

Supporting Information

Catalytic Redox-Neutral C–H functionalisation with TEMPO in Water to access Aminomethyl-substituted Pyrroles

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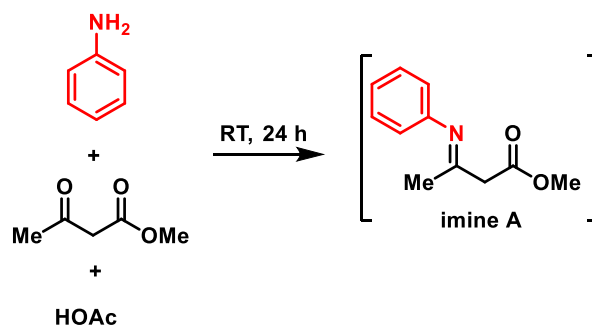
1. General Information

All reagents were purchased from commercial suppliers (Sigma-Aldrich, Oakwood and Combi-Blocks) and used without further purification, all solvents were analytical grade. Temperatures above room temperature were maintained by using an aluminium block heated on a hotplate, and reaction temperatures are reported as the temperature of the block surrounding the vessel. Thin layer chromatography (TLC) was performed using silica gel GF254, 0.25 mm thickness, visualization was accomplished with short wave UV light or KMnO_4 staining solution followed by heating. Hydrogen nuclear magnetic resonance spectra (^1H NMR) were obtained at 500 MHz in CDCl_3 solutions, at ambient temperature. Carbon-13 nuclear magnetic resonance spectra (^{13}C NMR) were obtained at 125 MHz in CDCl_3 solutions, at ambient temperature. Chemical shifts (δ) are given in ppm and the residual solvent signals were used as references for ^1H and ^{13}C NMR spectra (CDCl_3 : $\delta\text{H} = 7.27$ ppm, $\delta\text{C} = 77.00$ ppm). High resolution mass spectra were recorded on Thermo Scientific LTQ FT Ultra and Q Exactive Orbitrap spectrometers working with an electrospray ionization (ESI). The Gas Chromatography coupled to Mass Spectrometry (GC-MS) analyses were performed using a Network GC system 6890N (Agilent Technologies Inc., Palo Alto, CA, USA), equipped with a HP-5MS 5% Phenyl Methyl Silox (25.0 m \times 250 μm \times 0.25 μm nominal) capillary column. The GC analyses were carried out in split mode (ratio 150:1) using helium as carrier gas at a flow rate of 504 mL/min (7.65 psi). The injection port temperature was 250 $^\circ\text{C}$; the oven was maintained at an initial temperature of 50 $^\circ\text{C}$ for 3 minutes, then programmed at 40 $^\circ\text{C}/\text{min}$ to a temperature of 280 $^\circ\text{C}$, where it was held, post-run, for 2 minutes. The MS detector was at 250 $^\circ\text{C}$, using H_2 flow at 40.00 mL/min, air at 400 mL/min and He makeup flow at 45.0 mL/min.

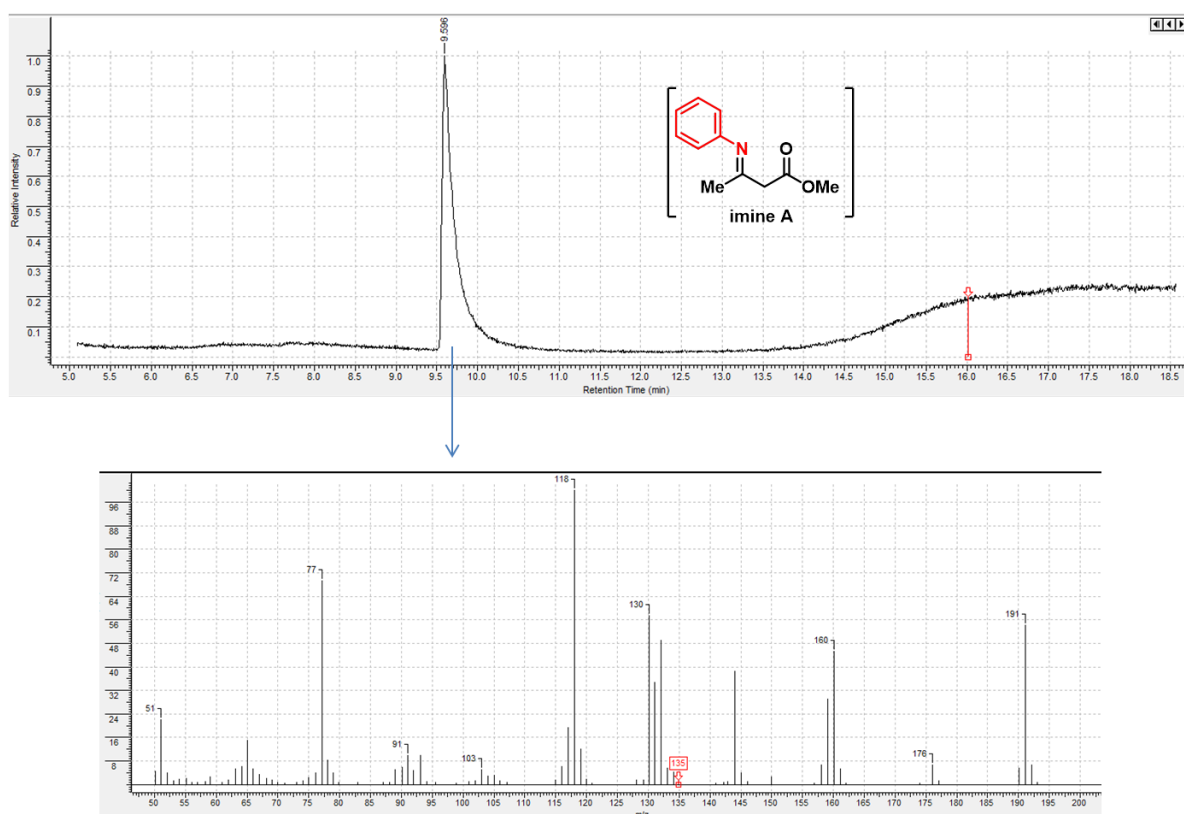
2. General Procedure

Following this sequence, acetic acid (1.0 mmol, 1.0 equiv), ketoesters (2.0 mmol, 2.0 equiv), anilines (2.0 mmol, 2.0 equiv) and TEMPO (30 mol%) were added to 4 ml of water. The resulting suspension was capped with a rubber septum and stirred at 70 $^\circ\text{C}$ for 72 h. The stirring speed of the reaction mixture was kept at 1150 rpm to ensure a proper diffusion of reaction components. Then, the suspension was extracted with ethyl acetate (2 \times 2 mL) and the combined organic layers were concentrated. The crude mixture was added to silica column and eluted using ethyl acetate/n-hexane (20:80) to afford the targeted product.

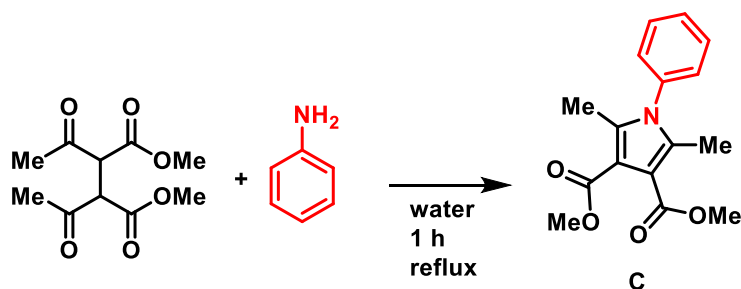
3. Synthesis of imine A



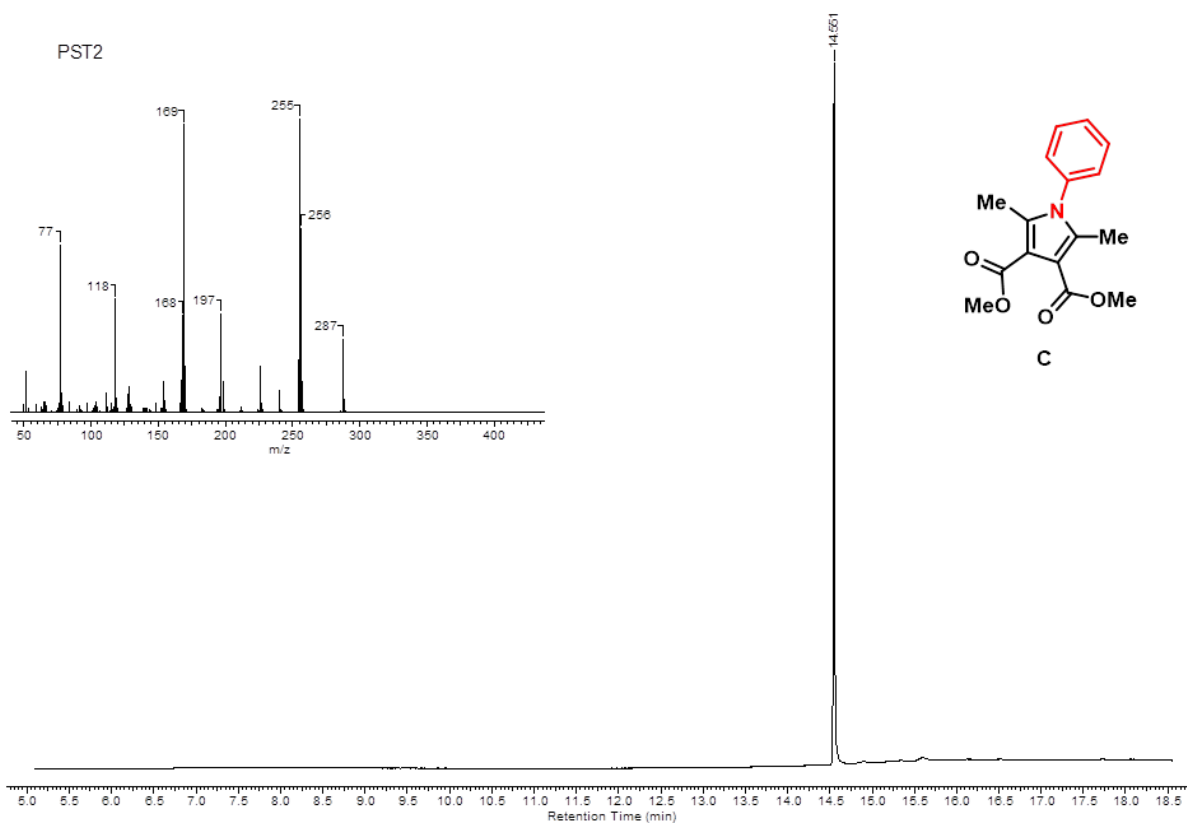
Aniline (1.0 mmol, 1.0 equiv), methyl acetoacetate (1.0 mmol, 1 equiv) and acetic acid (1.0 mmol, 1 equiv) were mixed together neat (solvent-free) and stirred at room temperature for 24 h. The crude mixture was added to the top of a silica gel column and eluted using ethyl acetate/n-hexane (20:80) to afford imine A:



4. Synthesis of pyrrole C



3,4-diacetylhexane-2,5-dione (1.0 mmol, 1 equiv) and aniline (1.0 mmol, 1 equiv) were added to 1mL of water and refluxed for 1h. Then, the mixture was extracted with ethyl acetate (3×5 mL) and dried with anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure and pyrrole C was recrystallized from ethanol:



5. Scheme 3 in the manuscript, item I, Figure S1

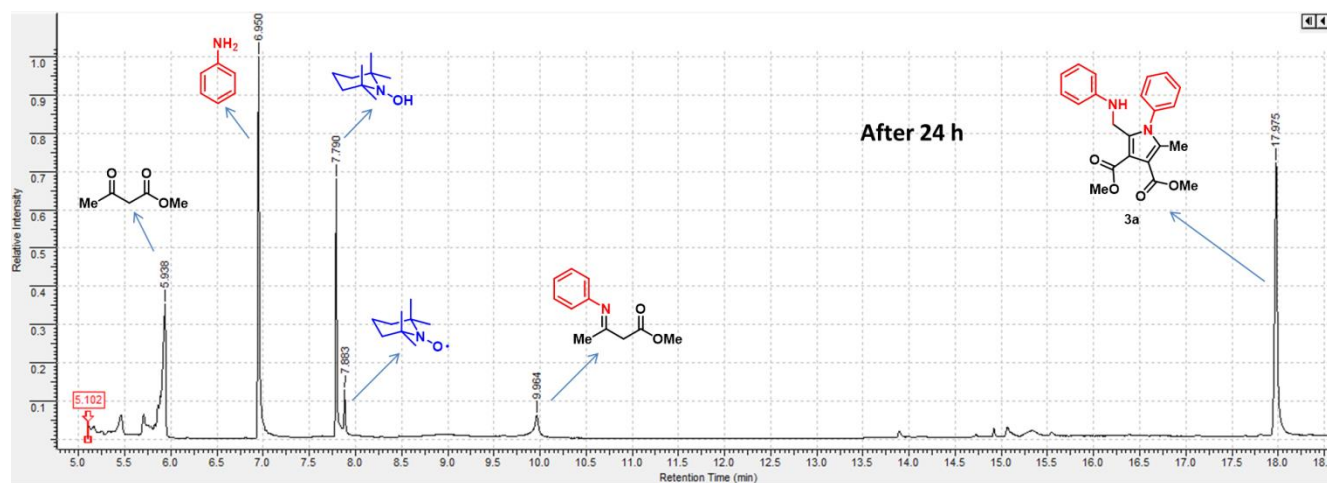
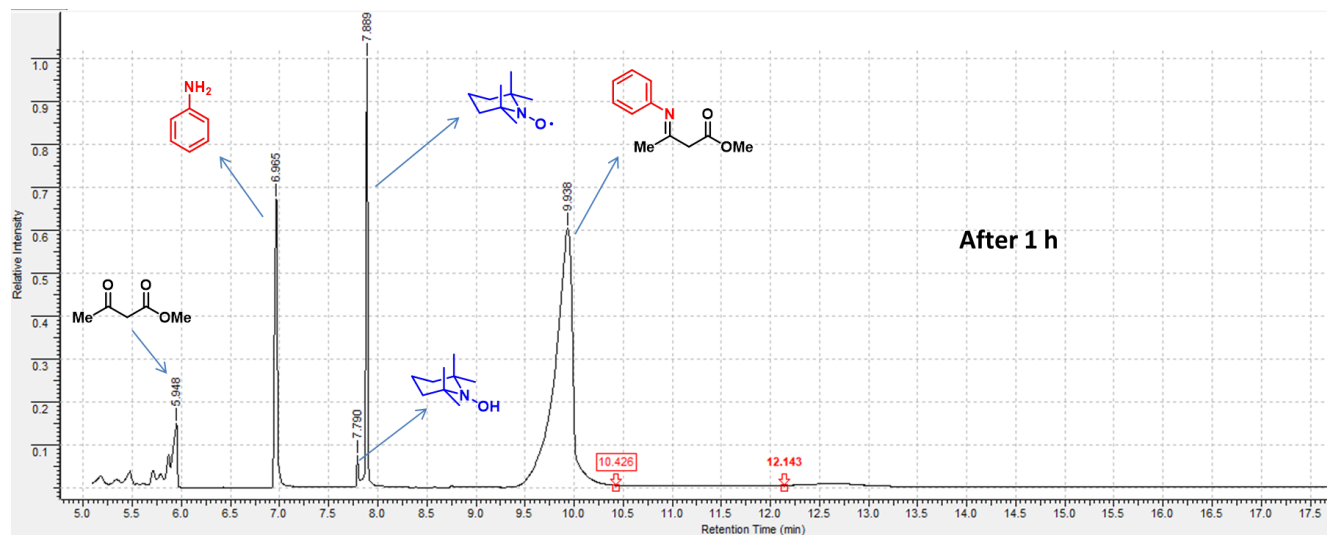


Figure S1

6. Scheme 3 in the manuscript, item III, Figure S2

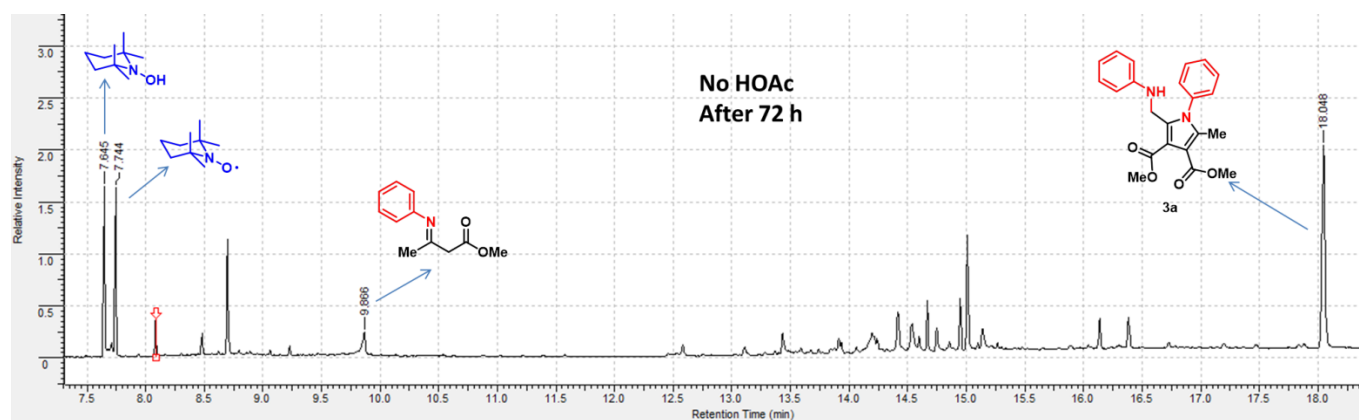


Figure S2

7. Scheme 4 in the manuscript, item III, Figure S3

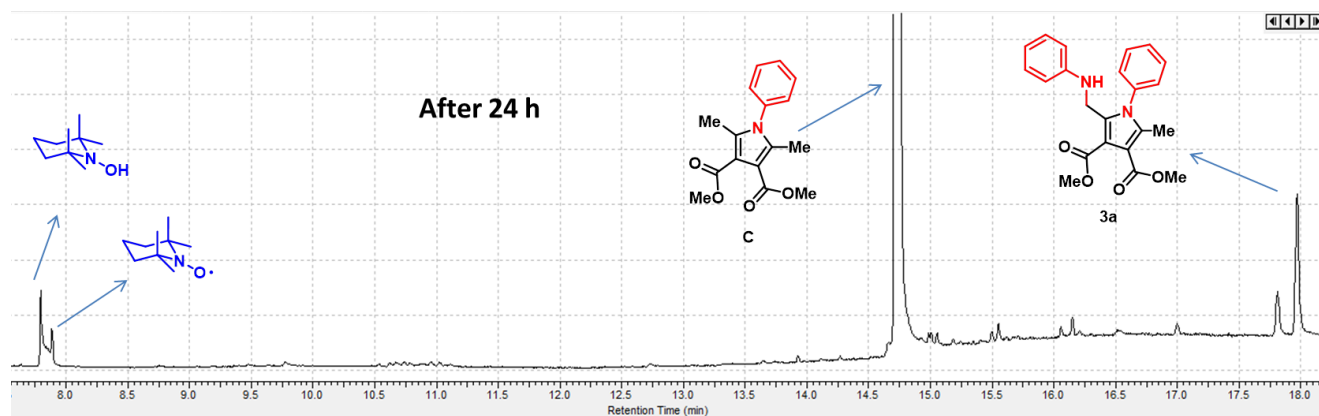


Figure S3

8. Scheme 5 in the manuscript, item II, Figure S4

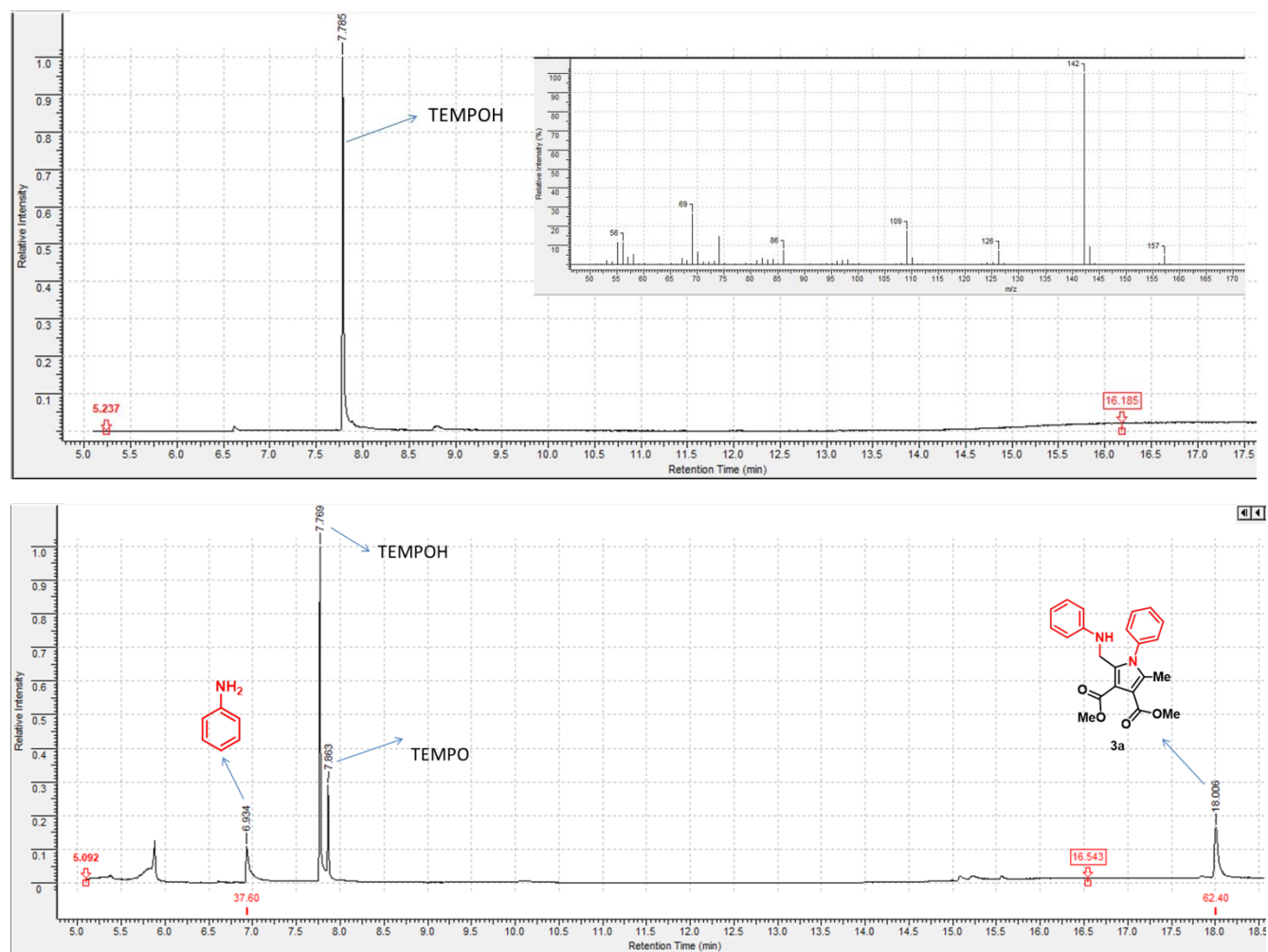


Figure S4

9. Scheme 5 in the manuscript, item III, Figure S5

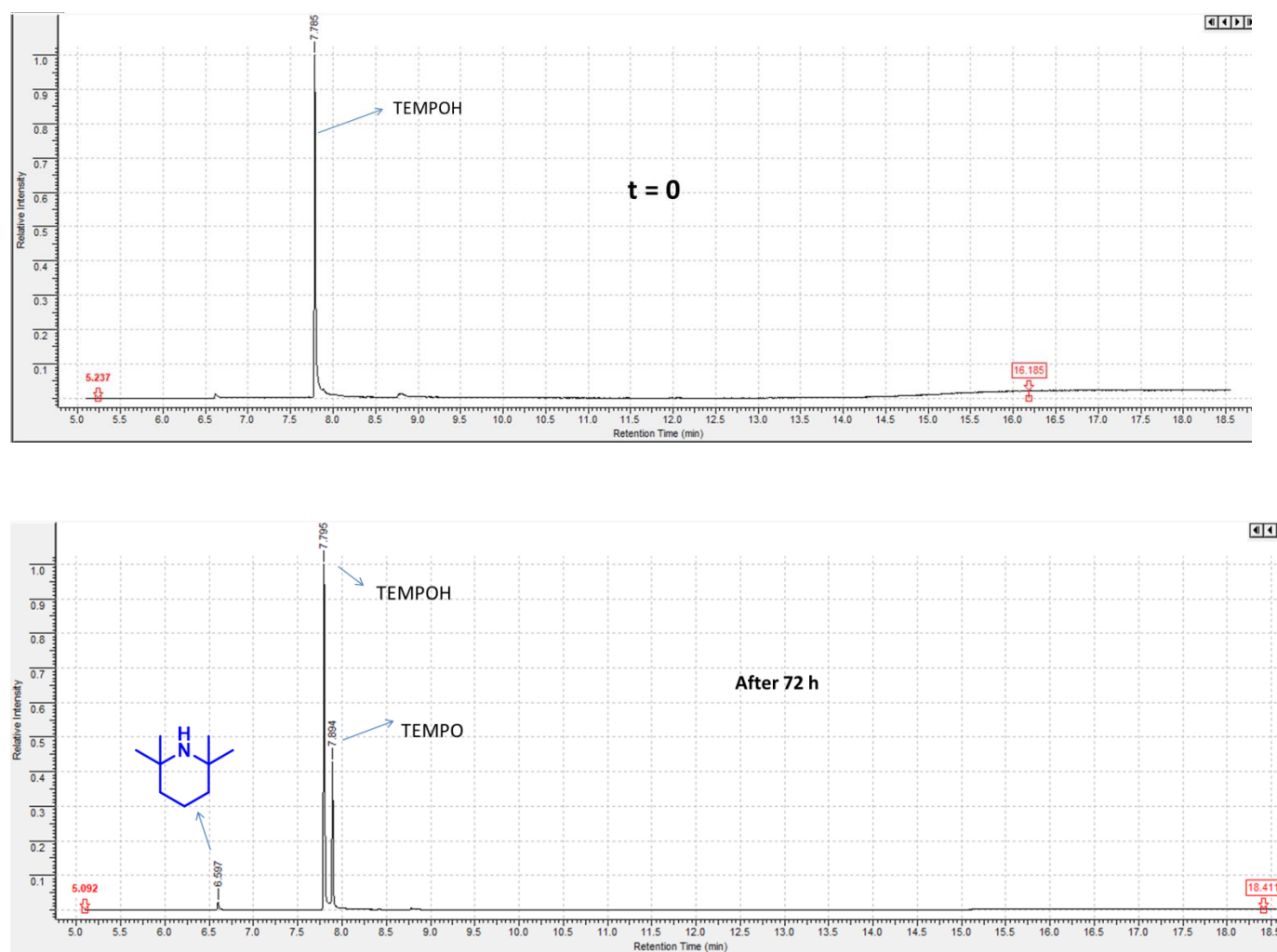
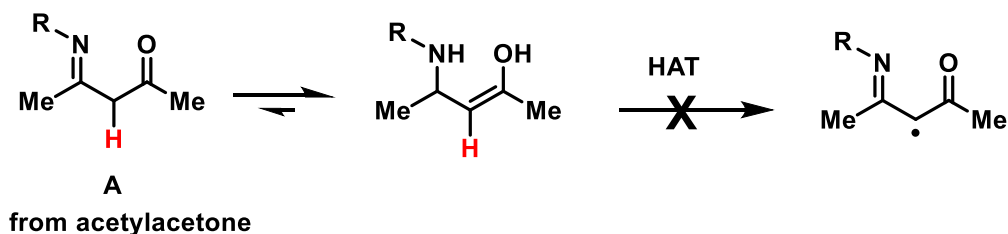


Figure S5

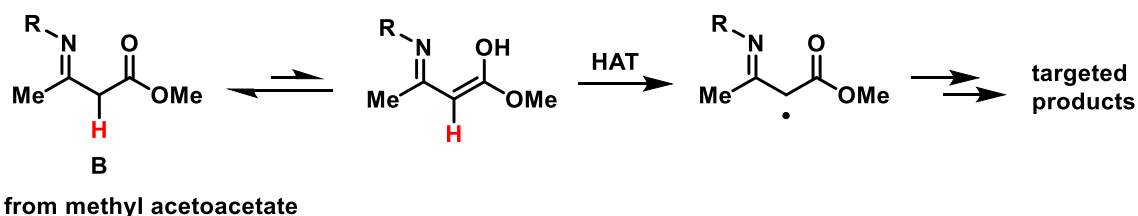
10. Possible reason for reaction failure when using either acetylacetone or dimethyl malonate.

Following the **General Procedure** with acetylacetone and aniline, we could observe by GC-MS the formation of the corresponding imine in accordance to the mechanistic studies described in the manuscript. However, no product was obtained after 72 h and the imine remained unreacted. We hypothesize that, under our conditions, the enol form of imine **A** is favoured and considering that HAT parallels bond dissociation energies¹, hydrogen transfer from sp^2 carbon becomes more difficult compared to the same transfer from sp^3 carbons:

¹ J. M. Mayer, *Acc. Chem. Res.*, 2011, **44**, 36–46.



On the other hand, the keto form of imine **B**, coming from methyl acetoacetate, might be favoured due to higher pK_a of α -hydrogens next to an ester group. In this scenario, imine **B** undergoes HAT more easily (from sp^3 carbon) allowing the transformation reach completion:

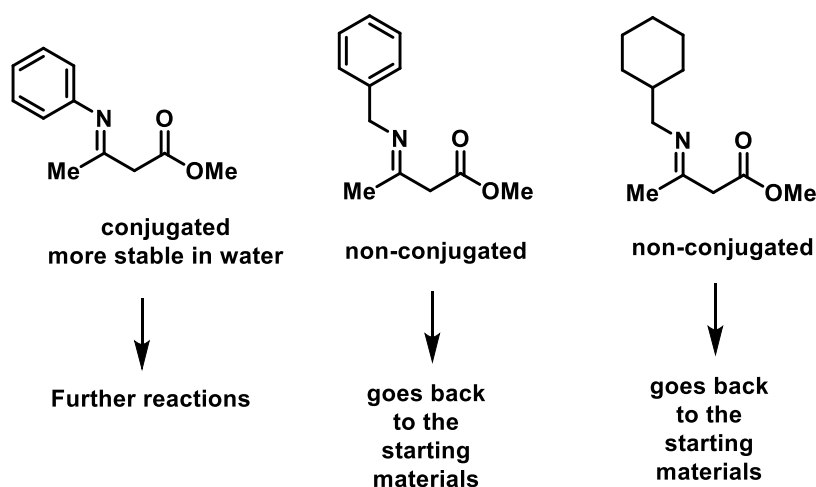


When employing dimethyl malonate, the imine is not formed at all and the process is shut down. This result emphasizes the necessity of imine formation to carry the transformation on.



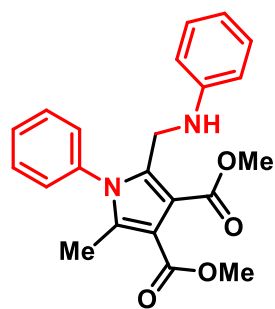
11. Possible reason for reaction failure when using either benzylamines or aliphatic amines.

The formation of imines takes place by a mechanism that is the reverse of the hydrolysis. Conjugated C=N systems are more stable to aqueous hydrolysis than non-conjugated imines and, for many cases, the equilibrium constants for formation are high, even in aqueous solution.² Therefore, we believe that, under our conditions, conjugated imines formed from anilines are very stable and allow the transformation to carry on. On the other hand, non-conjugated imines coming from benzylamines or aliphatic amines hydrolyse easily, thus preventing the reaction to move forward:



² W. P. Jencks, *Prog. Phys. Org. Chem.*, **1964**, 2, 63; J. M. Sayer, M. Peskin, and W. P. Jencks, *J. Am. Chem. Soc.*, **1973**, 95, 4277.

12. Characterization of the products

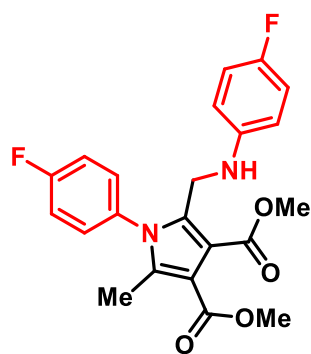


3a

Prepared from aniline and methyl acetoacetate following the general procedure to give the product as pale yellow oil (89% yield).³

¹H NMR (500 MHz, Chloroform-*d*): δ 7.51–7.48 (m, 3H), 7.23–7.19 (m, 2H), 7.09 (appt t, 2H, $J = 8.24$ Hz), 6.70 (appt t, 1H, $J = 7.32$ Hz), 6.41 (d, 2H, $J = 8.24$ Hz), 4.14 (s, 2H), 3.85 (s, 6H), 2.16 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.89, 165.68, 147.41, 136.15, 135.32, 135.05, 129.65, 129.04, 128.07, 118.02, 114.52, 113.64, 112.67, 51.83, 51.54, 38.92, 11.91.



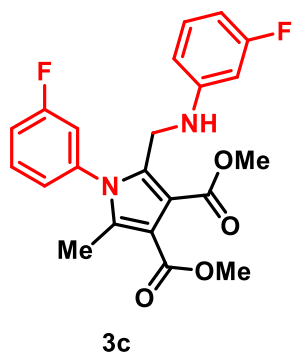
3b

Prepared from 4-fluoroaniline and methyl acetoacetate following the general procedure to give the product as pale yellow oil (92% yield).³

¹H NMR (500 MHz, Chloroform-*d*): δ 7.20–7.18 (m, 4H), 6.81 (t, 2H, $J = 8.80$ Hz), 6.38–6.38 (m, 2H), 4.05 (s, 2H), 3.86 (s, 3H), 3.85 (s, 3H), 2.15 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.76, 165.54, 163.75, 161.75, 157.23, 143.72, 135.33, 135.05, 132.01, 129.94, 129.87, 116.77, 116.58, 115.64, 115.45, 114.90, 114.86, 114.63, 112.88, 51.89, 51.60, 39.83, 11.89.

³ M. Zhao, Z. Zhang, Z. Ren, D. Yang and Z. Guan, *Org. Lett.*, 2018, **20**, 3088–3091.

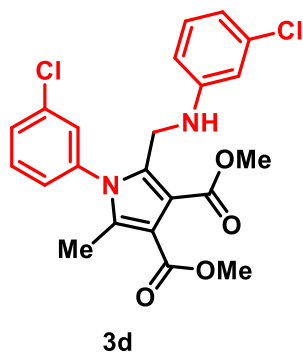


Prepared from 3-fluoroaniline and methyl acetoacetate following the general procedure to give the product as pale yellow oil (90% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.48 (m, 1H), 7.26–7.23 (m, 1H), 7.05–6.96 (m, 3H), 6.37 (td, 1H, $J = 8.49, 1.57$ Hz), 6.17 (dd, 1H, $J = 7.23, 1.26$ Hz), 6.04 (dt, 1H, $J = 11.32, 2.20$ Hz), 4.13 (s, 2H), 3.86 (s, 6H), 2.18 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 165.65, 165.40, 164.79, 163.70, 162.86, 161.71, 148.98, 148.90, 137.35, 137.27, 135.13, 134.47, 130.99, 130.14, 130.06, 124.01, 123.98, 116.93, 116.76, 115.86, 115.69, 114.87, 113.00, 109.39, 104.57, 104.41, 100.16, 99.95, 51.89, 51.59, 38.62, 11.80.

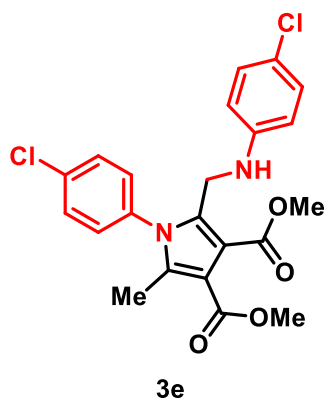
HRMS m/z (ESI): calcd. for $\text{C}_{22}\text{H}_{21}\text{F}_2\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 415.14639, found 415.14492.



Prepared from 3-chloroaniline and methyl acetoacetate following the general procedure to give the product as pale orange oil (85% yield).³

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.53–7.50 (m, 1H), 7.45 (t, 1H, $J = 7.86$ Hz), 7.23 (t, 1H, $J = 1.89$ Hz), 7.12–7.13 (m, 1H), 6.99 (t, 1H, $J = 7.86$ Hz), 6.65 (dd, 1H, $J = 7.86, 1.26$ Hz), 6.33 (t, 1H, $J = 1.89$ Hz), 6.29 (dd, 1H, $J = 7.86, 1.89$ Hz), 4.14 (d, 1H, $J = 14.46$ Hz), 4.13 (d, 1H, $J = 14.46$ Hz), 3.87 (s, 3H), 3.85 (s, 3H), 2.17 (s, 3H).

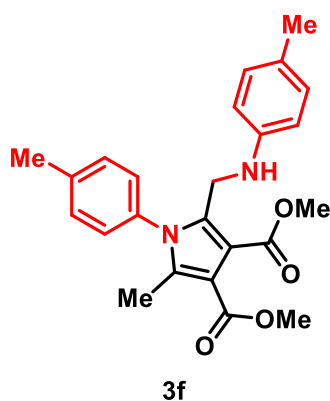
$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 165.70, 165.43, 148.15, 137.09, 135.45, 135.16, 134.87, 134.60, 130.72, 130.08, 129.98, 128.45, 126.42, 118.12, 114.89, 113.13, 113.10, 112.09, 51.97, 51.65, 38.65, 11.88.



Prepared from 4-chloroaniline and methyl acetoacetate following the general procedure to give the product as pale orange oil (86% yield).³

¹H NMR (500 MHz, Chloroform-*d*): δ 7.46 (d, 2H, $J = 8.80$ Hz), 7.14 (d, 2H, $J = 8.31$ Hz), 7.03 (d, 2H, $J = 8.80$ Hz), 6.35 (d, 2H, $J = 8.80$ Hz), 4.09 (s, 2H), 3.85 (s, 6H), 2.15 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.70, 165.40, 145.59, 135.65, 135.24, 134.46, 129.90, 129.31, 128.95, 123.02, 114.86, 113.04, 51.91, 51.60, 39.11, 11.86.

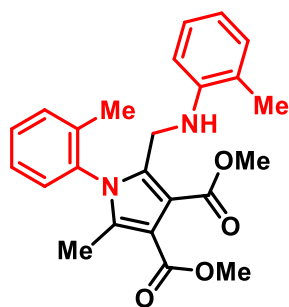


Prepared from 4-methylaniline and methyl acetoacetate following the general procedure to give the product as pale yellow oil (88% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.28 (d, 2H, $J = 8.17$ Hz), 7.09 (d, 2H, $J = 8.17$ Hz), 6.89 (d, 2H, $J = 8.17$ Hz), 6.34 (d, 2H, $J = 8.80$ Hz), 4.11 (s, 2H), 3.84 (s, 6H), 2.44 (s, 3H), 2.21 (s, 3H), 2.15 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.92, 165.75, 145.22, 139.49, 135.41, 133.51, 130.19, 129.50, 127.76, 127.24, 114.35, 113.93, 112.49, 51.78, 51.49, 39.32, 21.22, 20.37, 11.89.

HRMS *m/z* (ESI): calcd. for C₂₄H₂₇N₂O₄ [M+H]⁺ 407,19653, found 407,19525.

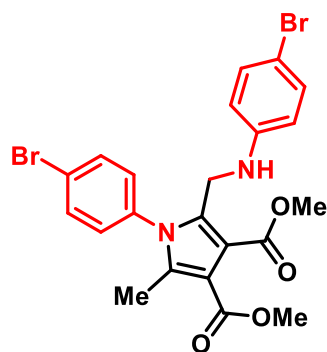


3g

Prepared from 2-methylaniline and methyl acetoacetate following the general procedure to give the product as pale yellow oil (84% yield).³

¹H NMR (500 MHz, Chloroform-*d*): δ 7.44–7.30 (m, 3H), 7.06 (d, 1H, $J = 7.02$ Hz), 6.98 (appt t, 2H, $J = 8.24$ Hz), 6.65 (t, 1H, $J = 7.02$ Hz), 6.32 (d, 1H, $J = 7.63$ Hz), 4.16 (d, 1H, $J = 14.04$ Hz), 4.01 (d, 1H, $J = 14.04$ Hz), 3.86 (s, 6H), 2.08 (s, 3H), 2.03 (s, 3H), 1.92 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 166.05, 165.60, 136.69, 135.27, 135.16, 131.28, 130.10, 129.84, 128.50, 127.17, 126.88, 114.63, 112.64, 51.87, 51.51, 38.97, 17.36, 17.16, 11.46.

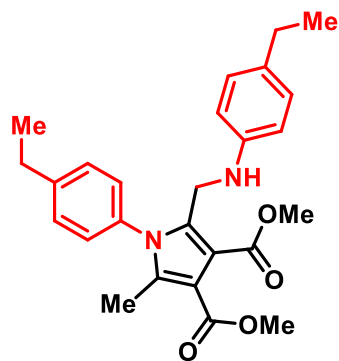


3h

Prepared from 4-bromoaniline and methyl acetoacetate following the general procedure to give the product as pale orange oil (87% yield).³

¹H NMR (500 MHz, Chloroform-*d*): δ 7.61 (d, 2H, $J = 8.80$ Hz), 7.17 (d, 2H, $J = 8.80$ Hz), 7.07 (d, 2H, $J = 8.80$ Hz), 6.31 (d, 2H, $J = 8.80$ Hz), 4.09 (s, 2H), 3.85 (s, 6H), 2.15 (s, 3H).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.73, 165.38, 145.86, 135.22, 134.95, 134.30, 132.94, 131.87, 129.60, 123.74, 115.45, 114.95, 113.10, 110.29, 51.95, 51.63, 39.10, 11.89.



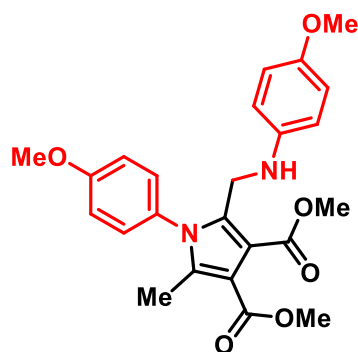
3i

Prepared from 4-ethyl aniline and methyl acetoacetate following the general procedure to give the product as pale brown oil (90% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.29 (d, 2H, $J = 8.31$ Hz), 7.09 (d, 2H, $J = 8.31$ Hz), 6.92 (d, 2H, $J = 8.31$ Hz), 6.39 (d, 2H, $J = 8.31$ Hz), 4.12 (s, 2H), 3.85 (s, 6H), 2.73 (q, 2H, $J = 7.34$ Hz), 2.51 (q, 2H, $J = 7.34$ Hz), 2.15 (s, 3H), 1.30 (t, 3H, $J = 7.83$ Hz), 1.17 (t, 3H, $J = 7.83$ Hz).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 166.02, 165.71, 145.70, 135.52, 133.56, 128.99, 128.37, 127.85, 114.37, 112.50, 51.83, 51.50, 39.67, 28.51, 27.91, 15.85, 15.29, 11.92.

HRMS m/z (ESI): calcd. for $\text{C}_{26}\text{H}_{31}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 435,22783, found 435,22632.



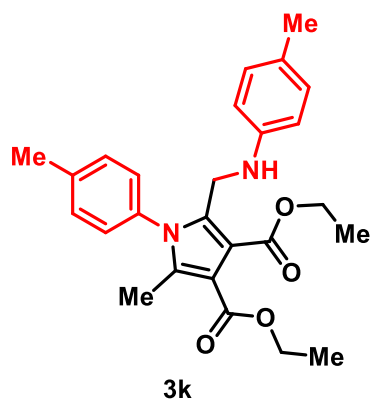
3j

Prepared from 4-methoxyaniline and methyl acetoacetate following the general procedure to give the product as pale yellow oil (83% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.11 (d, 2H, $J = 8.80$ Hz), 6.96 (d, 2H, $J = 8.80$ Hz), 6.68 (d, 2H, $J = 8.80$ Hz), 6.40 (d, 2H, $J = 8.80$ Hz), 4.07 (s, 2H), 3.87 (s, 3H), 3.84 (s, 6H), 3.72 (s, 3H), 2.14 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 165.76, 165.57, 159.84, 152.47, 141.42, 135.48, 135.40, 128.96, 128.47, 115.37, 114.44, 113.96, 112.25, 55.52, 55.39, 51.59, 51.31, 40.09, 11.70.

HRMS m/z (ESI): calcd. for $\text{C}_{24}\text{H}_{27}\text{N}_2\text{O}_6$ $[\text{M}+\text{H}]^+$ 439,18636, found 439,18489.

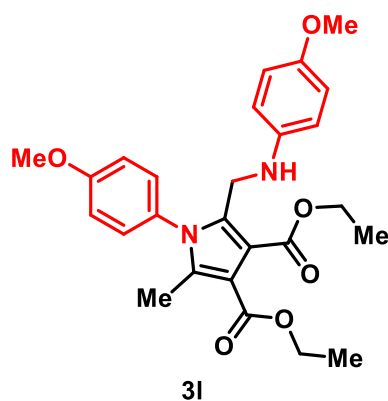


Prepared from 4-methyl aniline and ethyl acetoacetate following the general procedure to give the product as pale brown oil (89% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.26 (d, 2H, $J = 8.24$ Hz), 7.06 (d, 2H, $J = 8.24$ Hz), 6.89 (d, 2H, $J = 8.24$ Hz), 6.37 (d, 2H, $J = 8.24$ Hz), 4.30 (q, 4H, $J = 7.32$ Hz), 4.10 (s, 2H), 2.44 (s, 3H), 2.22 (s, 2H), 2.15 (s, 3H), 1.26–1.38 (t+t, 6H, $J = 7.32$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.56, 165.32, 139.44, 135.19, 134.66, 133.48, 130.18, 129.52, 127.76, 115.24, 114.70, 114.15, 112.78, 60.63, 60.26, 39.57, 21.20, 20.37, 14.29, 14.24, 11.84.

HRMS m/z (ESI): calcd. for C₂₄H₃₁N₂O₄ [M+H]⁺ 435,22783, found 435,22637.

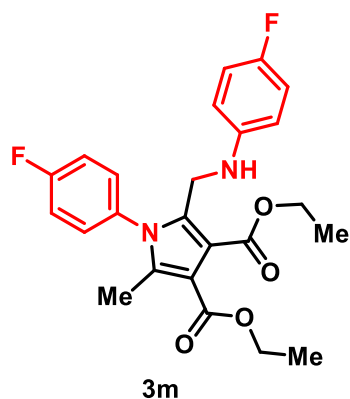


Prepared from 4-methoxy aniline and ethyl acetoacetate following the general procedure to give the product as pale yellow oil (86% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.10 (d, 2H, $J = 8.80$ Hz), 6.96 (d, 2H, $J = 8.80$ Hz), 6.69 (d, 2H, $J = 8.80$ Hz), 6.44 (d, 2H, $J = 8.80$ Hz), 4.30 (q, 4H, $J = 7.23$ Hz), 4.07 (s, 2H), 3.87 (s, 3H), 3.73 (s, 3H), 2.14 (s, 3H), 1.37–1.30 (t+t, 6H, $J = 7.23$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.56, 165.33, 159.98, 152.83, 135.33, 135.05, 129.14, 128.64, 115.70, 114.63, 114.49, 112.75, 60.61, 60.28, 55.71, 55.55, 40.44, 14.30, 14.24, 11.83.

HRMS m/z (ESI): calcd. for C₂₆H₃₁N₂O₆ [M+H]⁺ 467.21766, found 467.21656.

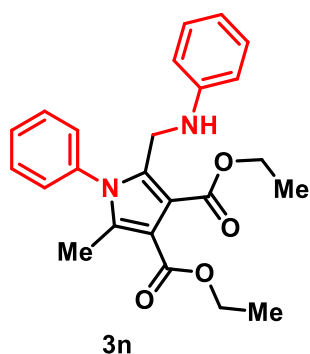


Prepared from 4-fluoroaniline and ethyl acetoacetate following the general procedure to give the product as pale yellow oil (91% yield).

¹H NMR (500 MHz, Chloroform-*d*): δ 7.22–7.16 (m, 4H), 6.81 (appt. t, 2H, $J = 8.80$ Hz), 6.38 (appt. dd, 2H, $J = 8.80, 4.40$ Hz), 4.34–4.29 (q+q, 4H, $J = 7.23$ Hz), 4.05 (s, 2H), 2.15 (s, 3H), 1.37–1.30 (t+t, 6H, $J = 7.23$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.35, 165.14, 163.74, 161.74, 157.33, 155.45, 143.51, 135.08, 134.55, 132.01, 129.95, 129.87, 116.74, 116.57, 115.66, 115.48, 115.02, 114.95, 113.21, 60.76, 60.44, 39.99, 14.30, 14.22, 11.84.

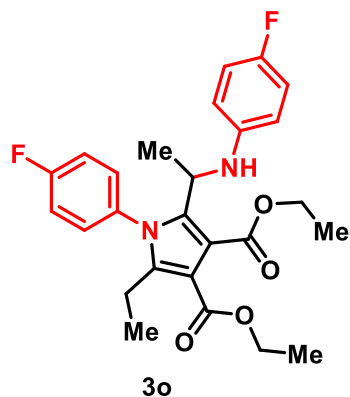
HRMS *m/z* (ESI): calcd. for C₂₄H₂₅F₂N₂O₄ [M+H]⁺ 443,17769, found 443,17607.



Prepared from aniline and ethyl acetoacetate following the general procedure to give the product as pale yellow oil (92% yield).³

¹H NMR (500 MHz, Chloroform-*d*): δ 7.51–7.47 (m, 3H), 7.22 (m, 2H), 7.08 (dd, 2H, $J = 8.17, 7.23$ Hz), 6.69 (t, 1H, $J = 7.23$ Hz), 6.41 (d, 2H, $J = 7.86$ Hz), 4.34–4.29 (q+q, 4H, $J = 7.23$ and 6.92 Hz), 4.13 (s, 2H), 2.16 (s, 3H), 1.37–1.30 (t+t, 6H, $J = 6.92$ Hz).

¹³C NMR (125 MHz, Chloroform-*d*): δ 165.48, 165.27, 147.19, 136.15, 135.06, 134.54, 129.62, 129.36, 129.04, 128.07, 118.16, 114.84, 113.77, 112.96, 60.68, 60.32, 39.10, 14.29, 14.22, 11.86.

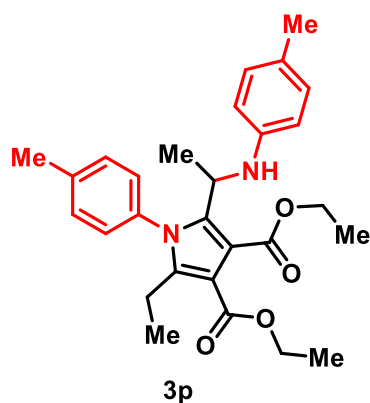


Prepared from 4-fluoroaniline and ethyl propionylacetate following the general procedure to give the product as pale yellow oil (74% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.25–7.10 (m, 4H), 6.80 (t, 2H, $J = 8.54$ Hz), 6.32 (dd, 2H, $J = 8.24, 3.66$ Hz), 4.37–4.24 (m, 5H), 2.47 (q, 2H, $J = 7.63$ Hz), 1.46 (d, 3H, $J = 6.71$ Hz), 1.34 (t+t, 6H, $J = 7.32$ Hz), 0.93 (t, 3H, $J = 7.63$ Hz).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 166.60, 165.07, 164.87, 160.88, 140.01, 138.82, 132.32, 132.26, 130.46, 130.40, 130.31, 130.27, 116.69, 116.33, 115.75, 115.40, 115.03, 112.97, 112.64, 60.90, 60.32, 47.21, 21.53, 18.76, 14.31, 14.27, 14.11.

HRMS m/z (ESI): calcd. for $\text{C}_{26}\text{H}_{29}\text{F}_2\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 471,20899, found 471,20854.

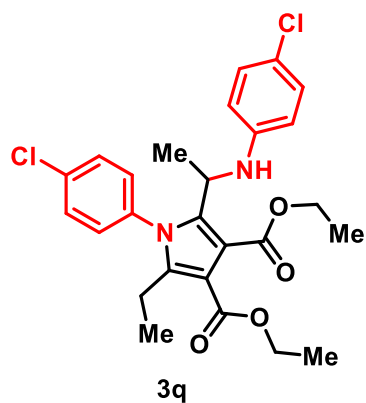


Prepared from 4-methylaniline and ethyl propionylacetate following the general procedure to give the product as pale yellow oil (70% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.30 (d, 2H, $J = 8.34$ Hz), 7.05 (m, 2H), 6.88 (d, 2H, $J = 7.93$ Hz), 6.32 (d, 2H, $J = 7.32$ Hz), 4.39–4.17 (m, 5H), 2.53–2.44 (s+m, 5H), 2.21 (s, 3H), 1.43 (d, 3H, $J = 6.71$ Hz), 1.32 (t+t, 6H, $J = 7.02$ Hz), 0.93 (t, 3H, $J = 7.32$ Hz).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 166.77, 165.29, 140.02, 139.72, 133.71, 130.03, 129.96, 129.53, 128.40, 128.30, 113.89, 112.53, 112.20, 60.71, 60.13, 46.29, 21.68, 21.27, 20.36, 18.77, 14.36, 14.27, 14.12.

HRMS m/z (ESI): calcd. for $\text{C}_{28}\text{H}_{35}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 463,25913, found 463,25848.



Prepared from 4-chloroaniline and ethyl propionylacetate following the general procedure to give the product as orange yellow oil (76% yield).

$^1\text{H NMR}$ (500 MHz, Chloroform-*d*): δ 7.54–7.47 (m, 2H), 7.19–7.02 (m, 4H), 6.26 (d, 2H, $J = 8.85$ Hz), 4.38–4.24 (m, 5H), 2.47 (q, 2H, $J = 7.32$ Hz), 1.42 (d, 3H, $J = 6.71$ Hz), 1.33 (t+t, 6H, $J = 7.02$ Hz), 0.93 (t, 3H, $J = 7.32$ Hz).

$^{13}\text{C NMR}$ (125 MHz, Chloroform-*d*): δ 166.47, 165.00, 145.15, 139.92, 138.51, 135.86, 134.89, 129.87, 129.83, 129.72, 128.99, 122.36, 114.67, 113.17, 112.76, 60.92, 60.33, 46.21, 21.61, 18.73, 14.34, 14.25, 14.11.

HRMS m/z (ESI): calcd. for $\text{C}_{26}\text{H}_{29}\text{Cl}_2\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 503,14989, found 503,14910.

13. Spectral data

