

## Supporting Information

### Effect of number and position of methoxy substituents on triphenylamine-based chalcone visible-light-absorbing photoinitiators

**Shih-Chieh Yen<sup>a</sup>, Zhong-Han Lee<sup>a</sup>, Jen-Shyang Ni<sup>a</sup>, Ching-Chin Chen<sup>b</sup>, Yung-Chung Chen<sup>a,c\*</sup>**

<sup>a</sup>. Department of Chemical and Materials Engineering, National Kaohsiung University of Science and Technology, Kaohsiung 80778, Taiwan

<sup>b</sup>. Department of Chemistry, National Chung Hsing University, Taichung, 40227, Taiwan

<sup>c</sup>. Photo-SMART (Photo-sensitive Material Advanced Research and Technology Center), National Kaohsiung University of Science and Technology, Kaohsiung City 80778, Taiwan

\*Corresponding authors: Y. C. Chen ([chenyc@nkust.edu.tw](mailto:chenyc@nkust.edu.tw))

## 1. Synthesis

### *4-Acetylphenyl)diphenylamine (CYA)*

TPA (17.19 g, 0.07 mol), anhydrous zinc chloride (10.19 g, 0.075 mol), and acetyl chloride (4.5 mL, 0.064 mol) were dissolved in dichloromethane (DCM) (40 mL). The solution was stirred at room temperature for 10 min and then heated to 60 °C for 24 h. Once the reaction was complete, a cooled solution comprising HCl<sub>(aq)</sub> (2 M) was added to terminate the reaction, and the solution was then extracted using DCM. The organic extract was collected and dried over anhydrous MgSO<sub>4</sub>. Finally, the crude residue was purified through silica gel chromatography (using a DCM/n-hexane = 1/1 eluent) to afford a light yellow powder of mass 8.93 g (yield = 44%). <sup>1</sup>H nuclear magnetic resonance (NMR; 400 MHz, CDCl<sub>3</sub>, δ, ppm): 7.81–7.78 (d, 2H, *J* = 9.2 Hz, Ar–H), 7.32–7.28 (4H, Ar–H), 7.16–7.08 (6H, Ar–H), 7.05–6.97 (2H, Ar–H), 2.53 (3H, C=O–H).

### *1-(4-(bis(4-methoxyphenyl)amino)phenyl)ethenone (CYB)*

**CYB** (yield = 60%) was synthesized using a similar procedure to that used to obtain **CYA** and was obtained as a yellow compound. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 7.76–7.74 (d, 2H, *J* = 9.2 Hz, Ar–H), 7.13–7.09 (4H, Ar–H), 6.90–6.86 (4H, Ar–H), 6.83–6.79 (2H, Ar–H), 3.81 (6H, OCH<sub>3</sub>), 2.53 (3H, C=O–H).

### *CY1–CY5 photoinitiators*

Taking **CY1** as an example. **CYA** (0.5g, 1.74 mmol), **CYC** (0.5g, 1.83 mmol), and NaOH (0.0836g, 2.09 mmol) were dissolved in mixing solvent comprising EtOH (20 mL) and H<sub>2</sub>O (10 mL). The solution was stirred at room temperature for 10 min and then heated to a reflux temperature of 90 °C for 5 h. Once the reaction was complete, the mixture was extracted using DCM, and the organic extract was collected and dried over anhydrous MgSO<sub>4</sub>. The crude product was purified through silica gel chromatography (using a DCM/n-hexane = 1/2 eluent) to afford the orange powder of interest in mass 0.34 g (yield = 34%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 7.90–7.88 (d, 2H, *J* = 9.2 Hz, Ar–H), 7.78–7.74 (1H, C=C–Ar), 7.49–7.47 (2H, *J* = 8.8 Hz, Ar–H), 7.41–7.38 (1H, Ar–C=O–H), 7.34–7.27 (8H, Ar–H), 7.18–7.07 (12H, Ar–H), 7.04–7.01 (4H, Ar–H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ, ppm): 188.2, 151.8, 149.9, 146.9, 146.5, 143.4, 131.1, 130.0,

129.6, 129.5, 129.4, 128.2, 125.9, 125.3, 124.5, 124.0, 121.7, 120.0, 119.3. FT-Mass (m/z) calcd. 543.24309; found 543.24306.

**CY2** (yield = 22%, orange powder), **CY3** (yield = 27%, orange powder), **CY4** (yield = 48%, orange powder), and **CY5** (yield = 12%, orange powder) were synthesized using a similar procedure to that for **CY1**. **CY2**:  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 7.90–7.88 (d, 2H,  $J = 8.8$  Hz, Ar–H), 7.77–7.73 (s, 1H, C=C–Ar), 7.46–7.44 (1H, Ar–C=O–H), 7.46–7.25 (8H, Ar–H), 7.17–6.95 (15H, Ar–H), 6.88–6.86 (2H, Ar–H), 3.82 (3H,  $-\text{OCH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 188.3, 156.8, 151.8, 150.2, 147.0, 146.6, 143.5, 139.6, 131.2, 129.9, 129.5, 129.3, 127.9, 127.4, 125.8, 124.6, 124.5, 123.5, 120.4, 120.0, 118.8, 114.9, 55.5. FT-Mass (m/z) calcd. 573.25365; found 573.25373.

**CY3**:  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 7.90–7.87 (d, 2H,  $J = 8.8$  Hz, Ar–H), 7.76–7.73 (s, 1H, C=C–Ar), 7.44–7.42 (1H, Ar–C=O–H), 7.37–7.26 (6H, Ar–H), 7.17–7.02 (12H, Ar–H), 6.88–6.85 (6H, Ar–H), 3.81 (6H,  $-\text{OCH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 188.3, 156.6, 151.7, 150.7, 146.6, 143.8, 139.7, 131.3, 129.9, 129.6, 129.5, 127.4, 126.4, 125.8, 124.4, 120.0, 118.7, 118.2, 114.8, 55.5. FT-Mass (m/z) calcd. 603.26422; found 603.26451.

**CY4**:  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 7.87–7.85 (2H, Ar–H), 7.76–7.72 (2H, C=C–Ar, Ar–C=O–H), 7.48–7.37 (2H, Ar–H), 7.31–7.26 (6H, Ar–H), 7.14–7.10 (6H, Ar–H), 7.10–7.01 (4H, Ar–H), 6.89–6.85 (4H, Ar–H), 6.82–6.80 (2H, Ar–H), 3.81 (6H,  $-\text{OCH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 188.0, 157.0, 156.9, 152.9, 152.6, 149.7, 147.7, 146.9, 143.0, 139.3, 139.2, 130.1, 129.9, 129.5, 129.4, 129.3, 129.2, 128.4, 128.2, 128.1, 127.8, 127.7, 125.3, 125.0, 124.1, 124.0, 123.9, 123.6, 122.7, 121.8, 119.4, 117.1, 116.8, 114.9, 114.8, 55.4. FT-Mass (m/z) calcd. 603.26422; found 603.26396.

**CY5**:  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 7.86–7.84 (d, 2H,  $J = 8.8$  Hz, Ar–H), 7.74–7.71 (1H, C=C–Ar), 7.43–7.40 (2H, Ar–H), 7.37–7.33 (1H, Ar–C=O–H), 7.13–7.07 (8H, Ar–H), 6.89–6.85 (12H, Ar–H), 3.81–3.80 (12H,  $-\text{OCH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ , ppm): 188.1, 156.9, 156.6, 152.5, 150.6, 143.3, 139.8, 139.4, 130.0, 129.6, 129.5, 127.7, 127.3, 126.6, 118.8, 118.4, 117.2, 114.9, 114.8, 55.5. FT-Mass (m/z) calcd. 663.28535; found 663.28558.

## 2. Measurement

The nuclear magnetic resonance (NMR) spectra of the samples were dissolved in deuterated chloroform and analyzed under room temperature by using an Agilent Unity plus-400 spectrometer. Fourier-transform mass spectrometry was performed using a JEOL AccuTOF GCx-plus instrument. The light absorption of the samples in dichloromethane (DCM) (concentration =  $1 \times 10^{-5}$  M) was measured using a PerkinElmer Lambda 35 UV-visible (UV-Vis) spectrometer. The photoluminescence (PL) spectra of samples in DCM (concentration =  $1 \times 10^{-5}$  M) were recorded using a Hitachi F-4500 spectrometer at an excitation wavelength of 420 nm. The highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) of the photoinitiators were optimized using the UB3LYP/6-31G\* level of theory. In addition, the geometries were frequency checked using Gaussian 09 software.<sup>1</sup> Cyclic voltammetry (CV) was performed using a BioLogic SP-150 model at a scan rate of  $100 \text{ mV s}^{-1}$  in the range of 0 to  $-2 \text{ V}$ . All measurements were conducted at room temperature in DCM solution (concentration =  $1 \times 10^{-3}$  M) with a conventional three-electrode configuration, in which the three electrodes were a platinum working electrode, a platinum wire auxiliary electrode, and a Ag/Ag<sup>+</sup> reference electrode. A Perkin Elmer DSC 6000A was used to determine the melting point ( $T_m$ ) under nitrogen atmosphere with a scan rate of  $10 \text{ }^\circ\text{C min}^{-1}$ . Thermogravimetric analysis (TGA) was performed using a TA Instruments SDT Q600 Simultaneous DTA-TGA device under nitrogen atmosphere at a heating rate of  $15 \text{ }^\circ\text{C min}^{-1}$ ; the samples analyzed had a mass of 3–5 mg. The thermal degradation temperature ( $T_d$ ) was taken as the temperature at which 5% weight loss had occurred.

### 3. Photolysis experiment

Photolysis was performed with the selected photoinitiation packages, including CYs (1 wt%) and TEOA (4 wt%) in DCM as the solvent (concentration =  $1 \times 10^{-5}$  M). The absorption intensity of the solution was determined with various durations of exposure under a Philips 16 W lamp (type Actinic BL;  $\lambda = 365 \text{ nm}$ ). The results were recorded using a PerkinElmer Lambda 35 UV-Vis spectrometer.

### 4. Electron spinning resonance (ESR) measurement

ESR experiments were conducted using a Bruker EMX Plus X-Band spectrometer. Ultra-high-pressure mercury (MUV-250U-L,  $\lambda = 250\text{--}450 \text{ nm}$ , intensity =  $5 \text{ mW}$ ) was selected as the radiation

source used to generate radicals at room temperature under nitrogen atmosphere. The radicals were trapped by N-tert-Butyl- $\alpha$ -phenylnitron (PBN), and *tert*-butylbenzene as a solvent in accordance with a procedure described in the literature.<sup>2</sup> The concentration of PBN and **CYs** was  $1 \times 10^{-2}$  and  $1 \times 10^{-3}$  M, respectively. In addition, the ratios between **CY** and TEOA or EDB was 1:4 wt%.

## 5. Photo-differential scanning photocalorimetry (Photo-DSC) experiment

The photopolymerization efficiency of the formulations was tested through photo-DSC analysis, performed using a Perkin Elmer DSC 6000 at 30 °C under nitrogen atmosphere (flow rate = 20 mL min<sup>-1</sup>). A UV lamp (intensity = 180 mW cm<sup>-2</sup>;  $\lambda$  = 250–450 nm) or LED@405 nm (intensity = 180 mW cm<sup>-2</sup>) was used as the radiation source. **CY** (the hydrogen acceptor), TEOA (the hydrogen donor), and TMPTA were mixed without additional solvent to obtain a photoinitiation system. Approximately 15 mg of a sample mixture (containing the hydrogen acceptor, hydrogen donor, and TMPTA under a certain weight ratio) was placed in an aluminum DSC pan, which was used to ensure the different formulations had similar thickness. Heat flow versus time curves were recorded to analyze the reaction of unsaturated moieties in the system during light irradiation. By integrating the area under the exothermic peak, the double bond conversion efficiency (DC; %) could be calculated using the following equation:<sup>3</sup>

$$DC = (\Delta H_t / \Delta H_o^{\text{theor}}) \times 100\%,$$

where  $\Delta H_t$  is the total reaction heat enthalpy within the exposure time, and  $\Delta H_o^{\text{theor}}$  is the theoretical reaction heat enthalpy of one acrylate for complete conversion.  $\Delta H_o^{\text{theor}}$  was 86 kJ mol<sup>-1</sup>.<sup>4</sup>

Furthermore, the rate of polymerization ( $R_p$ ) is directly related to the heat flow ( $dH/dt$ ) by the equation  $R_p = dC/dt$ .<sup>5</sup>

1. Æ. F. James B. Foresman. *Exploring Chemistry with Electronic Structure Methods*, Gaussian Inc., Pittsburgh, PA, 1996.
2. T. L. Huang, Y. H. Li and Y. C. Chen, *Journal of Polymer Science*, 2020, **58**, 2914-2925.
3. X. S. Jiang and J. Yin, *Macromolecules*, 2004, **37**, 7850-7853.
4. E. Andrzejewska and M. Andrzejewski, *Journal of Polymer Science Part a-Polymer Chemistry*, 1998, **36**, 665-673.
5. S. X. Chen, C. Qin, M. Jin, H. Y. Pan and D. C. Wan, *Journal of Polymer Science*, 2021, **59**, 578-593.

CY1  
 Pulse Sequence: s2pul  
 UNITYplus-400 "unity400"  
 Date: Aug 10 2020  
 Solvent: CDCl3  
 Ambient Temperature  
 Total 120 repetitions



Fig. S1 <sup>1</sup>H NMR of the CY-1.

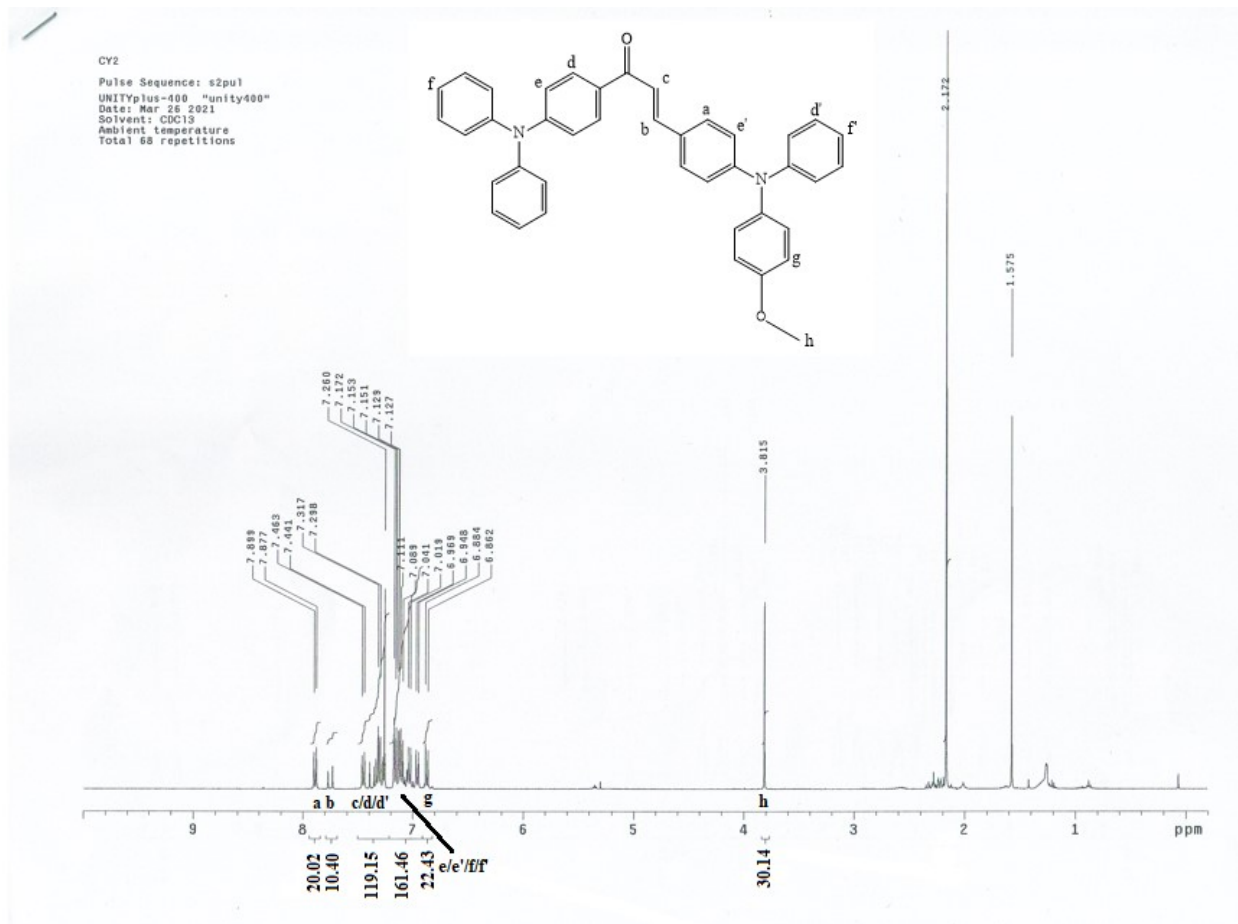


Fig. S2 <sup>1</sup>H NMR of the CY-2.

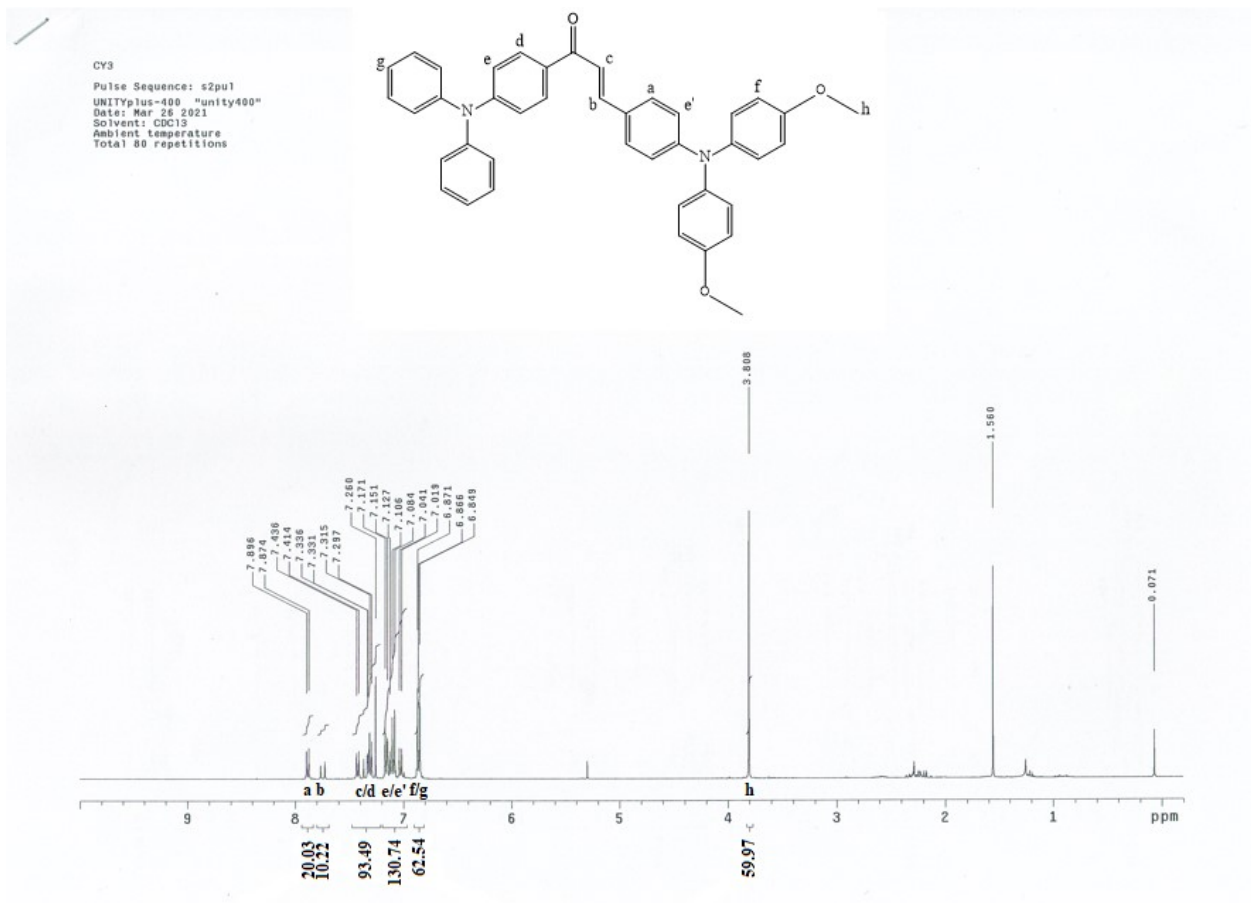


Fig. S3 <sup>1</sup>H NMR of the CY-3.



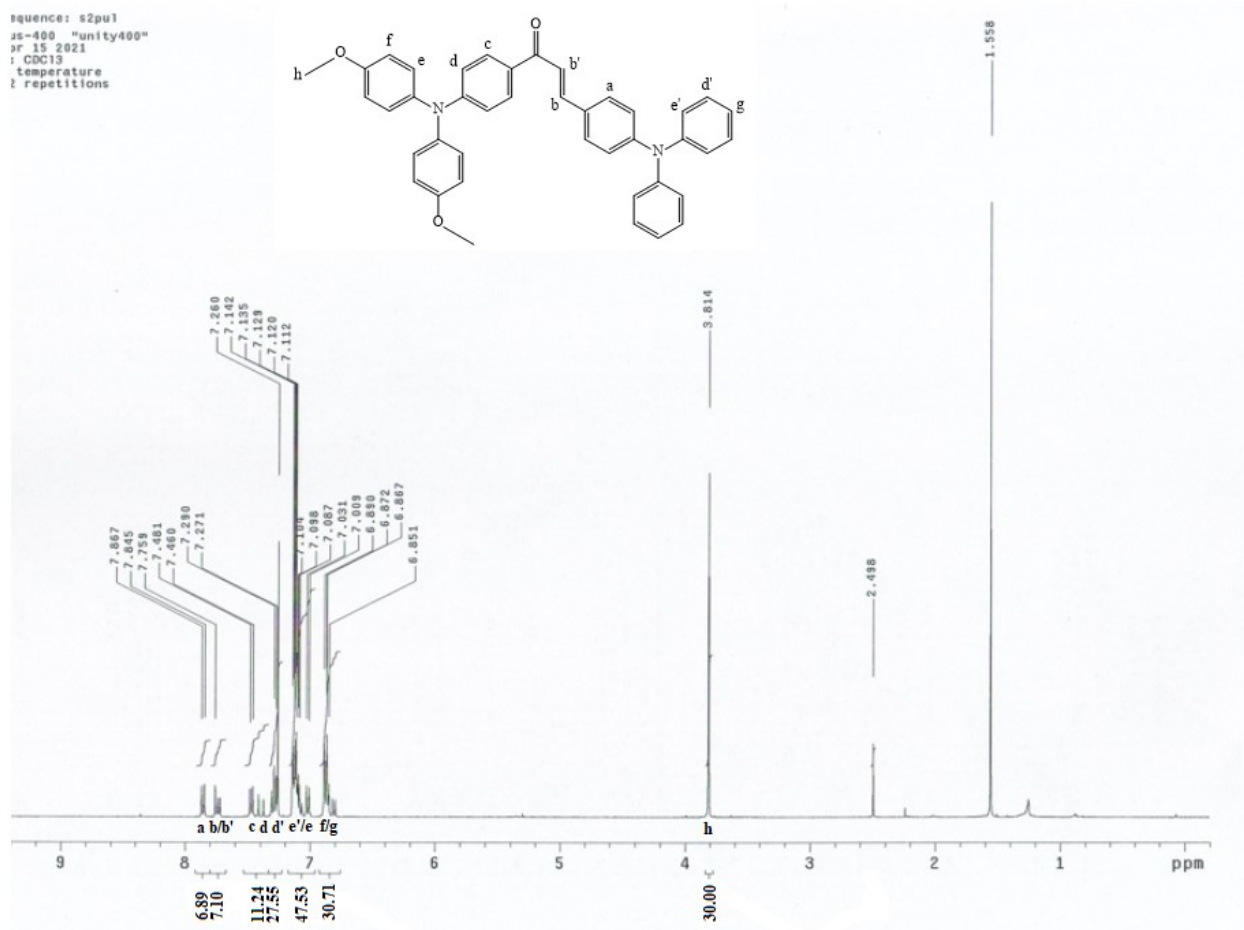


Fig. S4  $^1\text{H}$  NMR of the CY-4.

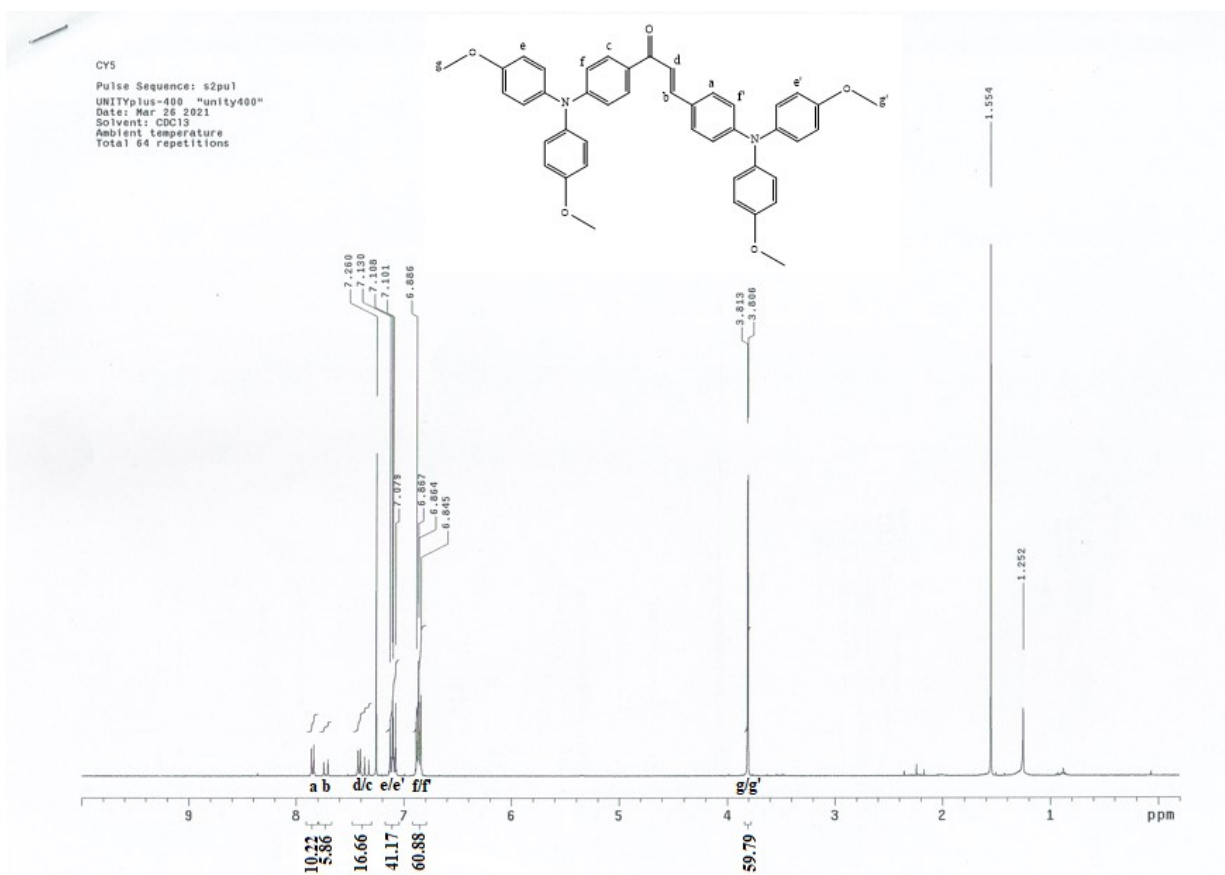


Fig. S5  $^1\text{H}$  NMR of the CY-5.

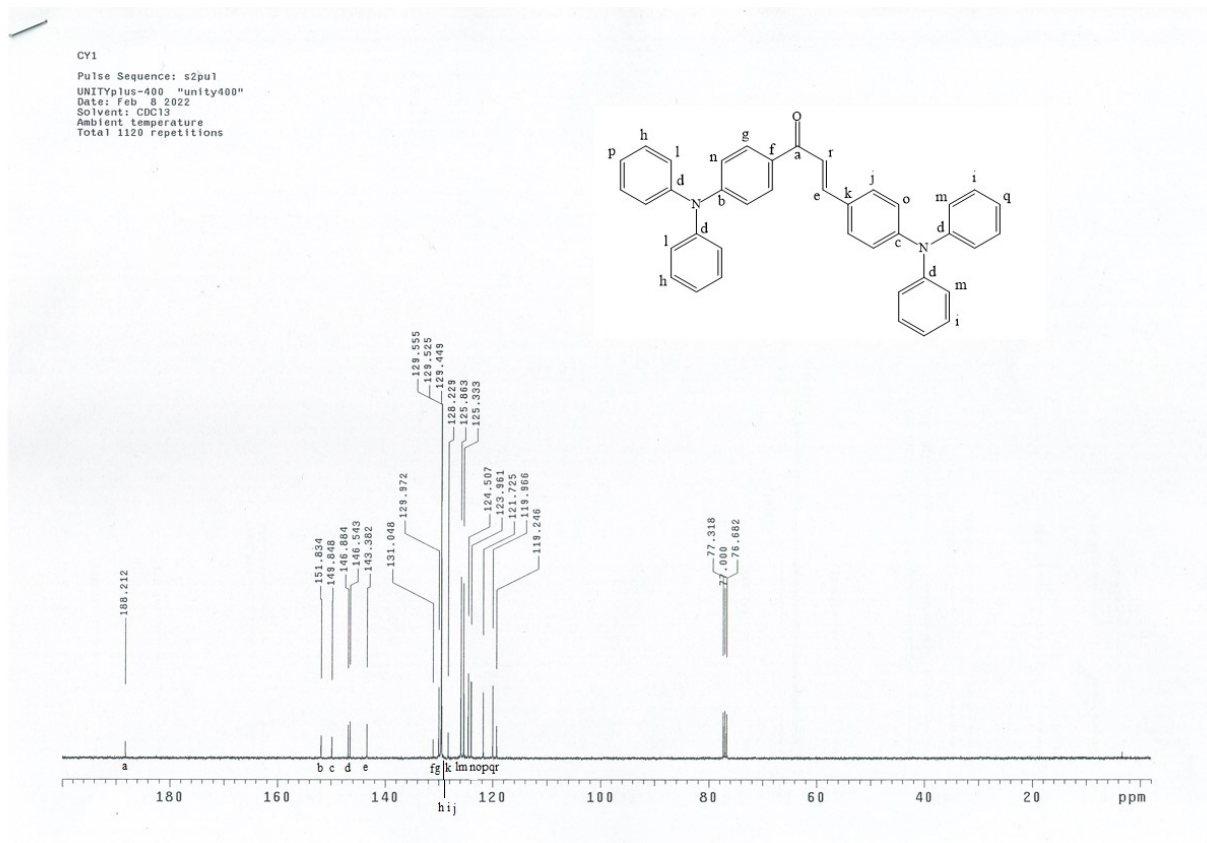


Fig. S6  $^{13}\text{C}$  NMR of the CY1.

CY2  
Pulse Sequence: s2pu1  
UNITYplus-400 "unity400"  
Date: Feb 8 2022  
Solvent: CDCl3  
Ambient temperature  
Total 1350 repetitions

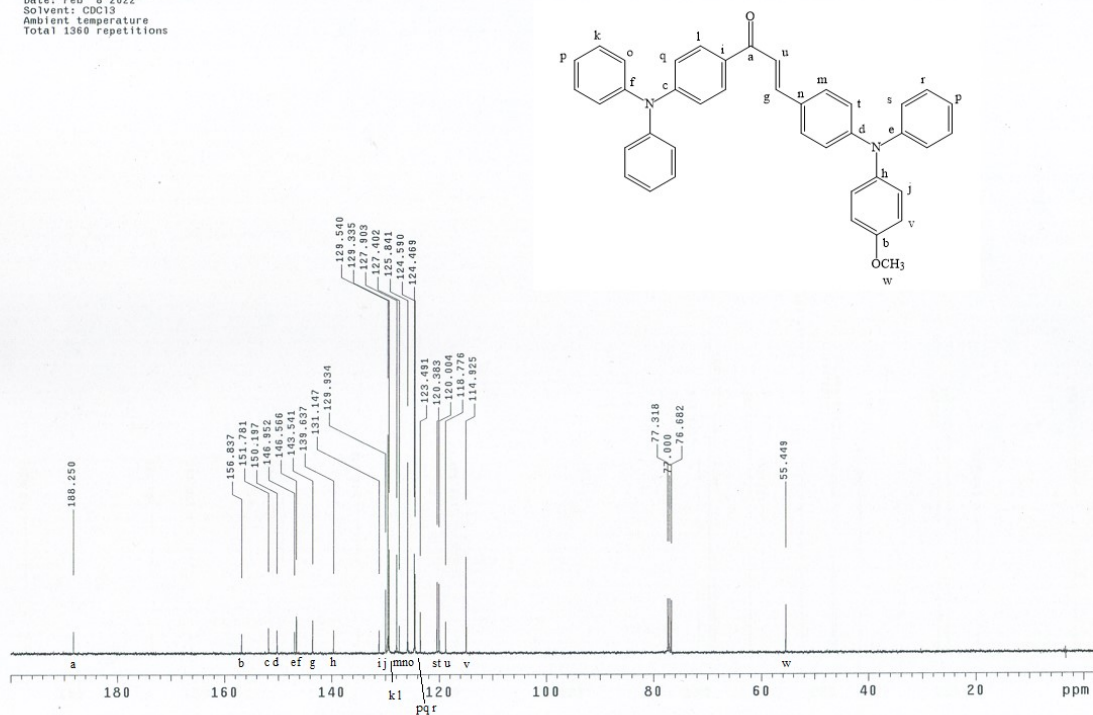


Fig. S7  $^{13}\text{C}$  NMR of the CY2.

CY3

Pulse Sequence: s2pu1  
UNITYplus-400 "unity400"  
Date: Feb 8 2022  
Solvent: CDCl3  
Ambient temperature  
Total 556 repetitions

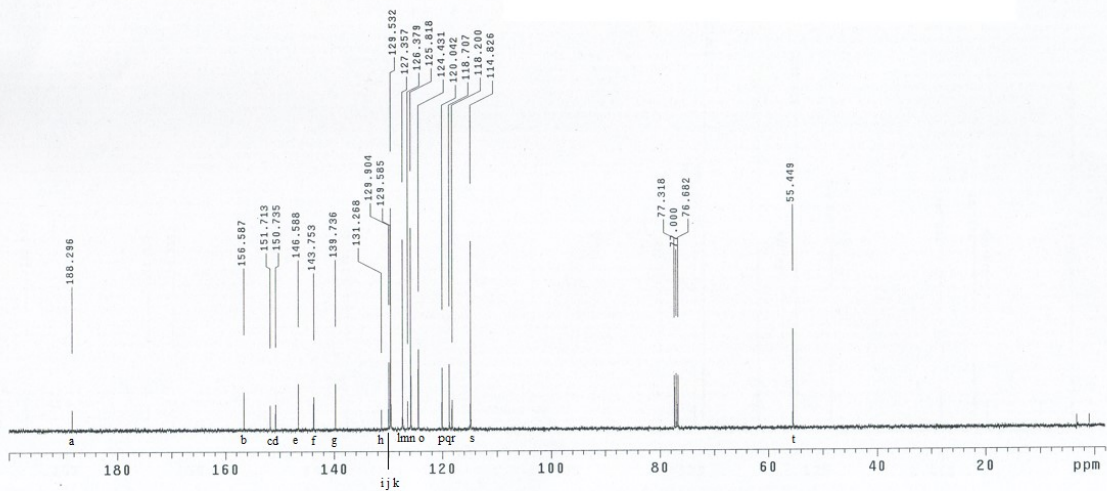
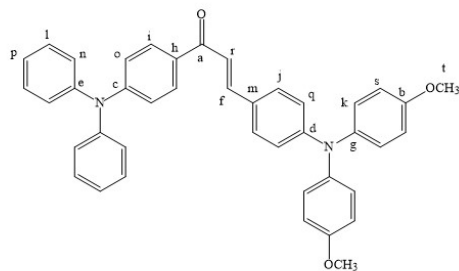


Fig. S8 <sup>13</sup>C NMR of the CY3.

CY4  
 Pulse Sequence: s2pul  
 UNITYplus-400 "unity400"  
 Date: Feb 18 2022  
 Solvent: CDCl<sub>3</sub>  
 Ambient Temperature  
 Total 656 repetitions

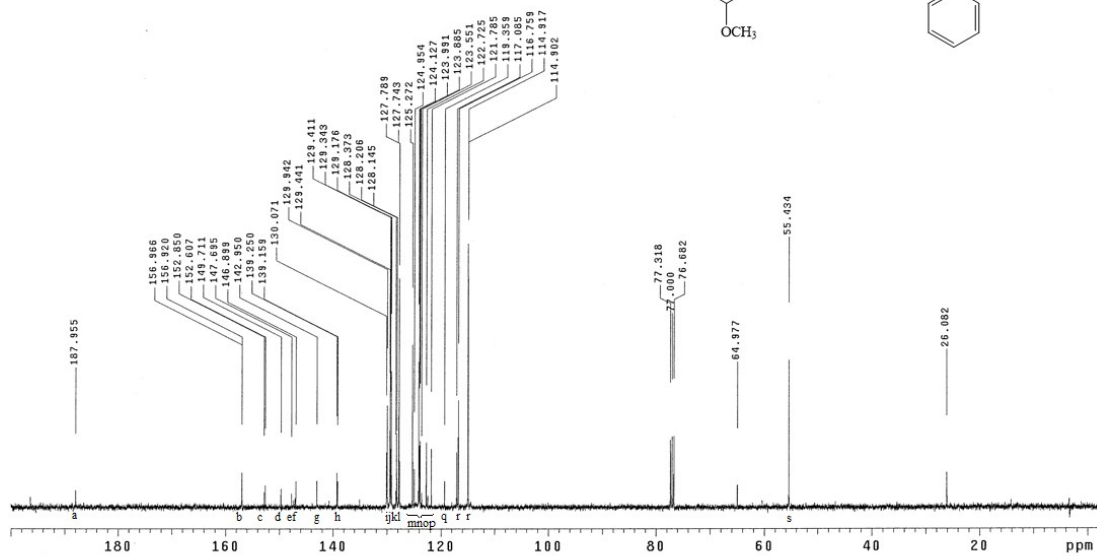
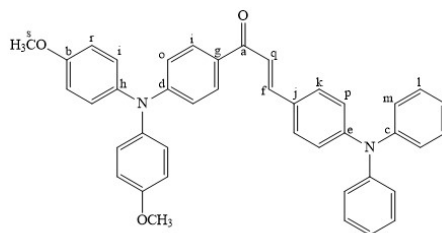


Fig. S9 <sup>13</sup>C NMR of the CY4.

CY5  
Pulse Sequence: s2pu1  
UNITYplus-400 "unity400"  
Date: Feb 8 2022  
Solvent: CDCl3  
Ambient temperature  
Total 1200 repetitions

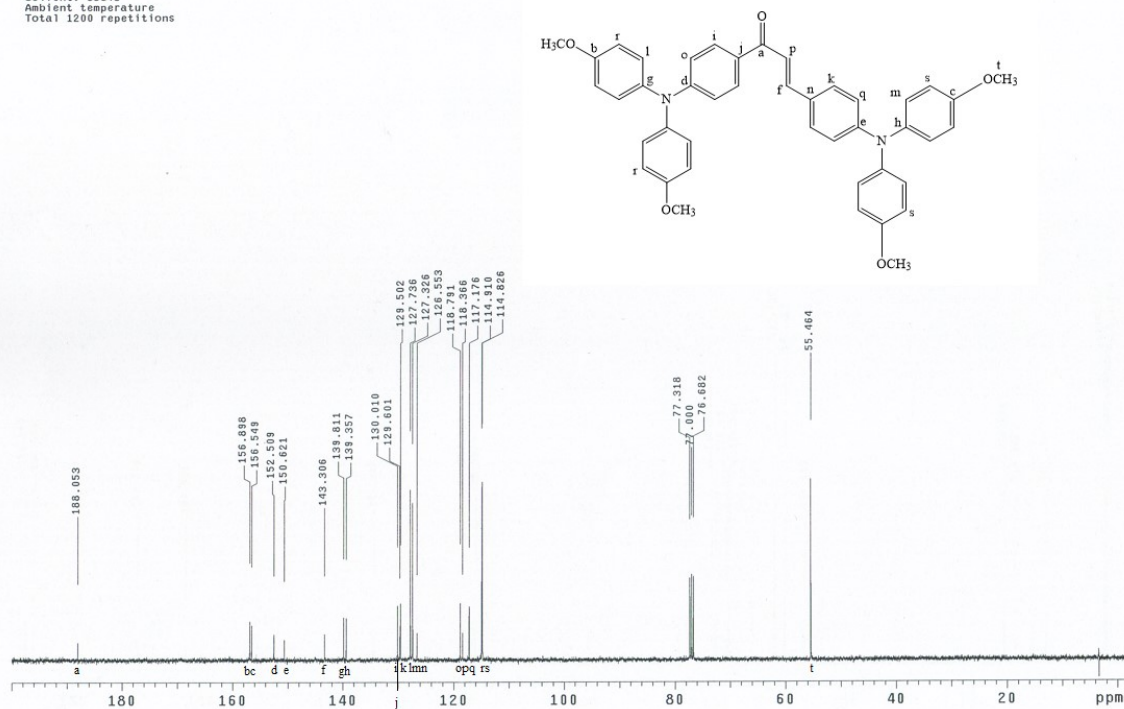
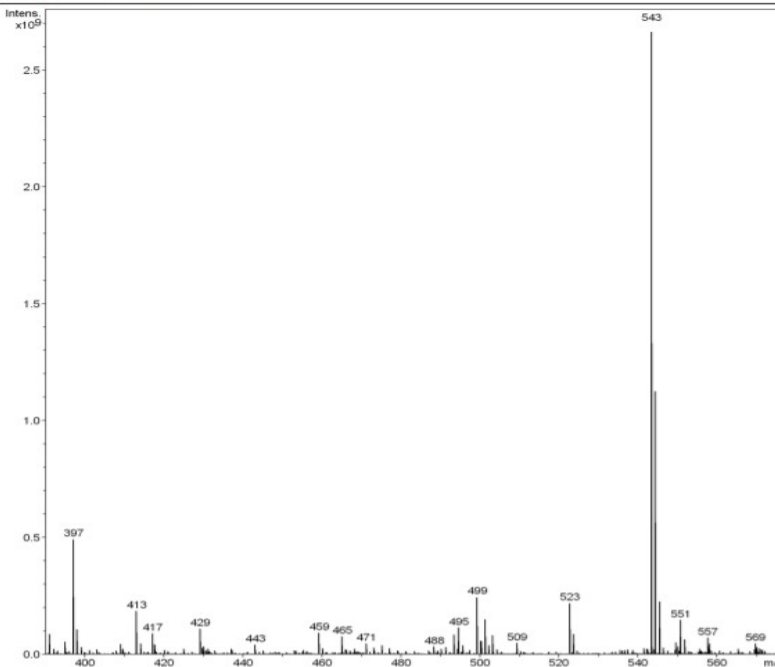


Fig. S10 <sup>13</sup>C NMR of the CY5.

FT-MS

Analysis Info

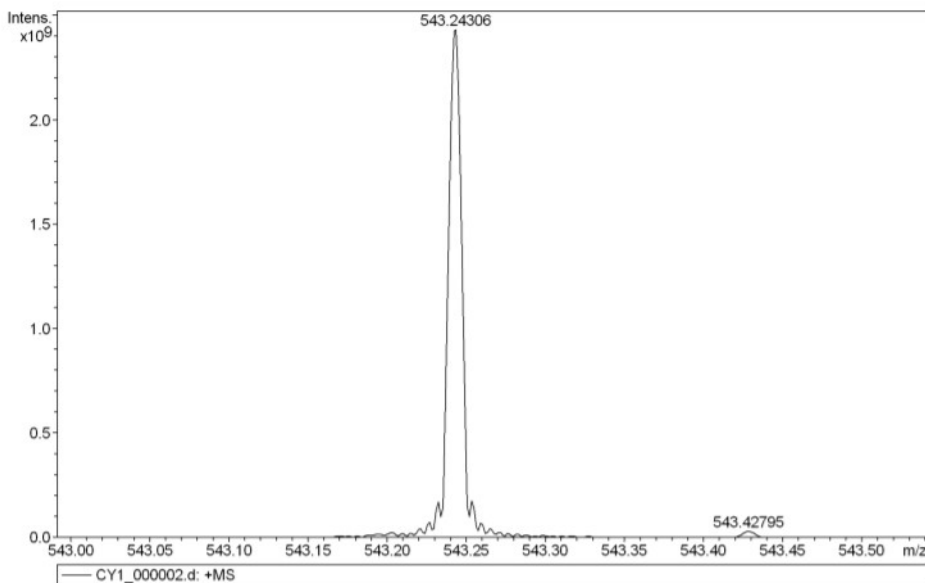
Analysis Name D:\Data\g8\CY1\_000003.d 9/9/2020 4:29:29 PM  
 Method broadband first signal  
 Sample Name CY1 Instrument: FT-MS solarix  
 Comment ESI Positive



Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name D:\Data\g8\CY1\_000002.d 9/9/2020 4:28:39 PM  
 Method broadband first signal Operator: YU HSIAO-CHING  
 Sample Name CY1 Instrument: BRUKER FT-MS solarix  
 Comment ESI Positive



Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
543.24306	1	C <sub>39</sub> H <sub>31</sub> N <sub>2</sub> O	100.00	543.24309	0.03	0.05	7.5	25.5	even	ok

Fig. S11 HR-MS of the CY-1

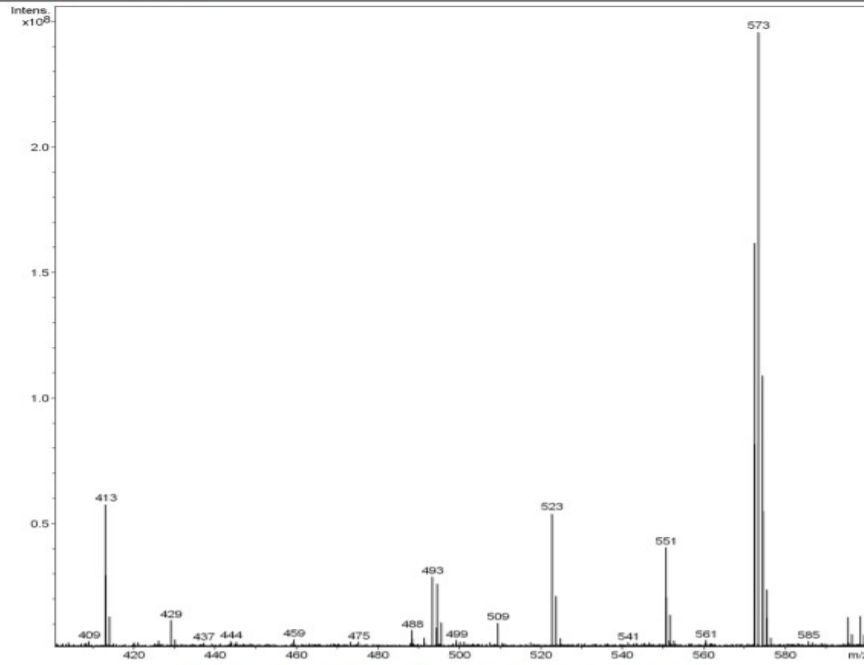


FT-MS

Analysis Info

Analysis Name D:\Data\g8\CY2\_000003.d  
 Method broadband first signal  
 Sample Name CY2  
 Comment ESI Positive

10/15/2020 3:54:33 PM  
 Instrument: FT-MS solariX

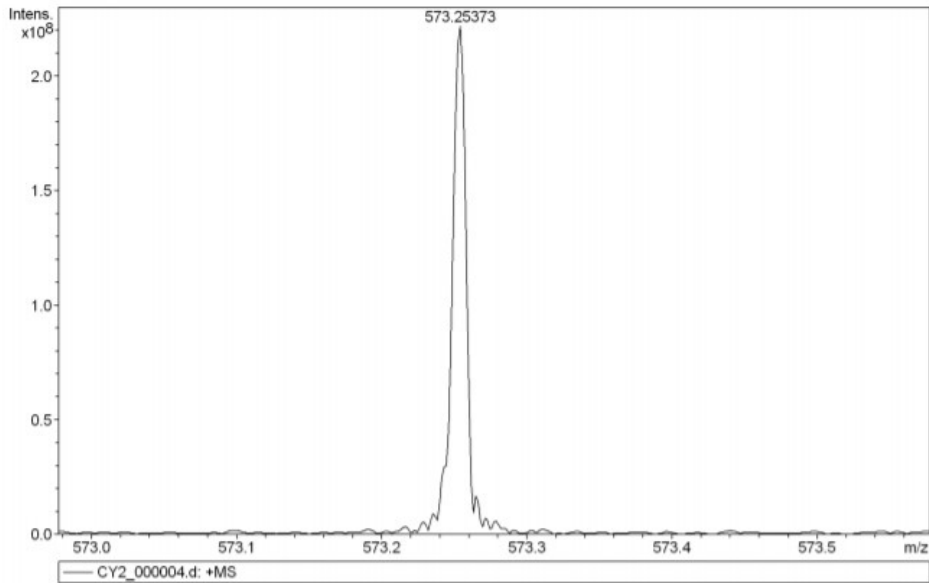


Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name D:\Data\g8\CY2\_000004.d  
 Method broadband first signal  
 Sample Name CY2  
 Comment ESI Positive

10/15/2020 3:53:34 PM  
 Operator: YU HSIAO-CHING  
 Instrument: BRUKER FT-MS solariX



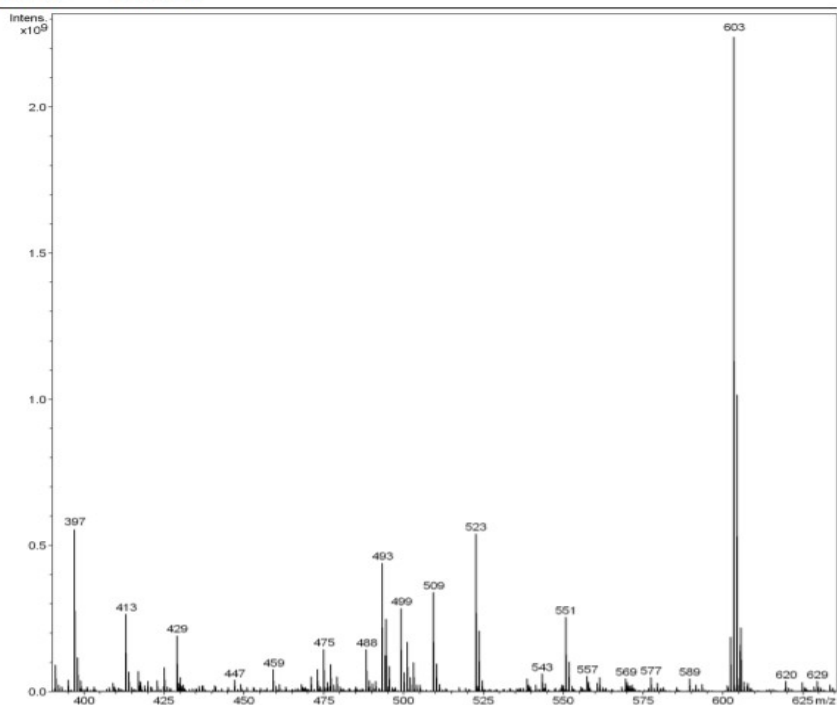
Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
573.25373	1	C <sub>40</sub> H <sub>33</sub> N <sub>2</sub> O <sub>2</sub>	100.00	573.25365	-0.08	-0.14	3.4	25.5	even	ok

Fig. S12 HR-MS of the CY-2.

FT-MS

Analysis Info

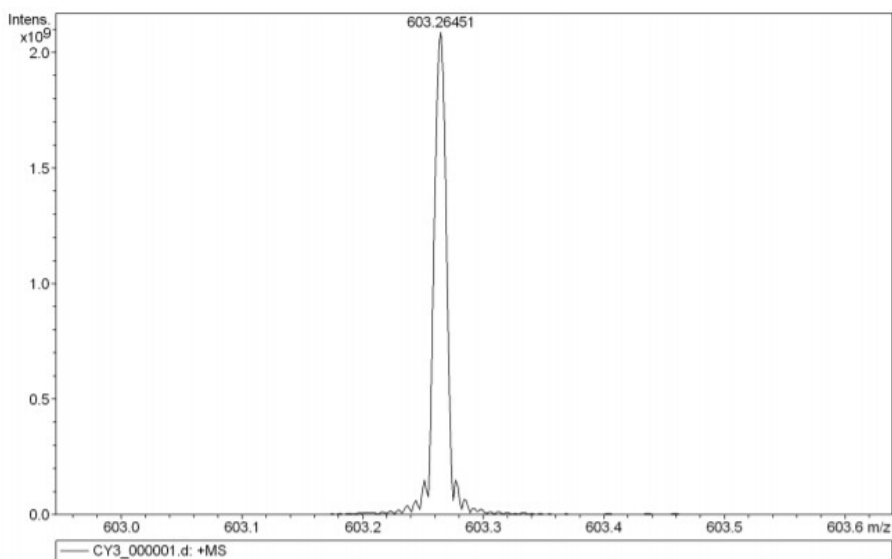
Analysis Name D:\Data\g8\CY3\_000003.d 9/9/2020 4:34:15 PM  
 Method broadband first signal  
 Sample Name CY3 Instrument: FT-MS solariX  
 Comment ESI Positive



Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name D:\Data\g8\CY3\_000001.d 9/9/2020 4:35:41 PM  
 Method broadband first signal Operator: YU HSIAO-CHING  
 Sample Name CY3 Instrument: BRUKER FT-MS solariX  
 Comment ESI Positive

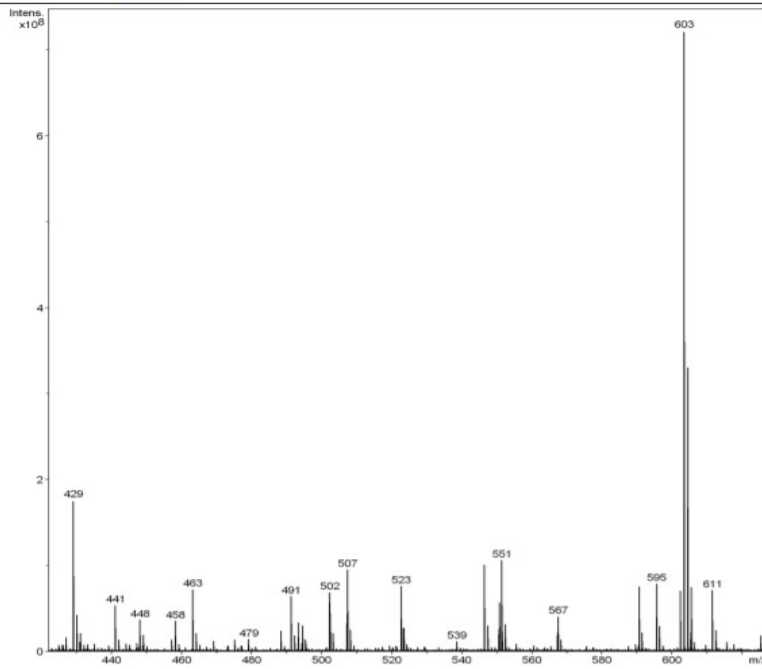


Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
603.26451	1	C 41 H 35 N 2 O 3	100.00	603.26422	-0.29	-0.48	8.3	25.5	even	ok

Fig. S13 HR-MS of the CY-3.

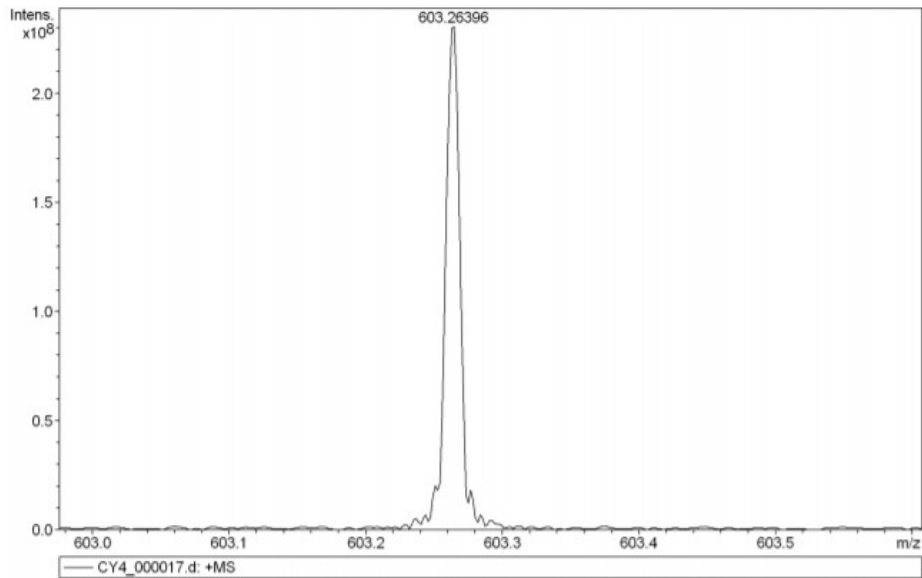
FT-MS

**Analysis Info**  
Analysis Name D:\Data\g8\CY4\_000003.d  
Method broadband first signal  
Sample Name CY4  
Comment ESI Positive  
10/15/2020 3:36:53 PM  
Instrument: FT-MS solariX



Mass Spectrum SmartFormula Report

**Analysis Info**  
Analysis Name D:\Data\g8\CY4\_000017.d  
Method broadband first signal  
Sample Name CY4  
Comment ESI Positive  
10/15/2020 3:47:13 PM  
Operator: YU HSIAO-CHING  
Instrument: BRUKER FT-MS solariX



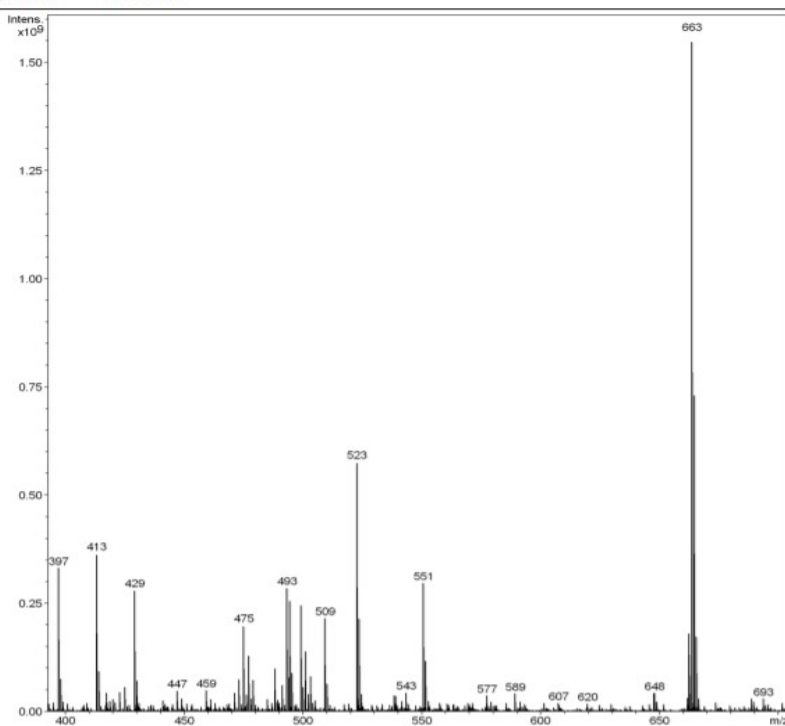
Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
603.26396	1	C 41 H 35 N 2 O 3	100.00	603.26422	0.26	0.43	5.8	25.5	even	ok

Fig. S14 HR-MS of the CY-4.

FT-MS

Analysis Info

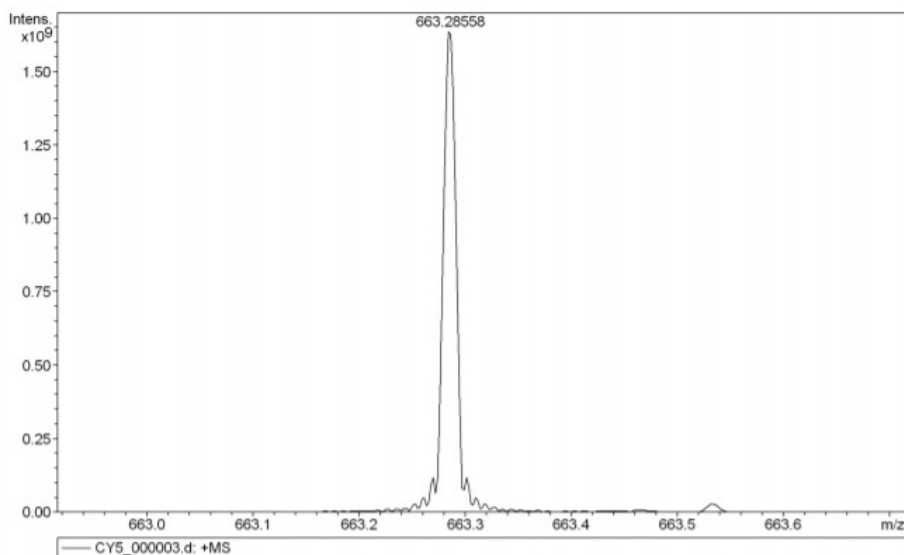
Analysis Name D:\Data\g8\CY5\_000001.d 9/9/2020 4:41:29 PM  
 Method broadband first signal  
 Sample Name CY5 Instrument: FT-MS solarIX  
 Comment ESI Positive



Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name D:\Data\g8\CY5\_000003.d 9/9/2020 4:42:51 PM  
 Method broadband first signal Operator: YU HSIAO-CHING  
 Sample Name CY5 Instrument: BRUKER FT-MS solarIX  
 Comment ESI Positive



Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
663.28558	1	C 43 H 39 N 2 O 5	100.00	663.28535	-0.23	-0.35	4.4	25.5	even	ok

Fig. S15 HR-MS of the CY-5.

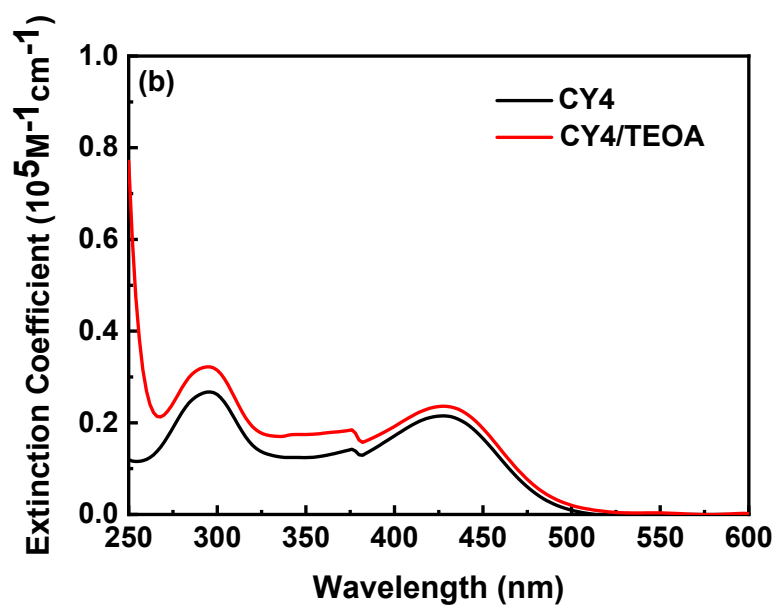
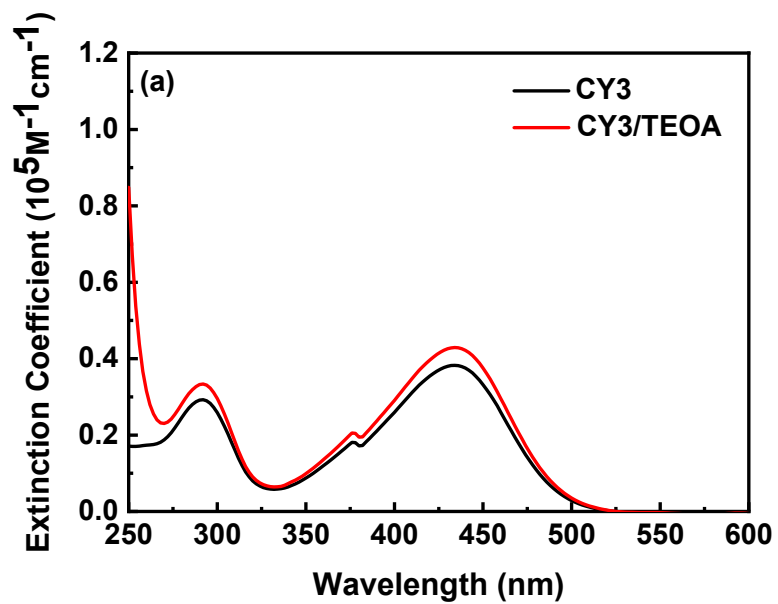
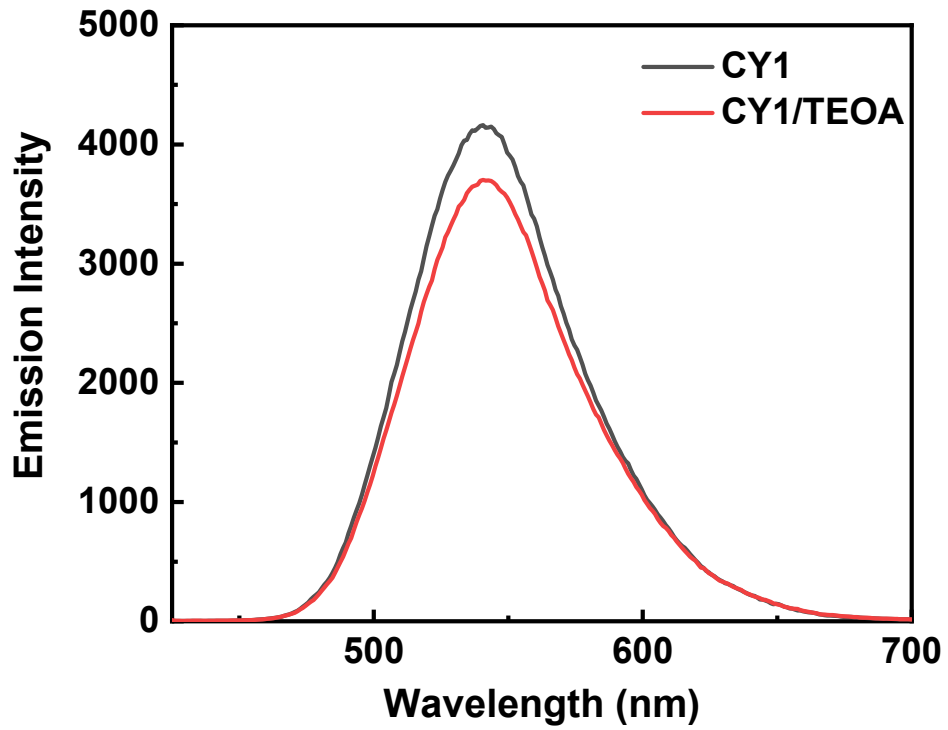


Fig. S16 UV-Vis spectra of (a) CY3, CY3/TEOA and (b) CY4, CY4/TEOA in DCM solution



**Fig. S17** PL spectra of **CY1** and **CY1/TEOA** (1:4 wt%) in DCM solutions (Conc. =  $1 \times 10^{-5}$  M).

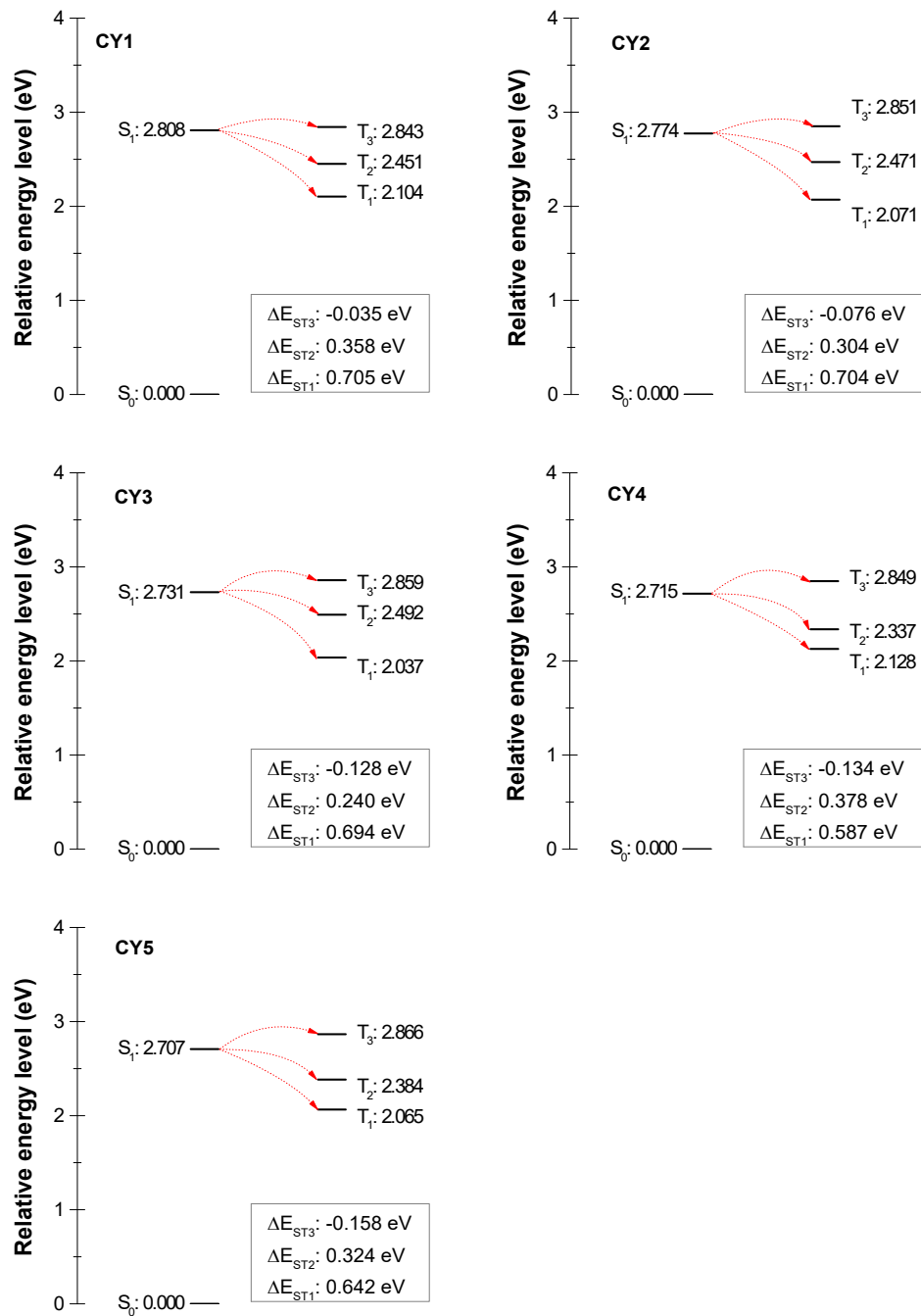
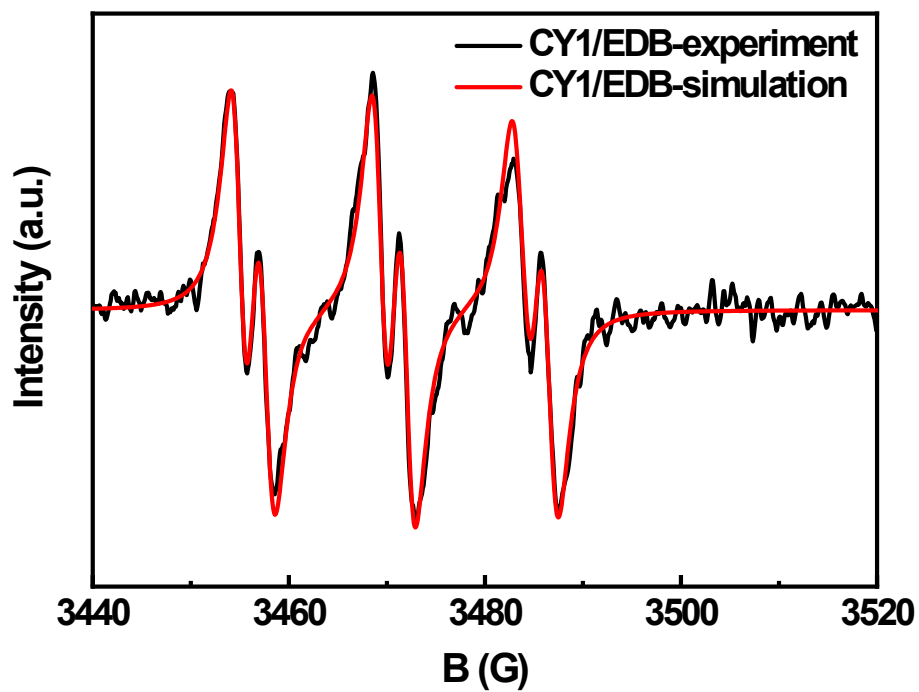
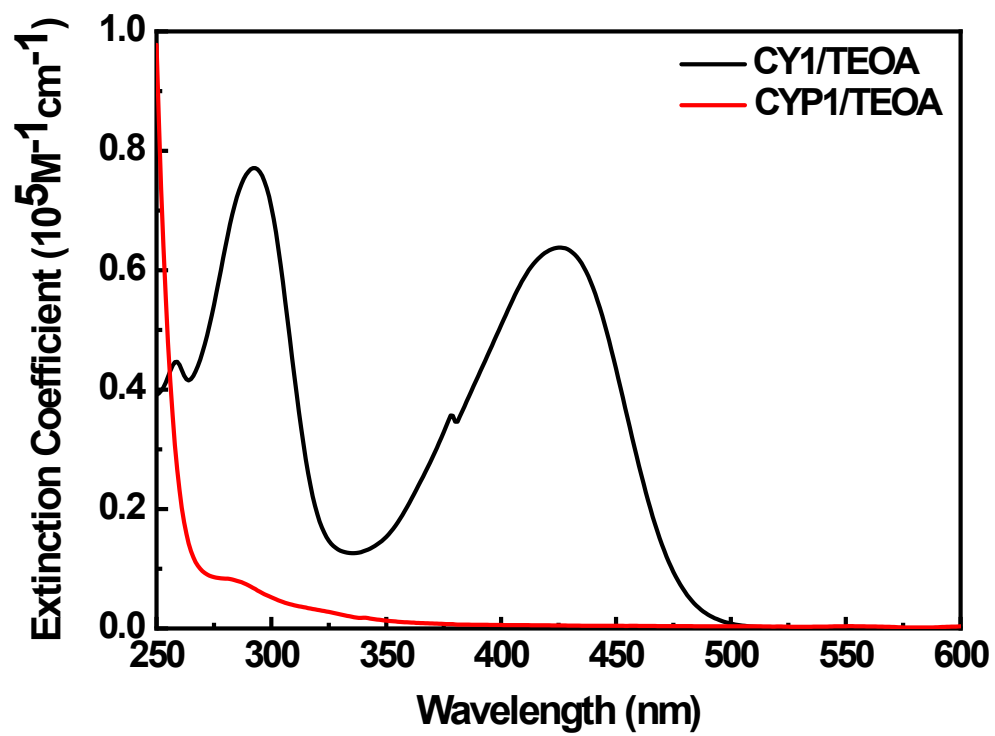


Fig. S18 Energy changes of the CYs between singlet and triplet states.



**Fig. S19** Experimented and simulated ESR spectra conducted by CY1/EDB (1:4 wt%) irradiated with a UV lamp for 2 min and trapped by PBN in *t*-butylbenzene solution.





**Fig. S20** UV-Vis spectra of CY1/TEOA and CYP1/TEOA (1:4 wt%) in DCM solution (concentration =  $1 \times 10^{-5}$  M).

**Table S1** Photo-DSC data derived from **CY1**/hydrogen donor packages<sup>a</sup>

<b>Sample</b>	<b>Final Conversion</b> (%)	<b><math>\Delta H_t</math></b> (kJ mol <sup>-1</sup> ) <sup>b</sup>	<b><math>H_{max}</math></b> (mW mg <sup>-1</sup> ) <sup>c</sup>	<b><math>R_{p_{max}}</math></b> (s <sup>-1</sup> ) <sup>d</sup>	<b><math>T_{max}</math></b> (s) <sup>e</sup>
<b>CY1/TEA (1:4)</b>	46	120	81	0.67	34
<b>CY1/TEOA (1:2)</b>	41	107	108	0.84	28
<b>CY1/TEOA (1:4)</b>	43	112	133	1.01	26
<b>CY1/TEOA (1:9)</b>	43	112	143	1.10	26

<sup>a</sup> Photo-DSC at various formulations measured with UV light of intensity 180 mW cm<sup>-2</sup> for 3 min and a total measurement time of 6 min.

<sup>b</sup>  $\Delta H_t$  is the total reaction heat enthalpy within 3 min.

<sup>c</sup>  $H_{max}$ : maximum heat flow.

<sup>d</sup>  $R_{p_{max}}$ : maximum rate of polymerization.

<sup>e</sup>  $T_{max}$ : time at maximum heat flow.

**Table S2** Photo-DSC data derived from **CY/TEOA** packages under UV light irradiation<sup>a</sup>

<b>PI package</b>	<b>Final Conversion</b> (%)	<b><math>\Delta H_t</math></b> (kJ mol <sup>-1</sup> ) <sup>b</sup>	<b><math>H_{max}</math></b> (mW mg <sup>-1</sup> ) <sup>c</sup>	<b><math>Rp_{max}</math></b> (s <sup>-1</sup> ) <sup>d</sup>	<b><math>T_{max}</math></b> (s) <sup>e</sup>
<b>CY1/TEOA</b>	43	112	133	1.01	26
<b>CY2/TEOA</b>	22	57	60	0.44	25
<b>CY3/TEOA</b>	10	23	27	0.20	28
<b>CY4/TEOA</b>	40	103	101	0.78	27
<b>CY5/TEOA</b>	18	45	42	0.30	28
<b>CYP1/TEOA</b>	38	98	258	1.98	23
<b>CYP1/TPA/TEOA</b> <sup>f</sup>	33	85	167	1.29	24

<sup>a</sup> Photo-DSC of the **CY/TEOA** blend at the ratio of **CY:TEOA:TMPTA = 1:4:95**), measured with UV light of intensity 180 mW cm<sup>-2</sup> for 3 min and a total measurement time of 6 min.

<sup>b</sup>  $\Delta H_t$  is the total reaction heat enthalpy within 3 min.

<sup>c</sup>  $H_{max}$ : maximum heat flow.

<sup>d</sup>  $Rp_{max}$ : maximum rate of polymerization.

<sup>e</sup>  $T_{max}$ : time at maximum heat flow.

<sup>f</sup> The weight ratio of **CYP1+TPA:TEOA:TMPTA** (w/w/w) was 1:4:95). The weight percentage of **CYP1** and **TPA** depended on their molecular weights.

**Table S3** Photo-DSC data derived from different photoinitiation packages under 405-nm LED light irradiation<sup>a</sup>

<b>PI package</b>	<b>Final Conversion</b>	<b><math>\Delta H_t</math></b>	<b><math>H_{max}</math></b>	<b><math>R_{p_{max}}</math></b>	<b><math>T_{max}</math></b>
	<b>(%)</b>	<b>(kJ mol<sup>-1</sup>)<sup>b</sup></b>	<b>(mW mg<sup>-1</sup>)<sup>c</sup></b>	<b>(s<sup>-1</sup>)<sup>d</sup></b>	<b>(s)<sup>e</sup></b>
<b>CY1/TEOA</b>	37	96	118	0.90	20
<b>CY2/TEOA</b>	22	58	46	0.35	25
<b>CY3/TEOA</b>	13	33	17	0.13	38
<b>CY4/TEOA</b>	43	112	188	1.38	22
<b>CY5/TEOA</b>	17	45	30	0.21	26
<b>CYP1/TEOA</b>	19	50	68	0.52	28
<b>CYP1/TPA/TEOA</b>	15	40	55	0.43	25

<sup>a</sup> Photo-DSC of the **CY/TEOA** blend at the ratio of **CY:TEOA:TMPTA** of 1:4:95, measured with 405 nm LED light of intensity 180 mW cm<sup>-2</sup> for 3 min and a total measurement time of 6 min.

<sup>b</sup>  $\Delta H_t$ : total reaction heat enthalpy within 3 min.

<sup>c</sup>  $H_{max}$ : maximum heat flow.

<sup>d</sup>  $R_{p_{max}}$ : maximum rate of polymerization.

<sup>e</sup>  $T_{max}$ : time at maximum heat flow.