

## Supplementary Information

### Fluorescent probes based on oxonium-coumarin scaffold for the detection of SO<sub>2</sub> derivatives

#### Authors:

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**Scheme S1.** (a) Synthesis route of **acceptor** and **donor**; (b) Normalized fluorescence emission spectrum of **donor** and normalized UV-vis absorption spectrum of **acceptor**.

**Fig. S1.** Fluorescence responses of probes in different solvents. (a) Fluorescence spectrum of CPO1; (b) Fluorescence ratio of CPO1; (c) Images of CPO1 under normal light and 365 nm UV light; (d) Fluorescence spectrum of CPO2; (e) Fluorescence ratio of CPO2; (f) Images of CPO2 under normal light and 365 nm UV light;

**Fig. S2.** (a) . Time-dependent experiment of CPO1 (10 μM) in the presence of Na<sub>2</sub>SO<sub>3</sub> (20 eq and 50 eq). (b) The effect of pH values on the ratio of fluorescence intensity ( $I_{508}/I_{645}$ ) of CPO1 (10 μM) in the absence and presence of Na<sub>2</sub>SO<sub>3</sub>.

**Fig. S3.** UV-Vis spectrum of CPO1 (10 μM) in the presence of Na<sub>2</sub>SO<sub>3</sub> (0 - 100 eq).

Inset: color changes of CPO1 with or without  $\text{Na}_2\text{SO}_3$  under normal light.

**Fig. S4.** HRMS of CPO1- $\text{HSO}_3^-$ .

**Fig. S5.**  $^1\text{H}$  NMR titration experiment of probe CPO1 towards  $\text{SO}_3^{2-}$ .

**Fig. S6.** MTT assay of HepG-2 cells with probe CPO1 at different concentration (0 - 30  $\mu\text{M}$ ) for 24 h.

**Figs. S7 - 9.**  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, and HRMS for Acceptor.

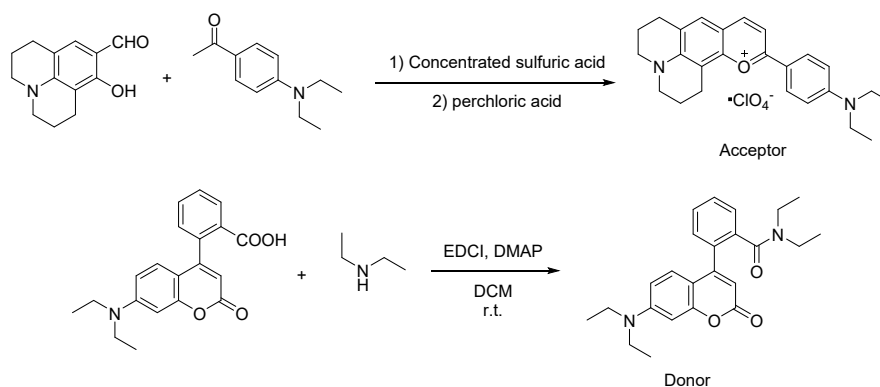
**Figs. S10 - 12.**  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, and HRMS for probe CPO1.

**Figs. S13 - 15.**  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, and HRMS for probe CPO2.

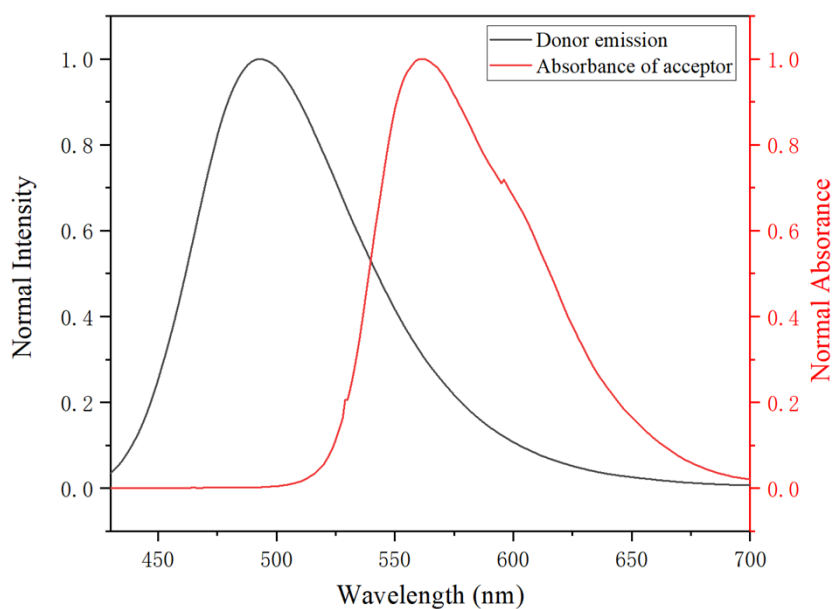
**Fig. S16.** Thermal analysis (DSC) of CPO1.

**Table S1.** Comparison of probe CPO1 with others.

(a)



(b)



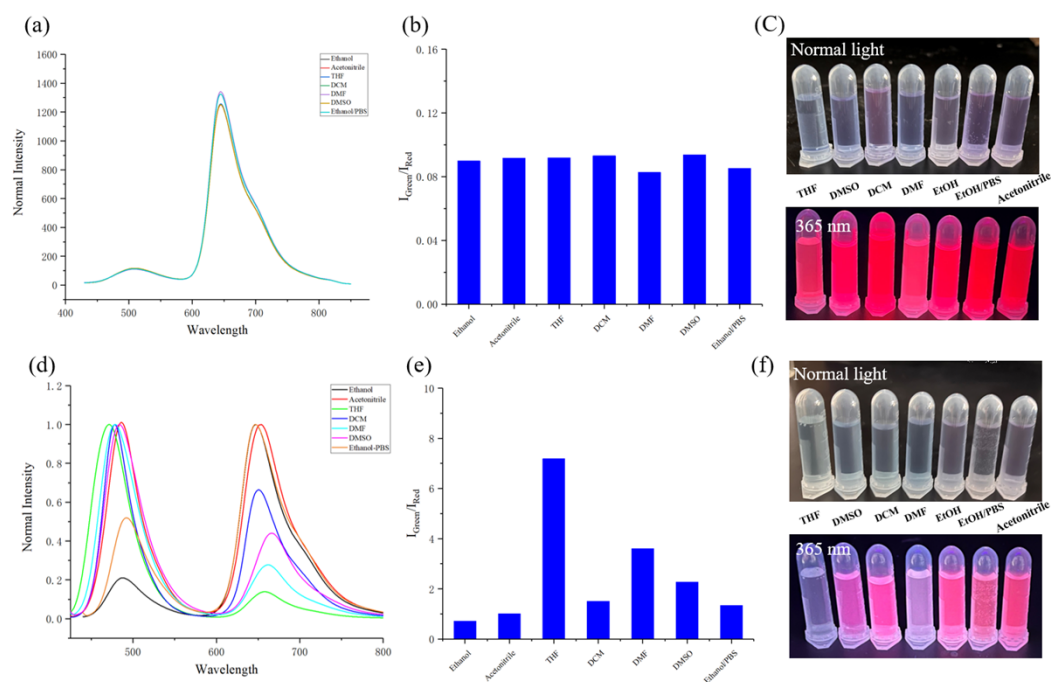
**Scheme S1.** (a) Synthesis route of **acceptor** and **donor**; (b) Normalized fluorescence emission spectrum of **donor** and normalized UV-vis absorption spectrum of **acceptor**.

### Synthesis of acceptor and donor

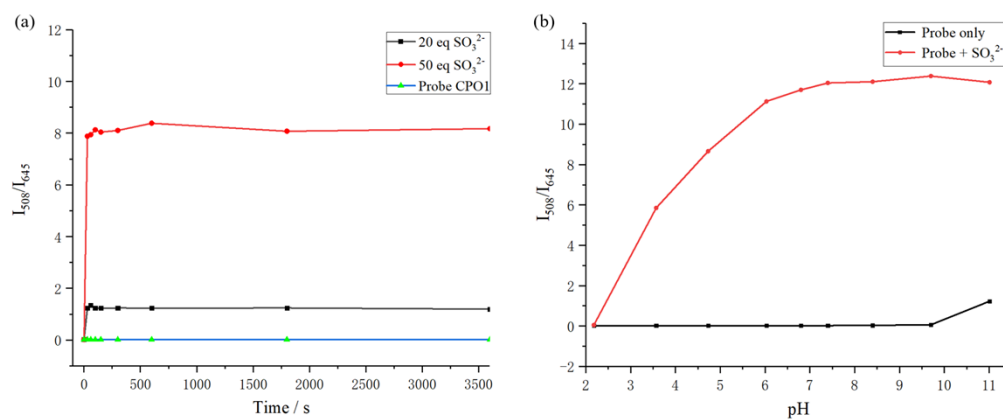
Acceptor was synthesized according to the synthetic route of intermediate 1 (390 mg, yield = 0.79). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.31 (s, 1H), 8.05 (s, 2H), 7.72 (t, *J* = 7.2 Hz, 1H), 7.52 – 7.35 (m, 1H), 6.91 (d, *J* = 8.6 Hz, 2H), 3.53 (p, *J* = 6.2, 5.5 Hz, 8H), 2.97 (d, *J* = 6.2 Hz, 2H), 2.82 (s, 2H), 2.12 - 1.87 (m, 4H), 1.18 (t, *J* = 7.0 Hz, 6H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 165.88, 152.69, 152.53, 151.09, 145.78, 130.71, 127.14, 126.33, 116.17, 112.64, 106.97, 104.89, 50.63, 50.14, 44.90, 27.42, 20.35,

19.49, 19.34, 12.94. HRMS (ESI+): m/z calculated 373.2274, found 373.22859.

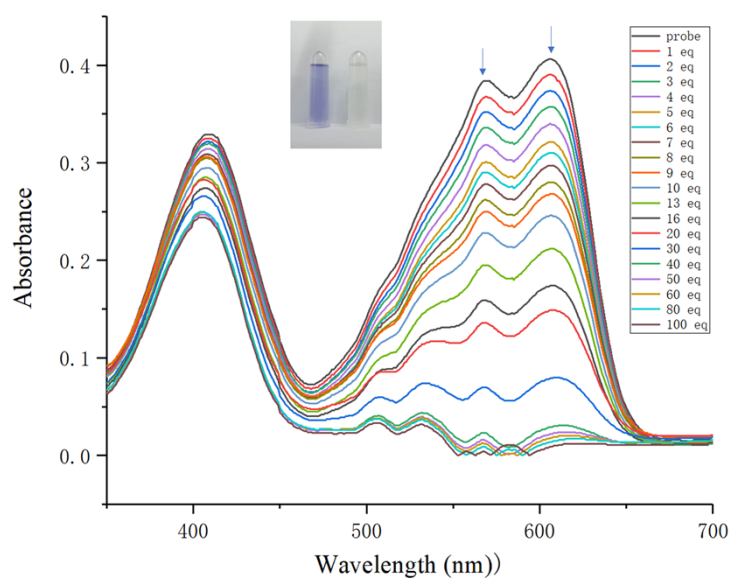
Donor was synthesized according to previous work[1].



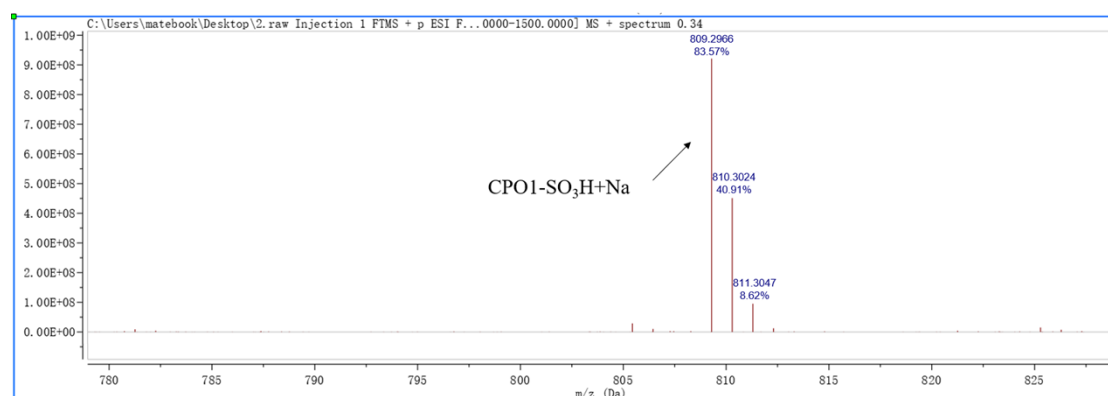
**Fig. S1.** Fluorescence responses of probes in different solvents. (a) Fluorescence spectrum of CPO1; (b) Fluorescence ratio of CPO1; (c) Images of CPO1 under normal light and 365 nm UV light; (d) Fluorescence spectrum of CPO2; (e) Fluorescence ratio of CPO2; (f) Images of CPO2 under normal light and 365 nm UV light;



**Fig. S2.** (a) . Time-dependent experiment of CPO1 (10  $\mu\text{M}$ ) in the presence of  $\text{Na}_2\text{SO}_3$  (20 eq and 50 eq). (b) The effect of pH values on the ratio of fluorescence intensity ( $I_{508}/I_{645}$ ) of CPO1 (10  $\mu\text{M}$ ) in the absence and presence of  $\text{Na}_2\text{SO}_3$  (100 eq).



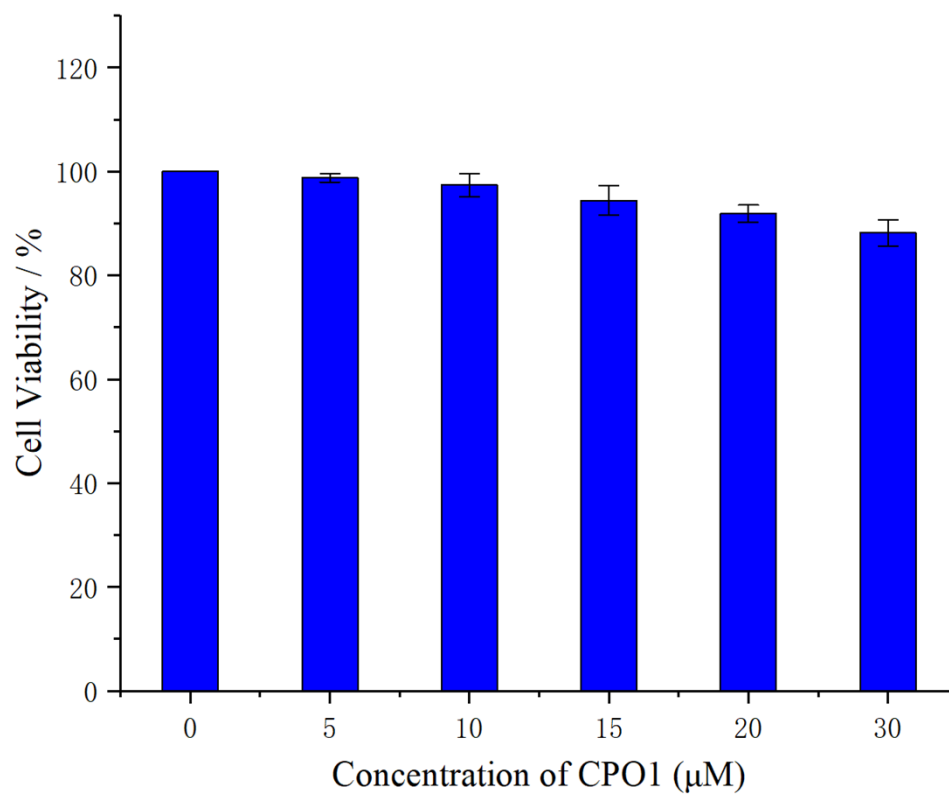
**Fig. S3.** UV-Vis spectrum of CPO1 (10 μM) in the presence of Na<sub>2</sub>SO<sub>3</sub> (0 - 100 eq). Inset: color changes of CPO1 with or without Na<sub>2</sub>SO<sub>3</sub> under normal light.



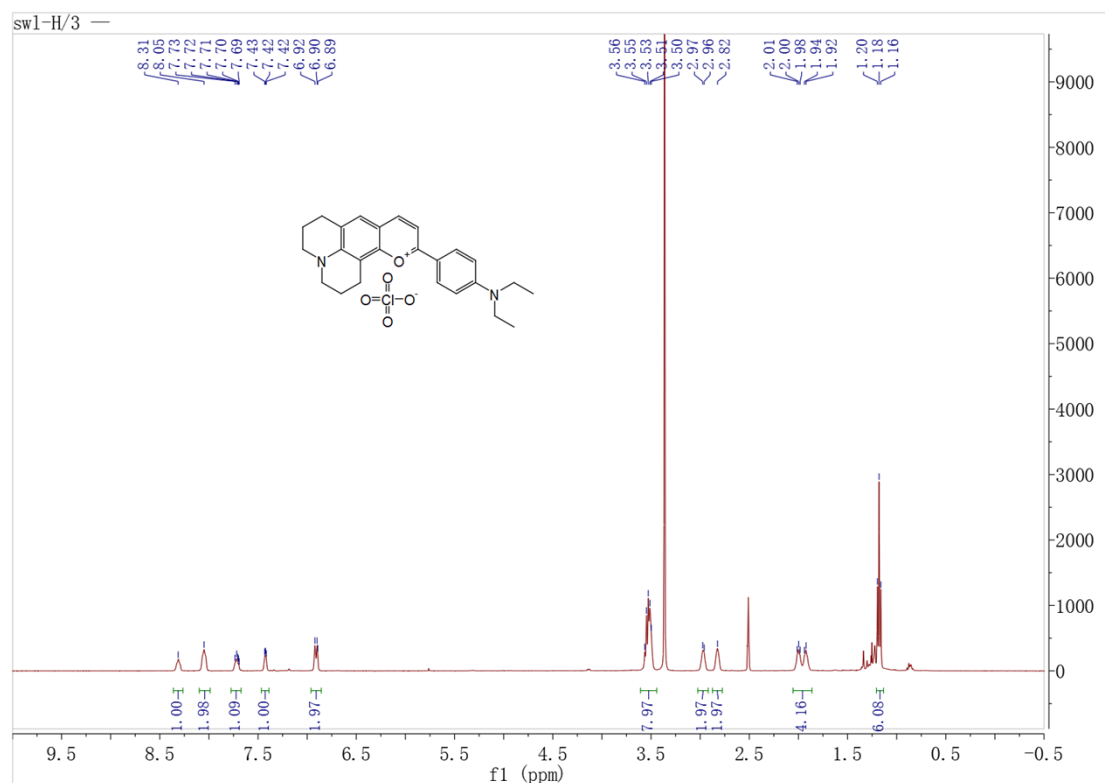
**Fig. S4.** HRMS of probe CPO1-HSO<sub>3</sub>.



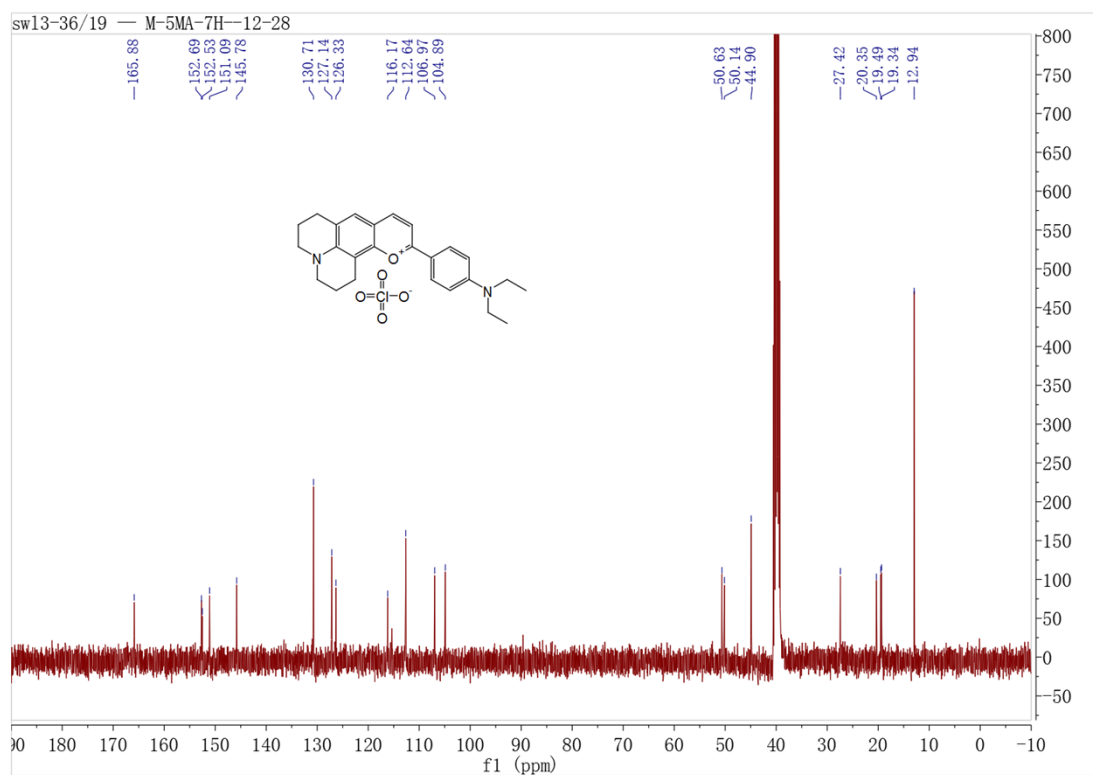
**Fig. S5.** <sup>1</sup>H NMR titration experiment of probe CPO1 towards SO<sub>3</sub><sup>2-</sup>.



**Fig. S6.** MTT assay of HepG-2 cells with probe CPO1 at different concentration (0 - 30 μM) for 24 h.



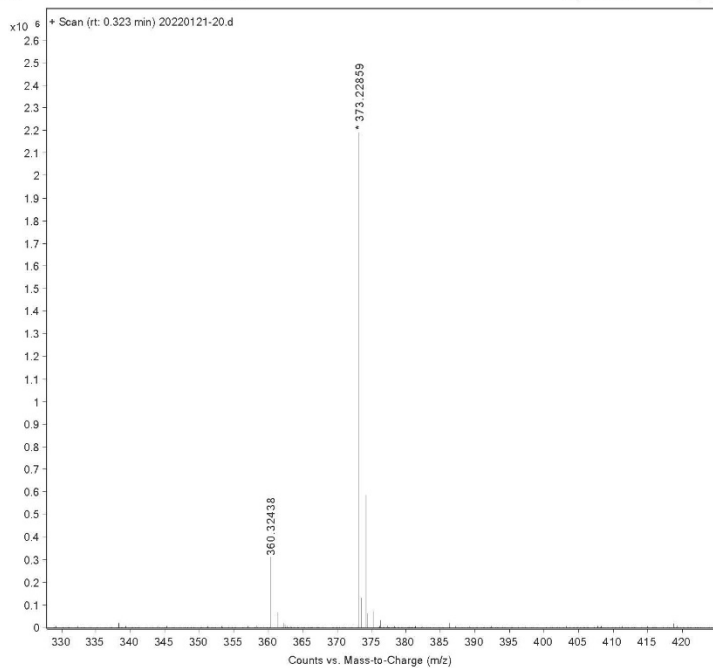
**Fig. S7.**  $^1\text{H}$  NMR for Acceptor.



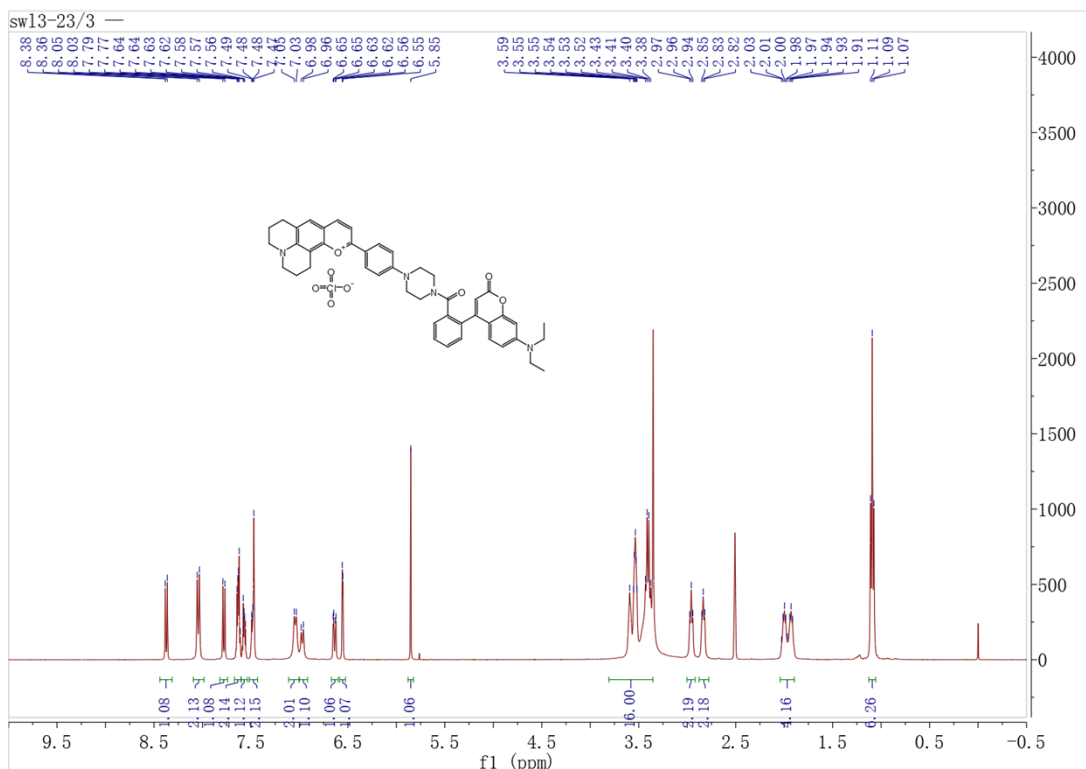
**Fig. S8.**  $^{13}\text{C}$  NMR for Acceptor.



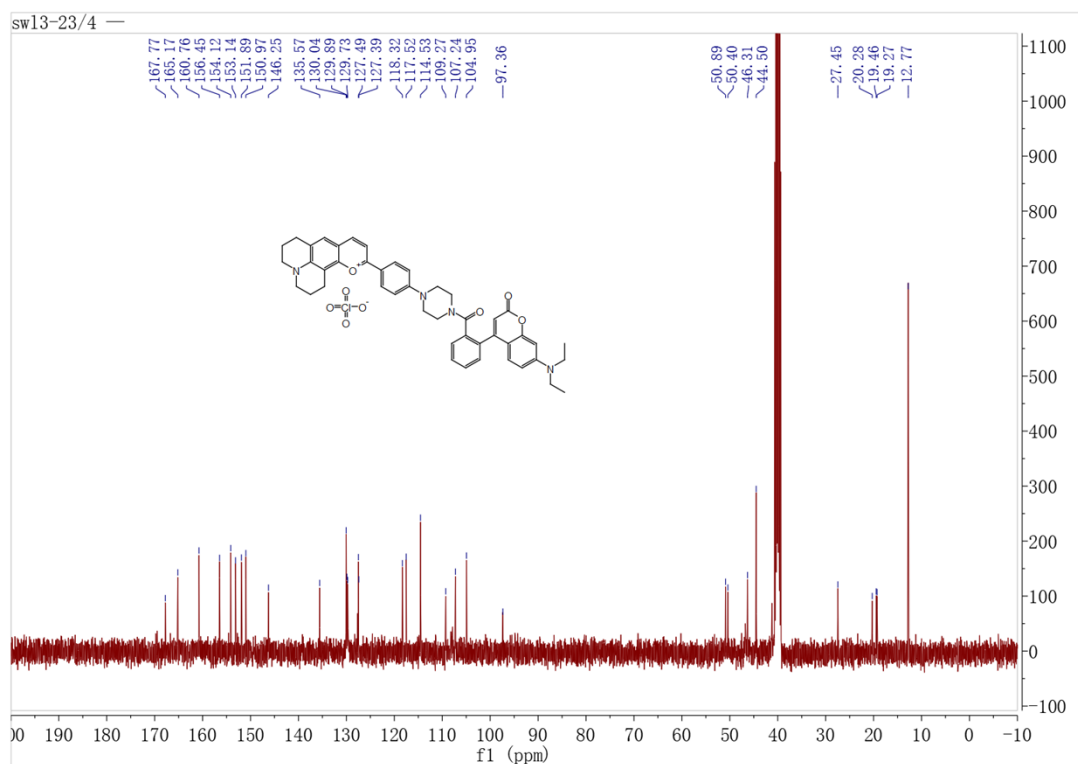
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**Fig. S9.** HRMS for Acceptor.

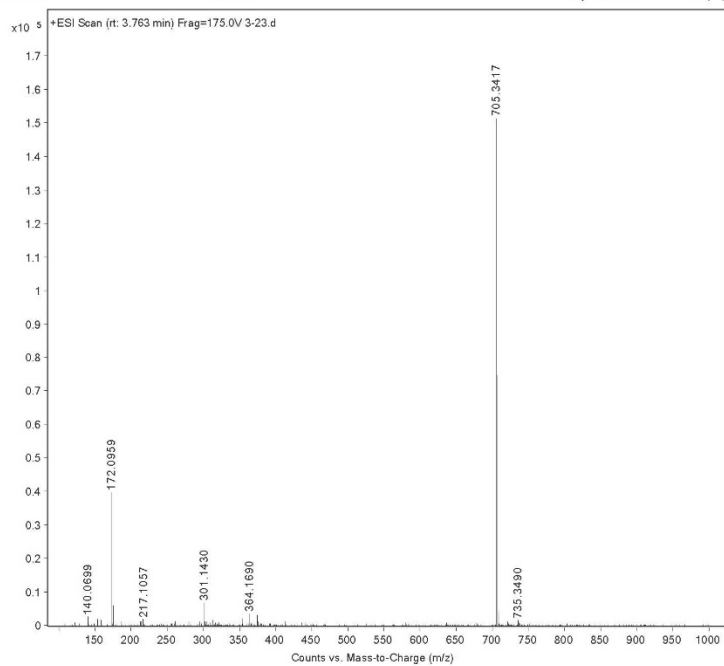


**Fig. S10.**  $^1\text{H}$  NMR for probe CPO1.

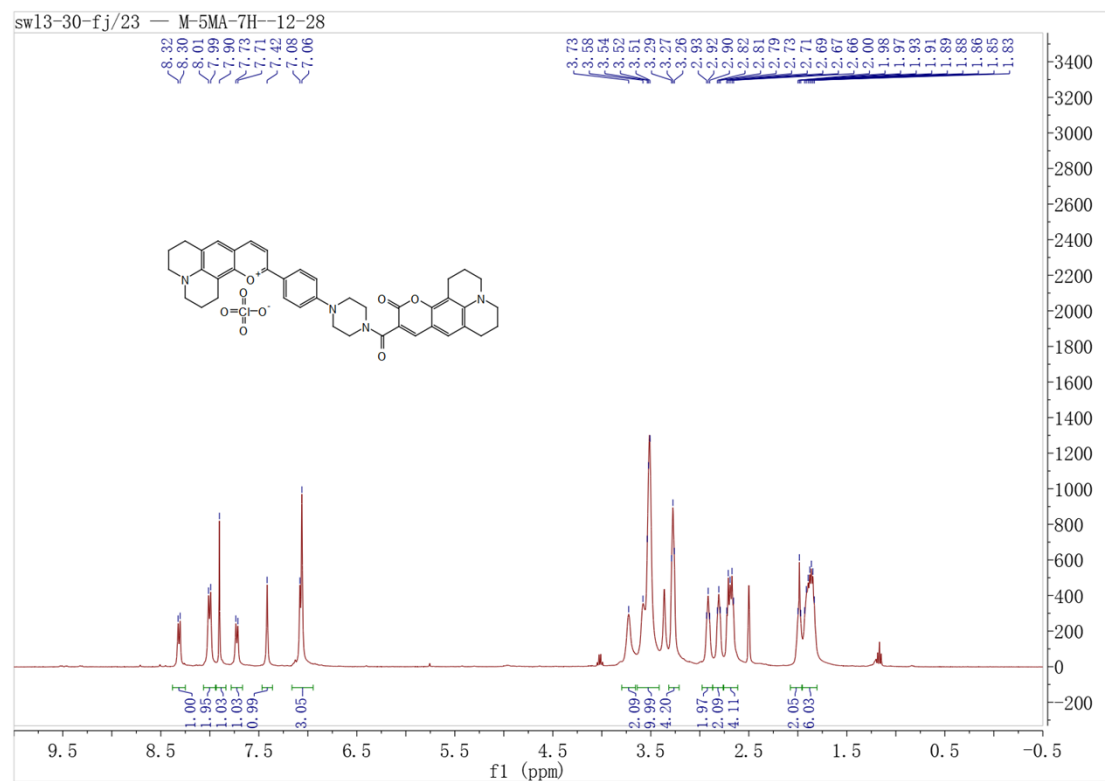


**Fig. S11.**  $^{13}\text{C}$  NMR for probe CPO1.

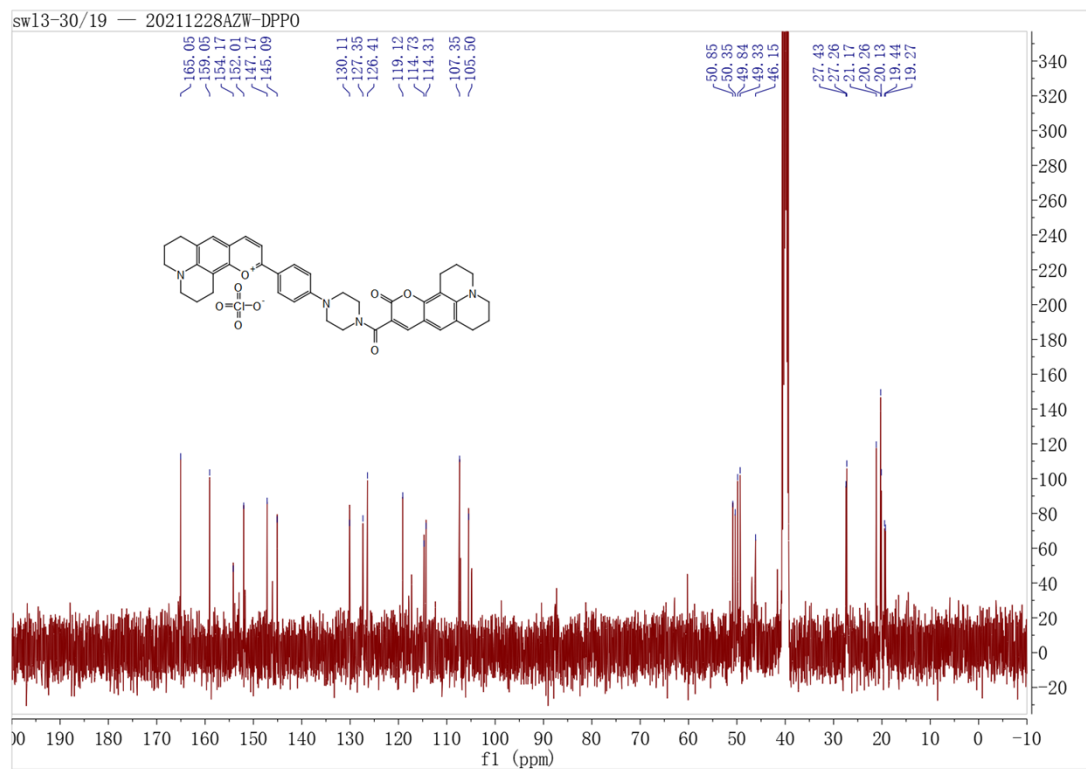
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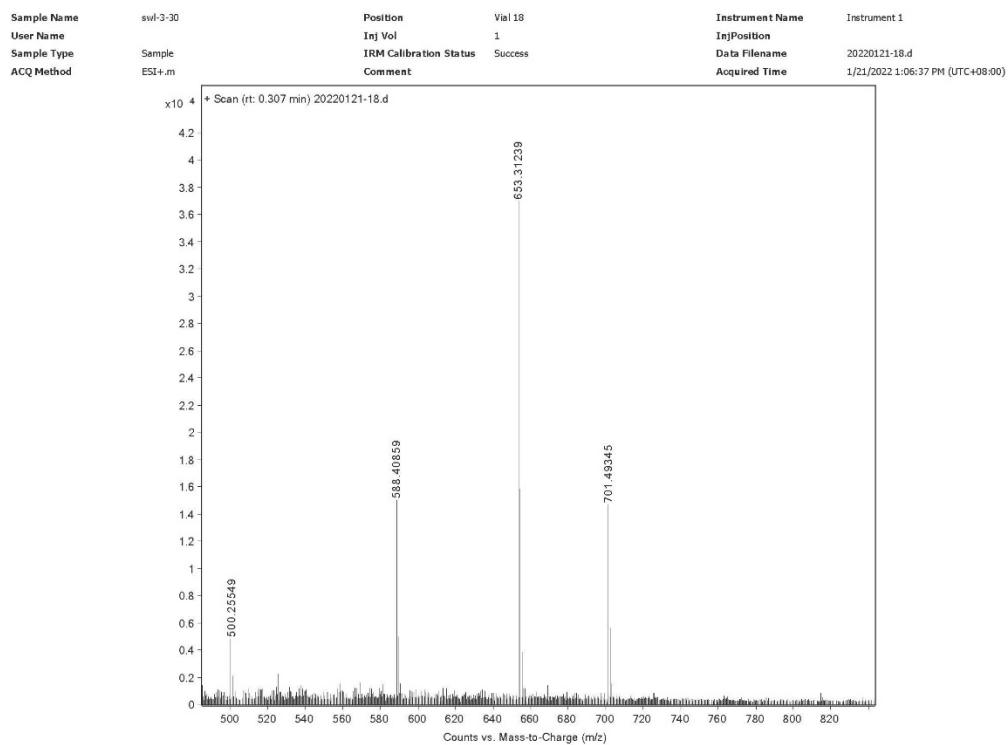
**Fig. S12.** HRMS for probe CPO1.



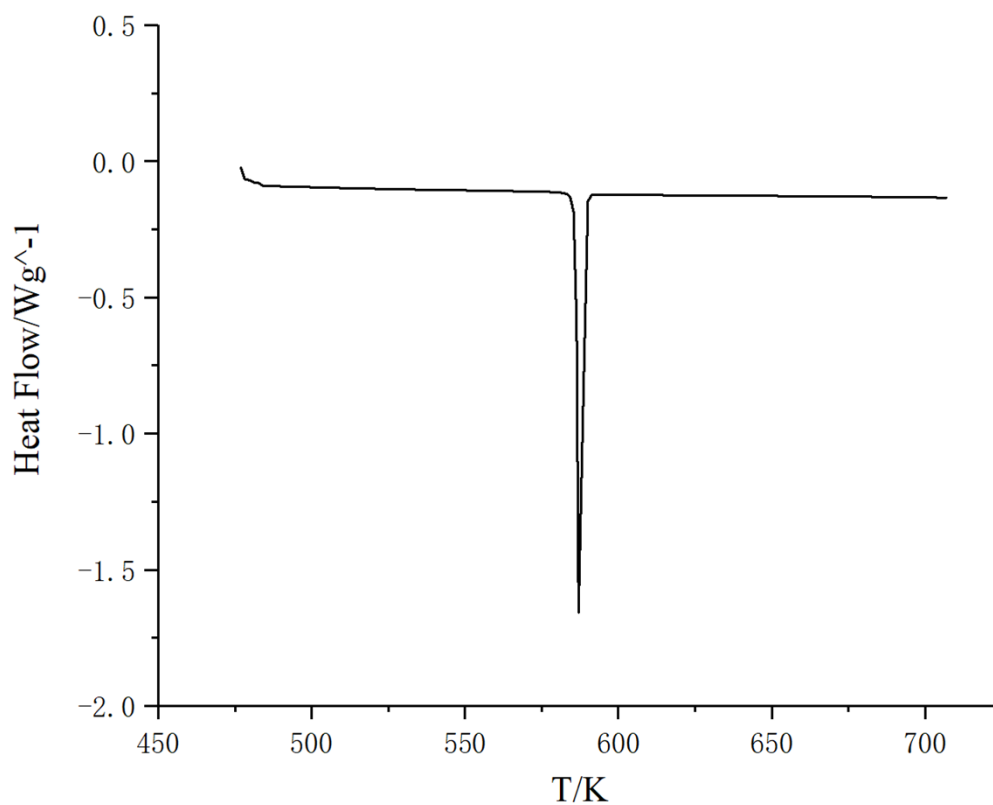
**Fig. S13.** <sup>1</sup>H NMR for probe CPO2.



**Fig. S14.**  $^{13}\text{C}$  NMR for probe CPO2.

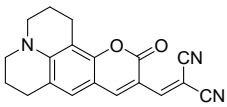
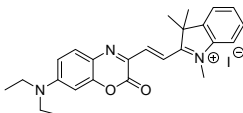
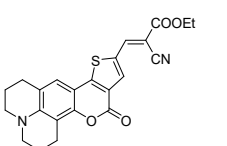
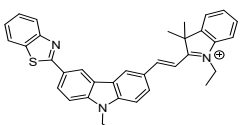


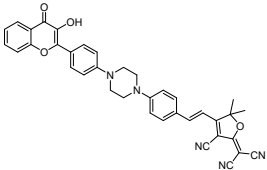
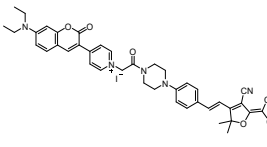
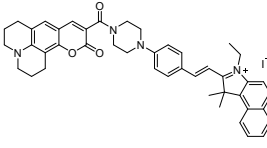
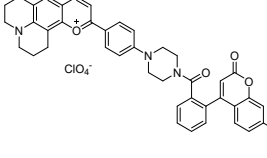
**Fig. S15.** HRMS for probe CPO2.



**Fig. S16.** Thermal analysis (DSC) of CPO1.

**Table S1.** Comparison of probe CPO1 with other probes.

| Probe   | Response time | Emission window gap<br>$\lambda_{em}$ (nm) | $\lambda_{ex}$ and $\lambda_{em}$ (nm) | LOD ( $\mu$ M) | Targeted subcellular | Quantitative detection interval | Ref. |
|---|---------------|--|--|----------------|----------------------|---------------------------------|------|
|  | 5 min         | No ratio                                   | 440<br>488                             | 0.87           | No                   | 0 - 200 $\mu$ M                 | [2]  |
|  | 30 s          | 157  | 500<br>560/717                         | 0.087          | No                   | 0 - 4 $\mu$ M                   | [3]  |
|  | 160 s         | 221  | 410/540<br>715/494                     | 0.055          | No                   | 0 - 55 $\mu$ M                  | [4]  |
|  | 3 min         | 138  | 350<br>458/596                         | 1.76           | Mitochondria         | 3 - 7 $\mu$ M                   | [5]  |

|   |        |     |                |       |                |                  |           |
|---|--------|-----|----------------|-------|----------------|------------------|-----------|
|  | 30 min | 111 | 345<br>530/641 | 0.017 | Mitochondria   | 0 - 20 $\mu$ M   | [6]       |
|  | 15 min | 80  | 435<br>568/648 | 0.080 | Lipid droplets | 0 - 5 $\mu$ M    | [7]       |
|  | 230 s  | 116 | 440<br>492/608 | 0.013 | Mitochondria   | 0 - 100 $\mu$ M  | [8]       |
|  | < 30 s | 137 | 410<br>508/645 | 0.06  | Mitochondria   | 50 - 400 $\mu$ M | This work |

## Reference:

- [1] W. Shen, H. Xu, J. Feng, W. Sun, G. Hu, Y. Hu, W. Yang, A ratiometric and colorimetric fluorescent probe designed based on FRET for detecting  $\text{SO}_3^{2-}/\text{HSO}_3^-$  in living cells and mice, *Spectrochim Acta A Mol Biomol Spectrosc* 263 (2021) 120183-120191.
- [2] Y. Yang, B. Bai, W. Xu, Z. Xu, J. Zhang, W. Li, A highly sensitive fluorescent probe for the detection of bisulfite ion and its application in living cells, *Dyes and Pigments* 136 (2017) 830-835.
- [3] M. Li, W. Feng, H. Zhang, G. Feng, An aza-coumarin-hemicyanine based near-infrared fluorescent probe for rapid, colorimetric and ratiometric detection of bisulfite in food and living cells, *Sensors and Actuators B: Chemical* 243 (2017) 51-58.
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- [6] D.-P. Li, X.-J. Han, Z.-Q. Yan, Y. Cui, J.-Y. Miao, B.-X. Zhao, A far-red ratiometric fluorescent probe for  $\text{SO}_2$  derivatives based on the ES IPT enhanced FRET platform with improved performance, *Dyes and Pigments* 151 (2018) 95-101.
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- [8] W. Shen, G. Hu, H. Xu, W. Sun, Y. Hu, W. Yang, Construction and evaluation of ratiometric fluorescent probes based on a 7-aminocoumarin scaffold for the detection of  $\text{SO}_2$  derivatives, *Dyes and Pigments* 198 (2022) 109971-109979.