCN/H ₂ O (10/1)	70
7	CeBr ₃	EtOH/H ₂ O (10/1)	75
8	CeBr₃	EtOH	77
9	FeBr ₂	EtOH	69
10	FeBr ₃	EtOH	73
11^b	CeBr₃	EtOH	86

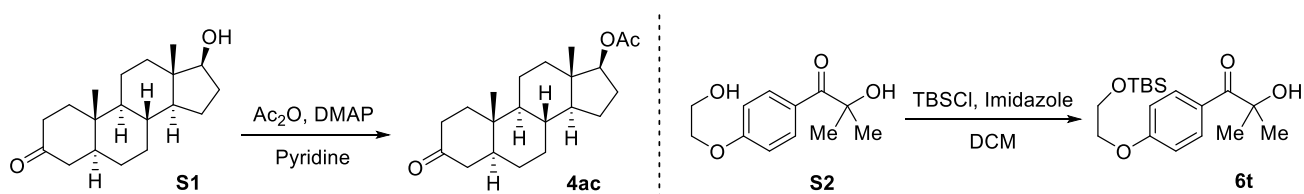
^aReaction was carried out at rt: **3a** (0.1 mmol) was dissolved in solvent (1 mL), then CeBr₃, FeBr₂ or FeBr₃ (0.01 mmol), and H_2O_2 (30%, 30 μ L, 0.3 mmol) were added and stirred at rt for 1.5 h. ^b0.005 mmol CeBr₃ and 0.2 mmol H_2O_2 were added, and the reaction was stirred at 40°C for 15 min. ^cYield was determined by 1H -NMR of the crude reaction mixture using CH_2Br_2 as the internal reference.

4. Preparation of thioacetals, thioketals and methylthiomethyl ether.

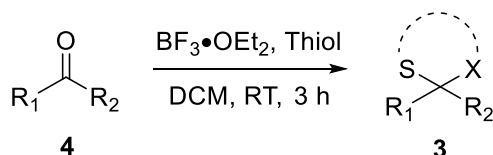
4.1. Substrates **3c**², **3g**³, **3h**⁴, **3k**⁵, **3l**⁶, **3n**⁷, **3p-t**⁸, **3u**⁹, **3v**¹⁰, **3x**¹¹, **3y**¹², **3ab**⁷, **3am**¹³, **5a**¹⁴, **5b**¹⁵ and **5n**¹⁶ were prepared according to the published procedure.



4.2. Synthesis of 4ac and 6t: To a stirred solution of compound **S1** or **S2** (20 mmol) in anhydrous pyridine (40 mL) or dichloromethane (DCM: 100 mL) were added acetic anhydride (Ac_2O : 9.4 mL, 100 mmol) and 4-dimethylaminopyridine (DMAP: 244 mg, 2 mmol) or *tert*-butyldimethylsilyl chloride (TBSCl: 3.01 g, 20 mmol) and imidazole (2.04 g, 30 mmol). After completion of the addition, the reaction mixture was allowed to stir at rt for 1 h. Then the mixture was quenched by cooled aqueous HCl or NaHCO_3 solution and DCM (200 mL). The organic fractions were collected, and the aqueous phase was extracted with DCM (2×100 mL). The combined organic fractions were washed with brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) to give **4ac** (4.72 g, 71%) or **6t** (5.42 g, 80%). The detail spectra data was described in detail below (Part 5).



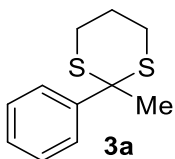
4.3. Others were synthesized according to “**General Procedure I or II**” as below:



General Procedure I: To a stirred solution of compound **4** (8 mmol) in DCM (40 mL) were added thiol (1,3-propanedithiol, 1,2-ethanedithiol, 2-mercaptoethanol, 3-mercaptopropanol, 2-mercaptophenol, 2-aminothiophenol or L-cysteine ethyl ester hydrochloride: 8 mmol; 4-methylbenzenethiol or benzyl

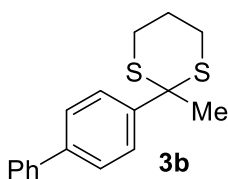
mercaptan: 16 mmol) and $\text{BF}_3 \cdot \text{OEt}_2$ (1.97 mL, 16 mmol) at rt. After completion of the addition, the reaction mixture was allowed to stir at rt for 3 h. Then the mixture was quenched by saturated aqueous NaHCO_3 solution (100 mL) and CH_2Cl_2 (100 mL). The organic fractions were collected, and the aqueous phase was extracted with CH_2Cl_2 (3 \times 50 mL). The combined organic fractions were washed with brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography to give compound **3**.

2-Methyl-2-phenyl-1,3-dithiane (**3a**)



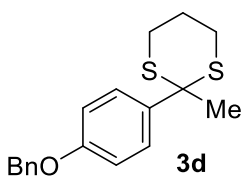
3a¹⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.26 g, 75%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.96–7.91 (m, 2H), 7.38–7.33 (m, 2H), 7.26–7.21 (m, 1H), 2.76–2.64 (m, 4H), 1.95–1.89 (m, 2H), 1.78 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 143.8, 128.5, 127.7, 127.0, 53.9, 32.8, 28.1, 24.6. IR 3055.6, 2907.4, 1593.1, 1485.7, 1440.9, 1368.0, 1275.1, 1179.9, 1062.5, 906.6, 866.5, 760.5 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{11}\text{H}_{15}\text{S}_2$ [M+H]⁺ 211.0610; found 211.0610.

2-Methyl-2-(4-phenylphenyl)-1,3-dithiane (**3b**)



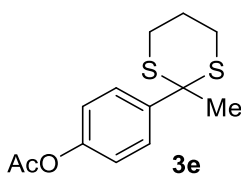
3b was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (1.83 g, 80%). M.P. 135–137 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 8.03–7.99 (m, 2H), 7.65–7.60 (m, 4H), 7.48–7.43 (m, 2H), 7.39–7.34 (m, 1H), 2.81–2.75 (m, 4H), 2.02–1.95 (m, 2H), 1.85 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 143.1, 140.7, 140.0, 128.9, 128.4, 127.5, 127.4, 127.2, 53.9, 32.9, 28.3, 24.8. IR 2878.7, 1591.8, 1478.2, 1390.6, 1275.8, 1175.3, 1069.9, 999.6, 845.7, 761.6 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{17}\text{H}_{19}\text{S}_2$ [M+H]⁺ 287.0923; found 287.0922.

2-Methyl-2-(4-benzyloxyphenyl)-1,3-dithiane (**3d**)



3d was purified by flash column chromatography (dichloromethane/hexane = 1:1 to 2:1) as a white solid (1.77 g, 70%). M.P. 98–100 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 7.87–7.82 (m, 2H), 7.49–7.36 (m, 5H), 7.00–6.95 (m, 2H), 5.08 (s, 2H), 2.81–2.69 (m, 4H), 1.99–1.93 (m, 2H), 1.81 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 158.0, 137.1, 136.2, 129.3, 128.7, 128.1, 127.7, 114.8, 70.2, 53.6, 32.3, 28.3, 24.9. IR 3030.9, 2906.3, 1600.2, 1573.8, 1498.1, 1453.7, 1383.7, 1287.4, 1236.1, 1169.8, 1070.4, 1007.1, 835.9, 736.5 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{18}\text{H}_{21}\text{OS}_2$ [M+H]⁺ 317.1028; found 317.1028.

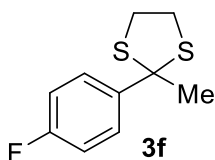
2-Methyl-2-(4-acetoxyphenyl)-1,3-dithiane (**3e**)



3e¹⁸ was purified by flash column chromatography (dichloromethane/hexane = 1:1 to 2:1) as a light-yellow oil (1.70 g, 79%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.96–7.92 (m, 2H), 7.10–7.05 (m, 2H), 2.75–2.69 (m, 4H), 2.30 (s, 3H), 1.99–1.88 (m, 2H), 1.77 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 169.5, 149.7, 141.5, 129.2, 121.6,

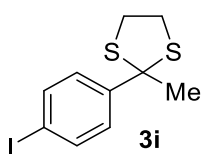
53.7, 33.1, 28.2, 24.7, 21.3. IR 2900.7, 1763.3, 1591.2, 1494.0, 1415.5, 1363.6, 1277.8, 1189.2, 1067.1, 1011.1, 903.9, 846.3 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{13}\text{H}_{17}\text{O}_2\text{S}_2$ [M+H]⁺ 269.0664; found 269.0666.

2-Methyl-2-(4-fluorophenyl)-1,3-dithiolane (3f)



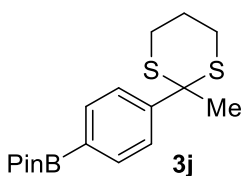
3f was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.37 g, 80%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.76–7.69 (m, 2H), 7.01–6.94 (m, 2H), 3.52–3.43 (m, 2H), 3.42–3.34 (m, 2H), 2.13 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 160.7 (d, $J = 245$ Hz), 141.9 (d, $J = 3.2$ Hz), 128.8 (d, $J = 8$ Hz), 114.7 (d, $J = 21.3$ Hz), 68.1, 40.6, 34.0. IR 2922.4, 1599.3, 1501.2, 1445.3, 1372.8, 1273.8, 1224.2, 1106.3, 1067.1, 1013.2, 833.6 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{12}\text{FS}_2$ [M+H]⁺ 215.0359; found 215.0357.

2-Methyl-2-(4-iodophenyl)-1,3-dithiolane (3i)



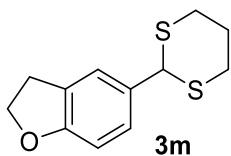
3i was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.86 g, 72%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.65–7.58 (m, 2H), 7.53–7.46 (m, 2H), 3.49–3.39 (m, 2H), 3.38–3.28 (m, 2H), 2.11 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 146.0, 137.0, 129.0, 92.9, 68.1, 40.5, 33.5. IR 2916.4, 1568.7, 1465.3, 1435.9, 1374.9, 1263.5, 1172.4, 1102.9, 1066.4, 817.3 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{12}\text{IS}_2$ [M+H]⁺ 322.9420; found 322.9412.

2-Methyl-2-[4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl]-1,3-dithiane (3j)



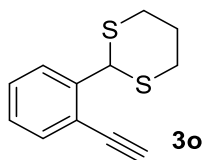
3j was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (1.78 g, 66%). M.P. 153–155 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 7.98–7.93 (m, 2H), 7.84–7.81 (m, 2H), 2.73–2.67 (m, 4H), 1.96–1.90 (m, 2H), 1.76 (s, 3H), 1.35 (s, 12H). ¹³C-NMR (100 MHz, CDCl_3) δ : 147.2, 135.3, 127.3, 84.0, 54.4, 33.0, 28.2, 25.0, 24.7. IR 2972.6, 2906.4, 1600.9, 1391.9, 1356.7, 1324.0, 1265.7, 1209.9, 1138.7, 1066.6, 1013.9, 849.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{17}\text{H}_{26}\text{BO}_2\text{S}_2$ [M+H]⁺ 337.1462; found 337.1468.

2-(2,3-dihydrobenzofuran-5-yl)-1,3-dithiane (3m)



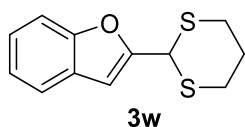
3m was purified by flash column chromatography (dichloromethane/hexane = 1:1 to 2:1) as a white solid (1.33 g, 70%). M.P. 167–169 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 7.32 (s, 1H), 7.18 (dd, $J = 8.2, 2.0$ Hz, 1H), 6.71 (d, $J = 8.2$ Hz, 1H), 5.11 (s, 1H), 4.52 (t, $J = 8.7$ Hz, 2H), 3.15 (t, $J = 8.7$ Hz, 2H), 3.08–2.99 (m, 2H), 2.91–2.84 (m, 2H), 2.18–2.11 (m, 1H), 1.95–1.83 (m, 1H). ¹³C-NMR (100 MHz, CDCl_3) δ : 160.3, 131.3, 127.8, 127.6, 124.4, 109.3, 71.5, 51.1, 32.3, 29.6, 25.1. IR 2926.3, 2891.1, 1602.5, 1483.4, 1417.2, 1319.8, 1275.6, 1237.0, 1174.2, 1099.0, 898.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{12}\text{H}_{15}\text{OS}_2$ [M+H]⁺ 239.0559; found 239.0550.

2-(2-ethynylphenyl)-1,3-dithiane (3o)



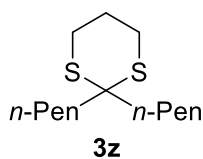
THF (40 mL) was used as the reaction solvent instead of DCM, and **3o** was purified by flash column chromatography (dichloromethane/hexane = 1:1 to 2:1) as a white solid (1.15 g, 65%). M.P. 102–104 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 7.65 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.47 (dd, *J* = 7.7, 1.4 Hz, 1H), 7.34 (td, *J* = 7.6, 1.4 Hz, 1H), 7.22 (td, *J* = 7.6, 1.4 Hz, 1H), 5.73 (s, 1H), 3.37 (s, 1H), 3.14–3.05 (m, 2H), 2.87 (dt, *J* = 14.4, 4.2 Hz, 2H), 2.20–2.10 (m, 1H), 1.99–1.86 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 141.0, 133.1, 129.6, 128.14, 128.09, 120.7, 82.2, 80.9, 48.9, 32.3, 25.2. IR 3236.8, 3048.4, 2945.9, 2891.1, 2103.2, 1475.4, 1443.1, 1271.2, 1243.2, 1191.3, 756.9 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₁₃S₂ [M+H]⁺ 221.0453; found 221.0452.

2-(benzofuran-2-yl)-1,3-dithiane (**3w**)



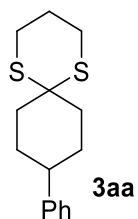
3w was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.40 g, 74%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.50 (d, *J* = 7.5 Hz, 1H), 7.45 (d, *J* = 8.1 Hz, 1H), 7.28–7.22 (m, 1H), 7.22–7.16 (m, 1H), 6.77 (s, 1H), 5.26 (s, 1H), 3.02–2.94 (m, 2H), 2.93–2.85 (m, 2H), 2.14–2.04 (m, 1H), 2.04–1.92 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 154.7, 154.6, 128.0, 124.5, 122.9, 121.0, 111.3, 104.7, 41.9, 29.8, 25.2. IR 3087.7, 2901.8, 1583.8, 1446.1, 1419.2, 1341.4, 1273.4, 1244.7, 1199.0, 1159.7, 1124.6, 1005.2, 748.4 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₁₃OS₂ [M+H]⁺ 237.0402; found 237.0399.

2,2-Dipentyl-1,3-dithiane (**3z**)



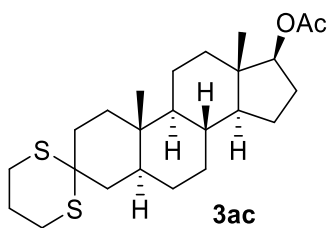
3z¹⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.50 g, 72%). ¹H-NMR (400 MHz, CDCl₃) δ: 2.77–2.74 (m, 4H), 1.92–1.88 (m, 2H), 1.82–1.79 (m, 4H), 1.40–1.35 (m, 4H), 1.32–1.27 (m, 4H), 1.27–1.22 (m, 4H), 0.85 (t, *J* = 4.0 Hz, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 52.4, 37.1, 31.0, 25.0, 24.7, 22.7, 21.6, 13.1. IR 2930.2, 2860.7, 1458.2, 1421.8, 1376.4, 1272.9, 1239.0, 1120.4, 907.5, 734.5 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₄H₂₉S₂ [M+H]⁺ 261.1705; found 261.1704.

9-Phenyl-1,5-dithiaspiro[5.5]undecane (**3aa**)



3aa²⁰ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (1.44 g, 68%). M.P. 110–112 °C; ¹H-NMR (400 MHz, CDCl₃) δ: 7.31–7.26 (m, 2H), 7.25–7.21 (m, 2H), 7.21–7.16 (m, 1H), 2.93–2.89 (m, 2H), 2.80–2.75 (m, 2H), 2.50 (tt, *J* = 12.1, 3.7 Hz, 1H), 2.50–2.44 (m, 2H), 2.05–1.94 (m, 4H), 1.87–1.78 (m, 2H), 1.76–1.69 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 146.7, 128.5, 127.0, 126.3, 49.8, 44.3, 38.2, 29.7, 26.4, 25.9. IR 3018.4, 2918.7, 1596.0, 1487.4, 1417.4, 1268.8, 1232.2, 1099.8, 1066.0, 993.0, 831.9, 755.2 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₅H₂₁S₂ [M+H]⁺ 265.1079; found 265.1076.

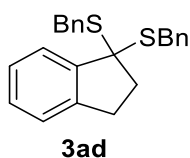
(5S)-17-Acetyl-4,5-dihydrotestosterone Cyclic 1,3-Propanediyl Dithioacetal (**3ac**)



3ac was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (2.03 g, 60%). M.P. 213–215 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 4.57 (t, *J* = 8.5 Hz, 1H), 2.90–2.84 (m, 2H), 2.76–2.71 (m, 2H), 2.23–2.08 (m, 2H), 2.02 (s, 3H), 2.00–1.93 (m, 2H), 1.92–1.78 (m, 2H), 1.73–1.53 (m, 6H), 1.52–1.38 (m, 4H), 1.33–1.19 (m, 4H), 1.18–0.88 (m, 4H), 0.80 (s, 3H), 0.76 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 171.3, 83.0,

54.1, 50.9, 50.6, 42.8, 41.5, 40.6, 37.0, 36.6, 35.4, 34.6, 34.0, 31.5, 28.1, 27.7, 26.41, 26.35, 26.1, 23.6, 21.3, 20.6, 12.2, 12.0. IR 2920.5, 2851.3, 1722.8, 1439.0, 1378.6, 1246.6, 1028.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₄H₃₉O₂S₂ [M+H]⁺ 423.2386; found 423.2388. [α]_D²⁵ = +21 (*c* 1, CHCl₃).

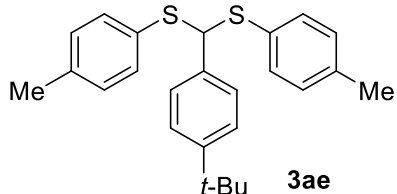
1-(Bisbenzylthio)indane (**3ad**)



InCl₃ (177 mg, 0.8 mmol) was used as the reaction solvent instead of BF₃•OEt₂, and **3ad** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.45 g, 50%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.28–7.16 (m, 14H), 3.89 (d, *J* = 12.0 Hz, 2H), 3.71 (d, *J* = 12.0 Hz, 2H), 2.96 (t, *J* = 6.9 Hz, 2H), 2.44 (t, *J* = 6.9

Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 144.0, 142.8, 137.4, 129.3, 128.6, 128.4, 127.1, 126.5, 125.5, 123.9, 67.7, 43.1, 36.0, 30.6. IR 2936.2, 2879.2, 1616.9, 1407.2, 1321.9, 1162.9, 1117.5, 1013.0, 758.9, 710.6 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₃H₂₃S₂ [M+H]⁺ 363.1236; found 363.1223.

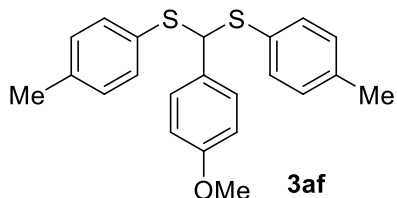
1-[Bis(4-tolylthio)methyl]-4-*tert*-butylbenzene (**3ae**)



3ae was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (2.39 g, 76%). M.P. 84–86 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 7.28–7.20 (m, 8H), 7.03 (d, *J* = 7.8 Hz, 4H), 5.31 (s, 1H), 2.30 (s, 6H), 1.29 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 151.0, 138.0, 137.0, 133.1, 131.4, 129.7, 127.6, 125.5,

61.3, 34.7, 31.5, 21.3. IR 2959.1, 2864.1, 1600.8, 1487.9, 1401.0, 1360.7, 1265.9, 1213.2, 1084.3, 1015.4, 800.4 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₅H₂₈NaS₂ [M+Na]⁺ 415.1525; found 415.1534.

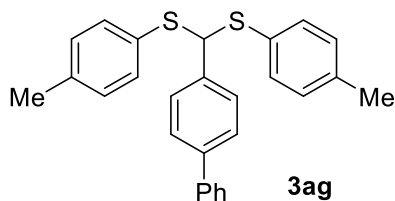
1-[Bis(4-tolylthio)methyl]-4-methoxybenzene (**3af**)



3af²¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (2.11 g, 72%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.31–7.24 (m, 6H), 7.05 (d, *J* = 7.9 Hz, 4H), 6.78–6.83 (m, 2H), 5.33 (s, 1H), 3.80 (s, 3H), 2.32 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 159.3, 138.0, 133.1, 132.1, 131.2,

129.7, 129.2, 113.8, 60.7, 55.4, 21.3. IR 2919.6, 1605.7, 1501.7, 1452.4, 1301.2, 1247.4, 1098.2, 1028.6, 835.9 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₂H₂₂NaOS₂ [M+Na]⁺ 389.1004; found 389.0998.

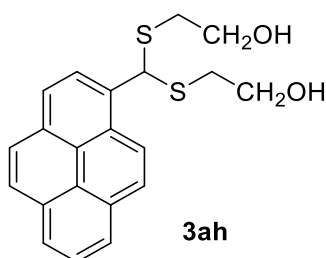
1-[Bis(4-tolylthio)methyl]-4-phenylbenzene (**3ag**)



3ag was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (2.48 g, 75%). M.P. 113–115 °C. ¹H-NMR (400 MHz, DMSO) δ: 7.67–7.58 (m, 4H), 7.56–7.51 (m, 2H), 7.43 (t, *J* = 7.4 Hz, 2H), 7.38–7.33 (m, 1H), 7.29 (d, *J* = 8.0 Hz, 4H), 7.10 (d, *J* = 8.0 Hz, 4H), 6.00 (s, 1H), 2.25 (s, 6H).

¹³C-NMR (100 MHz, DMSO) δ: 139.6, 139.4, 138.5, 137.2, 131.5, 130.5, 129.6, 128.9, 128.4, 127.6, 126.6, 57.1, 20.6. IR 3023.1, 2918.1, 1599.7, 1482.0, 1444.8, 1399.8, 1176.5, 1106.3, 1008.9, 838.2, 758.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₇H₂₄NaS₂ [M+Na]⁺ 435.1212; found 435.1208.

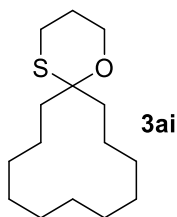
1-[Bis(2-Hydroxyethyl)thio]pyrene (**3ah**)



TsOH (800 mg, 4.64 mmol) was used instead of BF₃•OEt₂, and the mixture was refluxed with 2-mercaptoethanol (1.69 mL, 24 mmol) in EtOH for 24 h under nitrogen atmosphere. **3ah**²² was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 2:1) as a yellow solid (2.21 g, 75%). M.P. 132–134 °C. ¹H-NMR (400 MHz, DMSO) δ: 8.65 (d, *J* = 9.4 Hz, 1H), 8.40–8.24 (m, 5H), 8.21–8.14 (m, 2H), 8.06 (t, *J* = 7.6 Hz, 1H), 6.43 (s, 1H), 4.80 (t, *J* = 5.4 Hz, 2H), 3.60–3.49 (m, 4H), 2.85–2.76 (m, 2H), 2.70–

2.62 (m, 2H). ¹³C-NMR (100 MHz, DMSO) δ: 134.4, 130.8, 130.3, 130.2, 127.54, 127.46, 127.3, 126.4, 126.0, 125.5, 125.3, 124.9, 124.1, 123.9, 60.8, 35.0. IR 3221.3, 2912.0, 1592.9, 1394.2, 1284.6, 1231.6, 1161.3, 1004.6, 823.2 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₁H₂₀NaO₂S₂ [M+Na]⁺ 391.0797; found 391.0789.

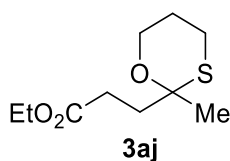
1-Oxa-5-thiaspiro[5.11]heptadecane (**3ai**)



THF (40 mL) was used as the reaction solvent instead of DCM, and **3ai** was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (1.23 g, 60%). M.P. 59–61 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 3.81 (t, *J* = 5.4 Hz, 2H), 2.86–2.82 (m, 2H), 2.17–2.08 (m, 2H), 1.83–1.68 (m, 4H), 1.34 (m, 18H). ¹³C-NMR (100 MHz, CDCl₃) δ: 85.5, 61.9, 33.3, 26.3, 26.2, 25.7, 24.1, 22.5, 22.1, 19.3. IR 2909.4, 2853.6, 1468.0, 1439.6, 1211.9, 1074.0, 1021.3, 722.1 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd.

for C₁₅H₂₉OS [M+H]⁺ 257.1934; found 257.1929.

2-[2-(Ethoxycarbonyl)ethyl]-2-methyl-1,3-oxathiane (**3aj**)

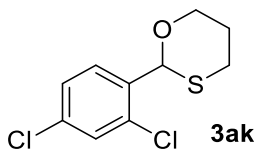


THF (40 mL) was used as the reaction solvent instead of DCM, and **3aj** was purified by flash column chromatography (ethyl acetate/petroleum ether = 1:20 to 1:5) as a light-yellow oil (1.36 g, 78%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.03 (q, *J* = 7.1 Hz, 2H), 3.83–3.72 (m, 2H), 2.82–2.76 (m, 2H), 2.46–2.32 (m, 2H), 2.23–2.03 (m, 2H),

1.74–1.67 (m, 2H), 1.48 (s, 3H), 1.17 (t, *J* = 7.2 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 173.5, 80.7, 61.9,

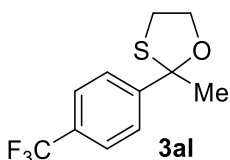
60.4, 35.4, 29.1, 26.1, 25.1, 24.3, 14.2. IR 2939.4, 2873.1, 1729.8, 1438.2, 1373.7, 1171.7, 1094.4, 1015.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{19}\text{O}_3\text{S}$ [M+H]⁺ 219.1049; found 219.1039.

2-(2,4-Dichlorophenyl)-1,3-oxathiane (3ak)



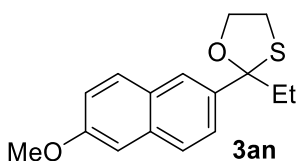
THF (40 mL) was used as the reaction solvent instead of DCM, and **3ak** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (1.46 g, 73%). M.P. 60–62 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 7.62 (d, $J = 8.4$ Hz, 1H), 7.34 (d, $J = 2.1$ Hz, 1H), 7.27 (dd, $J = 8.4, 2.2$ Hz, 1H), 6.03 (s, 1H), 4.36–4.29 (m, 1H), 3.79 (td, $J = 12.5, 2.2$ Hz, 1H), 3.21 (td, $J = 13.2, 2.8$ Hz, 1H), 2.86–2.79 (m, 1H), 2.18–2.05 (m, 1H), 1.82–1.74 (m, 1H). ¹³C-NMR (100 MHz, CDCl_3) δ : 135.9, 134.8, 132.0, 129.7, 129.2, 127.7, 80.7, 70.9, 29.3, 25.7. IR 2912.8, 2865.5, 1585.1, 1465.8, 1376.8, 1342.9, 1227.9, 1072.8, 852.5, 752.1 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{11}\text{Cl}_2\text{OS}$ [M+H]⁺ 248.9902; found 248.9896.

2-Methyl-2-(4-trifluoromethylphenyl)-1,3-oxathiolane (3al)



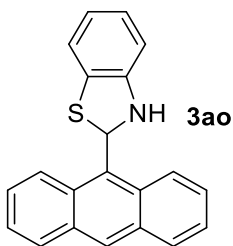
THF (40 mL) was used as the reaction solvent instead of DCM, and **3al** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.41 g, 71%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.63 (d, $J = 4.7$ Hz, 2H), 7.59 (d, $J = 5.1$ Hz, 2H), 4.38–4.34 (m, 1H), 4.04–4.00 (m, 1H), 3.25–3.21 (m, 1H), 3.09–3.05 (m, 1H), 1.92 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 151.1, 129.2 (q, $J = 18.4$ Hz), 125.4, 125.2 (q, $J = 2.1$ Hz), 121.9 (q, $J = 155$ Hz), 95.0, 71.0, 34.6, 32.2. IR 2980.1, 2879.2, 1616.9, 1407.2, 1321.9, 1162.8, 1117.6, 1067.9, 1013.0, 840.4 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{11}\text{H}_{12}\text{F}_3\text{OS}$ [M+H]⁺ 249.0555; found 249.0567.

2-Ethyl-2-(6-methoxynaphthalen-2-yl)-1,3-oxathiolane (3an)



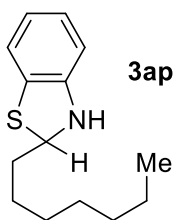
THF (40 mL) was used as the reaction solvent instead of DCM, and **3an** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.54 g, 70%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.93 (d, $J = 1.2$ Hz, 1H), 7.78 (d, $J = 5.2$ Hz, 1H), 7.74 (d, $J = 4.9$ Hz, 1H), 7.54 (dd, $J = 4.9, 1.1$ Hz, 1H), 7.20 (dd, $J = 5.1, 1.5$ Hz, 1H), 7.16 (d, $J = 1.6$ Hz, 1H), 4.42–4.39 (m, 1H), 4.04–4.00 (m, 1H), 3.92 (s, 3H), 3.21–3.16 (m, 1H), 3.08–3.04 (m, 1H), 2.26 (q, $J = 4.1$ Hz, 2H), 0.97 (t, $J = 4.2$ Hz, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 157.8, 140.4, 133.7, 129.7, 128.3, 126.8, 124.8, 123.8, 118.9, 105.5, 100.0, 70.7, 55.2, 37.4, 34.0, 9.8. IR 3055.1, 2927.4, 2871.1, 1599.1, 1473.2, 1378.9, 1335.2, 1220.3, 1122.3, 1024.7, 884.9 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{16}\text{H}_{19}\text{O}_2\text{S}$ [M+H]⁺ 275.1100; found 275.1096.

2-(Anthracen-9-yl)benzothiazoline (3ao)



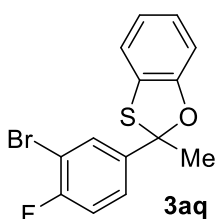
EtOH (40 mL) was used as the reaction solvent instead of DCM, and no $\text{BF}_3 \cdot \text{OEt}_2$ was needed. **3ao** was purified by flash column chromatography (dichloromethane/hexane = 1:5 to 1:2) as a yellow solid (1.63 g, 65%). M.P. 126–128 °C. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 8.75 (d, $J = 8.1$ Hz, 2H), 8.48 (s, 1H), 8.12 (s, 1H), 8.05–7.99 (m, 2H), 7.53–7.45 (m, 4H), 7.18 (dd, $J = 7.6, 1.2$ Hz, 1H), 7.01 (td, $J = 7.6, 1.3$ Hz, 1H), 6.85 (td, $J = 7.6, 1.2$ Hz, 1H), 6.67 (dd, $J = 7.8, 1.2$ Hz, 1H), 4.42 (s, 1H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 146.7, 131.7, 130.7, 130.1, 129.6, 128.0, 127.9, 126.2, 125.9, 125.1, 124.4, 122.1, 121.0, 110.5, 65.7. IR 3337.9, 3040.1, 1576.7, 1447.7, 1366.9, 1072.6, 1015.9, 844.1, 731.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{21}\text{H}_{16}\text{NS}$ $[\text{M}+\text{H}]^+$ 314.0998; found 314.1012.

2-(*n*-Heptyl)benzothiazoline (**3ap**)



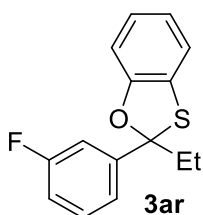
EtOH (40 mL) was used as the reaction solvent instead of DCM, and no $\text{BF}_3 \cdot \text{OEt}_2$ was needed. **3ap**²³ was purified by flash column chromatography (dichloromethane/hexane = 1:5 to 1:2) as a light-yellow oil (1.32 g, 70%). $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 7.06 (dd, $J = 7.6, 1.3$ Hz, 1H), 6.89 (td, $J = 7.6, 1.4$ Hz, 1H), 6.72 (td, $J = 7.5, 1.2$ Hz, 1H), 6.63 (dd, $J = 7.8, 1.2$ Hz, 1H), 5.25 (t, $J = 6.6$ Hz, 1H), 3.99 (s, 1H), 1.91–1.85 (m, 2H), 1.43–1.29 (m, 10H), 0.89 (t, $J = 7.0$ Hz, 3H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 146.7, 127.6, 125.2, 122.0, 120.8, 110.9, 69.0, 38.7, 31.8, 29.33, 29.25, 26.2, 22.7, 14.2. IR 3298.3, 2920.6, 2850.9, 1583.0, 1462.7, 1400.6, 1366.5, 1211.3, 1118.0, 1019.5, 736.5 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{14}\text{H}_{22}\text{NS}$ $[\text{M}+\text{H}]^+$ 236.1467; found 236.1472.

2-(3-Bromo-4-fluorophenyl)-2-methyl-1,3-benzoxathiole (**3aq**)



CHCl_3 (40 mL) was used as the reaction solvent instead of DCM, and the mixture was refluxed under nitrogen for 3 h. **3aq** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.56 g, 60%). $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 7.81 (dd, $J = 6.4, 2.4$ Hz, 1H), 7.49 (ddd, $J = 8.6, 4.5, 2.4$ Hz, 1H), 7.13–7.04 (m, 3H), 6.98–6.89 (m, 2H), 2.14 (s, 3H). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 157.5 (d, $J = 247$ Hz), 154.6, 142.2 (d, $J = 3.7$ Hz), 130.4, 126.3, 125.9, 125.8 (d, $J = 7.5$ Hz), 122.9, 122.2, 116.2 (d, $J = 22.5$ Hz), 111.0, 109.0 (d, $J = 21.1$ Hz), 98.3, 31.1. IR 1579.4, 1491.9, 1456.9, 1384.7, 1228.9, 1083.4, 1045.5, 1017.0, 852.7, 739.7 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{14}\text{H}_{11}\text{BrFOS}$ $[\text{M}+\text{H}]^+$ 324.9693; found 324.9690.

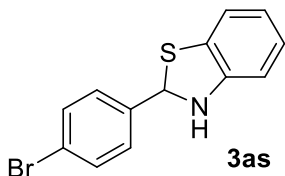
2-Ethyl-2-(3-fluorophenyl)-1,3-benzoxathiole (**3ar**)



CHCl_3 (40 mL) was used as the reaction solvent instead of DCM, and the mixture was refluxed under nitrogen for 3 h. **3ar** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.35 g, 65%). $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 7.30–7.19 (m, 3H), 7.04 (dd, $J = 7.6, 1.4$ Hz, 1H), 7.02–6.90 (m, 3H), 6.82 (td, $J = 7.5, 1.3$ Hz, 1H), 2.44–2.24 (m, 2H), 0.96 (t, $J = 7.3$ Hz, 3H). $^{13}\text{C-NMR}$

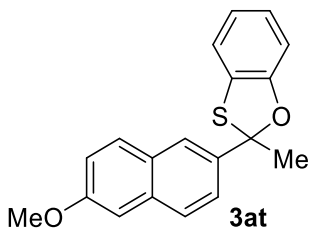
(100 MHz, CDCl₃) δ : 161.5 (d, J = 245 Hz), 155.2, 146.7 (d, J = 6.6 Hz), 129.9 (d, J = 8.1 Hz), 126.0, 125.8, 122.6, 122.1, 120.8 (d, J = 2.9 Hz), 114.8 (d, J = 21.1 Hz), 112.4 (d, J = 23.2 Hz), 110.6, 102.7 (d, J = 1.9 Hz), 36.7, 9.3. IR 2935.2, 1585.8, 1458.3, 1229.1, 1119.9, 862.9, 781.6, 739.4 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₃H₁₄FOS [M+H]⁺ 261.0744; found 261.0757.

2-(4-Bromophenyl)benzothiazoline (3as)



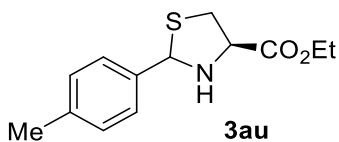
EtOH (40 mL) was used as the reaction solvent instead of DCM, and no BF₃•OEt₂ was needed. **3as** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a yellow solid (1.64 g, 70%). M.P. 81–83 °C. ¹H-NMR (400 MHz, CDCl₃) δ : 7.50–7.46 (m, 2H), 7.42–7.38 (m, 2H), 7.04 (dd, J = 7.6, 1.3 Hz, 1H), 6.96 (td, J = 7.7, 1.3 Hz, 1H), 6.77 (td, J = 7.5, 1.2 Hz, 1H), 6.65 (dd, J = 7.7, 1.2 Hz, 1H), 6.30 (s, 1H), 4.03 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ : 146.1, 140.9, 131.9, 128.3, 126.3, 125.7, 122.7, 121.7, 120.9, 110.0, 69.3. IR 3342.5, 1576.4, 1461.9, 1391.1, 1250.2, 1064.1, 826.1, 746.6 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₃H₁₁BrNS [M+H]⁺ 291.9790; found 291.9789.

2-Methyl-2-(6-methoxynaphthalen-2-yl)-1,3-benzoxathiole (3at)



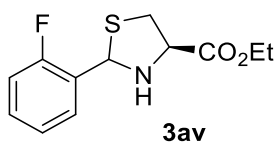
THF (40 mL) was used as the reaction solvent instead of DCM, and **3at** was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a white solid (1.83 g, 74%). M.P. 122–124 °C. ¹H-NMR (400 MHz, CDCl₃) δ : 7.99 (d, J = 2.0 Hz, 1H), 7.76 (d, J = 8.7 Hz, 2H), 7.68 (dd, J = 8.6, 2.0 Hz, 1H), 7.18 (dd, J = 8.9, 2.5 Hz, 1H), 7.16–7.13 (m, 2H), 7.11–7.06 (m, 1H), 7.02 (dd, J = 8.0, 1.4 Hz, 1H), 6.90 (td, J = 7.4, 1.4 Hz, 1H), 3.93 (s, 3H), 2.29 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ : 158.3, 155.1, 139.3, 134.3, 130.0, 128.2, 127.3, 126.6, 126.0, 124.0, 123.3, 122.5, 122.1, 119.3, 110.9, 105.8, 99.8, 55.4, 31.0. IR 2963.1, 1600.9, 1454.0, 1368.9, 1268.9, 1232.1, 1198.7, 1089.4, 1027.8, 841.8, 742.2 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₉H₁₇O₂S [M+H]⁺ 309.0944; found 309.0941.

(2*RS*,4*R*) Ethyl-2-(4-methylphenyl)thiazolidine-4-carboxylate (3au)



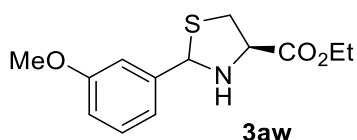
EtOH (40 mL) was used as the reaction solvent instead of DCM, and Et₃N (1.11 mL, 8 mmol) was added. **3au**²⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 1:2) as a light-yellow oil (1.51 g, 75%). ¹H-NMR (400 MHz, CDCl₃) δ : 7.44–7.35 (m, 2H), 7.19–7.11 (m, 2H), 5.81–5.52 (m, 1H), 4.30–4.21 (m, 2H), 4.21–3.93 (m, 1H), 3.48–3.35 (m, 1H), 3.22–3.07 (m, 1H), 2.69 (s, 1H), 2.36–2.32 (m, 3H), 1.33–1.27 (m, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ : 171.8, 171.2, 138.6, 138.3, 137.6, 135.2, 129.4, 129.1, 127.4, 126.9, 72.6, 70.8, 65.7, 64.4, 61.7, 61.6, 39.3, 38.2, 21.2, 21.1, 14.2. IR 3312.5, 2926.5, 1734.2, 1512.8, 1449.1, 1374.1, 1266.3, 1190.4, 1094.2, 1025.5, 763.0 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₃H₁₈NO₂S [M+H]⁺ 252.1053; found 252.1061.

(2*RS*,4*R*) Ethyl-2-(2-fluorophenyl)thiazolidine-4-carboxylate (3av)

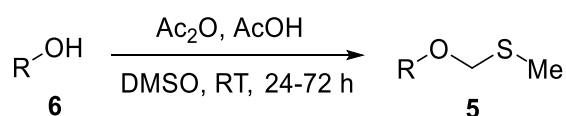


EtOH (40 mL) was used as the reaction solvent instead of DCM, and Et₃N (1.11 mL, 8 mmol) was added. **3av** was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 1:2) as a light-yellow oil (0.92 g, 45%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.64–7.49 (m, 1H), 7.35–7.21 (m, 1H), 7.19–7.00 (m, 2H), 6.04–5.78 (m, 1H), 4.30–4.24 (m, 2H), 4.24–3.93 (m, 1H), 3.50–3.37 (m, 1H), 3.17–3.08 (m, 1H), 2.82 (s, 1H), 1.34–1.29 (m, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 171.6, 171.1, 159.5 (d, *J* = 247 Hz), 159.2 (d, *J* = 246 Hz), 130.3 (d, *J* = 8.5 Hz), 129.4 (d, *J* = 12.3 Hz), 129.3 (d, *J* = 8.1 Hz), 128.6 (d, *J* = 3.3 Hz), 127.2 (d, *J* = 3.6 Hz), 125.3 (d, *J* = 12.5 Hz), 124.6 (d, *J* = 3.5 Hz), 124.1 (d, *J* = 3.5 Hz), 115.8 (d, *J* = 21.7 Hz), 115.5 (d, *J* = 21.1 Hz), 65.8, 65.7 (d, *J* = 3.7 Hz), 64.8, 64.3 (d, *J* = 2.2 Hz), 61.82, 61.79, 39.2, 38.0, 14.2. IR 3319.3, 2981.4, 1732.5, 1585.7, 1487.0, 1454.9, 1374.6, 1265.0, 1192.1, 1090.0, 1025.9, 753.6 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₁₅FNO₂S [M+H]⁺ 256.0802; found 256.0811.

(2*RS*,4*R*) Ethyl-2-(3-methoxyphenyl)thiazolidine-4-carboxylate (3aw)

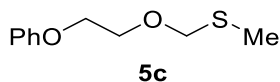


EtOH (40 mL) was used as the reaction solvent instead of DCM, and Et₃N (1.11 mL, 8 mmol) was added. **3aw** was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 1:2) as a light-yellow oil (1.07 g, 50%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.30–7.21 (m, 1H), 7.13–7.04 (m, 2H), 6.89–6.78 (m, 1H), 5.82–5.52 (m, 1H), 4.29–4.21 (m, 2H), 4.19–3.93 (m, 1H), 3.82–3.79 (m, 3H), 3.48–3.35 (m, 1H), 3.19–3.07 (m, 1H), 2.67 (s, 1H), 1.34–1.28 (m, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 171.8, 171.2, 159.9, 159.7, 143.2, 139.8, 129.8, 129.5, 119.8, 119.3, 114.4, 113.4, 113.0, 112.5, 72.6, 70.8, 65.8, 64.5, 61.8, 61.7, 55.4, 55.3, 39.3, 38.3, 14.2. IR 3313.8, 2934.5, 1733.3, 1595.0, 1458.4, 1373.2, 1255.3, 1154.5, 1039.1, 758.1 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₃H₁₈NO₃S [M+H]⁺ 268.1002; found 268.1012.



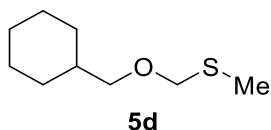
General Procedure II: To a solution of compound **6** (6.5 mmol) in dimethyl sulfoxide (20 mL) was added a mixture of acetic anhydride (14 mL) and acetic acid (2.5 mL) at rt. After completion of the addition, the mixture was allowed to stand for 24–72 h (primary alcohol: 24 h; secondary alcohol: 48 h; tertiary alcohol: 72 h). Then the mixture was quenched by saturated aqueous NaHCO₃ solution (200 mL) and ethyl acetate (100 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (3 × 50 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography to give compound **5**.

Methylthiomethyl 2-phenoxyethyl ether (5c)



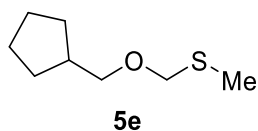
5c was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.97 g, 75%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.28–7.23 (m, 2H), 6.95–6.90 (m, 3H), 4.72 (s, 2H), 4.12 (t, *J* = 4.6 Hz, 2H), 3.88 (t, *J* = 4.8 Hz, 2H), 2.15 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 158.8, 129.5, 121.0, 114.7, 75.6, 67.1, 66.3, 13.9. IR 2922.2, 1742.0, 1594.6, 1493.0, 1430.0, 1369.6, 1299.1, 1241.8, 1212.4, 1174.3, 1085.1, 1022.9, 959.4, 922.2, 840.6, 752.9, 607.8, 509.7, 464.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₀H₁₅O₂S [M+H]⁺ 199.0787; found 199.0786.

Cyclohexylmethyl methylthiomethyl ether (**5d**)



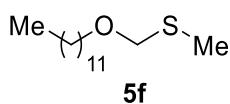
5d was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.76 g, 67%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.61 (s, 2H), 3.30 (d, *J* = 7.8 Hz, 2H), 2.13 (s, 3H), 1.77–1.67 (m, 5H), 1.60–1.54 (m, 1H), 1.26–1.16 (m, 3H), 0.99–0.89 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 75.4, 74.0, 37.9, 30.2, 26.7, 26.0, 13.9. IR 2919.9, 2850.1, 1445.0, 1299.1, 1260.0, 1068.3, 956.6, 882.2, 728.9, 678.9 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₉H₁₉OS [M+H]⁺ 175.1151; found 175.1154.

Cyclopentylmethyl methylthiomethyl ether (**5e**)



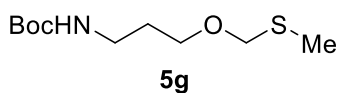
5e was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.89 g, 85%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.60 (s, 2H), 3.36 (d, *J* = 7.0 Hz, 2H), 2.14–2.08 (m, 4H), 1.76–1.68 (m, 2H), 1.60–1.47 (m, 4H), 1.24–1.19 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 75.4, 72.8, 39.3, 29.8, 25.6, 13.9. IR 2948.4, 2863.9, 1434.8, 1299.5, 1256.9, 1070.2, 960.0, 923.7, 728.7, 678.7 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₈H₂₀NOS [M+NH₄]⁺ 178.1260; found 178.1253.

Methylthiomethyl *n*-undecanyl ether (**5f**)



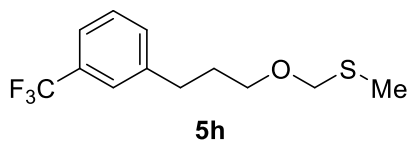
5f was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.35 g, 84%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.63 (s, 2H), 3.52 (t, *J* = 6.5 Hz, 2H), 2.15 (s, 3H), 1.62–1.55 (m, 2H), 1.32–1.26 (m, 18H), 0.89 (t, *J* = 6.5 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 75.3, 68.3, 32.0, 29.78, 29.75, 29.72, 29.56, 29.52, 29.47, 26.4, 22.8, 14.2, 14.0. IR 2921.6, 2853.3, 1461.8, 1381.1, 1299.4, 1081.6, 952.9, 726.3, 679.7 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₄H₃₁OS [M+H]⁺ 247.2090; found 247.2096.

3-[(*tert*-Butyloxycarbonyl)amino]propanyl methylthiomethyl ether (**5g**)



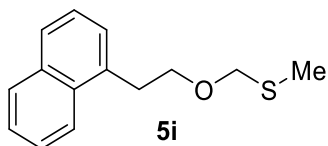
5g was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (1.16 g, 76%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.81 (s, 1H), 4.59 (s, 2H), 3.54 (t, *J* = 6.0 Hz, 2H), 3.23–3.18 (m, 2H), 2.12 (s, 3H), 1.78–1.72 (m, 2H), 1.41 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 156.1, 79.1, 75.5, 66.3, 38.5, 29.6, 28.5, 14.1. IR 3357.9, 2925.7, 1693.2, 1511.7, 1447.5, 1391.5, 1364.8, 1246.3, 1166.8, 1072.9, 859.7, 779.1, 728.6, 678.5 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₀H₂₂NO₃S [M+H]⁺ 236.1315; found 236.1313.

3-(3-Trifluoromethylphenyl)propanyl methylthiomethyl ether (**5h**)



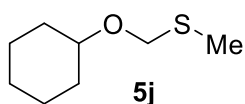
5h was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.53 g, 89%) as colorless oil. ¹H-NMR (400 MHz, CDCl₃) δ: 7.46–7.38 (m, 4H), 4.64 (s, 2H), 3.53 (t, *J* = 6.1 Hz, 2H), 2.76 (t, *J* = 7.7 Hz, 2H), 2.17 (s, 3H), 1.90 (p, *J* = 6.7 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 142.9, 132.0, 130.3 (q, *J* = 32.0 Hz), 128.9, 125.3 (q, *J* = 4.0 Hz), 122.8 (q, *J* = 4.0 Hz), 120.3 (q, *J* = 270.0 Hz), 75.5, 67.2, 32.4, 31.0, 14.2. IR 2924.4, 1744.4, 1442.0, 1326.4, 1210.6, 1162.4, 1121.3, 1068.2, 1017.9, 959.7, 907.4, 800.3, 732.7, 701.1, 458.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₁₆F₃OS [M+H]⁺ 265.0868; found 265.0876.

2-(Naphthalen-1-yl)ethyl methylthiomethyl ether (**5i**)



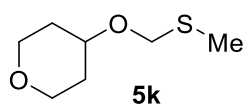
5i was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.22 g, 81%). ¹H-NMR (400 MHz, CDCl₃) δ: 8.09 (d, *J* = 8.3 Hz, 1H), 7.87 (d, *J* = 8.0 Hz, 1H), 7.75 (d, *J* = 9.4 Hz, 1H), 7.57–7.49 (m, 2H), 7.46–7.41 (m, 2H), 4.67 (s, 2H), 3.90 (t, *J* = 7.2 Hz, 2H), 3.40 (t, *J* = 7.2 Hz, 2H), 2.11 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 134.8, 133.9, 132.2, 128.9, 127.2, 126.9, 126.1, 125.61, 125.59, 123.8, 75.4, 68.3, 33.1, 14.0. IR 3046.2, 2916.7, 1727.2, 1594.8, 1510.1, 1431.4, 1392.6, 1299.2, 1258.9, 1163.6, 1062.2, 955.8, 862.1, 775.0, 728.8, 678.0, 596.5, 520.6, 478.0, 422.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₄H₁₆NaOS [M+Na]⁺ 255.0814; found 255.0810.

Cyclohexyl methylthiomethyl ether (**5j**)



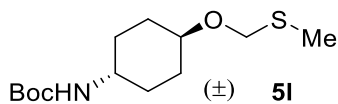
5j²⁵ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.78 g, 75%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.63 (s, 2H), 3.61–3.56 (m, 1H), 2.14 (s, 3H), 1.88–1.83 (m, 2H), 1.73–1.69 (m, 2H), 1.54–1.50 (m, 1H), 1.25 (t, *J* = 9.7 Hz, 5H). ¹³C-NMR (100 MHz, CDCl₃) δ: 74.5, 72.0, 32.1, 25.9, 24.3, 13.9. IR 2927.1, 2855.1, 1444.5, 1298.0, 1263.2, 1060.9, 931.2, 890.3, 792.7, 728.5, 677.5 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₈H₁₇OS [M+H]⁺ 161.0995; found 161.0993.

Tetrahydropyran-4-yl methylthiomethyl ether (**5k**)



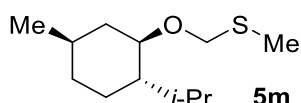
5k was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.97 g, 92%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.63 (s, 2H), 3.87 (dt, *J* = 11.7, 4.4 Hz, 2H), 3.85–3.81 (m, 1H), 3.45–3.39 (m, 2H), 2.12 (s, 3H), 1.86–1.81 (m, 2H), 1.60–1.51 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 71.8, 70.7, 65.7, 32.2, 13.8. IR 2922.6, 2846.8, 1743.6, 1435.3, 1366.7, 1296.9, 1155.5, 1067.9, 1004.2, 947.1, 858.9, 818.5, 728.7, 678.0, 599.6, 515.6 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₇H₁₅O₂S [M+H]⁺ 163.0787; found 163.0784.

trans-1-[(*tert*-Butyloxycarbonyl)amino]-4-methylthiomethoxy cyclohexane (**5l**)



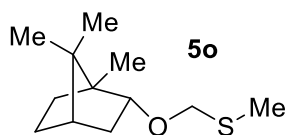
5I was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (1.32 g, 74%) as white solid. M.P. 92–94 °C; ¹H-NMR (400 MHz, CDCl₃) δ: 4.64 (s, 2H), 4.44 (s, 1H), 3.61–3.55 (m, 1H), 3.44 (s, 1H), 2.15 (s, 3H), 2.05–1.97 (m, 4H), 1.44 (s, 9H), 1.39–1.35 (m, 2H), 1.21–1.15 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 155.3, 79.3, 73.7, 72.3, 49.0, 31.2, 30.4, 28.5, 13.8. IR 3320.8, 2976.9, 2931.6, 2860.4, 1676.0, 1522.4, 1445.2, 1365.5, 1310.0, 1271.2, 1229.5, 1164.3, 1076.5, 1038.5, 962.2, 926.7, 896.2, 856.6, 785.9, 735.4, 677.4, 646.4, 463.0 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₃H₂₆NO₃S [M+H]⁺ 276.1628; found 276.1624.

(1*R*,2*S*,5*R*) 2-Isopropyl-5-methyl-1-methylthiomethoxy cyclohexane (**5m**)



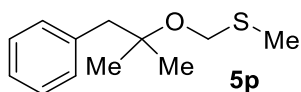
5m²⁶ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.01 g, 72%) as colorless oil. ¹H-NMR (400 MHz, CDCl₃) δ: 4.62 (dd, *J* = 15.4, 11.4 Hz, 2H), 3.36 (td, *J* = 10.5, 4.2 Hz, 1H), 2.21–2.15 (m, 1H), 2.14 (s, 3H), 2.10–2.05 (m, 1H), 1.65–1.61 (m, 2H), 1.36–1.33 (m, 1H), 1.23–1.20 (m, 1H), 1.00–0.88 (m, 8H), 0.86–0.84 (m, 1H), 0.79 (d, *J* = 7.0 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 75.9, 72.4, 48.4, 40.0, 34.6, 31.6, 25.4, 23.2, 22.4, 21.3, 15.9, 14.2. IR 2951.9, 2919.6, 2868.8, 1452.7, 1374.2, 1297.2, 1179.6, 1051.4, 1005.0, 959.3, 918.0, 839.5, 729.3, 678.5, 512.7 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₂₅OS [M+H]⁺ 217.1621; found 217.1613. [α]_D²⁵ = -228 (*c* 0.1, EtOH).

(1*S*,2*R*)-1,7,7-Trimethylbicyclo[2.2.1]heptan-2-yl methylthiomethyl ether (**5o**)



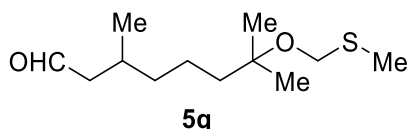
5o was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.53 g, 38%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.69–4.51 (m, 2H), 3.92 (ddd, *J* = 9.5, 3.2, 1.8 Hz, 1H), 2.14–2.10 (m, 4H), 1.97–1.90 (m, 1H), 1.74–1.68 (m, 1H), 1.62 (t, *J* = 4.6 Hz, 1H), 1.24–1.19 (m, 2H), 0.98 (dd, *J* = 13.0, 3.4 Hz, 1H), 0.86–0.83 (m, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 81.6, 74.0, 49.0, 47.9, 45.2, 35.9, 28.3, 27.0, 19.8, 19.0, 13.9, 13.7. IR 2947.3, 1450.2, 1368.8, 1301.0, 1261.9, 1112.7, 1062.9, 818.6, 728.7, 678.4, 473.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₂₂NaOS [M+Na]⁺ 237.1284; found 237.1282. [α]_D²⁵ = -160 (*c* 0.1, EtOH).

1,1-Dimethyl-2-phenylethyl methylthiomethyl ether (**5p**)



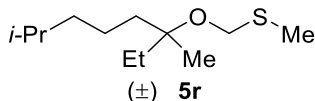
5p was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.74 g, 54%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.26–7.20 (m, 5H), 4.56 (s, 2H), 2.81 (s, 2H), 2.16 (s, 3H), 1.19 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 138.1, 130.7, 127.9, 126.3, 77.1, 66.9, 47.9, 25.2, 14.4. IR 3028.1, 2973.1, 2919.5, 1494.2, 1449.7, 1372.8, 1302.7, 1219.8, 1130.7, 1040.6, 856.3, 763.6, 698.8, 610.6, 514.6, 468.5 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₁₉OS [M+H]⁺ 211.1151; found 211.1149.

3,7,7-Trimethyl-7-methylthiomethoxy heptaldehyde (**5q**)



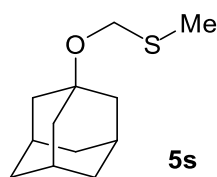
5q was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.91 g, 60%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.74 (t, *J* = 2.3 Hz, 1H), 4.48 (s, 2H), 2.36 (ddd, *J* = 16.1, 5.7, 2.0 Hz, 1H), 2.24 (td, *J* = 8.1, 2.5 Hz, 1H), 2.18 (s, 3H), 2.10–2.02 (m, 1H), 1.48–1.29 (m, 6H), 1.19 (s, 6H), 0.95 (d, *J* = 6.7 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 203.1, 76.6, 66.8, 51.2, 41.0, 37.4, 28.3, 25.73, 25.71, 21.4, 20.1, 14.6. IR 2926.4, 2716.2, 1723.0, 1462.3, 1371.9, 1304.9, 1213.1, 1151.5, 1045.1, 961.7, 733.9, 683.4 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₂₄NaO₂S [M+Na]⁺ 255.1389; found 255.1389.

1-Ethyl-1,5-dimethylhexyl methylthiomethyl ether (**5r**)



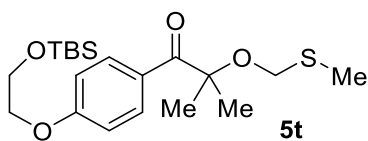
5r was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (0.75 g, 53%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.44 (s, 2H), 2.18 (s, 3H), 1.55–1.50 (m, 3H), 1.45–1.41 (m, 2H), 1.30–1.28 (m, 2H), 1.16–1.14 (m, 5H), 0.87–0.85 (m, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 78.9, 66.2, 39.7, 38.1, 30.8, 28.0, 22.8, 22.75, 22.73, 21.4, 14.6, 8.1. IR 2949.1, 1462.0, 1374.5, 1300.5, 1154.3, 1042.0, 872.3, 733.2, 683.2 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₂₇OS [M+H]⁺ 219.1777; found 219.1775.

1-Adamantyl methylthiomethyl ether (**5s**)



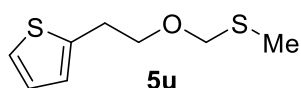
5s²⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (1.02 g, 74%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.58 (s, 2H), 2.17 (s, 3H), 2.14 (s, 3H), 1.79 (d, *J* = 2.8 Hz, 6H), 1.62–1.60 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 74.1, 65.7, 41.9, 36.4, 30.6, 14.4. IR 2906.3, 2850.6, 1446.8, 1353.4, 1298.5, 1104.9, 1073.4, 1035.9, 905.6, 875.8, 813.9, 777.6, 679.3, 548.9, 464.9 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₂₁OS [M+H]⁺ 213.1308; found 213.1301.

1-[4-[2-(*tert*-Butyldimethylsilyloxy)ethoxy]phenyl]-2-methyl-2-methylthiomethoxypropan-1-one (**5t**)



5t was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:10) as a light-yellow oil (2.31 g, 89%) as colorless oil. ¹H-NMR (400 MHz, CDCl₃) δ: 8.28 (d, *J* = 5.2 Hz, 2H), 6.92 (d, *J* = 5.1 Hz, 2H), 4.45 (s, 2H), 4.10 (d, *J* = 3.0 Hz, 2H), 3.99–3.98 (m, 2H), 2.06 (s, 3H), 1.57 (s, 6H), 0.90 (s, 9H), 0.10 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 201.0, 162.8, 132.6, 127.5, 114.1, 83.8, 69.9, 69.6, 62.0, 26.0, 25.6, 18.5, 15.3, -5.1. IR 2929.3, 1742.6, 1670.7, 1598.3, 1506.0, 1427.0, 1371.0, 1254.7, 1211.6, 1159.7, 1129.4, 1014.4, 958.7, 912.1, 834.5, 775.6, 697.5, 599.3, 560.5, 516.5, 463.1 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₂₀H₃₅O₄SSi [M+H]⁺ 399.2020; found 399.2008.

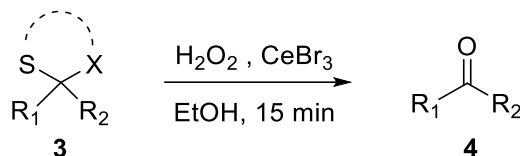
2-(Thiophen-2-yl)ethyl methylthiomethyl ether (**5u**)



5u was purified by flash column chromatography (dichloromethane/hexane = 1:2 to 1:1) as a light-yellow oil (0.43 g, 35%). ¹H-NMR (400 MHz, CDCl₃) δ:

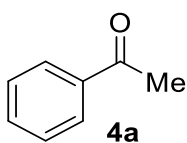
7.14 (dd, $J = 5.1, 1.2$ Hz, 1H), 6.93 (dd, $J = 5.1, 3.4$ Hz, 1H), 6.88–6.86 (m, 1H), 4.66 (s, 2H), 3.77 (t, $J = 6.6$ Hz, 2H), 3.11 (t, $J = 7.3$ Hz, 2H), 2.12 (s, 3H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 141.3, 126.8, 125.3, 123.8, 75.4, 68.6, 30.3, 14.1. IR 3071.5, 2856.1, 1761.7, 1431.4, 1300.7, 1201.8, 1083.0, 1050.1, 985.7, 954.2, 825.3, 688.3, 476.6 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_{13}\text{OS}_2$ $[\text{M}+\text{H}]^+$ 189.0402; found 189.0404.

5. Deprotections of thioacetals, thioketals and methylthiomethyl ether.



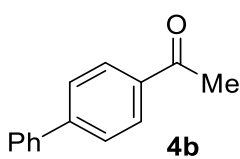
General Procedure III: To a stirred solution of compound **3** (1 mmol) in ethanol (EtOH: 10 mL) were added CeBr_3 (19.0 mg, 0.05 mmol) and 30% H_2O_2 (200 μL , 2 mmol) at rt. After completion of the addition, the resulting mixture was allowed to stir at 40 $^\circ\text{C}$ for 15 min. Then the mixture was quenched by aqueous $\text{Na}_2\text{S}_2\text{O}_3$ (0.1 M, 40 mL) and ethyl acetate (50 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (3 \times 20 mL). The combined organic fractions were washed with brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography to give compound **4**.

Acetophenone (**4a**)



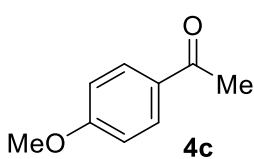
4a²⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (103 mg, 86%). ^1H -NMR (400 MHz, CDCl_3) δ : 7.98–7.93 (m, 2H), 7.59–7.53 (m, 1H), 7.44 (t, $J = 7.9$ Hz, 2H), 2.60 (s, 3H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 198.3, 137.3, 133.2, 128.7, 128.4, 26.7. IR 3062.1, 1680.2, 1593.2, 1445.1, 1357.9, 1179.6, 758.2 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_8\text{O}$ $[\text{M}+\text{H}]^+$ 121.0648; found 121.0649.

4'-Phenylacetophenone (**4b**)



4b²⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a white solid (165 mg, 84%). M.P. 118–120 $^\circ\text{C}$. ^1H -NMR (400 MHz, CDCl_3) δ : 8.06–8.02 (m, 2H), 7.71–7.67 (m, 2H), 7.65–7.61 (m, 2H), 7.50–7.45 (m, 2H), 7.43–7.38 (m, 1H), 2.64 (s, 3H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 197.9, 145.9, 140.0, 136.0, 129.10, 129.06, 128.4, 127.42, 127.38, 26.8. IR 1672.5, 1592.6, 1479.6, 1398.6, 1356.5, 1202.2, 1122.2, 835.3, 759.3 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{14}\text{H}_{10}\text{O}$ $[\text{M}+\text{H}]^+$ 197.0961; found 197.0957.

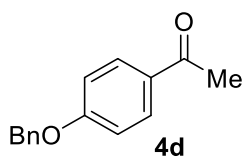
4'-Methoxyacetophenone (**4c**)



4c²⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (138 mg, 92%). ^1H -NMR (400 MHz, CDCl_3) δ : 7.96–7.91 (m, 2H), 6.95–6.91 (m, 2H), 3.87 (s, 3H), 2.55 (s, 3H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 196.9, 163.6, 130.7, 130.5, 113.8, 55.6, 26.5. IR 1667.0, 1597.7, 1505.8,

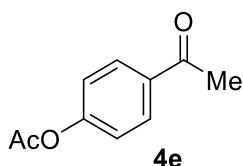
1416.7, 1356.4, 1248.1, 1171.5, 1018.7, 829.4 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_9\text{H}_{11}\text{O}_2$ [M+H]⁺ 151.0754; found 151.0755.

4'-Benzyloxyacetophenone (4d)



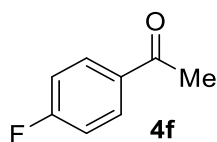
4d²⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a white solid (186 mg, 82%). M.P. 93–95 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 7.96–7.92 (m, 2H), 7.45–7.30 (m, 5H), 7.04–6.99 (m, 2H), 5.13 (s, 2H), 2.55 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 196.8, 162.7, 136.3, 130.7, 130.6, 128.8, 128.3, 127.6, 114.6, 70.2, 26.4. IR 3060.5, 2939.3, 2881.9, 1674.3, 1593.3, 1506.3, 1451.6, 1422.6, 1356.3, 1243.1, 1006.0, 826.9, 757.5 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{15}\text{H}_{15}\text{O}_2$ [M+H]⁺ 227.1067; found 227.1066.

4'-Acetyloxyacetophenone (4e)



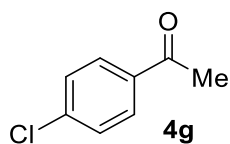
4e³⁰ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (142.55 mg, 80%). ¹H-NMR (400 MHz, CDCl_3) δ : 8.00–7.95 (m, 2H), 7.20–7.15 (m, 2H), 2.58 (s, 3H), 2.31 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 196.9, 168.9, 154.4, 134.8, 130.0, 121.9, 26.7, 21.2. IR 3071.8, 1749.0, 1670.8, 1591.3, 1503.6, 1418.0, 1265.5, 1161.4, 1007.5, 850.3 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{11}\text{O}_3$ [M+H]⁺ 179.0703; found 179.0700.

4'-Fluoroacetophenone (4f)



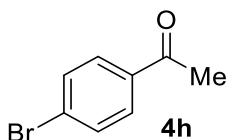
4f³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (117 mg, 85%). ¹H-NMR (400 MHz, CDCl_3) δ : 8.01–7.94 (m, 2H), 7.15–7.08 (m, 2H), 2.58 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 196.6, 164.6 (d, $J = 253$ Hz), 133.7 (d, $J = 2.9$ Hz), 131.0 (d, $J = 9.2$ Hz), 115.7 (d, $J = 21.6$ Hz), 26.6. IR 3073.3, 1682.2, 1594.3, 1504.5, 1408.4, 1357.5, 1227.6, 1155.1, 835.0 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_8\text{FO}$ [M+H]⁺ 139.0554; found 139.0549.

4'-Chloroacetophenone (4g)



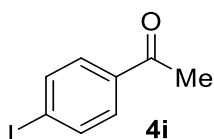
4g³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (139 mg, 90%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.89–7.84 (m, 2H), 7.43–7.38 (m, 2H), 2.56 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 196.9, 139.6, 135.5, 129.8, 129.0, 26.6. IR 3066.7, 1682.3, 1585.2, 1485.8, 1395.4, 1356.5, 1255.6, 1092.3, 1011.7, 823.6, 758.7 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_8\text{ClO}$ [M+H]⁺ 155.0258; found 155.0254.

4'-Bromoacetophenone (4h)



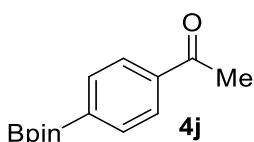
4h³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (175 mg, 88%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.83–7.78 (m, 2H), 7.61–7.57 (m, 2H), 2.57 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 197.1, 135.9, 132.0, 130.0, 128.4, 26.6. IR 1666.1, 1576.2, 1390.4, 1258.7, 1171.7, 1068.0, 818.4 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₈H₈BrO [M+H]⁺ 198.9753; found 198.9747.

4'-Iodoacetophenone (4i)



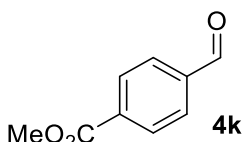
4i²⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a brown solid (224 mg, 91%). M.P. 80–82 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 7.84–7.80 (m, 2H), 7.67–7.63 (m, 2H), 2.56 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 197.4, 138.0, 136.5, 129.8, 101.2, 26.6. IR 1668.1, 1573.4, 1417.8, 1386.4, 1354.0, 1171.5, 813.5 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₈H₈IO [M+H]⁺ 246.9614; found 246.9611.

4'-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)acetophenone (4j)



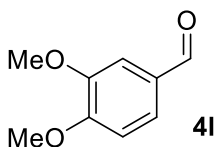
4j³² was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 2:1) as a light-yellow oil (172 mg, 70%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.88 (q, *J* = 8.0 Hz, 4H), 2.61 (s, 3H), 1.35 (s, 12H). ¹³C-NMR (100 MHz, CDCl₃) δ: 198.6, 139.1, 135.0, 127.4, 84.3, 26.9, 25.0. IR 1678.1, 1506.6, 1393.8, 1353.7, 1261.5, 1140.3, 1090.4, 854.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₄H₂₀BO₃ [M+H]⁺ 247.1500; found 247.1498.

4-Methoxycarbonylbenzaldehyde (4k)



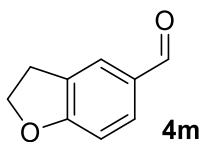
MeCN (10 mL) was used as the solvent instead of EtOH. **4k**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (148 mg, 90%). ¹H-NMR (400 MHz, CDCl₃) δ: 10.10 (s, 1H), 8.21–8.17 (m, 2H), 7.97–7.93 (m, 2H), 3.96 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 191.8, 166.2, 139.3, 135.2, 130.3, 129.7, 52.7. IR 1720.7, 1682.2, 1572.2, 1500.4, 1432.6, 1387.8, 1193.9, 1102.3, 1007.8, 849.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₉H₉O₃ [M+H]⁺ 165.0546; found 165.0542.

3,4-Dimethoxybenzaldehyde (4l)



MeCN (10 mL) was used as the solvent instead of EtOH. **4l**³³ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (143 mg, 86%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.81 (s, 1H), 7.41 (dd, *J* = 8.2, 1.9 Hz, 1H), 7.37 (d, *J* = 1.9 Hz, 1H), 6.93 (d, *J* = 8.2 Hz, 1H), 3.93 (s, 3H), 3.90 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 191.0, 154.5, 149.6, 130.1, 126.9, 110.4, 108.9, 56.2, 56.0. IR 3072.97, 2943.76, 2839.54, 2764.68, 1675.23, 1583.70, 1505.70, 1463.02, 1419.78, 1129.93, 1011.43, 869.50 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₉H₁₁O₃ [M+H]⁺ 167.0703; found 167.0700.

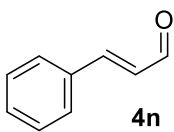
2,3-Dihydrobenzofuran-5-carboxaldehyde (4m)



MeCN (10 mL) was used as the solvent instead of EtOH. **4m**³⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (120 mg, 81%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.80 (s, 1H), 7.73–7.70 (m, 1H), 7.66–7.62 (m, 1H), 6.84 (d, *J* = 8.2 Hz, 1H), 4.64 (t, *J* = 8.8 Hz, 2H), 3.22 (t, *J* = 9.2 Hz, 2H).

¹³C-NMR (100 MHz, CDCl₃) δ: 190.7, 165.7, 133.1, 130.5, 128.5, 126.0, 109.7, 72.5, 28.8. IR 2906.3, 2833.3, 2742.8, 1677.6, 1596.9, 1486.1, 1441.2, 1327.5, 1239.7, 1091.9, 816.4 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₉H₉O₂ [M+H]⁺ 149.0597; found 149.0592.

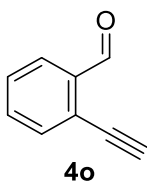
trans-Cinnamaldehyde (**4n**)



MeCN (10 mL) was used as the solvent instead of EtOH. **4n**³³ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (104 mg, 79%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.69 (d, *J* = 7.7 Hz, 1H), 7.58–7.52 (m, 2H), 7.49–7.40 (m, 4H), 6.68 (dd, *J* = 16.0, 7.7 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 193.7,

152.8, 134.1, 131.3, 129.2, 128.64, 128.55. IR 3057.9, 2813.5, 2741.8, 1668.6, 1621.7, 1448.8, 1296.4, 1247.8, 1118.7, 744.8 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₉H₉O [M+H]⁺ 133.0648; found 133.0647.

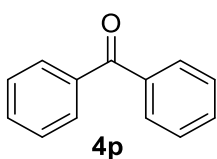
2-Ethynylbenzaldehyde (**4o**)



MeCN (10 mL) was used as the solvent instead of EtOH. **4o**³⁵ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (107 mg, 82%). ¹H-NMR (400 MHz, CDCl₃) δ: 10.53 (s, 1H), 7.91 (dd, *J* = 7.7, 1.4 Hz, 1H), 7.60 (dd, *J* = 7.8, 1.5 Hz, 1H), 7.54 (td, *J* = 7.3, 1.5 Hz, 1H), 7.50–7.44 (m, 1H), 3.46 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 191.5, 136.7, 134.0, 133.8, 129.3, 127.4, 125.6, 84.4, 79.3. IR 3345.2,

2847.6, 2753.7, 2091.9, 1681.6, 1584.3, 1445.5, 1388.2, 1342.0, 1264.4, 1195.3, 726.5 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₉H₇O [M+H]⁺ 131.0491; found 131.0490.

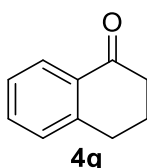
Benzophenone (**4p**)



4p³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (175 mg, 96%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.83–7.78 (m, 4H), 7.62–7.56 (m, 2H), 7.51–7.45 (m, 4H). ¹³C-NMR (100 MHz, CDCl₃) δ: 196.8, 137.7, 132.5, 130.2, 128.4. IR 3056.8, 1653.2, 1594.7, 1444.4, 1361.8, 1273.0, 756.5

cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₃H₁₁O [M+H]⁺ 183.0804; found 183.0797.

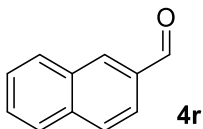
3,4-Dihydronaphthalen-1(2H)-one (**4q**)



4q³⁶ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (118 mg, 81%). ¹H-NMR (400 MHz, CDCl₃) δ: 8.02 (dd, *J* = 7.8, 1.5 Hz, 1H), 7.44 (td, *J* = 7.4, 1.5 Hz, 1H), 7.33–7.22 (m, 2H), 2.95 (t, *J* = 6.1 Hz, 2H), 2.64 (t, *J* = 6.3 Hz, 2H), 2.11 (p, *J* = 6.6 Hz, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 198.5, 144.6, 133.5, 132.7, 128.9, 127.3, 126.7, 39.3, 29.8, 23.4. IR 3025.1, 2941.2, 2871.7, 1678.7, 1599.1,

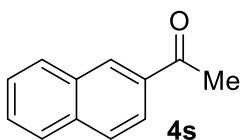
1452.1, 1323.3, 1186.2, 1024.3, 762.9 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{11}\text{O}$ $[\text{M}+\text{H}]^+$ 147.0804; found 147.0797.

2-Naphthaldehyde (4r)



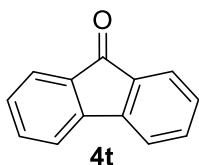
MeCN (10 mL) was used as the solvent instead of EtOH. **4r**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (131 mg, 84%). ¹H-NMR (400 MHz, CDCl_3) δ : 10.13 (s, 1H), 8.28 (s, 1H), 7.99–7.84 (m, 4H), 7.65–7.53 (m, 2H). ¹³C-NMR (100 MHz, CDCl_3) δ : 192.2, 136.5, 134.6, 134.1, 132.7, 129.6, 129.14, 129.10, 128.1, 127.1, 122.8. IR 2825.3, 1683.0, 1458.0, 1339.6, 1158.4, 870.0, 745.2 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{11}\text{H}_9\text{O}$ $[\text{M}+\text{H}]^+$ 157.0648; found 157.0646.

2-Acetonaphthone (4s)



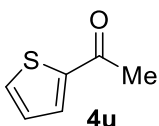
4s²⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (141 mg, 83%). ¹H-NMR (400 MHz, CDCl_3) δ : 8.46 (s, 1H), 8.02 (dd, $J = 8.6, 1.8$ Hz, 1H), 7.95 (d, $J = 8.2$ Hz, 1H), 7.91–7.85 (m, 2H), 7.63–7.52 (m, 2H), 2.72 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 198.2, 135.7, 134.6, 132.6, 130.3, 129.7, 128.6, 128.5, 127.9, 126.9, 124.0, 26.8. IR 3057.5, 1668.7, 1460.1, 1425.8, 1360.4, 1274.9, 1186.0, 1123.0, 867.0, 751.0 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{12}\text{H}_{11}\text{O}$ $[\text{M}+\text{H}]^+$ 171.0804; found 171.0800.

Fluorenone (4t)



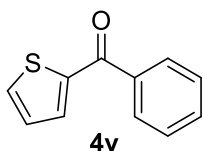
4t³⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a yellow solid (175 mg, 97%). M.P. 81–83 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 7.64 (d, $J = 7.4$ Hz, 2H), 7.53–7.44 (m, 4H), 7.32–7.24 (m, 2H). ¹³C-NMR (100 MHz, CDCl_3) δ : 194.0, 144.6, 134.8, 134.3, 129.2, 124.4, 120.4. IR 3054.2, 1706.1, 1596.8, 1443.0, 1291.6, 1185.2, 1088.9, 726.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{13}\text{H}_9\text{O}$ $[\text{M}+\text{H}]^+$ 181.0648; found 181.0641.

2-Acetylthiophene (4u)



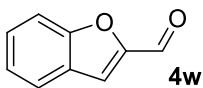
4u³⁸ was purified by flash column chromatography (dichloromethane/hexane = 1:1 to 3:1) as a light-yellow oil (105 mg, 83%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.69 (dd, $J = 3.8, 1.2$ Hz, 1H), 7.63 (dd, $J = 4.9, 1.1$ Hz, 1H), 7.12 (dd, $J = 5.0, 3.8$ Hz, 1H), 2.57 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 190.9, 144.7, 133.9, 132.6, 128.2, 27.1. IR 3094.1, 1655.9, 1516.1, 1410.9, 1355.6, 1267.9, 1028.9, 852.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_6\text{H}_7\text{OS}$ $[\text{M}+\text{H}]^+$ 127.0212; found 127.0211.

2-Benzoylthiophene (4v)



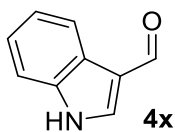
4v³⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (166 mg, 88%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.88–7.84 (m, 2H), 7.70 (dd, *J* = 4.9, 1.2 Hz, 1H), 7.63 (dd, *J* = 3.8, 1.2 Hz, 1H), 7.61–7.55 (m, 1H), 7.52–7.46 (m, 2H), 7.14 (dd, *J* = 5.0, 3.8 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 188.3, 143.7, 138.2, 134.9, 134.3, 132.4, 129.2, 128.5, 128.1. IR 3099.1, 1623.7, 1588.8, 1511.9, 1447.3, 1409.2, 1049.4, 708.0 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₁H₉OS [M+H]⁺ 189.0369; found 189.0368.

Benzofuran-2-carboxaldehyde (**4w**)



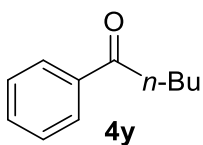
MeCN (10 mL) was used as the solvent instead of EtOH. **4w**⁴⁰ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (120 mg, 82%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.84 (s, 1H), 7.71 (dt, *J* = 7.9, 1.0 Hz, 1H), 7.60–7.53 (m, 2H), 7.52–7.46 (m, 1H), 7.34–7.29 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 179.8, 156.3, 152.8, 129.3, 126.7, 124.3, 123.7, 117.9, 112.7. IR 3089.8, 2835.9, 2726.2, 1675.7, 1609.3, 1552.3, 1476.2, 1445.9, 1323.6, 1286.7, 1117.9, 735.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₉H₇O₂ [M+H]⁺ 147.0441; found 147.0445.

Indole-3-carboxaldehyde (**4x**)



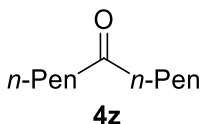
MeCN (10 mL) was used as the solvent instead of EtOH. **4x**⁴⁰ was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 1:1) as a brown solid (123 mg, 85%). M.P. 196–198 °C. ¹H-NMR (400 MHz, DMSO) δ: 12.13 (s, 1H), 9.93 (s, 1H), 8.30–8.25 (m, 1H), 8.08 (d, *J* = 8.0 Hz, 1H), 7.50 (d, *J* = 6.9 Hz, 1H), 7.29–7.18 (m, 2H). ¹³C-NMR (100 MHz, DMSO) δ: 184.9, 138.4, 137.0, 124.1, 123.4, 122.1, 120.8, 118.1, 112.4. IR 3103.9, 3041.4, 2815.6, 2737.4, 1627.1, 1571.4, 1518.6, 1436.7, 1385.6, 1237.8, 1122.7, 1078.5, 756.0 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₉H₈NO [M+H]⁺ 146.0600; found 146.0600.

1-Phenylpentan-1-one (**4y**)



4y³⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (140 mg, 86%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.98–7.94 (m, 2H), 7.57–7.52 (m, 1H), 7.48–7.42 (m, 2H), 2.94 (t, *J* = 7.4 Hz, 2H), 1.69 (p, *J* = 7.3 Hz, 2H), 1.37 (h, *J* = 7.4 Hz, 2H), 0.93 (t, *J* = 7.4 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 200.7, 137.2, 133.0, 128.7, 128.2, 38.4, 26.6, 22.6, 14.1. IR 3061.9, 2932.3, 2868.8, 1682.4, 1593.2, 1450.3, 1353.1, 1263.4, 1207.6, 1010.6, 747.4 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₁H₁₅O [M+H]⁺ 163.1117; found 163.1114.

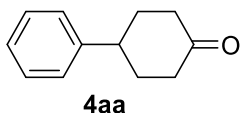
Undecan-6-one (**4z**)



4z⁴¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (160 mg, 94%). ¹H-NMR (400 MHz, CDCl₃) δ: 2.35 (t, *J* = 7.4 Hz, 4H), 1.52 (p, *J* = 7.5 Hz, 4H), 1.36–1.19 (m, 8H), 0.86 (t, *J* = 6.9 Hz, 6H). ¹³C-NMR

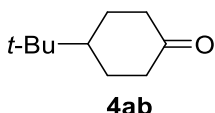
(100 MHz, CDCl₃) δ : 211.8, 42.9, 31.6, 23.7, 22.6, 14.0. IR 2928.7, 2863.7, 1712.4, 1460.6, 1411.1, 1371.7, 1131.7, 729.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₁H₂₃O [M+H]⁺ 171.1743; found 171.1737.

4-Phenylcyclohexanone (4aa)



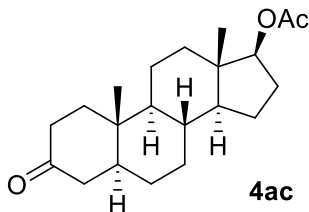
4aa⁴² was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (129 mg, 74%). ¹H-NMR (400 MHz, CDCl₃) δ : 7.35–7.29 (m, 2H), 7.27–7.19 (m, 3H), 2.98 (tt, *J* = 12.1, 3.5 Hz, 1H), 2.57–2.46 (m, 4H), 2.26–2.17 (m, 2H), 2.01–1.87 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ : 211.2, 144.9, 128.7, 126.8, 126.7, 42.8, 41.5, 34.1. IR 3026.3, 2937.1, 2866.9, 1698.4, 1599.6, 1489.8, 1450.2, 1416.9, 1163.6, 755.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₁₅O [M+H]⁺ 175.1117; found 175.1114.

4-*tert*-Butylcyclohexanone (4ab)



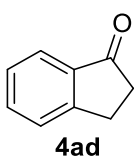
4ab³⁶ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (119 mg, 77%). ¹H-NMR (400 MHz, CDCl₃) δ : 2.41–2.23 (m, 4H), 2.10–2.02 (m, 2H), 1.51–1.36 (m, 3H), 0.89 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ : 212.6, 46.8, 41.4, 32.6, 27.7. IR 2945.7, 2868.3, 1719.2, 1460.4, 1420.3, 1362.1, 1218.3, 1160.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₀H₁₉O [M+H]⁺ 155.1430; found 155.1432.

(5S)-17-Acetyl-4,5-dihydrotestosterone (4ac)



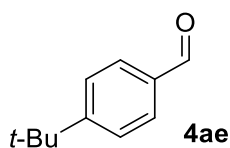
MeCN/AcOH (50/1, 10 mL) was used as the solvent instead of EtOH, and additional CeBr₃ (19 mg, 0.05 mmol) and H₂O₂ (204 μ L, 2 mmol) were needed. **4ac**⁴³ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a white solid (206 mg, 62%). M.P. 159–161 °C. ¹H-NMR (400 MHz, CDCl₃) δ : 4.62–4.56 (m, 1H), 2.42–2.09 (m, 5H), 2.03 (s, 3H), 1.77–1.24 (m, 14H), 1.20–1.12 (m, 1H), 1.01 (s, 3H), 0.96–0.85 (m, 1H), 0.80 (s, 3H), 0.78–0.71 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ : 212.0, 171.3, 82.9, 53.9, 50.7, 46.7, 44.8, 42.8, 38.6, 38.2, 37.0, 35.9, 35.3, 31.4, 28.9, 27.7, 23.7, 21.3, 21.0, 12.3, 11.6. IR 2929.7, 2848.3, 1730.7, 1710.5, 1436.6, 1371.2, 1241.9, 1029.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₂₁H₃₃O₃ [M+H]⁺ 333.2424; found 333.2416. $[\alpha]_D^{25} = +29$ (*c* 1, CHCl₃).

1-Indanone (4ad)



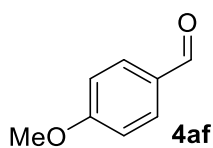
4ad³⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (83.3 mg, 63%). ¹H-NMR (400 MHz, CDCl₃) δ : 7.74 (d, *J* = 7.7 Hz, 1H), 7.60–7.55 (m, 1H), 7.46 (d, *J* = 7.7 Hz, 1H), 7.34 (t, *J* = 7.4 Hz, 1H), 3.12 (t, *J* = 5.8 Hz, 2H), 2.71–2.66 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ : 207.2, 155.3, 137.2, 134.7, 127.4, 126.8, 123.8, 36.3, 25.9. IR 3049.6, 2943.9, 2893.5, 1704.0, 1601.8, 1471.1, 1439.6, 1274.8, 1195.5, 1033.3, 756.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₉H₉O [M+H]⁺ 133.0648; found 133.0644.

4-*tert*-Butylbenzaldehyde (4ae)



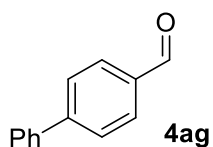
MeCN (10 mL) was used as the solvent instead of EtOH. **4ae**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (146 mg, 90%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.98 (s, 1H), 7.81 (d, *J* = 8.3 Hz, 2H), 7.54 (d, *J* = 8.2 Hz, 2H), 1.36 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 192.2, 158.6, 134.2, 129.8, 126.1, 35.5, 31.2. IR 2906.5, 2870.8, 2823.3, 2726.9, 1697.1, 1604.4, 1467.6, 1414.1, 1368.9, 1217.2, 1105.6, 825.9 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₁H₁₅O [M+H]⁺ 163.1117; found 163.1112.

4-Methoxybenzaldehyde (4af)



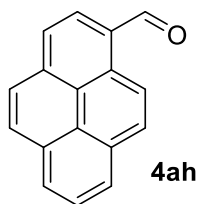
MeCN (10 mL) was used as the solvent instead of EtOH. **4af**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (128 mg, 94%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.88 (s, 1H), 7.82 (d, *J* = 8.4 Hz, 2H), 6.99 (d, *J* = 8.4 Hz, 2H), 3.88 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 190.9, 164.7, 132.1, 130.1, 114.4, 55.7. IR 3009.5, 2837.6, 2738.7, 1681.7, 1596.0, 1508.3, 1460.3, 1253.1, 1213.5, 1154.8, 1021.2, 827.1 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₈H₈O₂ [M+H]⁺ 137.0597; found 137.0597.

4-Phenylbenzaldehyde (4ag)



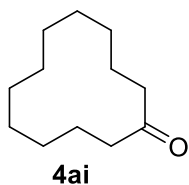
MeCN (10 mL) was used as the solvent instead of EtOH. **4ag**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (162 mg, 89%). ¹H-NMR (400 MHz, CDCl₃) δ: 10.06 (s, 1H), 7.98–7.93 (m, 2H), 7.78–7.74 (m, 2H), 7.66–7.62 (m, 2H), 7.52–7.46 (m, 2H), 7.45–7.40 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 192.0, 147.3, 139.8, 135.3, 130.4, 129.1, 128.6, 127.8, 127.5. IR 3025.1, 2833.8, 2742.7, 1687.7, 1598.2, 1560.8, 1382.7, 1213.0, 1166.7, 830.3, 758.2 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₃H₁₁O [M+H]⁺ 183.0804; found 183.0810.

1-Pyrenecarboxaldehyde (4ah)



MeCN (10 mL) was used as the solvent instead of EtOH. **4ah**⁴⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a yellow solid (200 mg, 87%). M.P. 124–126 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 10.77 (s, 1H), 9.39 (d, *J* = 5.3 Hz, 1H), 8.41 (d, *J* = 4.5 Hz, 1H), 8.31–8.27 (m, 3H), 8.20 (dd, *J* = 7.7, 4.5 Hz, 2H), 8.10–8.06 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 193.2, 135.7, 131.5, 131.2, 131.1, 131.0, 130.8, 130.5, 127.5, 127.3, 127.2, 127.0, 126.7, 124.8, 124.7, 124.2, 123.1. IR 2705.2, 1675.1, 1577.8, 1498.3, 1370.5, 1174.1, 1051.6, 778.8 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₇H₁₁O [M+H]⁺ 231.0804; found 231.0797.

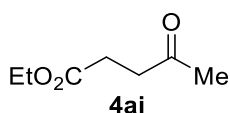
Cyclododecanone (4ai)



4ai³⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:5) as a light-yellow oil (168 mg, 92%). ¹H-NMR (400 MHz, CDCl₃) δ: 2.47–2.42 (m, 4H), 1.74–1.65 (m, 4H), 1.35–1.20 (m, 14H). ¹³C-NMR (100 MHz, CDCl₃) δ: 212.7, 40.4, 24.8, 24.6, 24.2, 22.6, 22.4. IR 2926.0, 2856.4, 1702.1, 1468.0, 1435.4, 1362.4, 1129.8, 1019.2, 722.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₂₃O [M+H]⁺ 183.1743; found

183.1741.

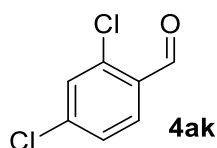
Ethyl levulinate (4aj)



4aj⁴⁵ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (123 mg, 85%). ¹H-NMR (400 MHz, CDCl₃) δ: 4.06 (q, *J* = 7.1 Hz, 2H), 2.69 (t, *J* = 6.5 Hz, 2H), 2.51 (t, *J* = 6.7 Hz, 2H), 2.15 (s, 3H), 1.19 (t, *J* = 7.2 Hz, 3H).

¹³C-NMR (100 MHz, CDCl₃) δ: 206.8, 172.8, 60.7, 38.0, 29.9, 28.1, 14.2. IR 2934.7, 1717.1, 1363.6, 1204.9, 1185.2, 1155.2, 1029.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₇H₁₃O₃ [M+H]⁺ 145.0859; found 145.0854.

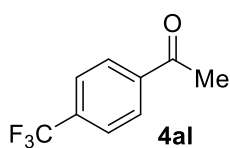
2,4-Dichlorobenzaldehyde (4ak)



MeCN (10 mL) was used as the solvent instead of EtOH. **4ak**⁴⁶ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (154 mg, 88%). ¹H-NMR (400 MHz, CDCl₃) δ: 10.38 (s, 1H), 7.83 (d, *J* = 8.4 Hz, 1H), 7.44 (s, 1H), 7.33 (d, *J* = 8.4 Hz, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 188.5, 141.2, 138.6,

131.0, 130.5, 130.4, 128.0. IR 3081.5, 2884.8, 2639.7, 1683.1, 1576.9, 1461.3, 1412.3, 1248.9, 1043.4, 820.6 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₇H₅Cl₂O [M+H]⁺ 174.9712; found 174.9719.

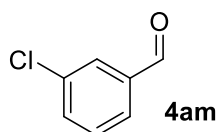
4'-Trifluoromethylacetophenone (4al)



MeCN (10 mL) was used as the solvent instead of EtOH. **4al**³⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (175 mg, 93%). ¹H-NMR (400 MHz, CDCl₃) δ: 8.02 (d, *J* = 5.0 Hz, 2H), 7.69 (d, *J* = 4.8 Hz, 2H), 2.62 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 197.0, 139.8, 134.2 (q, *J* = 18.5 Hz), 128.7, 125.7 (q, *J* = 2.2 Hz), 121.4 (q, *J* = 155 Hz), 26.8. IR 3010.9, 1690.9, 1409.6, 1320.2, 1258.6,

1165.8, 1116.9, 1058.8, 1013.0, 837.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₉H₈F₃O [M+H]⁺ 189.0522; found 189.0516.

3-Chlorobenzaldehyde (4am)

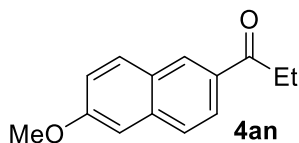


MeCN (10 mL) was used as the solvent instead of EtOH. **4am**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (128 mg, 91%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.97 (s, 1H), 7.84 (t, *J* = 1.1 Hz, 1H), 7.75 (dt, *J* = 4.3, 0.7 Hz, 1H), 7.58 (ddd, *J* = 4.6, 1.3, 0.6 Hz, 1H), 7.47 (t, *J* = 4.4 Hz, 1H).

¹³C-NMR (100 MHz, CDCl₃) δ: 191.0, 137.9, 135.6, 134.5, 130.5, 129.4, 128.1. IR 3070.5, 2830.1, 2729.5,

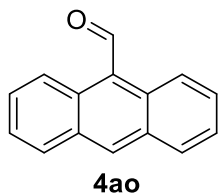
1695.6, 1572.4, 1470.1, 1432.1, 1191.0, 1070.9, 785.5 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_7\text{H}_6\text{ClO}$ [M+H]⁺ 141.0102; found 141.0099.

1-(6-methoxynaphthalen-2-yl)-propan-1-one (4an)



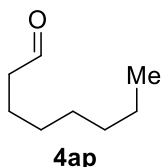
4an⁴⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a brown solid (201 mg, 94%). M.P. 110–112 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 8.38 (s, 1H), 7.99 (dd, J = 4.9, 1.0 Hz, 1H), 7.81 (d, J = 5.1 Hz, 1H), 7.73 (d, J = 4.9 Hz, 1H), 7.17 (dd, J = 5.1, 1.5 Hz, 1H), 7.12 (d, J = 1.5 Hz, 1H), 3.92 (s, 3H), 3.07 (q, J = 4.1 Hz, 2H), 1.26 (t, J = 4.1 Hz, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 200.6, 159.7, 137.2, 132.4, 131.2, 129.5, 127.9, 127.1, 124.7, 119.7, 105.8, 55.5, 31.7, 8.6. IR 3059.5, 1674.9, 1618.2, 1474.8, 1383.1, 1344.9, 1185.4, 1017.3, 860.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{14}\text{H}_{15}\text{O}_2$ [M+H]⁺ 215.1067; found 215.1060.

Anthracene-9-carbaldehyde (4ao)



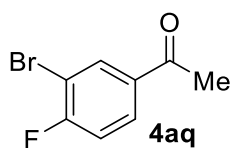
MeCN (10 mL) was used as the solvent instead of EtOH. **4ao**³³ was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow solid (134 mg, 65%). M.P. 103–105 °C; ¹H-NMR (400 MHz, CDCl_3) δ : 11.53 (s, 1H), 8.98 (dd, J = 9.0, 1.0 Hz, 2H), 8.70 (s, 1H), 8.06 (d, J = 8.4 Hz, 2H), 7.71–7.66 (m, 2H), 7.58–7.53 (m, 2H). ¹³C-NMR (100 MHz, CDCl_3) δ : 193.2, 135.4, 132.3, 131.2, 129.4, 129.3, 125.8, 124.9, 123.7. IR 1659.8, 1547.3, 1515.4, 1437.8, 1242.5, 1155.0, 1041.6, 843.8, 723.7 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{15}\text{H}_{11}\text{O}$ [M+H]⁺ 207.0804; found 207.0808.

Octanal (4ap)



MeCN (10 mL) was used as the solvent instead of EtOH. **4ap**⁴⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (64.1 mg, 50%). ¹H-NMR (400 MHz, CDCl_3) δ : 9.76 (t, J = 1.9 Hz, 1H), 2.39 (td, J = 7.4, 1.9 Hz, 2H), 1.67–1.59 (m, 2H), 1.34–1.24 (m, 8H), 0.86 (t, J = 7.0 Hz, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 203.2, 44.1, 31.8, 29.3, 29.2, 22.7, 22.2, 14.2. IR 2925.5, 2857.5, 2715.6, 1725.1, 1461.0, 1383.9, 1140.4, 724.2 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_{17}\text{O}$ [M+H]⁺ 129.1274; found 129.1275.

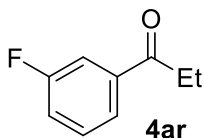
3'-Bromo-4'-fluoroacetophenone (4aq)



MeCN (10 mL) was used as the solvent instead of EtOH. **4aq** was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (176 mg, 81%). ¹H-NMR (400 MHz, CDCl_3) δ : 8.15 (dd, J = 6.6, 2.2 Hz, 1H), 7.88 (ddd, J = 8.6, 4.6, 2.2 Hz, 1H), 7.17 (t, J = 8.3 Hz, 1H), 2.58 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 195.4, 160.9 (d, J = 254 Hz), 134.7 (d, J = 3.4 Hz), 134.3 (d, J = 1.8 Hz), 129.6 (d, J = 8.5

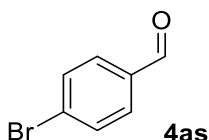
Hz), 116.7 (d, $J = 22.8$ Hz), 109.7 (d, $J = 21.5$ Hz), 26.6. IR 3088.9, 1677.3, 1583.0, 1484.9, 1391.0, 1351.1, 1248.8, 1040.3, 899.6 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_7\text{BrFO}$ [M+H]⁺ 216.9659; found 216.9668.

1-(3-Fluorophenyl)propan-1-one (4ar)



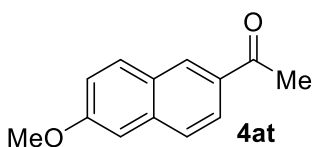
MeCN (10 mL) was used as the solvent instead of EtOH. **4ar**⁴⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (126 mg, 83%). ¹H-NMR (400 MHz, CDCl_3) δ : 7.73 (d, $J = 7.8$ Hz, 1H), 7.67–7.62 (m, 1H), 7.48–7.41 (m, 1H), 7.29–7.22 (m, 1H), 2.96 (q, $J = 7.2$ Hz, 2H), 1.21 (t, $J = 7.4$ Hz, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 199.6 (d, $J = 2.1$ Hz), 161.8 (d, $J = 246$ Hz), 139.1 (d, $J = 6.0$ Hz), 130.3 (d, $J = 7.6$ Hz), 123.8 (d, $J = 3.0$ Hz), 119.9 (d, $J = 21.3$ Hz), 114.8 (d, $J = 21.9$ Hz), 32.1, 8.2. IR 2940.6, 1687.8, 1586.4, 1439.3, 1350.4, 1244.9, 1165.4, 780.7 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_9\text{H}_{10}\text{FO}$ [M+H]⁺ 153.0710; found 153.0713.

4-Bromobenzaldehyde (4as)



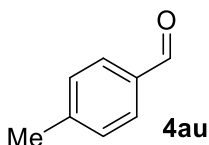
MeCN (10 mL) was used as the solvent instead of EtOH. **4as**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (124 mg, 67%). ¹H-NMR (400 MHz, CDCl_3) δ : 9.97 (s, 1H), 7.76–7.71 (m, 2H), 7.70–7.65 (m, 2H). ¹³C-NMR (100 MHz, CDCl_3) δ : 191.2, 135.2, 132.6, 131.1, 129.9. IR 1685.5, 1576.3, 1473.0, 1379.3, 1198.1, 1147.2, 1058.3, 806.8 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_7\text{H}_6\text{BrO}$ [M+H]⁺ 184.9597; found 184.9590.

2-Acetyl-6-methoxy naphthalene (4at)



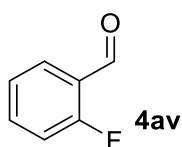
4at⁵⁰ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a white solid (170 mg, 85%). M.P. 109–111 °C. ¹H-NMR (400 MHz, CDCl_3) δ : 8.38 (s, 1H), 7.98 (dd, $J = 8.6, 1.8$ Hz, 1H), 7.83 (d, $J = 8.9$ Hz, 1H), 7.74 (d, $J = 8.6$ Hz, 1H), 7.18 (dd, $J = 8.9, 2.5$ Hz, 1H), 7.14 (d, $J = 2.5$ Hz, 1H), 3.94 (s, 3H), 2.69 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 197.9, 159.9, 137.4, 132.8, 131.2, 130.2, 128.0, 127.2, 124.8, 119.8, 105.9, 55.5, 26.6. IR 1669.45, 1614.38, 1468.06, 1355.57, 1267.16, 1194.54, 1014.38, 857.77 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{13}\text{H}_{13}\text{O}_2$ [M+H]⁺ 201.0910; found 201.0904.

4-Methylbenzaldehyde (4au)



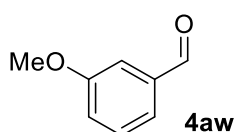
MeCN (10 mL) was used as the solvent instead of EtOH. **4au**³³ was purified by flash column chromatography (ethyl acetate/hexane = 1:20 to 1:5) as a light-yellow oil (76.9 mg, 64%). ¹H-NMR (400 MHz, CDCl_3) δ : 9.95 (s, 1H), 7.75 (d, $J = 8.1$ Hz, 2H), 7.31 (d, $J = 7.8$ Hz, 2H), 2.43 (s, 3H). ¹³C-NMR (100 MHz, CDCl_3) δ : 192.1, 145.6, 134.3, 129.9, 129.8, 22.0. IR 2824.6, 2733.2, 1691.5, 1602.8, 1386.5, 1301.7, 1208.8, 1109.2, 1040.0, 844.9 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_9\text{O}$ [M+H]⁺ 121.0648; found 121.0654.

2-Fluorobenzaldehyde (4av)

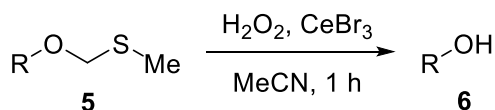


MeCN (10 mL) was used as the solvent instead of EtOH. **4av**³¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (62.1 mg, 50%). ¹H-NMR (400 MHz, CDCl₃) δ: 10.33 (s, 1H), 7.82 (td, *J* = 7.4, 2.0 Hz, 1H), 7.64–7.57 (m, 1H), 7.29–7.23 (m, 1H), 7.19–7.12 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 186.8 (d, *J* = 6.6 Hz), 163.2 (d, *J* = 257 Hz), 136.2 (d, *J* = 9.1 Hz), 128.5 (d, *J* = 1.9 Hz), 124.5 (d, *J* = 3.8 Hz), 124.0 (d, *J* = 7.8 Hz), 116.2 (d, *J* = 20.3 Hz). IR 1693.7, 1609.6, 1583.1, 1481.5, 1457.5, 1400.5, 1273.1, 1188.0, 1095.9, 758.7 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₇H₆FO [M+H]⁺ 125.0397; found 125.0398.

3-Methoxybenzaldehyde (4aw)

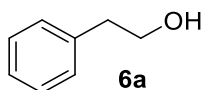


MeCN (10 mL) was used as the solvent instead of EtOH. **4aw**⁵¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:5) as a light-yellow oil (84.4 mg, 62%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.97 (s, 1H), 7.47–7.43 (m, 2H), 7.40–7.38 (m, 1H), 7.19–7.15 (m, 1H), 3.86 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 192.3, 160.3, 137.9, 130.2, 123.7, 121.7, 112.2, 55.6. IR 2837.8, 2730.0, 1697.0, 1589.8, 1482.7, 1459.9, 1389.1, 1146.2, 1038.2, 778.5 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₈H₉O₂ [M+H]⁺ 137.0597; found 137.0601.



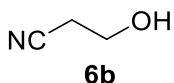
General Procedure IV: To a stirred solution of compound **5** (1 mmol) in MeCN (5 mL) were added CeBr₃ (19.0 mg, 0.05 mmol) and 30% H₂O₂ (100 μL, 1 mmol) at rt. After completion of the addition, the resulting mixture was allowed to stir at room temperature for 1 h. Then the mixture was quenched by aqueous Na₂S₂O₃ (0.1 M, 20 mL) and ethyl acetate (20 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (3 × 20 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography to give compound **6**.

2-Phenylethanol (6a)



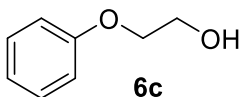
6a⁵² was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (85.5 mg, 70%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.35–7.31 (m, 2H), 7.27–7.23 (m, 3H), 3.81 (t, *J* = 6.7 Hz, 2H), 2.85 (t, *J* = 6.6 Hz, 2H), 1.98 (brs, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 138.6, 129.1, 128.6, 126.5, 63.7, 39.2. IR 3323.1, 3027.4, 2939.0, 2872.9, 1601.6, 1494.2, 1450.5, 1174.6, 1042.0, 907.3, 854.1, 742.1, 695.5, 570.2, 491.9 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₈H₁₀O [M+H]⁺ 123.0804; found 123.0809.

2-Cyanoethanol (6b)



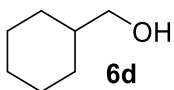
Solid Na₂S₂O₃ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6b**⁵³ was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 2:1) as a light-yellow oil (59.7 mg, 84%). ¹H-NMR (400 MHz, CDCl₃) δ: 3.84 (t, *J* = 6.0 Hz, 2H), 3.18 (s, 1H), 2.59 (t, *J* = 6.0 Hz, 2H). ¹³C-NMR (101 MHz, CDCl₃) δ: 118.5, 57.7, 21.5. IR 3417.4, 2899.2, 2254.3, 1723.4, 1638.2, 1414.3, 1326.5, 1171.0, 1119.1, 1051.1, 852.1, 576.1 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₃H₆NO [M+H]⁺ 72.0444; found 72.0447.

2-Phenoxyethanol (6c)



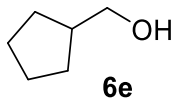
6c⁵⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (98.1 mg, 71%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.28 (t, *J* = 7.4 Hz, 2H), 7.00-6.92 (m, 3H), 4.06 (t, *J* = 4.2 Hz, 2H), 3.94 (t, *J* = 4.2 Hz, 2H), 2.53 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 158.7, 129.6, 121.2, 114.6, 69.2, 61.5. IR 3385.3, 2930.7, 1721.9, 1594.2, 1492.3, 1455.4, 1327.8, 1298.3, 1239.6, 1169.0, 1128.0, 1078.0, 1042.7, 957.2, 913.3, 791.7, 751.0, 690.3, 604.3, 554.6, 509.1, 482.1 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₈H₁₁O₂ [M+H]⁺ 139.0754; found 139.0751.

Cyclohexylmethanol (6d)



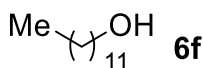
Solid Na₂S₂O₃ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6d**⁵⁵ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (74.2 mg, 65%). ¹H-NMR (400 MHz, CDCl₃) δ: 3.37 (d, *J* = 6.4 Hz, 2H), 2.26 (s, 1H), 1.73–1.61 (m, 5H), 1.47–1.39 (m, 1H), 1.26–1.11 (m, 3H), 0.93–0.83 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 69.0, 40.7, 29.7, 26.7, 26.0. IR 3334.1, 2922.1, 2851.7, 1718.9, 1448.9, 1338.8, 1171.1, 1089.1, 1019.2, 973.0, 943.1, 891.7, 835.9, 778.5, 744.9, 554.1, 524.5, 486.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₇H₁₅O [M+H]⁺ 115.1117; found 115.1121.

Cyclopentylmethanol (6e)



Solid Na₂S₂O₃ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6e**⁵⁶ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (73.1 mg, 73%). ¹H-NMR (400 MHz, CDCl₃) δ: 3.50–3.48 (m, 2H), 2.11–2.04 (m, 1H), 1.77–1.70 (m, 2H), 1.60–1.51 (m, 4H), 1.24–1.19 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 67.5, 42.2, 29.2, 25.6. IR 3318.8, 2946.4, 2864.3, 1449.4, 1157.9, 1030.7, 934.7, 624.7 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₆H₁₆NO [M+NH₄]⁺ 118.1226; found 118.1224.

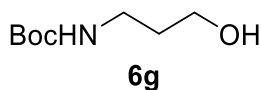
1-Undecanol (6f)



6f⁵⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (123 mg, 66%). ¹H-NMR (400 MHz, CDCl₃) δ: 3.61 (t, *J* = 6.6 Hz,

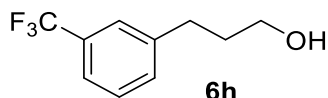
2H), 1.56–1.51 (m, 2H), 1.27–1.23 (m, 18H), 0.84 (t, $J = 6.3$ Hz, 3H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 63.3, 32.7, 32.1, 29.8, 29.77, 29.74, 29.6, 29.5, 25.9, 22.8, 14.2. IR 3353.3, 2921.8, 2853.4, 1461.6, 1350.9, 1173.6, 1046.6, 720.5, 528.6 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{12}\text{H}_{27}\text{O}$ [$\text{M}+\text{H}$]⁺ 187.2056; found 187.2065.

3-[(*tert*-Butyloxycarbonyl)amino]propanol (**6g**)



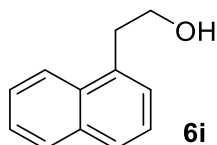
Solid $\text{Na}_2\text{S}_2\text{O}_3$ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6g**⁵⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:2) as a light-yellow oil (123 mg, 70%). ^1H -NMR (400 MHz, CDCl_3) δ : 4.91 (s, 1H), 3.61 (t, $J = 5.3$ Hz, 2H), 3.29–3.24 (m, 2H), 1.67–1.64 (m, 2H), 1.41 (s, 9H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 157.2, 79.6, 59.3, 37.1, 32.9, 28.5. IR 3348.7, 2974.1, 1682.7, 1519.6, 1452.0, 1393.9, 1365.4, 1163.9, 1046.1, 866.8, 777.9, 596.3 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_8\text{H}_{18}\text{NO}_3$ [$\text{M}+\text{H}$]⁺ 176.1281; found 176.1283.

3-(3-Trifluoromethylphenyl)propanol (**6h**)



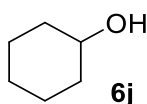
6h⁵⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (167 mg, 82%). ^1H -NMR (400 MHz, CDCl_3) δ : 7.45–7.38 (m, 4H), 3.67 (t, $J = 6.3$ Hz, 2H), 2.76 (t, $J = 7.8$ Hz, 2H), 1.94–1.87 (m, 2H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 142.9, 132.0, 130.7 (q, $J = 32.0$ Hz), 128.9, 125.2 (q, $J = 4.0$ Hz), 122.8 (q, $J = 4.0$ Hz), 120.3 (q, $J = 271.0$ Hz), 62.0, 34.1, 32.0. IR 3325.8, 2941.1, 1448.4, 1325.3, 1160.5, 1118.6, 1067.8, 903.7, 798.8, 740.0, 700.5, 658.7, 516.0 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{10}\text{H}_{12}\text{F}_3\text{O}$ [$\text{M}+\text{H}$]⁺ 205.0835; found 205.0839.

2-Naphthaleneethanol (**6i**)



6i⁶⁰ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (127 mg, 74%). ^1H -NMR (400 MHz, CDCl_3) δ : 8.05 (d, $J = 8.2$ Hz, 1H), 7.87 (d, $J = 7.8$ Hz, 1H), 7.76 (d, $J = 8.0$ Hz, 1H), 7.56–7.48 (m, 2H), 7.45–7.38 (m, 2H), 3.98 (t, $J = 6.6$ Hz, 2H), 3.34 (t, $J = 6.6$ Hz, 2H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 134.5, 134.1, 132.2, 129.0, 127.5, 127.3, 126.2, 125.8, 125.6, 123.8, 63.2, 36.3. IR 3327.4, 3046.4, 2943.2, 2875.6, 1716.1, 1593.9, 1509.5, 1391.4, 1164.1, 1034.7, 865.7, 772.7, 732.7, 597.7, 551.5, 422.3 cm^{-1} ; HRMS (ESI⁺) (m/z) calcd. for $\text{C}_{12}\text{H}_{12}\text{NaO}$ [$\text{M}+\text{Na}$]⁺ 195.0780; found 195.0781.

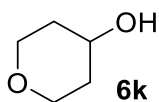
Cyclohexanol (**6j**)



Solid $\text{Na}_2\text{S}_2\text{O}_3$ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6j**⁶¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (90.1 mg, 90%). ^1H -NMR (400 MHz, CDCl_3) δ : 3.62–3.59 (m, 1H), 1.92–1.87 (m, 1H), 1.77–1.70 (m, 2H), 1.57–1.45

(m, 3H), 1.31–1.14 (m, 5H). ¹³C-NMR (100 MHz, CDCl₃) δ: 70.3, 35.6, 25.5, 24.1. IR 3341.3, 2997.5, 2855.4, 1715.5, 1449.6, 1367.0, 1174.8, 1059.1, 964.0, 889.9, 841.5, 781.7 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₆H₁₃O [M+H]⁺ 101.0961; found 101.0956.

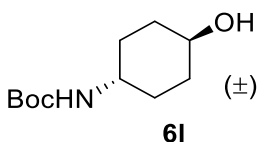
Tetrahydro-4-pyranol (**6k**)



Solid Na₂S₂O₃ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6k**⁶² was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 2:1) as a light-yellow oil (87.8 mg, 86%).

¹H-NMR (400 MHz, CDCl₃) δ: 3.92 (dt, *J* = 11.7, 4.4 Hz, 2H), 3.88–3.81 (m, 1H), 3.46–3.40 (m, 2H), 1.91–1.87 (m, 2H), 1.74 (s, 1H), 1.60–1.51 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 67.1, 65.8, 35.6. IR 3382.2, 2945.2, 2854.8, 1652.8, 1450.3, 1368.0, 1285.3, 1227.1, 1133.0, 1077.0, 1032.7, 1004.5, 984.6, 857.2, 813.7, 611.6, 517.5 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₅H₁₁O₂ [M+H]⁺ 103.0754; found 103.0756.

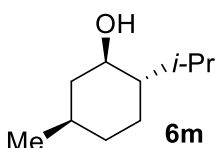
trans-4-[(*tert*-Butyloxycarbonyl)amino]cyclohexan-1-ol (**6l**)



Solid Na₂S₂O₃ (316 mg, 2 mmol) was used to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly. **6l**⁶³ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:2) as a white solid (129 mg, 60%). M.P. 172–174 °C.

¹H-NMR (400 MHz, CDCl₃) δ: 4.38 (s, 1H), 3.60–3.55 (m, 1H), 3.40 (s, 1H), 2.00–1.94 (m, 4H), 1.77 (s, 1H), 1.42 (s, 9H), 1.37–1.34 (m, 2H), 1.16–1.12 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ: 155.4, 79.4, 69.9, 49.0, 34.1, 31.3, 28.5. IR 3340.1, 2978.6, 2933.5, 2858.7, 2280.5, 2114.4, 1678.5, 1523.4, 1451.4, 1363.5, 1312.0, 1263.3, 1228.1, 1063.7, 948.5, 895.1, 782.9, 740.2, 619.0, 521.6, 461.4, 427.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₁H₂₂NO₃ [M+H]⁺ 216.1594; found 216.1592.

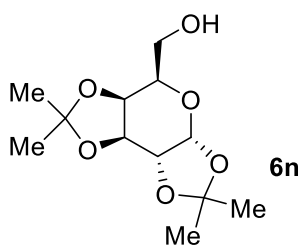
(1*R*,2*S*,5*R*) (-)-Menthol (**6m**)



6m⁶⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (127 mg, 81%). ¹H-NMR (400 MHz, CDCl₃) δ: 3.37 (td, *J* = 10.4, 4.3 Hz, 1H), 2.19–2.14 (m, 1H), 1.98–1.93 (m, 1H), 1.68–1.58 (m, 2H), 1.43–1.41 (m, 1H), 1.13–1.07 (m, 1H), 0.96–0.90 (m, 8H), 0.87–0.82 (m, 1H), 0.80 (d, *J* =

1.5 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 71.7, 50.3, 45.2, 34.7, 31.8, 26.0, 23.3, 22.4, 21.2, 16.2. IR 3359.0, 2952.5, 2921.3, 2867.8, 1723.4, 1454.6, 1368.9, 1176.1, 1099.4, 1027.0, 985.3, 914.5, 874.4, 842.6, 771.2, 545.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₀H₂₀NaO [M+Na]⁺ 179.1406; found 179.1407. [α]_D²⁵ = -51 (*c* 0.1, EtOH)

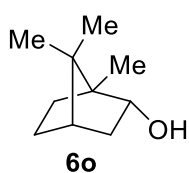
1,2:3,4-Di-*O*-isopropylidene- α -D-galactopyranose (**6n**)



6n

6n⁶⁵ was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 2:1) as a light-yellow oil (195 mg, 75%). ¹H-NMR (400 MHz, CDCl₃) δ: 5.56 (d, *J* = 5.0 Hz, 1H), 4.60 (d, *J* = 7.9 Hz, 1H), 4.34–4.26 (m, 2H), 3.87–3.83 (m, 2H), 3.77–3.73 (m, 1H), 1.53 (s, 3H), 1.46 (s, 3H), 1.33 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 109.6, 108.8, 96.4, 71.8, 70.9, 70.7, 68.2, 62.5, 26.2, 26.1, 25.1, 24.4. IR 3493.7, 2986.0, 2932.9, 1723.5, 1457.1, 1377.8, 1253.2, 1209.7, 1168.7, 1063.3, 997.7, 890.8, 858.7, 802.4, 769.0, 644.2, 552.8, 509.7 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₂H₂₁O₆ [M+H]⁺ 261.1333; found 261.1335. [α]_D²⁵ = -59 (*c* 0.1, CHCl₃)

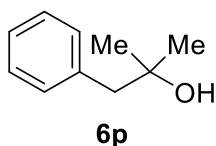
(-)-Borneol (6o)



6o

6o⁶⁶ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a white solid (128 mg, 83%). M.P. 207–210 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 4.01 (ddd, *J* = 10.0, 3.3, 1.6 Hz, 1H), 2.30–2.23 (m, 1H), 1.91–1.84 (m, 1H), 1.75–1.69 (m, 1H), 1.60 (t, *J* = 4.6 Hz, 1H), 1.27–1.20 (m, 2H), 0.91 (dd, *J* = 13.4, 3.4 Hz, 1H), 0.86–0.84 (m, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 77.5, 49.6, 48.2, 45.2, 39.2, 28.4, 26.0, 20.3, 18.8, 13.5. IR 3299.0, 2945.2, 2874.9, 1451.8, 1380.2, 1303.0, 1232.1, 1139.3, 1107.5, 1053.4, 1023.0, 941.3, 828.4, 656.6, 528.1, 461.2 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₀H₁₈KO [M+K]⁺ 193.0989; found 193.0980. [α]_D²⁵ = -34 (*c* 0.1, EtOH).

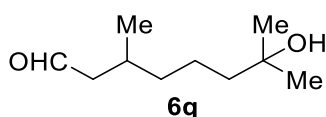
2-Methyl-1-phenylpropan-2-ol (6p)



6p

6p⁶⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (116 mg, 77%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.32–7.20 (m, 5H), 2.76 (s, 2H), 1.22 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 137.9, 130.6, 128.3, 126.6, 70.9, 49.9, 29.3. IR 3389.7, 3028.6, 2971.0, 2927.6, 1709.0, 1493.1, 1455.1, 1373.1, 1306.5, 1207.1, 1146.6, 1031.3, 974.3, 900.0, 777.7, 725.4, 698.2, 608.2, 516.3, 466.3 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₀H₁₄NaO [M+Na]⁺ 173.0937; found 173.0938.

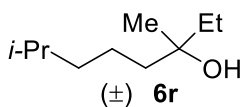
7-Hydroxy-3,7-dimethyloctanal (6q)



6q

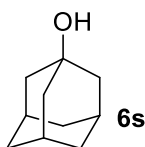
6q⁶⁷ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:2) as a light-yellow oil (141 mg, 82%). ¹H-NMR (400 MHz, CDCl₃) δ: 9.69 (t, *J* = 2.3 Hz, 1H), 2.36–2.32 (m, 1H), 2.22–2.15 (m, 1H), 2.04–1.99 (m, 1H), 1.45–1.30 (m, 6H), 1.19–1.15 (m, 6H), 0.90 (d, *J* = 6.7 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ: 203.2, 71.1, 51.1, 43.9, 37.4, 29.34, 29.25, 28.2, 21.7, 20.0. IR 3382.7, 2935.4, 1718.7, 1462.9, 1372.5, 1134.7, 937.1, 905.9, 761.9, 553.6, 482.7 cm⁻¹; HRMS (ESI⁺) (*m/z*) calcd. for C₁₀H₂₀NaO₂ [M+Na]⁺ 195.1356; found 195.1352.

3,7-Dimethyloctan-3-ol (6r)



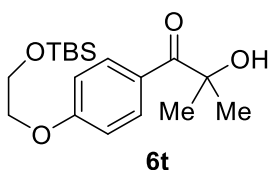
6r⁶⁸ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (125 mg, 79%). ¹H-NMR (400 MHz, CDCl₃) δ: 1.58–1.51 (m, 1H), 1.50–1.35 (m, 5H), 1.34–1.26 (m, 2H), 1.20–1.15 (m, 2H), 1.14 (s, 3H), 0.90–0.86 (m, 9H). ¹³C-NMR (100 MHz, CDCl₃) δ: 73.1, 41.7, 39.7, 34.4, 28.1, 26.5, 22.8, 21.7, 8.3. IR 3382.6, 2956.4, 1461.9, 1373.7, 1152.8, 1081.9, 972.8, 899.2, 840.1, 738.7, 612.3 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₀H₂₂KO [M+K]⁺ 197.1302; found 197.1301.

1-Adamantanol (6s)



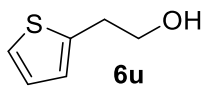
6s⁶⁴ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a white solid (129 mg, 85%). M.P. 246–249 °C. ¹H-NMR (400 MHz, CDCl₃) δ: 2.13 (s, 3H), 1.70 (d, *J* = 2.6 Hz, 6H), 1.64–1.56 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 68.4, 45.5, 36.2, 30.8. IR 3272.9, 2891.8, 2843.9, 1722.7, 1446.5, 1345.8, 1299.3, 1223.7, 1174.7, 1109.6, 1081.5, 976.6, 923.2, 809.3, 651.4, 548.4, 464.6, 432.5 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₀H₁₆NaO [M+Na]⁺ 175.1093; found 175.1090.

1-[4-[2-(*tert*-Butyldimethylsilyloxy)ethoxy]phenyl]-2-hydroxy-2-methylpropan-1-one (6t)



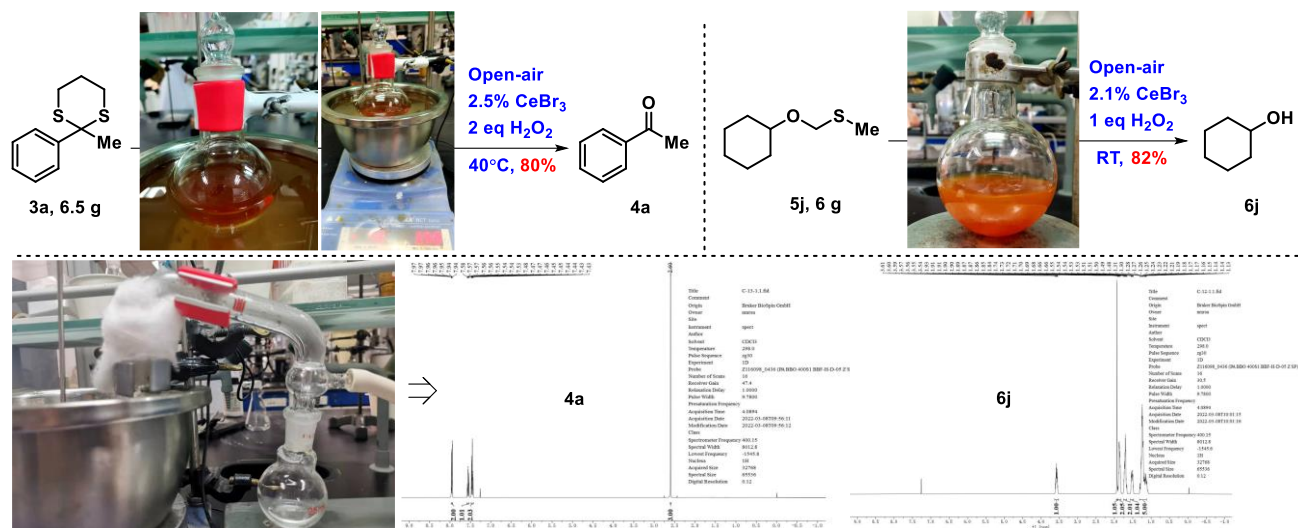
THF (5 mL) was used as the solvent instead of MeCN. **6t** was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (244 mg, 72%). ¹H-NMR (400 MHz, CDCl₃) δ: 8.03 (d, *J* = 8.5 Hz, 2H), 6.93 (d, *J* = 8.5 Hz, 2H), 4.30 (s, 1H), 4.10 (t, *J* = 4.8 Hz, 2H), 3.97 (t, *J* = 4.8 Hz, 2H), 1.62 (s, 6H), 0.90 (s, 9H), 0.09 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃) δ: 202.7, 163.0, 132.5, 126.0, 114.3, 75.9, 69.7, 61.9, 28.8, 26.0, 18.5, -5.1. IR 3454.2, 2931.6, 2858.5, 1736.9, 1663.6, 1598.3, 1571.8, 1507.4, 1463.8, 1416.8, 1368.4, 1306.8, 1252.4, 1161.5, 1128.4, 1053.9, 954.9, 832.0, 773.1, 728.5, 667.1, 586.8, 565.7, 510.9, 426.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₈H₃₁O₄Si [M+H]⁺ 339.1986; found 339.1998.

Thiophen-2-ylethanol (6u)



6u⁶⁹ was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) as a light-yellow oil (103 mg, 80%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.16 (dd, *J* = 5.1, 1.2 Hz, 1H), 6.95 (dd, *J* = 5.1, 3.4 Hz, 1H), 6.88–6.87 (m, 1H), 3.83 (t, *J* = 6.3 Hz, 2H), 3.06 (t, *J* = 6.4 Hz, 2H), 1.89 (s, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 140.9, 127.1, 125.7, 124.1, 63.6, 33.4. IR 3327.40, 2936.70, 1433.76, 1239.20, 1132.01, 1041.30, 845.97, 821.23, 690.27, 494.72 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₆H₉OS [M+H]⁺ 129.0369; found 129.0370.

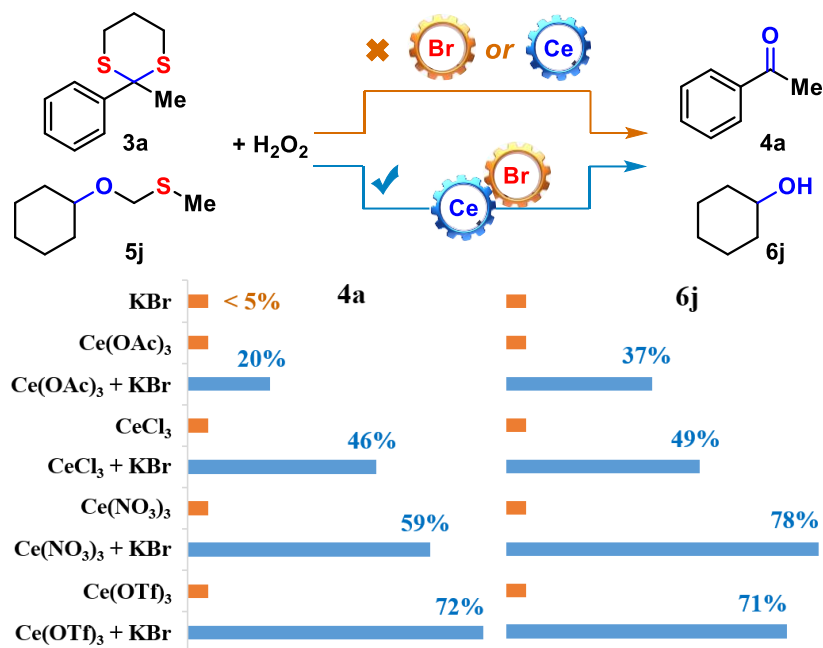
6. Gram-scale dethioacetalization of 3a and 5j under open-air condition



Gram-scale synthesis of 4a: To a stirred solution of compound **3a** (6.5 g, 30.9 mmol) in EtOH (150 mL) were added CeBr₃ (285 mg, 0.75 mmol) and H₂O₂ (30%, 6.2 mL, 60 mmol). After completion of the addition, the resulting mixture was allowed to stir at 40 °C for additional 1 h. Then the reaction was quenched by solid Na₂S₂O₃ (9.5 g, 60 mmol), and stirred for 30 min followed by filtration. The resulting filtrate was evaporated and purified by vacuum distillation (131-139 °C/~80 Torr) to give compound **4a** (2.97 g, 80%).

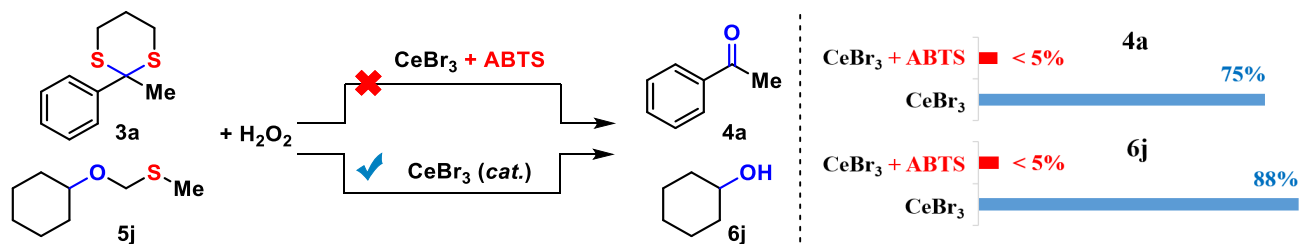
Gram-scale synthesis of 6j: To a stirred solution of compound **5j** (6.0 g, 37.4 mmol) in MeCN (90 mL) were added CeBr₃ (300 mg, 0.79 mmol) and H₂O₂ (30%, 3.9 mL, 37.4 mmol). After completion of the addition, the resulting mixture was allowed to stir at rt for additional 1 h. Then the reaction was quenched by solid Na₂S₂O₃ (5.9 g, 37.4 mmol), and stirred for 30 min followed by filtration. The resulting filtrate was evaporated and purified by vacuum distillation (107-113 °C/~80 Torr) to give compound **6j** (3.07 g, 82%).

7. Controlled experiments of bromide and cerium catalysts for dethioacetalization.



To a stirred solution of compound **3a** (21.0 mg, 0.1 mmol) in EtOH (1 mL) or **5j** (16.0 mg, 0.1 mmol) in MeCN (1 mL) were added KBr (with or without, 3.6 mg, 0.03 mmol), metal catalyst (0.01 mmol) and H₂O₂ (30%, 20 μ L, 0.2 mmol for **3a**; 10 μ L, 0.1 mmol for **5j**). After completion of the addition, the reaction mixture was allowed to stir at rt for 1.5 h. The reaction was quenched by dilute aqueous Na₂S₂O₃ solution (0.1 M, 4 mL) and ethyl acetate (10 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (2 \times 10 mL). The combined organic fractions were washed with H₂O, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. Yield was determined by ¹H-NMR of the crude reaction mixture using CH₂Br₂ as the internal reference. For compound **6j**, solid Na₂S₂O₃ (63.2 mg, 0.4 mmol) was added directly to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly.

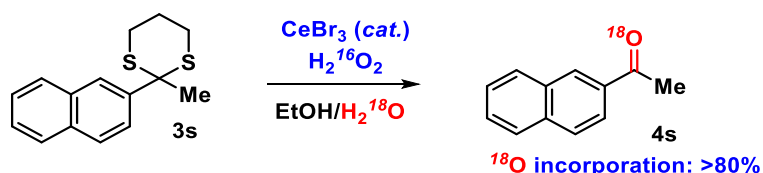
8. Radical trapping (ABTS) experiments.



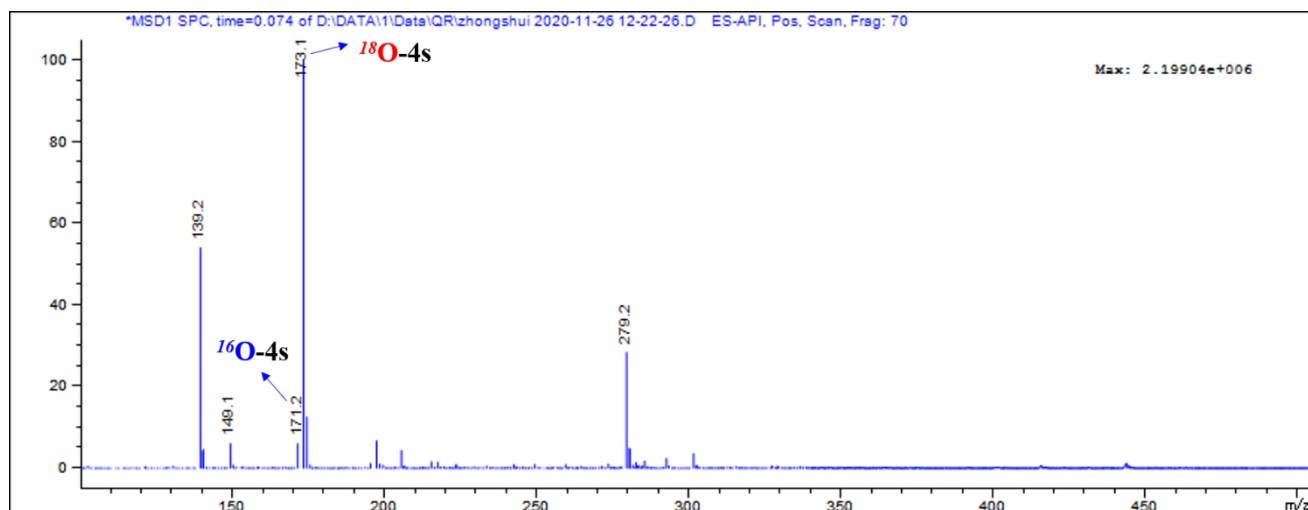
To a stirred solution of compound **3a** (21.0 mg, 0.1 mmol) in EtOH/H₂O (10/1, 1.1 mL) or compound **5j** (16.0 mg, 0.1 mmol) in MeCN/H₂O (10/1, 1.1 mL) were added ABTS (with or without, 110 mg, 0.2 mmol), and the mixture was stirred for 5 min. Then H₂O₂ (30%, 20 μ L, 0.2 mmol for **3a**; 10 μ L, 0.1 mmol for **5j**) and CeBr₃ (3.8 mg, 0.01 mmol) were added, and the resulting solution was stirred at rt for 1.5 h. The reaction was quenched by dilute aqueous Na₂S₂O₃ solution (0.1 M, 4 mL) and ethyl acetate (10 mL). The organic fractions

were collected, and the aqueous phase was extracted with ethyl acetate (2 × 4 mL). The combined organic fractions were washed with H₂O, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. Yield was determined by ¹H-NMR of the crude reaction mixture using CH₂Br₂ as the internal reference. For compound **6j**, solid Na₂S₂O₃ (63.2 mg, 0.4 mmol) was added to quench the reaction instead of an aqueous solution, and the resulting mixture was evaporated directly.

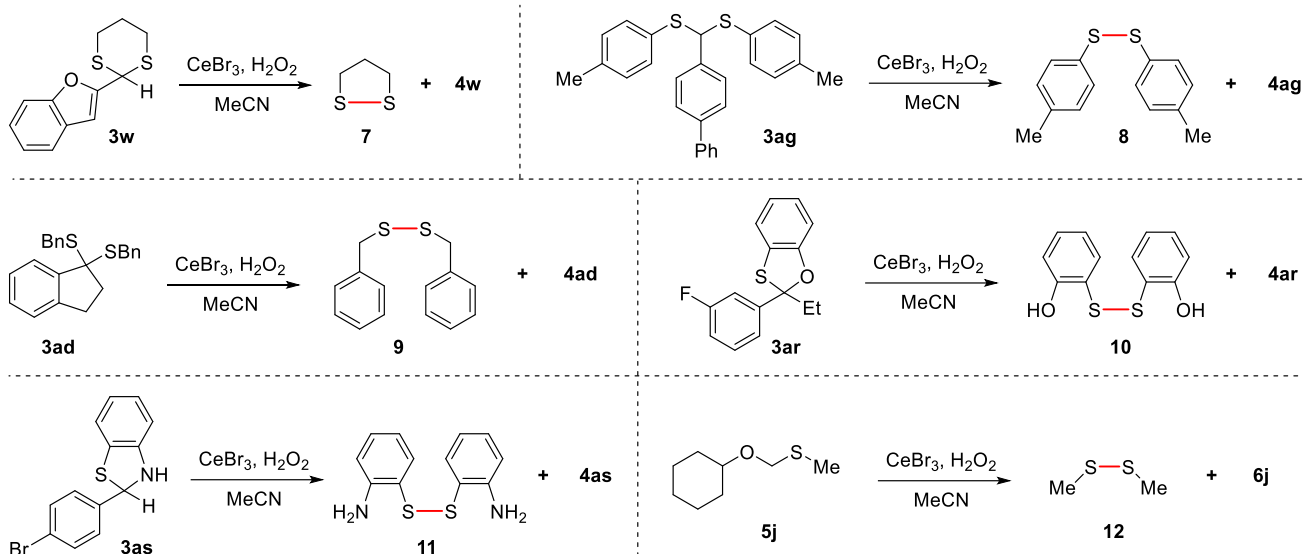
9. Isotopic labeling experiment.



To a stirred solution of compound **3s** (13.0 mg, 0.05 mmol) in ethanol (1 mL) and H₂¹⁸O (0.2 mL) were added CeBr₃ (1.9 mg, 0.005 mmol) and 30% H₂O₂ (10 μL, 0.1 mmol). After completion of the addition, the resulting mixture was allowed to stir at rt for 1.5 h. Then the mixture was quenched by aqueous Na₂S₂O₃ solution (0.1 M, 10 mL) and ethyl acetate (10 mL). The organic fractions were collected, and the aqueous phase was extracted with ethyl acetate (3 × 10 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography (ethyl acetate/hexane = 1:10 to 1:3) to give compound **4s** (6.0 mg, 70%). MS = 173.1 for ¹⁸O-**4s**, while MS = 171.2 for ¹⁶O-**4s**.

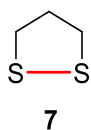


10. The isolation and confirmation of related side products.

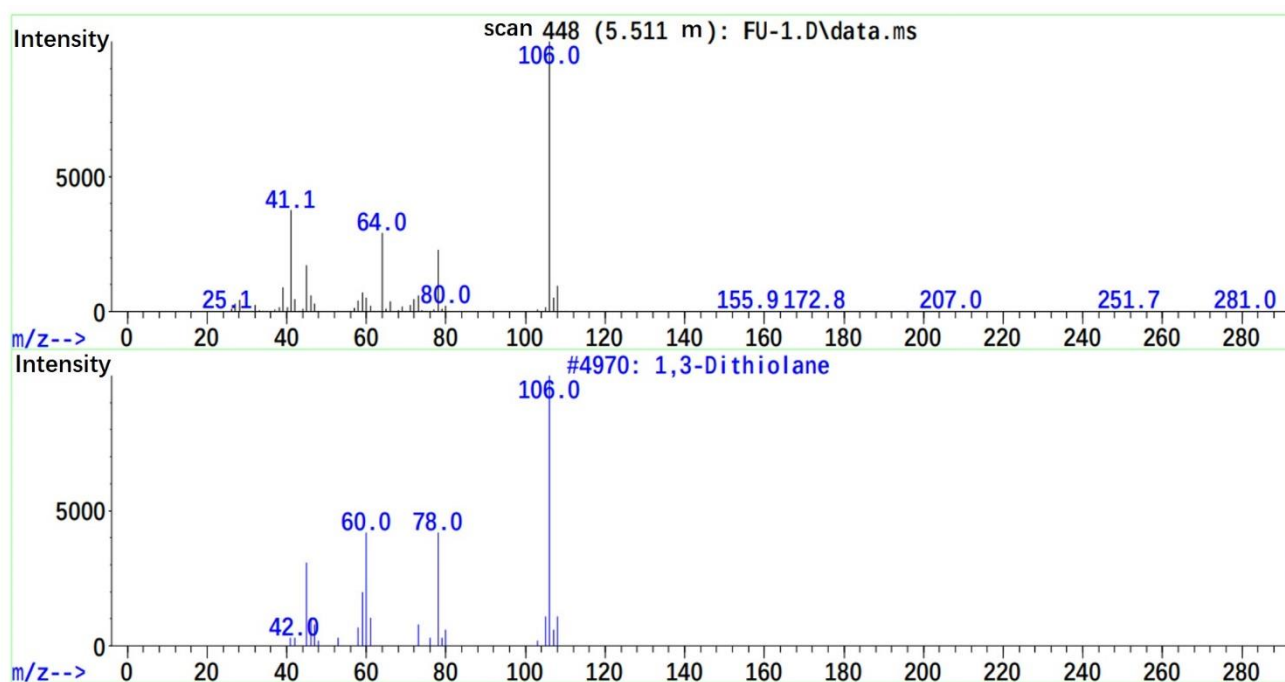


Disulfide **7-12** were isolated by flash column chromatography following the above **General Procedure III-IV** and confirmed by GC-MS, HRMS and NMR (^1H and ^{13}C).

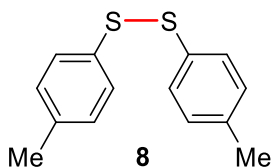
1,2-Dithiolane (7)



The purification of compound **7** always led to polymeric material⁷⁰, so the reaction mixture of **3w** was passed through millipore filter (0.45 μm) directly after 5 min at rt and analyzed by GC-MS directly. Mass spectrum and retrieval results were listed below respectively, and compound **7** ($m/z = 106.0$) was identified as 1,3-dithiolane.

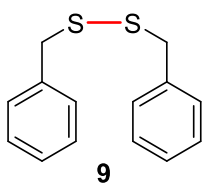


Bis(4-methylphenyl) disulfide (**8**)



8⁷¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:20) as a light-yellow oil (207 mg, 84%) following the deprotection of **3ag** (1 mmol). ¹H-NMR (400 MHz, DMSO) δ : 7.37 (d, J = 8.3 Hz, 4H), 7.16 (d, J = 8.0 Hz, 4H), 2.27 (s, 6H). ¹³C-NMR (100 MHz, DMSO) δ : 137.4, 132.6, 130.0, 128.1, 20.5. IR 1591.2, 1481.9, 1397.8, 1301.6, 1177.9, 1112.6, 1072.8, 1010.0, 794.3, 478.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₄H₁₅S₂ [M+H]⁺ 247.0610; found 247.0609.

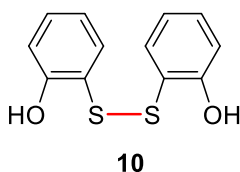
Dibenzyl disulfide (**9**)



9⁷¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:40 to 1:20) as a light-yellow oil (123 mg, 50%) following the deprotection of **3ad** (1 mmol). ¹H-NMR (400 MHz, CDCl₃) δ : 7.24–7.11 (m, 10H), 3.49 (s, 4H). ¹³C-NMR (100 MHz, CDCl₃) δ : 137.5, 129.5, 128.6, 127.5, 43.3. IR 3024.6, 2918.8, 1594.9, 1488.0, 1449.7, 1407.6, 1222.8, 1192.7, 1066.2, 1023.4, 758.7, 465.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd.

for C₁₄H₁₅S₂ [M+H]⁺ 247.0610; found 247.0604.

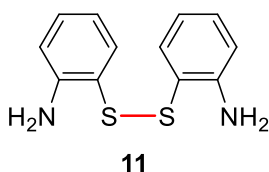
Bis(2-hydroxyphenyl) disulfide (**10**)



10⁷² was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 1:1) as a light-yellow oil (173 mg, 69%) following the deprotection of **3ar** (1 mmol). ¹H-NMR (400 MHz, CDCl₃) δ : 7.37–7.32 (m, 2H), 7.22 (dd, J = 7.8, 1.7 Hz, 2H), 7.00 (dd, J = 8.2, 1.3 Hz, 2H), 6.81 (td, J = 7.6, 1.3 Hz, 2H), 6.29 (s, 2H). ¹³C-NMR (100 MHz, CDCl₃) δ : 157.0, 136.3, 133.3, 121.2, 120.1, 115.9. IR 3449.7, 3399.5,

1566.5, 1459.4, 1331.8, 1291.5, 1236.4, 1174.4, 1021.1, 829.1, 749.4, 496.4, 428.1 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₁₁O₂S₂ [M+H]⁺ 251.0195; found 251.0189.

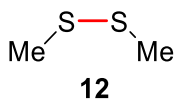
Bis(2-aminophenyl) disulfide (**11**)



11⁷¹ was purified by flash column chromatography (ethyl acetate/hexane = 1:5 to 1:1) as a light-yellow oil (109 mg, 44%) following the deprotection of **3as** (1 mmol). ¹H-NMR (400 MHz, CDCl₃) δ : 7.18–7.13 (m, 4H), 6.70 (dd, J = 8.4, 1.2 Hz, 2H), 6.57 (td, J = 7.4, 1.3 Hz, 2H), 4.22 (s, 4H). ¹³C-NMR (100 MHz, CDCl₃) δ : 148.8, 137.0, 131.7, 118.9, 118.4, 115.4. IR 3375.1, 3061.7, 1611.8, 1580.3,

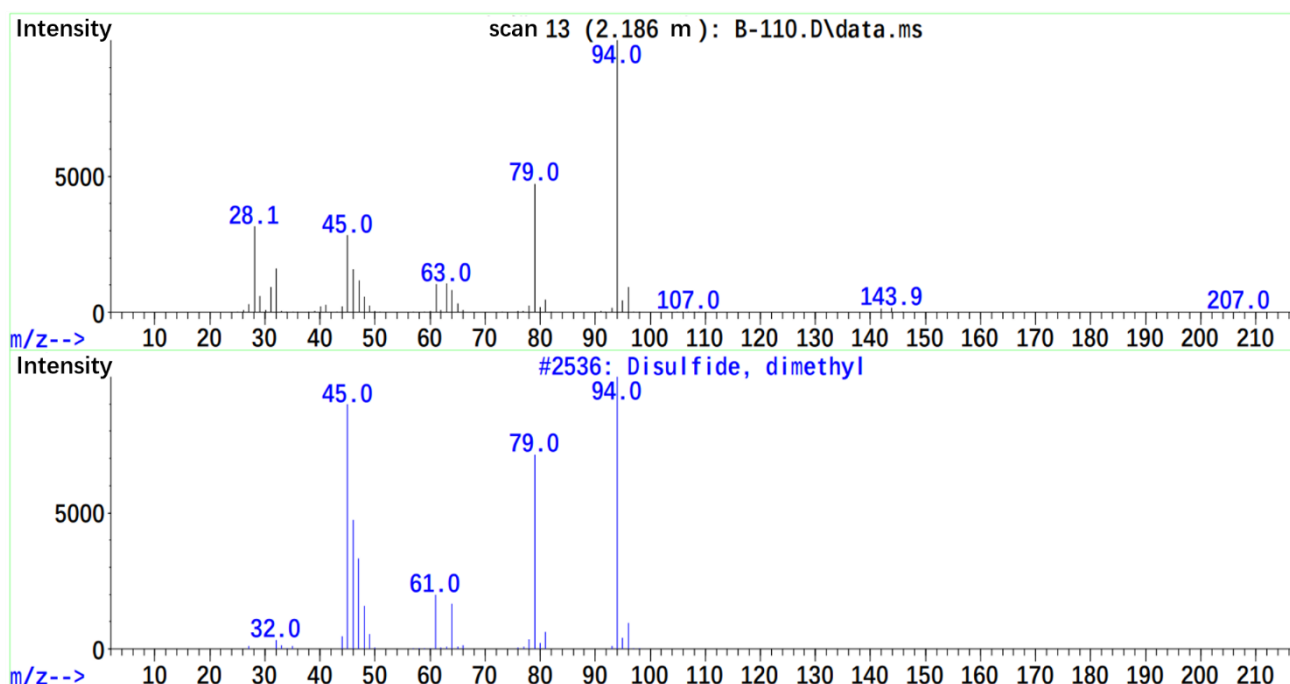
1468.2, 1441.1, 1299.5, 1242.9, 1151.1, 1020.8, 856.4, 745.5, 451.8 cm⁻¹; HRMS (ESI⁺) (m/z) calcd. for C₁₂H₁₃N₂S₂ [M+H]⁺ 249.0515; found 249.0525.

Dimethyl disulfide (**12**)



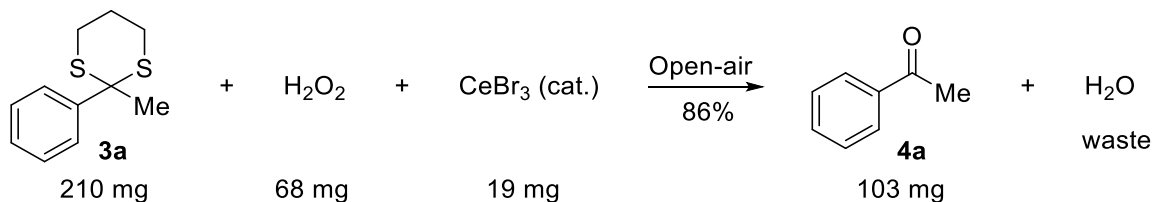
Compound **12** was too volatile to be isolated, so the reaction mixture of **5j** was passed through millipore filter (0.45 μ m) directly after 10 min and analyzed by GC-MS directly.

Mass spectrum and retrieval results were listed below respectively, and compound **12** ($m/z = 94.0$) was identified as dimethyl disulfide.



11. Green chemistry metrics analysis.

E-Factor for **3a** and **3p** using $\text{H}_2\text{O}_2/\text{CeBr}_3$:

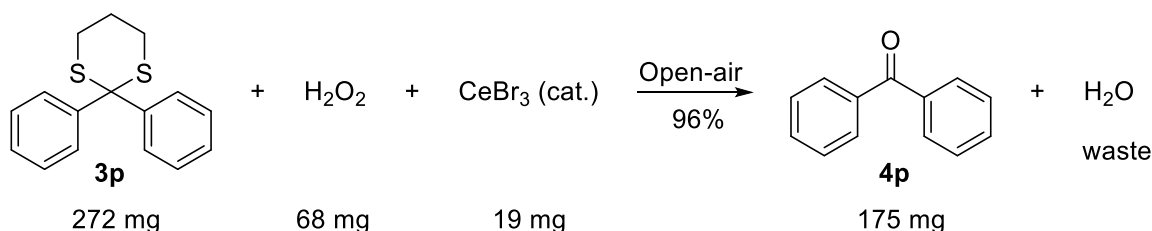


Total amount of reactants: 210 mg + 68 mg + 19 mg = 297 mg

Amount of final product: 103 mg

Amount of waste: 297 mg – 103 mg = 194 mg

E-Factor = Amount of waste/Amount of product = 194/103 = 1.88



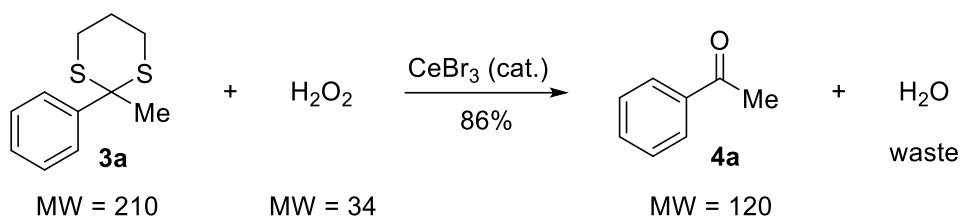
Total amount of reactants: 272 mg + 68 mg + 19 mg = 359 mg

Amount of final product: 175 mg

Amount of waste: 359 mg – 175 mg = 184 mg

E-Factor = Amount of waste/Amount of product = 184/175 = 1.05

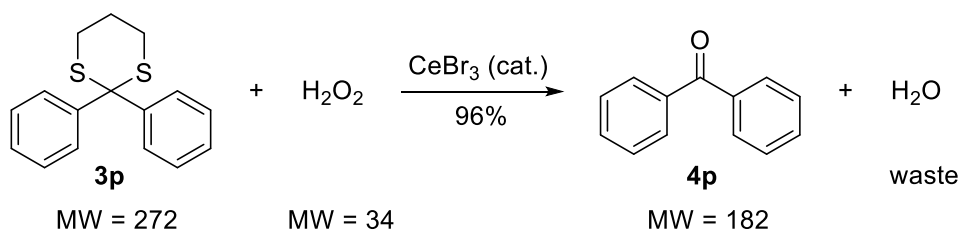
AE for 3a and 3p using H₂O₂/CeBr₃:



Molecular weight of product: 120

Sum of molecular weight of reagent: 210 + 34 = 244

Atom economy = Molecular weight of product/Sum of molecular weight of reagent = 120/244 = 49.2%

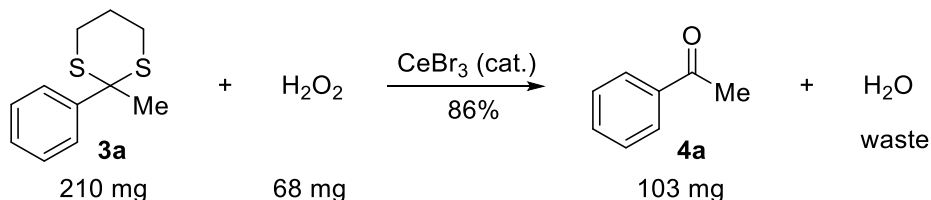


Molecular weight of product: 182

Sum of molecular weight of reagent: 272 + 34 = 306

Atom economy = Molecular weight of product/Sum of molecular weight of reagent = 182/306 = 59.5%

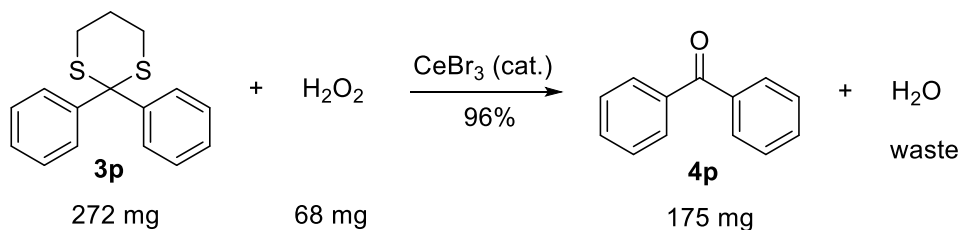
RME for 3a and 3p using H₂O₂/CeBr₃:



Mass of product: 103 mg

Total mass of reagent: 210 mg + 68 mg = 278 mg

RME = Mass of product/Total mass of reagent = 103/278 = 37.1%

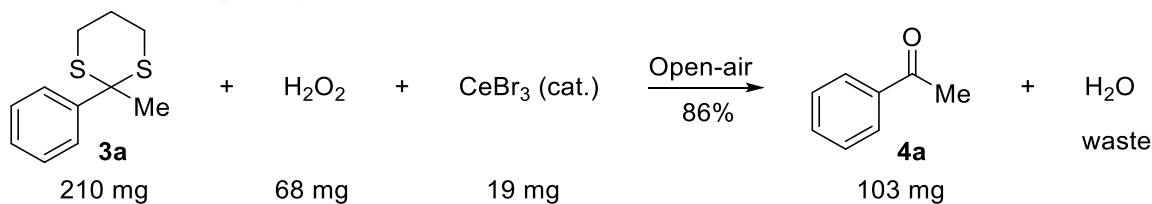


Mass of product: 175 mg

Total mass of reagent: 272 mg + 68 mg = 340 mg

RME = Mass of product/Total mass of reagent = 175/340 = 51.5%

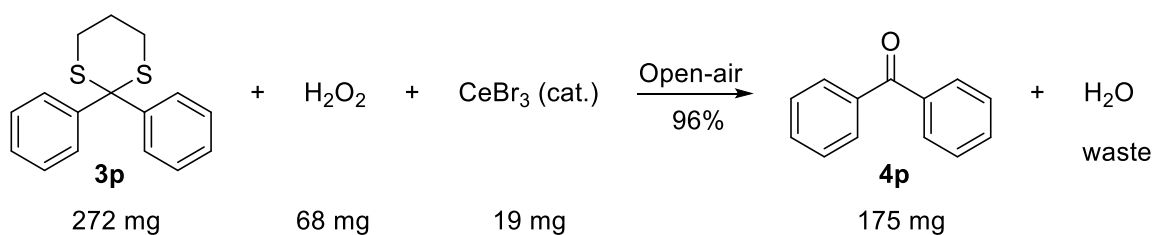
PMI for **3a** and **3p** using H₂O₂/CeBr₃:



Total mass in process: 210 mg + 68 mg + 19 mg = 297 mg

Mass of product: 103 mg

PMI = Total mass in process/Mass of product = 297/103 = 2.88



Total mass in process: 272 mg + 68 mg + 19 mg = 359 mg

Mass of product: 175 mg

PMI = Total mass in process/Mass of product = 359/175 = 2.05

Green chemistry metrics for **3a** and **3p** using other protocols (SDS-I₂/H₂O₂⁷³, TBHP⁷⁴, NBS⁷⁵, IBX⁷⁶, DDQ⁷⁷ and Oxone⁷⁸) were calculated according to the published literatures, and the results were listed below:

Reagent	Average							3p					
	CeBr ₃	IBX	DDQ	TBHP	NBS	Oxone	SDS, I ₂ /H ₂ O ₂	CeBr ₃	IBX	DDQ	TBHP	NBS	SDS, I ₂ /H ₂ O ₂
E-Factor	1.47	3.90	4.11	2.16	2.65	14.2	2.05	1.05	3.09	3.11	1.70	2.07	1.63
AE	54.4%	28.8%	32.0%	45.2%	35.7%	23.2%	54.4%	59.5%	33.0%	36.5%	50.3%	40.5%	59.5%
RME	44.3%	21.0%	20.4%	32.4%	28.2%	6.6%	39.7%	51.5%	24.5%	24.4%	37.0%	32.6%	44.6%
PMI	2.47	4.90	5.11	3.16	3.65	15.2	3.05	2.05	4.09	4.11	2.70	3.07	2.63

Reagent	3a						
	CeBr ₃	IBX	DDQ	TBHP	NBS	Oxone	SDS, I ₂ /H ₂ O ₂
E-Factor	1.88	4.70	5.11	2.61	3.22	14.2	2.47
AE	49.2%	24.5%	27.5%	40.0%	30.9%	23.2%	49.2%
RME	37.1%	17.5%	16.4%	27.7%	23.7%	6.6%	34.70%
PMI	2.88	5.70	6.11	3.61	4.22	15.2	3.47

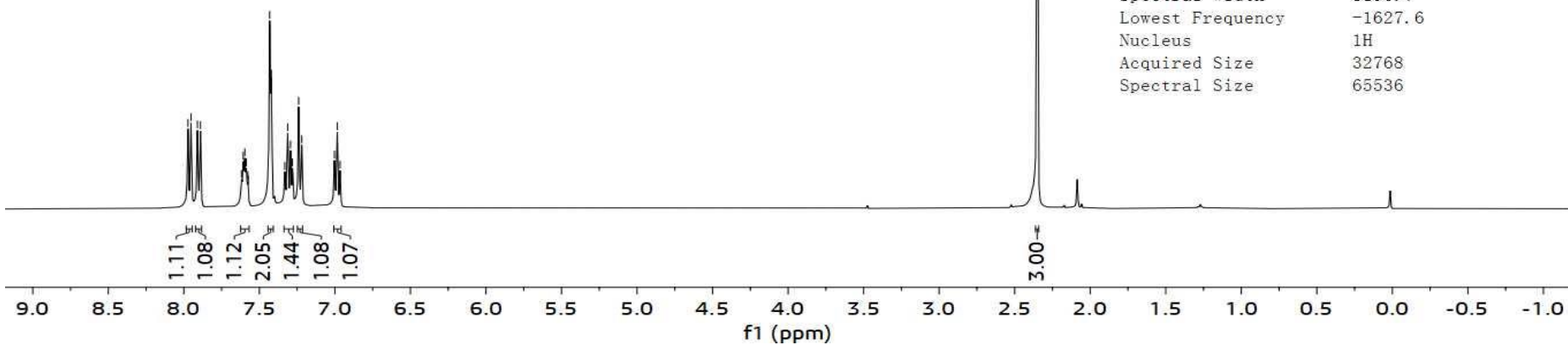
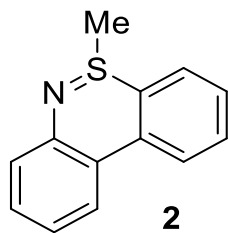
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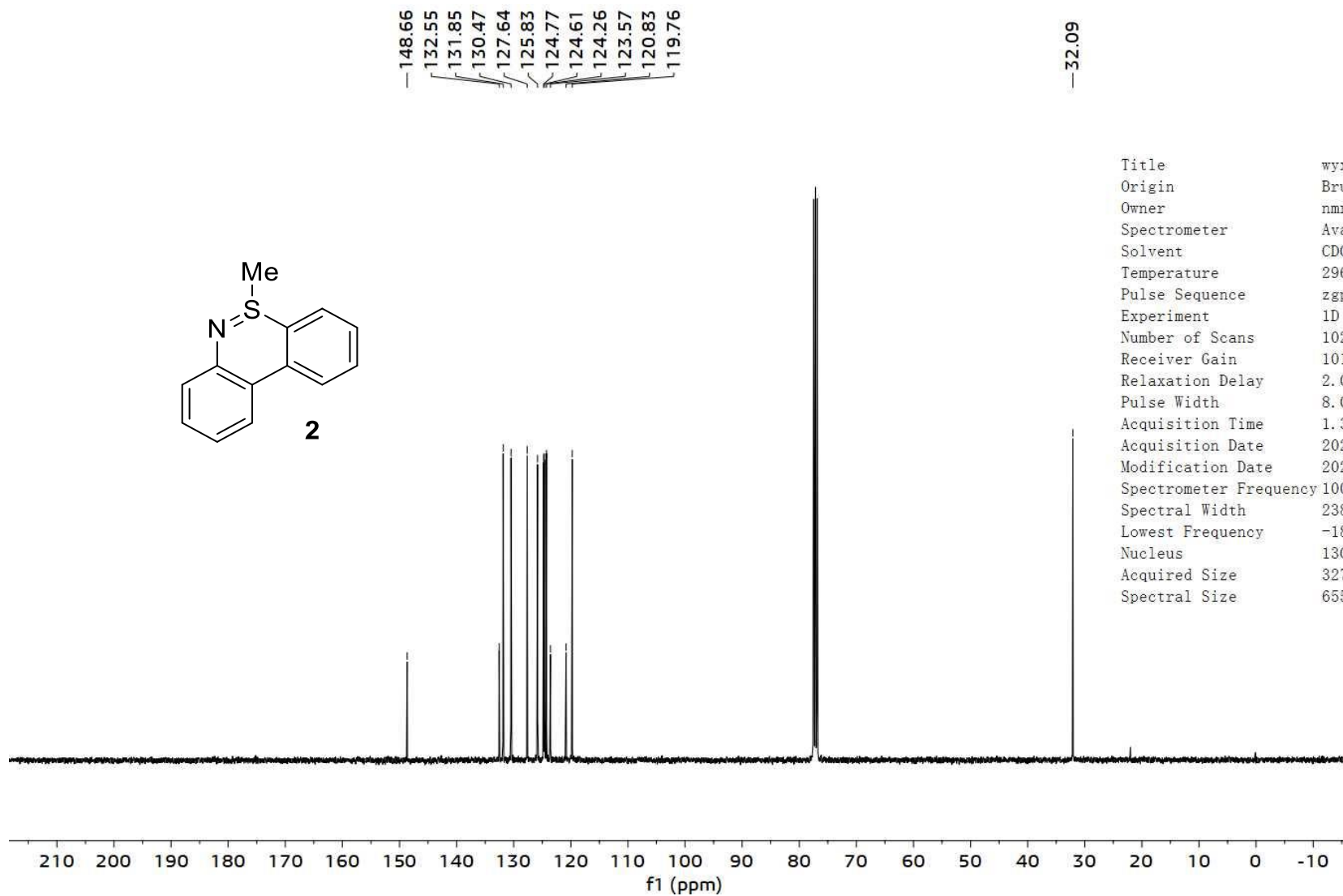
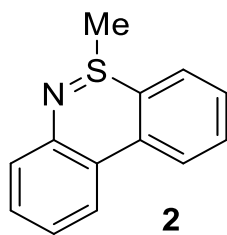
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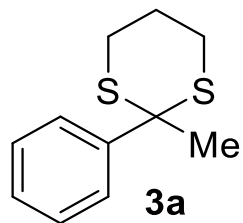
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Experiment	1D
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Relaxation Delay	1.0000
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Nucleus	1H
Acquired Size	32768
Spectral Size	65536



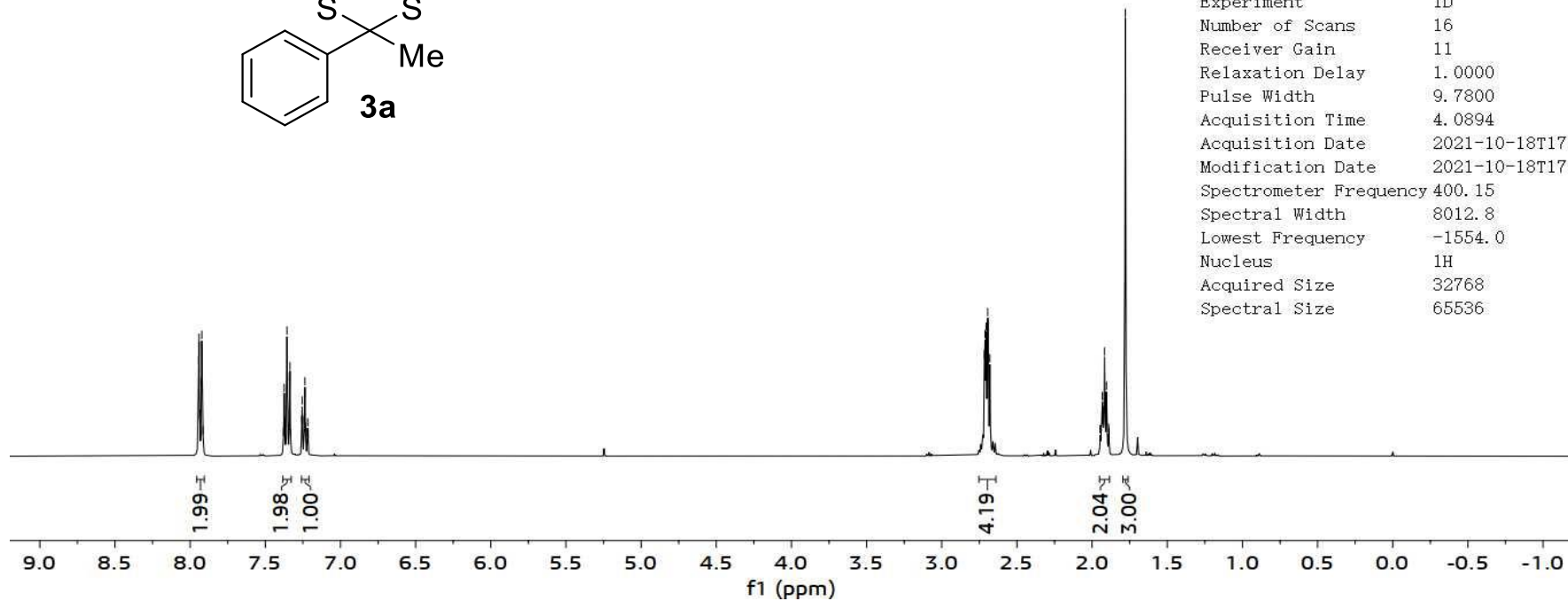
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Solvent	CDC13
Temperature	296.2
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
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Lowest Frequency	-1835.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

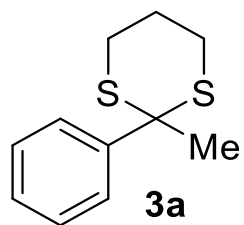
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Title	WYX-2-YL. 1. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	11
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
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Lowest Frequency	-1554.0
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



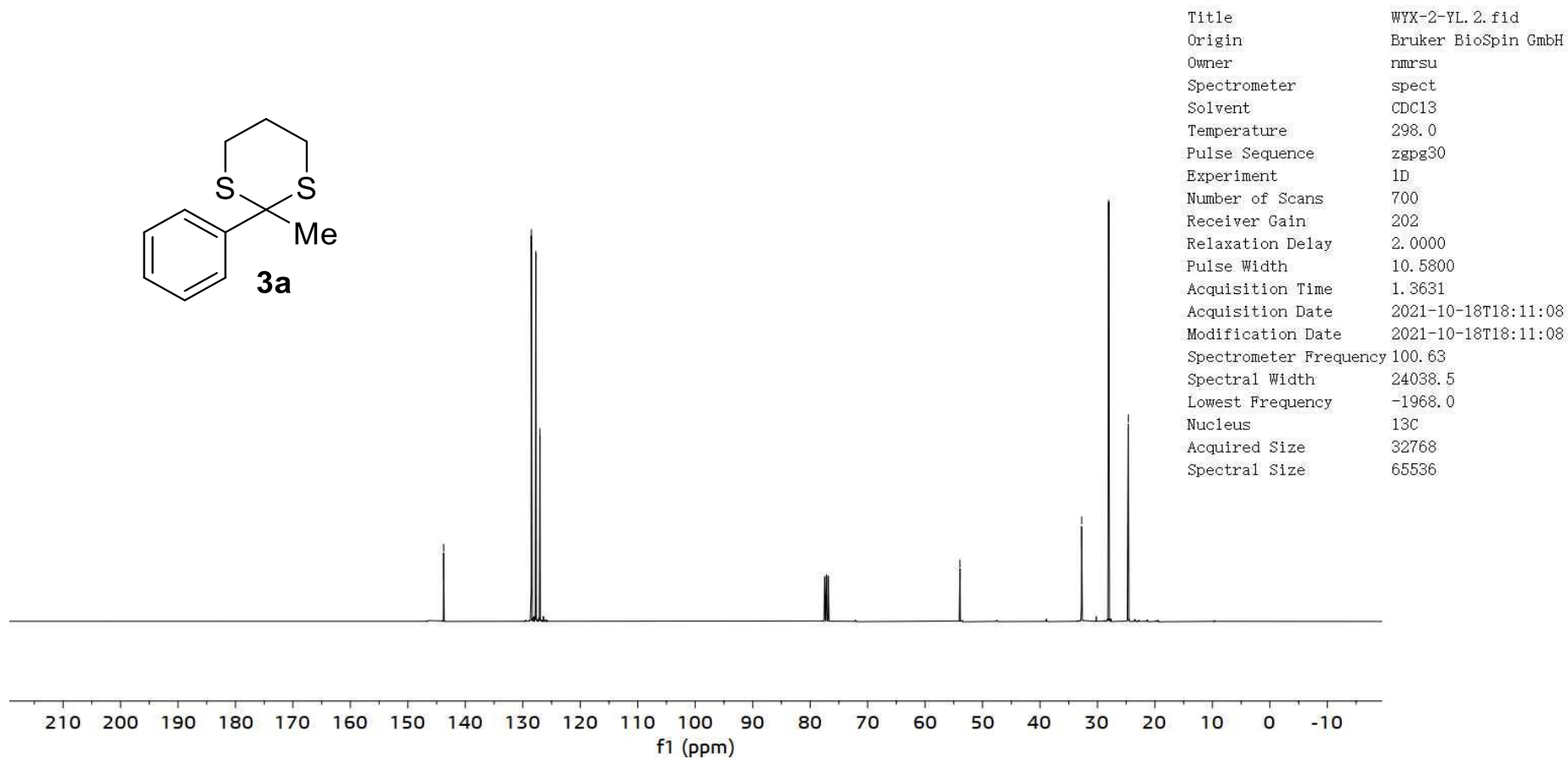


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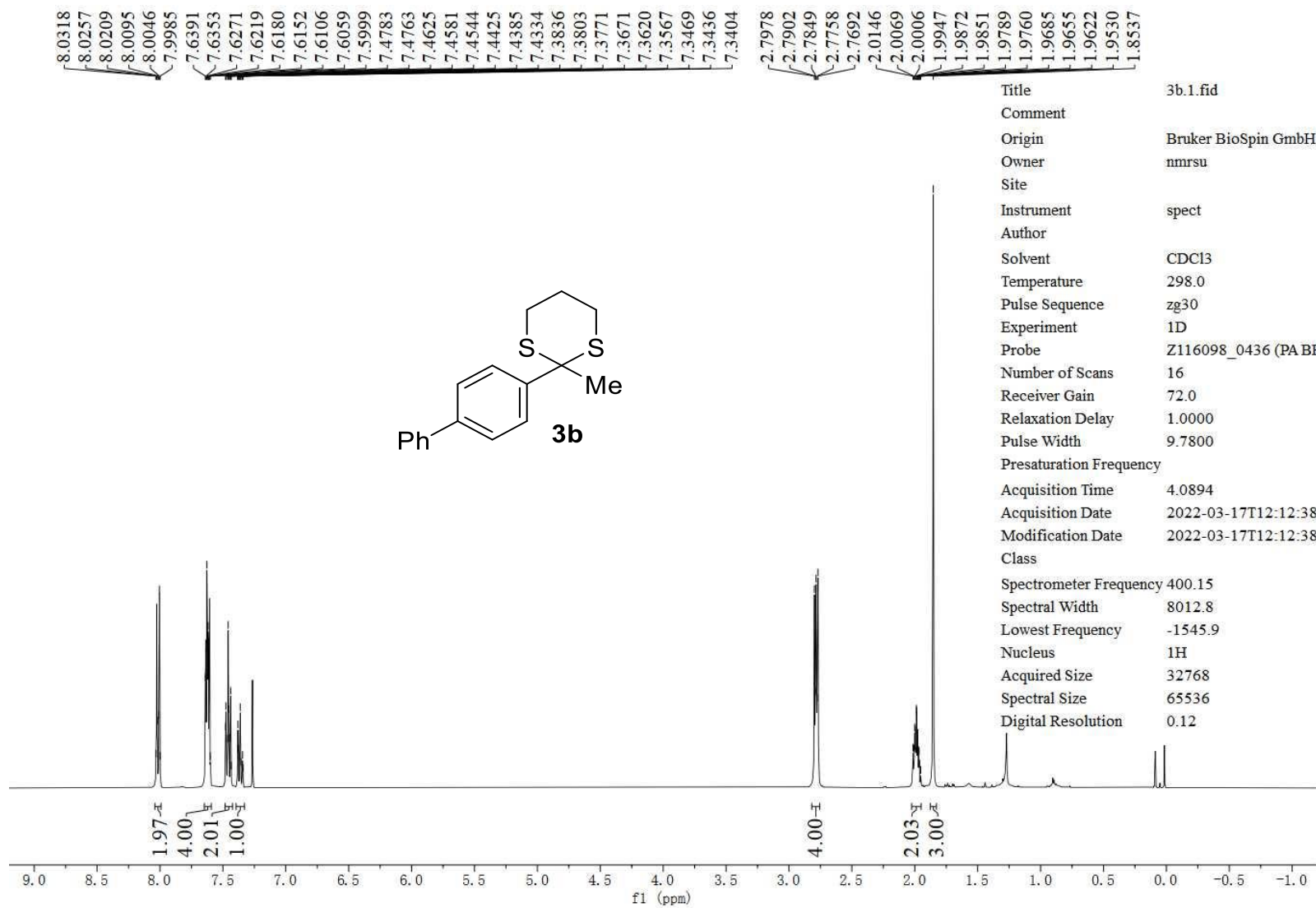
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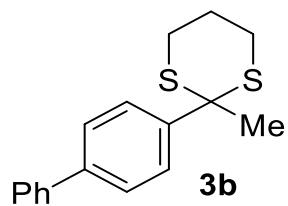


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Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-18T18:11:08
Modification Date	2021-10-18T18:11:08
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1968.0
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



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Title	3b.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl ₃
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	72.0
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Pulse Width	9.7800
Presaturation Frequency	
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Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12

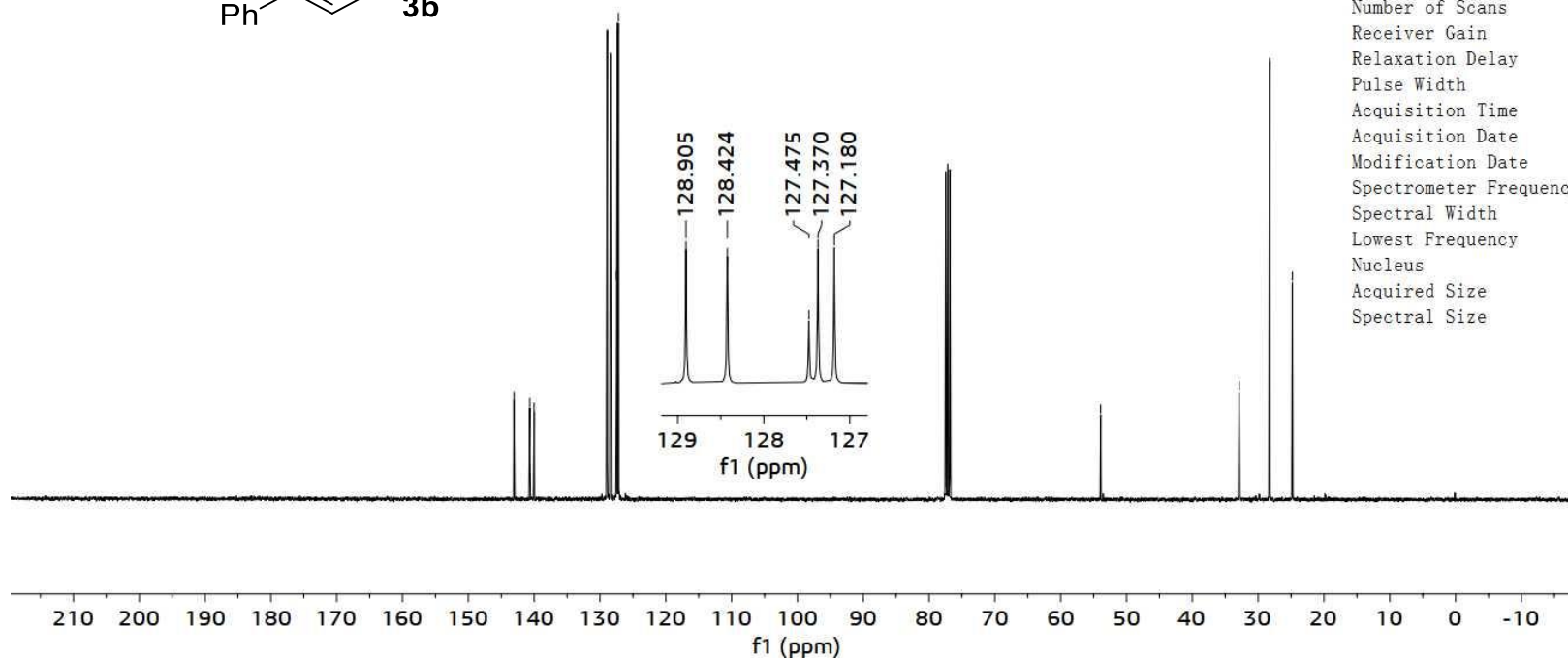


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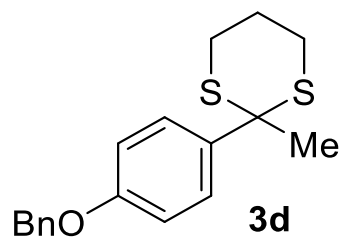
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Owner	nMrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-16T02:00:49
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Spectral Width	24038.5
Lowest Frequency	-1948.9
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



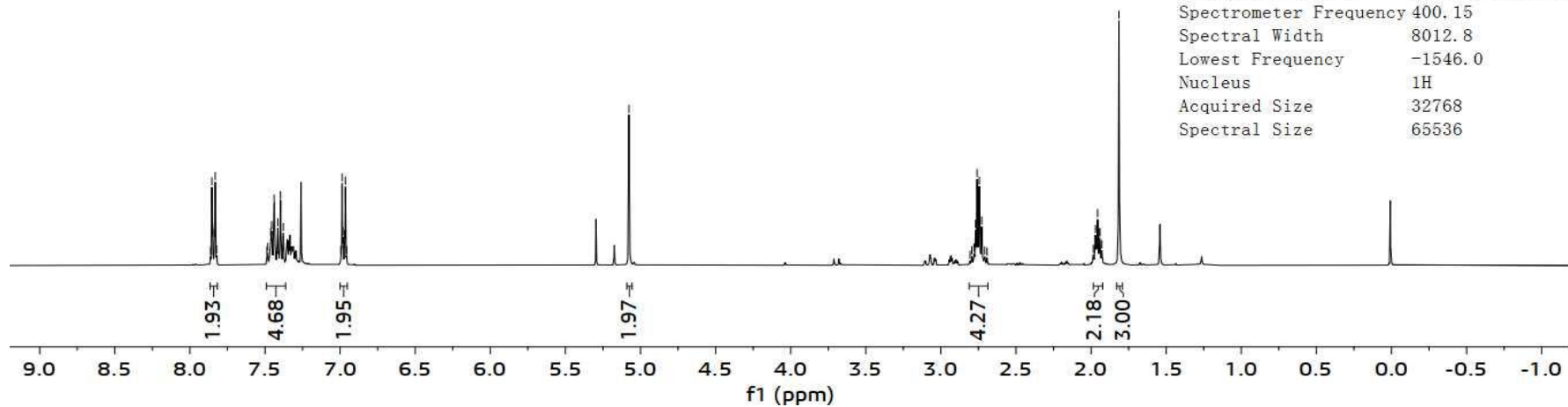
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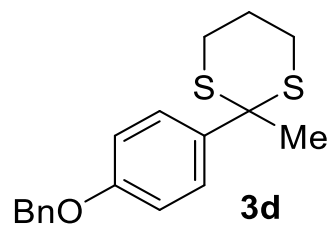
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Title	WYX-16-YL-dian.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	89
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
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Lowest Frequency	-1546.0
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536





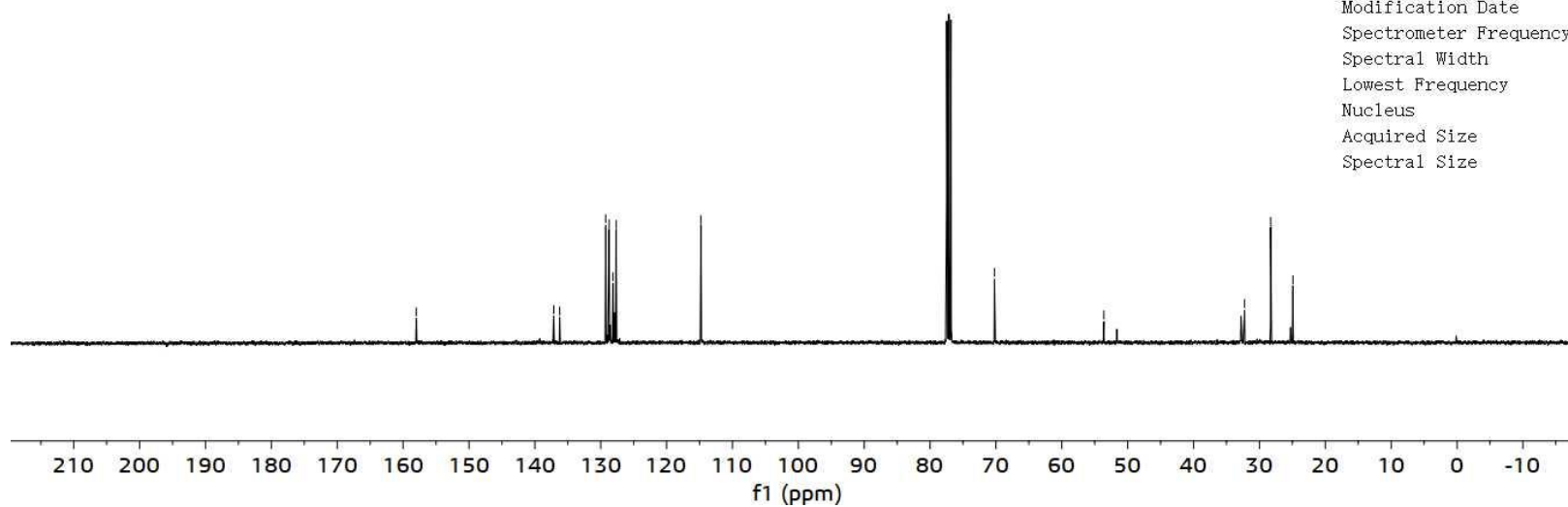
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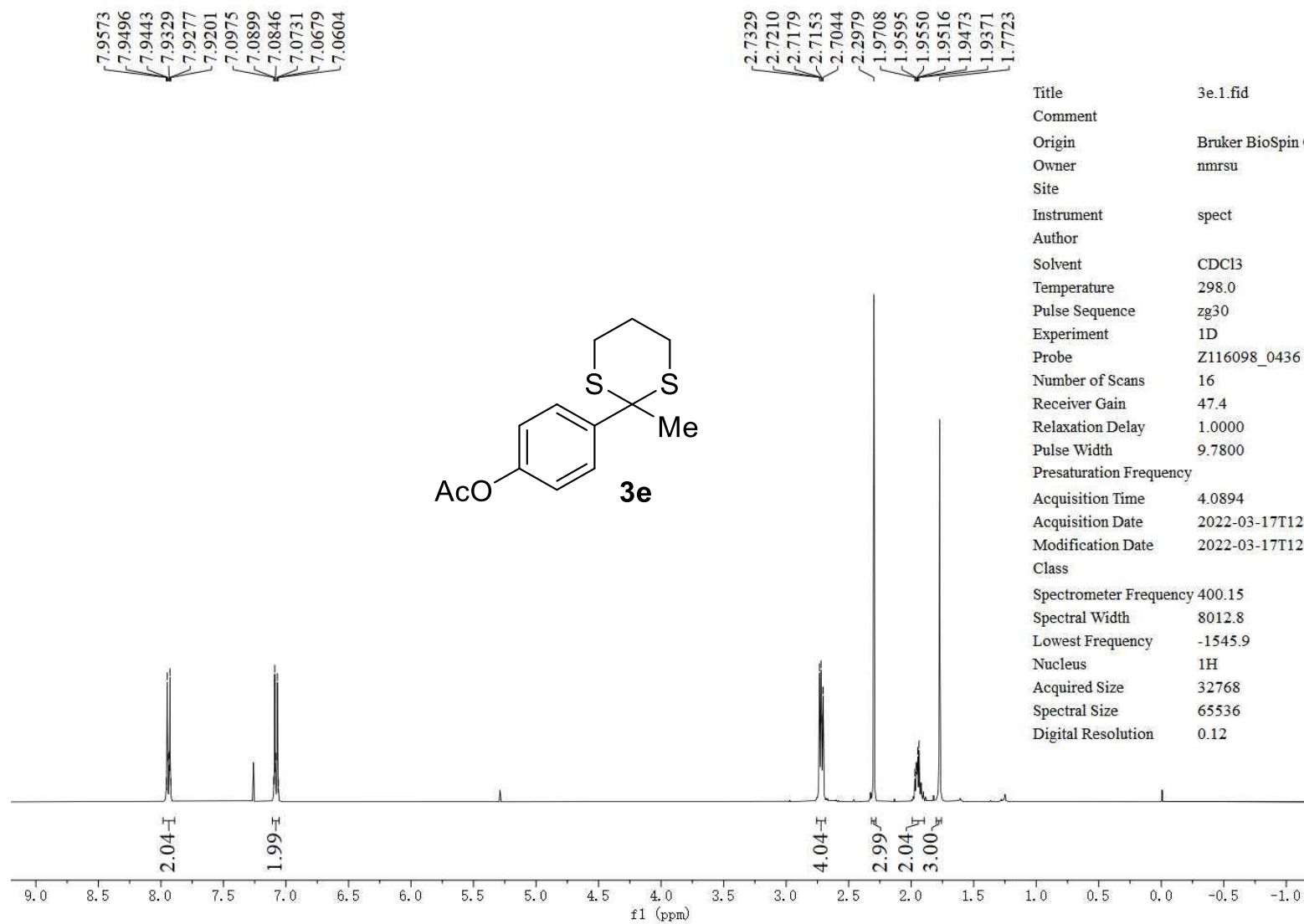
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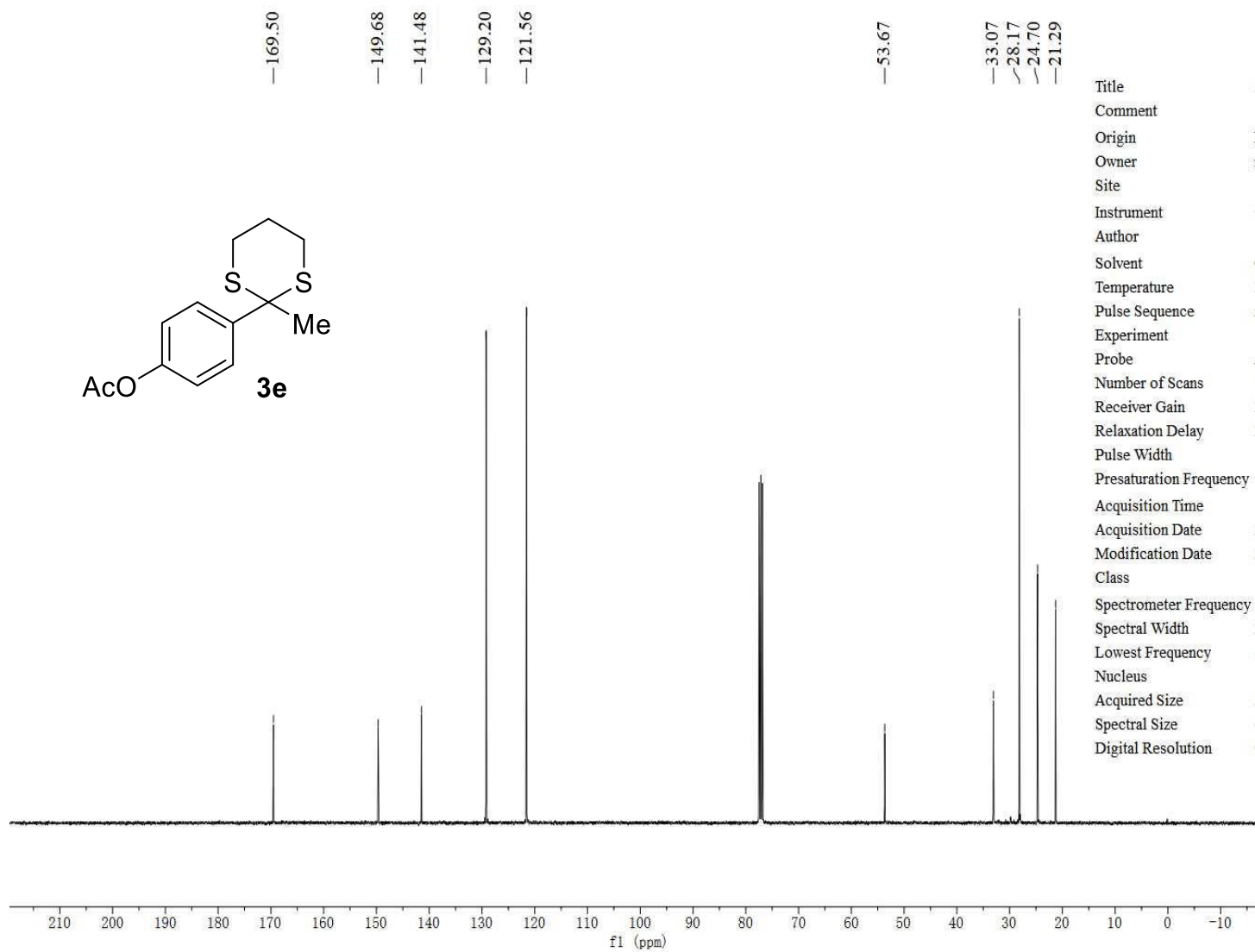
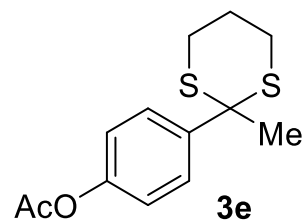
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Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
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Lowest Frequency	-1944.5
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536





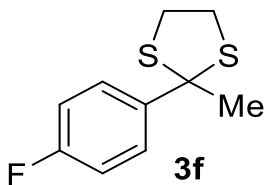
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Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl ₃
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	47.4
Relaxation Delay	1.0000
Pulse Width	9.7800
Presaturation Frequency	
Acquisition Time	4.0894
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Class	
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Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12



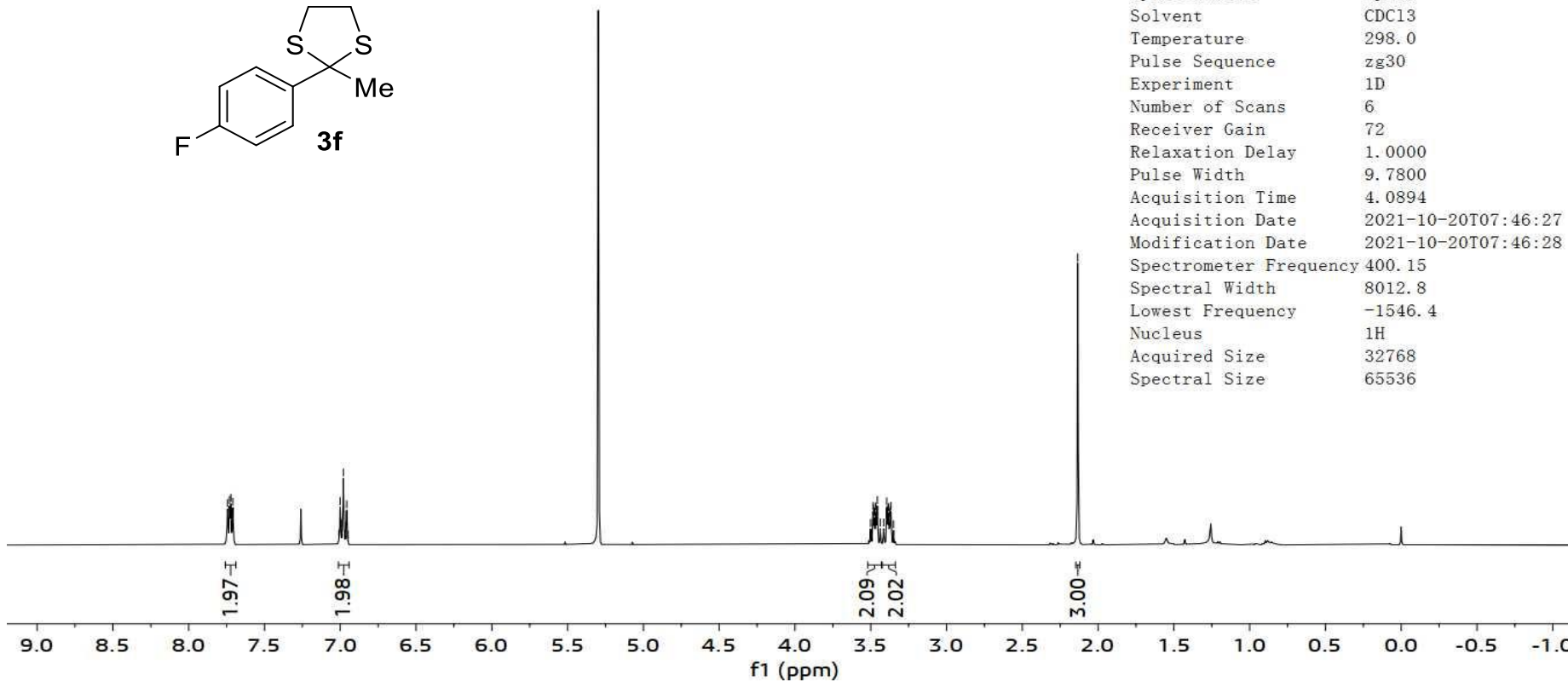
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Site	
Instrument	spect
Author	
Solvent	CDCl3
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Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
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Relaxation Delay	2.0000
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Spectral Size	65536
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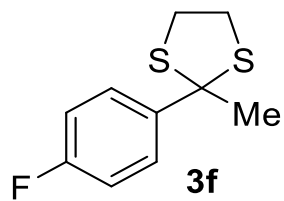
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Owner	nmrsu
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Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
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Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
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Modification Date	2021-10-20T07:46:28
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Lowest Frequency	-1546.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536





163.12
160.67

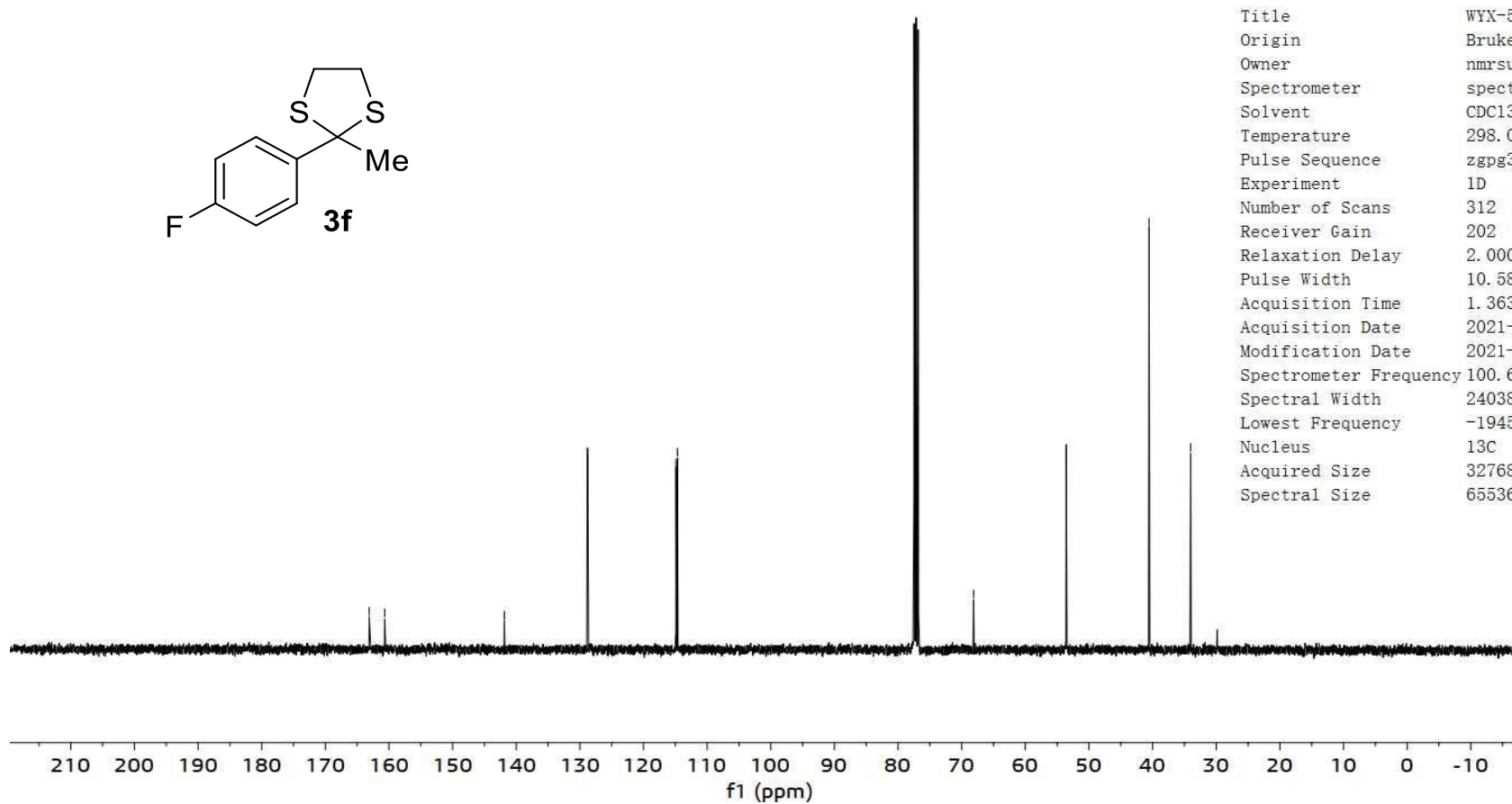
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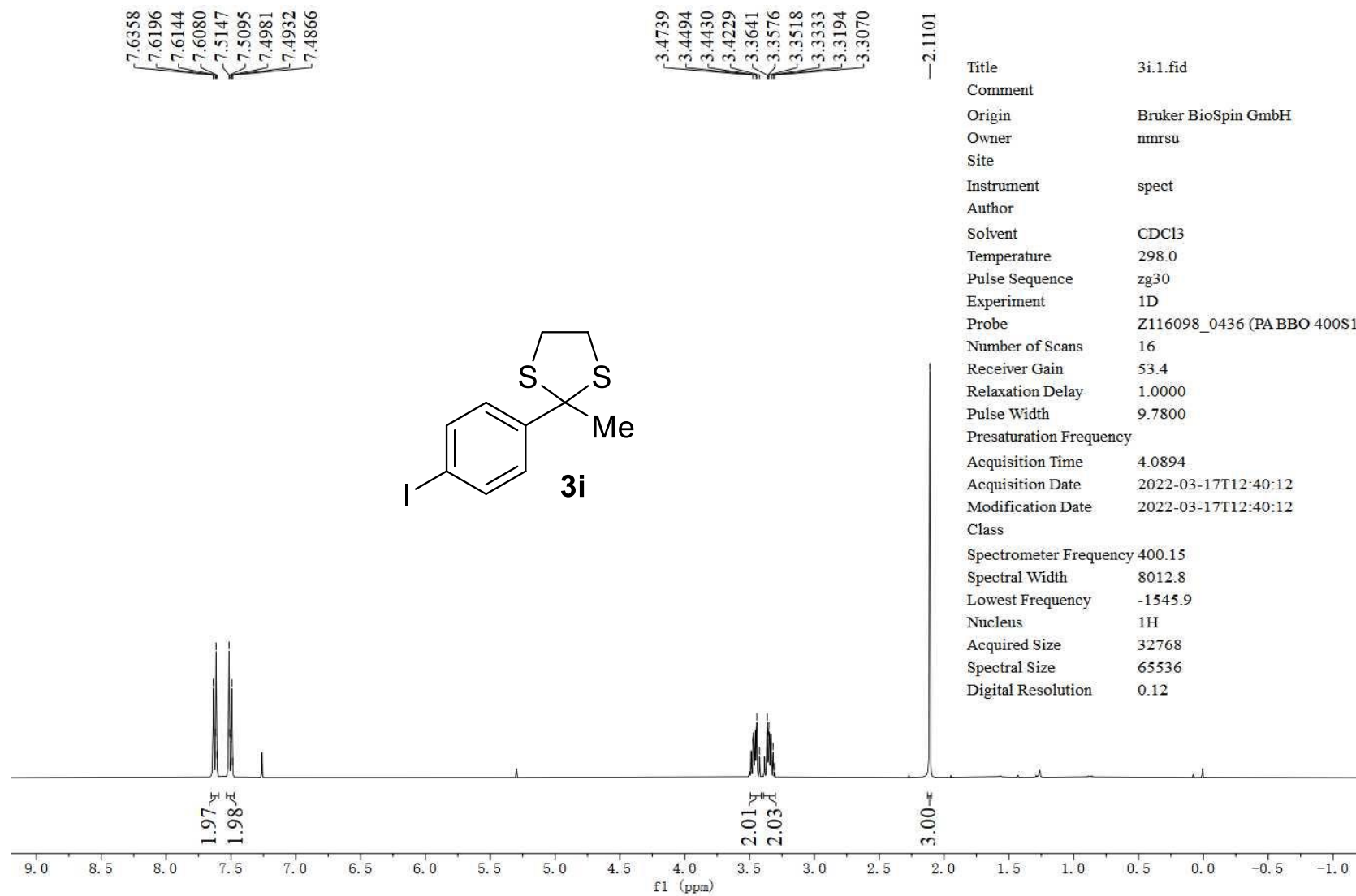
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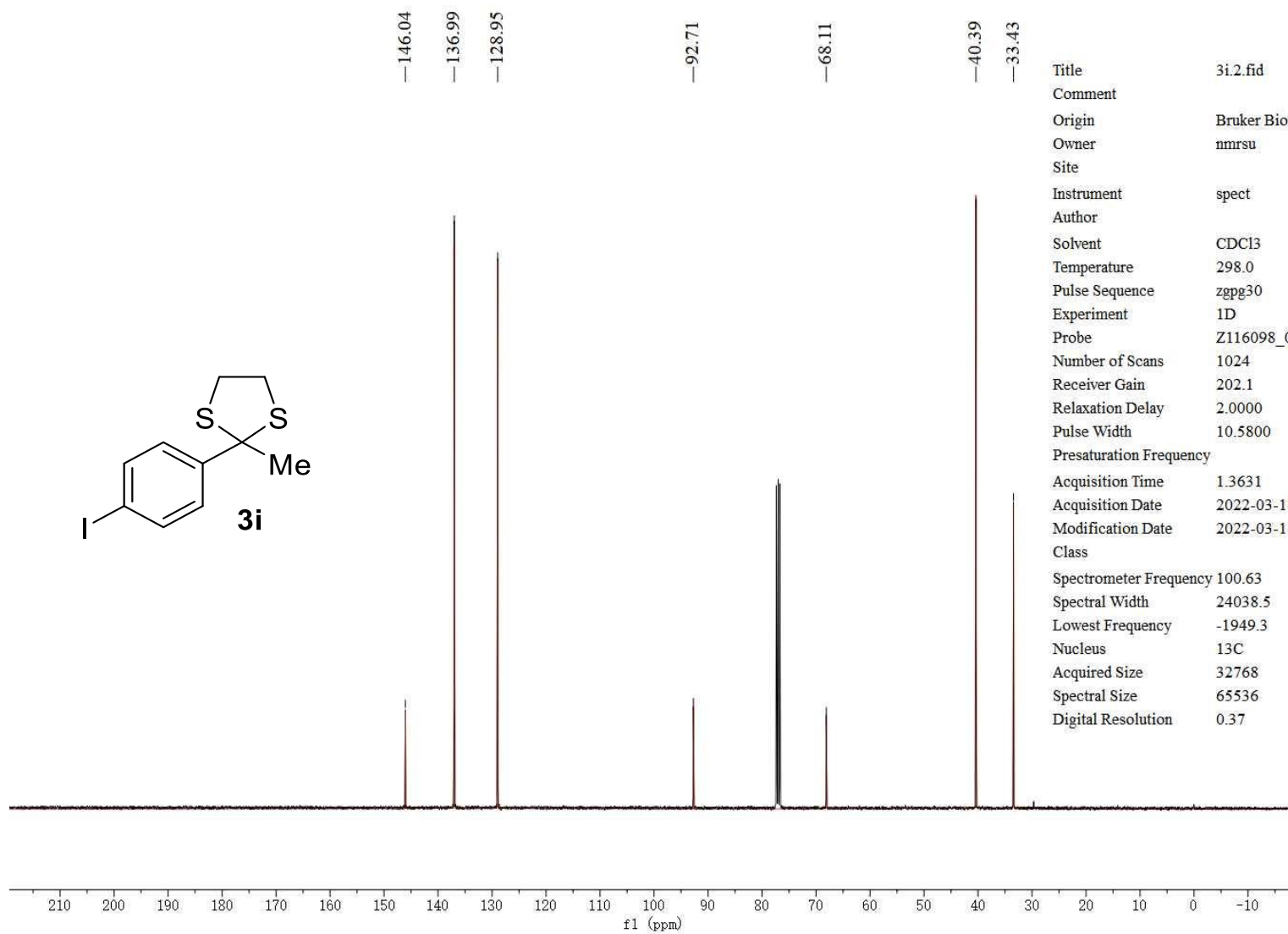
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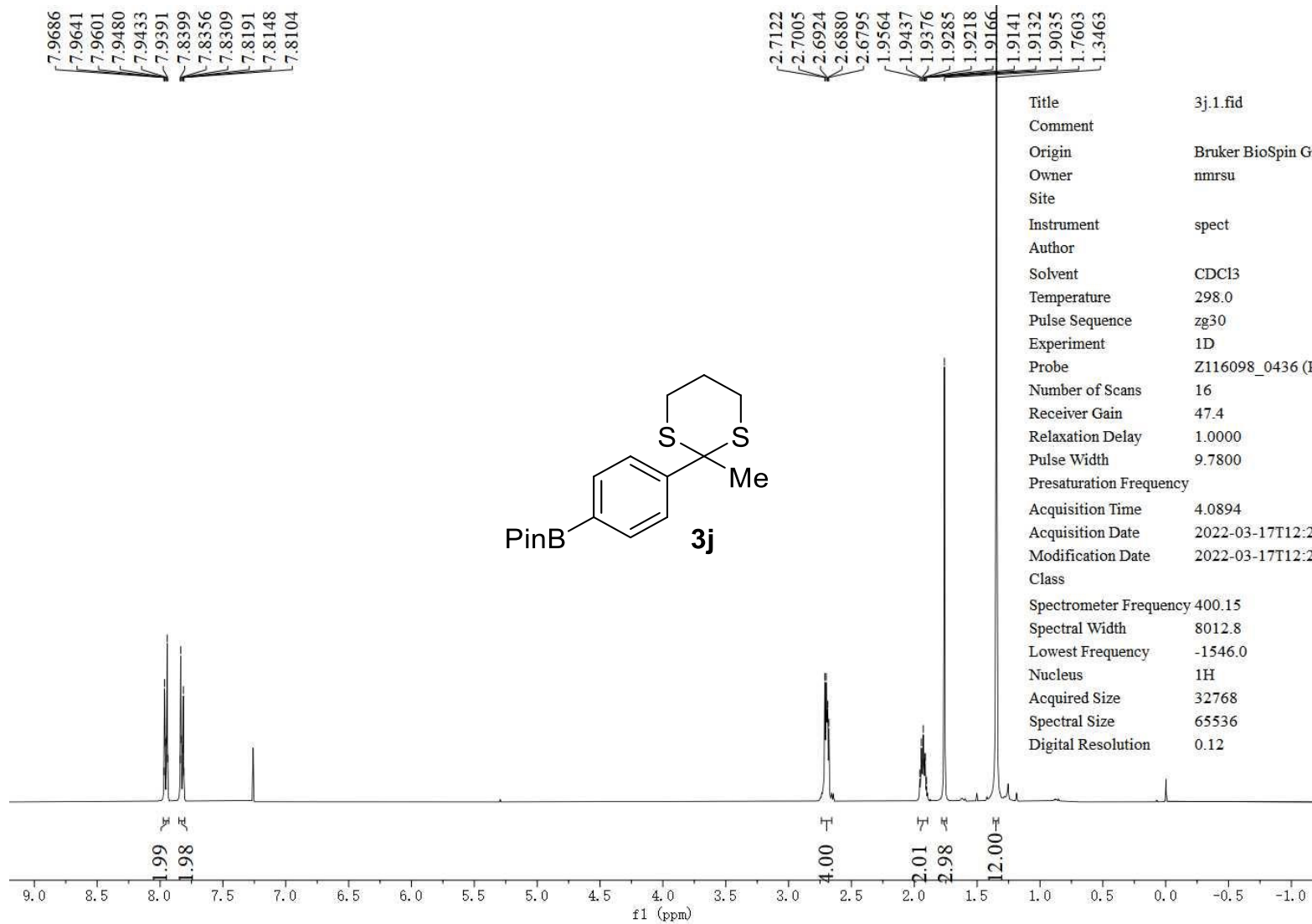
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Experiment	1D
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Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-20T08:05:27
Modification Date	2021-10-20T08:05:28
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1945.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Title	3i.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	53.4
Relaxation Delay	1.0000
Pulse Width	9.7800
Presaturation Frequency	
Acquisition Time	4.0894
Acquisition Date	2022-03-17T12:40:12
Modification Date	2022-03-17T12:40:12
Class	
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.9
Nucleus	1H
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12



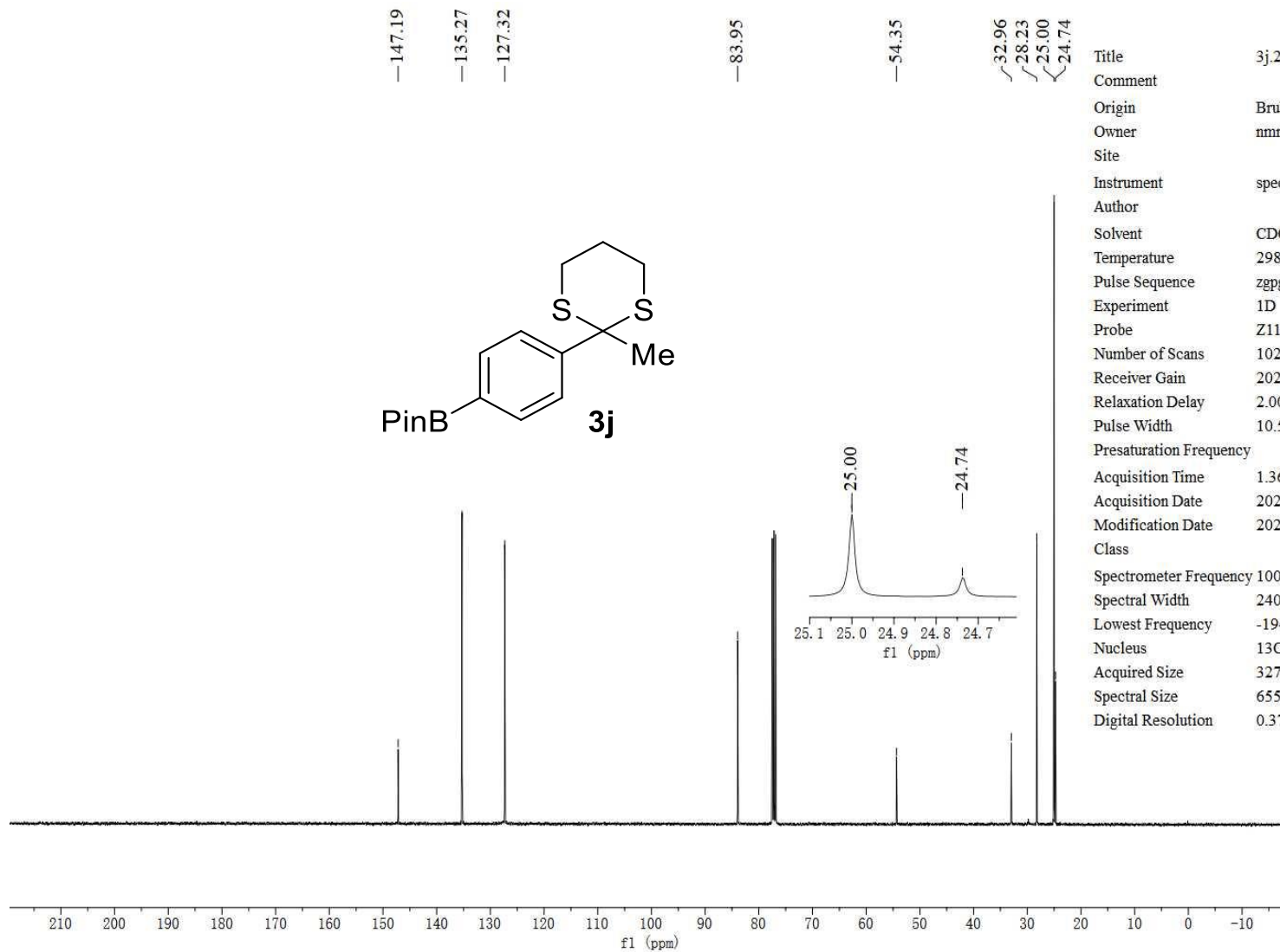
Title	3i.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrstu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PABBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-18T06:31:03
Modification Date	2022-03-18T06:31:02
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1949.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37



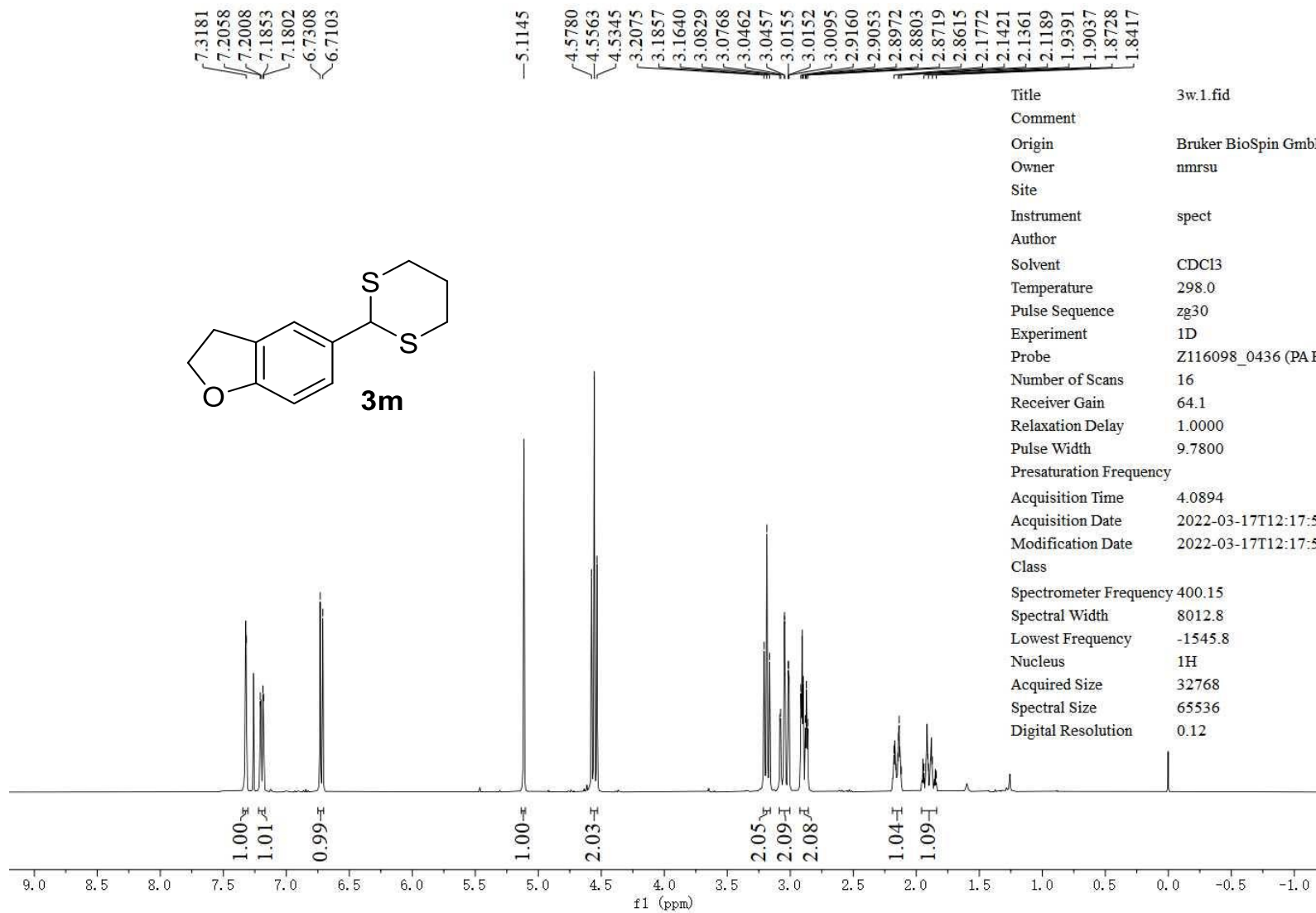
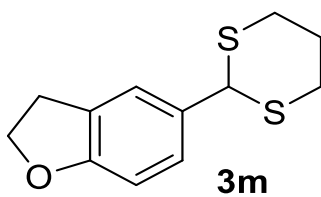
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7.9433
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7.8399
7.8356
7.8309
7.8191
7.8148
7.8104

2.7122
2.7005
2.6924
2.6880
2.6795
1.9564
1.9437
1.9376
1.9285
1.9218
1.9166
1.9141
1.9132
1.9035
1.7603
1.3463

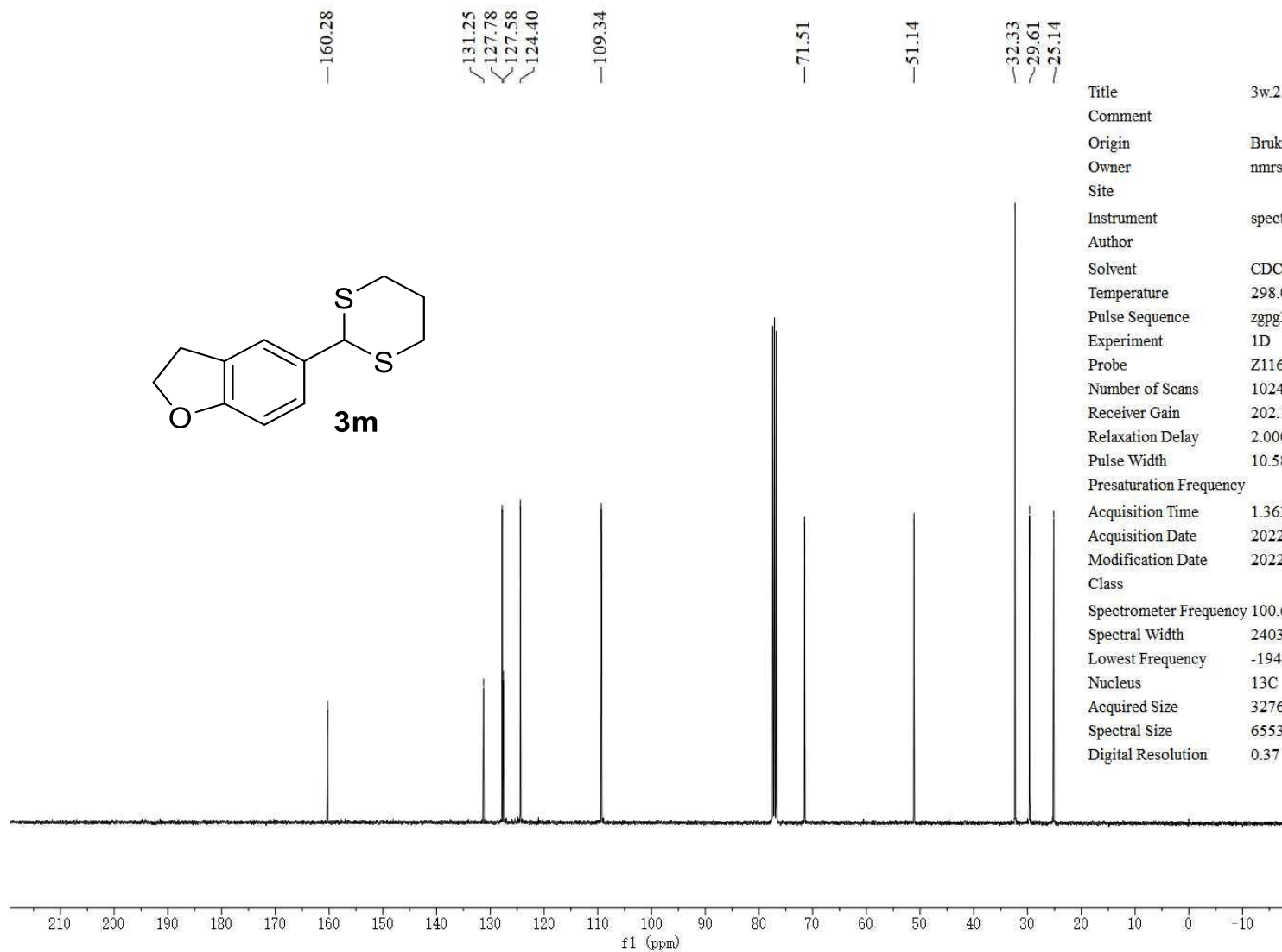
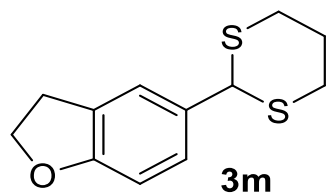
Title	3j.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	47.4
Relaxation Delay	1.0000
Pulse Width	9.7800
Presaturation Frequency	
Acquisition Time	4.0894
Acquisition Date	2022-03-17T12:23:33
Modification Date	2022-03-17T12:23:32
Class	
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1546.0
Nucleus	1H
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12



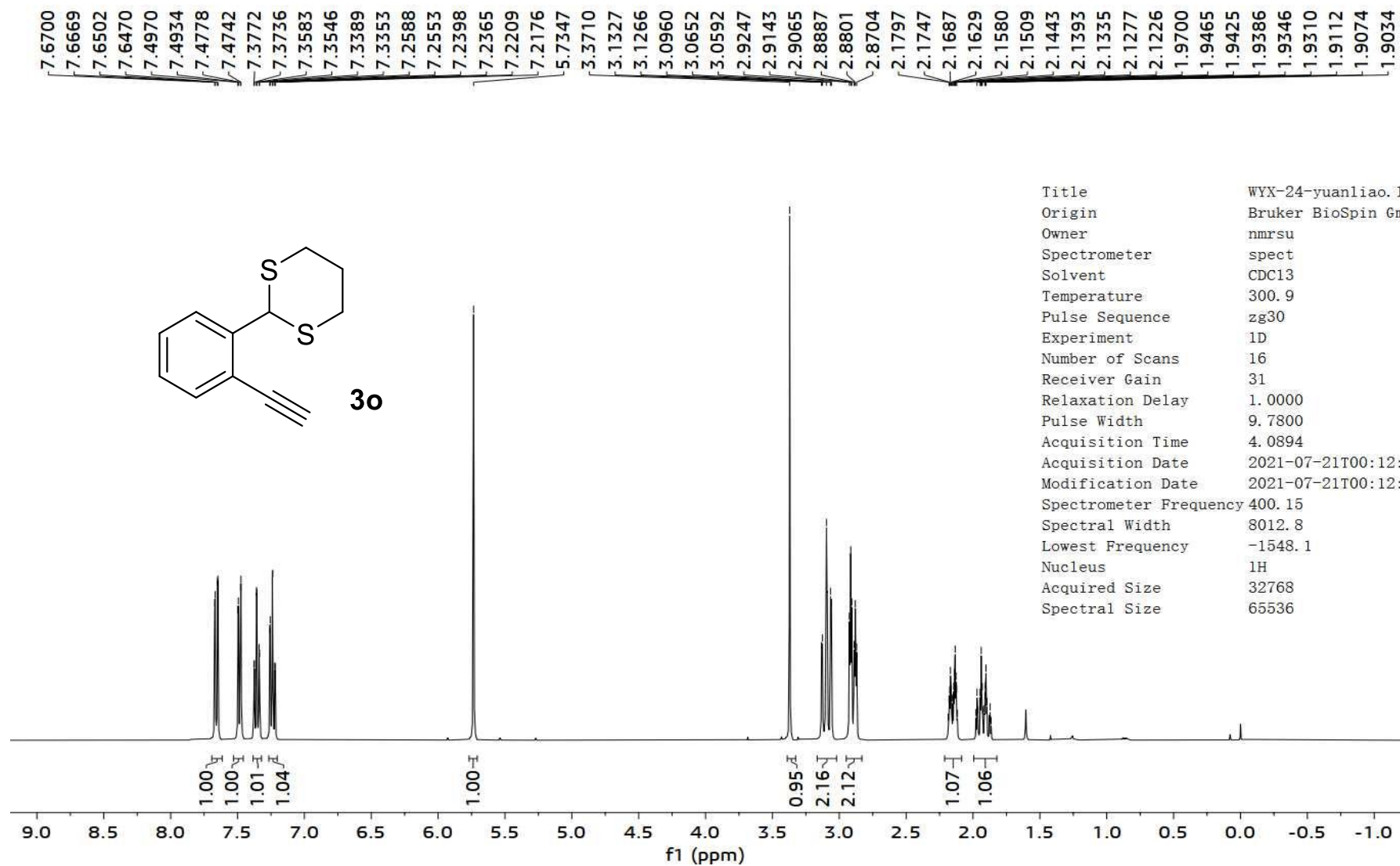
Title	3j.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-18T03:22:46
Modification Date	2022-03-18T03:22:46
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1946.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37



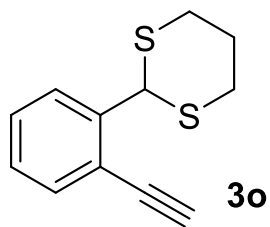
Title	3w.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	64.1
Relaxation Delay	1.0000
Pulse Width	9.7800
Presaturation Frequency	
Acquisition Time	4.0894
Acquisition Date	2022-03-17T12:17:57
Modification Date	2022-03-17T12:17:56
Class	
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12



Title	3w.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-18T02:19:51
Modification Date	2022-03-18T02:19:50
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1949.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37



Title	WYX-24-yuanliao.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	300.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-21T00:12:11
Modification Date	2021-07-21T00:12:12
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1548.1
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



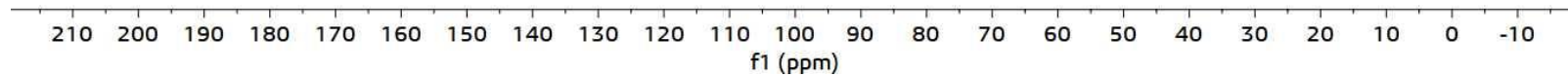
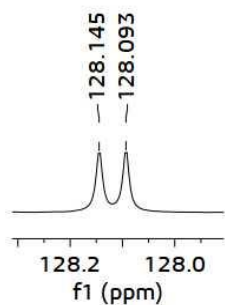
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120.65

82.24
80.93

48.91

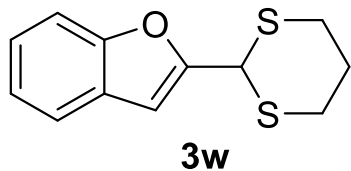
32.34
25.20

Title	WYX-24-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nrsu
Spectrometer	spect
Solvent	CDC13
Temperature	301.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-21T00:56:06
Modification Date	2021-07-21T00:56:08
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1957.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

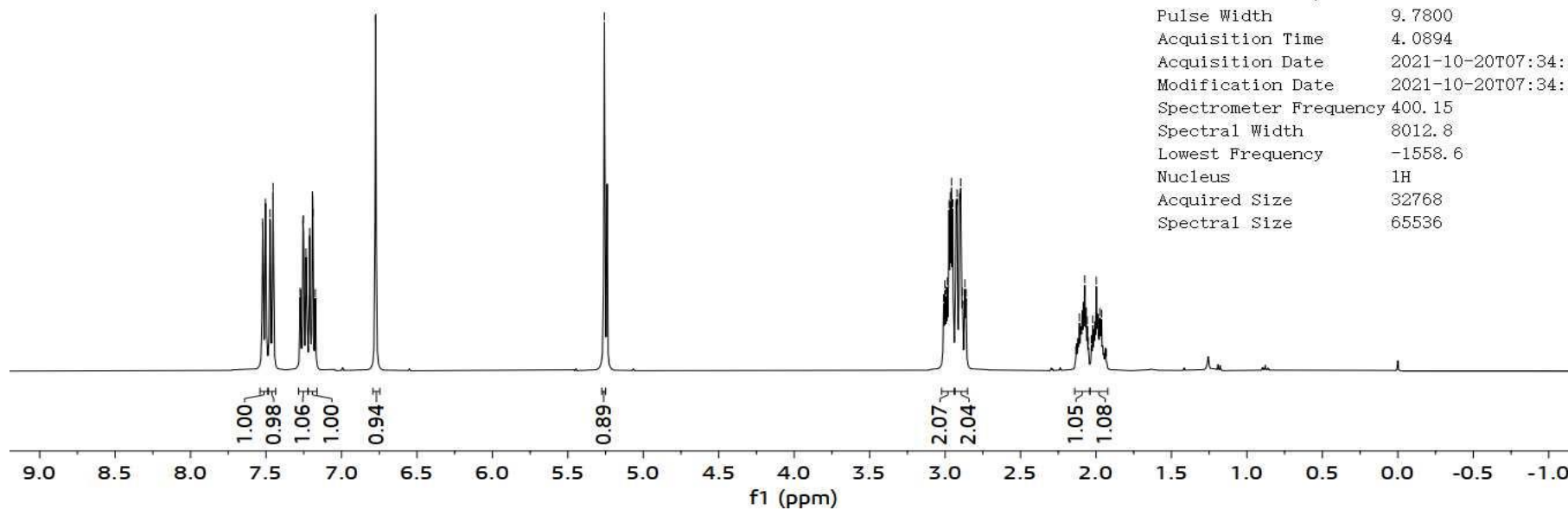


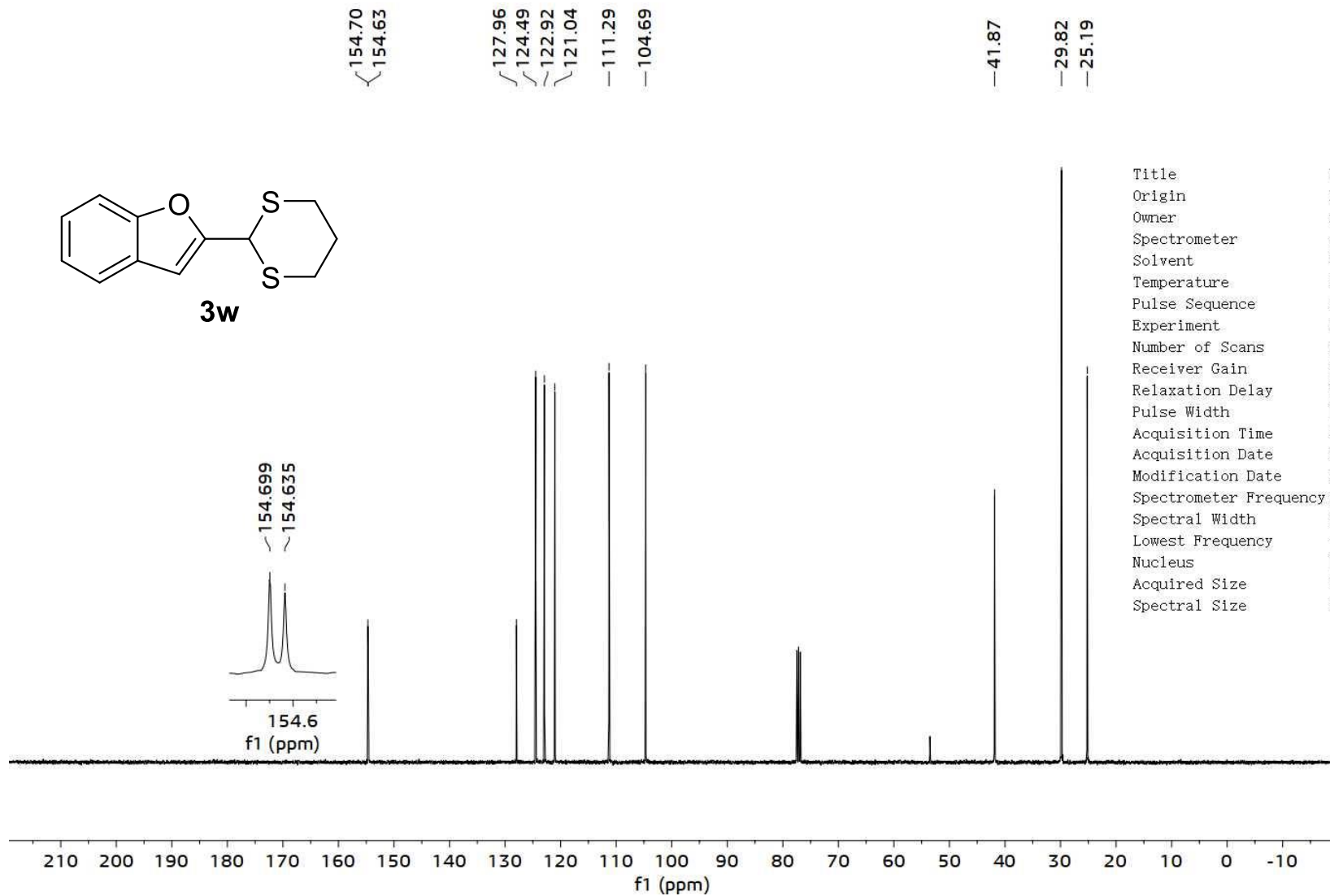
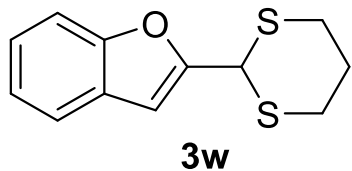
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7.5040
7.4736
7.4534
7.2554
7.2518
7.2352
7.2312
7.2110
7.2080
7.1924
6.7737

5.2580
3.0097
3.0015
2.9931
2.9849
2.9739
2.9659
2.9577
2.9493
2.9282
2.9206
2.9043
2.8967
2.8855
2.8686
2.8612
2.1336
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2.0584
2.0509
2.0303
2.0220
1.9980
1.9739
1.9635
1.9308

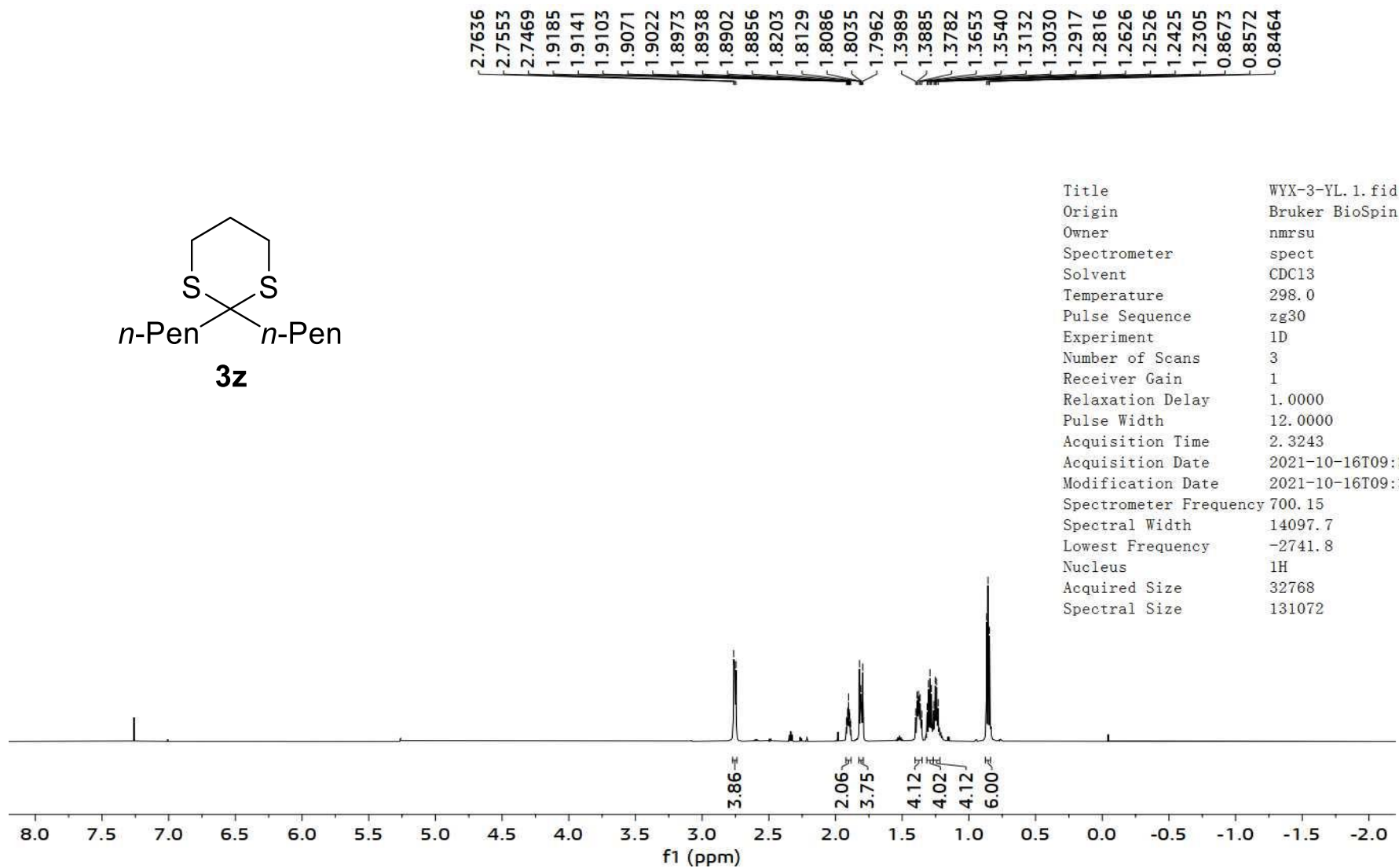
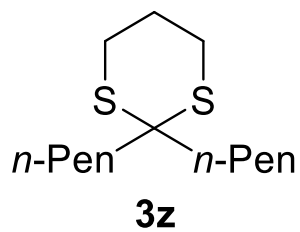


Title	WYX-21-YL.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	17
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-20T07:34:30
Modification Date	2021-10-20T07:34:30
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1558.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

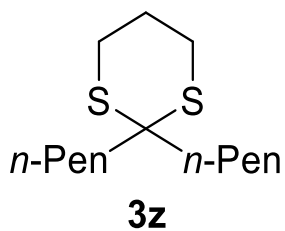




Title	WYX-21-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	100
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-20T07:41:21
Modification Date	2021-10-20T07:41:22
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1970.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

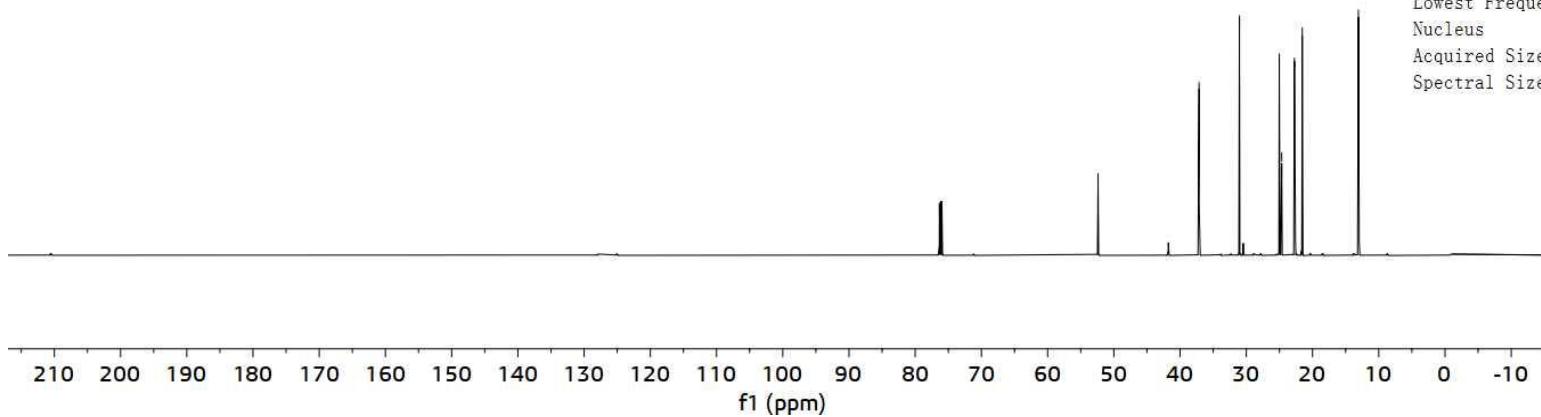


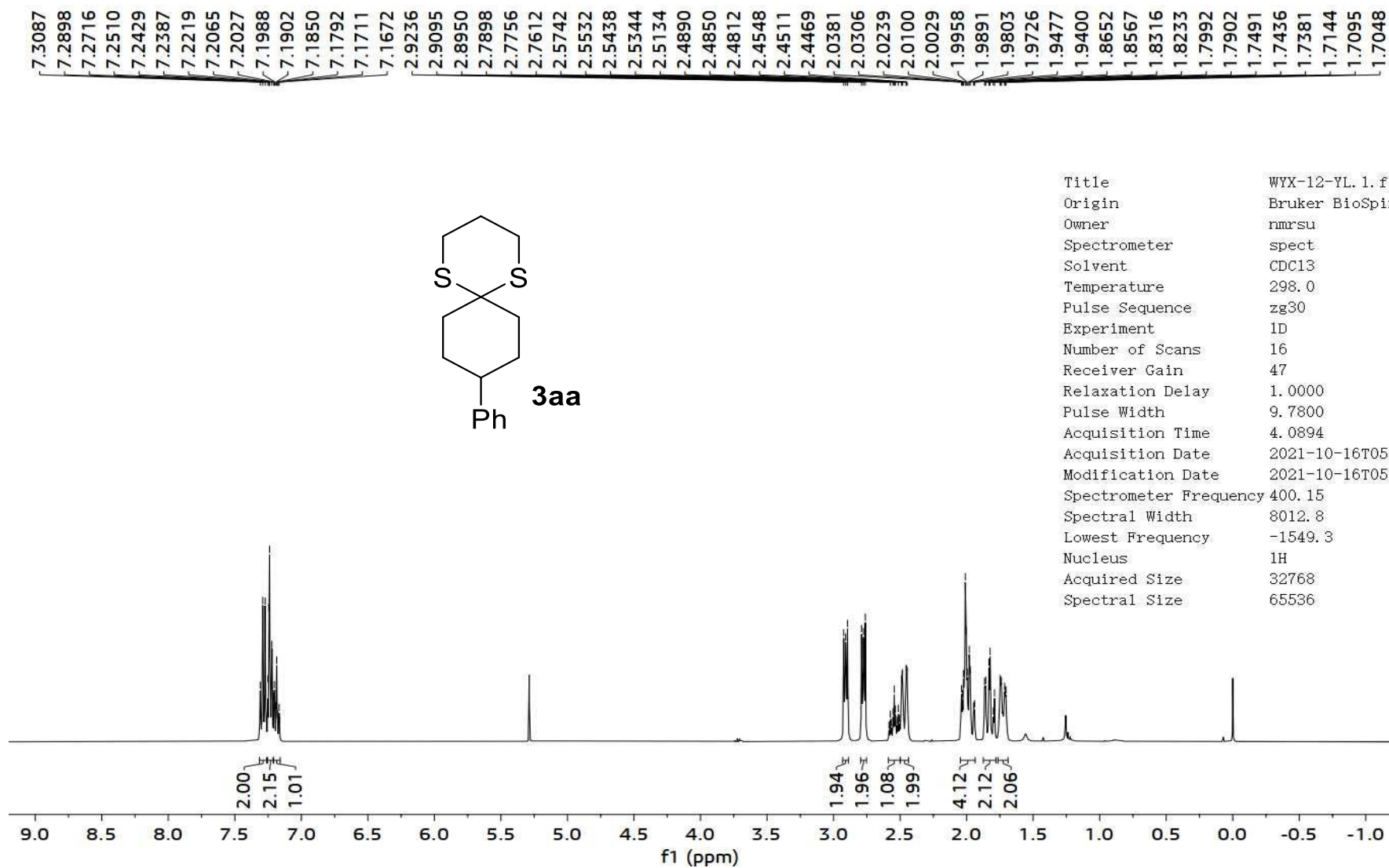
Title	WYX-3-YL. 1. fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	3
Receiver Gain	1
Relaxation Delay	1.0000
Pulse Width	12.0000
Acquisition Time	2.3243
Acquisition Date	2021-10-16T09:30:42
Modification Date	2021-10-16T09:30:44
Spectrometer Frequency	700.15
Spectral Width	14097.7
Lowest Frequency	-2741.8
Nucleus	¹ H
Acquired Size	32768
Spectral Size	131072

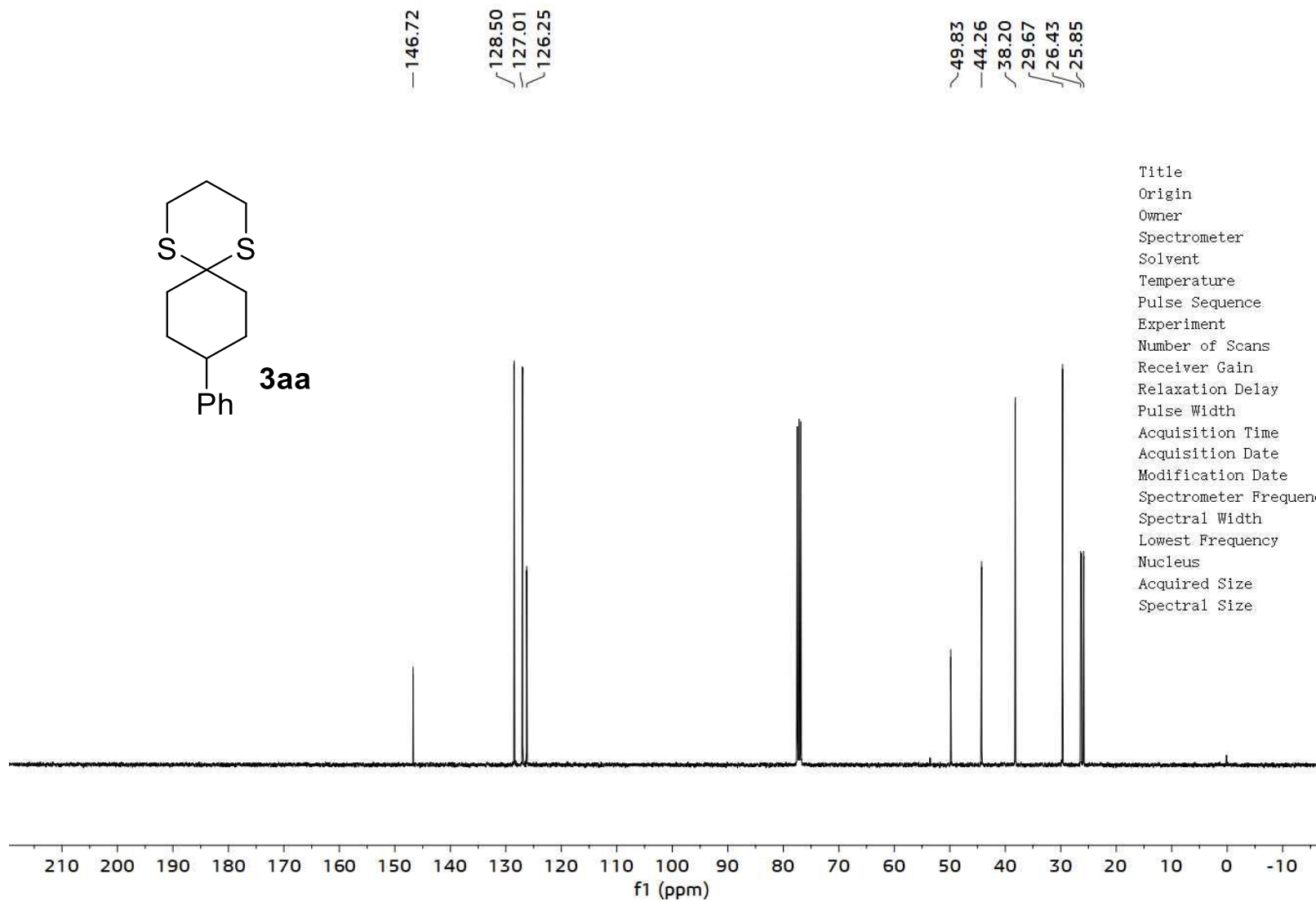
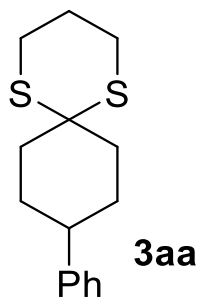


— 52.39
 / 37.13
 / 31.04
 / 25.02
 / 24.65
 / 22.73
 / 21.55
 — 13.07

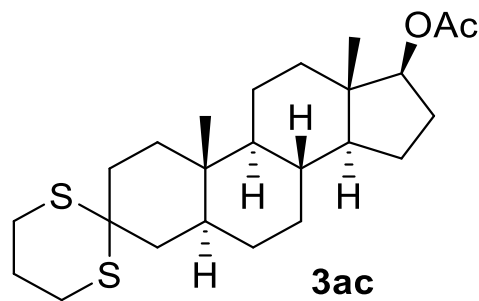
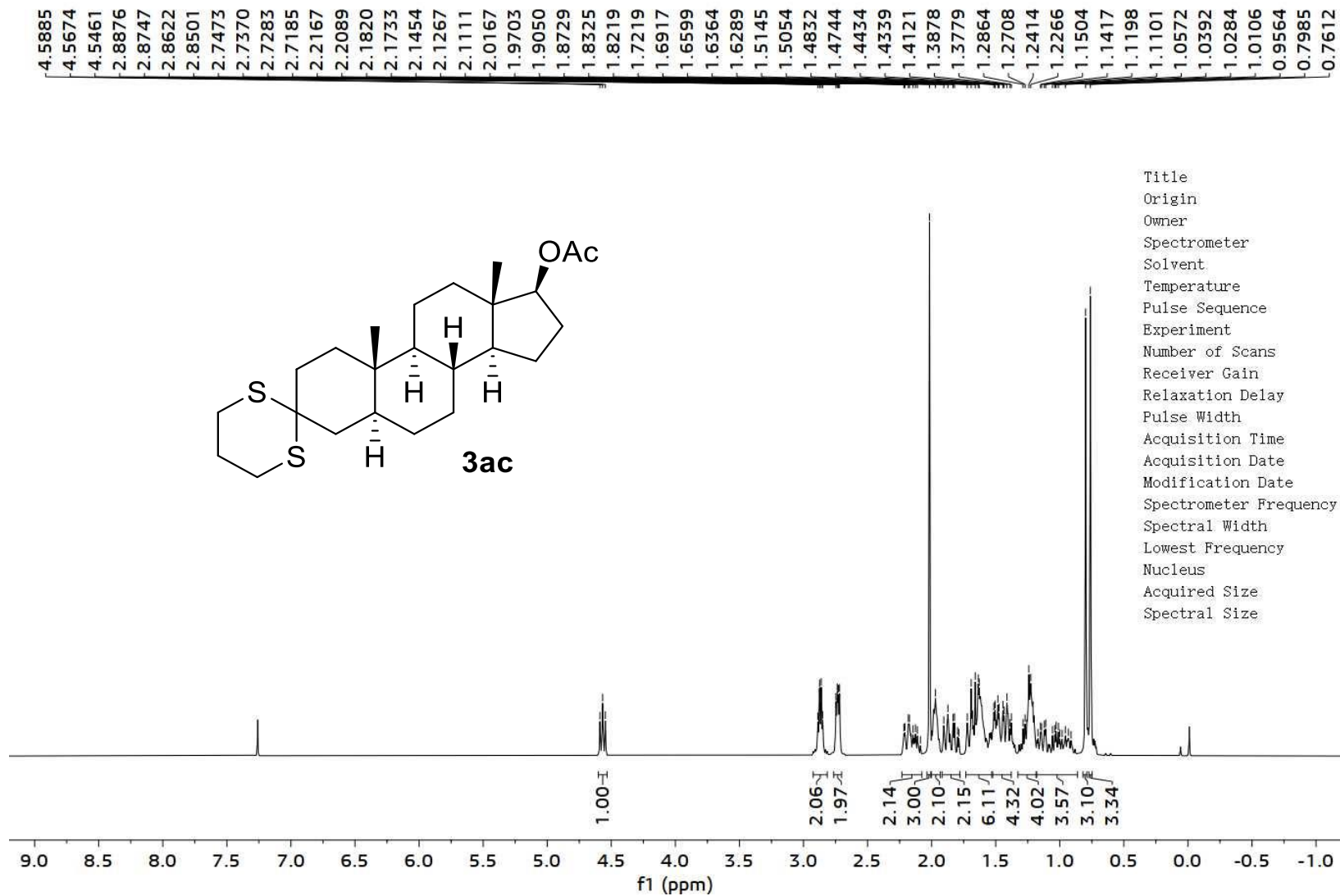
Title	WYX-3-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	26
Receiver Gain	188
Relaxation Delay	2.0000
Pulse Width	12.0000
Acquisition Time	0.7690
Acquisition Date	2021-10-16T09:31:42
Modification Date	2021-10-16T09:32:18
Spectrometer Frequency	176.07
Spectral Width	42613.6
Lowest Frequency	-3870.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536







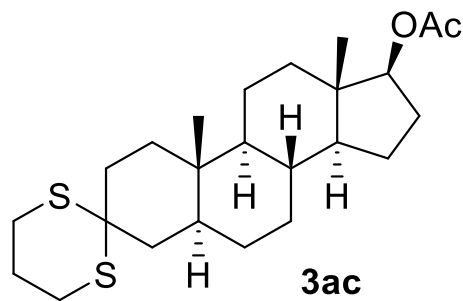
Title	WYX-12-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-16T05:46:08
Modification Date	2021-10-16T05:46:10
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1947.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



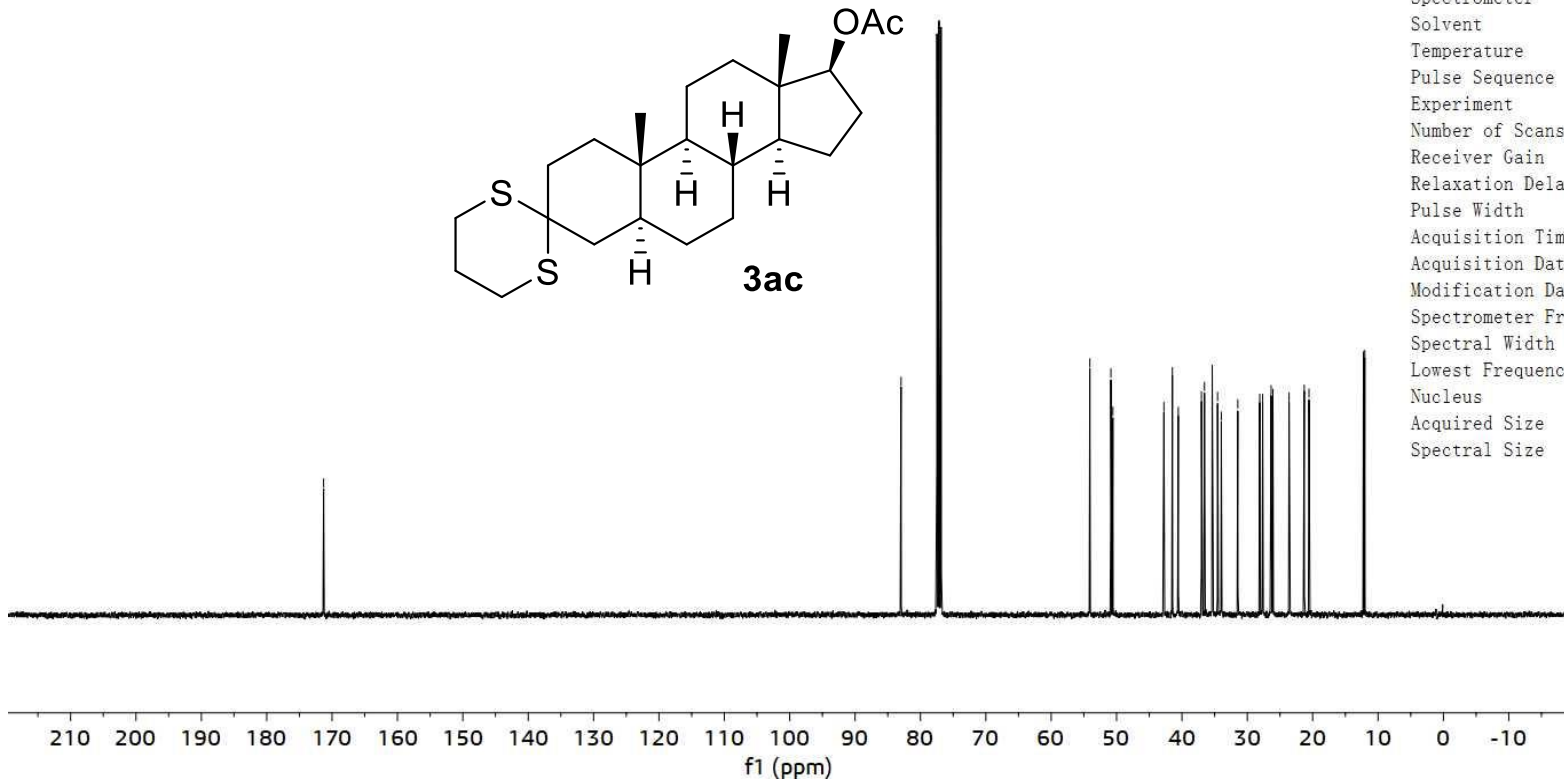
Title	WYX-22. 3. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	303. 0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	31
Relaxation Delay	1. 0000
Pulse Width	9. 7800
Acquisition Time	4. 0894
Acquisition Date	2021-06-11T18:57:43
Modification Date	2021-06-11T18:57:44
Spectrometer Frequency	400. 15
Spectral Width	8012. 8
Lowest Frequency	-1545. 5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

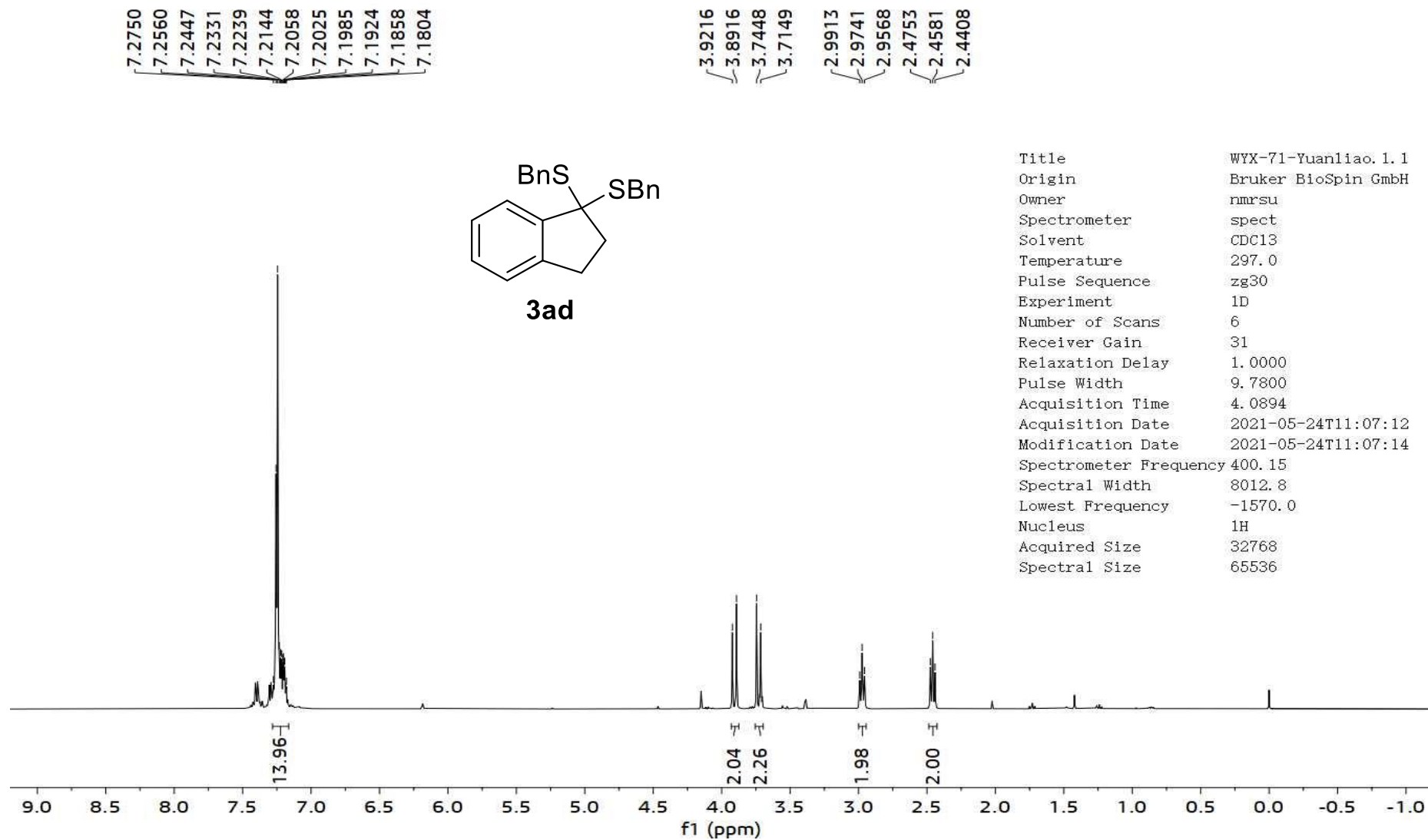
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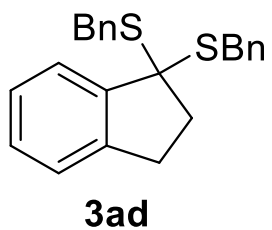
82.97
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50.56
42.76
41.46
40.56
37.02
36.55
35.37
34.55
33.98
31.46
28.10
27.65
26.41
26.35
26.08
23.61
21.29
20.56
12.24
12.03



Title	WYX-22.4.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	303.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-06-11T19:57:17
Modification Date	2021-06-11T19:57:18
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1945.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



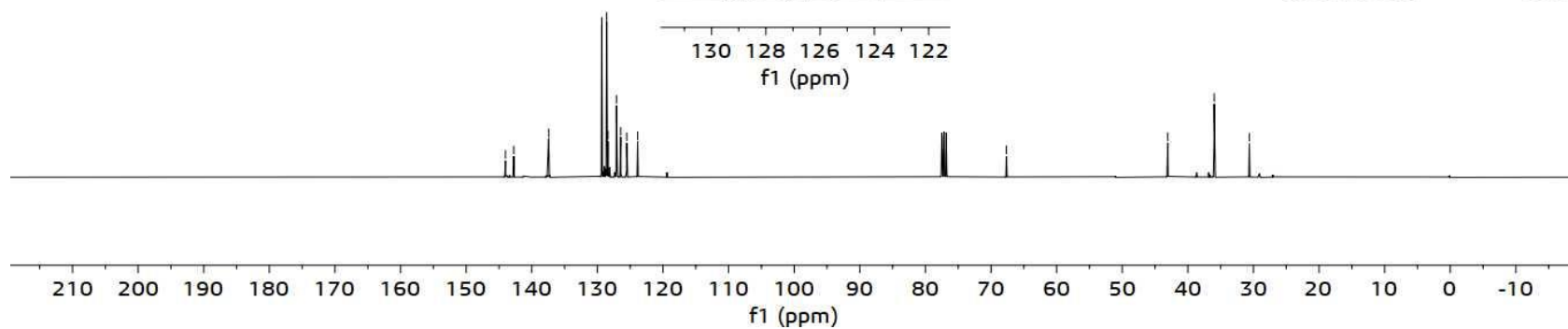
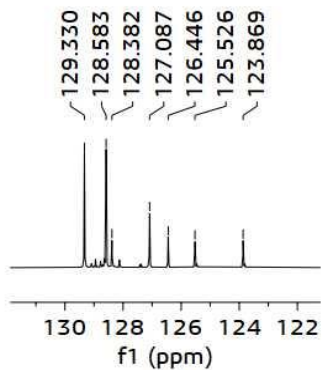




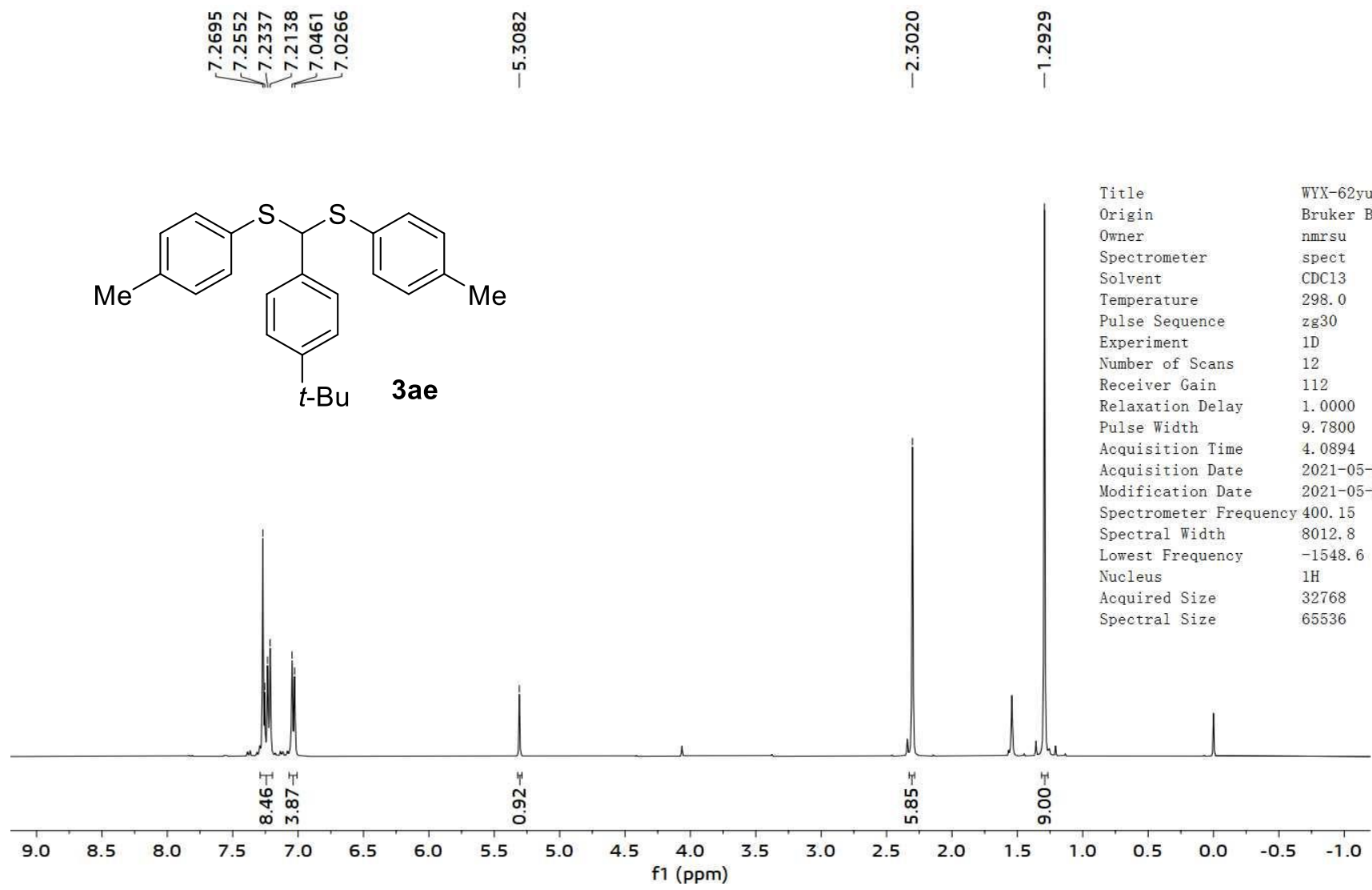
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 129.33
 128.58
 128.38
 127.09
 126.45
 125.53
 123.87

—67.66

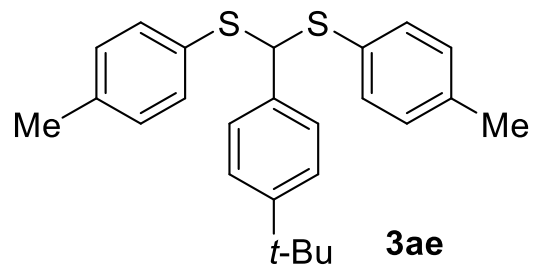
43.06
 35.97
 30.63



Title	WYX-71-Yuanliao. 2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	297.6
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-24T12:06:48
Modification Date	2021-05-24T12:06:50
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1953.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Title	WYX-62yuanliao.1.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	12
Receiver Gain	112
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-17T22:01:29
Modification Date	2021-05-17T22:01:28
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1548.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

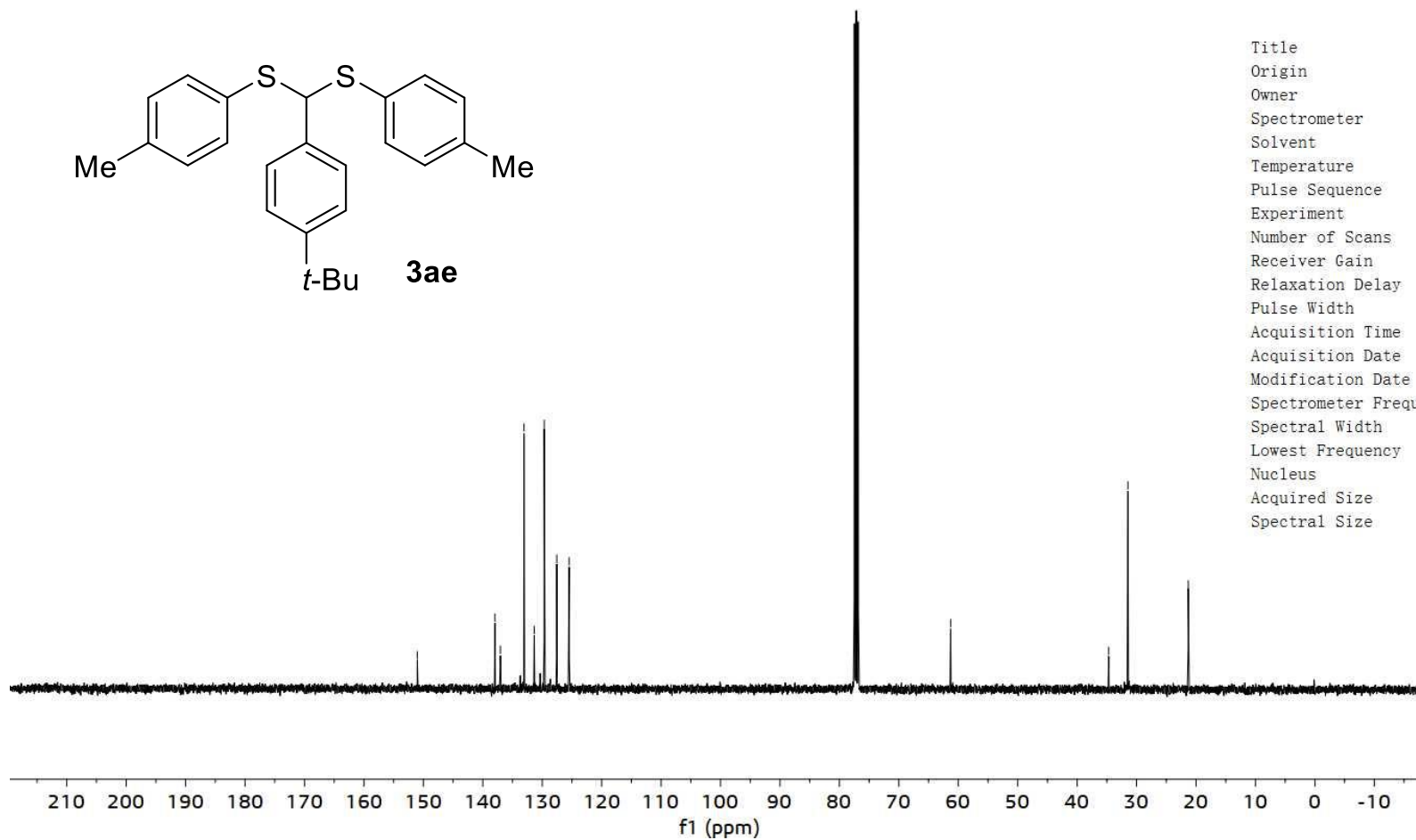


—151.02
 137.95
 137.04
 133.06
 131.35
 129.65
 127.55
 125.47

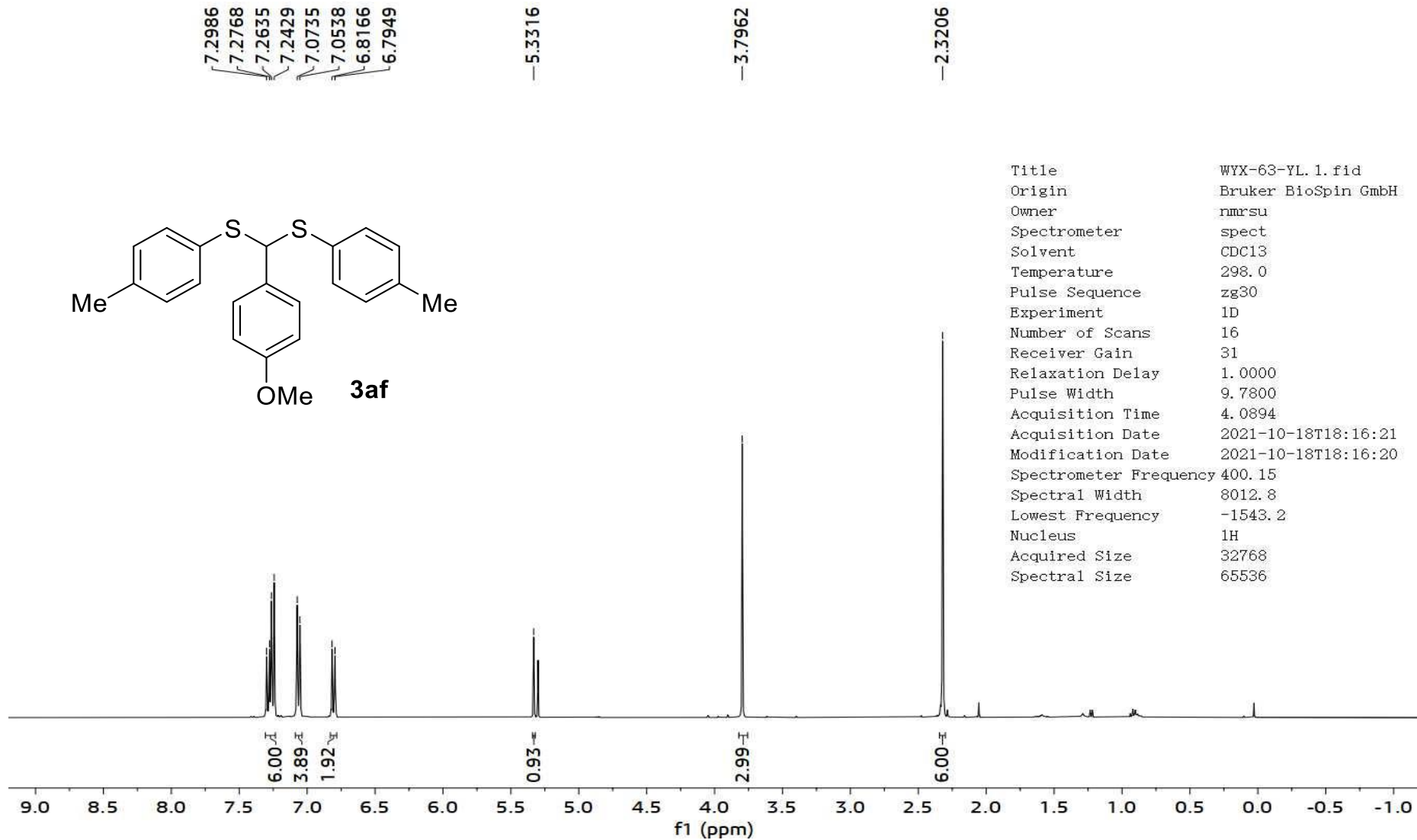
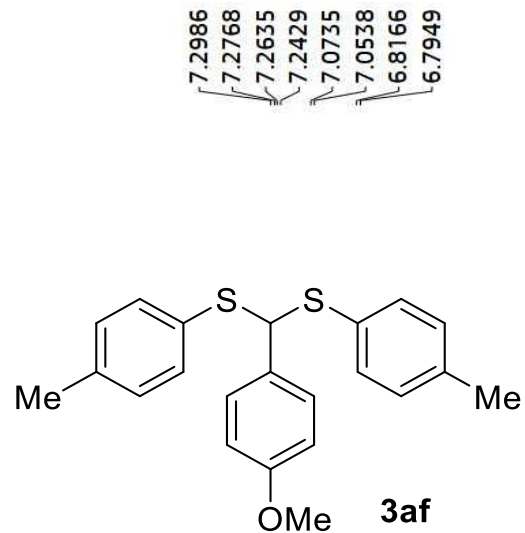
—61.28

—34.70
 —31.45

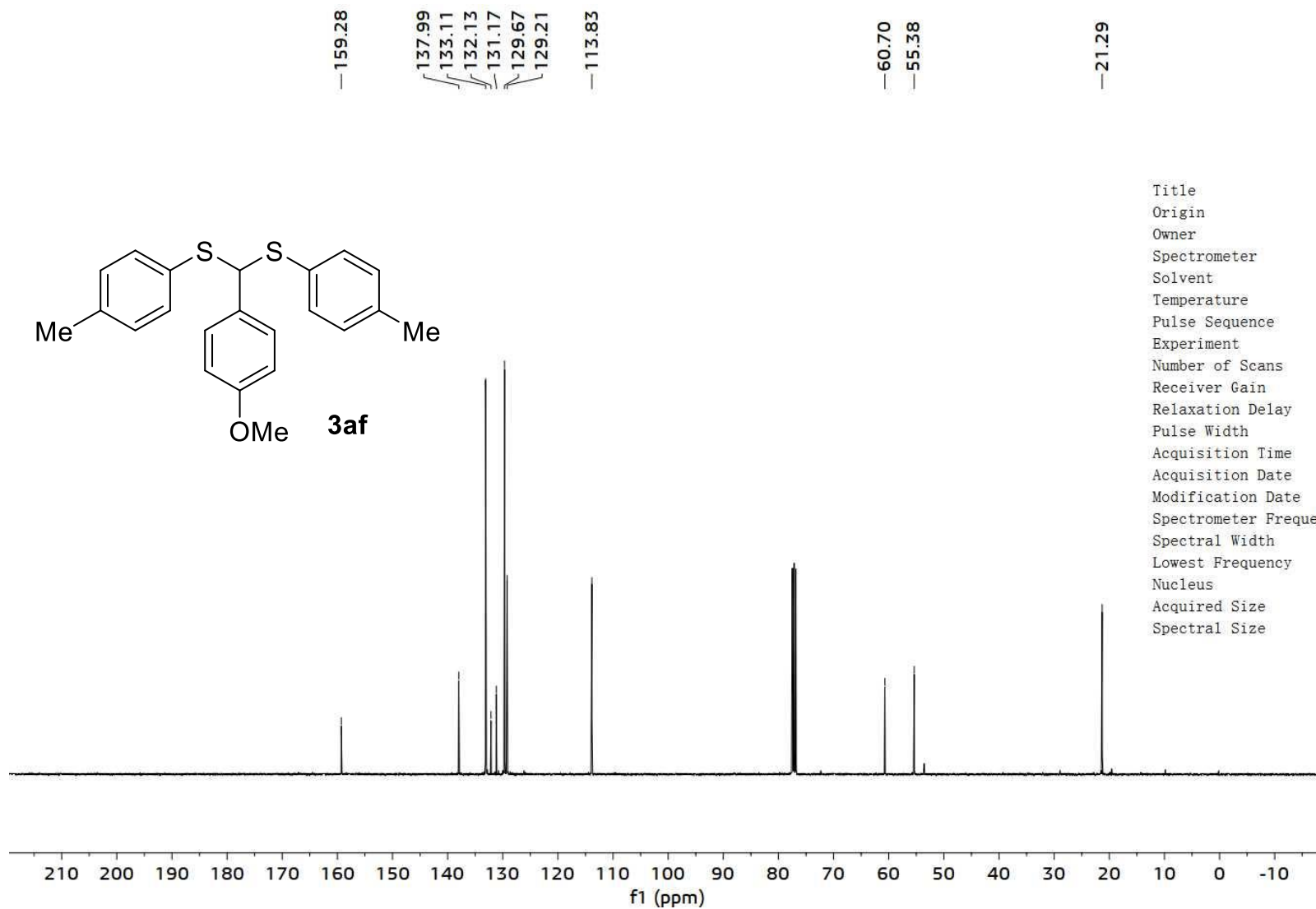
—21.31



Title	WYX-62yuanliao. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	298. 0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2. 0000
Pulse Width	10. 5800
Acquisition Time	1. 3631
Acquisition Date	2021-05-17T22:31:12
Modification Date	2021-05-17T22:31:13
Spectrometer Frequency	100. 62
Spectral Width	24038. 5
Lowest Frequency	-1944. 7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

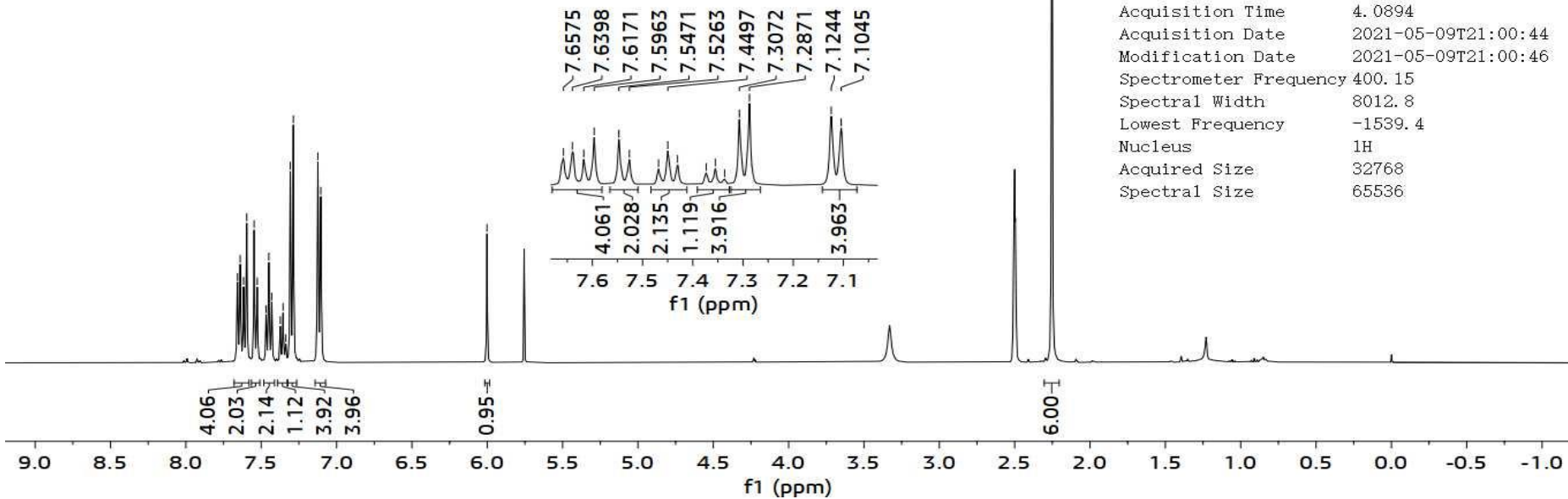
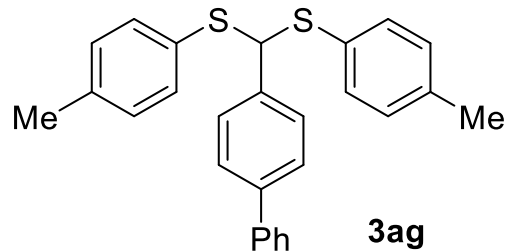


Title	WYX-63-YL.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-18T18:16:21
Modification Date	2021-10-18T18:16:20
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1543.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

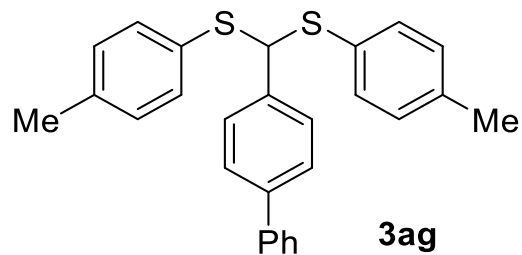


Title	WYX-63-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-18T18:57:27
Modification Date	2021-10-18T18:57:26
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1948.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

7.6575
7.6398
7.6171
7.5963
7.5471
7.5263
7.4682
7.4497
7.4304
7.3733
7.3551
7.3366
7.3072
7.2871
7.1244
7.1045
—6.0019



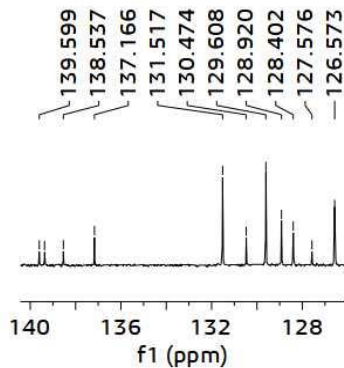
Title	WYX-61-yuanliao.1.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	127
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-09T21:00:44
Modification Date	2021-05-09T21:00:46
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1539.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



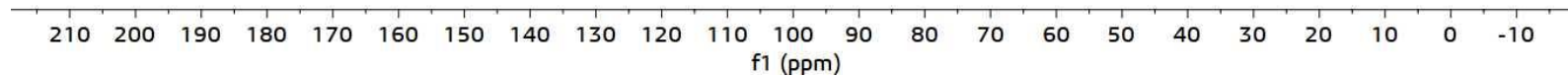
139.60
139.37
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137.17
131.52
130.47
129.61
128.92
128.40
127.58
126.57

—57.12

—20.62

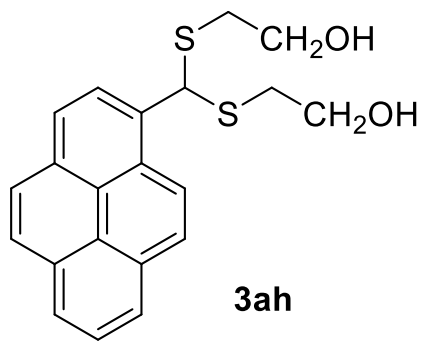


Title	WYX-61-yuanliao. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	298. 0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2. 0000
Pulse Width	10. 5800
Acquisition Time	1. 3631
Acquisition Date	2021-05-09T22:00:22
Modification Date	2021-05-09T22:00:24
Spectrometer Frequency	100. 62
Spectral Width	24038. 5
Lowest Frequency	-2007. 2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

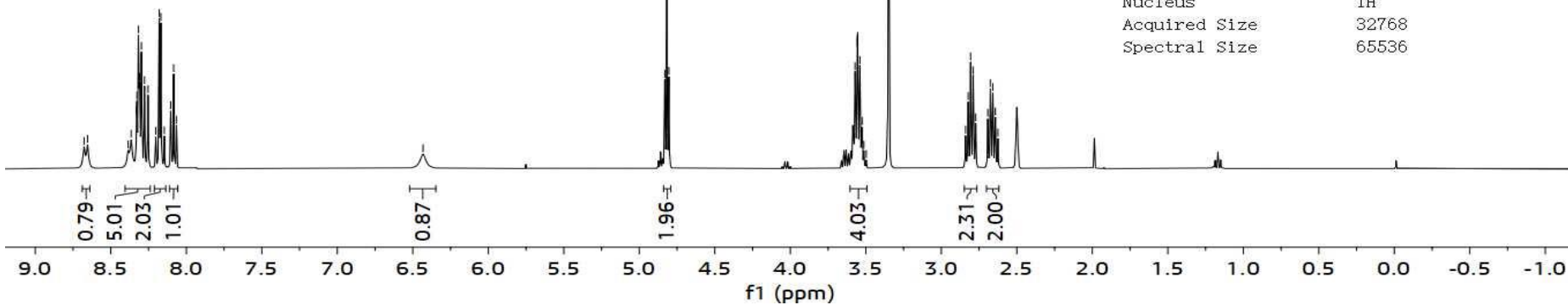


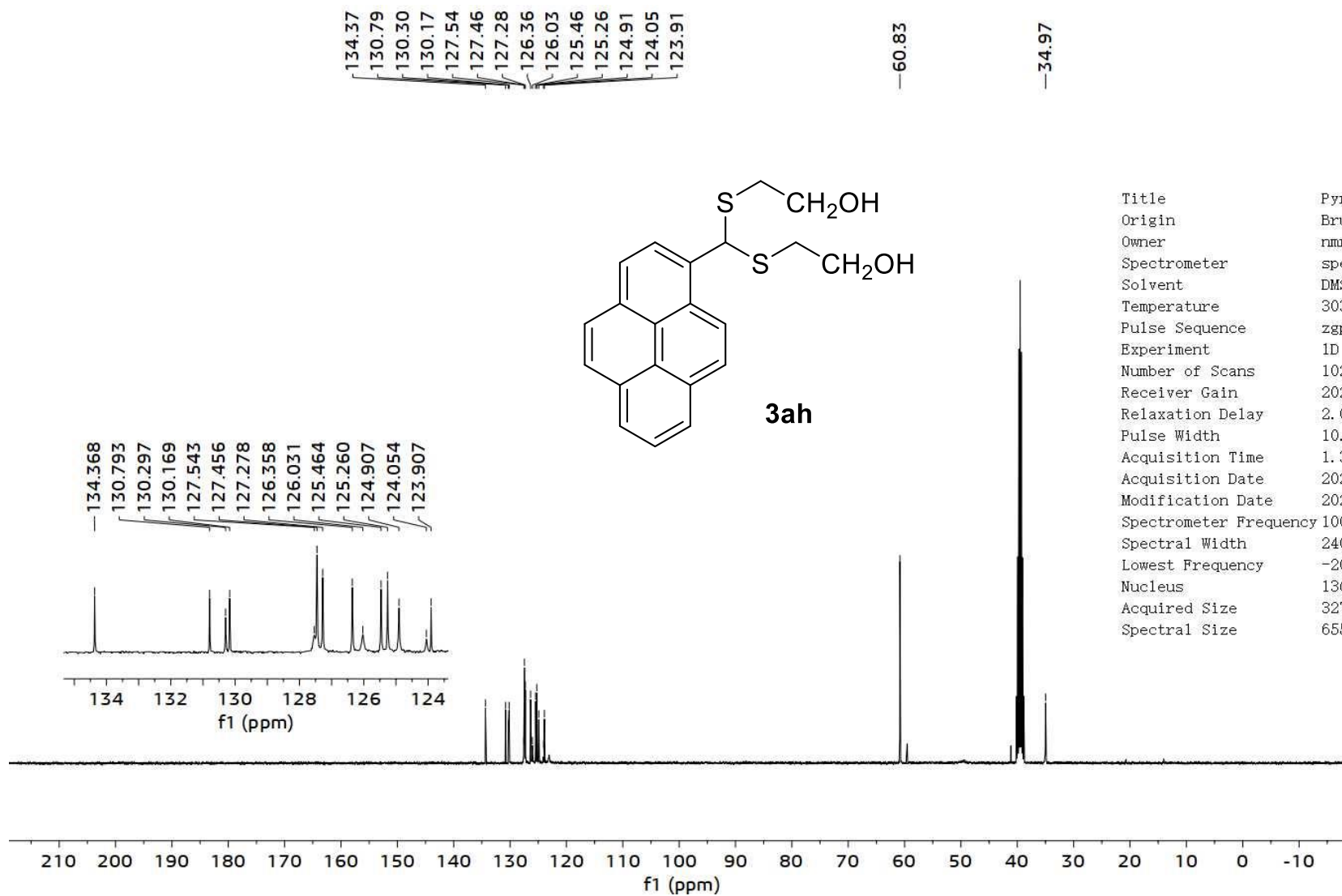
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8.6535
8.3848
8.3646
8.3298
8.3270
8.3166
8.3089
8.2972
8.2943
8.2766
8.2532
8.2017
8.1794
8.1682
8.1459
8.1029
8.0839
8.0648
6.4321

4.8312
4.8176
4.8042
3.5713
3.5569
3.5539
3.5390
3.5242
3.5130
3.4970
2.8395
2.8225
2.8059
2.7900
2.7724
2.6922
2.6752
2.6587
2.6422
2.6254

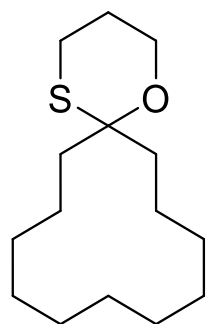


Title	Pyr2-2. 1. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	303.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	12
Receiver Gain	64
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-06-24T23:24:38
Modification Date	2021-06-24T23:24:40
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1539.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

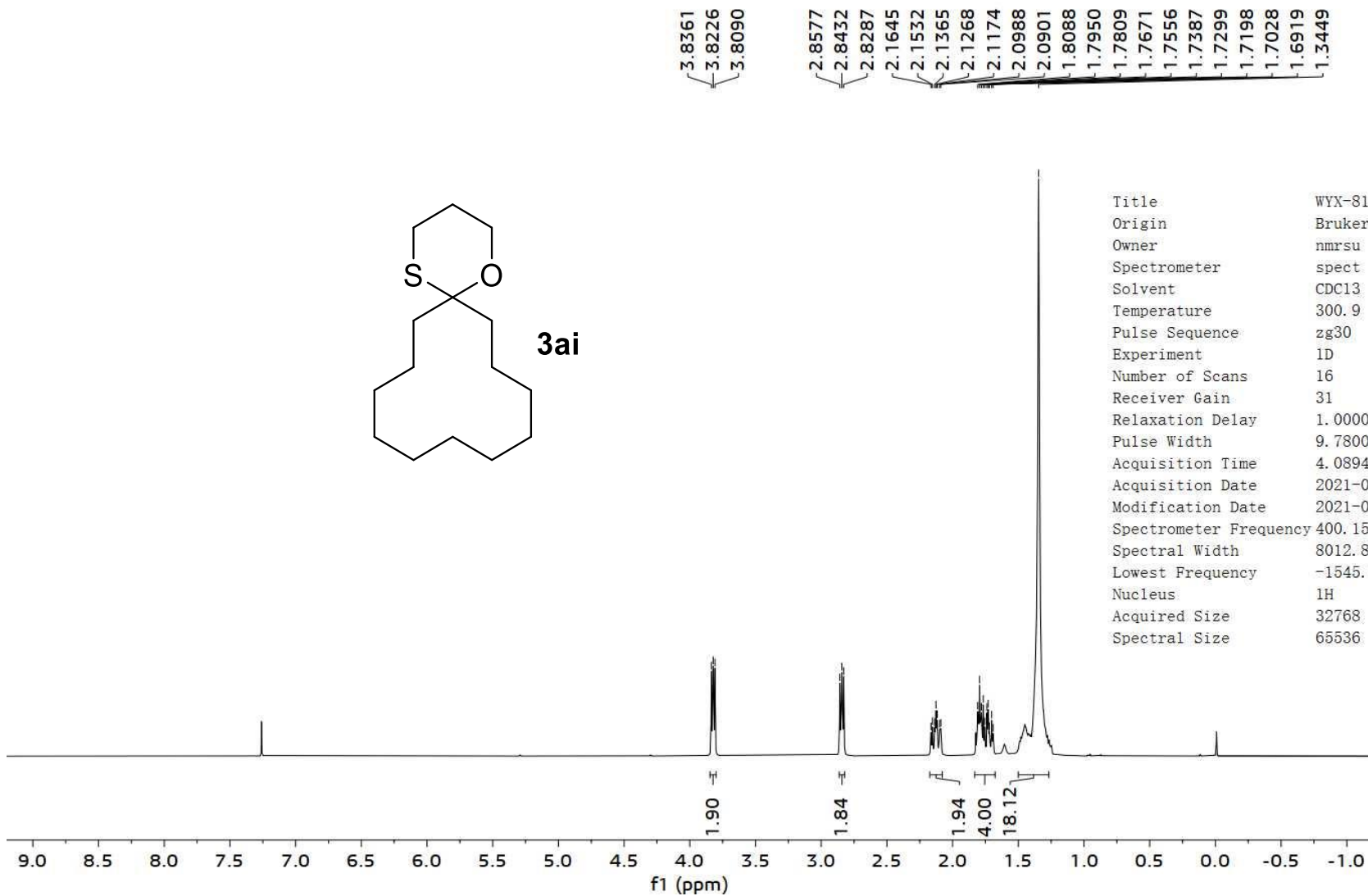




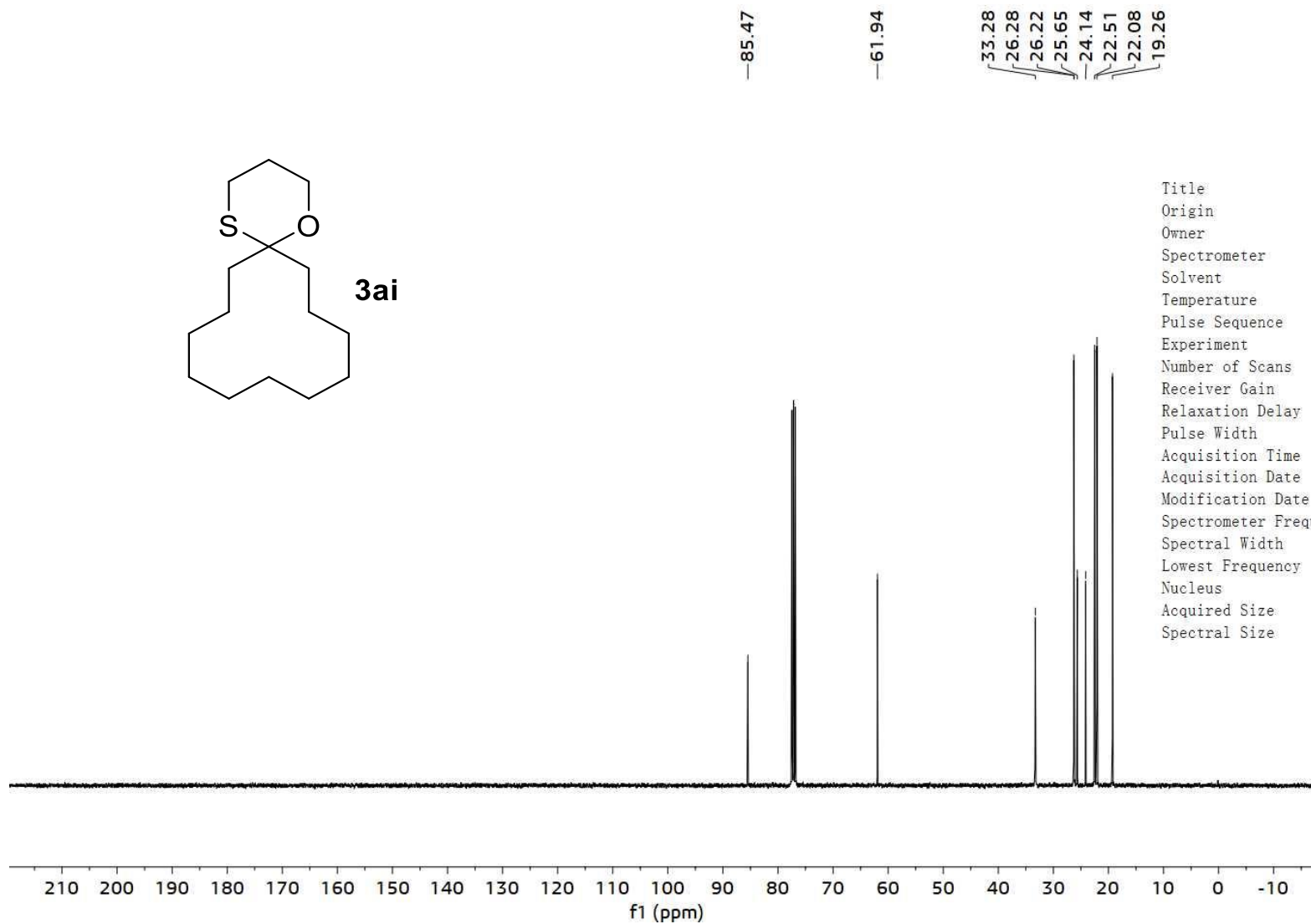
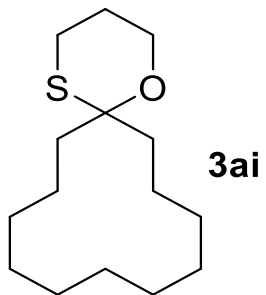
Title	Pyr2-2. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	303.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-06-25T00:24:17
Modification Date	2021-06-25T00:24:18
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-2011.0
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



3ai

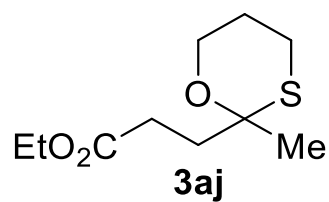


Title	WYX-81-yuanliao.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	300.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-20T23:23:12
Modification Date	2021-07-20T23:23:14
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

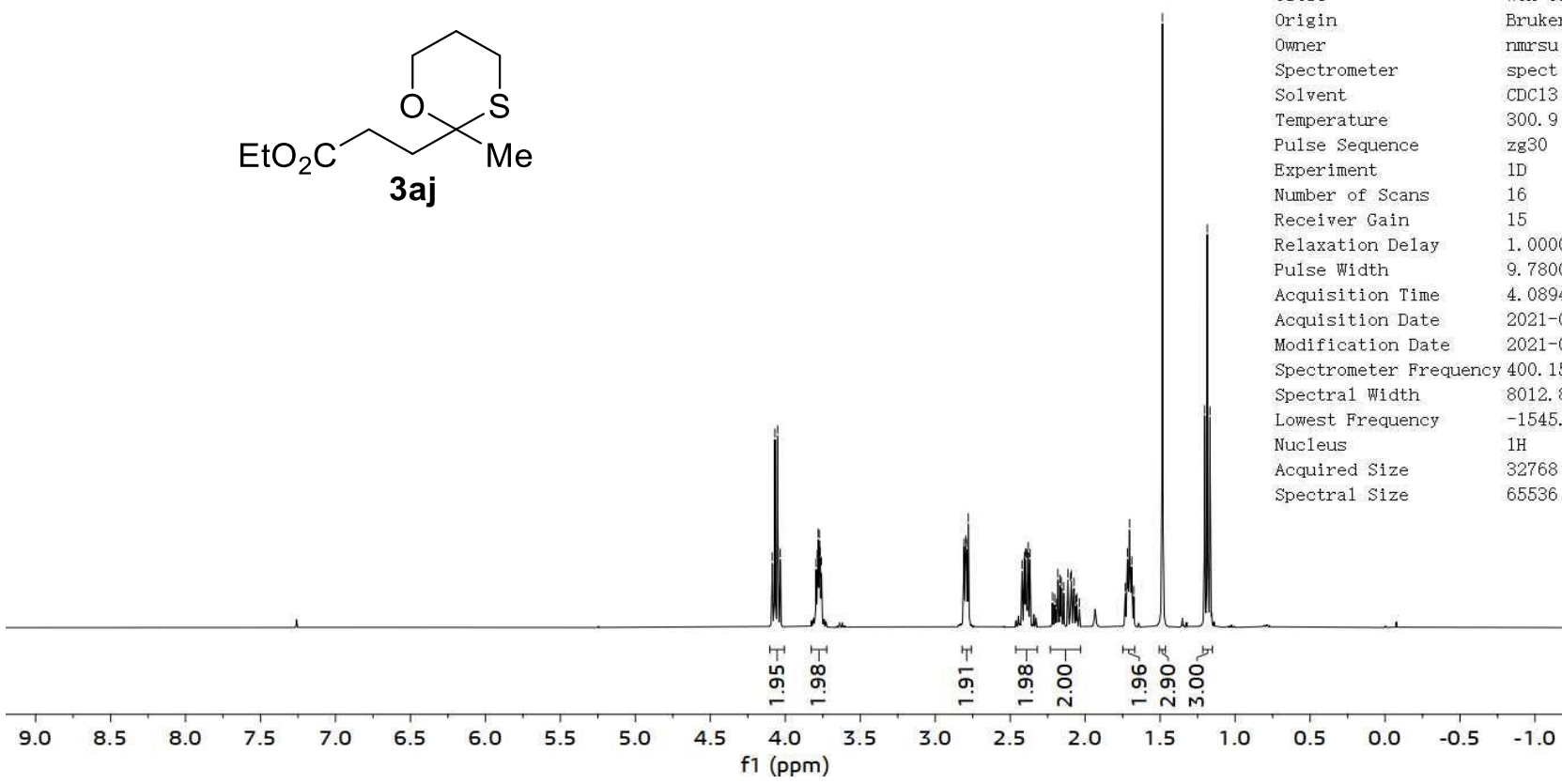


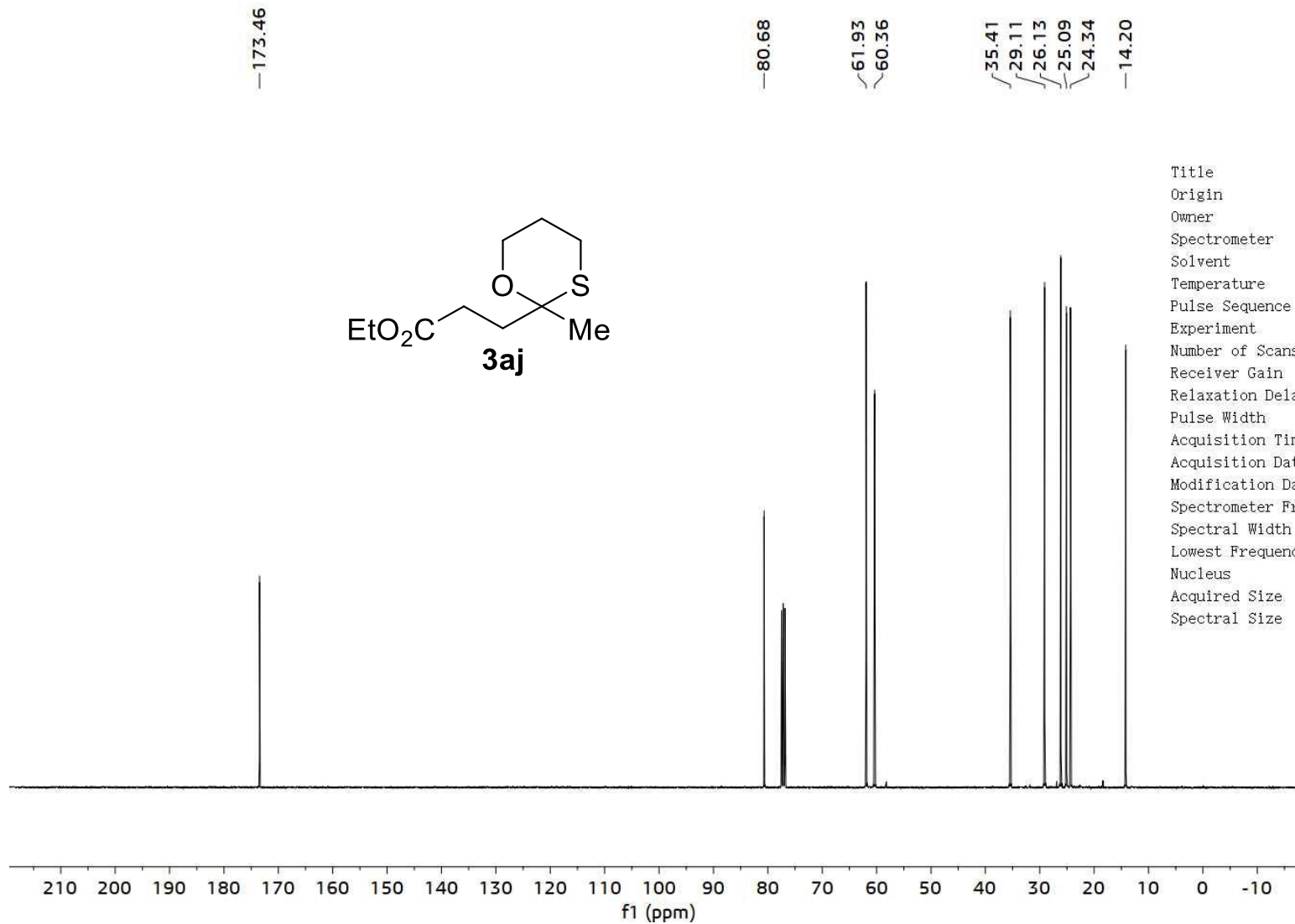
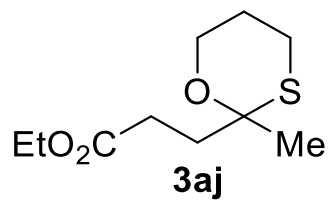
Title	WYX-81-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	301.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-21T00:07:07
Modification Date	2021-07-21T00:07:08
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1944.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

4.0870
 4.0692
 4.0512
 4.0335
 3.7942
 3.7863
 3.7807
 3.7724
 3.7664
 3.7597
 2.8075
 2.7998
 2.7960
 2.7924
 2.7884
 2.7795
 2.4208
 2.4072
 2.4040
 2.3975
 2.3917
 2.3834
 2.3798
 2.3686
 2.2180
 2.2035
 2.1937
 2.1823
 2.1669
 2.1589
 2.1433
 2.1152
 2.0977
 2.0922
 2.0795
 2.0746
 2.0627
 2.0556
 2.0388
 1.7325
 1.7180
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 1.6897
 1.6761
 1.4849
 1.2034
 1.1855
 1.1677

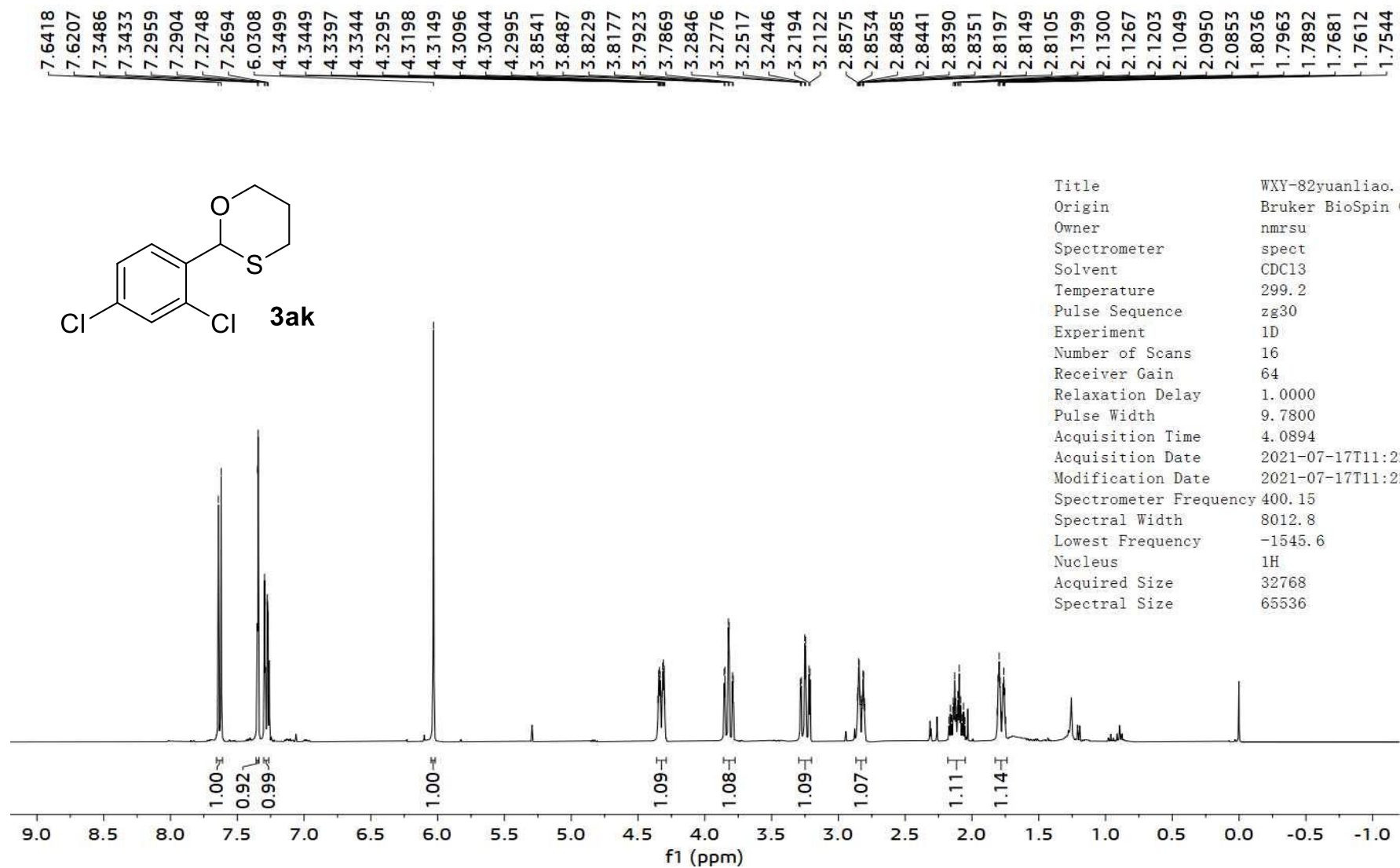


Title	WYX-83-yuanliao.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	300.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	15
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-20T22:34:21
Modification Date	2021-07-20T22:34:22
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.4
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

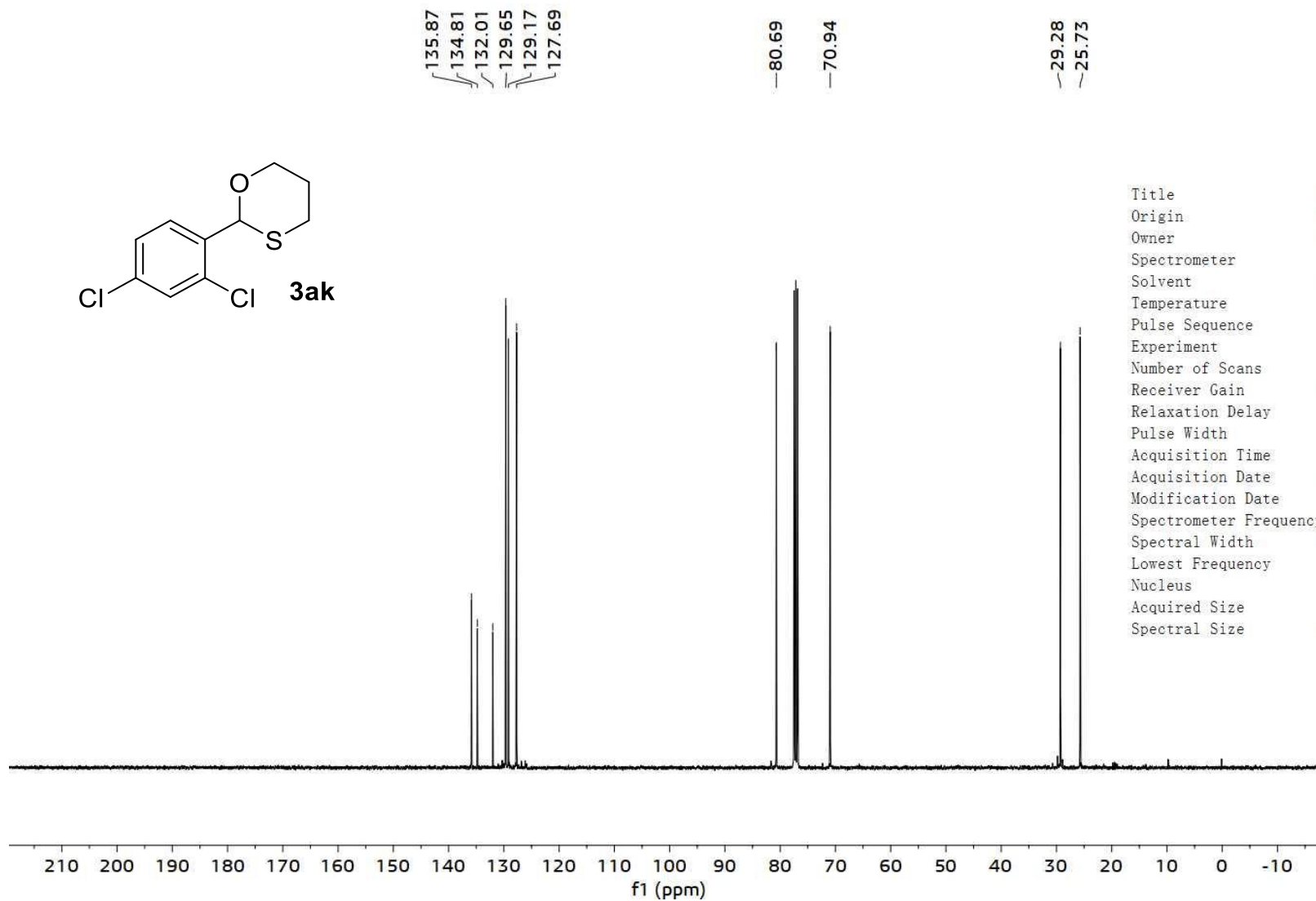
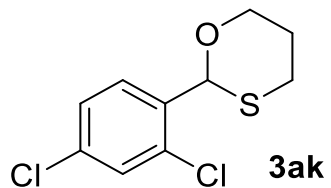




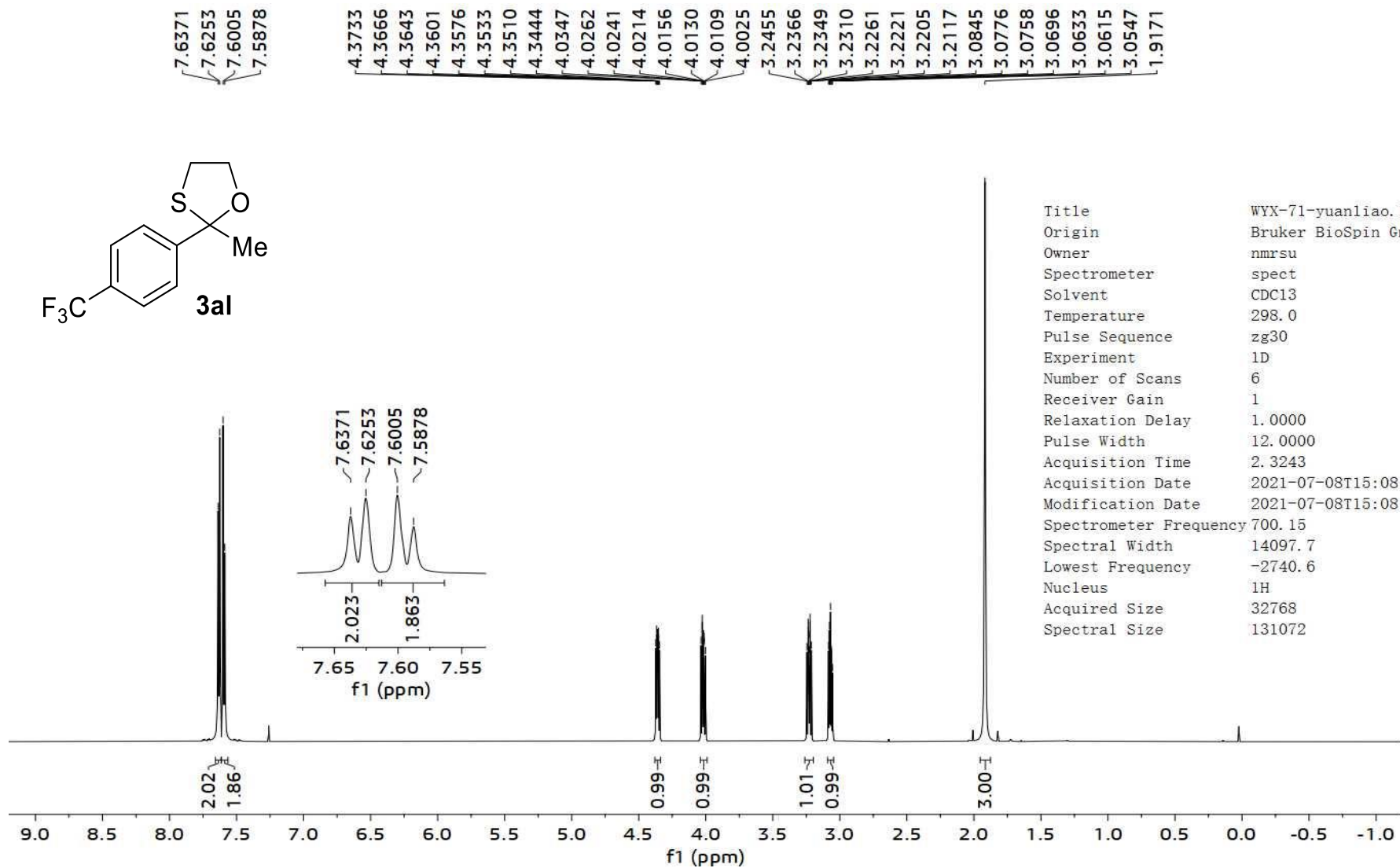
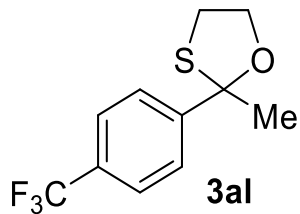
Title	WYX-83-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	301.5
Pulse Sequence	zpgp30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-20T23:18:15
Modification Date	2021-07-20T23:18:16
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1954.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



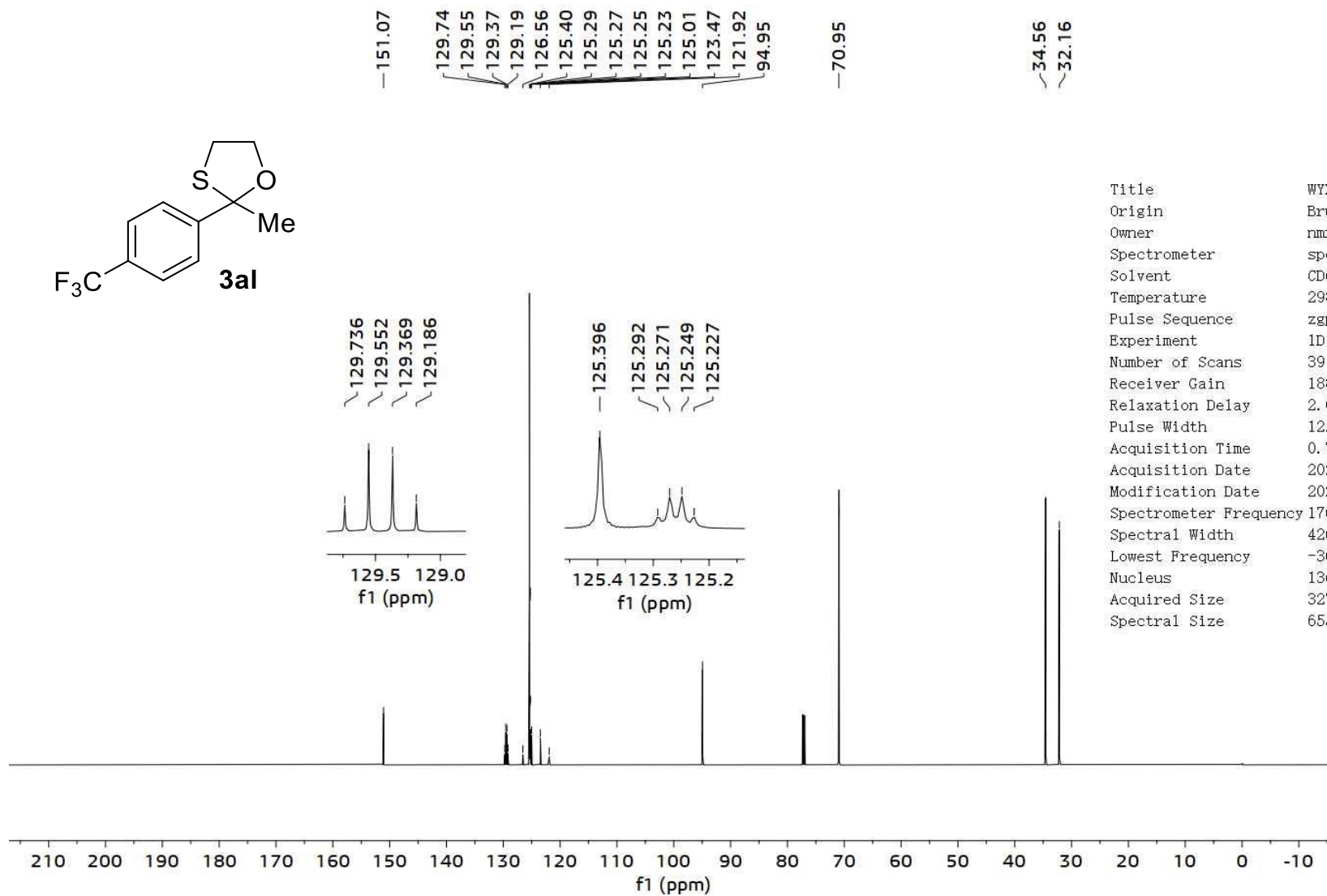
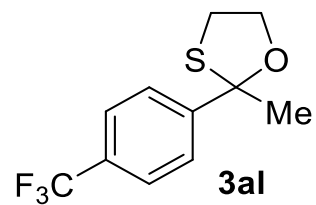
Title	WXY-82yuanliao.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	299.2
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	64
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-17T11:22:05
Modification Date	2021-07-17T11:22:06
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



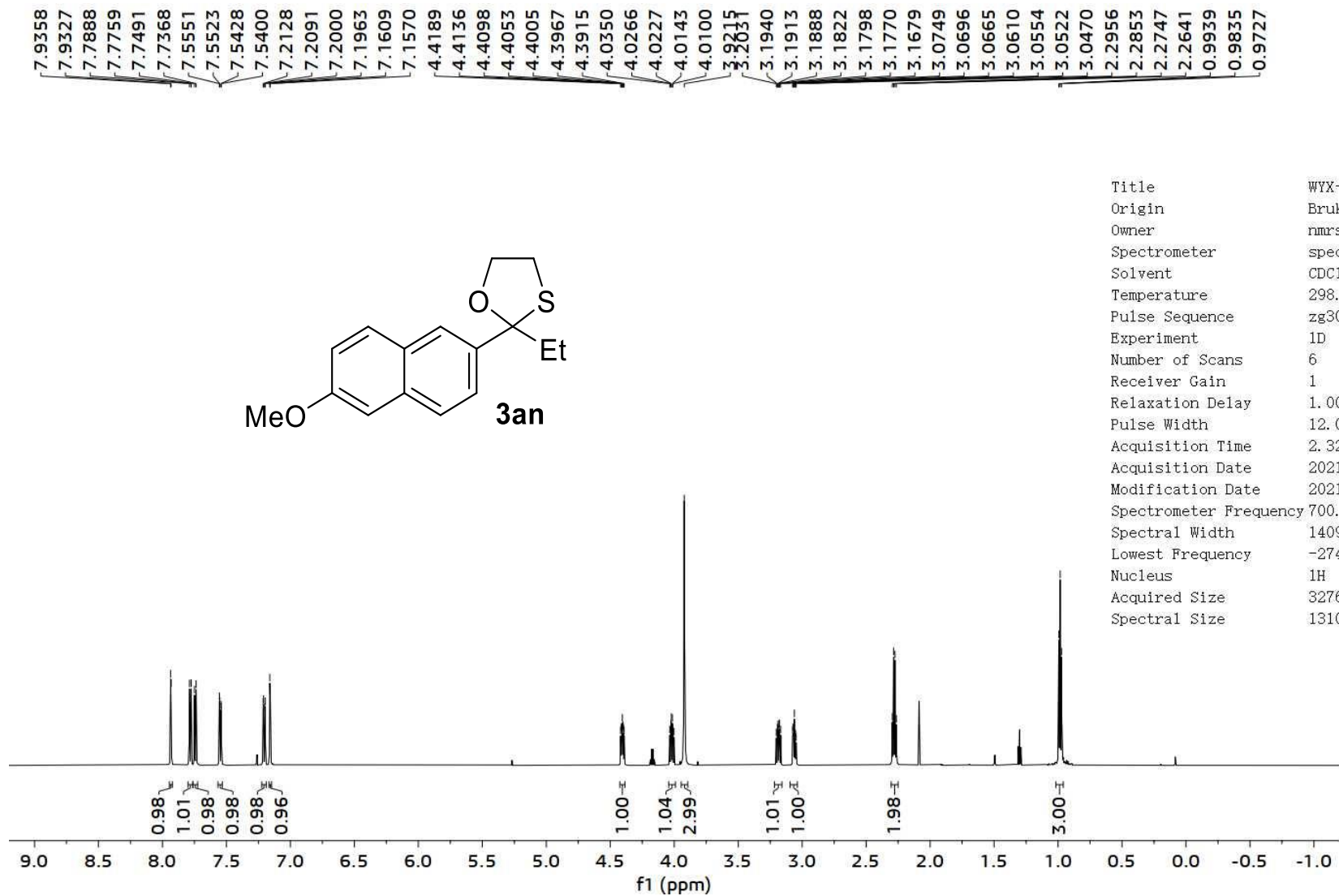
Title	WXY-82yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	300.1
Pulse Sequence	zpg30
Experiment	1D
Number of Scans	1700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-17T13:00:15
Modification Date	2021-07-17T13:00:16
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1947.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



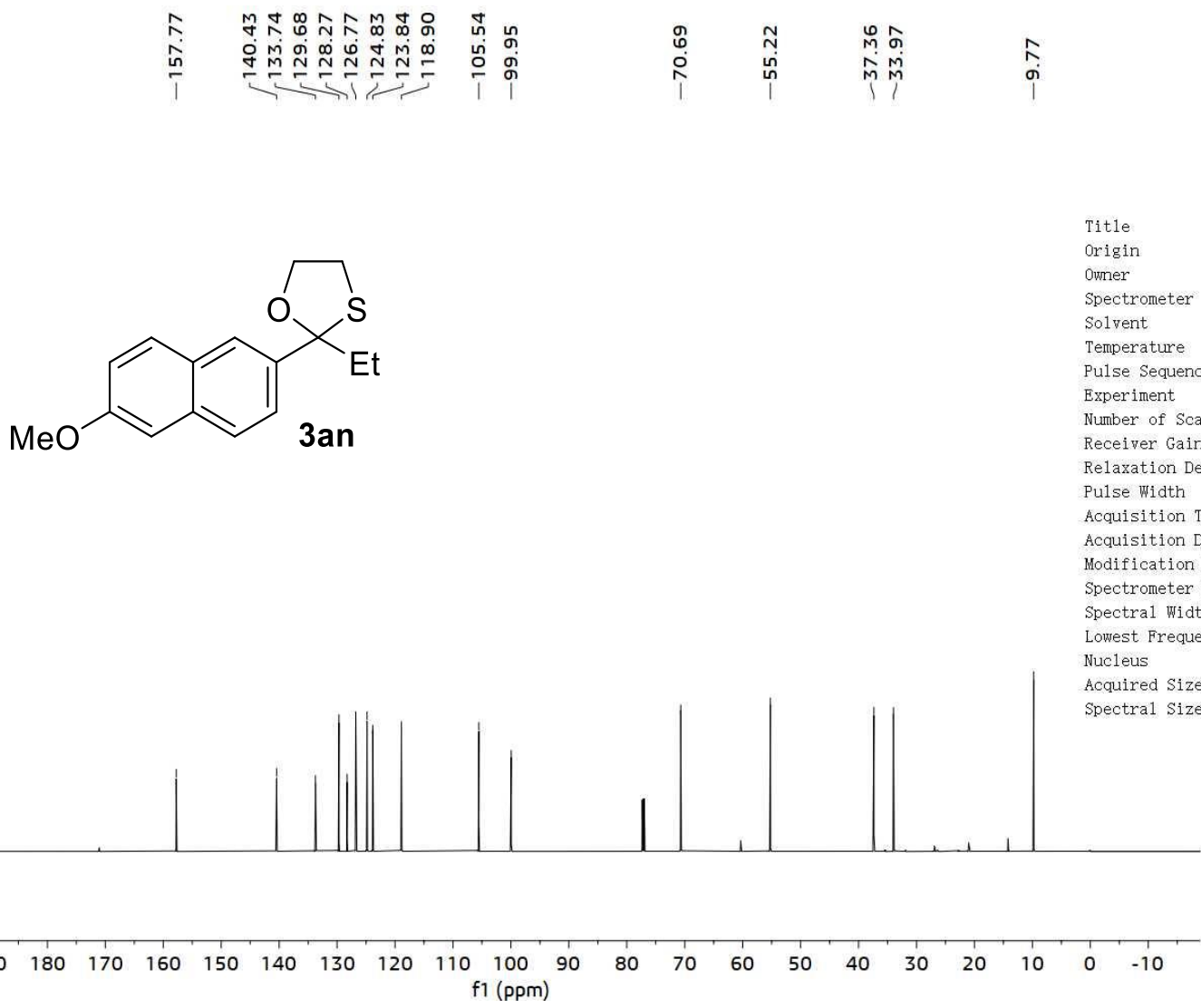
Title	WYX-71-yuanliao.1. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	1
Relaxation Delay	1.0000
Pulse Width	12.0000
Acquisition Time	2.3243
Acquisition Date	2021-07-08T15:08:38
Modification Date	2021-07-08T15:08:40
Spectrometer Frequency	700.15
Spectral Width	14097.7
Lowest Frequency	-2740.6
Nucleus	1H
Acquired Size	32768
Spectral Size	131072



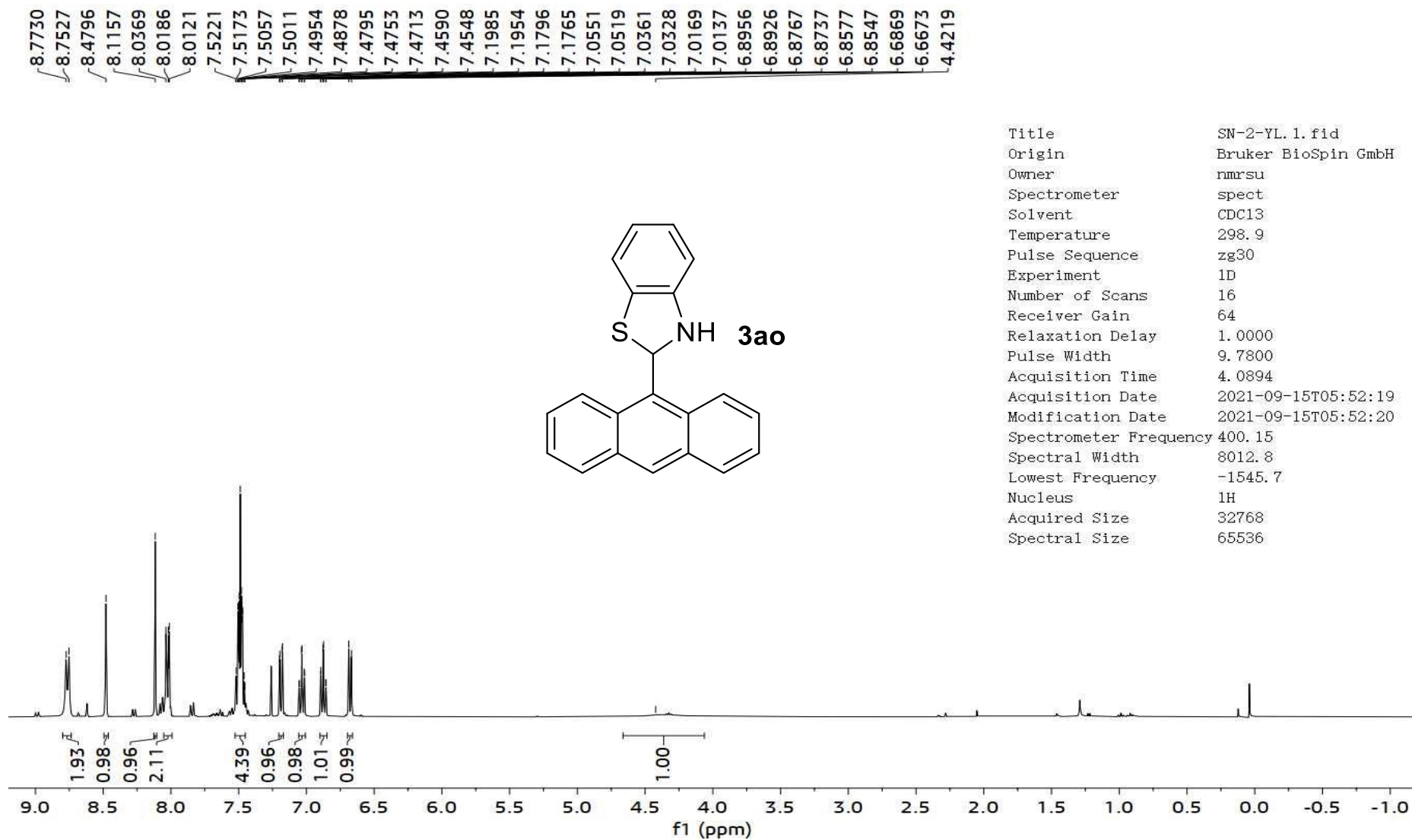
Title	WYX-71-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	39
Receiver Gain	188
Relaxation Delay	2.0000
Pulse Width	12.0000
Acquisition Time	0.7690
Acquisition Date	2021-07-08T15:10:41
Modification Date	2021-07-08T15:10:52
Spectrometer Frequency	176.07
Spectral Width	42613.6
Lowest Frequency	-3692.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

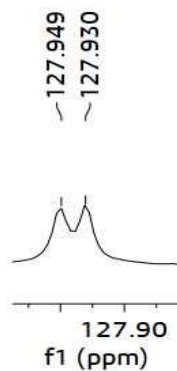
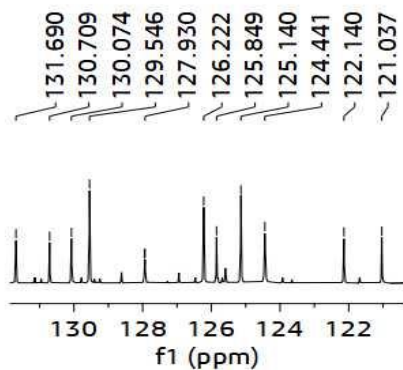
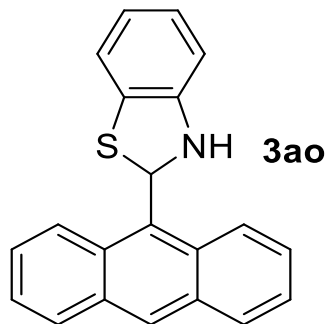


Title	WYX-72-yuanliao.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	1
Relaxation Delay	1.0000
Pulse Width	12.0000
Acquisition Time	2.3243
Acquisition Date	2021-07-08T15:14:46
Modification Date	2021-07-08T15:14:50
Spectrometer Frequency	700.15
Spectral Width	14097.7
Lowest Frequency	-2740.1
Nucleus	1H
Acquired Size	32768
Spectral Size	131072



Title	WYX-72-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	65
Receiver Gain	188
Relaxation Delay	2.0000
Pulse Width	12.0000
Acquisition Time	0.7690
Acquisition Date	2021-07-08T15:18:06
Modification Date	2021-07-08T15:18:14
Spectrometer Frequency	176.07
Spectral Width	42613.6
Lowest Frequency	-3718.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





— 65.65

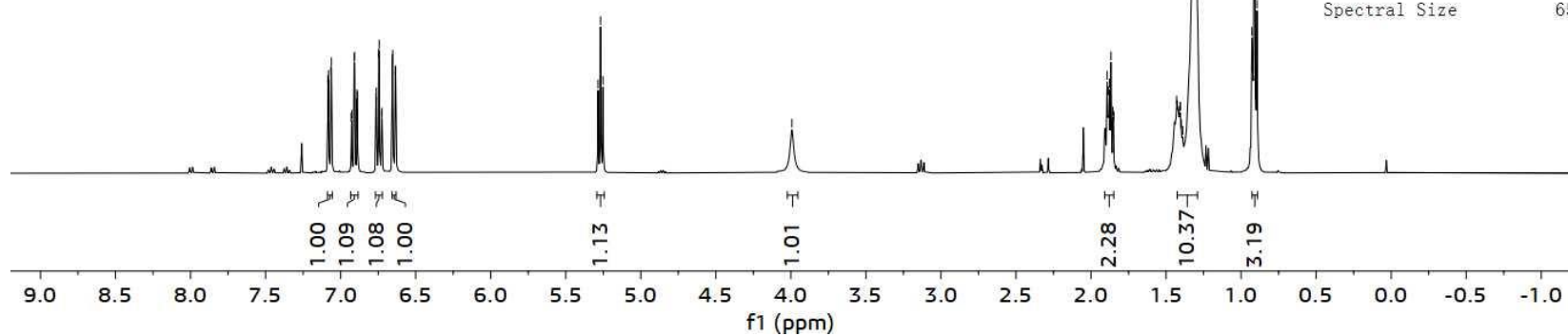
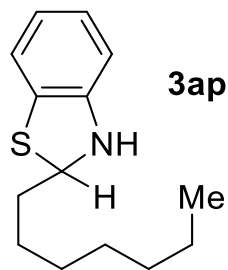
Title	SN-2-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	299.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-15T06:36:19
Modification Date	2021-09-15T06:36:20
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1950.0
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



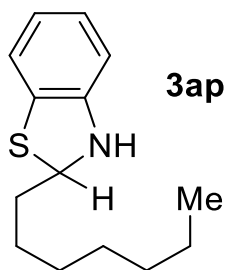
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6.7464
6.7434
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6.7246
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6.6527
6.6362
6.5837
5.2682
5.2520

— 3.9930

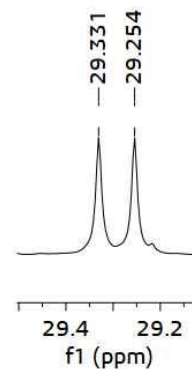
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1.8497
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1.4051
1.4048
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1.2914
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0.8939



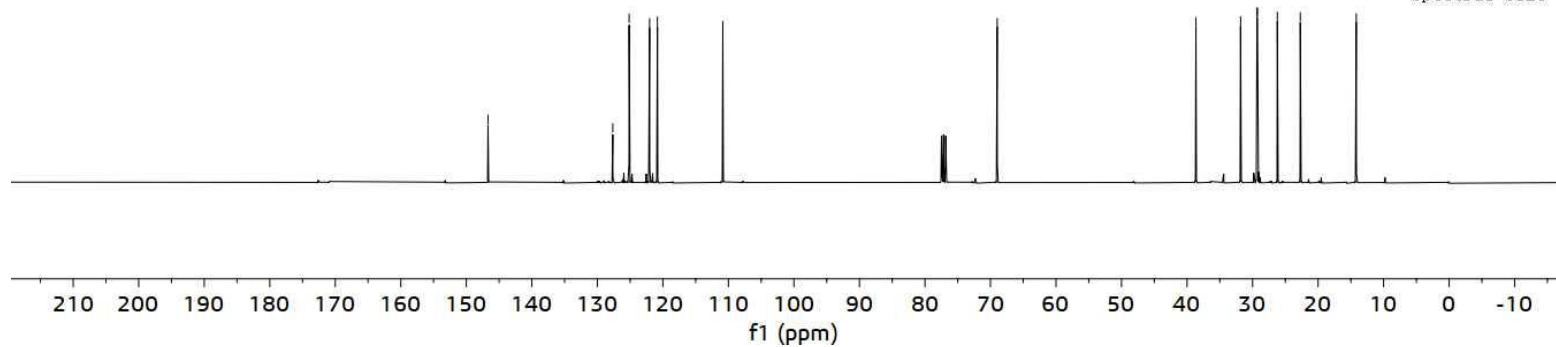
Title	SN-3-YL. 1. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	17
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-12T00:42:37
Modification Date	2021-10-12T00:42:38
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



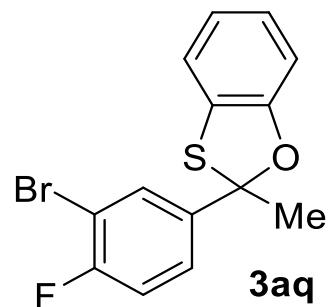
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 —127.64
 —125.15
 —122.03
 —120.83
 —110.86
 —68.98
 —38.65
 —31.83
 —29.33
 —29.25
 —26.22
 —22.71
 —14.18



Title	SN-3-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T01:23:40
Modification Date	2021-10-12T01:23:42
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1953.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

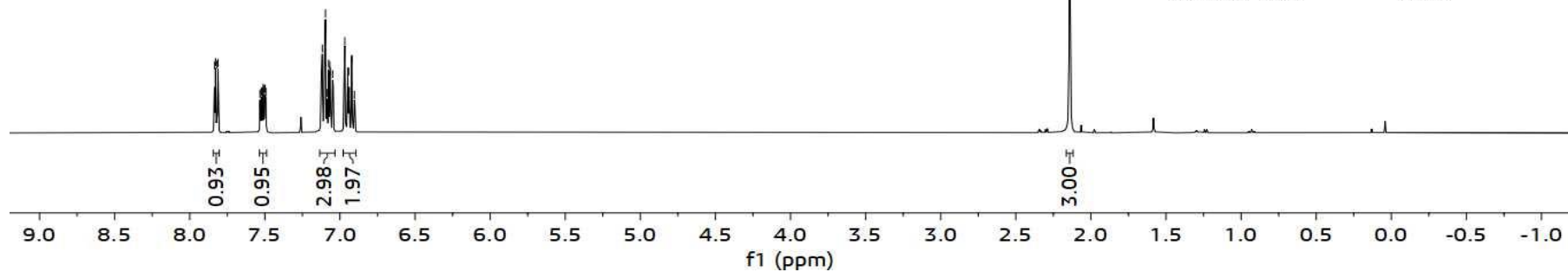


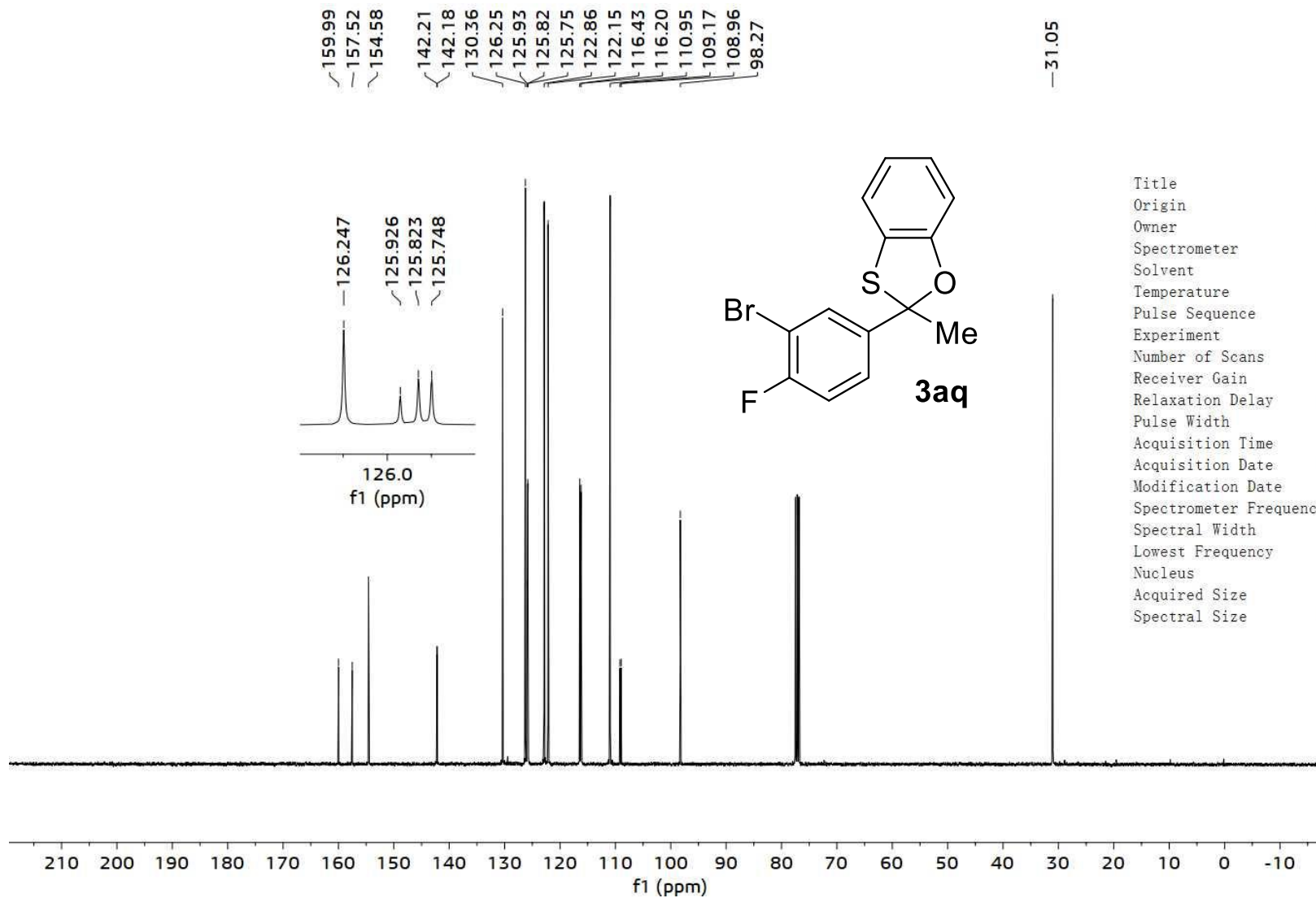
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7.0965
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6.9486
6.9416
6.9203
6.9039



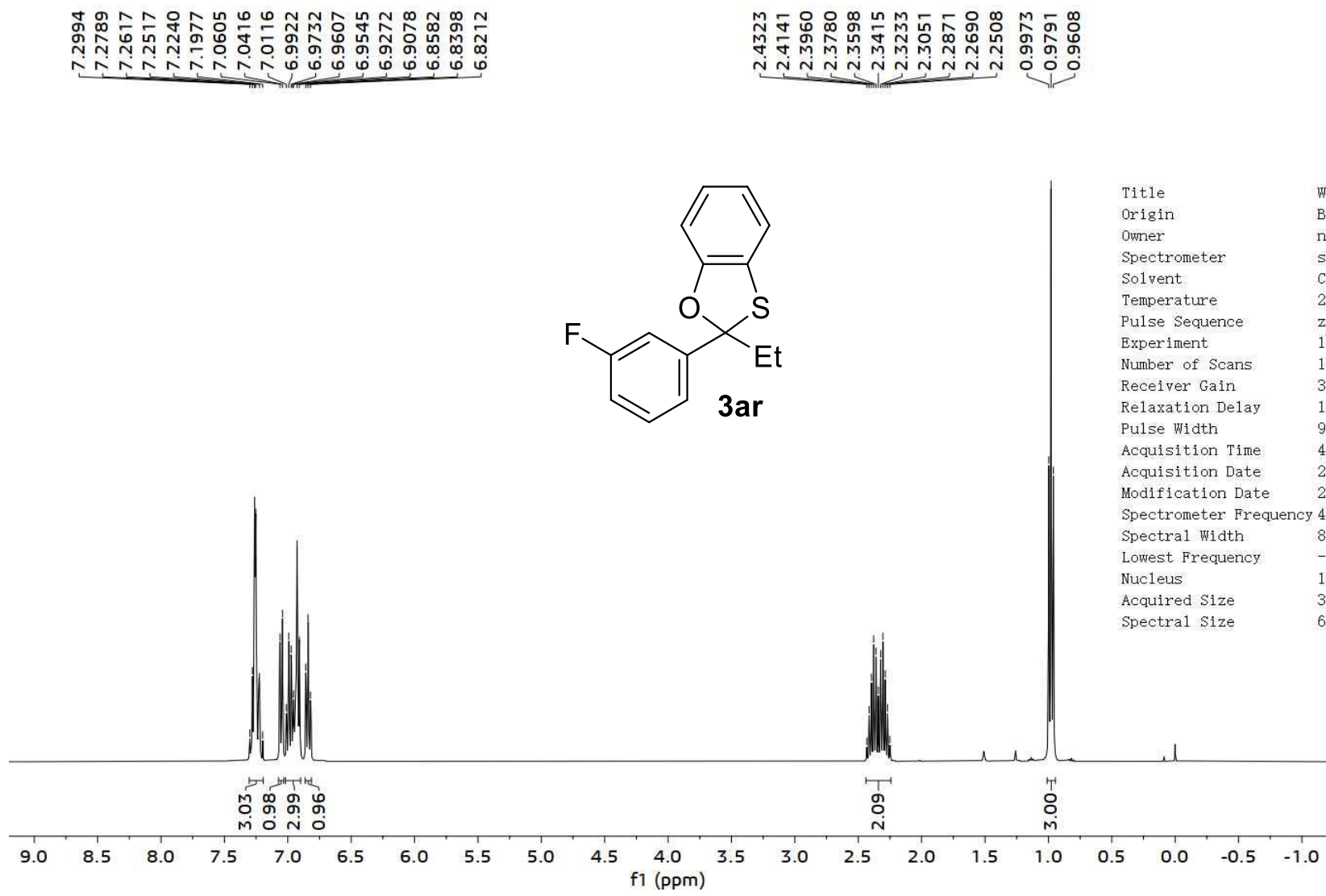
-2.1411

Title	WYX-91-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-15T23:52:13
Modification Date	2021-09-15T23:52:14
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

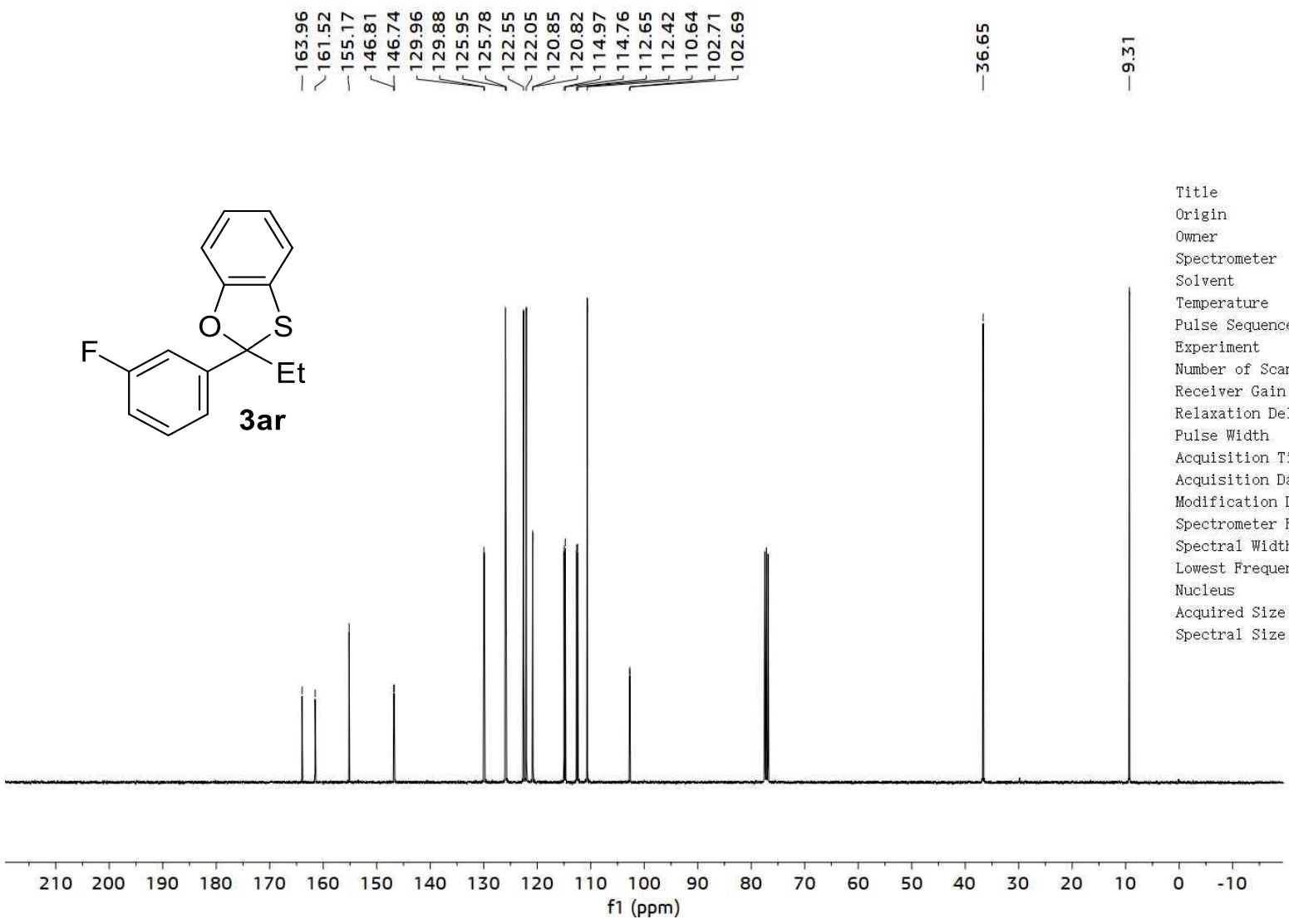
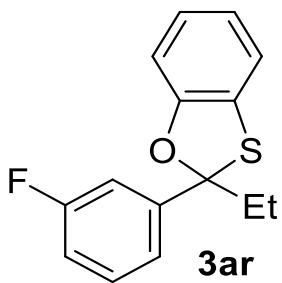




Title	WYX-91-YL.3.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	550
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-16T00:24:43
Modification Date	2021-09-16T00:24:44
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1952.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



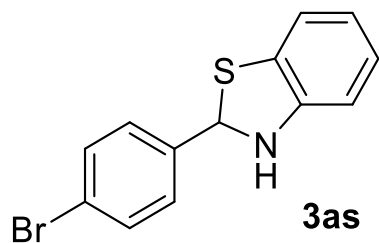
Title	WYX-93-YL. 2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-16T01:07:09
Modification Date	2021-09-16T01:07:10
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1570.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



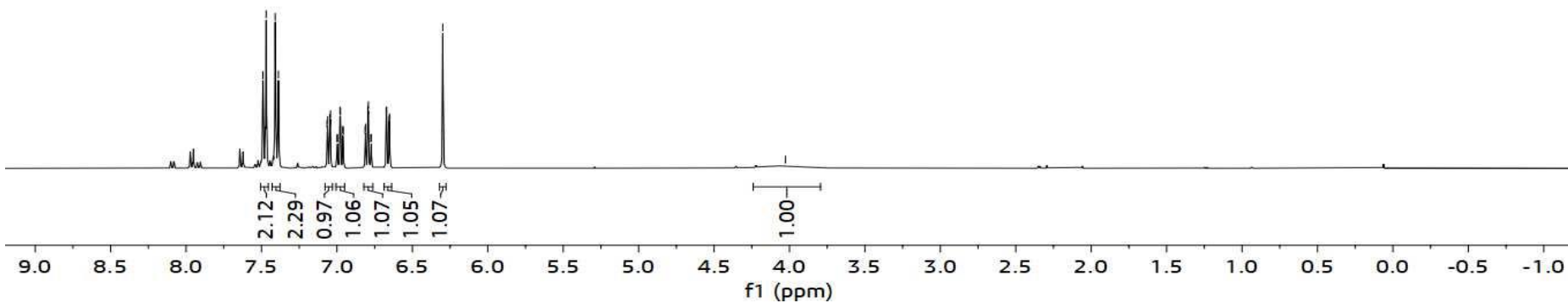
Title	WYX-93-YL. 3. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.3
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	550
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-16T01:39:39
Modification Date	2021-09-16T01:39:40
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1953.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

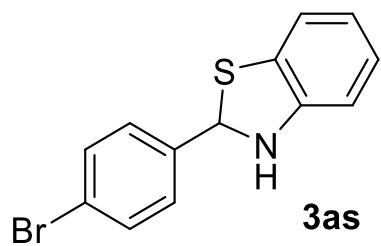
7.4904
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7.4096
7.3883
7.0654
7.0622
7.0464
7.0431
6.9992
6.9960
6.9800
6.9768
6.9610
6.9577
6.8132
6.8103
6.7945
6.7915
6.7756
6.7726
6.6734
6.6705
6.6542
6.6509
6.2985

—4.0268



Title	SN-1-yuanliao.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	301.7
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-08-02T15:23:39
Modification Date	2021-08-02T15:23:40
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.7
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

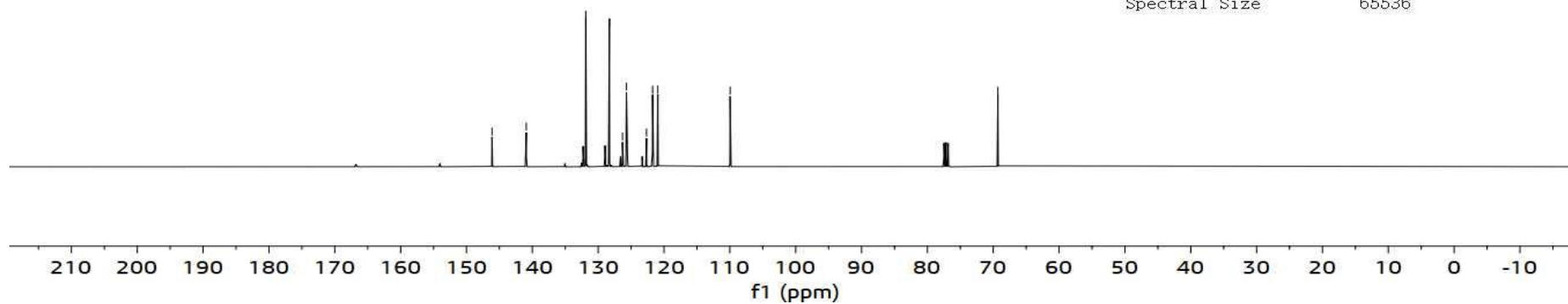


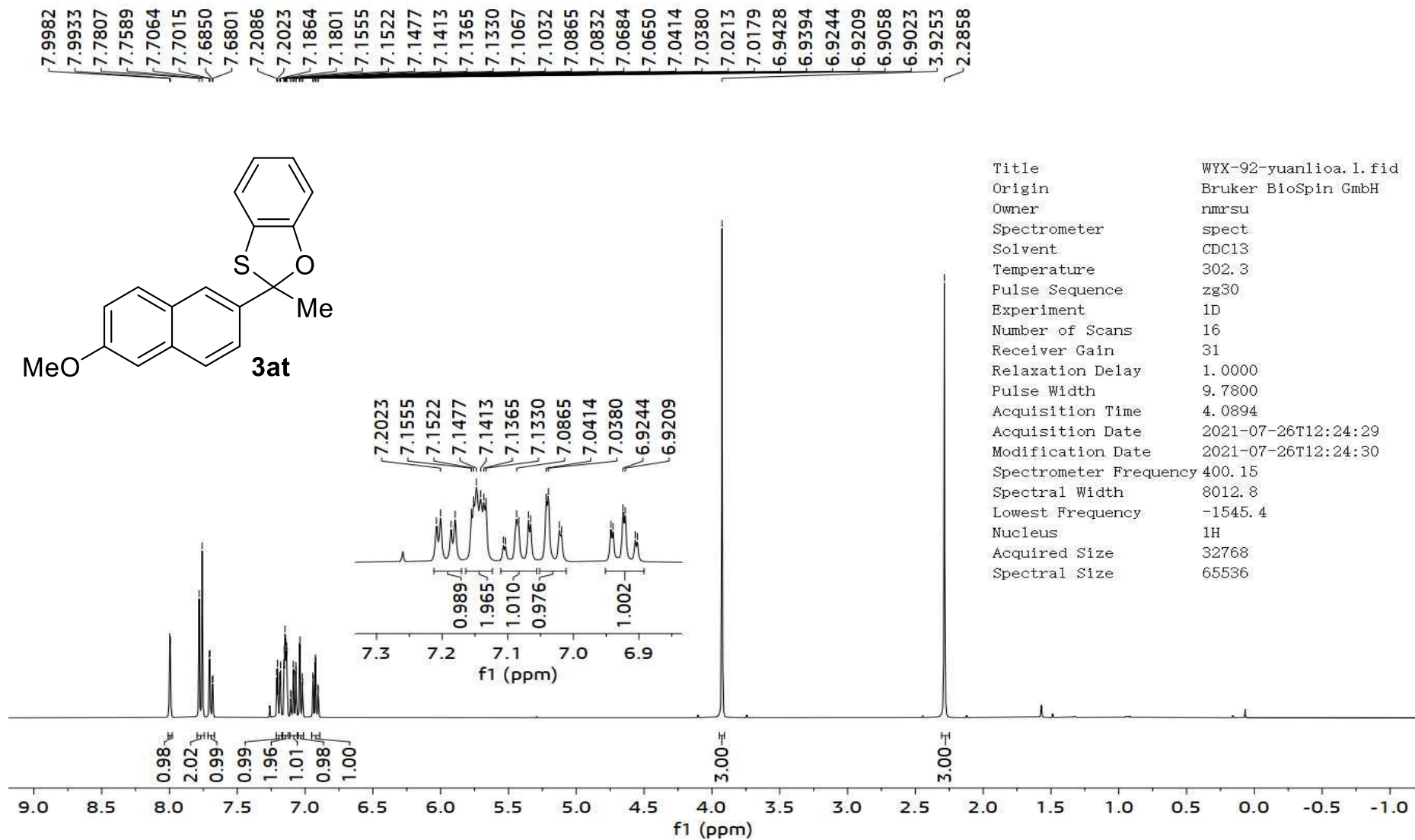


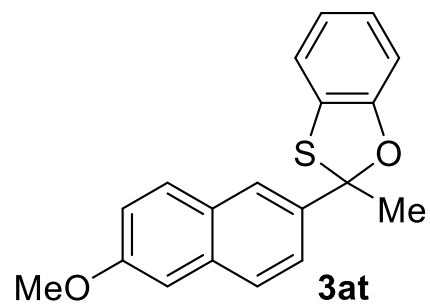
146.12
 140.92
 131.88
 128.30
 126.31
 125.71
 122.66
 121.73
 120.94
 -109.95

-69.30

Title	SN-1-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	302.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-08-02T16:23:13
Modification Date	2021-08-02T16:23:14
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1959.9
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536





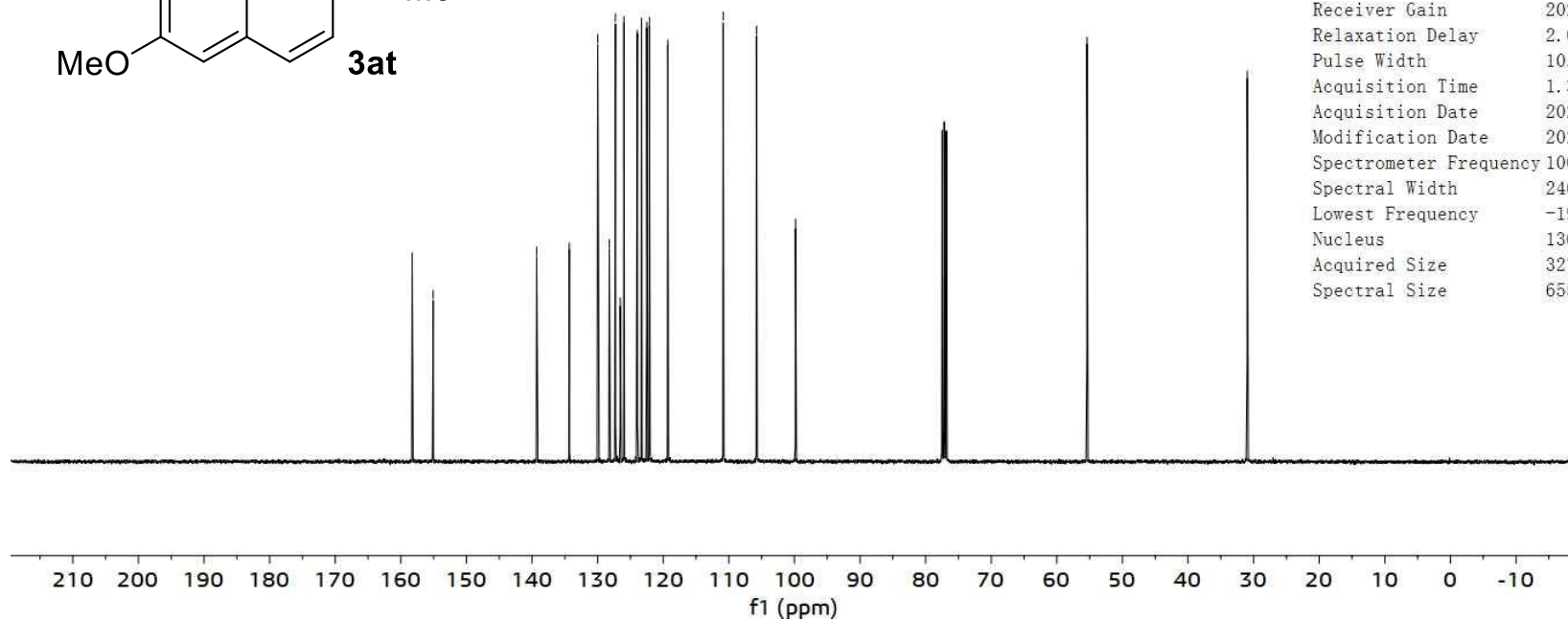


158.27
 155.07
 139.30
 134.32
 129.99
 128.23
 127.29
 126.55
 125.99
 123.95
 123.31
 122.51
 122.12
 119.30
 110.87
 105.77
 99.84

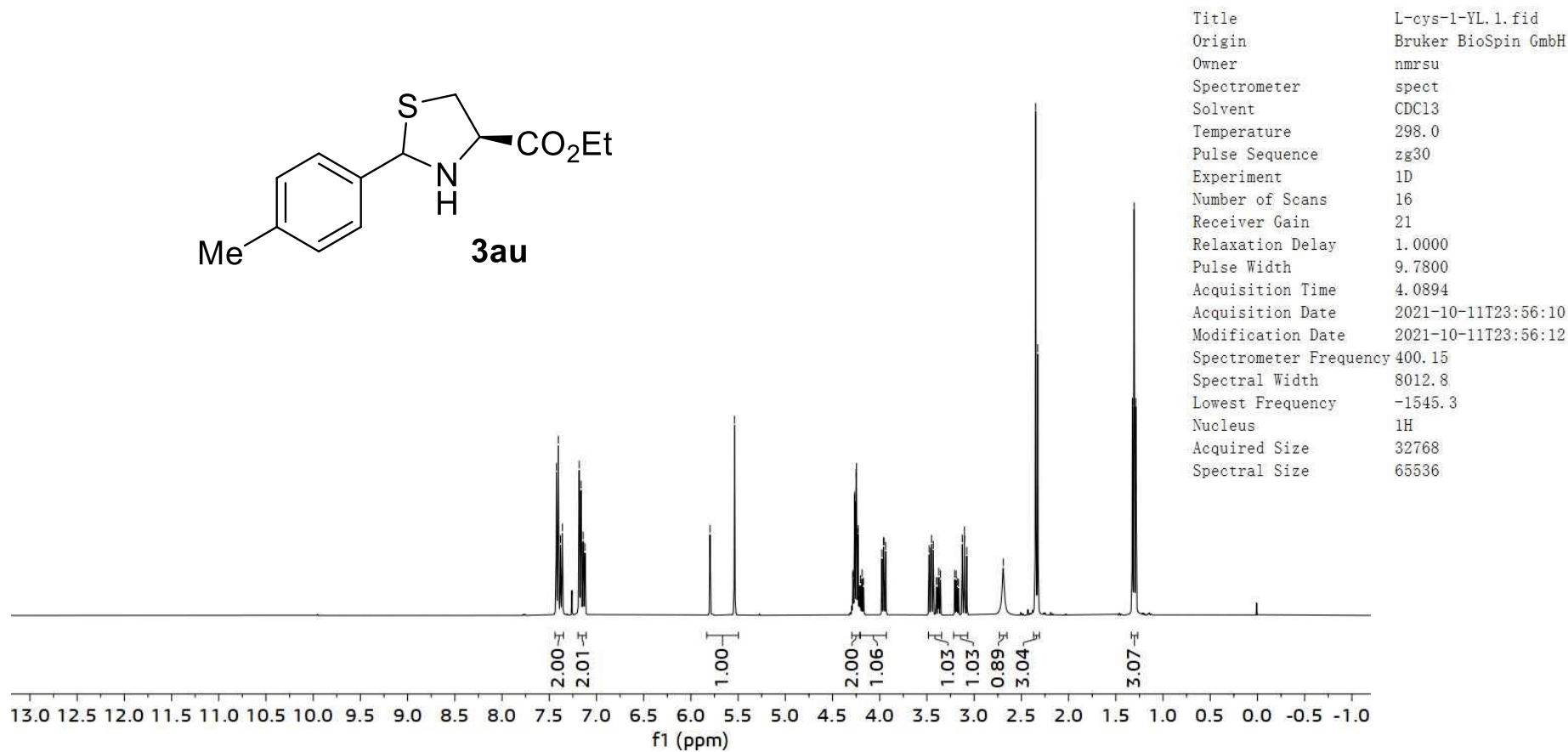
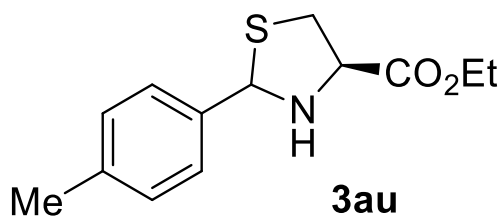
55.40

30.98

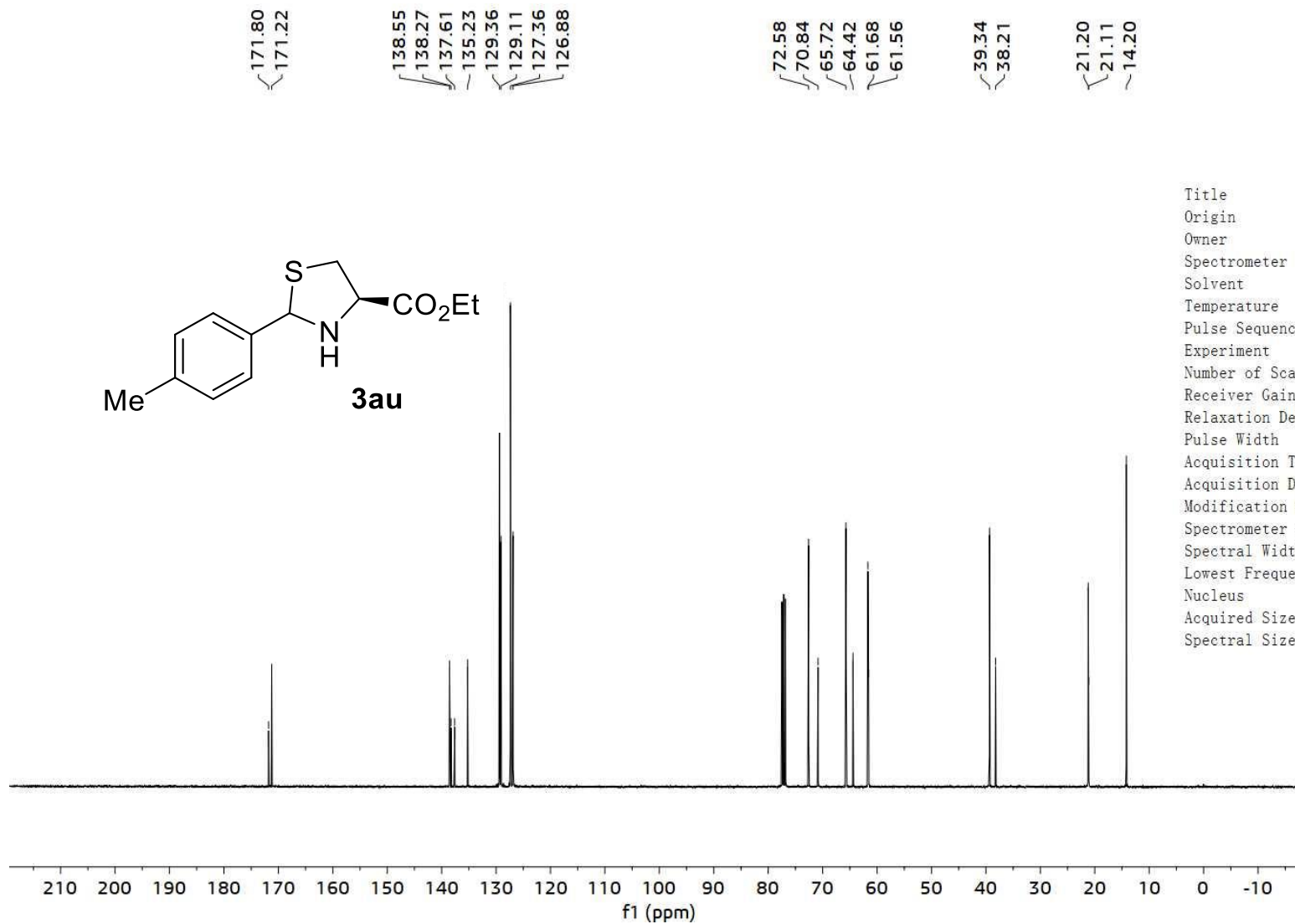
Title	WYX-92-yuanliao.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	303.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-26T13:08:25
Modification Date	2021-07-26T13:08:26
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1953.0
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



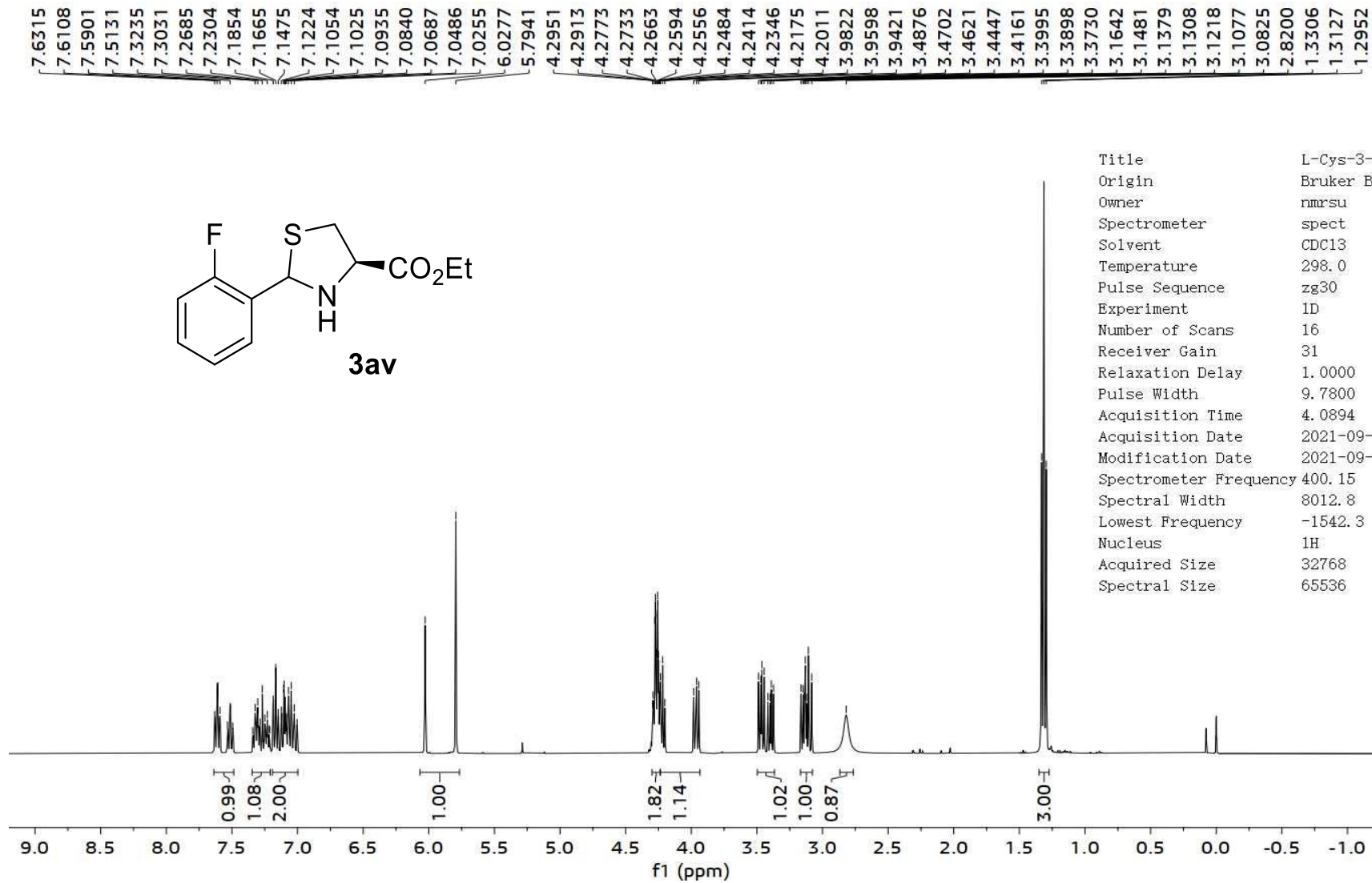
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7.3813
7.3612
7.1832
7.1634
7.1417
7.1218
5.7976
5.5373
4.2859
4.2816
4.2681
4.2641
4.2505
4.2464
4.2332
4.2289
4.2191
4.2157
4.2115
4.2036
4.1877
4.1715
3.9772
3.9593
3.9548
3.9371
3.4767
3.4589
3.4510
3.4333
3.4010
3.3831
3.3746
3.3567
3.2060
3.1915
3.1796
3.1650
3.1263
3.1039
3.1005
3.0785
2.6924
2.3489
2.3273
1.3243
1.3199
1.3064
1.3032
1.2885
1.2845

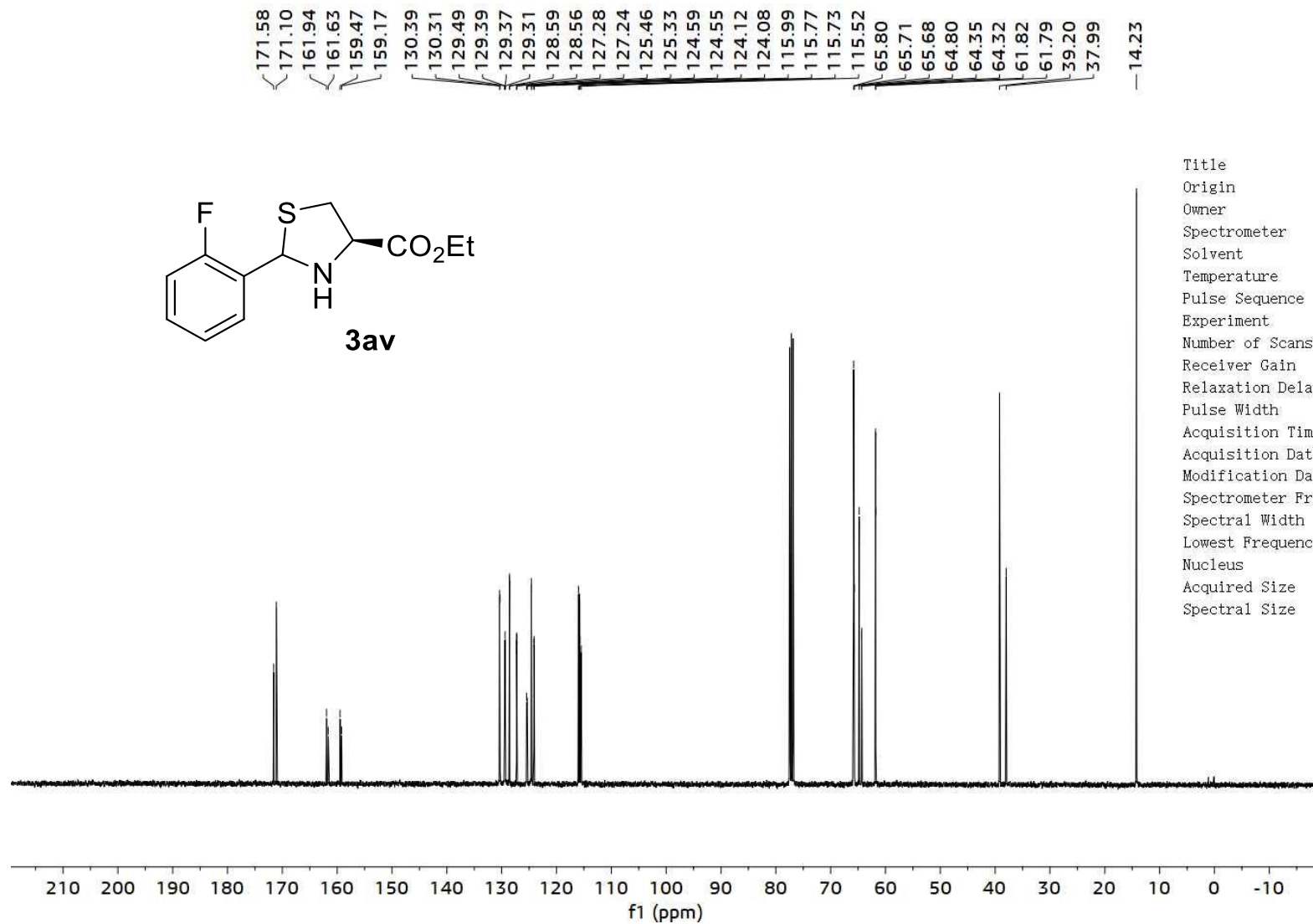


Title	L-cys-1-YL.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	21
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-11T23:56:10
Modification Date	2021-10-11T23:56:12
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.3
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

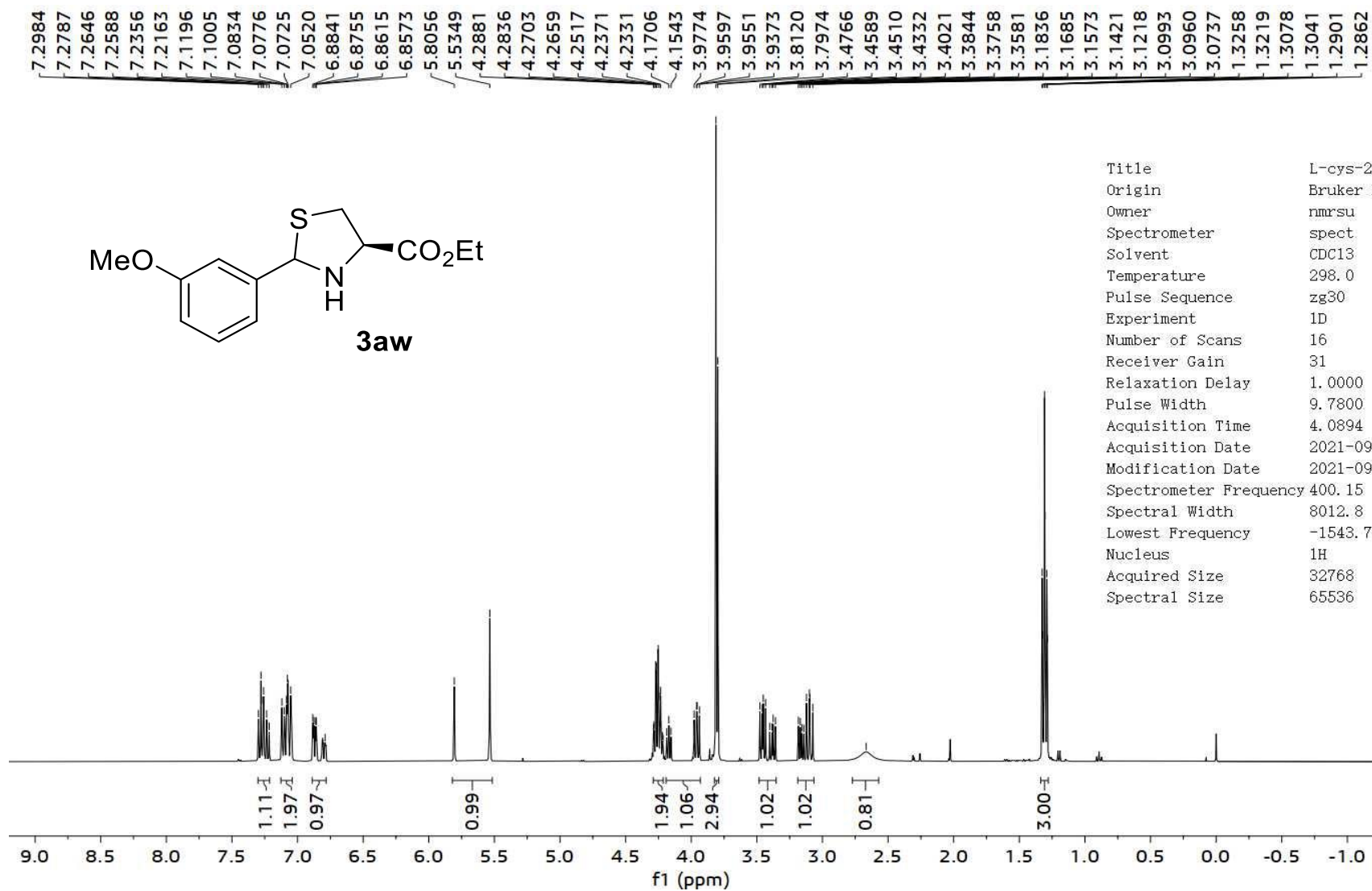


Title	L-cys-1-YL.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T00:37:13
Modification Date	2021-10-12T00:37:14
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1957.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

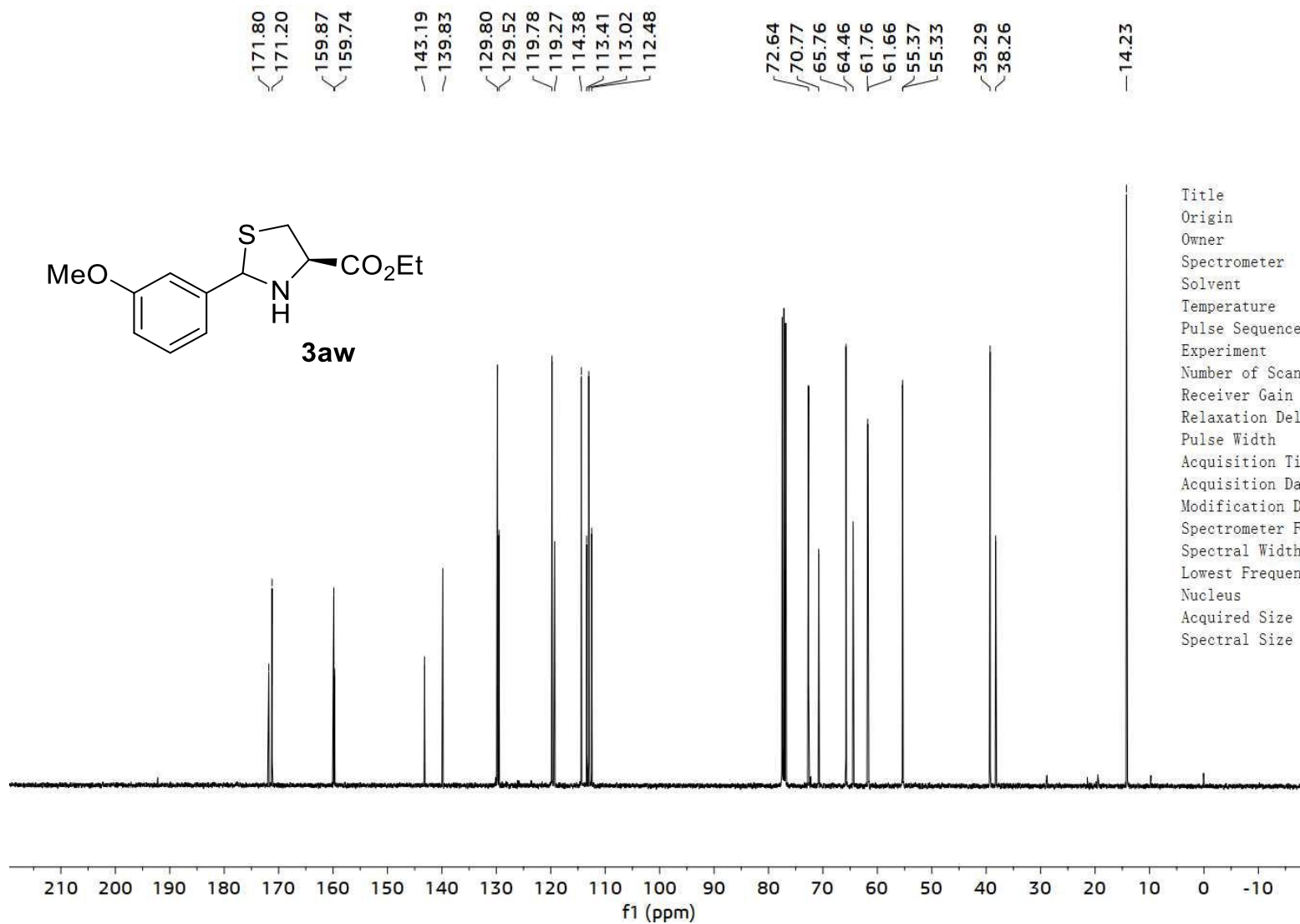




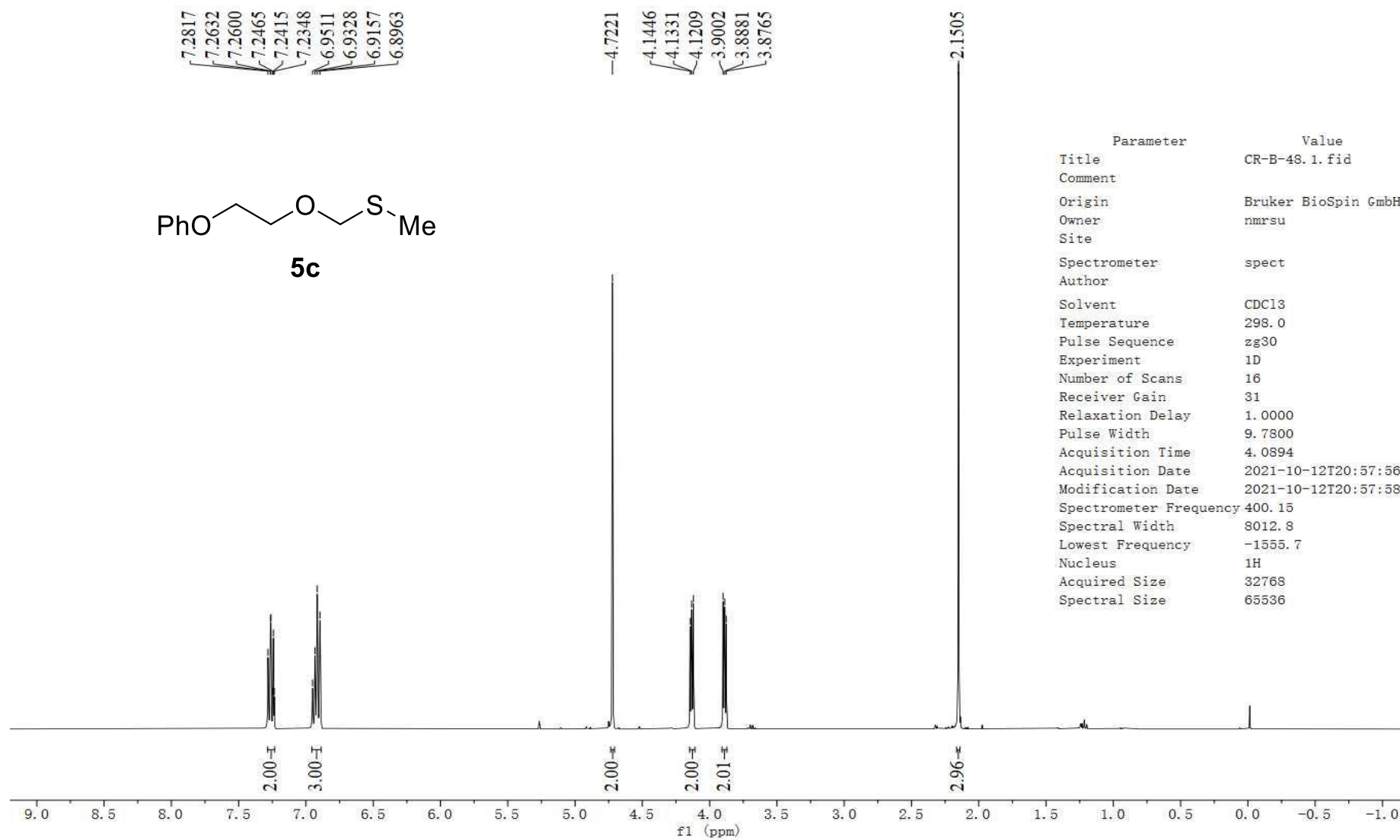
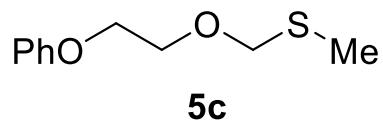
Title	L-Cys-3-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-23T23:58:46
Modification Date	2021-09-23T23:58:46
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1951.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



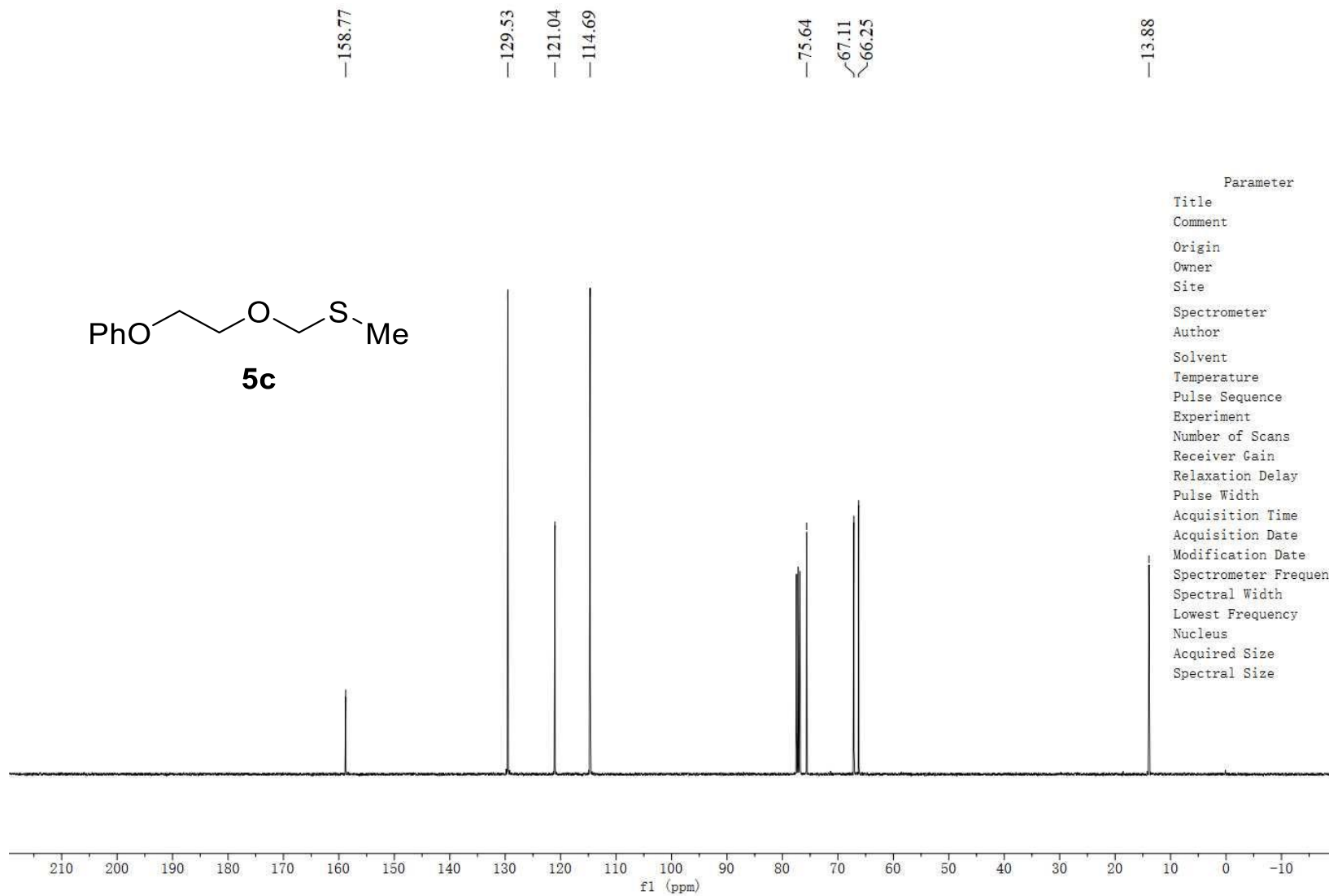
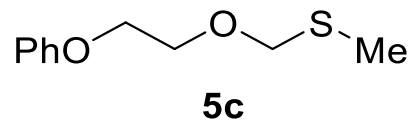
Title	L-cys-2.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-22T23:37:58
Modification Date	2021-09-22T23:37:58
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1543.7
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



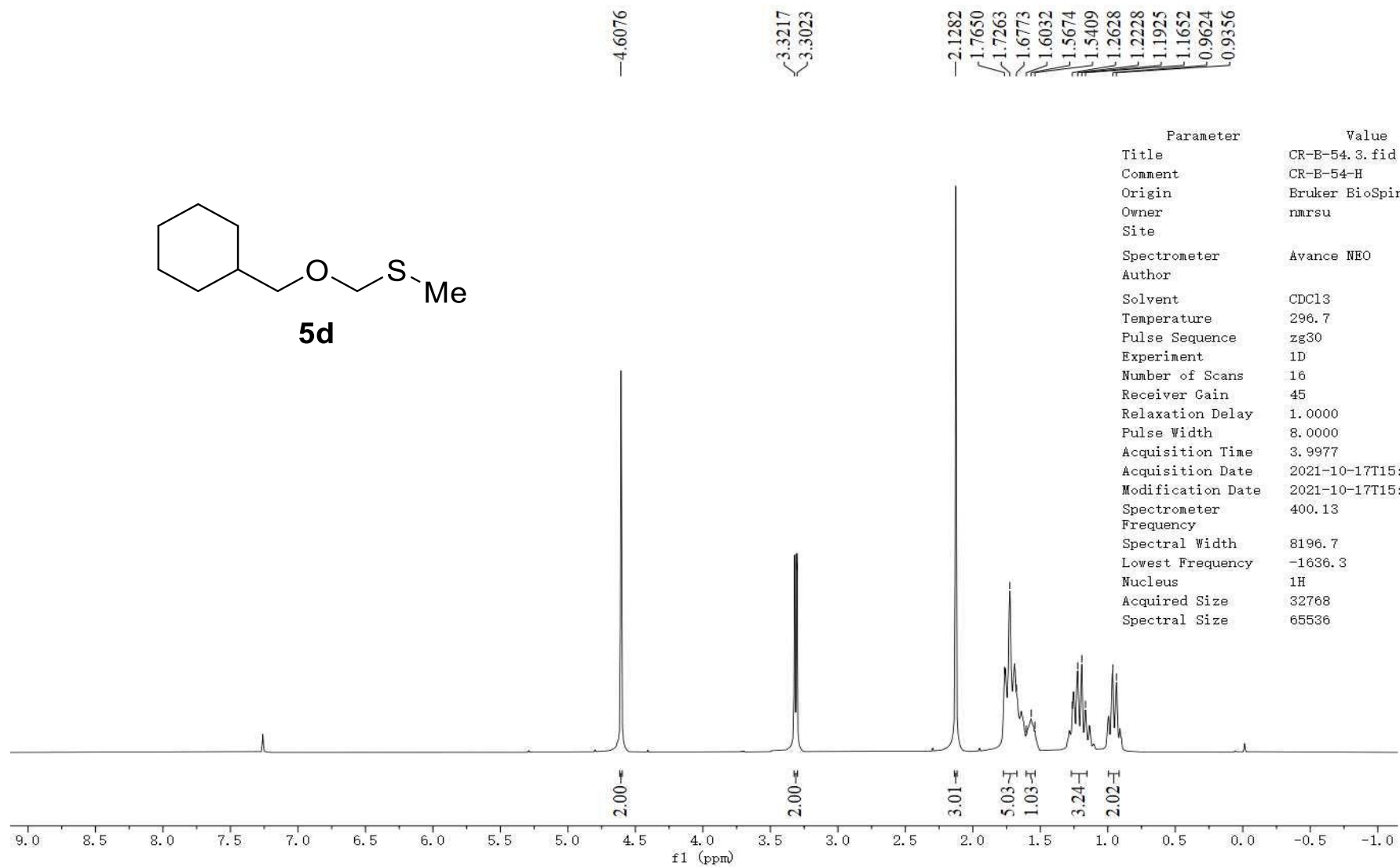
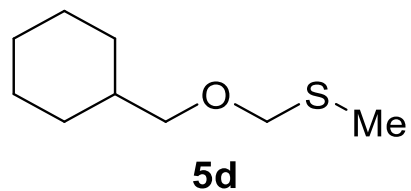
Title	L-cys-2.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-23T00:37:33
Modification Date	2021-09-23T00:37:32
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1952.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



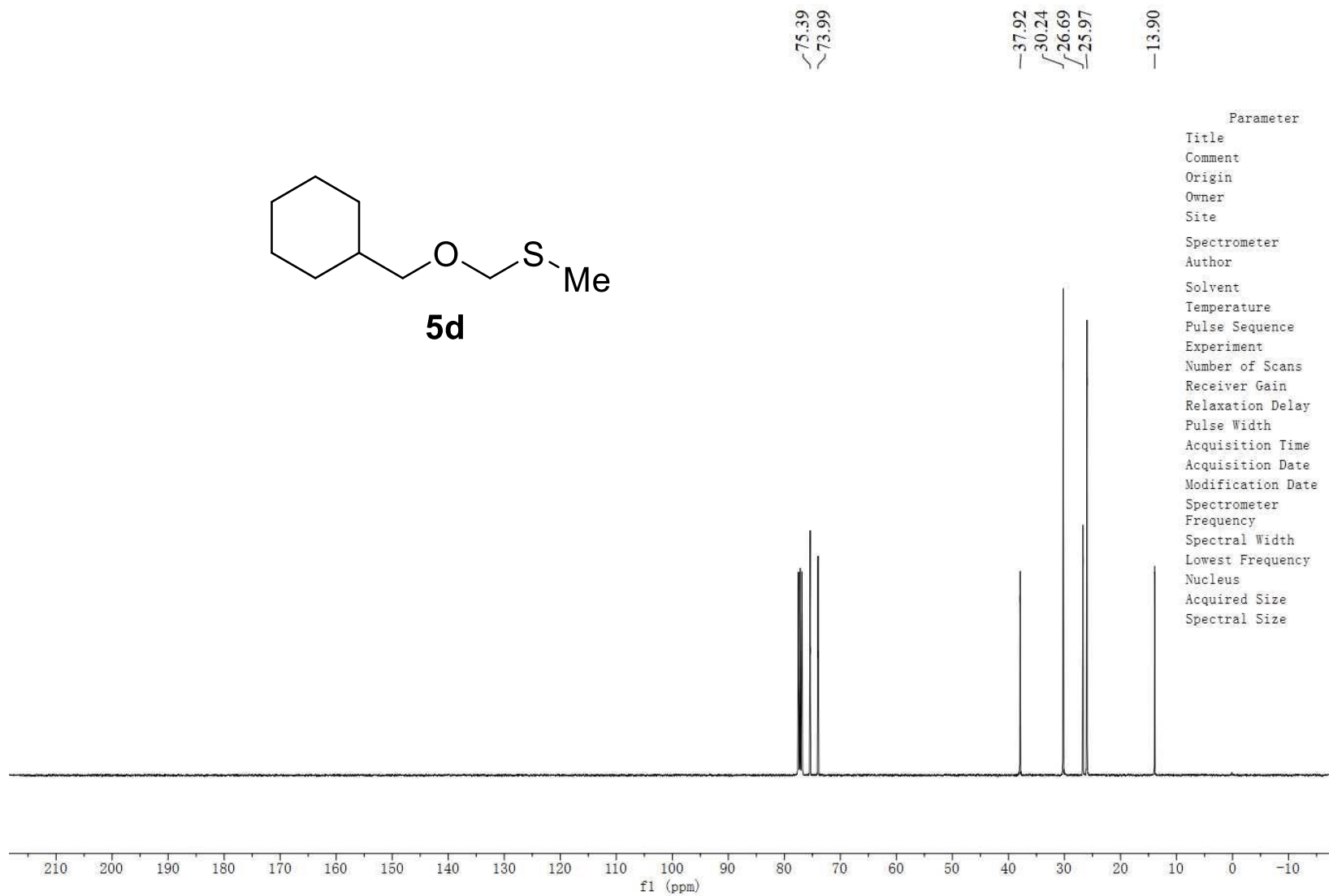
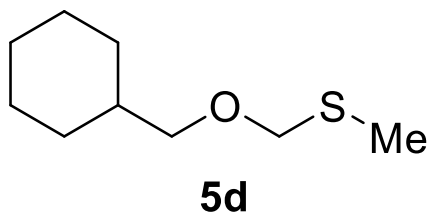
Parameter	Value
Title	CR-B-48. 1. fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-12T20:57:56
Modification Date	2021-10-12T20:57:58
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1555.7
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



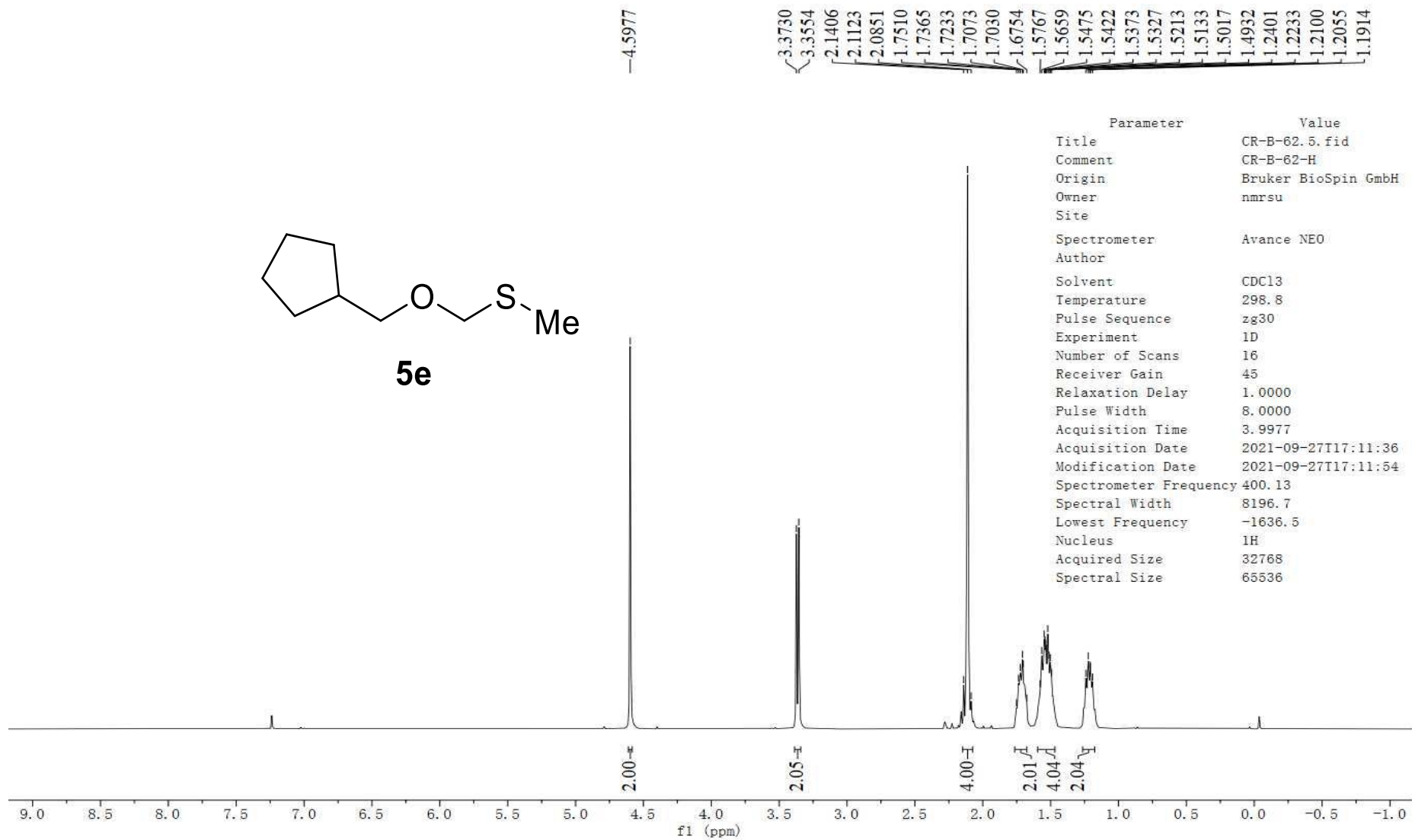
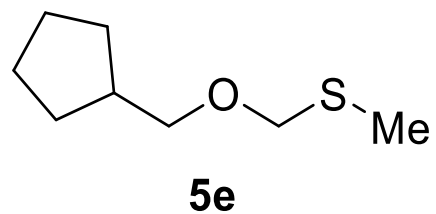
Parameter	Value
Title	CR-B-48.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T21:33:16
Modification Date	2021-10-12T21:33:18
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1949.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

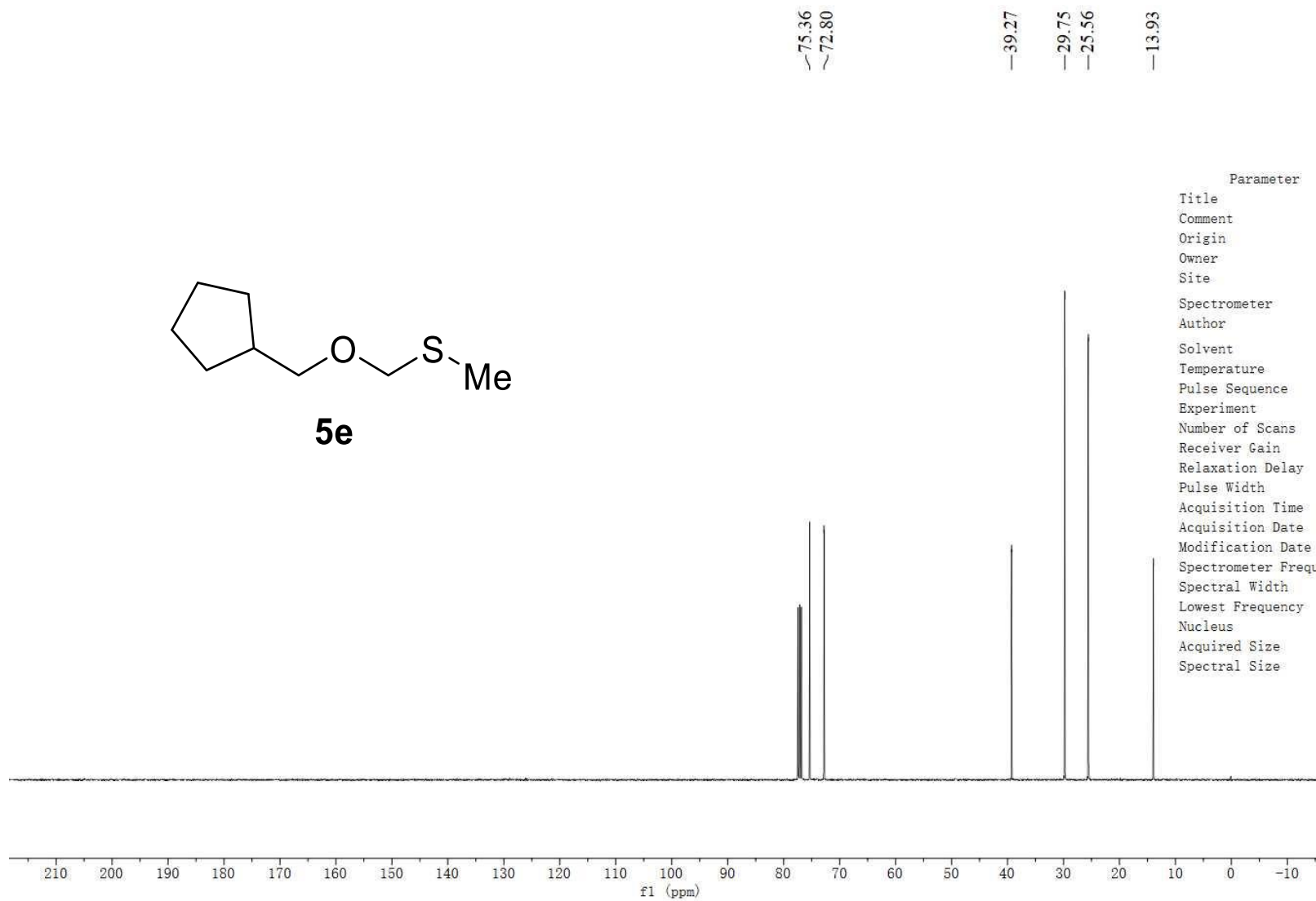
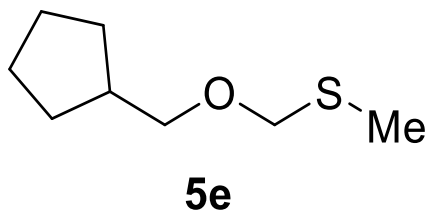


Parameter	Value
Title	CR-B-54.3.fid
Comment	CR-B-54-H
Origin	Bruker BioSpin GmbH
Owner	mrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	296.7
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	45
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-10-17T15:12:30
Modification Date	2021-10-17T15:11:50
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

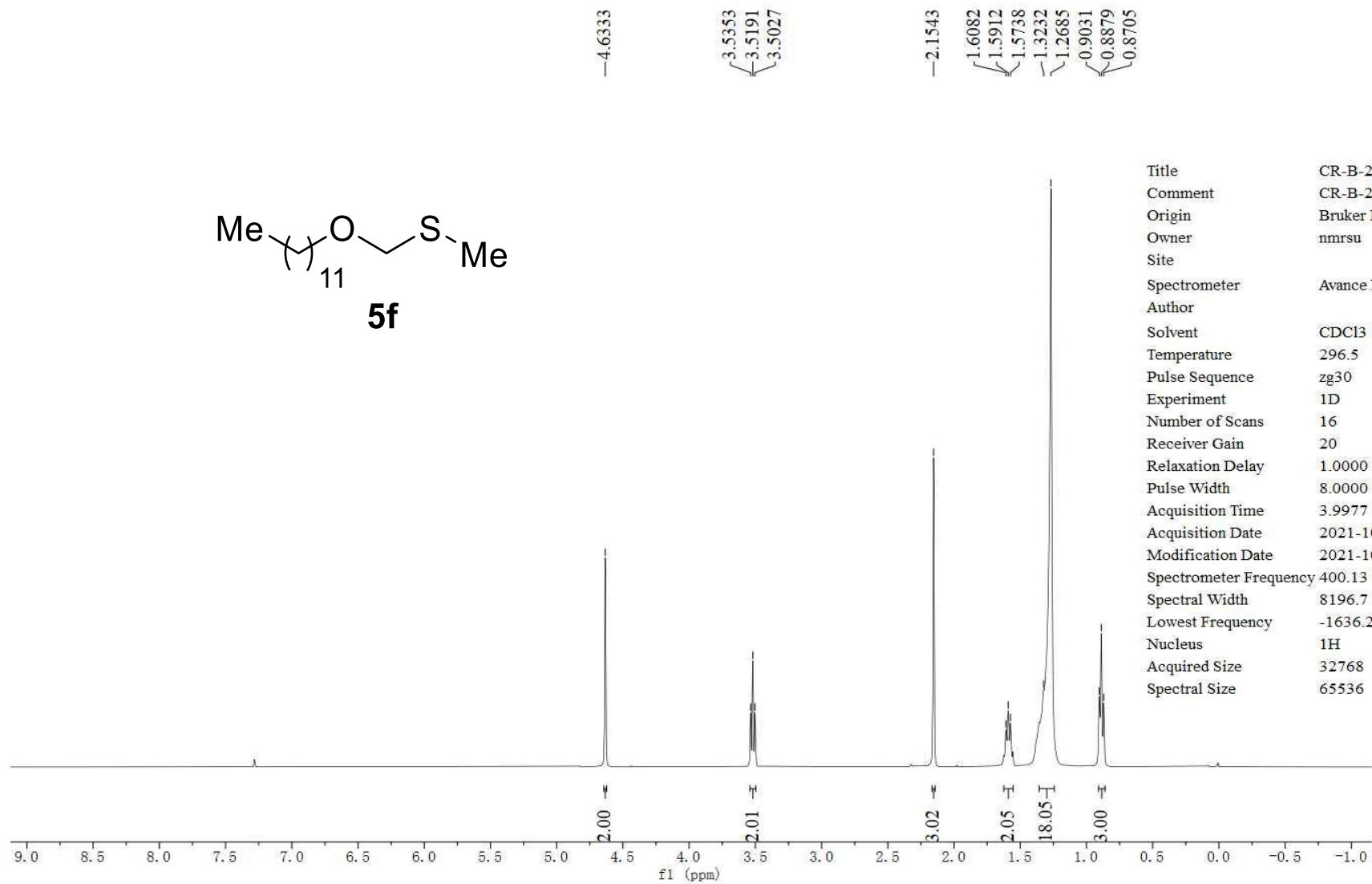
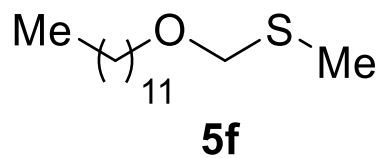


Parameter	Value
Title	CR-B-54. 4. fid
Comment	CR-B-54-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	297.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-10-17T16:11:46
Modification Date	2021-10-17T16:11:06
Spectrometer	100.62
Frequency	
Spectral Width	23809.5
Lowest Frequency	-1831.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

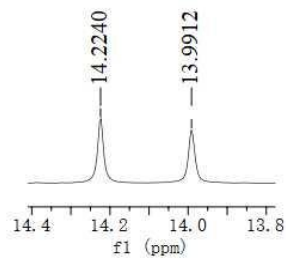
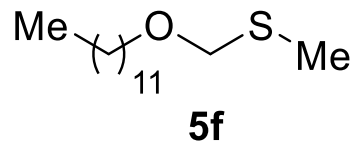




Parameter	Value
Title	CR-B-62.6.fid
Comment	CR-B-62-C
Origin	Bruker BioSpin GmbH
Owner	nmsru
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-27T18:11:06
Modification Date	2021-09-27T18:11:24
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1831.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

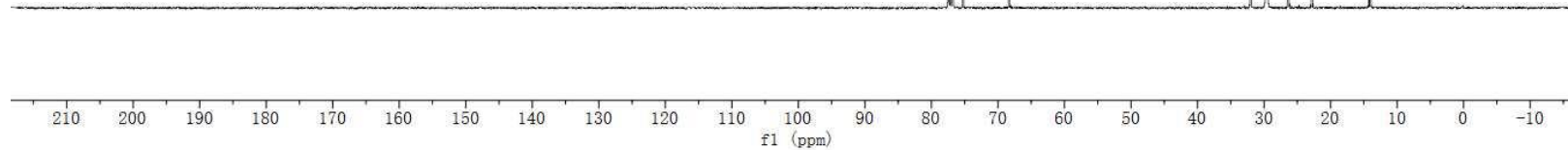


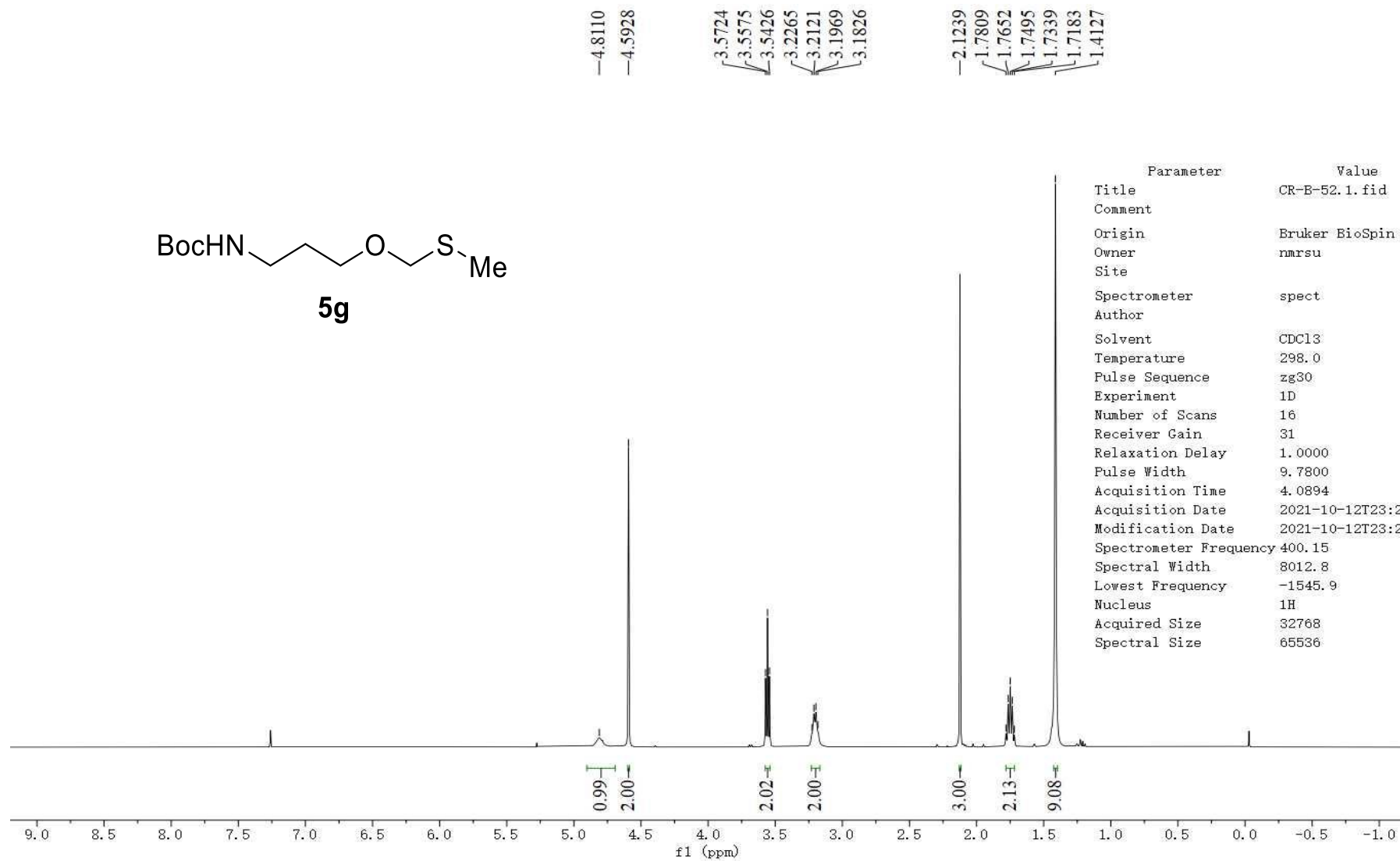
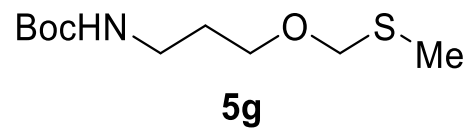
Title	CR-B-24.1.fid
Comment	CR-B-24-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	296.5
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	20
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-10-17T14:07:58
Modification Date	2021-10-17T14:07:18
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1636.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



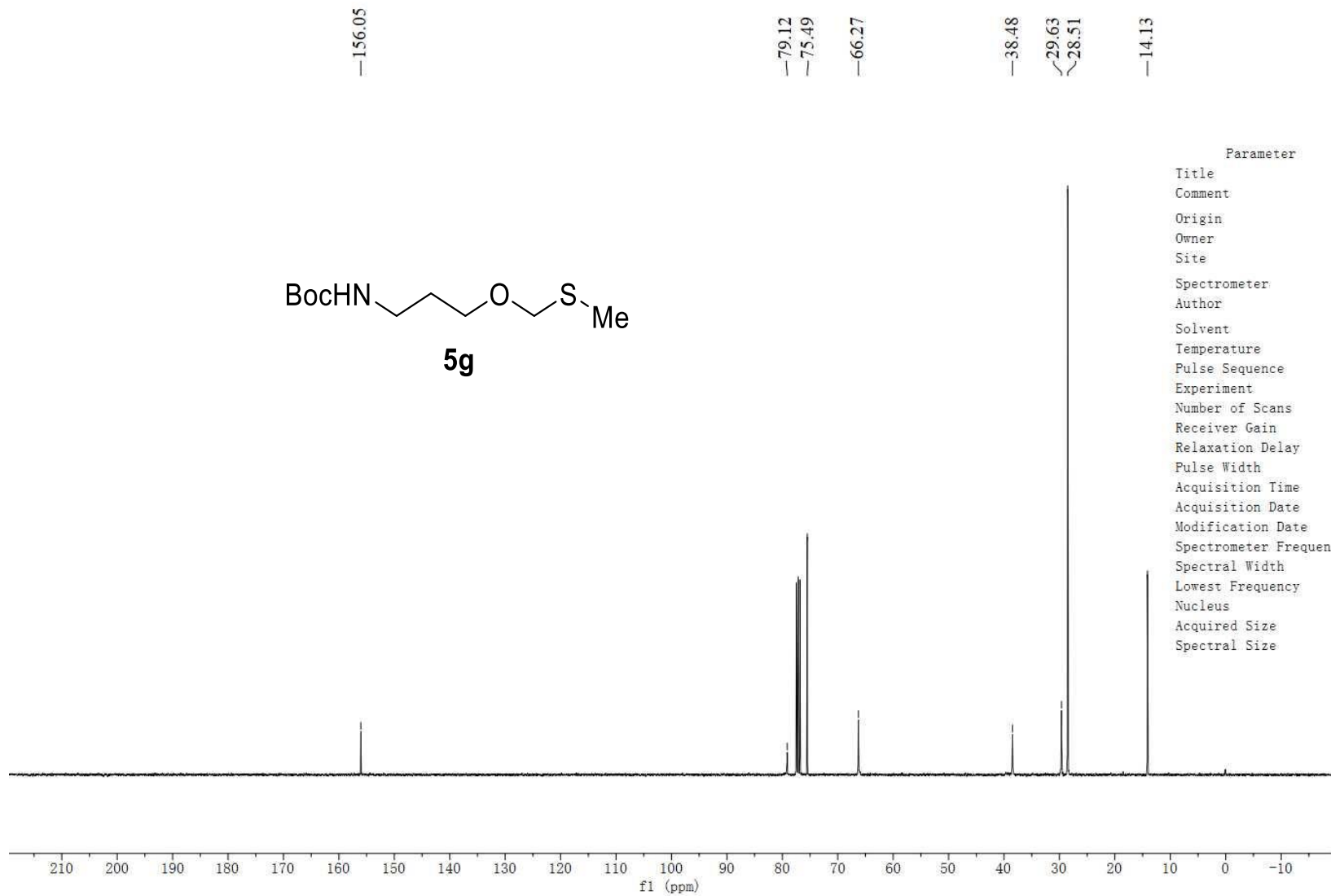
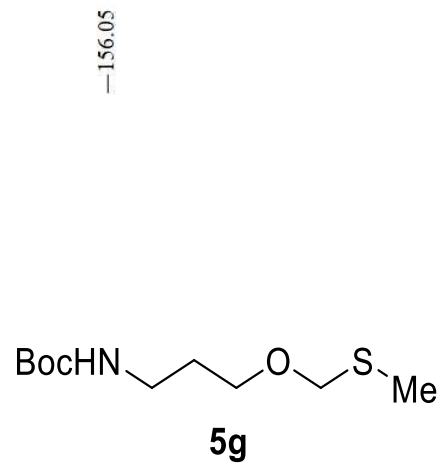
— 75.25
 — 68.32
 32.04
 29.78
 29.75
 29.72
 29.56
 29.52
 29.47
 26.37
 22.81
 14.22
 13.99

Title	CR-B-24.2.fid
Comment	CR-B-24-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	297.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-10-17T15:07:17
Modification Date	2021-10-17T15:06:36
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1831.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

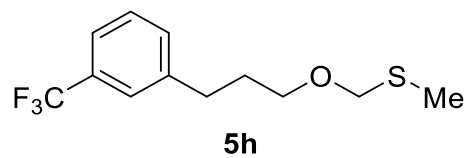




Parameter	Value
Title	CR-B-52.1.fid
Comment	
Origin	Eruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-12T23:23:38
Modification Date	2021-10-12T23:23:40
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.9
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Title	CR-B-52.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl ₃
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T23:58:59
Modification Date	2021-10-12T23:59:00
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1947.8
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



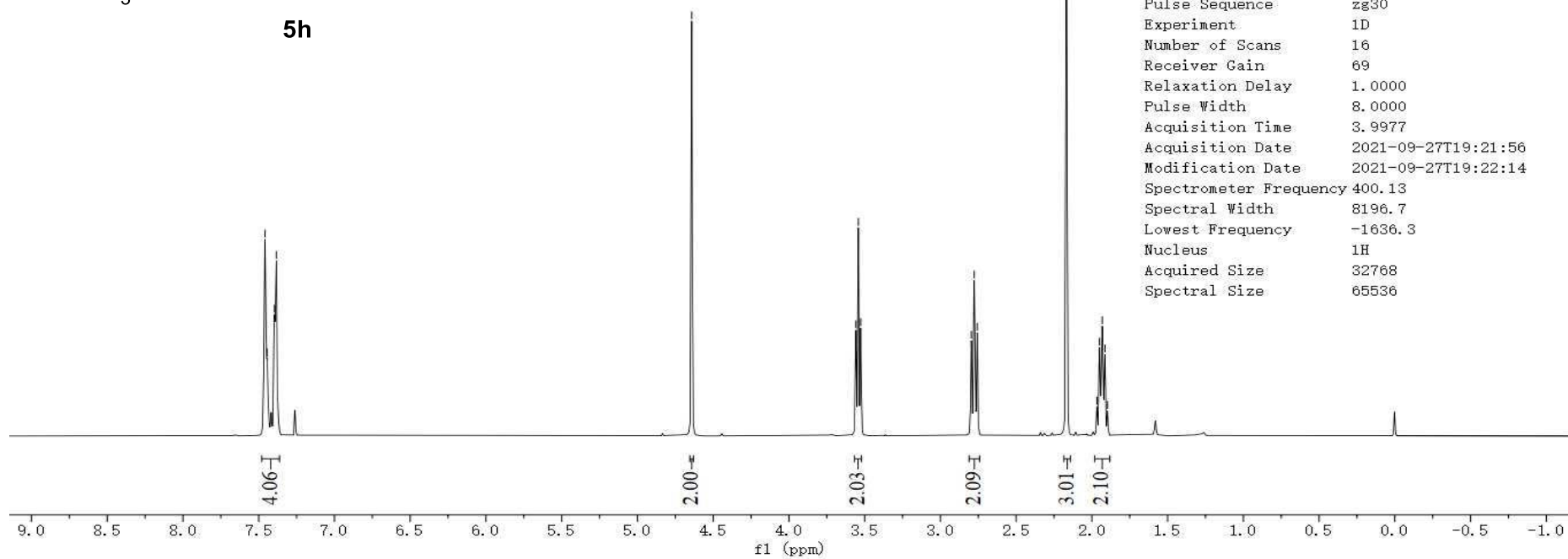
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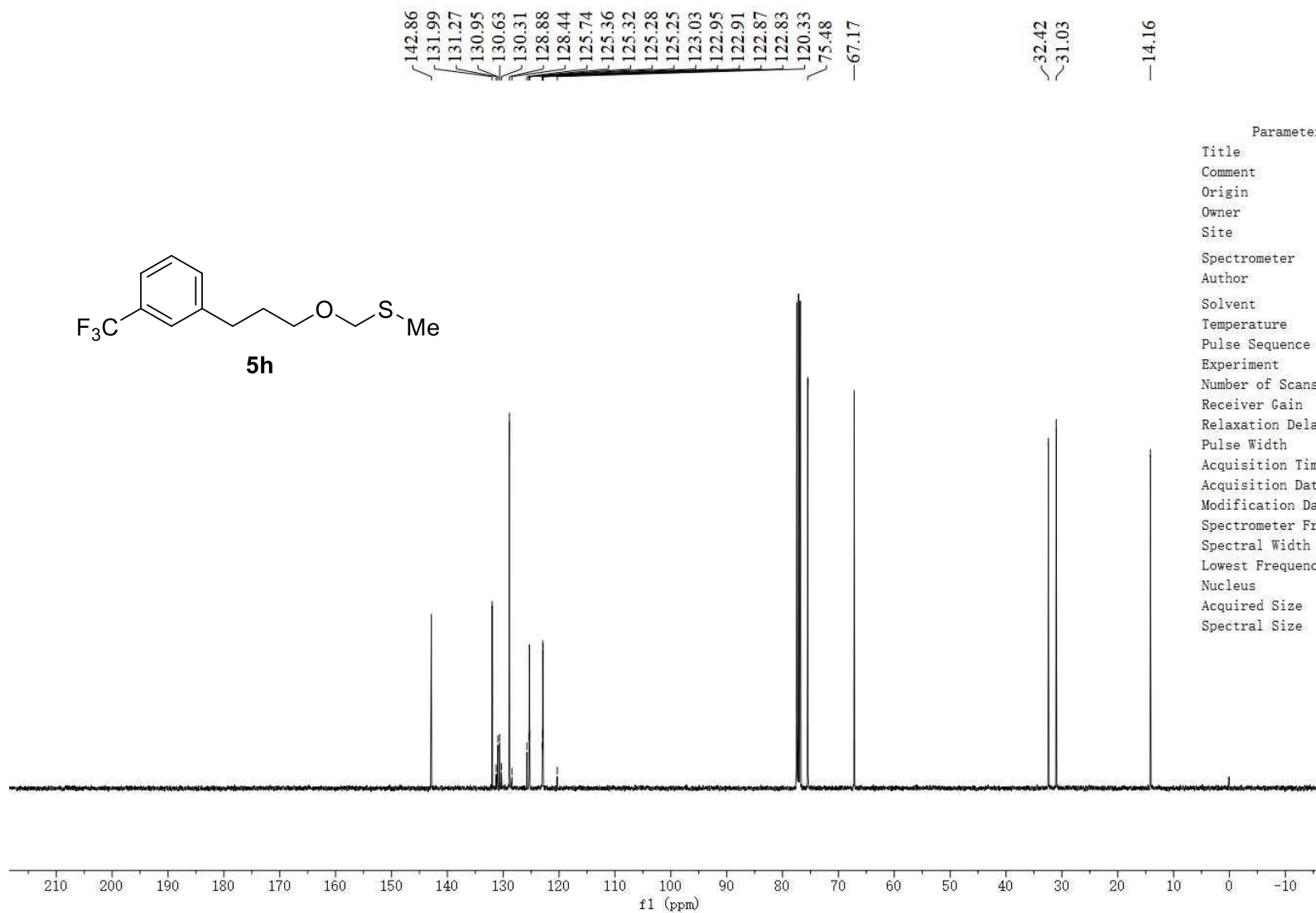
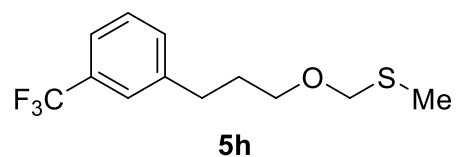
4.6417

3.5570
3.5417
3.5263

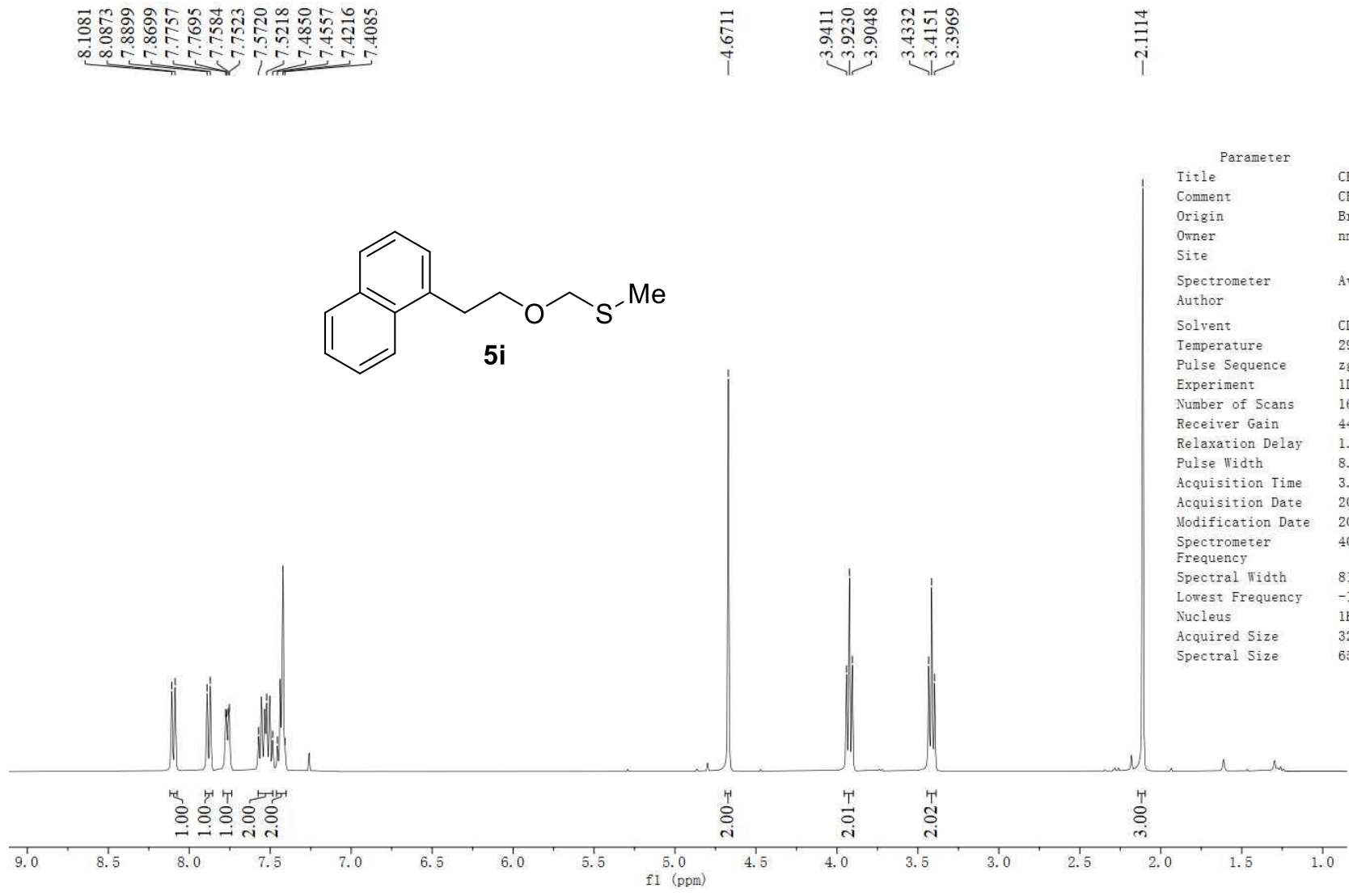
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2.7570
2.1682
1.9663
1.9498
1.9315
1.9138
1.8969

Parameter	Value
Title	CR-B-60.9.fid
Comment	CR-B-60-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	298.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	69
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-27T19:21:56
Modification Date	2021-09-27T19:22:14
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1636.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

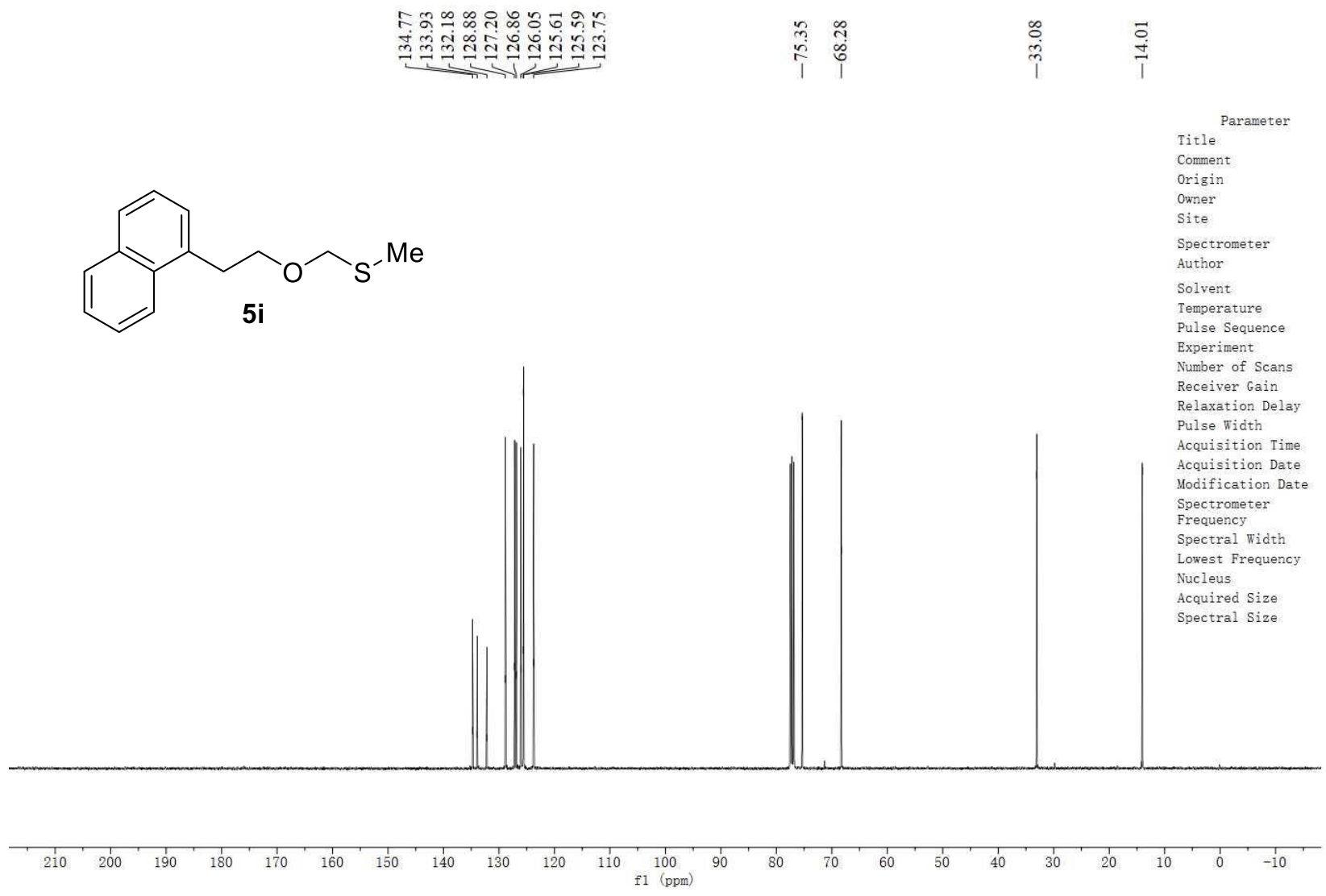
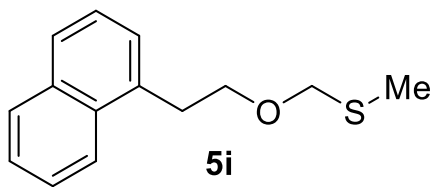




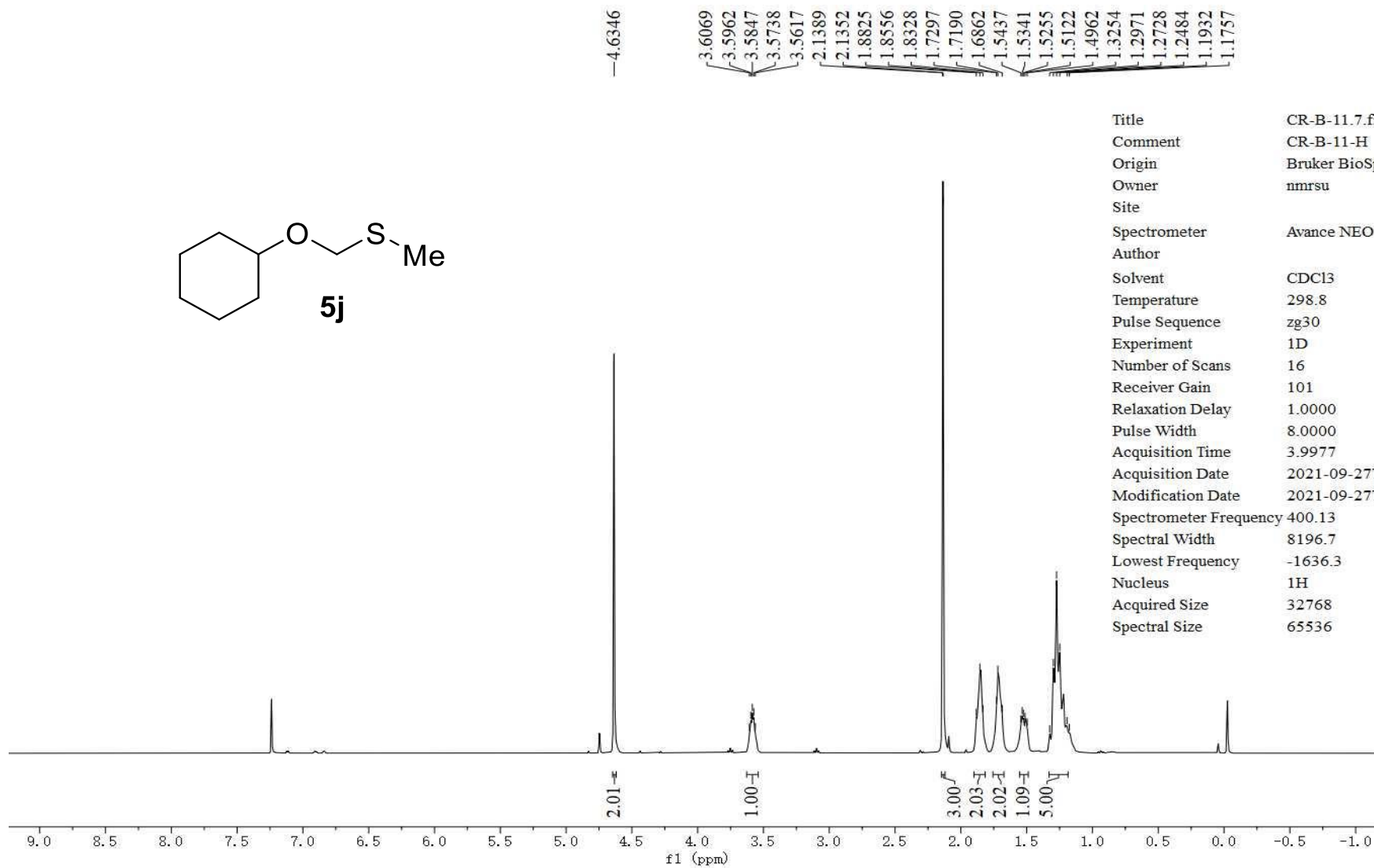
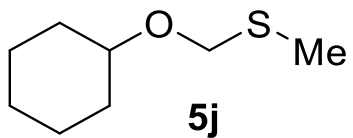
Parameter	Value
Title	CR-B-60.10.fid
Comment	CR-B-60-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-27T20:21:19
Modification Date	2021-09-27T20:21:38
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1829.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



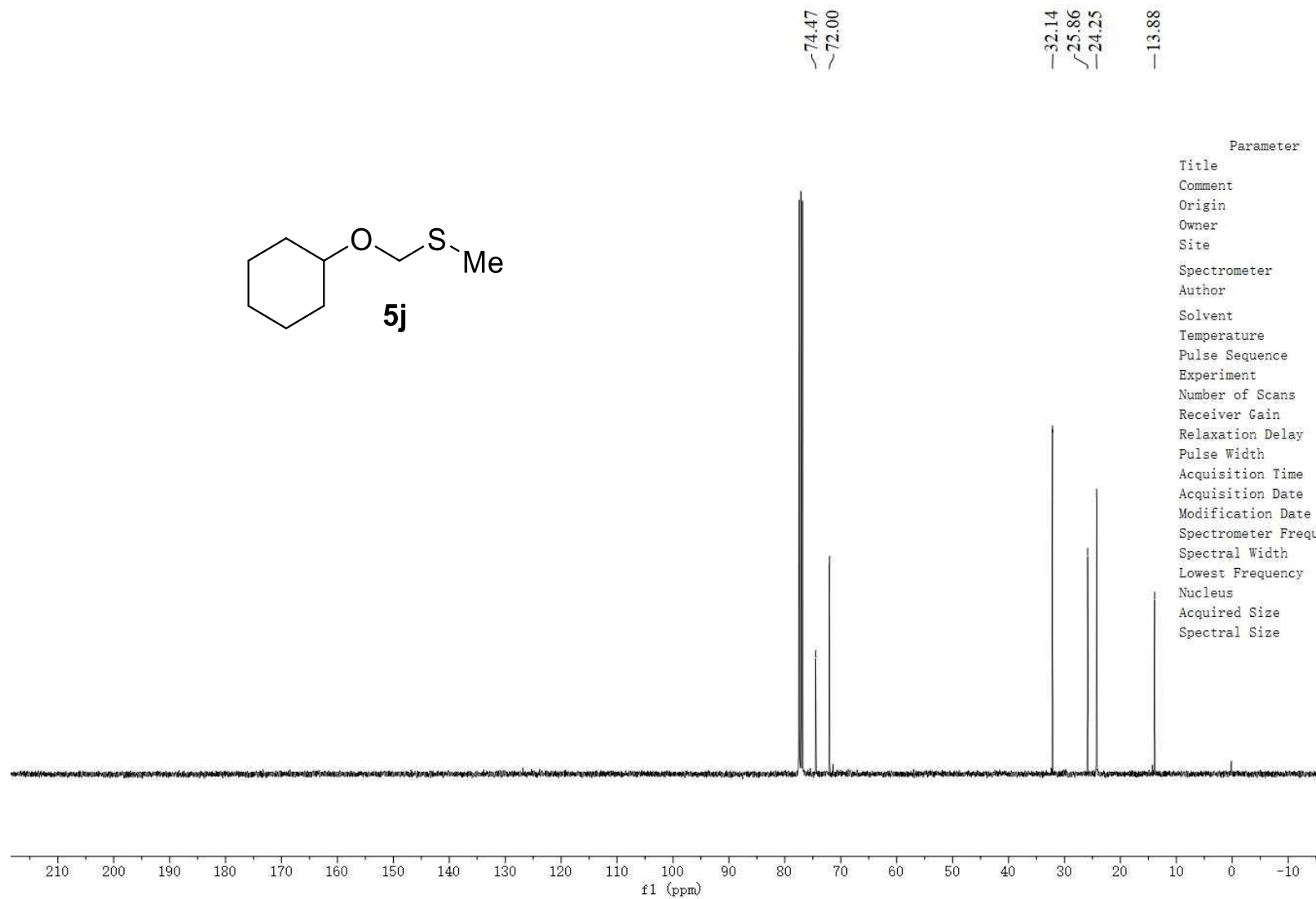
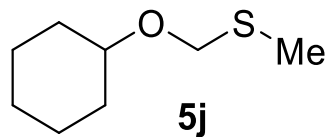
Parameter	Value
Title	CR-B-86.9.fid
Comment	CR-B-86-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl ₃
Temperature	296.6
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	44
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-10-18T08:39:47
Modification Date	2021-10-18T08:39:08
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.1
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



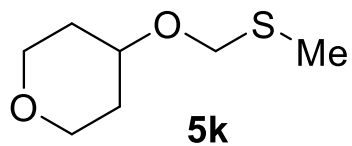
Parameter	Value
Title	CR-B-86.10.fid
Comment	CR-B-86-C
Origin	Bruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDC13
Temperature	297.3
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-10-18T09:39:14
Modification Date	2021-10-18T09:38:36
Spectrometer	100.62
Frequency	
Spectral Width	23809.5
Lowest Frequency	-1838.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Title	CR-B-11.7.fid
Comment	CR-B-11-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	298.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	101
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-27T18:17:23
Modification Date	2021-09-27T18:17:42
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1636.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

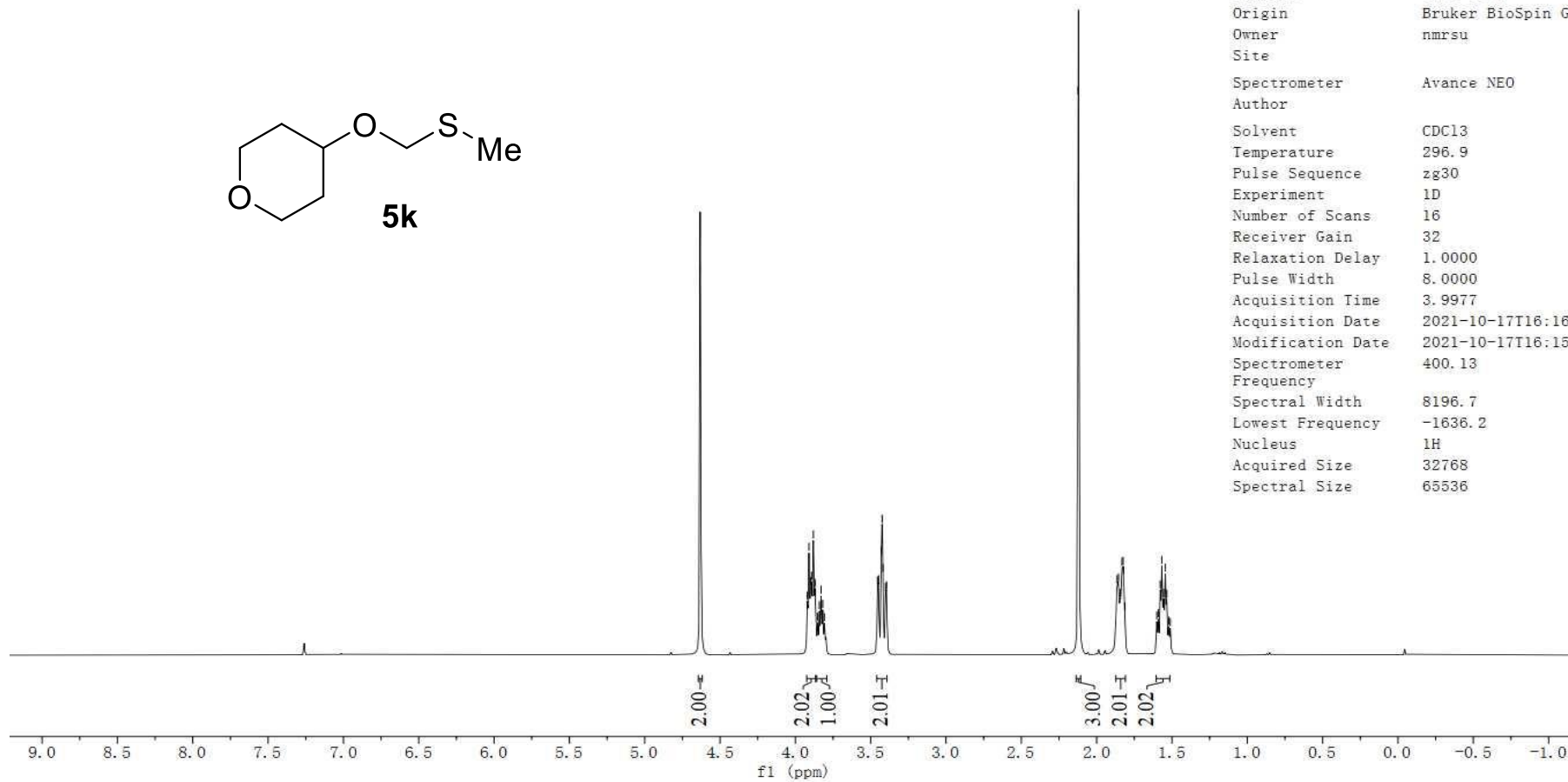


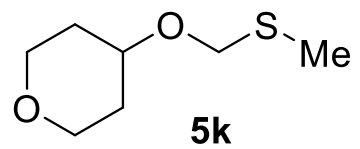
Parameter	Value
Title	CR-B-11.8.fid
Comment	CR-B-11-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-27T19:16:41
Modification Date	2021-09-27T19:17:00
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1829.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



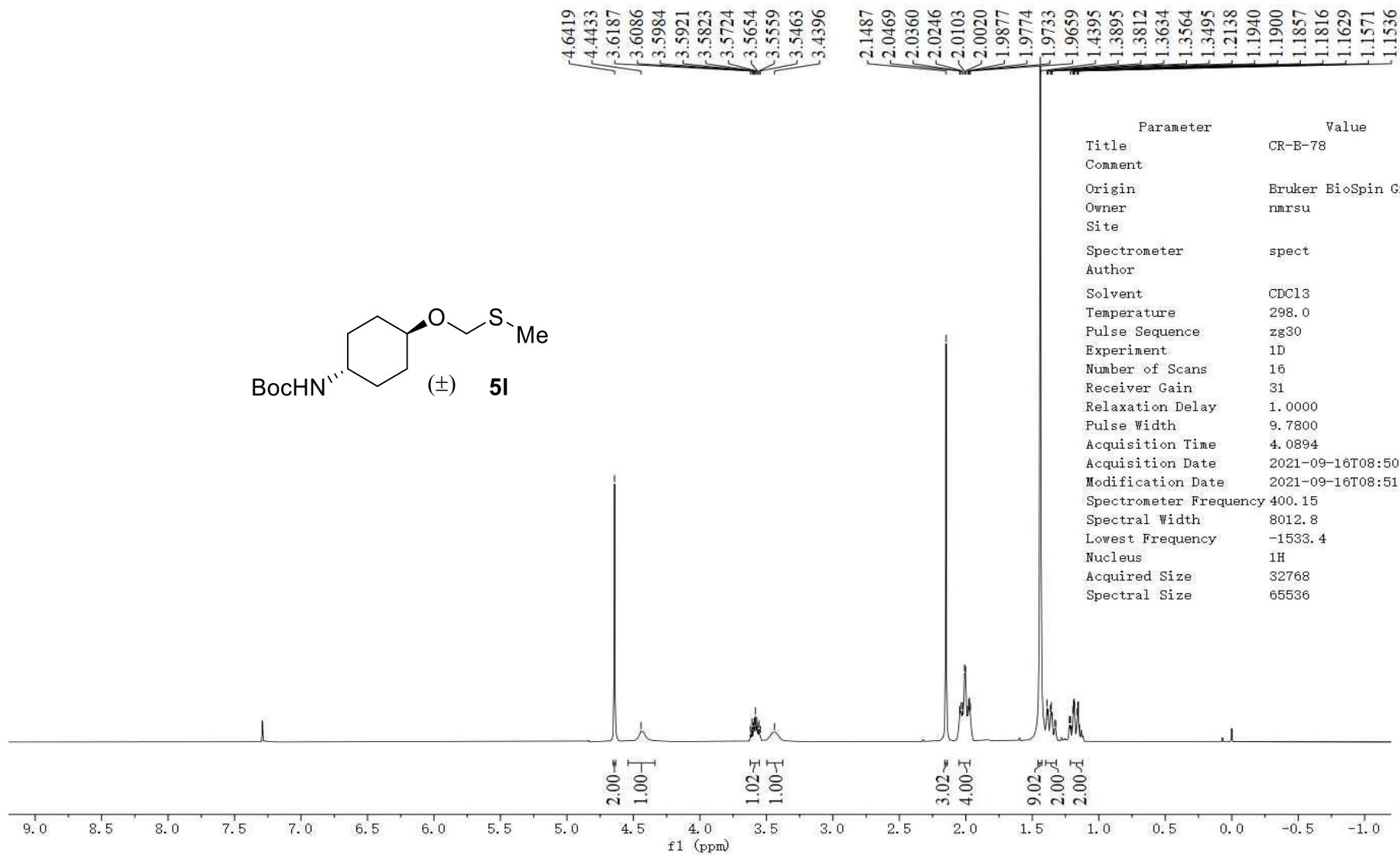
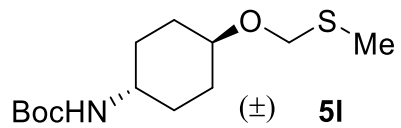
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3.9098
3.8985
3.8920
3.8804
3.8694
3.8523
3.8405
3.8294
3.8184
3.8068
3.4545
3.4476
3.4299
3.4240
3.4182
3.4014
3.3945
2.1233
1.8644
1.8549
1.8436
1.8317
1.8243
1.8206
1.8133
1.6008
1.5910
1.5779
1.5677
1.5561
1.5445
1.5344
1.5215
1.5112

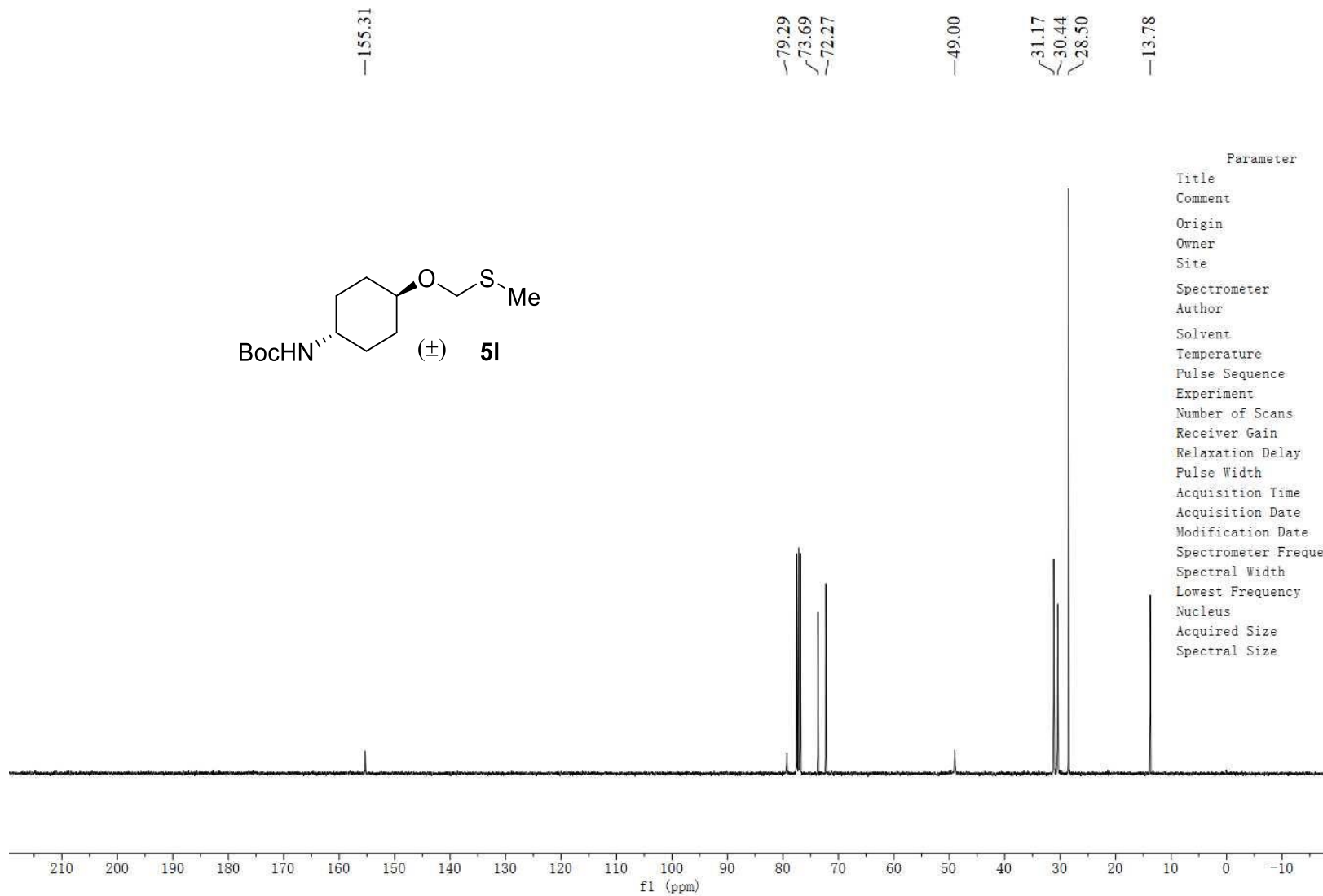
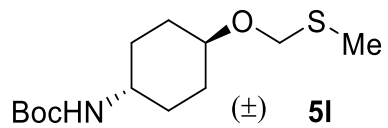
Parameter	Value
Title	CR-B-58.5.fid
Comment	CR-B-58-H
Origin	Bruker BioSpin GmbH
Owner	nmsru
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDC13
Temperature	296.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-10-17T16:18:24
Modification Date	2021-10-17T16:15:44
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



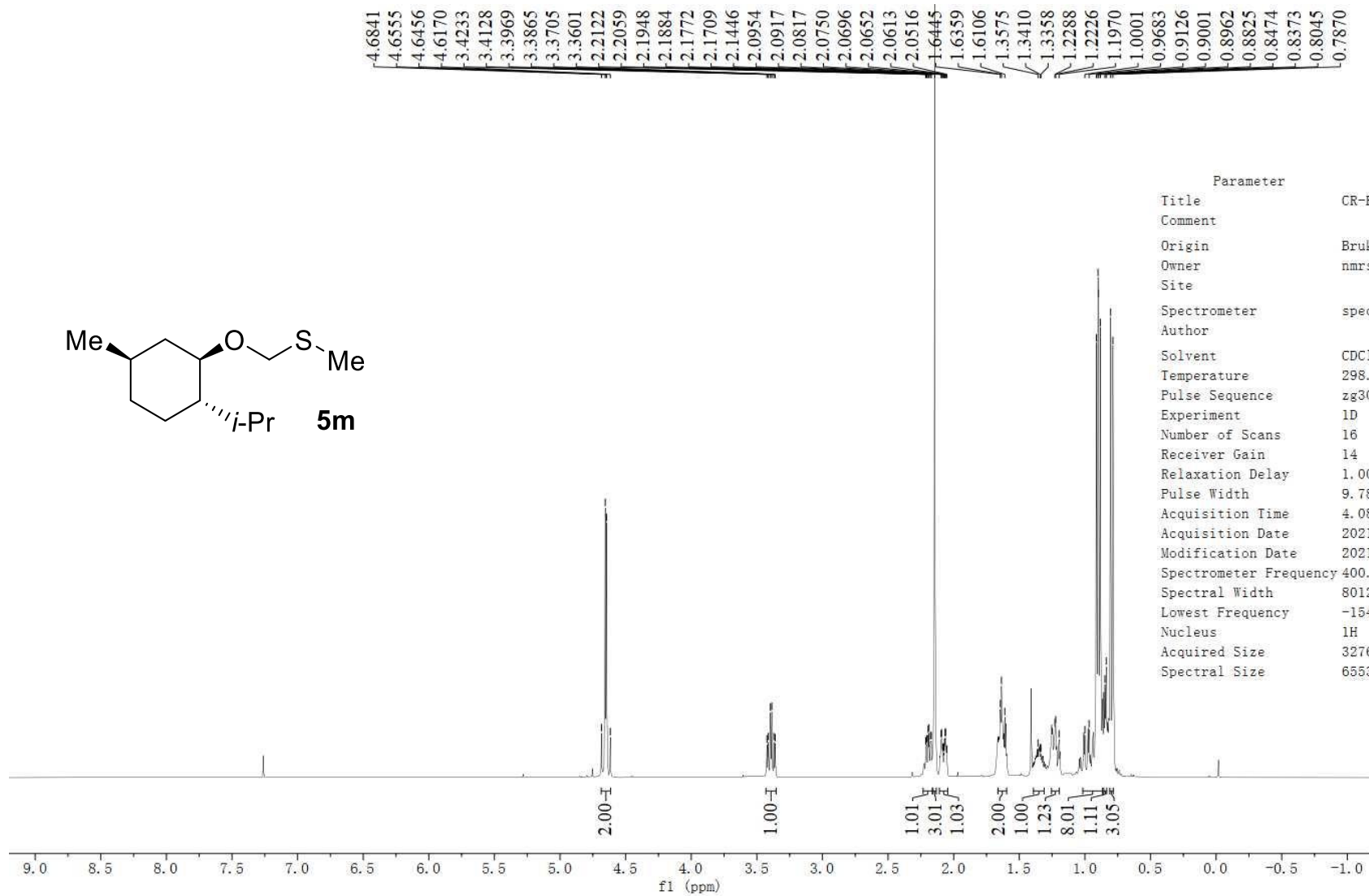
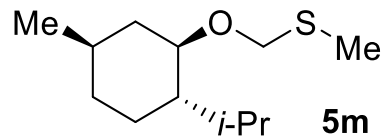


Parameter	Value
Title	CR-B-58.6.fid
Comment	CR-B-58-C
Origin	Eruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	297.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-10-17T17:16:24
Modification Date	2021-10-17T17:15:44
Spectrometer	100.62
Frequency	
Spectral Width	23809.5
Lowest Frequency	-1838.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

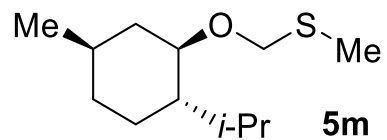




Parameter	Value
Title	CR-B-78
Comment	
Origin	Bruker BioSpin GmbH
Owner	nrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	300
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-16T09:09:12
Modification Date	2021-09-16T09:09:14
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1948.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

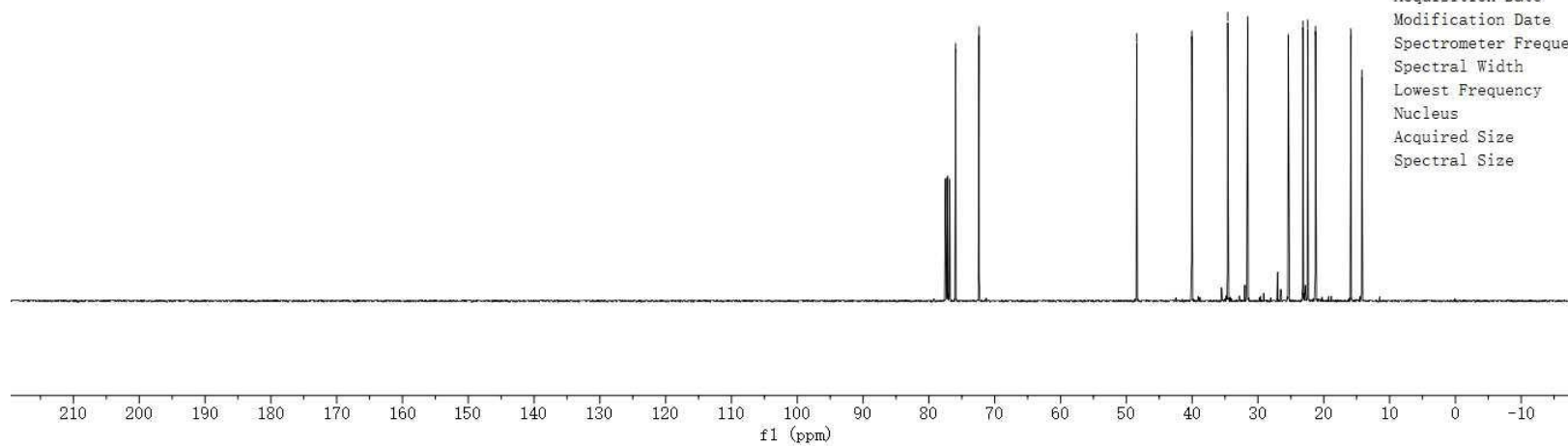


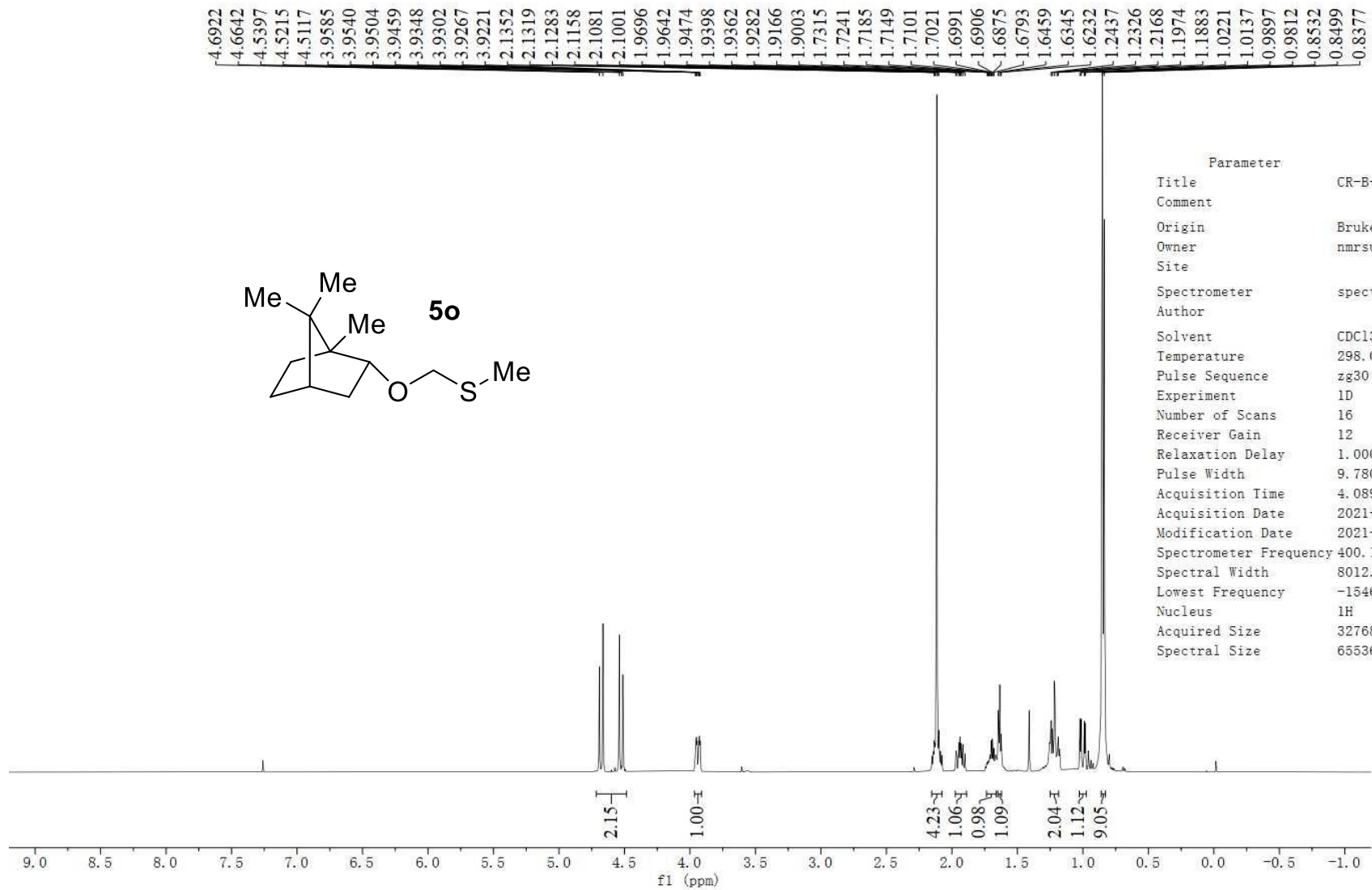
Parameter	Value
Title	CR-B-82.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	14
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-26T00:34:30
Modification Date	2021-09-26T00:34:32
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.7
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



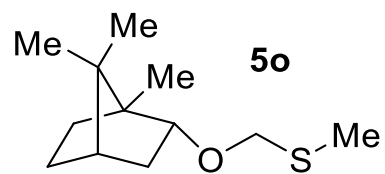
—75.94
 —72.39
 —48.43
 /40.04
 /34.59
 /31.56
 /25.39
 /23.17
 /22.44
 /21.25
 /15.88
 /14.20

Parameter	Value
Title	CR-B-82.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-26T01:09:52
Modification Date	2021-09-26T01:09:54
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1946.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



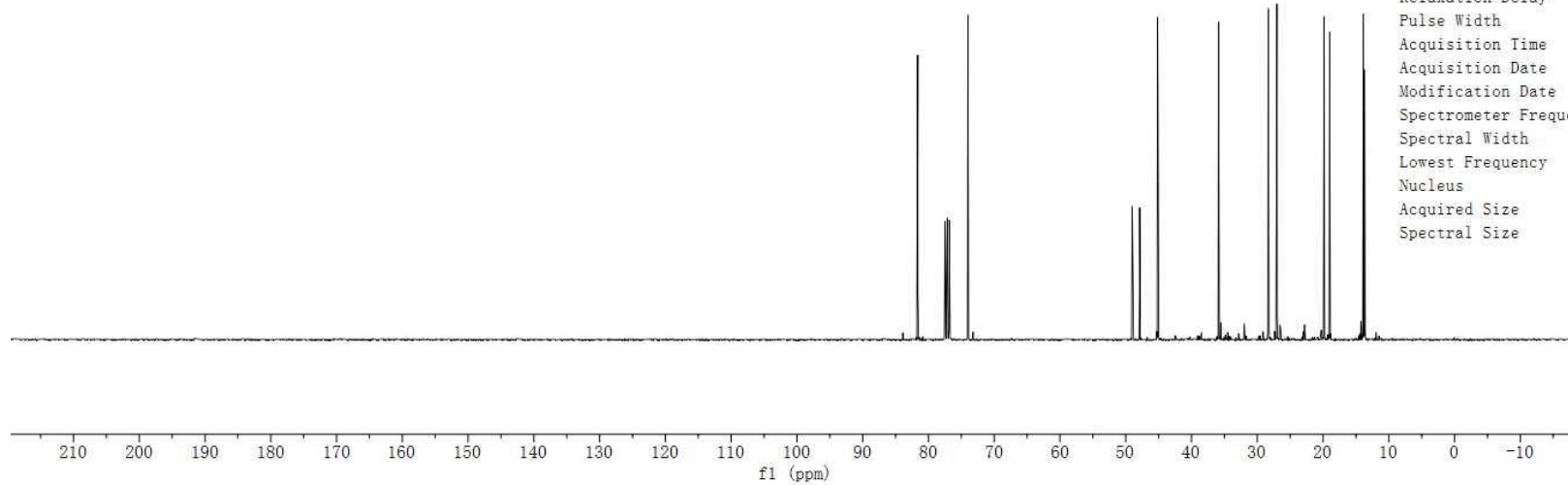


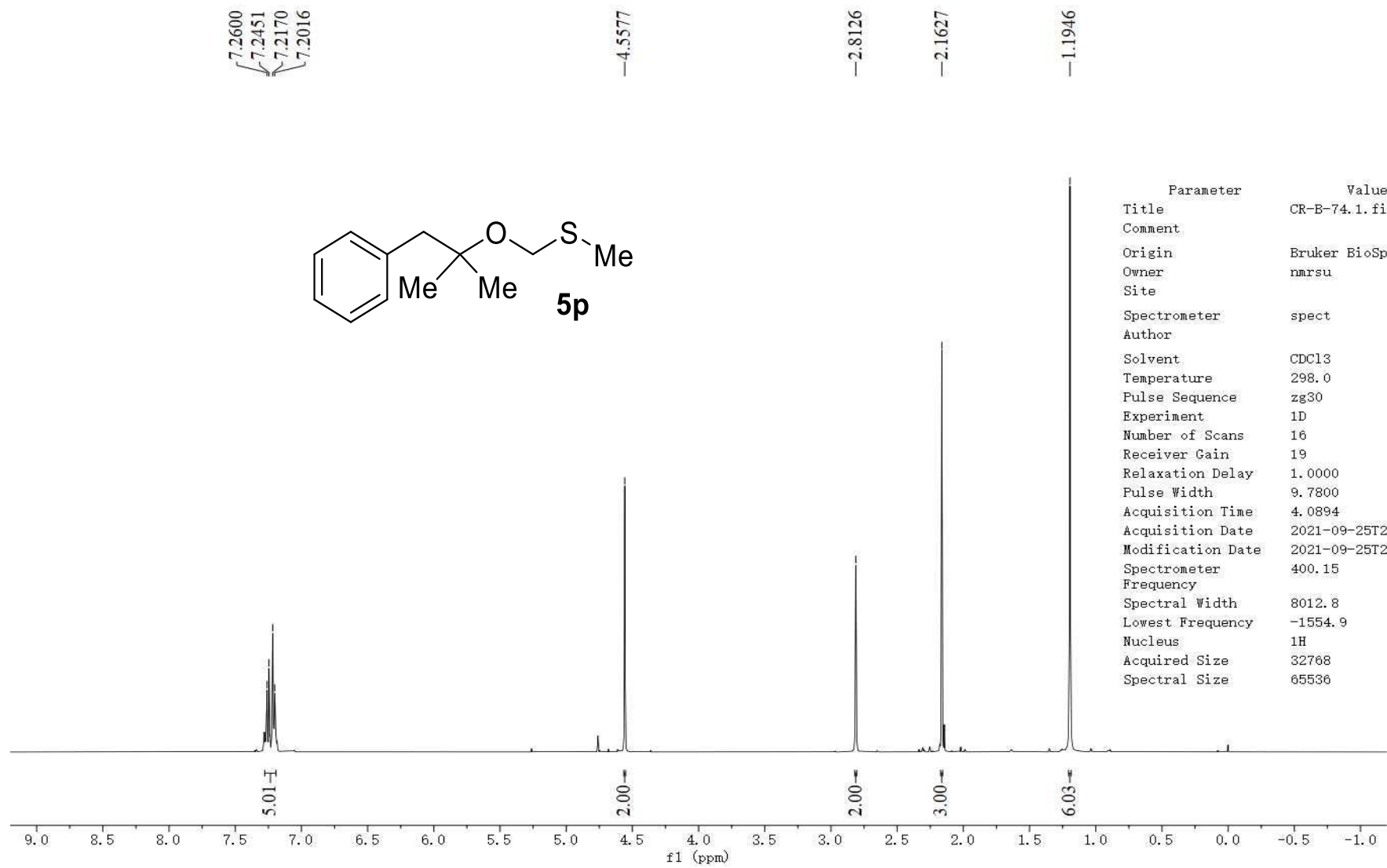
Parameter	Value
Title	CR-B-84
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	12
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-26T03:17:55
Modification Date	2021-09-26T03:17:56
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1546.0
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



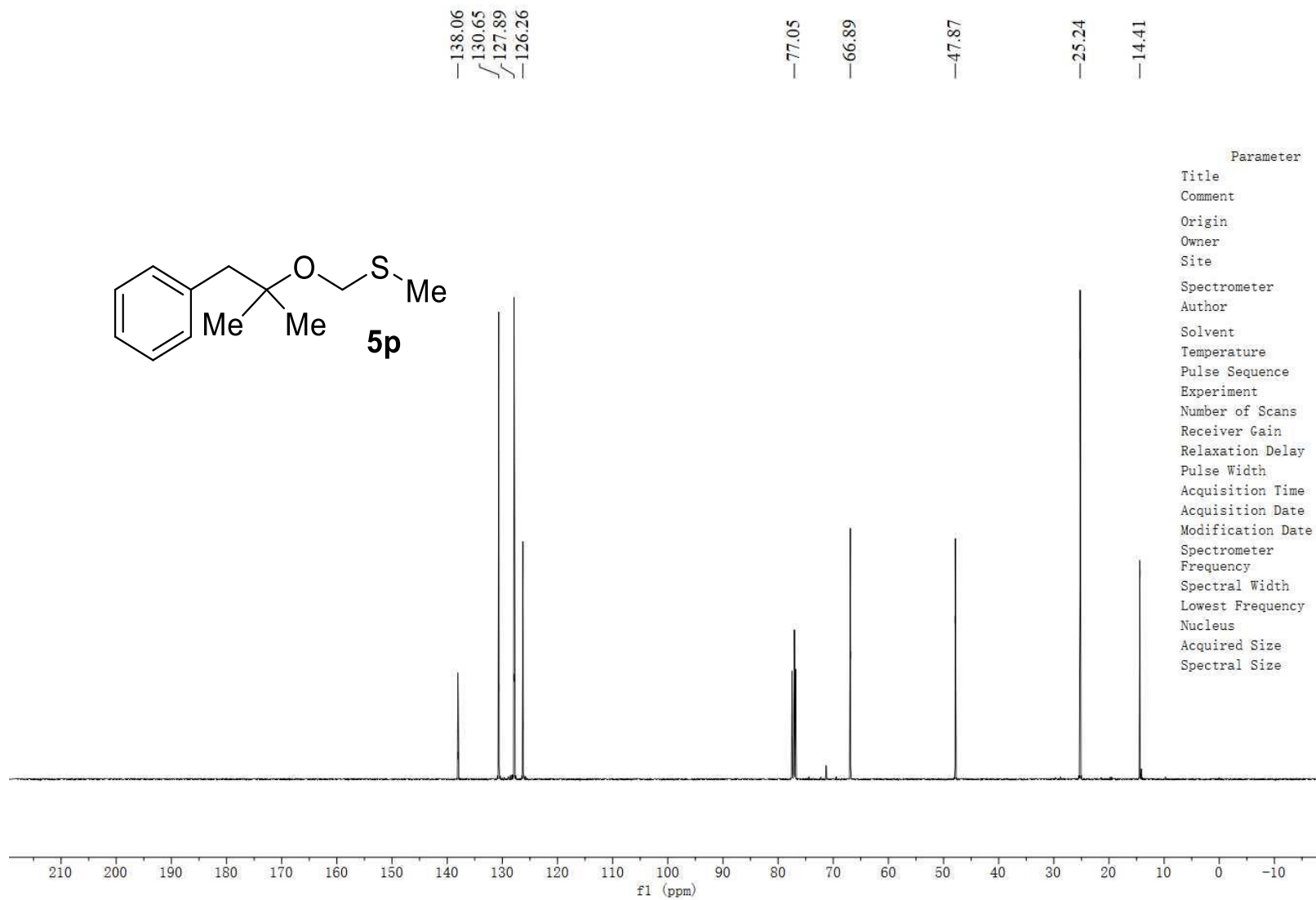
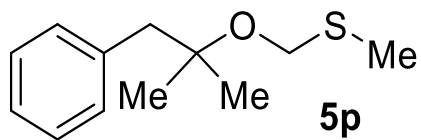
—81.64
 —73.99
 ~49.02
 ~47.88
 ~45.15
 ~35.89
 ~28.33
 ~27.00
 ~19.84
 ~19.01
 ~13.87
 ~13.69

Parameter	Value
Title	CR-B-84
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmsru
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-26T03:53:16
Modification Date	2021-09-26T03:53:18
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1947.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

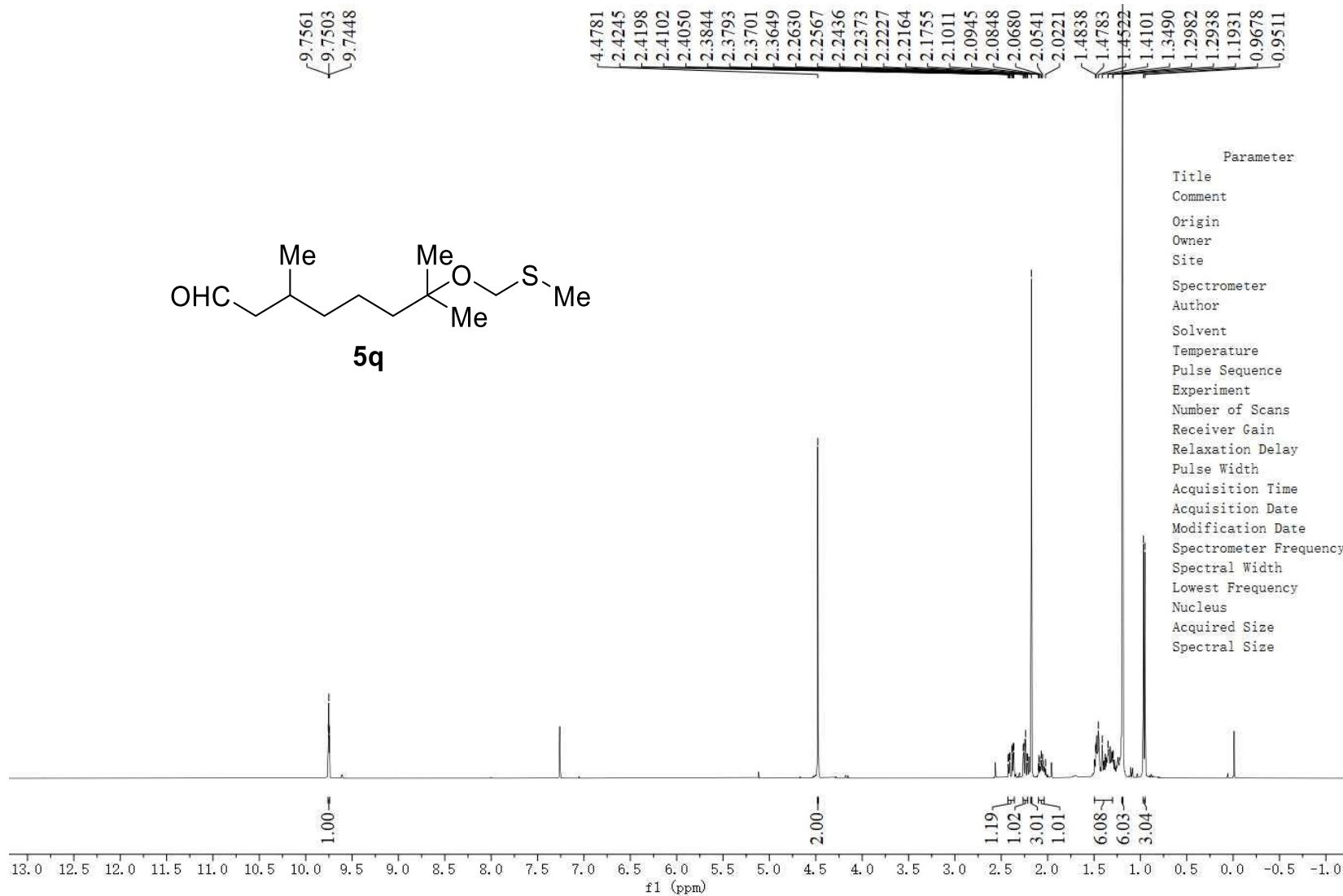
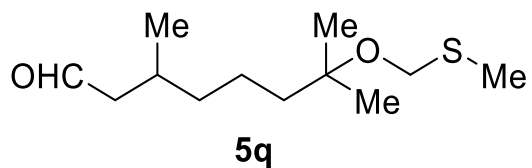




Parameter	Value
Title	CR-B-74.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	mrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	19
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-25T23:53:28
Modification Date	2021-09-25T23:53:30
Spectrometer	400.15
Frequency	
Spectral Width	8012.8
Lowest Frequency	-1554.9
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

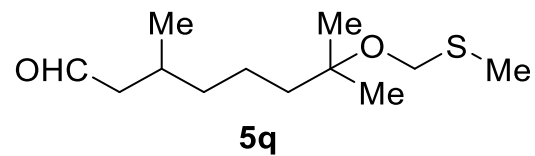


Parameter	Value
Title	CR-B-74. 2. fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-26T00:28:49
Modification Date	2021-09-26T00:28:50
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1955.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



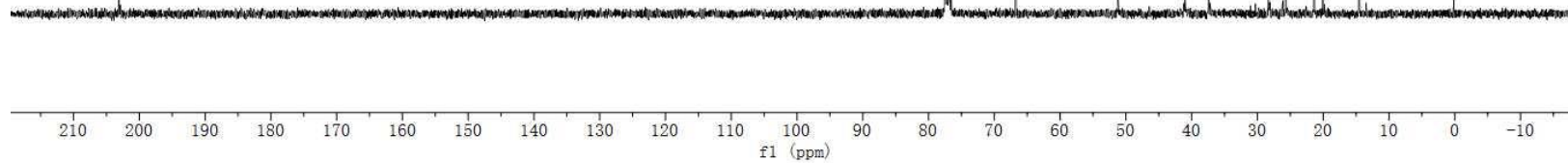
Parameter	Value
Title	CR-B-88
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	64
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-25T01:14:48
Modification Date	2021-09-25T01:14:50
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.7
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

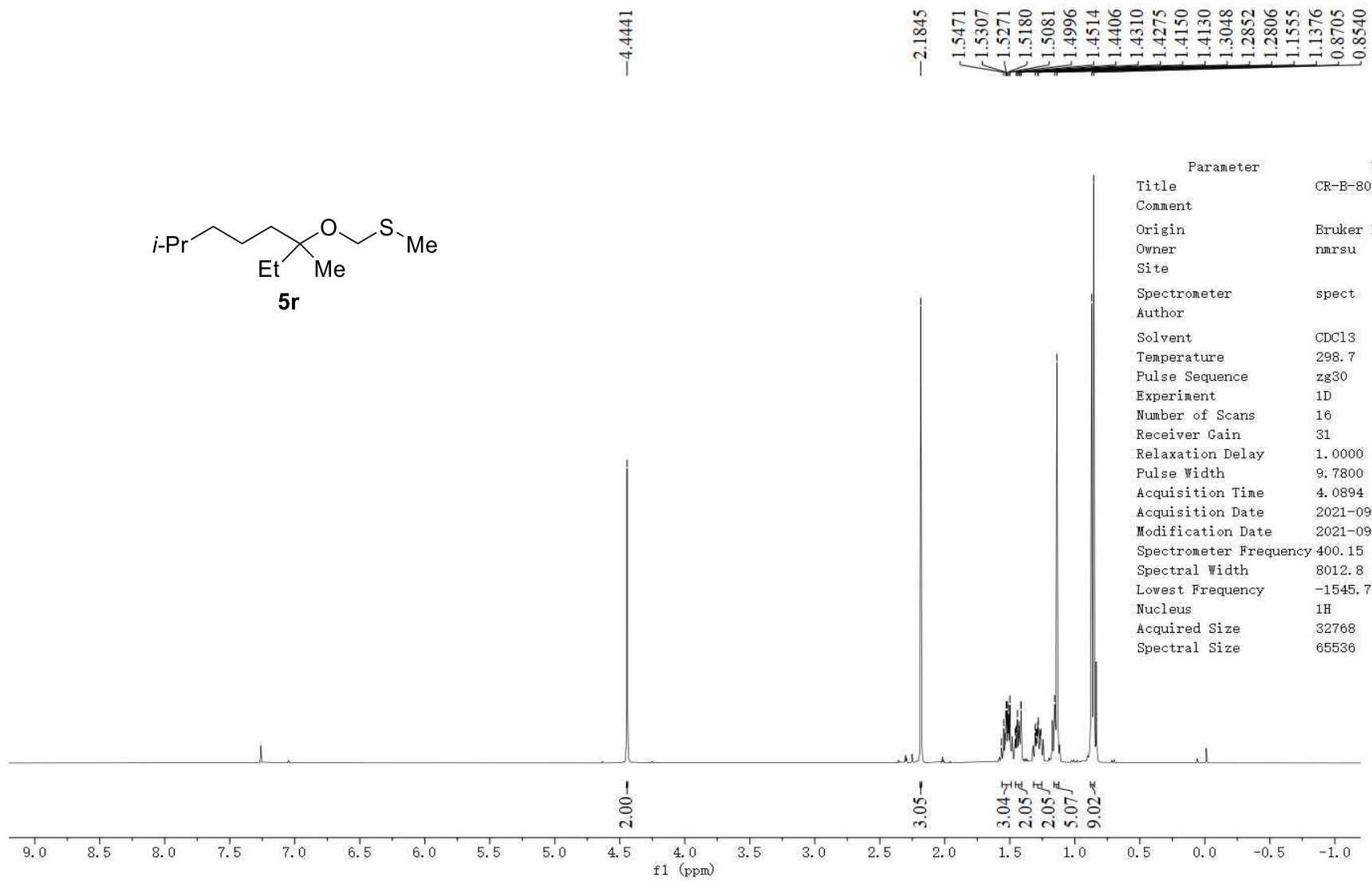
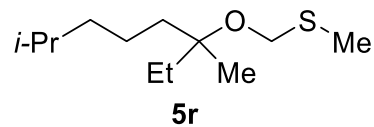
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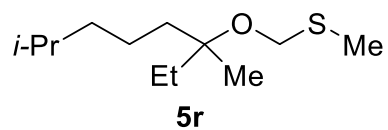


-76.59
-66.76
-51.19
-41.02
-37.42
28.25
25.73
25.71
21.35
20.07
14.56

Parameter	Value
Title	CR-B-88
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-25T01:50:09
Modification Date	2021-09-25T01:50:10
Spectrometer	100.62
Frequency	
Spectral Width	24038.5
Lowest Frequency	-1945.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

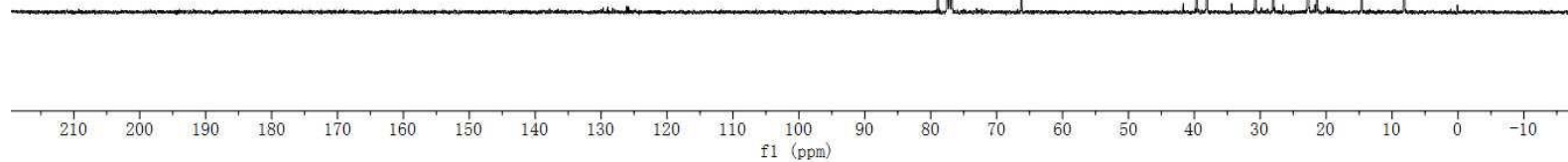


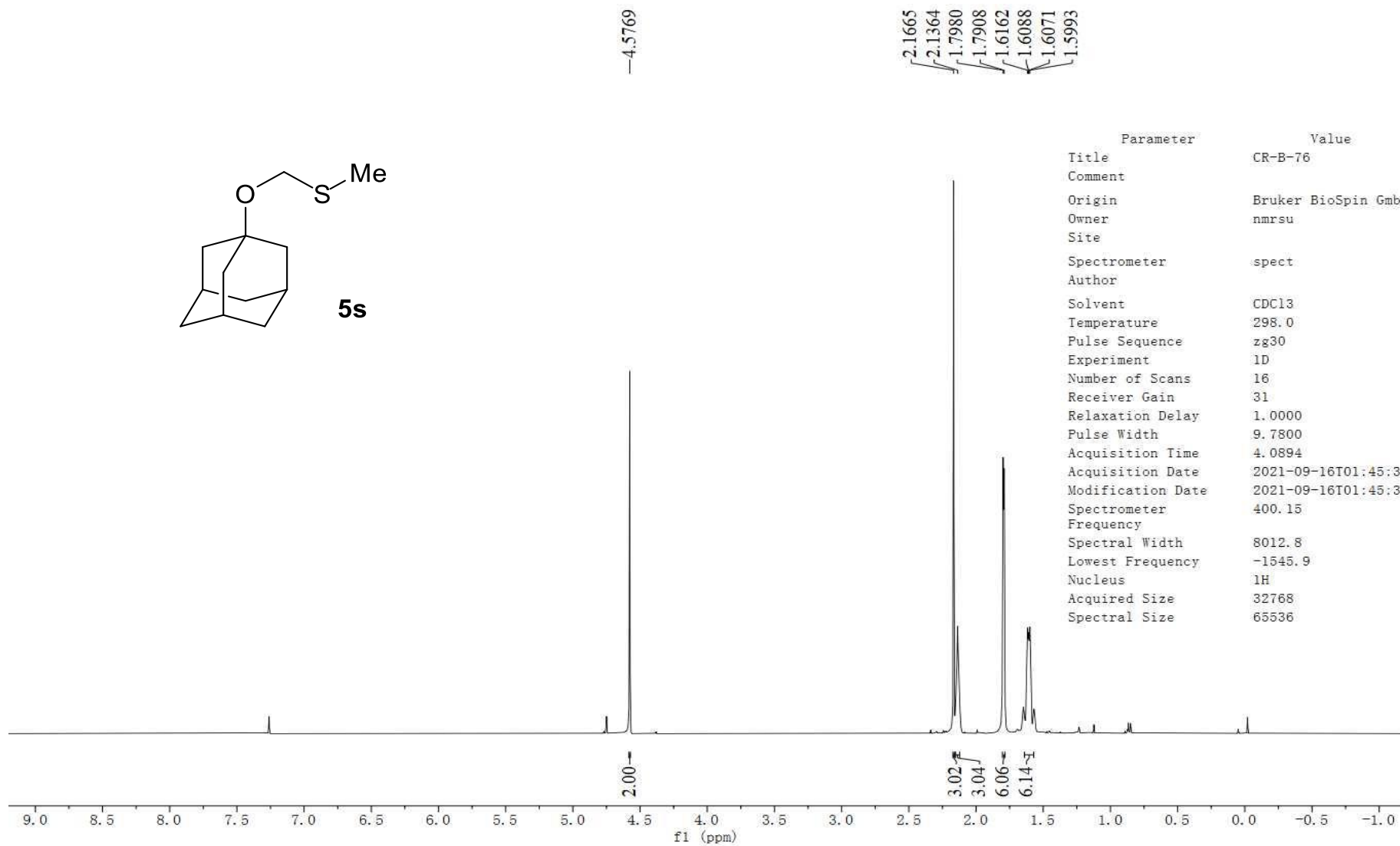
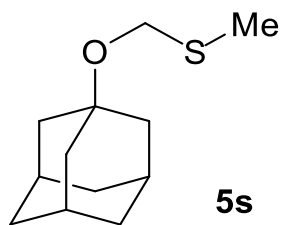


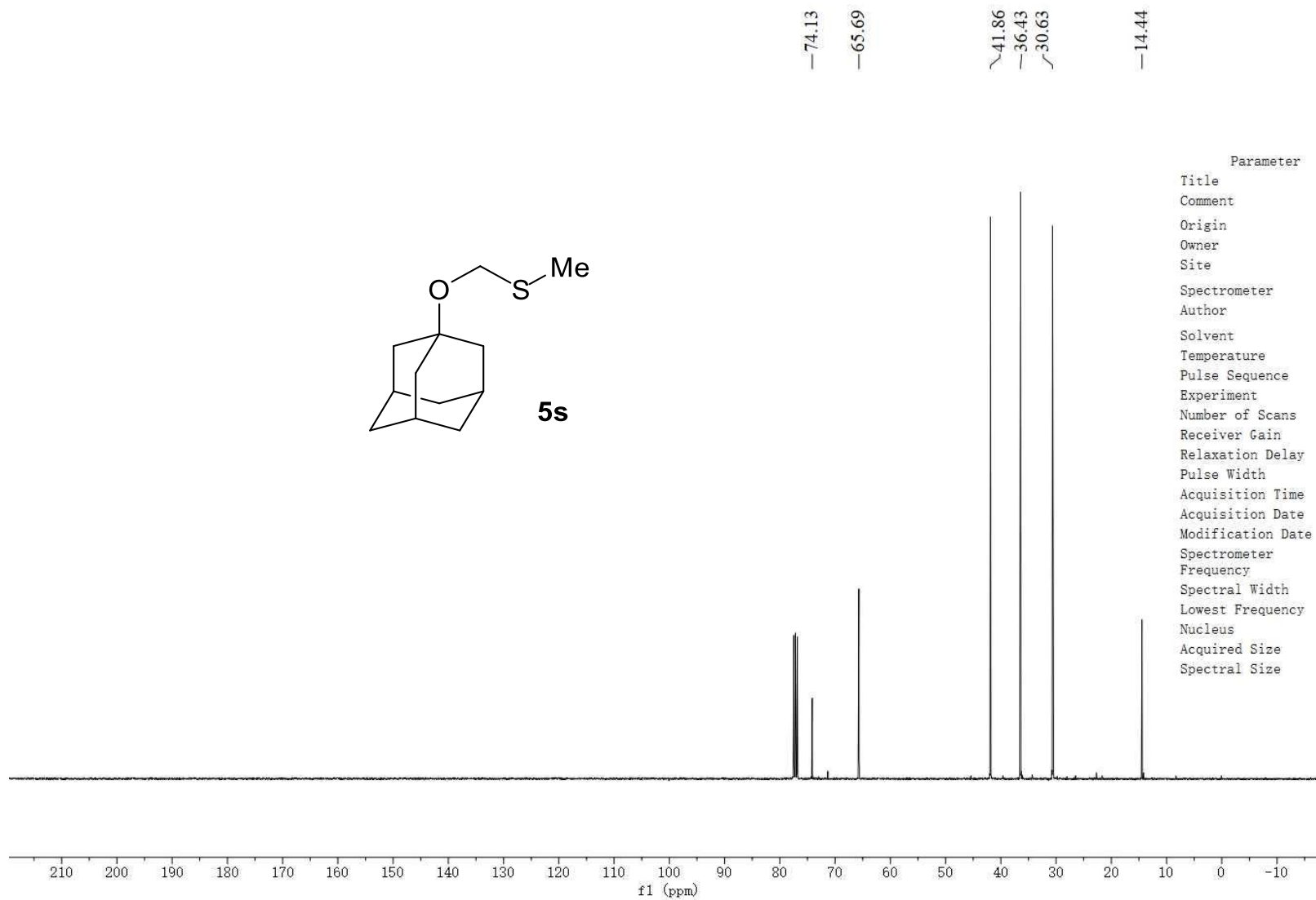
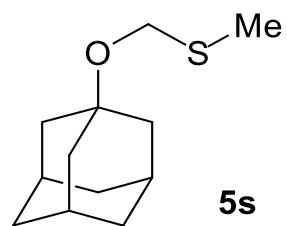


—78.90
—66.22
39.65
38.14
30.78
28.04
22.82
22.75
22.73
21.38
14.64
8.13

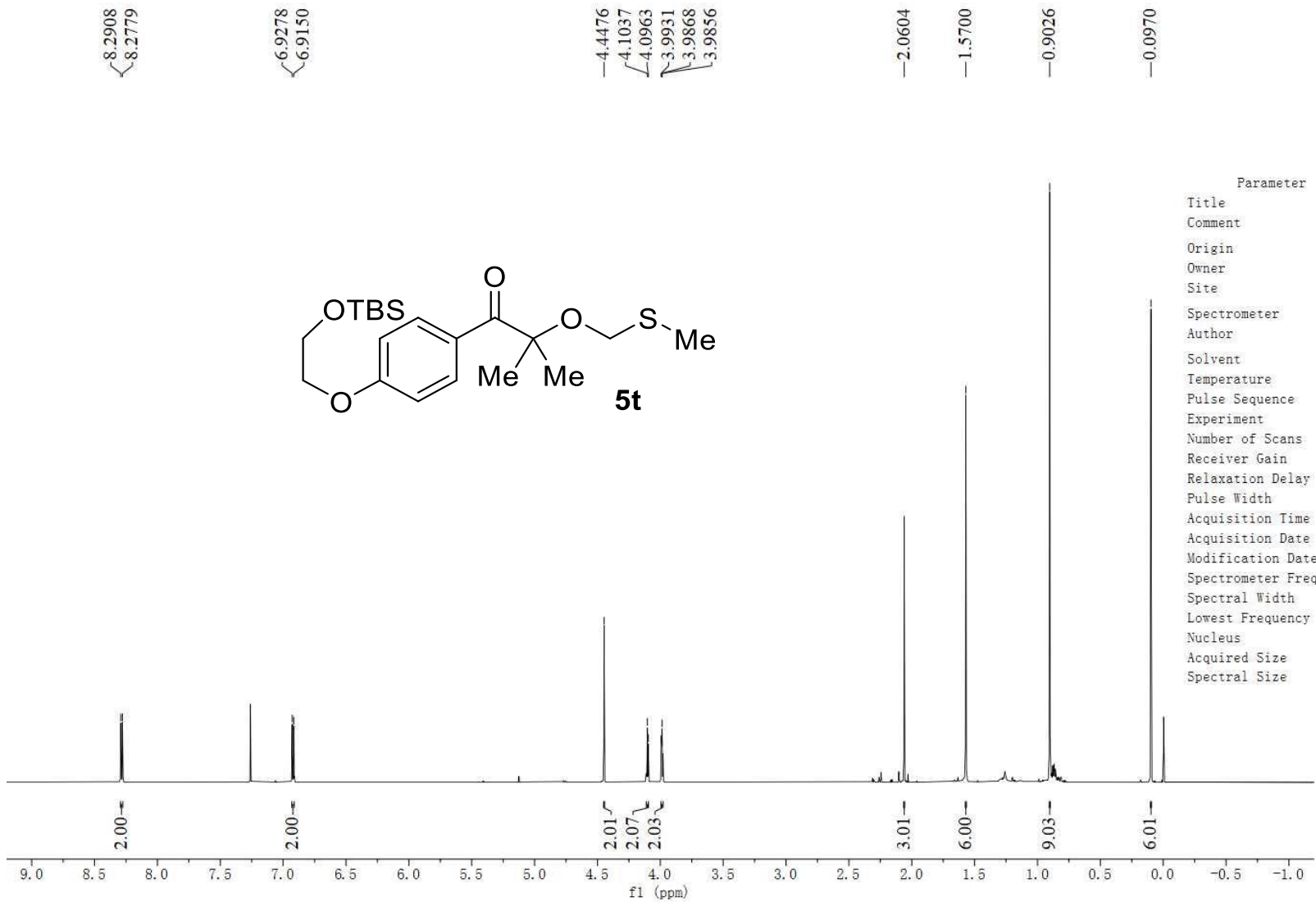
Parameter	Value
Title	CR-B-80. 2. fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl ₃
Temperature	299.3
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-15T07:22:22
Modification Date	2021-09-15T07:22:24
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1946.0
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



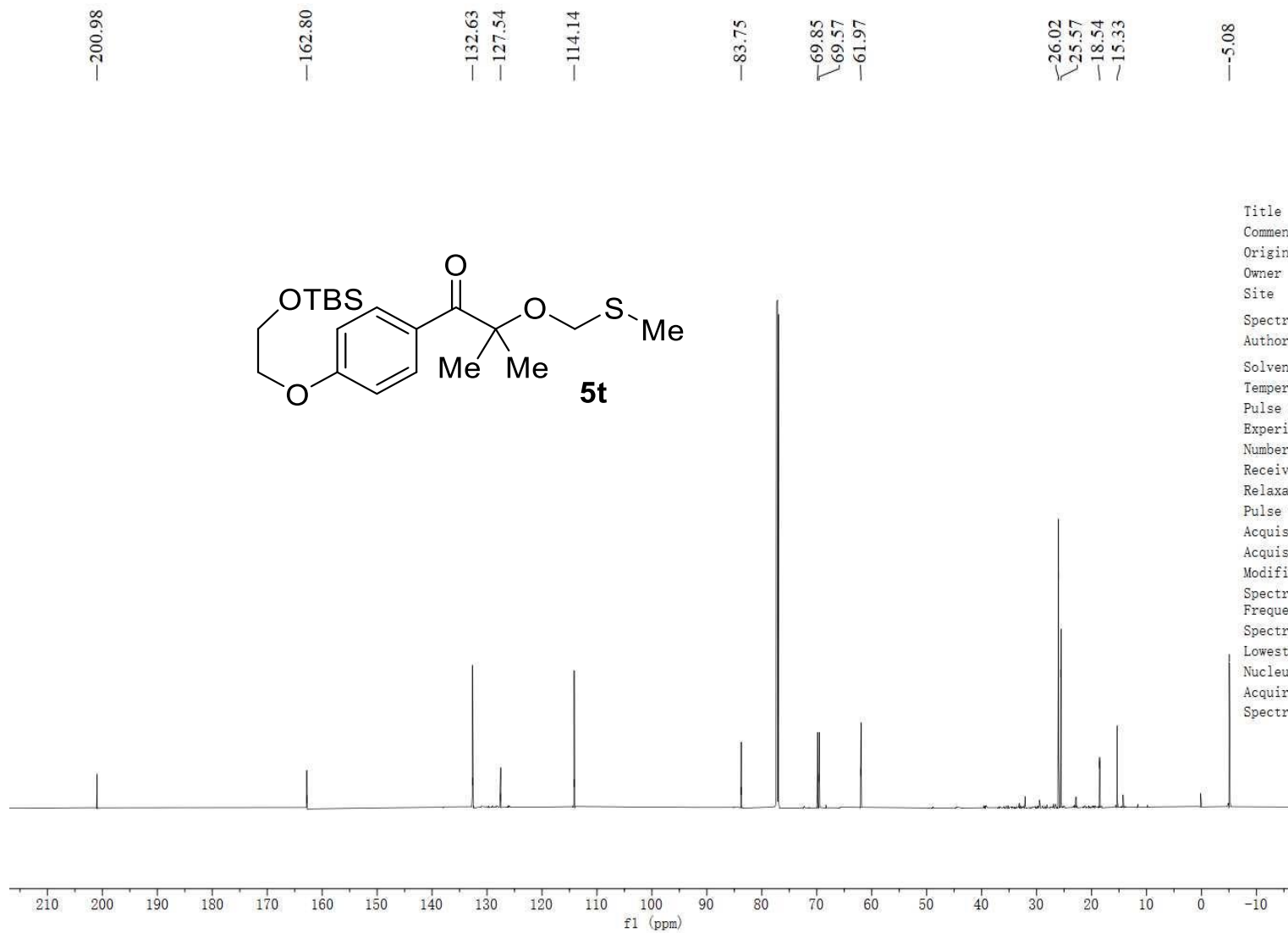




Parameter	Value
Title	CR-B-76
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.3
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-16T02:15:09
Modification Date	2021-09-16T02:15:10
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1949.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

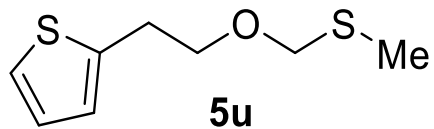


Parameter	Value
Title	CR-B-101.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	1
Relaxation Delay	1.0000
Pulse Width	12.0000
Acquisition Time	2.3243
Acquisition Date	2021-10-16T07:39:51
Modification Date	2021-10-16T07:39:54
Spectrometer Frequency	700.15
Spectral Width	14097.7
Lowest Frequency	-2741.2
Nucleus	¹ H
Acquired Size	32768
Spectral Size	131072



Parameter	Value
Title	CR-B-101.2.fid
Comment	13C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	188
Relaxation Delay	2.0000
Pulse Width	12.0000
Acquisition Time	0.7690
Acquisition Date	2021-10-16T08:03:47
Modification Date	2021-10-16T08:03:50
Spectrometer	176.07
Frequency	
Spectral Width	42613.6
Lowest Frequency	-3678.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

7.1551
7.1522
7.1423
7.1394
6.9470
6.9384
6.9342
6.9256
6.8759
6.8734
6.8709
6.8674
6.8649

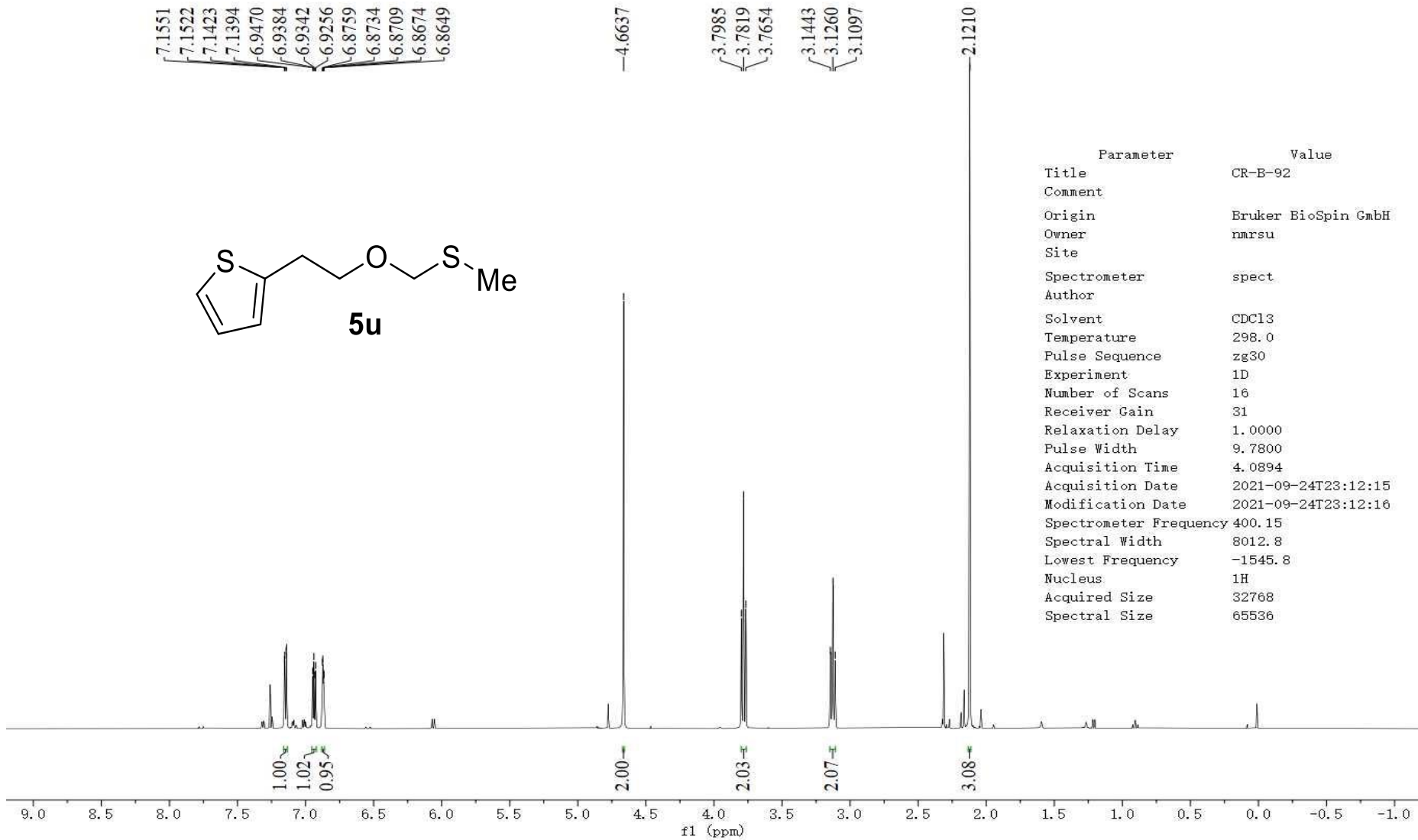


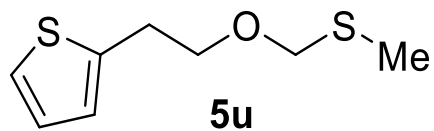
4.6637

3.7985
3.7819
3.7654
3.1443
3.1260
3.1097

2.1210

Parameter	Value
Title	CR-B-92
Comment	
Origin	Bruker BioSpin GmbH
Owner	narsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-24T23:12:15
Modification Date	2021-09-24T23:12:16
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536





—141.31

—126.80

—125.28

—123.79

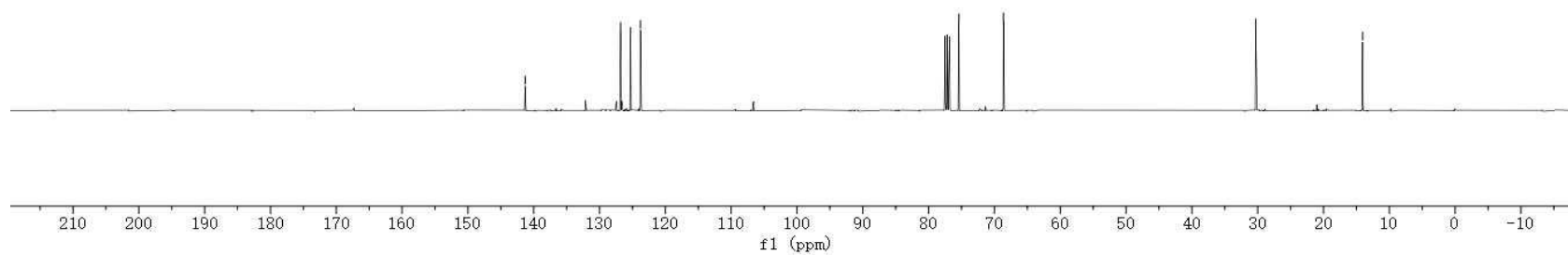
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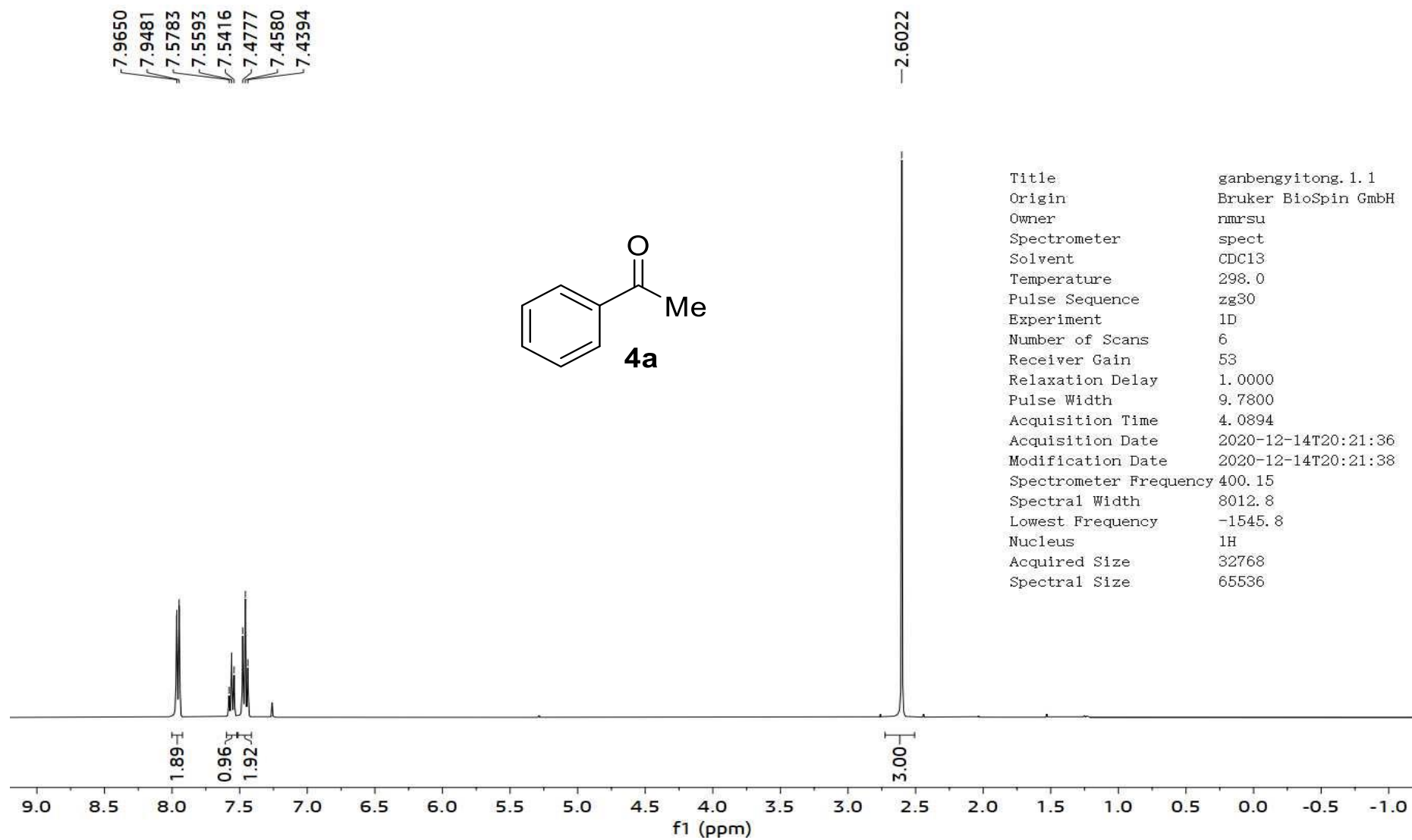
—68.63

—30.30

—14.06

Parameter	Value
Title	CR-B-92
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl ₃
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-24T23:47:36
Modification	2021-09-24T23:47:38
Date	
Spectrometer	100.62
Frequency	
Spectral Width	24038.5
Lowest Frequency	-1949.4
Nucleus	13C
Acquired Size	32768

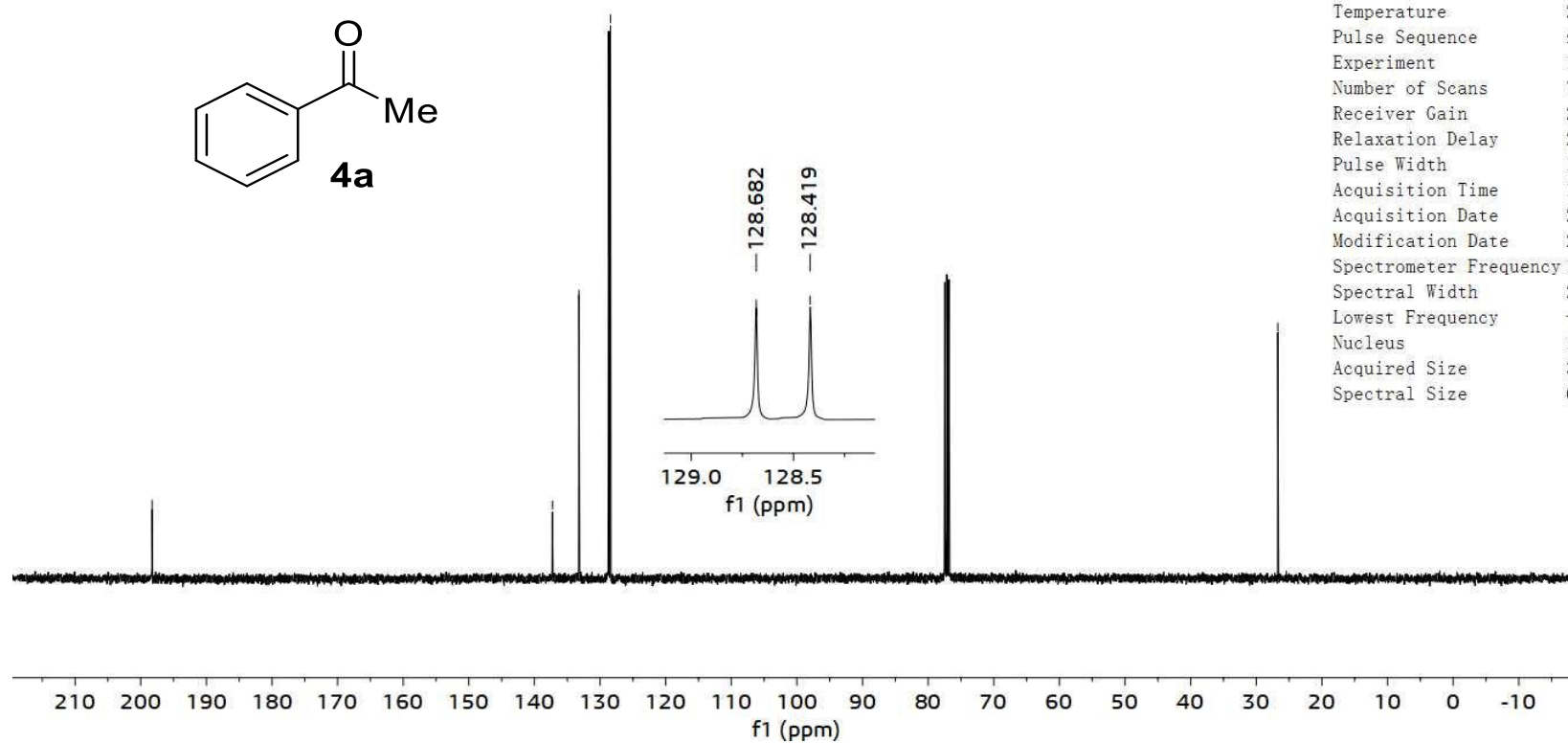
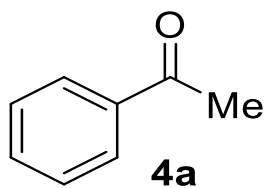




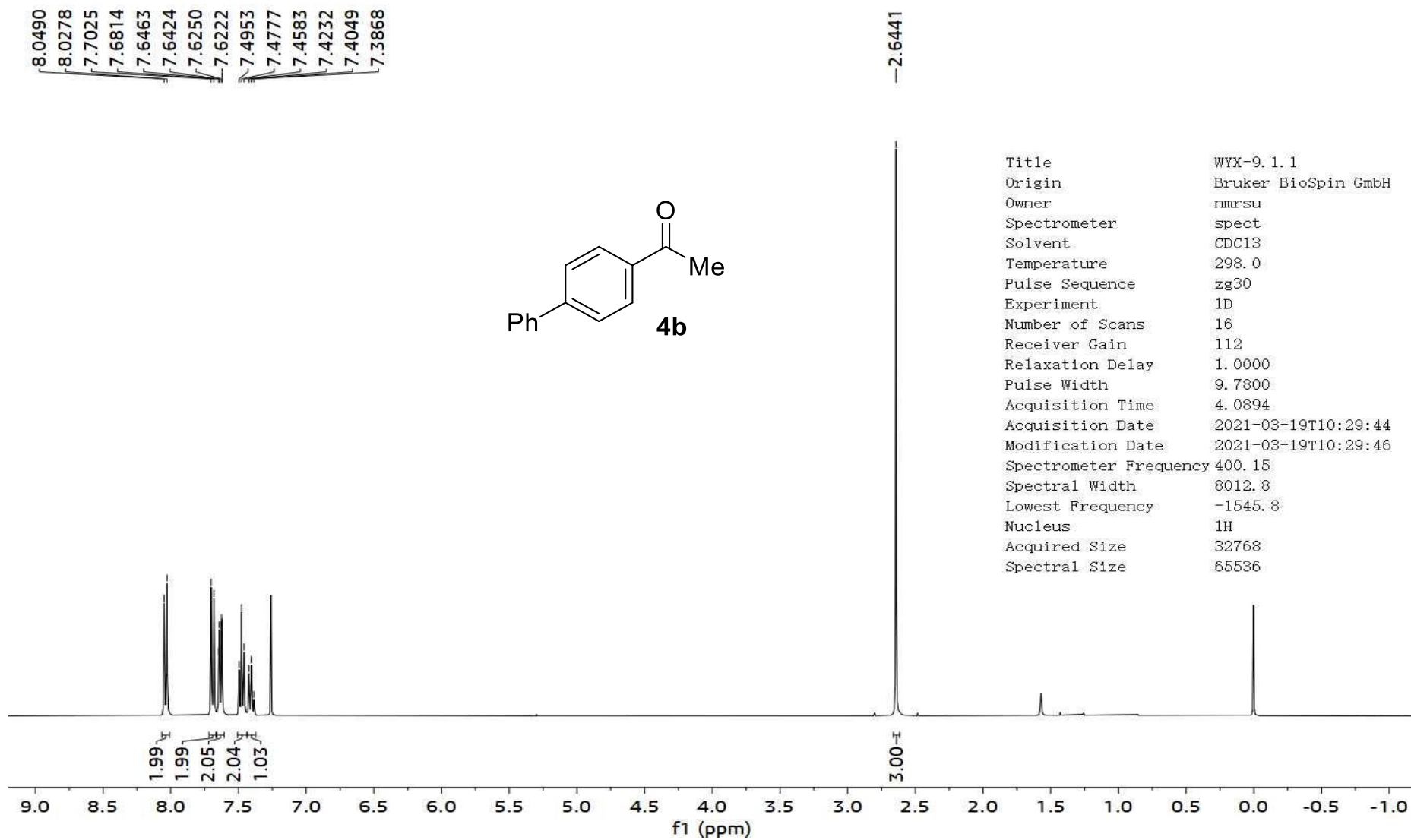
—198.26

✓ 137.25
✓ 133.21
✓ 128.68
✓ 128.42

—26.72



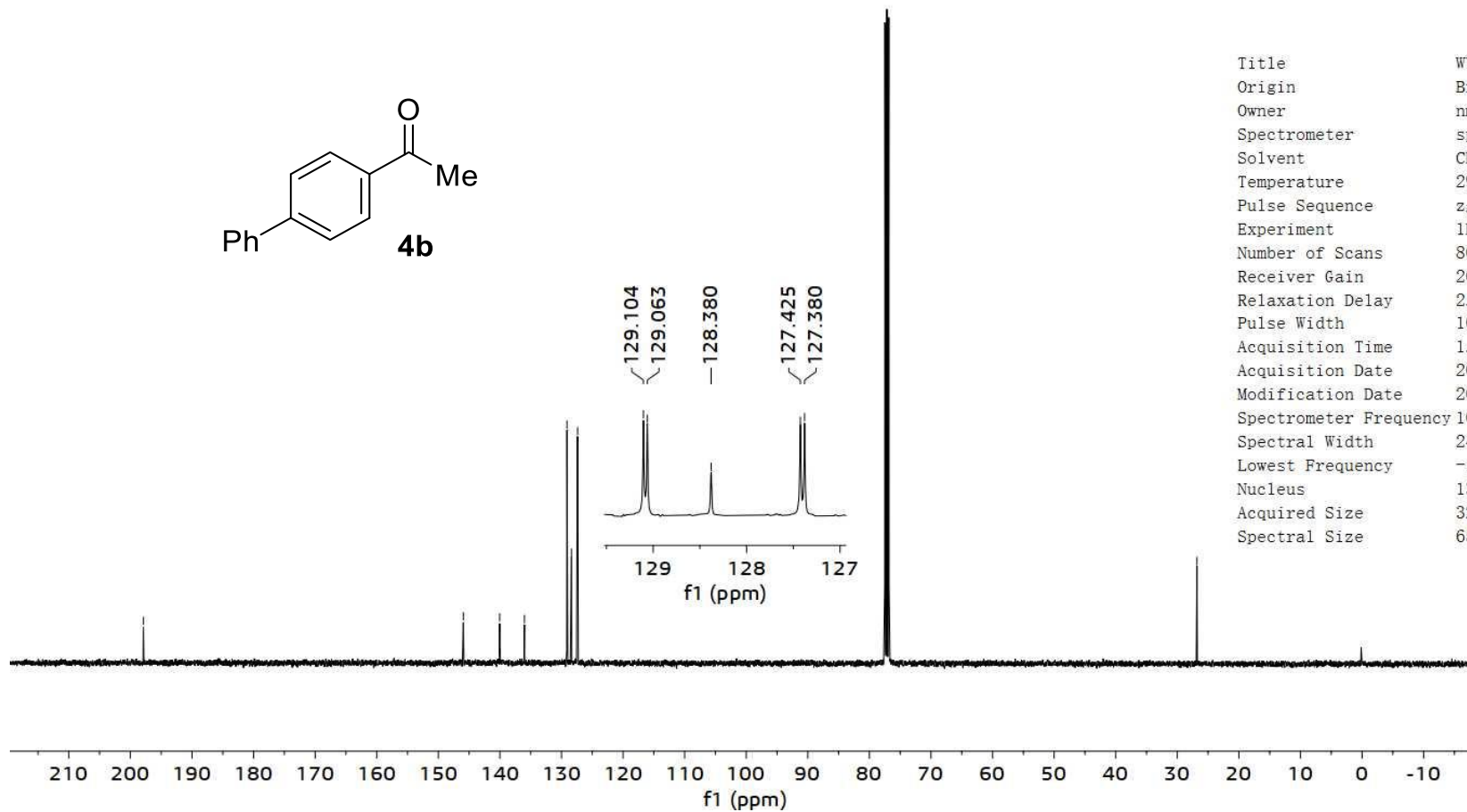
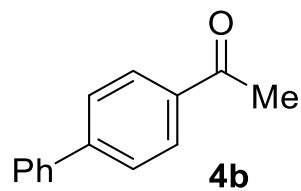
Title	ganbengyitong.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	70
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2020-12-14T20:26:42
Modification Date	2020-12-14T20:26:44
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1947.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



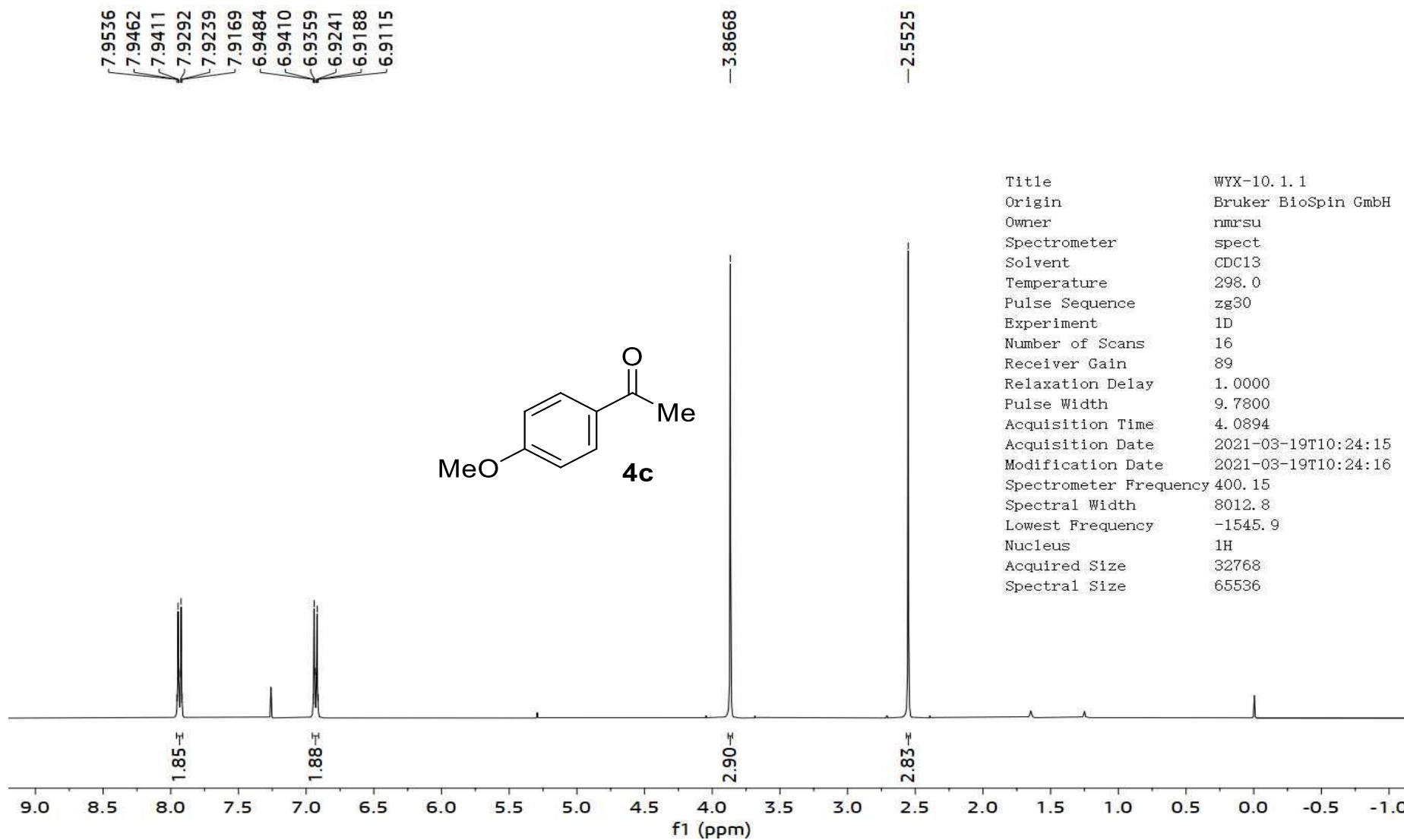
—197.90

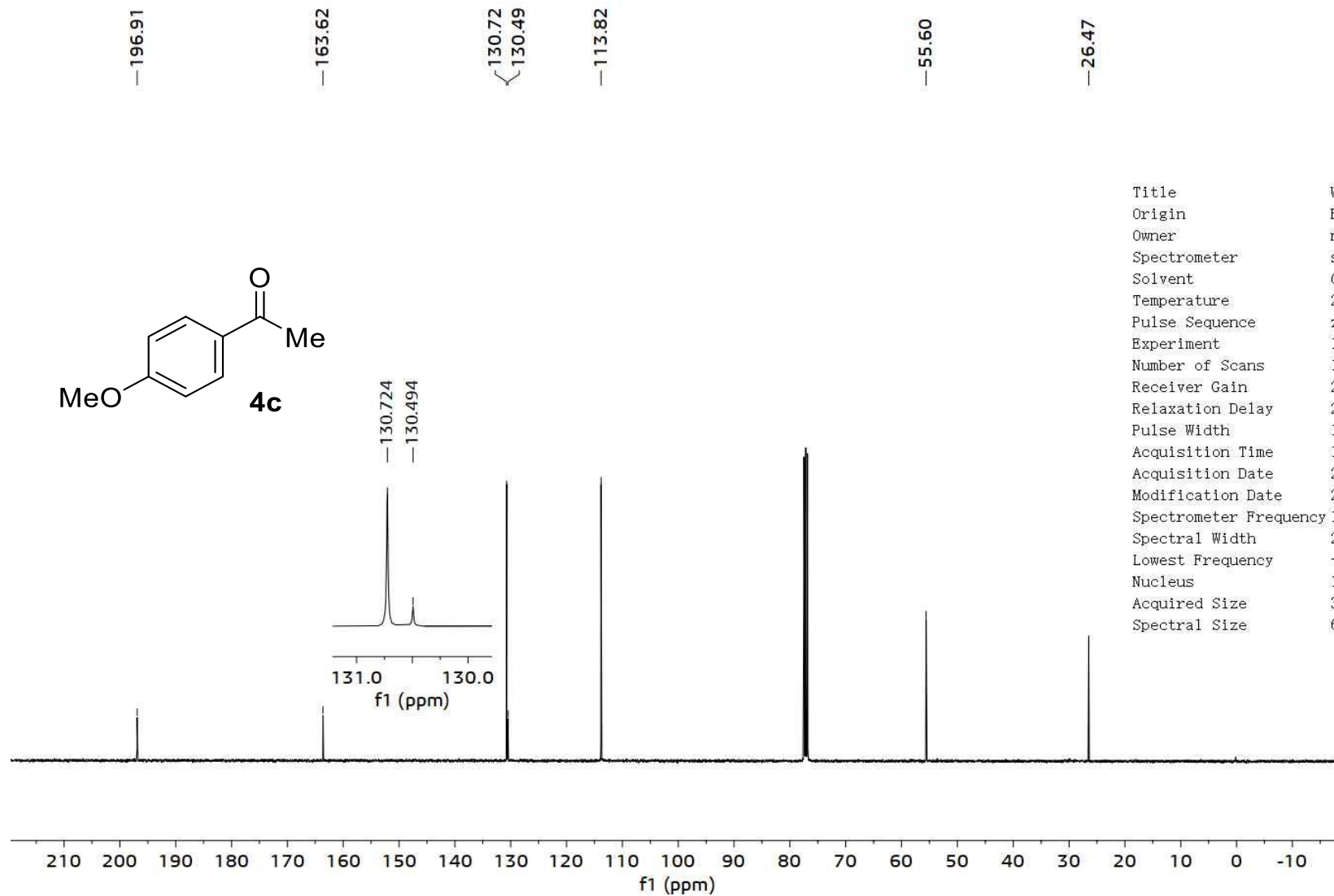
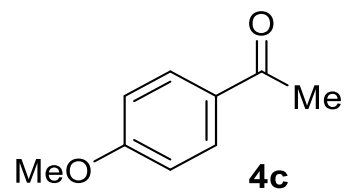
145.94
140.04
136.02
129.10
129.06
128.38
127.42
127.38

—26.82

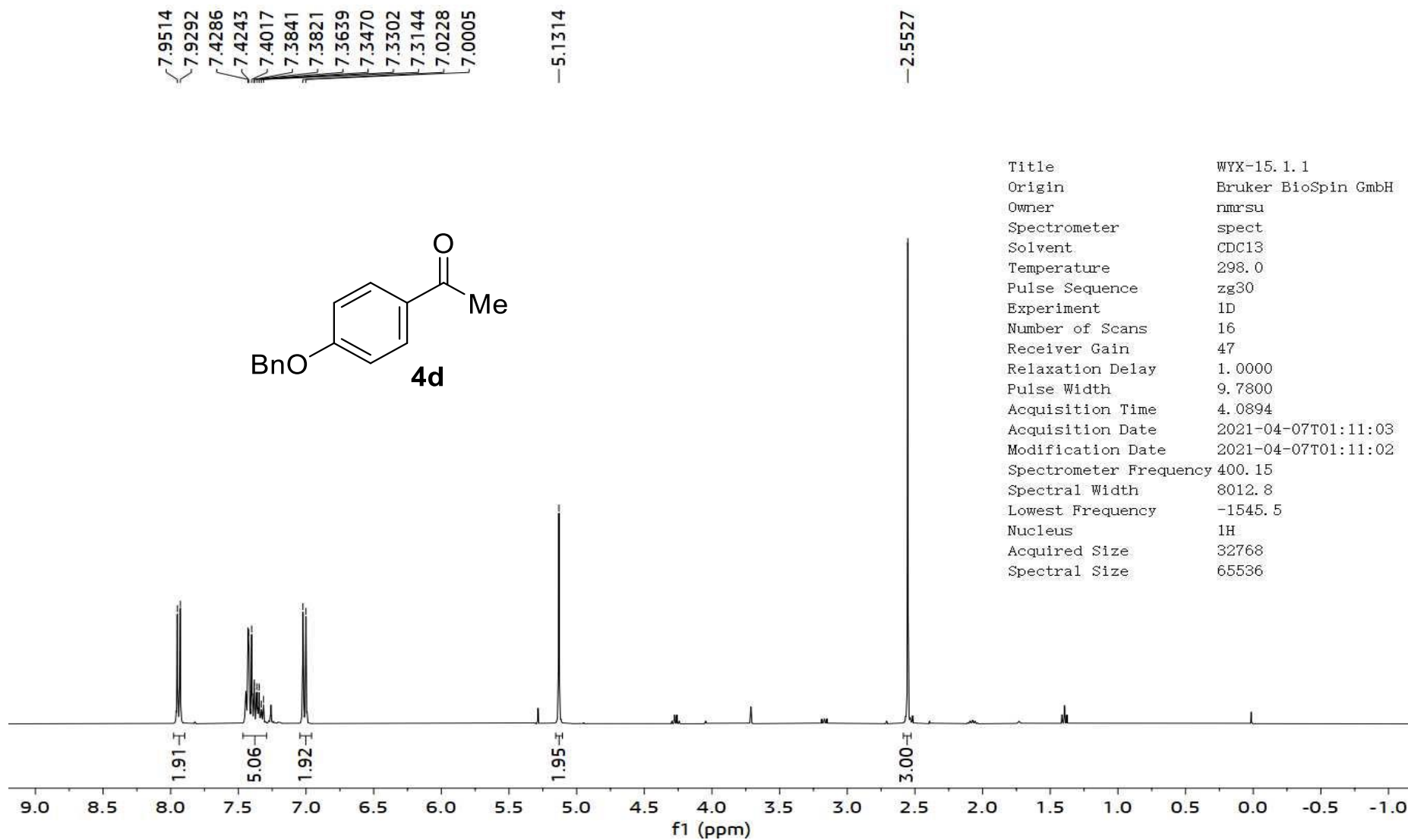


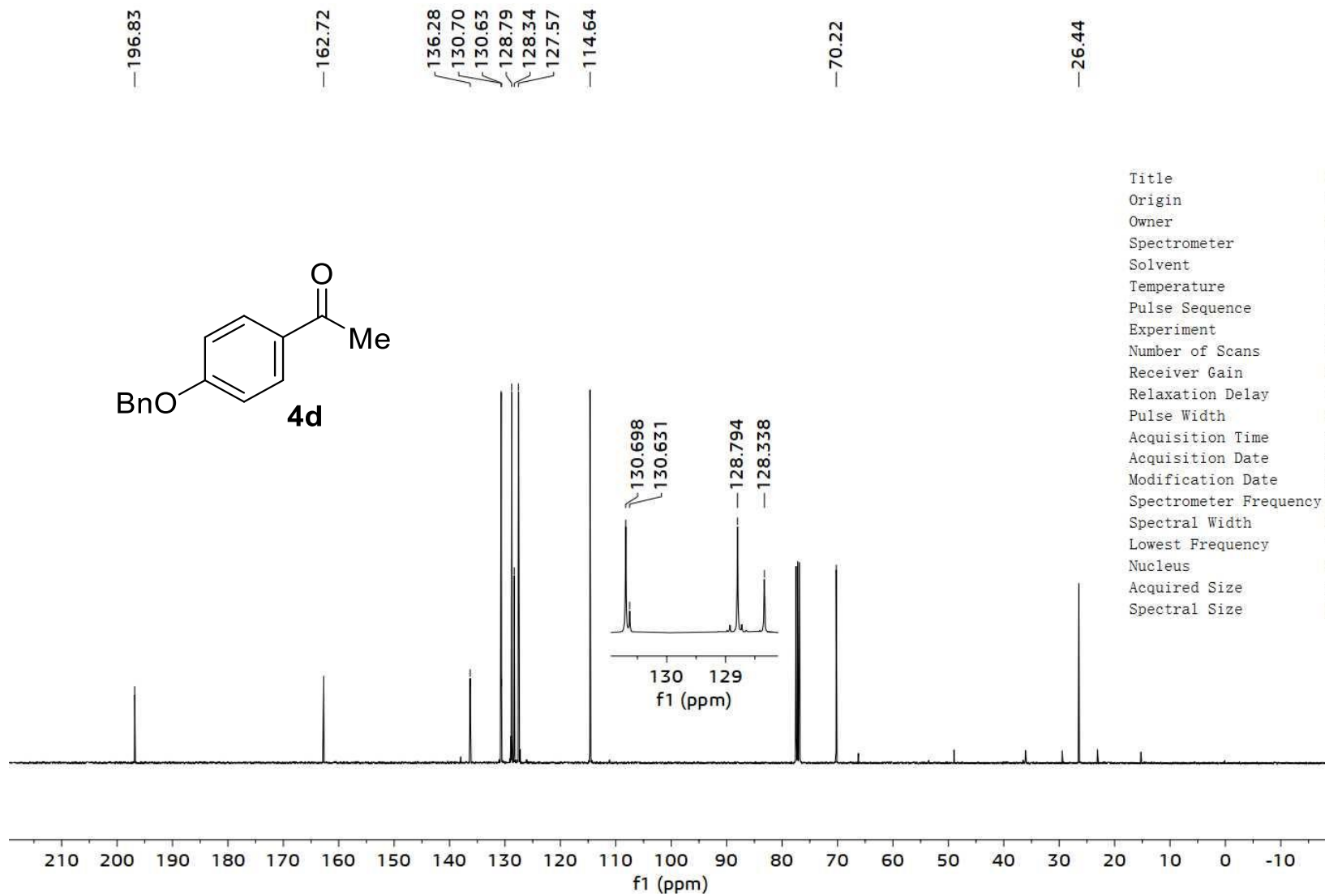
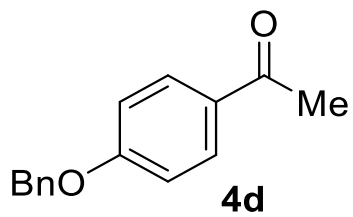
Title	WYX-9. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmr-su
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	800
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-19T14:00:01
Modification Date	2021-03-19T14:00:04
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1943.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



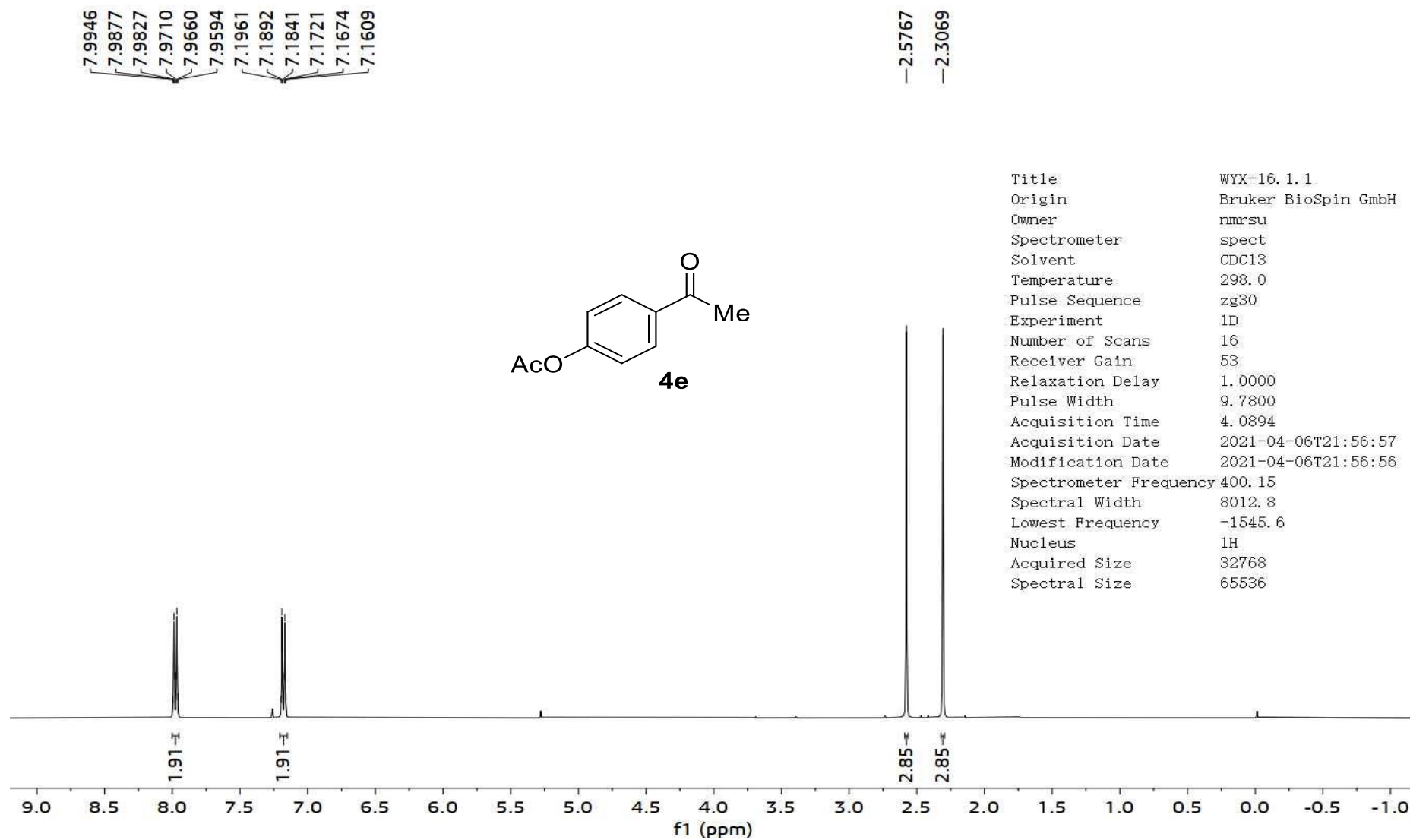


Title	WYX-10.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-19T13:09:55
Modification Date	2021-03-19T13:09:56
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1945.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





Title	WYX-15.2.1
Origin	Bruker BioSpin GmbH
Owner	nmr-su
Spectrometer	spect
Solvent	CDCl ₃
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-07T02:10:42
Modification Date	2021-04-07T02:10:42
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1950.6
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



—196.90

—168.92

—154.43

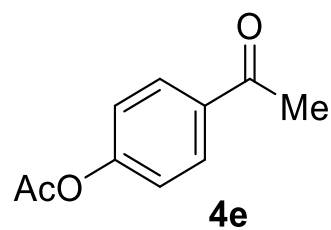
^134.81

^130.02

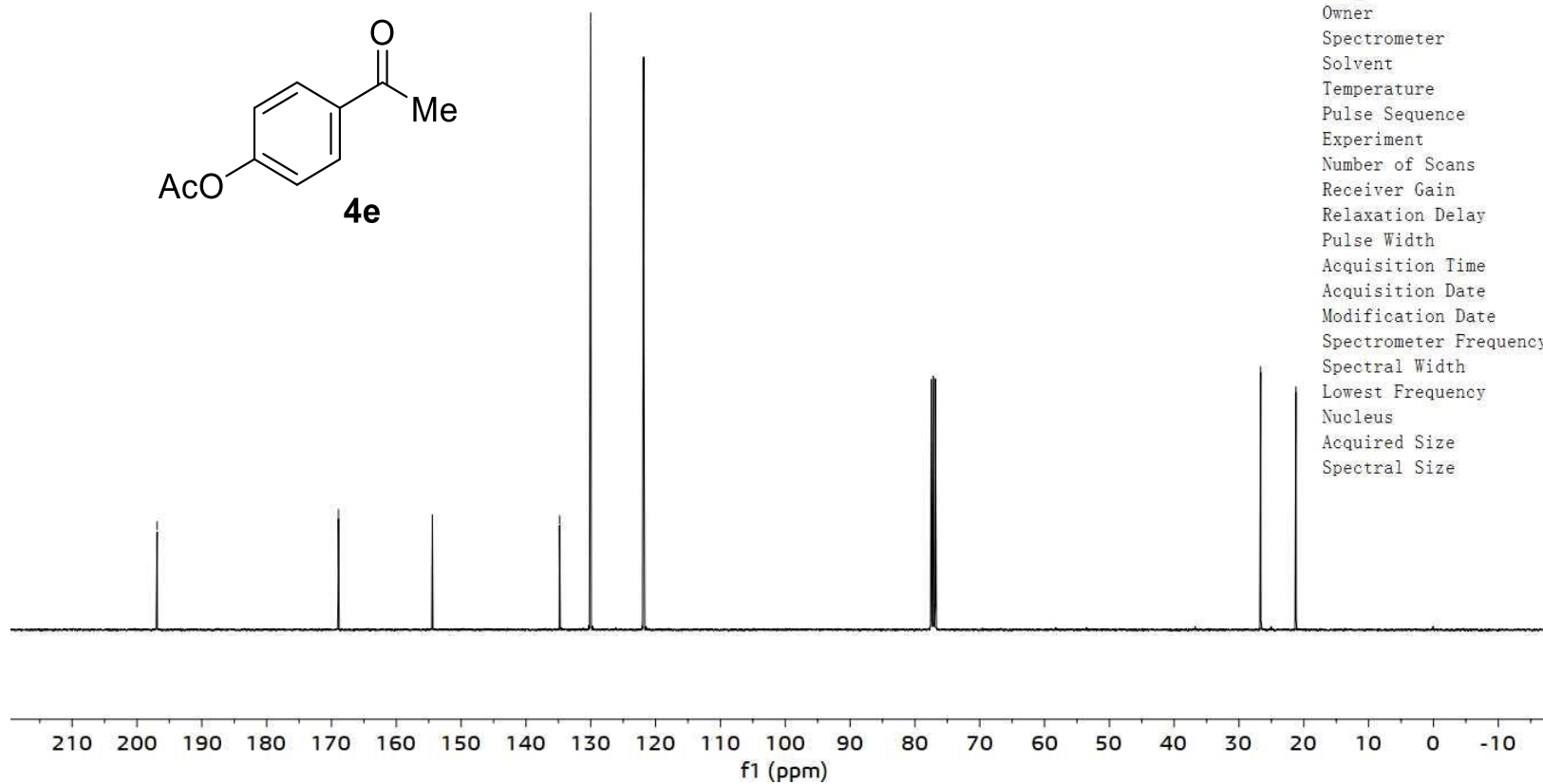
^121.85

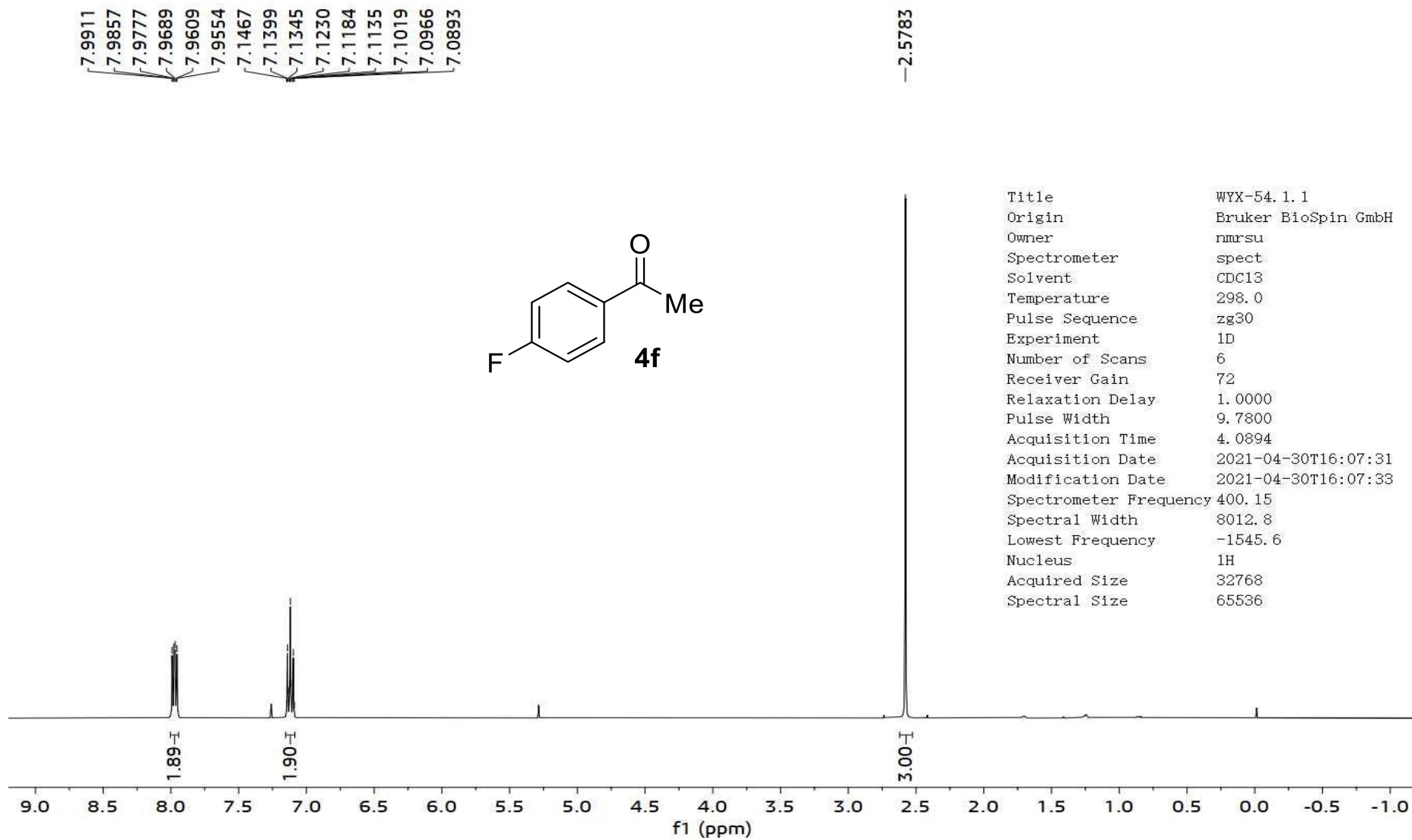
—26.66

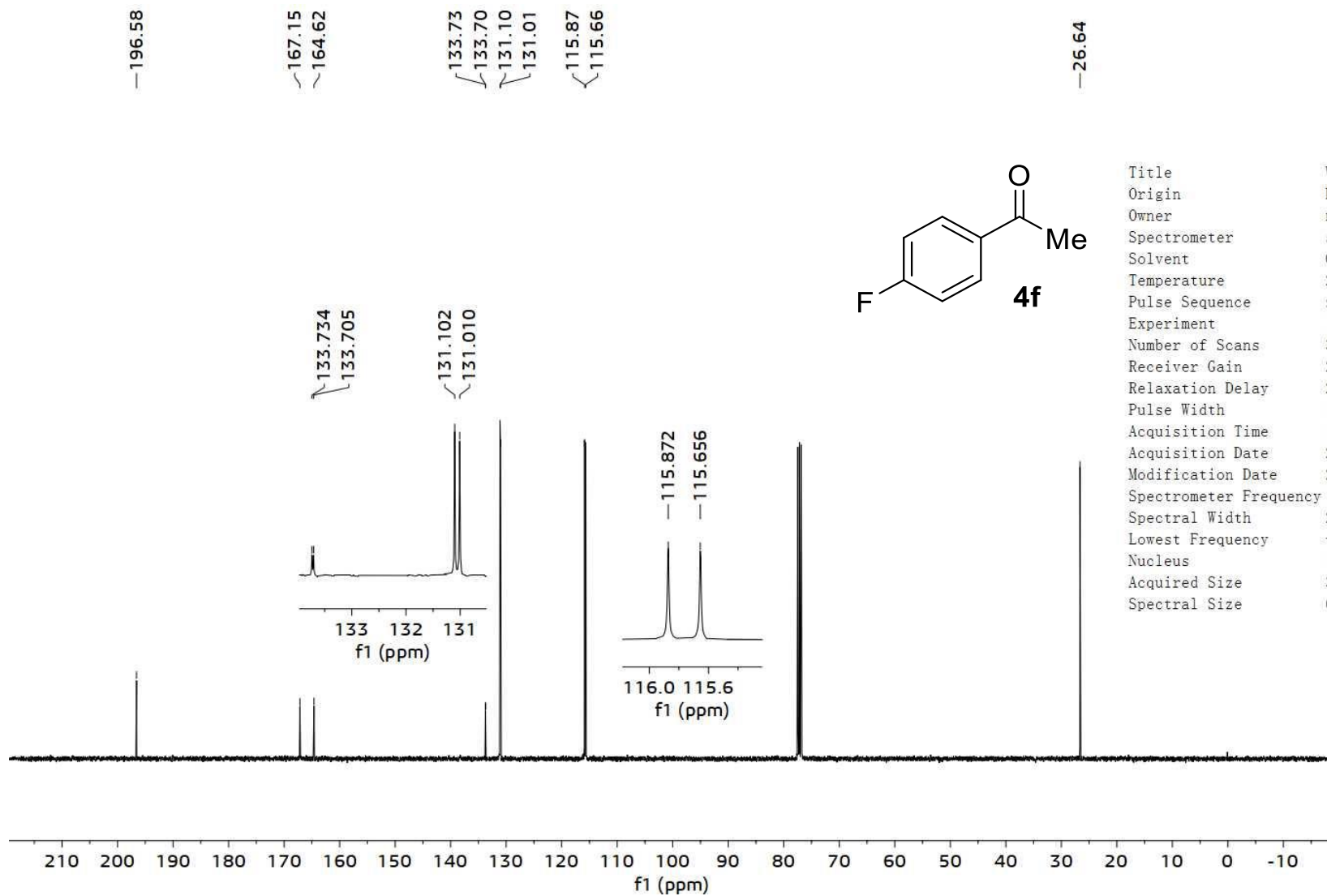
—21.21



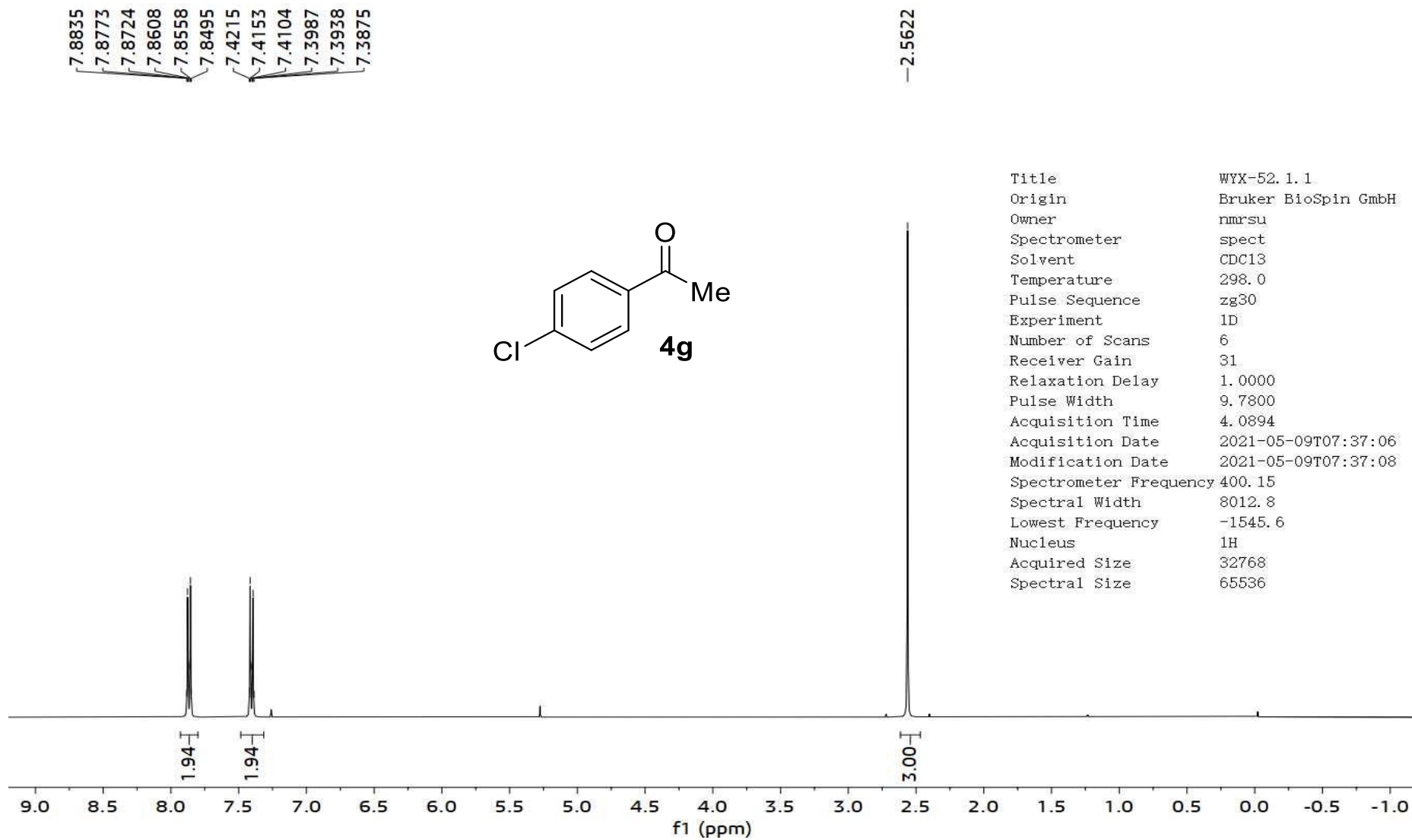
Title	WYX-16.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-06T22:56:31
Modification Date	2021-04-06T22:56:30
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1950.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

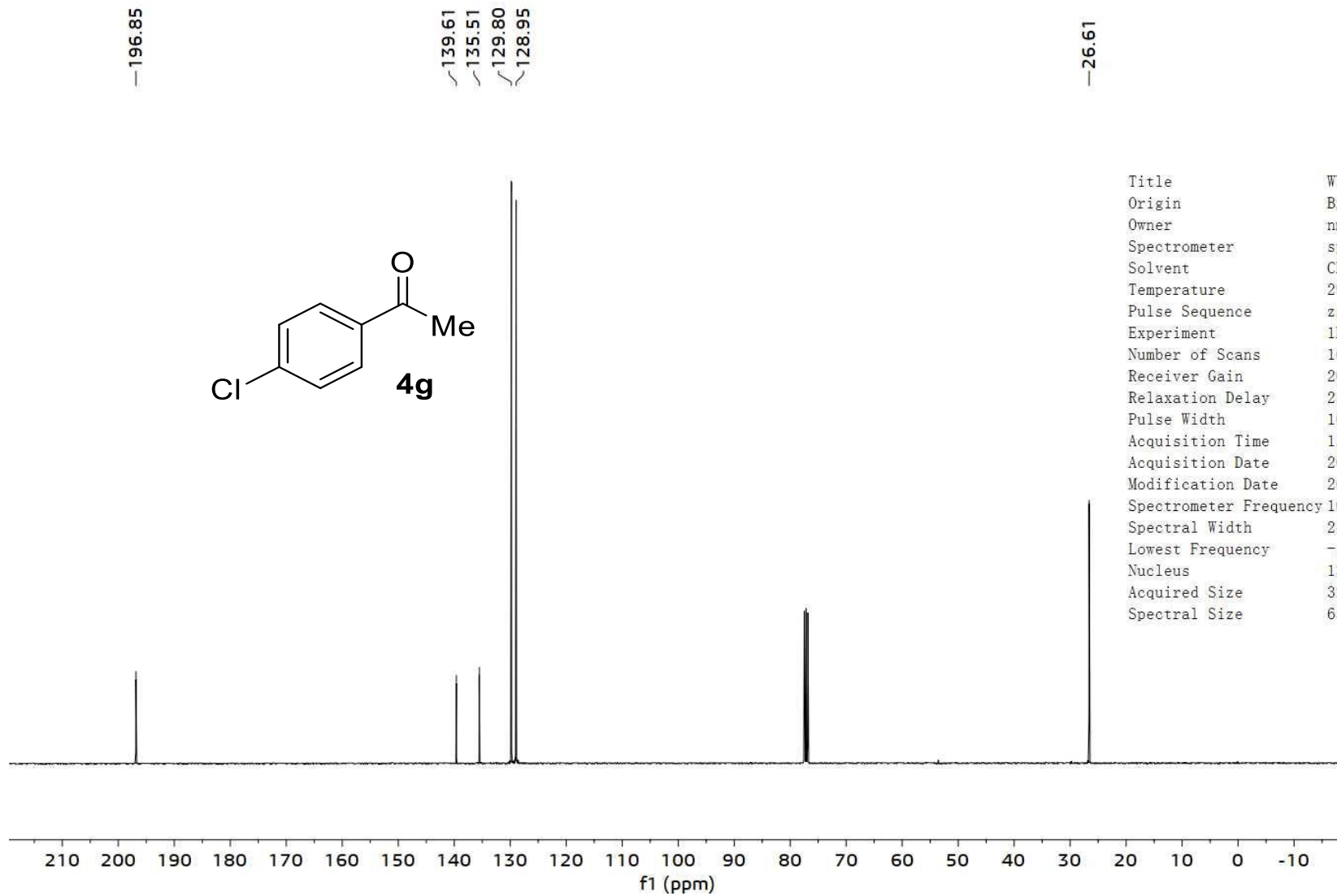




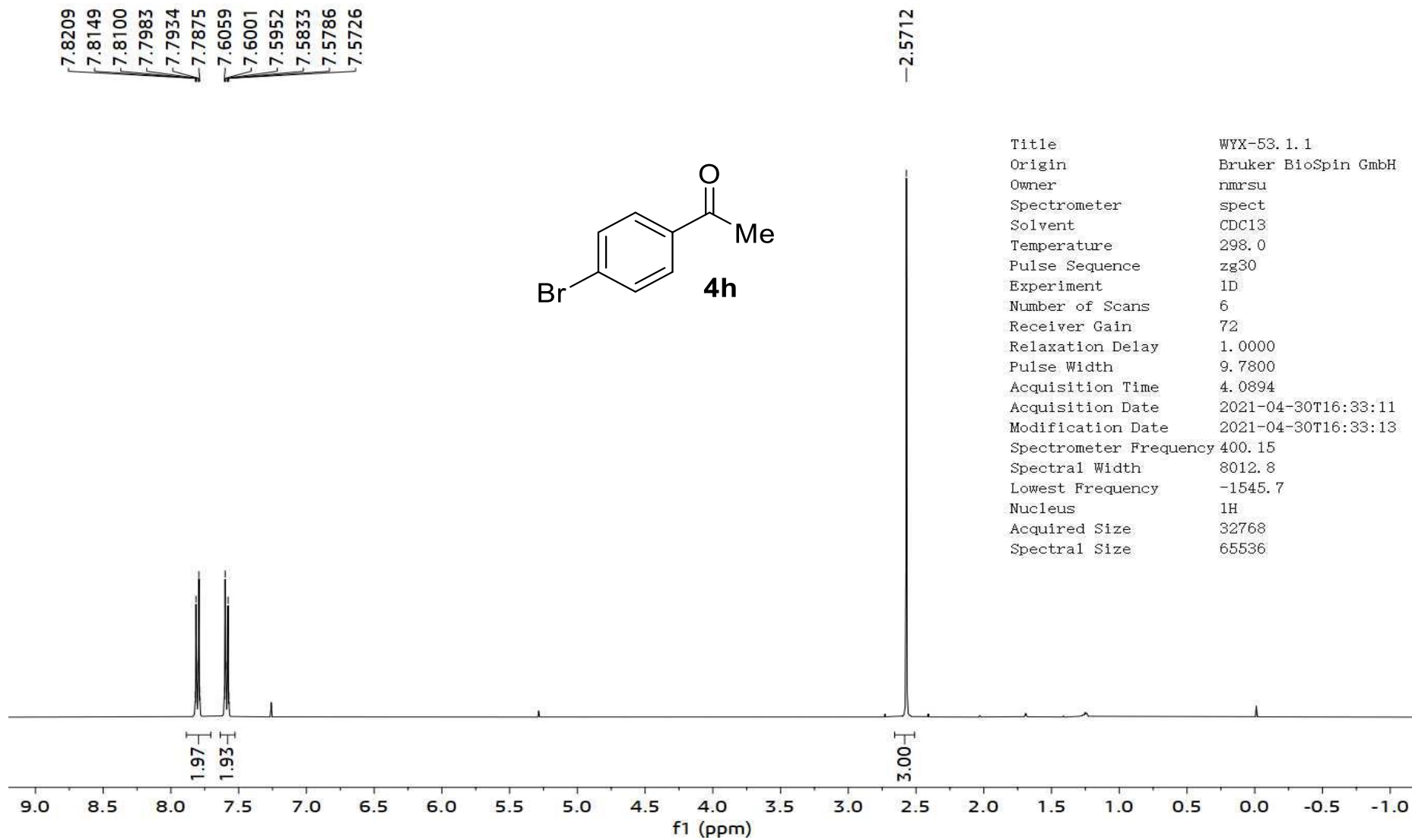


Title	WYX-54.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	350
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-30T16:28:38
Modification Date	2021-04-30T16:28:41
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1946.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





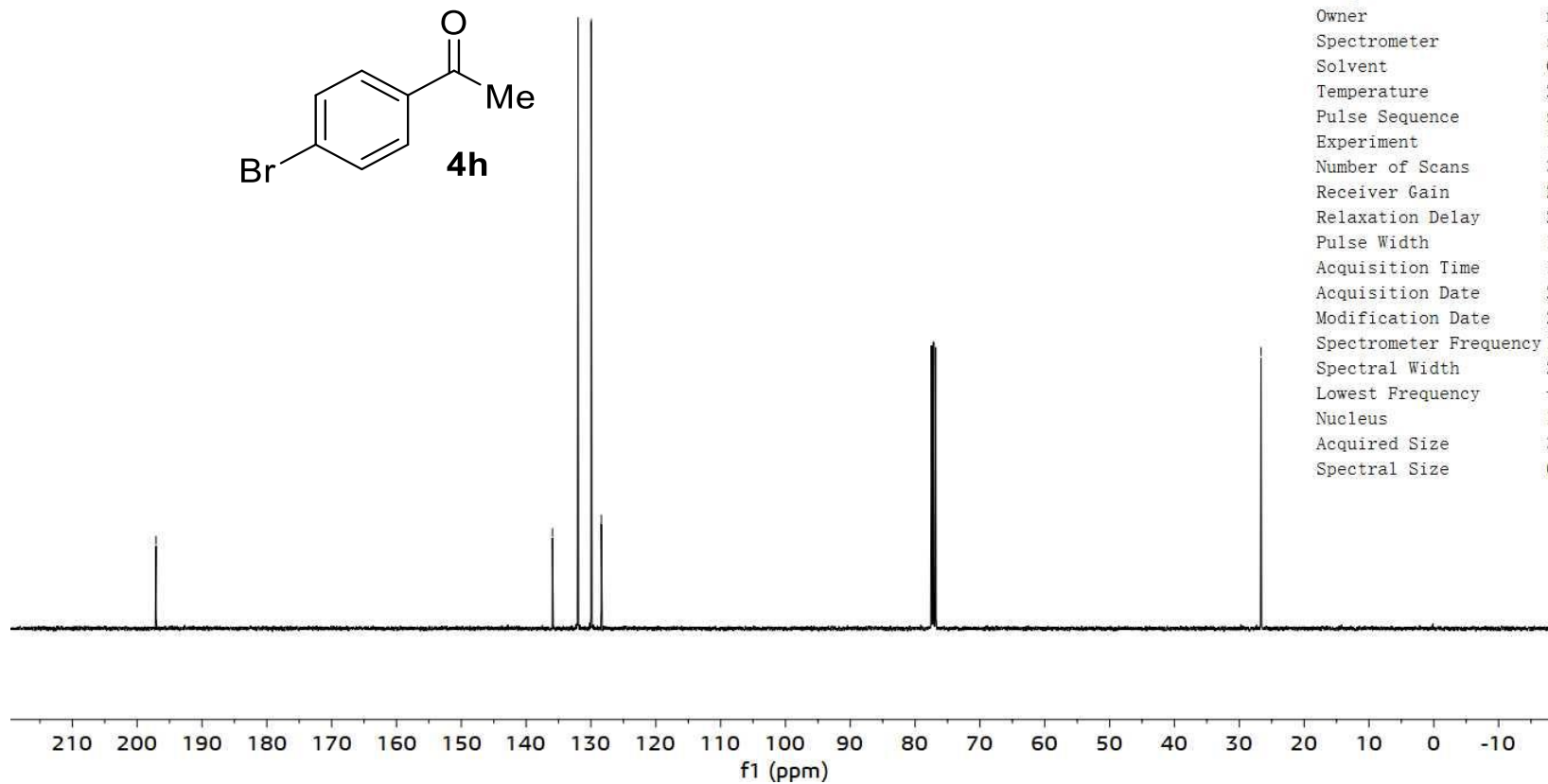
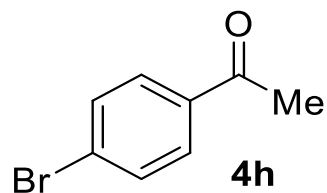
Title	WYX-52.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-09T08:36:40
Modification Date	2021-05-09T08:36:42
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1950.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



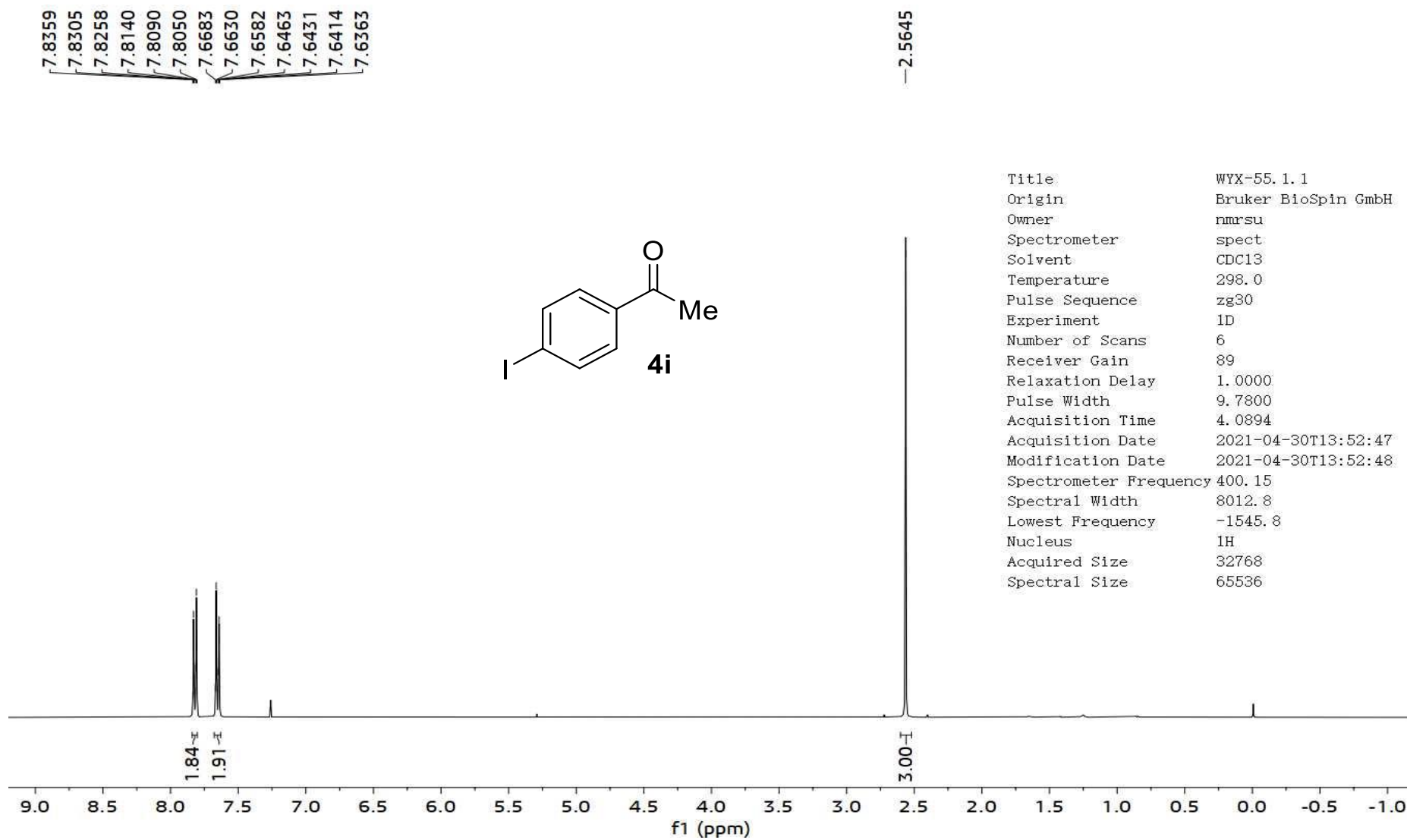
-197.09

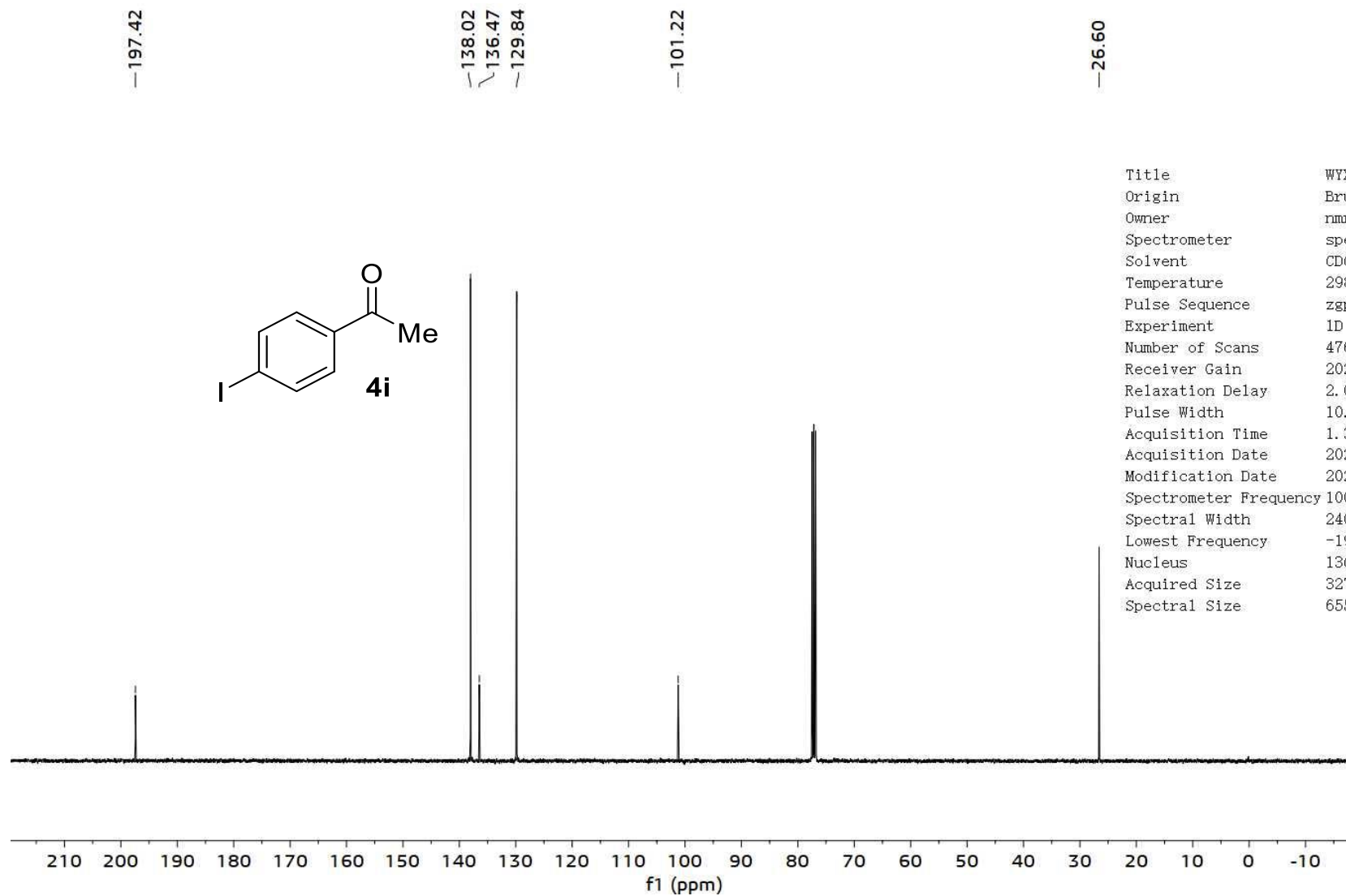
135.94
132.00
129.95
128.40

-26.64

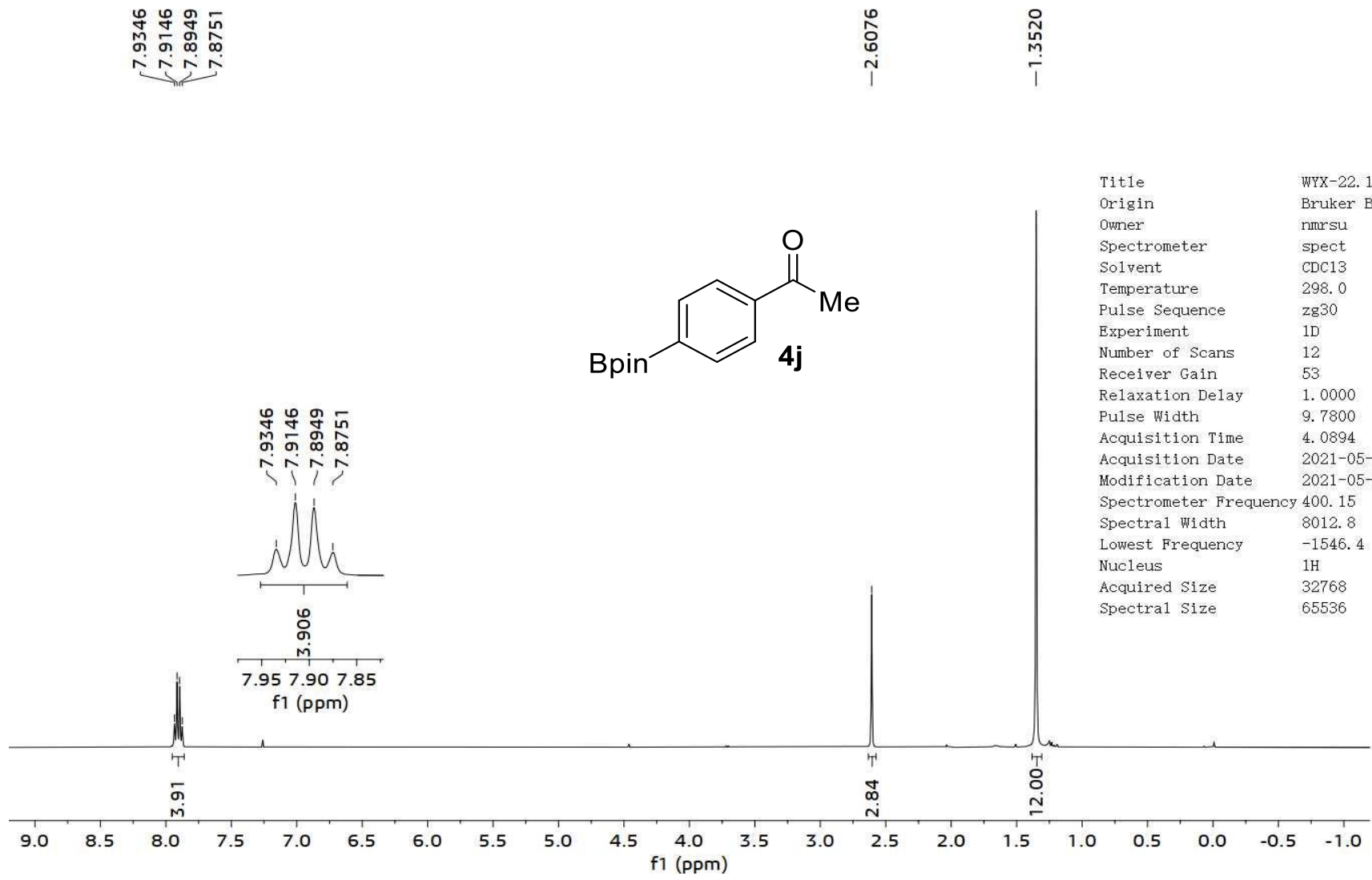


Title	WYX-53.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	350
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-30T16:54:18
Modification Date	2021-04-30T16:54:21
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1947.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





Title	WYX-55.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	476
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-30T14:21:07
Modification Date	2021-04-30T14:21:11
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1947.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



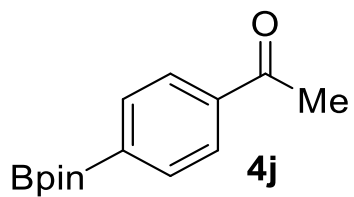
Title	WYX-22. 1. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	12
Receiver Gain	53
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-17T14:13:59
Modification Date	2021-05-17T14:13:58
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1546.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

—198.57

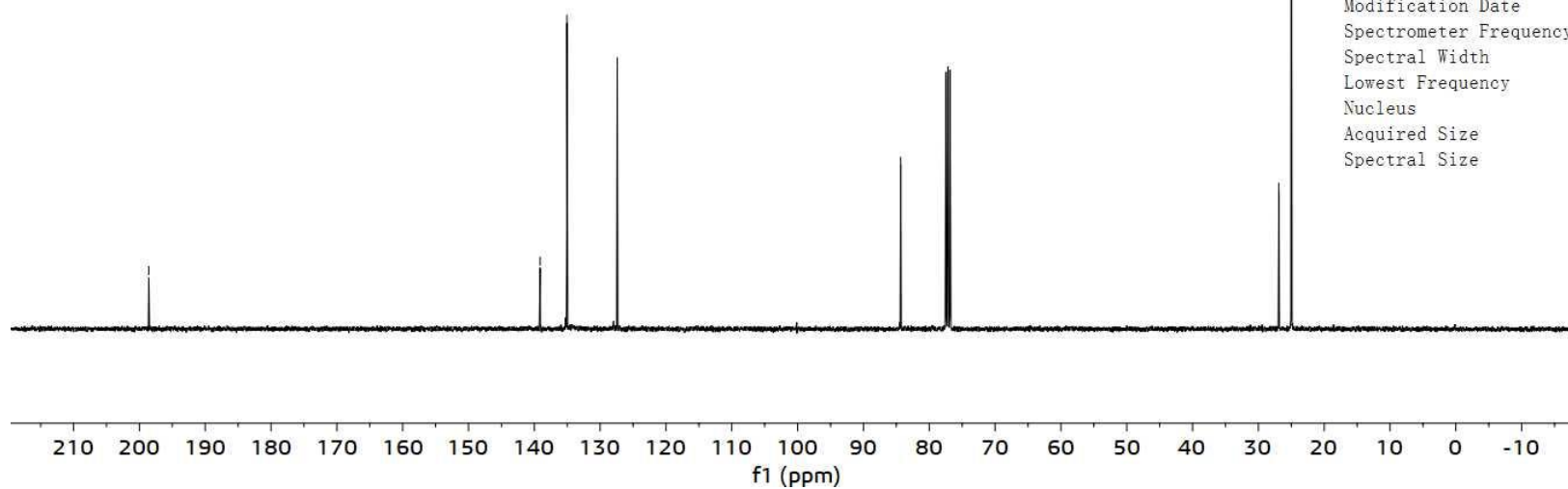
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~135.04
~127.40

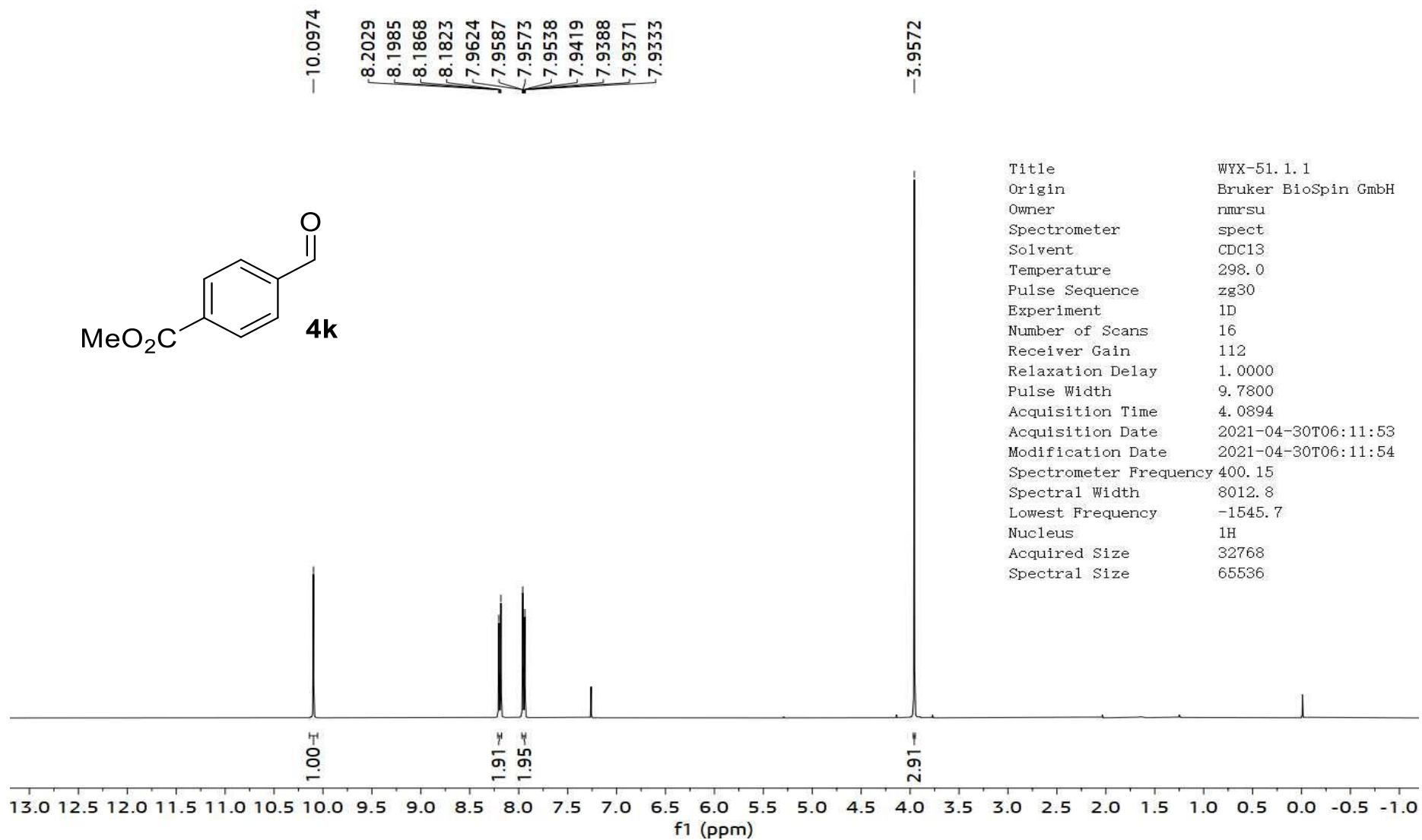
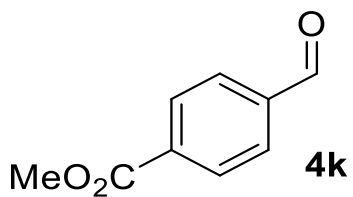
—84.33

~26.89
~25.00



Title	WYX-22.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.1
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	250
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-17T14:29:21
Modification Date	2021-05-17T14:29:22
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1946.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





—191.75

—166.19

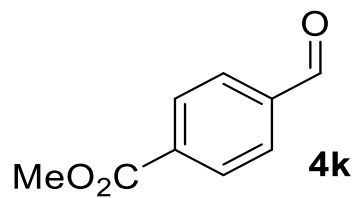
✓139.29

✓135.23

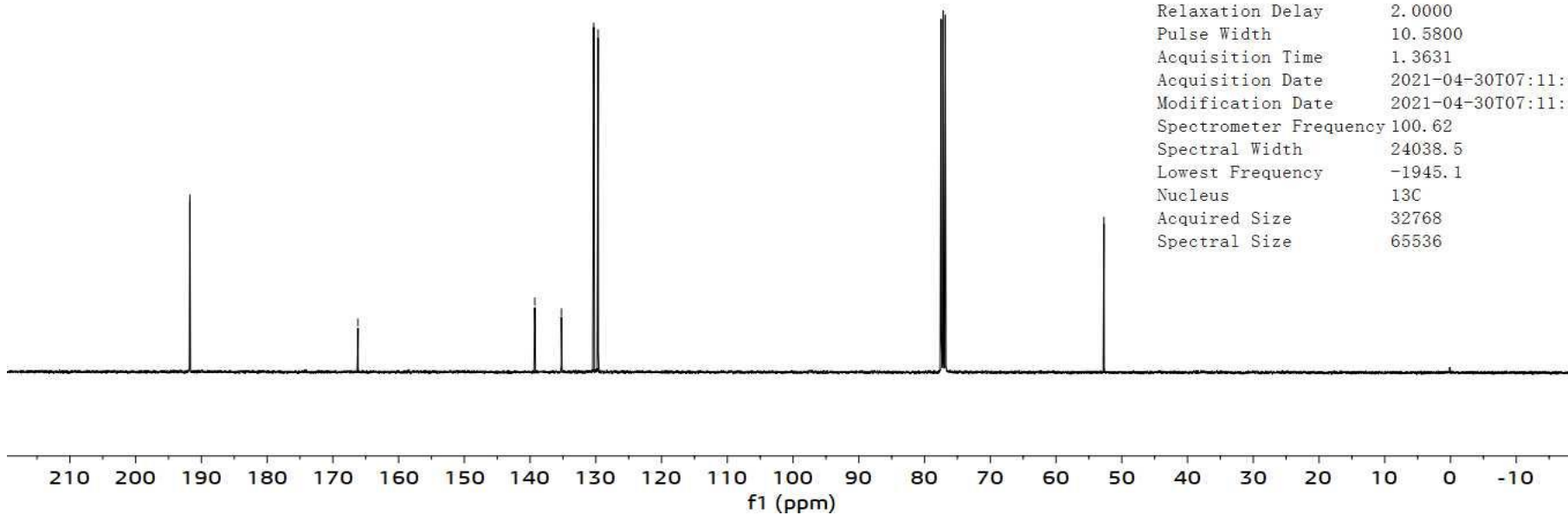
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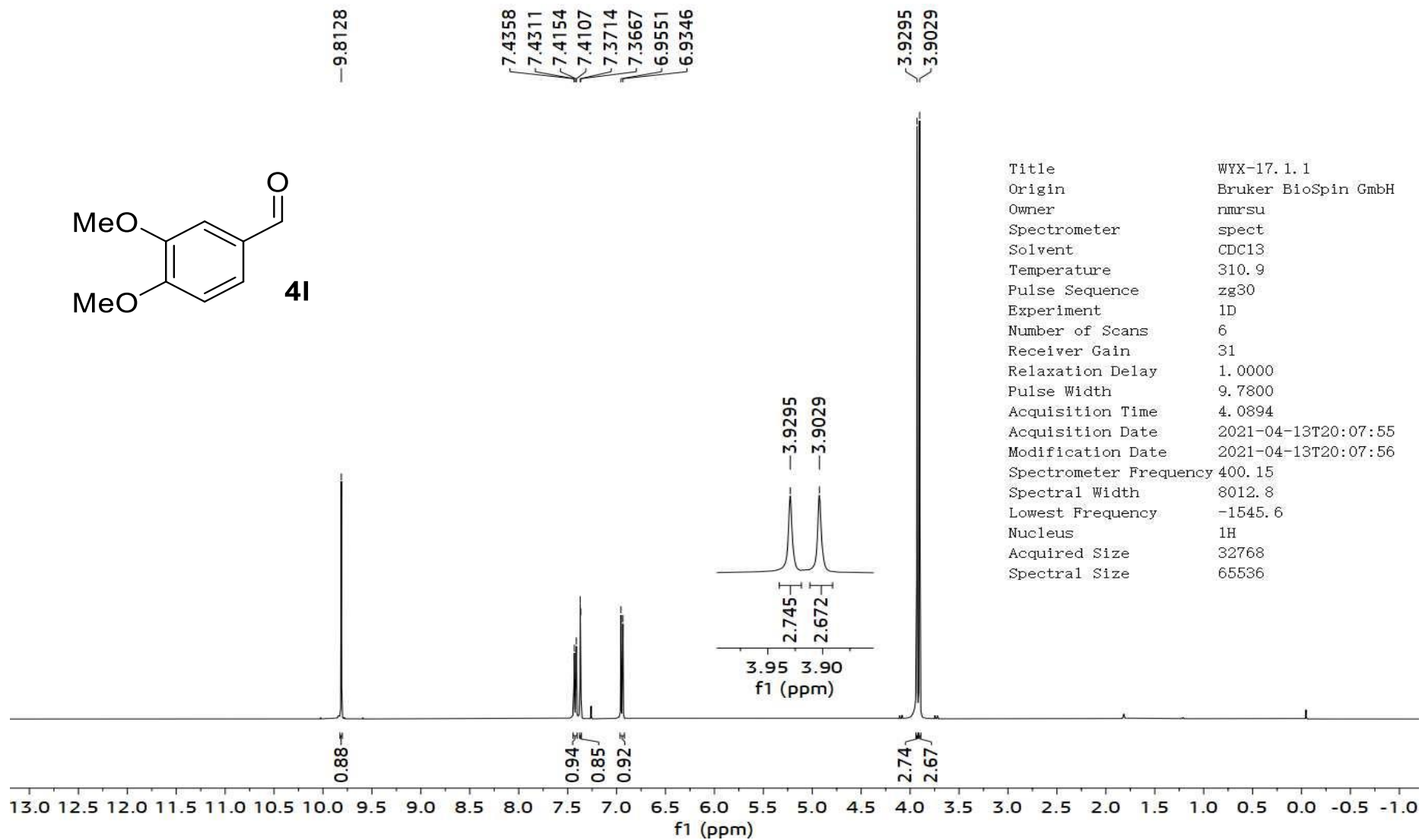
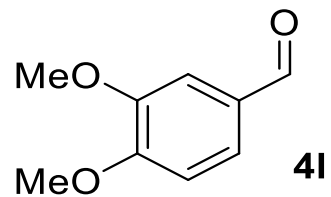
✓129.65

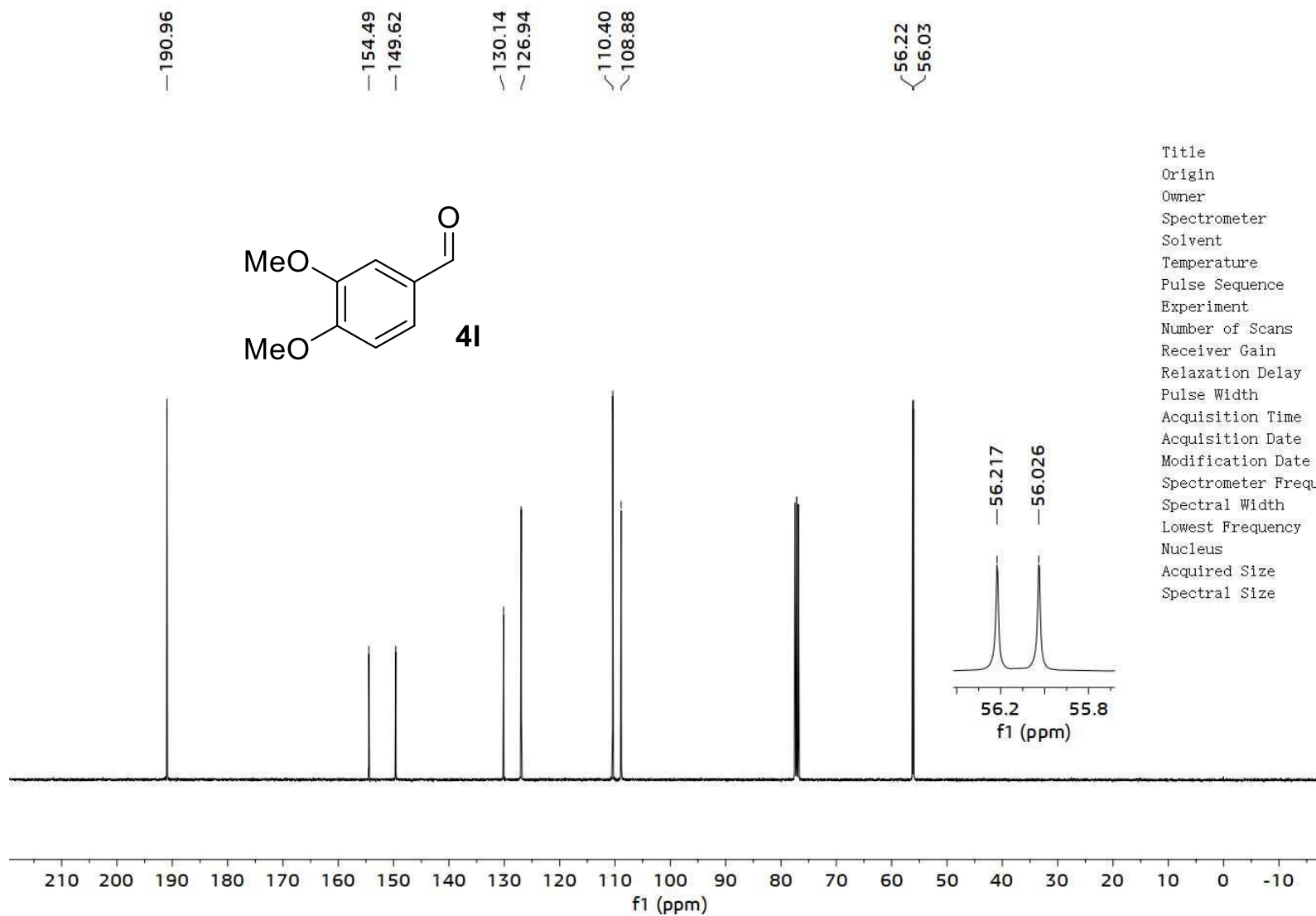
—52.71



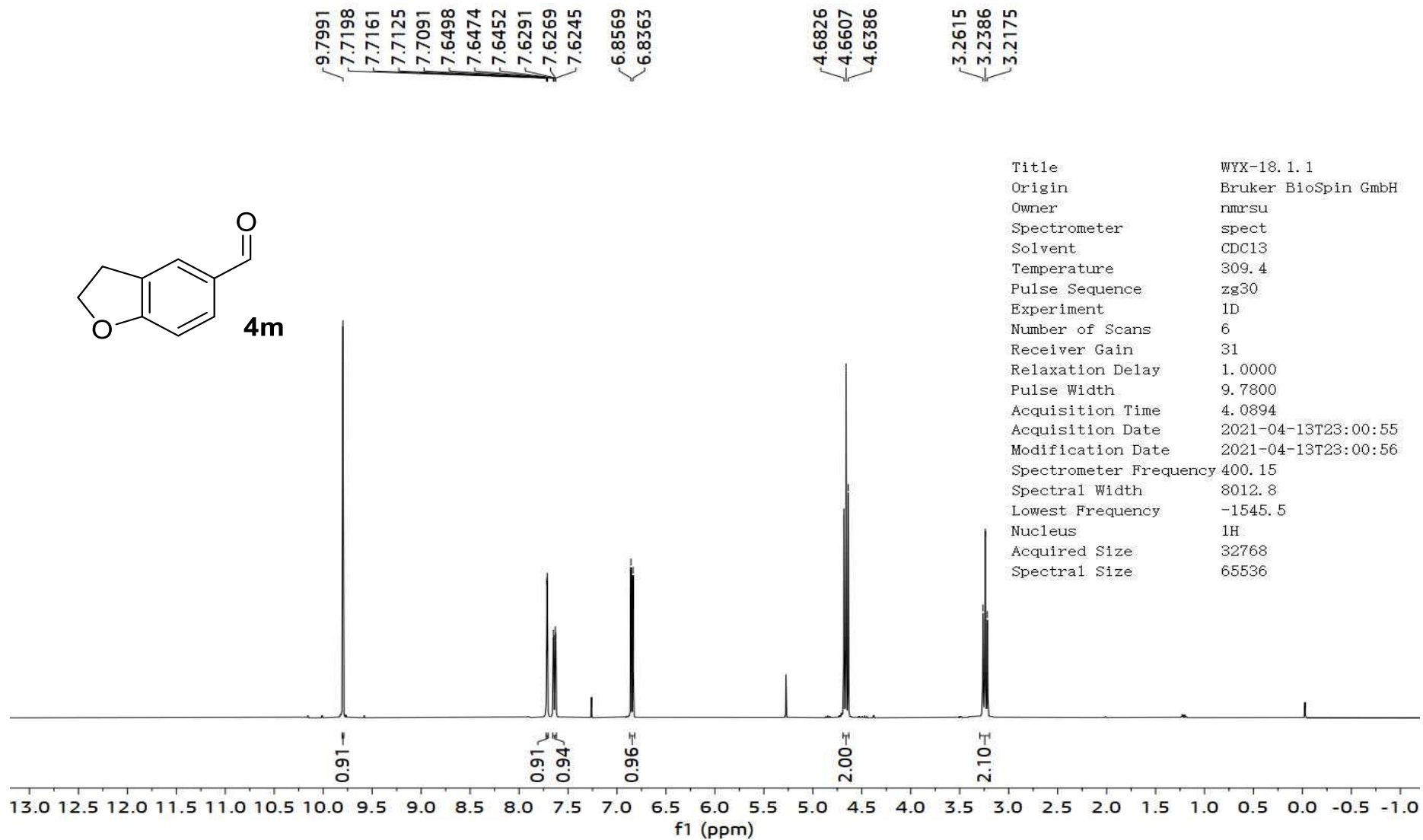
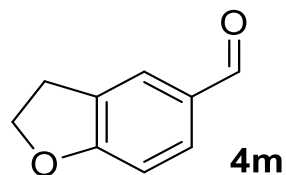
Title	WYX-51.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-30T07:11:30
Modification Date	2021-04-30T07:11:32
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1945.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



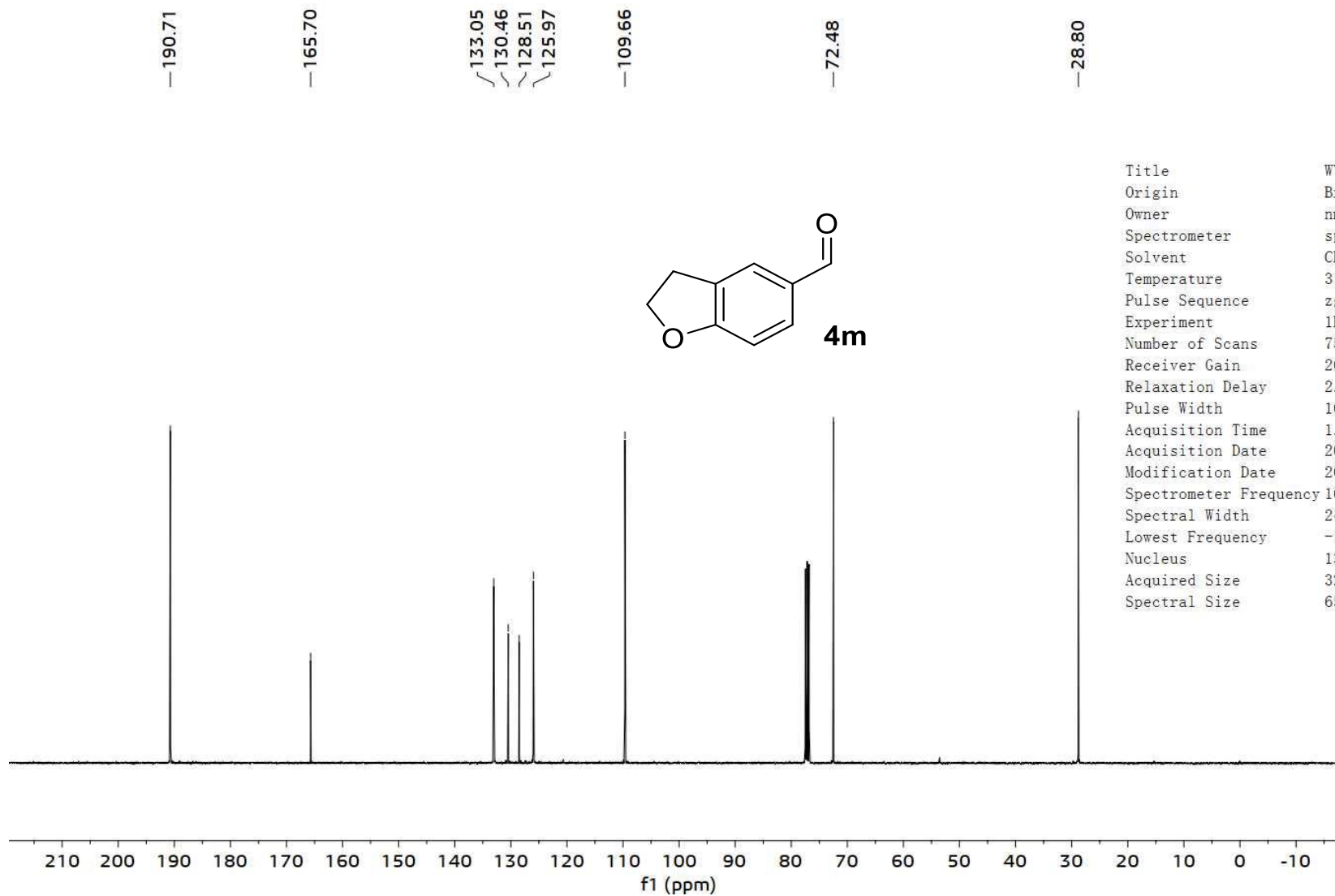




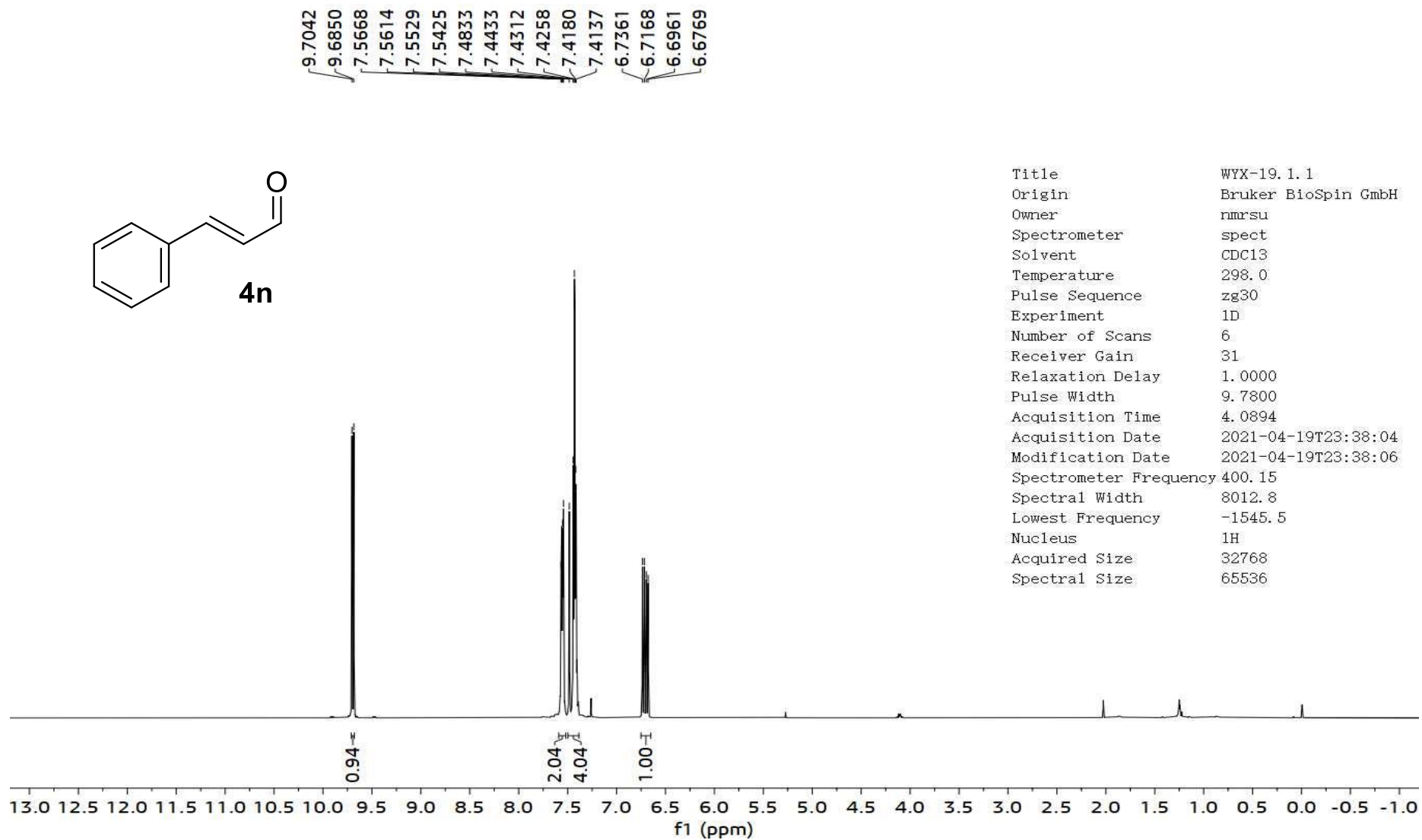
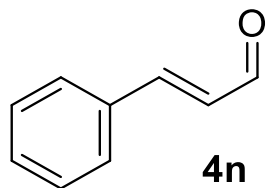
Title	WYX-17. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	310. 8
Pulse Sequence	zpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2. 0000
Pulse Width	10. 5800
Acquisition Time	1. 3631
Acquisition Date	2021-04-13T20:51:55
Modification Date	2021-04-13T20:51:56
Spectrometer Frequency	100. 62
Spectral Width	24038. 5
Lowest Frequency	-1953. 9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

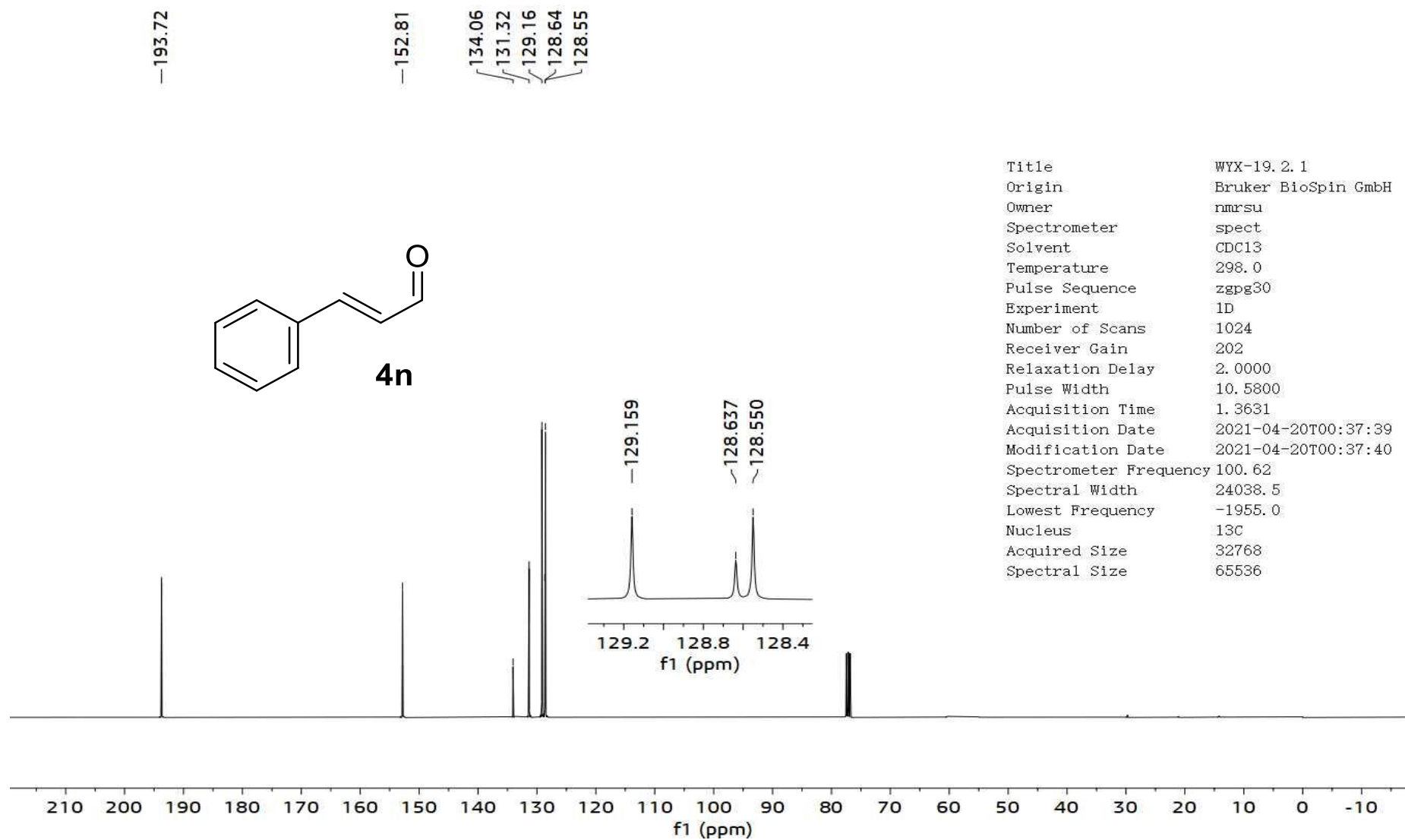


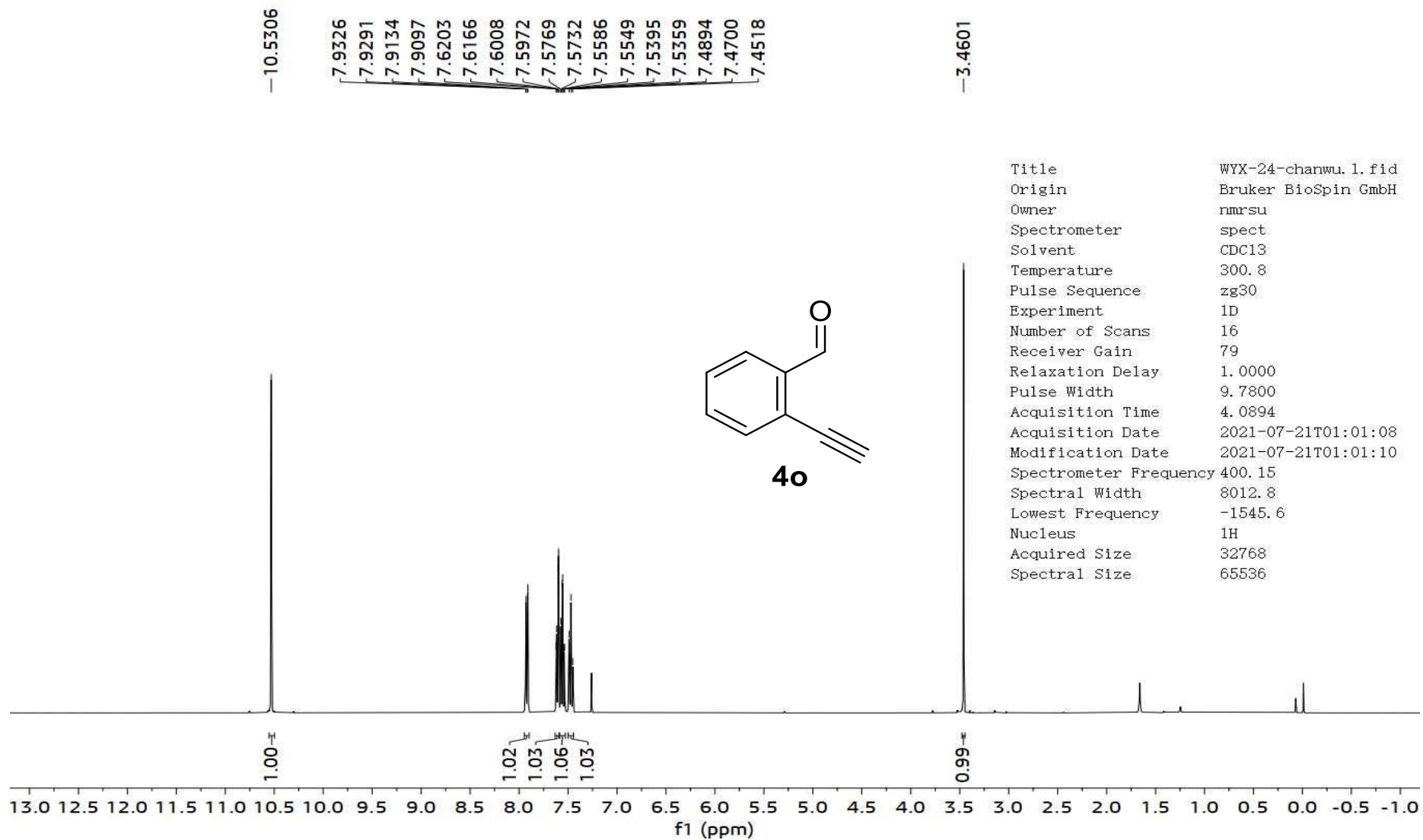
Title	WYX-18.1.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	309.4
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-04-13T23:00:55
Modification Date	2021-04-13T23:00:56
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



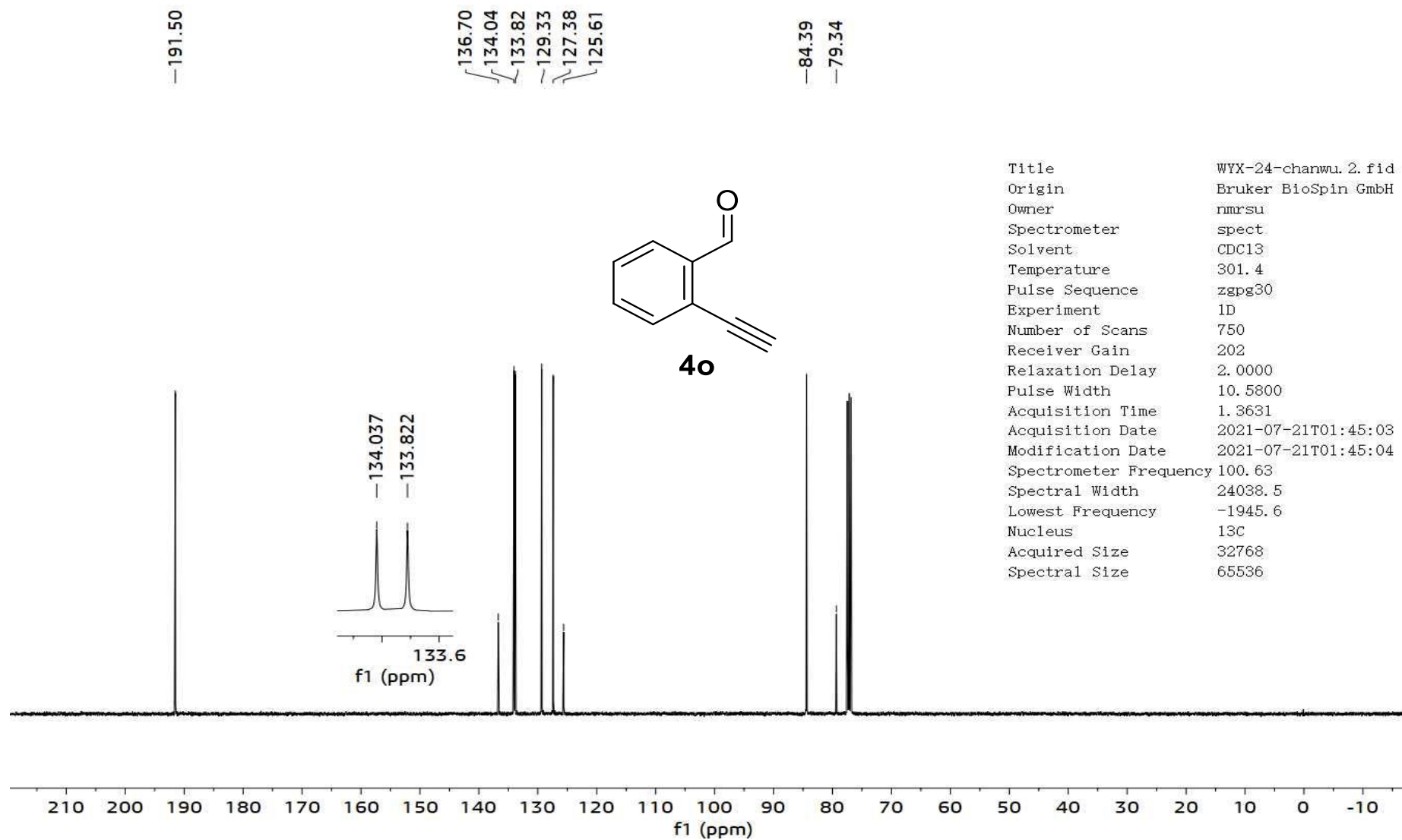
Title	WYX-18.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	312.2
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-13T23:44:55
Modification Date	2021-04-13T23:44:56
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1953.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

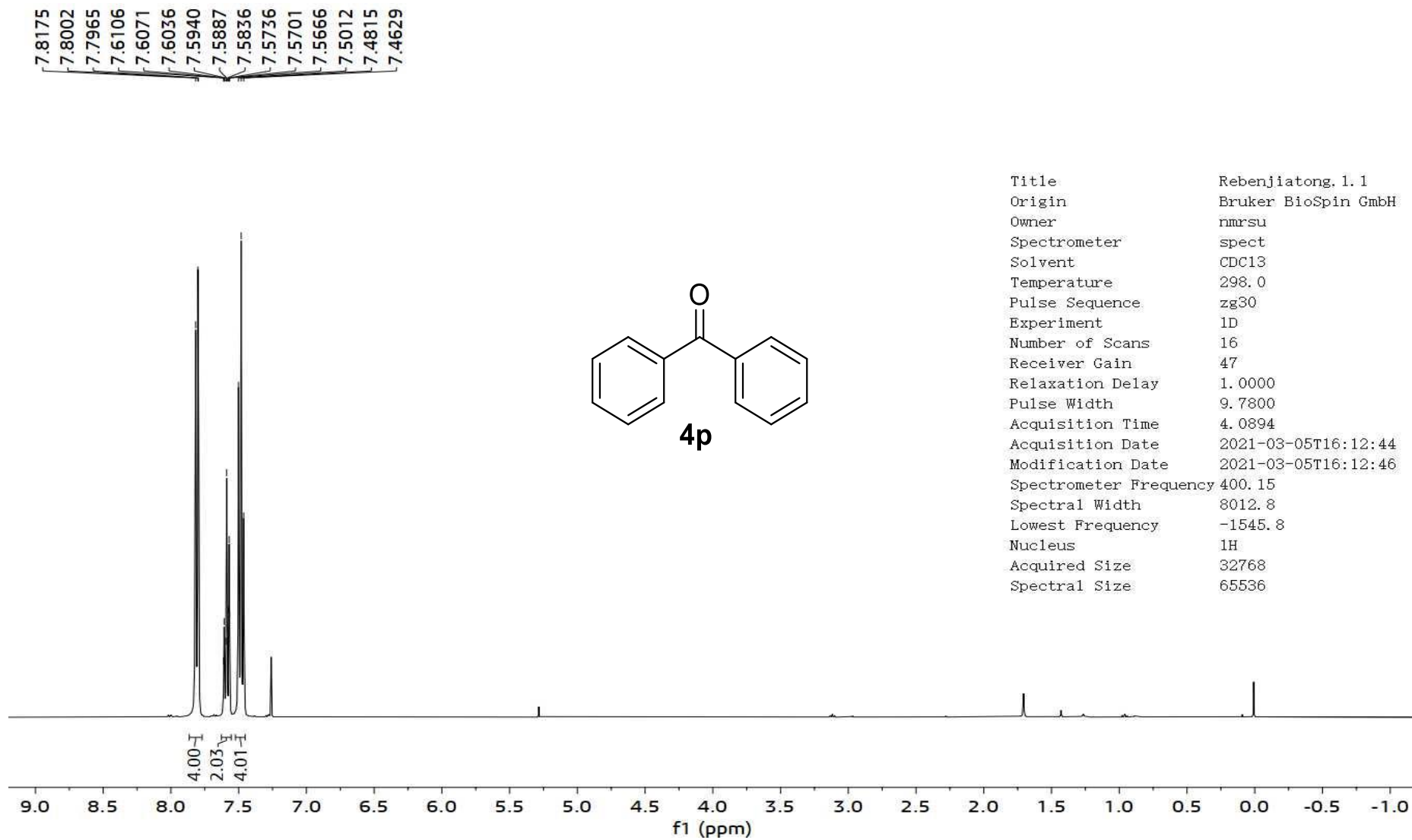






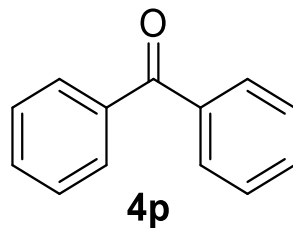
Title	WYX-24-charwu.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	300.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	79
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-21T01:01:08
Modification Date	2021-07-21T01:01:10
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



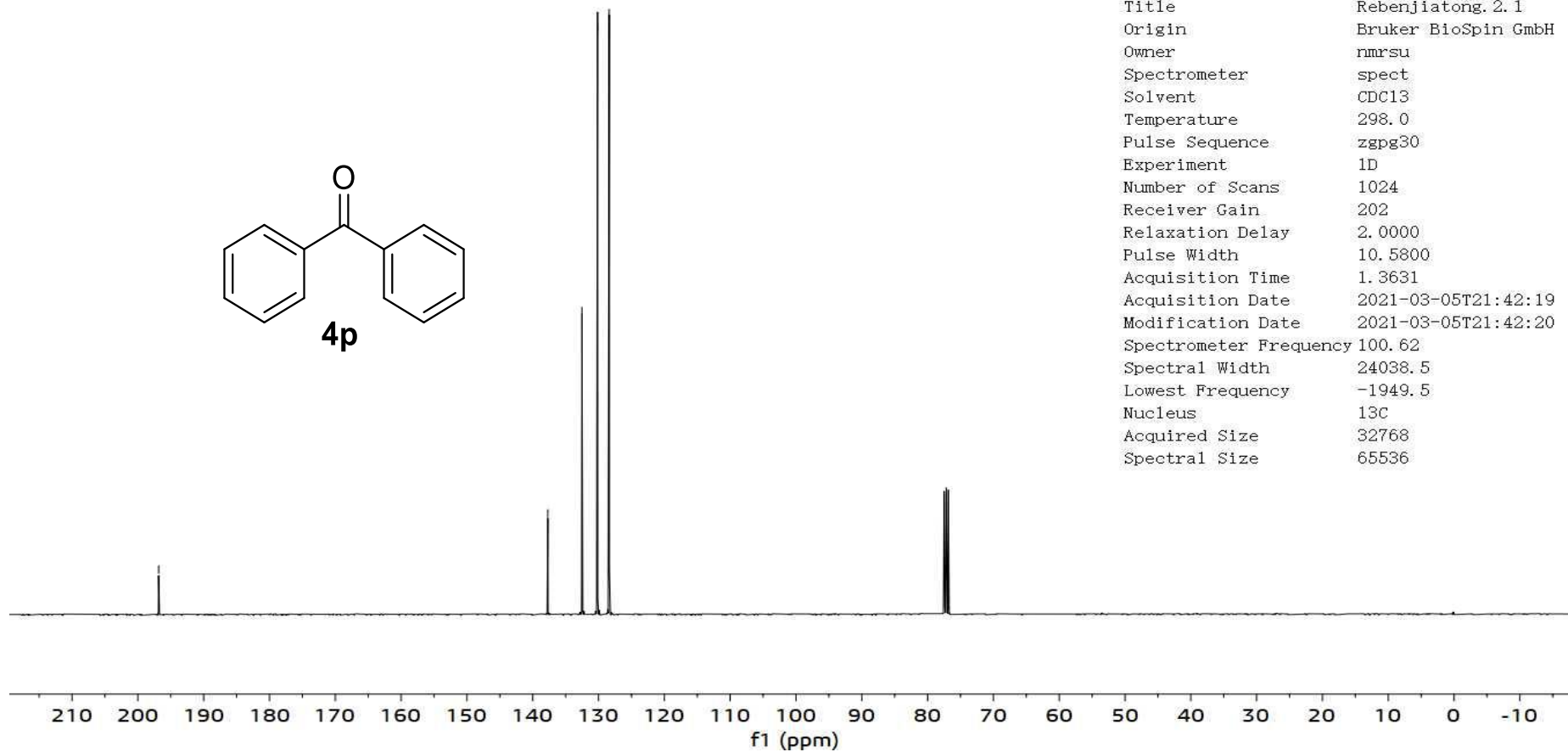


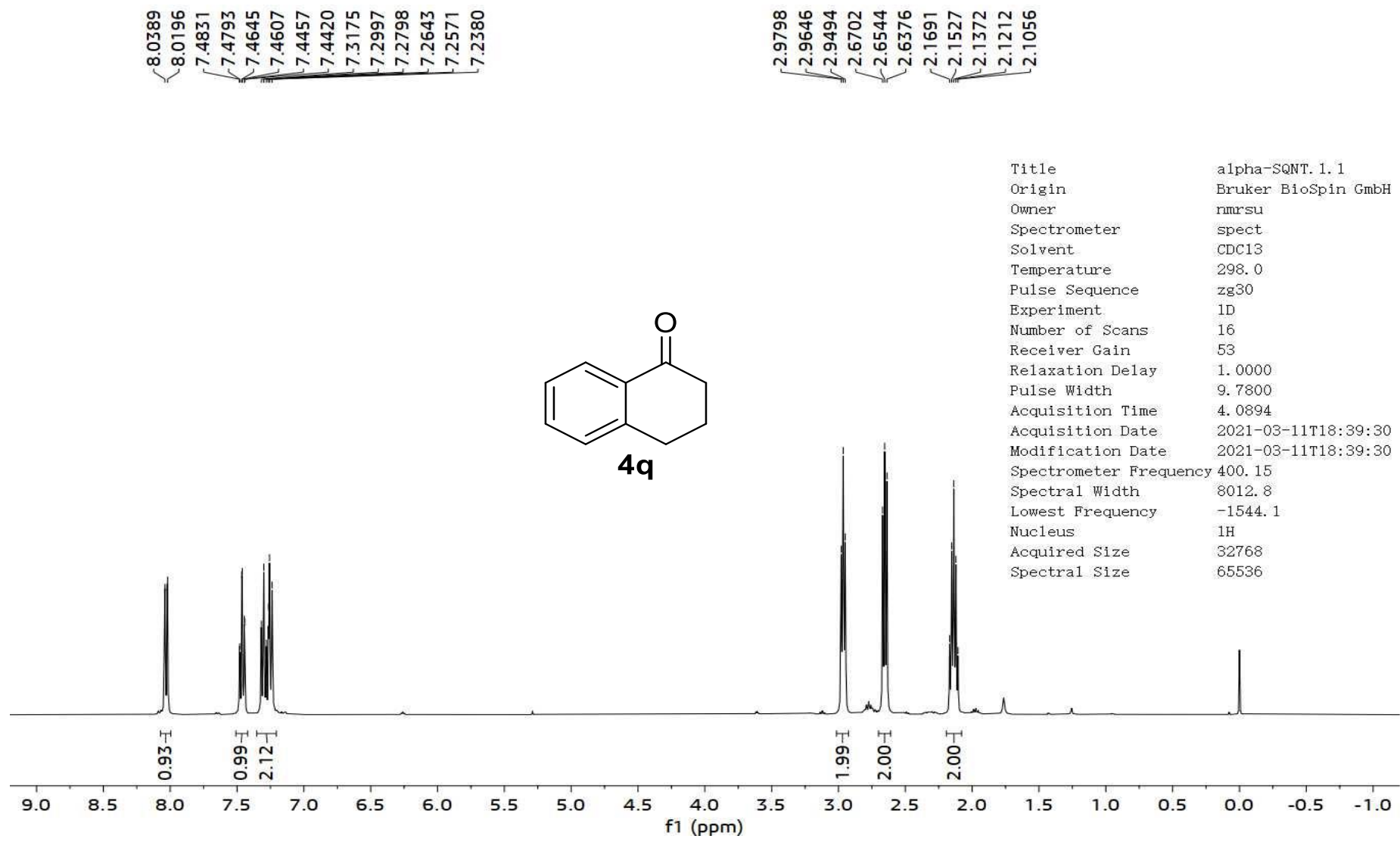
—196.84

✓137.71
✓132.52
✓130.16
✓128.38



Title	Rebenjiatong. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298. 0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2. 0000
Pulse Width	10. 5800
Acquisition Time	1. 3631
Acquisition Date	2021-03-05T21:42:19
Modification Date	2021-03-05T21:42:20
Spectrometer Frequency	100. 62
Spectral Width	24038. 5
Lowest Frequency	-1949. 5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





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Title          alpha-SQNT. 1. 1
Origin         Bruker BioSpin GmbH
Owner          nmrsu
Spectrometer   spect
Solvent        CDC13
Temperature    298.0
Pulse Sequence zg30
Experiment     1D
Number of Scans 16
Receiver Gain  53
Relaxation Delay 1.0000
Pulse Width    9.7800
Acquisition Time 4.0894
Acquisition Date 2021-03-11T18:39:30
Modification Date 2021-03-11T18:39:30
Spectrometer Frequency 400.15
Spectral Width 8012.8
Lowest Frequency -1544.1
Nucleus        1H
Acquired Size  32768
Spectral Size   65536
  
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-198.48

-144.59

133.49

132.72

128.87

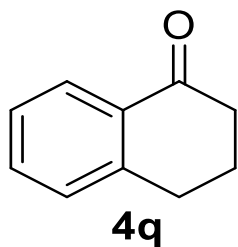
127.26

126.72

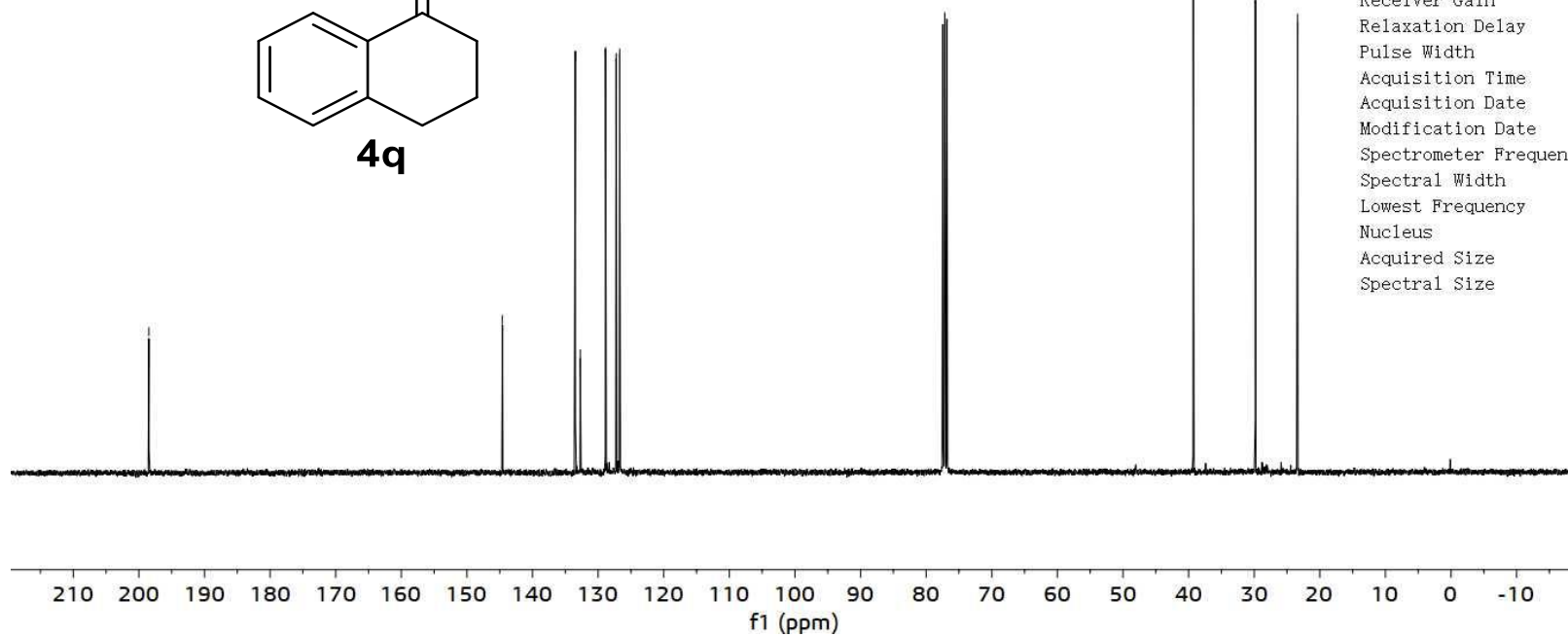
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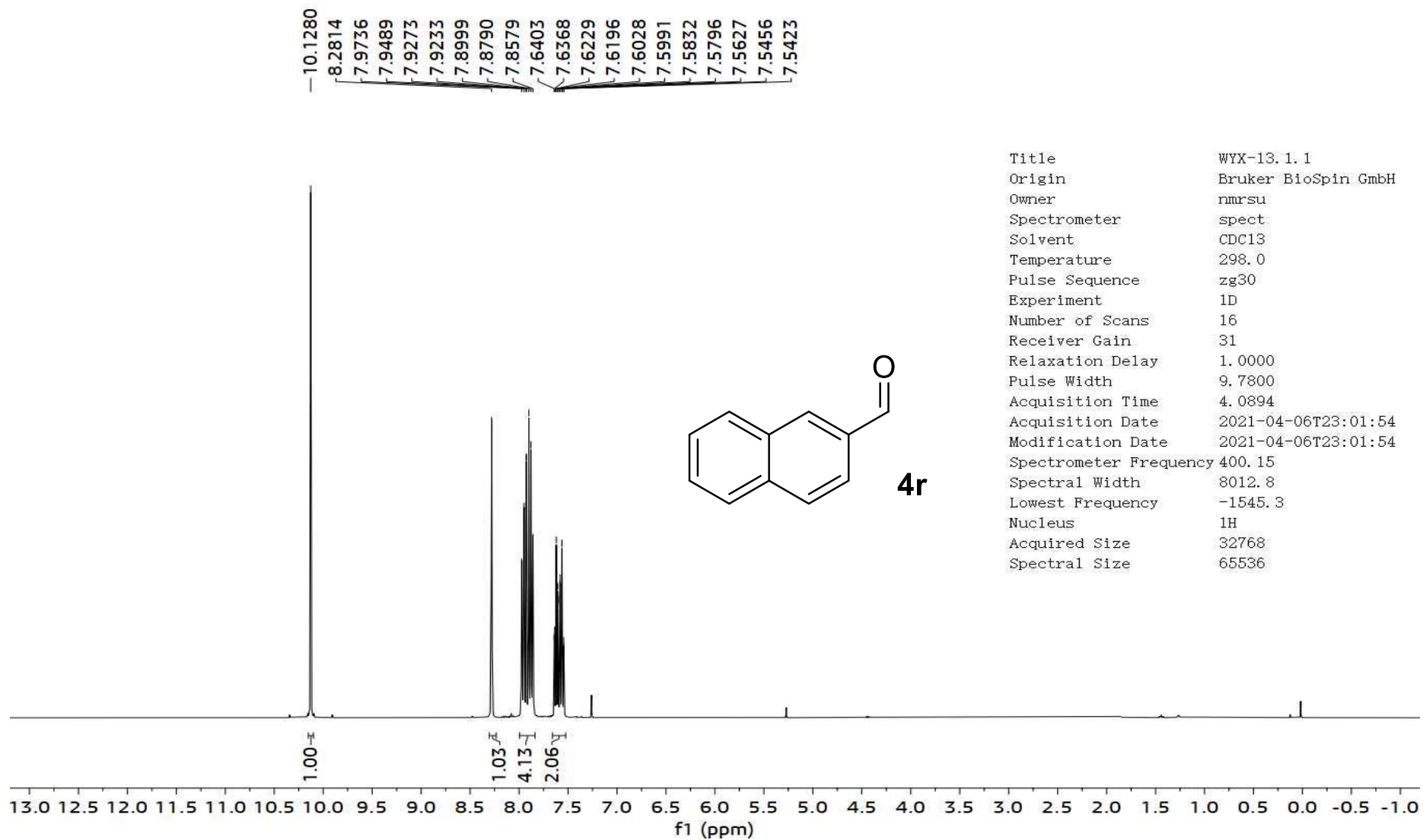
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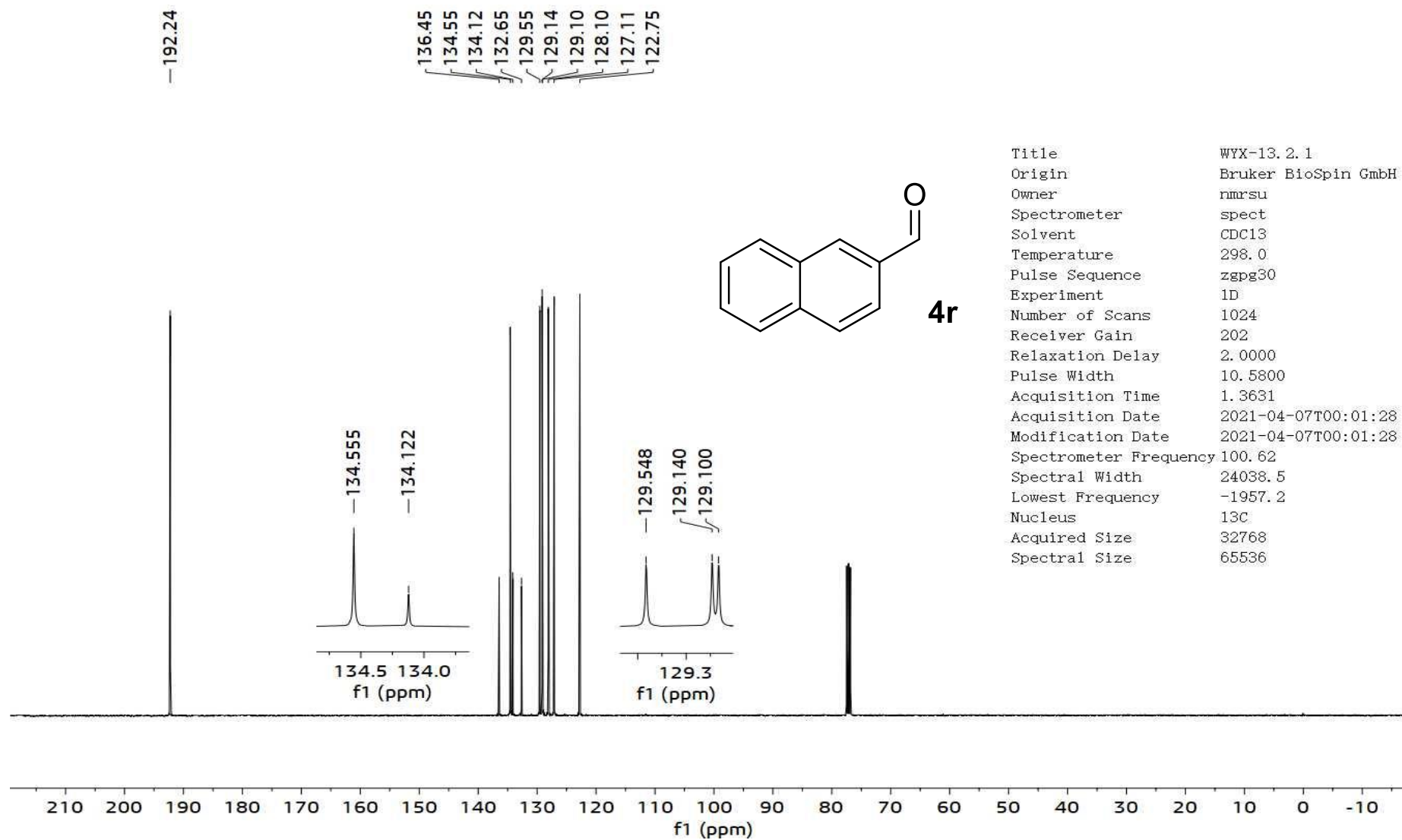
-23.39

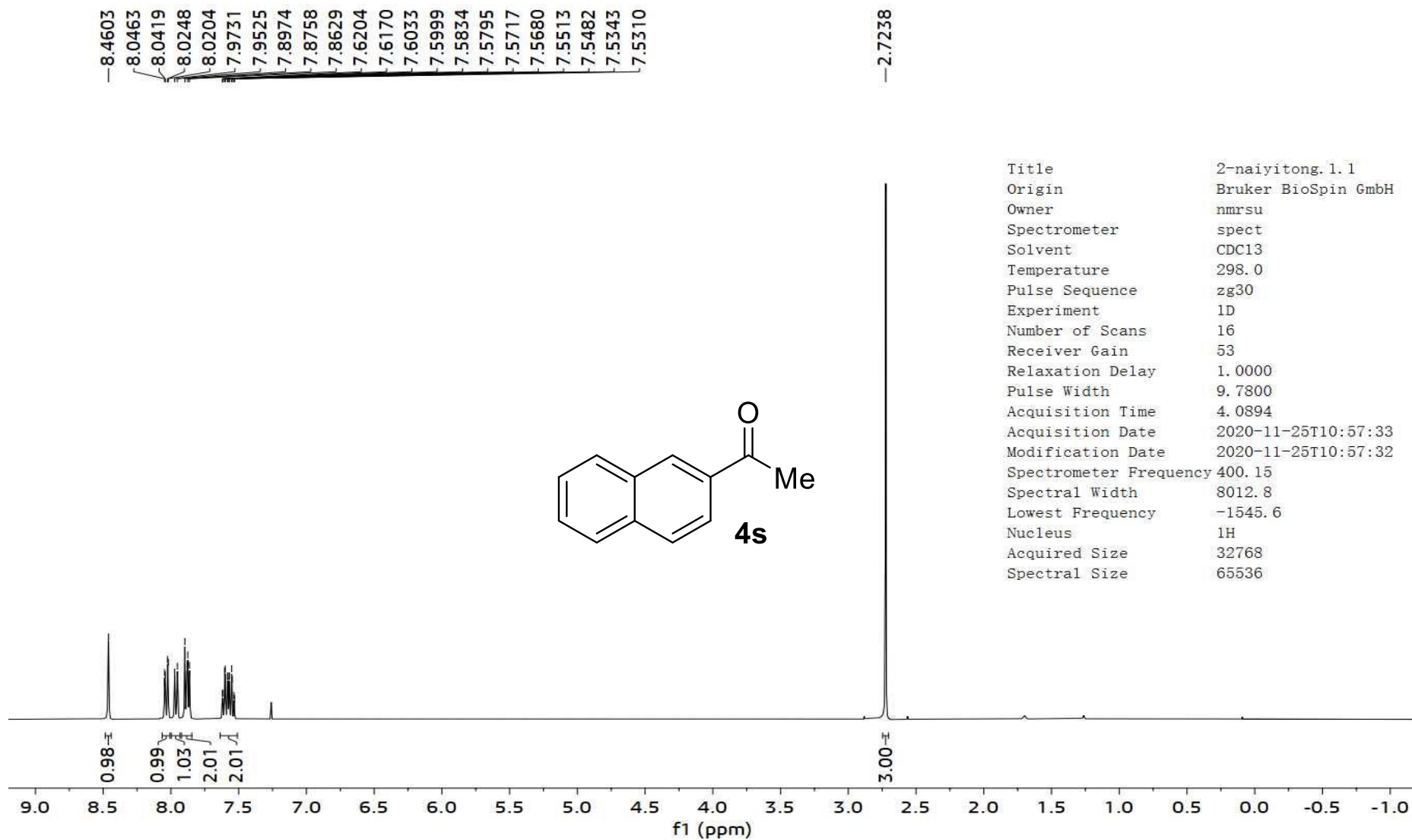


Title	alpha-SQNT. 2.1
Origin	Bruker BioSpin GmbH
Owner	rmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	512
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-11T19:09:57
Modification Date	2021-03-11T19:09:57
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1948.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





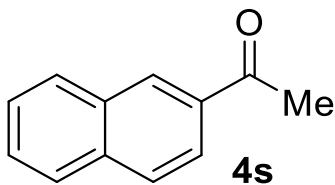




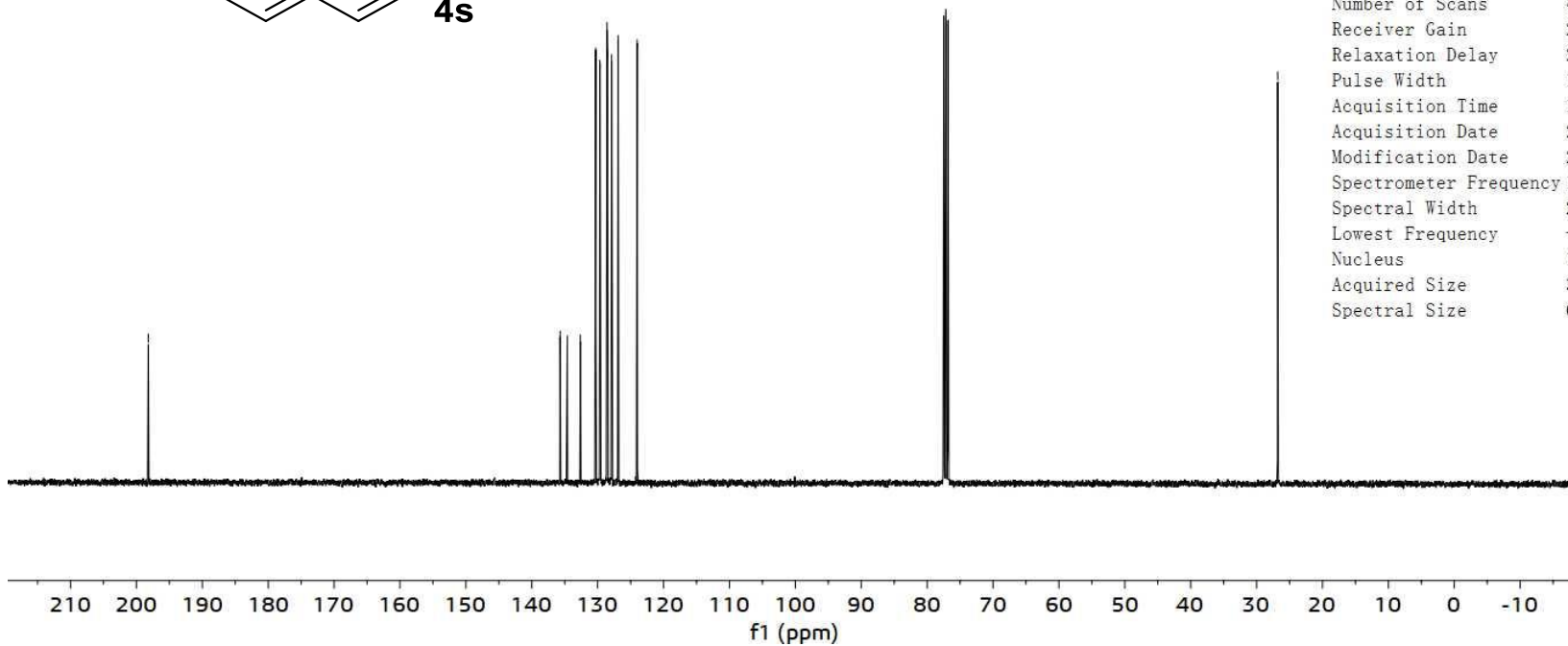
—198.20

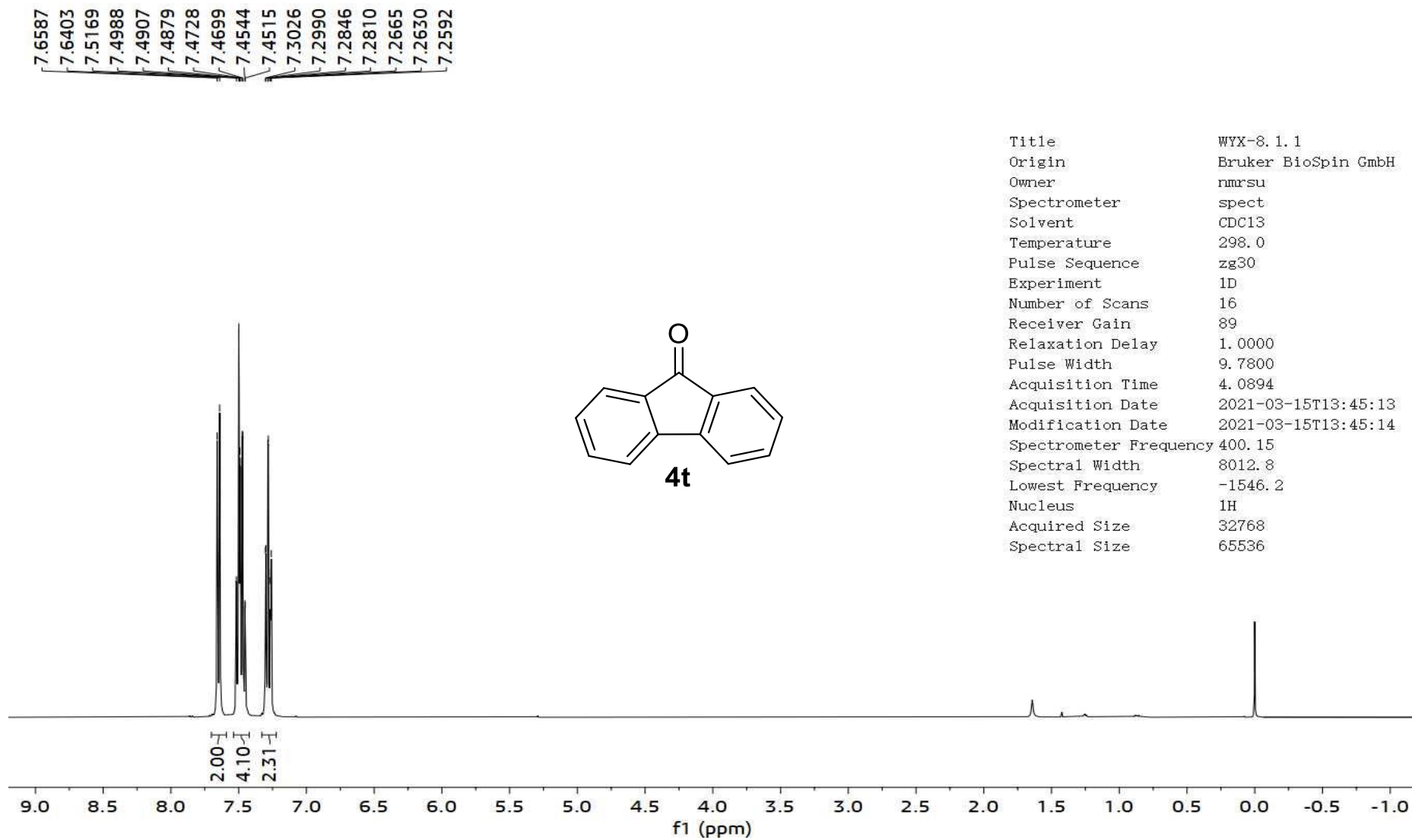
135.70
134.61
132.63
130.30
129.66
128.58
128.53
127.89
126.88
124.01

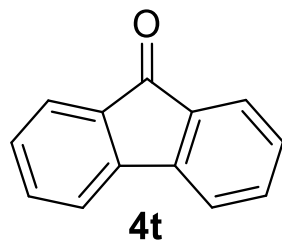
—26.79



Title	2-naiyitong. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	400
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2020-11-25T11:21:34
Modification Date	2020-11-25T11:21:34
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1948.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536







4t

—194.03

—144.55

134.80

134.28

129.20

124.43

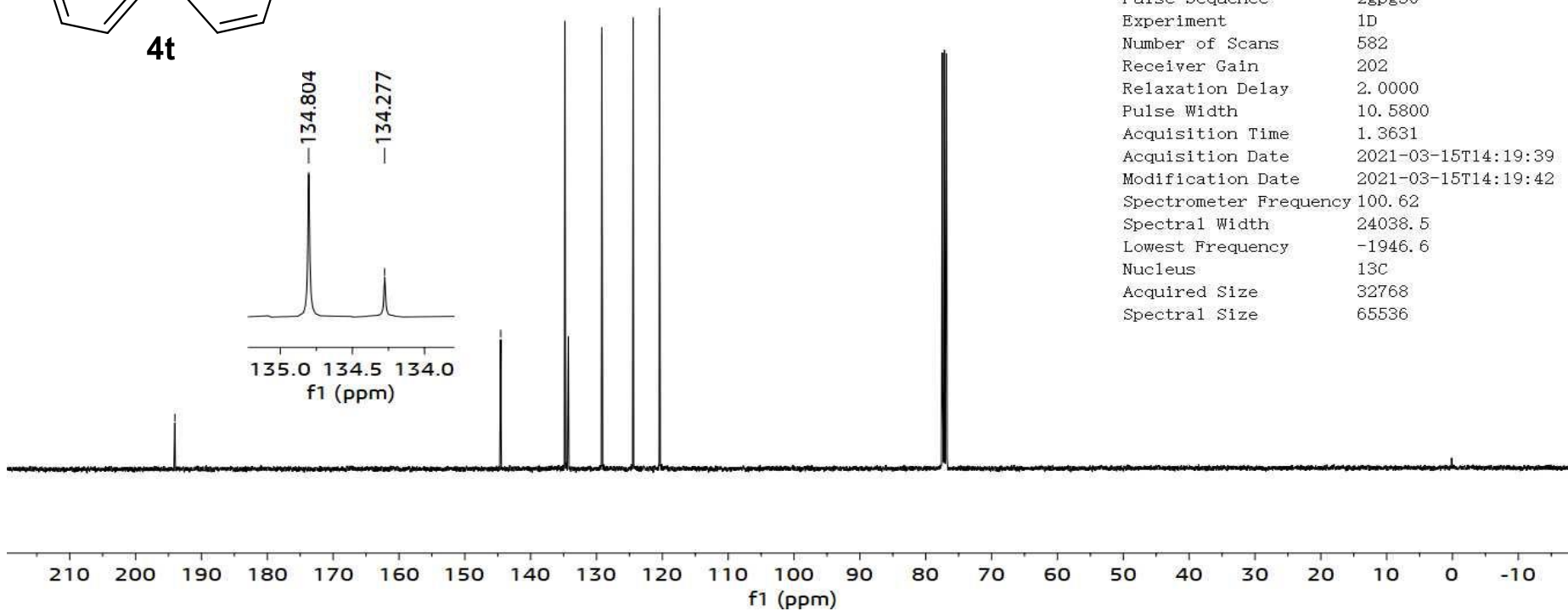
120.43

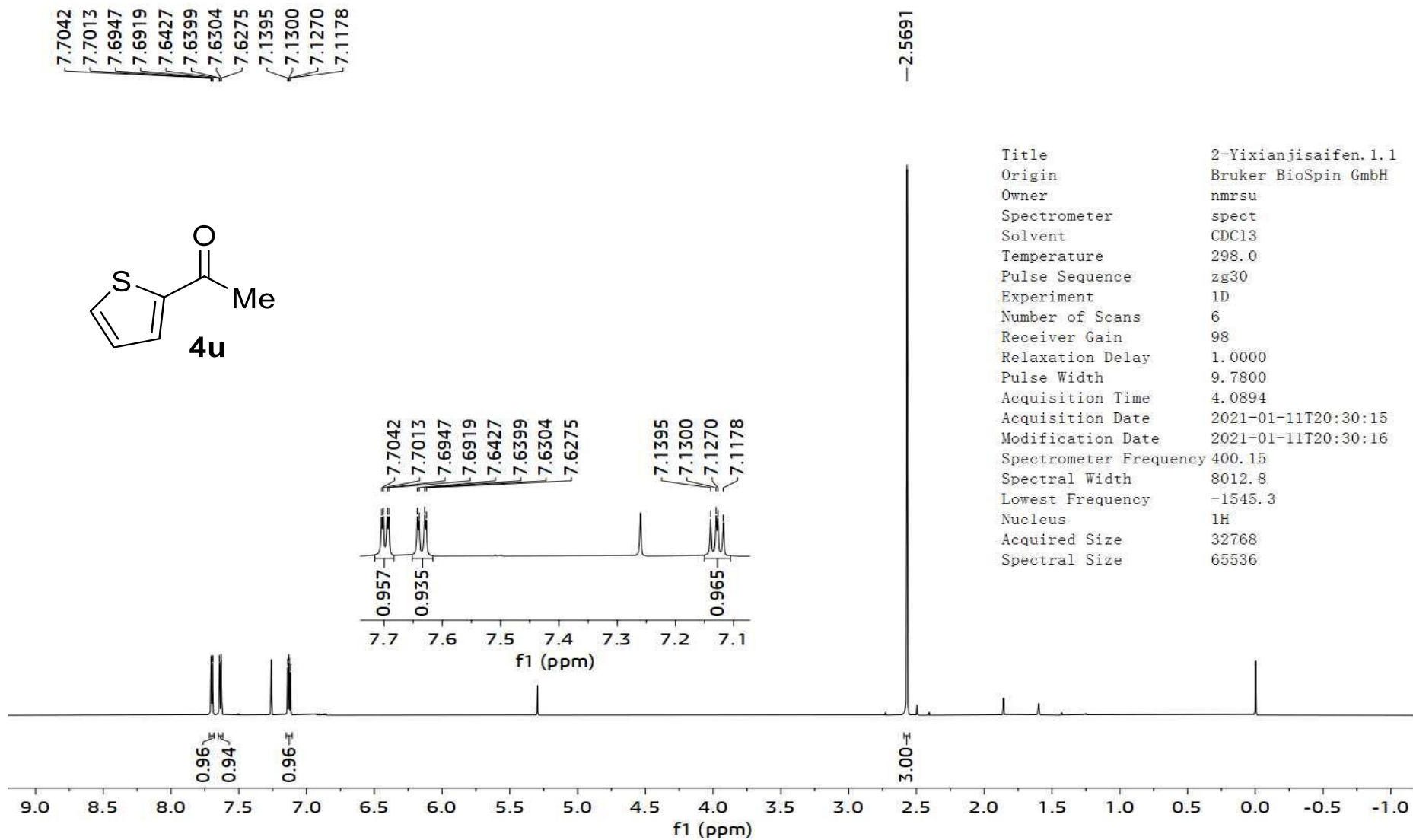
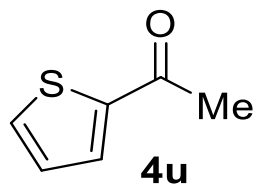
134.804

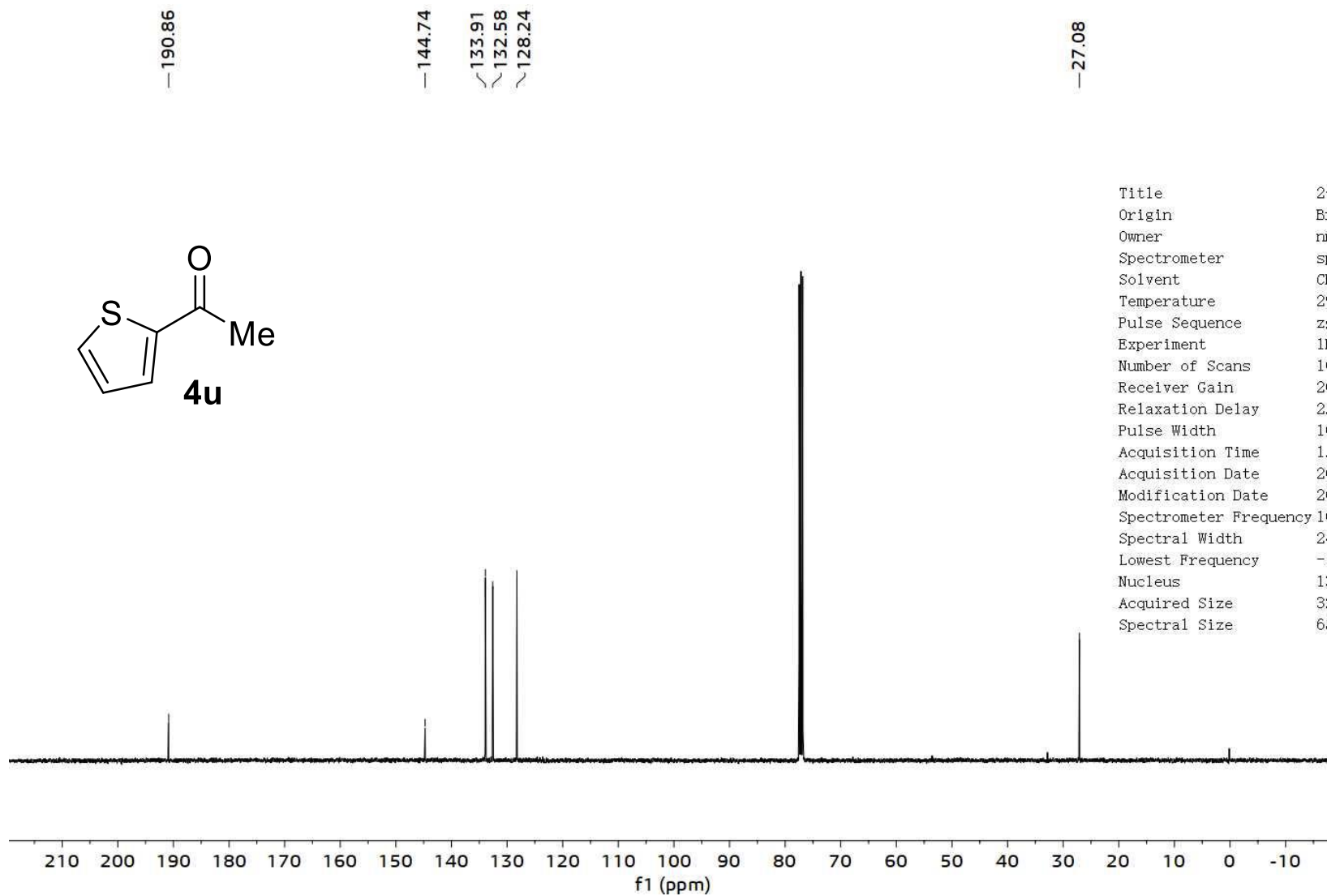
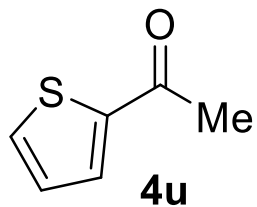
134.277

135.0 134.5 134.0
f1 (ppm)

Title	WYX-8.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zpg30
Experiment	1D
Number of Scans	582
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-15T14:19:39
Modification Date	2021-03-15T14:19:42
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1946.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

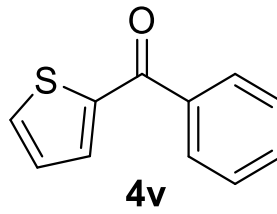




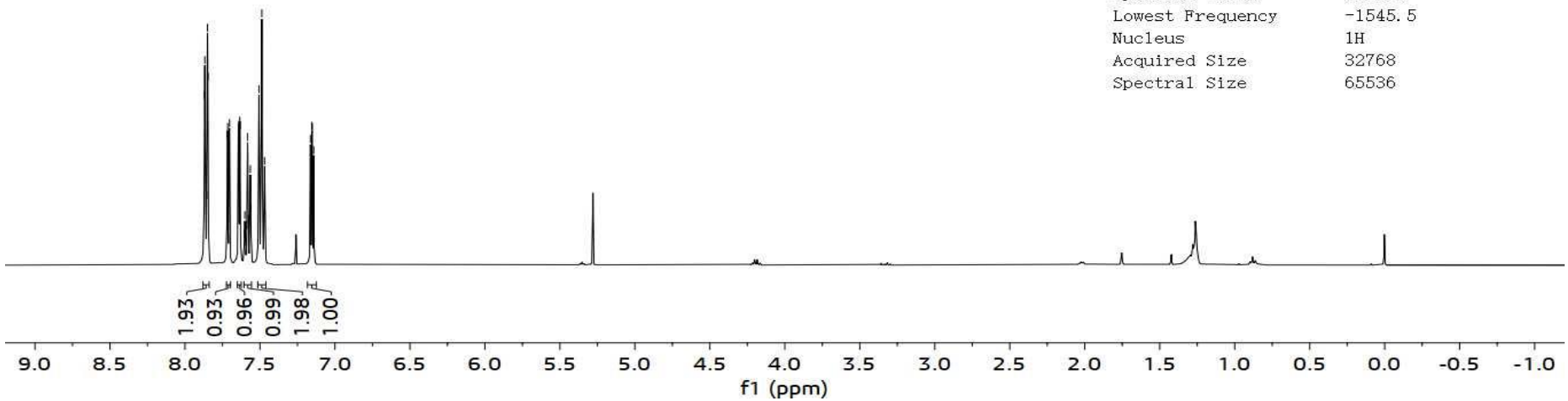


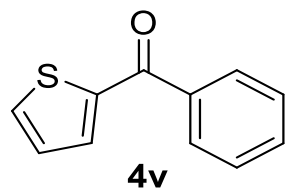
Title	2-Yixianjisaifen. 2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-01-11T21:29:50
Modification Date	2021-01-11T21:29:52
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1944.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

7.8704
7.8675
7.8637
7.8553
7.8500
7.8462
7.7180
7.7151
7.7058
7.7029
7.6441
7.6411
7.6346
7.6317
7.6044
7.6010
7.5977
7.5881
7.5825
7.5773
7.5674
7.5640
7.5606
7.5068
7.4874
7.4694
7.1634
7.1540
7.1510
7.1416



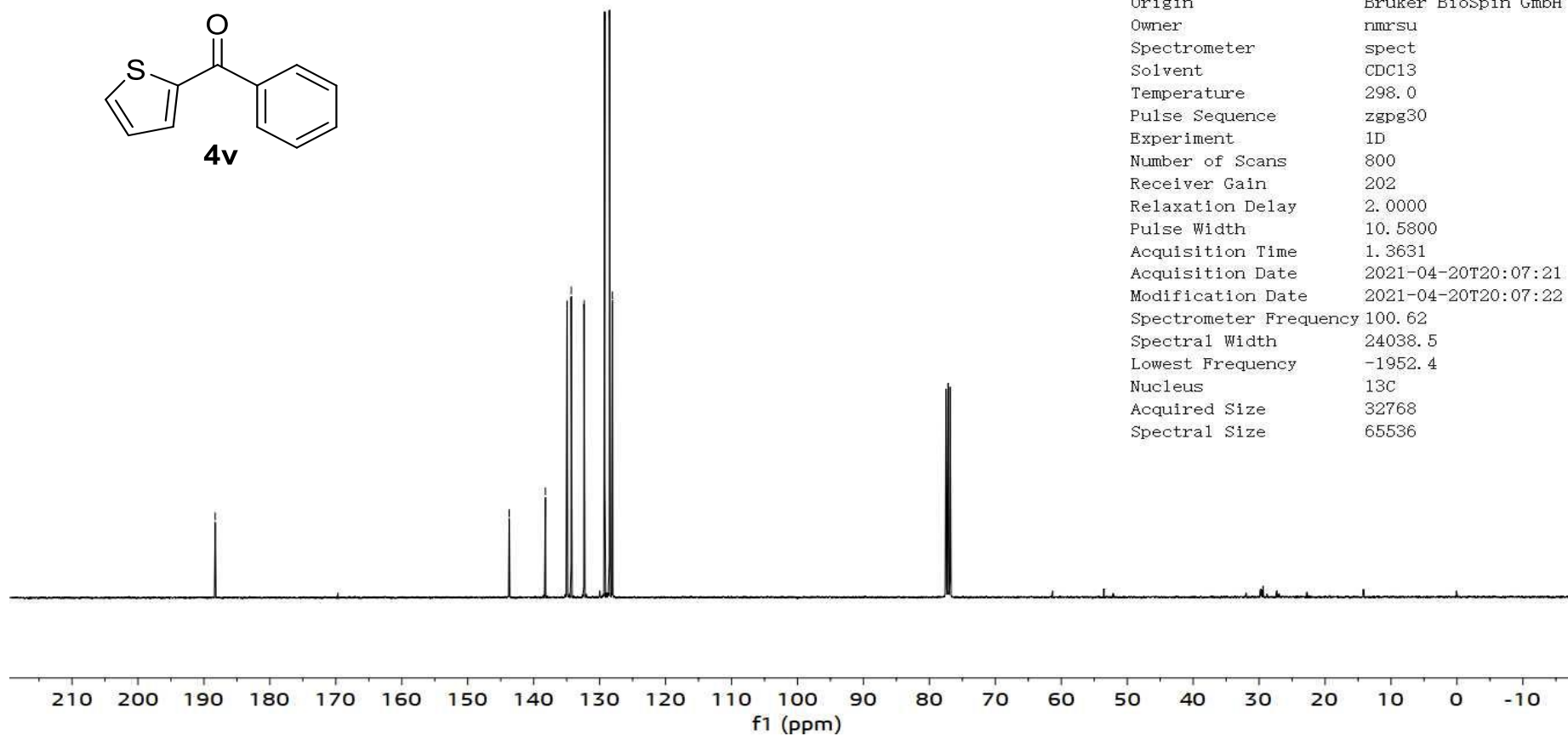
Title	WYX-20.1.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-04-20T19:20:30
Modification Date	2021-04-20T19:20:32
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



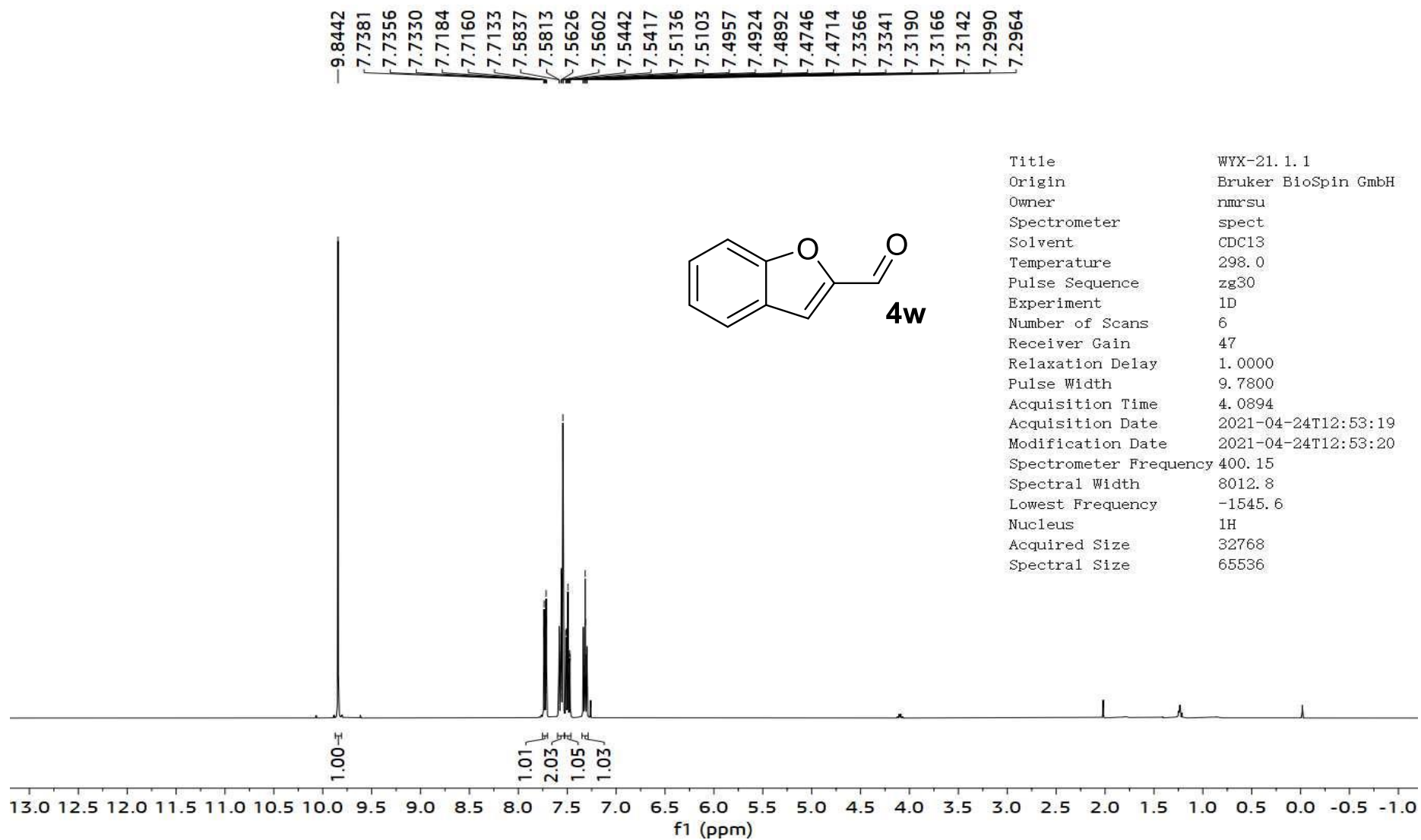


— 188.29

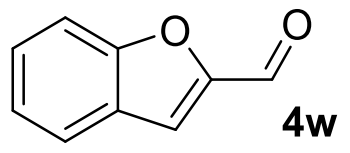
143.70
 138.22
 134.94
 134.30
 132.35
 129.24
 128.50
 128.06



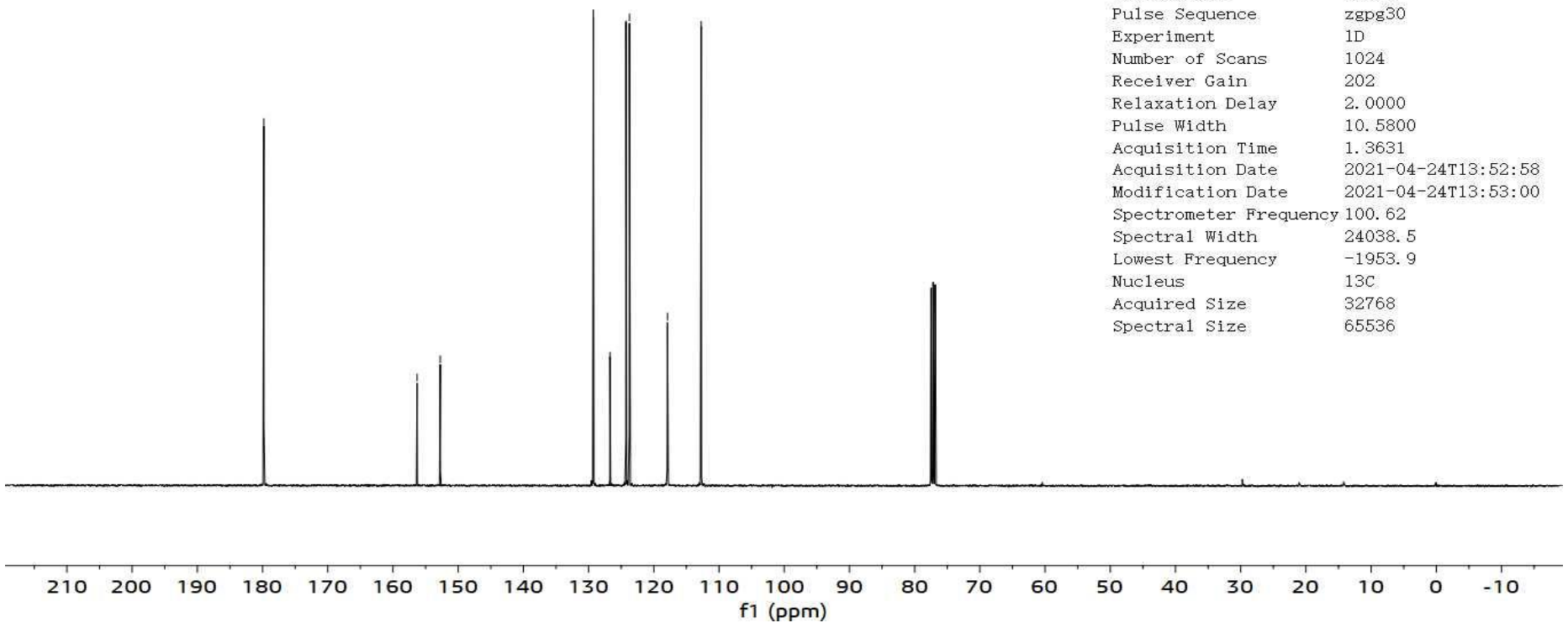
Title	WYX-20. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298. 0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	800
Receiver Gain	202
Relaxation Delay	2. 0000
Pulse Width	10. 5800
Acquisition Time	1. 3631
Acquisition Date	2021-04-20T20:07:21
Modification Date	2021-04-20T20:07:22
Spectrometer Frequency	100. 62
Spectral Width	24038. 5
Lowest Frequency	-1952. 4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



179.80
 156.29
 152.75
 129.27
 126.71
 124.26
 123.73
 117.90
 112.73



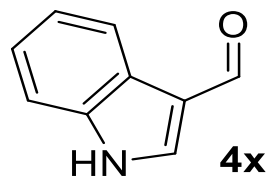
Title	WYX-21. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-04-24T13:52:58
Modification Date	2021-04-24T13:53:00
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1953.9
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



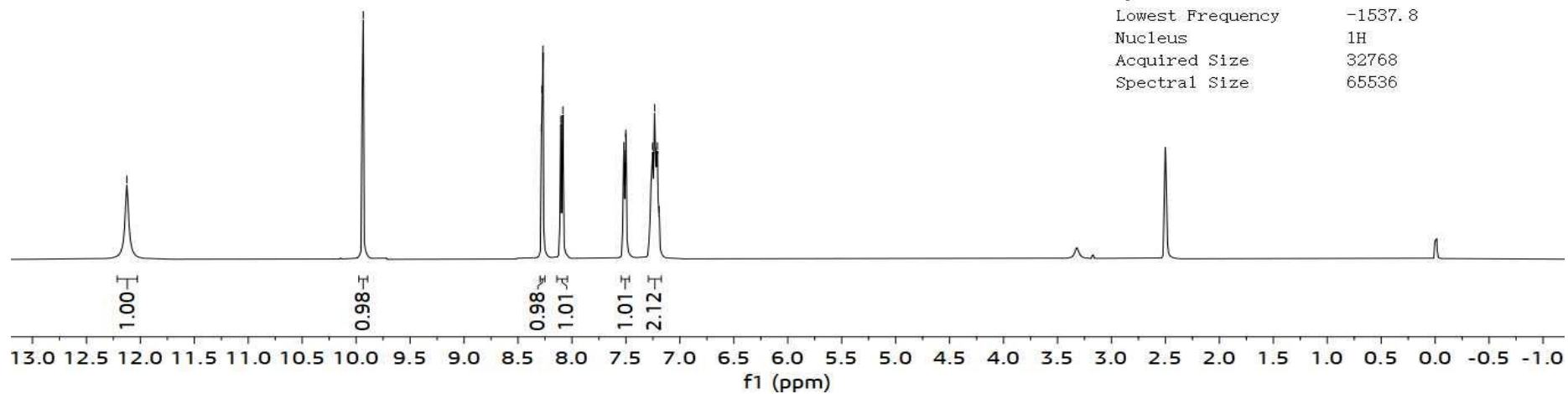
-12.1252

-9.9346

8.2832
8.2774
8.2686
8.1041
8.0840
7.5189
7.5017
7.2528
7.2338
7.2226
7.2078
7.1908

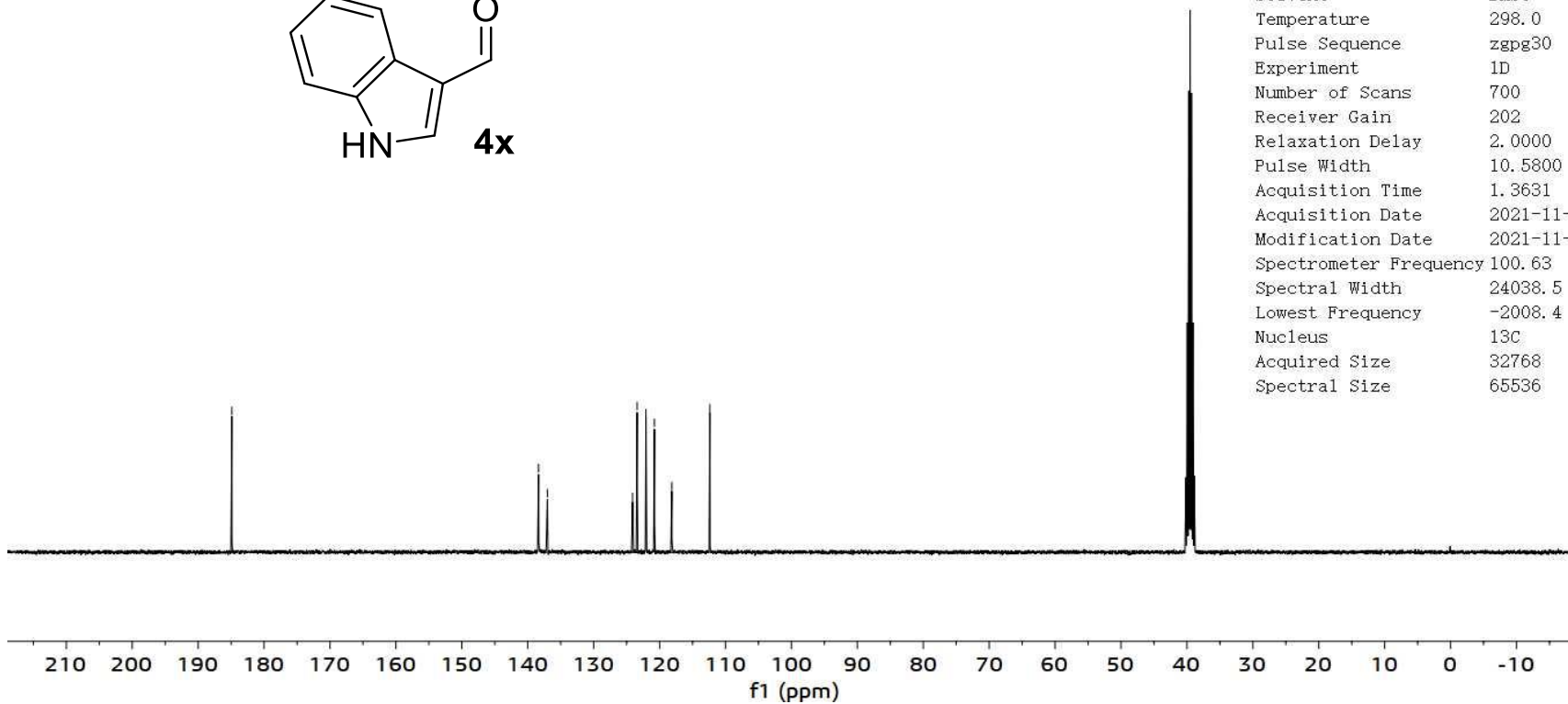
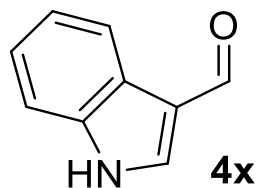


Title	WYX-25YD.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	10
Receiver Gain	112
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-11-10T17:18:37
Modification Date	2021-11-10T17:18:36
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1537.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



— 184.90

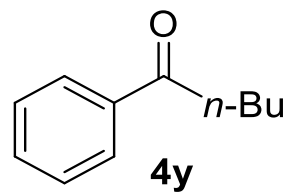
138.37
137.02
124.10
123.41
122.07
120.78
118.14
112.38



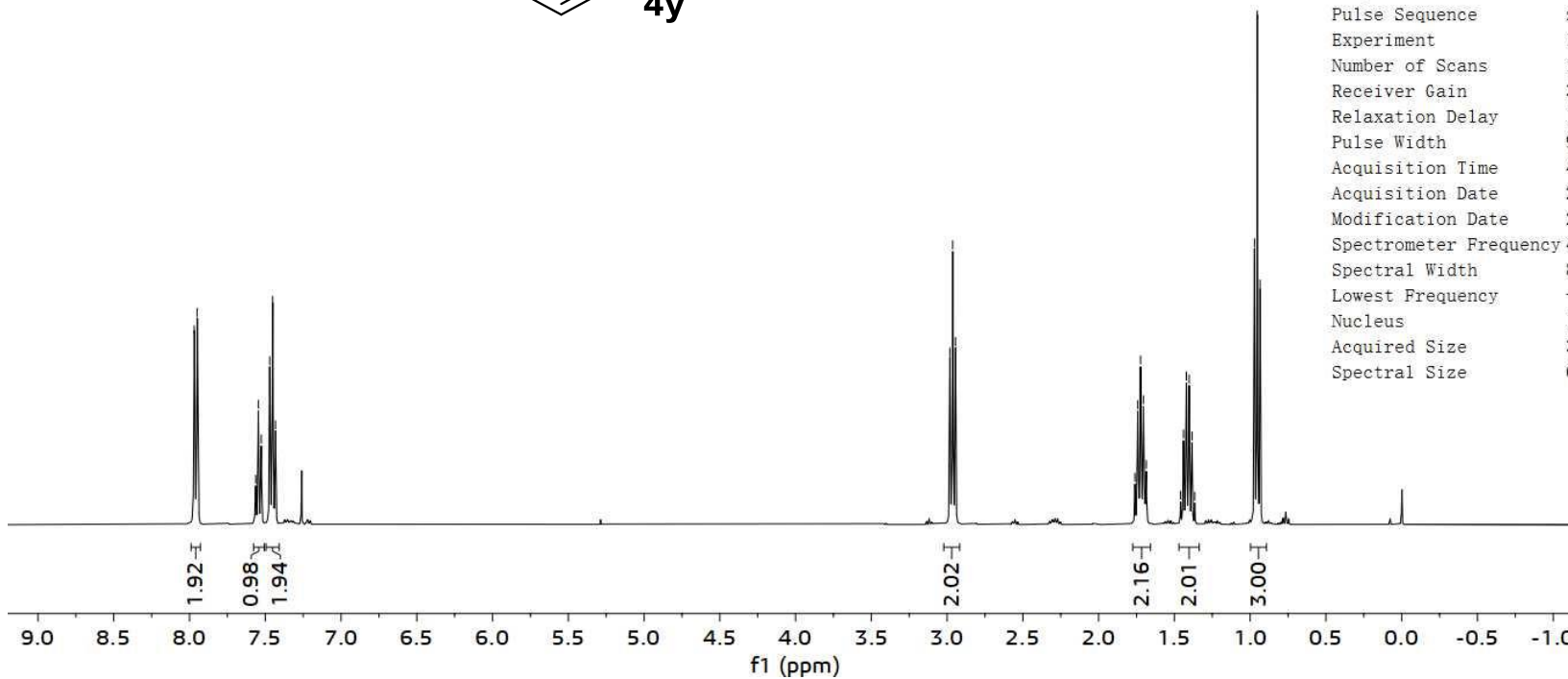
Title	WYX-25YD. 2. fid
Origin	Bruker BioSpin GmbH
Owner	rmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-11-11T02:02:49
Modification Date	2021-11-11T02:02:48
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-2008.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

7.9689
7.9493
7.5628
7.5455
7.5261
7.4706
7.4513
7.4324

2.9818
2.9633
2.9447
1.7604
1.7421
1.7234
1.7045
1.6855
1.4586
1.4400
1.4213
1.4025
1.3840
1.3657
0.9715
0.9531
0.9348



Title	Benwutong.1.1
Origin	Bruker BioSpin GmbH
Owner	nrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-03-05T15:55:20
Modification Date	2021-03-05T15:55:22
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

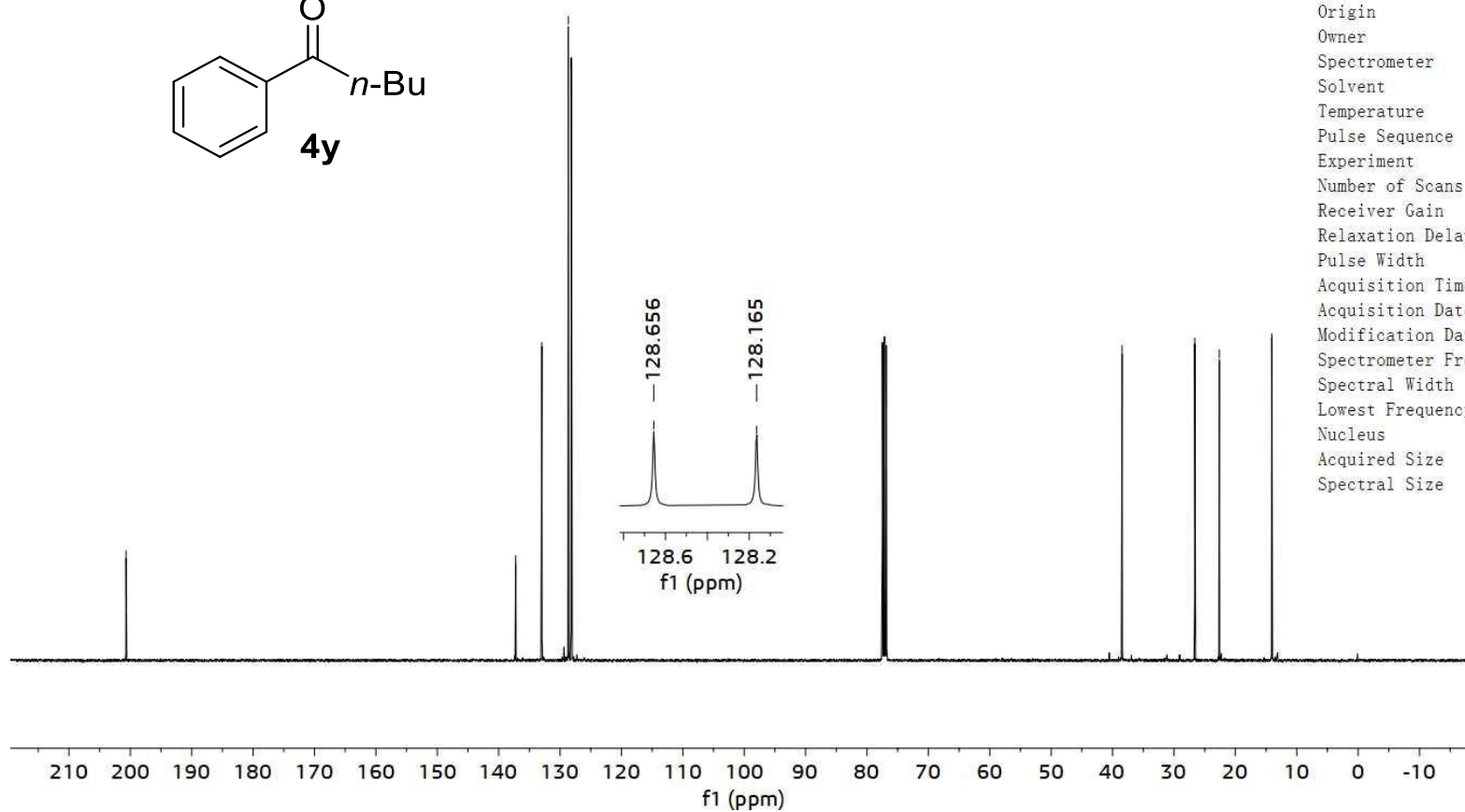
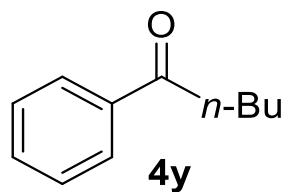


—200.69

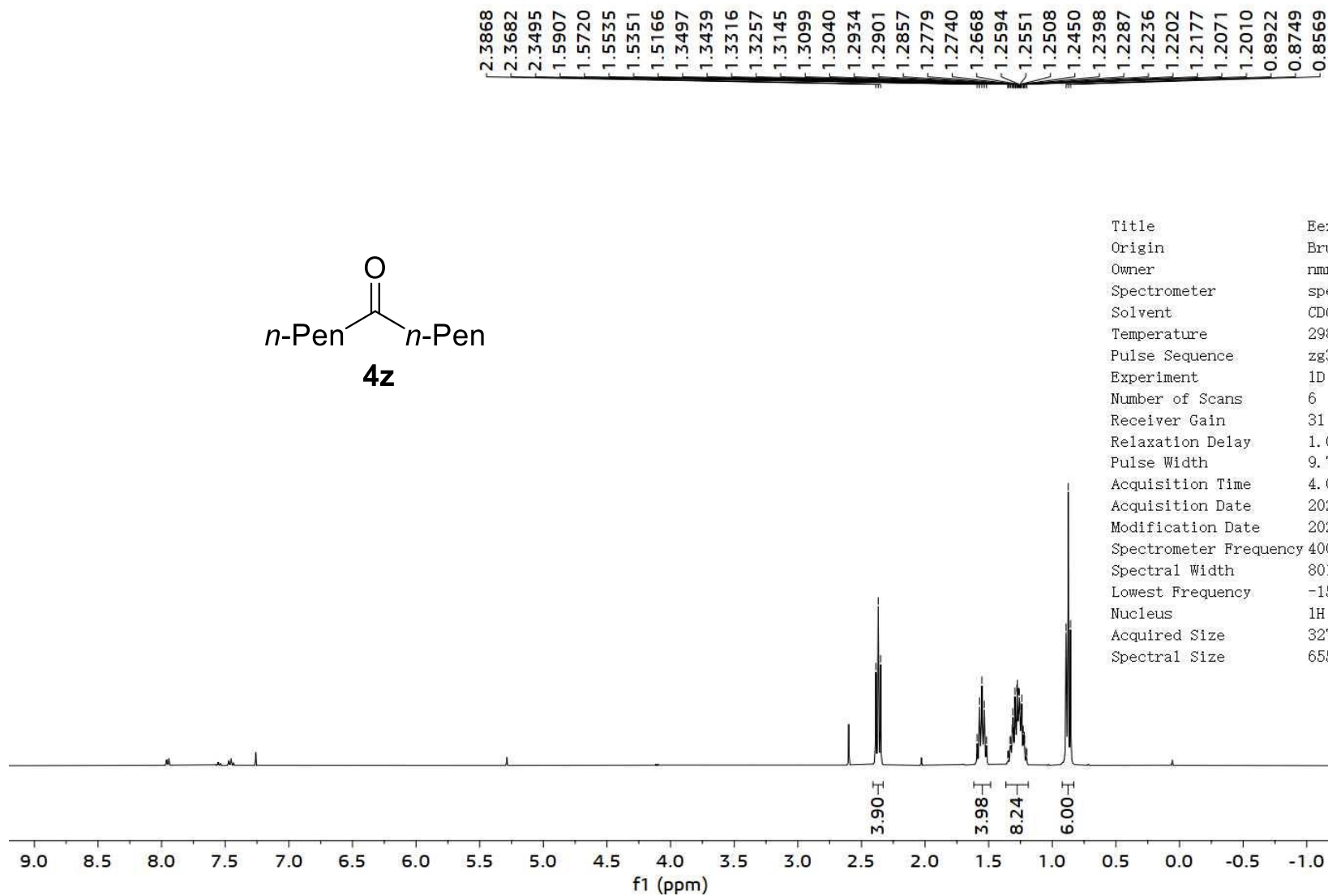
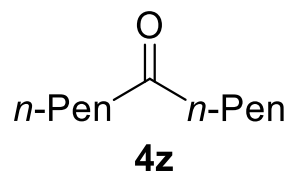
✓137.22
✓132.96
✓128.66
✓128.16

—38.44

✓26.60
✓22.60
✓14.06

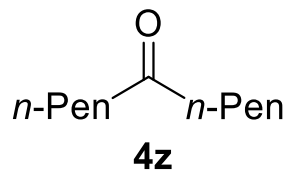


Title	Benwutong. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-05T18:33:43
Modification Date	2021-03-05T18:33:44
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1947.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Title	Bezhengwujitong.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2020-12-14T19:47:26
Modification Date	2020-12-14T19:47:28
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

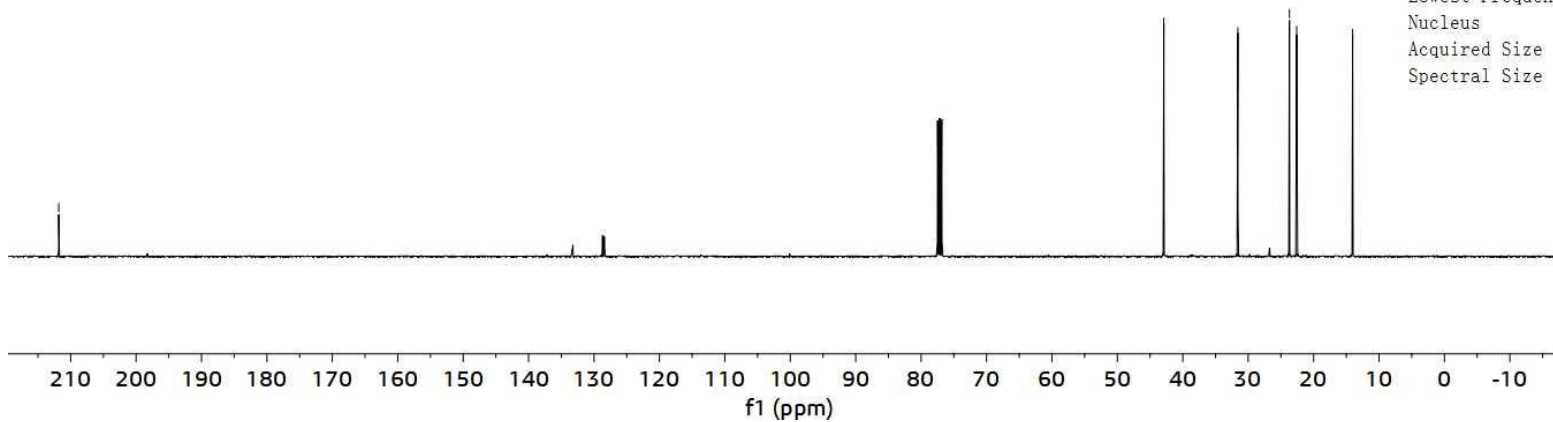
— 211.82

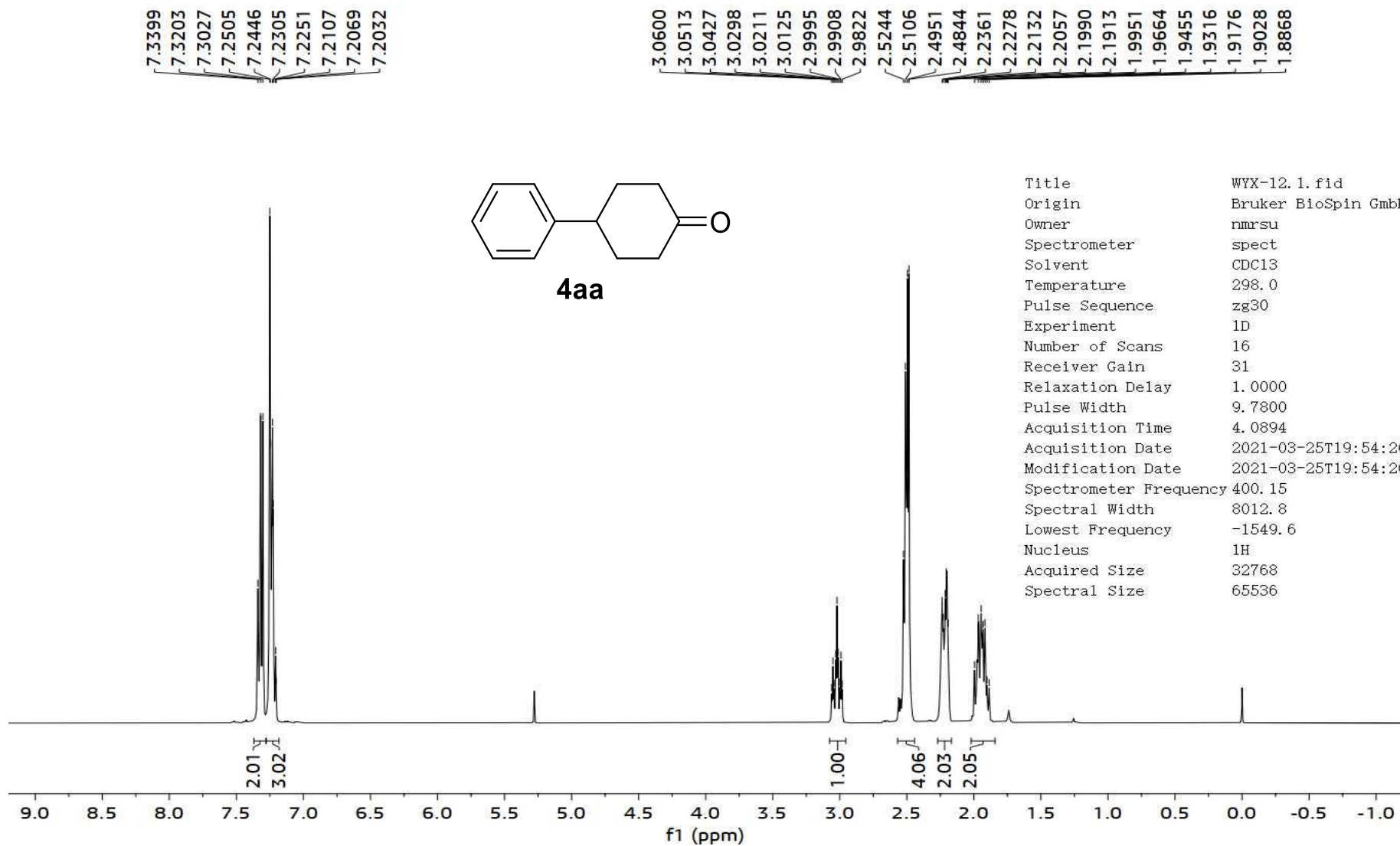


— 42.90

31.58
23.70
22.59
14.04

Title	Ezhenhwujitong.2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2020-12-14T20:17:09
Modification Date	2020-12-14T20:17:12
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1944.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



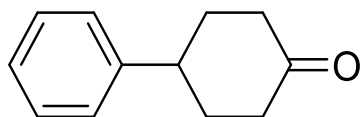


-211.19

-144.88

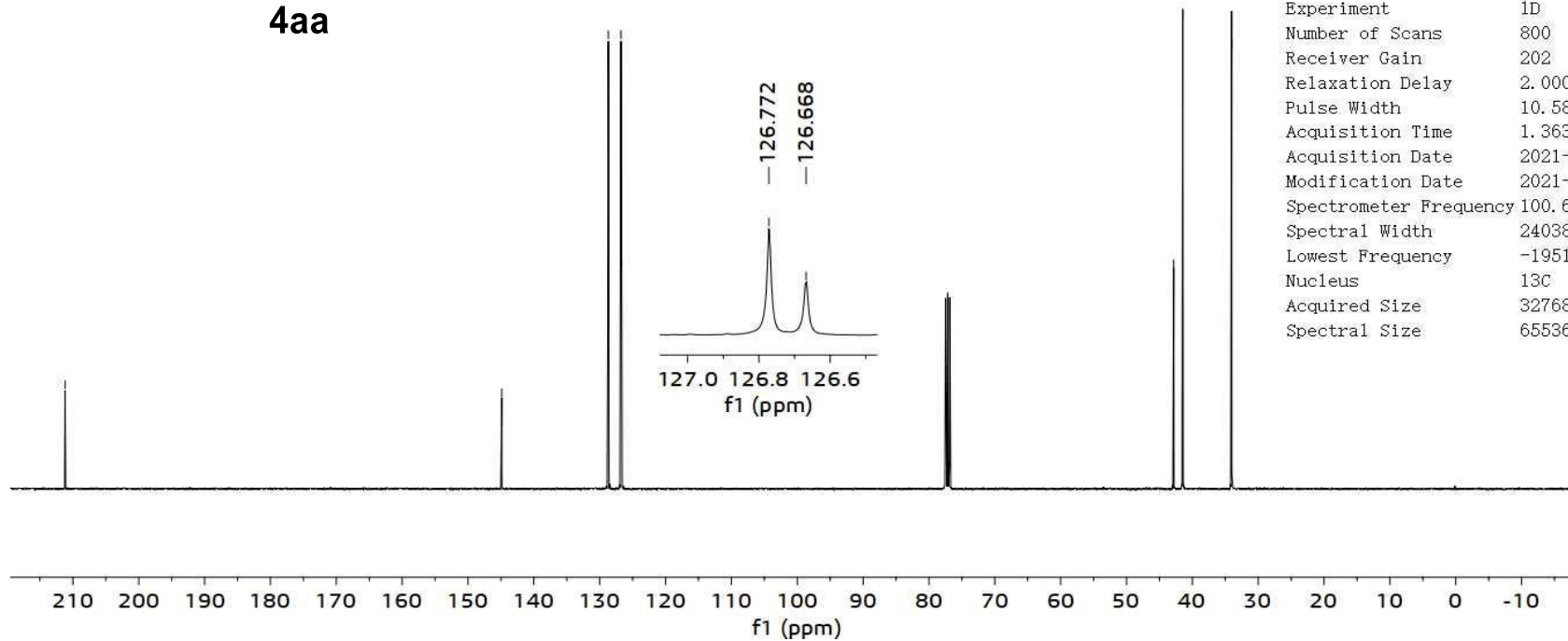
128.69
126.77
126.67

42.84
41.45
34.05

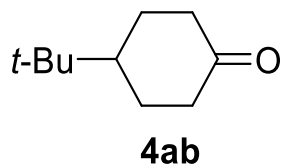


4aa

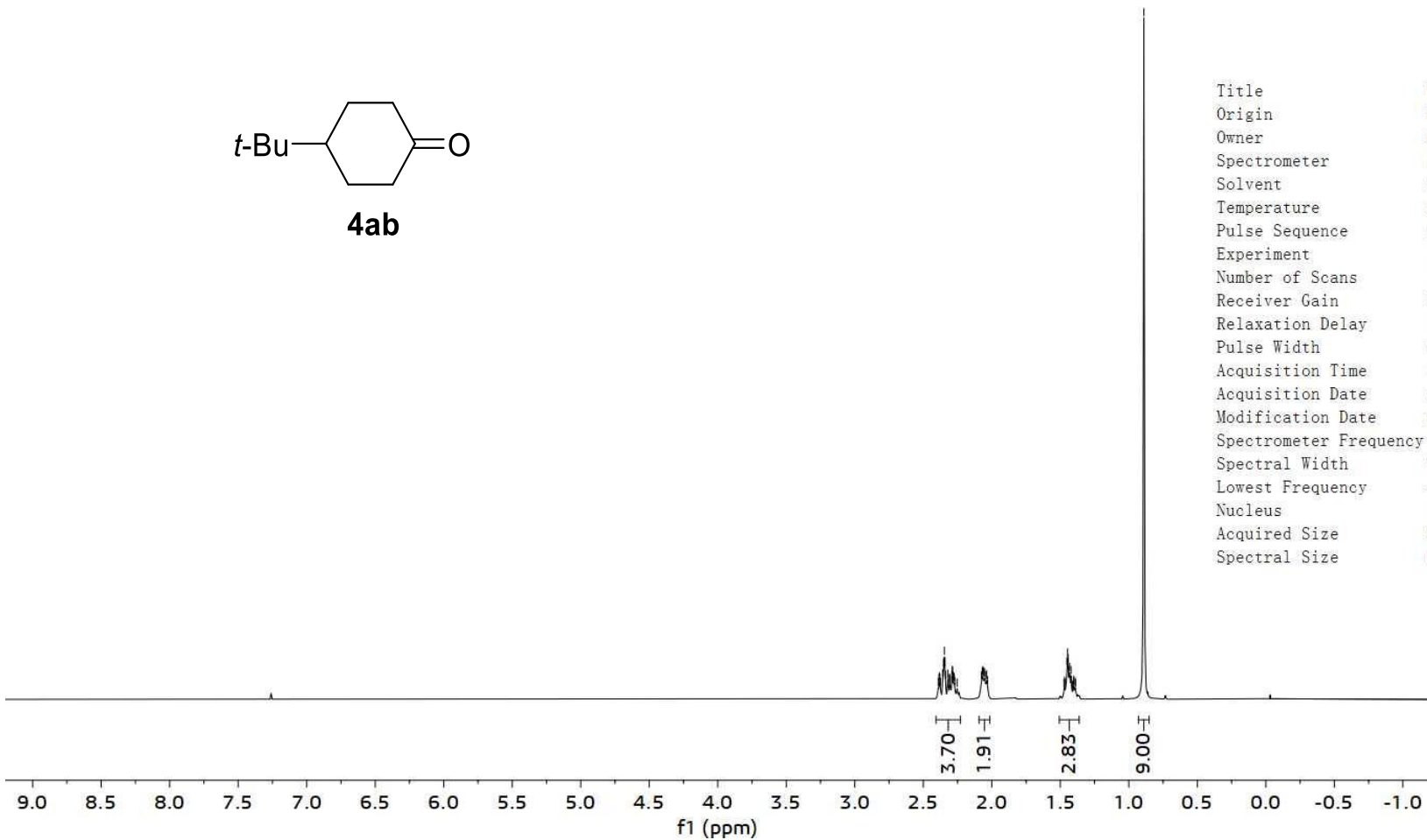
Title	WYX-12. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	800
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-25T20:41:07
Modification Date	2021-03-25T20:41:06
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1951.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



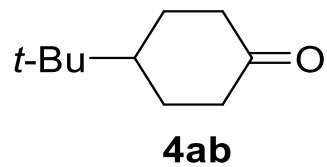
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2.3826
2.3790
2.3745
2.3584
2.3530
2.3464
2.3418
2.3378
2.3212
2.3178
2.3086
2.3061
2.3036
2.2920
2.2888
2.2864
2.2830
2.2779
2.2737
2.2703
2.2532
2.0760
2.0699
2.0663
2.0616
2.0574
2.0510
2.0426
2.0366
2.0316
1.4735
1.4693
1.4621
1.4520
1.4467
1.4437
1.4387
1.4316
1.4220
1.4102
1.4032
1.4011
1.3923
1.3883
0.8902



Title	WYX-11.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsm
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-03-25T20:50:29
Modification Date	2021-03-25T20:50:28
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

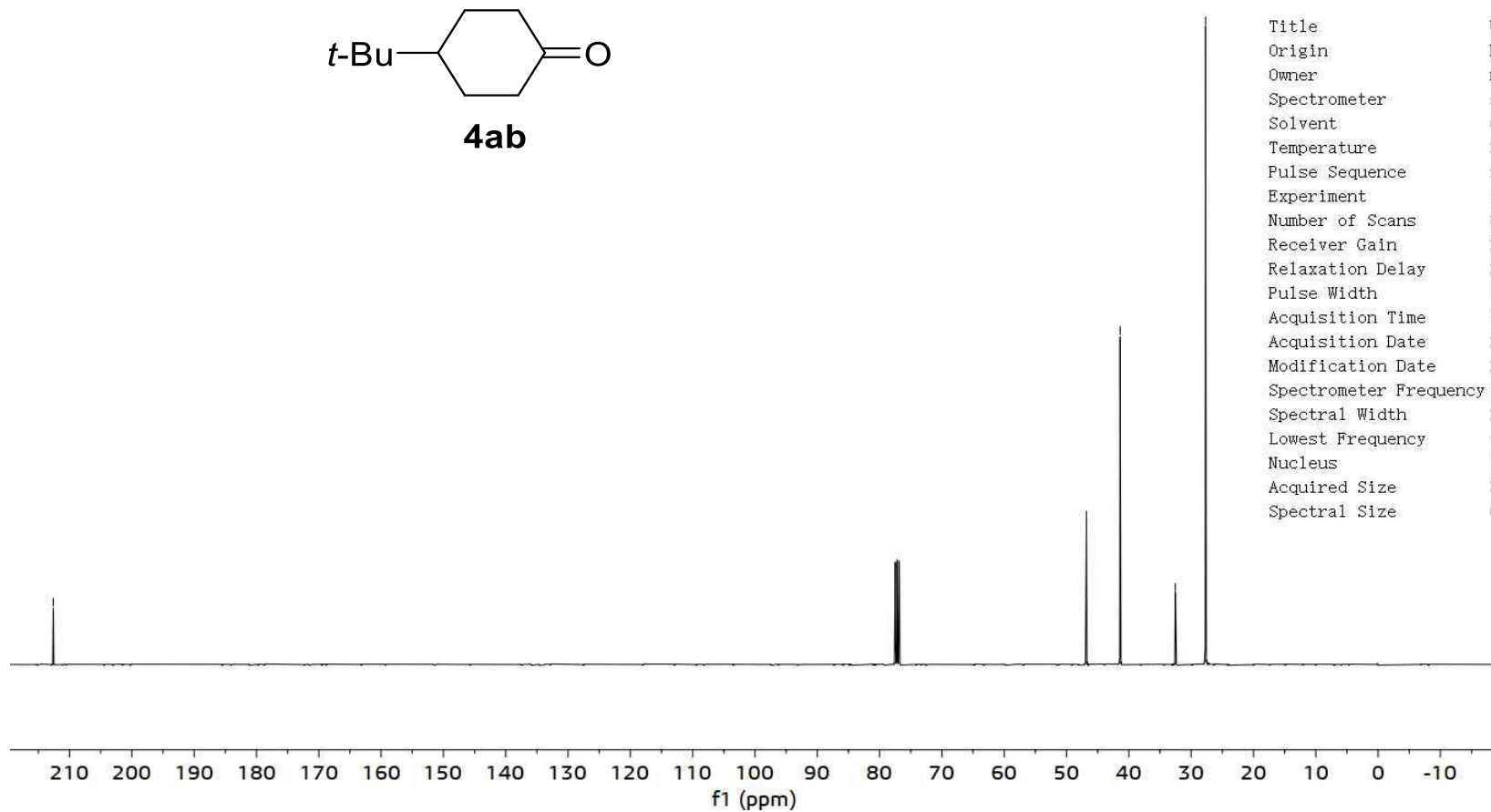


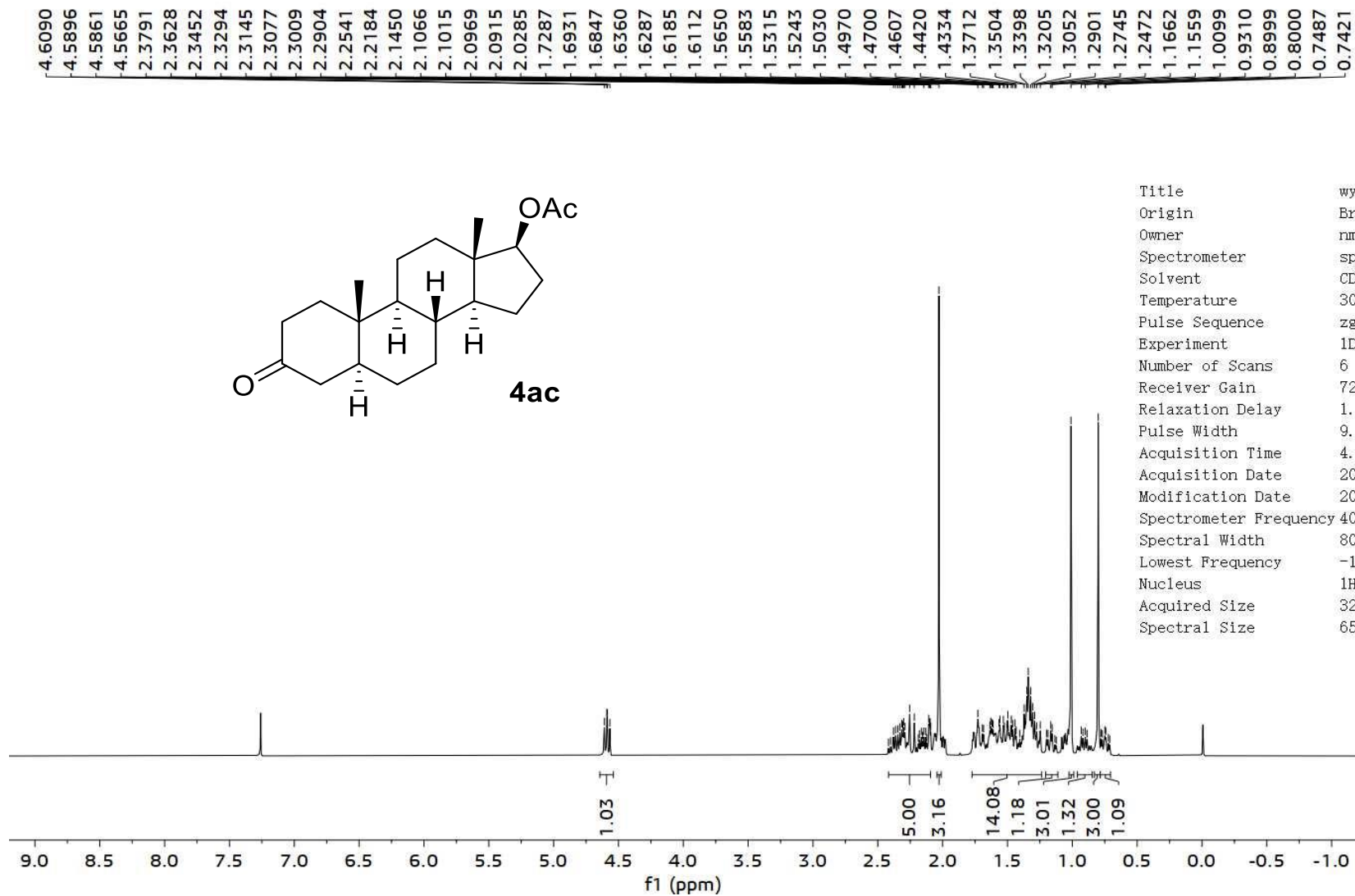
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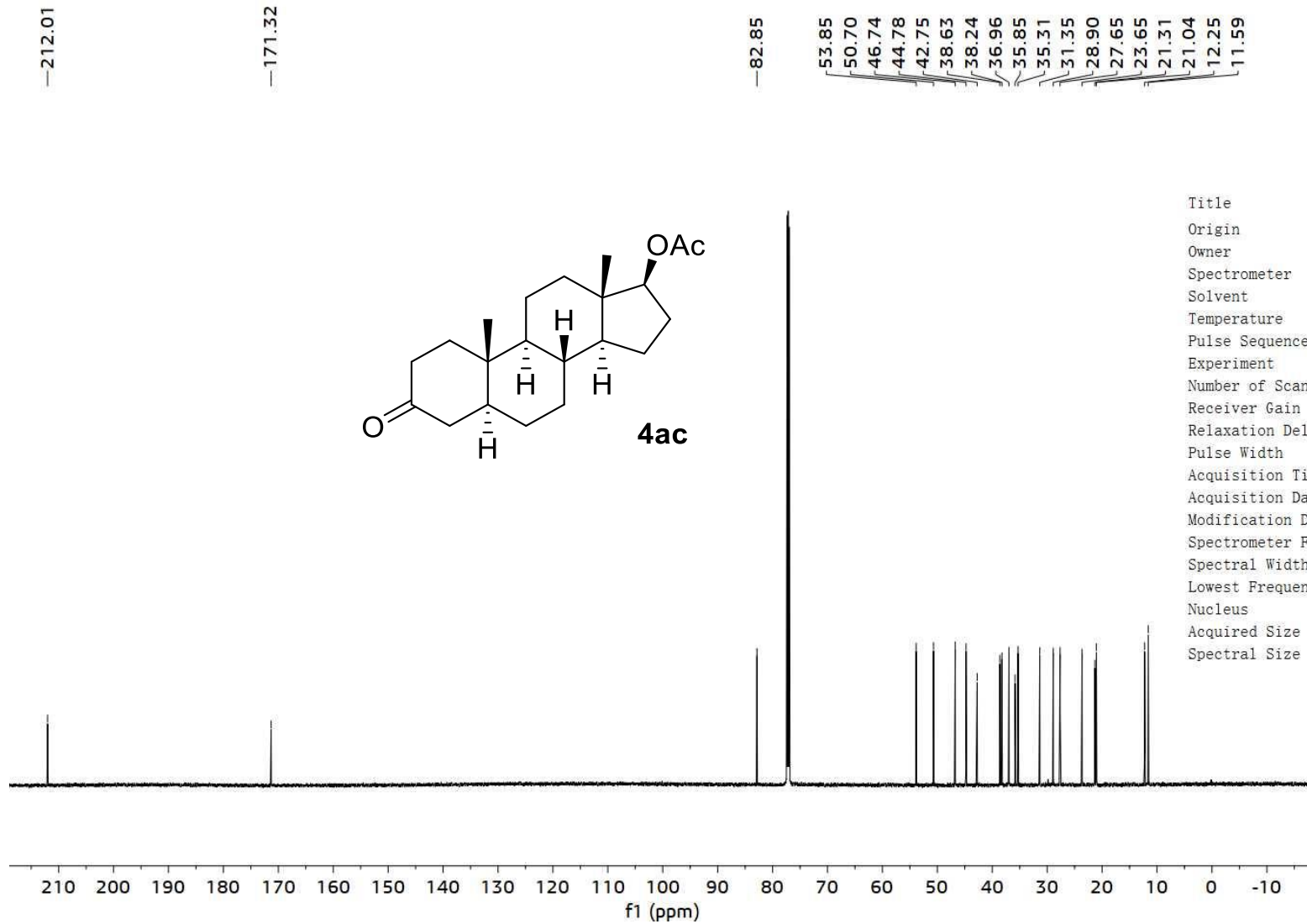


—46.80
—41.40
—32.55
—27.69

Title	WYX-11.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	800
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-03-25T21:37:16
Modification Date	2021-03-25T21:37:16
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1947.8
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536

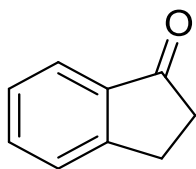






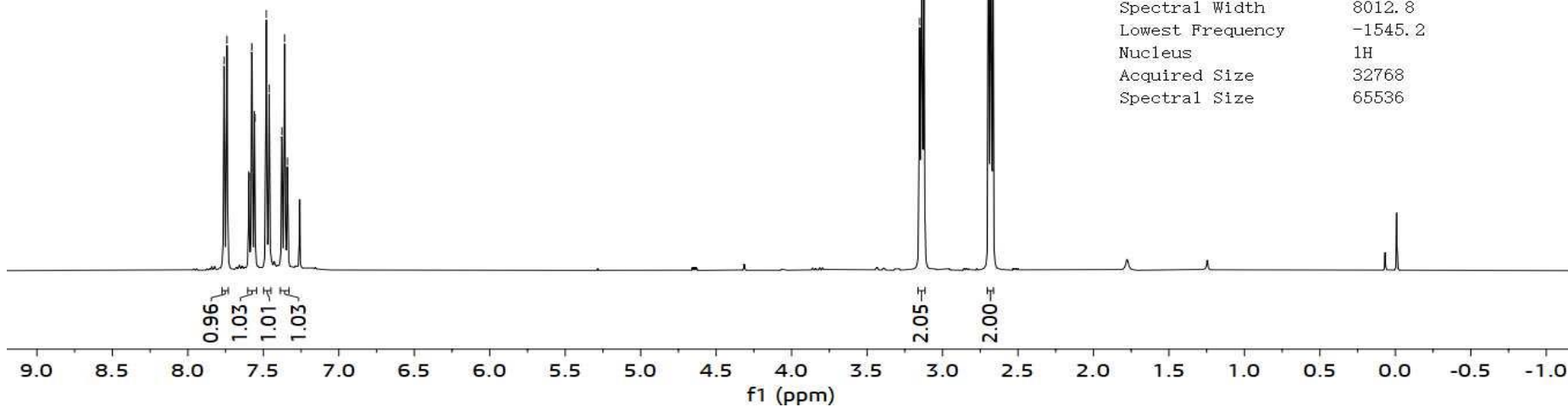
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7.3776
7.3590
7.3405

3.1521
3.1376
3.1226
2.6960
2.6859
2.6810
2.6767
2.6663



4ad

Title	WYX-71-charwu. 1. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	296.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	72
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-27T15:55:16
Modification Date	2021-05-27T15:55:18
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



—207.17

—155.27

✓137.20

—134.70

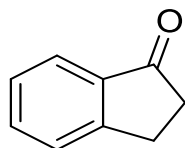
✓127.39

✓126.81

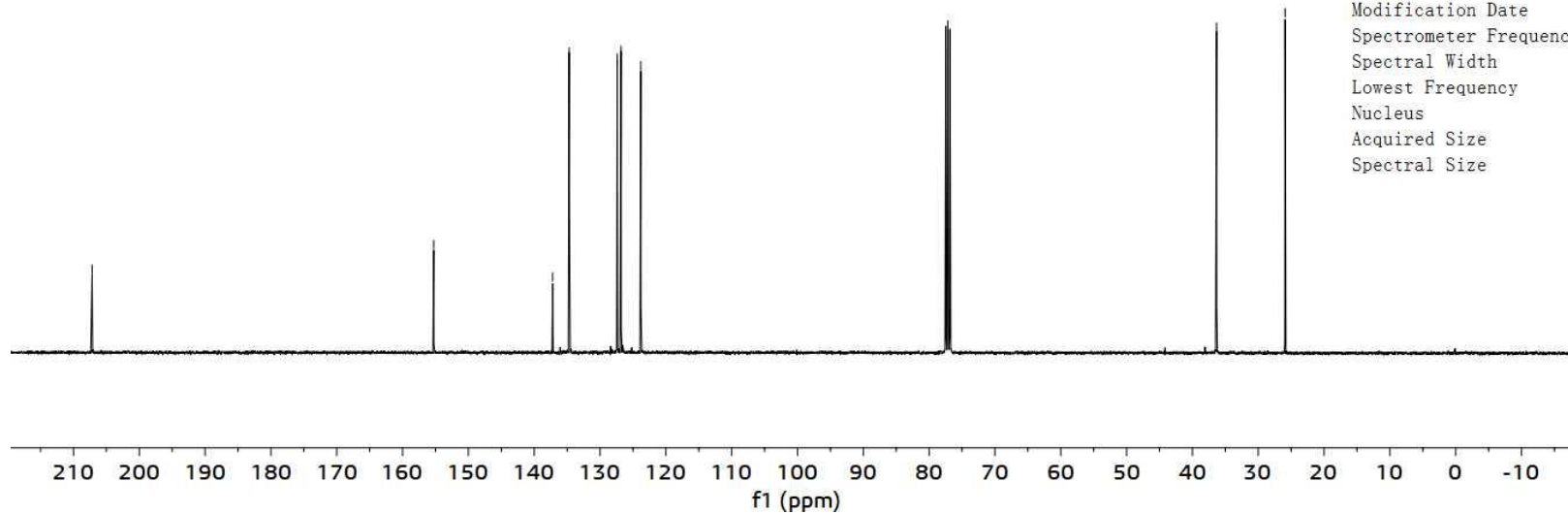
✓123.82

—36.33

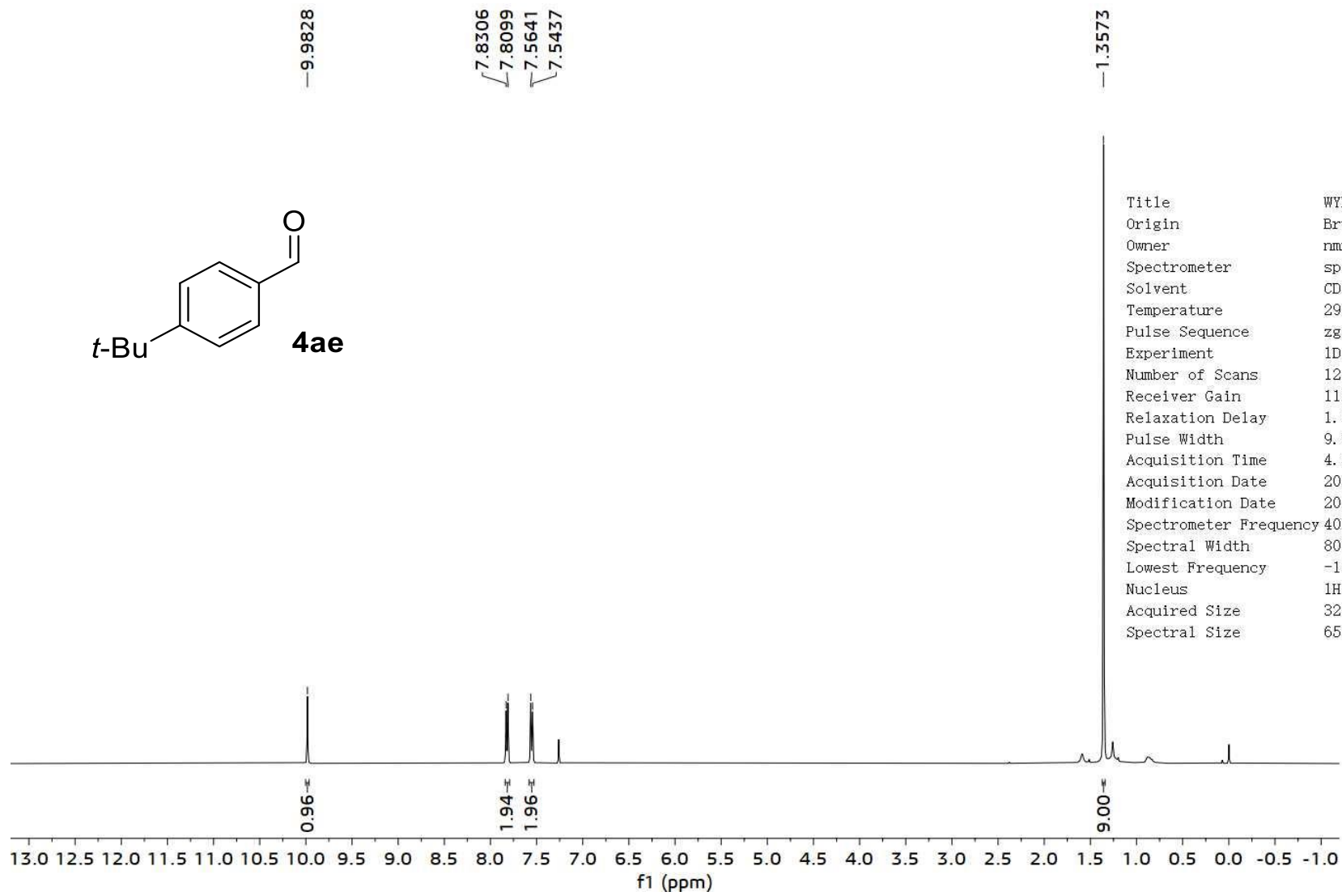
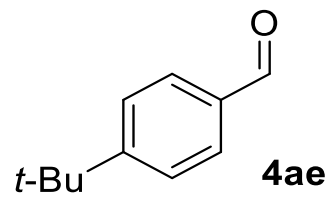
—25.91



4ad



Title	WYX-71-chanwu. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	296. 9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2. 0000
Pulse Width	10. 5800
Acquisition Time	1. 3631
Acquisition Date	2021-05-28T02:54:29
Modification Date	2021-05-28T02:54:30
Spectrometer Frequency	100. 62
Spectral Width	24038. 5
Lowest Frequency	-1947. 9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Title	WYX-62chanwu.1.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	12
Receiver Gain	112
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-17T14:58:01
Modification Date	2021-05-17T14:58:00
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1546.5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

—192.20

—158.60

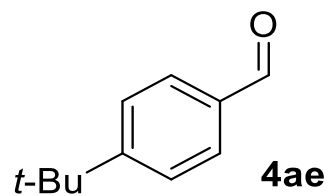
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—129.84

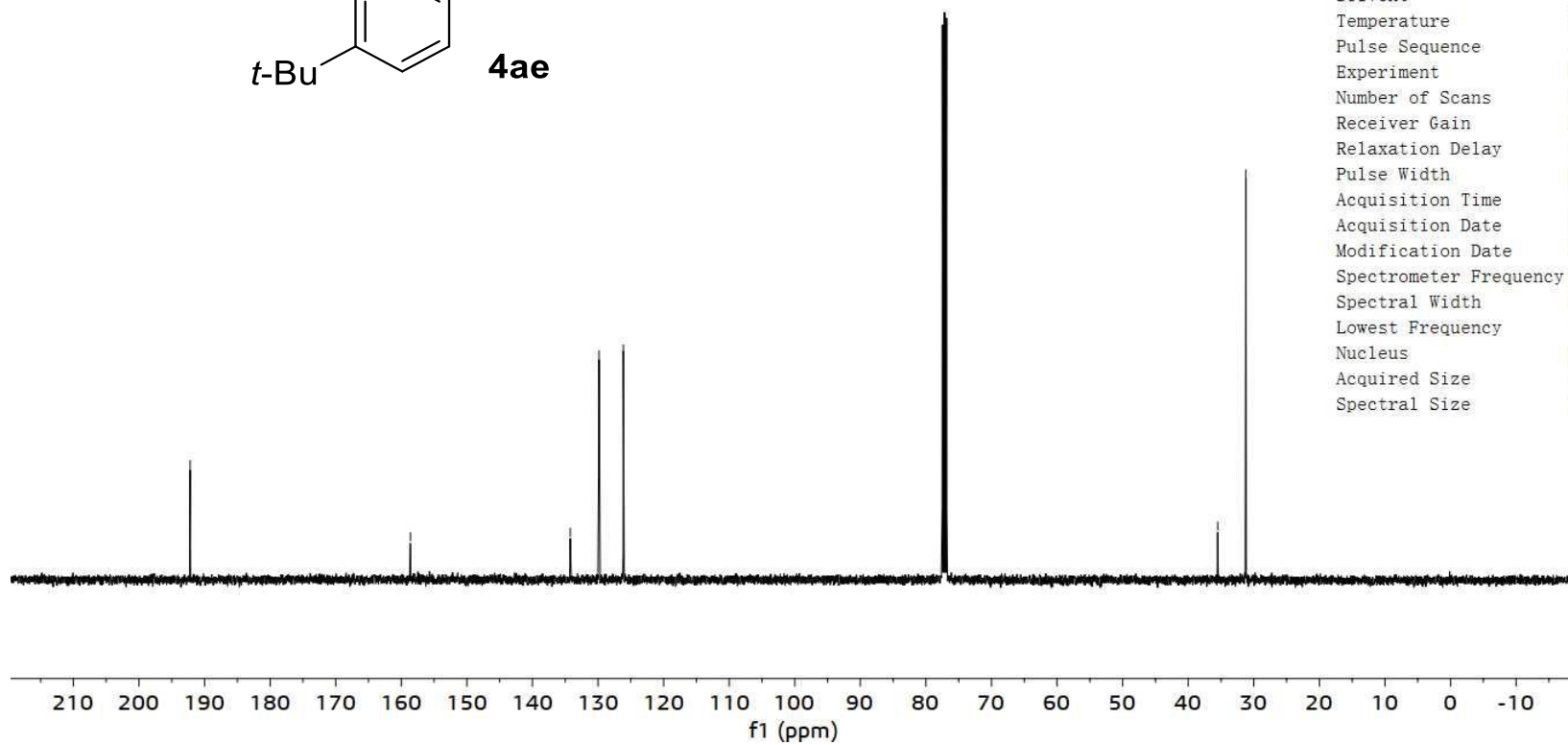
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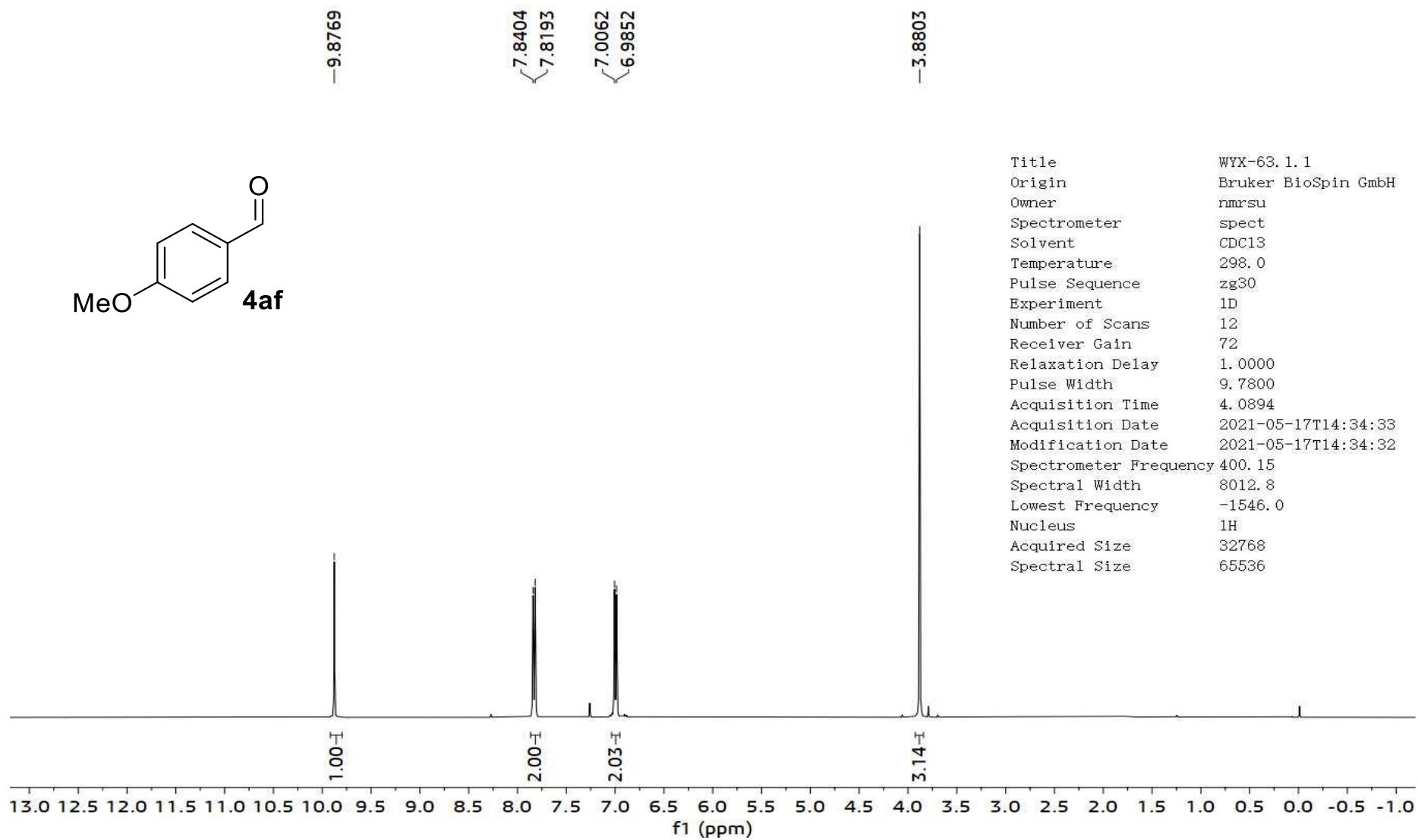
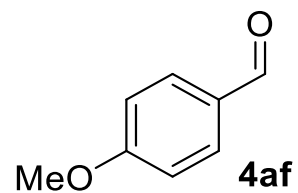
—35.50

—31.22



Title	WYX-62chanwu. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	300
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-17T15:16:14
Modification Date	2021-05-17T15:16:14
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1944.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





Title	WYX-63. 1. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	12
Receiver Gain	72
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-17T14:34:33
Modification Date	2021-05-17T14:34:32
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1546.0
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

—190.93

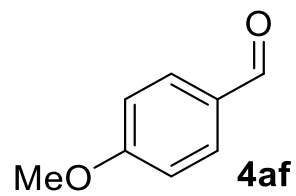
—164.73

✓132.10

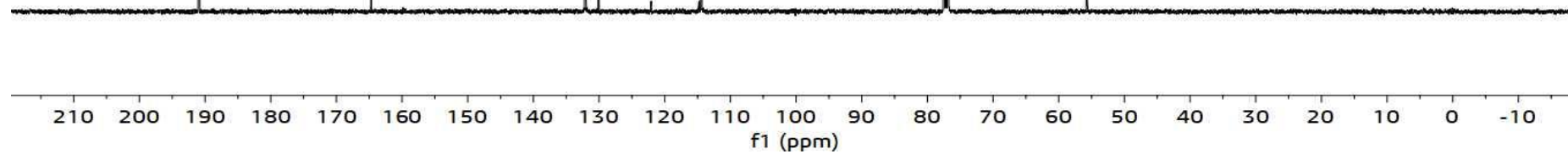
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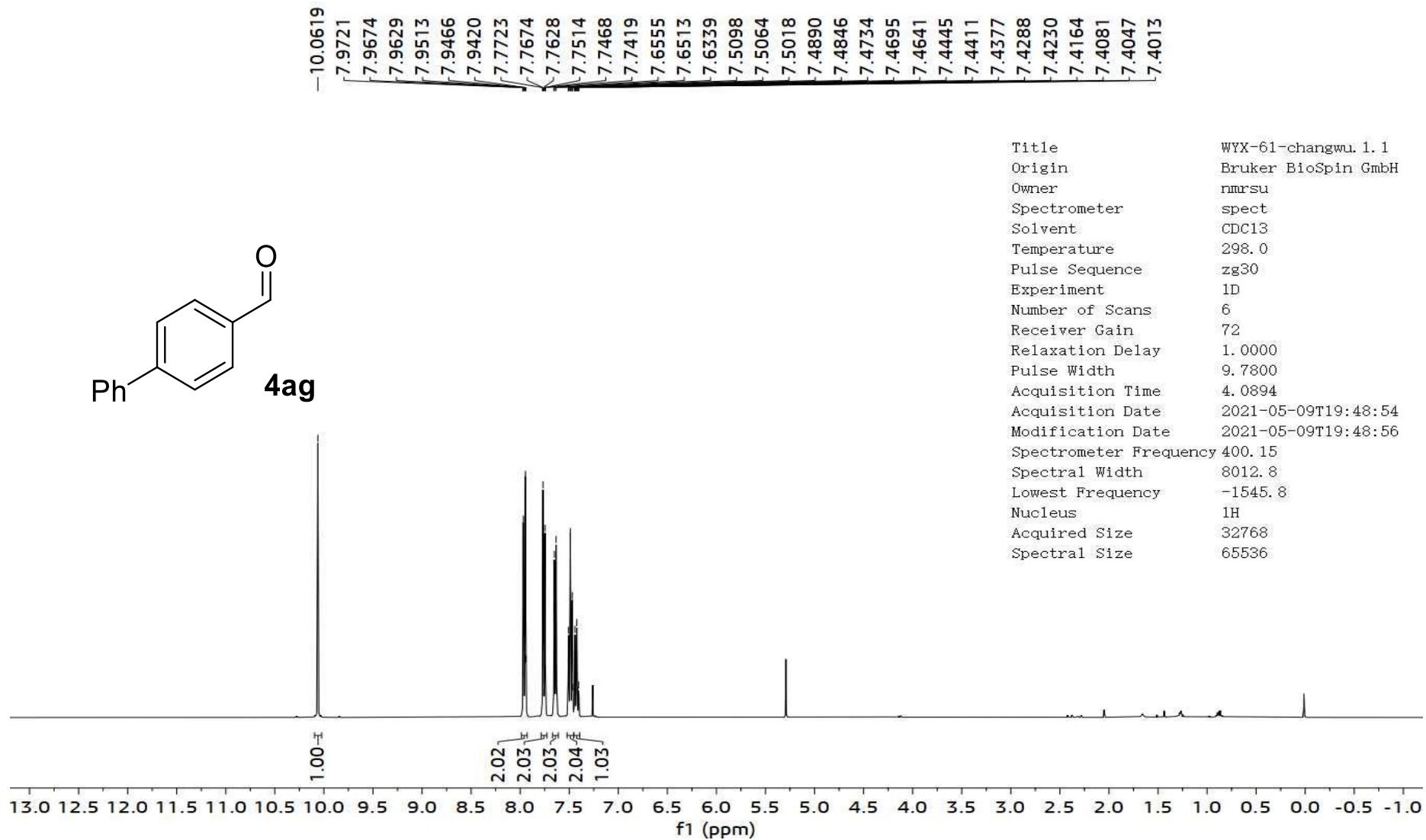
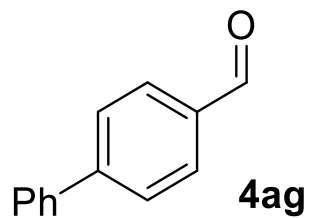
—114.43

—55.70

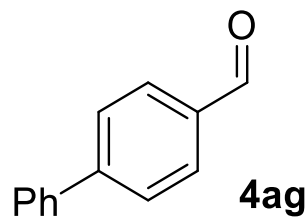


Title	WYX-63. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	300
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-17T14:52:46
Modification Date	2021-05-17T14:52:46
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1947.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

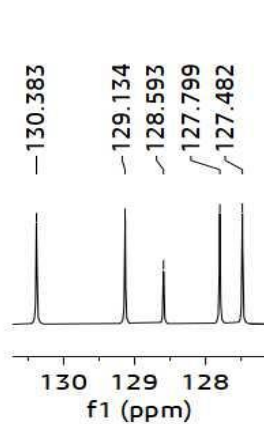




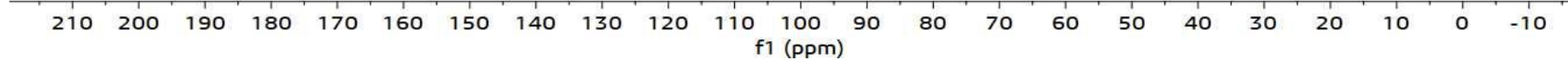
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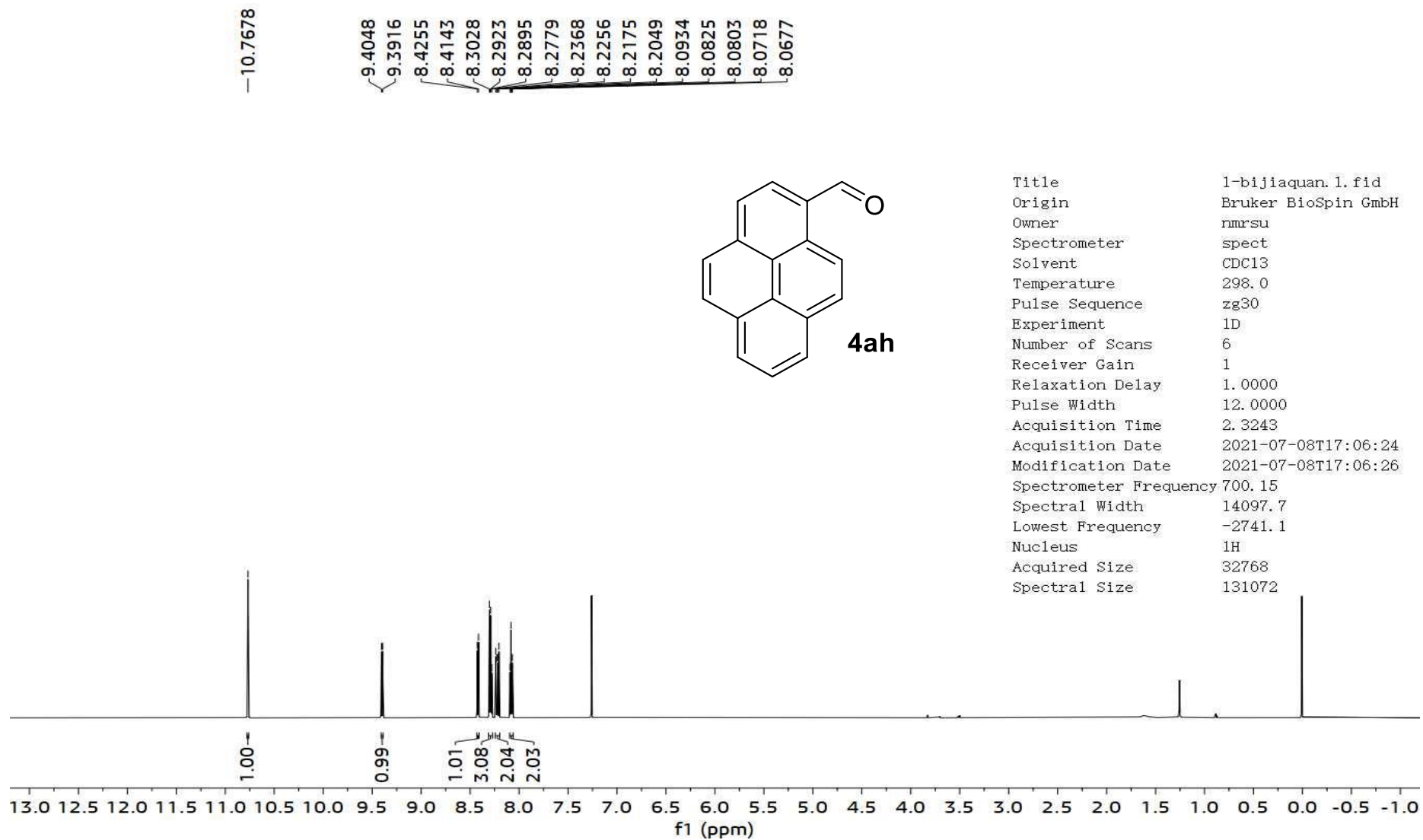


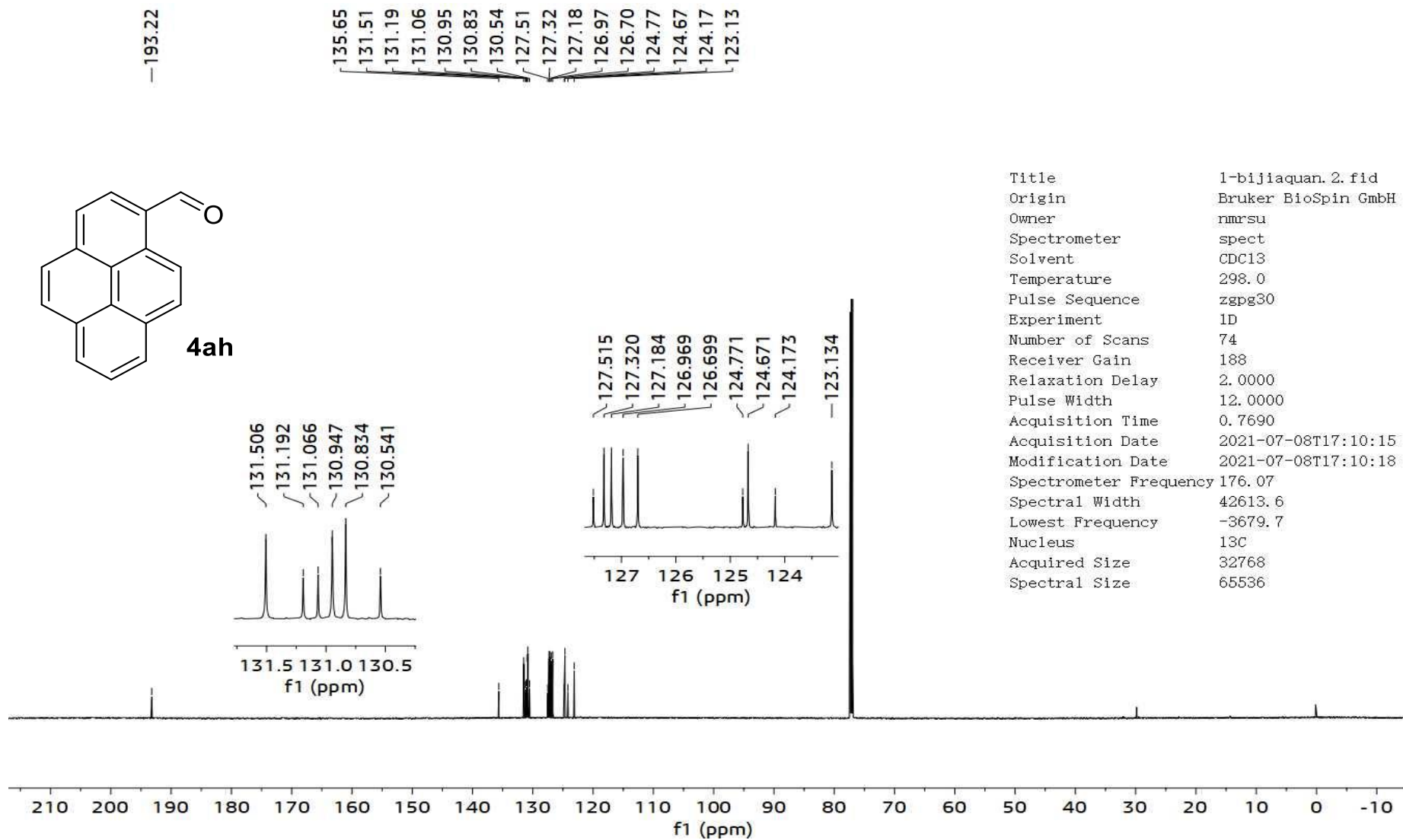
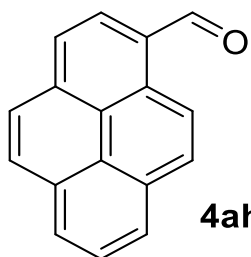
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130.38
129.13
128.59
127.80
127.48



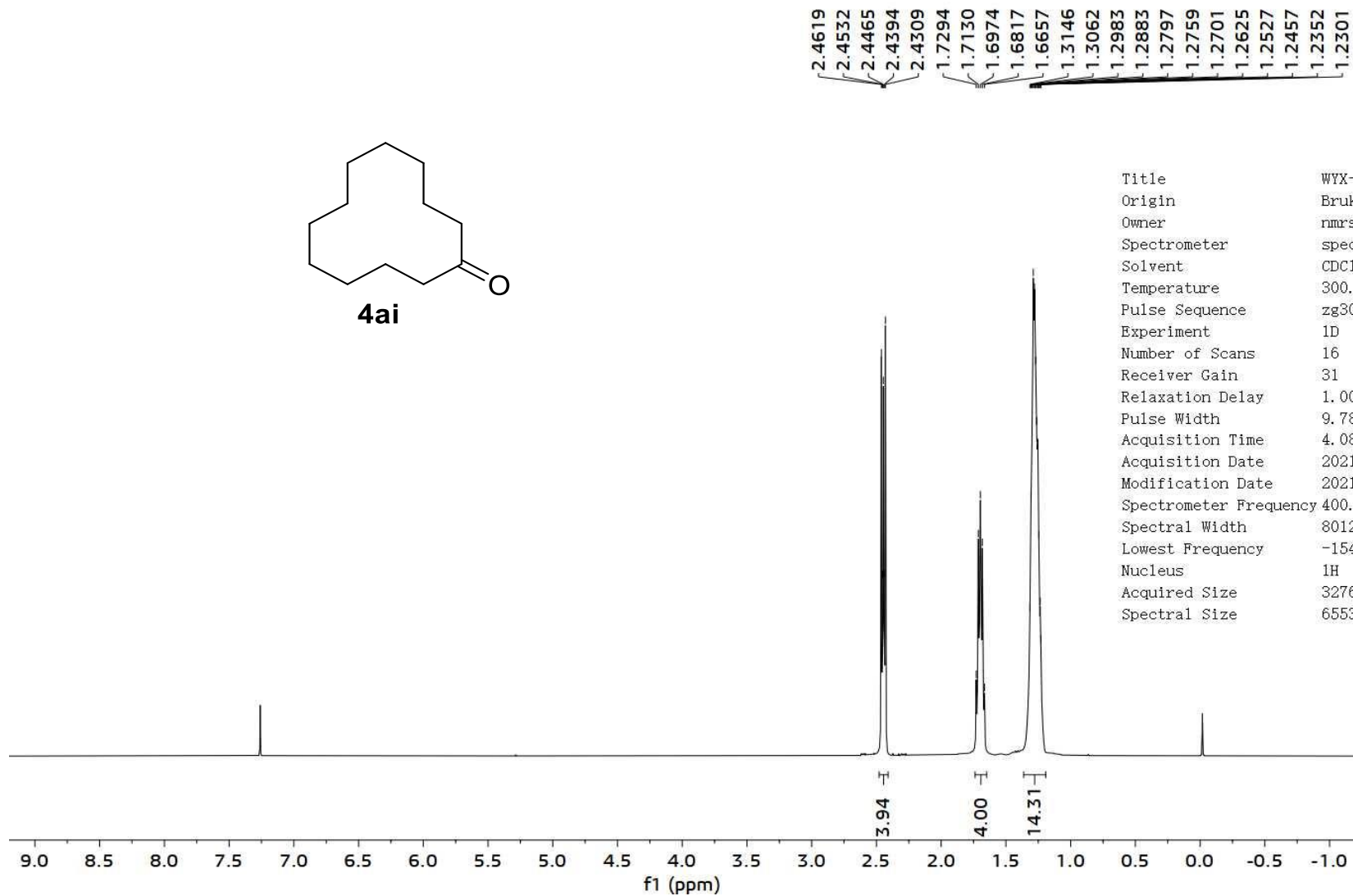
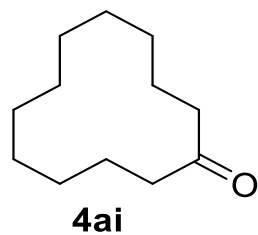
Title	WYX-61-changwu. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-09T20:48:28
Modification Date	2021-05-09T20:48:30
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1948.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



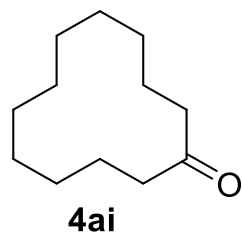




Title	1-bijiaquan.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	74
Receiver Gain	188
Relaxation Delay	2.0000
Pulse Width	12.0000
Acquisition Time	0.7690
Acquisition Date	2021-07-08T17:10:15
Modification Date	2021-07-08T17:10:18
Spectrometer Frequency	176.07
Spectral Width	42613.6
Lowest Frequency	-3679.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

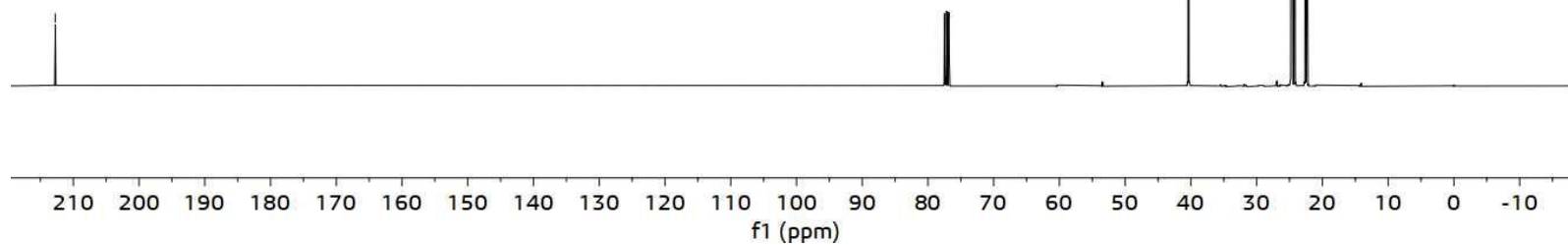


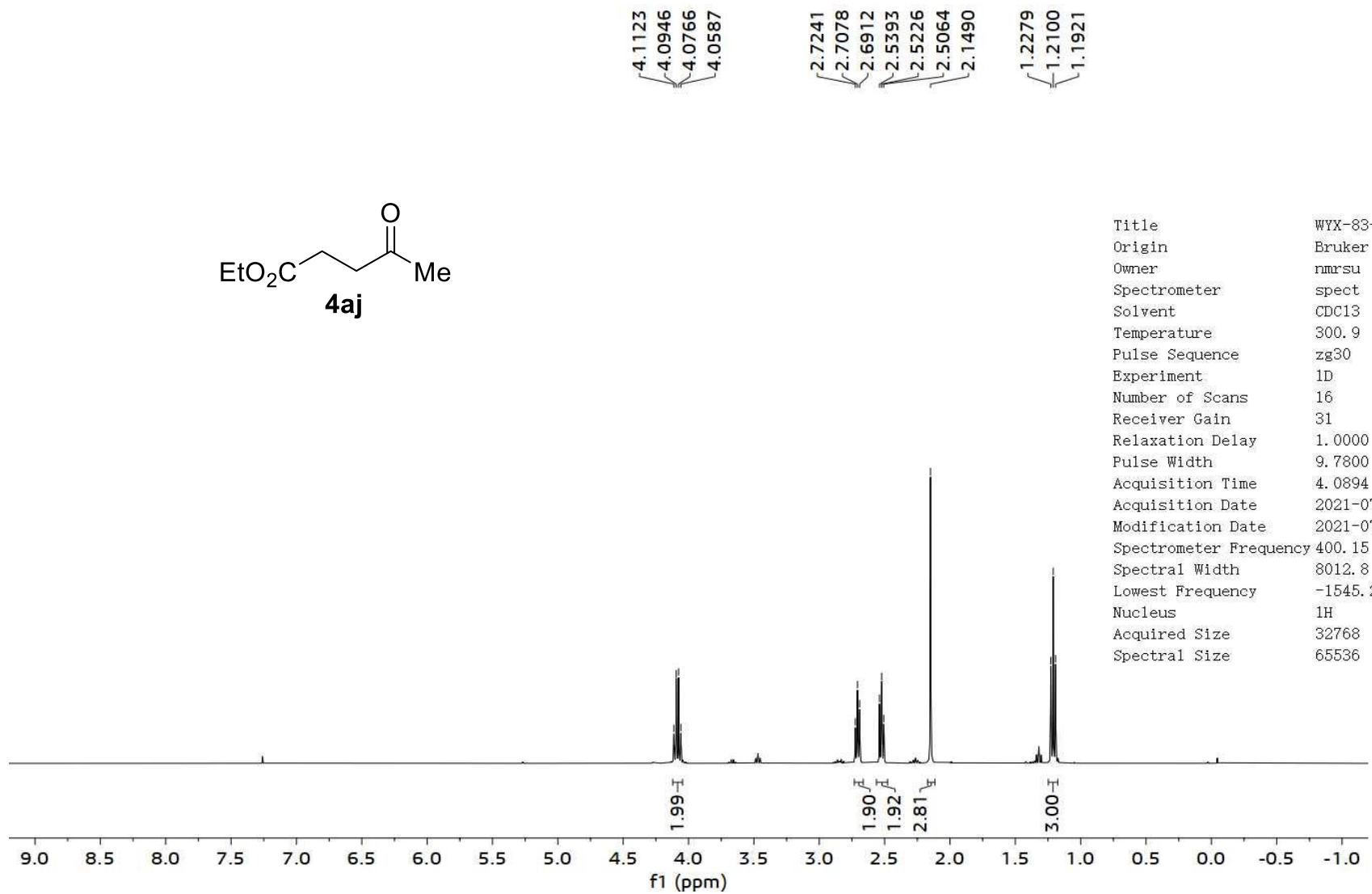
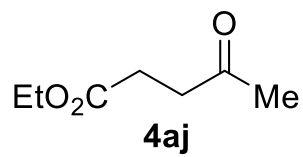
-212.73



-40.36
24.76
24.62
24.24
22.56
22.37

Title	WYX-81chanwu. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	299.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-15T21:24:01
Modification Date	2021-07-15T21:24:00
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1951.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





Title	WYX-83-charwu.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	300.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-21T01:50:28
Modification Date	2021-07-21T01:50:30
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

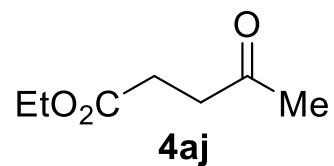
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—172.80

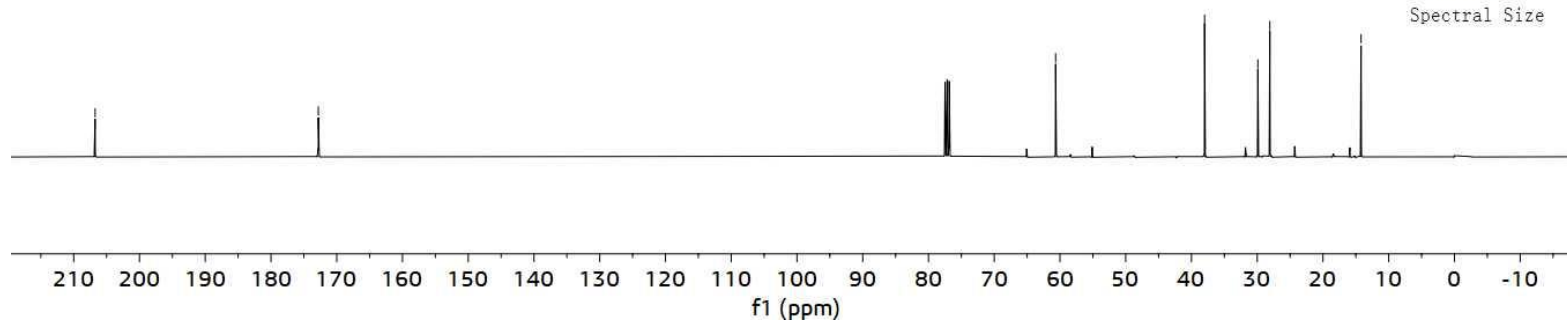
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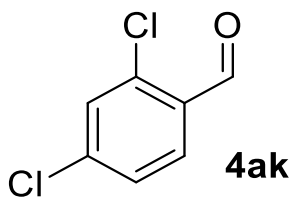
—38.00
—29.90
—28.08

—14.21



Title	WYX-83-chanwu.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDCl3
Temperature	301.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-21T02:34:23
Modification Date	2021-07-21T02:34:24
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1948.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

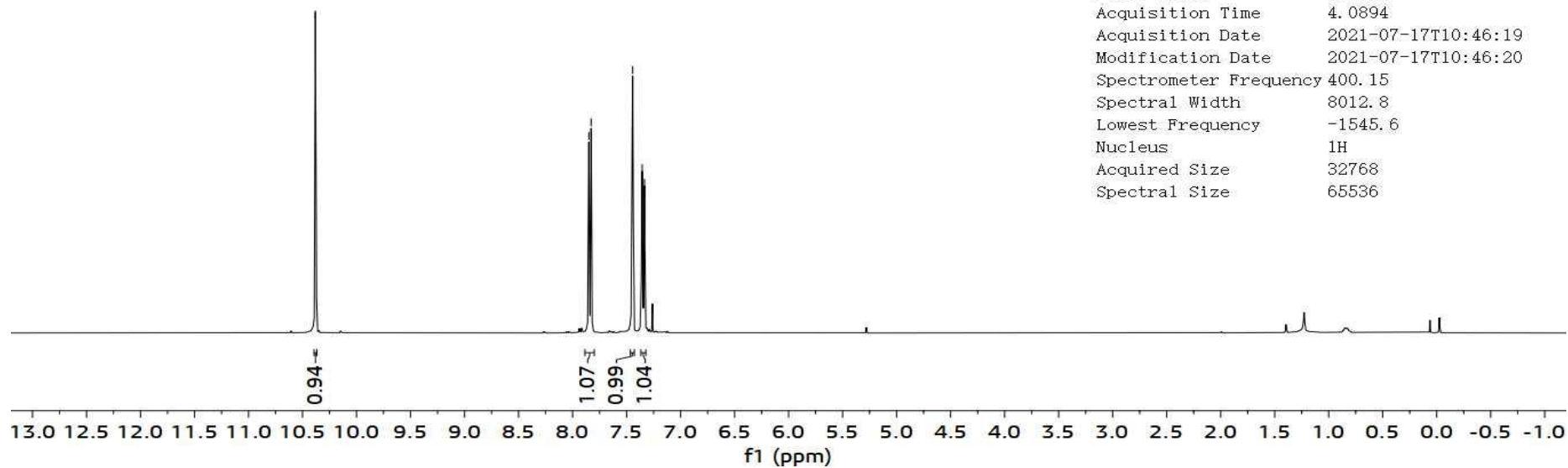


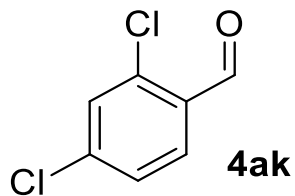


10.3813

7.8497
7.8286
7.4433
7.3553
7.3344

Title	WYX-82-fuchanwu.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	299.3
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	53
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-07-17T10:46:19
Modification Date	2021-07-17T10:46:20
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

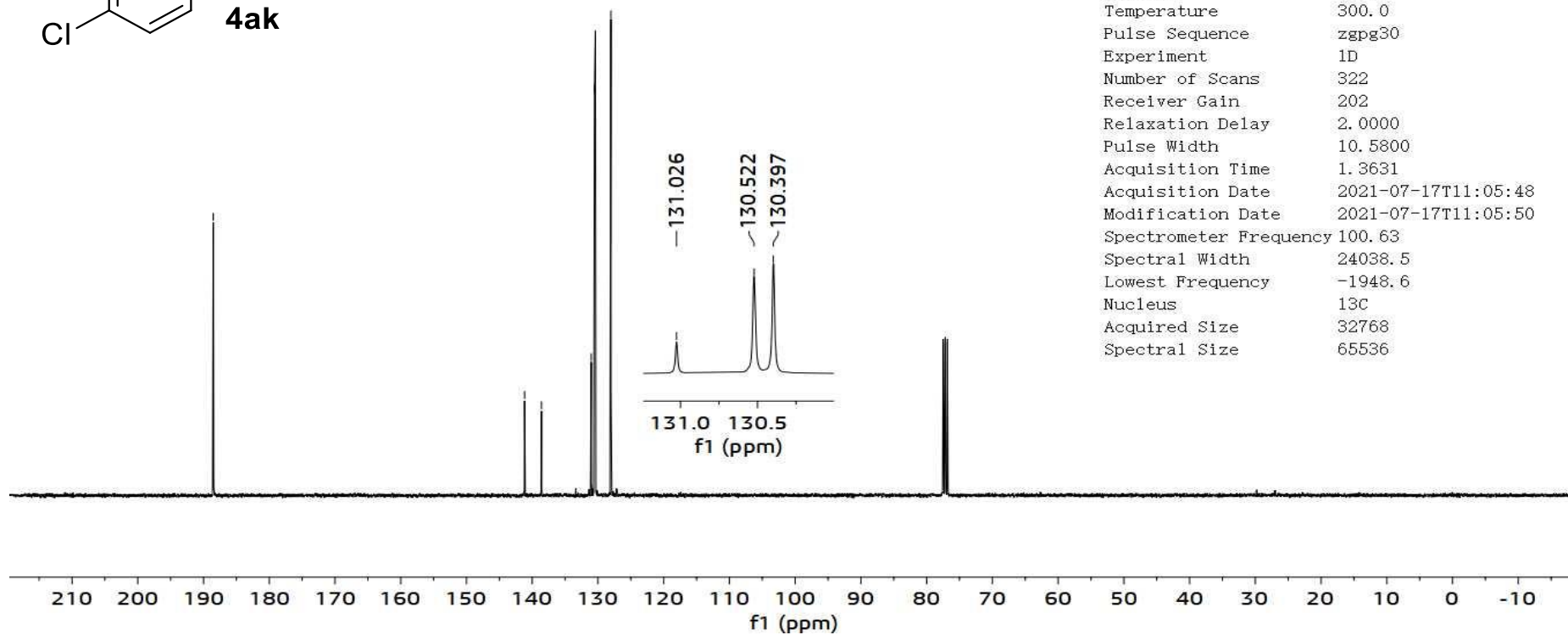


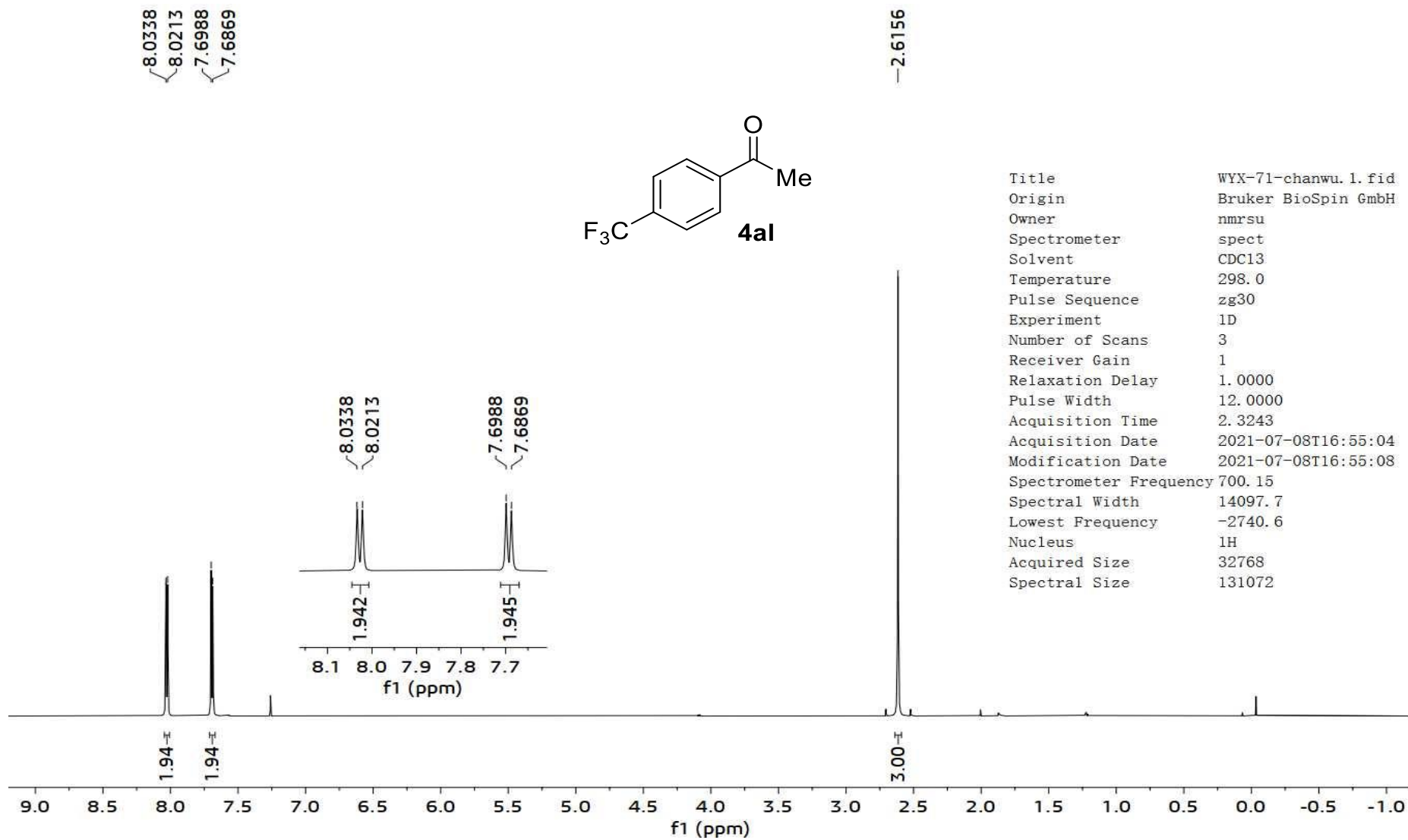


—188.52

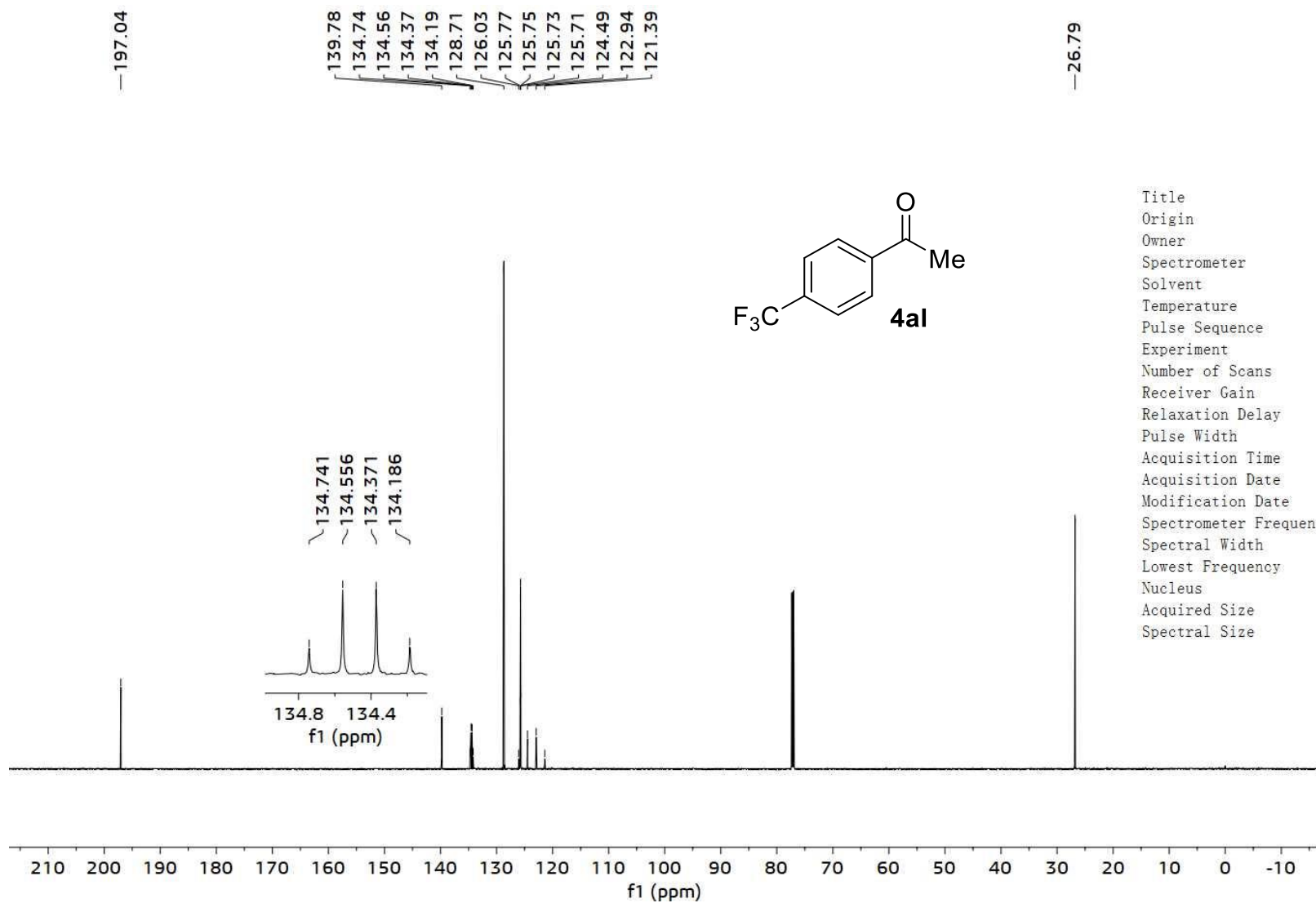
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128.03

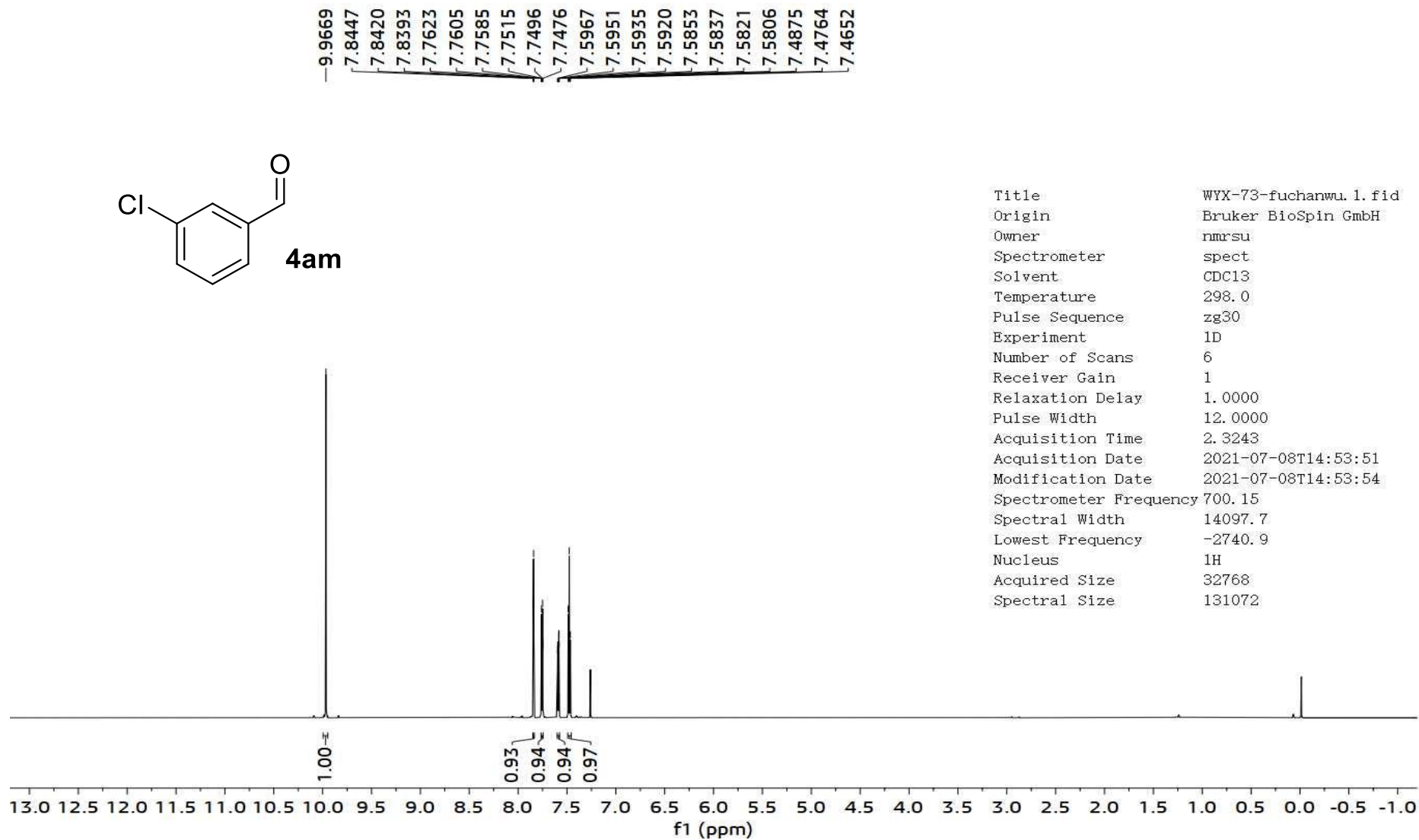
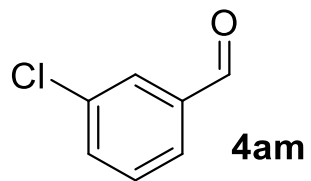
Title	WYX-82-fuchanwu. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	300.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	322
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-17T11:05:48
Modification Date	2021-07-17T11:05:50
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1948.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

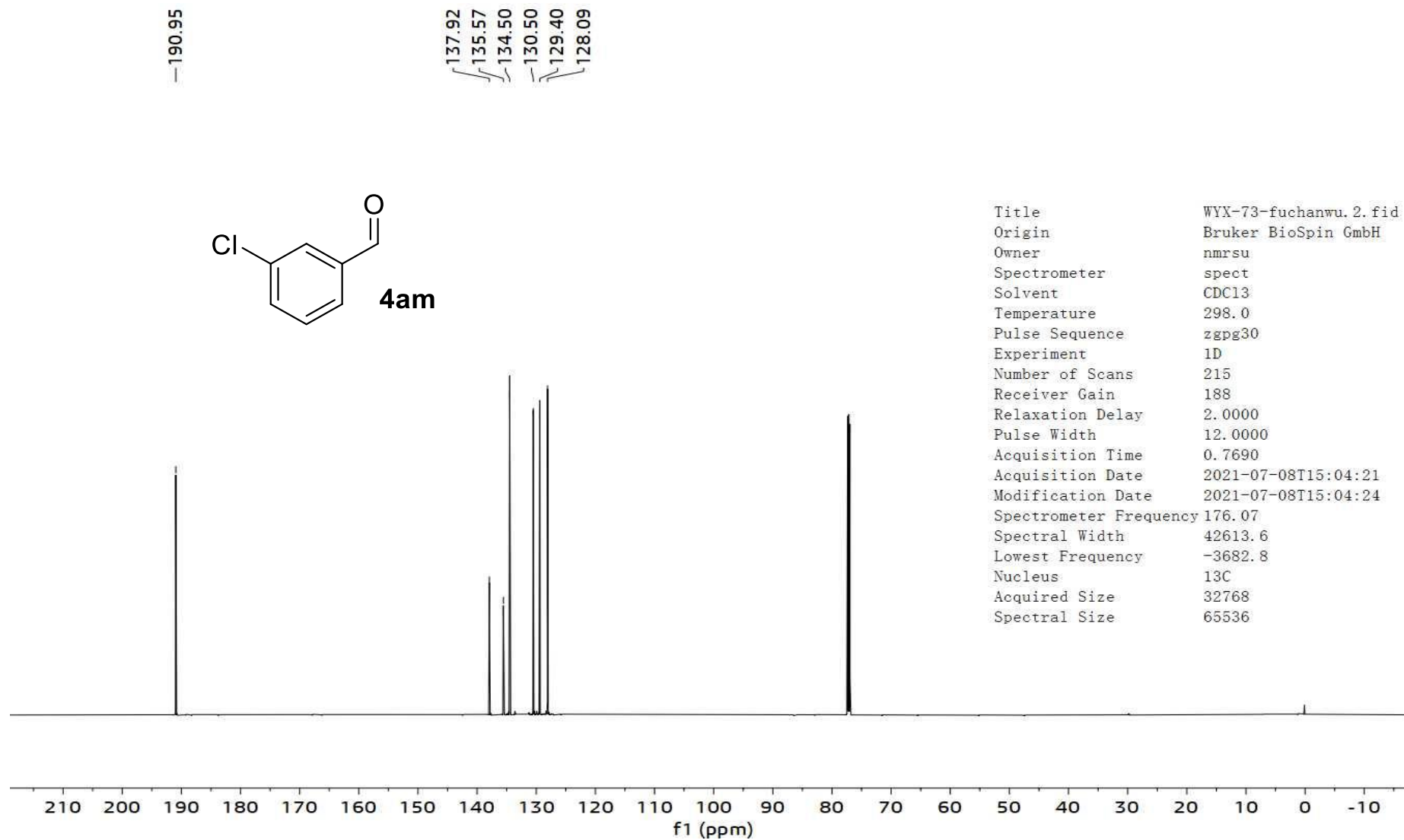




Title	WYX-71-ghanwu.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	3
Receiver Gain	1
Relaxation Delay	1.0000
Pulse Width	12.0000
Acquisition Time	2.3243
Acquisition Date	2021-07-08T16:55:04
Modification Date	2021-07-08T16:55:08
Spectrometer Frequency	700.15
Spectral Width	14097.7
Lowest Frequency	-2740.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	131072





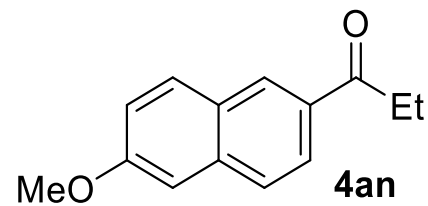


8.3773
8.0042
8.0016
7.9920
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7.1897
7.1860
7.1769
7.1733
7.1272
7.1235

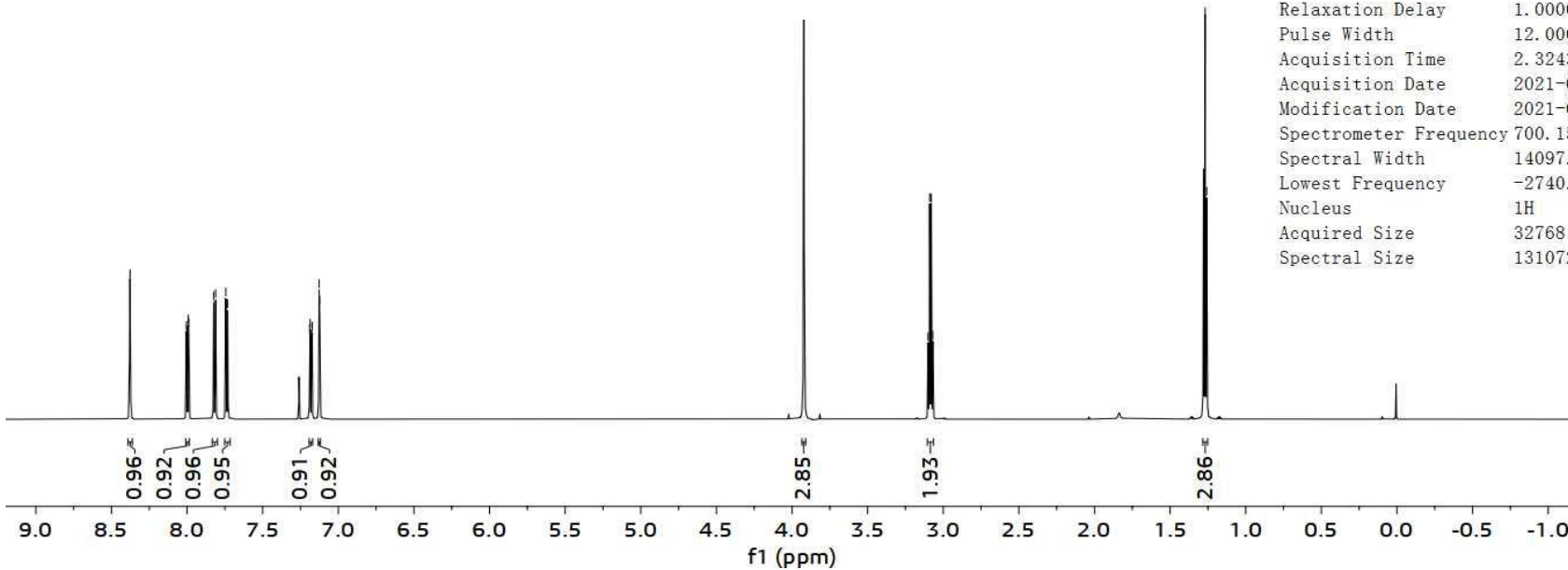
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3.0794
3.0690

1.2788
1.2685
1.2579



Title	WYX-72-chanwu.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	1
Relaxation Delay	1.0000
Pulse Width	12.0000
Acquisition Time	2.3243
Acquisition Date	2021-07-08T16:48:30
Modification Date	2021-07-08T16:48:32
Spectrometer Frequency	700.15
Spectral Width	14097.7
Lowest Frequency	-2740.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	131072



—200.61

—159.70

137.24

132.38

131.15

129.45

127.92

127.12

124.70

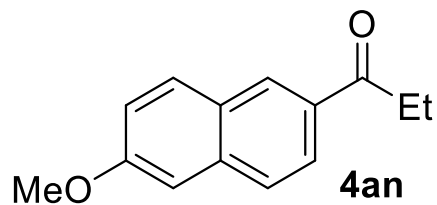
119.71

—105.78

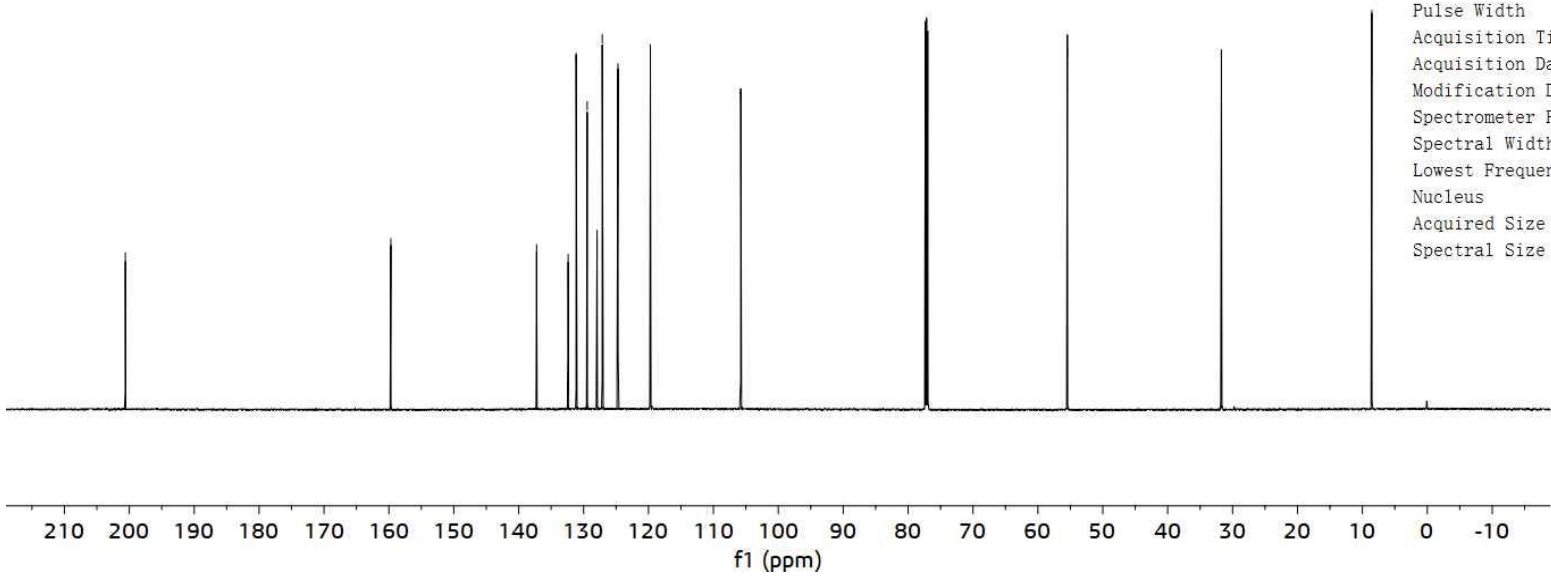
—55.46

—31.73

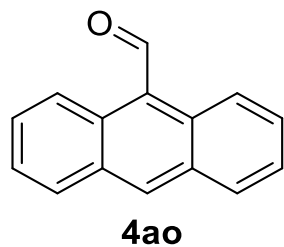
—8.55



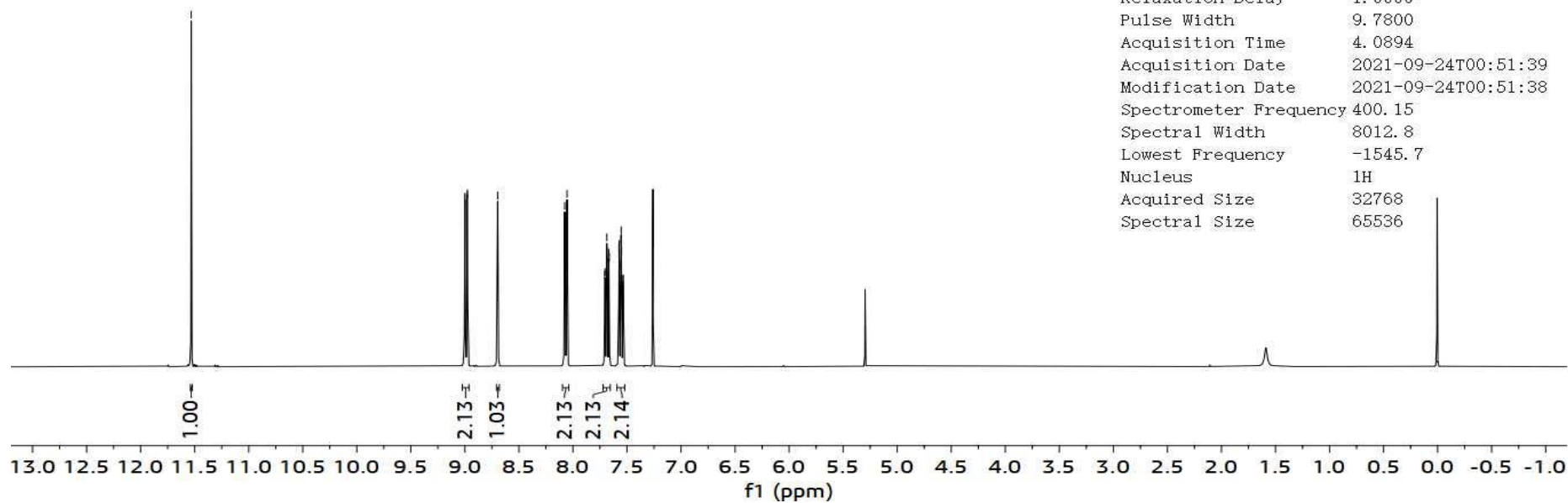
Title	WYX-72-chanwu.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	40
Receiver Gain	188
Relaxation Delay	2.0000
Pulse Width	12.0000
Acquisition Time	0.7690
Acquisition Date	2021-07-08T16:50:39
Modification Date	2021-07-08T16:50:48
Spectrometer Frequency	176.07
Spectral Width	42613.6
Lowest Frequency	-3690.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



— 11.5314

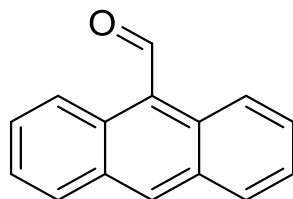


9.0011
8.9986
8.9787
8.9761
8.6965
8.0761
8.0551
7.7069
7.7034
7.6869
7.6842
7.6679
7.6644
7.5746
7.5720
7.5582
7.5553
7.5533
7.5507
7.5370
7.5344



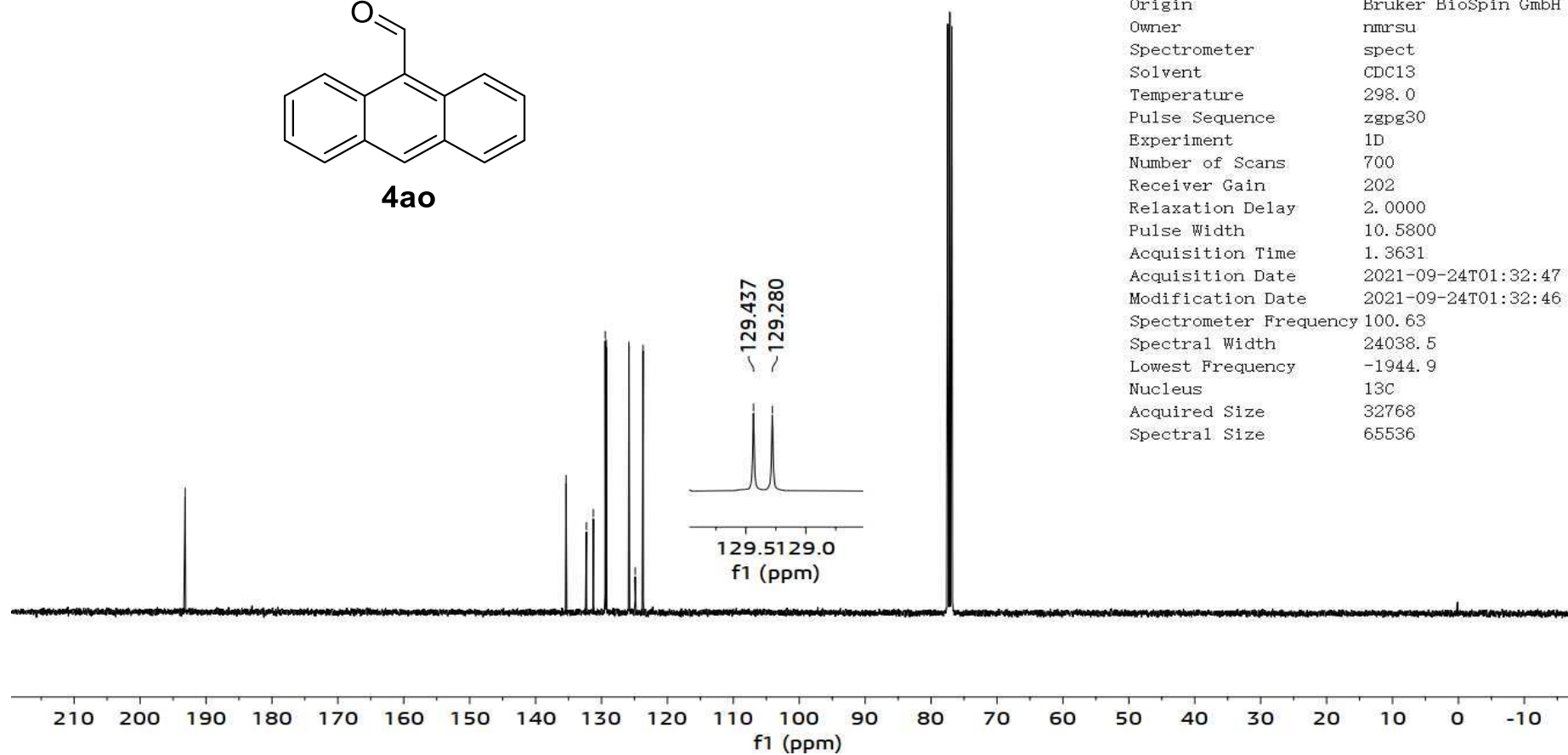
Title	SN-2-20210923.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	127
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-24T00:51:39
Modification Date	2021-09-24T00:51:38
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.7
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

—193.16

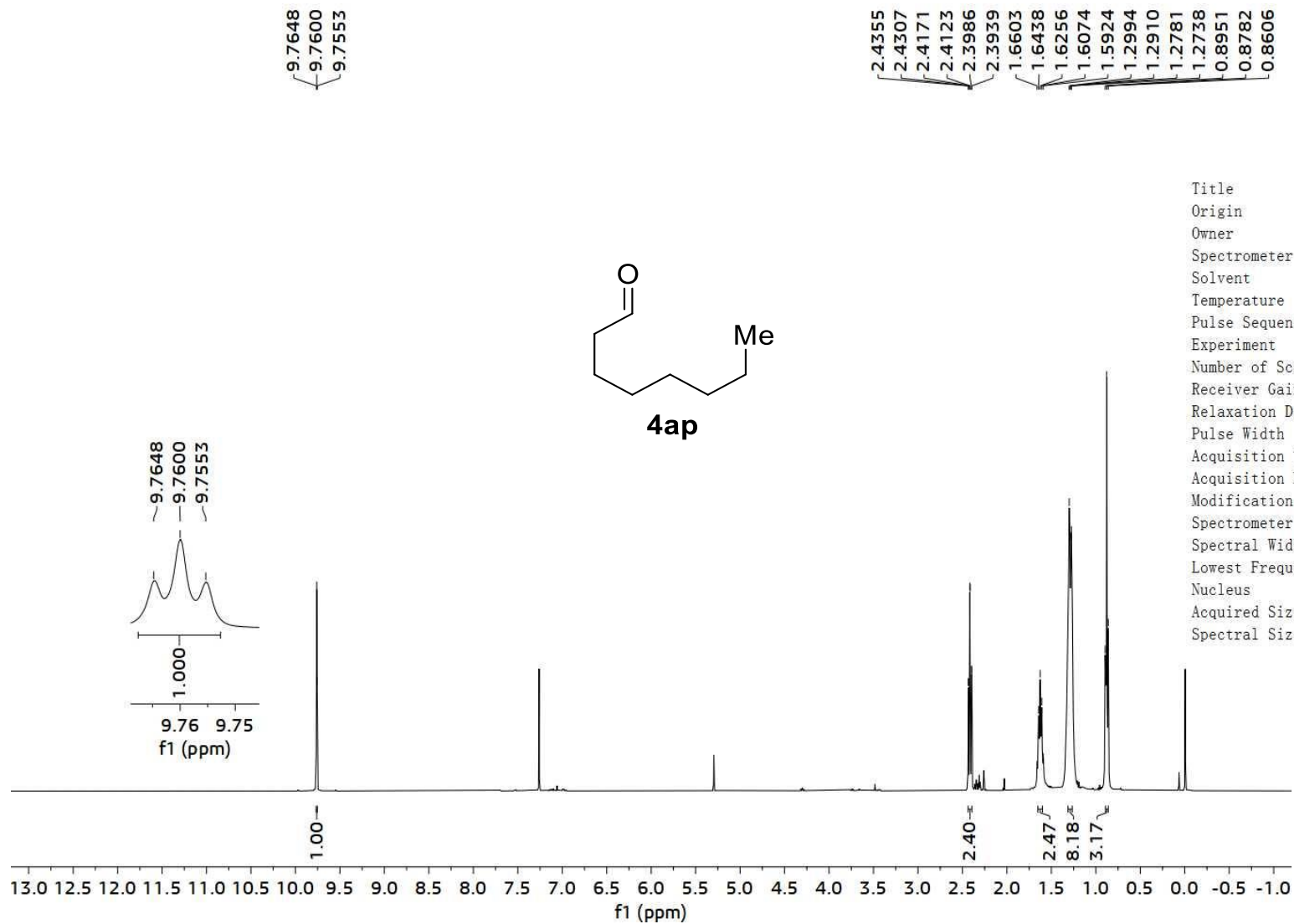


4ao

135.38
132.29
131.23
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129.28
125.84
124.89
123.70

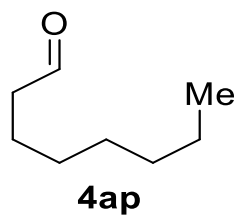


Title	SN-2-20210923.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-24T01:32:47
Modification Date	2021-09-24T01:32:46
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1944.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



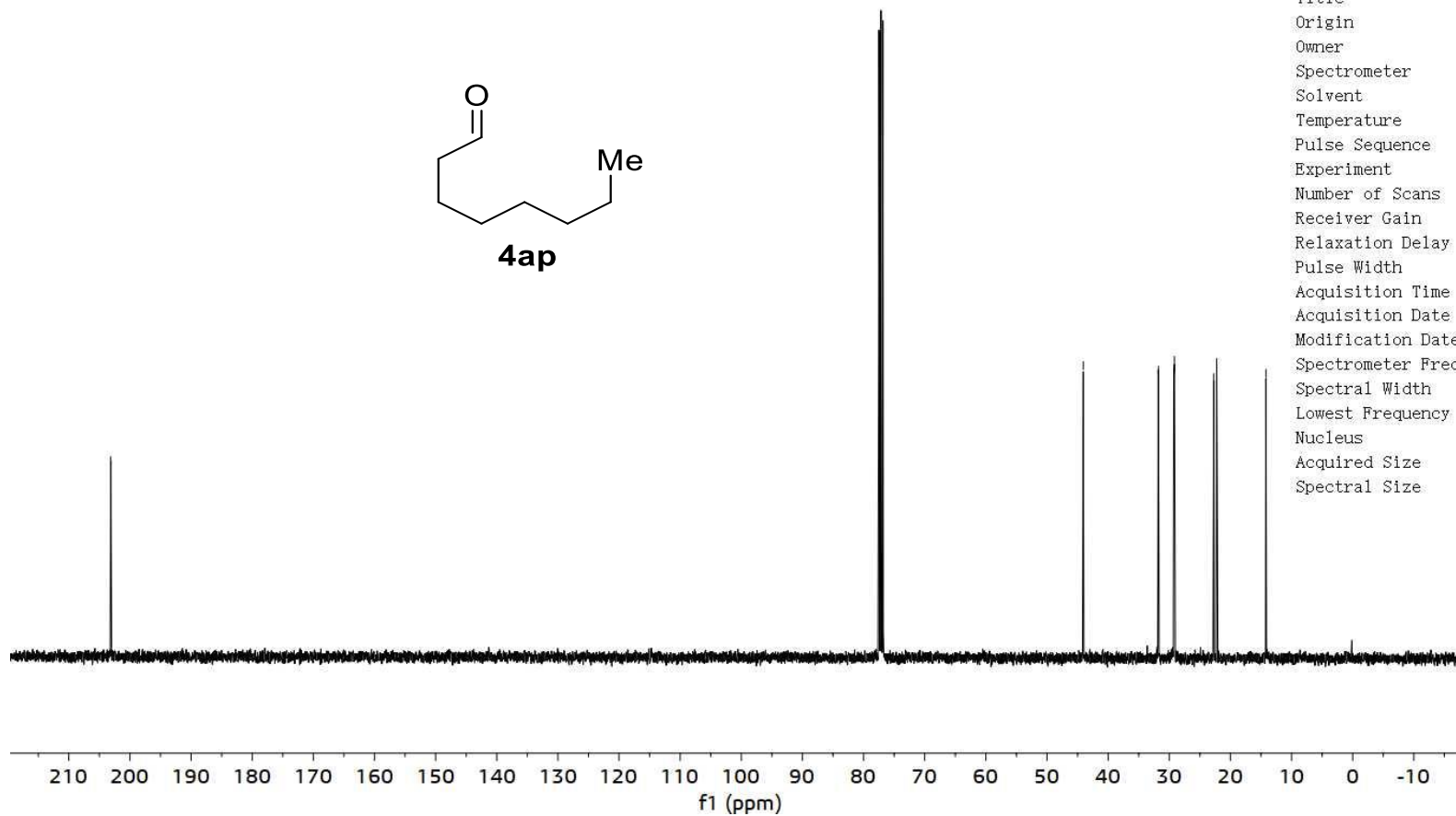
Title	SN-3.3.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	79
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-23T08:27:55
Modification Date	2021-09-23T08:27:54
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

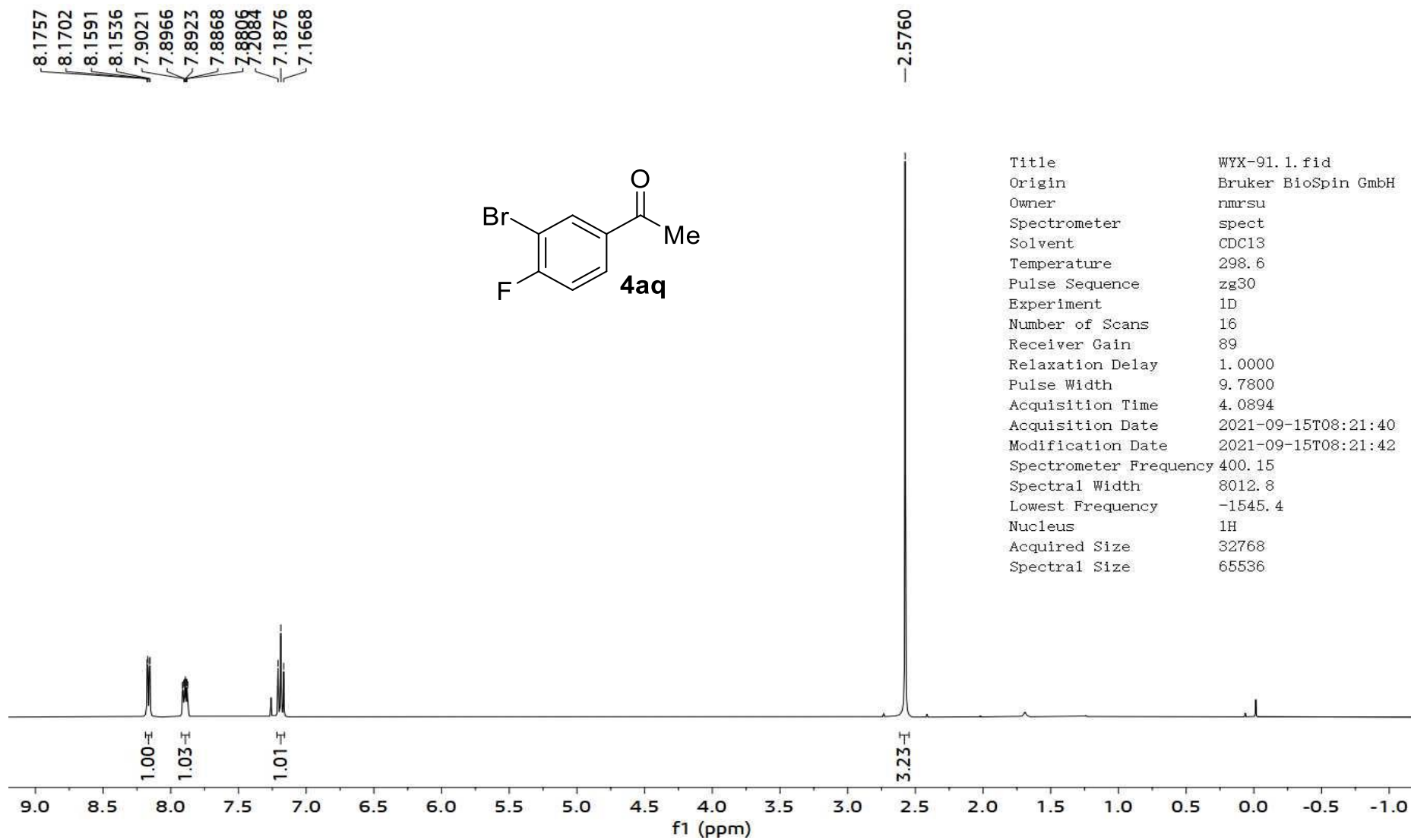
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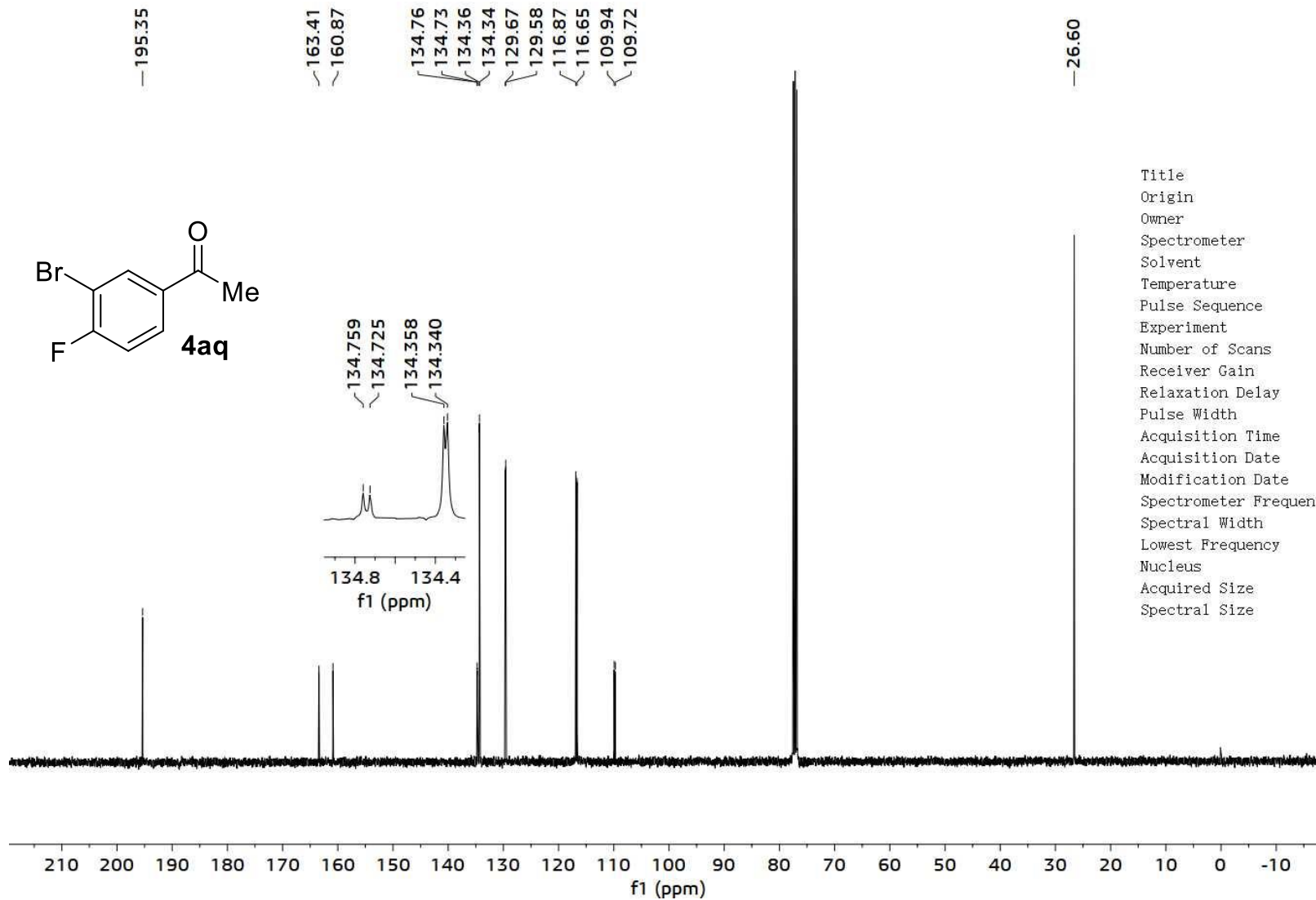


—44.07
31.77
29.27
29.16
22.73
22.24
14.19

Title	SN-3.4.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-23T08:57:37
Modification Date	2021-09-23T08:57:36
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1943.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





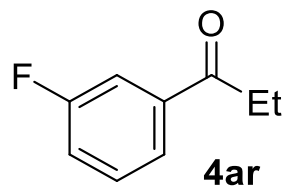


Title	WYX-91.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	299.2
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	441
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-15T08:47:57
Modification Date	2021-09-15T08:48:00
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1946.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

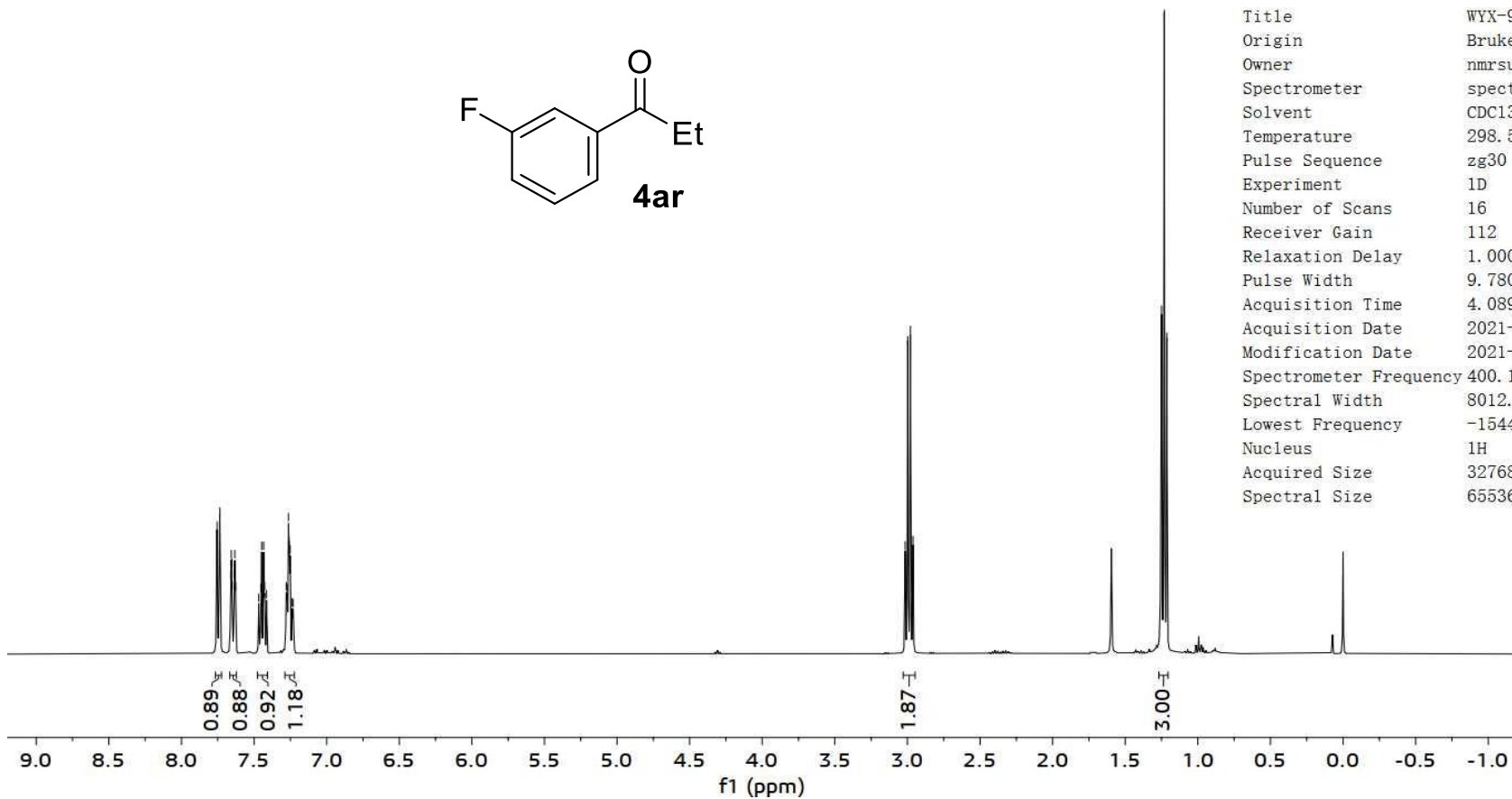
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7.4535
7.4474
7.4335
7.4273
7.4137
7.2782
7.2718
7.2632
7.2583
7.2512
7.2369
7.2306

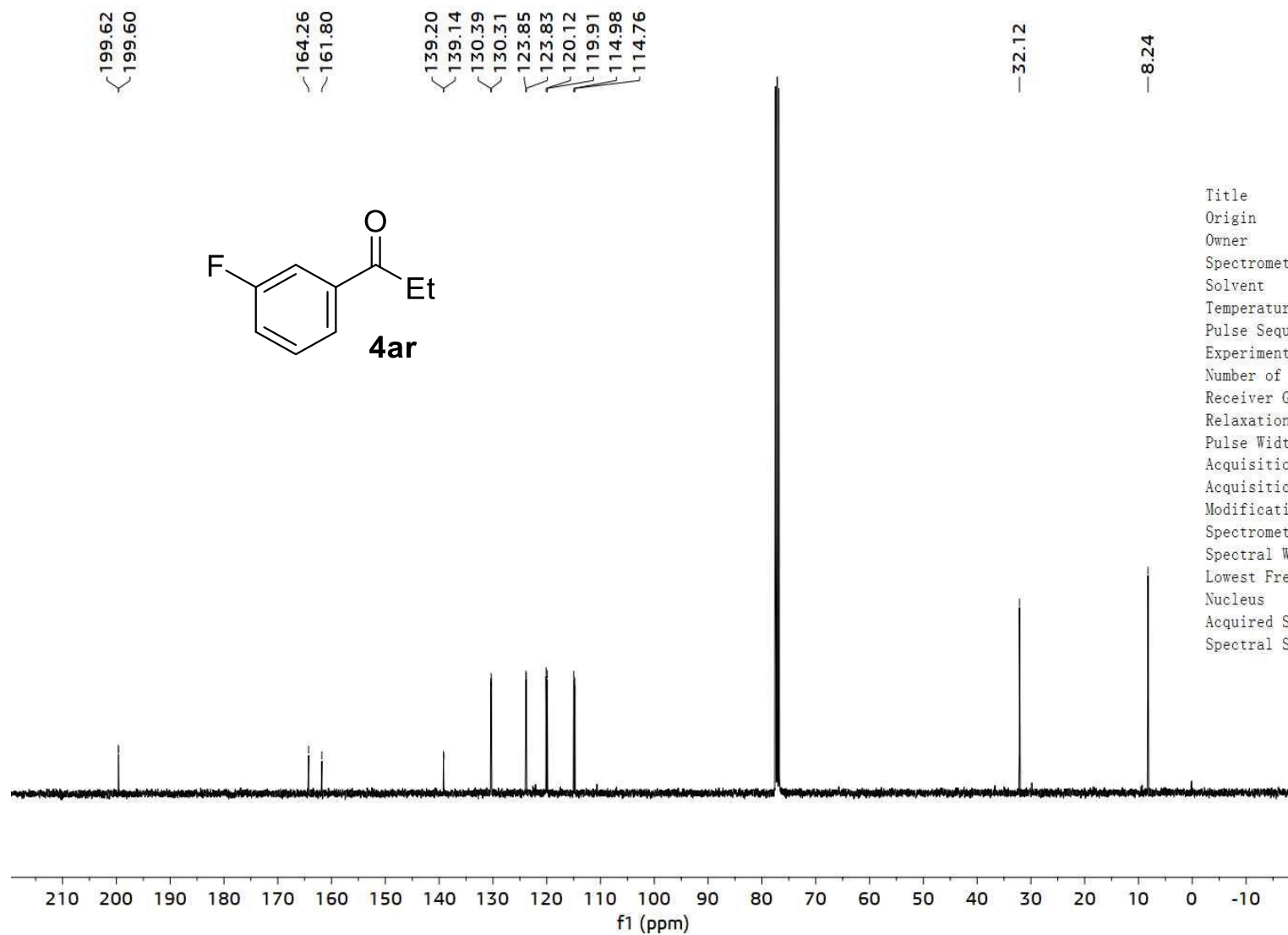
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2.9617

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1.2313
1.2132

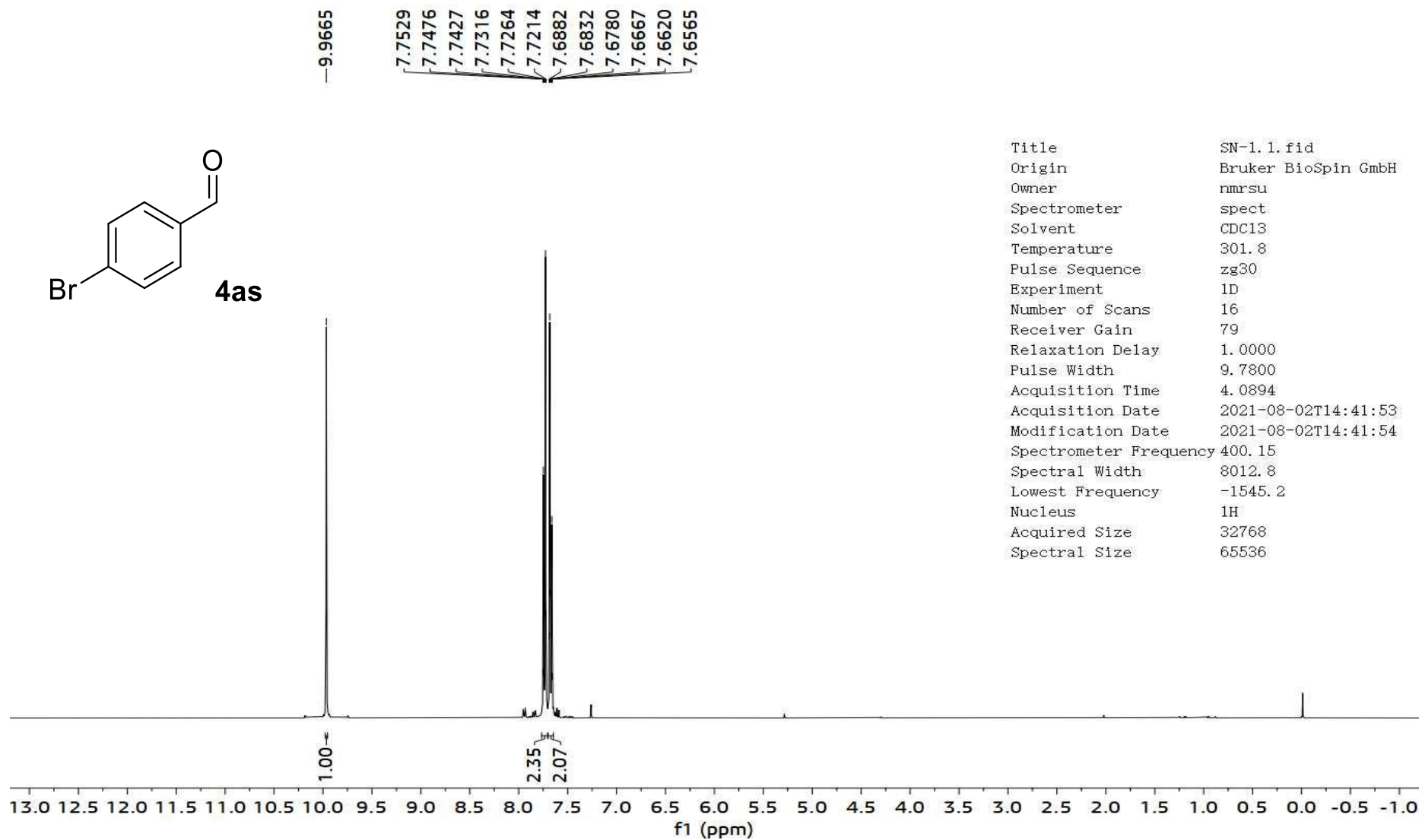
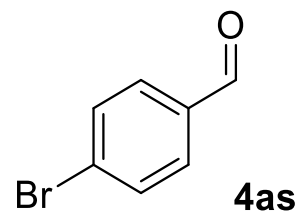


Title	WYX-93.3.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.5
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	112
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-15T08:59:12
Modification Date	2021-09-15T08:59:14
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1544.2
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

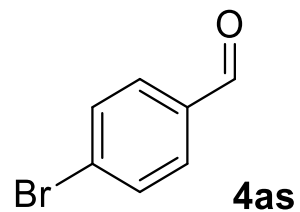




Title	WYX-93.4.fid
Origin	Bruker BioSpin GmbH
Owner	nmsu
Spectrometer	spect
Solvent	CDC13
Temperature	299.2
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-15T09:28:50
Modification Date	2021-09-15T09:28:52
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1944.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

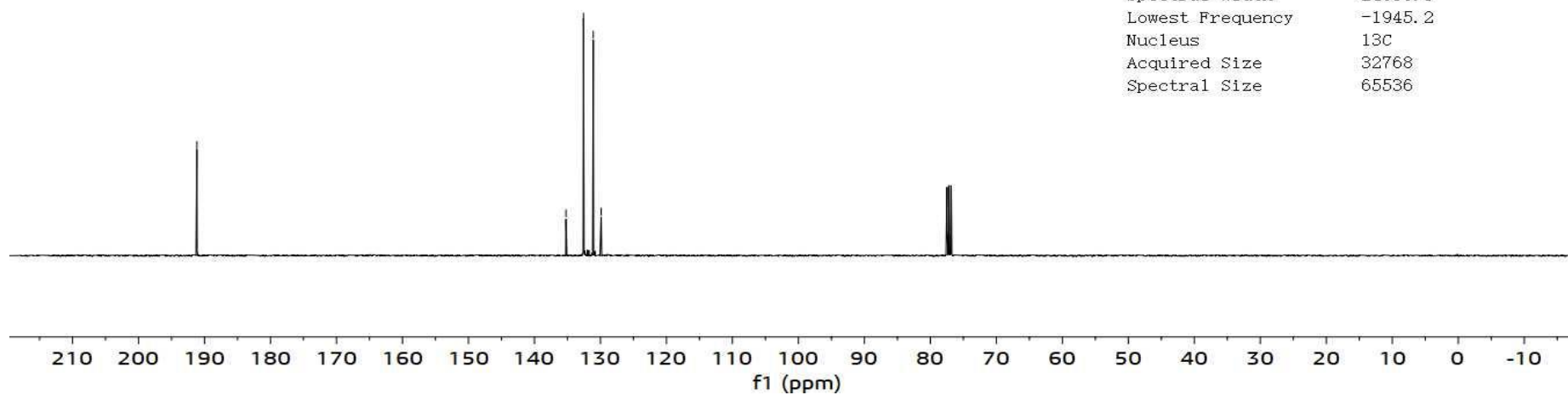


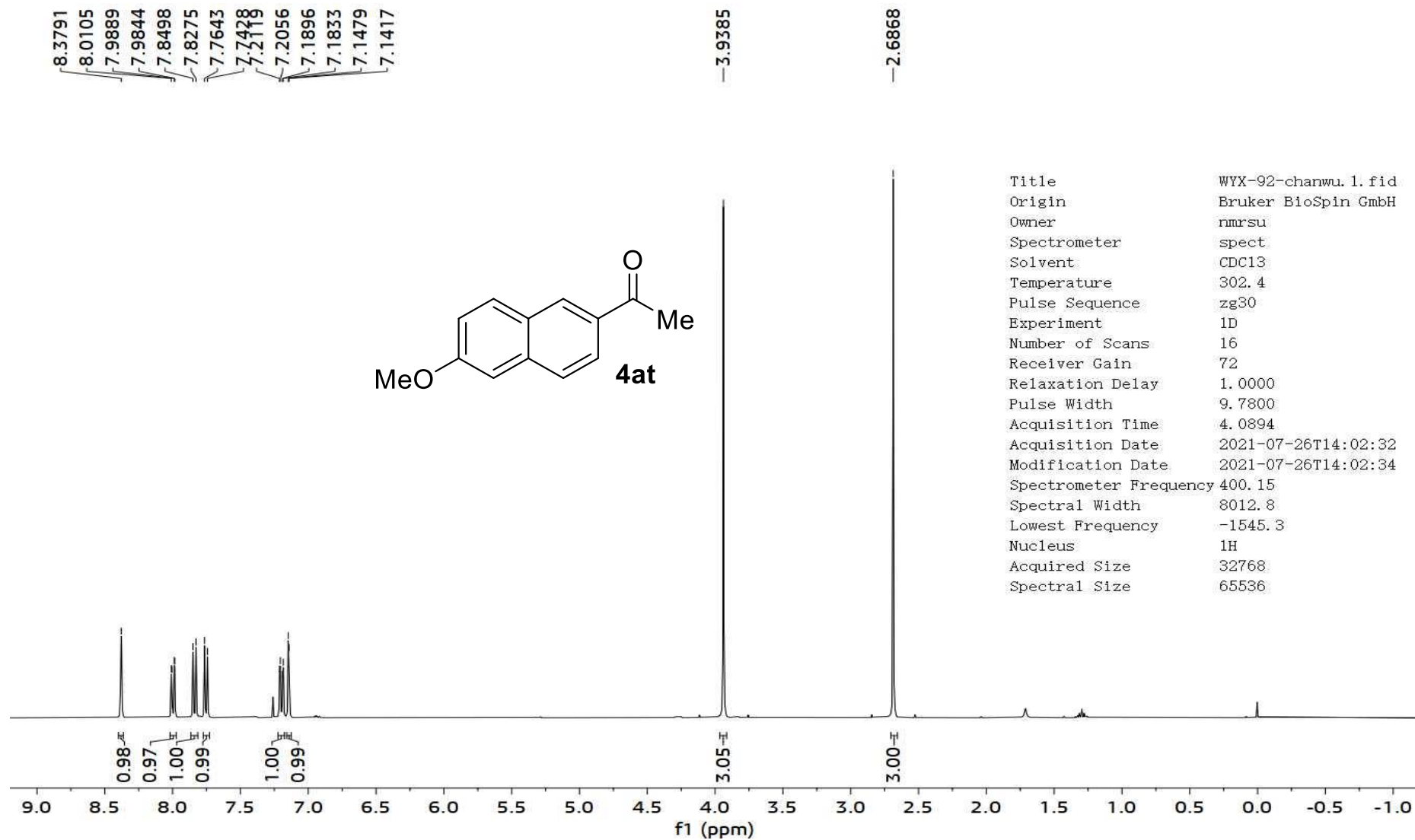
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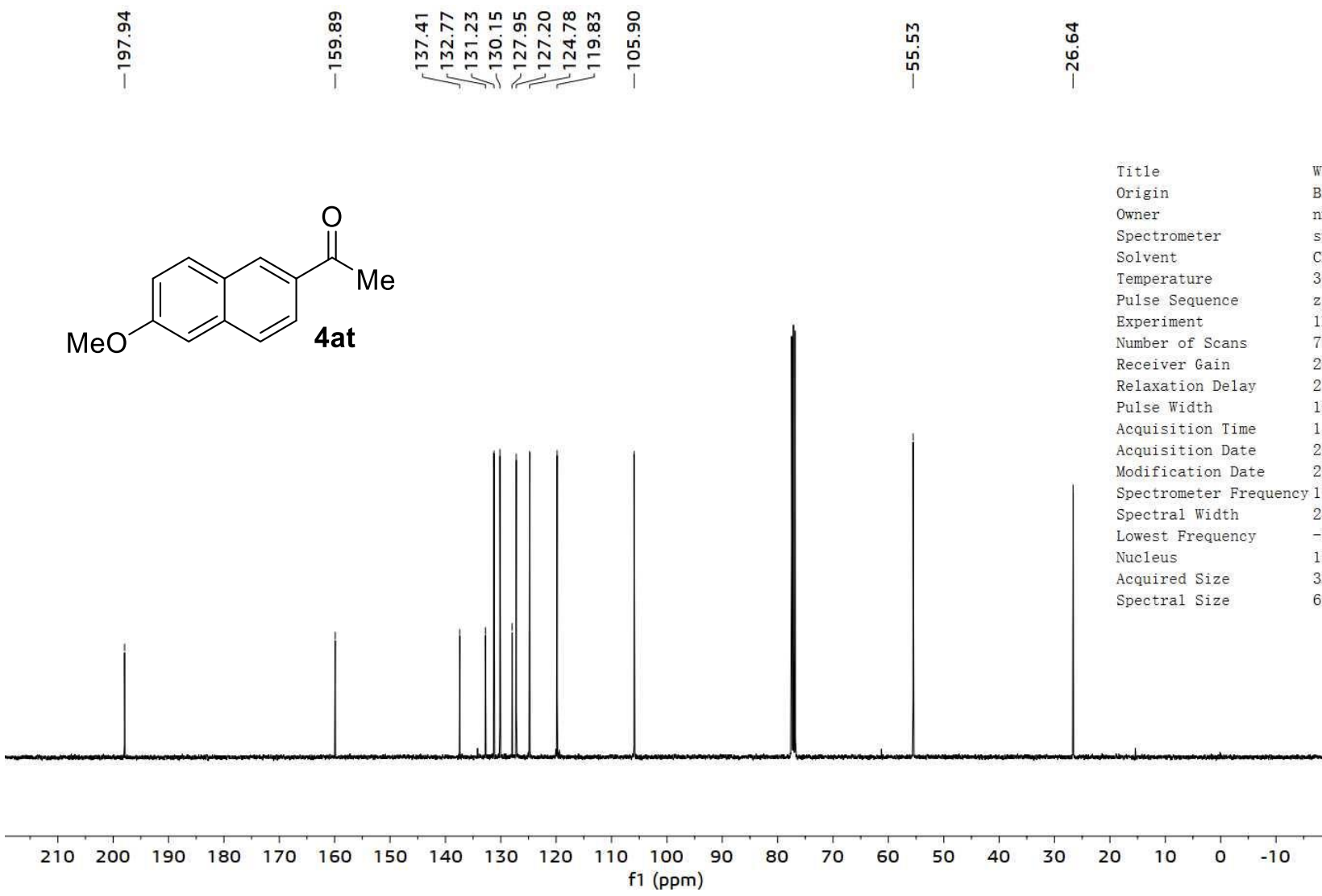
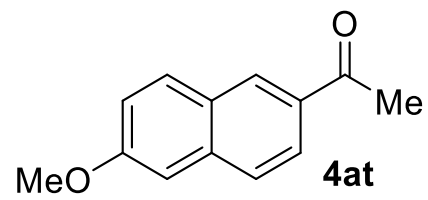


135.23
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131.09
129.89

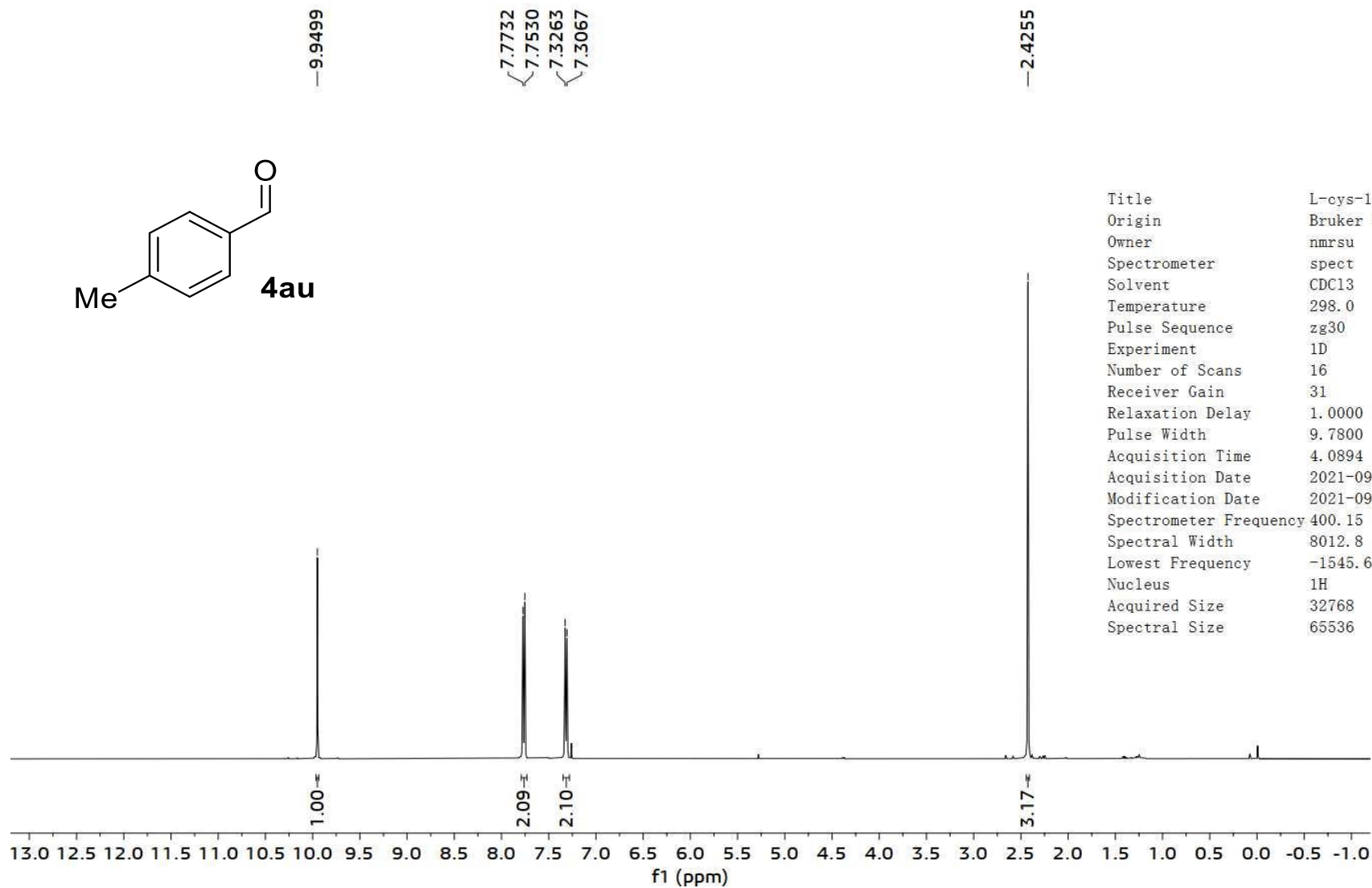
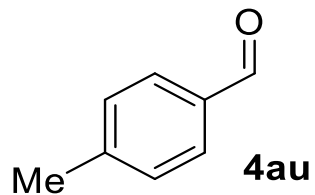
Title	SN-1.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	302.4
Pulse Sequence	zpgp30
Experiment	1D
Number of Scans	507
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-08-02T15:11:58
Modification Date	2021-08-02T15:12:00
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1945.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536







Title	WYX-92-chanwu. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	303.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	750
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-07-26T14:46:27
Modification Date	2021-07-26T14:46:28
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1946.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Title	L-cys-1.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-23T09:02:44
Modification Date	2021-09-23T09:02:44
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

— 192.07

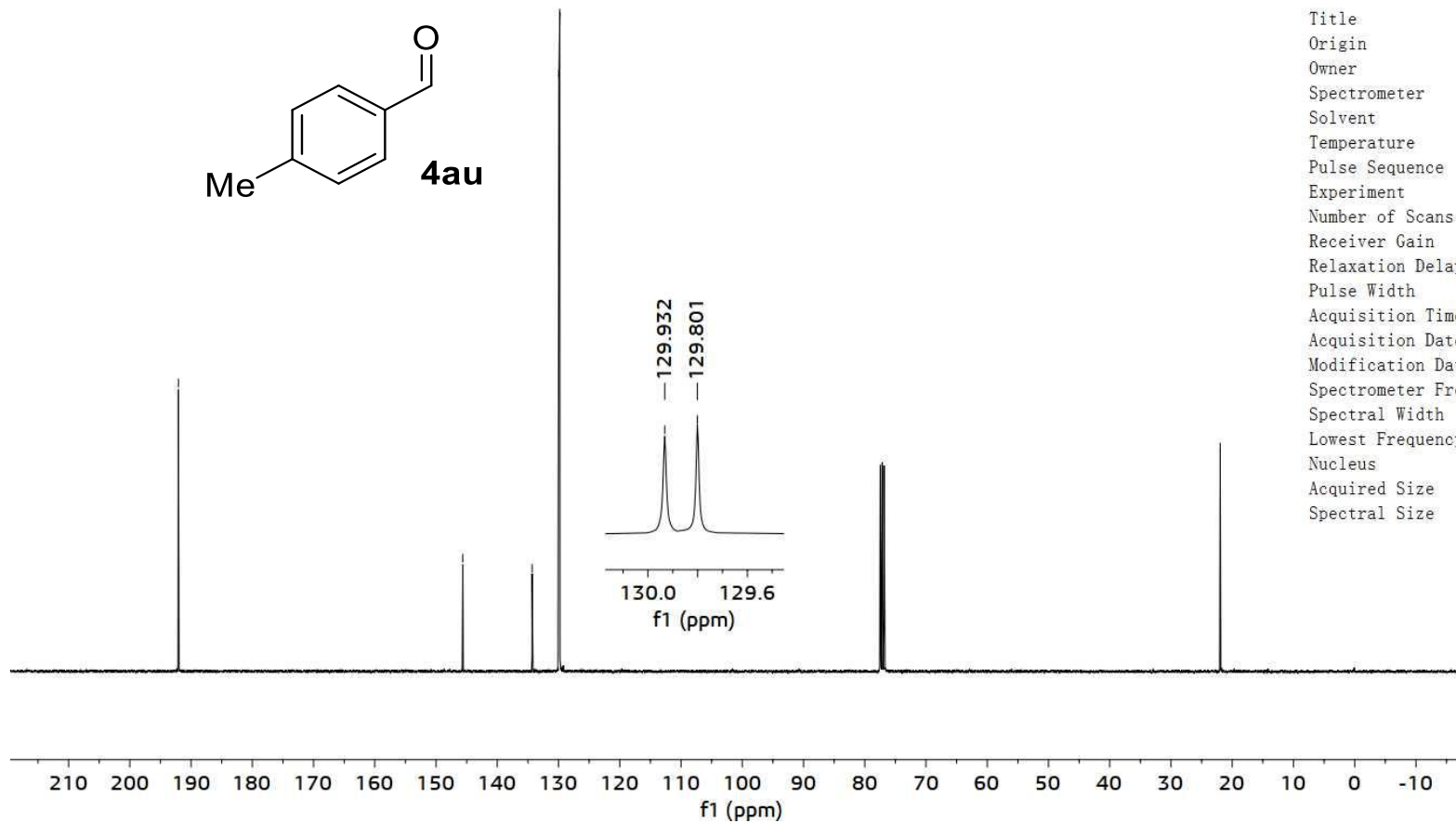
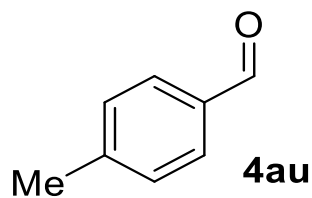
— 145.63

✓ 134.30

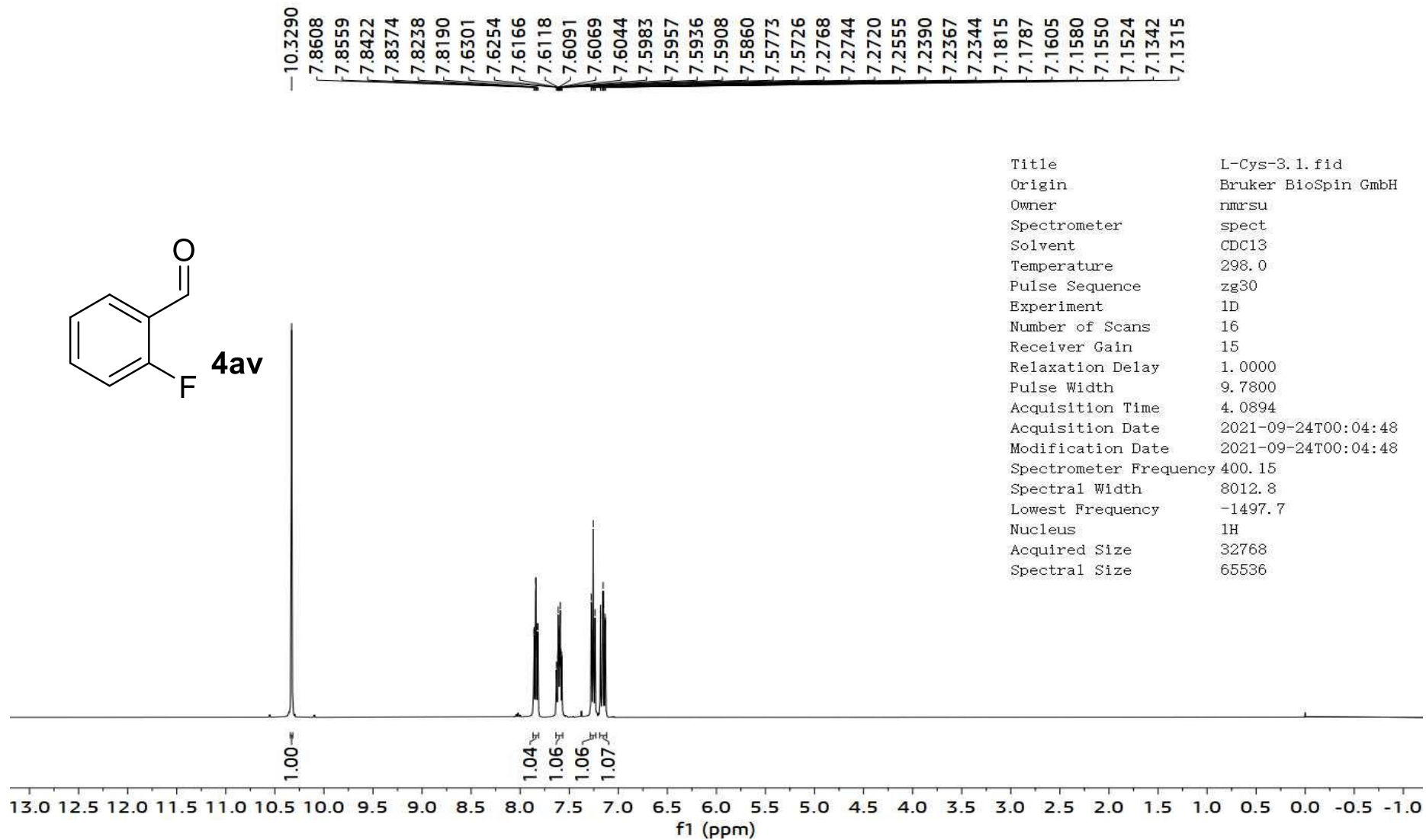
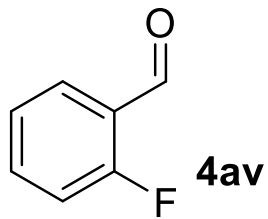
✓ 129.93

✓ 129.80

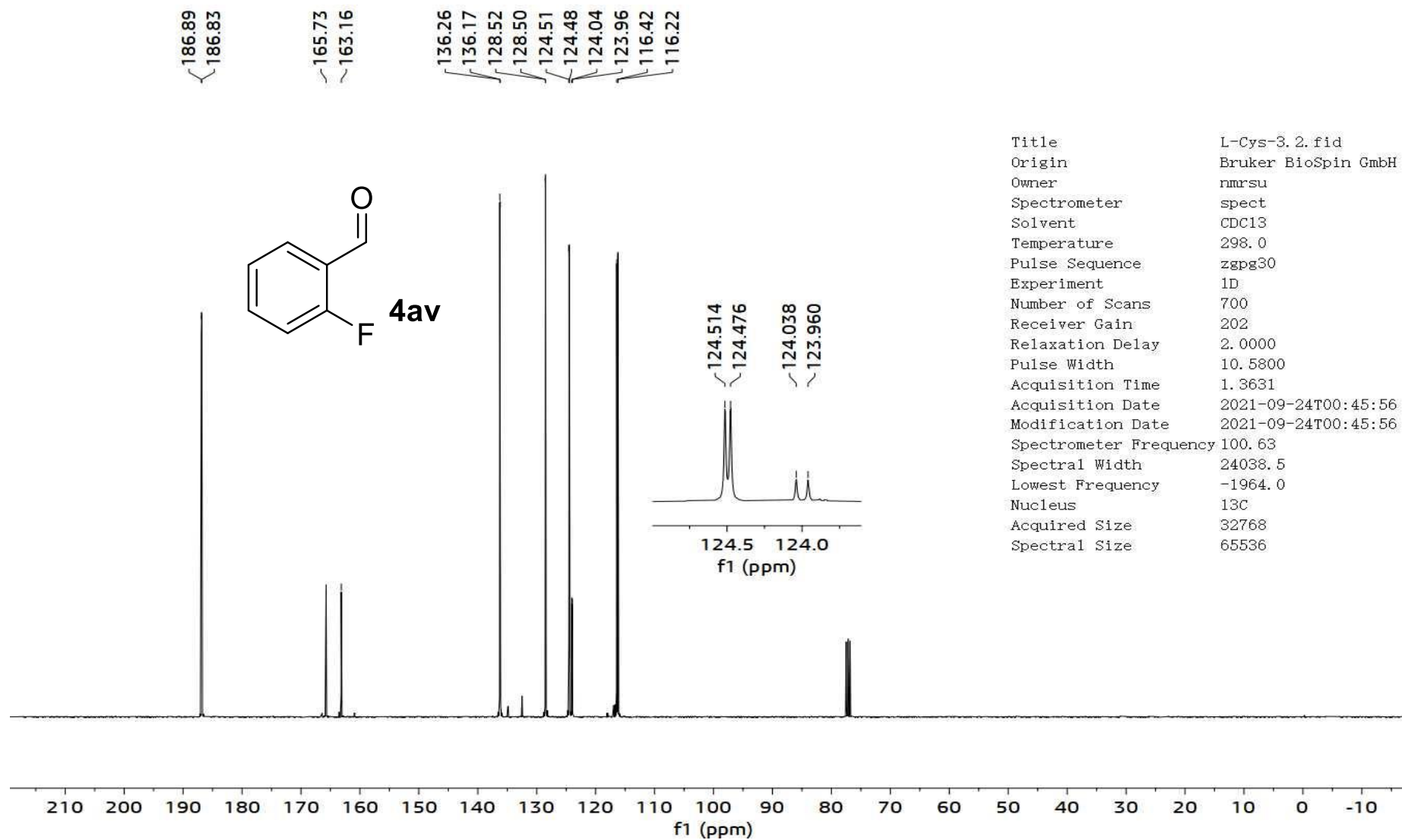
— 21.96

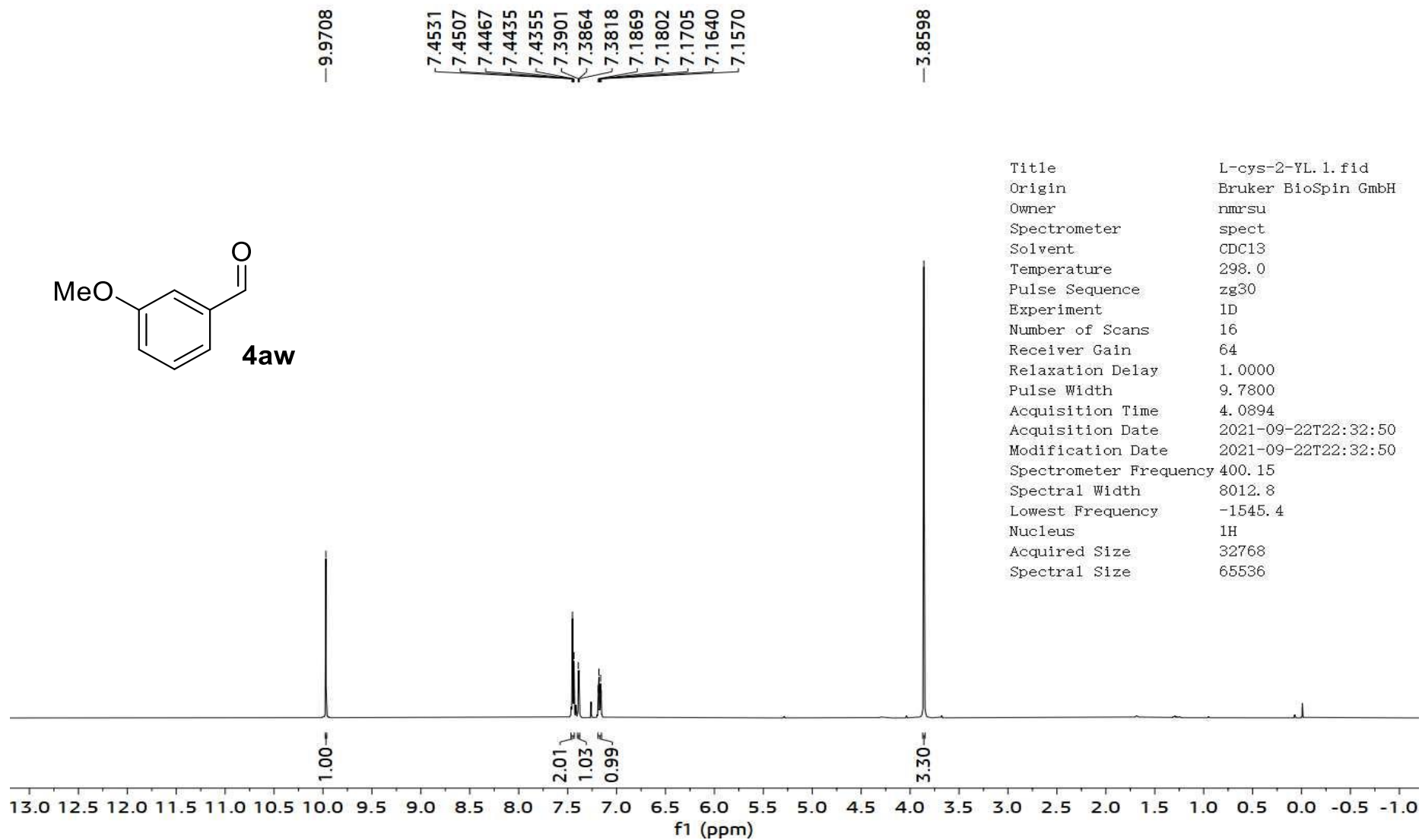
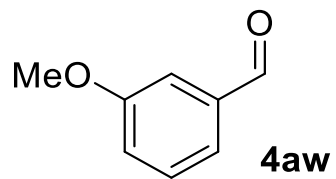


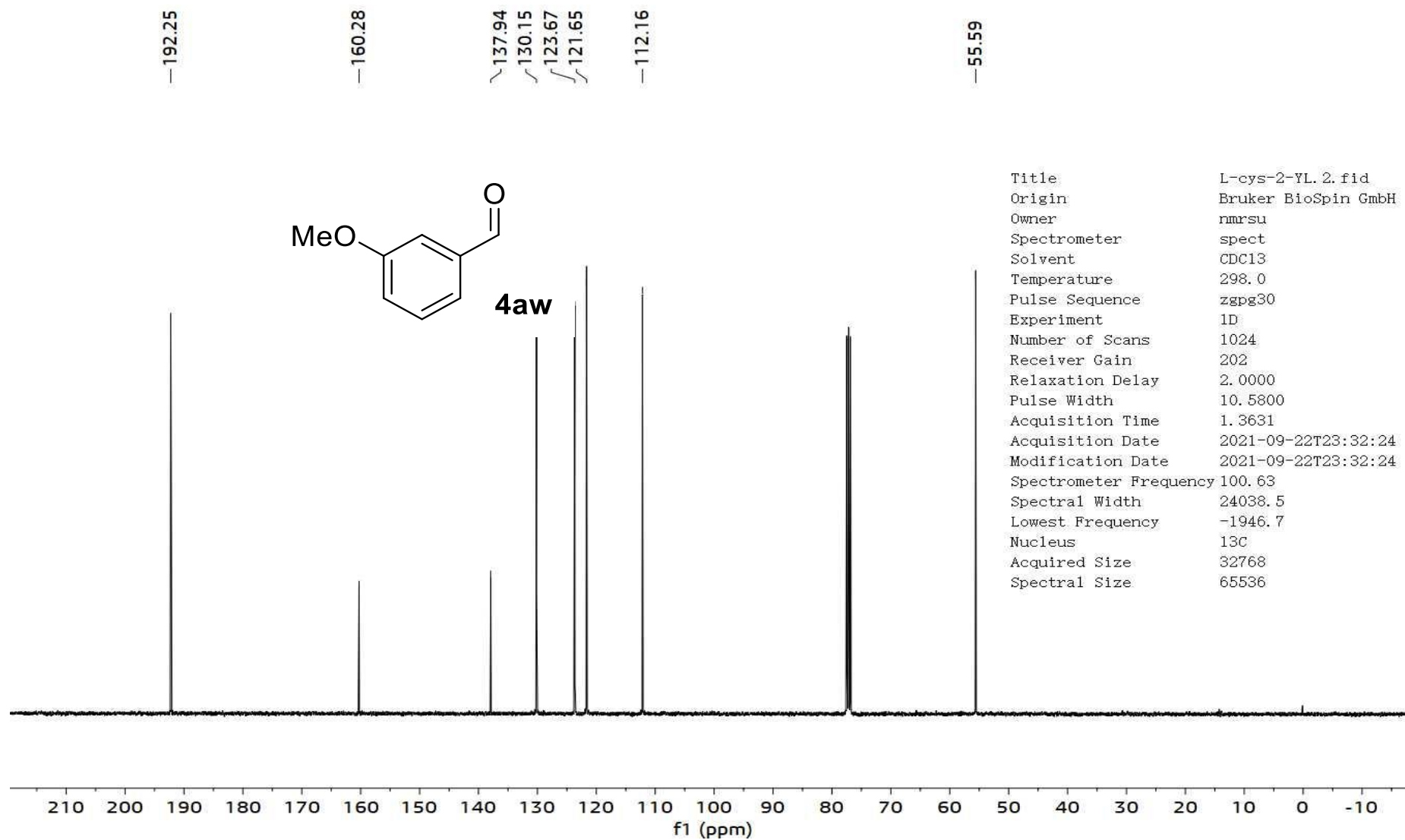
Title	L-cys-1.2.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-23T09:32:22
Modification Date	2021-09-23T09:32:22
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1949.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



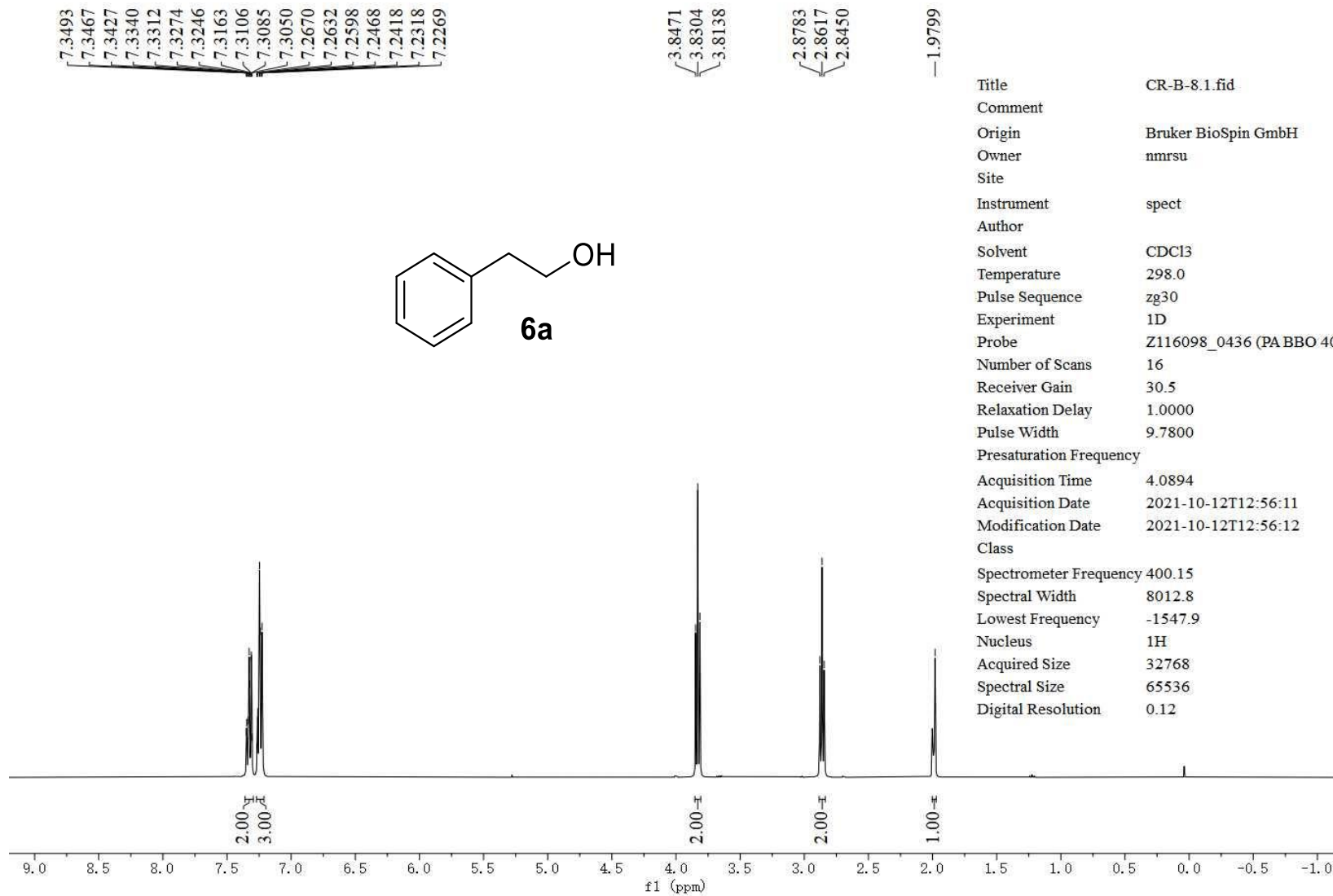
Title	L-Cys-3.1.fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	15
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-24T00:04:48
Modification Date	2021-09-24T00:04:48
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1497.7
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

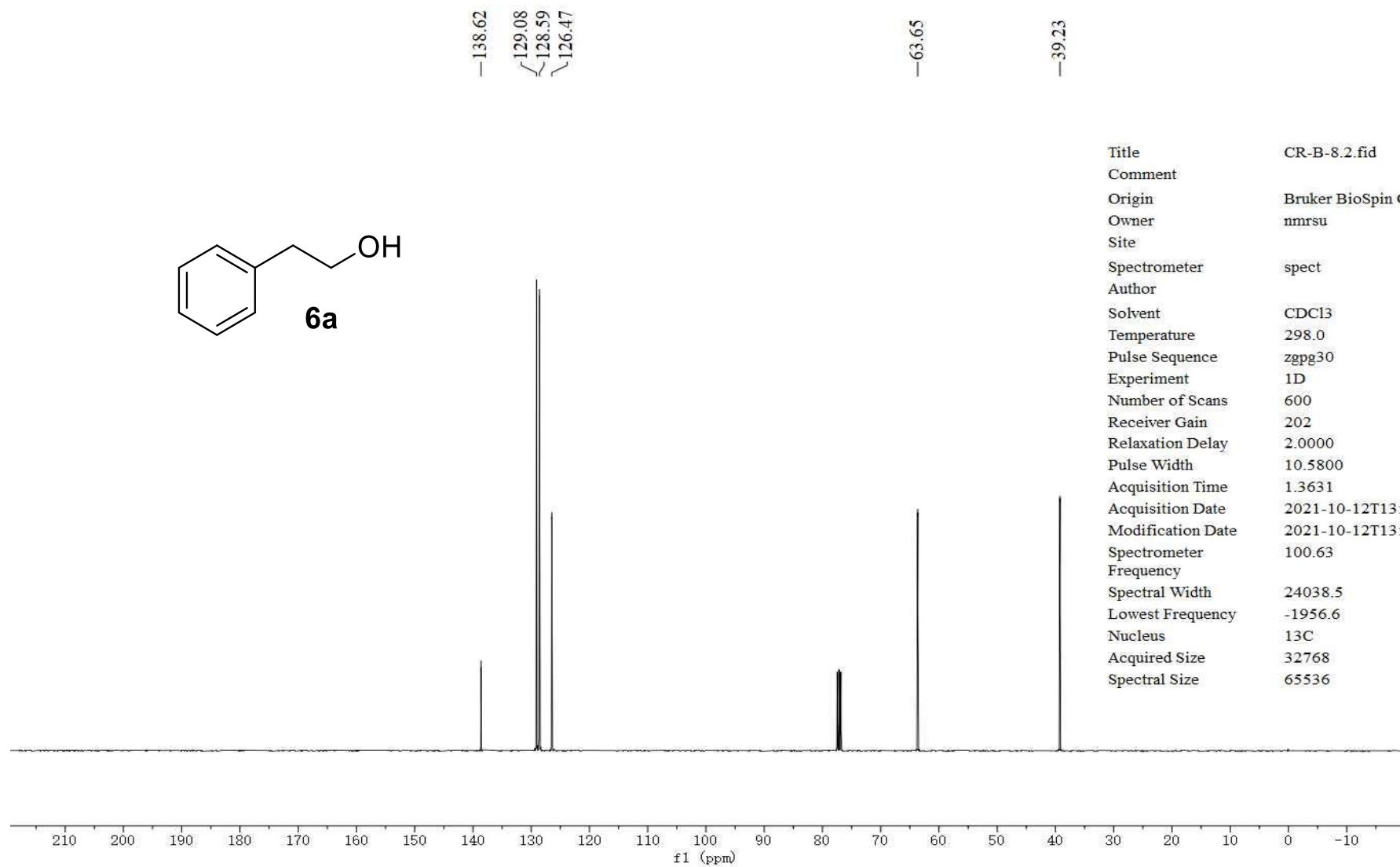
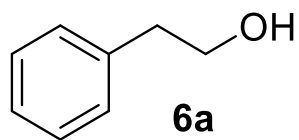




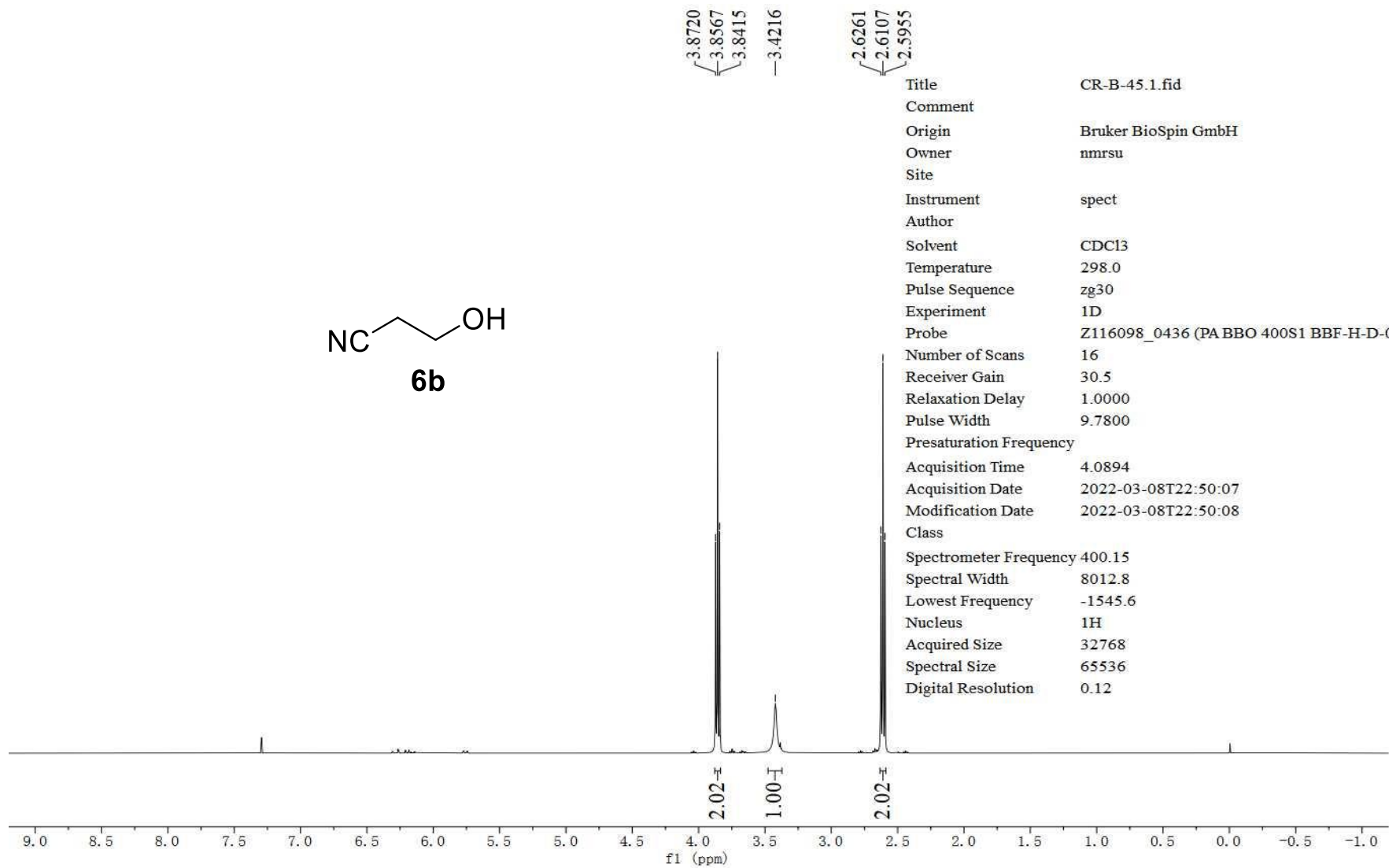
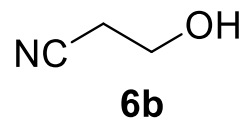


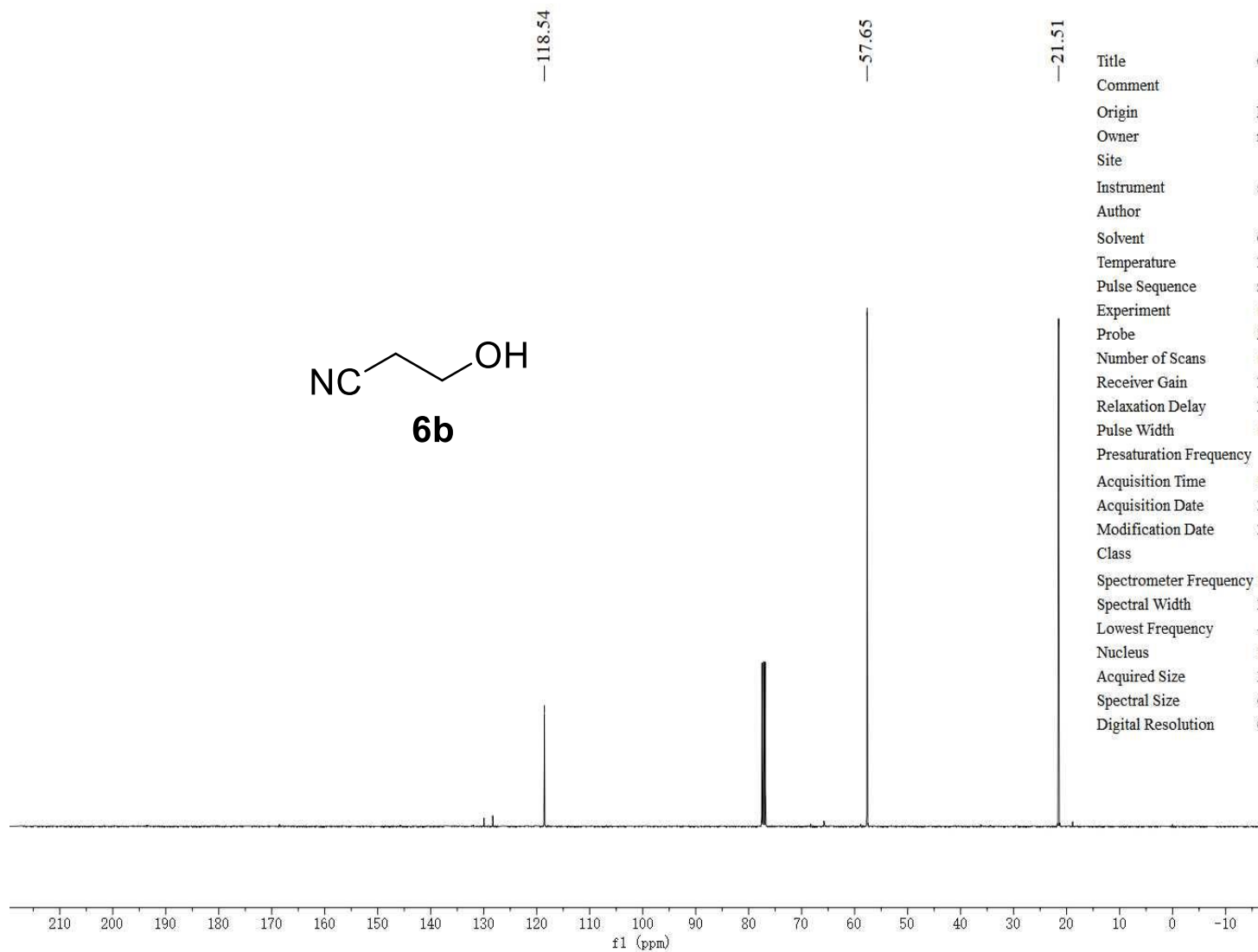
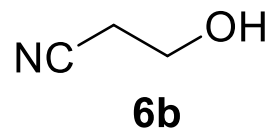
Title	L-cys-2-YL. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-22T23:32:24
Modification Date	2021-09-22T23:32:24
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1946.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



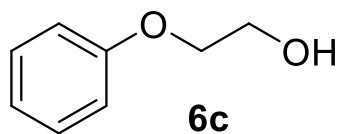


Title	CR-B-8.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T13:31:33
Modification Date	2021-10-12T13:31:34
Spectrometer	100.63
Frequency	
Spectral Width	24038.5
Lowest Frequency	-1956.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





Title	CR-B-45.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-08T23:49:46
Modification Date	2022-03-08T23:49:48
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1955.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37

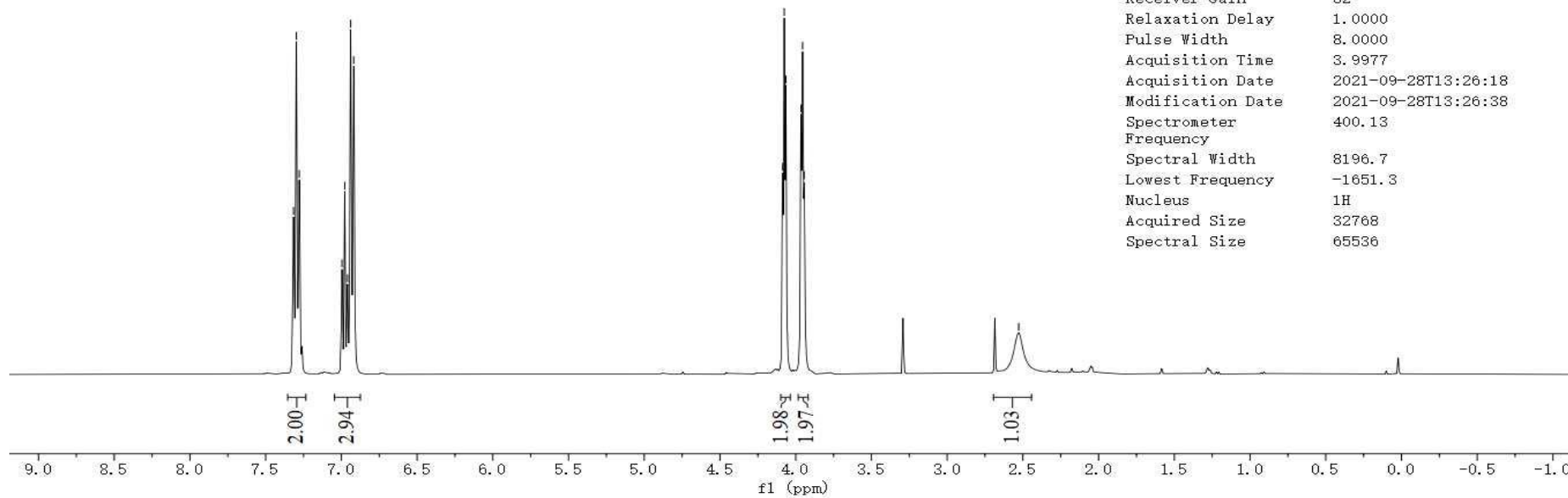


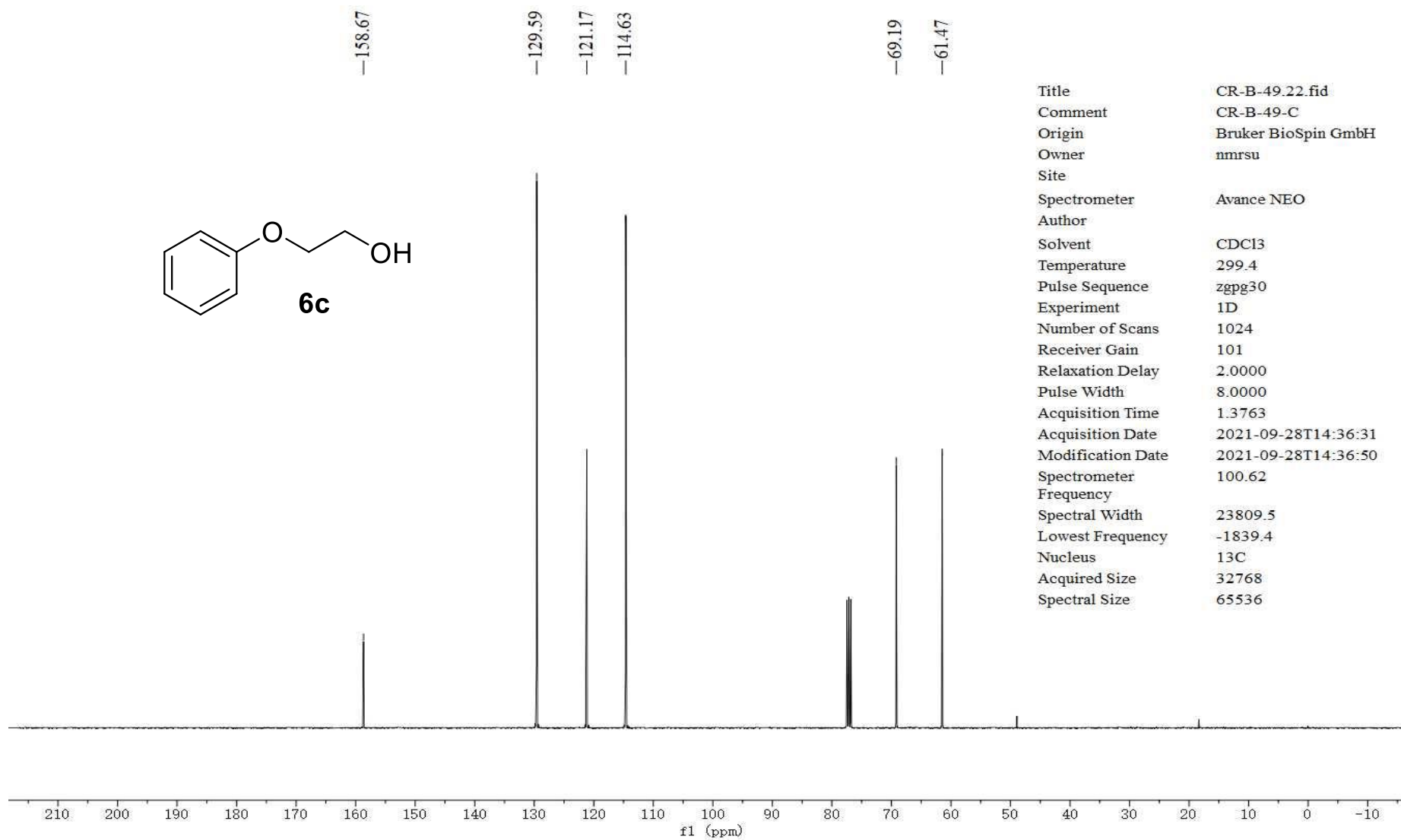
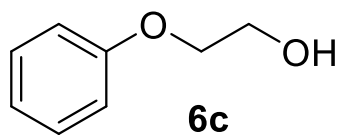
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6.9187

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4.0647
3.9649
3.9543
3.9439

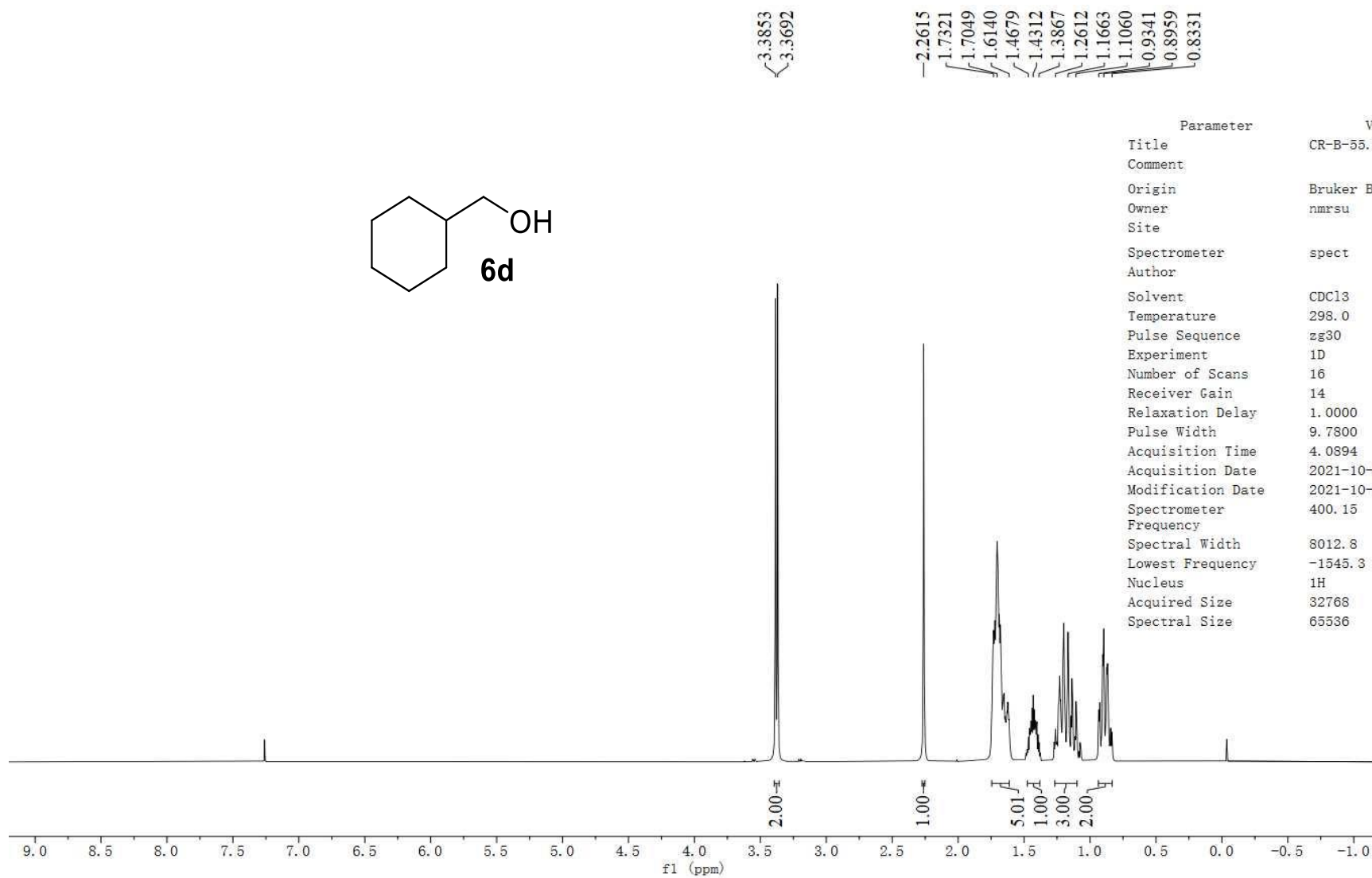
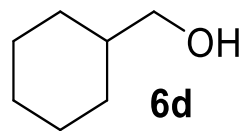
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Parameter	Value
Title	CR-B-49.21.fid
Comment	CR-B-49-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	298.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-28T13:26:18
Modification Date	2021-09-28T13:26:38
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1651.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

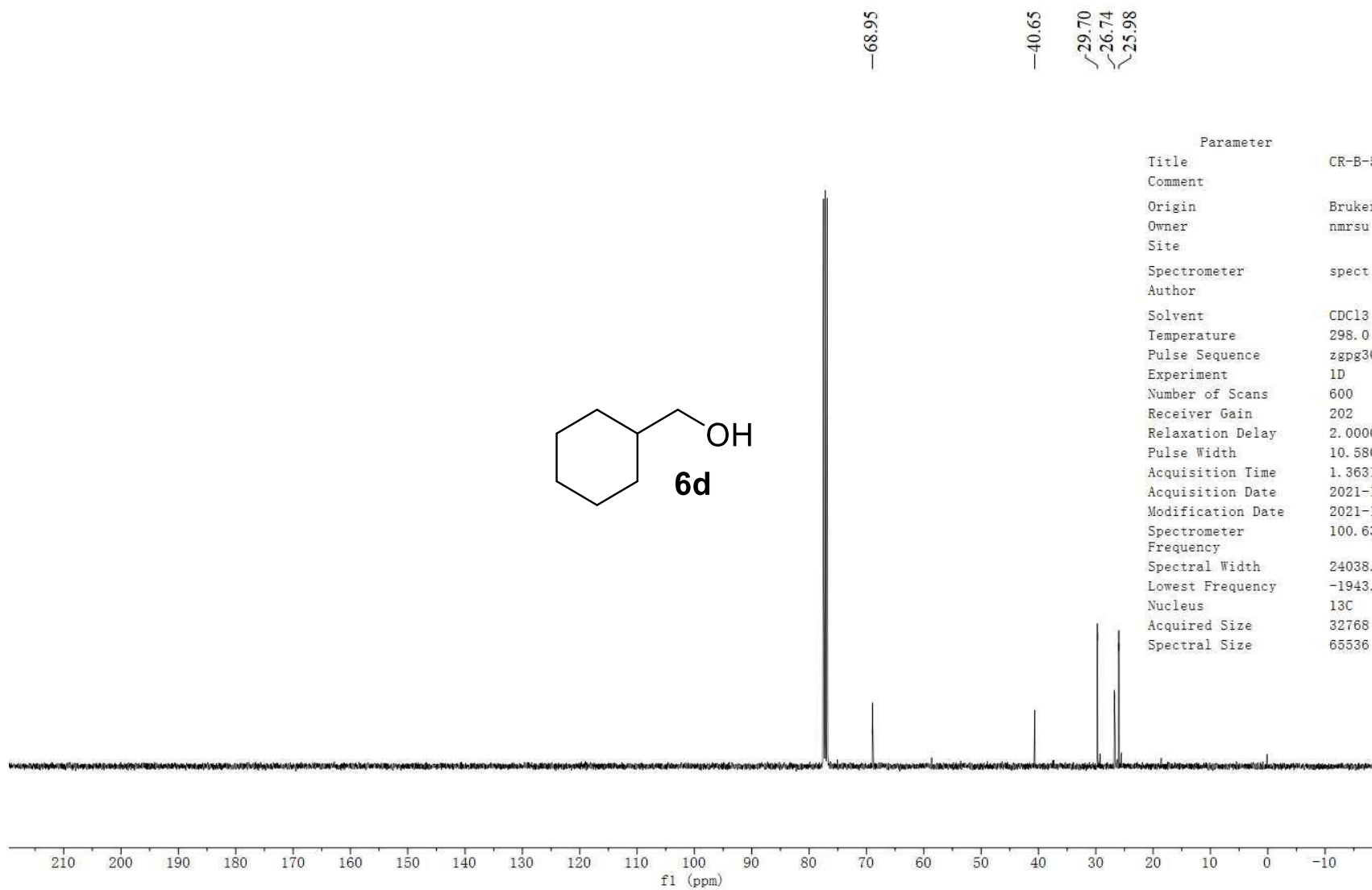
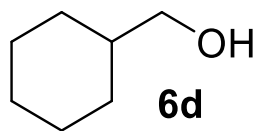




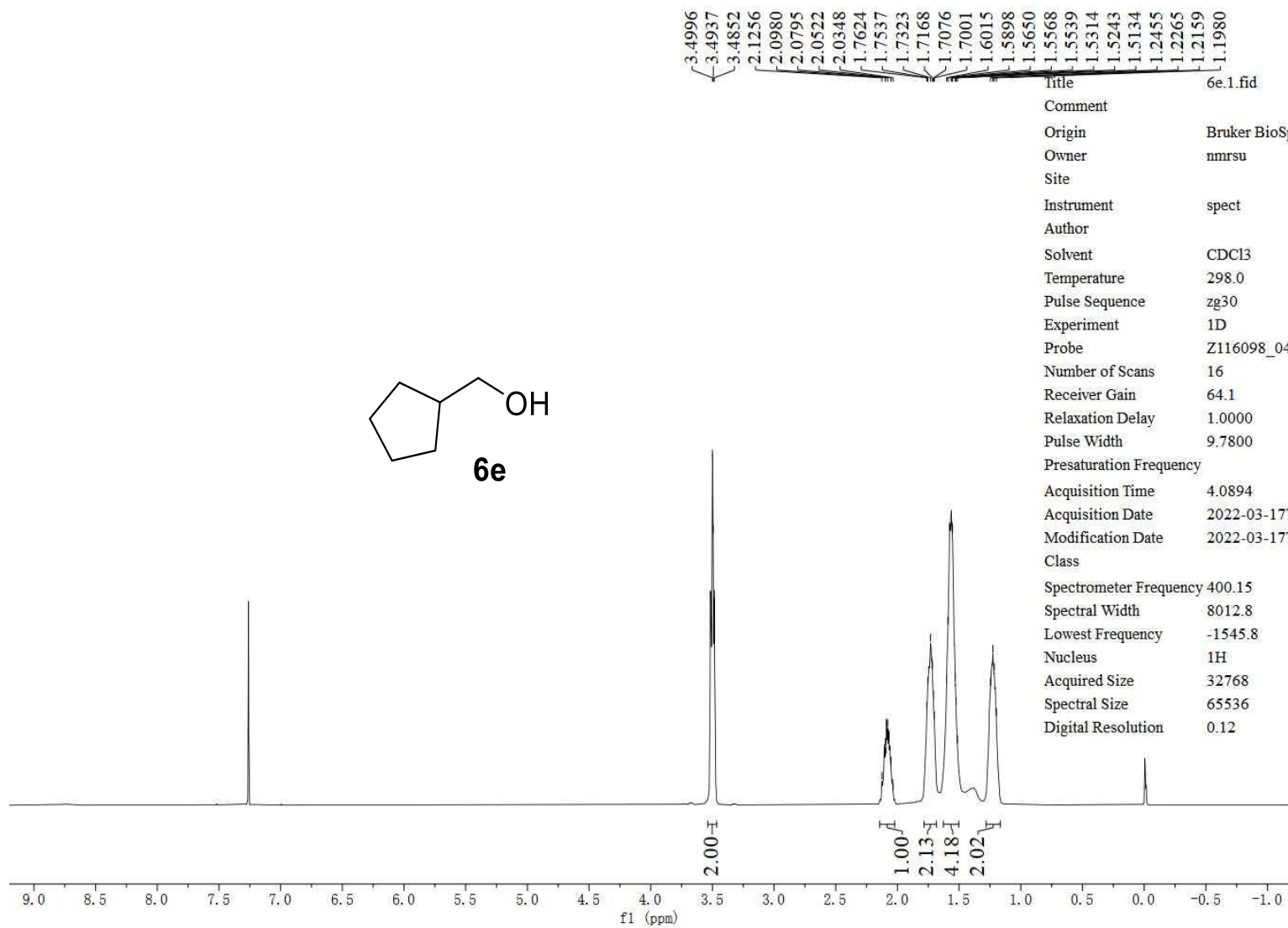
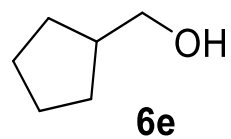
Title	CR-B-49.22.fid
Comment	CR-B-49-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-28T14:36:31
Modification Date	2021-09-28T14:36:50
Spectrometer	100.62
Frequency	
Spectral Width	23809.5
Lowest Frequency	-1839.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

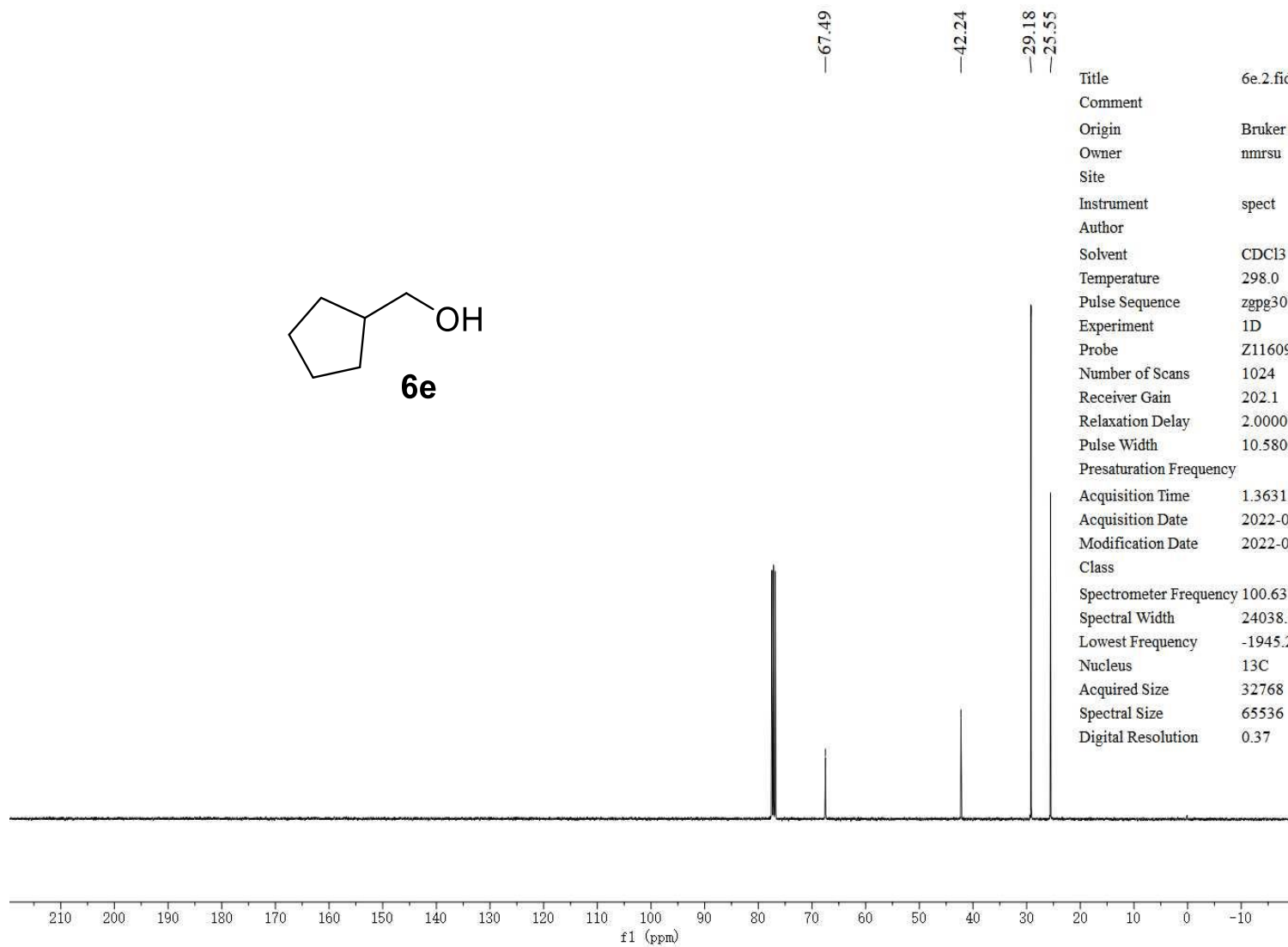
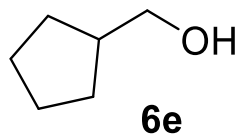


Parameter	Value
Title	CR-B-55.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	14
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-15T13:07:18
Modification Date	2021-10-15T13:07:20
Spectrometer	400.15
Frequency	
Spectral Width	8012.8
Lowest Frequency	-1545.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

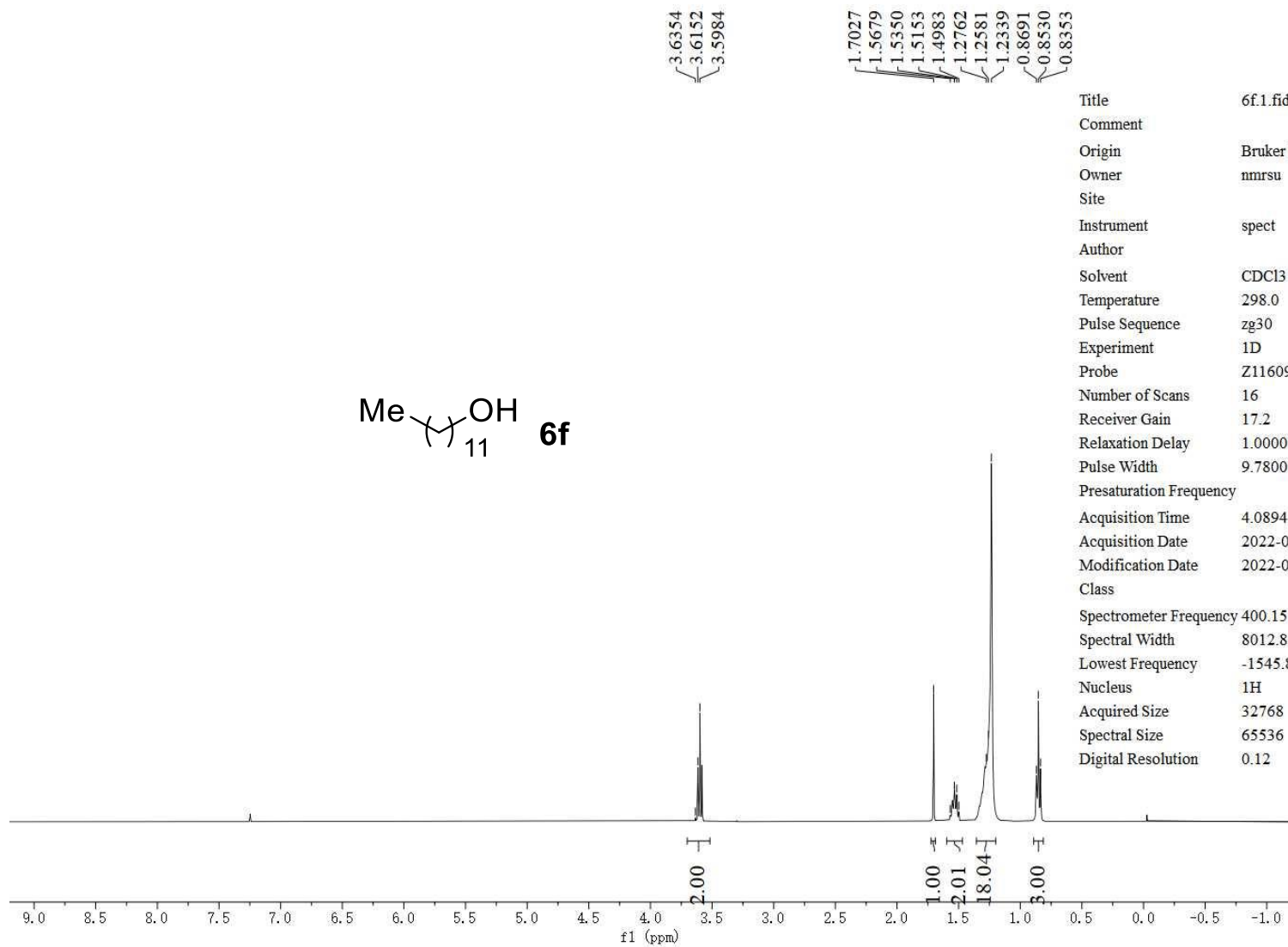
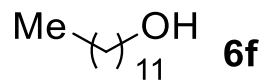


Parameter	Value
Title	CR-B-55.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T23:17:56
Modification Date	2021-10-12T23:17:58
Spectrometer	100.63
Frequency	
Spectral Width	24038.5
Lowest Frequency	-1943.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

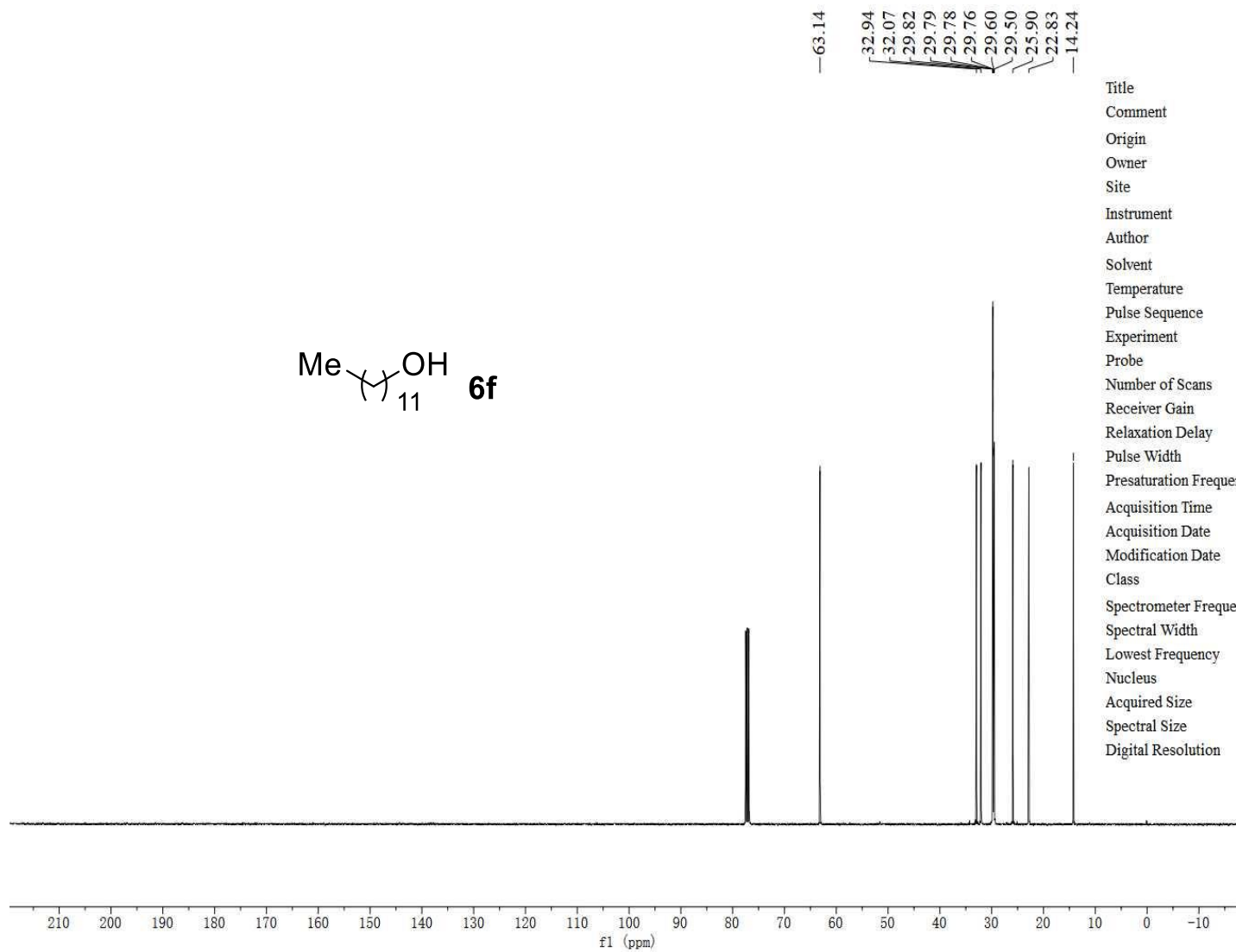
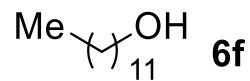




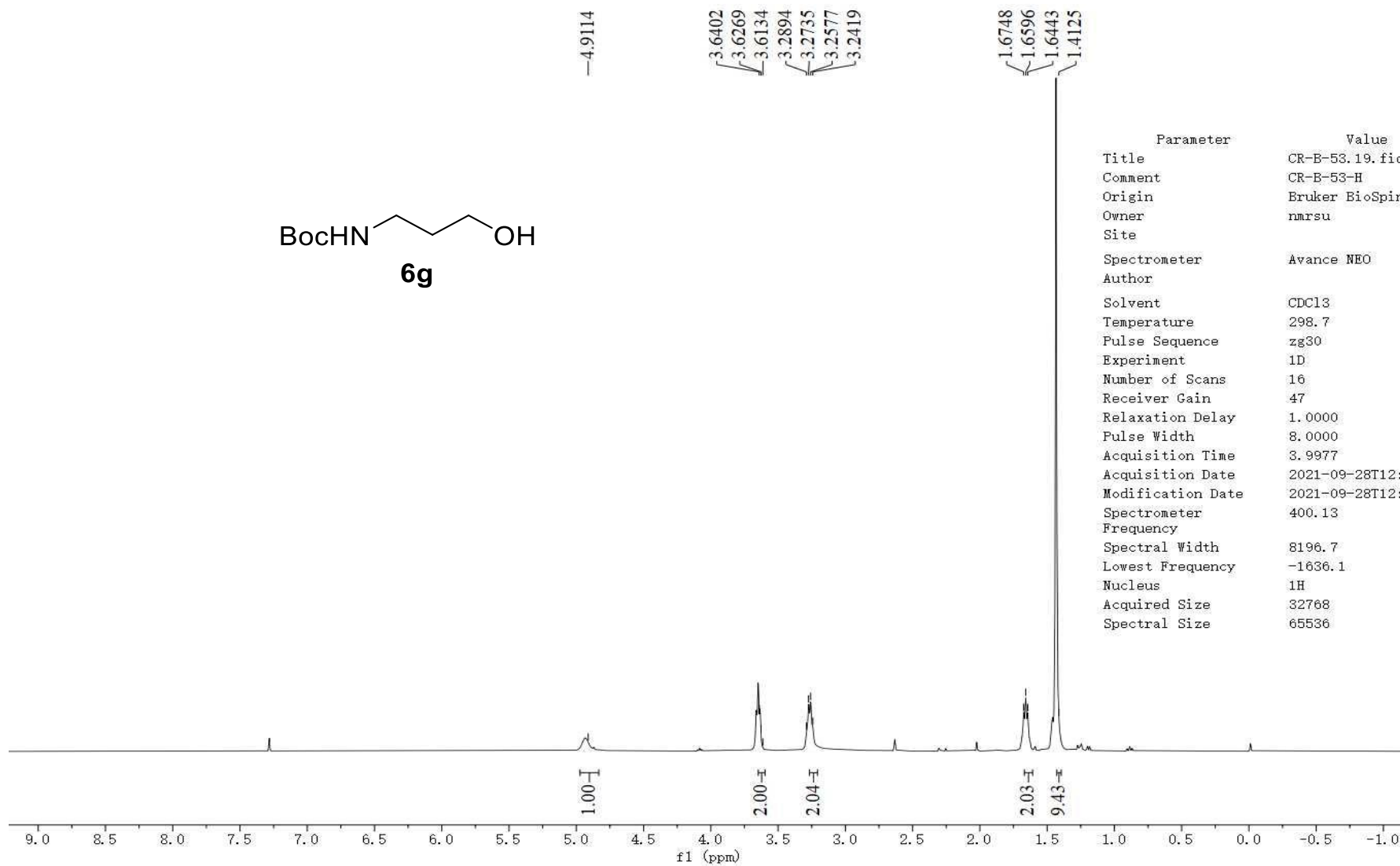
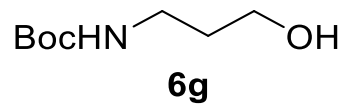
Title	6e.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-18T04:25:38
Modification Date	2022-03-18T04:25:38
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1945.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37



Title	6f.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	17.2
Relaxation Delay	1.0000
Pulse Width	9.7800
Presaturation Frequency	
Acquisition Time	4.0894
Acquisition Date	2022-03-17T11:56:31
Modification Date	2022-03-17T11:56:30
Class	
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12



Title	6f.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-17T21:38:27
Modification Date	2022-03-17T21:38:26
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1944.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37

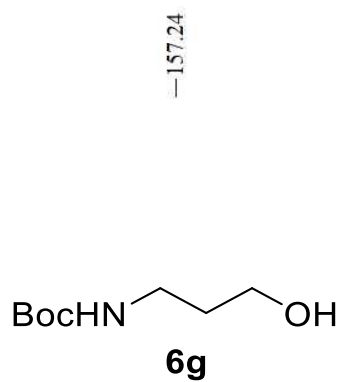


4.9114

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3.6134
3.2894
3.2735
3.2577
3.2419

1.6748
1.6596
1.6443
1.4125

Parameter	Value
Title	CR-B-53.19.fid
Comment	CR-B-53-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl ₃
Temperature	298.7
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	47
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-28T12:21:36
Modification Date	2021-09-28T12:21:56
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.1
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

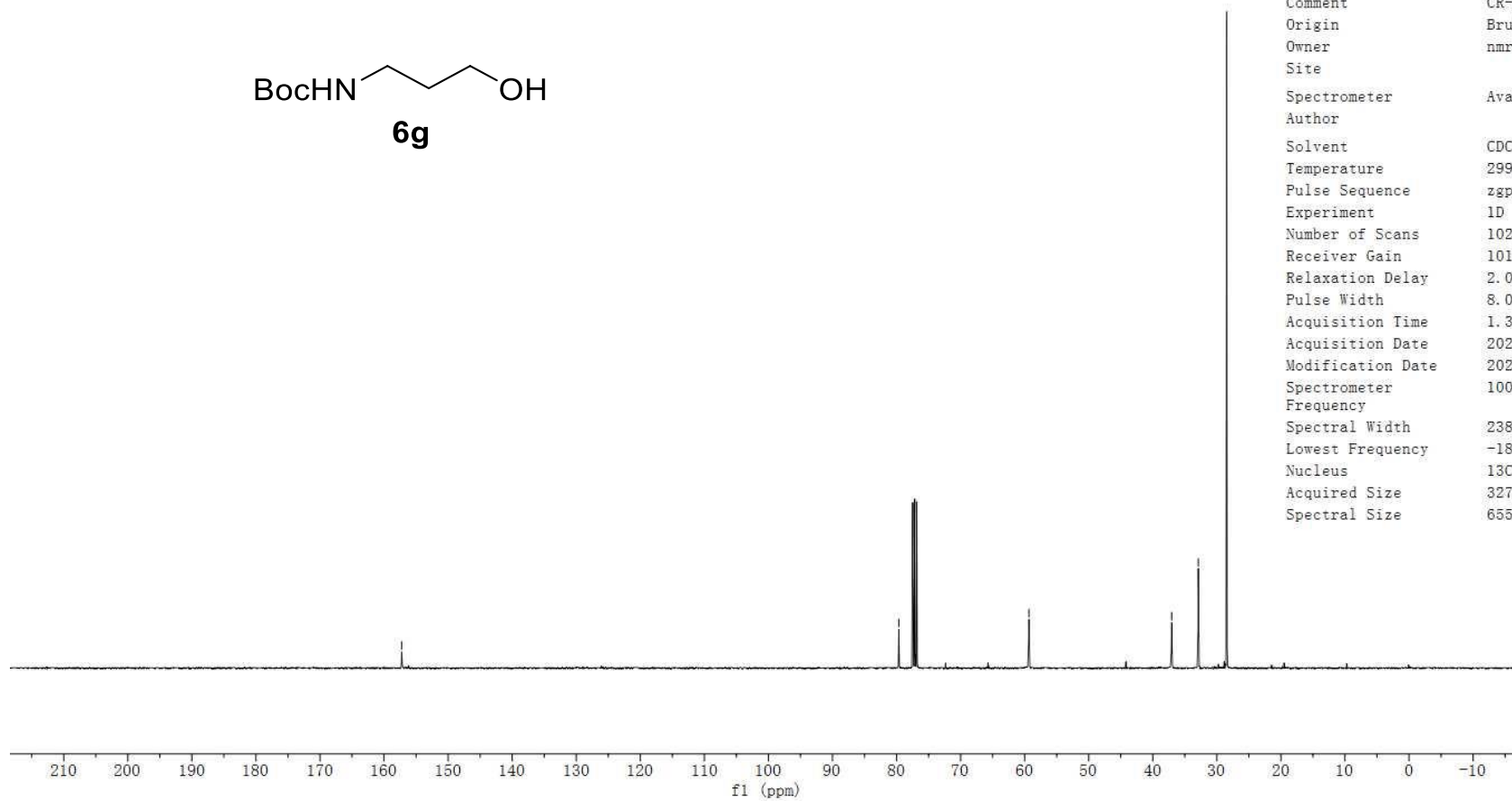


-157.24

-79.64

-59.34

~37.05
 ~32.90
 ~28.46



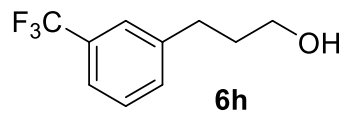
Parameter	Value
Title	CR-B-53.20.fid
Comment	CR-B-53-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-28T13:20:57
Modification Date	2021-09-28T13:21:16
Spectrometer	100.62
Frequency	
Spectral Width	23809.5
Lowest Frequency	-1832.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

7.4547
7.3948
7.3861

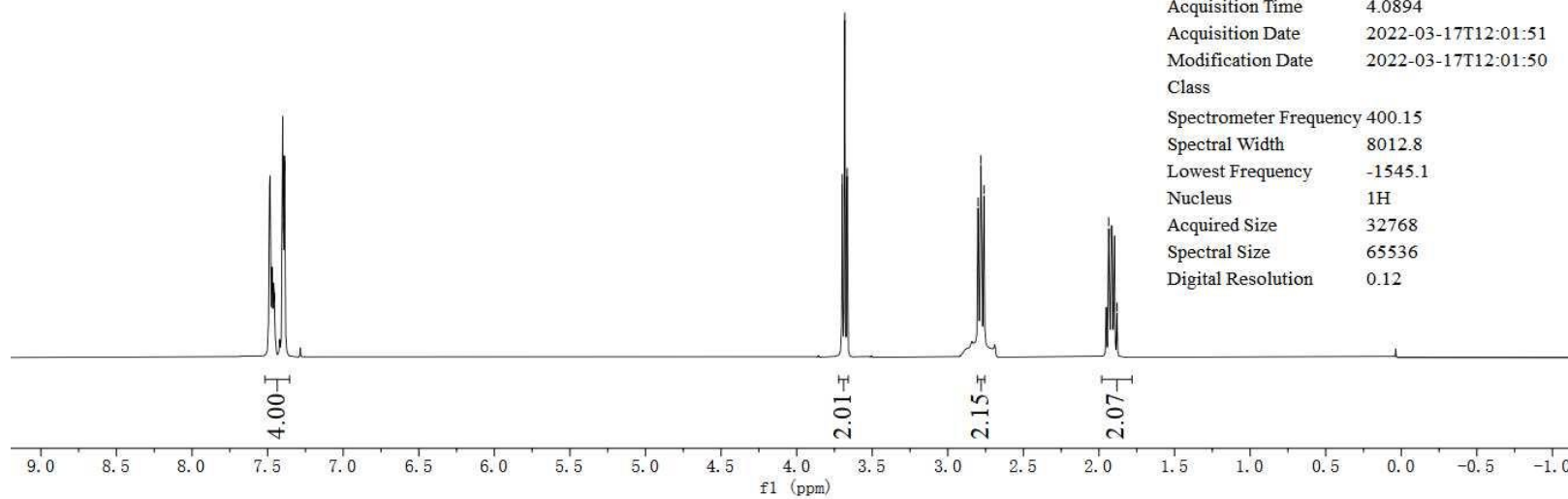
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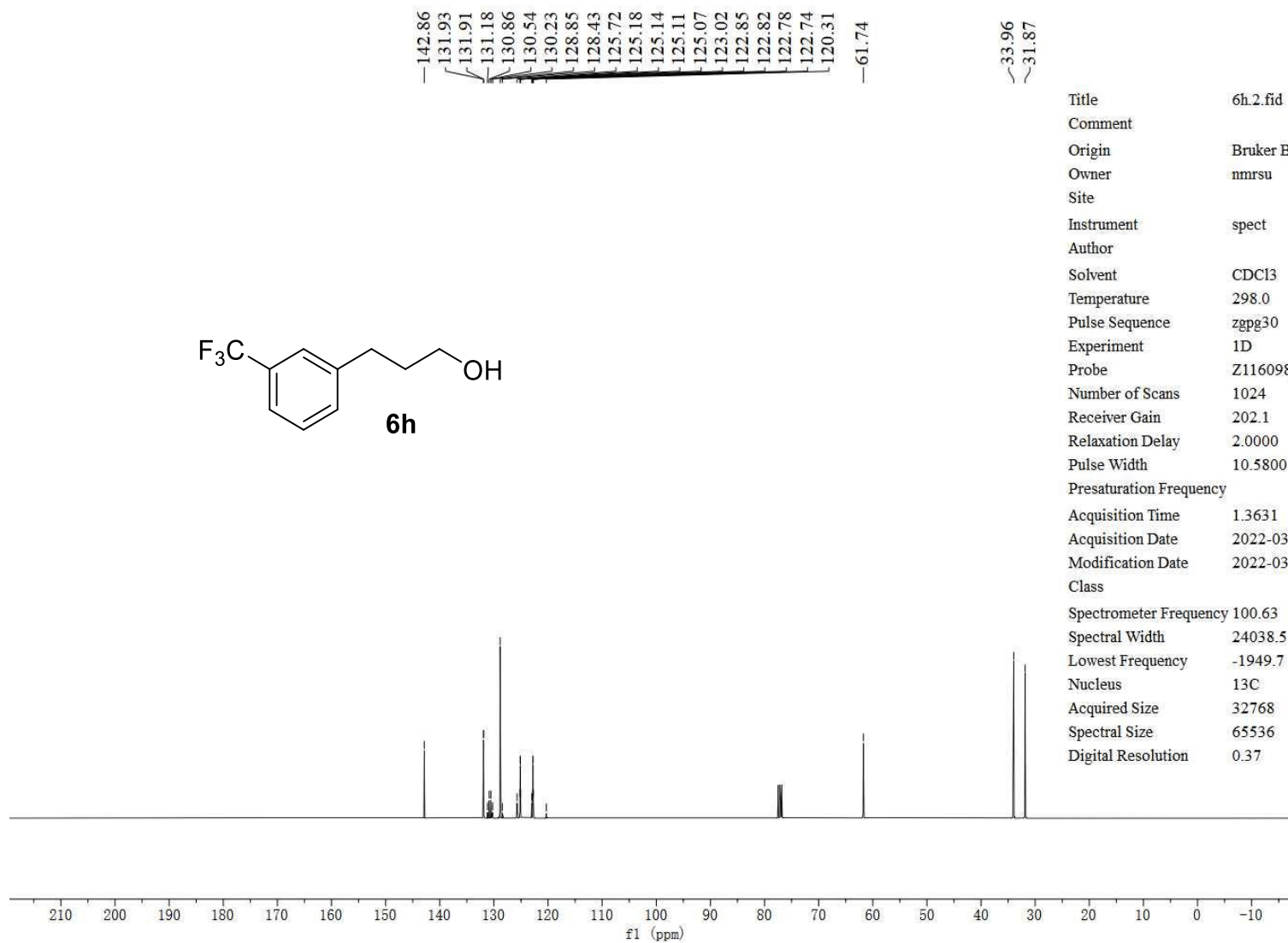
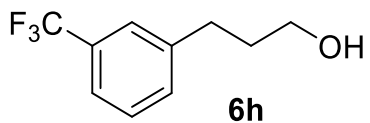
2.8001
2.7812
2.7610

1.9359
1.9200
1.9044
1.8809



Title	6h.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	16
Receiver Gain	13.8
Relaxation Delay	1.0000
Pulse Width	9.7800
Presaturation Frequency	
Acquisition Time	4.0894
Acquisition Date	2022-03-17T12:01:51
Modification Date	2022-03-17T12:01:50
Class	
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.1
Nucleus	1H
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.12

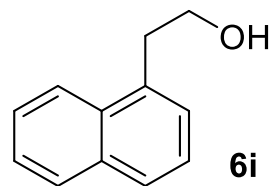




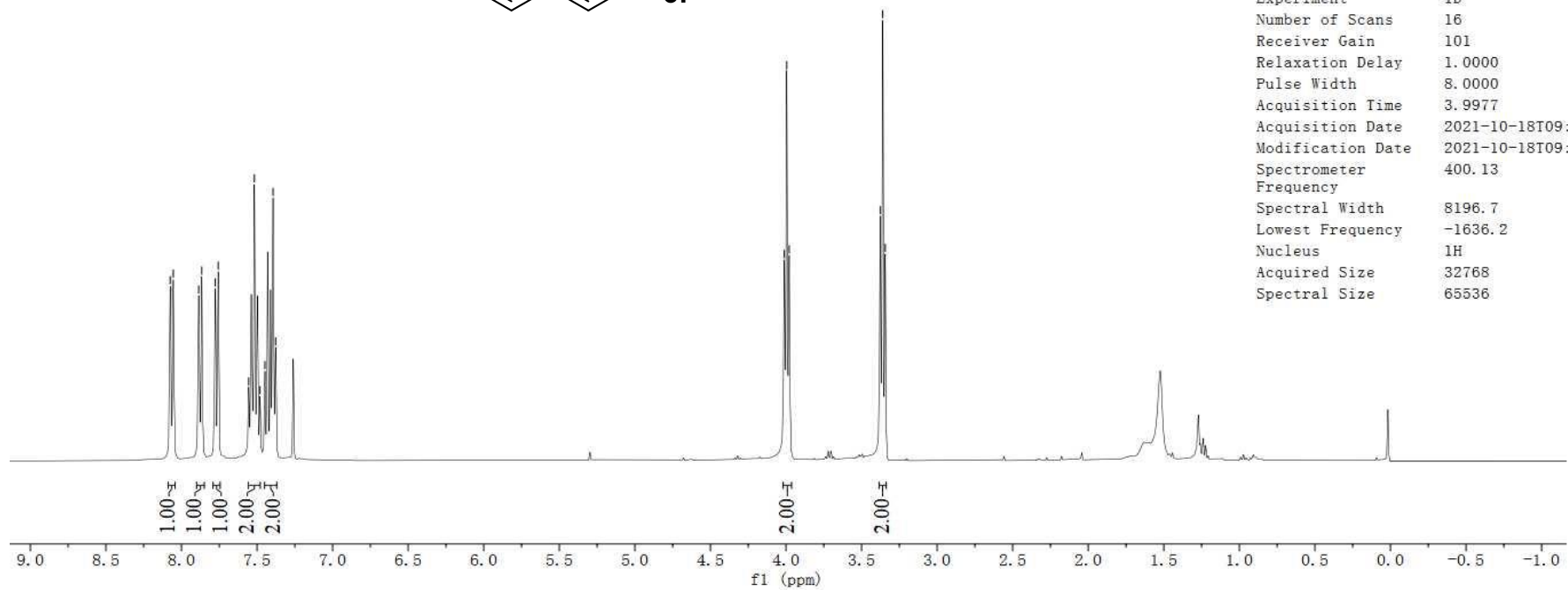
Title	6h.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Instrument	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Probe	Z116098_0436 (PA BBO 400S1 BBF-H-D-05 Z SP)
Number of Scans	1024
Receiver Gain	202.1
Relaxation Delay	2.0000
Pulse Width	10.5800
Presaturation Frequency	
Acquisition Time	1.3631
Acquisition Date	2022-03-17T22:59:55
Modification Date	2022-03-17T22:59:54
Class	
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1949.7
Nucleus	13C
Acquired Size	32768
Spectral Size	65536
Digital Resolution	0.37

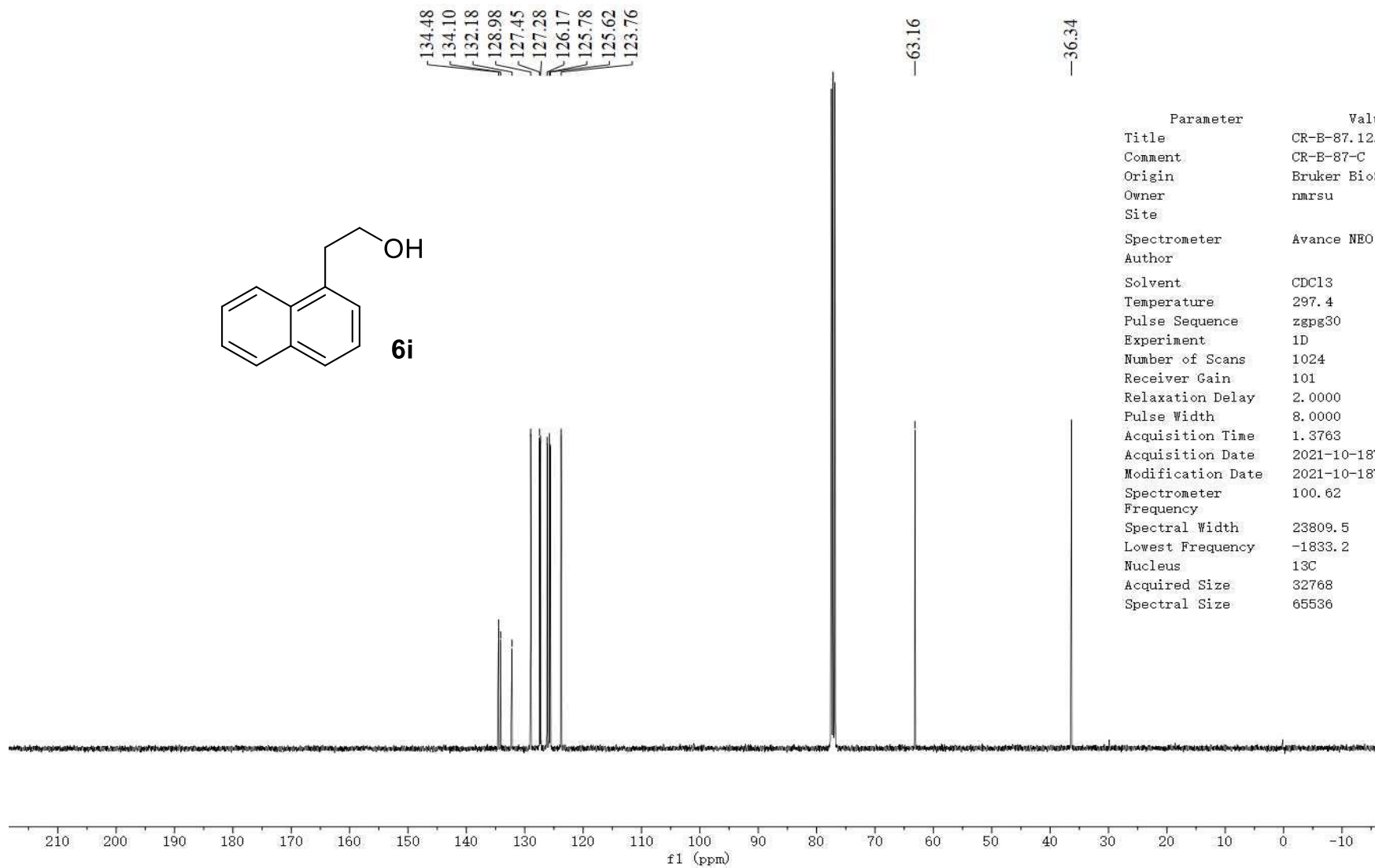
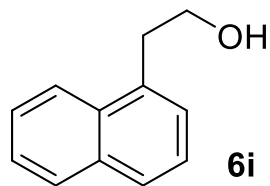
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7.3766

4.0116
3.9950
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3.3597
3.3429



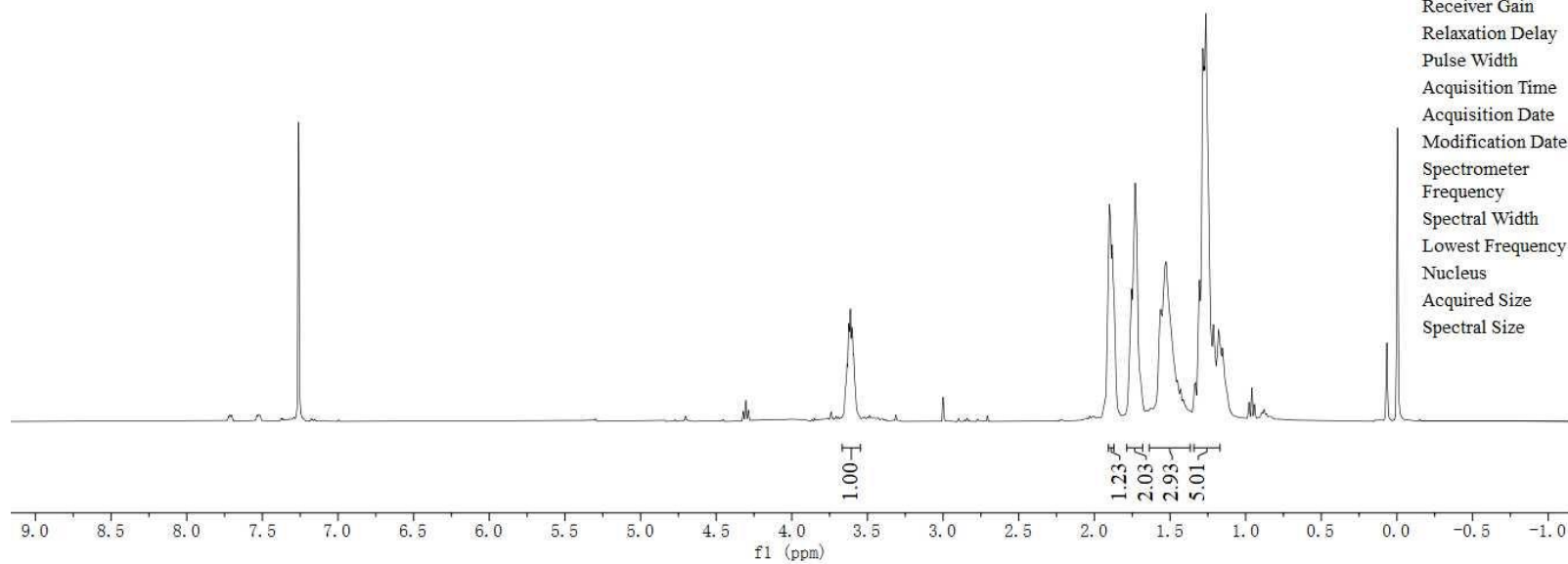
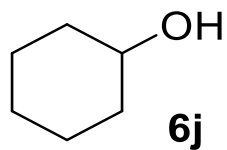
Parameter	Value
Title	CR-B-87.11.fid
Comment	CR-B-87-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	296.7
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	101
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-10-18T09:44:04
Modification Date	2021-10-18T09:43:24
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



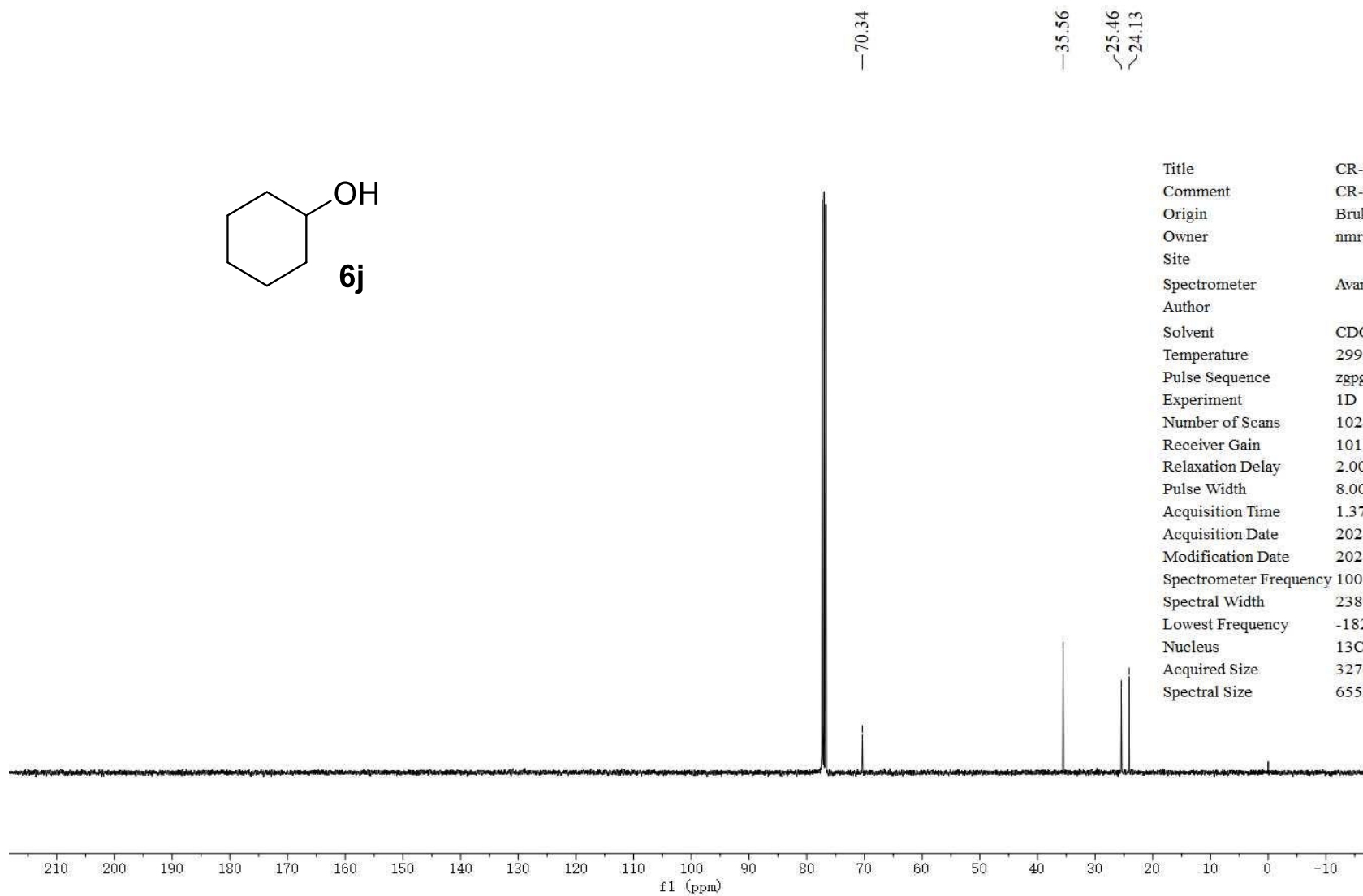
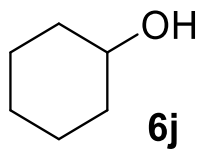


Parameter	Value
Title	CR-B-87.12.fid
Comment	CR-B-87-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	297.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-10-18T10:43:15
Modification Date	2021-10-18T10:42:36
Spectrometer	100.62
Frequency	
Spectral Width	23809.5
Lowest Frequency	-1833.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

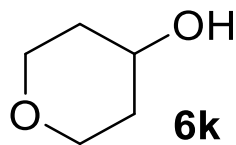
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 1.7637
 1.7542
 1.7416
 1.7304
 1.7197
 1.6950
 1.5735
 1.5622
 1.5456
 1.5339
 1.5240
 1.5070
 1.4869
 1.4684
 1.4496
 1.3117
 1.3045
 1.2838
 1.2737
 1.2625
 1.2517
 1.2429
 1.2285
 1.2125
 1.1836
 1.1783
 1.1648
 1.1521
 1.1436



Title	CR-B-12.29.fid
Comment	CR-B-12-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	298.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	101
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-28T18:01:26
Modification Date	2021-09-28T18:01:46
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

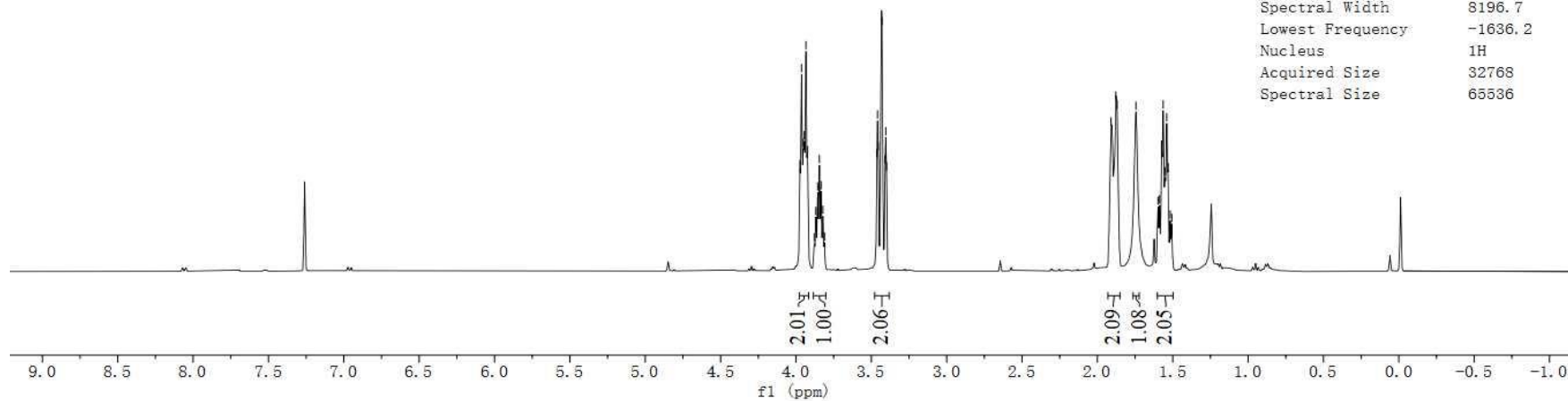


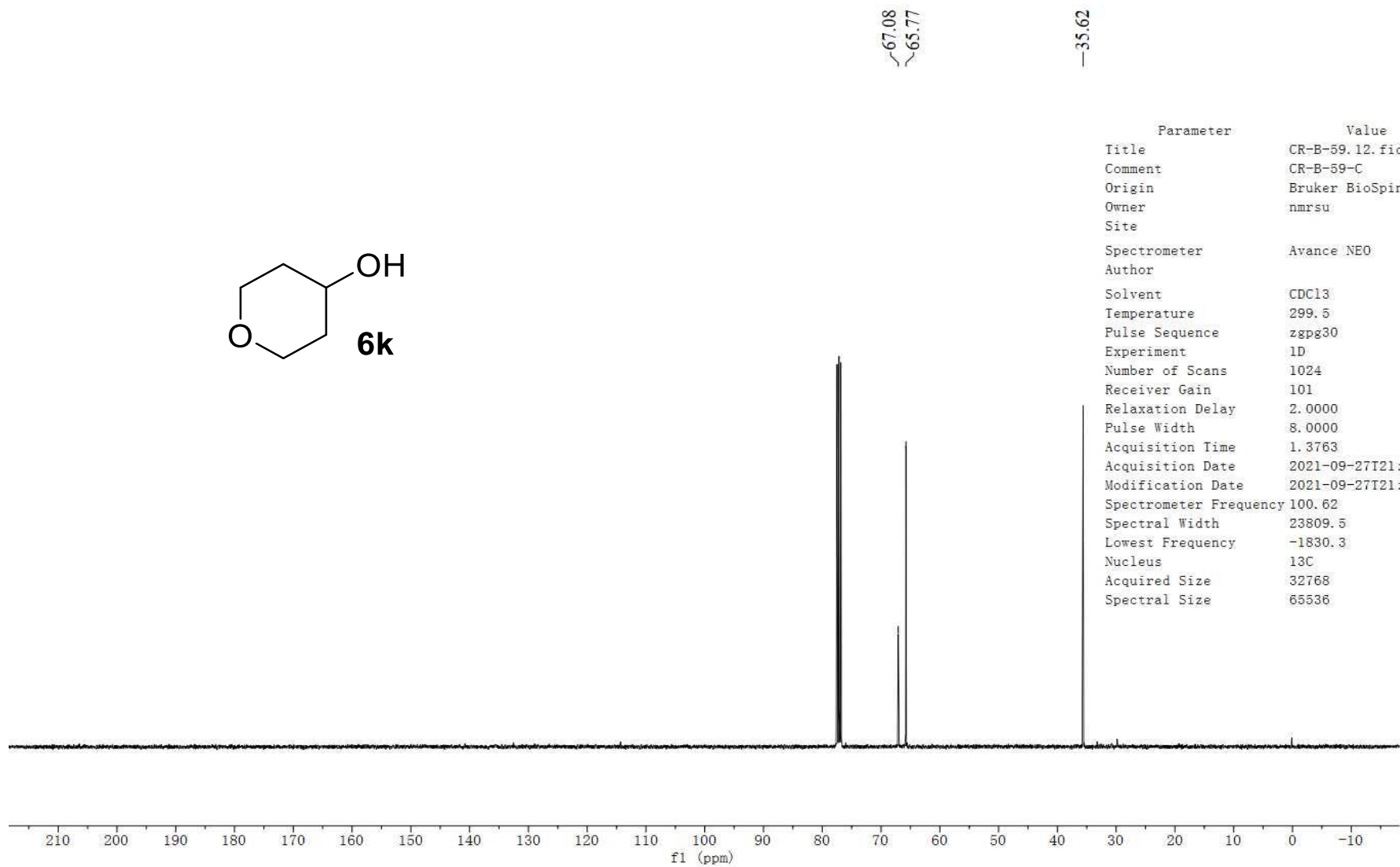
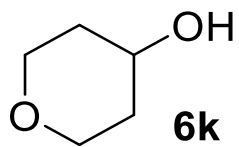
Title	CR-B-12.30.fid
Comment	CR-B-12-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-28T19:00:46
Modification Date	2021-09-28T19:01:06
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1828.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



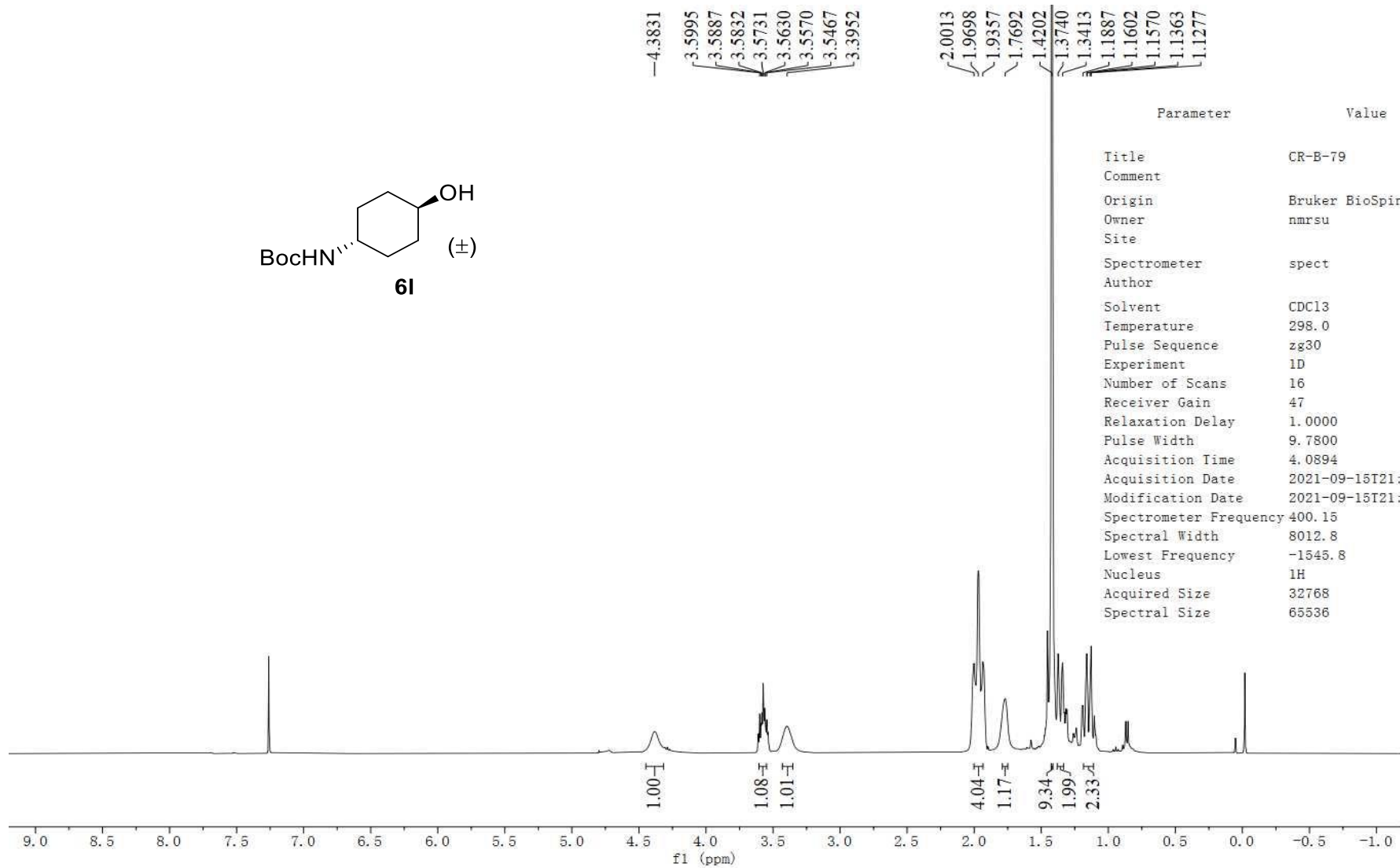
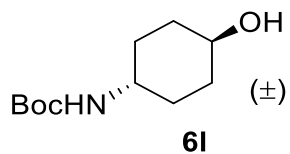
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3.8680
3.8561
3.8446
3.8333
3.8213
3.8105
3.4636
3.4588
3.4539
3.4342
3.4287
3.4089
3.4039
3.3991
1.9093
1.8780
1.8689
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1.5739
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1.5302
1.5167
1.5062

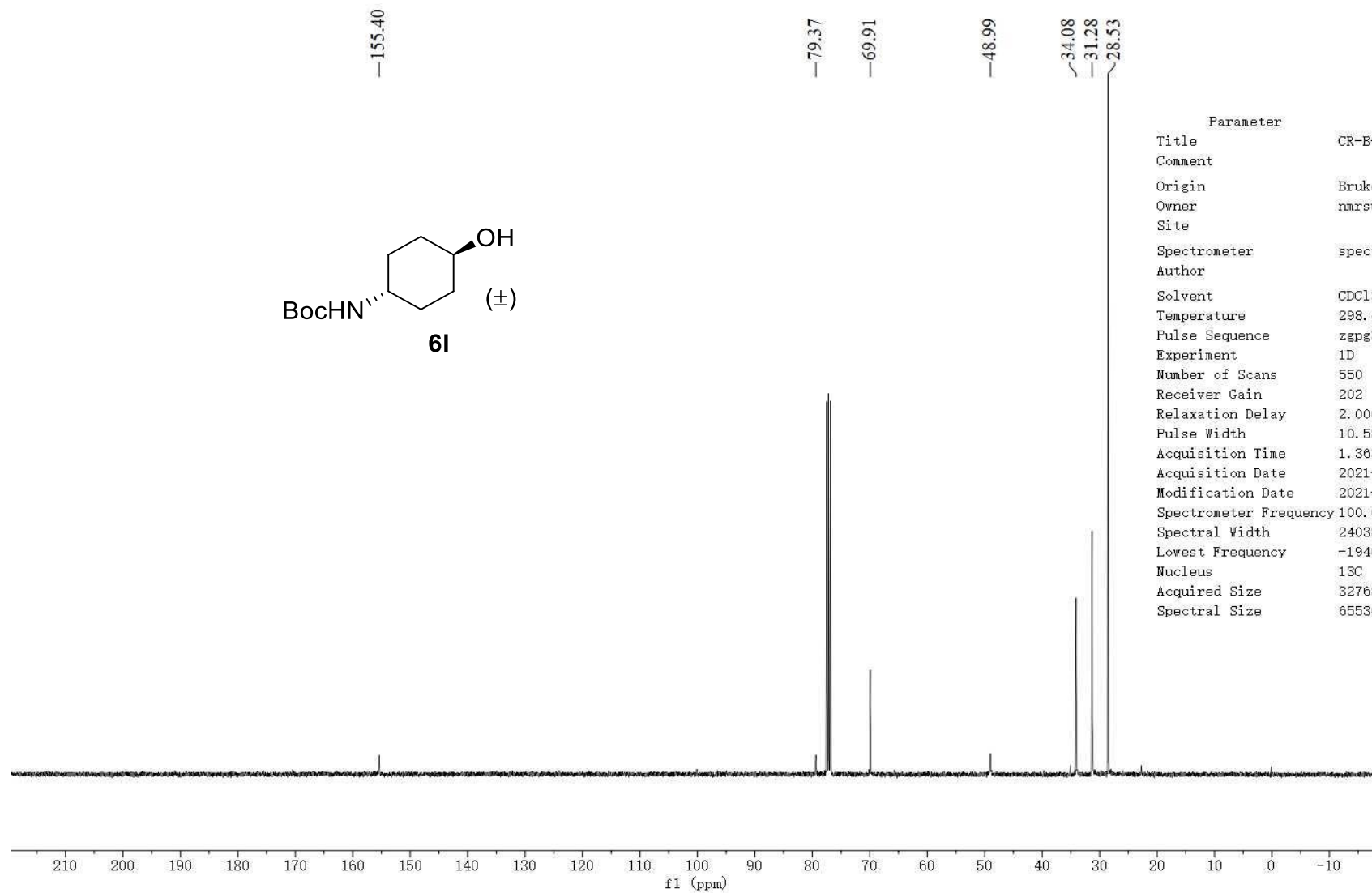
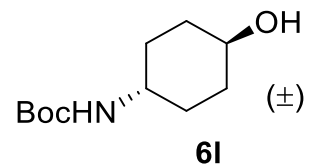
Parameter	Value
Title	CR-B-59.11.fid
Comment	CR-B-59-H
Origin	Bruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDC13
Temperature	298.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	101
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-27T20:28:47
Modification Date	2021-09-27T20:29:06
Spectrometer	400.13
Frequency	
Spectral Width	8196.7
Lowest Frequency	-1636.2
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



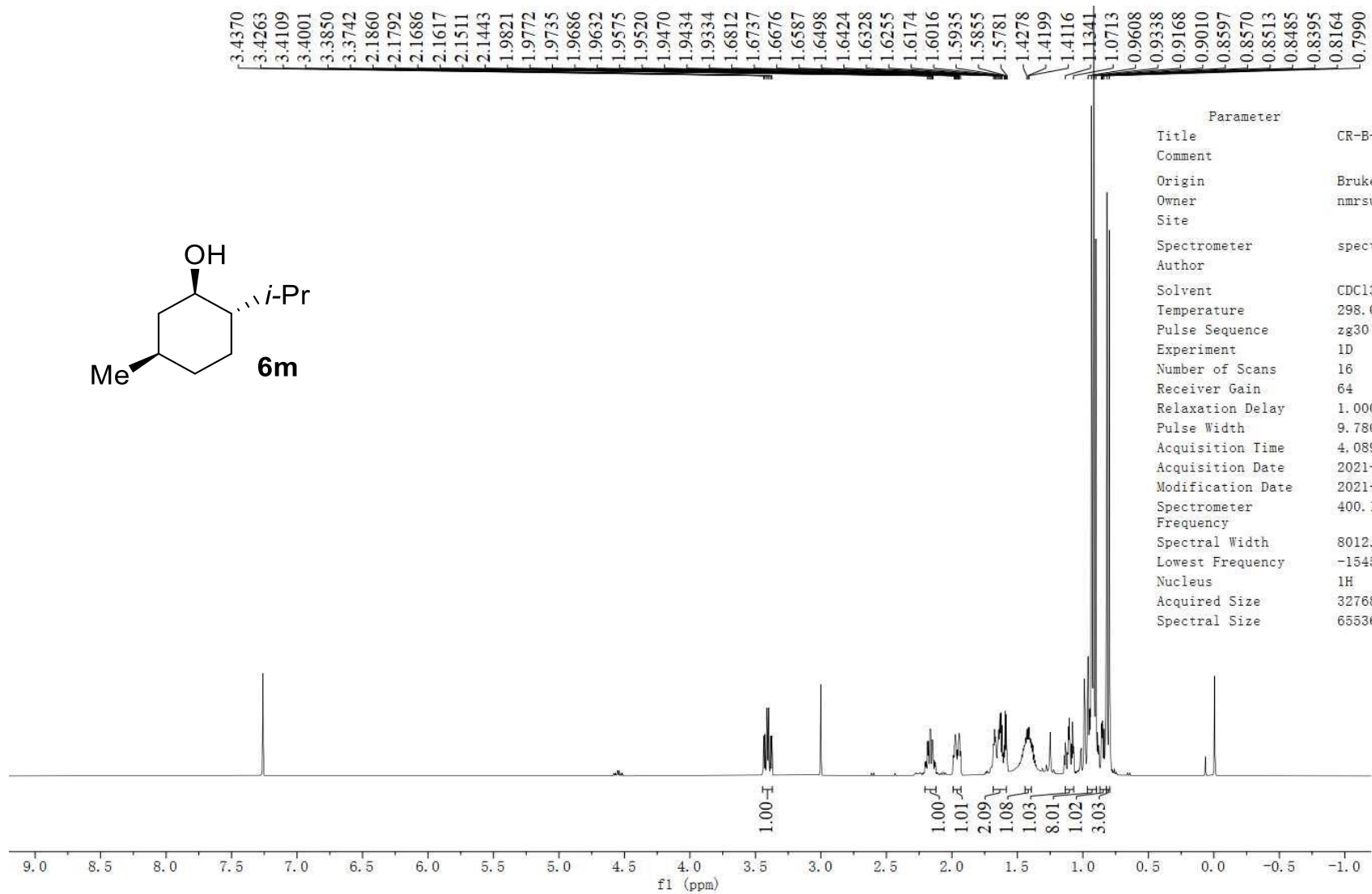
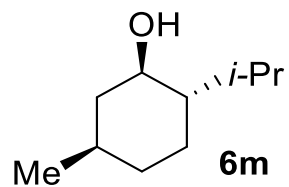


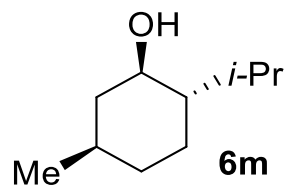
Parameter	Value
Title	CR-B-59.12.fid
Comment	CR-B-59-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-27T21:28:27
Modification Date	2021-09-27T21:28:46
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1830.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





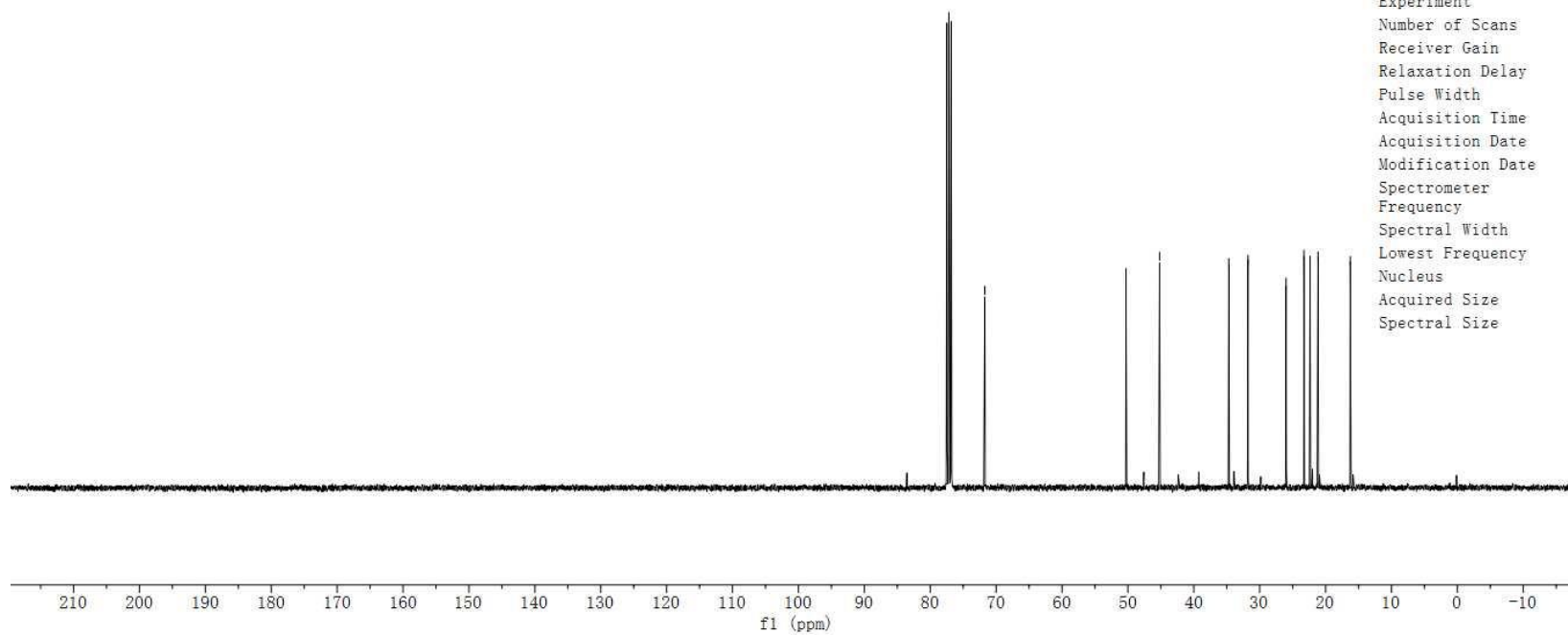
Parameter	Value
Title	CR-B-79
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	550
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-15T22:30:53
Modification Date	2021-09-15T22:30:54
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1946.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

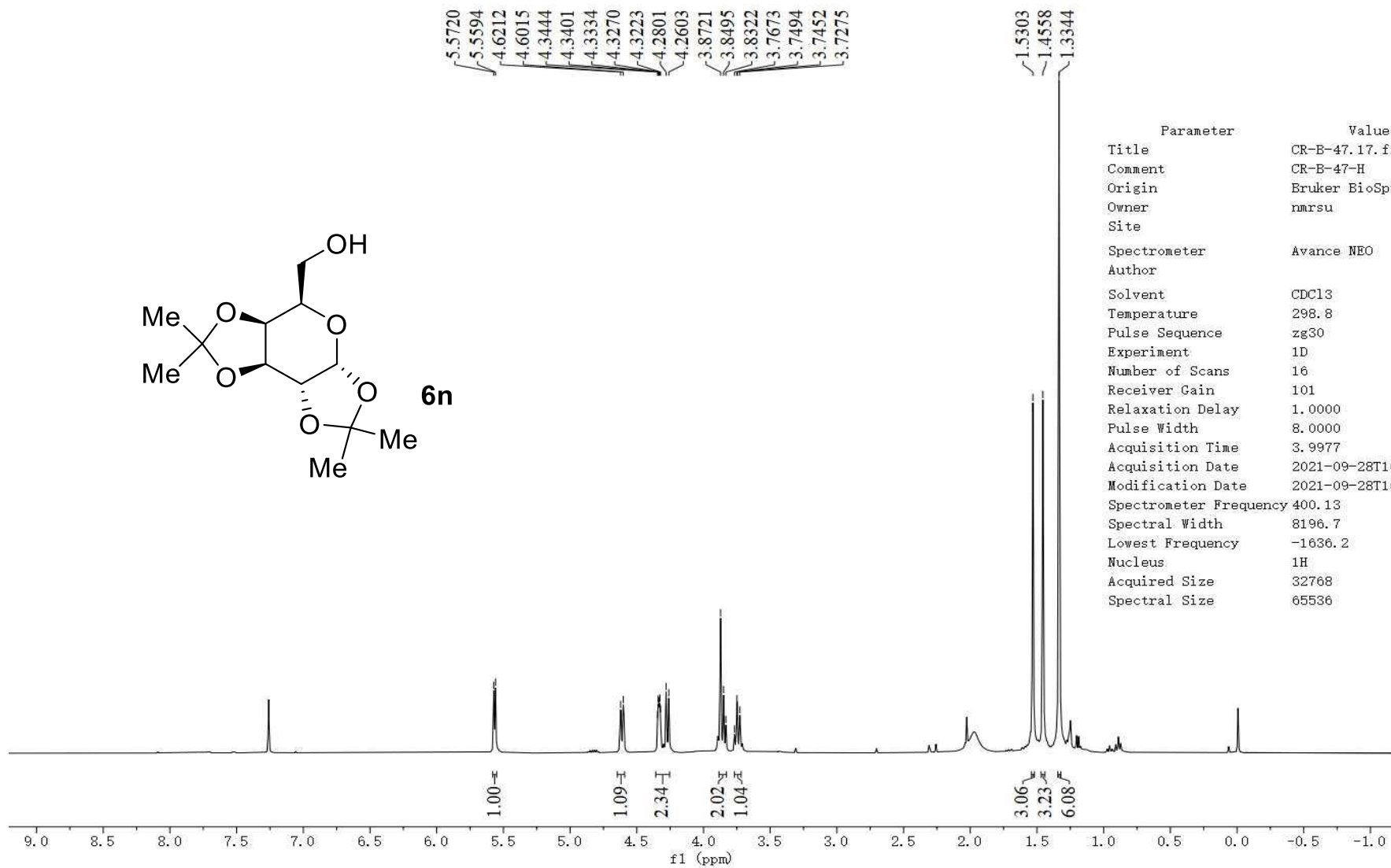
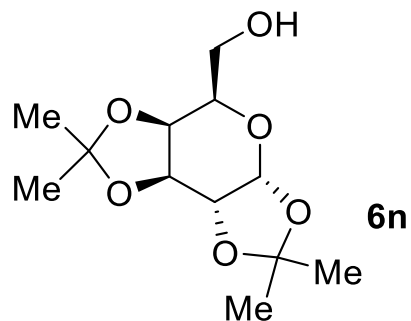




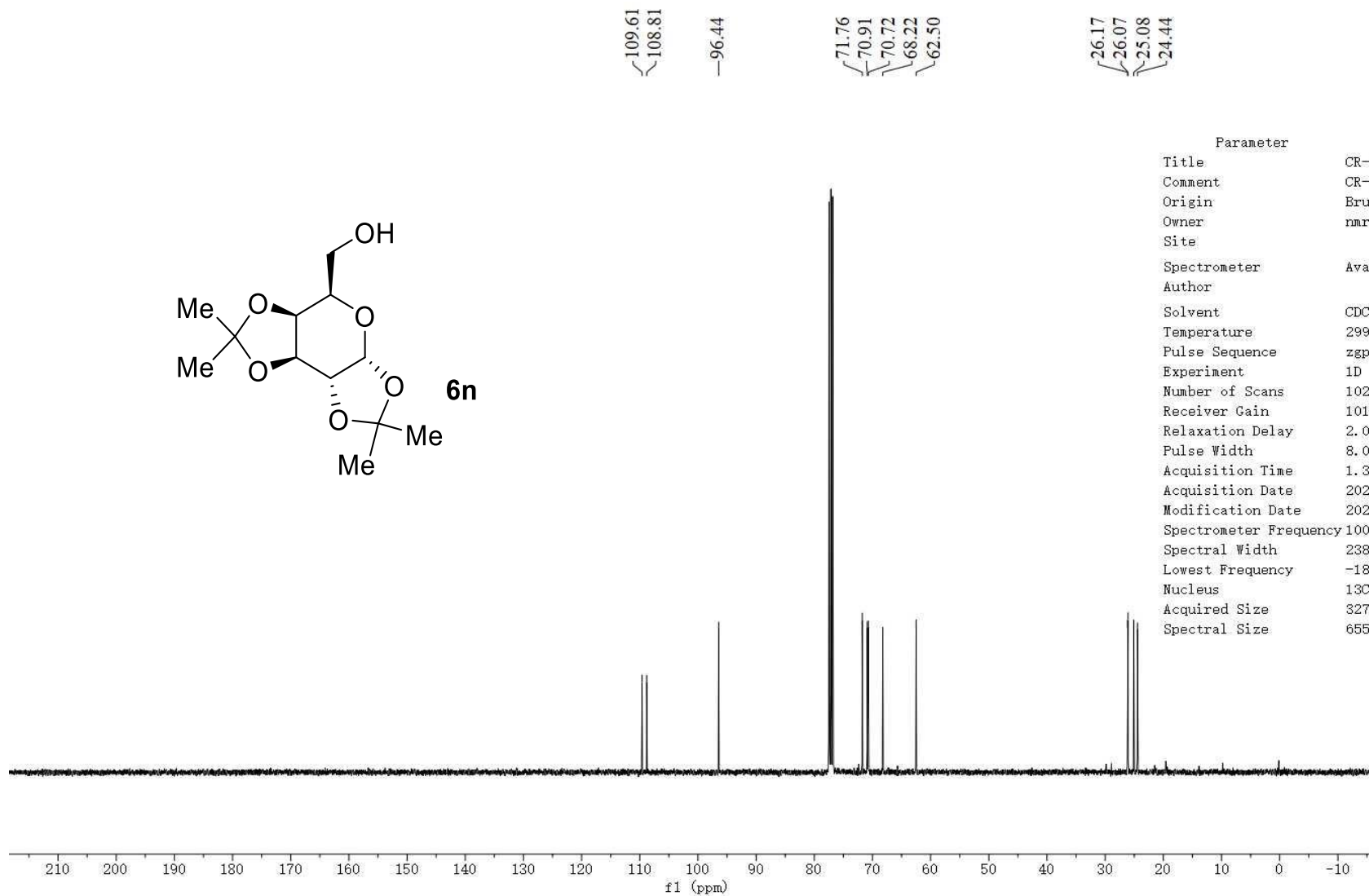
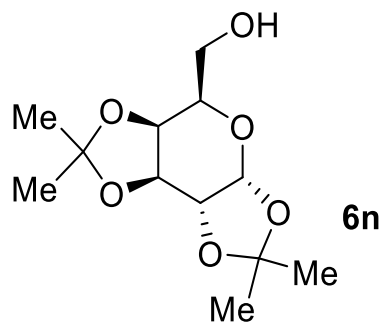
—71.70
 —50.29
 —45.20
 /34.68
 /31.78
 /25.99
 /23.28
 /22.35
 /21.15
 /16.24

Parameter	Value
Title	CR-B-83.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-26T04:34:06
Modification Date	2021-09-26T04:34:08
Spectrometer	100.63
Frequency	
Spectral Width	24038.5
Lowest Frequency	-1944.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



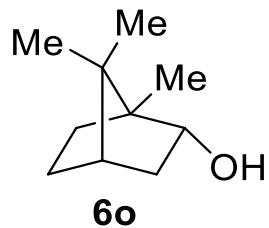


Parameter	Value
Title	CR-B-47.17.fid
Comment	CR-B-47-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDC13
Temperature	298.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	101
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-28T10:50:20
Modification Date	2021-09-28T10:50:38
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1636.2
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

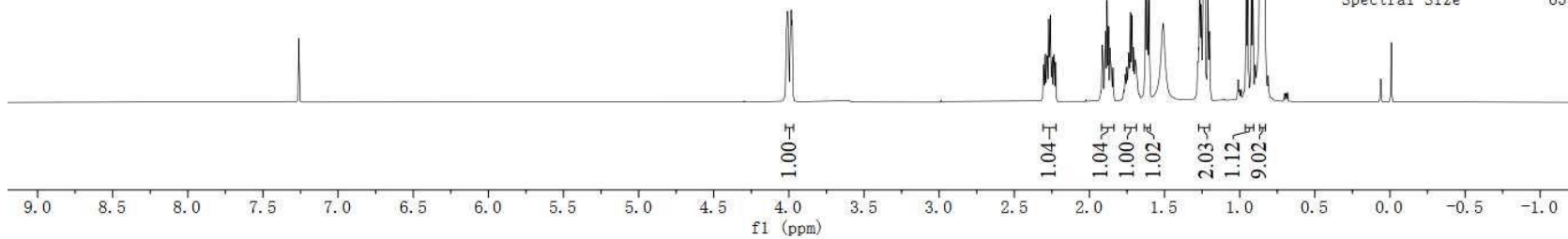


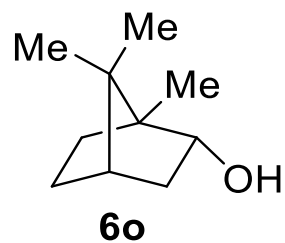
Parameter	Value
Title	CR-B-47.18.fid
Comment	CR-B-47-C
Origin	Bruker BioSpin GmbH
Owner	nrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl ₃
Temperature	299.5
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-28T11:49:46
Modification Date	2021-09-28T11:50:06
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1829.6
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

4.0159
4.0118
4.0076
4.0035
3.9911
3.9868
3.9827
3.9785
2.3042
2.2957
2.2926
2.2840
2.2794
2.2708
2.2592
2.2506
2.2460
2.2373
2.2342
2.2257
1.9138
1.8927
1.8823
1.8727
1.8621
1.8551
1.8433
1.7518
1.7381
1.7270
1.7180
1.7069
1.6927
1.6261
1.6147
1.6033
1.2673
1.2625
1.2566
1.2371
1.2113
1.2006
0.9554
0.9468
0.9220
0.9133
0.8589
0.8476
0.8380



Parameter	Value
Title	CR-B-85
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-26T01:56:12
Modification Date	2021-09-26T01:56:14
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



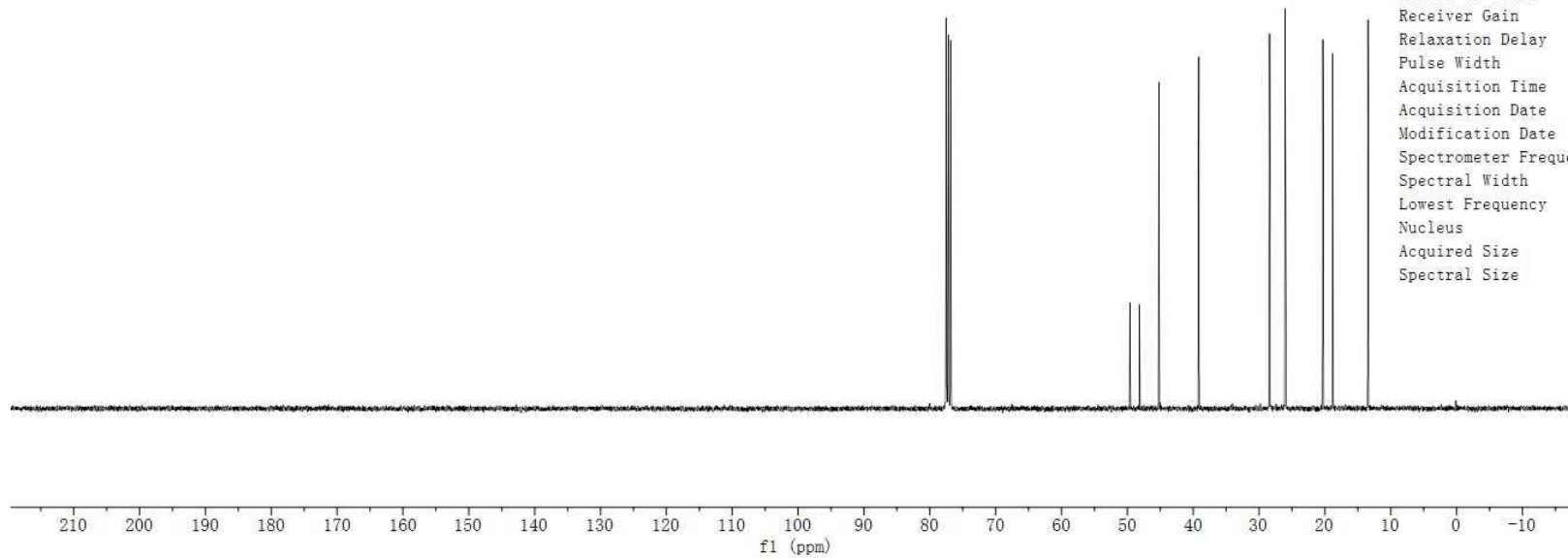


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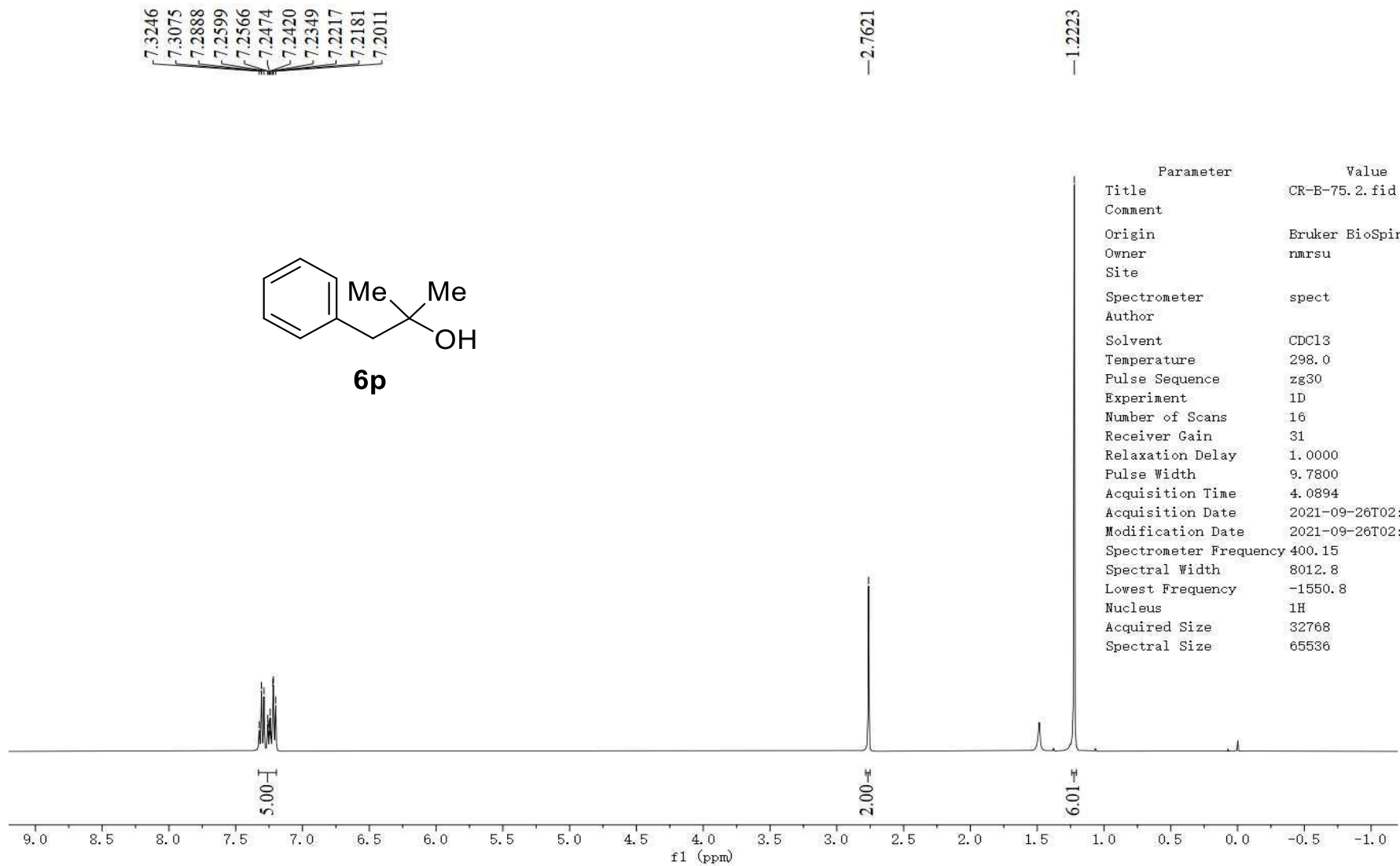
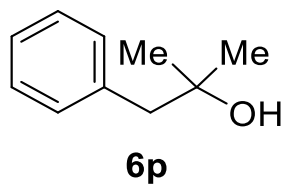
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~48.15
~45.22
~39.15

~28.41
~26.04
~20.32
~18.81
~13.46

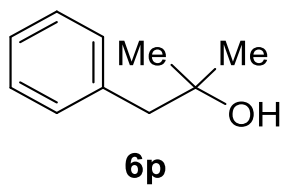
Parameter	Value
Title	CR-B-85
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zpg30
Experiment	1D
Number of Scans	600
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-26T02:31:33
Modification Date	2021-09-26T02:31:34
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1945.2
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



7.3246
7.3075
7.2888
7.2599
7.2566
7.2474
7.2420
7.2349
7.2217
7.2181
7.2011



Parameter	Value
Title	CR-B-75. 2. fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-26T02:37:07
Modification Date	2021-09-26T02:37:08
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1550.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

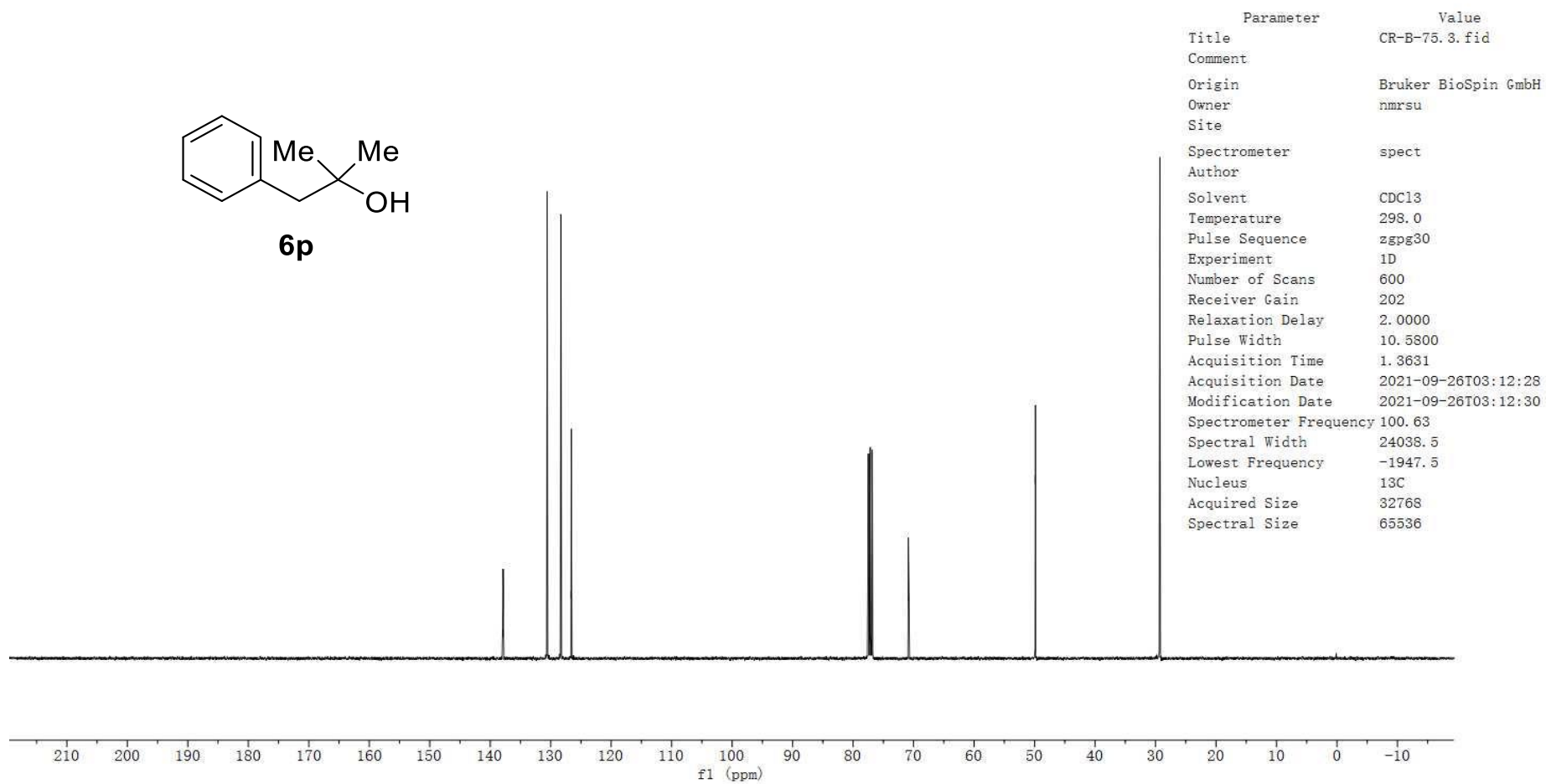


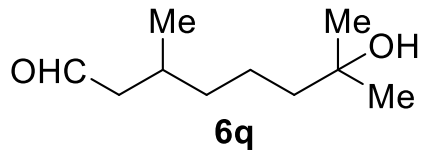
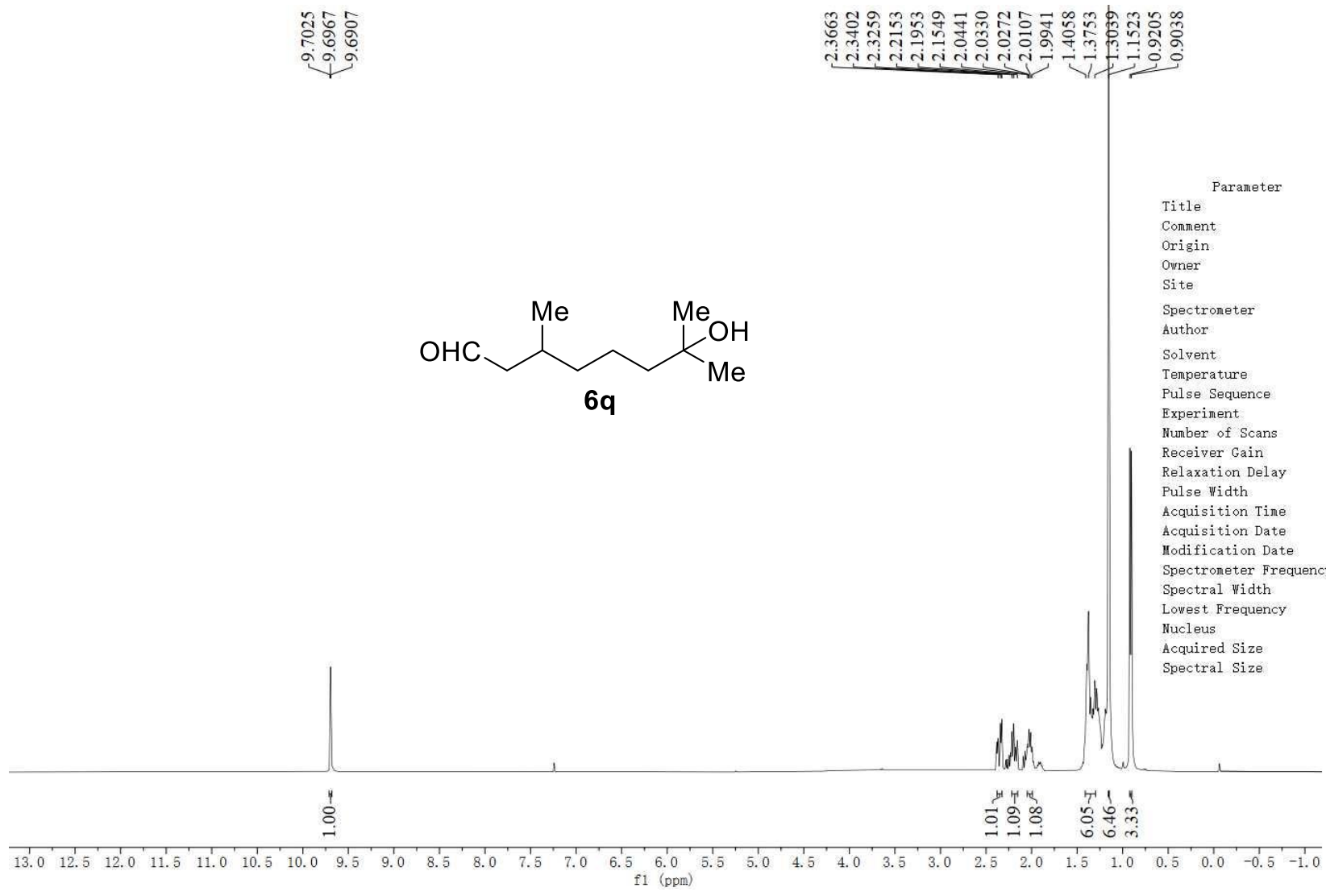
137.89
130.59
128.32
126.61

70.87

49.87

29.29



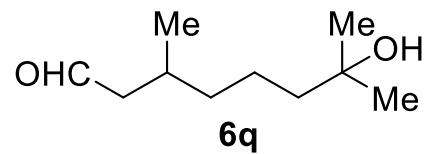


Parameter	Value
Title	CR-B-89.13.fid
Comment	CR-B-89-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	296.8
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-10-18T10:48:06
Modification Date	2021-10-18T10:47:28
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1635.3
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

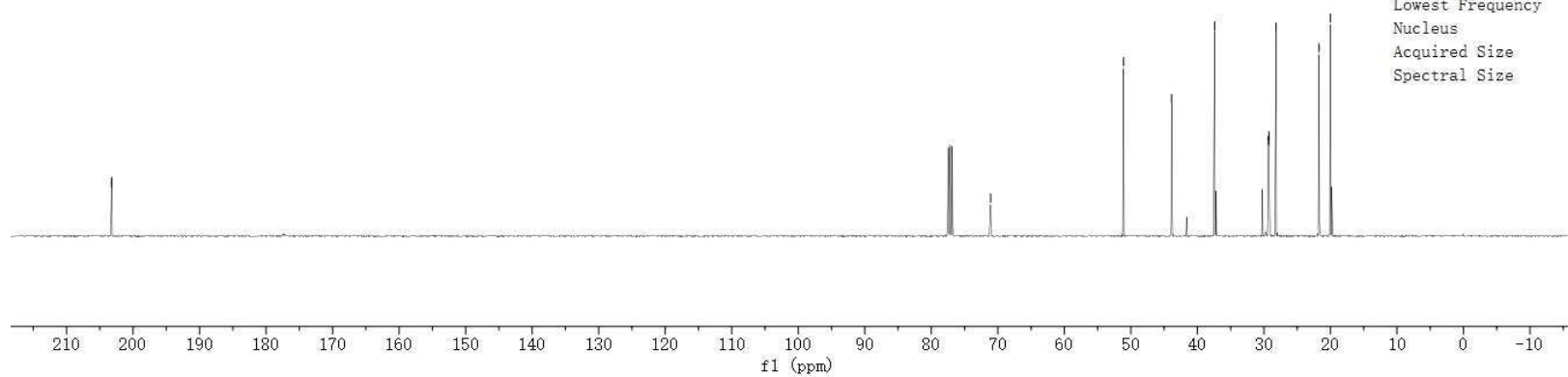
-203.24

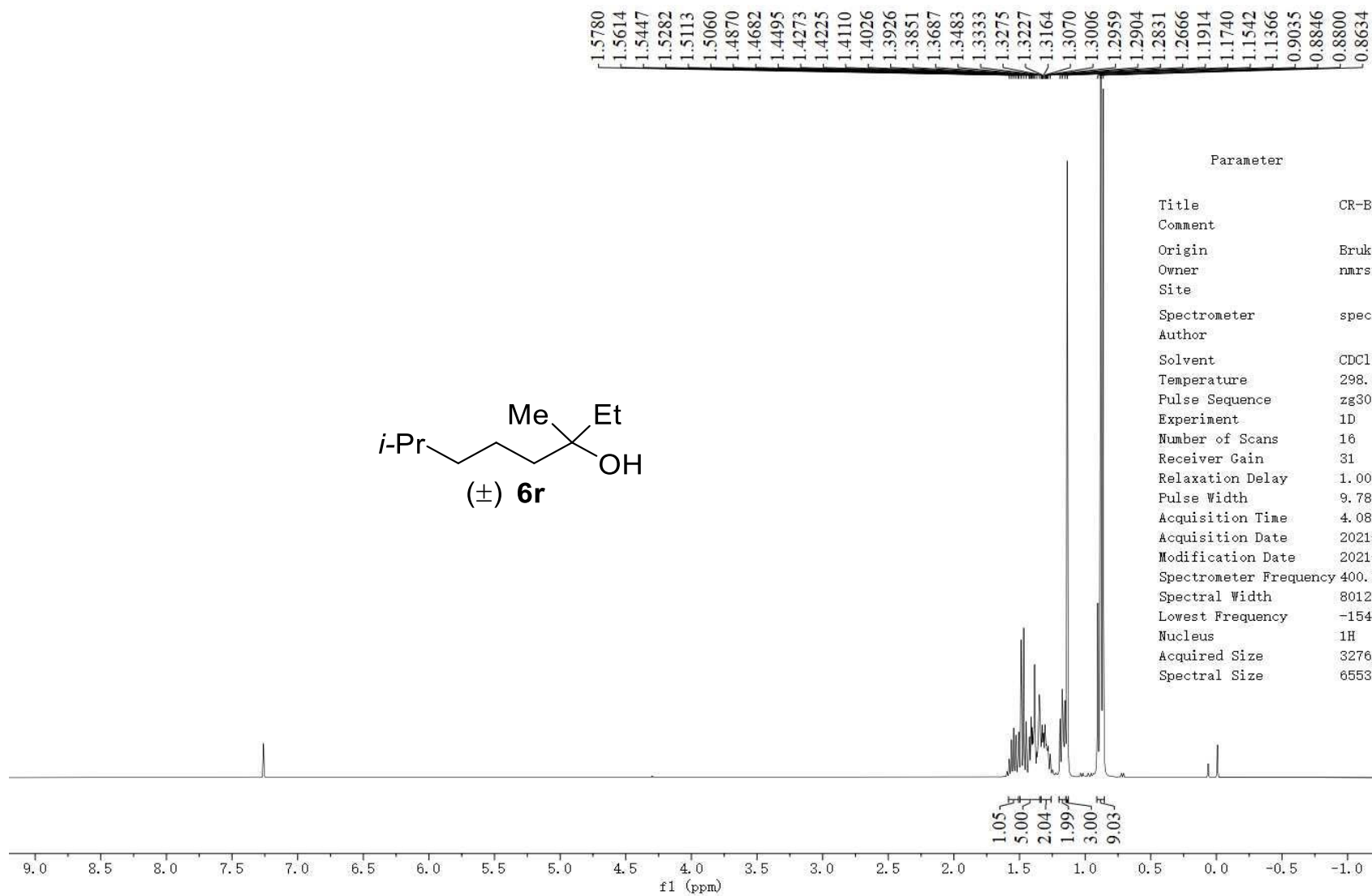
-71.07

51.11
43.89
37.40
29.34
29.25
28.18
21.72
20.02

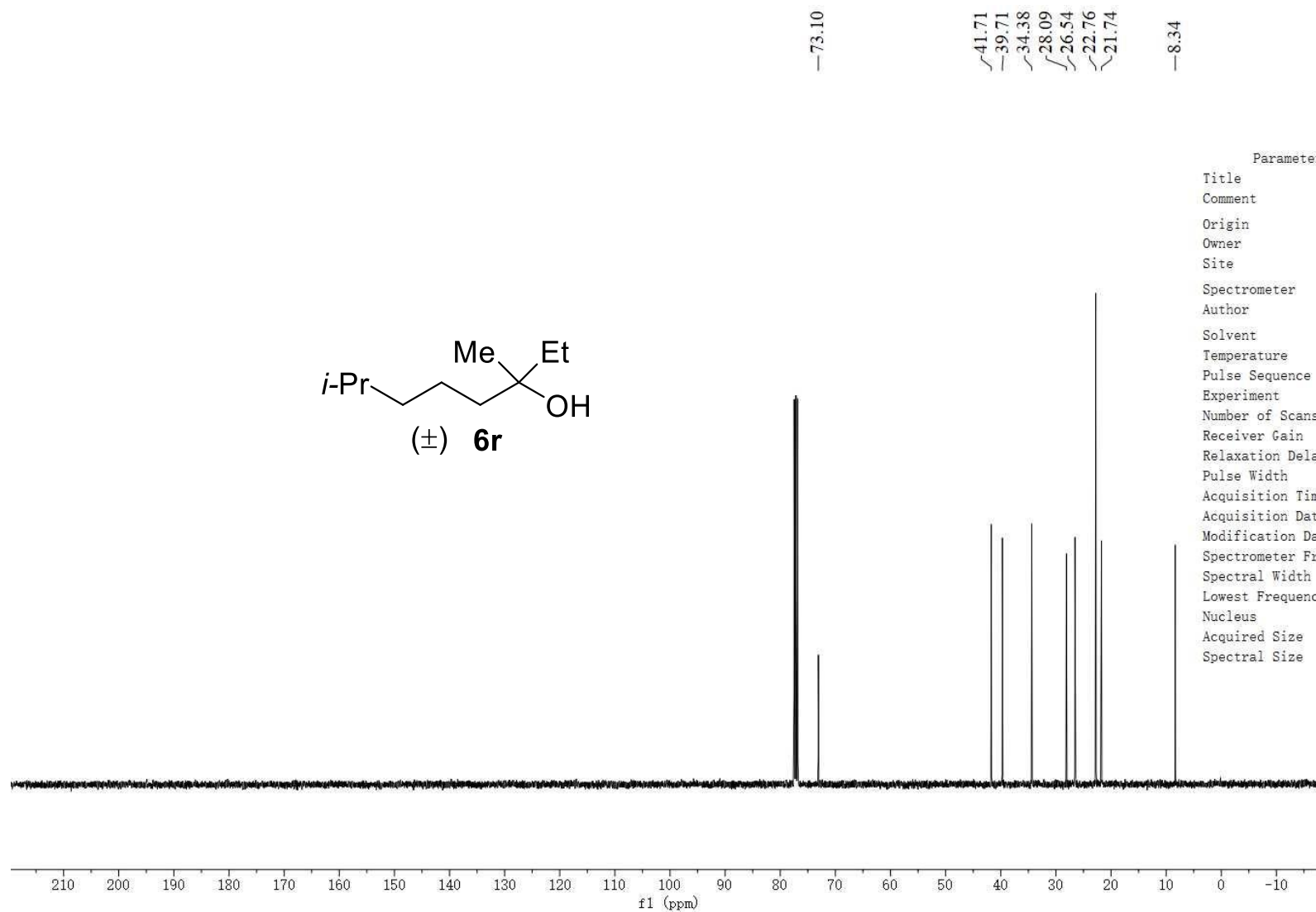
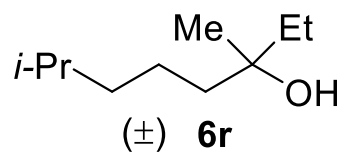


Parameter	Value
Title	CR-B-89. 14. fid
Comment	CR-B-89-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDC13
Temperature	297.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-10-18T11:47:20
Modification Date	2021-10-18T11:46:40
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1833.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

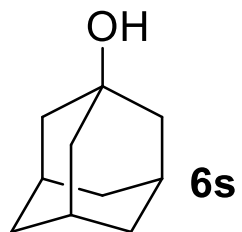




Parameter	Value
Title	CR-B-81
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.7
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-15T07:53:01
Modification Date	2021-09-15T07:53:02
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

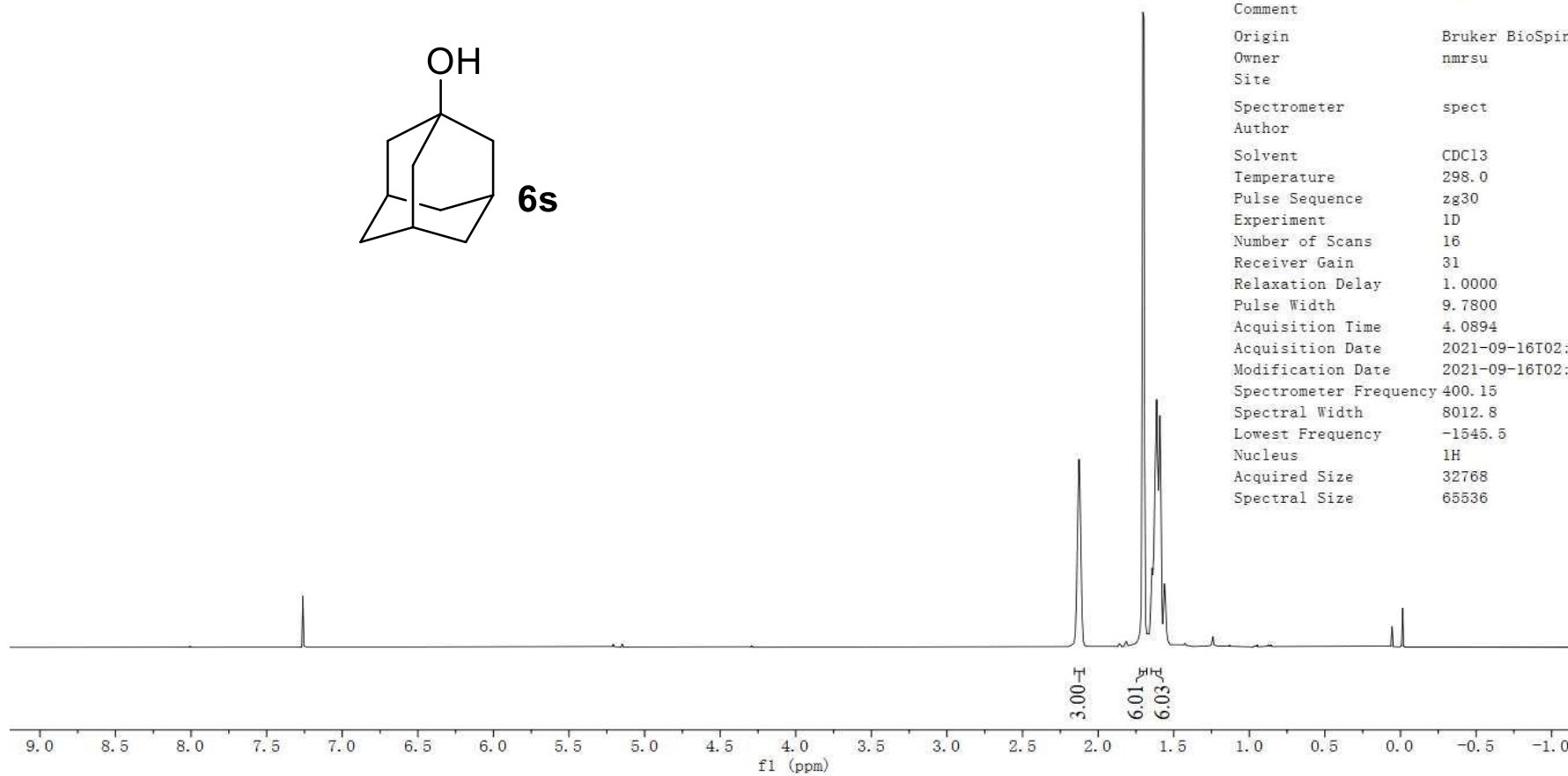


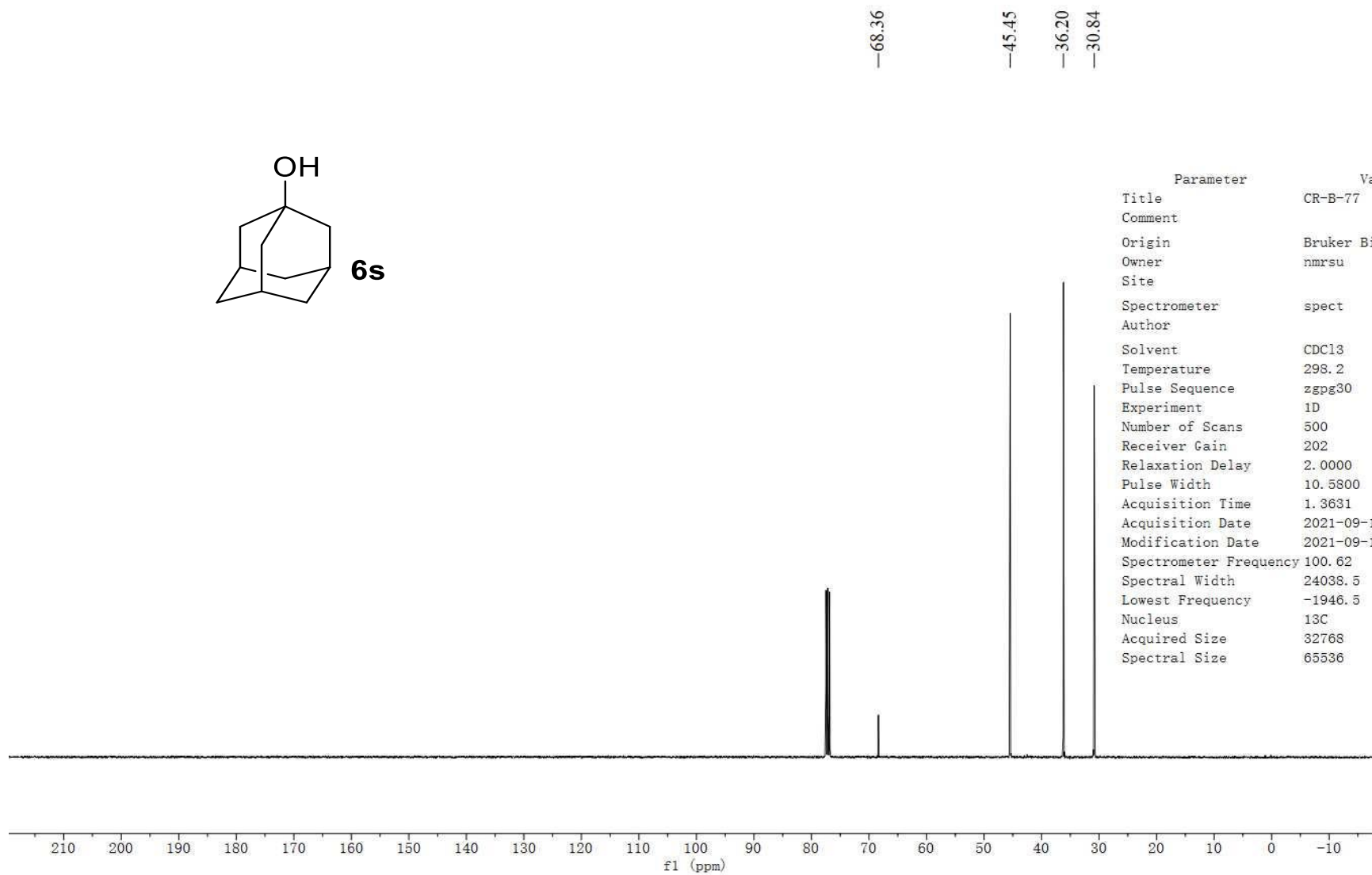
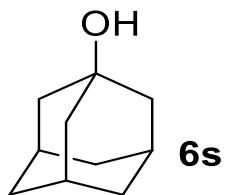
Parameter	Value
Title	CR-B-81
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl ₃
Temperature	299.3
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	198
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-15T08:12:50
Modification Date	2021-09-15T08:12:52
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1943.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



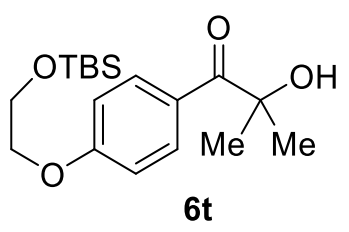
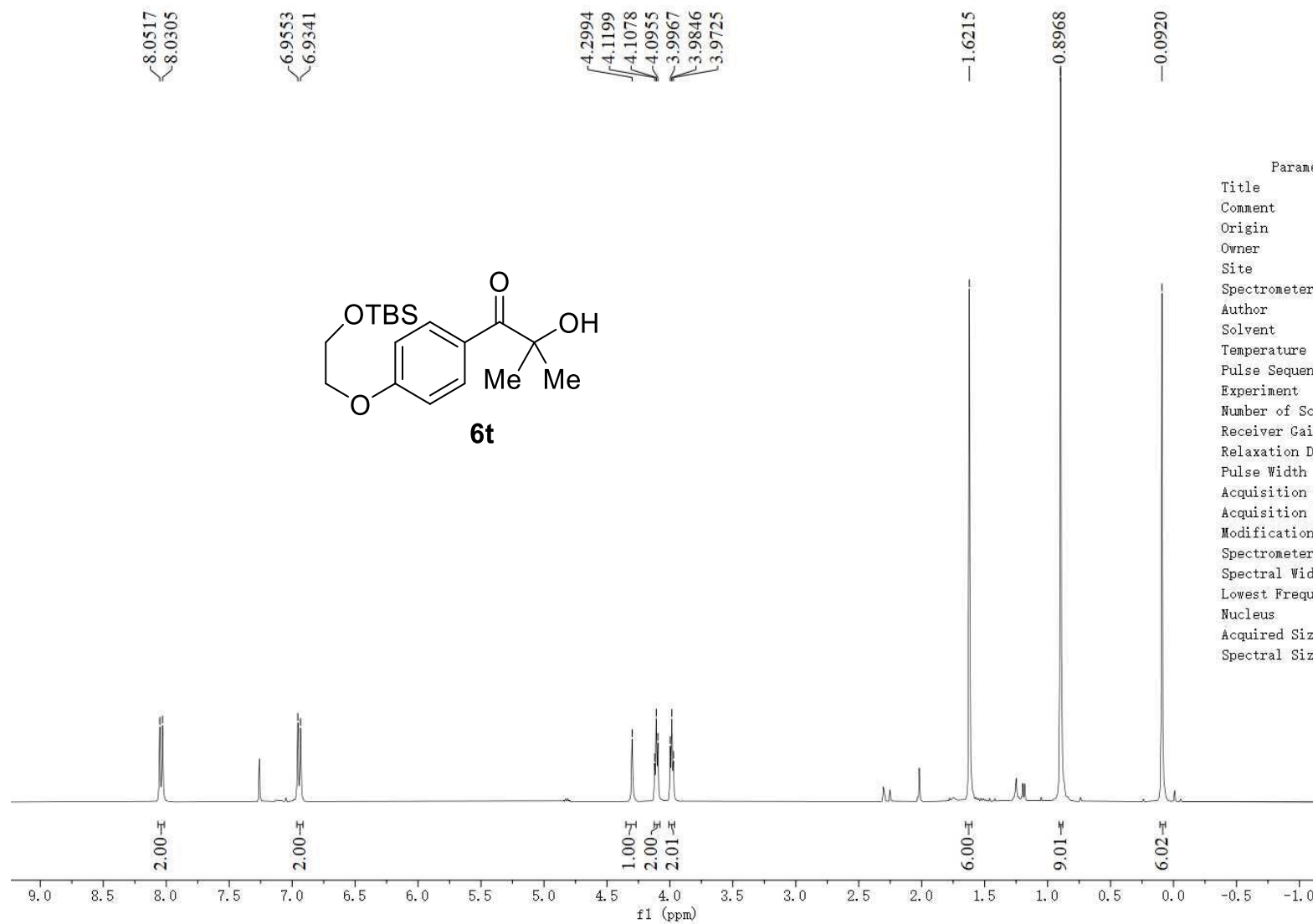
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 1.6973
 1.6430
 1.6129
 1.5916
 1.5610

Parameter	Value
Title	CR-B-77
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-09-16T02:20:18
Modification Date	2021-09-16T02:20:20
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

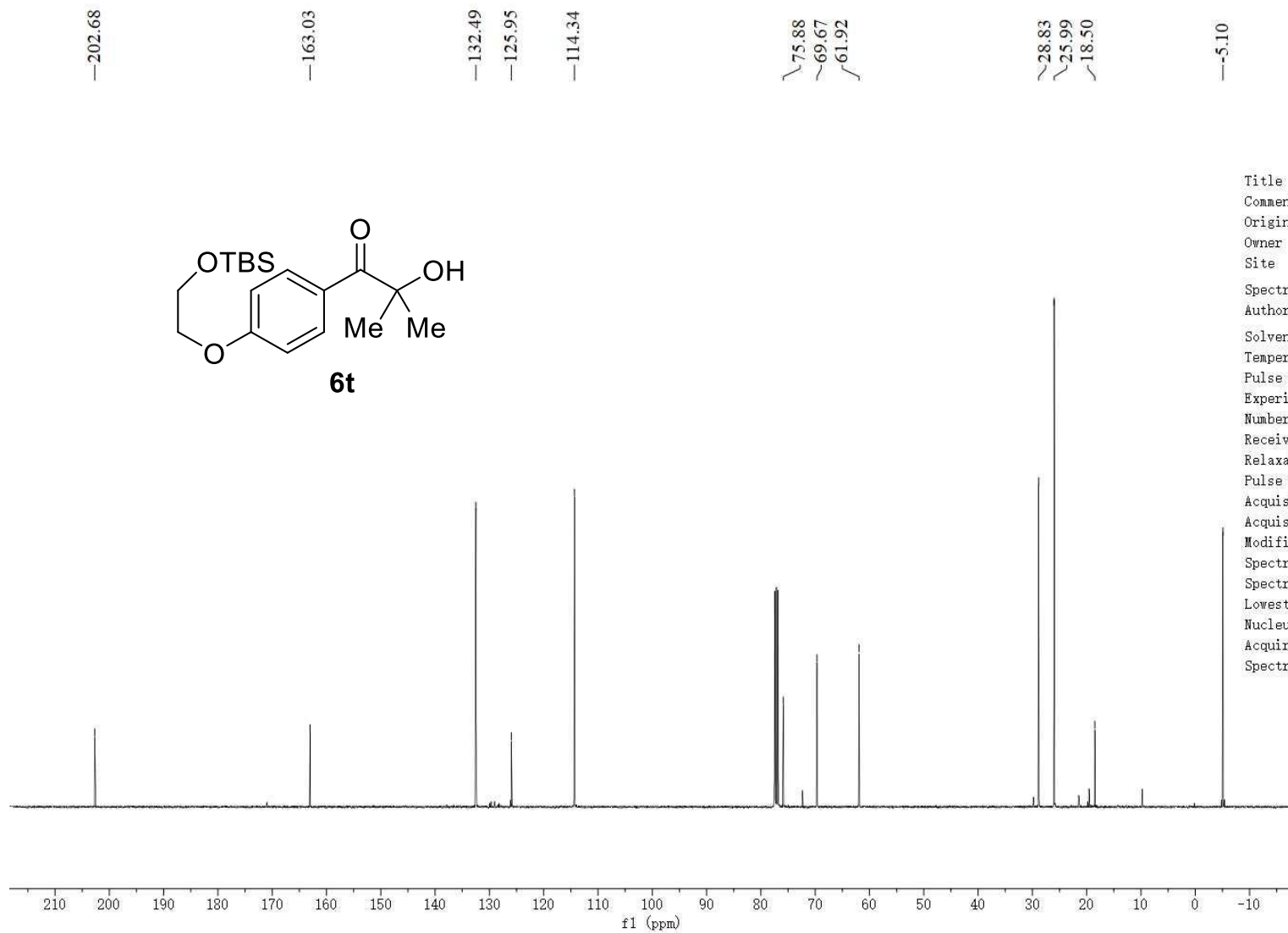




Parameter	Value
Title	CR-B-77
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.2
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	500
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-09-16T02:49:57
Modification Date	2021-09-16T02:49:58
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1946.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Title	CR-B-100.23.fid
Comment	CR-B-100-H
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	298.7
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	43
Relaxation Delay	1.0000
Pulse Width	8.0000
Acquisition Time	3.9977
Acquisition Date	2021-09-28T14:44:14
Modification Date	2021-09-28T14:44:34
Spectrometer Frequency	400.13
Spectral Width	8196.7
Lowest Frequency	-1636.1
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

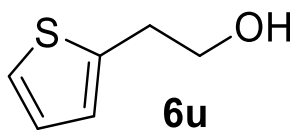


Parameter	Value
Title	CR-B-100.24.fid
Comment	CR-B-100-C
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	Avance NEO
Author	
Solvent	CDCl3
Temperature	299.4
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	101
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	1.3763
Acquisition Date	2021-09-28T15:45:51
Modification Date	2021-09-28T15:46:10
Spectrometer Frequency	100.62
Spectral Width	23809.5
Lowest Frequency	-1831.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

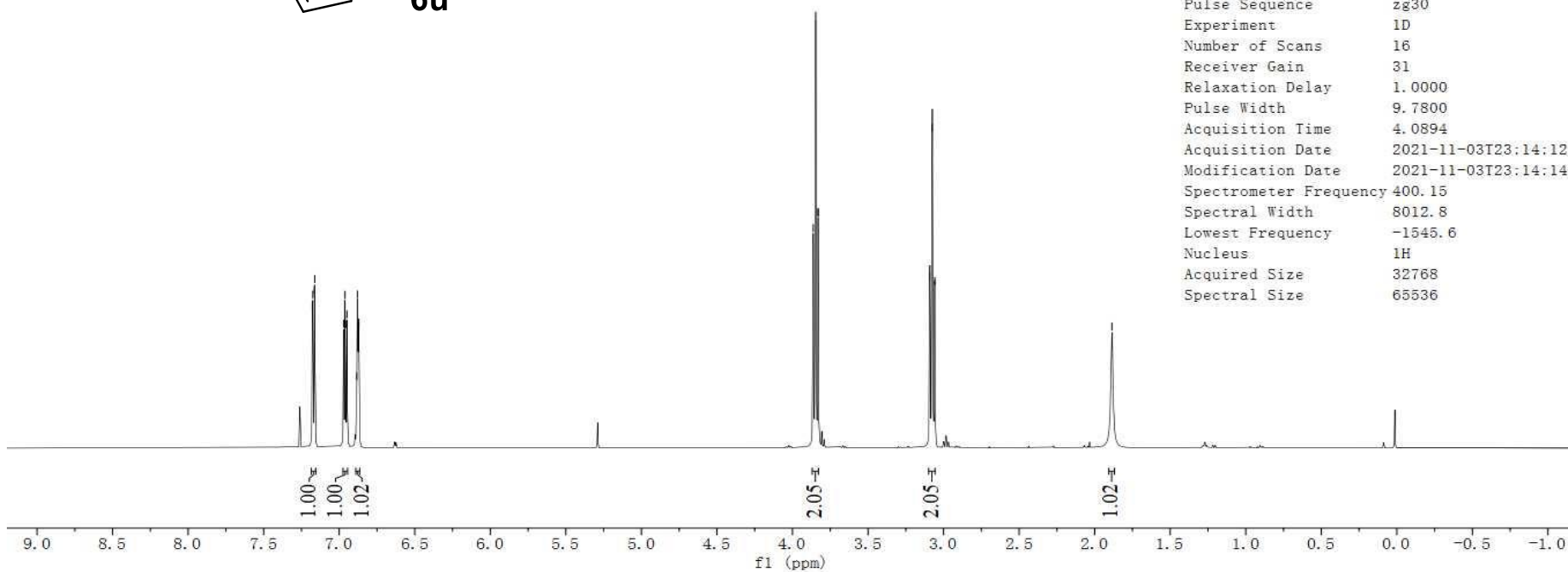
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7.1647
7.1616
6.9700
6.9615
6.9572
6.9486
6.8842
6.8788
6.8701

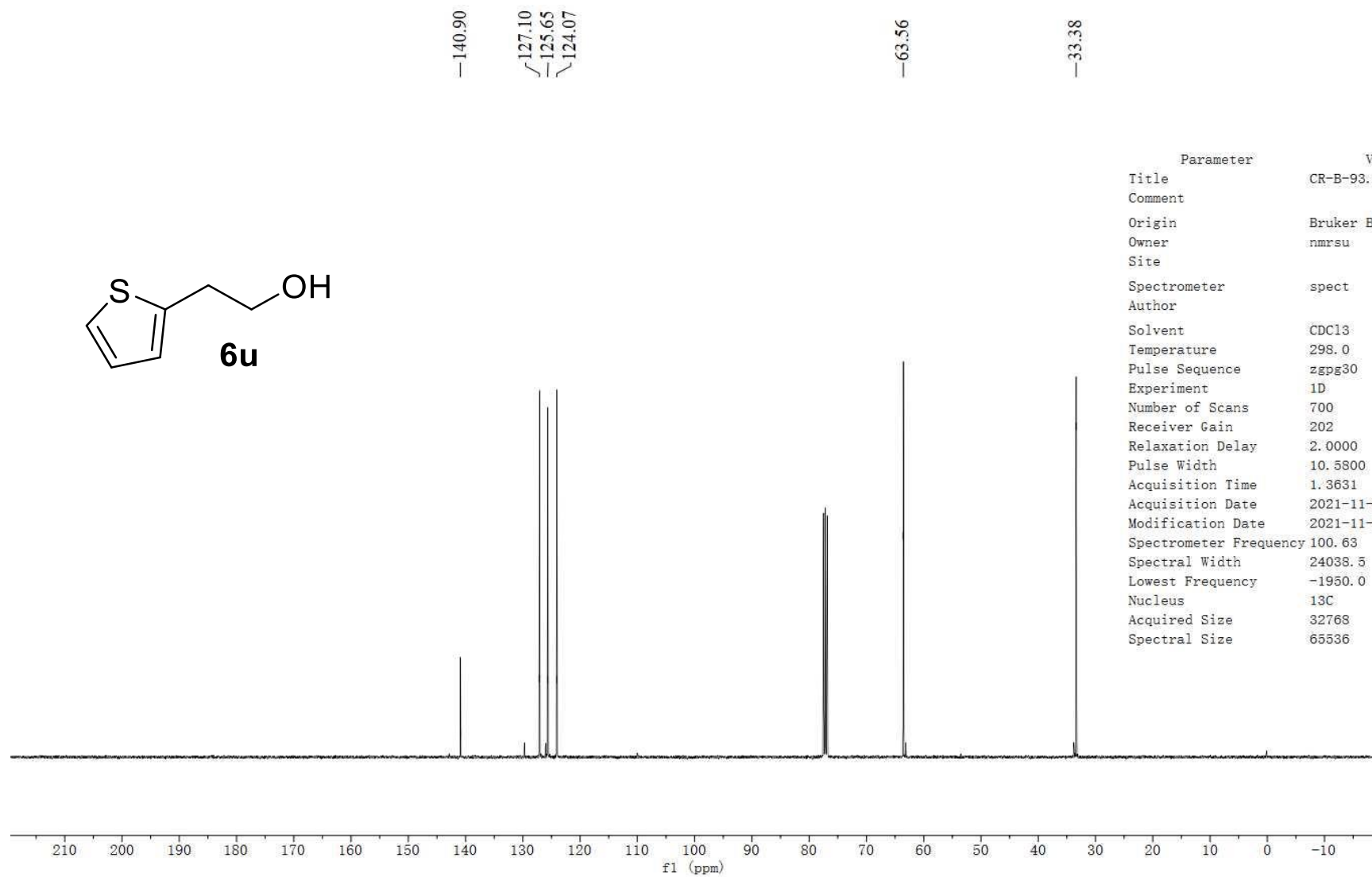
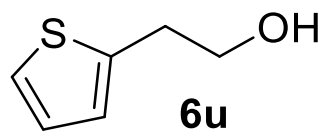
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3.8465
3.8308
3.8305
3.0916
3.0757
3.0577

-1.8856

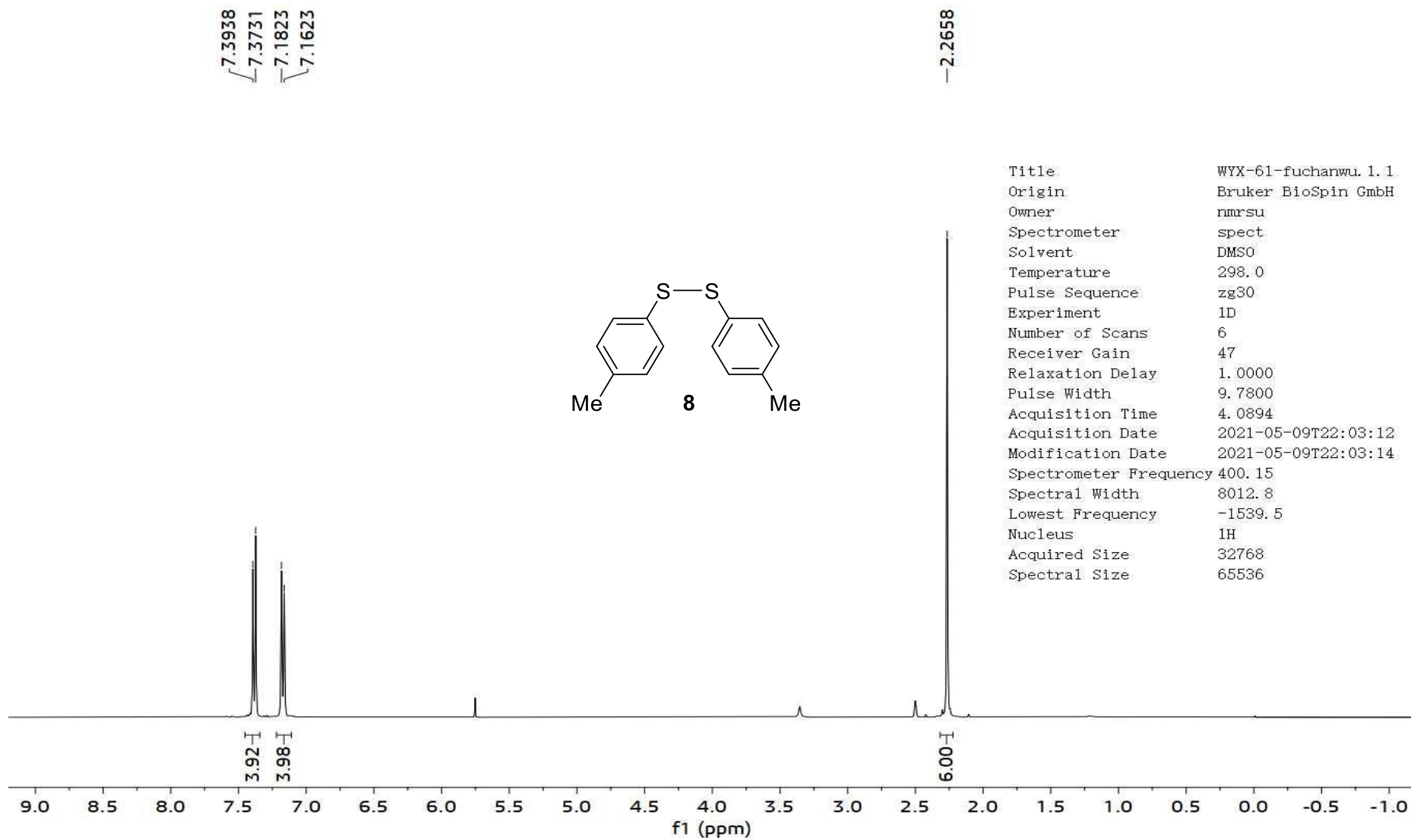


Parameter	Value
Title	CR-B-93. 2. fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDCl3
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-11-03T23:14:12
Modification Date	2021-11-03T23:14:14
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536





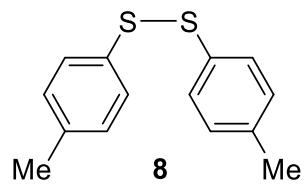
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Title	CR-B-93.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Site	
Spectrometer	spect
Author	
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-11-03T23:55:16
Modification Date	2021-11-03T23:55:18
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1950.0
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



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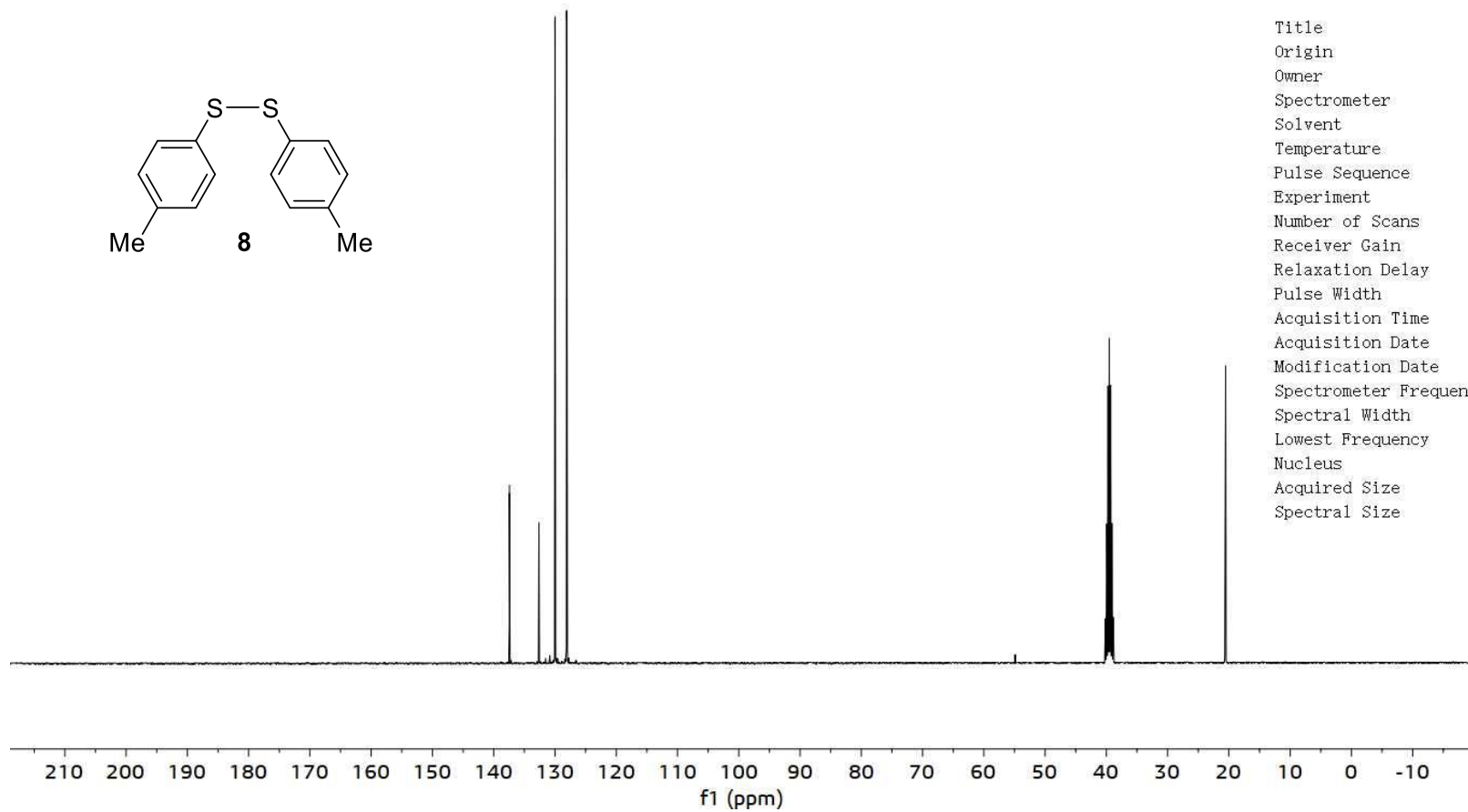
Title      WYX-61-fuchanwu.1.1
Origin     Bruker BioSpin GmbH
Owner      nmrsu
Spectrometer spect
Solvent    DMSO
Temperature 298.0
Pulse Sequence zg30
Experiment 1D
Number of Scans 6
Receiver Gain 47
Relaxation Delay 1.0000
Pulse Width 9.7800
Acquisition Time 4.0894
Acquisition Date 2021-05-09T22:03:12
Modification Date 2021-05-09T22:03:14
Spectrometer Frequency 400.15
Spectral Width 8012.8
Lowest Frequency -1539.5
Nucleus    1H
Acquired Size 32768
Spectral Size 65536

```

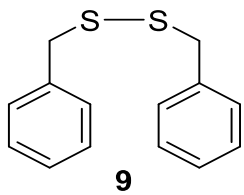
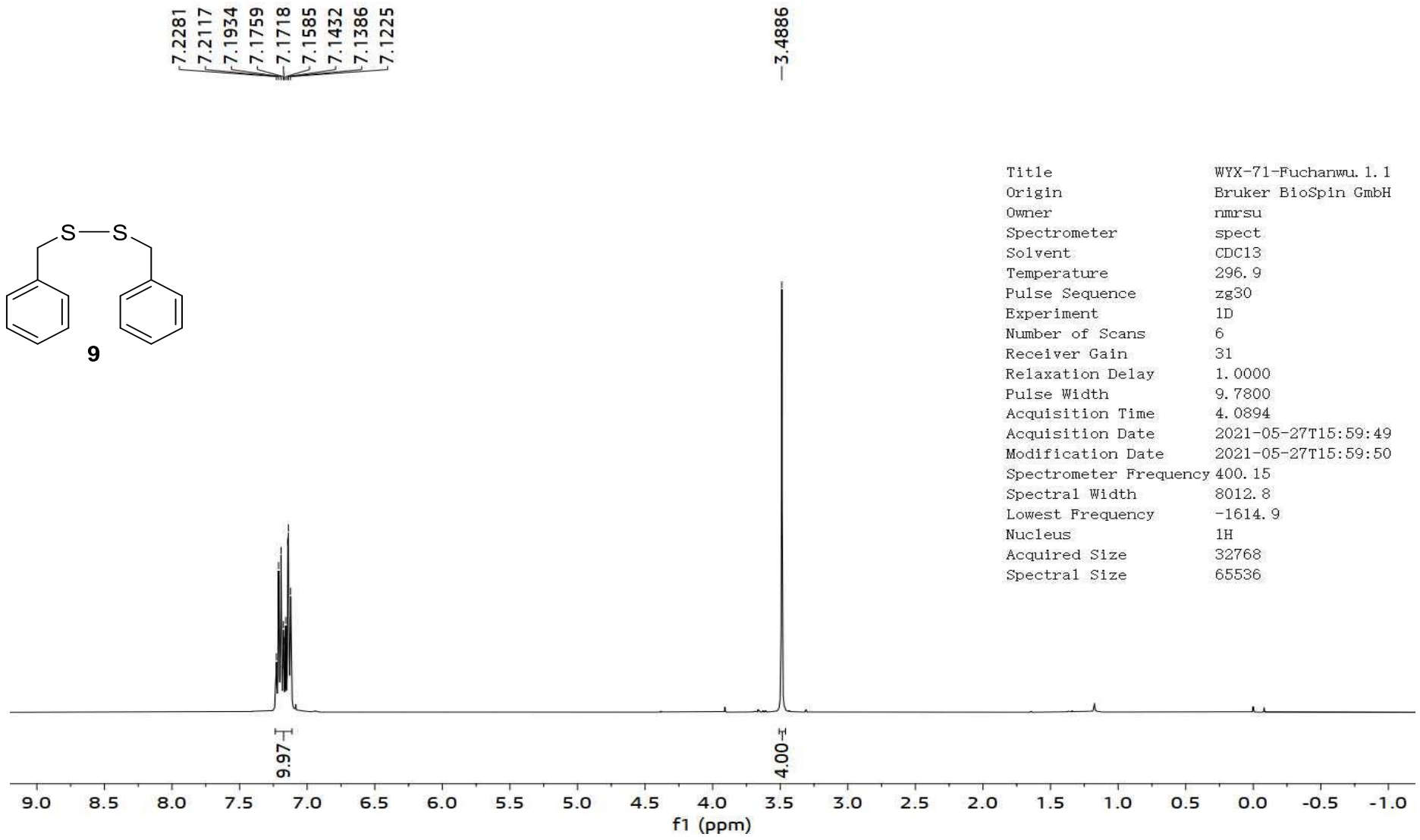



137.43
132.61
129.99
128.09

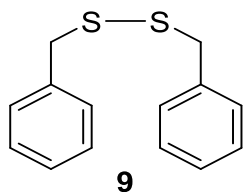
20.54



Title	WYX-61-fuchanwu. 2. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	DMSO
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-09T23:02:46
Modification Date	2021-05-09T23:02:48
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-2008.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



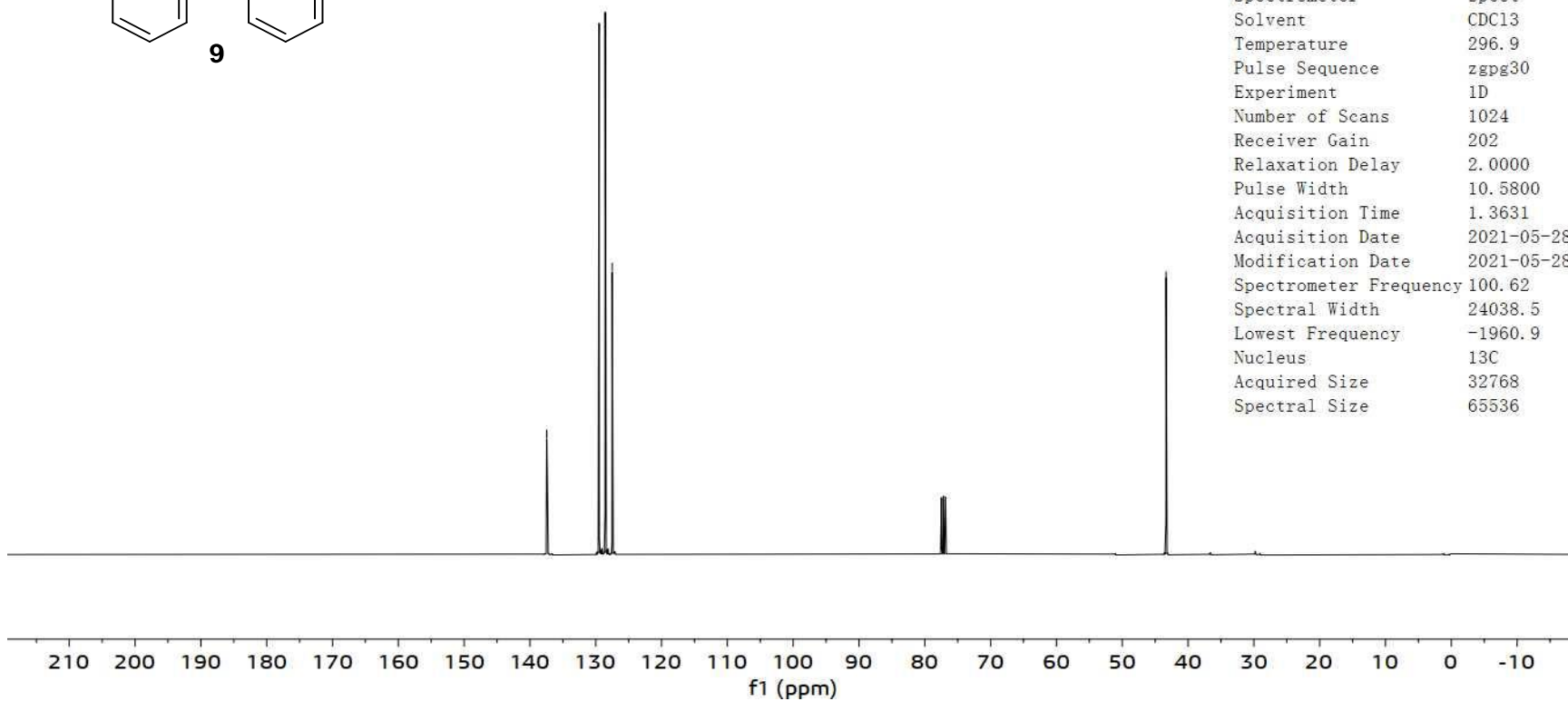
Title	WYX-71-Fuchanwu. 1. 1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDCl3
Temperature	296.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	6
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-05-27T15:59:49
Modification Date	2021-05-27T15:59:50
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1614.9
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



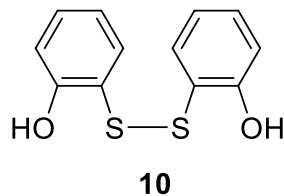
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128.56
127.50

-43.34

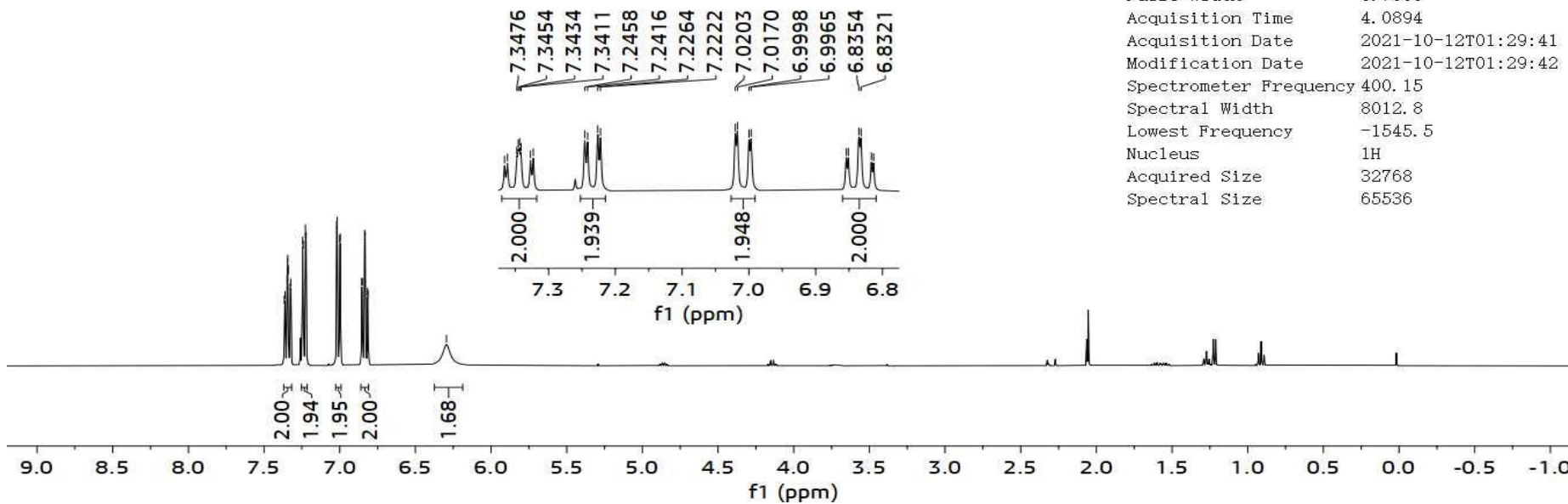
Title	WYX-71-Fuchanwu. 2.1
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	296.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-05-28T03:57:58
Modification Date	2021-05-28T03:58:00
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1960.9
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

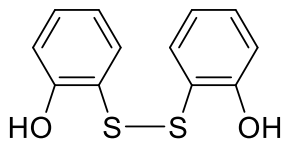


7.3476
7.3454
7.3434
7.3411
7.2458
7.2416
7.2264
7.2222
7.0203
7.0170
6.9998
6.9965
6.8510
6.8354
6.8321
6.2943



Title	Fu-OH. 1. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	31
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-12T01:29:41
Modification Date	2021-10-12T01:29:42
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536

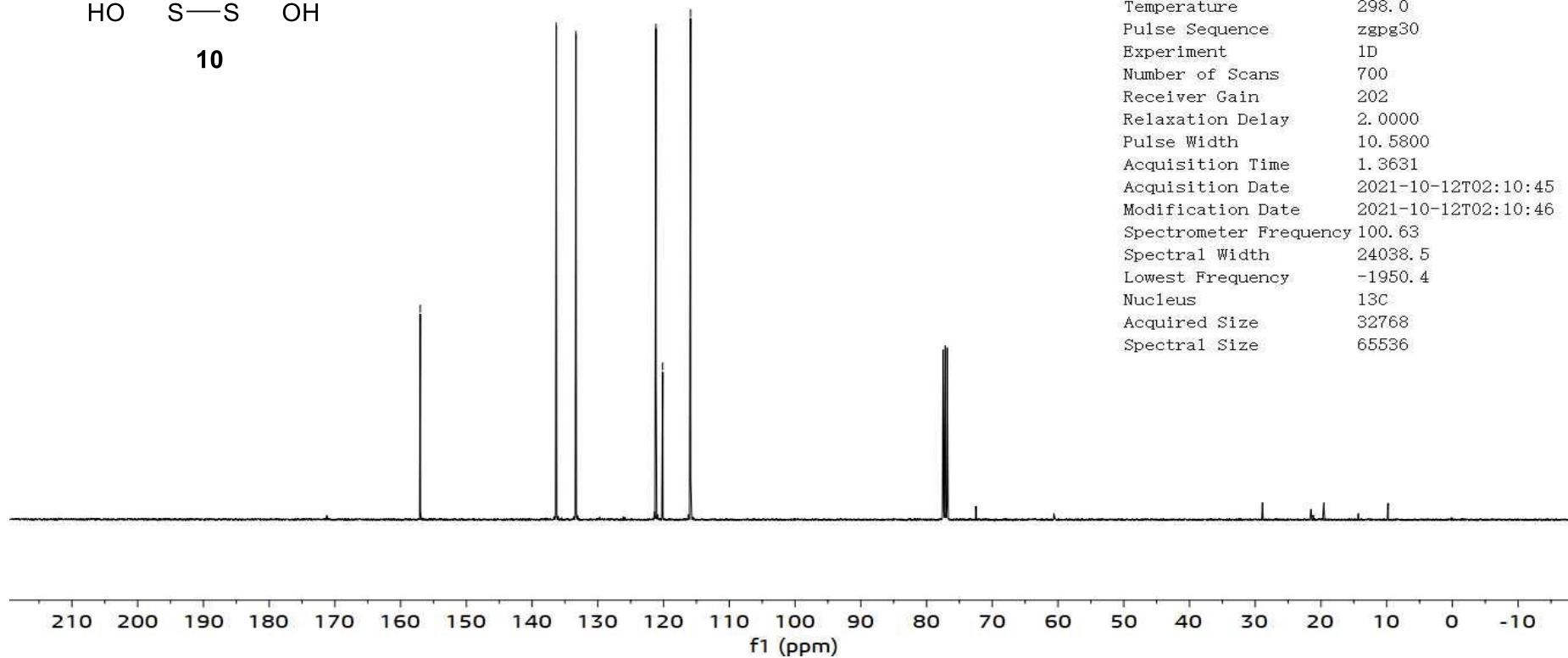




10

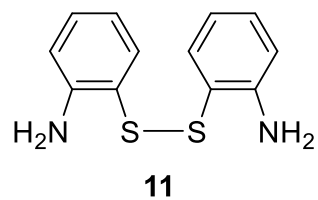
— 157.01
~ 136.32
~ 133.33
~ 121.16
~ 120.13
~ 115.90

Title	Fu-OH. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T02:10:45
Modification Date	2021-10-12T02:10:46
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1950.4
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

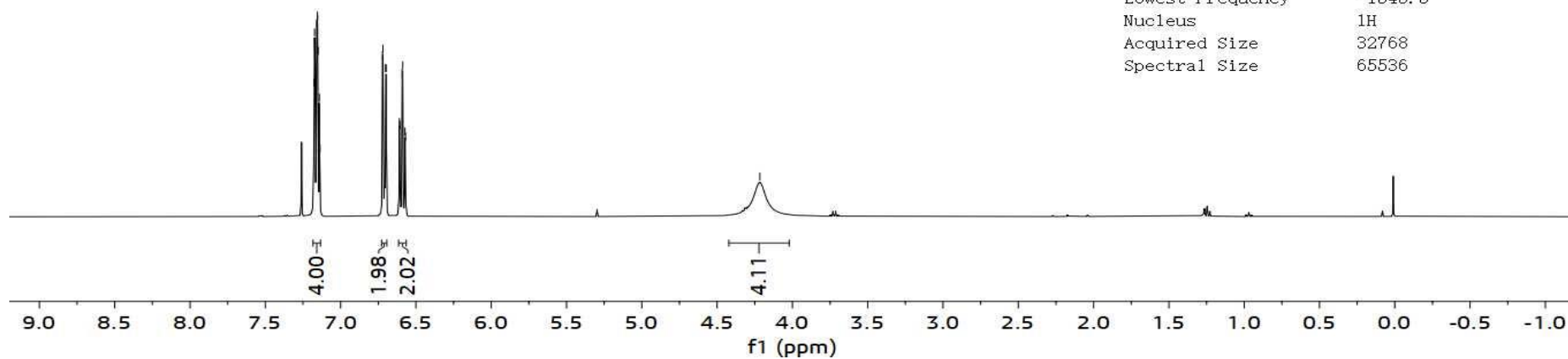


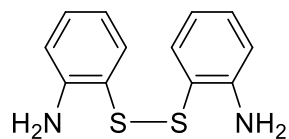
7.1763
7.1729
7.1689
7.1589
7.1538
7.1496
7.1412
7.1372
6.7220
6.7189
6.7011
6.6978
6.6100
6.6067
6.5914
6.5880
6.5725
6.5692

4.2164



Title	Fu-NH2. 1. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	79
Relaxation Delay	1.0000
Pulse Width	9.7800
Acquisition Time	4.0894
Acquisition Date	2021-10-12T02:16:44
Modification Date	2021-10-12T02:16:46
Spectrometer Frequency	400.15
Spectral Width	8012.8
Lowest Frequency	-1545.5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536





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— 148.75
 — 136.95
 — 131.73
 { 118.87
 { 118.36
 { 115.36

Title	Fu-NH2. 2. fid
Origin	Bruker BioSpin GmbH
Owner	nmrsu
Spectrometer	spect
Solvent	CDC13
Temperature	298.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	700
Receiver Gain	202
Relaxation Delay	2.0000
Pulse Width	10.5800
Acquisition Time	1.3631
Acquisition Date	2021-10-12T02:57:48
Modification Date	2021-10-12T02:57:50
Spectrometer Frequency	100.63
Spectral Width	24038.5
Lowest Frequency	-1947.8
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

