# Water enables the tunable electrochemical synthesis of heterocyclic 3a- and 5a-bromoindolines

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### **Supporting Information**

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### **1.** General information

Unless stated, otherwise all reactions were carried out under an atmosphere of nitrogen using standard Schlenk techniques. All the reactions that need to be heated, the oil bath is used as a heating source. All solvents and reagents were obtained from commercial sources and were purified according to standard procedures before use. Column chromatography was performed on silica gel (Qingdao, 300 - 400 mesh) using the indicated eluents. NMR spectra were recorded on an Agilent Mercury 600 MHz spectrometer (<sup>1</sup>H: 600 MHz and <sup>13</sup>C: 150 MHz) in chloroform-*d*. <sup>1</sup>H and <sup>13</sup>C NMR spectra were internally referenced to the proton (<sup>1</sup>H) of the internal TMS signal at 0.00 ppm or the solvent residue of DMSO at 2.54 ppm and the residual carbon nuclei (<sup>13</sup>C) of the solvent at 77.0 or 39.5 ppm, respectively. Data for <sup>1</sup>H NMR were recorded as follows: chemical shift ( $\delta$ , ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet or unresolved, coupling constant(s) in Hz, integration). High resolution mass spectra were obtained using Bruker ESI-QTOF mass spectrometry.

### 2. Preparations of tryptophol and tryptamine substrates



#### 2.1 Preparations of tryptophol substrates

Substrates **1a-1d**, **1f-1o** are known compounds, the general procedure for synthesis of these substrates are described as below according to previous literatures.<sup>[1-3, 7]</sup>

**General procedures:** To a solution of indole (10.0 mmol) in dry Et<sub>2</sub>O (50 mL) at 0  $^{\circ}$ C was added dropwise methyl oxalyl chloride (2.7 mL, 30.0 mmol). The ice bath was removed and the resultant yellow slurry was stirred at room temperature for 6 h. The crude reaction mixture was filtered with celite and washed with cold Et<sub>2</sub>O. The solid **S1** was used directly for the next step without further purification. A solution of **S1** in THF (20 mL) was added dropwise to a suspension of LiAlH<sub>4</sub> (1.52 g, 40 mmol) in THF (40 mL) at 0  $^{\circ}$ C. The solution was stirred at 80  $^{\circ}$ C for 2 h and quenched carefully by H<sub>2</sub>O (1.5 mL), 10% aqueous NaOH (3.0 mL), H<sub>2</sub>O (4.5 mL) at 0  $^{\circ}$ C. The solution was then filtered and washed with EtOAc. The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub> and the solvent was removed under reduced pressure to give the crude product. The crude product was purified by silica gel column chromatography (EtOAc/hexane = 1:3) to afford the tryptophol **S2.** 

TBSCl (1.66 g, 11 mmol) was added to a solution of S2 (10.0 mmol) and imidazole (1.36 g, 20.0 mmol) in DMF (50 mL) at 0 °C. The ice bath was then removed and the reaction mixture was stirred at room temperature for 3 h. The mixture was quenched with water and extracted with EtOAc (3×30 mL), then the combined organic layers were washed with water, brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was used directly for the next step without further purification.

To a solution of **S3** in THF (50 mL) was added NaH (400 mg, 10.0 mmol) at 0 °C. After stirring at 0 °C for 15 min and then at room temperature for 1 h, the reaction mixture was cooled to 0 °C, treated with Boc<sub>2</sub>O or TsCl or CbzCl or 4-NsCl (11.0 mmol), and then allowed to stir at room temperature for 6-12 h. After the reaction was complete (monitored by TLC), aqueous saturated NaHCO<sub>3</sub> (30 mL) was added slowly. The organic layer was separated and the aqueous layer was extracted with EtOAc ( $3 \times 30$  mL). The combined organic layers were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product **S4** was further treated with tetrabutylammonium fluoride (15 mmol), after 24 h, the mixture was worked up and purified by silica gel column chromatography (EtOAc/hexane = 1/2) to afford the desired product **1**.

The physical datas for new compounds were provided below:

#### 2-(1-(ethylsulfonyl)-1H-indol-3-yl)ethan-1-ol (1e)

Following the general procedure for preparations of tryptophol substrates. The procedure was performed on tryptophol (0.4836 g, 2 mmol) to afford **1e**, Light yellow oil, 290.6 mg, 57% yield; <sup>1</sup>H SO<sub>2</sub>Et **NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.79 (d, *J* = 8.2 Hz, 1H), 7.49 **1e** (d, *J* = 7.8 Hz, 1H), 7.26–7.17(m, 3H), 3.79 (t, *J* = 6.6 Hz, 2H), 3.16 (q, *J* = 7.4 Hz, 2H), 2.84 (t, *J* = 6.6 Hz, 2H), 1.09 (t, *J* = 7.4 Hz, 3H) ppm; <sup>13</sup>C **NMR** (150 MHz, Chloroform-*d*)  $\delta$  135.2, 130.5, 124.7, 124.0, 123.0, 119.5, 118.3, 113.0, 61.5, 48.3, 28.1, 7.8 ppm; **HRMS** (**ESI**) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>12</sub>H<sub>15</sub>NO<sub>3</sub>SNa 276.0665; Found 276.0654.

### 2.2 Preparations of tryptamine substrates



Substrates **4a**, **4b**, **4c**, **4f**, **4g**, **4h**, **4i**, **4j**, **4l**, **4m** were prepared according to the published procedures<sup>[1, 5]</sup>.

**General procedures**: To a solution of 1-dimethylamino-2-nitroethylene (20.0 mmol) in trifluoroacetic acid (15 mL) was added indole (20.0 mmol) at 0 °C. The mixture was allowed to warm to room temperature and stirred for 0.5 hours. The reaction mixture was basified to pH 9~10 using saturated NaHCO<sub>3</sub> in aqueous solution and extracted with DCM. The combined organic layers were washed with saturated brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo to furnish the desired crude nitroolefin. The crude nitroolefin was purified by flash column chromatography (Petroleum ether/ EtOAc = 3/1) to give the tryptophan derivative **S5**.

Under a nitrogen atmosphere, the THF solution (10 mL/mmol of **S5**) of **S5** (1.0 equiv.) was added to a stirred slurry of LiAlH<sub>4</sub> (6.0 equiv.) in THF at 0  $^{\circ}$ C. The mixture was allowed to warm to room temperature and after completion as detected by TLC. After the reaction mixture was cooled to 0°C, the reaction was quenched by dropwise addition of H<sub>2</sub>O until effervescence ceased and followed by three times the amount of H<sub>2</sub>O of 15% aqueous sodium hydroxide. After being stirred at room temperature for 12 hours, the solid was removed by filtration (or the mixture was filtered through Celite pad to remove by-product). The filtrate was concentrated to dryness to give crude product **S6**.

The tryptamine derivative **S6** was dissolved in  $CH_2Cl_2$  (2 mL/mmol of **S6**) and 10% aqueous  $Na_2CO_3$  (1 mL/mmol of **S6**), and then to the reaction mixture was added Boc<sub>2</sub>O or CbzCl or isobutyl chlorocarbonate (2 equiv.). After completion as detected by TLC, the reaction mixture was diluted with water. The organic layer was collected

and the aqueous phase was extracted with EtOAc. The combined extracts were washed with brine, dried over  $Na_2SO_4$ , filtered and concentrated *in vacuo* to give the desired tryptamine derivative **S7**, which was used for the next step without further purification.

To a stirred solution of tryptamine derivative **S7** in CH<sub>2</sub>Cl<sub>2</sub> (3 mL/mmol of **S6**) were sequentially added Bu<sub>4</sub>NHSO<sub>4</sub> (0.1 equiv.) and NaOH (4.5 equiv.) at 0 °C. The resulting mixture was stirred at room temperature for 5 minutes before addition of TsCl or Boc<sub>2</sub>O (2 equiv.). The reaction mixture was allowed to stir at room temperature and after completion as detected by TLC. Water was added, and the mixture was extracted with EtOAc. The combined organic phases were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was purified by flash column chromatography (Petroleum ether/EtOAc = 10/1) to give the tryptophan derivative **4**.

The physical datas for new compounds were provided below:

## *tert*-butyl-3-(2-((tert-butoxycarbonyl)amino)ethyl)-6-chloro-1*H*-indole-1-carboxy late (4d)



Following the general procedure for synthesis of the tryptophan derivative. The procedure was performed on 6-Chloroindole (1.51 g, 10 mmol) to afford **4d**. Yellow solid, 2.12 g, 54% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  8.17 (s, 1H), 7.44 (d, *J* = 8.2 Hz, 1H), 7.39 (s, 1H), 7.21 (dd, *J* = 8.2, 1.8 Hz, 1H), 4.63

(s, 1H), 3.45–3.41 (m, 2H), 2.86 (t, J = 7.0 Hz, 2H), 1.67 (s, 9H), 1.44 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  155.8, 149.3, 130.5, 128.9, 123.6, 123.0, 119.7, 117.6, 115.6, 84.1, 79.4, 40.2, 28.4, 28.1, 25.5 ppm; HRMS (ESI) m/z: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>27</sub>N<sub>2</sub>O<sub>4</sub>ClNa 417.1552; Found 417.1551.

# *tert*-butyl-3-(2-((tert-butoxycarbonyl)amino)ethyl)-6-methyl-1H-indole-1-carboxy late(4e)



Following the general procedure for synthesis of the tryptophan derivative. The procedure was performed on 6-Methylindole (1.31 g, 10 mmol) to afford **4e**. Brown oil, 717.9 mg, 19% yield; <sup>1</sup>H NMR

(600 MHz, Chloroform-*d*)  $\delta$  8.00 (s, 1H), 7.40 (d, J = 8.0 Hz, 1H), 7.33 (s, 1H), 7.07 (d, J = 7.8 Hz, 1H), 4.69 (s, 1H), 3.46–3.43 (m, 2H), 2.87 (t, J = 7.0 Hz, 2H), 2.48 (s, 3H), 1.66 (s, 9H), 1.44 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  155.9, 149.7, 136.0, 134.4, 128.1, 123.8, 122.4, 118.5, 117.7, 115.5, 83.2, 79.1, 40.2, 28.3, 28.1, 25.6, 21.9 ppm; **HRMS (ESI)** m/z: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>30</sub>N<sub>2</sub>O<sub>4</sub>Na 397.2098; Found 397.2095.

#### tert-Butyl 3-(2-((isopropoxycarbonyl)amino)ethyl)-1H-indole-1-carboxylate (4k)



J = 6.8 Hz, 2H), 3.53–3.49 (m, 2H), 2.91 (t, J = 7.0 Hz, 2H), 1.91–1.87 (m, 1H), 1.67 (s, 9H), 0.91 (d, J = 6.8 Hz, 6H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  156.7, 149.6, 135.5, 130.3, 124.4, 123.1, 122.5, 118.9, 117.6, 115.3, 83.5, 70.9, 40.5, 28.2, 28.0, 25.6, 19.0 ppm; HRMS (ESI) m/z: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>28</sub>N<sub>2</sub>O<sub>4</sub>Na 383.1941; Found 383.1938.

### 3. Optimization of the reaction conditions

### 3.1 Electrolysis general informations

Electrochemical reactions were performed with ElectraSyn 2.0 package (IKA) using the constant current or constant voltage modes. The reactions were conducted in a 10 mL vial for 4 mL of solvent with a stir bar and a carbon graphite-SK-50 ( $5.0 \times 0.8 \times 0.2$  cm) working electrode (anode and cathode) with a distance of 0.6 cm between the two electrodes.



	R A (0.2 mm	`ОН с∎ <sup>  </sup> 5 m/	$ \xrightarrow{A, rt} \xrightarrow{X} \xrightarrow{R} $	-0 +	×	R
	I (0.2 mm	01)	2	X = Br or	U 3	
Enter	Entry R Electroly	Flootroluto	Solvent	t (b)	Yield 2	Yield 3
Entry		Electrolyte	(mL)	<i>t</i> (n)	(%)	(%)
1	Н	NaBr	MeCN (4)	24	0	0
2	Boc	NaBr	MeCN (4)	24	69	0
3	Boc	NaBr	MeCN/EtOH (4/0.1)	24	NR	NR
4	Boc	NaBr	MeCN/H <sub>2</sub> O (4/0.1)	4	83	0
5	Boc	NaCl	MeCN (4)	24	Trace	NR
6	Boc	NH <sub>4</sub> Cl	MeCN (4)	24	NR	NR

### 3.2 Screening of protecting group, solvent and electrolyte <sup>a</sup>

<sup>a</sup> Reaction conditions: undivided cell, carbon cloth anode and cathode, **1** (0.2 mmol), electrolyte (0.4 mmol), CH<sub>3</sub>CN (4 mL), CCE = 5.0 mA, 4 h, rt, under air. Yields were isolated yields. CCE = constant current electrolysis.

(1 - 1) $(1 - 1)$ $(1 -$						
Entry	Solvent (mL)	<i>t</i> (h)	Yield <b>2a</b> (%)	Yield <b>3a</b> (%)		
1	MeCN/H <sub>2</sub> O (4/0.1)	4	83	0		
2	MeCN/H <sub>2</sub> O (4/0.3)	7	77	0		
3	MeCN/H <sub>2</sub> O (4/0.5)	7	68	0		
4	MeCN/H <sub>2</sub> O (3/1)	12	39	31		
5	MeCN/H <sub>2</sub> O (2.5/1.5)	12	trace	75		
6	MeCN/H <sub>2</sub> O (2/2)	12	trace	34		
7	MeCN/H <sub>2</sub> O (1/3)	12	trace	trace		
8	H <sub>2</sub> O (4)	12	trace	0		

### **3.3 Screening of the amounts of water** <sup>*a*</sup>

<sup>a</sup> Reaction conditions: undivided cell, carbon cloth anode and cathode, **1a** (0.2 mmol), electrolyte (0.4 mmol), CCE = 5.0 mA, rt, under air. Yields were isolated yields. CCE = constant current electrolysis.

### 3.4 Screening of the amounts of NaBr<sup>a</sup>

	OH N Boc	C NaBr (x eq.) 5 mA, rt CH <sub>3</sub> CN/H <sub>2</sub> O (2.5/1.5)	Br N Boc	+ Br N Boc	
1a (	(0.2 mmol)		2a	3a	
Entry	х	Yield <b>2a</b> (%	<b>b</b> )	Yield <b>3a</b> (%)	
1	1.5	trace		64	
2	2	trace		75	
3	3	10		67	

<sup>a</sup> Reaction conditions: undivided cell, carbon cloth anode and cathode, **1a** (0.2 mmol), CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5), CCE = 5.0 mA, 12 h, rt, under air. Yields were isolated yields. CCE = constant current electrolysis.

### 4. Mechanistic studies

### **4.1 Kinetic experiments**



**Reaction procedure**: A mixture of **1a** (0.2 mmol) and NaBr (41.2 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5 mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA between a carbon graphite anode and carbon graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring. The reactions were stopped respectively at 0.5 h, 1 h, 1.5 h, 2 h, 2.5 h, 3 h, 4 h, 5 h, 8 h and 12 h. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. Yields were determined by <sup>1</sup>H NMR with TTCE as internal standard. The plots of the percentage yield of the product displayed in Figure S1.

Entry	<i>t</i> (h)	Yield <b>2a</b> (%)	Yield 11 (%)	Yield <b>3a</b> (%)	Recoved yield 1a (%)
1	0	0	0	0	100
2	0.5	13	6	0	80
3	1	18	10	3	65
4	1.5	28	13	5	53
5	2	36	20	7	42
6	2.5	28	25	13	33
7	3	22	28	18	27
8	4	18	40	24	15
9	5	14	31	30	8
10	6.5	10	23	43	0
11	8	8	18	55	0
12	10	4	9	68	0
13	12	2	4	75	0



Figure S1 Results of kinetic experiments

# 4.2 Investigation of the hydrodebromination process and isotopic experiment using $D_2O$



General Procedure for deuterium labelling experiments: A mixture of 2a (0.2 mmol) and NaBr (41.2 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA a carbon graphite anode and carbon graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring for 10 h. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to give compound **3a** and **12**.

*tert*-Butyl-5-bromo-2,3,3a,8a-tetrahydro-8H-furo[2,3-*b*]indole-8-carboxylate-3a-d (d-3a)



Chloroform-*d*)  $\delta$  152.2, 131.0, 127.4, 115.8, 115.1, 93.5, 66.4, 33.6, 28.3 ppm. **HRMS** (ESI) m/z:  $[M + Na]^+$  Calcd for C<sub>15</sub>H<sub>17</sub>DBrNO<sub>3</sub>Na 363.0425; Found 363.0418.

## *tert*-Butyl-3a,5-dibromo-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxylate (12)



ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

### 4.3 TEMPO trapping experiments and HRMS-ESI analysis



Procedure for HRMS-ESI analysis: A mixture of 2a (0.2 mmol), NaBr (41.2 mg,

0.4 mmol) and TEMPO (62.5 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5 mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA between a carbon graphite anode and carbon graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring. After 4 hours of reaction, send it to HRMS-ESI for monitoring.

#### 4.4 Investigation of 3-bromopyrroloindoline

The reaction activities of 3-bromopyrroloindoline 5a were also investigated. When the 3-bromopyrroloindoline 5a was employed in the standard conditions, 51% yield of 5-bromopyrroloindoline 6a and 13% yield of 3,5-dibromopyrroloindoline were obtained, providing further evidence for the formation of 5-bromoindoline via key 3-bromoindoline intermediate.



**General Procedures:** A mixture of **5a** (0.2 mmol) and NaBr (41.2 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5 mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA a carbon graphite anode and carbon graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/ EtOAc = 10/1) to give compound **6a** and **16**.

# Di*-tert*-Butyl-3a,5-dibromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dicarbo xylate (16)



Yellow oil, 6 h, 13.7 mg, 13% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.50–7.47 (m, 2H), 7.39 (dd, J = 8.6, 2.2 Hz, 1H), 6.42 (s, 1H), 3.78–3.75 (m, 1H), 2.85–3.80 (m, 1H), 2.75– 2.67 (m, 2H), 1.58 (s, 9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150

MHz, Chloroform-*d*)  $\delta$  153.3, 151.8, 141.2, 134.7, 133.2, 126.9, 118.7, 116.2, 84.2, 82.5, 80.9, 61.0, 46.2, 28.3, 28.2 ppm; **HRMS** (**ESI**) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub>Br<sub>2</sub>Na 539.0152; Found 539.0142.

### 4.5 Investigation of 5-bromonation of indolines



**Procedures for substrate 11**: A mixture of **3a** (68.9 mg, 0.2 mmol), Triphenylphosphine (5.3 mg, 0.02 mmol),  $Pd(OAc)_2$  (2.2 mg, 0.01 mmol),  $Cs_2CO_3$ (97.8 mg, 0.3 mmol) was dissolved in Toluene/MeOH (4:1) under argon atmosphere. Then, the mixture was stirred at 80 °C and when the reaction was completed as monitored by TLC. The reaction was cooled to room temperature. The resulting solid was filtered off and the solvent was concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to obtain **11**.

#### tert-butyl -2, 3, 3a, 8a-tetrahydro-8H-furo[2, 3-b]indole-8-carboxylate (11)

Colorless oil, 16 h, 39.2 mg, 75%; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.81 (s, 1H), 7.20–7.15 (m, 2H), 6.99 (t, *J* = 7.4 Hz, 1H), 6.15 (br, 1H), 3.97 (t, *J* = 8.0 Hz, 1H), 3.93 (t, *J* = 7.4 Hz, 1H), 3.46–3.41 (m, 11 1H), 3.35–2.28 (m, 1H), 2.10–2.07 (m, 1H), 1.59 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  152.4, 142.9, 131.1, 128.1, 124.3, 122.7, 114.3, 93.3, 81.2, 66.3, 44.8, 33.8, 28.3 ppm; HRMS (ESI) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>19</sub>O<sub>3</sub>NNa 284.1257; Found 284.1255.



**Procedures for 5-bromination of 11**: A mixture of **11** (0.2 mmol) and NaBr (41.2 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5 mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA between a carbon graphite anode and carbon graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to give compound **3a**.

#### tert-Butyl 5-bromoindoline-1-carboxylate (15)



White solid, m.p. 102.7 – 103.9 °C, 10 h, 21.0 mg, 35% yield; <sup>1</sup>H **NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.71 (br, 1H), 7.25–7.23 (m, 2H), <sup>c</sup> 3.97 (t, *J* = 8.8, 2H), 3.06 (t, *J* = 8.6 Hz, 2H), 1.55 (s, 9H) ppm; <sup>13</sup>C **NMR** (150 MHz, Chloroform-*d*)  $\delta$  152.4, 142.3, 130.2, 127.6, 116.0,

114.4, 47.7, 28.4 ppm; **HRMS (ESI)** m/z:  $[M + Na]^+$  Calcd for  $C_{13}H_{16}O_2NBrNa$  320.0257; Found 320.0249.

### 4.6 P(OEt)<sub>3</sub> trapping experiments

In order to explore the oxidative activities between bromide and **1a** in the current conditions, several control experiments have been performed. The electrochemical indole radical cation intermediate could be trapped by  $P(EtO)_3$  have been frequently identified.<sup>8</sup> According to Lei's work<sup>8a</sup>, when 6 equivalent of  $P(EtO)_3$  was added in the reaction, 20% yield of 3a-bromoindoline **2a** and 63% yield of 5a-bromoindoline **3a** was obtained, but none of indole-phosphorylation adduct was detected by HRMS analysis. Besides, when 3-methyl indole was employed in the standard condition, the reaction did not take place and none of indole-phosphorylation product was detected. These results suggested the preferred oxidation of **1a** to indole radical cation intermediate may not involve in this reaction.



### 4.7 Investigation of NaCl and NaI

When NaCl was used in the standard conditions instead of NaBr, only 28% yield of 3a-chloroindolines was obtained in the less water conditions, and none of 5achloroindoline was isolated, and most substrates decomposed. When the NaI was used in the standard conditions instead of NaBr, none of any 3a- or 5a-chloroindoline was detected, and part of substrates decomposed. We assumed that the reactivity of Br<sup>-</sup> is the key to the reaction.



*tert*-butyl-3a-chloro-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carboxylate

(2a')

Light yellow oil, 3.5 h, 16.7 mg, 28% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.87 (br, 1H), 7.40 (d, *J* = 7.6 Hz, 1H), 7.31 (t, *J* = 7.8 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.08 (s, 1H), 4.09 (t, *J* = 8.2 Hz, 1H), 3.60–3.49 (m, 1H), 2.82–2.77 (m, 1H), 2.71–2.68 (m, 1H), 1.60 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[3]</sup>.

### 4.8 Investigation N-H/Me substrate

When the N-Me and N-H substituted tryptophol and tryptamine substrates were employed in the standard conditions, none of 3a-bromoindolines or 5a-bromoindolines was obtained, and the substrates decomposed after a period of time. We assumed that the electron-rich of indole was unstable under the current electronic oxidative conditions, an electron-withdrawing group is necessary for the reaction.



### 5. Cyclic voltammetry studies

Cyclic voltammetry were performed with CORRTEST potentiostat/galvanostat CS310H. A glassy carbon disc (diameter 3 mm) working electrode, a platinum wire counter electrode and a saturated silver chloride electrode as reference electrode were used at scan rate of 100 mV/s. The experiments were conducted in a 50 mL vial without stirring in CH<sub>3</sub>CN/H<sub>2</sub>O (10 mL/0.25 mL) or CH<sub>3</sub>CN/H<sub>2</sub>O (6.25 mL/3.73 mL) with **1a** (5 mmol/L) and/or NaBr (5 mmol/L) and *n*Bu<sub>4</sub>NBF<sub>4</sub> (0.05 M).



Figure S2 Cyclic voltammograms of *n*-Bu<sub>4</sub>NBF<sub>4</sub> (0.05 M) in CH<sub>3</sub>CN/H<sub>2</sub>O (4/0.1)



Figure S3 Cyclic voltammograms of *n*-Bu<sub>4</sub>NBF<sub>4</sub> (0.05 M) in CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5)

# 6. General Procedure for cyclic 3a- and 5a-bromoindolins and product characterizations



General Procedure for cyclic 3a-bromoindolins: A mixture of 1a (0.2 mmol) and NaBr (41.2 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (4/0.1 mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA a carbon graphite anode and carbon graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to give compound **2a**.



General Procedure for cyclic 5a-bromoindolins: A mixture of 1a (0.2 mmol) and NaBr (41.2 mg, 0.4 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (2.5/1.5 mL) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA between a carbon graphite anode and carbon graphite cathode (1.4  $\times$  0.8  $\times$  0.2 cm submerged in solution) with stirring. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to give compound **3a**.

*tert*-Butyl-3a-bromo-2,3,3a,8a-tetrahydro-8H-furo[2,3-*b*]indole-8-carboxylate (2a)



2b

White solid, 4 h, 56.3 mg, 83% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.83 (br, 1H), 7.40 (d, J = 7.4 Hz, 1H), 7.28 (t, J = 7.8 Hz, 1H), 7.07 (td, J = 7.6, 0.9 Hz, 1H), 6.19 (br, 1H), 3.99 (t, J = 8.6 Hz, 1H), 3.50–3.46 (m, 1H), 2.87–2.92(m, 1H), 2.81–2.78 (m, 1H),

1.60 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

#### Benzyl-3a-bromo-2,3, 3a, 8a-tetrahydro-8H-furo[2, 3-b]indole-8-carboxylate (2b)

Colorless oil, 5 h, 58.4 mg, 79% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.88 (br, 1H), 7.46–7.31 (m, 7H), 7.10 (t, *J* = 7.4 Hz, 1H), 6.29 (s, 1H), 5.39–5.29 (m, 2H), 4.01 (t, *J* = 8.0 Hz, 1H), 3.53– 3.48 (m, 1H), 2.92–2.87 (m, 1H), 2.82–2.79 (m, 1H) ppm. All spectral

data were in agreement with the literature <sup>[1]</sup>.

#### 3a-Bromo-8-tosyl-3, 3a, 8, 8a-tetrahydro-2H-furo[2,3-b]indole (2c)

#### 3a-Bromo-8-(phenylsulfonyl)-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (2d)



Light yellow oil, 6 h, 54.8 mg, 72% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.91 (d, *J* = 7.4 Hz, 2H), 7.54 (t, *J* = 7.6 Hz, 1H), h 7.51 – 7.42 (m, 3H), 7.34 (d, *J* = 7.6 Hz, 1H), 7.30 – 7.25 (m, 1H), 7.10 (t, *J* = 7.6 Hz, 1H), 6.23 (s, 1H), 3.99 (t, *J* = 8.2 Hz, 1H), 3.44 –

3.37 (m, 1H), 2.85 – 2.80 (m, 1H), 2.73 (dd, J = 12.7, 4.1 Hz, 1H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  140.3, 138.5, 133.4, 132.4, 130.6, 129.0, 127.2, 125.2, 124.9, 114.2, 103.1, 68.0, 61.2, 44.5 ppm; HRMS (ESI) *m*/*z*: [M + Na]<sup>+</sup> Calcd for

C<sub>16</sub>H<sub>14</sub>NO<sub>3</sub>BrSNa 401.9770; Found 401.9755.

#### 3a-Bromo-8-(ethylsulfonyl)-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (2e)

white solid, m.p. 97.3 – 98.5 °C, 6 h, 43.8 mg, 66% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.45 (dd, *J* = 7.6, 0.8 Hz, 1H), 7.38 (d, *J* = 8.2 Hz, 1H), 7.30 (td, *J* = 7.8, 1.4 Hz, 1H), 7.14 (td, *J* = 7.6, 1.0 Hz, **2e** 1H), 6.19 (s, *J* = 0.4 Hz, 1H), 4.07–4.04 (m, 1H), 3.56 –3.52 (m, 1H), 3.34 – 3.21 (m, 2H), 2.94 –2.89 (m, 1H), 2.86 – 2.80 (m, 1H), 1.42 (t, *J* = 7.4 Hz, 3H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  140.8, 131.8, 130.8, 125.4, 124.5, 113.1, 102.7, 68.1, 61.4, 47.8, 44.4, 7.8 ppm; **HRMS (ESI)** *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>12</sub>H<sub>14</sub>NO<sub>3</sub>BrSNa 353.9770; Found 353.9757.

# *Tert*-butyl-3a-bromo-4-methyl-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carbo xylate (2f)



Colorless oil, 3.5 h, 43.2 mg, 61% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.71 (br, 1H), 7.20 (t, *J* = 7.8 Hz, 1H), 6.85 (d, *J* = 7.6 Hz, 1H), 6.21 (s, 1H), 4.04–3.96 (m, 1H), 3.61–3.57 (m, 1H), 2.91–

2.83 (m, 2H), 2.49 (s, 3H), 1.60 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[4]</sup>.

### *Tert*-butyl-3a-bromo-4-fluoro-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carbo xylate (2g)



Colorless oil, 3 h, 54.0 mg, 75% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.64 (s, 1H), 7.32–7.24 (m, 2H), 6.73 (t, *J* = 8.8 Hz, 2H), 6.18 (s, 1H), 4.03 (t, *J* = 7.8 Hz, 2H), 3.58–3.54 (m, 2H), 3.02–2.98 (m, 2H), 2.87–2.82 (m, 2H), 1.60 (s, 9H) ppm. All spectral data

were in agreement with the literature<sup>[3]</sup>.

3a-Bromo-5-chloro-8-tosyl-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (2h)



Yellow solid, 7 h, 52.6 mg, 61% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.77 (d, J = 8.4 Hz, 2H), 7.42 (d, J = 8.6 Hz, 1H), 7.30 (d, J = 2.2 Hz, 1H), 7.26 (d, J = 8.2 Hz, 2H), 7.24 (dd, J = 8.6, 2.2 Hz, 1H), 6.21 (s, 1H), 4.04–4.01 (m, 1H), 3.47–3.43 (m, 1H),

2.85-2.79 (m, 1H), 2.71-2.68 (m, 1H), 2.38 (s, 3H) ppm. All spectral data were in agreement with the literature <sup>[3]</sup>.

## *tert*-Butyl-3a-bromo-5-chloro-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carbo xylate (2i)



Light yellow oil, 6 h, 49.2 mg, 66% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.78 (s, 1H), 7.36 (s, 1H), 7.24 (d, J = 8.2 Hz, 1H), 6.16 (s, 1H), 4.01 (t, J = 8.4 Hz, 1H), 3.51–3.47 (m, 1H), 2.90–2.85 (m, 1H), 2.77–2.75 (m, 1H), 1.59 (s, 9H) ppm. All

spectral data were in agreement with the literature <sup>[3]</sup>.

#### Tert-butyl-3a-bromo-6-chloro-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carbo

xylate (2j)



ppm. All spectral data were in agreement with the literature<sup>[3]</sup>.

# *tert*-butyl-3a-bromo-6-fluoro-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carbox vlate (2k)



9H). All spectral data were in agreement with the literature <sup>[4]</sup>.

# *tert*-Butyl-3a-bromo-7-methyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carbo xylate (2l)



White solid, 9 h, 65.6 mg, 93% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.25 (dd, J = 7.4, 1.6 Hz, 1H), 7.14–7.09 (m, 2H), 6.18 (s, 1H), 3.97–3.94 (m, 1H), 3.43–3.39 (m, 1H), 2.89–2.84 (m, 1H), 2.80–2.76 (m, 1H), 2.32 (s, 3H), 1.57 (s, 9H) ppm. All spectral

data were in agreement with the literature <sup>[2]</sup>.

## *tert*-Butyl-3a-bromo-7-ethyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carbox vlate (2m)



Yellow oil, 6 h, 68.5 mg, 93% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.25 (d, J = 7.4 Hz, 1H), 7.21 (d, J = 7.4 Hz, 1H), 7.16 (t, J = 7.6 Hz, 1H), 6.18 (s, 1H), 3.96–3.93 (m, 1H), 3.42–3.38 (m, 1H), 2.89–2.84 (m, 1H), 2.80–2.60 (m, 3H), 1.57 (s, 9H), 1.18 (t, J = 7.4 Hz, 3H) ppm. All spectral data were in agreement with the

literature<sup>[2]</sup>.

### tert-butyl-3a,7-dibromo-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carboxylate (2n)



Colorless oil, 4 h, 38.3 mg, 46% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.51 (dd, J = 8.2, 1.2 Hz, 1H), 7.37 (dd, J = 7.6, 1.0 Hz, 1H), 7.06 (t, J = 7.8 Hz, 1H), 6.15 (s, 1H), 4.00–3.98 (m, 1H), 3.48–3.44 (m, 1H), 2.89–2.84 (m, 1H), 2.78–2.75 (m, 1H), 1.58 (s,

9H) ppm. All spectral data were in agreement with the literature<sup>[3]</sup>.

# *Tert*-Butyl-3a-bromo-8a-phenyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-car boxylate (20)



Colorless oil, 9 h, 75.0 mg, 90% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.97–7.87 (m, 1H), 7.65–7.30 (m, 6H), 7.20–7.08 (m, 2H), 4.21 (t, *J* = 8.0 Hz, 1H), 3.69–3,65 (m, 1H), 2.93–2.86 (m, 1H), 2.83–2.77 (m, 1H), 1.16 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

tert-Butyl -5-bromo-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxylate (3a)



Light yellow oil, 12 h, 50.8 mg, 75% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.70 (br, 1H), 7.31–7.28 (m, 2H), 6.14 (br, 1H), 3.99 (t, *J* = 8.0 Hz, 1H), 3.92 (t, *J* = 7.6 Hz, 1H), 3.47–3.43 (m, 1H), 2.35–2.28 (m, 1H), 2.10–2.07 (m, 1H), 1.58 (s, 9H) ppm.<sup>13</sup>C

**NMR** (150 MHz, Chloroform-*d*)  $\delta$  152.2, 131.0, 127.3, 115.8, 115.1, 93.5, 66.3, 58.4, 33.7, 28.3, 18.4 ppm; **HRMS** (**ESI**) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>18</sub>O<sub>3</sub>NBrNa 362.0362; Found 362.0357.

#### Benzyl-5-bromo-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxylate (3b)



Colorless oil, 20 h, 64.5 mg, 86% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.74 (br, 1H), 7.45–7.29 (m, 7H), 6.23 (s, 1H), 5.41–5.25 (m, 2H), 4.01 (t, *J* = 8.2 Hz, 1H), 3.94 (t, *J* = 7.6 Hz, 1H), 3.49–3.45 (m, 1H), 2.36–2.29 (m, 1H), 2.11–2.08 (m, 1H) ppm; <sup>13</sup>C

**NMR** (151 MHz, Chloroform-*d*)  $\delta$  152.7, 141.8, 136.0, 133.4, 131.2, 128.6, 128.2, 127.9, 127.4, 115.8, 115.6, 93.35, 67.36, 66.52, 44.99, 33.67 ppm; **HRMS** (**ESI**) *m/z*:  $[M + Na]^+$  Calcd for C<sub>18</sub>H<sub>16</sub>O<sub>3</sub>NBrNa 396.0206; Found 396.0205.

#### 5-Bromo-8-tosyl-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (3c)



White solid, m.p. 88.0 – 89.2 °C, 20 h, 40.2 mg, 51% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.83 (d, J = 8.2 Hz, 2H), 7.29– 7.25 (m, 6H), 6.24 (d, J = 6.6 Hz, 1H), 3.98 (t, J = 8.2 Hz, 1H), 3.88 (t, J = 7.6 Hz, 1H), 3.33–2.28 (m, 1H), 2.39 (s, 3H), 2.32–2.26

(m, 1H), 2.02–1.99 (m, 1H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  144.2, 140.8, 136.2, 133.7, 131.3, 129.7, 128.0, 127.4, 116.0, 114.3, 96.0, 66.4, 45.3, 33.5, 21.5 ppm; **HRMS (ESI)** *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>17</sub>H<sub>16</sub>O<sub>3</sub>NBrNaS 415.9926; Found 415.9916.

5-Bromo-8-(phenylsulfonyl)-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (3d)



3.30 (m, 1H), 2.34 – 2.25 (m, 1H), 2.00 (dd, J = 12.2, 4.8 Hz, 1H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  140.6, 139.2, 133.7, 133.2, 131.3, 129.0, 128.0, 127.3, 116.1, 114.21, 96.0, 66.4, 45.3, 33.5 ppm; HRMS (ESI) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>14</sub>NO<sub>3</sub>BrSNa 401.9770; Found 401.9757.

#### 5-Bromo-8-(ethylsulfonyl)-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (3e)



3e

white solid, m.p. 93.0 – 94.1 °C, 15 h, 34.4 mg, 52% yield; <sup>1</sup>H **NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.33 – 7.29 (m, 2H), 7.22 (d, J = 8.5 Hz, 1H), 6.18 (d, J = 6.4 Hz, 1H), 4.06 (t, J = 7.8 Hz, 1H), 4.02 (t, J = 7.4 Hz, 1H), 3.51 – 3.47 (m, 1H), 3.33 – 3.23 (m, 2H),

2.40 – 2.32 (m, 1H), 2.12 (dd, J = 12.4, 4.7 Hz, 1H), 1.41 (t, J = 7.4 Hz, 3H) ppm; <sup>13</sup>C **NMR** (150 MHz, Chloroform-*d*)  $\delta$  141.5, 133.0, 131.4, 128.1, 115.6, 113.6, 95.7, 66.8, 48.7, 45.4, 33.4, 7.6 ppm; **HRMS** (**ESI**) m/z: [M + Na]<sup>+</sup> Calcd for C<sub>12</sub>H<sub>14</sub>NO<sub>3</sub>BrSNa 353.9770; Found 353.9757.

# *Tert*-butyl-5-bromo-4-methyl-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carbox ylate (3f)



Colorless oil, 10.5 h, 25.2 mg, 36% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.53 (br, 1H), 7.37 (d, J = 8.4 Hz, 1H), 6.19 (s, 1H), 4.00–3.96 (m, 1H), 3.92 (t, J = 8.0 Hz, 1H), 3.51–3.46 (m, 1H), 2.35 (s, 3H), 2.34–2.29 (m, 1H), 2.03–2.01 (m, 1H), 1.59 (s,

9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  152.3, 142.5, 133.5, 131.8, 118.2, 113.1, 93.6, 81.7, 65.7, 44.6, 32.8, 28.3, 19.2 ppm; **HRMS (ESI)** *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>20</sub>O<sub>3</sub>NBrNa 376.0519; Found 376.0515.

tert-butyl-5-bromo-4-fluoro-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxy late (3g)

Colorless oil, 24 h, 50.3 mg, 70% yield; <sup>1</sup>H NMR (600 MHz,



Chloroform-*d*)  $\delta$  7.51 (s, 1H), 7.36 (t, *J* = 7.8 Hz, 1H), 6.18 (s, 1H), 4.06-4.02 (m, 2H), 3.50-3.46 (m, 1H), 2.33-2.18 (m, 2H), 1.58 (s, 3g 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  155.0 (d, <sup>1</sup>*J*<sub>C-F</sub> = 244.6 Hz), 151.9, 136.8, 133.1, 111.2 (d,  ${}^{3}J_{C-F} = 3.0$  Hz), 101.4 (d,  ${}^{2}J_{C-F} = 19.6$  Hz), 96.6, 93.9, 83.0, 66.5, 42.4, 31.9, 28.3 ppm; <sup>19</sup>F NMR (565 MHz, Chloroform-d) δ -115.49 ppm; **HRMS (ESI)** m/z:  $[M + Na]^+$  Calcd for C<sub>15</sub>H<sub>17</sub>O<sub>3</sub>NFBrNa 380.0268; Found 380.0260.

#### 5-Chloro-8-tosyl-3,3a,8,8a-tetrahydro-2H-furo[2,3-b]indole (3h)



Solid, m.p. 106.6 – 107.0 °C, 20 h, 37.7 mg, 54% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.83 (d, J = 8.4 Hz, 2H), 7.31 (d, J =8.6 Hz, 1H), 7.26 (d, J = 8.0 Hz, 2H), 7.14 (dd, J = 8.4, 2.2 Hz, 1H), 7.10 (s, 1H), 6.25 (d, J = 6.6 Hz, 1H), 3.98 (t, J = 8.0 Hz,

1H), 3.87 (t, J = 7.6 Hz, 1H), 3.33-3.29 (m, 1H), 2.39 (s, 3H), 2.32-2.26 (m, 1H), 2.01–1.99 (m, 1H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-d)  $\delta$  144.2, 140.3, 136.2, 133.3, 129.7, 128.7, 128.4, 127.4, 113.9, 96.1, 66.4, 45.4, 33.6, 21.5 ppm; HRMS (ESI) m/z:  $[M + Na]^+$  Calcd for C<sub>17</sub>H<sub>16</sub>O<sub>3</sub>NClNaS 372.0432; Found 372.0428.

#### tert-Butyl-5-Chloro-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxylate (3i)



Yellow solid, m.p. 66.3 - 67.5 °C, 8 h, 35.3 mg, 60% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-d) δ 7.75 (br, 1H), 7.16–7.14 (m, 2H), 6.15 (br, 1H), 3.99 (t, J = 8.2 Hz, 1H), 3.91 (t, J = 7.4 Hz, 1H), 3.47-3.42 (m, 1H), 2.35-2.29 (m, 1H), 2.09-2.07 (m, 1H),

1.59 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-d) δ 152.2, 141.7, 133.1, 128.1, 127.7, 124.4, 115.3, 93.5, 81.6, 66.3, 44.8, 33.7, 28.3 ppm; HRMS (ESI) m/z: [M + Na]<sup>+</sup> Calcd for  $C_{15}H_{18}O_3NCINa$  318.0867; Found 318.0865.

*tert*-Butyl-5-bromo-6-chloro-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carbox ylate (3j)



Colorless oil, 16 h, 42.8 mg, 57% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.97 (br, 1H), 7.37 (s, 1H), 6.15 (s, 1H), 4.00 (t, *J* = 8.2 Hz, 1H), 3.90 (t, *J* = 7.4 Hz, 1H), 3.47–3.43 (m, 1H), 2.35–2.28 (m, 1H), 2.08–2.05 (m, 1H), 1.59 (s, 9H) ppm; <sup>13</sup>C NMR

(150 MHz, Chloroform-*d*)  $\delta$  151.9, 143.2, 133.8, 128.8, 127.3, 116.0, 114.8, 93.8, 82.1, 66.4, 44.4, 33.7, 28.3 ppm; **HRMS** (**ESI**) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>17</sub>O<sub>3</sub>NClBrNa 395.9972; Found 395.9968.

# *tert*-Butyl-5-bromo-6-fluoro-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carboxy late (3k)



Light yellow oil, 22 h, 31.2 mg, 44% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.66 (s, 1H), 7.28 (d, *J* = 6.8 Hz, 1H), 6.16 (s, 1H), 4.00 (t, *J* = 8.0 Hz, 1H), 3.90 (t, *J* = 7.6 Hz, 1H), 3.48–3.43 (m, 1H), 2.34–2.28 (m, 2H), 2.07–2.05 (m, 1H), 1.59 (s, 9H) ppm;

<sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  158.8 (d, <sup>1</sup>*J*<sub>C-F</sub> = 241.5 Hz), 151.9, 128.1, 103.5 (d, <sup>2</sup>*J*<sub>C-F</sub> = 29.8 Hz), 101.2 (d, <sup>3</sup>*J*<sub>C-F</sub> = 22.5 Hz), 94.0, 66.4, 44.3, 33.8, 28.3 ppm; <sup>19</sup>**F NMR** (565 MHz, Chloroform-*d*)  $\delta$  -106.65 ppm; **HRMS** (**ESI**) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>15</sub>H<sub>17</sub>O<sub>3</sub>NFBrNa 380.0268; Found 380.0258.

## *tert*-Butyl-5-bromo-7-methyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carbox vlate (3l)



White solid, 14 h, 61.1 mg, 86% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.19 (s, 1H), 7.14 (s, 1H), 6.11 (d, J = 5.9 Hz, 1H), 4.02 (t, J = 7.0 Hz, 1H), 3.95 – 3.93 (m, 1H), 3.40 – 3.36 (m, 1H), 2.33 – 2.28 (m, 1H), 2.27 (s, 3H), 2.09 – 2.03 (m, 1H), 1.56

(s, 9H) ppm. <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  152.8, 140.6, 136.0, 133.4, 129.1, 124.4, 117.1, 95.7, 81.9, 66.7, 45.6, 33.0, 28.2, 20.2; HRMS (ESI) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>20</sub>O<sub>3</sub>NBrNa 376.0519; Found 376.0508.

*tert*-Butyl-5-bromo-7-ethyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxyl ate (3m)



**NMR** (150 MHz, Chloroform-*d*) δ 152.9, 139.8, 136.2, 135.1, 131.4, 124.4, 117.6, 95.7, 81.8, 66.6, 45.5, 33.1, 28.2, 26.0, 13.3 ppm; **HRMS** (**ESI**) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>17</sub>H<sub>22</sub>O<sub>3</sub>NBrNa 390.0675; Found 390.0671.

### *tert*-Butyl-5,7-dibromo-2,3,3a,8a-tetrahydro-8*H*-furo[2,3-*b*]indole-8-carboxylate (3n)



**NMR** (150 MHz, Chloroform-*d*)  $\delta$  151.9, 141.2, 138.3, 135.2, 126.2, 117.3, 111.8, 96.3, 82.6, 66.7, 46.2, 32.9, 28.1 ppm; **HRMS** (**ESI**) m/z:  $[M + Na]^+$  Calcd for C<sub>15</sub>H<sub>17</sub>O<sub>3</sub>NBr<sub>2</sub>Na 439.9467; Found 439.9455.

# *tert*-Butyl-5-bromo-8a-phenyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carbo xylate (30)



Colorless oil, 20 h, 56.2 mg, 68% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.83 (br, 1H), 7.38–7.36 (m, 3H), 7.31 (t, *J* = 7.4 Hz, 2H), 7.28–7.25 (m, 2H), 4.27 (t, *J* = 8.0 Hz, 1H), 3.79–3.72 (m, 2H), 2.40–2.34 (m, 1H), 2.03 (dd, *J* = 12.4, 5.0 Hz, 1H),

1.16 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ135.3, 132.0, 131.2, 128.1, 127.4, 127.2, 125.3, 124.4, 115.8, 115.0, 104.7, 68.1, 58.5, 55.6, 33.7, 27.8, 27.5 ppm; HRMS (ESI) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>22</sub>O<sub>3</sub>NBrNa 438.0675; Found 438.0668.

Di*-tert*-butyl-3a-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dicarboxyla te (5a)



1.49 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

# Di*-tert*-butyl-3a-bromo-4-fluoro-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (5b)



Light yellow oil, 12 h, 60.7 mg, 66% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.42 (s, 1H), 7.29–7.25 (m, 1H), 6.74 (t, *J* = 8.8 Hz, 1H), 6.44 (s, 1H), 3.79–3.76 (m, 1H), 3.07–3.04 (m, 1H), 2.92–2.87 (tm, 1H), 2.67–2.62 (m, 1H), 1.59 (s, 9H), 1.49 (s, 9H)

ppm. All spectral data were in agreement with the literature <sup>[5]</sup>.

# Di*-tert*-butyl-3a-bromo-4-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-di carboxylate (5c)



Following the general Procedure for cyclic 3a-bromoindolins. The procedure was performed on **4c** (0.2 mmol) and NaBr (24.7 mg, 0.24 mmol) to afford **5c**. Light yellow oil, 3 h, 70.8 mg, 78 % yield. <sup>1</sup>**H NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.47 (d, *J* = 8.6 Hz, 1H),

7.18 (t, J = 7.8 Hz, 1H), 6.85 (d, J = 7.6 Hz, 1H), 6.48 (s, 1H), 3.73 (dd, J = 11.2, 7.8 Hz, 1H), 3.00 (dd, J = 12.6, 4.8 Hz, 1H), 2.84 – 2.78 (m, 1H), 2.67-2.61 (m, J = 12.4, 7.8 Hz, 1H), 2.48 (s, 3H), 1.57 (s, 9H), 1.49 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[5]</sup>.

### Di*-tert*-butyl-3a-bromo-6-chloro-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (5d)


J = 8.2, 1.8 Hz, 1H), 6.44 (s, 1H), 3.77–3.74 (m, 1H), 2.84–2.79 (m, 1H), 2.75–2.67 (m, 2H), 1.59 (s, 9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 153.2, 151.7, 143.0, 136.1, 131.1, 124.7, 124.1, 117.6, 84.4, 82.7, 80.9, 61.4, 46.2, 28.3, 28.2 ppm; **HRMS (ESI)** m/z:  $[M + Na]^+$  Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>4</sub>N<sub>2</sub>BrClNa 495.0657; Found 495.0651.

### Di-tert-butyl-3a-bromo-6-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-di carboxylate (5e)



Following the general procedure for cyclic 3a-bromoindolines. The procedure was performed on 4e (0.2 mmol) and NaBr (24.7 mg, 0.24 mmol) to afford 5e. Colorless oil, 3 h, 47.3 mg, 58% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.46 (br, 1H), 7.24

(d, J = 7.8 Hz, 1H), 6.91 (d, J = 7.2 Hz, 1H), 6.42 (s, 1H), 3.71 (dd, J = 10.8, 7.6 Hz, 1H), 2.83 – 2.74 (m, 2H), 2.72– 2.67 (m, 1H), 2.35 (s, 3H), 1.59 (s, 9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-d) δ 153.4, 152.3, 142.2, 140.8, 129.8, 124.9, 123.4, 84.2, 82.0, 80.7, 62.5, 46.2, 28.4, 28.2, 21.9 ppm; **HRMS (ESI)** *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>29</sub>BrN<sub>2</sub>O<sub>4</sub>Na 475.1203; Found 475.1197.

### Di-tert-butyl-3a-bromo-7-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-di carboxylate (5f)



White solid, 9 h, 74.0 mg, 82% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.20 (dd, J = 6.6, 2.2 Hz, 1H), 7.14–7.11 (m, 2H), 6.24 (s, 1H), 3.55–3.52 (m, 1H), 2.82–2.81 (m, 1H), 2.76–2.67 (m, 2H), 2.30 (s, 3H), 1.54 (s, 9H), 1.50 (s, 9H) ppm; All spectral data

were in agreement with the literature<sup>[3]</sup>.

### Di-tert-butyl-3a-bromo-7-chloro-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (5g)

Light yellow oil, 5.5 h, 71.5 mg, 75% yield; <sup>1</sup>H NMR (600 MHz,  
Cl Boc Chloroform-d ) 
$$\delta$$
 7.32 (d,  $J = 8.0$  Hz, 1H), 7.28 (d,  $J = 7.8$  Hz, 1H),  
<sup>5g</sup> S-37

7.14 (t, J = 7.8 Hz, 1H), 6.21 (s, 1H), 3.60–3.55 (m, 1H), 2.82–2.75 (m, 2H), 2.74–2.67 (m, 1H), 1.53 (s, 9H), 1.50 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[5]</sup>.

# Di*-tert*-butyl-3a,7-dibromo-2,3,3a,8a-tetrahydropyrrolo[2,3-*b*]indole-1,8-dicarbo xylate (5h)



All spectral data were in agreement with the literature <sup>[5]</sup>.

## 1,8-Di*-tert*-butyl-2-methyl-3a-bromo-2,3,3a,8a-tetrahydropyrrolo-[2,3-b]indole-1, 2,8-tricarboxylate (5i)



Following the general procedure for cyclic 3a-bromoindolins, the procedure was performed on **4i** (0.2 mmol) and NaBr (24.7 mg, 0.24 mmol); Colorless oil, 9 h, 82.1 mg, 83% yield; **<sup>1</sup>H NMR** (600 MHz, Chloroform-*d*) δ 7.55 (s, 1H), 7.36 (d, *J* 

= 7.6 Hz, 1H), 7.31 (t, J = 7.6 Hz, 1H), 7.13 (t, J = 7.4 Hz,

1H), 6.40 (s, 1H), 3.91–3.88 (m, 1H), 3.75 (s, 3H), 3.23–3.20 (m, 1H), 2.84–2.81 (m, 1H), 1.59 (s, 9H), 1.40 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

## Di*-tert*-butyl-3a-bromo-8a-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dicarbox ylate (5j)



= 9.4 Hz, 1H), 2.91–2.81 (m, 2H), 2.69–2.61 (m, 1H), 2.16 (s, 3H), 1.59 (s, 9H), 1.42

(s, 9H) ppm. All spectral data were in agreement with the literature <sup>[6]</sup>.

### 8-(*tert*-Butyl)-1-isopropyl-3a-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8 -dicarboxylate (5k)



2.85–2.82 (m, 1H), 2.78–2.73 (m, 1H), 1.98 (s, 1H), 1.59 (s, 9H), 0.94 (t, J = 6.2 Hz, 6H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  154.3, 152.1, 142.0, 132.4, 130.4, 124.1, 123.7, 117.4, 84.0, 82.1, 71.9, 62.0, 46.2, 28.2, 27.8, 19.1, 19.0 ppm; HRMS (ESI) m/z; [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>27</sub>BrN<sub>2</sub>O<sub>4</sub>Na 461.1046; Found 461.1034.

## 1-Benzyl-8-(*tert*-butyl)-3a-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-di carboxylate (5l)



Following the general procedure for cyclic 3a-bromoindolins, the procedure was performed on **4l** (0.2 mmol) and NaBr (24.7 mg, 0.24 mmol); Light Coral oil, 4 h, 51.3 mg, 54% yield; <sup>1</sup>H **NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.63 (s, 1H), 7.38–7.29 (m, 7H), 7.10 (t, *J* = 7.6 Hz, 1H), 6.46 (s, 1H), 5.19–5.15 (m, 2H),

3.80–3.77 (m, 1H), 2.92–2.87 (m, 1H), 2.84–2.81 (m, 1H), 2.77–2.72 (m, 1H), 1.54 (s, 9H) ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

# Benzyl-3a-bromo-8-tosyl-3,3a,8,8a-tetrahydropyrrolo[2,3-b]indole-1(2H)-carbox ylate (5m)



Following the general procedure for cyclic 3a-bromoindolins, the procedure was performed on **4m** (0.2 mmol) and NaBr (24.7 mg, 0.24 mmol); Light yellow oil, 16 h, 45.4 mg, 43% yield; <sup>1</sup>H **NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.65–7.46 (m, 5H), 7.38 (t, *J* 

= 7.2 Hz, 2H), 7.34–7.31 (m, 2H), 7.28 (d, *J* = 7.6 Hz, 1H), 7.17–7.12 (m, 3H), 6.30 (s, 1H), 5.30–5.18 (m, 2H), 3.82–3.79 (m, 1H), 2.84–2.80 (m, 1H), 2.71–2.63 (m, 2H), 2.32 (s, 3H) ppm. All spectral data were in agreement with the literature <sup>[1]</sup>.

### Di*-tert*-butyl-5-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dicarboxylat e (6a)



2.06 (m, 1H), 1.99–1.96 (m, 1H), 1.56 (s, 9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  153.9, 152.2, 142.3, 134.3, 130.9, 127.0, 117.1, 115.2, 81.8, 80.1, 76.3, 44.9, 31.9, 29.6, 28.3, 28.3 ppm; **HRMS (ESI)** *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>27</sub>BrN<sub>2</sub>O<sub>4</sub>Na 461.1046; Found 461.1030.

# Di*-tert*-butyl-5-bromo-4-fluoro-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dica rboxylate (6b)



White solid, m.p. 106.3 – 107.6 °C, 12 h, 60.4 mg, 66% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.37–7.31 (m, 2H), 6.51 (d, *J* = 7.6 Hz, 1H), 4.08 (t, *J* = 7.8 Hz, 1H), 3.91–3.88 (m, 1H), 2.88–2.83 (m, 1H), 2.13–2.10 (m, 1H), 2.05–1.99 (m, 1H), 1.54

(s, 9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  **13C NMR** (150 MHz, Chloroform-*d*)  $\delta$ 154.9 (d, <sup>1</sup>*J*<sub>C-F</sub>= 244.4 Hz), 153.7, 152.0, 144.6 (d, <sup>3</sup>*J*<sub>C-F</sub>= 7.6 Hz), 132.9, 119.3, 112.2, 101.4 (d, <sup>2</sup>*J*<sub>C-F</sub>= 19.6 Hz), 82.2, 81.8, 80.2, 76.9, 45.0, 42.5, 28.3, 28.3 ppm; <sup>19</sup>F NMR (565 MHz, Chloroform-*d*)  $\delta$  -114.1 ppm; HRMS (ESI) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>26</sub>BrFN<sub>2</sub>O<sub>4</sub>Na 479.0952; Found 479.0942.

## Di*-tert*-butyl-5-bromo-4-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (6c)



8.6 Hz, 1H), 6.53 (d, J = 7.8 Hz, 1H), 3.98 – 3.88 (m, 2H), 2.85 (td, J = 12.2, 5.4 Hz, 1H), 2.31 (s, 3H), 2.12 – 2.04 (m, 1H), 1.87 (dd, J = 12.4, 5.4 Hz, 1H), 1.55 (s, 9H), 1.50 (s, 9H) ppm. <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  153.8, 152.4, 142.2, 133.2, 131.7, 118.1, 113.8, 81.7, 80.0, 76.6, 44.3, 32.4, 28.4, 28.3, 19.3 ppm; **HRMS (ESI)** m/z: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>29</sub>BrN<sub>2</sub>O<sub>4</sub>Na 475.1203; Found 475.1193.

## Di*-tert*-butyl-5-bromo-6-chloro-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dica rboxylate (6d)



9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  153.7, 151.9, 143.3, 133.6, 132.4, 128.4, 124.6, 114.9, 82.3, 81.9, 80.2, 76.8, 44.9, 44.3, 28.3, 28.2 ppm; HRMS (ESI) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>4</sub>N<sub>2</sub>BrClNa 495.0657; Found 495.0644.

## Di-tert-butyl-5-bromo-6-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (6e)



Light yellow oil, 9 h, 20.9 mg, 23% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.60 (br, 1H), 7.27 (s, 1H), 6.43 (d, J = 7.0 Hz, 1H), 3.93 (t, J = 7.6 Hz, 1H), 3.83–3.80 (m, 1H), 2.85–2.80 (m, 1H), 2.36 (s, 3H), 2.09–2.03 (m, 1H), 2.00–

1.90 (m, 1H), 1.57 (s, 9H), 1.48 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$ 13C NMR (151 MHz, CDCl3)  $\delta$  153.9, 152.4, 142.6, 137.3, 131.5, 127.4, 117.7, 81.7, 80.0, 76.6, 44.9, 44.4, 29.7, 28.4, 28.3, 23.3 ppm; **HRMS (ESI)** *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>29</sub>BrN<sub>2</sub>O<sub>4</sub>Na 475.1203; Found 475.1195.

Di-tert-butyl-5-bromo-7-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (6f)

Br 
$$H$$
 NBoc  
Me Boc  
6f

Chloroform-*d*)  $\delta$  7.16 (s, 1H), 7.11 (s, 1H), 6.20 (s, 1H), 3.95 (t, J = 5.8 Hz, 1H), 3.55 (dd, J = 10.6, 7.8 Hz, 1H), 2.80 – 2.75 (m, 1H), 2.24 (s, 3H), 2.15 – 2.04 (m, 2H), 1.48 (d, J = 7.0 Hz, 18H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  154.0, 153.2, 141.4, 137.2, 132.8, 131.8, 123.9, 117.9, 81.7, 80.0, 78.9, 46.4, 44.8, 28.5, 28.2, 19.5 ppm; **HRMS (ESI)** *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>29</sub>BrN<sub>2</sub>O<sub>4</sub>Na 475.1203; Found 475.1193.

# Di*-tert*-butyl-5-bromo-7-chloro-2,3,3a,8a-tetrahydropyrrolo[2,3-*b*]indole-1,8-dica rboxylate (6g)



Colorless oil, 10.5 h, 9.5 mg, 10% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.38 (s, 1H), 7.21 (s, 1H), 6.18 (s, 1H), 4.03 (t, J = 6.0 Hz, 1H), 3.64–3.55 (m, 1H), 2.85–2.81 (m, 1H), 2.20– 2.10 (m, 2H), 1.50 (s, 9H), 1.49 (s, 9H) ppm; <sup>13</sup>C NMR (150

MHz, Chloroform-*d*)  $\delta$  154.0, 152.3, 140.2, 139.3, 132.0, 125.2, 117.8, 115.5, 82.2, 80.5, 79.3, 47.1, 44.8, 28.5, 28.1 ppm; **HRMS** (**ESI**) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>4</sub>N<sub>2</sub>ClBrNa 495.0657; Found 495.0651.

# Di*-tert*-butyl-5,7-dibromo-2,3,3a,8a-tetrahydropyrrolo[2,3*-b*]indole-1,8-dicarbox ylate (6h)



MHz, Chloroform-*d*)  $\delta$  153.9, 152.2, 142.0, 139.3, 134.7, 125.8, 118.1, 115.6, 82.4, 80.5, 79.4, 47.3, 44.8, 28.5, 28.1 ppm; **HRMS (ESI)** *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>4</sub>N<sub>2</sub>Br<sub>2</sub>Na 539.0152; Found 539.0145.

1,8-Di*-tert*-butyl-2-methyl-5-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,2, 8-tricarboxylate (6i)



Colorless oil, 20 h, 47.5 mg, 48% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.44 (br, 1H), 7.34 (dd, J = 8.6, 2.0 Hz, 1H), 7.28–7.27 (m, 1H), 6.35 (d, J = 5.9 Hz, 1H), 4.00–3.94 (m, 2H), 3.74 (s, 3H), 2.53–2.49 (m, 1H),

2.31–2.26 (m, 1H), 1.58/1.57 (s, 9H), 1.39 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  173.0, 152.1, 141.6, 134.4, 131.3, 126.8, 126.6, 119.0, 115.9, 81.9, 80.9, 58.9, 52.1, 44.6, 32.8, 29.7, 28.2, 28.1 ppm; HRMS (ESI) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>22</sub>H<sub>29</sub>BrN<sub>2</sub>O<sub>6</sub>Na 519.1101; Found 519.1092.

## Di*-tert*-butyl-5-bromo-8a-methyl-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-di carboxylate (6j)



Light yellow oil, 9 h, 53.0 mg, 58% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.56 (d, J = 8.7 Hz, 1H), 7.29–7.26 (m, 1H), 7.22 (s, 1H), 3.73–3.70 (m, 1H), 3.49 (d, J = 7.8 Hz, 1H), 3.08–3.03 (m, 1H), 2.25–2.18 (m, 1H), 2.04 (s, 3H), 1.93–1.90

(m, 1H), 1.57 (s, 9H), 1.45 (s, 9H) ppm. For the mixture of two conformationl isomers: <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  155.8, 152.9, 151.8, 150.3, 142.4, 135.6, 134.5, 133.3, 130.8, 126.5, 126.1, 120.3, 118.0, 116.9, 115.8, 114.8, 87.8, 84.1, 81.6, 79.7, 54.6, 46.7, 28.5, 28.4, 28.2, 24.2, 14.0 ppm. HRMS (ESI) *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>21</sub>H<sub>29</sub>BrN<sub>2</sub>O<sub>4</sub>Na 475.1203; Found 475.1194.

#### 8-(*tert*-Butyl)-1-isopropyl-5-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8dicarboxylate (6k)



Light yellow oil, 9 h, 38.2 mg, 43% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.53 (br, 1H), 7.30 (dd, *J* = 8.6, 2.0 Hz, 1H), 7.26 (s, 1H), 6.44 (d, *J* = 6.8 Hz, 1H), 3.98 (t, *J* = 7.2 Hz, 1H), 3.93–3.84 (m, 3H), 2.94–2.89 (m, 1H),

2.17–2.11 (m, 1H), 2.04–2.02 (m, 1H), 1.99–1.95 (m, 1H), 1.57/1.56 (s, 9H), 0.94– 0.92 (m, 6H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  154.8, 152.2, 142.2, 134.2, 130.9, 128.0, 126.9, 115.4, 81.9, 81.4, 76.6, 71.6, 44.8, 28.3, 28.3, 27.9, 19.1 ppm; **HRMS (ESI)** *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>27</sub>BrN<sub>2</sub>O<sub>4</sub>Na 461.1046; Found 461.1034.

## 1-Benzyl-8-(tert-butyl)-5-bromo-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dic arboxylate (6l)



**NMR** (150 MHz, Chloroform-*d*) δ 154.4, 152.2, 142.1, 136.7, 134.1, 131.0, 128.4, 127.9, 127.8, 126.9, 123.8, 115.4, 82.0, 76.7, 67.0, 66.9, 44.9, 28.3, 28.2 ppm; **HRMS** (**ESI**) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>23</sub>H<sub>25</sub>O<sub>4</sub>N<sub>2</sub>BrNa 495.0890; Found 495.0885.

## Benzyl-5-bromo-8-tosyl-3,3a,8,8a-tetrahydropyrrolo[2,3-b]indole-1(2H)-carboxyl ate (6m)



3H), 2.11–2.04 (m, 1H), 1.97–1.91 (m, 1H) ppm; <sup>13</sup>**C NMR** (150 MHz, Chloroform-*d*) δ 13C NMR (151 MHz, CDCl3) δ 154.1, 144.1, 141.3, 136.3, 135.1, 131.5, 129.6, 129.4, 128.4, 128.4, 128.1, 128.0, 127.3, 127.1, 79.7, 67.5, 44.5, 29.6, 21.5 ppm; HRMS (ESI) *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>25</sub>H<sub>23</sub>O<sub>4</sub>N<sub>2</sub>BrNaS 549.0454; Found 549.0449.

#### 7. X-ray Dates

Single-crystal X-ray diffraction measurements were carried out on an Agilent Gemini E diffractometer using graphite monochromated Mo radiation ( $\lambda = 0.71073$  Å, 50kV 40mA). The crystal data was collected on a Agilent Gemini E diffractometer (Mo, 50kV 40mA) and reducted by CrysAlisPro (Rigaku). The structures were solved by direct methods using SHELXS-97. Refinements were performed with SHELXL-2013 using fullmatrix least-squares calculations on F2, with anisotropic displacement parameters for all the nonhydrogen atoms.

The single crystals of compound 3c suitable for X-ray diffraction analysis were obtained by slow evaporation of a mixed Dichloromethane/Petroleum ether solution of 3c at room temperature.

Molecular structure of 3c, ellipsoids shown at 35% probability.





#### Crystal data and structure refinement for CCDC:2173146.

Identification code	exp_10608
Empirical formula	C <sub>17</sub> H <sub>16</sub> BrNO <sub>3</sub> S
Formula weight	394.28
Temperature/K	293(2)
Crystal system	triclinic
Space group	P-1
a/Å	8.2932(10)
b/Å	8.8298(11)
c/Å	12.4718(14)

$\alpha/^{\circ}$	95.863(10)
β/°	90.752(10)
γ/°	116.824(12)
Volume/Å <sup>3</sup>	808.94(18)
Z	2
$\rho_{calc}g/cm^3$	1.619
$\mu/\text{mm}^{-1}$	2.684
F(000)	400.0
Crystal size/mm <sup>3</sup>	$0.22 \times 0.18 \times 0.09$
Radiation	Mo Ka ( $\lambda = 0.71073$ )
$2\Theta$ range for data collection/°	5.208 to 48.994
Index ranges	$-9 \le h \le 9, -6 \le k \le 10, -13 \le l \le 14$
Reflections collected	5512
Independent reflections	2687 [ $R_{int} = 0.0339$ , $R_{sigma} = 0.0567$ ]
Data/restraints/parameters	2687/0/209
Goodness-of-fit on F <sup>2</sup>	1.074
Final R indexes [I>= $2\sigma$ (I)]	$R_1 = 0.0572, wR_2 = 0.1393$
Final R indexes [all data]	$R_1 = 0.0813, wR_2 = 0.1524$
Largest diff. peak/hole / e Å <sup>-3</sup>	0.37/-0.64

#### 8. Scale-up experiments



**Procedure:** A mixture of **1a** (261.1 mg, 1.0 mmol) and NaBr (123.5 mg, 1.2 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (8/0.2ml) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at rt using a constant current of 5.0 mA between a graphite anode and graphite cathode  $(1.4 \times 0.8 \times 0.2 \text{ cm} \text{ submerged in solution})$  with stirring. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to give compound **2a** (15 h, 214.2 mg, 63% yield).



**Procedure:** A mixture of **1a** (261.1 mg, 1.0 mmol) and NaBr (123.5 mg, 1.2 mmol) were added in a 10 mL vial with a stir bar, then CH<sub>3</sub>CN/H<sub>2</sub>O (5/3ml) were added under air atmosphere. The vial was covered with the electrode holder. The electrolysis was carried out at RT using a constant current of 5.0 mA between a graphite anode and graphite cathode ( $1.4 \times 0.8 \times 0.2$  cm submerged in solution) with stirring. After completion as detected by TLC. The reaction mixture was diluted with EtOAc. The organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 10/1) to give compound **3a** (28 h, 223.2 mg, 66 %).

#### 9. Transformation and applications



A mixture of **3a** (34.0 mg, 0.1 mmol),  $K_3PO_4$  (64.1 mg, 0.30 mmol), PdCl<sub>2</sub> (dppf) (7.3 mg, 0.01 mmol), and B<sub>2</sub>Pin<sub>2</sub> (28.2 mg, 0.11 mmol) was dissolved in DMF (3 mL) under argon atmosphere. Then, the mixture was stirred at 80 °C and when the reaction was completed as monitored by TLC. The reaction was cooled to room temperature. The resulting solid was filtered off and adsorbed onto the minimal amount of silica gel using CH<sub>2</sub>Cl<sub>2</sub>. the solvent was concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 4/1) to give **7**. The same procedure was performed on **6a** to afford **10**.

#### Di*-tert*-butyl-2,2',3,3a,3',3'a,8a,8'a-octahydro-8H,8'H-[5,5'-bifuro[2,3-b]indole]-8, 8'-dicarboxylate (7)



Colorless oil, 2.5 h, 18.1 mg, 70 % yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.85 (br, 1H), 7.41–7.37 (m, 5H), 6.19 (br, 2H), 4.02–3.94 (m, 4H), 3.52–3.47 (m, 2H), 2.39–2.33 (m, 2H), 2.18–2.15 (m, 2H), 1.63/1.61 (s, 18H); <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 152.4, 142.1,

135.8, 126.7, 122.6, 114.5, 93.6, 66.4, 33.9, 28.4, 24.8 ppm; **HRMS (ESI)** *m/z*: [M + Na]<sup>+</sup> Calcd for C<sub>30</sub>H<sub>36</sub>N<sub>2</sub>O<sub>6</sub>Na 543.2466; Found 543.2465.

#### Di-*tert*-Butyl-2,2',3,3a,3',3'a,8a,8'a-octahydro-[5,5'-bipyrrolo[2,3-b]indole]-1,1',8, 8'-tetracarboxylate (10)



Reaction of **6a** (35.1 mg, 0.08 mmol) with  $K_3PO_4$  (51.3 mg, 0.24 mmol),  $PdCl_2(dppf)$  (5.8 mg, 0.008 mmol), and  $B_2Pin_2$  (22.5 mg, 0.089 mmol) provided **10.** Light yellow oil, 1 h, 18.9 mg, 66 % yield; <sup>1</sup>H NMR (600 MHz,

Chloroform-*d*)  $\delta$  7.66 (br, 2H), 7.39 (d, *J* = 7.0 Hz, 2H), 7.33 (d, *J* = 7.0 Hz, 2H), 6.48 (d, *J* = 7.2 Hz, 2H), 4.02 (t, *J* = 7.4 Hz, 2H), 3.85–3.81 (m, 2H), 2.93–2.86 (m, 2H), 2.17–2.05 (m, 4H), 1.58 (s, 18H), 1.50 (s, 18H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  144.7, 143.3, 134.3, 128.7, 126.0, 120.6, 120.5, 116.7, 81.0, 79.7, 76.5, 48.8, 34.3, 10.2 ppm; **HRMS (ESI)** *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>40</sub>H<sub>54</sub>N<sub>4</sub>O<sub>8</sub>Na 741.3834; Found 741.3836.



General Procedures for 8a: A mixture of 3a (34.0 mg, 0.1 mmol), 3,6-dihydro-2H-pyran-4-boronic acid pinacol ester (23.1 mg, 0.11 mmol), K<sub>3</sub>PO<sub>4</sub> (25.5 mg, 0.112 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (5.8 mg, 0.005 mmol) dissolved in MeOH (0.8 mL) under argon atmosphere. Then, the mixture was stirred at 80 °C and when the reaction was completed as monitored by TLC. The reaction was cooled to room temperature. the solvent was concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate =40/1 to 10/1) to give **8a**.

# tert-butyl-5-(3,6-dihydro-2H-pyran-4-yl)-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]ind ole-8-carboxylate (8a)



Light yellow oil, 22 h, 16.3 mg, 48 % yield; <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.76 (br, 1H), 7.25 – 7.22 (m, 2H), 6.17 (br, 1H), 6.06 (s, 1H), 4.32 (d, J = 2.9 Hz, 2H), 3.99 (t, J = 8.1 Hz, 1H), 3.93 (q, J = 5.3 Hz, 3H), 3.95 – 3.92 (m, 1H), 2.57 –

2.44 (m, 2H), 2.38 – 2.28 (m, 1H), 2.11 (dd, J = 12.2, 4.7 Hz, 1H), 1.60 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  152.1, 141.7, 135.2, 133.6, 131.2, 124.7, 121.1, 120.6, 114.0, 93.6, 81.3, 66.3, 65.8, 64.4, 45.0, 33.8, 28.4, 27.3 ppm; HRMS (ESI) m/z: [M + Na]<sup>+</sup> Calcd for C<sub>20</sub>H<sub>25</sub>NO<sub>4</sub>Na 366.1676; Found 366.1661.

#### tert-Butyl-5-phenyl-2,3,3a,8a-tetrahydro-8H-furo[2,3-b]indole-8-carboxylate (8b)



Following the general Procedure for 8a. The procedure was performed on 3a (68.0 mg, 0.2 mmol), phenylboronic acid (26.8 mg, 0.22 mmol), K<sub>3</sub>PO<sub>4</sub> (50.9 mg, 0.24 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (11.6 mg, 0.01 mmol) to afford **8b**. Light yellow oil, 22 h, 51.0 mg, 76% yield; <sup>1</sup>**H NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.88 (br, 1H), 7.56 (d, *J* = 7.8 Hz, 2H),

7.46–7.41 (m, 4H), 7.32 (t, J = 7.4 Hz, 1H), 6.20 (s, 1H), 4.00 (q, J = 9.4, 8.8 Hz, 2H), 3.53–3.49 (m, 1H), 2.41–2.31 (m, 1H), 2.17–2.14 (m, 1H), 1.62 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-d) δ 152.4, 140.8, 136.0, 131.8, 128.7, 127.1, 126.8, 126.7, 114.5, 93.6, 81.5, 66.4, 44.9, 33.8, 28.4 ppm; **HRMS (ESI)** m/z:  $[M + Na]^+$ Calcd for C<sub>21</sub>H<sub>23</sub>O<sub>3</sub>NNa 360.1570; Found 360.1556.



A mixture of **6a** (17.56 mg, 40 µmol), DavePhos (2.3 mg, 6 µmol), Pd<sub>2</sub>(dba)<sub>3</sub> (3.7 mg, 4 µmol), NaOt-Bu (7.7 mg, 80 µmol), and morpholine (10.5 µL, 120 µmol) was dissolved in 1,4-dioxane (315 µL) under argon atmosphere .Then, the mixture was stirred at 100  $\,^{\circ}$ C and when the reaction was completed as monitored by TLC. The reaction was cooled to room temperature and diluted with CH<sub>2</sub>Cl<sub>2</sub> (2 mL). The solution was washed with water (2 mL) and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 mL x 3). The combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated under reduced pressure, purified by flash column chromatography (Petroleum ether/Ethyl acetate = 3/1) to give 9.

### Di-tert-butyl-5-morpholino-2,3,3a,8a-tetrahydropyrrolo[2,3-b]indole-1,8-dicarbo xylate (9)



Yellow oil, 4.5 h, 14.4 mg, 81% yield; <sup>1</sup>H NMR (600 MHz, Chloroform-d) δ 7.50 (br, 1H), 6.77–6.75 (m, 2H), 6.40 (d, J =

S-50

5.2 Hz, 1H), 3.94 (t, J = 7.3 Hz, 1H), 3.86 (t, J = 4.8 Hz, 4H), 3.79 (dd, J = 11.3, 7.4 Hz, 1H), 3.13–3.05 (m, 4H), 2.84 (td, J = 11.7, 5.2 Hz, 1H), 2.13–1.98 (m, 2H), 1.56 (s, 9H), 1.49 (s, 9H) ppm; <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  154.0, 152.6, 147.9, 136.7, 133.1, 116.6, 115.8, 112.3, 81.1, 76.2, 71.6, 71.1, 66.9, 61.8, 50.5, 44.9, 28.4, 28.4 ppm; **HRMS (ESI)** *m*/*z*: [M + Na]<sup>+</sup> Calcd for C<sub>24</sub>H<sub>35</sub>O<sub>5</sub>N<sub>3</sub>Na 468.2469; Found 468.2467.

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## 11 <sup>1</sup>H NMR, <sup>13</sup>C NMR and <sup>19</sup>F NMR Spectra of compounds



Parameters	
Parameter	value
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Origin	Bruker BioSpin GmbH
Solvent	CDC13
Temperature	298.1
Number of Scans	1
Acquisition Date	2022-05-31T10:17:15
Spectrometer Frequency	150 MHz
Nucleus	13C



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$$7.7218$$

$$7.207$$

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$$-8.002$$

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$$7.7.075$$

$$7.7.061$$

$$-4.688$$

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$$3.463$$

$$3.463$$

$$3.463$$

$$3.463$$

$$-2.884$$

$$-2.884$$

$$-2.884$$

$$-2.884$$

$$-2.482$$

$$-1.663$$



## $-\frac{8.137}{7.548}$ $-\frac{8.137}{7.548}$ $-\frac{7.548}{7.331}$ $-\frac{7.548}{7.237}$ $-\frac{7.248}{7.223}$ $-\frac{7.224}{7.223}$ $-\frac{7.224}{7.223}$ $-\frac{7.224}{7.223}$ $-\frac{7.224}{7.223}$ $-\frac{7.223}{7.223}$ $-\frac{1.206}{1.906}$ $-\frac{1.906}{1.1878}$ $-\frac{1.906}{1.1809}$ $-\frac{1.906}{1.1809}$



### $\begin{array}{c} 7.833 \\ 7.408 \\ 7.295 \\ 7.292 \\ 7.279 \\ 7.279 \\ 7.279 \\ 7.279 \\ 7.279 \\ 7.279 \\ 7.073 \\ 7.073 \\ 7.061 \\ 7.061 \\ 7.061 \\ 7.061 \\ 8.88 \\ 9.88 \\ 7.061 \\ 7.061 \\ 8.88 \\ 9.88$

#### 4,007 3.993 3.978 3.993 3.494 3.494 3.461 3.461 3.461 2.319 2.319 2.319 2.219 2.289 2.299 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.289 2.297 2.297 2.289 2.289 2.297 2.297 2.289 2.289 2.297 2.297 2.289 2.289 2.297 2.297 2.289 2.289 2.297 2.297 2.297 2.289 2.289 2.297 2.289 2.299



## -7.8497.404 7.392 7.322 7.309 7.099 7.099 7.074 -6.076

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Number of Scans	8
Acquisition Date	2022-08-03T04:46:42
Spectrometer Frequency	600 MHz
Nucleus	1H

### 4,099 4,085 4,085 3,556 3,556 3,558 3,558 3,558 3,558 3,558 3,558 3,558 3,558 3,558 2,580 2,768 2,280 7,270 2,768 2,768 2,770 2,768 2,770 2,768 2,770 2,770 2,770 2,770 2,770 2,770 2,770 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,556 2,566 2,556 2,566 2,556 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,566 2,576 2,566 2,576 2,566 2,576



### $\begin{array}{c} 7.875 \\ 7.459 \\ 7.459 \\ 7.7459 \\ 7.7459 \\ 7.7459 \\ 7.7459 \\ 7.7350 \\ 7.7350 \\ 7.7350 \\ 7.7350 \\ 7.7350 \\ 7.7350 \\ 7.7350 \\ 7.7350 \\ 7.3534 \\ 7.7350 \\ 7.3534 \\ 7.3534 \\ 7.3534 \\ 7.3536 \\ 7.3534 \\ 7.3534 \\ 7.3536 \\ 7.3526 \\$



## 

### 

Parameter	value
Title	RV-0399-1.1.fid
Origin	Bruker BioSpin GmbH
Solvent	CDC13
Temperature	298.0
Number of Scans	8
Acquisition Date	2021-12-13T23:50:45
Spectrometer Frequency	600 MHz
Nucleus	1H





### 7.918 7.555 7.555 7.543 7.543 7.543 7.543 7.471 7.449 7.449 7.449 7.449 7.449 7.449 7.449 7.436 7.7336 7.7336 7.7336 7.7116 7.71

Parameters		
Parameter	value	
Title	Ra-641.1.fid	
Origin	Bruker BioSpin GmbH	
Solvent	CDCl3	
Temperature	298.0	
Number of Scans	8	
Acquisition Date	2022-05-31T00:34:19	
Spectrometer Frequency	600 MHz	
Nucleus	1H	

### 



### $\begin{array}{c} 7.453\\ 7.453\\ 7.453\\ 7.439\\ 7.338\\ 7.338\\ 7.338\\ 7.338\\ 7.338\\ 7.336\\ 7.336\\ 7.336\\ 7.139\\ 7.139\\ 7.139\\ 7.130\\ 7.$



f1 (ppm)

## -7.715 -7.115 -6.213 -6.223 -6.233 -6.233 -6.233 -6.233 -



$$\begin{array}{c} 7,645\\ 7,7291\\ 7,7291\\ 7,727\\ 7,7267\\ 7,7277\\ 7,7267\\ 7,7267\\ 6,719\\ 6,$$



## $\begin{array}{c} 7.779\\ 7.776\\ 7.7427\\ 7.427\\ 7.427\\ 7.427\\ 7.429\\ 7.7300\\ 7.7300\\ 7.7257\\ 7.7257\\ 7.7257\\ 7.7257\\ 7.7257\\ 7.7254\\ 7.7256\\ 7.725$





## -7.900 -7.900 -7.900 -7.10316 -6.173



## 

Parameters	
Parameter	value
Title	Ra-762.1.fid
Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	298.0
Number of Scans	8
Acquisition Date	2022-08-05T00:43:10
Spectrometer Frequency	600 MHz
Nucleus	1H

### 4,020 4,020 3,517 3,517 3,517 3,476 3,476 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,888 2,277 3 2,773 1,601 1,601 1,601 1,601 1,601 1,602 1,60



### -6.182 = -6.182

### 





### $\int_{-1.53}^{7.526} \int_{-1.505}^{7.518} \int_{-1.505}^{7.505} \int_{-1.505}^{7.375} \int_{-1.342}^{7.362} \int_{-1.047}^{7.060} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.153} \int_{-0.153}^{0.152} \int_{-0.153}^{$

#### 3,3,9983,3,9983,3,9923,3,9823,3,9823,3,9823,3,4673,3,4763,3,4763,3,4533,4433,3,4612,3,4533,3,4612,3,4433,3,4433,3,4433,3,4433,3,4433,3,4433,3,4433,3,4433,3,4432,2,8432,2,8432,2,8432,2,8432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7432,2,7772,2,77



#### 7.974 7.960 7.586 7.586 7.586 7.586 7.586 7.586 7.582 7.582 7.582 7.403 7.582 7.403 7.569 7.107 7.423 7.433 7.532 7.232 7.5327 7.532 7.532 7.532 7.5327 7.5327 7.5327 7.5327 7.5327 7.5327 7.5327 7.5327 7.5327 7.


# -7.698 -7.593 -6.139 -6.139 -6.139 -6.139 -3.929 -2.929 -



### -6226 -7266 -7

Parameter	value
Title	rv-0561-2.1.fid
Origin	Bruker BioSpin GmbH
Solvent	CDC13
Temperature	298.0
Number of Scans	8
Acquisition Date	2022-06-02T14:30:32
Spectrometer Frequency	600 MHZ
Nucleus	1H





### $\begin{array}{c} 7.840\\ 7.781\\ 7.782\\ 7.7287\\ 7.7287\\ 7.7264\\ 7.7264\\ 7.7264\\ 7.7264\\ 7.7247\\ 7.7247\\ 7.7247\\ 7.7247\\ 7.7247\\ 7.2343\\ 3.3996\\ 6.6239\\ 3.3996\\ 6.6239\\ 3.3996\\ 5.3339\\ 3.3996\\ 5.3339\\ 3.3996\\ 5.3339\\ 3.3396\\ 5.3339\\ 3.3396\\ 5.3339\\ 3.3396\\ 5.3339\\ 3.3396\\ 5.3339\\ 5.$



### 3,3,9883,3,9743,3,9743,3,9083,3,9083,3,9083,3,9083,3,9083,3,9083,3,9083,3,2953,2,2953,2,







# $\begin{array}{c} & 7,532\\ & 7,361\\ & 7,375\\ & 7,375\\ & 7,375\\ & 7,375\\ & 3,992\\ & 3,9$





Parameters		
Parameter	value	
Title	RV-0619-4.3.fid	
Origin	Bruker BioSpin GmbH	
Instrument	Avance	
Solvent	CDC13	
Number of Scans	16	
Acquisition Time	0.4981	
Spectrometer Frequency	565MHZ	
Nucleus	19F	





### 7,787 7,787 7,787 7,787 7,787 7,787 7,787 7,725 7,7145 7,7



### - 6.148





S-83



S-84



Parameters		
Parameter	value	
Title	RV-0630.3.fid	
Origin	Bruker BioSpin GmbH	
Solvent	CDC13	
Temperature	298.0	
Number of Scans	16	
Acquisition Date	2022-08-06T15:52:35	
Spectrometer Frequency	565MHZ	
Nucleus	19F	





### 7.187 7.187 6.103 6.103 6.103 7.3942 7.3942 7.3942 7.3952 7.3942 7.3953 7.3952 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.3552 7.2352 7.2352 7.2352 7.2252 7.2252 7.2252 7.2252 7.22562 7.25562



### $<_{6.105}^{6.115}$



### 20MG

## 



### 7,829 7,381 7,376 7,368 7,368 7,368 7,368 7,368 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,315 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,316 7,317 7,217,

## 



### 7.288 7.288 7.298 7.298 7.294 7.294 7.272 7.272 7.272 7.091 7.091 7.078

## 3.746 3.739 3.739 3.716 3.716 2.827 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.821 2.733 2.710 2.710 2.710 2.710 2.710 2.710 2.710 2.701 2.810 2.810 2.810 2.810 2.810 2.701 2.810 2.810 2.810 2.810 2.810 2.810 2.810 2.810 2.810</





 $\int \frac{7.477}{7.196}$   $\int \frac{7.196}{7.110}$   $\int \frac{7.183}{7.170}$   $\int \frac{6.851}{6.839}$  -6.476

### -3.747 -3.728 -3.728 -3.728 -3.716 -3.016 -3.016 -3.016 -3.016 -3.016 -3.016 -3.728 -3.016 -2.835 -2.987 -2.8336 -2.8336 -2.8336 -2.8336 -2.8336 -2.8336 -2.8336 -2.8336 -2.8336 -2.655











# $\int_{-1}^{7.355} \int_{-1.285}^{7.311} \int_{-1.285}^{7.285} \int_{-1.145}^{7.132} \int_{-1.132}^{7.132} \int_{-0.206}^{-1.325} \int_{-0.206}^{-$

### **3.587 3.587 3.576 2.819 2.811 2.777 2.779 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.709 2.700**



# $\int_{-7.059}^{-7.496} 7.496$ 7.327 7.315 7.084 7.072 7.059

-6.193

## 3.586 3.579 3.579 3.556 3.556 3.558 2.5814 2.5814 2.5179 2.5779 2.7776 2.7776 2.7776 2.7776 2.7756 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655 2.6655</



### 7.552 7.366 7.366 7.3336 7.3338 7.113 7.113 7.113 7.113 7.113 7.113 7.3138 7.3218 7.3218 7.3218 7.3218 7.3218 7.3218 7.3218 7.3218 7.3218 7.3218 7.3218 7.3219





### 7.6217.5377.5377.53677.313677.313677.31906.44016.44016.44016.39107.30267.20267.





### 



## $\begin{array}{c} 7.525 \\ 7.294 \\ 7.291 \\ 7.277 \\ 7.248 \\ 7.248 \\ 7.248 \\ 6.436 \end{array}$

### 





Parameter	value
Title	Ra-66.2.fid
Origin	Bruker BioSpin GmbH
Solvent	CDC13
Temperature	298.4
Number of Scans	16
Acquisition Date	2022-05-18T12:29:38
Spectrometer Frequency	565 MHz
Nucleus	19F





--114.07



### .....










7,2457,2457,2457,2737,2737,2727,2227,22



## $\int_{7.281}^{7.567} 7.553 \\ 7.286 \\ 7.282 \\ 7.221 \\ 7.221$

### $\begin{array}{c} 3.732\\ 3.3.719\\ 3.3.779\\ 3.3.796\\ 3.3.3.976\\ 3.3.3.056\\ 3.3$



### 7.532 7.295 7.291 7.2010

### 3,3,988 3,3,976 3,3,8976 3,3,8976 3,3,8976 3,3,8976 3,3,897 3,3,897 3,3,897 3,3,897 3,3,897 3,3,897 3,3,897 3,3,897 3,3,897 3,3,597 3,5977 3,5977 3,59775 3,59775 3,597755,59775 5,597755 5,597755555555555



## 

### (55202) (555154) (555154) (555154) (555154) (555154) (555154) (555154) (55323) (553333) (5533333) (5533333) (553333) (553333) (553333) (553333) (553333) (55333



## 

Parameter Title Origin Solvent Temperature Number of Scans Acquisition Date Spectrometer Frequency Nucleus	value RV-0518-4.1.fid Bruker BioSpin GmbH CDC13 298.0 8 2022-05-01T05:45:12 600 MHz 1H				1	Br	N Ts 6m	V Cbz
						Mul	А	Li
10.5 10.0 9.5 9.0	8.5 8.0 7.5 7.0	6.5 6.0	5.5 5.0 4 f1 (ppr	.5 4.0 3.5 n)	3.0 2.5	2.0 1.5 1	.0 0.5	0.0 -0.5 -1.0
	- [54,09 [44,10] [136,30 [135,13 [135,13 [135,13] [136,30] [135,13] [136,30] [137,13] [129,38] [129,39] [129,39]	128.10 128.10 127.27 127.10		79.72 77.21 77.00 76.79	- 67.49		— 29.63 — 21.49	
Parameter Title Origin Solvent Temperature Number of Scans Acquisition Date Spectrometer Frequency Nucleus	value RV-0518-4.2.fid Bruker BioSpin GmbI CDC13 298.0 800 2022-05-01T06:24:07 / 150 MHz 13C	ł				Br	N Ts 6m	`Cbz
					J			

180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1 (ppm)











# $\begin{array}{c} 7.660\\ 7.385\\ 7.335\\ 7.336\\ 7.336\\ 6.473\\ 6.473\\ 6.473\\ 6.473\\ 6.473\\ 6.473\\ 6.473\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.389\\ 7.2876\\ 7.2866\\ 7.2876\\ 7.286\\ 7.2866\\$

ParameterTitleROriginBSolventCTemperature2Number of Scans8Acquisition Date2Spectrometer Frequency6Nucleus1	value a-597.1.fid ruker BioSpin Gr DC13 98.0 022-04-18T23:24 00 MHz H	nbH :36		Boc		10		Boc	Зос					
10.0 9.5 9.0 8.5		۸٫۲ ۲۹ ۲۰ ۲۰ ۲.0 7.0 6.5	6.0	5.5	5.0 4.5 f1 (ppm)		3.5 3.	0 2.5	2.0	1.5	1.0	0.5 0.	0 -0.5	-
Parameter	×144.68	Z 125.95 Z 120.58 Z 120.56 - 116.74	٦		$\int_{77.19}^{80.97}$	76.81			— 34.29			- 10.21		
Title Origin Solvent Temperature Number of Scans Acquisition Date Spectrometer Frequency Nucleus	Ra-597.2.1.1r Bruker BioSpi CDC13 300.0 800 1970-01-01T0 150 MHz 13C	n GmbH 8:00:00								Boc	Boc	1	0	N Boc
~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~														<b>197</b>



### S-123

80 70

60 50

40 30

10

0 -10

20

150 140 130 120 110 100 90 fl (ppm)

210 200

190 180

170 160

# $\begin{array}{c} & 7.730\\ 7.7375\\ 7.3759\\ 7.3759\\ 7.3759\\ 7.3759\\ 7.3759\\ 7.3759\\ 7.3759\\ 7.3759\\ 7.3779\\ 3.3097\\ 3.3097\\ 3.3091\\ 3.3519\\ 3.3519\\ 3.3519\\ 3.3599\\ 3.3519\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.359\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\ 3.3599\\$



$$-7.713 < 7.253 < 7.232$$

 $\left\{\begin{array}{c} 3.980 \\ 3.965 \\ 3.951 \\ 3.076 \\ 4.3.076 \\ 3.047 \\ 3.047 \\ \end{array}\right.$ 



Parameter	value					
Title	RV-0546-1.1.fid					
Origin	Bruker BioSpin GmbH					
Solvent	CDCl3					
Temperature	298.0					
Number of Scans	8					
Acquisition Date	2022-05-07T19:37:15					
Spectrometer Frequency	600 MHz					
Nucleus	1H					







