

Supporting Information

Short and efficient synthesis of alkylresorcinols: a route for the preparation of cannabinoids

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General Remarks

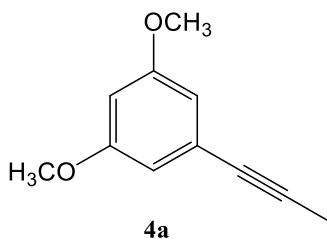
All reagents were purchased from Merck / Sigma-Aldrich and used without further purification. THF was distilled from 3Å MS and stored under an atmosphere of nitrogen. TLC was carried out on silica gel 60 F254 plates (Merck) and visualized with UV light and *p*-anisaldehyde solution followed by heating. Flash chromatography was performed on Merck silica gel (60, particle size: 0.040-0.063 mm). NMR spectra were recorded in CDCl₃ by a Varian 400 spectrometer or a Bruker 400 spectrometer, chemical shifts (δ) are reported in ppm and are internally referenced to residual solvent (¹H-NMR: 7.26 ppm and ¹³C-NMR: 77.00 ppm for CDCl₃). Multiplicities are indicated by s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) and bs (broad signal). Coupling constants (J) are reported in Hz. Yields are given for isolated products showing one spot on a TLC plate and no impurities detectable in the NMR spectrum. HRESIMS were performed on an OrbiTrap high-resolution mass spectrometer (Q Exactive) equipped with the heated electrospray ionization probe HESI-II (Thermo Fisher Scientific, Bremen, Germany) operating in both positive and negative ion modes.

Experimental Protocols and Characterization data

General procedure for the synthesis of **4**

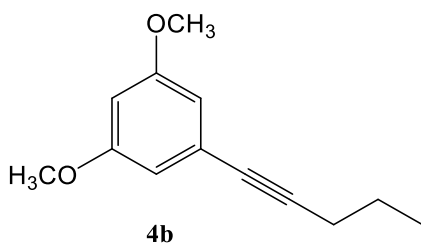
In a dry round-bottom flask, a solution of diisopropylamine (1.5 eq, 0.37 mmol, 52.6 μ L) in dry THF (0.5 mL) was stirred at -10°C for 15 min under N₂ atmosphere. Then *n*BuLi (solution 2.5 M in hexanes, 2.0 eq, 0.50 mmol, 0.2 mL) was added dropwise followed by a solution of ethynyl-3,5-dimethoxybenzene **2** (0.25 mmol, 40.5 mg) in dry THF (1.0 mL). After 20 min, the appropriate alkyl iodide **3** (2.0 eq, 0.50 mmol) was added and the resulting mixture was allowed to stir for 3h (24h for **3b**) at rt. Then the mixture was diluted with H₂O (6.0 mL) and it was extracted with EtOAc (5.0 mL x 3). The organic layers were combined and dried over anhydrous Na₂SO₄. After filtration and evaporation of the solvent under reduced pressure, no purification of the crude product was required.

1,3-dimethoxy-5-(prop-1'-ynyl)benzene 4a



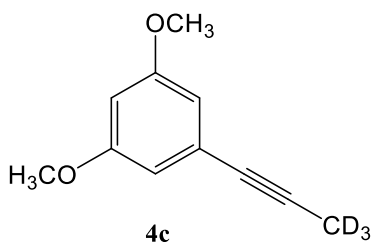
The spectroscopic data of **4a** matched the ones reported in the literature.¹ Pale yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.55 (d, J=2.4 Hz, 2H), 6.40 (t, J=2.4 Hz, 1H), 3.77 (s, 6H), 2.04 (s, 3H); ¹³C-NMR (CDCl₃): δ 160.4, 125.3, 109.3, 101.0, 85.5, 79.7, 55.3, 4.3.

1,3-dimethoxy-5-(pent-1'-ynyl)benzene 4b



The spectroscopic data of **4b** matched the ones reported in the literature.² Yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.56 (d, J=2.4 Hz, 2H), 6.40 (t, J=2.4 Hz, 1H), 3.77 (s, 6H), 2.37 (t, J=7.2 Hz, 2H), 1.67-1.58 (m, 2H), 1.04 (t, J=7.6 Hz, 3H); ¹³C-NMR (CDCl₃): δ 160.4, 125.3, 109.2, 101.0, 89.9, 80.6, 55.3, 22.1, 21.3, 13.5.

1,3-dimethoxy-5-(prop-1'-ynyl[3',3',3'-d₃])benzene 4c

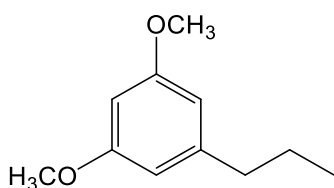


Pale yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.55 (d, J=2.4 Hz, 2H), 6.40 (t, J=2.4 Hz, 1H), 3.77 (s, 6H); ¹³C-NMR (CDCl₃): δ 160.4, 125.3, 109.3, 101.0, 85.0, 79.8, 55.3. HRMS (ESI) m/z for C₁₁H₁₀D₃O [M+H]⁺ calcd 180.1098, found 180.1112.

General procedure for the synthesis of 5a-c

Pd/C (30 w %) and EtOH (3 mL) or H₂O (3 mL) were placed in a dry 50 mL two-neck round-bottom flask. After five vacuum/H₂ cycles, the mixture was stirred at room temperature overnight, then a solution of **4** (0.25 mmol) in THF (1.5 mL) was added. After 24 h stirring at room temperature, the reaction mixture was filtered or alternatively centrifuged, then, EtOAc and H₂O were added and the aqueous layer was extracted with EtOAc (2 x 5 mL). The organic layers were combined and dried over anhydrous Na₂SO₄. After filtration and evaporation of the solvent under reduced pressure, no purification of the crude product was required.

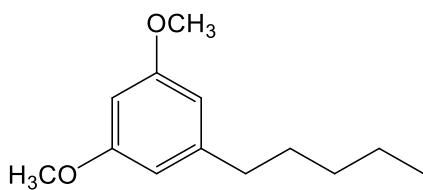
1,3-dimethoxy-5-propylbenzene 5a



5a

The spectroscopic data of **5a** matched the ones reported in the literature.^{3,4} Yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.35 (d, J=2.4 Hz, 2H), 6.30 (t, J=2.4 Hz, 1H), 3.78 (s, 6H), 2.55-2.51 (m, 2H), 1.69-1.59 (m, 2H), 0.95 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 160.6, 145.1, 106.5, 97.5, 55.2, 38.4, 24.3, 13.8.

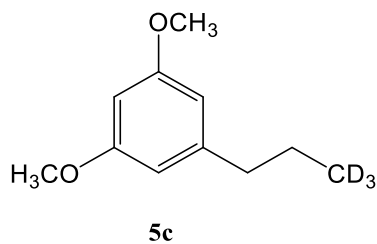
1,3-dimethoxy-5-pentylbenzene 5b



5b

The spectroscopic data of **5b** matched the ones reported in the literature.³ Yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.35 (d, J=2.4 Hz, 2H), 6.30 (t, J=2.4 Hz, 1H), 3.78 (s, 6H), 2.54 (t, J=7.6 Hz, 2H), 1.65-1.57 (m, 2H), 1.37-1.29 (m, 4H), 0.89 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 160.6, 145.2, 106.5, 97.5, 55.2, 36.5, 31.4, 31.0, 22.1, 13.9.

1,3-dimethoxy-5-[3',3',3'-d₃]propylbenzene 5c

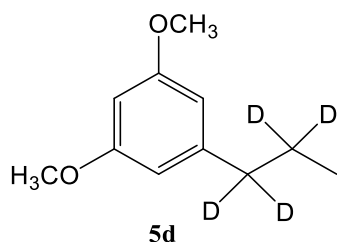


The spectroscopic data of **5c** matched the ones reported in the literature.⁵ Pale yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.35 (d, J=2.4 Hz, 2H), 6.30 (t, J=2.4 Hz, 1H), 3.78 (s, 6H), 2.53 (t, J=7.6 Hz, 2H), 1.62 (t, J=7.6 Hz, 2H); ¹³C-NMR (CDCl₃): δ 160.6, 145.1, 106.5, 97.5, 55.2, 38.3, 24.1.

General procedure for the preparation of 5d-e

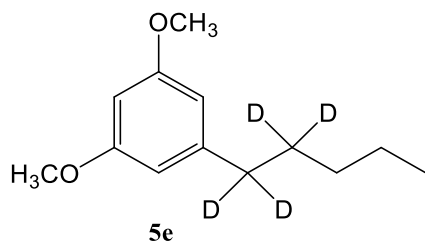
Pd/C (30 w %) and D₂O (3 mL) were placed in a dry 50 mL two-neck round-bottom flask. After five vacuum/H₂ cycles, the system was closed and the mixture was stirred at room temperature for 24 h, then a solution of **4** (0.25 mmol) in THF (1.5 mL) was added. After 6 h stirring at room temperature the reaction mixture was filtered or alternatively centrifuged, then EtOAc and H₂O were added and the aqueous layer was extracted with EtOAc (2 x 5 mL). The organic layers were combined and dried over anhydrous Na₂SO₄. After filtration and evaporation of the solvent under reduced pressure, no purification of the crude product was required.

1,3-dimethoxy-5-[1',1',2',2'-d₄]propylbenzene 5d



Yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.35 (d, J=2.4 Hz, 2H), 6.31 (t, J=2.4 Hz, 1H), 3.79 (s, 6H), 0.93 (s, 3H); ¹³C-NMR (CDCl₃): δ 160.6, 145.0, 106.5, 97.5, 55.2, 13.6. HRMS (ESI) m/z for C₁₁H₁₃D₄O₂ [M+H]⁺ calcd 185.1474, found 185.1487.

1,3-dimethoxy-5-[1,1',2',2'-d₄]pentylbenzene 5e

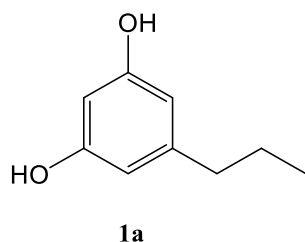


Yellow oil. Crude product. ¹H-NMR (CDCl₃): δ 6.36 (d, J=2.4 Hz, 2H), 6.31 (t, J=2.4 Hz, 1H), 3.79 (s, 6H), 1.36-1.32 (m, 4H), 0.91 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 160.6, 145.3, 106.4, 97.5, 55.1, 35.5 (m), 31.2, 30.0 (m), 22.5, 14.0. HRMS (ESI) m/z for C₁₃H₁₇D₄O₂ [M+H]⁺ calcd 213.1787, found 213.1792.

General procedure for the preparation of 1

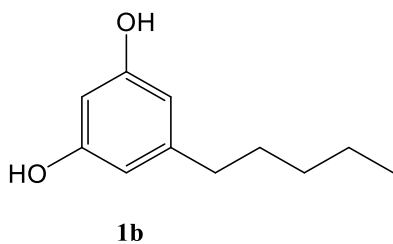
The adduct **5** (0.25 mmol) was dissolved in dry CH₂Cl₂ (5 mL) and the resulting solution was cooled to -78°C. A solution of BBr₃ in CH₂Cl₂ (sol. 1M, 4 eq, 1.0 mmol, 1.0 ml) was added dropwise. After warming at rt, the mixture was stirred overnight and then, after cooling the mixture to 0°C, H₂O (5 mL) was added dropwise until the evolution of gas ceased; the aqueous layer was extracted with CH₂Cl₂ (1 x 5 ml) and EtOAc (2 x 5 mL). The organic layers were combined and dried over anhydrous Na₂SO₄. After filtration and evaporation of the solvent under reduced pressure, the crude mixture was directly purified by column chromatography (silica gel, petroleum ether/EtOAc 7/3) to obtain pure products **1**.

5-propylbenzene-1,3-diol 1a



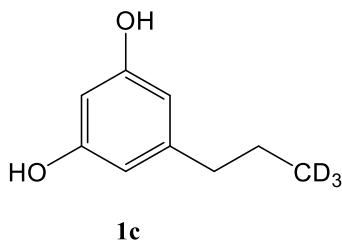
The spectroscopic data of **1a** matched the ones reported in the literature.⁴ Yellow oil. 33.3 mg (0.219 mmol, 88% yield from **2**). R_f 0.3 in Etp/AcOEt 7/3. ¹H-NMR (CDCl₃): δ 6.25 (d, J=2.4 Hz, 2H), 6.19 (t, J=2.4 Hz, 1H), 5.07 (bs, 1H), 2.42 (t, J=7.2 Hz, 2H), 1.60-1.50 (m, 2H), 0.89 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 156.3, 146.0, 108.2, 100.3, 37.8, 24.1, 13.7.

5-pentylbenzene-1,3-diol (olivetol) **1b**



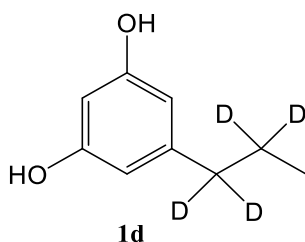
The spectroscopic data of **1b** matched the ones reported in the literature.² Yellow wax. 39.6 mg (0.22 mmol, 88% yield from **2**). Rf 0.3 in Etp/AcOEt 7/3. ¹H-NMR (CDCl₃): δ 6.24 (m, 2H), 6.17 (m, 1H), 4.65 (bs, 2H), 2.49 (t, J=7.6 Hz, 2H), 1.62-1.54 (m, 2H), 1.38-1.25 (m, 4H), 0.89 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 156.6, 108.2, 100.5, 35.8, 31.4, 30.8, 22.5, 13.9.

5-(propyl-3,3,3-d₃)benzene-1,3-diol **1c**



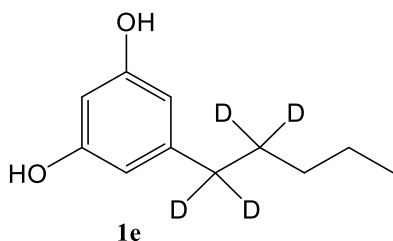
The spectroscopic data of **1c** matched the ones reported in the literature.^{5,6} Pale yellow oil. 33.7 mg (0.22 mmol, 87% yield from **2**). Rf 0.3 in Etp/AcOEt 7/3. ¹H-NMR (CDCl₃): δ 6.80 (bs, 2H), 6.27 (d, J=2.4 Hz, 2H), 6.22 (t, J=2.4 Hz, 1H), 2.35 (t, J=7.6 Hz, 2H), 1.47 (t, J=7.6 Hz, 2H); ¹³C-NMR (CDCl₃): δ 155.9, 146.2, 108.4, 100.4, 37.7, 23.7.

5-(propyl-1,1,2,2-d₄)benzene-1,3-diol **1d**



Yellow oil. 33.5 mg (0.21 mmol, 86% yield from **2**). Rf 0.3 in Etp/AcOEt 7/3. ¹H-NMR (CDCl₃): δ 6.25 (d, J=2.4 Hz, 2H), 6.19 (t, J=2.4 Hz, 1H), 5.29 (bs, 2H), 0.87 (bs, 3H); ¹³C-NMR (CDCl₃): δ 156.4, 145.9, 108.1, 100.2, 13.5. HRMS (ESI) m/z for C₉H₈D₄NaO₂ [M+Na]⁺ calcd 179.0981, found 179.0970.

5-(pentyl-1,1,2,2-d₄)benzene-1,3-diol **1e**

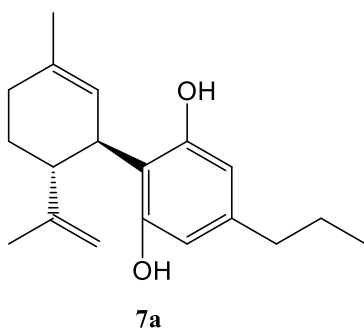


Yellow oil. 42 mg (0.23 mmol, 91% yield from **2**). R_f 0.3 in Etp/AcOEt 7/3. ¹H-NMR (CDCl₃): δ 6.41 (bs, 2H), 6.26 (d, J=2.4 Hz, 2H), 6.21 (t, J=2.4 Hz, 1H), 1.32-1.21 (m, 4H), 0.86 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 156.4, 146.0, 108.0, 100.2, 35.0 (m), 31.2, 29.7 (m), 22.4, 13.9. HRMS (ESI) m/z for C₁₁H₁₂D₄NaO₂ [M+H]⁺ calcd 207.1294, found 207.1301.

General procedure for the preparation of **7** and **8**

To a solution of alkylresorcinol **1** (0.19 mmol) and *p*TSA (33 mol %, 11.9 mg) in toluene (1.5 mL), a solution of terpene **6** (1 eq, 0.19 mmol, 28.9 mg) in toluene (0.5 mL) was added dropwise at 0°C. The mixture was stirred at this temperature for 3 h, then H₂O (5 mL) was added. After warming at rt, the aqueous layer was extracted with EtOAc (3 x 5 mL). The organic layers were combined and dried over anhydrous Na₂SO₄. After filtration and evaporation of the solvent under reduced pressure, the crude mixture was directly purified by column chromatography (silica gel, petroleum ether/EtOAc 9/1) to obtain pure products **7** and **8**.

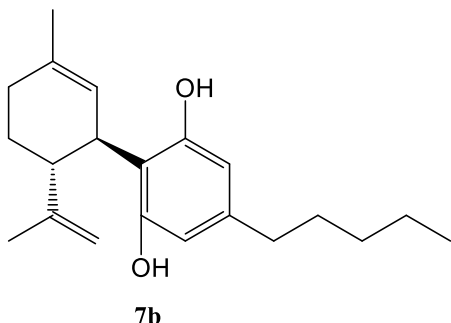
Cannabidivarin (CBDV) **7a**



The spectroscopic data of **7a** matched the ones reported in the literature.⁴ Yellow wax. 13.6 mg (0.047 mmol, 25% yield). R_f 0.6 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.28-6.15 (m, 2H), 5.98 (bs, 1H), 5.57 (bs, 1H), 4.67-4.64 (m, 2H), 4.56-4.55 (m, 1H), 3.87-3.81 (m, 1H), 2.44-2.36 (m, 3H), 2.27-2.18 (m, 1H), 2.12-2.06 (m, 1H), 1.86-1.75 (m and s overlapping, total 5H, especially

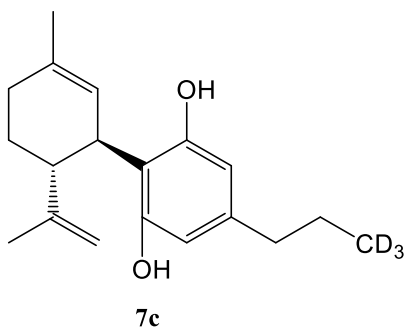
1.79, s, 3H), 1.66 (s, 3H), 1.63-1.53 (m, 2H), 0.90 (t, J=7.2 Hz, 3H); ^{13}C -NMR (CDCl_3): δ 149.4, 142.7, 140.1, 124.1, 113.7, 110.8, 46.1, 37.5, 37.2, 30.4, 28.4, 24.0, 23.7, 20.5, 13.8.

Cannabidiol (CBD) 7b



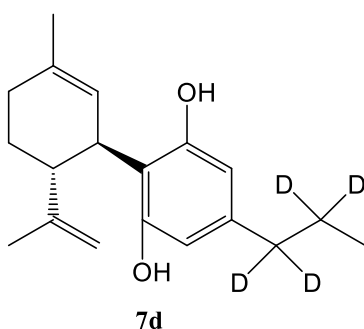
The spectroscopic data of **7b** matched the ones reported in the literature.^{4,7,8} Pale yellow wax. 15.4 mg (0.049 mmol, 26% yield). Rf 0.6 in Etp/AcOEt 9/1. ^1H -NMR (CDCl_3): δ 6.29-6.13 (m, 2H), 5.98 (bs, 1H), 5.57-5.56 (m, 1H), 4.67-4.62 (m, 2H), 4.56 (m, 1H), 3.88-3.81 (m, 1H), 2.46-2.36 (m, 3H), 2.29-2.18 (m, 1H), 2.13-2.05 (m, 1H), 1.86-1.72 (m and s overlapping, total 5H, especially 1.79, s, 3H), 1.65 (s, 3H), 1.60-1.51 (m, 2H), 1.35-1.22 (m, 4H), 0.88 (t, J=7.2 Hz, 3H); ^{13}C -NMR (CDCl_3): δ 149.4, 143.0, 140.1, 124.1, 113.7, 110.8, 46.1, 37.2, 35.4, 31.5, 30.6, 30.4, 28.4, 23.7, 22.5, 20.5, 14.0.

Cannabidivarin- d_3 (CBDV- d_3) 7c



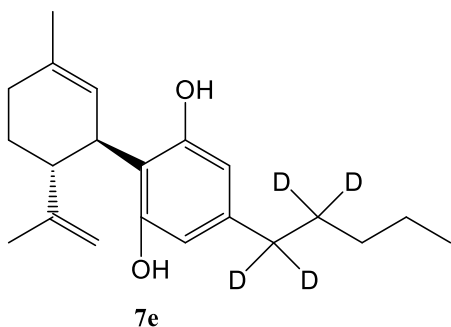
The spectroscopic data of **7c** matched the ones reported in the literature.⁸ Yellow wax. 13.2 mg (0.045 mmol, 24% yield). Rf 0.6 in Etp/AcOEt 9/1. ^1H -NMR (CDCl_3): δ 6.28-6.15 (m, 2H), 5.99 (bs, 1H), 5.57-5.56 (m, 1H), 4.80 (m, 1H), 4.67-4.65 (m, 1H), 4.56-4.55 (m, 1H), 3.89-3.82 (m, 1H), 2.44-2.36 (m, 3H), 2.27-2.18 (m, 1H), 2.12-2.06 (m, 1H), 1.86-1.75 (m and s, overlapping, total 5H, especially 1.79, s, 3H), 1.66 (s, 3H), 1.56 (t, J=7.2 Hz, 2H); ^{13}C -NMR (CDCl_3): δ 149.4, 142.8, 140.1, 124.1, 113.7, 110.8, 46.1, 37.5, 37.2, 30.4, 28.4, 23.7, 20.5.

***Cannabidivarin-d₄* (CBDV-d₄) 7d**



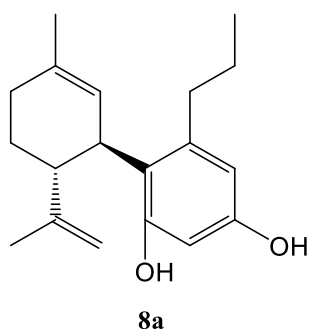
Yellow wax. 11.6 mg (0.040 mmol, 21% yield). Rf 0.6 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.30-6.13 (m, 2H), 5.98 (bs, 1H), 5.57 (m, 1H), 4.67-4.60 (m, 2H), 4.56-4.55 (m, 1H), 3.89-3.80 (m, 1H), 2.39 (td, J=11.2, 3.6 Hz, 1H), 2.30-2.18 (m, 1H), 2.13-2.04 (m, 1H), 1.86-1.74 (m and s overlapping, total 5H, especially 1.79, s, 3H), 1.65 (bs, 3H), 0.88 (s, 3H); ¹³C-NMR (CDCl₃): δ 149.4, 142.7, 140.1, 124.1, 113.7, 110.8, 46.1, 37.2, 30.4, 28.4, 23.7, 20.5, 13.5; HRMS (ESI) m/z for C₁₉H₂₃D₄O₂ [M+H]⁺ calcd 291.2257, found 291.2221.

***Cannabidiol-d₄* (CBD-d₄) 7e**



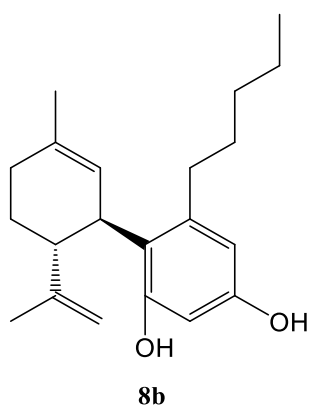
Pale yellow wax. 14.5 mg (0.045 mmol, 24% yield). Rf 0.6 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.28-6.15 (m, 2H), 6.00 (bs, 1H), 5.57 (m, 1H), 5.09 (bs, 1H), 4.65-4.63 (m, 1H), 4.54 (m, 1H), 3.90-3.83 (m, 1H), 2.46-2.36 (m, 1H), 2.29-2.18 (m, 1H), 2.13-2.05 (m, 1H), 1.86-1.72 (m and s overlapping, total 5H, especially 1.79, s, 3H), 1.66 (s, 3H), 1.35-1.22 (m, 4H), 0.86 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 149.0, 142.8, 139.9, 124.1, 113.7, 110.8, 109.5, 107.9, 46.1, 36.9, 31.2, 30.3, 28.3, 23.6, 22.4, 20.2, 14.0. HRMS (ESI) m/z for C₁₉H₂₃D₄O₂ [M+H]⁺ calcd 319.2570, found 319.2579.

Abn-Cannabidiarin (abn-CBDV) 8a



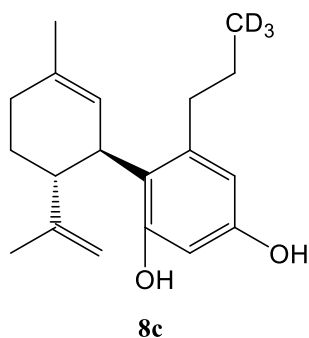
The spectroscopic data of **8a** matched the ones reported in the literature.⁴ Yellow oil. 16.2 mg (0.057 mmol, 30% yield). R_f 0.2 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.21 (d, J=2.8 Hz, 1H), 6.19 (d, J=2.8 Hz, 1H), 6.05 (bs, 1H), 5.52 (bs, 1H), 4.65-4.62 (m, 2H), 4.46 (bs, 1H), 3.54-3.51 (m, 1H), 2.62-2.54 (m, 1H), 2.51-2.44 (td, J=11.6, 3.2 Hz, 1H), 2.27-2.20 (m, 2H), 2.12-2.06 (m, 1H), 1.87-1.72 (m and s overlapping, total 5H, especially 1.79, s, 3H), 1.53 (s, 3H), 1.51-1.41 (m, 2H), 0.92 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 156.4, 154.5, 147.6, 143.7, 139.8, 124.7, 120.0, 111.4, 108.6, 102.1, 44.9, 39.9, 36.1, 30.2, 28.1, 24.5, 23.6, 21.3, 14.2.

abn-Cannabidiol (abn-CBD) 8b



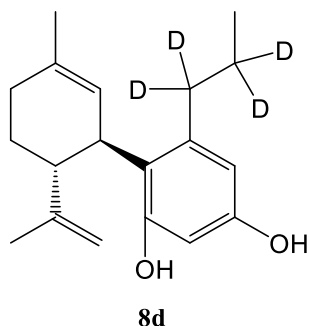
The spectroscopic data of **8b** matched the ones reported in the literature.^{4,8} Yellow oil. 22.7 mg (0.072 mmol, 38% yield). R_f 0.2 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.21 (d, J=2.8 Hz, 1H), 6.19 (d, J=2.8 Hz, 1H), 6.06 (bs, 1H), 5.52 (bs, 1H), 4.67-4.62 (m, 2H), 4.45 (bs, 1H), 3.56-3.50 (m, 1H), 2.62-2.55 (m, 1H), 2.50-2.44 (m, 1H), 2.29-2.16 (m, 2H), 2.12-2.06 (m, 1H), 1.87-1.72 (m and s overlapping, total 5H, especially 1.79, s, 3H), 1.53 (s, 3H), 1.52-1.42 (m, 2H), 1.37-1.24 (m, 4H), 0.92 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 156.4, 154.5, 147.6, 143.9, 139.8, 124.7, 119.9, 111.4, 108.5, 102.1, 44.9, 39.9, 33.9, 31.9, 31.1, 30.2, 28.1, 23.6, 22.5, 21.3, 14.0.

Abn-Cannabidivarin-*d*₃ (CBDV-*d*₃) 8c



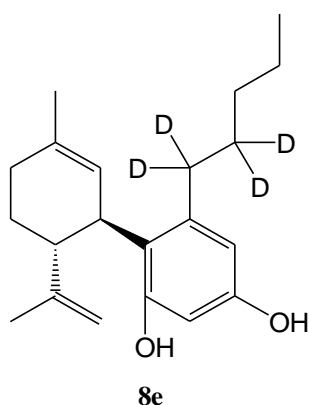
The spectroscopic data of **8c** matched the ones reported in the literature.⁸ Yellow oil. 11.0 mg (0.038 mmol, 20% yield). R_f 0.2 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.21 (d, J=2.8 Hz, 1H), 6.19 (d, J=2.8 Hz, 1H), 6.05 (bs, 1H), 5.52 (bs, 1H), 4.65-4.62 (m, 2H), 4.46 (bs, 1H), 3.56-3.51 (m, 1H), 2.62-2.54 (m, 1H), 2.51-2.44 (td, J=11.6, 3.2 Hz, 1H), 2.27-2.19 (m, 2H), 2.12-2.06 (m, 1H), 1.87-1.72 (m and s, overlapping, total 5H, especially 1.79, s, 3H), 1.53 (s, 3H), 1.51-1.41 (m, 2H); ¹³C-NMR (CDCl₃): δ 156.4, 154.5, 147.6, 143.7, 139.8, 124.7, 120.0, 111.4, 108.6, 102.1, 44.9, 39.9, 36.0, 30.2, 28.1, 24.2, 23.6, 21.3.

Abn-Cannabidivarin-*d*₄ (abn-CBDV-*d*₄) 8d



Yellow oil. 14.9 mg (0.051 mmol, 27% yield). R_f 0.2 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.21 (d, J=2.8 Hz, 1H), 6.18 (d, J=2.8 Hz, 1H), 6.04 (bs, 1H), 5.52 (bs, 1H), 4.65-4.62 (m, 1H), 4.46 (bs, 1H), 3.56-3.51 (m, 1H), 2.62-2.54 (m, 1H), 2.51-2.44 (m, 1H), 2.27-2.18 (m, 1H), 2.12-2.06 (m, 1H), 1.87-1.72 (m and s overlapping, total 5H, especially 1.78, bs, 3H), 1.53 (s, 3H), 0.90 (s, 3H); ¹³C-NMR (CDCl₃): δ 156.4, 154.5, 147.6, 143.7, 139.8, 124.7, 120.0, 111.4, 108.5, 102.1, 44.9, 39.9, 30.2, 28.1, 23.6, 21.3, 13.9. HRMS (ESI) m/z for C₁₉H₂₃D₄O₂ [M+H]⁺ calcd 291.2257, found 291.2228.

***abn*-Cannabidiol-*d*₄ (*abn*-CBD-*d*₄) 8e**



Yellow oil. 15.7 mg (0.049 mmol, 26% yield). R_f 0.2 in Etp/AcOEt 9/1. ¹H-NMR (CDCl₃): δ 6.21 (d, J=2.8 Hz, 1H), 6.19 (d, J=2.8 Hz, 1H), 6.09 (bs, 1H), 5.52 (bs, 1H), 5.33 (bs, 1H), 4.67-4.62 (m, 1H), 4.46 (bs, 1H), 3.56-3.50 (m, 1H), 2.50-2.44 (m, 1H), 2.29-2.16 (m, 1H), 2.12-2.06 (m, 1H), 1.87-1.72 (m and s overlapping, total 5H, especially 1.79, s, 3H), 1.53 (s, 3H), 1.37-1.24 (m, 4H), 0.89 (t, J=7.2 Hz, 3H); ¹³C-NMR (CDCl₃): δ 156.2, 154.6, 147.6, 143.9, 139.8, 124.7, 119.8, 111.4, 108.6, 102.1, 44.9, 40.0, 31.6, 30.2, 28.0, 23.6, 22.4, 21.3, 14.0. HRMS (ESI) m/z for C₁₉H₂₃D₄O₂ [M+H]⁺ calcd 319.2570, found 319.2582.

References

- 1 J. H. Kim, T. Song and Y. K. Chung, *Org. Lett.*, 2017, **19**, 1248-1251.
- 2 M. Sisa, M. Dvorakova and T. Vanek, *Tetrahedron*, 2017, **73**, 5297-5301; M. Sisa, M. Dvorakova, V. Temml, V. Jarosova, T. Vanek and P. Landa, *Eur. J. Med. Chem.*, 2020, **204**, 112620.
- 3 J. Poldy, R. Peakall and R. A. Barrow, *Org. Biomol. Chem.*, 2009, **7**, 4296-4300.
- 4 E. Chiurchiù, S. Sampaolesi, P. Allegrini, D. Ciceri, R. Ballini, A. Palmieri, *EurJOC*, 2021, 1286-1289.
- 5 S. P. Nikas, G. A. Thakura and A. Makriyannis, *J. Chem. Soc. Perkin Trans 1*, 2002, 2544-2548.
- 6 S. P. Nikas, G. A. Thakur, D. Parrish, S. O. Alapafuja, M. A. Huestis and A. Makriyannis, *Tetrahedron*, 2007, **63**, 8112-8123.
- 7 Y. H. Choi, A. Hazekamp, A. M G Peltenburg-Looman, M. Frédérich, C. Erkelens, A. W. M. Lefeber and R. Verpoorte, *Phytochem. Anal.*, 2004, **15**, 345-354.

- 8 A. Makriyannis, S. P. Nikas and S. O. Alapafuja, Angiogenic resorcinol derivatives, Northeastern University in Boston, Massachusetts-WO2011/6099, Patent Family members: *US2012/172339 A1; WO2011/6099 A1*

¹H-NMR, ¹³C-NMR, gHSQC spectra

