Supporting information

A novel NIR fluorescent probe with ratiometric imaging of cysteine

in Endoplasmic Reticulum

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Experimental part

1. Apparatus

Absorption spectra were accurately measured on a HP-8453 UV/Vis (Agilent) spectrometer. Fluorescence spectra were measured on the F-4500 Spectrophotometer and the EX Slit and EM Slit were both set at 10.0 nm. The pH was measured by a Model PHs-3C meter (Shanghai, China). ¹H NMR and ¹³C NMR spectra were measured on a Bruker DTX-400 spectrometer using TMS as internal reference. HR-MS (high-resolution mass spectrometry) spectra were collected using the Q-T of HR-MS spectrometer (Waters Micromass). Cells were imaged using LEICA TCS SP8 laser scanning confocalmicroscope.

2. Materials

All the reagents were purchased from reagent companies without further purification and directly used in the experiment. The deionized water was purified by Milli-Q. The probe was put into 10 mL DMSO to get a 1mM stock solution. Then it was diluted to 10 μ M in 10 mM HEPES solution with 1 mM CTAB for spectroscopic determination. The interfering ions include: Na⁺, K⁺, Fe²⁺, F⁻, Br⁻, I⁻, CH₃COO⁻, CO₃²⁻, HCO₃⁻, HPO₄²⁻, H₂PO₄⁻, SO₄²⁻, NO₃⁻, NO₂⁻, HS⁻, SO₃²⁻, HSO₃⁻, GSH, Hcy, Cys, NO, HNO, ¹O₂, H₂O₂, HCIO, ·OH,ONOO⁻.

3. Cell culture

HeLa cells were inoculated in 96-well plates, cultured in 5% CO₂, 37°C for 24 h, removed the old medium, washed with PBS for 3 times. Then, the preprepared medium containing different concentrations of probe **HL-Cys** (0, 5 μ M, 10 μ M, 15 μ M and 20 μ M) was added and placed in an incubator for further culture for 24 h. Cell Counting Kit (CCK-8) staining was used to evaluate the cytotoxicity of probe **HL-Cys**. Cell survival rate = experimental group/control group × 100%.

4. Synthesis

Compound 1 and Compound 4 was synthesized according to the previous $\mbox{literatures}^{[a,b]}$

Synthesis of Compound **2**:Compound **1** (242 mg, 1.3 mmol), 5-Formylsalicylic Acid (166 mg, 1 mmol) and piperidine (0.1mL) were dissolved in 7 mL EtOH under argon atmosphere, the mixture was refluxed 10 h. At the end of the reaction, the solvent was removed under reduced pressure and the residue was purified by column chromatography with dichloromethane: methanol (30:1) as the eluent to afford the desire product as a red solid (230 mg, yield 69%). ¹H NMR (400 MHz, DMSO d_6) δ (ppm):8.06(d,J=2.12,1H),7.73(dd,J=8.6,2.16Hz,1H),7.24(q,J=16.08Hz, 2H),6.83(t,J=4.64Hz,2H),2.59(s,2H),2.53(s,2H),1.01(s,6H).¹³CNMR(100MHz,DMSO d_6) δ (ppm):171.9,170.7,164.8,157.2,138.7,133.3,131.4,126.7,126.0,121.8,118.1,117.6, 114.6, 113.8, 75.2, 42.8, 38.6, 32.1, 27.9.HR-MS: *m/z* calcd for C₂₀H₁₈N₂O₃[M - H]⁻: =333.1244, found: 333.1250.

Synthesis of Compound **3**: Compound **2** (100 mg, 0.3 mmol) and N-hydroxysuccinimide (41.4mg,0.36 mmol) were dissolved in THF (4 mL) was added DCC (74.2 mg, 0.36 mmol). The mixture was stirred at room temperature for 30 min. At the end of the reaction, the solvent was removed under reduced pressure and the residue was purified by column chromatography with Petroleum ether: dichloromethane (1:1) as the eluent to afford the desire product as a yellow solid (40 mg, yield 31%). ¹H NMR (400 MHz, DMSO-*d*₆) δ (ppm): 12.21 (s, 1H), 8.12 (d, *J* = 1.88 Hz, 1H), 8.00 (dd, *J* = 1.88, 1.92 Hz, 1H), 7.38 - 7.29 (m, 2H), 7.11 (d, *J* = 8.72 Hz, 1H), 6.87 (s, 1H), 2.90 (s, 4H).2.61 (s, 2H), 2.54 (s, 2H), 1.01 (s, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ (ppm): 170.9, 170.8, 161.1, 161.0, 156.5, 136.8, 135.0, 132.4, 128.6, 128.0, 122.8, 119.0, 114.4, 113.6, 112.0, 76.4, 42.7, 38.6, 32.1, 27.9, 26.0.HR-MS: *m/z* calcd for C₂₄H₂₁N₃O₅[M - H]⁻: = 430.1408, found: 430.1413.

Synthesis of Compound **5**: Compound **3** (86.2 mg, 0.2 mmol) and Compound **4** (55.6mg, 0.3 mmol) were dissolved in DCM (6 mL) was added triethylamine(0.06 mL). The mixture was stirred at room temperature for 2 h. At the end of the reaction, the solvent was removed under reduced pressure and the residue was purified by column chromatography with dichloromethane: methanol (100 :1) as the eluent to afford the desire product as an orange solid (91 mg, yield 86%). ¹H NMR (400 MHz, DMSO-*d*₆) δ (ppm): 12.77 (s, 1H), 8.87 (t, *J* = 5.56 Hz, 1H), 8.15 (d, *J* = 1.88 Hz,1H) 7.76 – 7.72 (m, 2H), 7.69 (s, 1H), 7.67 (s, 1H), 7.36 (s, 1H).7.34 (s, 1H), 7.23 (s, 2H),

6.95 (d, J =8.60 Hz, 1H), 6.81 (s, 1H), 3.37 (q, J = 6.28 Hz, 2H), 2.94 (q, J = 6.28 Hz, 2H),2.54 (s,2H),2.34(s,3H),1.02(s,6H).¹³CNMR(100MHz,DMSO-2H), 2.62 (s, d_6) δ (ppm):170.6,169.1,161.7,156.4,143.1,137.9,137.6,134.0,130.7,127.9,127.8,127.3,127 .0,122.4,118.5,116.1,114.4,76.2,42.7,42.0,38.6,32.1,27.9,21.4.HR-MS:*m*/zcalcdforC₂₉H₃₀N₄O₄S[M-H]⁻:=529.1915,found: 529.1919.

5. References

- Ma Y, Gao W and Ma S, Anal. Chem., 2020, 92, 13405-13410. a)
- b) Song W, Dong B and Lu Y, New J. Chem., 2019, 43, 12103-12108.



Scheme S1. Synthetic route of HL-Cys

Structure	$\lambda_{ex}/\lambda_{em}$ (nm)	Stock's Shifts(nm)	Detectio n Limit	Respo nse time	Targeti ng	Rational	References
NC ₄ CN C C C C C C C C C C C C C	590/770	180	0.4µM	10min	-	-	Anal Chimica Acta 2021, 1171, 33865
Ly COCH	445/500	55	0.122µM	200s	_	-	5 Sens. Actuators B: Chem. 2018, 267, 76-82
	540/714	174	28.6 nM	5min			Sens. Actuators B: Chem. 2022, 357, 131430
QH st	700/770	70	16 nM	10min	Lyso-	_	J. Mater. Chem. B, 2020, 8, 2269
qt & to	560/640	80	0.2 μM	5min	Mito-	F ₆₄₀ /F ₇₈₅	Biosensors and Bioelectroni cs 2015, 74 156-164
9 4 -6-49	535/635	100	0.09 µM	30min	Mito-	F ₆₃₅ /F ₇₉₄	Sens. Actuators B: Chem. 2019, 282, 69-77
	600/760	160	48 nM	5min	-	-	Anal. Chem. 2018 , 90, 1014– 1020

 Table S1 Comparison of reported probes for recognition of Cys

- Anginiza	410/650	240	490 nM	6min	ER	F ₆₅₀ /F ₅₄₅	This work
	390/550	160	1.8 μM	20min	ER	F ₅₅₀ /F ₄₄₀	Anal. Chem. 2019 , 91, 5513– 5516
	441/514	73	8.95 nM	30min	ER	-	Chem Commun 2019, 55, 9629.
ter and the second seco	610/716	116	0.083 μM	-	Mito-	F ₇₁₆ /F ₆₆₀	Sens. Actuators B: Chem. 2018, 259, 219-225
han the	481/675	194	0.2 μM	5min	-	F ₆₇₅ /F ₅₃₀	Dyes. Pigments, 2017, 146 103-111

Characterization of compounds



Figure S1. ¹H NMR spectra of 2 in DMSO- d_6 .



Figure S2. ¹³C NMR spectra of 2 in DMSO- d_6 .



Figure S3. ESI-HRMS spectra of 2.



Figure S5. ¹³C NMR spectra of 3 in DMSO- d_6 .







Figure S8. ¹³C NMR spectra of 5 in DMSO- d_6 .



Figure S9. ESI-HRMS spectra of 5.



Figure S10. ¹H NMR spectra of HL-Cys in DMSO-*d*₆.



Figure S11. ¹³C NMR spectra of HL-Cys in DMSO-d₆.



Figure S12. ESI-HRMS spectra of HL-Cys.



Figure S13. UV–vis absorption spectra of **HL-Cys** (10 μ M) before and after reaction with Cys (100 μ M)in PBS buffer solution (containing 30% DMSO, 10mM, pH=7.4). Inset: left,**HL-Cys**(10 μ M); right, **HL-Cys** (10 μ M) + Cys (100 μ M).



Figure S14. The linear fitting of fluorescence intensity ratio (F_{650}/F_{545}) against concentrations of Cys (0-9 μ M) in PBS buffer solution (containing 30% DMSO, 10mM, pH = 7.4).



Figure S15. HR-MS spectra of HL-Cys in response to Cys.



Figure S16. The fluorescence emission intensity of HL-Cys (10 μ M) in solution of different pH value (4-12).



Figure S17. Time-dependent fluorescence intensity ratio (F_{650}/F_{545}) of the HL-Cys (10 μ M) without (black) and with (red) 100 μ M Cys in PBS buffer solution (containing 30% DMSO, 10 mM, pH = 7.4).



Figure S18. The cytotoxicity test of different concentrations HL-Cys in living HeLa cells for 24 h.