

Supporting Information

Bromo- and Glycosyl-Substituted BODIPYs for an Application in Photodynamic Therapy and

Imaging

Benjamin F. Hohlfeld, Dorika Steen, Gerhard D. Wieland, Katharina Achazi, Nora Kulak, Rainer Haag, and Arno Wiehe*

Table of Contents

S1 General remarks	S2
S2 Cellular testing	S2
S2.1 Cellular assays	S2
S2.2 Cellular tests of core-unsubstituted BODIPYs against A-253 and HT-29.....	S4
S2.3 Cellular tests of 1,3,5,7-tetramethyl-unsubstituted BODIPYs against A-253 and HT-29	S5
S2.4 Cellular tests of 2,6-dibromo-unsubstituted BODIPYs against A-253 and HT-29.....	S6
S2.5 Cellular tests of 2-bromo-substituted BODIPYs against A-253 and HT-29	S7
S2.6 Overview on the phototoxic activity of the four series of BODIPYs regarding the different substitutions	S8
S3 Confocal laser scanning microscopy (cLSM) for cellular uptake analysis	S9
S4 Nucleophilic substitution of BODIPY 31 with amines (51 – 53)	S9
S4.1 General synthetic procedure	S9
S4.2 8-[3-Nitro-4-(<i>N</i> -2-prop-2-enylamino)phenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a- diazas-indacene (51):	S9
S4.3 8-[4-(<i>N</i> -6-Methoxy-6-oxohexylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora- 3a,4a-diazas-indacene (52):	S15
S4.4 8-[4-(<i>N,N</i> -Dibutylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diazas- indacene (53):	S20
S5 Bromination of tetramethyl-BODIPYs (54 – 55)	S25
S5.1 General synthetic procedure	S25
S5.2 2,6-Dibromo-8-[3-nitro-4-(<i>N</i> -2-prop-2-enylamino)phenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4- bora-3a,4a-diazas-indacene (54):	S25
S5.3 2,6-Dibromo-8-[4-(<i>N</i> -6-methoxy-6-oxohexylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4- difluoro-4-bora-3a,4a-diazas-indacene (55):	S30
S5.4 2,6-Dibromo-8-[4-(<i>N,N</i> -dibutylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora- 3a,4a-diazas-indacene (56):	S35
S6 References	S40

S1 General remarks

All reactions were performed in standard round bottom flasks. DCM, HFIP, and methanol were purchased and used as received. Other solvents were purchased and distilled at reduced pressure. Purchased chemicals were used as received without further purification. All liquid reagents were added through syringes. Reactions were monitored by thin-layer chromatography (Merck, TLC Silica gel 60 F₂₅₄) and visualized under UV light (254 nm and 366 nm). Flash column chromatography was performed on silica gel (Fluka silica gel 60M, 40-63µm). NMR spectra were recorded with JEOL ECX400 and JEOL ECP500 Instruments. Multiplicity of the signals was assigned as follows: s = singlet, br s = broad singlet, d = doublet, t = triplet, dd = doublet of doublets, ddd = doublet of doublets of doublets, ddt = doublet of doublets of triplets, m = multiplet, m_c = centered multiplet. Chemical shifts are reported relative to CDCl₃ (¹H: δ = 7.26 ppm, ¹³C: δ = 77.2 ppm) and THF-d₈ (¹H: δ = 3.58 ppm, ¹³C: δ = 67.6 ppm). All ¹³C NMR spectra are proton-decoupled and coupling constants are given in hertz (Hz). For a detailed peak assignment 2D spectra were measured (COSY, HMBC, and HMQC). HRMS analyses were carried out on an Agilent Technologies 6210 ESI-TOF (electrospray ionization, time of flight) instrument. IR spectra were measured with a JASCO FT/IR 4100 spectrometer equipped with a PIKE MIRacle™ ATR instrument. UV/Vis spectra were recorded on a SPECORD S300 UV/Vis spectrometer (Analytik Jena) in quartz cuvettes (1 cm length). The fluorescence spectra of the BODIPYs were recorded with a JASCO FP 6500 spectrofluorometer in quartz cuvettes (1 cm length). Specified melting points were recorded on a Reichert Thermovar Apparatus and are not corrected.

Core-unsubstituted BODIPYs **1** – **16**^[1], 1,3,5,7-tetramethyl-substituted BODIPYs **17** – **29**^[2,3], 2-bromo-substituted BODIPYs **30** – **37**^[3], and 2,6-dibromo-substituted BODIPYs **38** – **50**^[3] were prepared according to the literature.

S2 Cellular testing

S2.1 Cellular assays

All cell lines used were purchased from DSMZ, except for the A-253 cell line which was purchased from ATCC. Human epidermoid carcinoma (A-431), human epithelial tongue squamous cell carcinoma (CAL-27), human colorectal adenocarcinoma (HT-29), and submaxillary salivary gland epidermoid carcinoma (A-253) were grown in Dulbecco's modified eagle medium (DMEM) with 10% heat inactivated FCS, 1% penicillin (10,000 IU) and

streptomycin (10,000 $\mu\text{g mL}^{-1}$). A stock solution (2 mM) of the BODIPY was prepared in DMSO and kept in the dark at 4 °C. DMEM (without phenol red) with 10% fetal calf serum (FCS) was used for further dilution to reach a concentration of 2 or 10 μM of the BODIPY, respectively. In microplates 2×10^4 cells per well were seeded in fresh medium (DMEM without phenol red containing 10% FCS) with 2 μM or 10 μM solution of the BODIPY and incubated for 24 h. After an exchange of medium (to remove any BODIPY not taken up by the cells), the irradiation was performed at rt with a white light source (KI 2500 LCD, Schott) at an energy dose of approximately 50 J cm^{-2} . The absorbance was measured at a wavelength of 490 nm.

S2.2 Cellular tests of core-substituted BODIPYs

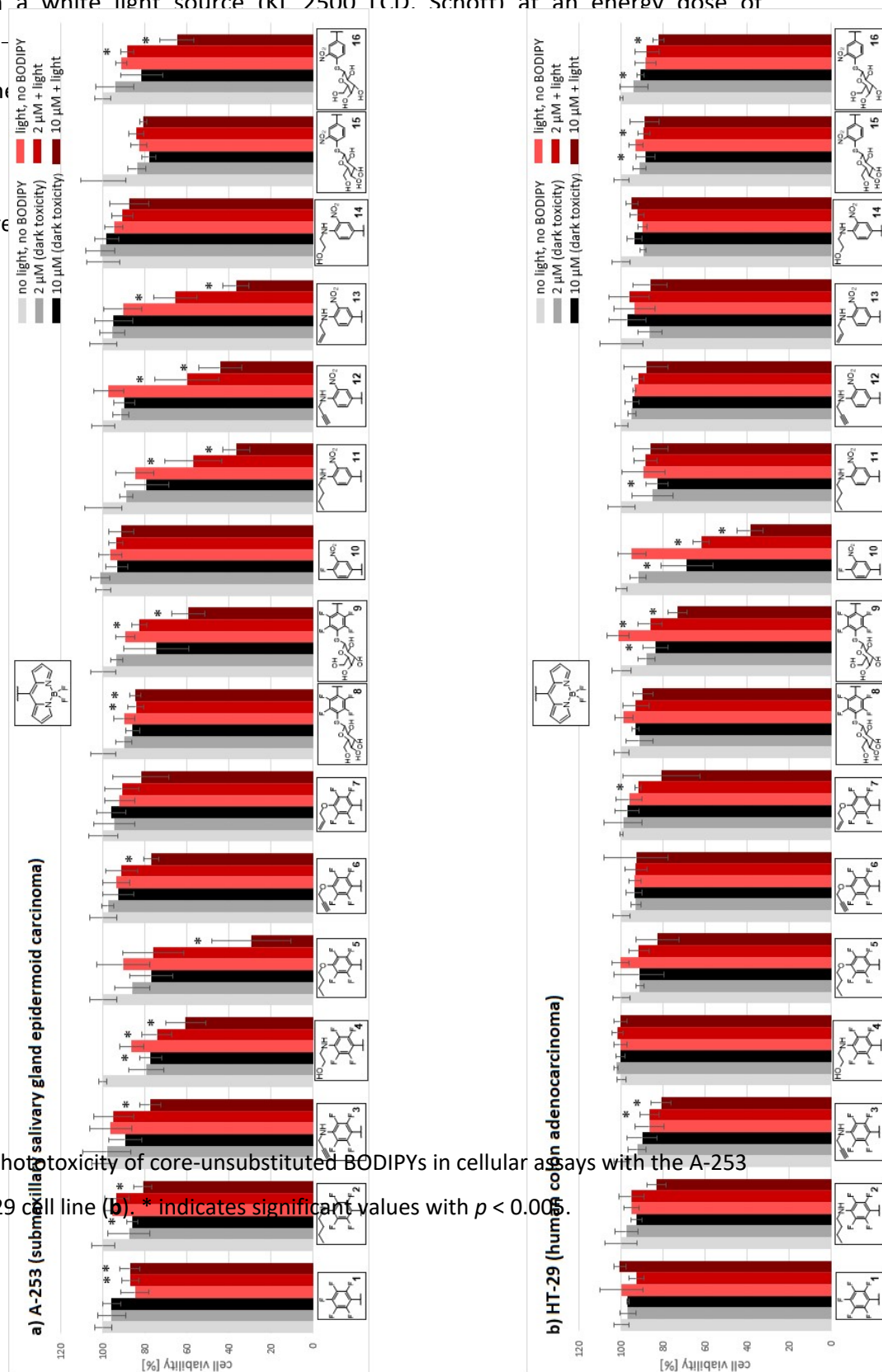


Figure S2.2: Dark- and phototoxicity of core-substituted BODIPYs in cellular assays with the A-253 cell line (a) and the HT-29 cell line (b). * indicates significant values with $p < 0.001$.

S2.3 Cellular tests of 1,3,5,7-tetramethyl-substituted BODIPYs

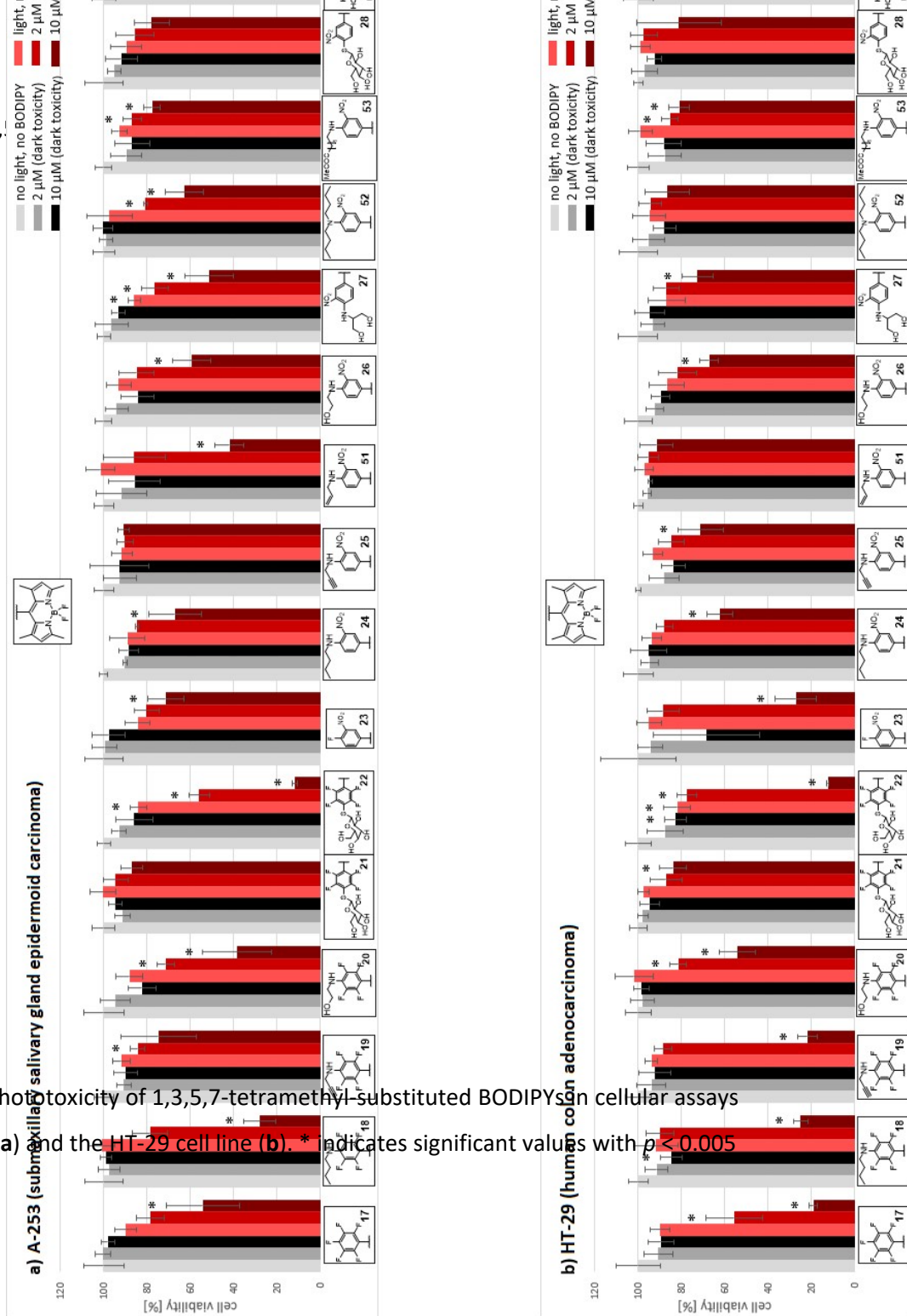


Figure S2.3: Dark- and phototoxicity of 1,3,5,7-tetramethyl-substituted BODIPYs on cellular assays with the A-253 cell line (a) and the HT-29 cell line (b). * indicates significant values with $p < 0.005$

S2.4 Cellular tests of 2,6-

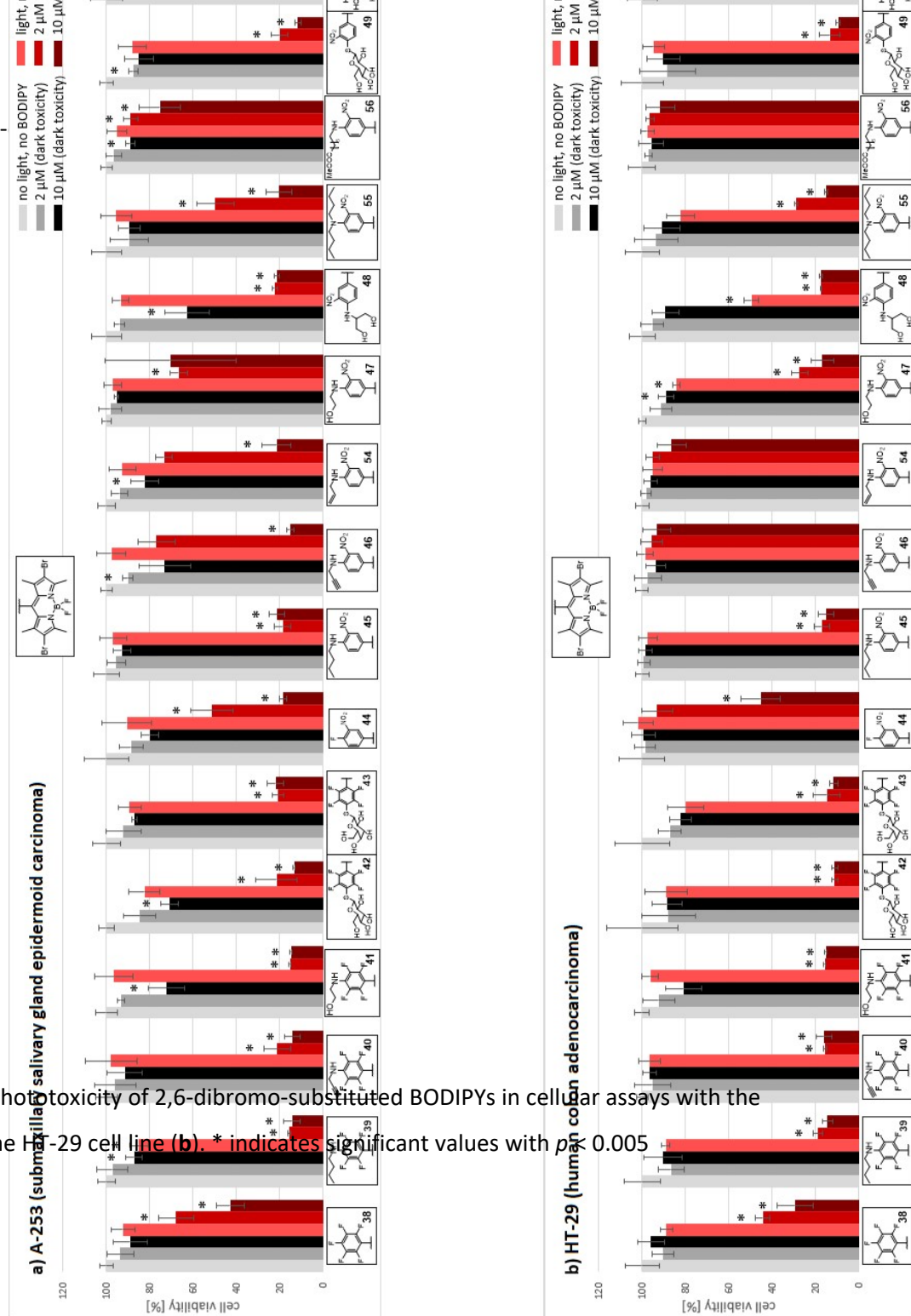


Figure S2.4: Dark- and phototoxicity of 2,6-dibromo-substituted BODIPYs in cellular assays with the A-253 cell line (a) and the HT-29 cell line (b). * indicates significant values with $p < 0.005$.

S2.5 Cellular tests of 2-bromo-substituted BODIPYs against A-253 and HT-29

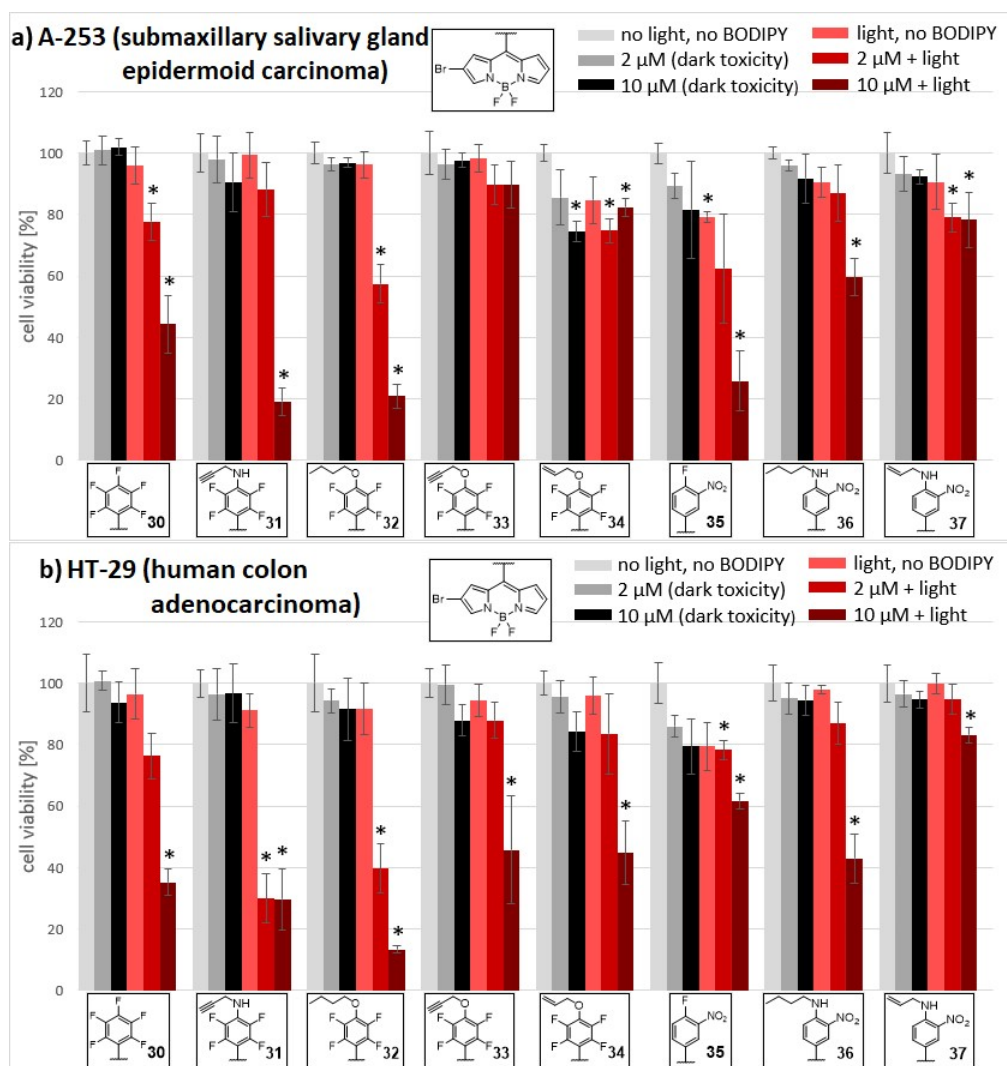


Figure S2.5: Dark- and phototoxicity of 2-bromo-substituted BODIPYs in cellular assays with the A-253 cell line (a) and the HT-29 cell line (b). * indicates significant values with $p < 0.005$.

S2.6 Overview on the phototoxic activity of the four series of BODIPYs regarding the different substitutions. *Highlighted compounds showed a cell viability reduction of > 40 % at 10 μ M in at least two cell lines under irradiation (X indicates structural combinations that have not been synthesized).*

Aryl substituent at the 8-position	BODIPY	1,3,5,7-Tetramethyl-BODIPY	2,6-Dibromo-1,3,5,7-tetramethyl-BODIPY	2-Bromo-BODIPY
	1	17	38	30
	2	18	39	X
	3	19	40	31
	4	20	41	X
	5	X	X	32
	6	X	X	33
	7	X	X	34
	8	21	42	X
	9	22	43	X
	10	23	44	35
	11	24	45	36
	12	51	54	X
	13	25	46	37
	14	26	47	X
	X	27	48	X
	X	52	55	X
	X	53	56	X
	15	28	49	X
	16	29	50	X

S3 Confocal laser scanning microscopy (cLSM) for cellular uptake analysis

The uptake of the BODIPYs in A-431 cells was analyzed after 4 h and 24 h by confocal laser scanning microscopy (cLSM). A-431 cells were routinely cultivated as described above (S2.1). For the uptake study, 270 μL of a A-431 cell suspension in DMEM were seeded in each well of an 8-well ibidi μ -slide (50,000 cells mL^{-1}). After 1 day, 30 μL of selected BODIPYs were added at final test concentration of 10 μM and incubated for 4 h and 24 h. After 24 h cell nuclei were stained with 1 $\mu\text{g mL}^{-1}$ Hoechst 33342 (Life Technologies GmbH, Darmstadt, Germany). Then confocal images were taken by using an inverted confocal laser scanning microscope Leica DMI6000CSB SP8 (Leica, Wetzlar, Germany) with a 63x/1.4 HC PL APO CS2 oil immersion objective and the LAS X software.

S4 Nucleophilic substitution of BODIPY 31 with amines (51 – 53)

S4.1 General synthetic procedure

BODIPY **23** (1 equiv.) was dissolved in DCM and the corresponding amine (10 – 20 equiv.) was added. The mixture was stirred for 24 h. Afterwards, the mixture was diluted with EtOAc and washed with water several times. The organic layer was dried with Na_2SO_4 , filtered, and evaporated to dryness. The crude product was purified by column chromatography; the main fraction was collected and evaporated to dryness.

S4.2 8-[3-Nitro-4-(*N*-2-prop-2-enylamino)phenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene (**51**):

BODIPY **51** was prepared according to the general synthetic procedure. BODIPY **23** (132 mg, 0.34 mmol) and allylamine (0.3 mL, 3.40 mmol, 10 equiv.) were dissolved in 10 mL of DCM. After column purification (chromatography, silica gel, *n*-hexane/EtOAc = 9/1, v/v); BODIPY **51** was obtained as an orange solid (124 mg, 0.29 mmol, 86%).

$^1\text{H NMR}$ (500 MHz, CDCl_3): δ (ppm) = 1.53 (s, 6H, Me), 2.54 (s, 6H, Me), 4.05 (t, J = 5.0 Hz, 2H, CH_2), 5.29–5.36 (m, 2H, $\text{H}_2\text{C}=\text{CH}$ -), 5.95–6.02 (m, 3H, $\text{H}_{\text{pyrrole}}$ + $\text{H}_2\text{C}=\text{CH}$ -), 6.98 (d, J = 8.8 Hz, 1H, Ar- H_{meta}), 7.30 (dd, J = 8.8, 2.2 Hz, 1H, Ar- H_{ortho}), 8.14 (d, J = 2.1 Hz, 1H, Ar- H_{ortho}), 8.28 (t, J = 5.8 Hz, 1H, NH).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3): δ (ppm) = 14.7 (Me), 15.3 (Me), 45.6 (CH_2), 115.2 (Ar- C_{meta}), 117.7 ($\text{H}_2\text{C}=\text{CH}$), 121.6 ($\text{CH}_{\text{pyrrole}}$), 121.8*, 126.8 (Ar- C_{ortho}), 131.9 ($\text{C}_{\text{pyrrole}}$), 132.1 (C_{meso}), 132.7 ($\text{H}_2\text{C}=\text{CH}$ -), 136.0 (Ar- C_{ortho}),

139.2*, 142.7 (C_{pyrrole}), 145.3 (Ar-C_{para}), 156.1 (C_{pyrrole}). *These signals could not be assigned exactly to corresponding carbon atoms. They belong to the Ar-C_{ipso} and the Ar-C_{nitro} of the aryl moiety.

¹⁹F NMR (376 MHz, CDCl₃): δ (ppm) = -145.57 – -146.59 (m_c, 2F, BF₂).

HRMS (ESI-TOF, MeOH): m/z calcd. for C₂₂H₂₃BF₂N₄O₂Na⁺ [M+Na]⁺: 447.1774, found: 447.1796, m/z calcd. for C₂₂H₂₃BF₂N₄O₂K⁺ [M+K]⁺: 463.1514, found: 463.1543, m/z calcd. for C₄₄H₄₆B₂F₄N₈O₄Na⁺ [2M+Na]⁺: 871.3656, found: 871.3699, m/z calcd. for C₄₄H₄₆B₂F₄N₈O₄K⁺ [2M+K]⁺: 887.3396, found: 887.3442.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 3590 [ν (NH)], 2920 [ν (CH₂)], 1628 [ν (C=N), ν (C=C)], 1543 [ν_{as} (NO₂)], 1527 [δ (Ar-C)], 1466 [δ (CH₂), δ (Me)], 1349 [ν_{sym} (NO₂)], 1061 [ν (BF)], 758 [δ (HC=CH)].

UV/vis (DCM): λ_{max} (nm) [log (ϵ /L mol⁻¹ cm⁻¹)] = 504 [4.83].

Fluorescence (DCM): λ_{max} (nm) = 517 at $\lambda_{excitation}$ (nm) 490.

M.P. (°C): 176-179.

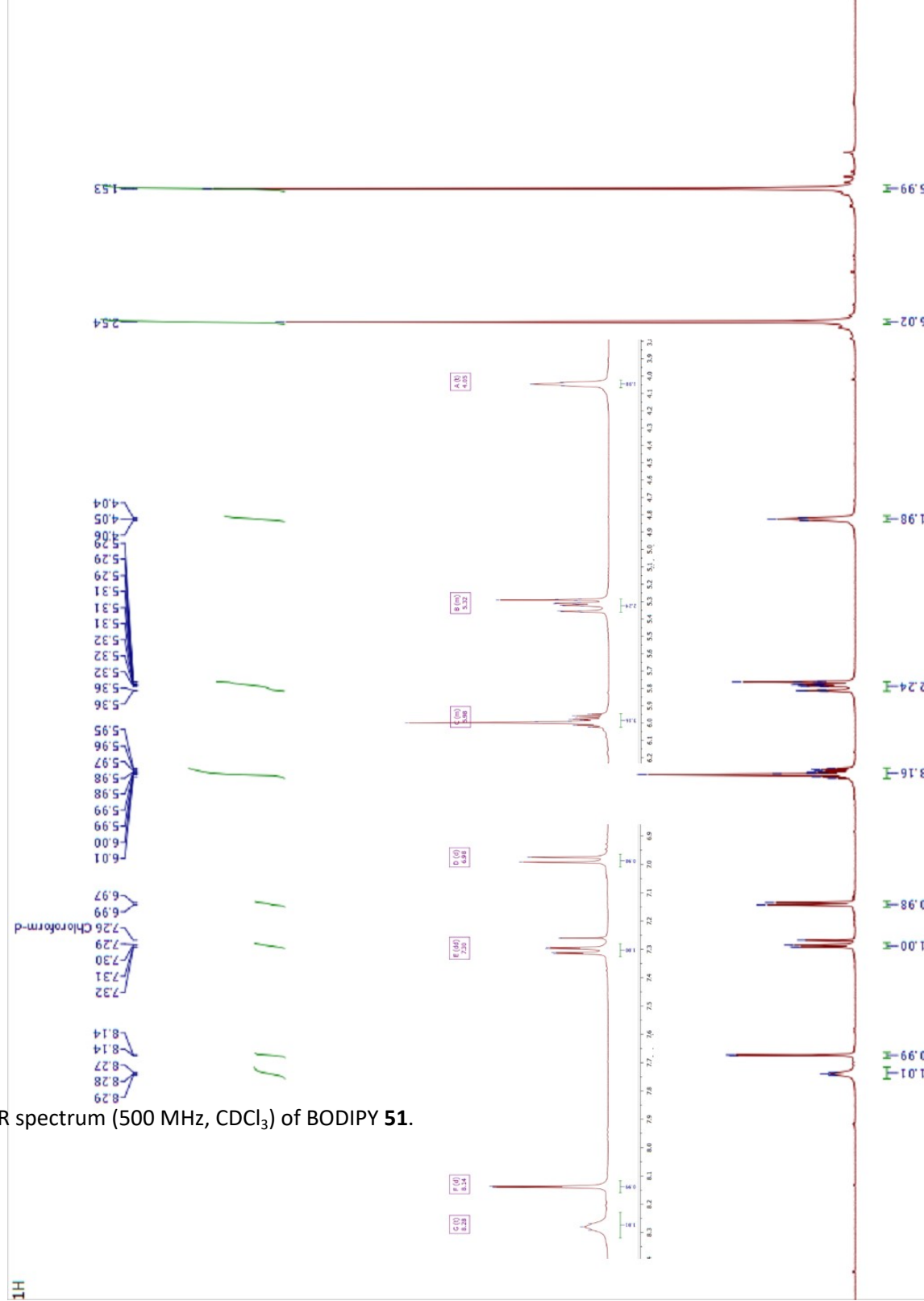
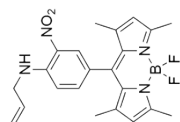


Figure S4.2.1: ^1H NMR spectrum (500 MHz, CDCl_3) of BODIPY 51.

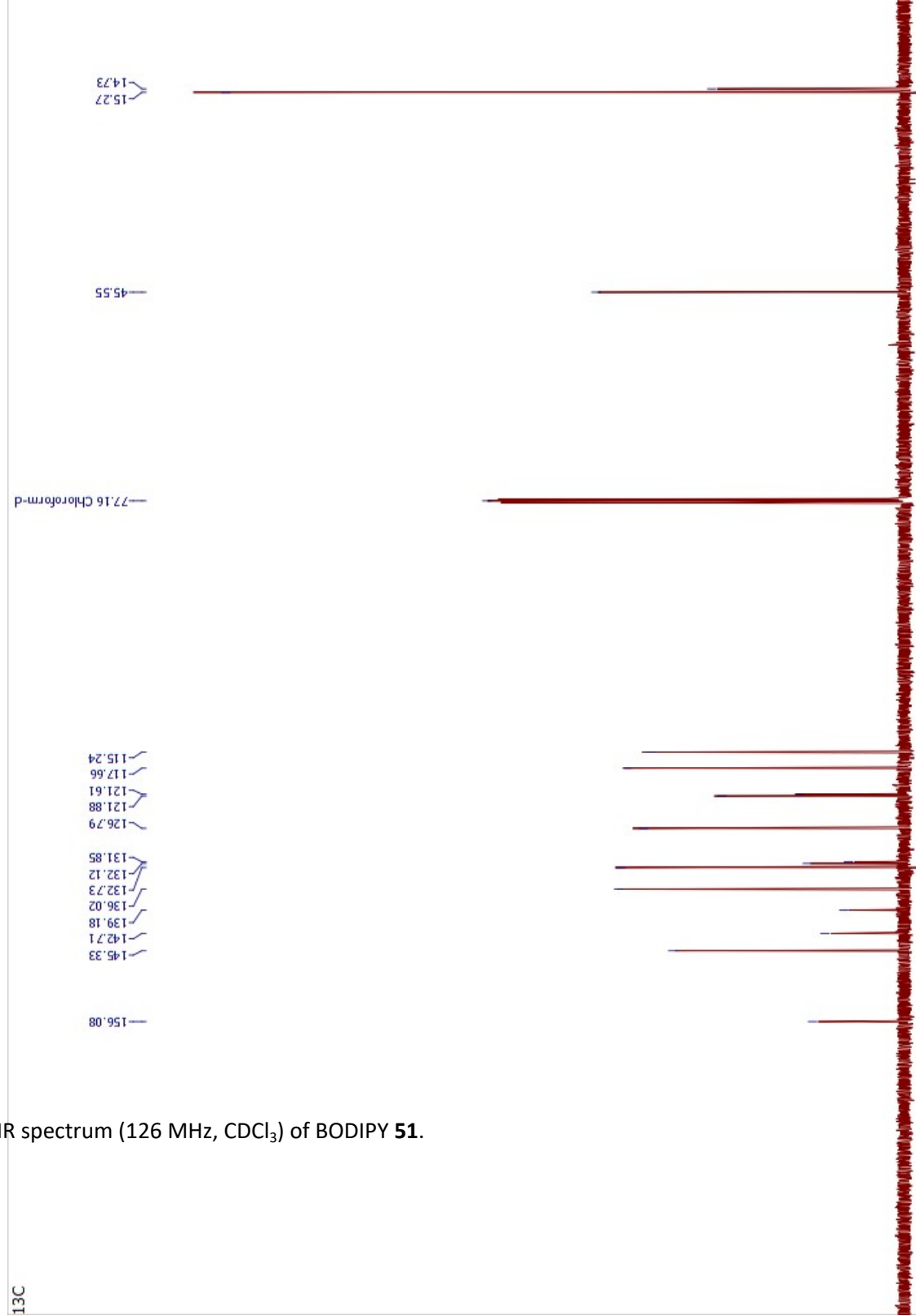
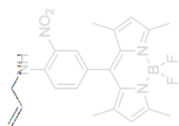


Figure S4.2.2: ^{13}C NMR spectrum (126 MHz, CDCl_3) of BODIPY 51.



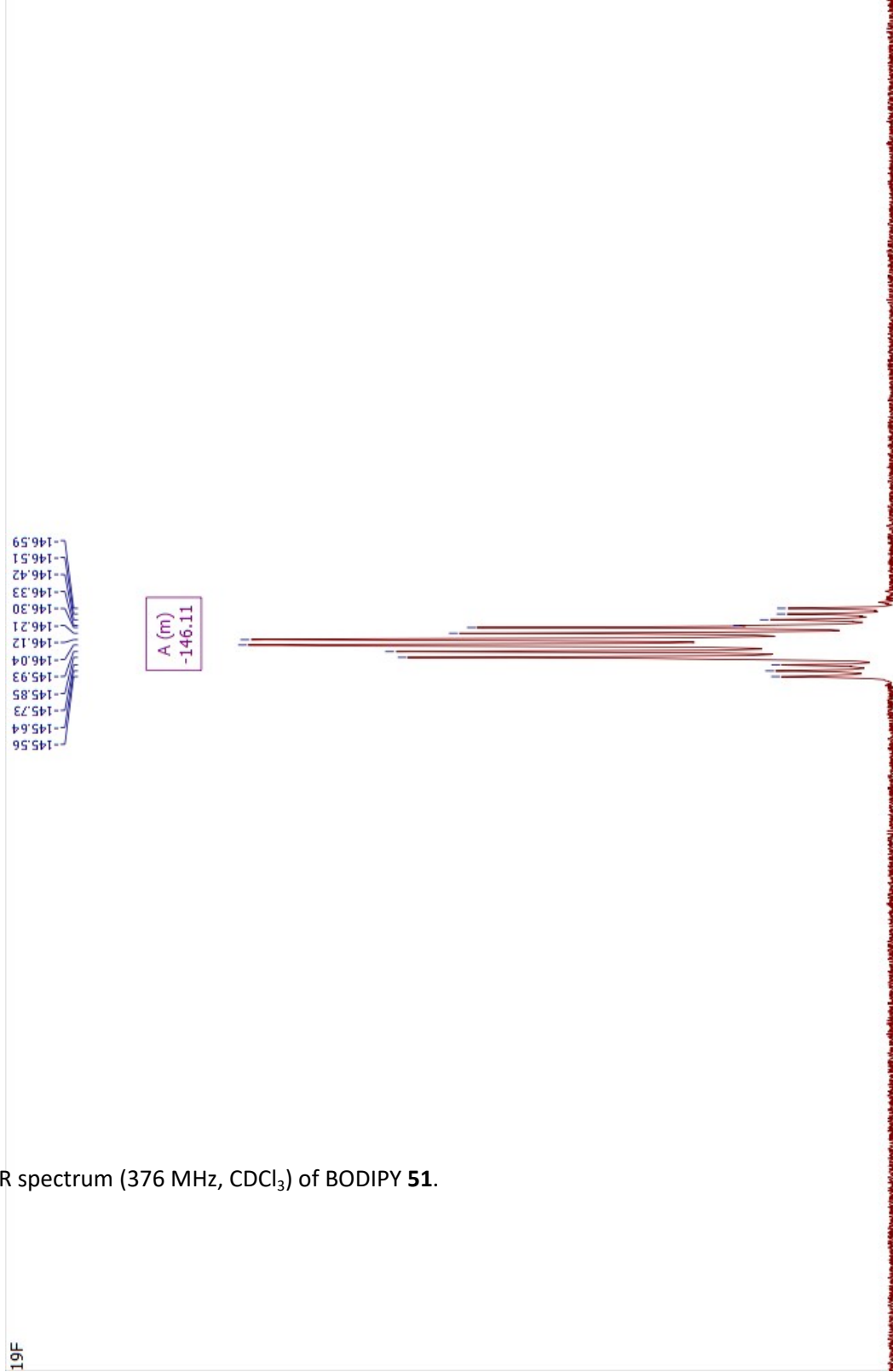
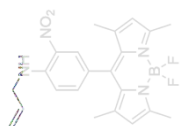


Figure S4.2.3: ^{19}F NMR spectrum (376 MHz, CDCl_3) of BODIPY 51.



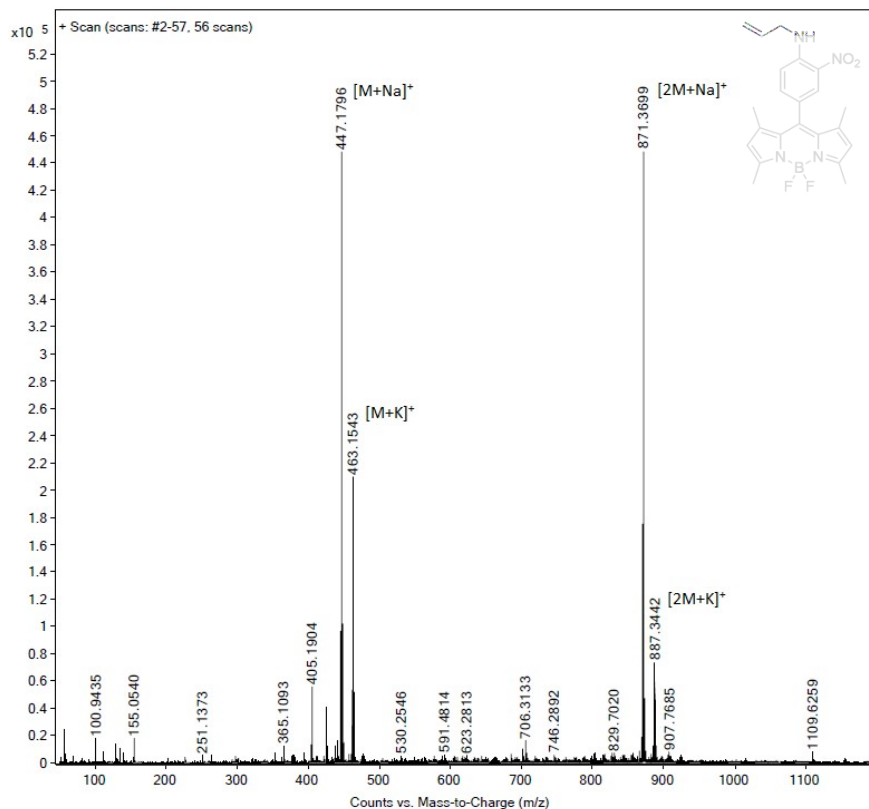


Figure S4.2.4: HRMS spectrum (ESI+) of BODIPY 51.

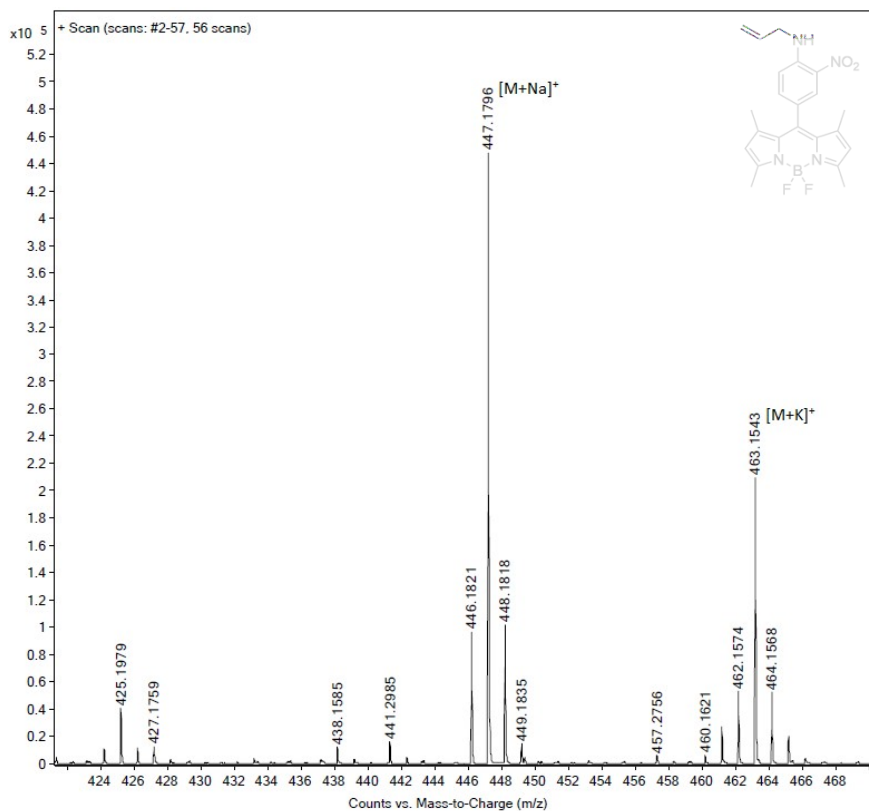


Figure S4.2.5: HRMS spectrum (ESI+) of BODIPY 51.

S4.3 8-[4-(*N*-6-Methoxy-6-oxohexylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene (**52**):

BODIPY **52** was prepared according to the general synthetic procedure. BODIPY **23** (200 mg, 0.52 mmol), *N,N*-diisopropylethylamine (1.8 mL, 10.33 mmol, 20 equiv.), and methyl-6-aminohexanoate hydrochloride (939 mg, 5.17 mmol, 10 equiv.) were dissolved in 20 mL of DCM. After purification (column chromatography, silica gel, *n*-hexane/EtOAc = 9/1, v/v, then *n*-hexane/EtOAc = 4/1, v/v); BODIPY **52** was obtained as an orange solid (80 mg, 0.16 mmol, 30%).

¹H NMR (500 MHz, CDCl₃): δ (ppm) = 1.49–1.57 (m, 8H, Me + CH₂), 1.70–1.76 (m, 2H, CH₂), 1.77–1.83 (m, 2H, CH₂), 2.36 (t, *J* = 7.4 Hz, 2H, CH₂), 2.54 (s, 6H, Me), 3.36 (td, *J* = 7.1, 5.0 Hz, 2H, CH₂), 3.67 (s, 3H, Me_{COOMe}), 5.99 (s, 2H, H_{pyrrole}), 6.98 (d, *J* = 8.8 Hz, 1H, Ar-H_{meta}), 7.31 (dd, *J* = 8.7, 2.2 Hz, 1H, Ar-H_{ortho}), 8.11–8.14 (m, 2H, Ar-H_{ortho} + NH).

¹³C NMR (126 MHz, CDCl₃): δ (ppm) = 14.7 (Me), 15.3 (Me), 24.6 (CH₂), 26.7 (CH₂), 28.7 (CH₂), 33.9 (CH₂), 43.1 (CH₂), 51.7 (Me_{COOMe}), 114.8 (Ar-C_{meta}), 121.5*, 121.6 (CH_{pyrrole}), 126.9 (Ar-C_{ortho}), 131.9 (C_{pyrrole}), 131.9 (C_{meso}), 136.1 (Ar-C_{ortho}), 139.3*, 142.7 (C_{pyrrole}), 145.5 (Ar-C_{para}), 156.1 (C_{pyrrole}), 173.95 (CO). *These signals could not be assigned exactly to corresponding carbon atoms. They belong to the Ar-C_{ipso} and the Ar-C_{nitro} of the aryl moiety.

¹⁹F NMR (376 MHz, CDCl₃): δ (ppm) = -145.57 – -146.63 (m_c, 2F, BF₂).

HRMS (ESI-TOF, MeOH): *m/z* calcd. for C₂₆H₃₁BFN₄O₄⁺ [M-F]⁺: 493.2417, found: 493.2410, *m/z* calcd. for C₂₆H₃₁BF₂N₄O₄Na⁺ [M+Na]⁺: 535.2299, found: 535.2298.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 3377 [ν (NH)], 2929 [ν (CH₂), ν (Me)], 2858 [ν (CH)], 1734 [ν (COOMe)], 1626 [ν (C=N), ν (C=C)], 1542 [ν_{as} (NO₂)], 1526 [δ (Ar-C)], 1468 [δ (CH₂), δ (Me)], 1304 [ν_{sym} (NO₂)], 1075 [ν (BF)], 758 [δ (HC=CH)]. **UV/vis (DCM):** λ_{max} (nm) [log (ϵ /L mol⁻¹ cm⁻¹)] = 504 [4.66].

Fluorescence (DCM): λ_{max} (nm) = 518 at $\lambda_{excitation}$ (nm) 490.

M.P. (°C): 123-125.

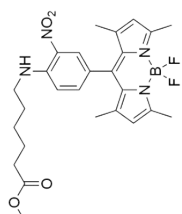
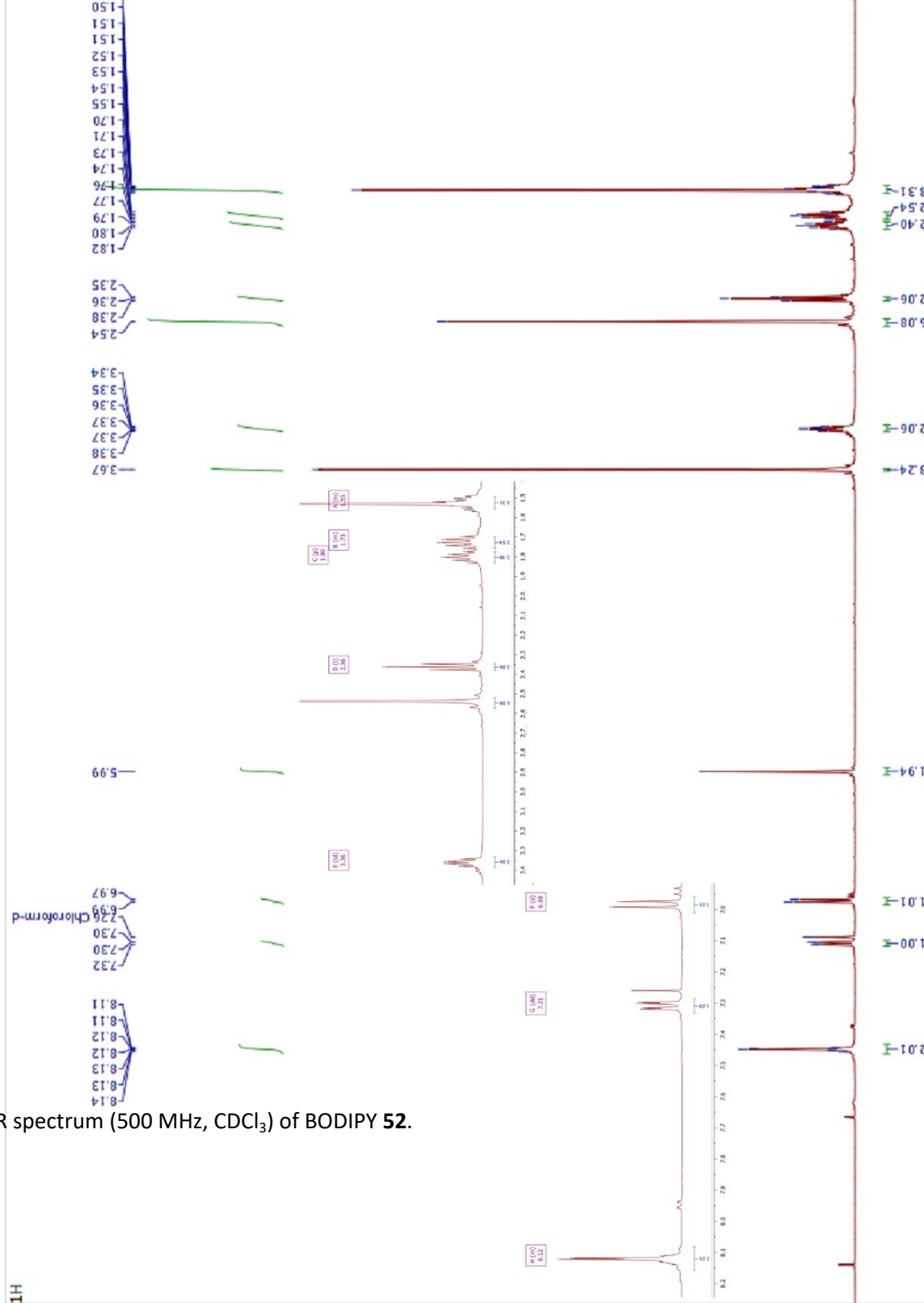
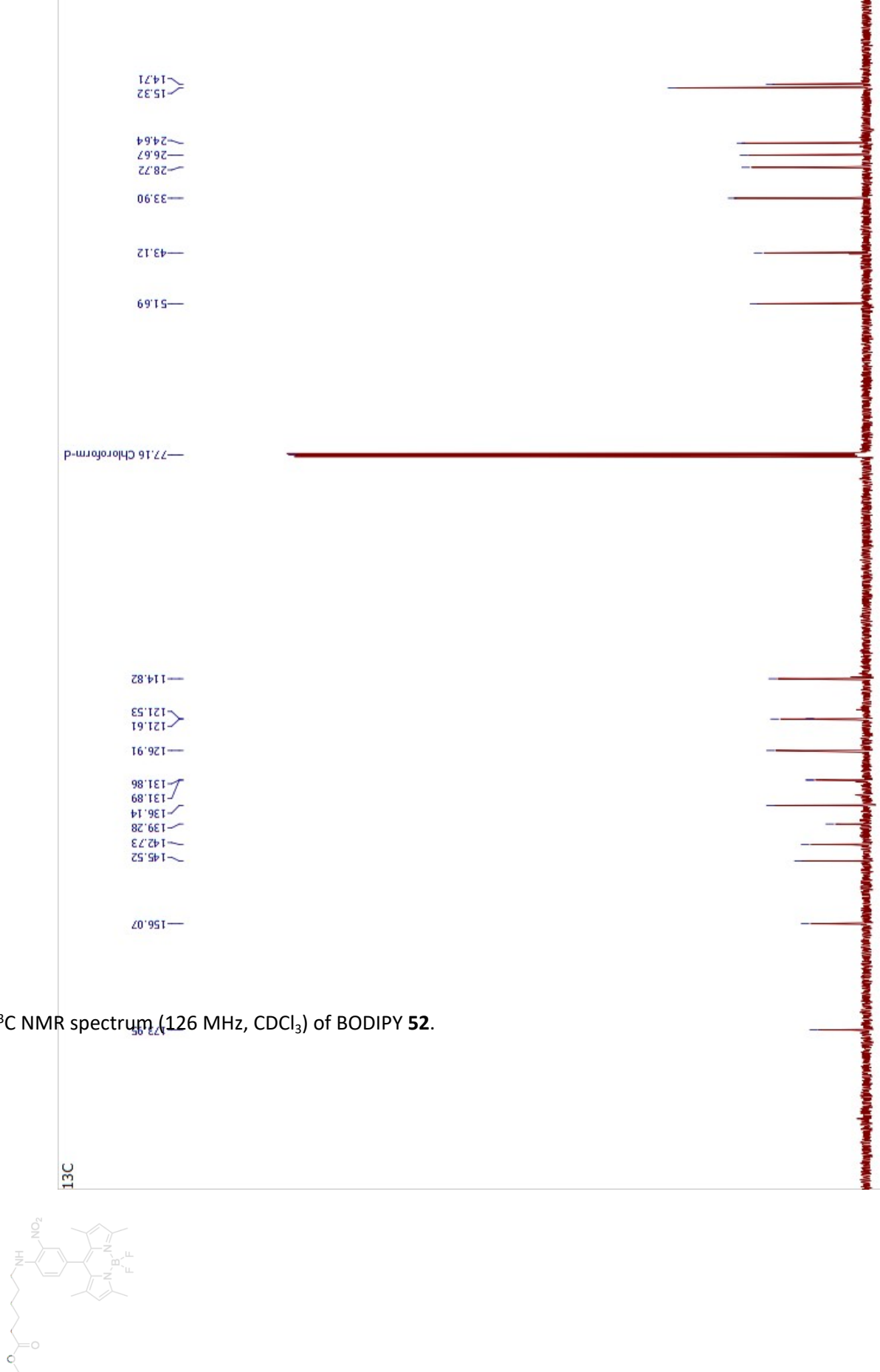


Figure S4.3.1: ^1H NMR spectrum (500 MHz, CDCl_3) of BODIPY 52.





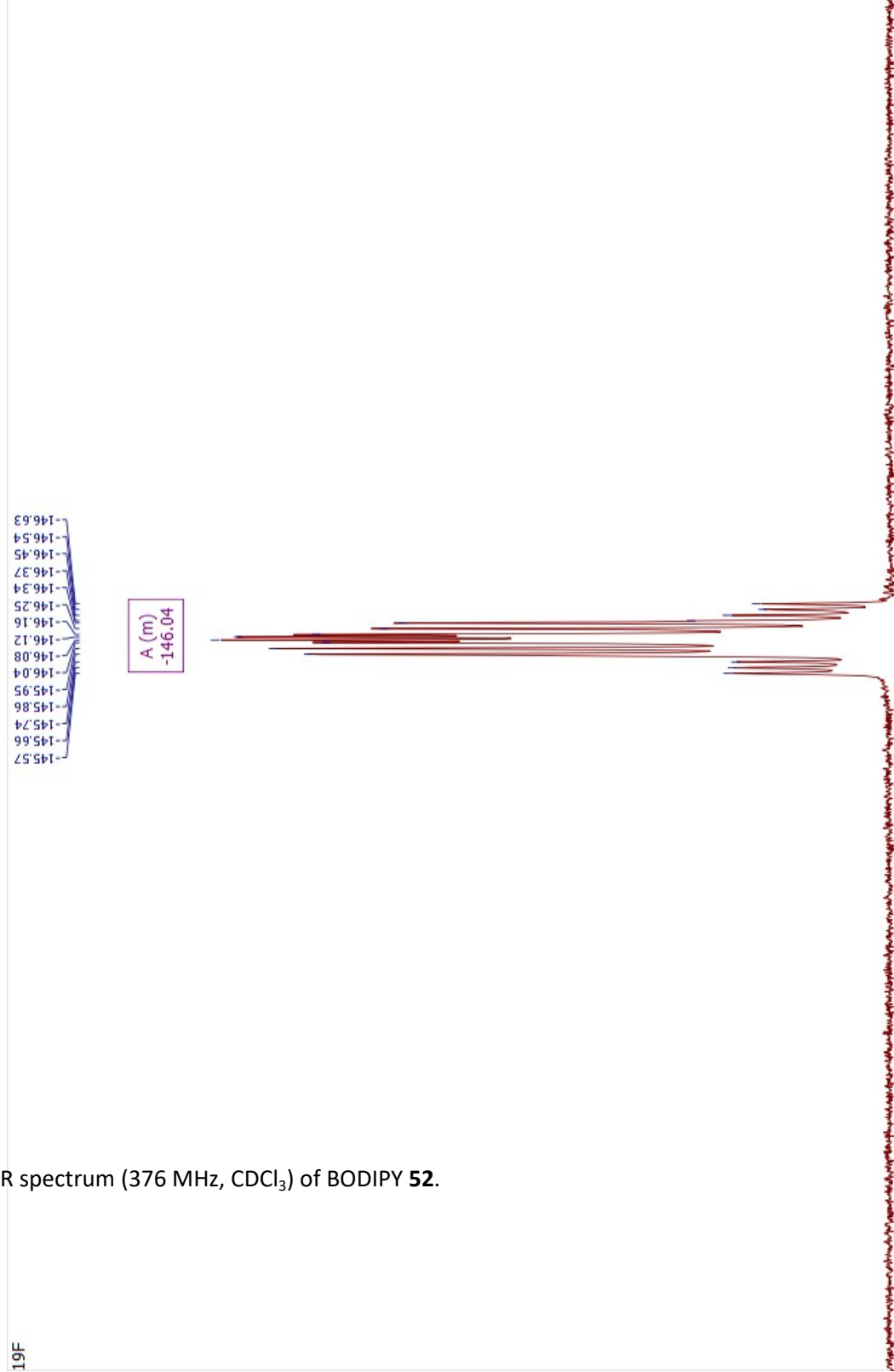
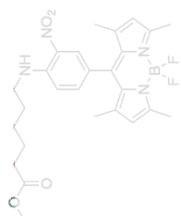


Figure S4.3.3: ^{19}F NMR spectrum (376 MHz, CDCl_3) of BODIPY 52.

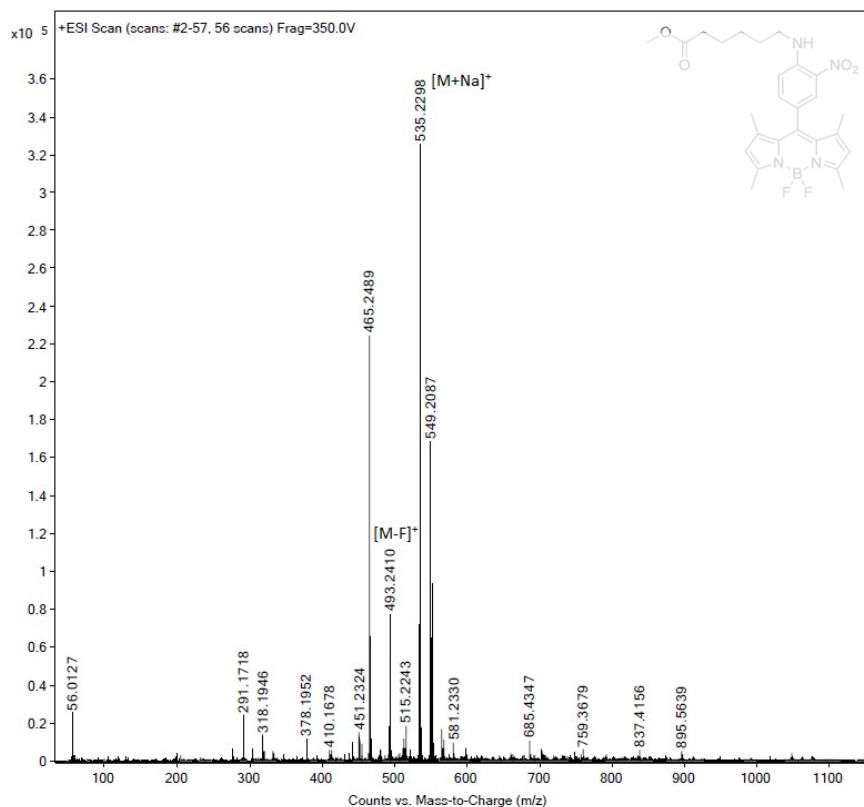


Figure S4.3.4: HRMS spectrum (ESI+) of BODIPY 52.

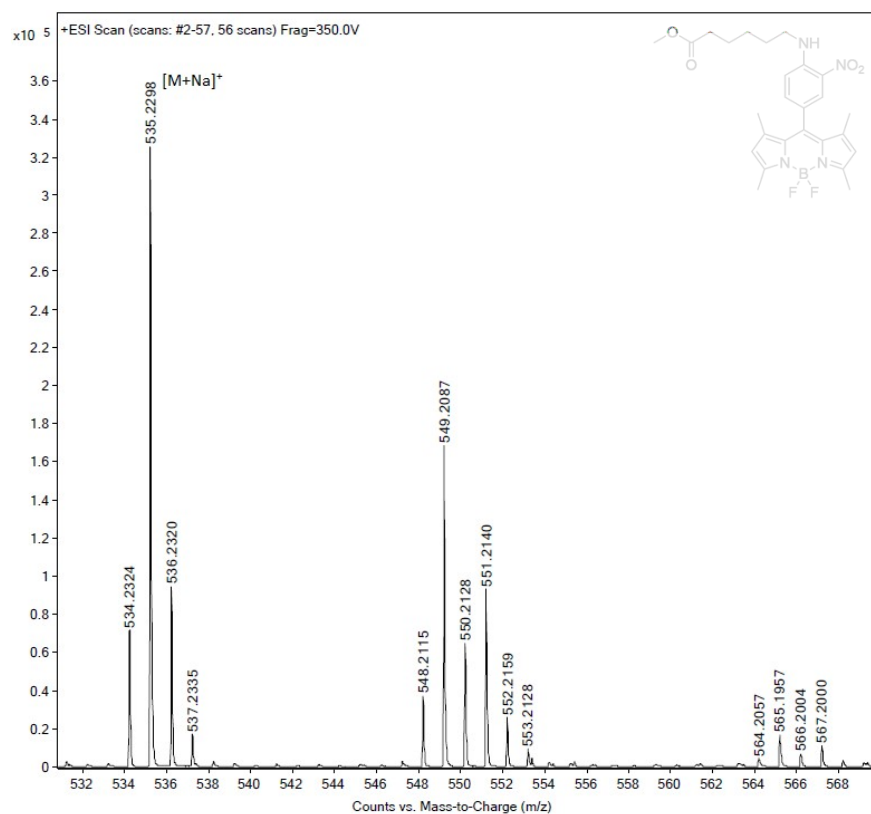


Figure S4.3.5: HRMS spectrum (ESI+) of BODIPY 52.

S4.4 8-[4-(*N,N*-Dibutylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene (**53**):

BODIPY **53** was prepared according to the general synthetic procedure. BODIPY **23** (200 mg, 0.52 mmol) and *N,N*-dibutylamine (1.60 mL, 9.66 mmol, 15 equiv.) were dissolved in 10 mL of DCM. After column purification (chromatography, silica gel, *n*-hexane/EtOAc = 9/1, v/v); BODIPY **53** was obtained as an orange solid (90 mg, 0.18 mmol, 35%).

¹H NMR (500 MHz, CDCl₃): δ (ppm) = 0.87 (t, J = 7.4 Hz, 6H, Me_{Butyl}), 1.25–1.32 (m, 4H, CH₂), 1.50–1.56 (m, 10H, CH₂ + Me), 2.55 (s, 6H, Me), 3.20 (t, J = 7.3 Hz, 1H, CH₂), 6.00 (s, 2H, H_{Pyrrrole}), 7.28 (br s, 2H, Ar-H_{meta} + Ar-H_{ortho}), 7.65 (t, J = 1.2 Hz, 1H, Ar-H_{ortho}).

¹³C NMR (126 MHz, CDCl₃): δ (ppm) = 13.9 (Me_{Butyl}), 14.8 (Me), 14.99 (Me), 20.2 (CH₂), 29.5 (CH₂), 52.2 (CH₂), 121.7 (CH_{Pyrrrole}), 122.6 (Ar-C_{meta}), 126.3 (Ar-C_{ortho}), 131.7 (C_{Pyrrrole}), 132.7 (Ar-C_{ortho}), 138.96*, 142.1*, 142.8 (C_{Pyrrrole}), 144.9 (Ar-C_{para}), 156.2 (C_{Pyrrrole}). *These signals could not be assigned exactly to corresponding carbon atoms. They belong to the Ar-C_{ipso} and the Ar-C_{nitro} of the aryl moiety.

¹⁹F NMR (376 MHz, CDCl₃): δ (ppm) = -145.61 – -146.64 (m_c, 2F, BF₂).

HRMS (ESI-TOF, MeOH): m/z calcd. for C₂₇H₃₅BF₂N₄O₂Na⁺ [M+Na]⁺: 519.2713, found: 519.2713.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2957 and 2929 [ν (CH₂), ν (Me)], 2871 [ν (CH)], 1613 [ν (C=N), ν (C=C)], 1545 [ν_{as} (NO₂)], 1512 [δ (Ar-C)], 1467 [δ (CH₂), δ (Me)], 1348 [ν_{sym} (NO₂)], 1082 [ν (BF)], 760 [δ (HC=CH)].

UV/vis (DCM): λ_{max} (nm) [log (ϵ /L mol⁻¹ cm⁻¹)] = 505 [4.62].

Fluorescence (DCM): λ_{max} (nm) = 515 at $\lambda_{excitation}$ (nm) 490.

M.P. (°C): 95-99.

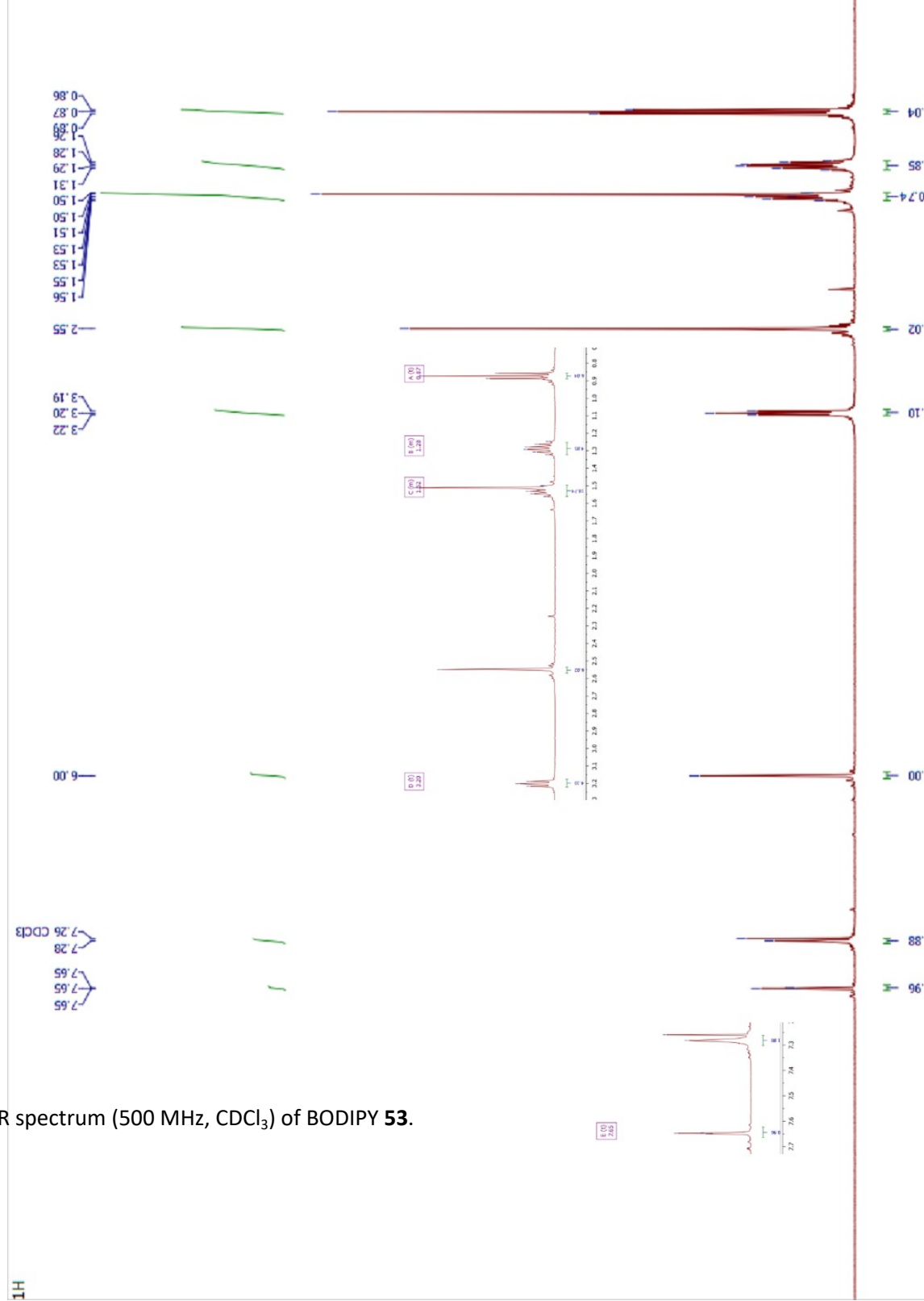
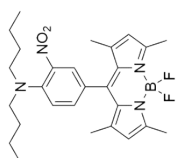


Figure S4.4.1: ¹H NMR spectrum (500 MHz, CDCl₃) of BODIPY 53.

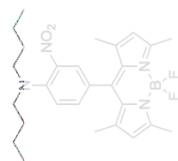
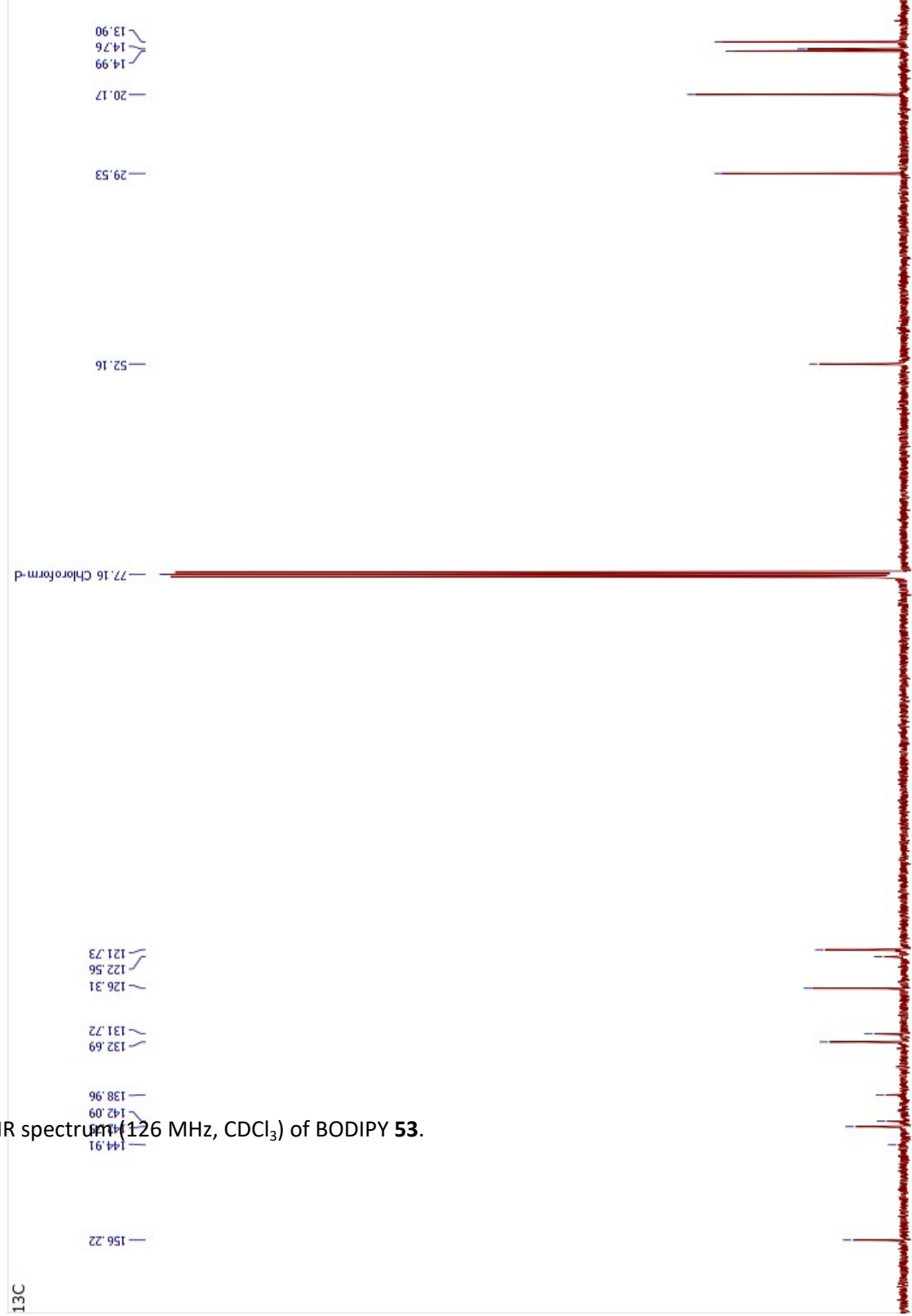


Figure S4.4.2: ^{13}C NMR spectrum (126 MHz, CDCl_3) of BODIPY **53**.



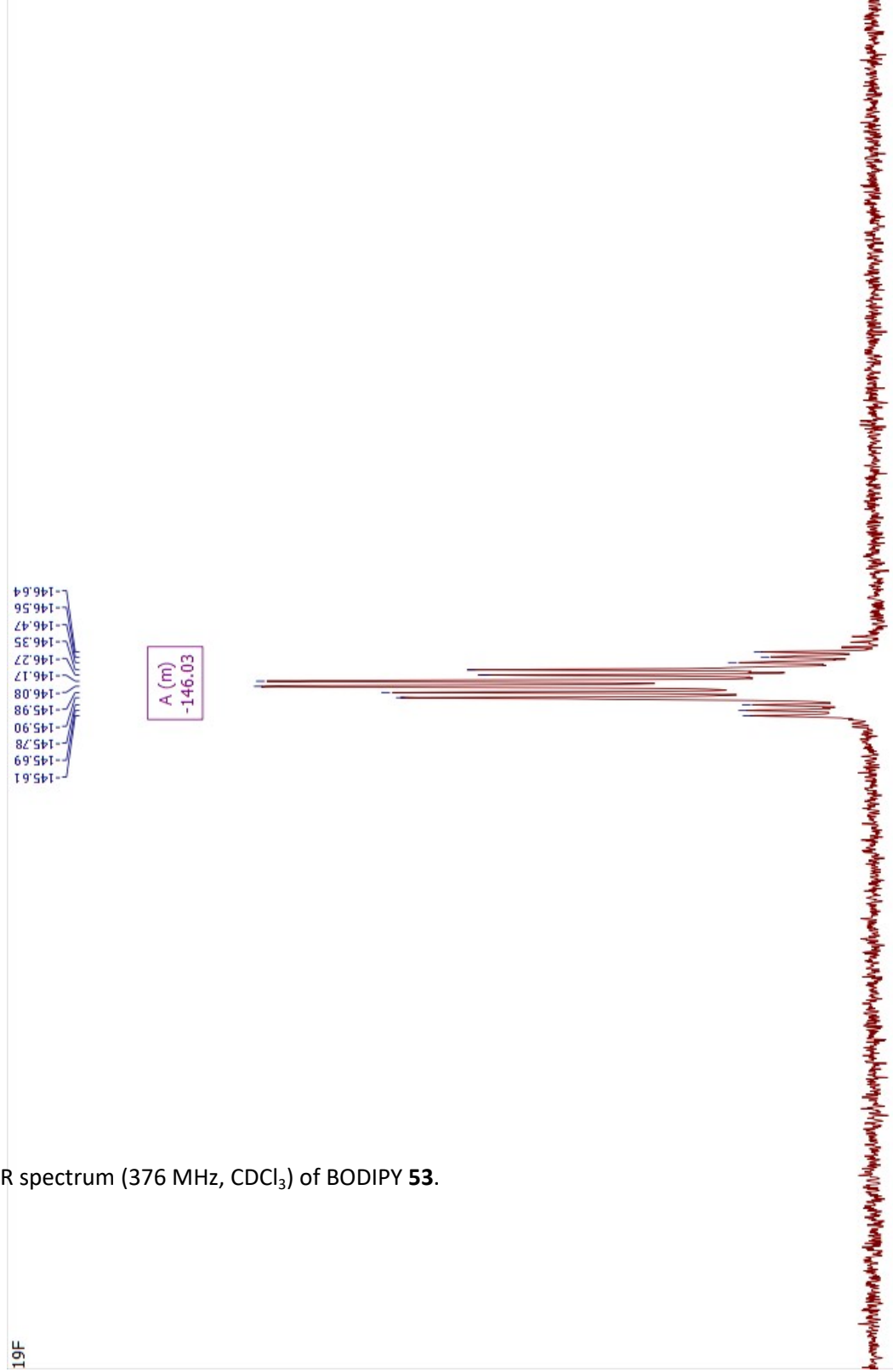
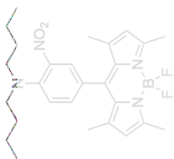


Figure S4.4.3: ^{19}F NMR spectrum (376 MHz, CDCl_3) of BODIPY 53.

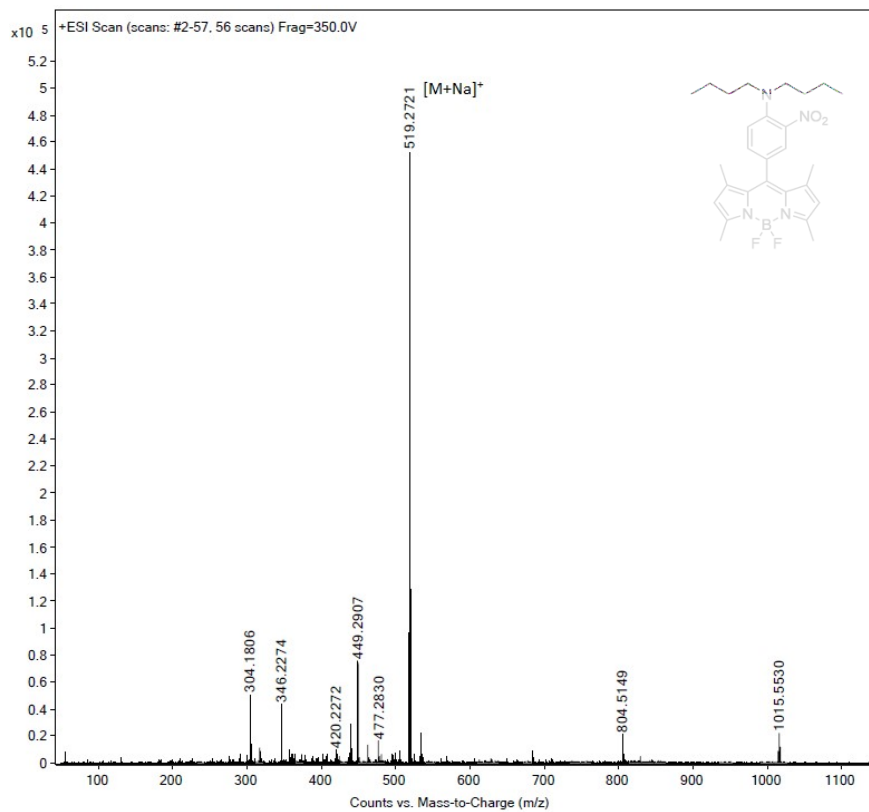


Figure S4.4.4: HRMS spectrum (ESI+) of BODIPY 53.

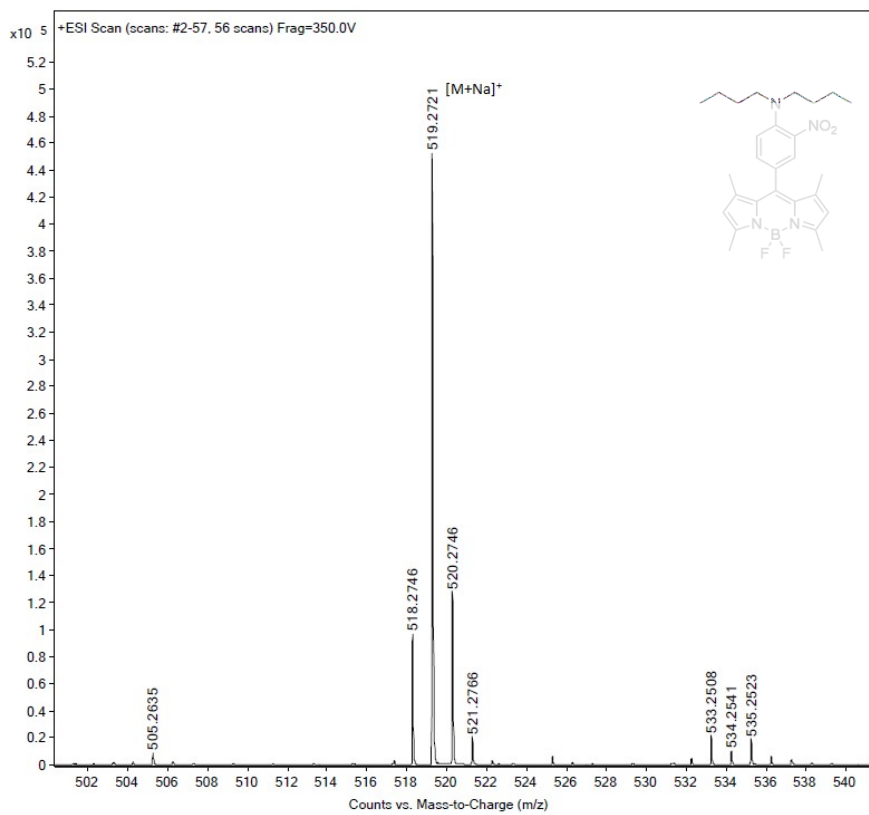


Figure S4.4.5: HRMS spectrum (ESI+) of BODIPY 53.

S5 Bromination of tetramethyl-BODIPYs (54 – 55)

S5.1 General synthetic procedure

The corresponding 1,3,5,7-tetramethyl substituted BODIPY (1 equiv.) was dissolved in 2 mL of HFIP, NBS (2.5 equiv.) was added, and the mixture was stirred for 1 min at rt. Afterwards, the reaction mixture was diluted with EtOAc and washed with water several times. The organic layer was dried with Na₂SO₄, filtered, and evaporated to dryness. The crude product was purified by column chromatography; the main fraction was collected and evaporated to dryness.

S5.2 2,6-Dibromo-8-[3-nitro-4-(*N*-2-prop-2-enylamino)phenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene (**54**):

BODIPY **54** was prepared according to the general synthetic procedure. BODIPY **51** (87 mg, 0.21 mmol) and NBS (91 mg, 0.51 mmol) were dissolved in HFIP. After purification (column chromatography, silica gel, EtOAc/*n*-hexane = 1/9, v/v); BODIPY **54** was obtained as a red solid (53 mg, 92 μmol, 44%). Despite thorough chromatographic purification, this compound still shows tiny signals from an impurity in the range of 3.75 – 4.00 ppm and at 8.25 ppm.

¹H NMR (500 MHz, THF-*d*₈): δ (ppm) = 1.58 (m, 6H, Me), 2.56 (s, 6H, Me), 4.12–4.15 (m, 2H, CH₂), 5.23 (ddd, *J* = 10.4, 1.5 Hz, 1H, H₂C=CH-), 5.31 (ddd, *J* = 17.2, 1.6 Hz, 1H, H₂C=CH-), 6.01 (ddt, *J* = 17.2, 10.1, 4.9 Hz, 1H, H₂C=CH-), 7.20 (d, *J* = 8.8 Hz, 1H, Ar-H_{meta}), 7.41 (dd, *J* = 8.8, 2.1 Hz, 1H, Ar-H_{ortho}), 8.19 (d, *J* = 2.2 Hz, 1H, Ar-H_{ortho}), 8.45 (t, *J* = 5.9 Hz, 1H, NH).

¹³C NMR (126 MHz, THF-*d*₈): δ (ppm) = 13.9 (Me), 14.8 (Me), 46.1 (CH₂), 112.3 (C_{Br}), 116.9 (H₂C=CH-), 117.0 (Ar-C_{meta}), 121.5*, 127.7 (Ar-C_{ortho}), 131.99 (C_{pyrrole}), 133.5*, 134.9 (H₂C=CH-), 136.4 (Ar-C_{ortho}), 141.1 (C_{pyrrole}), 142.1 (C_{meso}), 146.8 (Ar-C_{para}), 154.9 (C_{pyrrole}). *These signals could not be assigned exactly to corresponding carbon atoms. They belong to the Ar-C_{ipso} and the Ar-C_{nitro} of the aryl moiety.

¹⁹F NMR (376 MHz, THF-*d*₈): δ (ppm) = -146.56 – -146.27 (m_c, 2F, BF₂).

HRMS (ESI-TOF, MeOH): *m/z* calcd. for C₂₂H₂₁BBBr₂FN₄O₂⁺ [M-F]⁺: 563.0082, found: 563.0083, *m/z* calcd. for C₂₂H₂₁BBBr₂F₂N₄O₂Na⁺ [M+Na]⁺: 604.9964, found: 604.9968, *m/z* calcd. for C₂₂H₂₁BBBr₂F₂N₄O₂K⁺ [M+K]⁺: 620.9703, found: 620.9707, *m/z* calcd. for C₄₄H₄₂B₂Br₄F₄N₈O₄Na⁺ [2M+Na]⁺: 1187.0036, found: 1187.0044.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 3358 [ν (NH)], 2925 [ν (CH₂)], 2847 [ν (CH)], 1631 [ν (C=N), ν (C=C)], 1535 [ν_{as} (NO₂)], 1460 [δ (CH₂), δ (Me)], 1346 [ν_{sym} (NO₂)], 1170 and 1119 [ν (BF), ν (CBr)], 755 [δ (HC=CH)].

UV/vis (DCM): λ_{max} (nm) [log (ε/L mol⁻¹ cm⁻¹)] = 399 [4.17], 528 [4.85].

Fluorescence (DCM): λ_{max} (nm) = 550 at $\lambda_{excitation}$ (nm) = 380, 510

M.P.(°C): 209-214.

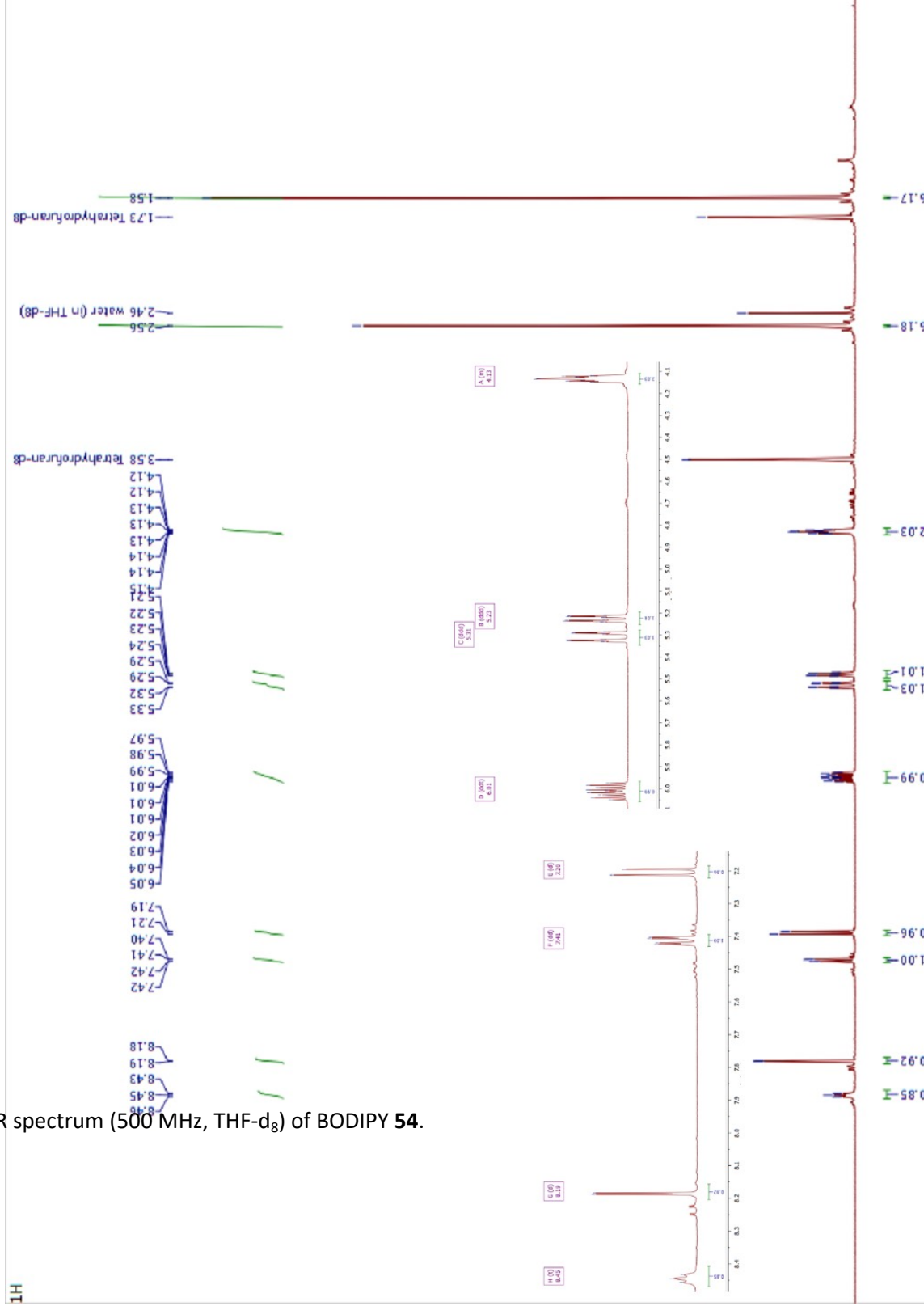
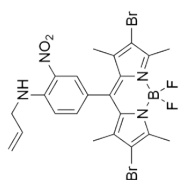


Figure S5.2.1: ^1H NMR spectrum (500 MHz, THF-d_8) of BODIPY 54.

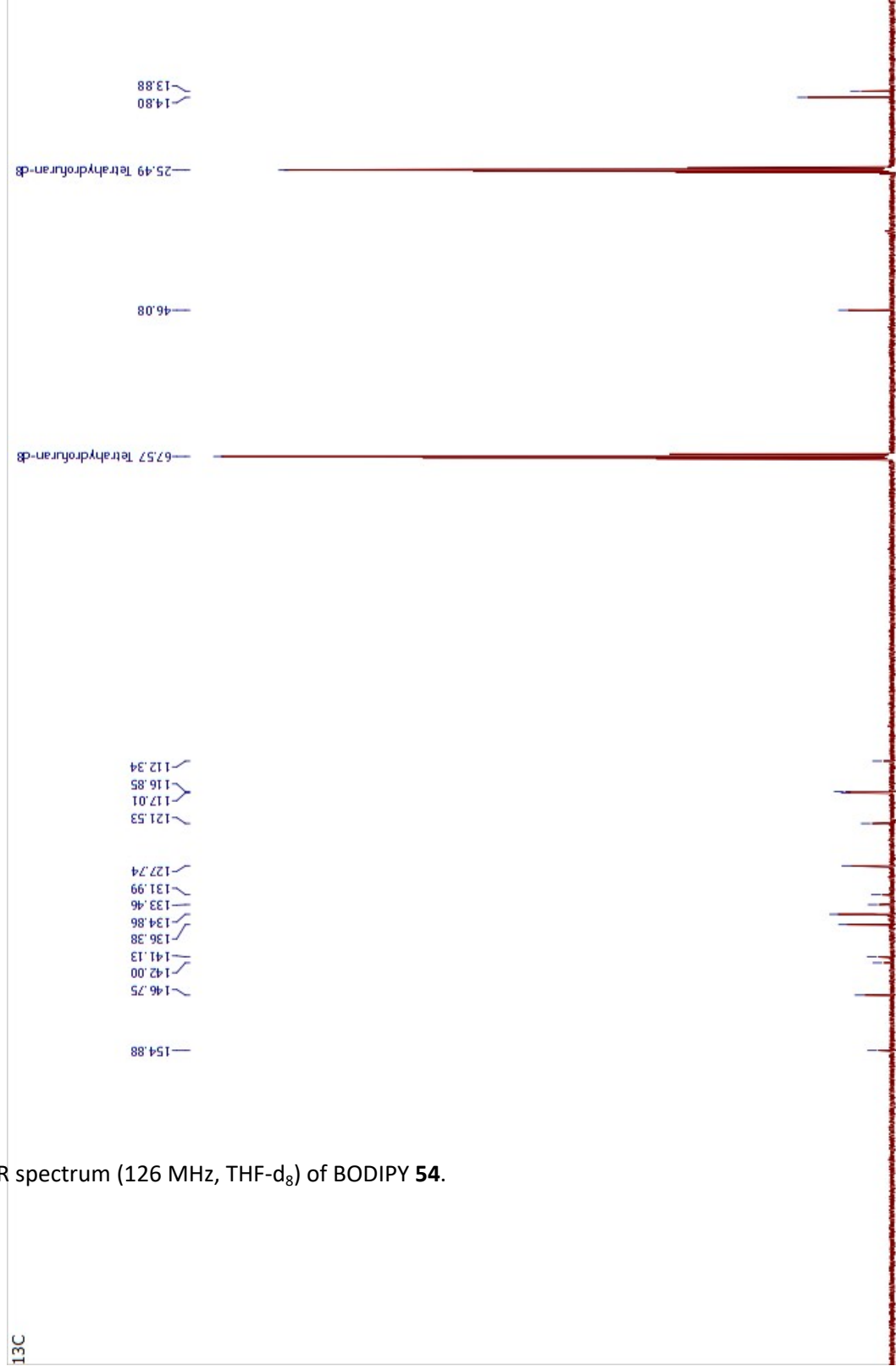
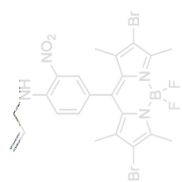


Figure S5.2.2: ^{13}C NMR spectrum (126 MHz, THF-d_8) of BODIPY 54.



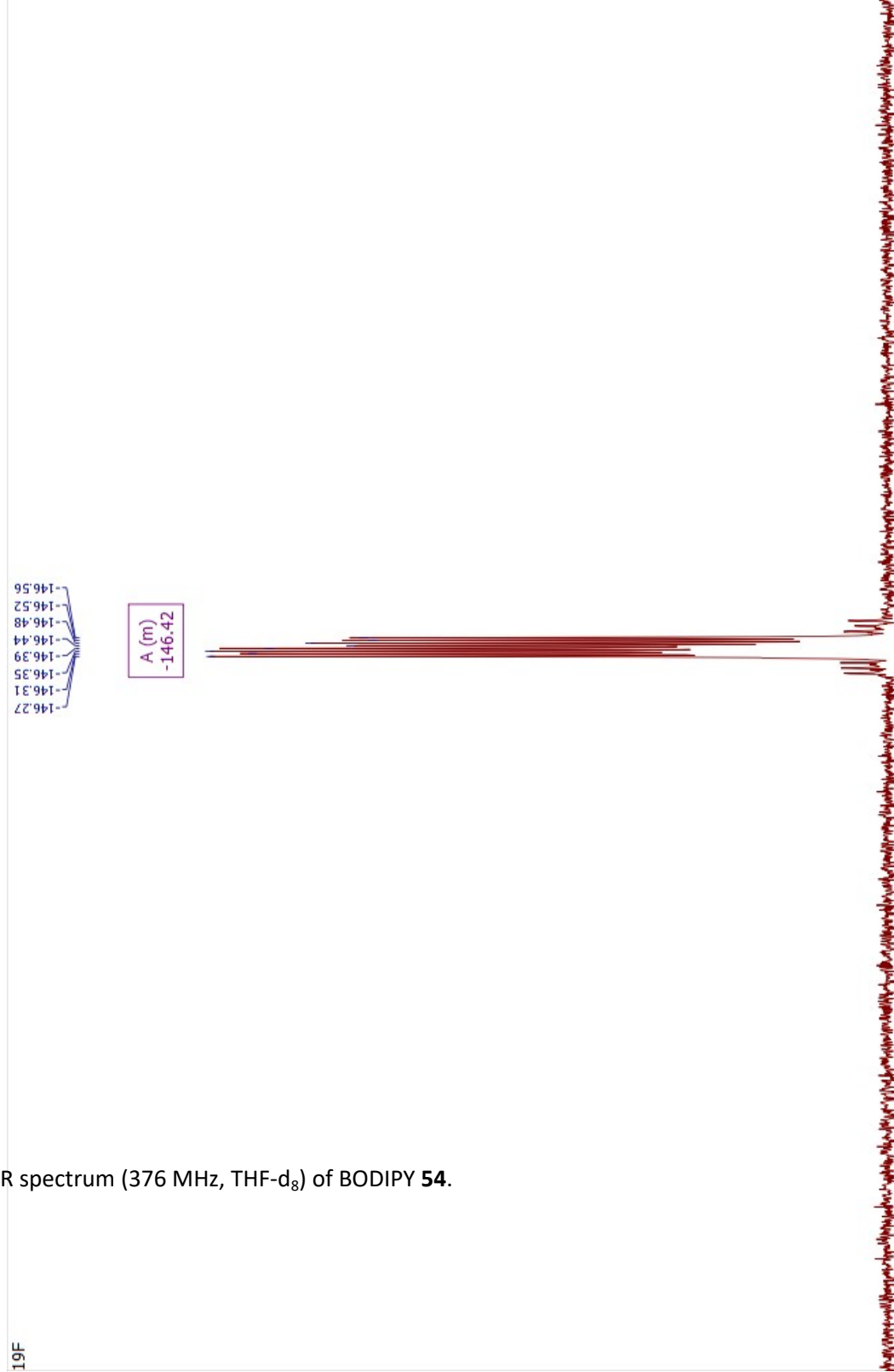
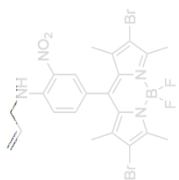


Figure S5.2.3: ^{19}F NMR spectrum (376 MHz, THF-d_8) of BODIPY 54.

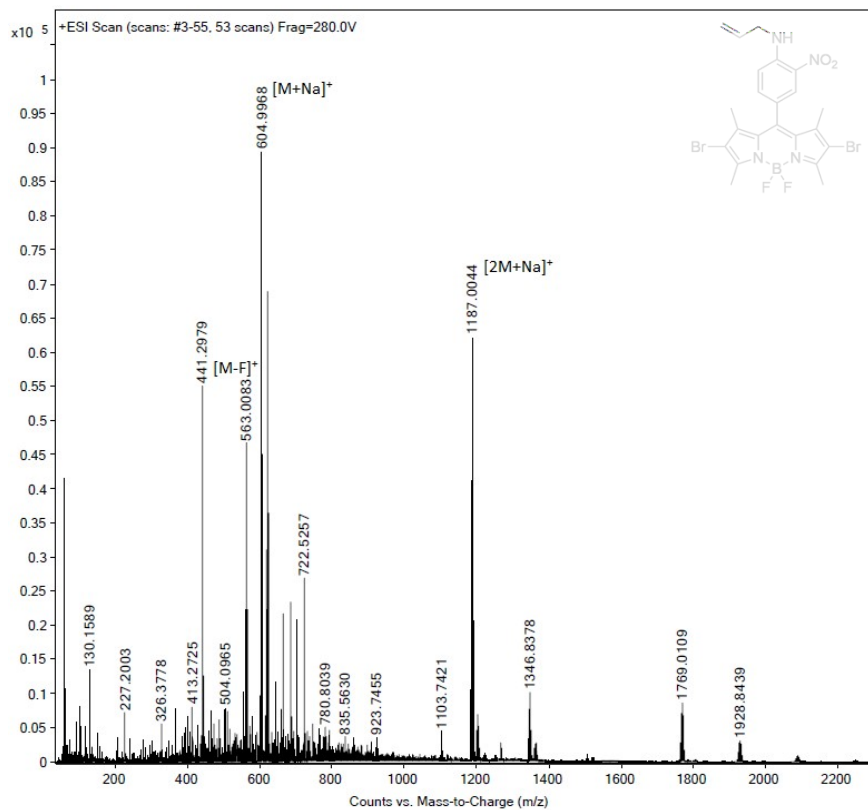


Figure S5.2.4: HRMS spectrum (ESI+) of BODIPY 54.

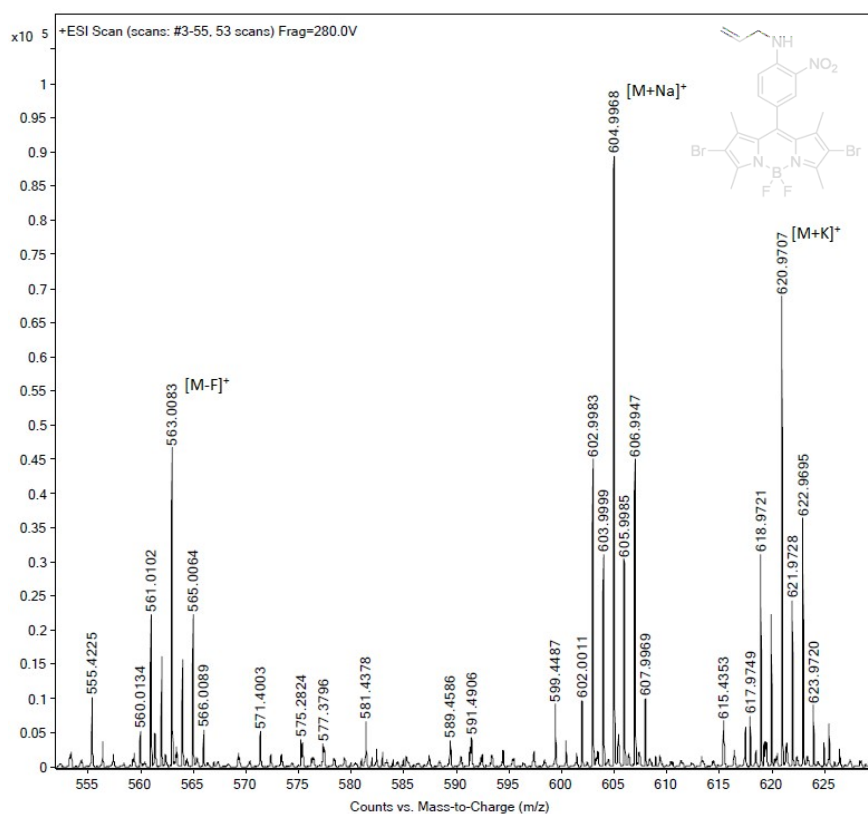


Figure S5.2.5: HRMS spectrum (ESI+) of BODIPY 54.

S5.3 2,6-Dibromo-8-[4-(*N*-6-methoxy-6-oxohexylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene (**55**):

BODIPY **55** was prepared according to the general synthetic procedure. BODIPY **52** (95 mg, 0.19 mmol) and NBS (83 mg, 0.46 mmol) were dissolved in HFIP. After purification (column chromatography, silica gel, EtOAc/*n*-hexane = 1/9, v/v); BODIPY **55** was obtained as a red solid (67 mg, 0.10 mmol, 54%). After purification, the ¹H-NMR spectrum still showed tiny amounts of *n*-hexane resulting from chromatography.

¹H NMR (500 MHz, THF-*d*₈): δ (ppm) = 1.48–1.54 (m, 2H, CH₂), 1.59 (s, 6H, Me), 1.67–1.71 (m, 2H, CH₂), 1.76–1.82 (m, 2H, CH₂), 2.33 (t, *J* = 7.3 Hz, 2H, CH₂), 2.56 (s, 6H, Me), 3.46 (td, *J* = 7.3, 5.6 Hz, 2H, CH₂), 3.60 (s, 3H, Me_{COOMe}), 7.27 (d, *J* = 8.8 Hz, 1H, Ar-H_{meta}), 7.43 (dd, *J* = 8.8, 2.2 Hz, 1H, Ar-H_{ortho}), 8.18 (d, *J* = 2.1 Hz, 1H, Ar-H_{ortho}), 8.27 (t, *J* = 5.5 Hz, 1H, NH).

¹³C NMR (126 MHz, THF-*d*₈): δ (ppm) = 13.9 (Me), 14.9 (Me), 25.98 (CH₂), 27.6 (CH₂), 29.4 (CH₂), 34.3 (CH₂), 43.9 (CH₂), 51.5 (Me_{COOMe}), 112.6 (C_{Br}), 116.5 (Ar-C_{meta}), 121.2*, 127.9 (Ar-C_{ortho}), 132.0 (C_{pyrrole}), 133.2*, 136.6 (Ar-C_{ortho}), 141.1 (C_{pyrrole}), 142.1 (C_{meso}), 146.9 (Ar-C_{para}), 154.9 (C_{pyrrole}), 173.8 (CO). *These signals could not be assigned exactly to corresponding carbon atoms. They belong to the Ar-C_{ipso} and the Ar-C_{nitro} of the aryl moiety.

¹⁹F NMR (376 MHz, THF-*d*₈): δ (ppm) = -146.56 – -146.25 (m_c, 2F, BF₂).

HRMS (ESI-TOF, MeOH): *m/z* calcd. for C₂₆H₂₉BBr₂F₂N₄O₄Na⁺ [M+Na]⁺: 693.0488, found: 693.0503, *m/z* calcd. for C₂₆H₂₉BBr₂F₂N₄O₄K⁺ [M+K]⁺: 709.0228, found: 709.0235, *m/z* calcd. for C₅₂H₅₈B₂Br₄F₄N₈O₈Na⁺ [2M+Na]⁺: 1363.1085, found: 1363.1112.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 3379 [ν (NH)], 2926 [ν (CH₂), ν (Me)], 2860 [ν (CH)], 1733 [ν (COOMe)], 1632 [ν (C=N), ν (C=C)], 1532 [ν_{as} (NO₂)], 1457 [δ (CH₂), δ (Me)], 1346 [ν_{sym} (NO₂)], 1163 and 1118 [ν (BF), ν (CBr)], 755 [δ (HC=CH)].

UV/vis (DCM): λ_{max} (nm) [log (ε/L mol⁻¹ cm⁻¹)] = 404 [4.19], 532 [4.87].

Fluorescence (DCM): λ_{max} (nm) = 550 at $\lambda_{excitation}$ (nm) = 390, 520.

M.P.(°C): 152-155.

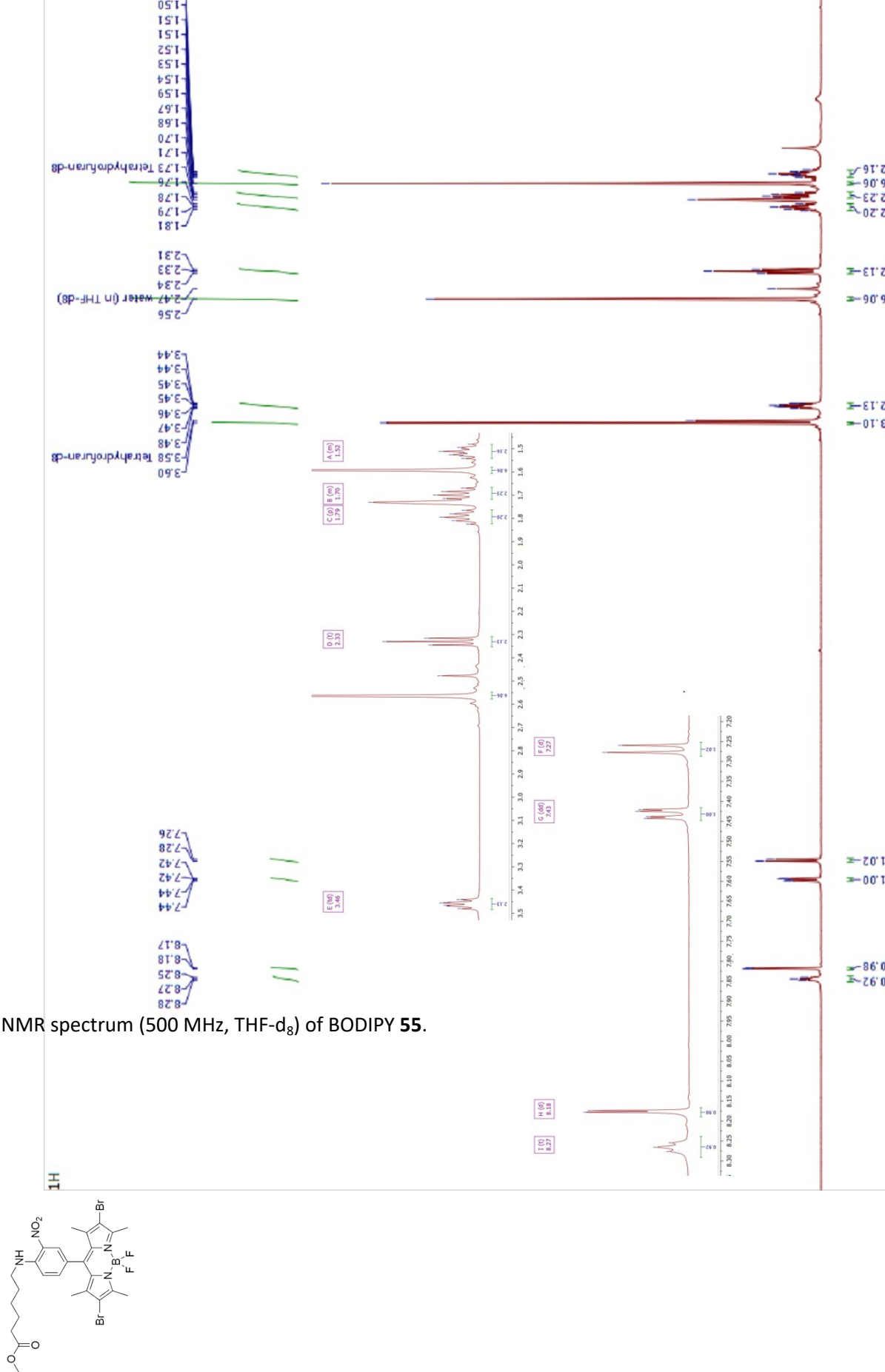


Figure S5.3.1: ¹H NMR spectrum (500 MHz, THF-d₈) of BODIPY 55.

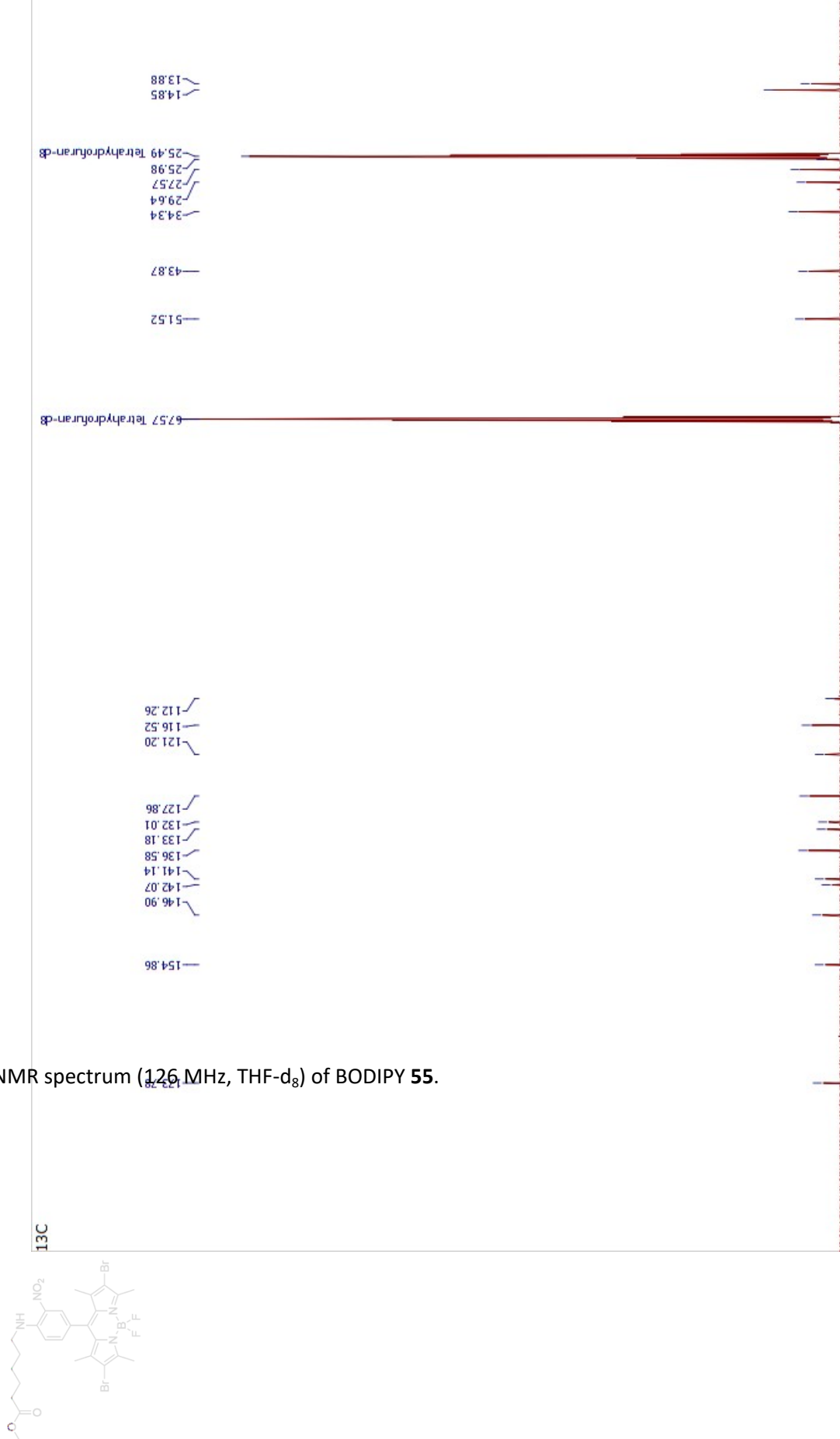


Figure S5.3.2: ^{13}C NMR spectrum (126 MHz, THF-d_8) of BODIPY 55.

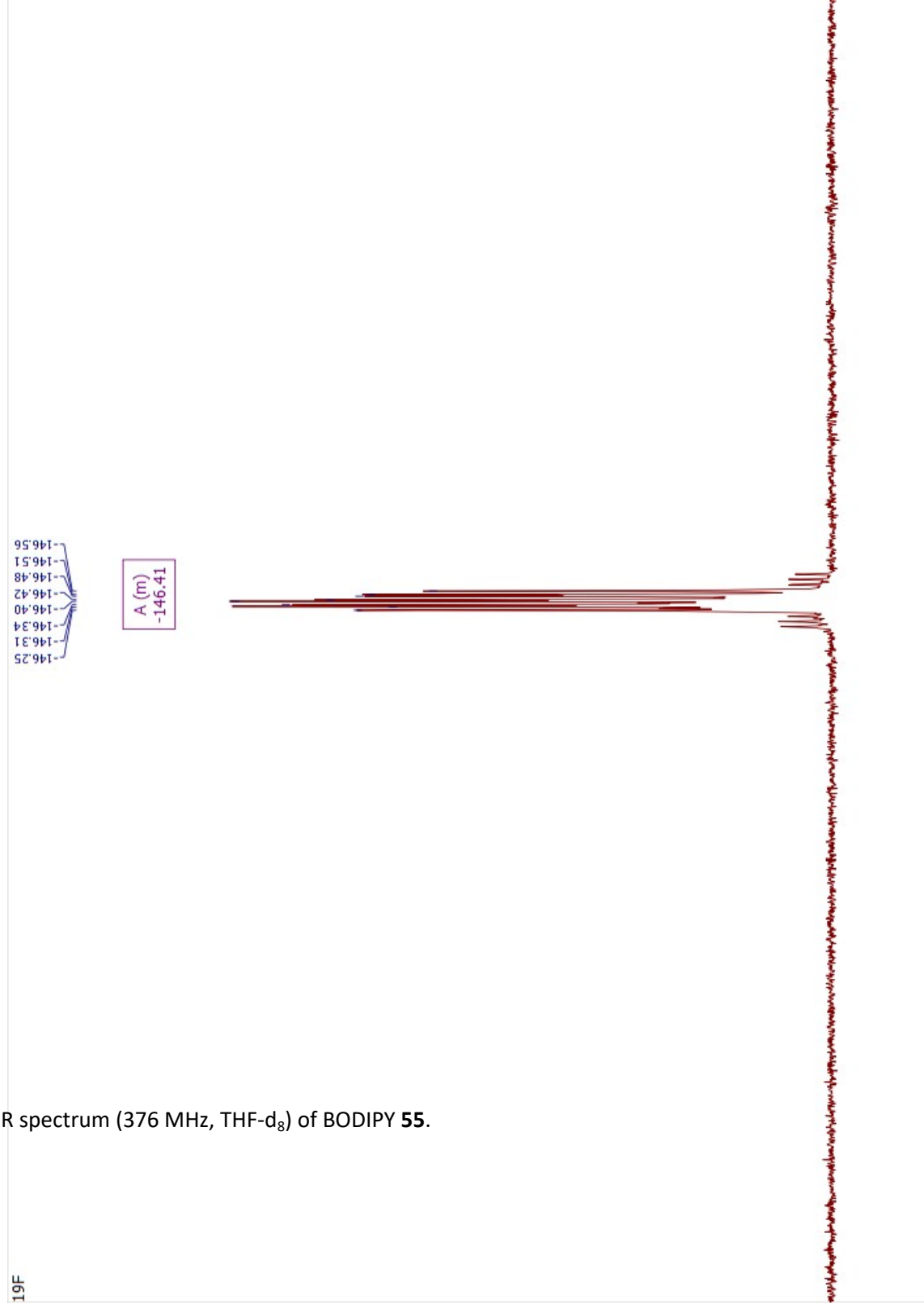
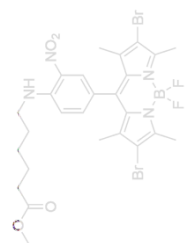


Figure S5.3.3: ^{19}F NMR spectrum (376 MHz, THF-d_8) of BODIPY 55.

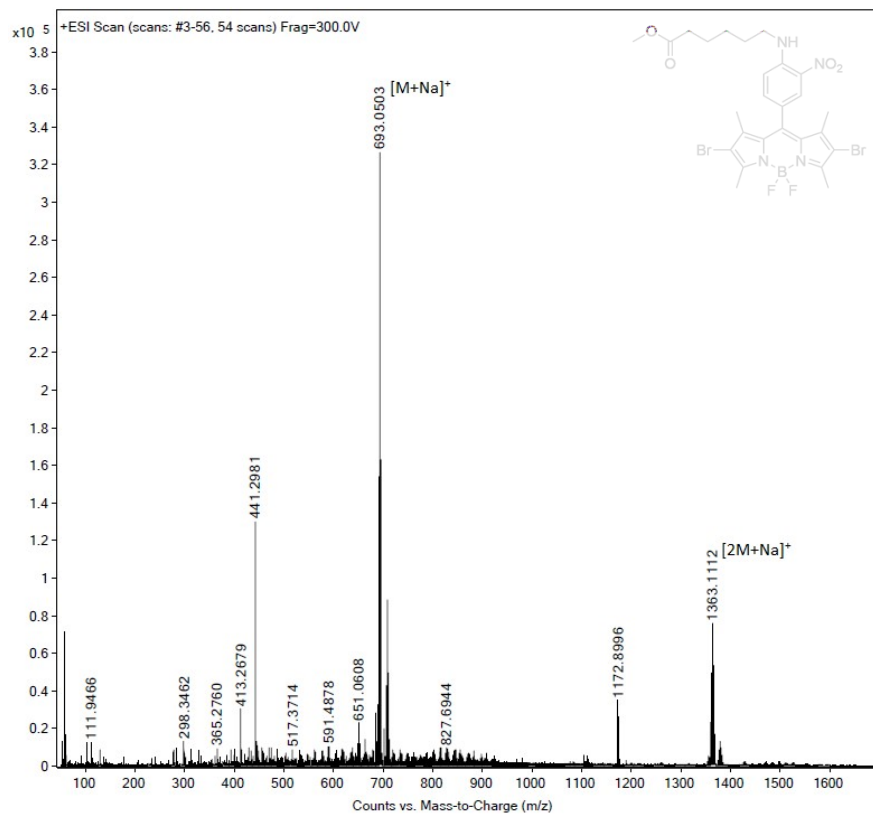


Figure S5.3.4: HRMS spectrum (ESI+) of BODIPY 55.

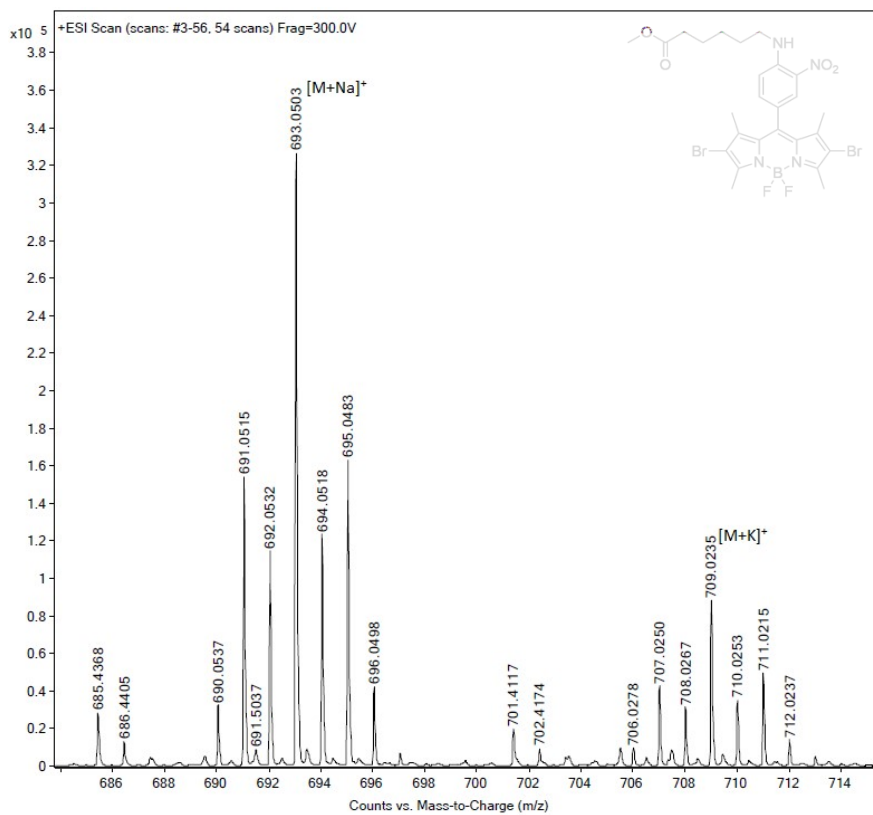


Figure S5.3.5: HRMS spectrum (ESI+) of BODIPY 55.

S5.4 2,6-Dibromo-8-[4-(*N,N*-dibutylamino)-3-nitrophenyl]-1,3,5,7-tetramethyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene (**56**):

BODIPY **56** was prepared according to the general synthetic procedure. BODIPY **53** (90 mg, 0.18 mmol) and NBS (80 mg, 0.45 mmol) were dissolved in HFIP. After purification (column chromatography, silica gel, EtOAc/*n*-hexane = 1/9, v/v); BODIPY **56** was obtained as a red solid (68 mg, 0.10 mmol, 57%).

¹H NMR (500 MHz, THF-*d*₈): δ (ppm) = 0.89 (t, J = 7.3 Hz, 6H, Me_{Butyl}), 1.28–1.35 (m, 4H, CH₂), 1.53–1.59 (m, 10H, Me + CH₂), 2.56 (s, 6H, Me), 3.25 (t, J = 7.1 Hz, 4H, CH₂), 7.39 (dd, J = 8.7, 2.0 Hz, 1H, Ar-H_{meta}), 7.46 (dd, J = 8.7, 1.4 Hz, 1H, Ar-H_{ortho}), 7.77 (d, J = 2.0 Hz, 1H, Ar-H_{ortho}).

¹³C NMR (126 MHz, THF-*d*₈): δ (ppm) = 13.99 (Me_{Butyl}), 14.3 (Me), 14.6 (Me), 21.0 (CH₂), 30.6 (CH₂), 52.8 (CH₂), 112.3–112.4 (m, C_{Br}), 123.7 (Ar-C_{meta}), 125.4*, 127.2 (Ar-C_{ortho}), 131.8 (C_{Pyrrrole}), 133.2 (Ar-C_{ortho}), 141.1 (C_{Pyrrrole}), 141.7 (C_{meso}), 143.9*, 146.4 (Ar-C_{para}), 154.97 (C_{Pyrrrole}). *These signals could not be assigned exactly to corresponding carbon atoms. They belong to the Ar-C_{ipso} and the Ar-C_{nitro} of the aryl moiety.

¹⁹F NMR (376 MHz, THF-*d*₈): δ (ppm) = -146.52 – -146.31 (m_c, 2F, BF₂).

HRMS (ESI-TOF, MeOH): m/z calcd. for C₂₇H₃₄BBr₂F₂N₄O₂⁺ [M+H]⁺: 655.1084, found: 655.1087, m/z calcd. for C₂₇H₃₃BBr₂F₂N₄O₂Na⁺ [M+Na]⁺: 677.0903, found: 677.0899, m/z calcd. for C₅₄H₆₆B₂Br₄F₄N₈O₄Na⁺ [2M+Na]⁺: 1331.1914, found: 1331.1930.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2957 and 2929 [ν (CH₂), ν (Me)], 2871 [ν (CH)], 1615 [ν (C=N), ν (C=C)], 1533 [ν_{as} (NO₂)], 1461 [δ (CH₂), δ (Me)], 1346 [ν_{sym} (NO₂)], 1171 and 1118 [ν (BF), ν (CBr)], 756 [δ (HC=CH)].

UV/vis (DCM): λ_{max} (nm) [log (ϵ /L mol⁻¹ cm⁻¹)] = 389 [4.03], 533 [4.86].

Fluorescence (DCM): λ_{max} (nm) = 551 at $\lambda_{excitation}$ (nm) = 360, 520.

M.P.(°C): 157-159.

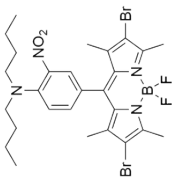
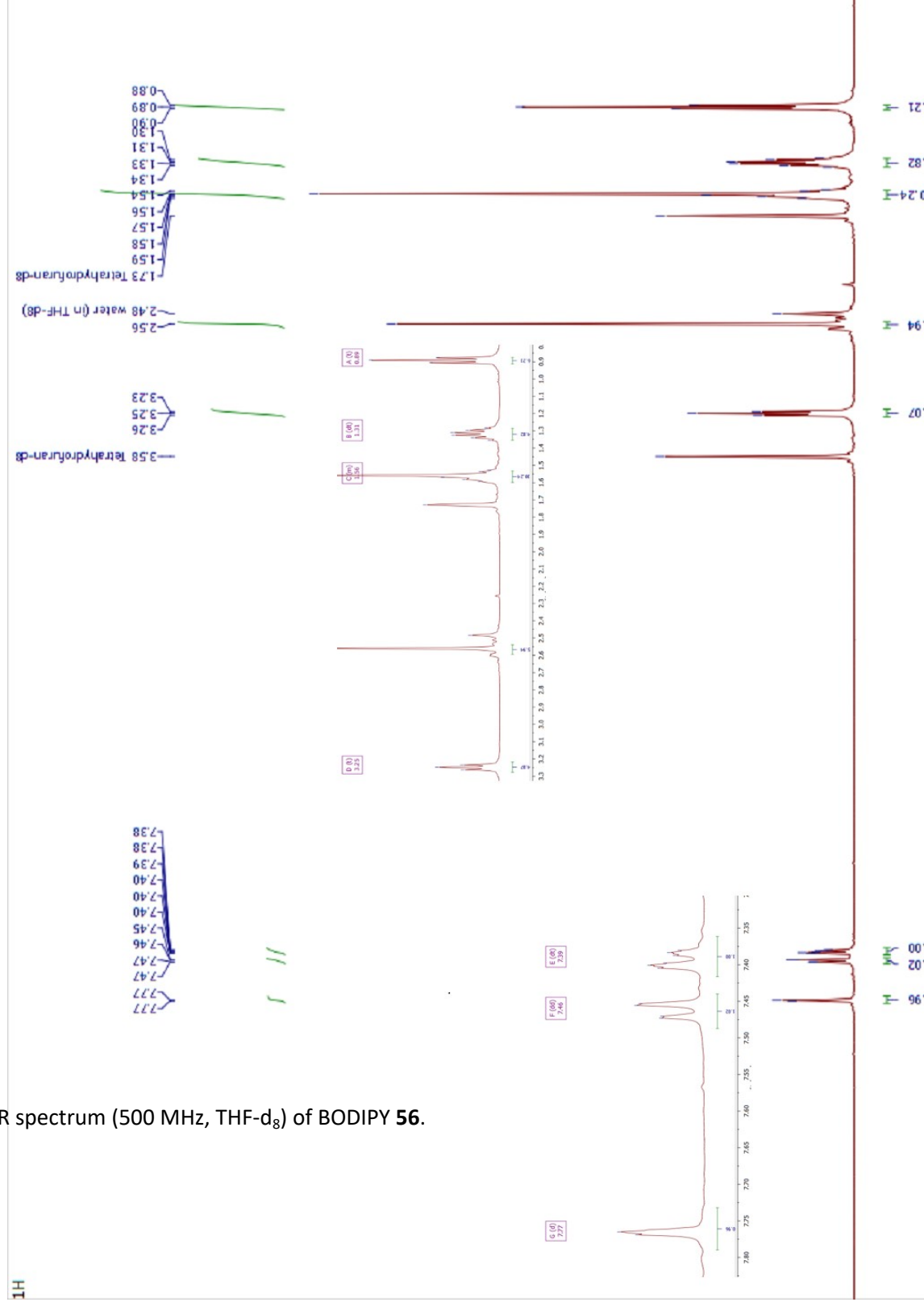


Figure S5.4.1: ^1H NMR spectrum (500 MHz, THF-d_8) of BODIPY 56.



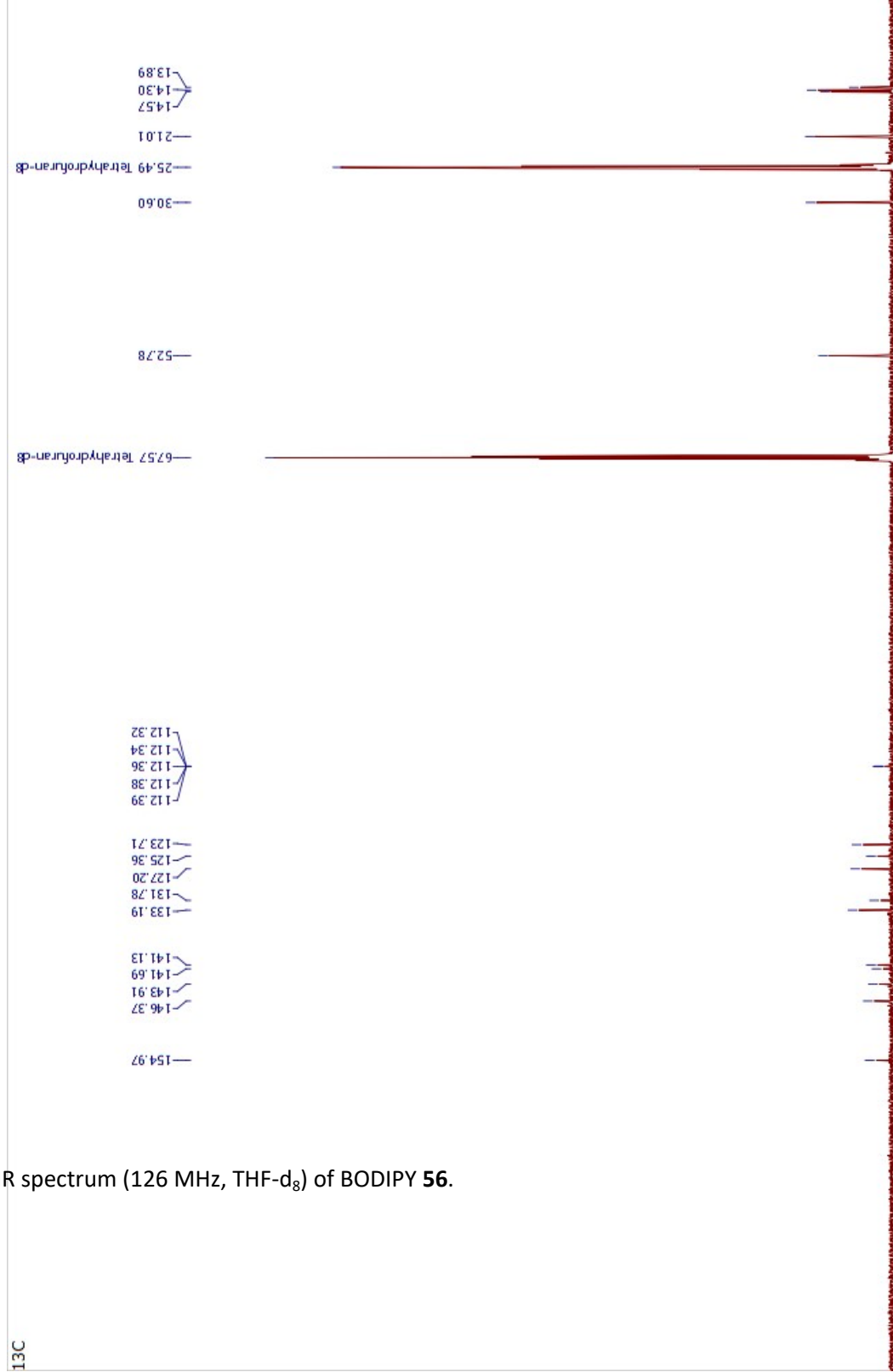
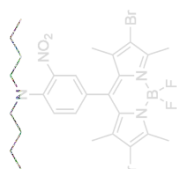


Figure S5.4.1: ^{13}C NMR spectrum (126 MHz, THF- d_8) of BODIPY 56.



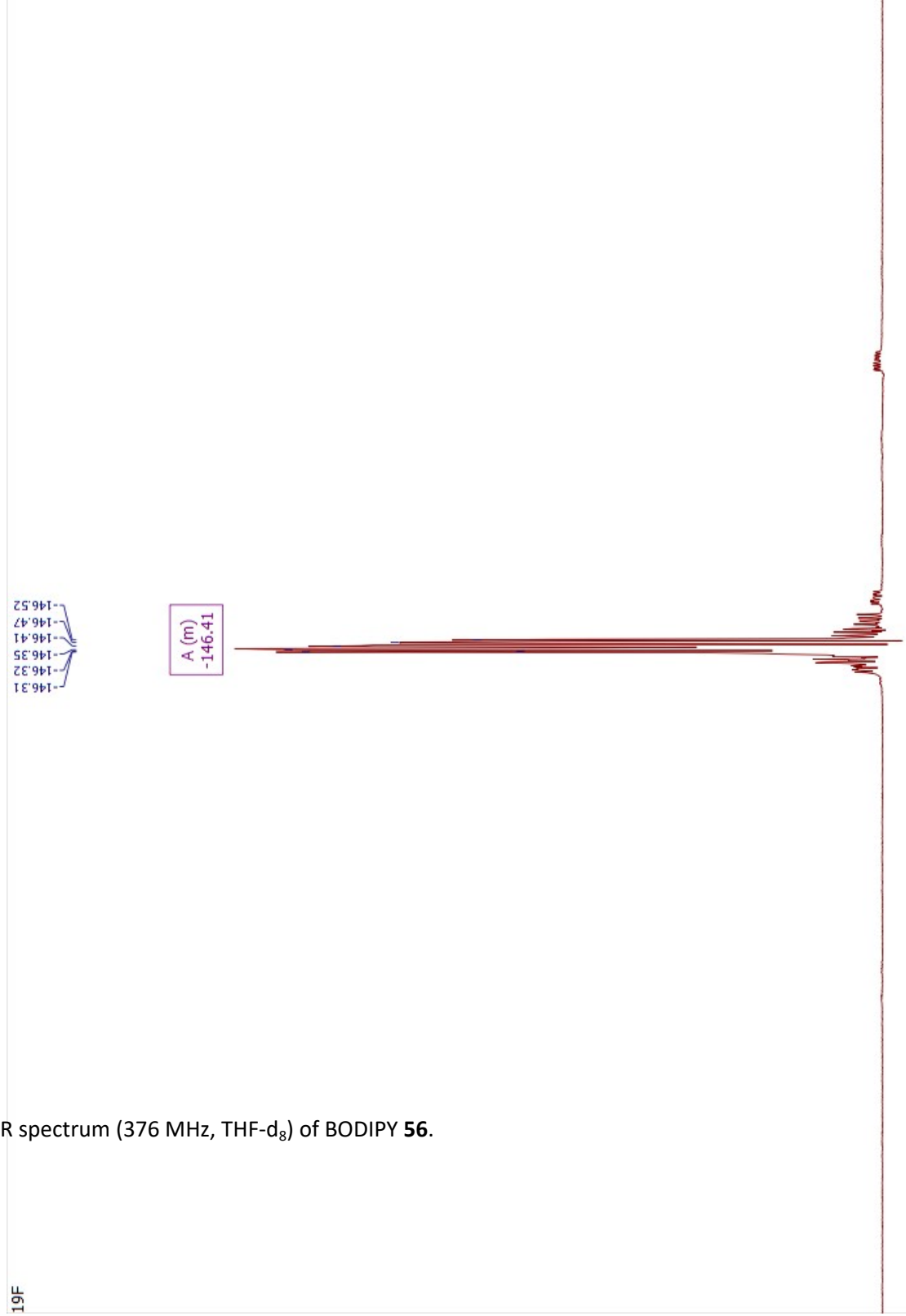
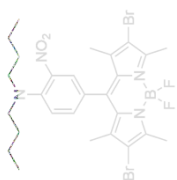


Figure S5.4.3: ^{19}F NMR spectrum (376 MHz, THF-d_8) of BODIPY **56**.



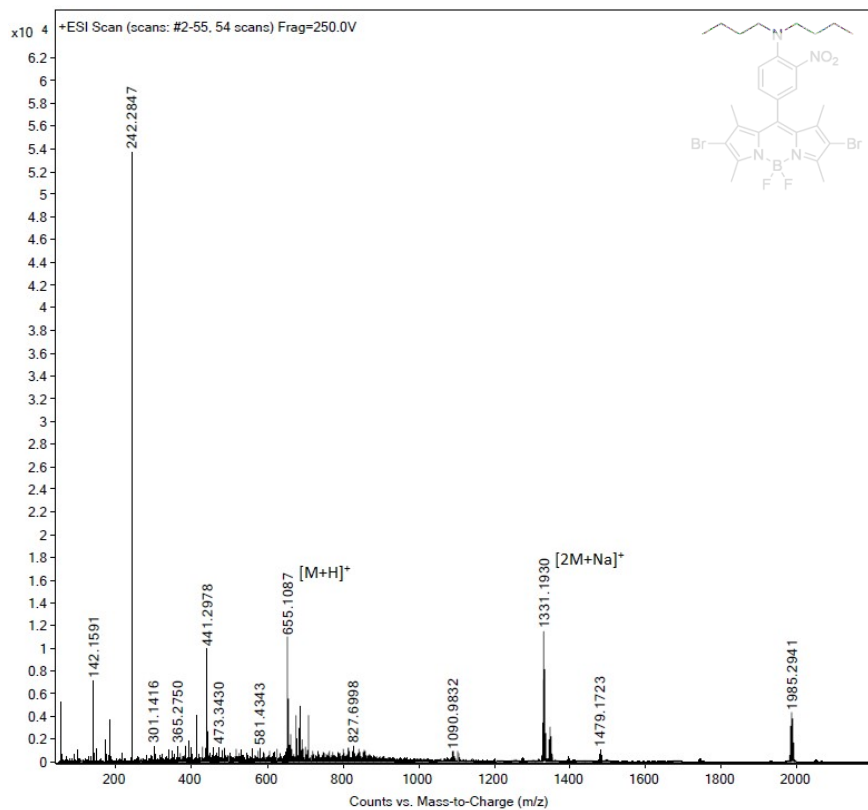


Figure S5.4.4: HRMS spectrum (ESI+) of BODIPY 56.

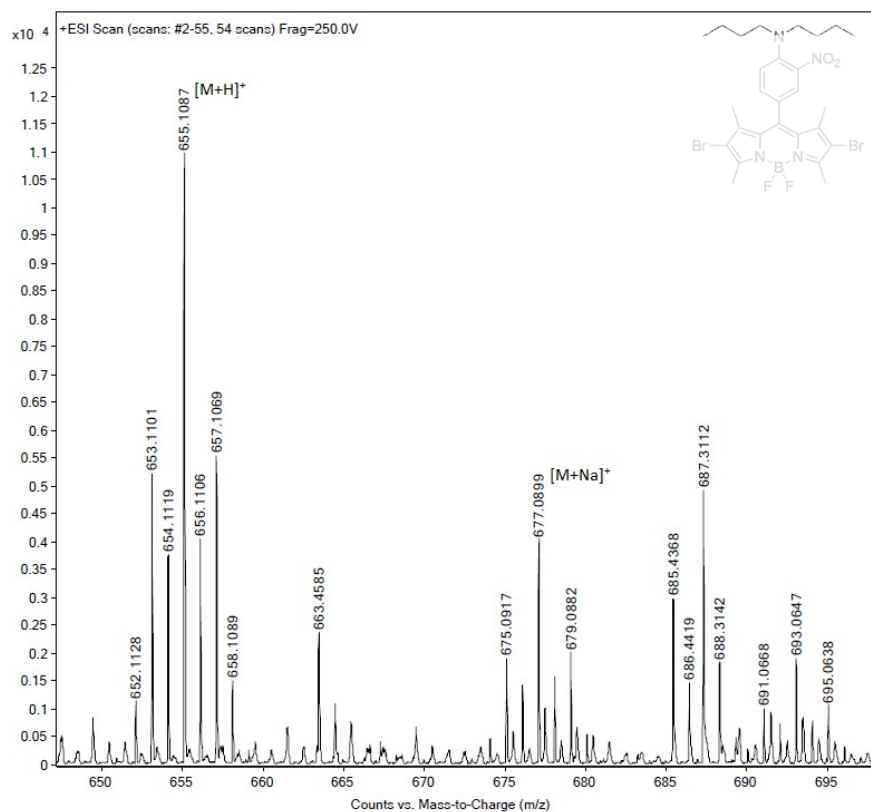


Figure S5.4.5: HRMS spectrum (ESI+) of BODIPY 56.

S6 References

- [1] (a) B. F. Hohlfeld, K. F. Flanagan, N. Kulak, M. O. Senge, M. Christmann and A. Wiehe, *Eur. J. Org. Chem.*, 2019, 4020–4033; (b) C. S. Gutsche, B. F. Hohlfeld, K. J. Flanagan, M. O. Senge, N. Kulak and A. Wiehe, *Eur. J. Org. Chem.*, 2017, 3187–3196; (c) C. S. Gutsche, M. Ortwerth, S. Gräfe, K. J. Flanagan, M. O. Senge, H.-U. Reissig, N. Kulak and A. Wiehe, *Chem. Eur. J.*, 2016, **22**, 13953–13964; (d) H. G. A. Golf, U.-H. Reissig and A. Wiehe, *Org. Lett.*, 2015, **17**, 982–985.
- [2] G. Vives, C. Giansante, R. Bofinger, G. Raffy, A. Del Guerzo, B. Kauffmann, P. Batat, G. Jonusauskas, N. D. McClenaghan, *Chem. Commun.* 2011, **47**, 10425-100427.
- [3] B. F. Hohlfeld, B. Gitter, K. J. Flanagan, C. J. Kingsbury, N. Kulak, M. O. Senge, A. Wiehe, *Org. Biomol. Chem.* 2020, **18**, 2416-2431.