Cross-Linked Supramolecular Polymer Networks Constructed by Pillar[5]arene-Based Host–Guest Recognition and Coordination/Oxidation of Catechol

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1. Materials and instrumentations

All reagents were commercially available and used as supplied without further purification. ¹H NMR spectra, ¹³C NMR spectra, NOESY and DOSY were recorded with an Agilent 600 MHz DirectDrive2 with use of the deuterated solvent as the lock and the residual solvent as the internal reference. UV-vis spectra were taken on a PerkinElmer Lambda 35 UV-vis spectrophotometer. Low-resolution electrospray ionization (LRESI) mass spectra were obtained on a Bruker Esquire 3000 plus mass spectrometer (Bruker-Franzen Analytik GmbH Bremen, Germany) equipped with an ESI interface and an ion trap analyzer. High-resolution mass spectrometric experiments were performed with a Bruker 7-Tesla FT-ICR mass spectrometer equipped with an electrospray source (Billerica, MA, USA). Matrix-assisted laser desorption/ionization time of flight mass spectrometric experiments were performed on a Bruker Ultraflex MALDI-TOF mass spectrometer with a 355 nm Nd: YAG laser (Smartbeam II) and 25 kV ion source voltage. Scanning electron microscopy (SEM) investigations were carried out on a JEOL 6390LV instrument. SEM samples were prepared at the concentration of 1.00 mM via the vacuum freeze-drying methodology. The compounds 2 and 3 were prepared according to previous work.^{S1,S2}

2. Synthesis and characterizations of compounds



Scheme S1 The synthetic route to compound 1.

The compound **3** was prepared according to previous work.^{S1} The ¹H NMR spectrum of **3** is shown in Fig. S1. ¹H NMR spectrum of **3** (600 MHz, 298 K) in CDCl₃ δ (ppm): 6.79–6.82 (m, 10 H), 4.09–4.12 (t, J = 9 Hz, 4 H), 3.78–3.79 (br, 10 H), 3.71 (s, 18 H), 3.68 (s, 6 H), 3.50–3.52 (t, J = 6 Hz, 4 H).



Fig. S1 ¹H NMR spectrum (600 MHz, CDCl₃, 298 K) of **3**.

Synthesis of compound **4**: A mixture of compound **3** (3.00 g, 3.20 mmol) and potassium phthalimide (3.72 g, 20.0 mmol) was stirred in *N*, *N*-dimethylformamide at 90 °C for 24 h. The solution was evaporated under vacuum. The crude product was dissolved in CH₂Cl₂ (400 mL) and washed three times with H₂O (100 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated to afford a yellow solid, which was further purified by column chromatography using petroleum ether/ethyl acetate (v : v = 3 : 1). The fractions containing the product were concentrated to give **4** as a yellow solid. The ¹H NMR spectrum of **4** is shown in Fig. S2. ¹H NMR spectrum of **4** (600 MHz, 298 K) in CDCl₃ δ (ppm): 7.82–7.83 (m, 4 H), 7.67–7.69 (m, 4 H), 6.69 (s, 2 H), 6.79–6.80 (d, J = 6 Hz, 4 H), 6.75 (s, 2 H), 6.72 (s, 2 H), 4.21–4.25 (m, 2 H), 4.03–4.10 (m, 4 H), 3.97–4.01 (m. 2 H), 3.57–3.78 (m, 34 H). The ¹³C NMR spectrum of **4** is shown in Fig. S3. ¹³C NMR spectrum of **4** (150 MHz, 298 K) in CDCl₃ δ (ppm): 170.8, 153.4, 153.3, 153.2, 136.6, 134.7, 131.0, 130.8, 125.9, 117.1, 116.7, 116.6, 116.4, 68.0, 58.5, 40.5, 32.1. LRESIMS is shown in Fig. S4: m/z 1086.23 [M + NH₄]⁺.



30 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 $\mathcal{S}(\mathrm{ppm})$

Fig. S3 ¹³C NMR spectrum (150 MHz, CDCl₃, 298 K) of 4.



Fig. S4 LRESIMS spectrum of 4 in CHCl₃.

Synthesis of compound **5**: A mixture of **4** (2.00 g, 2.48 mmol) and hydrazine hydrate (25 ml) was heated at reflux in methanol (40 mL) for 10 h. Then the mixture was filtered and washed with methanol to give **5** as a white solid. The ¹H NMR spectrum of **5** is shown in Fig. S5. ¹H NMR spectrum of **5** (600 MHz, 298 K) in CDCl₃ δ (ppm): 6.78 (s, 2 H), 6.73 (d, J = 6 Hz, 4 H), 6.67 (s, 2 H), 6.65 (s, 2 H), 3.77–3.78 (br, 14 H), 3.67 (s, 6 H), 3.64 (s, 6 H), 3.60 (s, 12 H), 2.85–2.87 (m, J = 6 Hz, 4 H), 1.39 (s, 4 H). The ¹³C NMR spectrum of **5** is shown in Fig. S6. ¹³C NMR spectrum of **5** (150 MHz, 298 K) in CDCl₃ δ (ppm): 153.5, 152.5, 131.2, 116.9, 73.6, 58.6, 44.3, 32.5. LRESIMS is shown in Fig. S7: m/z 809.48 [M + H]⁺. m/z calcd for [M + H]⁺ C₄₇H₅₇N₂O₁₀⁺, 809.4013; found 809.4011, error –0.25 ppm.



Fig. S5 1 H NMR spectrum (600 MHz, CDCl₃, 298 K) of 5.



Fig. S6 ¹³C NMR spectrum (150 MHz, CDCl₃, 298 K) of 5.



Fig. S7 LRESIMS spectrum of 5 in CHCl₃



Synthesis of compound 1: A mixture of compound 5 (2.00 g, 2.47 mmol), DOPAC (1.18 g, 7.00 mmol), EDC (1.34 g, 7.00 mmol) and DMAP (catalytic amount) was stirred at room temperature in dry THF (200 ml) for 24 hours. After the solid was filtered off, the solvent was concentrated by rotary evaporation. The crude product was dissolved in CH₂Cl₂ (100 mL) and washed three times with H₂O (50 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated to afford a brown solid, further purified by column which was chromatography using dichloromethane/methanol (v: v = 25: 1). The fractions containing the product were concentrated to give 1 as a white solid. The ¹H NMR spectrum of 1 is shown in Fig. S9. ¹H NMR spectrum of **1** (600 MHz, 298 K) in CDCl₃ δ (ppm): 6.40–6.70 (m, 18 H), 3.31-3.70 (m, 46 H). The ¹³C NMR spectrum of 1 is shown in Fig. S10. ¹³C NMR spectrum of 1 (150 MHz, 298 K) in CDCl₃ δ (ppm): 172.4, 150.0, 149.7, 148.7, 143.7, 142.8, 127.8, 127.5, 127.0, 125.2, 120.2, 115.3, 114.2, 55.0, 52.4, 41.7, 38.5, 28.6. LRESIMS is shown in Fig. S11: m/z 1107.45 [M - H]⁻. m/z calcd for [M - H]⁻ C₆₃H₆₇N₂O₁₆⁻, 1107.4496; 1107.4483, error -1 ppm.



Fig. S9 ¹H NMR spectrum (600 MHz, CDCl₃, 298 K) of 1.



Fig. S10 ¹³C NMR spectrum (150 MHz, CDCl₃, 298 K) of 1.



Fig. S11 LRESIMS spectrum of 1 in CHCl₃



Fig. S12 HRESIMS spectrum of 1 in CHCl₃.

3. pH-dependent catechol-Fe³⁺ coordination



Scheme S2 pH-dependent catechol-Fe³⁺ coordination.

4. 2D NOESY NMR spectrum of a mixture of 1 and 2



Fig. S13 2D NOESY NMR spectrum (600 MHz, 298 K) in CDCl₃/CD₃OD (*v* : *v* = 10 : 1) of 10.0 mM **1** and 5.00 mM **2**.

5. MALDI-TOF mass spectrum of a mixture of compound 1 and BPO



Fig. S14 MALDI-TOF mass spectrum of 1 (1.00 mM) and BPO (2.00 mM) after mixing for 3 hours.



Fig. S15 MALDI-TOF mass spectrum of 1 (1.00 mM) and BPO (2.00 mM) after mixing for 12 hours.

6. Concentration-variant 2D DOSY NMR spectra of $1 \cdot Fe^{3+} \cdot 2$



Fig. S16 2D DOSY NMR spectrum (600 MHz, 298 K) of CHCl₃ in CDCl₃ (99.9%).



Fig. S17 2D DOSY NMR spectrum (600 MHz, 298 K) of polymer **a** at 50.0 mM of **1** in CDCl₃/CD₃OD (v : v = 10 : 1).



Fig. S18 2D DOSY NMR spectrum (600 MHz, 298 K) of polymer **a** at 5.00 mM of 1 in $CDCl_3/CD_3OD$ (v : v = 10 : 1).

We first measured the diffusion constant *D* of CHCl₃ in CDCl₃ (99.9%) resulting in a value of 52.5×10^{-10} m²/s at 298 K. And then 2D DOSY NMR experiments on polymer **a** at 50.0 mM and 5.00 mM of **1** in CDCl₃/CD₃OD (v : v = 10 : 1) were carried out, respectively. The values for the diffusion constant *D* of residual CHCl₃ in solution deviated from the diffusion constant *D* measured in CDCl₃ (99.9%). Therefore, we corrected the measured values according to the measured value of the diffusion constant *D* of CHCl₃ in CDCl₃ (99.9%).^{S3}



Fig. S19 Concentration dependence of diffusion coefficient *D* (600 MHz, 298 K) in $CDCl_3/CD_3OD$ (v : v = 10 : 1) of polymer **a** at different concentrations of **1**: 50.0 mM, 5.00 mM.



7. Digital photos of supramolecular glues

Fig. S20 Digital photos of the supramolecular glues: (a) glue a; (b) glue b.



8. The temperature-variant NMR spectra of a mixture of 1 and 2

Fig. S21 ¹H NMR spectra (600 MHz) of a 1 : 2 molar ratio mixture of 2 and 1 at 5.00 mM 1 in CDCl₃/CD₃OD (v : v = 10 : 1) at various temperatures: (a) 298 K; (b) 303 K;
(c) 308 K; (d) 313 K; (e) 318 K; (f) 323 K; (g) 328 K.

9. References

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