

Synthesis of chemical tools to label the mycomembrane of corynebacteria using a modified Iron (III) chloride-mediated protection of trehalose

Supporting Information

Table of contents:

I. Abbreviations in the manuscript and supporting information	S2
II. Methods	S3
1. Chemical Synthesis	S3
General methods	S3
Synthesis of probes I, II and III	S4
2. Labeling of bacteria with trehalose-based probes	S14
Bacterial strains, growth conditions and reagents	S14
General procedure for labeling <i>C. glutamicum</i>	S14
Flow cytometry quantification	S14
General procedures for confocal imaging of labeled bacteria	S15
3. <i>C. glutamicum</i> cell growth assays	S15
III. Supplementary figures (S1-S2)	S16
IV. NMR spectra	S18
V. References in the supporting information	S32

I. Abbreviations in the manuscript and supporting information

AG	Arabinogalactan
Ag85	Antigen 85 complex
AGM	Arabinogalactan-linked mycolate
Alk	Alkyne
Az	Azide
BHI	Brain Heart Infusion
Bodipy	4,4-Difluoro-4-bora-3a,4a-diaza- <i>s</i> -indacene
CDG	Cephalosporinase-dependent green
CmpL	Corynebacterial membrane proteins large
CR110-CCH	5/6-Carboxyrhodamine 110-PEG ₄ -alkyne
CuAAC	Copper(I)-catalyzed azide-alkyne cycloaddition
Dabcyl	4-(Dimethylaminoazo)benzene-4-carboxylic acid
DCM	Dichloromethane
DMN	4- <i>N,N</i> -Dimethylamino-1,8-naphthalimide
DMF	<i>N,N</i> -Dimethylformamide
DMSO	Dimethylsulfoxide
Fbp	Fibronectin binding protein
FI	Fluorescein
FITC	Fluorescein isothiocyanate
FRET	Fluorescence resonance energy transfer
MA	Mycolic acid
MFI	Mean fluorescence intensity
MM	Mycomembrane
MmpL	Mycobacterial membrane proteins large
Myt	Mycoloyl transferase
PBS	Phosphate-buffered saline
PEG	Polyethylene glycol
PG	Peptidoglycan
Pks	Polyketide synthase
QTF	Quencher trehalose fluorophore
RND	Resistance nodulation division
SPAAC	Strain-promoted alkyne-azide cycloaddition
TBAF	<i>n</i> -Tetrabutylammonium fluoride
TBAN ₃	<i>n</i> -Tetrabutylammonium azide
TCO	<i>trans</i> -Cyclooctene
TDM	Trehalose dimycolate
Tf	Trifluoromethanesulfonyl
TGTA	Tris[(1-glucosyl-1 <i>H</i> -1,2,3-triazol-4-yl)methyl]amine
THF	Tetrahydrofuran
TMM	Trehalose monomycolate
TMS	Trimethylsilyl
TMR	Tetramethylrhodamine
Tre	Trehalose

II. Methods

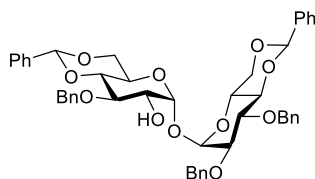
1. Chemical Synthesis

General methods

All air sensitive reactions were carried out in oven-dried glassware under a slight positive pressure of argon. Solvents were dried by standard methods. THF was distilled from sodium benzophenone ketyl. All chemical reagents and dry solvents were of analytical grade, purchased from chemical suppliers, and used without further purifications. TLC (Silica Gel 60 F₂₅₄) were visualized under UV (254 nm) and by staining either in 5% ethanolic sulfuric acid or orcinol or phosphomolybdic acid. Silica gel SDS 60 ACC 35-70 μm was used for column chromatography. NMR spectra were recorded on Bruker DRX 300, AV 360 or Advance 300 or 500 spectrometers. Chemical shifts (in ppm) were determined relative to residual protonated solvent as an internal reference. Abbreviations of multiplicity were as follows: s (singlet), d (doublet), dd (doublet of doublet), t (triplet), at (apparent triplet), m (multiplet), b (broad). Coupling constants in hertz (Hz) were measured from one-dimensional spectra. High-resolution mass spectra (positive or negative mode ESI) were recorded on a Bruker Daltonics micrOTOF-QII spectrometer or on a Waters LCT Premier XE (ToF). Optical rotations were measured on an Anton Paar MCP 150 polarimeter or with an Anton Paar MCP 300 polarimeter in a 10-cm cell at 20 °C and 589 nm (c in g / 100 mL). IR-FT spectra were recorded on a Perkin Elmer Spectrum 100 spectrometer. Characteristic absorptions are reported in cm^{-1} .

Synthesis of probes I, II and III

2',3,3'-Tri-*O*-benzyl-4,6;4',6'-di-*O*-benzylidene- α,α -D-trehalose (**1**)

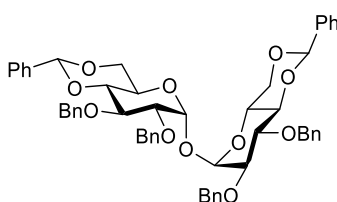


To a solution of per-*O*-silylated α,α -D-trehalose **2**^{i,ii} (200 mg, 0.217 mmol, 1.0 equiv) in dry CH_2Cl_2 (180 μL) at 0 °C were added benzaldehyde (220 μL , 2.174 mmol, 10.0 equiv) and a solution of $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (45 μL of a 242 mM solution in CH_3CN , 5.0 mol%, dropwise). Triethylsilane (140 μL , 0.870 mmol, 4.0 equiv) was then added and the reaction mixture was warmed to room temperature and stirred overnight. Then, the reaction mixture was diluted with EtOAc and quenched by addition of an ice-cold saturated aqueous NaHCO_3 solution. The reaction mixture was extracted three times with EtOAc, and the combined organic layers were dried over anhydrous Na_2SO_4 and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (cyclohexane/EtOAc, 85:15 to 70:30) to give the expected product **1** (68 mg, 40%). The di- and tetra-*O*-benzylated derivatives (i.e. compounds **3** and **5** respectively) were also obtained as by-products, see table 1 of the main text. For larger scales (i. e. 0.5 - 2.0 g of starting material), a TBAF treatment could be required, and the yield of expected compound **1** can decrease to 30-36%.

M = 788.89 g/mol. Chemical formula: $\text{C}_{47}\text{H}_{48}\text{O}_{11}$

Rf = 0.65 (cyclohexane/EtOAc 6/4). IR (cm^{-1}): 3478, 3064, 3032, 2955, 2867, 1496, 1453, 1370, 1250, 1154, 1105, 1071, 1010, 984, 874, 841, 749, 696. ^1H NMR (CDCl_3 , 360 MHz) δ (ppm): 7.55-7.10 (m, 25H, H_{Ar}), 5.58 (s, 2H, 2CH-Ph), 5.19 (d, 1H, $J = 3.8$ Hz, H_1), 5.15 (d, 1H, $J = 3.8$ Hz, H_1'), 5.02 (d, 1H, $J = 11.4$ Hz, CH_2 -Ph), 4.99 (d, 1H, $J = 11.4$ Hz, CH_2 -Ph), 4.87 (d, 1H, $J = 11.4$ Hz, CH_2 -Ph), 4.85 (d, 1H, $J = 11.8$ Hz, CH_2 -Ph), 4.76 (d, 1H, $J = 11.4$ Hz, CH_2 -Ph), 4.74 (d, 1H, $J = 11.8$ Hz, CH_2 -Ph), 4.29 (dd, 1H, $J = 10.0$ Hz, $J = 4.8$ Hz, $\text{H}_{6'a}$), 4.27 (td, 1H, $J = 10.0$ Hz, $J = 5.0$ Hz, H_5), 4.16 (dd, 1H, $J = 10.0$ Hz, $J = 5.0$ Hz, H_{6a}), 4.14 (td, 1H, $J = 10.0$ Hz, $J = 4.8$ Hz, H_5'), 4.09 (at, 1H, $J = 9.2$ Hz, H_3'), 3.98 (at, 1H, $J = 9.2$ Hz, H_3), 3.78 (dd, 1H, $J = 9.2$ Hz, $J = 3.8$ Hz, H_2), 3.75-3.64 (m, 4H, H_{6b} , $\text{H}_{6b'}$, H_4 , H_4'), 3.65 (dd, 1H, $J = 9.2$ Hz, $J = 3.8$ Hz, H_2'). ^{13}C NMR (CDCl_3 , 90 MHz) δ (ppm): 138.8, 138.5, 138.0, 137.5, 137.4 (5C, 5 Cq-Ar), 128.9, 128.8, 128.6, 128.5, 128.4, 128.3, 128.2, 128.2, 128.0, 128.0, 127.9, 127.8, 127.6, 126.2, 126.0 (25C, 25 CH-Ar), 101.3, 101.2 (2C, 2 CH-Ph), 95.5 (C_1), 94.7 (C_1'), 82.3, 82.3 (2C, C_4 , C_4'), 78.8, 78.7, 78.7 (3C, C_3 , C_3' , C_2'), 75.3, 75.0, 73.9 (3C, 3 CH_2 -Ph), 71.8 (C_2), 69.0, 68.9 (2C, C_6 , C_6'), 63.3 (C_5), 63.1 (C_5'). HRMS (ESI⁺): calcd for $\text{C}_{47}\text{H}_{48}\text{NaO}_{11}$ [$\text{M}+\text{Na}$]⁺ 811.3089, found 811.3082. $[\alpha]_{\text{D}}^{20}$: + 61 (c 1.0, CHCl_3).

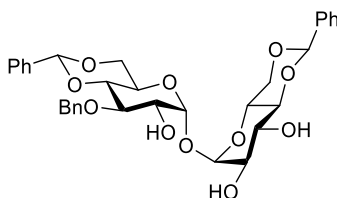
2,2',3,3'-Tetra-*O*-benzyl-4,6;4',6'-di-*O*-benzylidene- α,α -D-trehalose (**5**)



Compound **5** was obtained in 16 % yield as a by-product during the synthesis of **1**.

$C_{54}H_{54}O_{11}$; $M = 879.02 \text{ g}\cdot\text{mol}^{-1}$; $R_f = 0.70$ (cyclohexane/ethyl acetate: 7/3); $^1\text{H NMR}$ (CDCl_3 , 360 MHz) δ (ppm): 7.56-7.28 (m, 15H, H_{ar}), 5.58 (s, 1H, CH-Ph), 5.14 (d, 1H, $J = 3.9 \text{ Hz}$, H_1), 4.99 (d, 1H, $J = 11.0 \text{ Hz}$, $\text{CH}_2\text{-Ph}$), 4.88 (d, 1H, $J = 11.0 \text{ Hz}$, $\text{CH}_2\text{-Ph}$), 4.87 (d, 1H, $J = 12.0 \text{ Hz}$, $\text{CH}_2\text{-Ph}$), 4.76 (d, 1H, $J = 12.0 \text{ Hz}$, $\text{CH}_2\text{-Ph}$), 4.31 (td, 1H, $J = 4.8 \text{ Hz}$, $J = 10.0 \text{ Hz}$, H_5), 4.17 (at, 1H, $J = 9.3 \text{ Hz}$, H_3), 4.15 (dd, 1H, $J = 10.0 \text{ Hz}$, $J = 4.8 \text{ Hz}$, H_{6a}), 3.69 (at, 1H, $J = 10.0 \text{ Hz}$, H_{6b}), 3.67 (dd, 1H, $J = 10.0 \text{ Hz}$, $J = 9.3 \text{ Hz}$, H_4), 3.65 (dd, 1H, $J = 9.3 \text{ Hz}$, $J = 3.9 \text{ Hz}$, H_2); $^{13}\text{C NMR}$ (CDCl_3 , 90 MHz) δ (ppm): 138.9, 138.2, 137.6 (3C, Cq-Ar), 129.0, 128.6, 128.4, 128.3, 128.1, 127.9, 127.7, 126.3 (15C, CH-Ar), 101.3 (CH-Ph), 95.1 (C_1), 82.4 (C_4), 78.8 (2C, C_2 , C_3), 75.2 ($\text{CH}_2\text{-Ph}$), 73.9 ($\text{CH}_2\text{-Ph}$), 69.1 (C_6), 63.0, (C_5); ESI HRMS for $C_{54}H_{55}O_{11}$ $[\text{M}+\text{H}]^+$: Calcd, 879.3739, Found 879.3706; $[\alpha]_D^{20} = +47$ (c 0.75, CHCl_3).

3-*O*-benzyl-4,6;4',6'-di-*O*-benzylidene- α,α -D-trehalose (**4**)



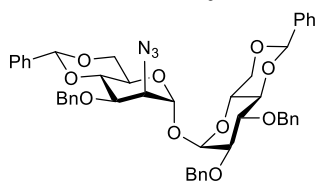
To a solution of per-*O*-silylated α,α -D-trehalose **2**^{i,iii} (100 mg, 0.109 mmol, 1.0 equiv) in dry CH_2Cl_2 (180 μL) at 0 °C were added benzaldehyde (35 μL , 0.354 mmol, 3.2 equiv) and a solution of $\text{FeCl}_3\cdot 6\text{H}_2\text{O}$ (45 μL of a 136 mM solution in CH_3CN , 5.0 mol%, dropwise). Triethylsilane (20 μL , 0.125 mmol, 1.1 equiv) was then added and the reaction mixture was warmed to room temperature and stirred for 2 h. Then, the reaction mixture was diluted with EtOAc and quenched by addition of an ice-cold saturated aqueous NaHCO_3 solution. The reaction mixture was extracted three times with EtOAc, and the combined organic layers were dried over anhydrous Na_2SO_4 and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (cyclohexane/EtOAc, 60:40) to give compound **4** (24 mg, 35%). The di-*O*-benzylated derivative **3** was also obtained as by-product, see table 1 of the main text. NMR data were in agreement with those reported.ⁱⁱⁱ

$M = 608.63 \text{ g}\cdot\text{mol}^{-1}$. Chemical formula: $C_{33}H_{36}O_{11}$

$R_f = 0.14$ (cyclohexane/ethyl acetate: 6/4); $^1\text{H NMR}$ (CDCl_3 , 360 MHz) δ (ppm): 7.52-7.28 (m, 15H, H_{ar}), 5.58 (s, 1H, CH-Ph), 5.50 (s, 1H, CH-Ph), 5.19 (d, 1H $J = 4.0 \text{ Hz}$, H_1), 5.13 (d, 1H, $J = 4.0 \text{ Hz}$, H_1'), 5.01 (d, 1H, $J = 11.5 \text{ Hz}$, $\text{CH}_2\text{-Ph}$), 4.75 (d, 1H, $J = 11.5 \text{ Hz}$, $\text{CH}_2\text{-Ph}$), 4.31

(dd, 1H, $J = 5.0$ Hz, $J = 10.4$ Hz, H_{6b}), 4.27 (dd, 1H, $J = 5.0$ Hz, $J = 10.4$ Hz, 1H, H_{6'b}), 4.14-3.98 (m, 3H, H₅, H_{5'}, H_{3'}), 3.93 (t, 1H, $J = 9.0$ Hz, H₃), 3.81-3.64 (m, 5H, H_{6a}, H₂, H_{6'a}, H_{2'}, H₄), 3.47 (t, 1H, $J = 9.4$ Hz, 1H, H_{4'}), 3.20 (s, 1H, OH), 2.55 (d, 1H $J = 5.8$ Hz, OH), 2.46 (d, 1H, $J = 5.4$ Hz, OH); ¹³C NMR (CDCl₃, 90 MHz) δ (ppm): 138.6, 137.5, 137.3 (3C, Cq-Ar), 129.4, 129.2, 128.7, 128.5, 128.2, 128.1, 126.5, 126.3 (15C, CH-Ar), 102.1, 101.6 (CH-Ph), 95.2, 95.1 (C₁, C_{1'}), 82.4 (C₄), 81.3 (C_{4'}), 78.9 (C₃), 75.2 (CH₂-Ph), 72.6 (C_{2'}), 71.9 (C₂), 71.5 (C_{3'}), 69.1, 69.0 (C₆, C_{6'}), 63.7, 63.2 (C₅, C_{5'}); ESI HRMS for C₃₃H₃₆NaO₁₁ [M+Na]⁺: Calcd 631.2150, Found 631.2144; $[\alpha]_D^{24} = +75$ (*c* 1.6, CHCl₃).

2-Azido-2',3,3'-tri-*O*-benzyl-4,6;4',6'-di-*O*-benzylidene-2-deoxy-2-epi- α,α -D-trehalose (7)



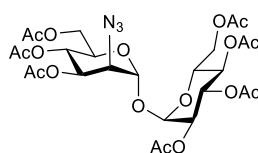
To a solution of compound **1** (500 mg, 0.63 mmol, 1.0 equiv) and anhydrous pyridine (0.5 mL, 6.1 mmol, 10.0 equiv) in anhydrous CH₂Cl₂ (10 mL, 0.06 M) at 0 °C under an argon atmosphere was added triflic anhydride (250.0 μ L, 1.4 mmol, 2.2 equiv). The reaction mixture was stirred at room temperature under an argon atmosphere for 3 hours then cooled to 0 °C. The reaction was quenched by addition of ice-cold saturated aqueous NaHCO₃. The reaction mixture was extracted three times with CH₂Cl₂, and the combined organic layer was dried over anhydrous sodium sulfate and co-evaporated with toluene three times under reduced pressure. The crude material was purified by flash chromatography over silica gel (cyclohexane/EtOAc, 7:3) to give the crude triflate **6** as a white powder. This compound was dissolved in toluene (12.0 mL, 0.06 M) at room temperature under an argon atmosphere and treated with tetrabutylammonium azide (600 mg, 2.1 mmol, 4.0 equiv). The reaction mixture was stirred overnight at 50 °C. After cooling to room temperature, the reaction mixture was diluted with EtOAc and quenched by addition of ice-cold saturated aqueous NaHCO₃. The reaction mixture was extracted three times with EtOAc, and the combined organic layer was dried over anhydrous sodium sulfate and concentrated under vacuum. The crude residue was purified by flash chromatography over silica gel (cyclohexane/EtOAc, 7:3) to give compound **7** (506 mg, 0.62 mmol, 98%) as a white powder.

M = 813.90 g/mol. Chemical formula: C₄₇H₄₇N₃O₁₀

R_f = 0.7 (heptane/EtOAc, 7:3). IR (cm⁻¹): 3064, 3032, 2931, 2867, 2108, 1496, 1453, 1371, 1250, 1129, 1105, 1088, 998, 916, 737. ¹H NMR (CDCl₃, 500 MHz) δ (ppm): 7.56-7.25 (m, 25H, 25 H_{ar}), 5.63 (s, 1H, CH-Ph), 5.58 (s, 1H, CH-Ph), 5.09 (d, 1H, $J = 3.8$ Hz, H_{1'}), 4.99 (d, 1H, $J = 1.5$ Hz, H₁), 4.97 (d, 1H, $J = 11.4$ Hz, CH₂-Ph), 4.94 (d, 1H, $J = 12.0$ Hz, CH₂-Ph), 4.86 (d, 1H, $J = 11.4$ Hz, CH₂-Ph), 4.83 (d, 1H, $J = 11.9$ Hz, CH₂-Ph), 4.73 (d, 1H, $J = 12.0$ Hz, CH₂-Ph), 4.72 (d, 1H, $J = 11.9$ Hz, CH₂-Ph), 4.29 (dd, 1H, $J = 9.0$ Hz, $J = 3.0$ Hz, H_{6'a}), 4.22 (dd, 1H, $J = 9.0$ Hz, $J = 3.6$ Hz, H₃), 4.20-4.10 (m, 3H, H₄, H₅, H_{6a}), 3.99 (at, 1H, $J = 9.5$ Hz, H_{3'}), 3.86 (dd, 1H, $J = 3.6$ Hz, $J = 1.5$ Hz, H₂), 3.79 (at, 1H, $J = 9.7$ Hz, H_{6b}), 3.78-3.70 (m, 2H,

H₅[•], H_{6b}[•]), 3.66 (at, 1H, *J* = 9.5 Hz, H₄[•]), 3.61 (dd, 1H, *J* = 9.5 Hz, *J* = 3.8 Hz, H₂[•]). ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 138.7, 138.1, 137.9, 137.7, 137.5 (5C, 5 C_q-Ar), 129.2, 129.1, 128.8, 128.7, 128.6, 128.5, 128.4, 128.2, 128.2, 128.2, 128.1, 128.0, 127.9, 126.4, 126.2 (25C, 25 CH-Ar), 101.8, 101.5 (2 C, CH-Ph), 95.4 (C₁), 94.4 (C₁[•]), 82.3 (C₄[•]), 79.1 (C₄), 78.5 (C₂[•]), 78.4 (C₃[•]), 75.9 (C₃), 75.4 (CH₂-Ph), 74.4 (CH₂-Ph), 74.1 (CH₂-Ph), 69.1 (C₆[•]), 68.7 (C₆), 64.6 (C₅), 63.8 (C₅[•]), 63.0 (C₂). HRMS (ESI⁺): calcd for C₄₇H₄₇N₃NaO₁₀ [M+Na]⁺ 836.3154, found 836.3192. [α]_D²⁰: + 57 (*c* 0.4, CHCl₃).

2',3,3',4,4',6,6'-Hepta-*O*-acetyl-2-azido-2-deoxy-2-epi-α,α-D-trehalose (8)

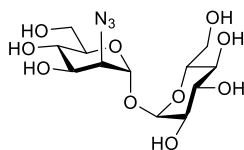


A dry 100 mL round bottom flask was outfitted with a stir bar and a rubber septum. The assembly was put in a drier and cooled under vacuum. The flask was charged with anhydrous iron (III) chloride (542 mg, 3.34 mmol, 21.0 equiv), three cycles of vacuum – argon were done. Iron (III) chloride was suspended in dry CH₂Cl₂ (20.0 mL, 0.17 M) and sonicate during 10 min. A solution of compound **7** (130 mg, 0.16 mmol, 1.0 equiv) in anhydrous CH₂Cl₂ (10 mL, 0.016 M) was added. The suspension was stirred at 0 °C and anhydride acetic (600 μL, 648 mg, 6.35 mmol, 40.0 equiv) was added. The reaction mixture was stirred under an argon atmosphere for 48 hours. Then, the reaction was quenched by addition of ice-cold saturated aqueous NaHCO₃ and extracted three times with CH₂Cl₂, and the combined organic layers were dried over anhydrous sodium sulfate and concentrated under vacuum. The crude product diluted in CH₃OH (≈ 15 mL) was passed rapidly through a short pad of resin (Dowex 50WX8 hydrogen form), concentrated and then purified by flash chromatography on silica gel (CH₂Cl₂/EtOAc, 100:0 to 80:20) to give compound **8** (64 mg, 0.097 mmol, 61%) as a white powder after lyophilization.

M = 661.57 g/mol. Chemical formula: C₂₆H₃₅N₃O₁₇

R_f = 0.4 (heptane/EtOAc 7:3). IR (cm⁻¹): 2960, 2111, 1743, 1433, 1368, 1214, 1143, 1034, 1010, 913, 754. ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 5.42 (dd, 1H, *J* = 10.3 Hz, *J* = 9.4 Hz, H₃[•]), 5.35 (dd, 1H, *J* = 9.7 Hz, *J* = 3.8 Hz, H₃), 5.31 (d, 1H, *J* = 3.9 Hz, H₁[•]), 5.26 (dd, 1H, *J* = 9.8 Hz, *J* = 9.7 Hz, H₄), 5.07 (d, 1H, *J* = 1.8 Hz, H₁), 5.06 (dd, 1H, *J* = 10.0 Hz, *J* = 9.4 Hz, H₄[•]), 4.99 (dd, 1H, *J* = 10.3 Hz, *J* = 3.9 Hz, H₂[•]), 4.24 (dd, 1H, *J* = 12.4 Hz, *J* = 4.8 Hz, H_{6a}[•]), 4.14 (dd, 1H, *J* = 12.3 Hz, *J* = 6.1 Hz, H_{6a}), 4.10-4.02 (m, 3H, H₂, H_{6b}[•], H_{6b}), 4.01 (ddd, 1H, *J* = 10.0 Hz, *J* = 4.8 Hz, *J* = 2.4 Hz, H₅[•]), 3.85 (ddd, 1H, *J* = 9.8 Hz, *J* = 6.1 Hz, *J* = 2.3 Hz, H₅), 2.09-1.99 (6 s, 21H, 7 CH₃). ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 170.8, 170.7, 170.1, 170.1, 169.8, 169.7 (7C, 7 C=O), 94.2 (C₁), 92.1 (C₁[•]), 71.0 (C₃), 70.0 (2 C, C₃[•], C₂[•]), 69.8 (C₅), 68.6 (2C, C₅[•], C₄[•]), 66.0, (C₄), 62.3, 62.0 (2C, C₆, C₆[•]), 61.2 (C₂), 20.8, 20.8, 20.7 (7C, 7 CH₃). HRMS (ESI⁺): calcd for C₂₆H₃₅N₃NaO₁₇ [M+Na]⁺ 684.1859, found 684.1852. [α]_D²⁰: + 149 (*c* 1.0, CHCl₃).

2-azido-2-deoxy-2-epi- α,α -D-trehalose (**I**) (2-epi-TreAz)

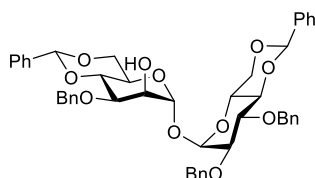


To a solution of compound **8** (46.5 mg, 0.07 mmol, 1.0 equiv) in anhydrous CH₃OH (10 mL, 0.007 M) was added anhydrous K₂CO₃ (5 mg, 0.035 mmol, 0.5 equiv). The reaction mixture was stirred at room temperature under an argon atmosphere overnight. The reaction was quenched by addition of CH₃OH (10.0 mL) and Dowex® H⁺ resin. After 10 min, the resin was filtered and the reaction mixture was concentrated under vacuum to give compound **I** (26.1 mg, 0.07 mmol, 99%) as a white powder after lyophilization.

M = 367.31 g/mol. Chemical formula: C₁₂H₂₁N₃O₁₀

R_f = 0.4 (EtOAc/CH₃OH 6:4). IR (cm⁻¹): 3353, 2936, 2116, 1567, 1407, 1267, 1145, 1078, 1051, 1033, 1001. ¹H NMR (CD₃OD, 500 MHz) δ (ppm): 5.14-5.11 (m, 2H, H₁, H_{1'}), 4.11 (dd, 1H, *J* = 9.4 Hz, *J* = 3.9 Hz, H₃), 3.89 (dd, 1H, *J* = 3.9 Hz, *J* = 1.7 Hz, H₂), 3.86-3.75 (m, 3H, H_{6a}, H_{6a'}, H₅), 3.69-3.57 (m, 5H, H_{6b}, H_{6b'}, H_{3'}, H₄, H_{5'}), 3.48 (dd, 1H, *J* = 9.8 Hz, *J* = 3.8 Hz, H_{2'}), 3.28 (at, 1H, *J* = 9.5 Hz, H_{4'}). ¹³C NMR (CD₃OD, 125 MHz) δ (ppm): 95.5, 95.0 (2C, C_{1'}, C₁), 75.1, 74.8, 74.7 (3C, C_{5'}, C₅, C_{3'}), 73.0, (C_{2'}), 72.4 (C₃), 72.0 (C_{4'}), 68.9 (C₄), 65.7 (C₂), 62.9, 62.9 (2C, C₆, C_{6'}). HRMS (ESI⁺): calcd for C₁₂H₂₁N₃NaO₁₀ [M+Na]⁺ 390.1119, found 390.1133, calcd for C₁₄H₂₄N₄NaO₁₀ [M+Na+CH₃CN]⁺ 431.1385, found 431.1395. [α]_D²⁰: + 157 (*c* 0.5, CH₃OH).

2',3,3'-tri-*O*-benzyl-4,6;4',6'-di-*O*-benzylidene-2-epi- α,α -D-trehalose (**9**)



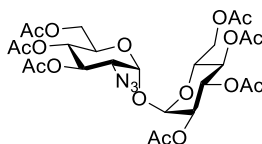
To a solution of compound **1** (500 mg, 0.63 mmol, 1.0 equiv) and anhydrous pyridine (0.5 mL, 6.1 mmol, 10.0 equiv) in anhydrous CH₂Cl₂ (10 mL, 0.03 M) at 0 °C under an argon atmosphere was added triflic anhydride (Tf₂O, 250.0 μ L, 1.4 mmol, 2.2 equiv). The reaction mixture was stirred at room temperature under an argon atmosphere for 3 hours then cooled to 0 °C. The reaction was quenched by addition of ice-cold saturated aqueous NaHCO₃. The reaction mixture was extracted three times with CH₂Cl₂, and the combined organic layer was dried over anhydrous sodium sulfate and co-evaporated with toluene three times under reduced pressure. The crude material was purified by flash chromatography over silica gel (cyclohexane/EtOAc, 7:3) to give the crude triflate **6** as a white powder. This compound was dissolved in dimethylformamide (8.0 mL, 0.07 M) at room temperature under an argon atmosphere and treated with sodium nitrite (750 mg, 10.8 mmol, 18.0 equiv). The reaction mixture was stirred overnight at 65 °C. After cooling to room temperature, the reaction mixture was diluted with EtOAc and quenched by addition of ice-cold saturated aqueous NaHCO₃. The reaction mixture was extracted three times with EtOAc, and the combined organic layer was dried over anhydrous sodium sulfate and concentrated under vacuum. The crude residue was purified by

flash chromatography over silica gel (cyclohexane/EtOAc, 8:2) to give compound **9** (231 mg, 0.29 mmol, 46%) as a white powder.

M = 788.89 g/mol Chemical formula: C₄₇H₄₈O₁₁

Rf = 0.4 (cyclohexane/EtOAc 6:4). IR (cm⁻¹): 3448, 3064, 3032, 2920, 2867, 1497, 1454, 1372, 1214, 1088, 998749, 697. ¹H NMR (CDCl₃, 500 MHz) δ (ppm): 7.55-7.26 (m, 25H, 25 H_{ar}), 5.63 (s, 1H, CH-Ph), 5.60 (s, 1H, CH-Ph), 5.19-5.16 (m, 2H, H₁, H_{1'}), 4.97 (d, 1H, J = 11.1 Hz, CH₂-Ph), 4.91 (d, 1H, J = 11.1 Hz, CH₂-Ph), 4.86 (d, 1H, J = 11.1 Hz, CH₂-Ph), 4.83 (d, 1H, J = 11.1 Hz, CH₂-Ph), 4.75 (d, 1H, J = 11.1 Hz, CH₂-Ph), 4.74 (d, 1H, J = 11.1 Hz, CH₂-Ph), 4.29 (dd, 1H, J = 10.1 Hz, J = 4.7 Hz, H_{6a'}), 4.22 (dt, 1H, J = 9.5 Hz, J = 4.8 Hz, H₅), 4.21-4.11 (m, 2H, H_{6a}, H₄), 4.07-4.01 (m, 3H, H₂, H_{3'}, H₃), 3.87-3.80 (m, 2H, H_{6b}, H_{5'}), 3.74 (at, 1H, J = 10.1 Hz, H_{6b'}), 3.68 (at, 1H, J = 9.3 Hz, H_{4'}), 3.64 (dd, 1H, J = 9.3 Hz, J = 3.8 Hz, H_{2'}), 2.72 (bs, 1H, OH). ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 138.8, 138.2, 138.0, 137.9, 137.5 (5C, 5 C_q-Ar), 129.2, 129.1, 128.8, 128.7, 128.6, 128.5, 128.4, 128.2, 128.2, 128.2, 128.1, 127.9, 126.4, 126.1 (25C, 25 CH-Ar), 101.9 (CH-Ph), 101.4 (CH-Ph), 96.1 (C₁), 94.1 (C_{1'}), 82.4 (C_{4'}), 78.9 (C₄), 78.7, 78.6 (2C, C₂, C_{3'}), 75.8 (C₃), 75.5, 74.2, 73.6 (3C, 3 CH₂-Ph), 70.3 (C₂), 69.2, 68.9 (2C, C₆, C₆), 63.9 (C₅), 63.6 (C_{5'}). HRMS (ESI⁺): calcd for C₄₇H₄₉O₁₁, calcd for C₄₇H₄₈NaO₁₁ [M+Na]⁺ 811.3089, found 811.3116. [α]_D²⁰: + 42 (c 1.0, CHCl₃).

2',3,3',4,4',6,6'-Hepta-O-acetyl-2-azido-2-deoxy-α,α-D-trehalose (**12**)



To a solution of compound **9** (318 mg, 0.40 mmol, 1.0 equiv) and anhydrous pyridine (0.5 mL, 6.18 mmol, 15.0 equiv) in anhydrous CH₂Cl₂ (2.6 mL, 0.15 M) at 0 °C, under an argon atmosphere was added triflic anhydride (Tf₂O, 148 μL, 0.88 mmol, 2.2 equiv). The reaction mixture was stirred at room temperature under an argon atmosphere for 3 hours then cooled to 0 °C. The reaction was quenched by addition of ice-cold saturated aqueous NaHCO₃. The reaction mixture was extracted three times with CH₂Cl₂, and the combined organic layer was dried over anhydrous sodium sulfate and co-evaporated with toluene three times under reduced pressure. The crude material was purified by flash chromatography over silica gel (cyclohexane/EtOAc, 7:3) to give the crude triflate **10** as a white powder. This compound was dissolved in toluene (10.0 mL, 0.04 M) at room temperature under an argon atmosphere and treated with tetrabutylammonium azide (455 mg, 1.6 mmol, 4.0 equiv). The reaction mixture was stirred overnight at 50 °C. After cooling to room temperature, the reaction mixture was diluted with EtOAc and neutralized by addition of ice-cold saturated aqueous NaHCO₃. The reaction mixture was extracted three times with EtOAc, and the combined organic layer was dried over anhydrous sodium sulfate and concentrated under vacuum. The crude residue was purified by flash chromatography over silica gel (heptane/EtOAc, 8:2) to give a mixture of compound **11** with the by-product of dehydration with a ratio of 2:1 (215 mg).

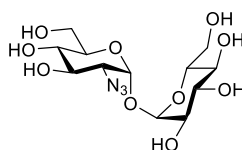
A dry 100 mL round bottom flask was outfitted with a stir bar and a rubber septum. The assembly was put in a drier and cooled under vacuum. The flask was charged with anhydrous

iron (III) chloride (834 mg, 5.1 mmol, 21.0 equiv), three cycles of vacuum – argon were done. Iron (III) chloride was suspended in dry CH₂Cl₂ (30.0 mL, 0.17 M) and sonicate during 10 min. A solution of compound **11** with the by-product of dehydration with a ratio of 2:1 (200 mg) in anhydrous CH₂Cl₂ (18 mL, 0.014 M) was added. The suspension was stirred at 0 °C and anhydride acetic (1 mL, 1.09 g, 8.5 mmol) was added. The reaction mixture was stirred under an argon atmosphere for 48 hours. Then, the reaction was quenched by addition of ice-cold saturated aqueous NaHCO₃ and extracted three times with CH₂Cl₂, and the combined organic layer was dried over anhydrous sodium sulfate and concentrated under vacuum. The crude product diluted in CH₃OH (≈ 15 mL) was passed rapidly through a short pad of resin (Dowex 50WX8 hydrogen form), concentrated and then purified by flash chromatography on silica gel (heptane/EtOAc, 90:10 to 60:40) to give compound **12** (66.2 mg, 0.1 mmol, 25%) as a white powder after lyophilisation.

M = 661.57 g/mol. Chemical formula: C₂₆H₃₅N₃O₁₇

Rf = 0.3 (heptane/EtOAc, 7:3). IR (cm⁻¹): 2954, 2923, 2113, 1749, 1434, 1369, 1221, 1141, 1038, 1017. ¹H NMR (CDCl₃, 500 MHz) δ (ppm): 5.45 (at, 1H, *J* = 10.0 Hz, H_{3'}), 5.43 (at, 1H, *J* = 10.0 Hz, H₃), 5.27 (d, 1H, *J* = 4.0 Hz, H_{1'}), 5.17 (d, 1H, *J* = 4.0 Hz, H₁), 5.09 (at, 1H, *J* = 10.0 Hz, H_{4'}), 5.00 (dd, 1H, *J* = 10.0 Hz, *J* = 4.0 Hz, H_{2'}), 4.99 (at, 1H, *J* = 10.0 Hz, H₄), 4.27 (ddd, 1H, *J* = 10.0 Hz, *J* = 4.0 Hz, *J* = 1.8 Hz, H_{5'}), 4.25-4.19 (m, 2H, H_{6a'}, H_{6a}), 4.10 (dd, 1H, *J* = 12.2 Hz, *J* = 1.8 Hz, H_{6b'}), 4.00 (ddd, 1H, *J* = 10.2 Hz, *J* = 5.5 Hz, *J* = 2.1 Hz, H₅), 3.98 (dd, 1H, *J* = 12.2 Hz, *J* = 2.1 Hz, H_{6b}), 3.71 (dd, 1H, *J* = 10.4 Hz, *J* = 3.8 Hz, H₂), 2.08, 2.07, 2.06, 2.05, 2.04, 2.03, 2.00 (7s, 21H, 7 CH₃). ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 170.8, 170.7, 170.0, 170.0, 169.9, 169.9, 169.9 (7C, 7 C=O), 93.6 (C₁), 92.6 (C_{1'}), 71.5 (C₃), 70.2, 70.2 (2C, C₂, C_{3'}), 68.8, 68.7, 68.5, 68.4 (4C, C₅, C_{5'}, C₄, C_{4'}), 62.0, 61.9 (2C, C₆, C_{6'}), 61.3 (C₂), 20.9, 20.9, 20.8, 20.8 (7C, 7 CH₃). HRMS (ESI⁺): calcd for C₂₆H₃₅N₃NaO₁₇ [M+Na]⁺ 684.1859, found 684.1891. [α]_D²⁰: +75 (*c* 0.7, CHCl₃).

2-Azido-2-deoxy-α,α-D-trehalose (**II**) (2-TreAz)



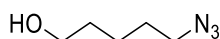
To a solution of compound **12** (69.1 mg, 0.10 mmol, 1.0 equiv) in anhydrous CH₃OH (10 mL, 0.01 M) was added anhydrous K₂CO₃ (10 mg, 0.072 mmol, 0.5 equiv). The reaction mixture was stirred overnight at room temperature under an argon atmosphere. The reaction was quenched by addition of CH₃OH (10.0 mL) and Dowex® H⁺ resin. After 10 min, the resin was filtered and the reaction mixture was concentrated under vacuum to give compound **II** (37.2 mg, 0.10 mmol, 99%) as a white powder after lyophilisation. NMR data were in agreement with those reported.^{iv}

M = 367.31 g/mol. Chemical formula: C₁₂H₂₁N₃O₁₀

Rf = 0.4 (EtOAc/CH₃OH, 6:4). IR (cm⁻¹): 3352, 2929, 2111, 1643, 1568, 1406, 1317, 1259, 1146, 1050, 995. ¹H NMR (CD₃OD, 500 MHz) δ (ppm): 5.21 (d, 1H, *J* = 3.6 Hz, H₁), 5.12 (d, 1H, *J* = 3.8 Hz, H_{1'}), 3.99 (dd, 1H, *J* = 10.0 Hz, *J* = 9.0 Hz, H₃), 3.85 (ddd, 1H, *J* = 10.0 Hz, *J*

= 5.2 Hz, $J = 2.2$ Hz, H₅), 3.84-3.78 (m, 3H, H₅[′], H_{6a}[′], H_{6a}), 3.76 (at, 1H $J = 9.5$ Hz, H₃[′]), 3.74-3.68 (m, 2H, H_{6b}[′], H_{6b}), 3.49 (dd, 1H, $J = 9.5$ Hz, $J = 3.8$ Hz, H₂[′]), 3.40 (dd, 1H, $J = 10.0$ Hz, $J = 9.0$ Hz, H₄), 3.36 (at, 1H, $J = 9.5$ Hz, H₄[′]), 3.27 (dd, 1H, $J = 10.0$ Hz, $J = 3.6$ Hz, H₂). ¹³C NMR (CD₃OD, 125MHz) δ (ppm): 95.3 (C₁[′]), 94.2 (C₁), 74.8 (C₃[′]), 74.5, 74.2 (2C, C₅[′], C₅), 73.2 (C₂[′]), 72.8 (C₃), 72.2 (C₄), 71.7 (C₄[′]), 64.8 (C₂), 62.7, 62.5 (2C, C₆, C₆[′]). HRMS (ESI⁺): calcd for C₁₂H₂₁N₃NaO₁₀ [M+Na]⁺ 390.1119, found 390.1133. $[\alpha]_D^{20}$: + 91 (c 0.7, CH₃OH).

5-Azidopentan-1-ol (S1)^v

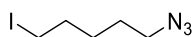


To a solution of commercially available 5-bromopentan-1-ol (725 μ L, 5.99 mmol) in H₂O (17 mL), were added NaN₃ (778 mg, 11.96 mmol, 2 equiv) and the reaction mixture was stirred at reflux for 18 hours. After cooling to room temperature, the reaction mixture was extracted three times with CH₂Cl₂, the combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash silica gel chromatography (diethyl ether/pentane, 40:60) to give the desired product **S1** (400 mg, 52%) as a colourless oil. NMR data were in agreement with those reported.ⁱⁱⁱ

M = 129.2 g/mol. Chemical formula: C₅H₁₁N₃O

R_f = 0.35 (cyclohexane/EtOAc, 7:3). ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 3.66 (t, 2H, $J = 6.4$ Hz), 3.29 (t, 2H, $J = 6.8$ Hz), 1.70-1.54 (m, 4H), 1.52-1.41 (m, 2H). ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 62.8, 51.5, 32.3, 28.8, 23.2.

1-Azido-5-iodopentane (S2)

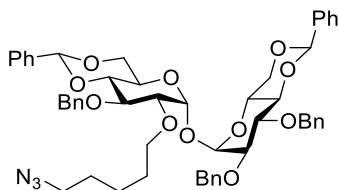


A solution of compound **S1** (320 mg, 2.48 mmol) in CH₃CN/THF (1/1, 3 mL) was cooled to 0 °C. Imidazole (202 mg, 2.97 mmol, 1.2 equiv), PPh₃ (845 mg, 3.22 mmol, 1.3 equiv) and iodine (880 mg, 3.47 mmol, 1.4 equiv) were then added and the reaction mixture was stirred at room temperature for 30 min. A saturated aqueous solution of Na₂S₂O₃ was added and the reaction mixture was extracted three times with EtOAc. The combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash silica gel chromatography (cyclohexane/EtOAc, 95:5) to give the desired product **S2** (539 mg, 91%) as a colourless oil.

M = 239.1 g/mol. Chemical formula: C₅H₁₀IN₃

R_f = 0.85 (cyclohexane/EtOAc, 7:3). ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 3.29 (t, 2H, $J = 6.7$ Hz), 3.19 (t, 2H, $J = 6.8$ Hz), 1.86 (quint, 2H, $J = 6.8$ Hz), 1.62 (quint, 2H, $J = 6.8$ Hz), 1.51 (m, 2H). ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 51.3, 33.1, 28.0, 27.8, 6.4.

2-O-5-Azidopentyl-2',3,3'-tri-O-benzyl-4,6;4',6'-di-O-benzylidene- α,α -D-trehalose (**13**)

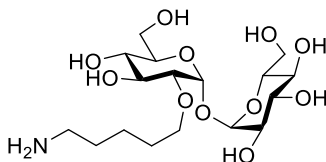


To a solution of compound **1** (400 mg, 0.507 mmol) in dry DMF (2 mL) at 0 °C was added NaH (45.6 mg, 1.52 mmol, 3 equiv). Then, a solution of compound **S2** (364 mg, 1.52 mmol, 3 equiv) in DMF (1.2 mL) was added at 0 °C and the reaction mixture was stirred at room temperature for 18 hours. The reaction was diluted with CH₃OH (2 mL), and a saturated aqueous solution of NaCl and Et₂O was added. The reaction mixture was extracted three times with Et₂O. The combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash silica gel chromatography (cyclohexane/EtOAc, 95:5 to 80:20) to give the desired product **13** (395.7 mg, 87%) as a colourless oil.

M = 900.0 g/mol. Chemical formula: C₅₂H₅₇N₃O₁₁

Rf = 0.57 (cyclohexane/EtOAc, 7:3). ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 7.56-7.24 (m, 25H, H_{Ar}), 5.57 (s, 1H, CH-Ph), 5.56 (s, 1H, CH-Ph), 5.18 (d, 1H, *J* = 3.7 Hz, H₁), 5.12 (d, 1H, *J* = 3.7 Hz, H_{1'}), 4.97 (d, 1H, *J* = 11.4 Hz, CH₂-Ph), 4.93 (d, 1H, *J* = 11.4 Hz, CH₂-Ph), 4.87 (d, 1H, *J* = 11.4 Hz, CH₂-Ph), 4.84 (d, 1H, *J* = 12.0 Hz, CH₂-Ph), 4.81 (d, 1H, *J* = 11.4 Hz, CH₂-Ph), 4.73 (d, 1H, *J* = 12.0 Hz, CH₂-Ph), 4.31-4.11 (m, 4H, H₅, H_{5'}, H_{6a}, H_{6a'}), 4.13 (at, 1H, *J* = 9.2 Hz, H_{3'}), 4.04 (at, 1H, *J* = 9.2 Hz, H₃), 3.73-3.58 (m, 7H, CH₂-O, H_{2'}, H₄, H_{4'}, H_{6b}, H_{6b'}), 3.47 (dd, 1H, *J* = 9.2 Hz, *J* = 3.7 Hz, H₂), 3.10 (t, 2H, *J* = 6.8 Hz, CH₂-N₃), 1.66-1.38 (m, 6H, 3 CH₂). ¹³C NMR (CD₃OD, 75 MHz) δ (ppm): 139.0, 138.9, 138.2, 137.7, 137.6 (5C, C_q-Ar), 129.1, 128.7, 128.5, 128.4, 128.3, 128.1, 127.9, 127.7, 126.3, 126.1 (25C, CH-Ar), 101.4, 101.3 (2C, CH-Ph), 95.0 (C_{1'}), 94.6 (C₁), 82.5 (2C, C₄, C_{4'}), 80.1 (C₂), 78.9 (C_{2'}), 78.8 (C_{3'}), 78.7 (C₃), 75.5, 75.4, 74.0 (3C, CH₂-Ph), 72.0 (CH₂-O), 69.3, 69.2 (2C, C₆, C_{6'}), 63.1, 63.0 (2C, C₅, C_{5'}), 51.3 (CH₂-N₃), 29.9, 28.9, 23.6 (3C, 3 CH₂). HRMS (ESI): calcd for C₅₂H₅₇NaN₃O₁₁ [M+Na]⁺ 922.3885, found 922.3868. [α]_D²⁰: + 52 (*c* 1.1, CHCl₃).

2-O-(5-Aminopentyl)- α,α -D-trehalose (**14**)



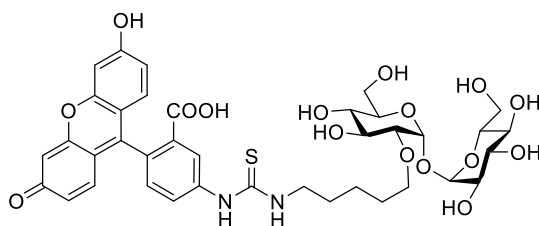
To a solution of compound **13** (67 mg, 0.07 mmol) in dioxane/H₂O (2:1, 3 mL) were added 70 mg of Pd(OH)₂. The reaction mixture was purged with dihydrogen and vigorously stirred at 40 °C for 3h30 under 1 atm of dihydrogen. The reaction mixture was filtered, washed three times with CH₃OH and concentrated under reduced pressure to give derivative **14** (31.8 mg, quantitative) as a syrup, which was engaged in the next step without further purification.

M = 427.4 g/mol. Chemical formula C₁₇H₃₃NO₁₁

Rf = 0.1 (EtOAc/CH₃OH/H₂O, 3:3:1). ¹H NMR (CD₃OD, 300 MHz) δ (ppm): 5.30 (d, 1H, *J* = 3.5 Hz, H₁), 5.11 (d, 1H, *J* = 3.8 Hz, H_{1'}), 3.89-3.64 (m, 9H, H₃, H_{3'}, H₅, H_{5'}, H_{6a}, H_{6a'}, H_{6b},

H_{6b'}, 1H of CH₂-O), 3.59 (m, 1H, 1H of CH₂-O), 3.48 (dd, 1H, *J* = 9.8 Hz, *J* = 3.8 Hz, H_{2'}), 3.37-3.30 (m, 2H, H₄, H_{4'}), 3.23 (dd, 1H, *J* = 9.7 Hz, *J* = 3.5 Hz, H₂), 2.94 (t, 2H, *J* = 7.5 Hz, CH₂-N), 1.75-1.58 (m, 4H, 2 CH₂), 1.50 (m, 2H, CH₂). ¹³C NMR (CD₃OD, 90 MHz) δ (ppm): 95.4 (C_{1'}), 92.9 (C₁), 81.3 (C₂), 74.5, 73.9, 73.8, 73.6 (4C, C₃, C_{3'}, C₅, C_{5'}), 73.1 (C_{2'}), 71.9, 71.8, (2C, C₄, C_{4'}), 71.7 (CH₂-O) 62.8, 62.5 (2C, C₆, C_{6'}), 40.7 (CH₂-N), 30.4, 28.3, 24.1 (3C, 3 CH₂). HRMS (ESI): calcd for C₁₇H₃₄NO₁₁ [M+H]⁺ 428.2126, found 428.2112. [α]_D²⁰: + 144 (*c* 1.05, CH₃OH).

2-O-(5-[(Fluorescein-5-yl)thioureido]pentyl)-α,α-D-trehalose (III) (2-FIC5Tre)



To a solution of compound **14** (10.0 mg, 0.023 mmol) in 400 μL of NaHCO₃ buffer (75 mM, pH = 9) and CH₃CN (200 μL) was added fluorescein isothiocyanate (11.0 mg, 0.028 mmol, 1.2 equiv). The reaction mixture was stirred at 60 °C for 4 hours. The crude mixture was purified by HPLC with a C18 column (10% CH₃CN in H₂O to 100% CH₃CN, 2 mL/min) to give derivative **III** (12.6 mg, 66%) as an orange solid.

M = 816.8 g/mol. Chemical formula C₃₈H₄₄N₂O₁₆S

R_f = 0.9 (EtOAc/iPrOH/H₂O, 2:2:1). ¹H NMR (D₂O, 400 MHz) δ (ppm): 7.69 (m, 1H, H_{ar}), 7.52 (m, 1H, H_{ar}), 7.31-7.17 (m, 3H, H_{ar}), 6.70-6.60 (m, 4H, H_{ar}), 5.38 (m, 1H, H₁), 5.18 (d, 1H, *J* = 3.8 Hz, H_{1'}), 3.88-3.80 (m, 6H, H₃, H_{3'}, H₅, H_{5'}, H_{6a}, H_{6b}), 3.79-3.70 (m, 3H, H_{6a'}, H_{6b'}, 1H of CH₂-O), 3.68-3.56 (m, 3H, 1H of CH₂-O, CH₂-N), 3.63 (dd, 1H, H_{2'}, *J* = 10.0 Hz, *J* = 3.8 Hz), 3.51-3.39 (m, 3H, H₂, H₄, H_{4'}), 1.79-1.58 (m, 4H, 2 CH₂), 1.42 (m, 2H, CH₂). ¹³C NMR (D₂O, 100 MHz) δ (ppm): 180.8, 179.5, 159.9 (C_q), 133.2, 127.2, 126.2, 123.8 (CH_{ar}), 115.3 (C_q), 105.2 (CH_{ar}) 95.3 (C_{1'}), 92.9 (C₁), 80.8 (C₂), 74.2, 74.1, 73.7, 73.5 (4C, C₃, C_{3'}, C₅, C_{5'}), 73.2 (CH₂-O), 72.8 (C_{2'}), 71.6, 71.2, (2C, C₄, C_{4'}), 62.3 (C₆), 62.0 (C_{6'}), 46.5 (CH₂-N), 30.5 (2C, 2 CH₂), 24.3 (CH₂). HRMS (ESI): calcd for C₃₈H₄₅N₂O₁₆S [M+H]⁺ 817.2484, found 817.2464.

2. Labeling of bacteria with trehalose-based probes

Bacterial strains, growth conditions and reagents

Dimethylsulfoxide (DMSO) was purchased from Sigma-Aldrich. Phosphate-buffered saline solution (PBS), Gene Frame seals, Microscope Slides and Microscope Coverslips were purchased from Thermo Fisher Scientific. 5/6-Carboxyrhodamine 110-PEG₄-Alkyne (CR110-CCH) was purchased from Jena Bioscience. Tris((1-(β-D-glucopyranosyl)-1H-1,2,3-triazol-4-yl)methyl)amine (TGTA) was synthesized according to a reported procedure.^{vi}

Solutions stocks of organic and inorganic compounds were prepared as followed: sodium ascorbate: 50 mM in PBS 1X; TGTA: 40 mM in PBS 1X; Copper sulfate (CuSO₄): 20 mM in H₂O; CR110-CCH: 2 mM in DMSO. Stock solutions of trehalose analogues were prepared in milliQ water. *Corynebacterium glutamicum* ATCC13032 were inoculated into BHI growth medium and cultured at 30 °C under agitation (180 rpm).

General procedure for labeling *C. glutamicum*

C. glutamicum was inoculated (1:100) in BHI medium (final volume 900 μL). Bacteria were cultured 8 h with probe **I** or **II** (250 μM); or were cultured until mid-log phase growth (4 h, OD ≈ 6) at which point probe **III** (100 μM) was added (control experiments were treated with probe vehicle i.e. water) and the growth was pursued for 4 h at 30 °C.

Then, 400 μL of these suspensions were transferred in a 1.5 mL microtube and the cells were harvested by centrifugation at 3200 rpm for 5 min. The pellets were washed three times with PBS buffer (200 μL, 3200 rpm, 5 min, rt). The pellets of bacteria cultured with probe **III** were kept at 4 °C between 2 and 4 hours in the dark before being analyzed. The pellets of bacteria cultured with probe **I** or **II** were re-suspended in 200 μL of the "click" solution (0.05 mM CR110-CCH, 2.5 mM sodium ascorbate, 2.0 mM TGTA and 1.0 mM copper sulfate pentahydrate in PBS/DMSO (98:2) buffer) or in 200 μL of PBS/DMSO (97:3) buffer, and transferred in a 2.0 mL microtube for a better agitation. This suspension was vigorously shaken (550 rpm) in the dark for 30 min at room temperature, then transferred back in a 1.5 mL microtube and the cells were harvested by centrifugation (3200 rpm, 5 min, rt). Bacterial pellets were washed one times with PBS/DMSO (97:3) buffer (200 μL, 3200 rpm, 5 min, rt) and two times with PBS buffer (200 μL, 3200 rpm, 5 min, rt). The pellets were re-suspended in PBS buffer (200 μL) and kept at 4 °C between 1 and 3 hours in the dark before being analyzed.

Flow cytometry quantification

Fluorescence of the samples (diluted 100 times in PBS buffer) were analyzed using a Cytoflex flow cytometer (Beckman-Coulter). A minimum of 20000 events were counted for each data set (10 μL/min). Data were analyzed using CytExpert software (Beckman-Coulter). Representative histograms for *C. glutamicum* experiments labeling with probes **I**, **II** or **III** at 250 μM or 100 μM, followed by reaction with alkyne functionalized carboxyrhodamine 110 (CR110-CCH) using CuAAC.

General procedures for confocal imaging of labeled bacteria

Samples of 2 μL were deposited on agar (1%) pads placed on glass slides and then covered by coverslips. Cell morphology was observed with bright field light. Fluorescence microscopy experiments were performed on a confocal straight Leica SP8 microscope (DM 6000), using a 63x PLAN APO oil immersion lens (Leica), an argon laser and a GaAsP Hybrid detector (Hamamatsu). CR110 were excited at 488 nm and collected with a 500-550 nm band pass filter. The experiments were imaged with identical exposure setting. The microscope was operated with LAS-X program. Images were visualized and montages were constructed with ImageJ using Fiji without further processing.

3. *C. glutamicum* cell growth assays

C. glutamicum strain was grown in BHI medium overnight at 30 °C. The culture was then diluted in BHI medium to a final absorbance at 600 nm of 0.9. Sample aliquots (9 μL) of this diluted cell culture and 181 μL of BHI medium were placed in wells of a microtiter plate. The probe solution (10 μL) at various concentrations was then added to the wells and the plate was incubated at 30 °C for 16 h in a microtiter plate reader (MultiskanEX system microplate reader from Labsystem). Optical density readings (600 nm) were taken at 30 min intervals. The plate was shaken for 30 s before each reading.

III. Supplementary figures (S1-S2)

Figure S1. Growth curves of *C. glutamicum* treated with 2-TreAz **II** (A), 2-epi-TreAz **I** (B) and 2-FIC5Tre **III** (C) at 7 different concentrations. Experiments were performed in duplicate and data represent an average of two independent assays with error bars indicating standard deviations.

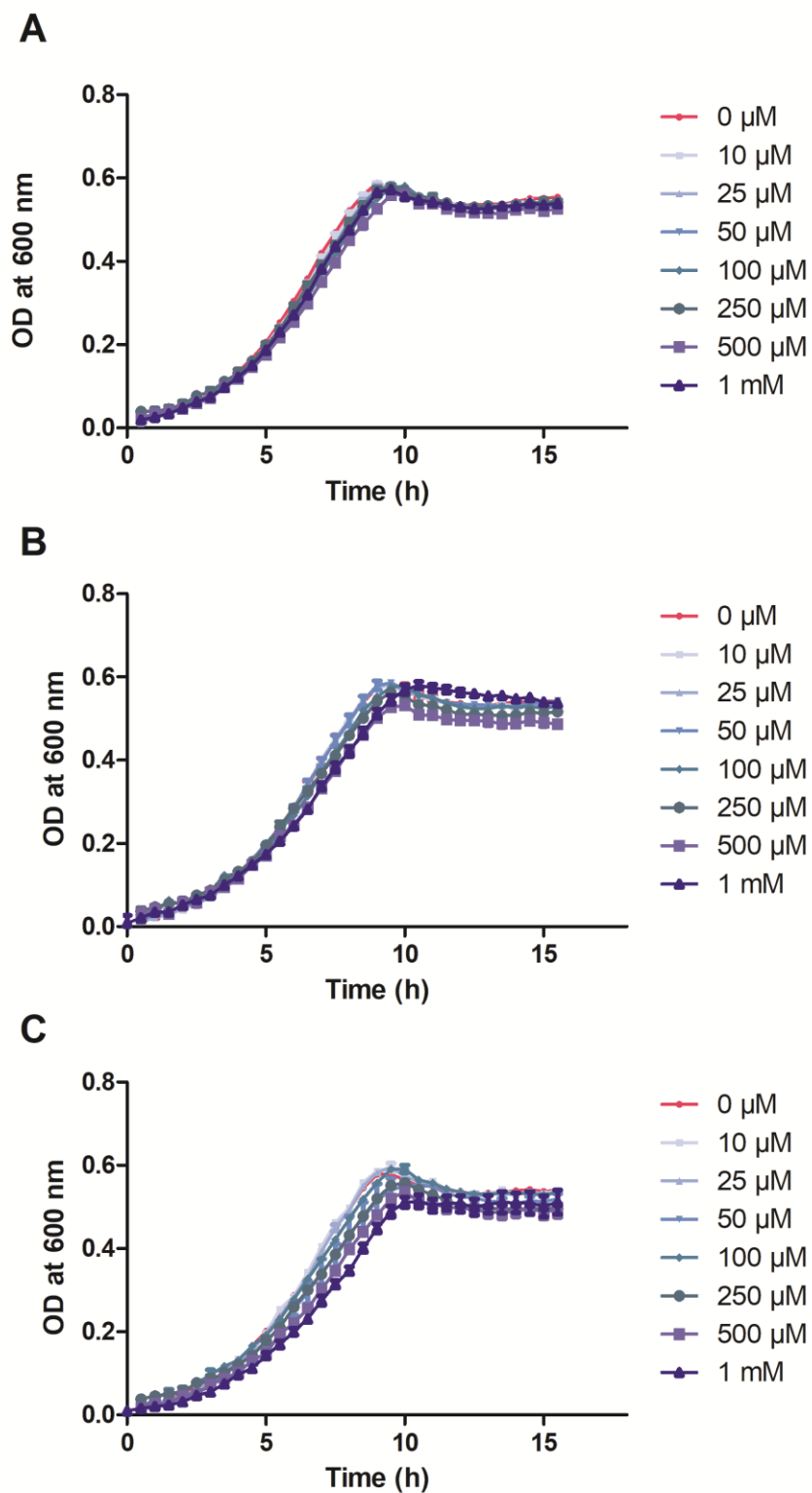
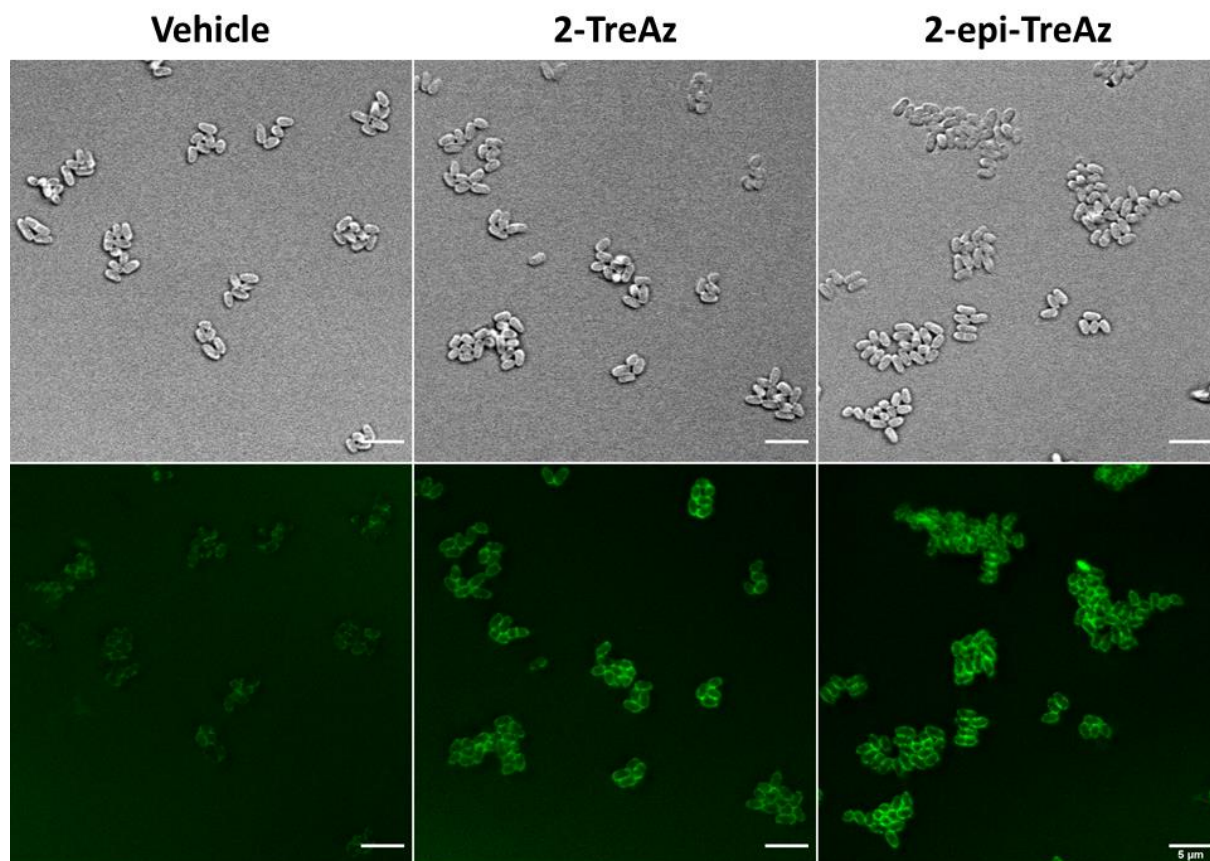
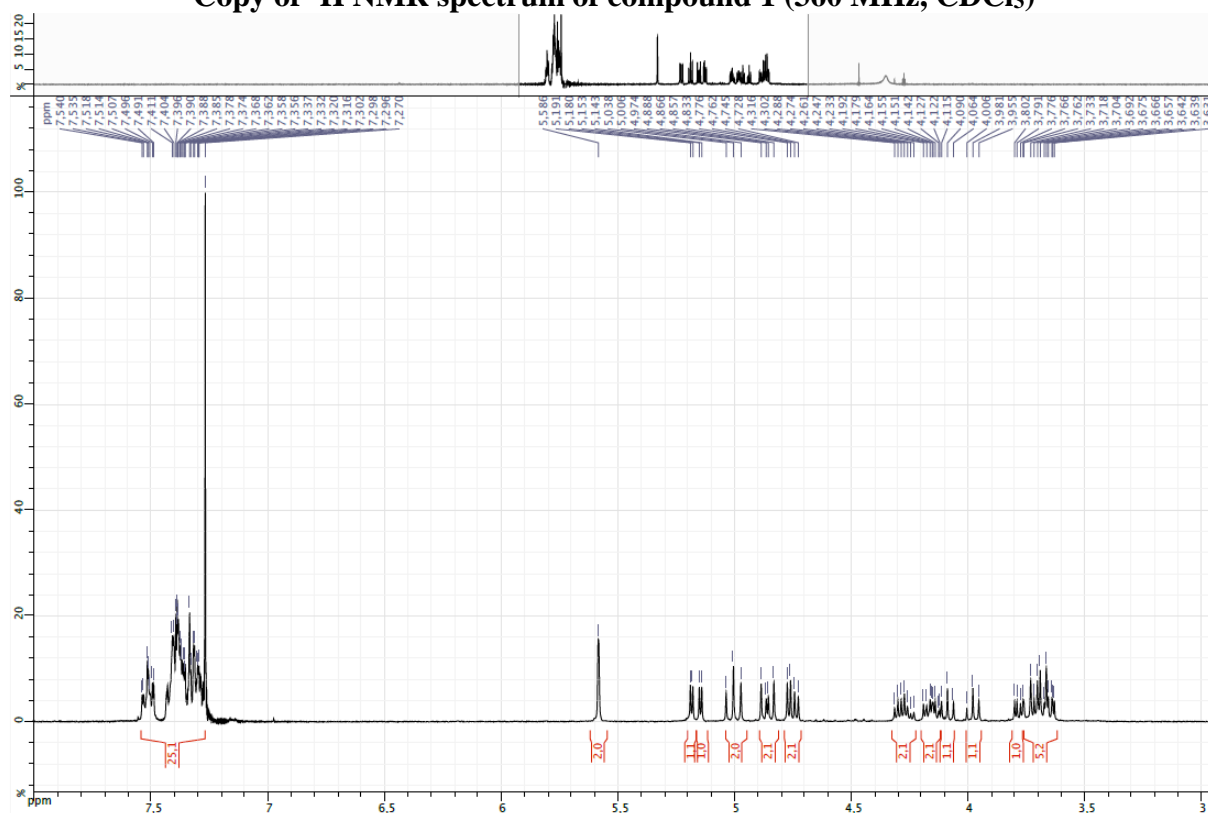


Figure S2. *C. glutamicum* incubated with with 250 μ M of 2-TreAz **II** or 2-epi-TreAz **I** (or untreated) for 8 h, then reacted with CR110-CCH and analyzed by fluorescence microscopy with bright field images on top. Scale bars = 5 μ m.

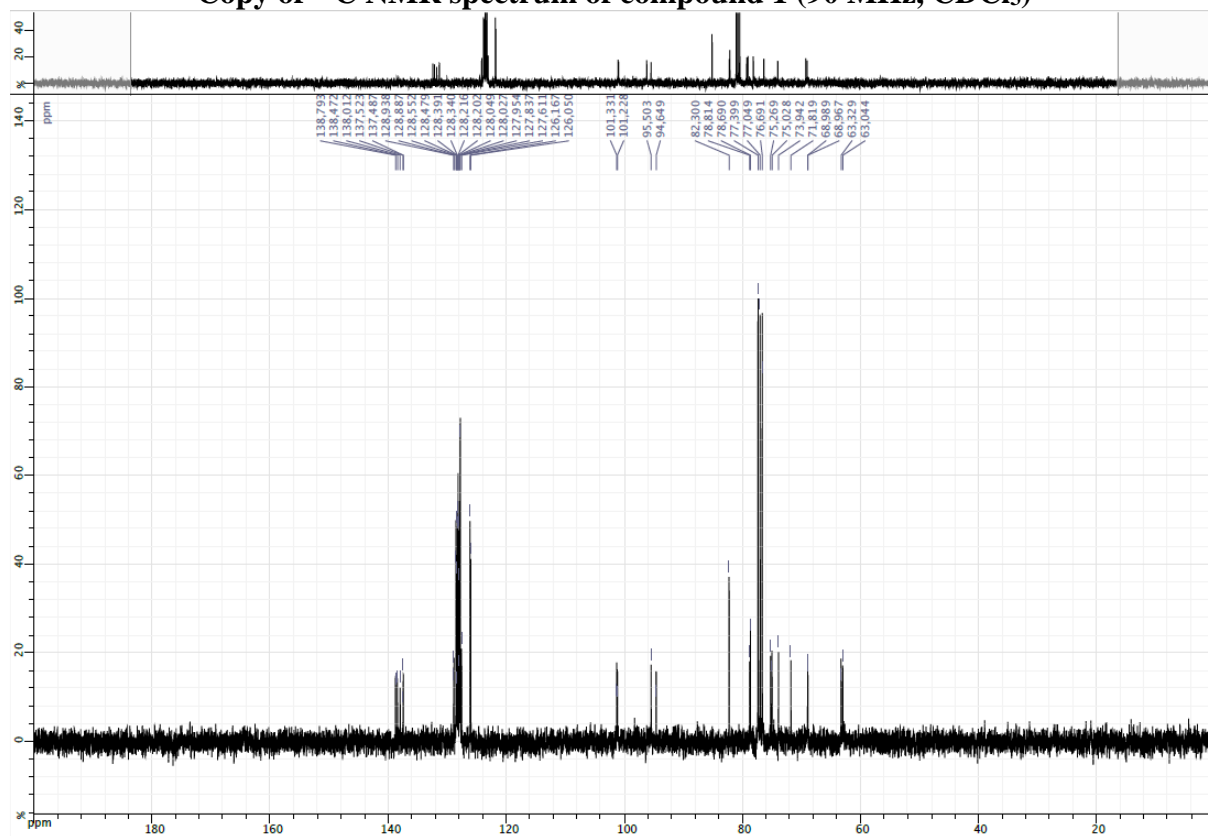


IV. NMR spectra

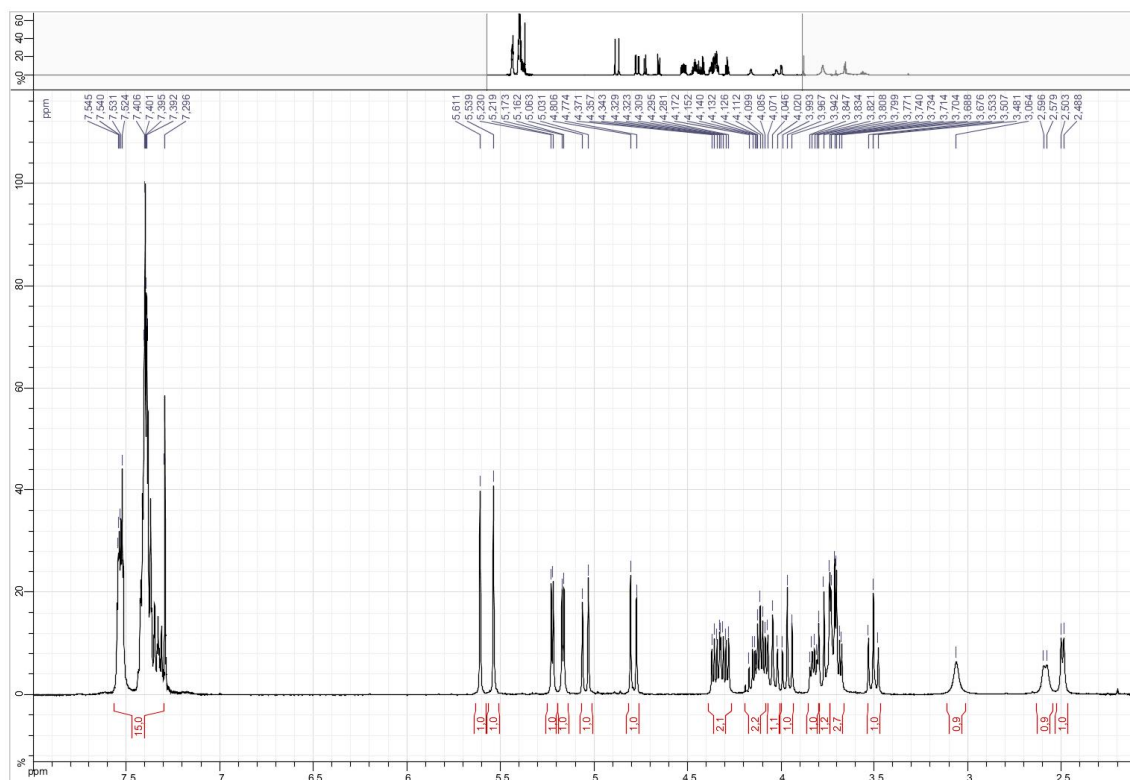
Copy of ^1H NMR spectrum of compound 1 (360 MHz, CDCl_3)



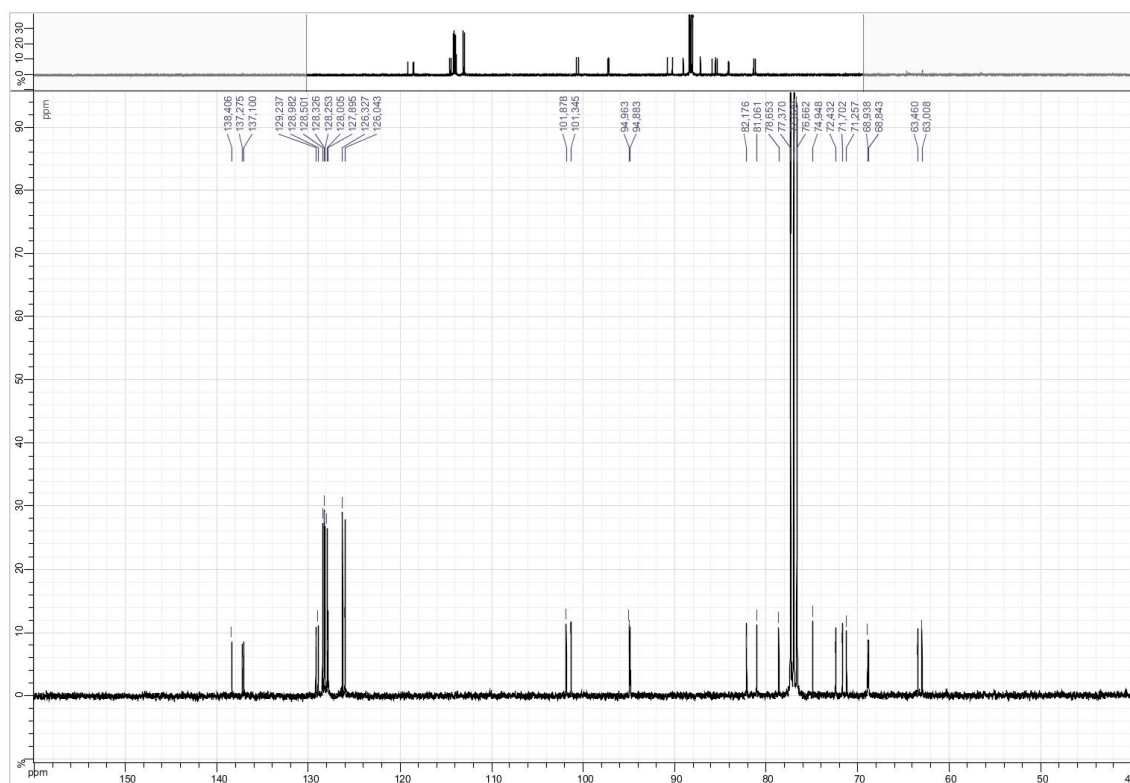
Copy of ^{13}C NMR spectrum of compound 1 (90 MHz, CDCl_3)



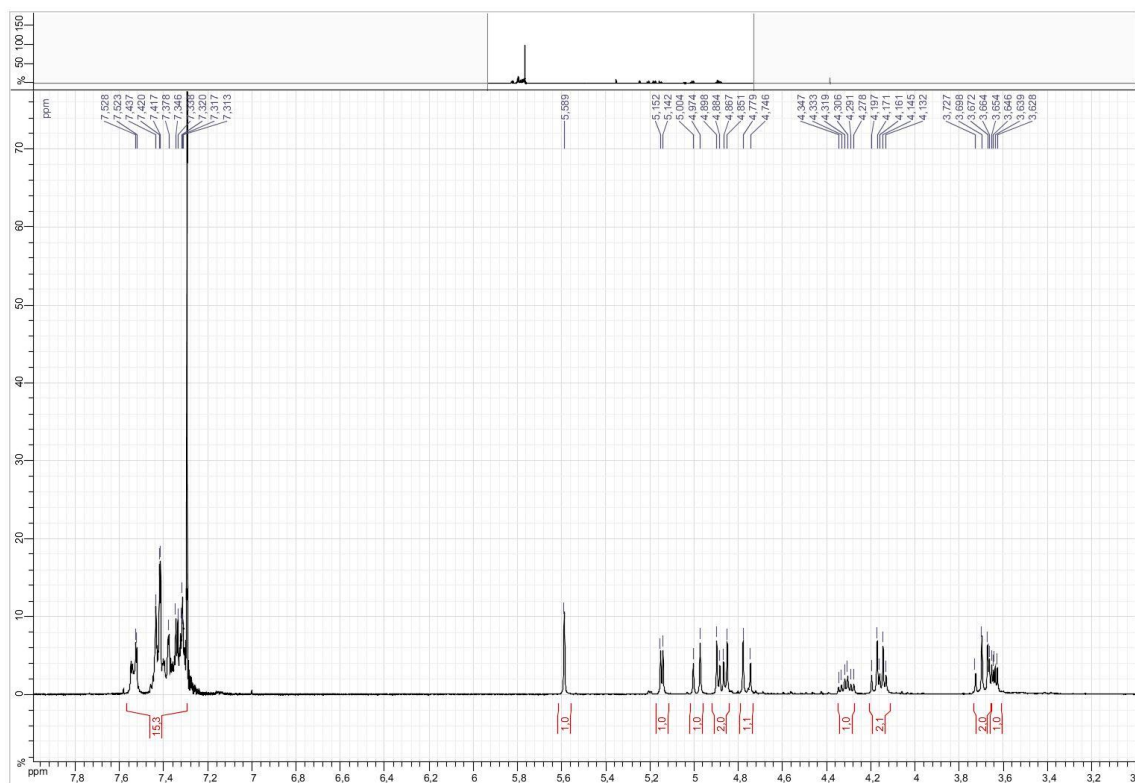
Copy of ^1H NMR spectrum of compound 4 (360 MHz, CDCl_3)



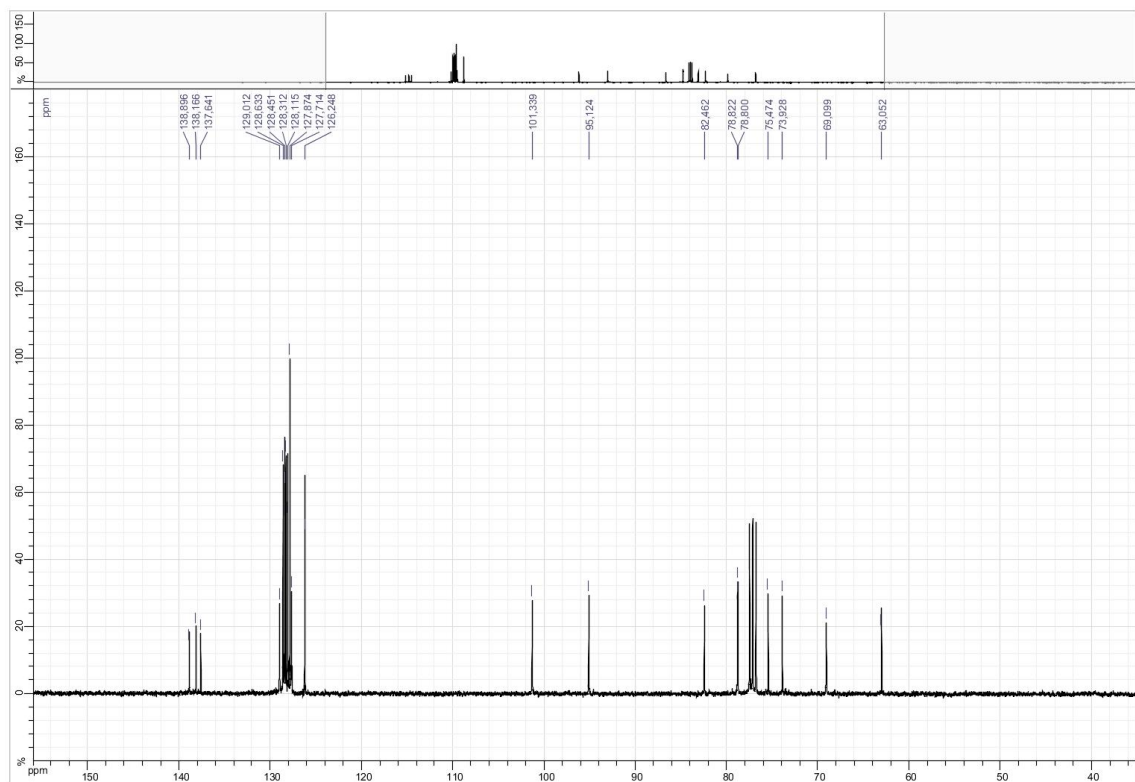
Copy of ^{13}C NMR spectrum of compound 4 (90 MHz, CDCl_3)



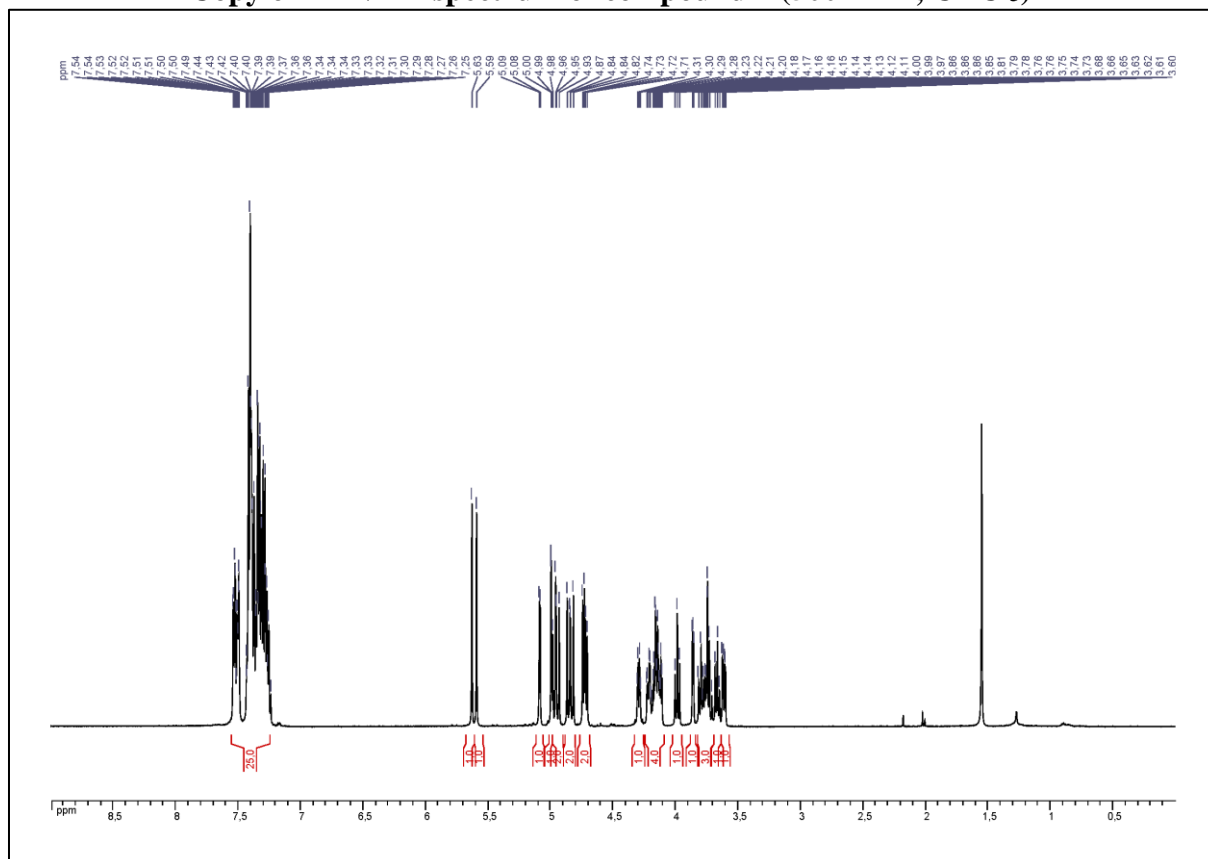
Copy of ^1H NMR spectrum of compound 5 (360 MHz, CDCl_3)



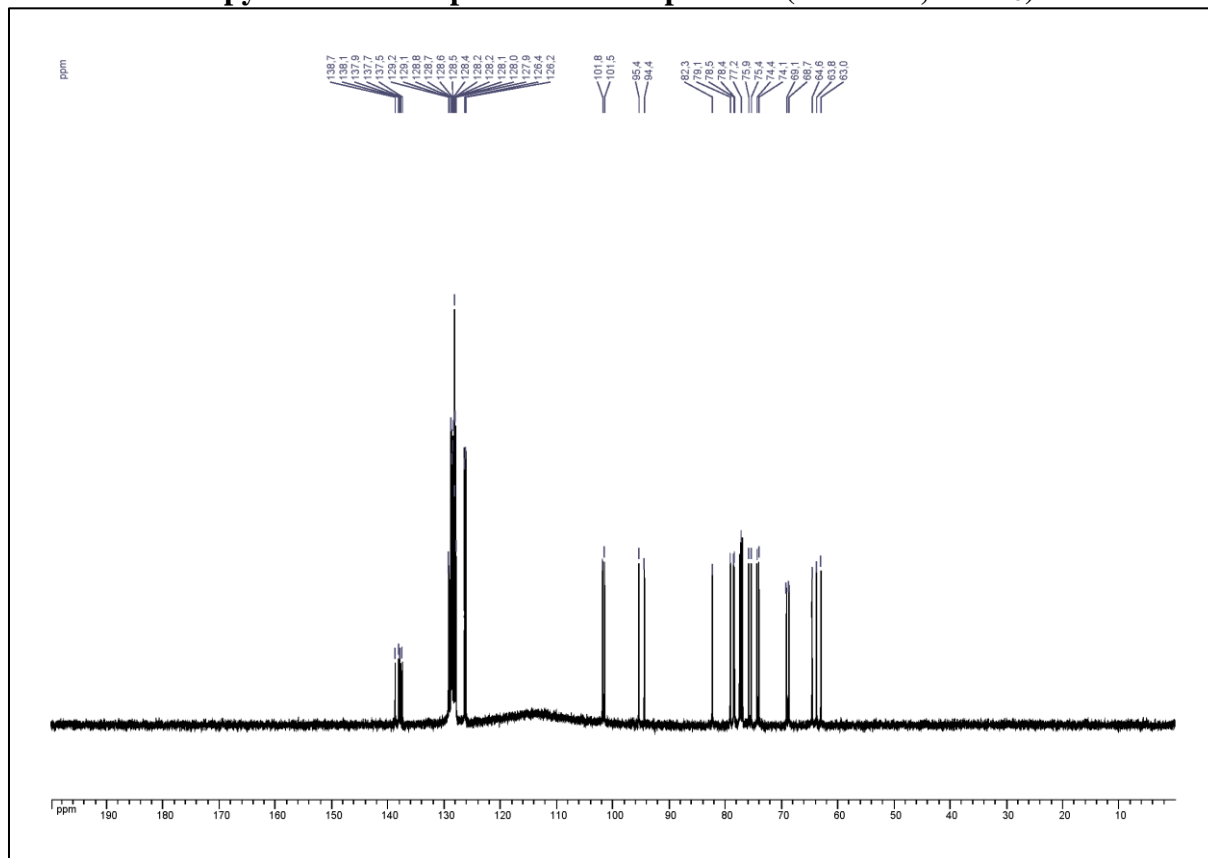
Copy of ^{13}C NMR spectrum of compound 5 (90 MHz, CDCl_3)



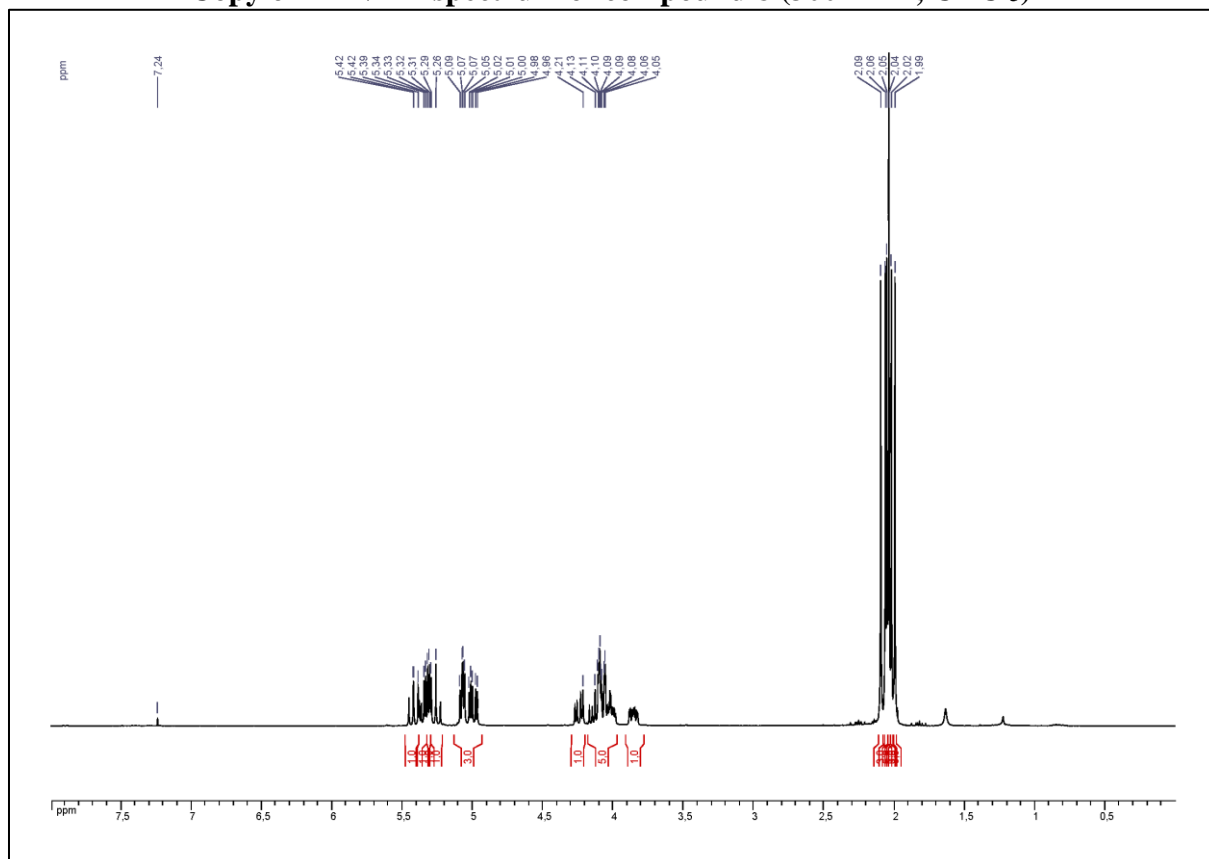
Copy of ^1H NMR spectrum of compound 7 (500 MHz, CDCl_3)



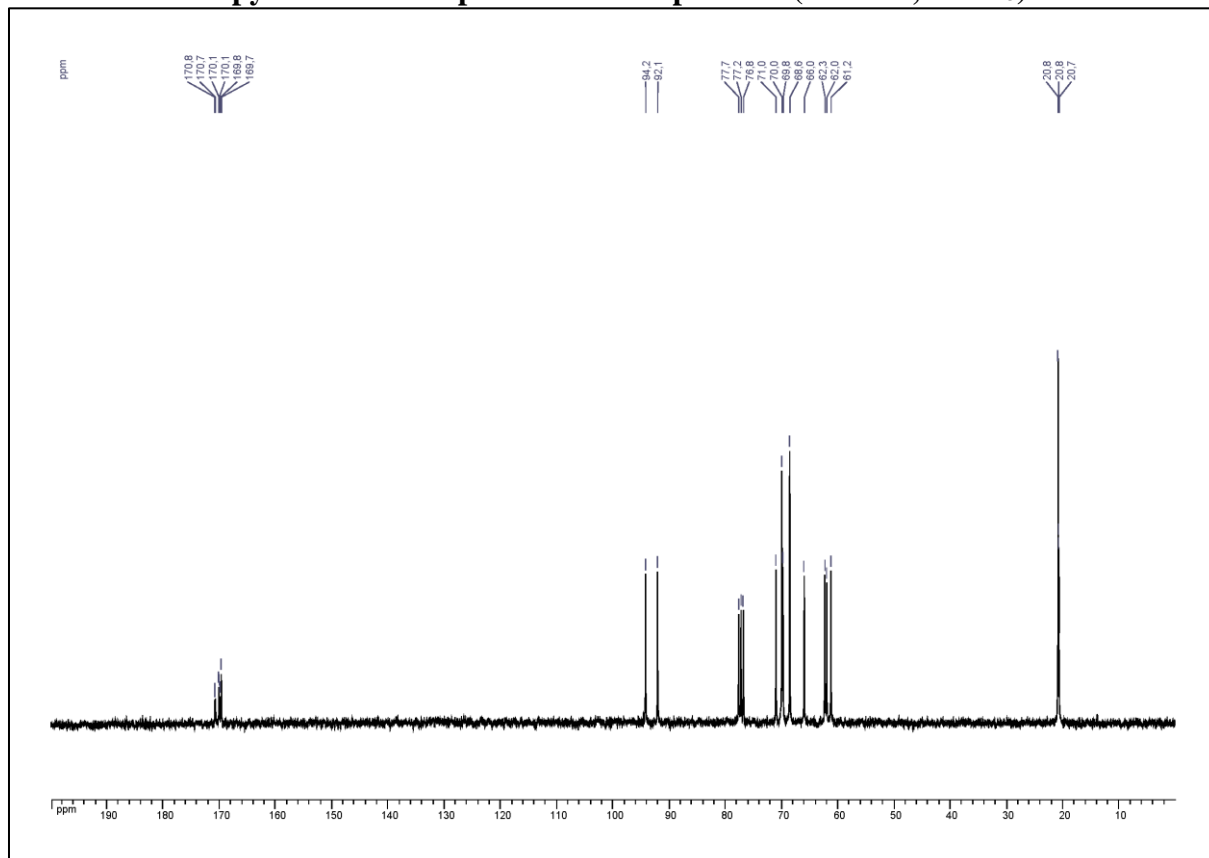
Copy of ^{13}C NMR spectrum of compound 7 (125 MHz, CDCl_3)



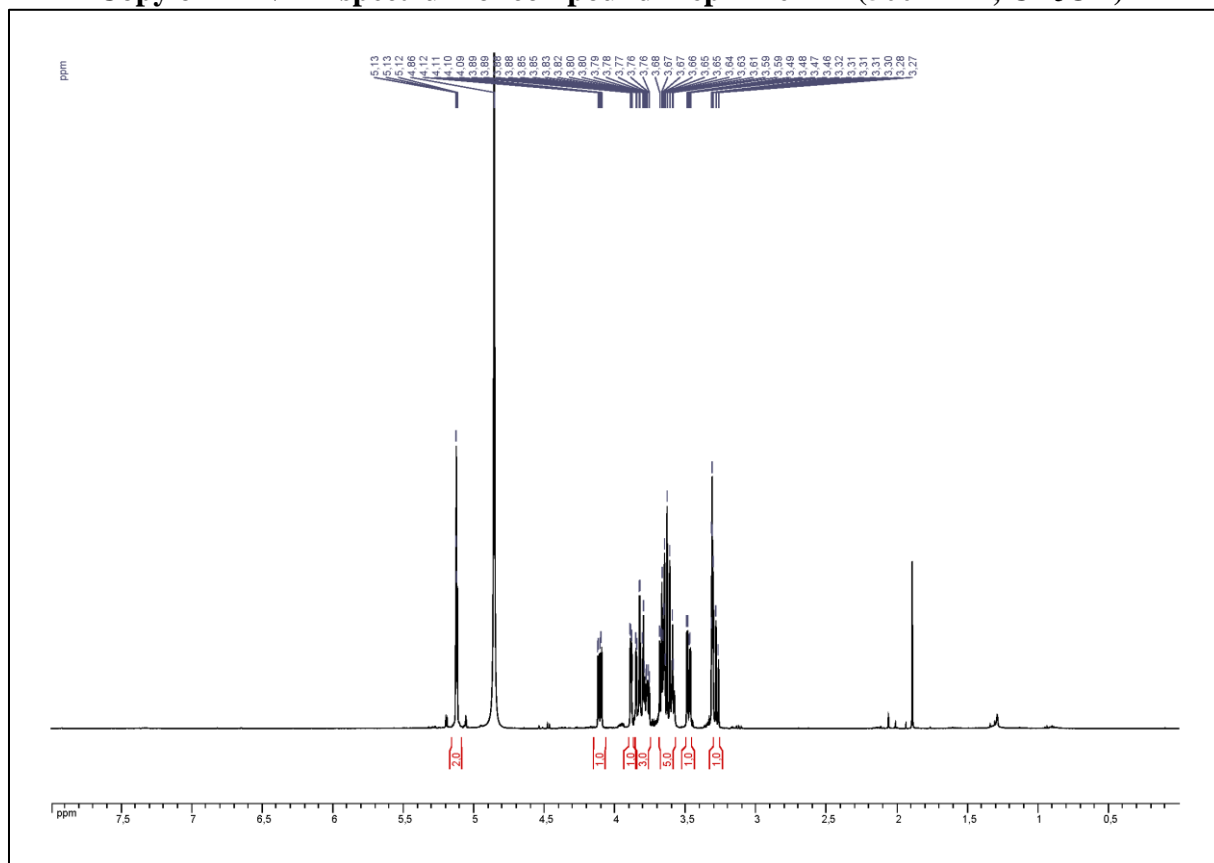
Copy of ^1H NMR spectrum of compound 8 (300 MHz, CDCl_3)



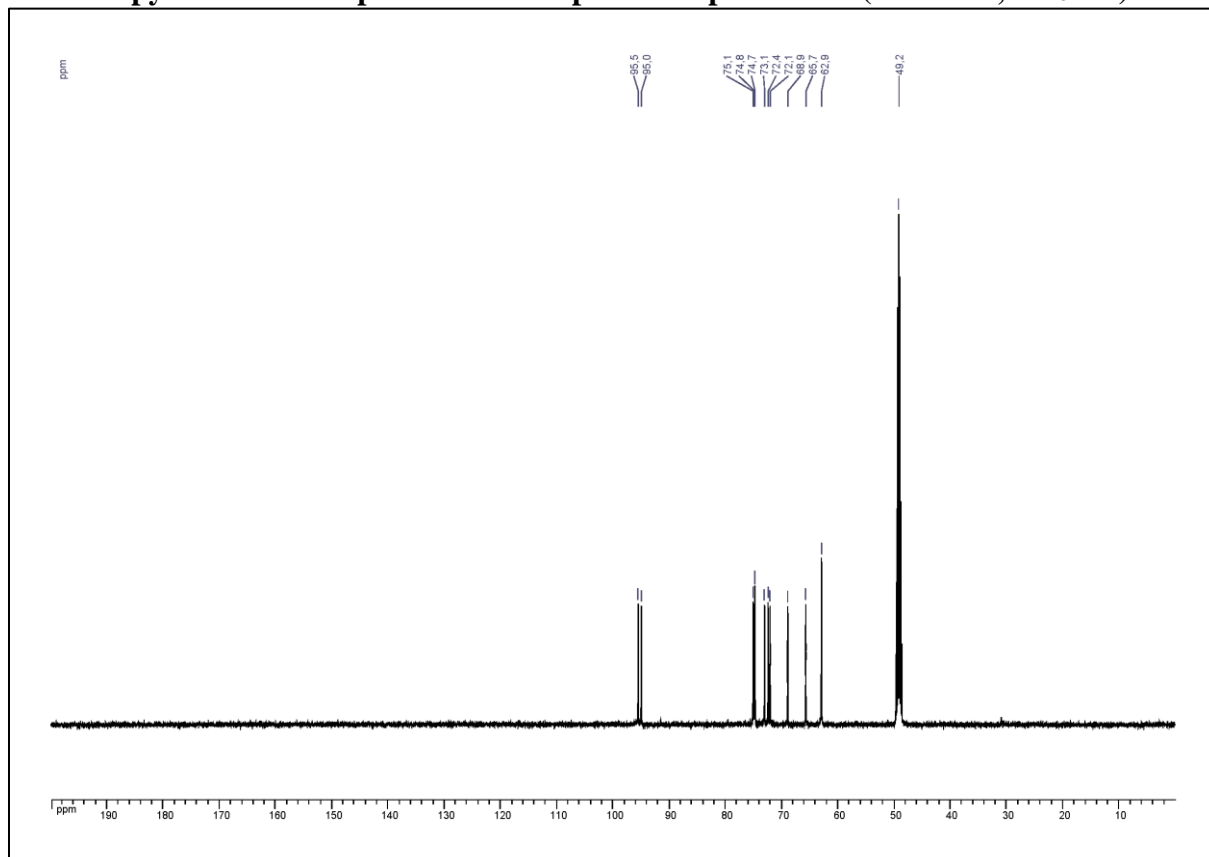
Copy of ^{13}C NMR spectrum of compound 8 (75 MHz, CDCl_3)



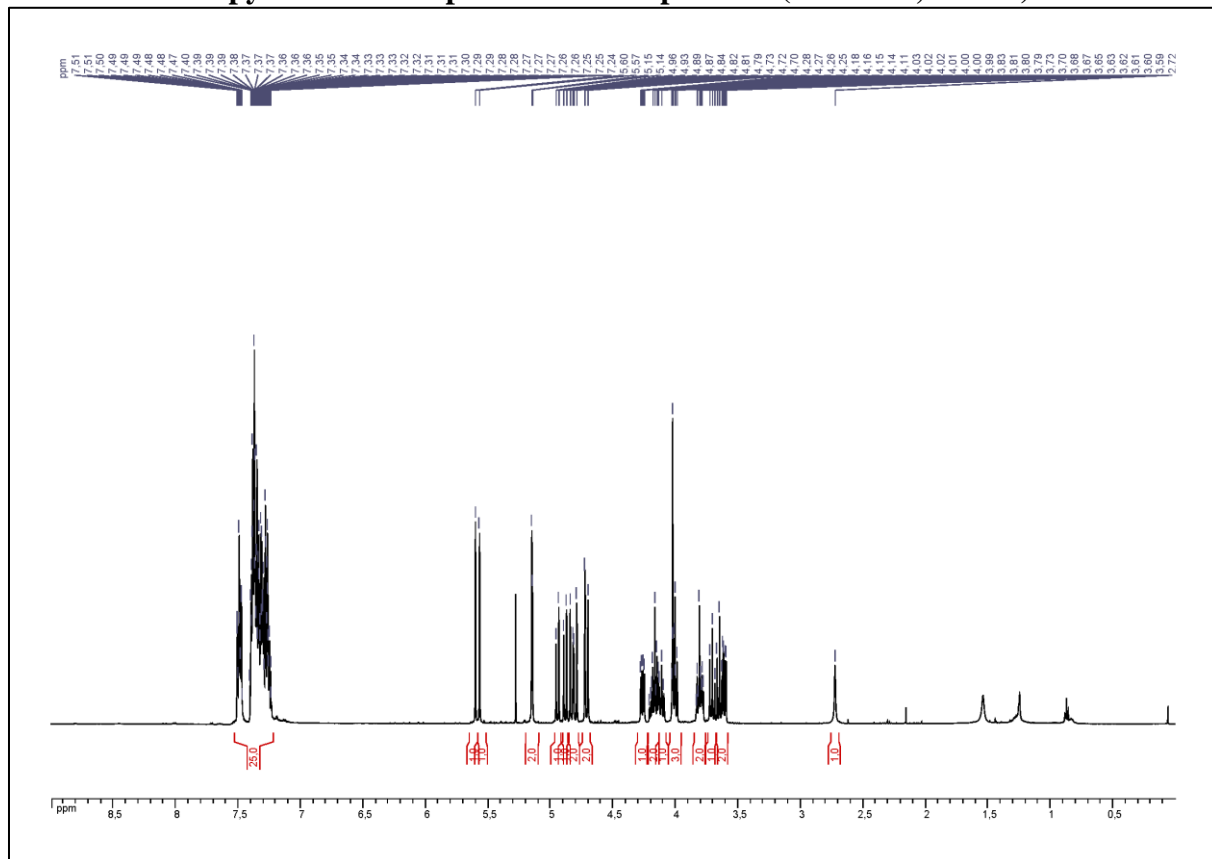
Copy of ^1H NMR spectrum of compound 2-*epi*-TreAz I (500 MHz, CD_3OD)



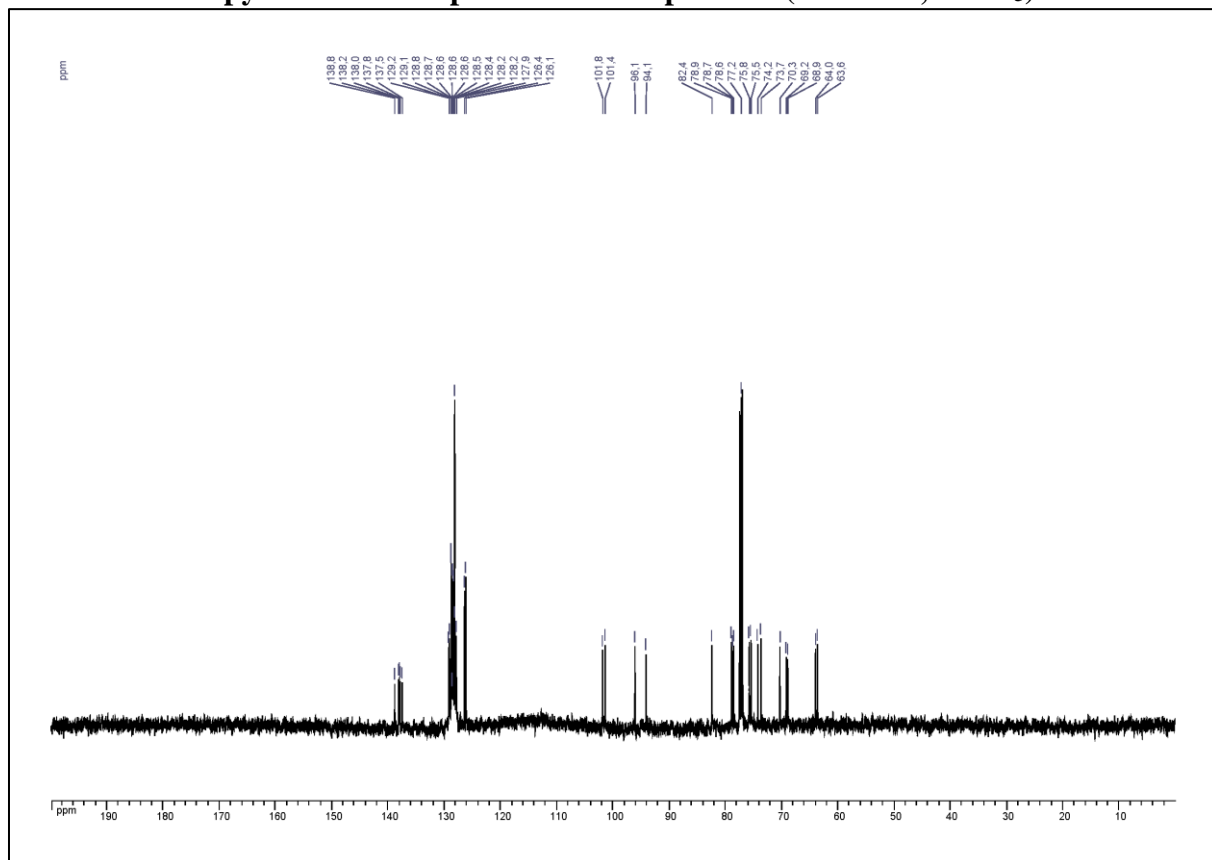
Copy of ^{13}C NMR spectrum of compound 2-*epi*-TreAz I (125 MHz, CD_3OD)



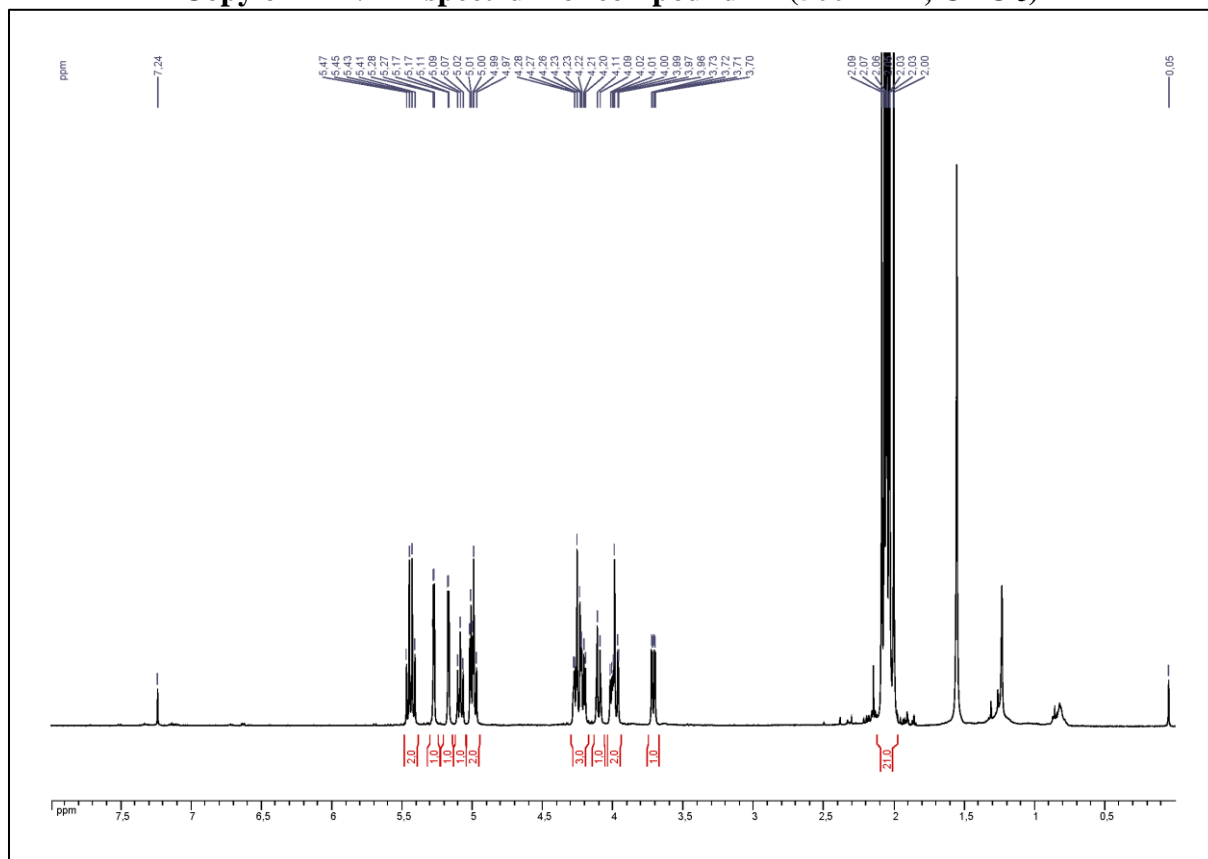
Copy of ¹H NMR spectrum of compound 9 (500 MHz, CDCl₃)



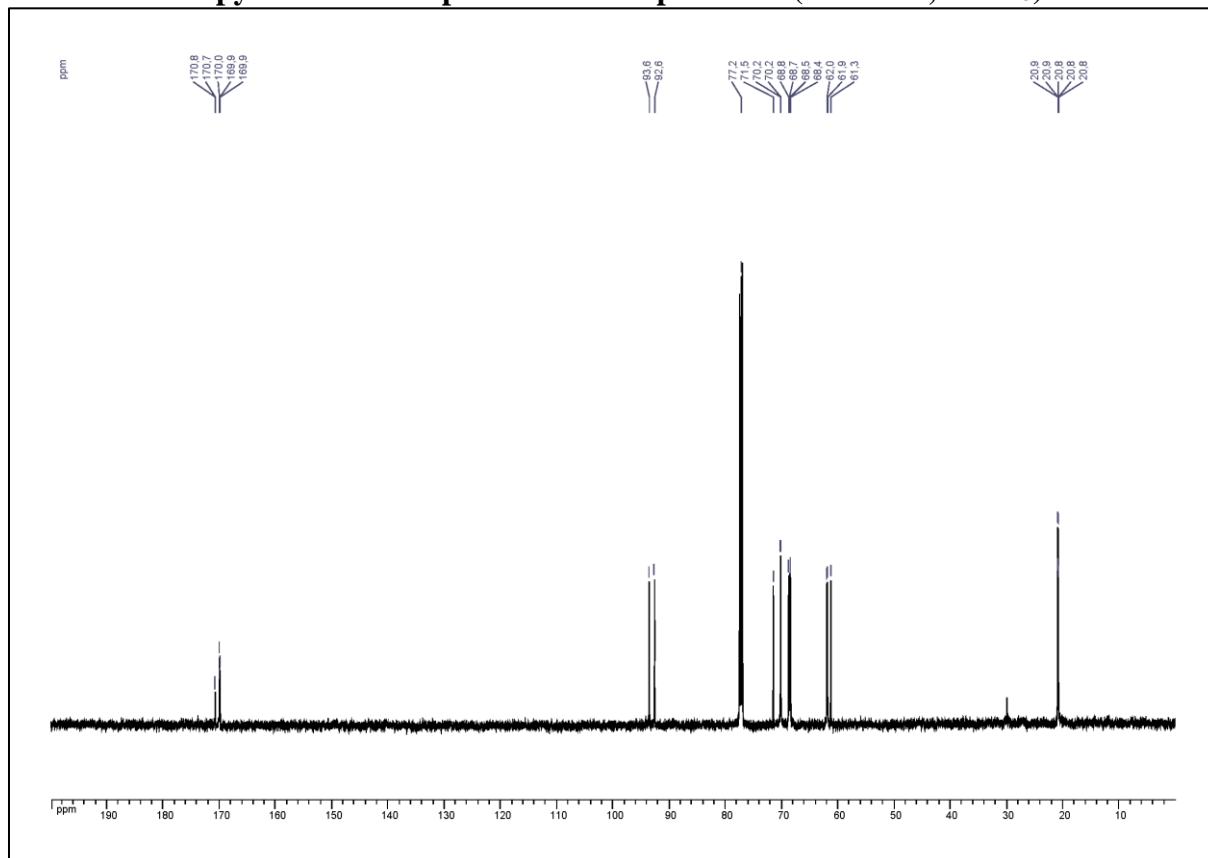
Copy of ¹³C NMR spectrum of compound 9 (125 MHz, CDCl₃)



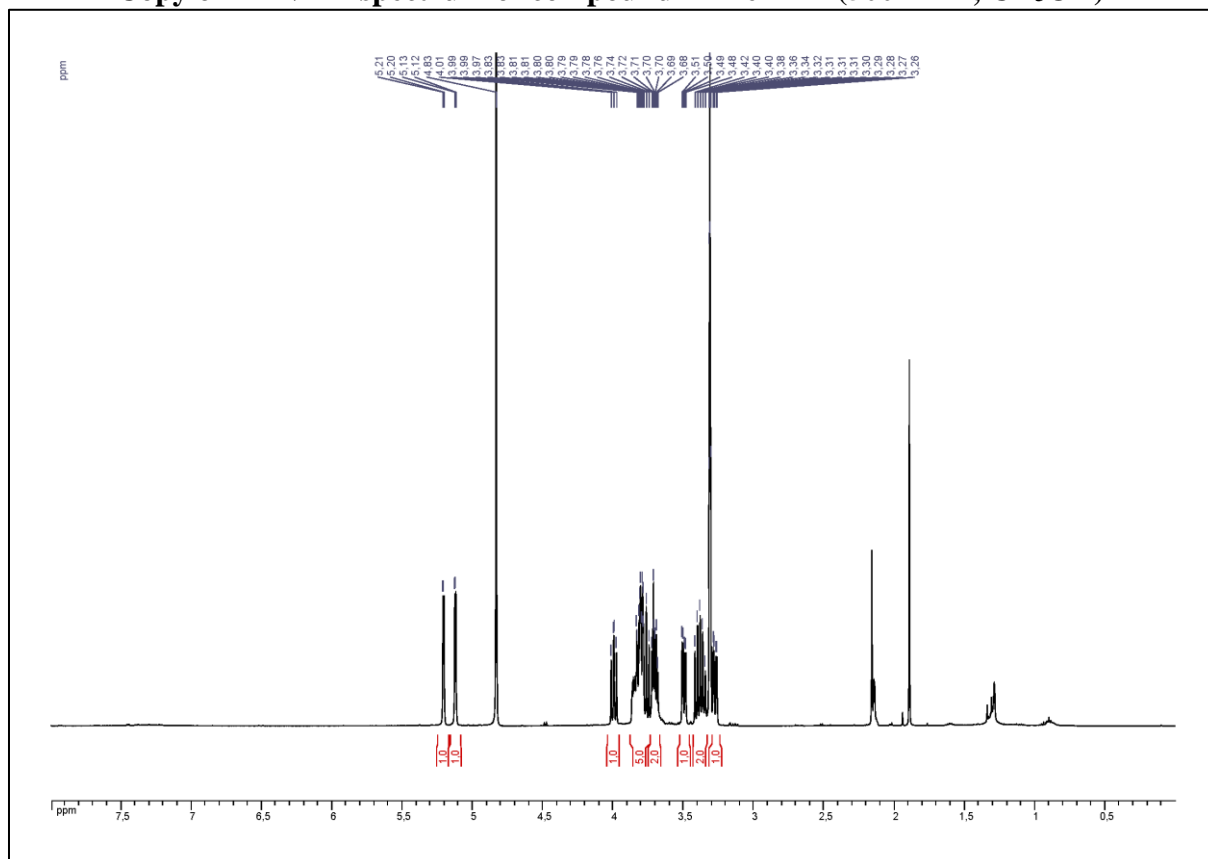
Copy of ^1H NMR spectrum of compound 12 (500 MHz, CDCl_3)



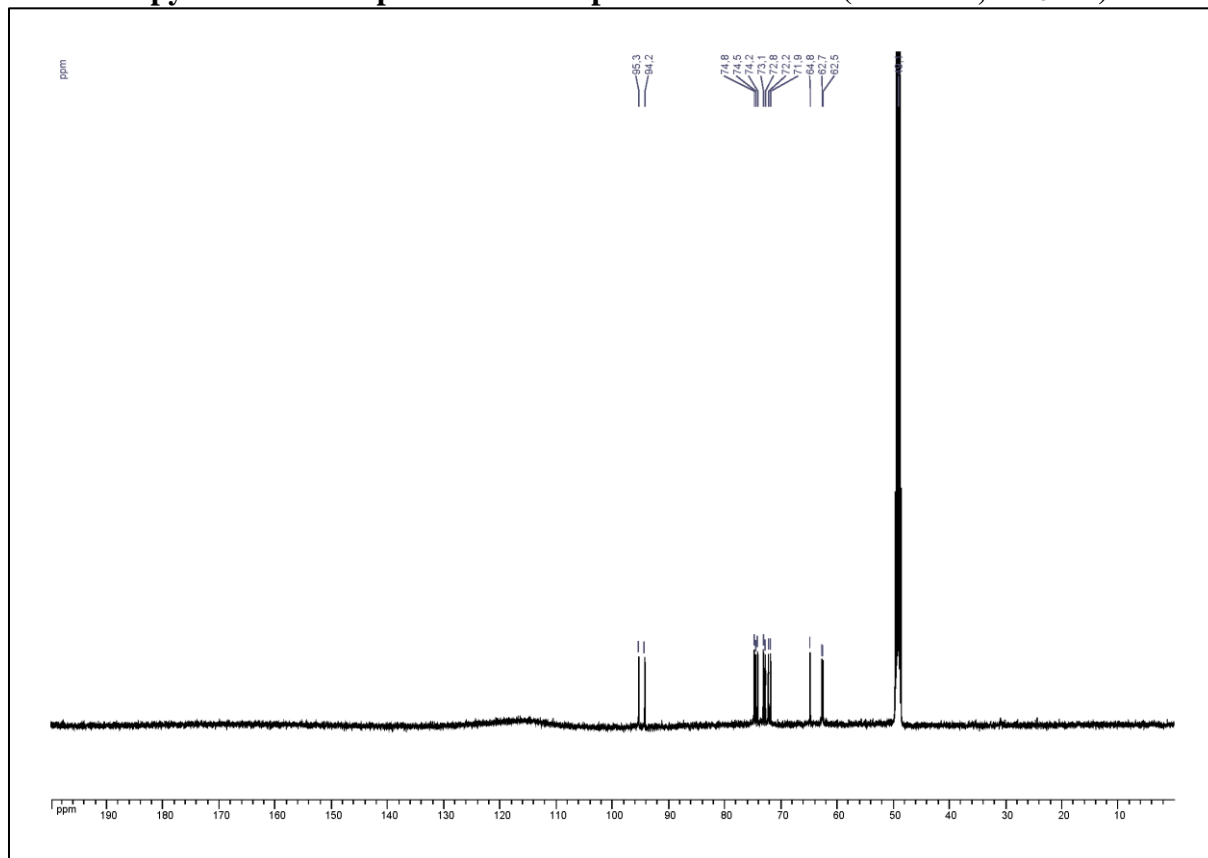
Copy of ^{13}C NMR spectrum of compound 12 (125 MHz, CDCl_3)



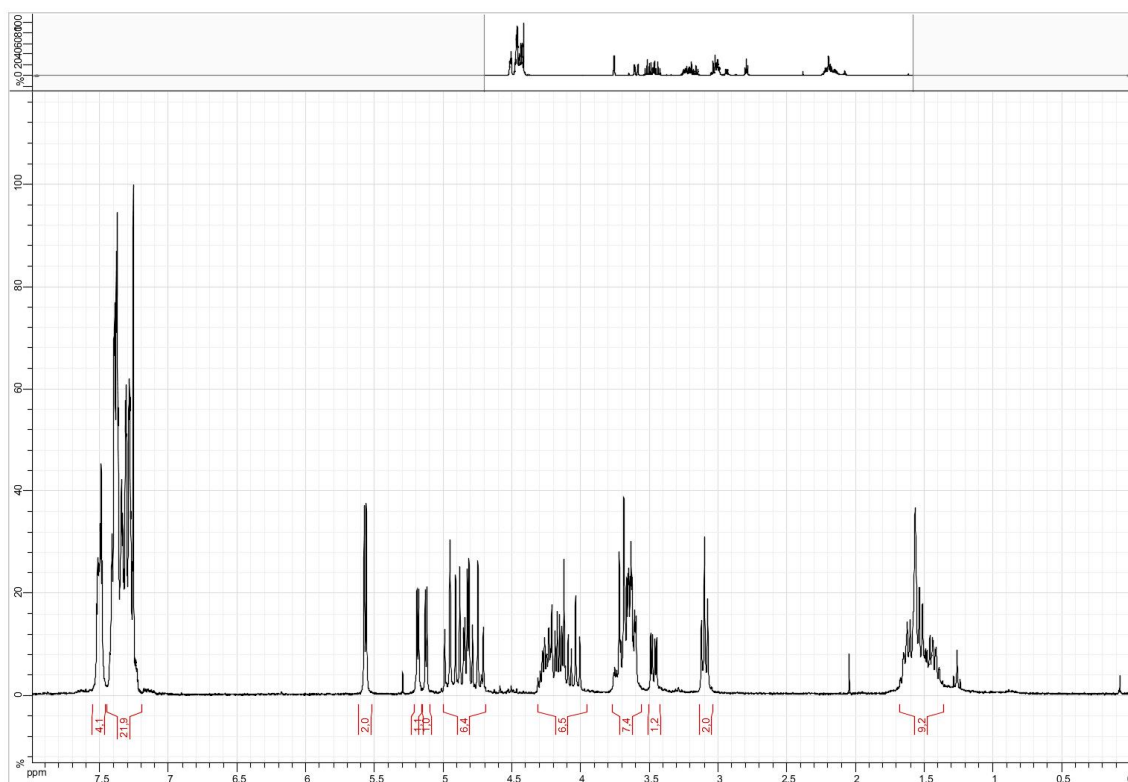
Copy of ^1H NMR spectrum of compound 2-TreAz II (500 MHz, CD_3OD)



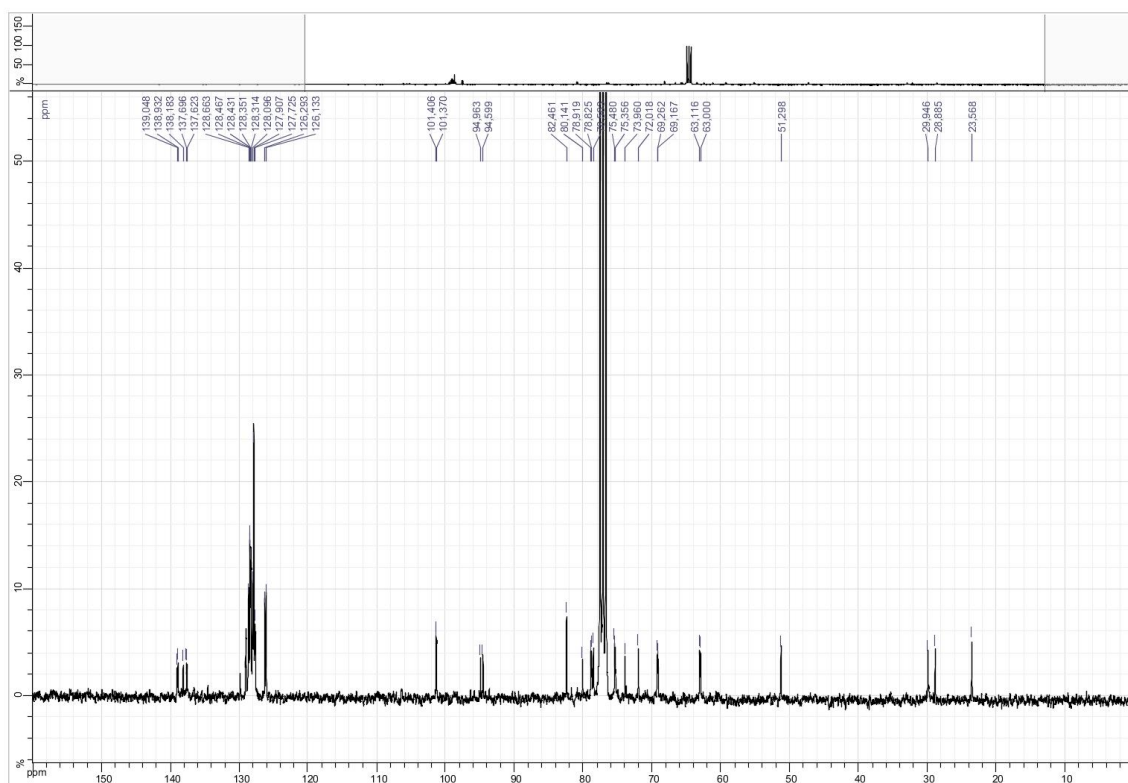
Copy of ^{13}C NMR spectrum of compound 2-TreAz II (125 MHz, CD_3OD)



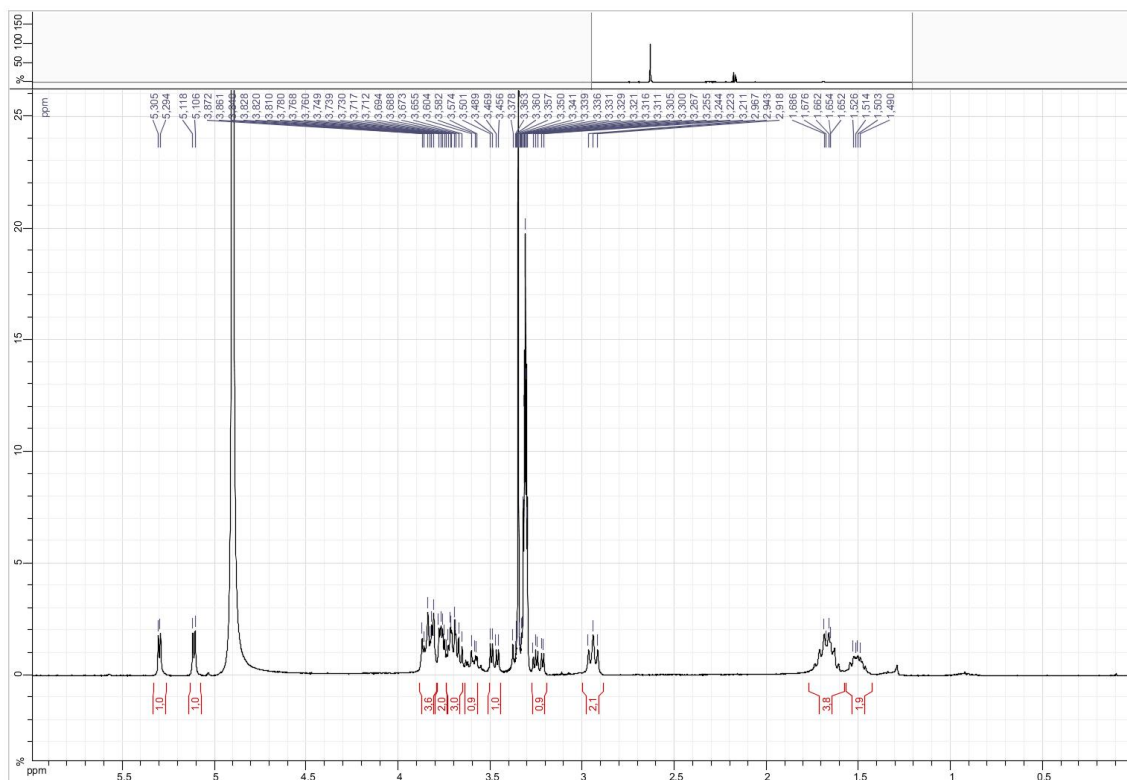
Copy of ^1H NMR spectrum of compound 13 (300 MHz, CDCl_3)



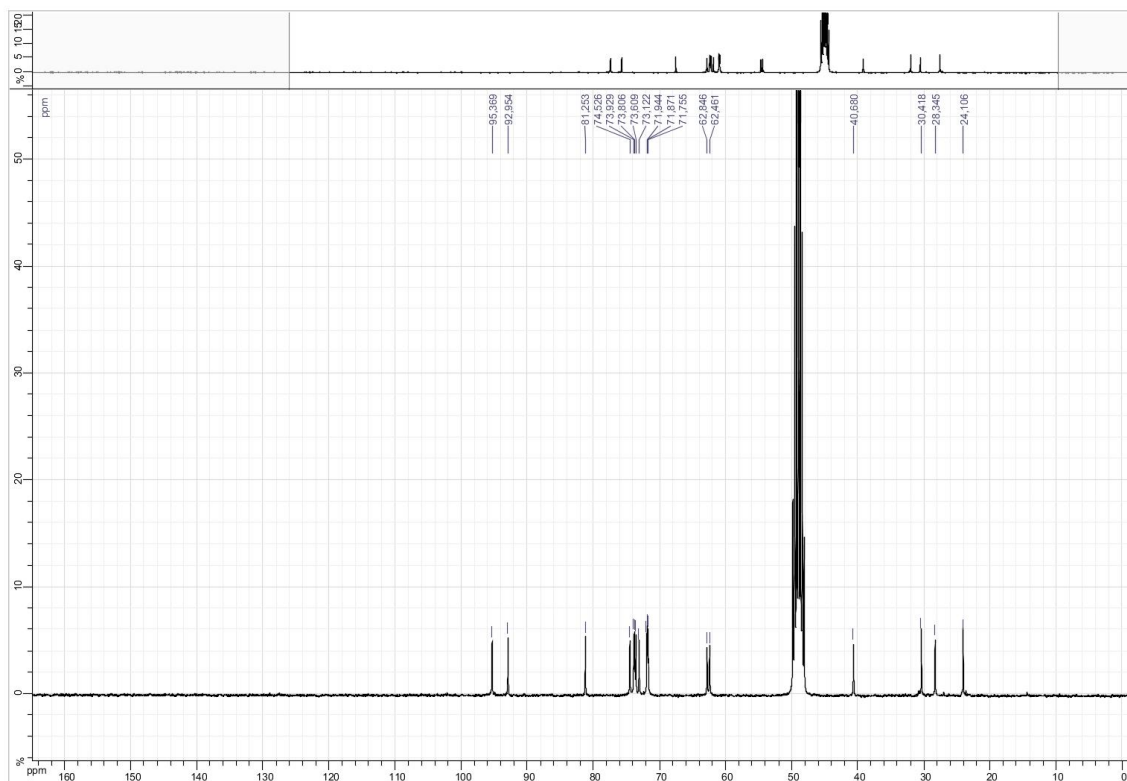
Copy of ^{13}C NMR spectrum of compound 13 (75 MHz, CDCl_3)



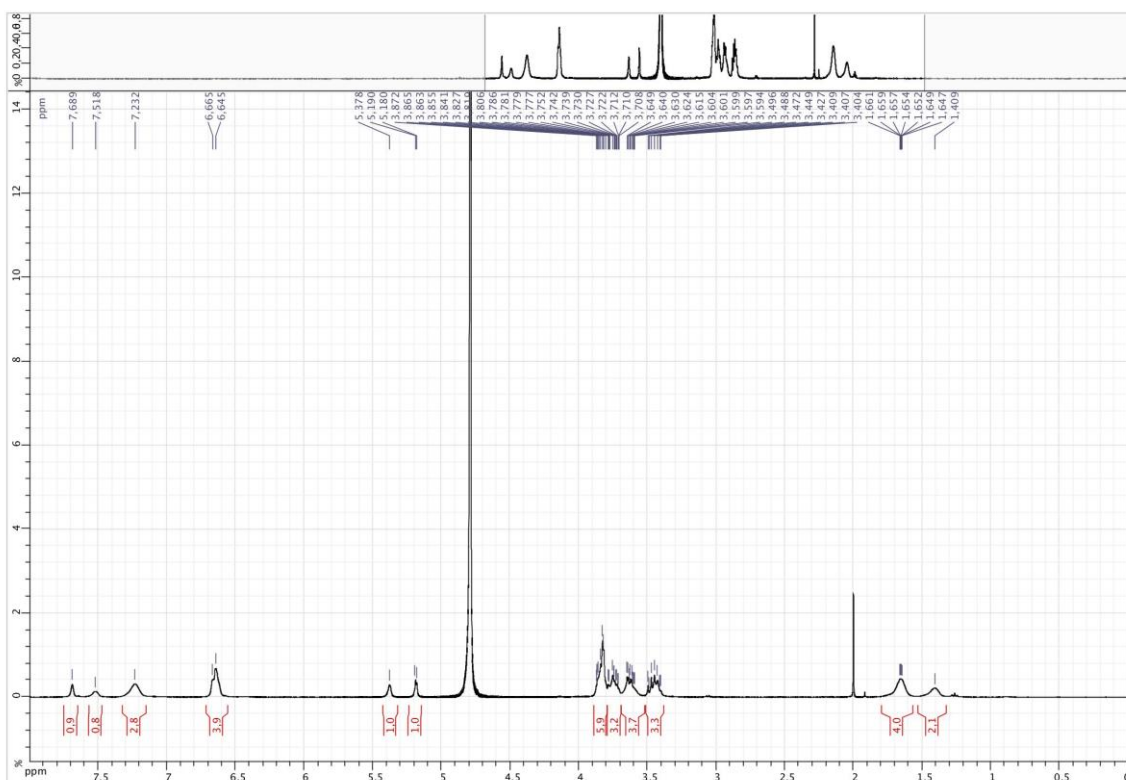
Copy of ^1H NMR spectrum of compound 14 (300 MHz, CD_3OD)



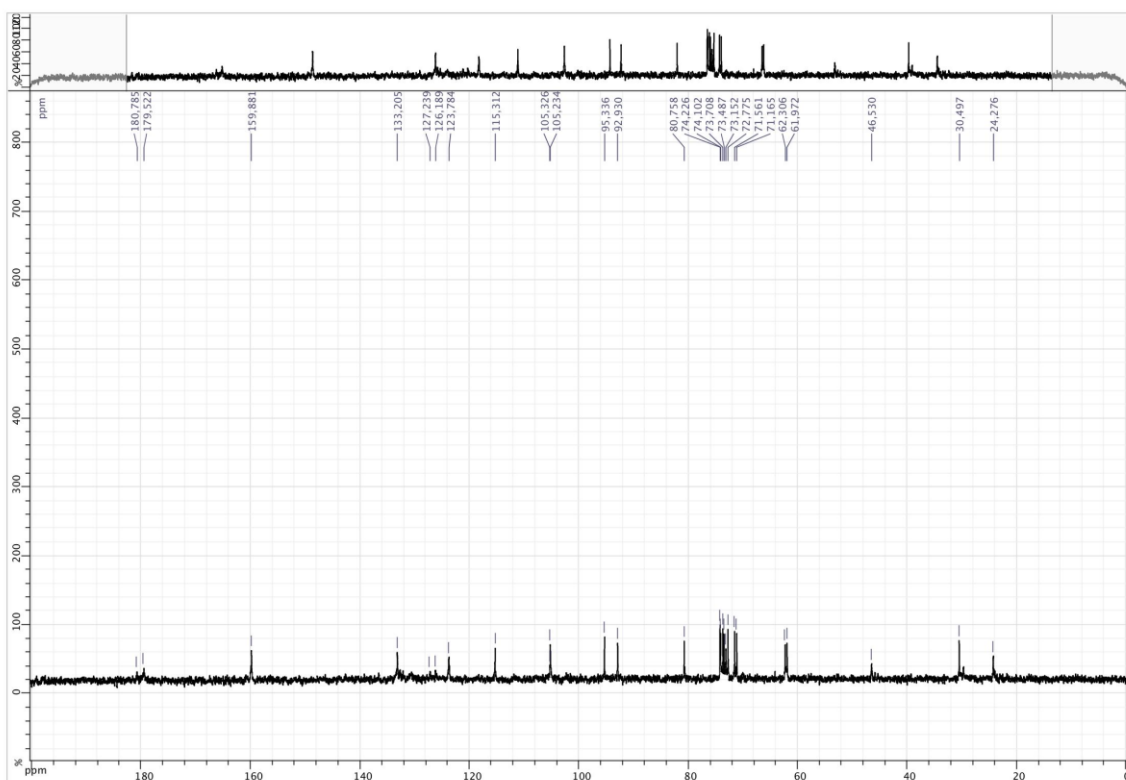
Copy of ^{13}C NMR spectrum of compound 14 (75 MHz, CD_3OD)



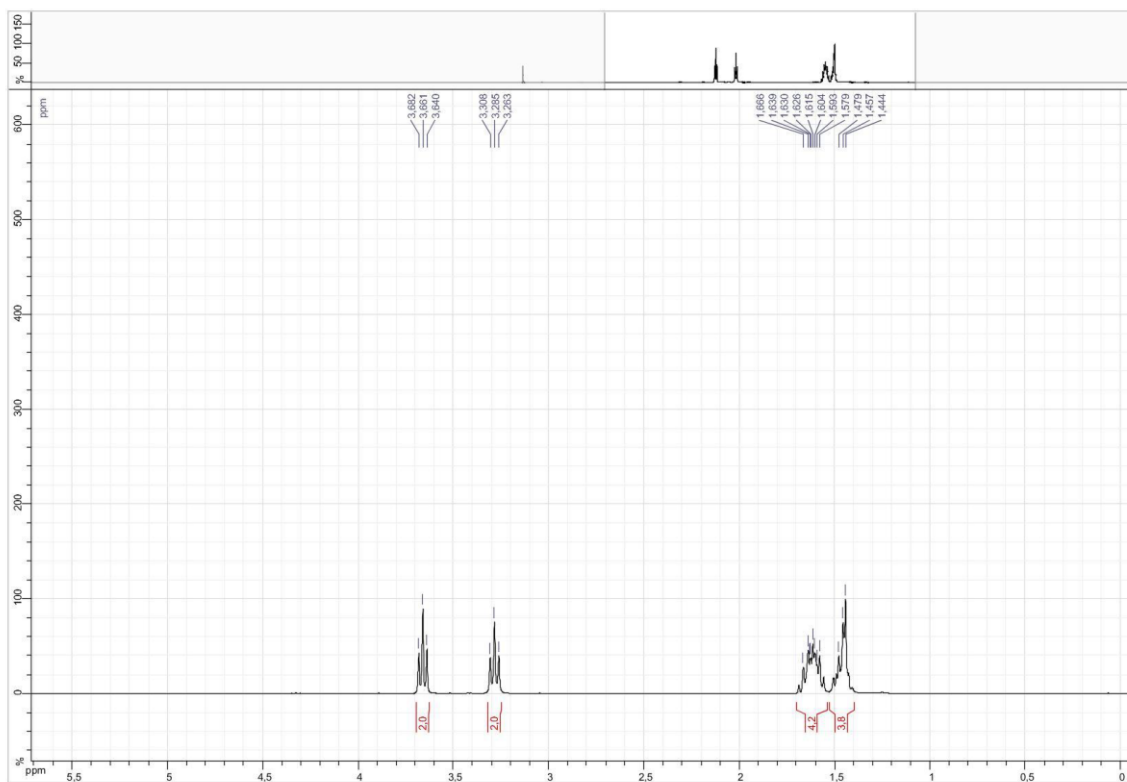
Copy of ^1H NMR spectrum of compound 2-FIC5Tre III (400 MHz, D_2O)



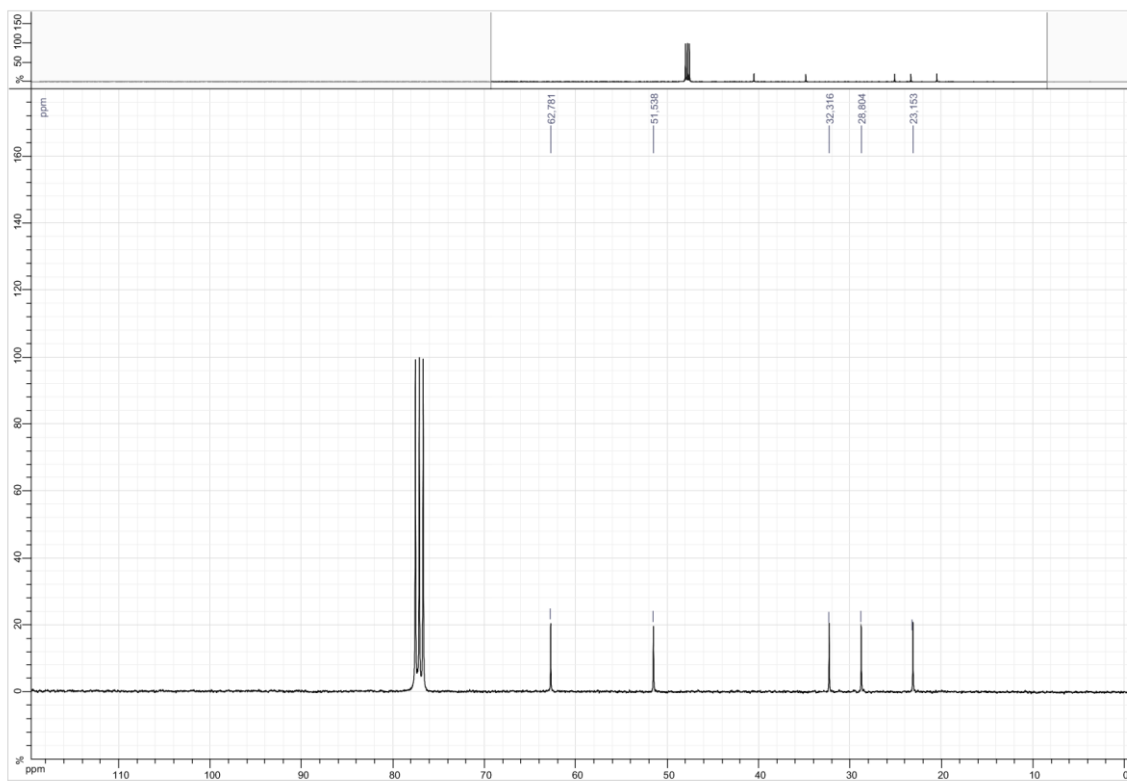
Copy of ^{13}C NMR spectrum of compound 2-FIC5Tre III (100 MHz, D_2O)



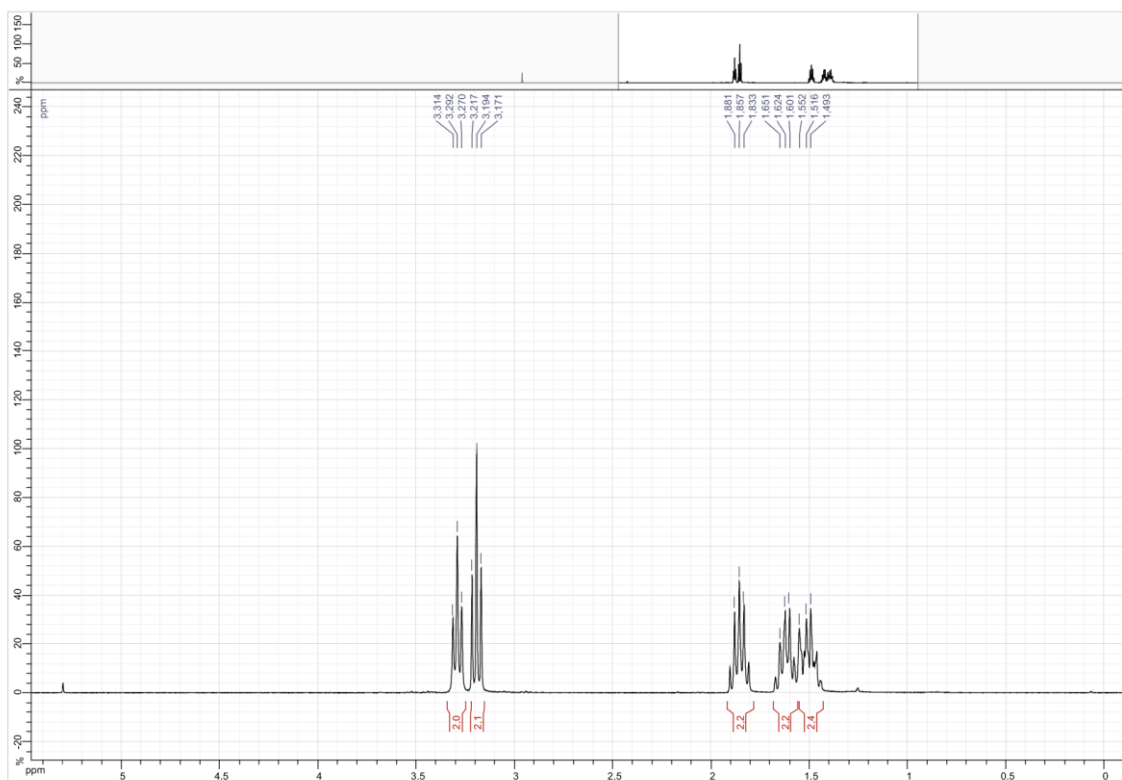
Copy of ^1H NMR spectrum of compound S1 (300 MHz, CDCl_3)



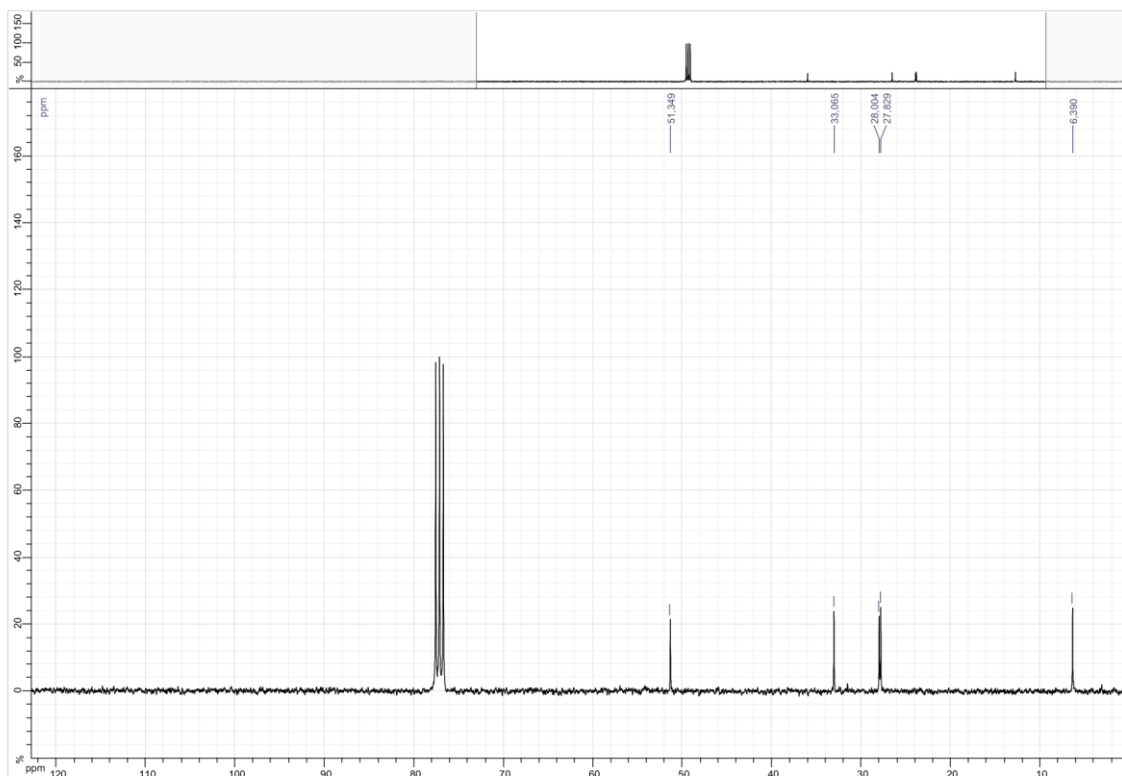
Copy of ^{13}C NMR spectrum of compound S1 (75 MHz, CDCl_3)



Copy of ^1H NMR spectrum of compound S2 (300 MHz, CDCl_3)



Copy of ^{13}C NMR spectrum of compound S2 (75 MHz, CDCl_3)



V. References in the supporting information

-
- ⁱ R. Toubiana, B. C. Das, J. Defaye, B. Mompon and M.-J. Toubiana, *Carbohydr. Res.*, 1975, **44**, 308-312.
- ⁱⁱ Y. Bourdreux, A. Lemétais, D. Urban and J.-M. Beau, *Chem. Commun.*, 2011, **47**, 2146-2148.
- ⁱⁱⁱ A. Attouche, D. Urban and J.-M. Beau, *Angew. Chem. Int. Ed.*, 2013, **52**, 9572-9575.
- ^{iv} B. M. Swarts, C. M. Holsclaw, J. C. Jewett, M. Alber, D. M. Fox, M. S. Siegrist, J. A. Leary, R. Kalscheuer and C. R. Bertozzi, *J. Am. Chem. Soc.*, 2012, **134**, 16123-16126.
- ^v K. Antien, A. Lacambra, F. P. Cossío, S. Massip, D. Deffieux, L. Pouységu, P. A. Peixoto and S. Quideau, *Chem. Eur. J.*, 2019, **25**, 11574–11580.
- ^{vi} A. Baron, Y. Blériot, M. Sollogoub and B. Vauzeilles, *Org. Biomol. Chem.*, 2008, **6**, 1898-1901.