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Supporting Information for

The Synthesis and Properties Research of Functionalized Polyolefins

Zhanshan Ma,^{a, ‡} Minghang Ji,^{a, ‡} Wenmin Pang,^b Guifu Si,^{b,*}and Min Chen^{a,*}

^a Institutes of Physical Science and Information Technology, Anhui University, Hefei, Anhui 230601, China.
 ^b CAS Key Laboratory of Soft Matter Chemistry, Department of Polymer Science and Engineering, University of Science and Technology of China, Hefei, 230026, China.

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1 Experimental Section

1.1 General methods.

Tetrahydrofuran (THF) and toluene (both from Avantor) were passed through an activated alumina column under nitrogen to remove protic impurities. All other reagent grade solvents were used without further purification. *cis*-cyclooctene (**COE**), (IMesH₂)-(IPrO)RuCl₂(CHPh) (Hoveyda-Grubbs second generation catalyst, **HGII**), dimethyl maleate were purchased from Sigma-Aldrich. The deuterated solvents CD₂Cl₂, CDCl₃, and 1,1,2,2-tetradhloroethane-d₂ were purchased from Cambridge Isotope Laboratories. **COE** was distilled over CaH₂ prior to use. HDPE: DMDA-8008H, produced by the China National Petroleum Corporation, PET: Item No. P875573, purchase with Shanghai Makclin Biochemical Co., Ltd.; All other chemicals were used as received. **M1**, **M2**, **M3** and **M5** were synthesized according to literature¹⁻⁴; **M4** is commercially available.

All experiments were carried out under a dry nitrogen atmosphere using standard Schlenk techniques or in a glove-box. ¹H NMR spectra were recorded by a Bruker Ascend Tm 400 spectrometer at ambient temperature. ¹H NMR chemical shifts were referenced to residual deuterated solvent peaks or the tetramethylsilane signal (0 ppm). Molecular weights and molecular weight distributions were determined by gel permeation chromatography (GPC) employing a series of two linear Styragel columns (HR2 and HR4) at an oven temperature of 45 °C. A Waters 1515 pump and Waters 2414 differential refractive index detector (30 °C) were used. The eluent was THF at a flow rate of 1.0 mL/min. A series of low polydispersity polystyrene standards was used for calibration. Molecular weight and molecular weight distribution of the hydrogenated polymer were determined by gel permeation chromatography (GPC) with a PL-220 equipped with two Agilent PLgel Olexis columns at 150 °C using 1, 2, 4-trichlorobenzene as a solvent, and the calibration was made using polystyrene standard. DSC measurements were performed on a TA Instruments DSC Q20. Samples (ca. 5 mg) were annealed by heating to 150 °C at 20 °C/min, cooled to 40 °C at 20 °C/min, and then analyzed while being heated to 150 °C at 20 °C/min.

A standard test method, ASTM 638, was followed to measure the tensile properties of the polyethylene samples. Polymers were melt-pressed at 30 to 35°C above their melting point to obtain the dog-bone-shaped tensile-test specimens. The test specimens showed 25-mm gauge length, 2-mm

width, and thickness of 0.4 mm. Stress/strain experiments were performed at 10 m/min using a Universal Test Machine (UTM2502) at room temperature. At least three specimens of each copolymer were tested.

Hydrogenation of Polymer: Hydrogenation by chemical hydrogenation procedure. An example of the chemical hydrogenation procedure is described for the hydrogenation of poly(1,4-divinylbenzene-COE) (entry 5 in Table S1). A mixture of poly(1,4-divinylbenzene-COE) (1.00 g, 15.00 mmol of olefin), *p*-toluenesulfonhydrazide (2.32 g, 16 mmol), tributylamine (2.32 ml, 16 mmol) and o-xylene (40 mL) was refluxed for 8 h, and then allowed to cool to room temperature. The reaction mixture was poured into methanol and the polymer precipitated. The precipitated polymer was isolated by decantation and purified by repeating reprecipitation using dichloromethane/methanol system. The polymer was freeze dried overnight from methanol solution to afford hydrogenated **copolymer-1**. The dried hydrogenated **copolymer-1** need to repeat the above chemical hydrogenation procedure. The hydrogenation of **copolymer-1** (0.98g, 98wt%) as a white solid.

1.2 Polymerization.

Polymerization of *cis*-cyclooctene with diene/triene monomers and chemical hydrogenation.



The preparation of sample is described in detail: anhydrous dichloromethane (40 mL), **M1** (0.15 g, 0.77 mmol) and **COE** (1.70 g, 15.4 mmol) were transferred via injector into a 100 mL Schlenk flask with a constant nitrogen purge. The flask and its contents were place under vacuum and then recharged with nitrogen. The flask was immersed in an oil-bath at 40 °C immediately before transferring **HGII** catalyst (12.53 mg, 20 µmol) as a solution in 1 mL of dichloromethane. After a desired amount of time, it was evaporated to dryness under vacuum, then precipitating into 200 mL methanol. The unsaturated **copolymer-1** was isolated, washed with methanol (3×50 mL) and dried under vacuum for 12 hours at 45 °C. The polymer was characterized by ¹H NMR spectrum and GPC. And the unsaturated **copolymer-2**, **copolymer-3**, **copolymer-4** and **copolymer-5** were synthesized by the similar method except different diene/triene monomers.



An example of the chemical hydrogenation procedure is described for the hydrogenation of polymer. A mixture of the above-mentioned unsaturated **copolymer-1** (5.66 mmol of olefin), p-toluenesulfonyl hydrazide (4.67g, 25 mmol), tripropylamine (4.01g, 28 mmol), small amount of BHT (5 mg), and o-xylene (50 mL) were refluxed for 8 h, and then allowed to cool to room temperature. The reaction mixture was poured into methanol and the polymer precipitated. The precipitated polymer was isolated by decantation and purified by repeating reprecipitation using dichloromethane /methanol system. The polymer was drained of solvent in a vacuum oven to afford hydrogenated **copolymer-1** as a solid. The polymer was characterized by high temperature ¹H NMR spectrum and high temperature GPC. The other hydrogenated polymers (**copolymer-2, copolymer-3, copolymer-4 and copolymer-5**) were prepared by corresponding unsaturated copolymers.

2. ¹H NMR and ¹H-¹H COSY NMR of the copolymers.



Figure S1. (a) ¹H NMR spectrum before degradation of copolymer- $3^{1/20}$ (C₂D₂Cl₄, 120°C)

(b) ¹H NMR spectrum after degradation of copolymer- $3^{1/20}$ (C₂D₂Cl₄, 120°C).



Figure S2. (a) ¹H NMR spectrum before degradation of copolymer-3^{1/100} (C₂D₂Cl₄, 120°C)
(b) ¹H NMR spectrum after degradation of copolymer-3^{1/100} (C₂D₂Cl₄, 120°C).



Figure S4. ¹H COSY NMR spectrum of the copolymer-1^{1/20} (CDCl₃).



Figure S5. ¹H NMR spectrum of the copolymer- $1^{1/20}$ (C₂D₂Cl₄, 120°C).



 $Incorporation(\%) = \frac{I(H-1)/4}{I(H-1)/4 + I(H-4)/2} \times 100\% = \frac{4.00/4}{4.00/4 + 42.39/2} \times 100\% = 4.5\%$



Figure S7. ¹H COSY NMR spectrum of the copolymer-2^{1/20} (CDCl₃).



Figure S8. ¹H NMR spectrum of the copolymer- $2^{1/20}$ (C₂D₂Cl₄, 120°C).

Incorporation(%)

$$=\frac{I(H-1)/4}{\frac{I(H-1)}{4} + I(H-4 + H-5)/4} \times 100\% = \frac{4.00/4}{4.00/4 + (339.29 + 6)/4} \times 100\% = 1.0\%$$



Figure S10. ¹H COSY NMR spectrum of the copolymer-3^{1/20} (CDCl₃).











Figure S12. ¹H NMR spectrum of the copolymer-4^{1/20} (CDCl₃).

 $Incorporation(\%) = \frac{I(H-1)/2}{I(H-1)/2 + I(H-6)/2} \times 100\% = \frac{2.08/3}{2.08/2 + 46.41/2} \times 100\% = 4.3\%$



Figure S13. ¹H COSY NMR spectrum of the copolymer-4^{1/20} (CDCl₃).





Incorporation(%)

$$=\frac{I(H-1)/2}{I(H-1)/2 + I(H-6+H-7)/4} \times 100\% = \frac{2.00/2}{2.00/2 + (340.64+6)/4} \times 100\% =$$

1.1%



Figure S16. ¹H COSY NMR spectrum of the copolymer-5^{1/20} (CDCl₃).



Figure S17. ¹H NMR spectrum of the copolymer- $5^{1/20}$ (C₂D₂Cl₄, 120°C). Incorporation(%)

$$=\frac{I(H-1)/3}{I(H-1)/3 + I(H-6+H-7)/4} \times 100\% = \frac{3.00/3}{3.00/3 + (384.72+9.02)/4} \times 100\%$$

= 1.0%

3. GPC and DSC Spectrum of the Copolymers.



Figure S18. GPC trace of the copolymer-1^{1/5} from Table 1, Entry 1.



Figure S19. GPC trace of the copolymer-1^{1/20} from Table 1, Entry 2.



Figure S20. GPC trace of the copolymer-1^{1/100} from Table 1, Entry 3.



Figure S21. GPC trace of the copolymer-1^{1/200} from Table 1, Entry 4.



Figure S22. GPC trace of the copolymer-2^{1/5} from Table 1, Entry 5.



Figure S23. GPC trace of the copolymer- $2^{1/20}$ from Table 1, Entry 6.



Figure S24. GPC trace of the copolymer-2^{1/100} from Table 1, Entry 7.



Figure S25. GPC trace of the copolymer-2^{1/200} from Table 1, Entry 8.



Figure S26. GPC trace of the copolymer-3^{1/5} from Table 1, Entry 9.



Figure S27. GPC trace of the copolymer-3^{1/20} from Table 1, Entry 10.



Figure S28. GPC trace of the copolymer-3^{1/100} from Table 1, Entry 11.



Figure S29. GPC trace of the copolymer-3^{1/200} from Table 1, Entry 12.



Figure S30. GPC trace of the copolymer-4^{1/5} from Table 1, Entry 13.



Figure S31. GPC trace of the copolymer-4^{1/20} from Table 1, Entry 14.



Figure S32. GPC trace of the copolymer-4^{1/100} from Table 1, Entry 15.



Figure S33. GPC trace of the copolymer-4^{1/200} from Table 1, Entry 16.



Figure S34. GPC trace of the copolymer-5^{1/5} from Table 1, Entry 17.



Figure S35. GPC trace of the copolymer- $5^{1/20}$ from Table 1, Entry 18.



Figure S36. GPC trace of the copolymer-5^{1/100} from Table 1, Entry 19.



Figure S37. GPC trace of the copolymer-5^{1/200} from Table 1, Entry 20.



Figure S38. DSC trace of the copolymer-1^{1/20} from Table 1, Entry 2.



Figure S39. DSC trace of the copolymer-1^{1/100} from Table 1, Entry 3.



Figure S40. DSC trace of the copolymer-1^{1/200} from Table 1, Entry 4.



Figure S41. DSC trace of the copolymer- $2^{1/20}$ from Table 1, Entry 6.



Figure S42. DSC trace of the copolymer-2^{1/200} from Table 1, Entry 8.



Figure S43. DSC trace of the copolymer-3^{1/5} from Table 1, Entry 9.



Figure S44. DSC trace of the copolymer-3^{1/20} from Table 1, Entry 10.



Figure S45. DSC trace of the copolymer-3^{1/100} from Table 1, Entry 11.



Figure S46. DSC trace of the copolymer-3^{1/200} from Table 1, Entry 12.



Figure S47. DSC trace of the copolymer-5^{1/100} from Table 1, Entry 19.

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