Supplementary information

Ethynylene-linked multifunctional benzoxazines: effect of ethynylene group and packing on thermal behavior

Masanobu Muraoka,[†] Masahide Goto,[†] Masaki Minami,[‡] Dayang Zhou,[§] Takeyuki Suzuki,[§] Tatsuo Yajima,[†] Jun'ichi Hayashi,[⊥] Hiromitsu Sogawa,[†] Fumio Sanda^{*,†}

⁺ Department of Chemistry and Materials Engineering, Faculty of Chemistry, Materials and Bioengineering, Kansai University, 3-3-35 Yamate-cho, Suita, Osaka 564-8680, Japan

⁺ Chemicals R&D Group, HPM Research and Development Department, High Performance Materials Company, ENEOS Corporation, 8 Chidori-cho, Naka-ku, Yokohama, Kanagawa 231-0815, Japan

[§] Comprehensive Analysis Center, SANKEN, Osaka University, Ibaraki, Osaka 567-0047, Japan

¹ Department of Chemical, Energy and Environmental Engineering, Faculty of Environmental and Urban Engineering, Kansai University, 3-3-35 Yamate-cho, Suita, Osaka 564-8680, Japan

* Corresponding Author

Experimental section

Measurements. Solution state ¹H and ¹³C NMR spectra were recorded on JEOL JNM-ECZ400, JNM-ECS400 and BRUKER AVANCE II 700 spectrometers. Solid state ¹³C NMR spectra were recorded on a BRUKER AVANCE III 600WB. Melting points (mp) were measured on a Yanaco micro melting point apparatus. IR absorption spectra were measured on a JASCO FT/IR-4100 spectrophotometer. ESI-TOF mass spectra were measured on an Applied Biosystems APCI-3000 and a Shimadzu LCMS-IT-TOF. Single crystal X-ray analysis was performed using a Rigaku RAXIS imaging plate area detector with graphite-monochromated Mo-K α radiation (λ = 0.71075 Å). A crystal was mounted on a nylon loop at 123 K. The structure was solved and refined using the Olex2^{S1} and SHELX^{S2,S3} programs. XRD spectra were measured on a BRUKER D2 PHASER with Cu-Kα radiation. The ring-opening crosslinking reaction of the samples was carried out in a Nitto small electric furnace mini-BSI. DSC was measured on a Rigaku Thermoplus EVO DSC 8230 under air. TGA was measured with a Shimadzu thermogravimetric analyzer TGA-50 with a heating rate of 10 °C min⁻¹ from 50 °C to 800 °C under nitrogen at a flow rate of 50 mL min⁻¹. Densities were measured with a Shimadzu gas pycnometer AccuPyc II 1340 at 26 °C, in which averages of 10 measurements were employed. BET specific surface area and micropore volume were measured by using N2 adsorption isotherm at 77 K on a Microtrac BEL, BELSORP-mini II. CO₂ adsorption isotherms were measured at 298 K on a Microtrac BEL, BELSORP-28. Field emission scanning electron microscopy (FE-SEM) observation was performed on a JEOL JSM 6700f.

Reagents. Pd-Bz was obtained from Seika Corporation. The other reagents were purchased and used without purification unless otherwise stated. p-Ethynylaniline (FUJIFILM Wako Pure Chemical Corporation), salicylaldehyde (Tokyo Chemical Industry Co., TCI), ethanol (EtOH, FUJIFILM Wako), sodium borohydride (NaBH₄, TCI), paraformaldehyde (FUJIFILM Wako), 1,4-dioxane (FUJIFILM Wako), iodobenzene (FUJIFILM Wako), triphenylphosphine (FUJIFILM Wako), copper(I) iodide (Cul(I), NACALAI TESQUE, INC.), bis(triphenylphosphine)palladium(II)dichloride, tetrahydrofuran (THF) with stabilizer (FUJIFILM Wako), 2-dicyclohexylphosphino-2',4',6'-triisopropyl-1,1'-biphenyl (XPhos, TCI), triethylamine (Et₃N, NACALAI), 1,4-diiodobenzene (TCI), 1,3,5-tribromobenzene (TCI), potassium iodide (KI, NACALAI), 1,3-dimethyl-2-imidazolidinone (DMI, FUJIFILM Wako), anhydrous magnesium sulfate (MgSO₄, FUJIFILM Wako), hexane (NACALAI), dichloromethane (CH₂Cl₂, NACALAI), sodium hydroxide (NaOH, NACALAI) were used without purification. Solvents used in the reaction under Ar and N₂ were dehydrated over molecular sieves 4A1/16 and degassed by Ar and N₂ gas bubbling, respectively.

Synthesis of 3-(4-Ethynyl)phenyl-3,4-dihydro-1,3-benzoxazine (6)

Compound 6 was synthesized according to Scheme S1. The synthetic procedures follow the scheme.



Scheme S1 Synthesis of 6.

(E)-2-(((4-ethynylphenyl)imino)methyl)phenol (4)

p-Ethynylaniline (10.6 g, 90.3 mmol) was fed in a glass flask. It was degassed and filled with Ar. EtOH (180 mL) was added to the flask, and the resulting solution was stirred at room temperature for 5 min. It was heated to 30 °C; then salicylaldehyde (9.60 mL, 90.3 mmol) was added to the solution. The resulting mixture was stirred at 30 °C for 4 h, and then stirred at 60 °C for 0.5 h. The mixture was cooled with ice water to precipitate a solid mass. It was separated by filtration using a Kiriyama funnel, and dried under reduced pressure overnight to obtain yellow solid **4** (14.4 g, 64.9 mmol). Yield 72%. ¹H NMR (400 MHz, CDCl₃): δ 3.12 (s, 1H, CH=C), 6.92–7.03 (m, 1H, Ar), 7.01–7.03 (m, 1H, Ar), 7.15–7.28 (m, 3H, Ar), 7.33–7.44 (m, 2H, Ar), 7.51–7.59 (m, 2H, Ar), 8.60 (s, 1H, N=CH) ppm.

2-(((4-Ethynylphenyl)amino)methyl)phenol (5)

Compound **4** (14.4 g, 64.9 mmol) was fed in a glass flask, EtOH (300 mL) was added to the flask, and the resulting solution was cooled to 0 °C and stirred for 5 min. NaBH₄ (1.84 g, 64.9 mmol) was added to the solution, and the resulting mixture was stirred at room temperature for 2 h. Pure water (10 mL) was added to the mixture, and the resulting mixture was stirred for a while to dissolve residual NaBH₄. Then the reaction mixture was sequentially washed with CH₂Cl₂ (300 mL), 1 M NaOH aq (100 mL × 3). Solvents were evaporated under reduced pressure overnight to obtain brownish white solid **5** (13.3 g, 59.4 mmol). Yield 92%. ¹H NMR (400 MHz, CDCl₃): δ 2.98 (s, 1H, CH=C), 4.10 (s, 1H, –NH–), 4.40 (s, 2H, N–CH₂–), 6.66–6.78 (m, 2H, Ar), 6.87–6.97 (m, 2H, Ar), 7.16–7.27 (m, 2H, Ar), 7.34–7.38 (m, 2H, Ar), 7.46 (s, 1H, –OH) ppm.

3-(4-Ethynylphenyl)-3,4-dihydro-2H-[1,3]benzoxazine (6)⁵⁴

Compound **5** (13.3 g, 59.4 mmol) and paraformaldehyde (2.05 g, 67.4 mmol) were fed in a glass flask. It was degassed and filled with N₂. 1,4-Dioxane (200 mL) was added to the flask, and the resulting mixture was stirred at 100 °C for 4 h. The mixture was cooled to room temperature, and CH₂Cl₂ (90 mL) was added to the mixture. The resulting solution was washed with 1 M NaOH aq (30 mL × 3) to obtain a clear yellow solution. The organic phase was separated, and the solvents were evaporated under reduced pressure to obtain yellow solid **6** (13.4 g, 57.0 mmol). Yield 96%. ¹H NMR (400 MHz, CDCl₃): δ 2.98 (s, 1H, CH=C), 4.63 (s, 2H, Ar–CH₂–N), 5.35 (s, 2H, O–CH₂–N), 6.80–6.93 (m, 2H, Ar), 6.98–7.17 (m, 4H, Ar), 7.37–7.45 (m, 2H, Ar) ppm.

1,3,5-Triiodobenzene^{S5,S6}

The title compound was synthesized according to Scheme S2. The synthetic procedure follows the scheme.



Scheme S2 Synthesis of 1,3,5-triiodobenzene.

1,3,5-Tribromobenzene (4.00 g), CuI (5.78 g), KI (6.71 g) were fed in a glass flask. It was degassed and filled with N₂. DMI (70 mL) was added to the flask, and the resulting mixture was stirred at 200°C for 2 h. The reaction mixture was cooled with ice water to precipitate a solid mass. It was separated by filtration and washed with diethyl ether/brine to obtain 1,3,5-tribromobenzene as white solid (2.45 g). Yield 42%. ¹H NMR (400 MHz, CDCl₃): δ 7.19 (s, 3H) ppm.

3-(4-(Phenylethynyl)phenyl)-3,4-dihydro-2*H*-[1,3]benzoxazine (1)

The title compound was synthesized according to Scheme 1. The synthetic procedure is as follows. Compound **6** (4.24 g, 17.9 mmol), PPh₃ (0.967 g, 3.59 mmol), Cul (6.99×10⁻² g, 0.359 mmol) and (PPh₃)₂PdCl₂ (0.132 g, 0.179 mmol) were fed in a glass flask. It was degassed and filled with Ar. THF (30 mL), Et₃N (25 mL) and iodobenzene (2.0 mL, 17.9 mmol) were added to the flask, and the resulting mixture was stirred at 80 °C for 24 h under light shielding. Then the reaction mixture was sequentially washed with CH₂Cl₂ (90 mL), 1 M NaOH aq (30 mL × 3), and pure water (30 mL). The organic phase was separated, and solvents were evaporated under reduced pressure overnight. a yellow solid was obtained, which was purified by recrystallization with CH₂Cl₂/hexane to give a yellowish-white solid **1** (4.10 g, 13.2 mmol). Yield 73%, mp = 134–137 °C. IR (KBr disk): 3279, 3042, 2858, 2818, 2212, 1884, 1609, 1523, 1259, 1008, 816, 751, 690, 544 cm⁻¹. ¹H NMR (400 MHz, DMSO- *d*₆): δ 4.71 (s, 2H, Ar–CH₂–N), 5.49 (s, 2H, O–CH₂–N), 6.70–6.73 (m, 1H, Ar), 6.83–6.87 (m, 1H, Ar), 7.04–7.15 (m, 4H, Ar), 7.31–7.40 (m, 5H, Ar), 7.43–7.49 (m, 2H, Ar) ppm. ¹³C NMR (100 MHz, DMSO*d*₆): δ 48.9, 78.3, 88.5, 90.3, 113.8, 116.9, 117.2, 121.2, 121.8, 123.3, 127.8, 128.3, 128.8, 129.2, 131.6, 133.0, 148.4, 154.4 ppm. ESI-MS (*m/z*): calcd. 312.1383 ([C₂₂H₁₇NO + H]⁺), found 312.1362.

1,4-Bis((4-(2H-3(4H)-[1,3]benzoxazinyl)phenyl)ethynyl)benzene (2)

Compound **2** was synthesized from **6** and 1,4-diiodobenzene in a manner similar to **1**. XPhos was used instead of PPh₃. Yield 70%, yellowish-white solid, mp = 254–258 °C. IR (KBr disk): 3435, 3043, 2918, 2882, 2850, 2213, 1893, 1605, 1522, 1256, 1048, 817, 748, 532 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 4.65 (s, 4H, Ar–CH₂–N), 5.36 (s, 4H, O–CH₂–N), 6.80–6.91 (m, 4H, Ar), 6.99–7.14 (m, 8H, Ar), 7.40–7.43 (m, 8H, Ar) ppm. ¹³C NMR (175 MHz, CDCl₃): δ 50.1, 78.6, 88.2, 91.3, 115.2, 117.1, 117.3, 120.7, 121.1, 123.0, 126.8, 128.0, 131.3, 132.8, 148.2, 154.2 ppm. ESI-MS (*m/z*): calcd. 545.2224 ([C₃₈H₂₈N₂O₂ + H]⁺), found 545.2212.

1,3,5-Tris((4-(2H-3(4H)-[1,3]benzoxazinyl)phenyl)ethynyl)benzene (3)

Compound **3** was synthesized from **6** and 1,3,5-triiodobenzene in a manner similar to **1**. It was purified by recrystallization with CH_2Cl_2 /hexane. Yield 64%, yellow solid, mp = 136–140 °C. IR (KBr disk): 3398, 3040, 2893, 2851, 2567, 2360, 2203, 1894, 1782, 1607, 1577, 1512, 1226, 947, 754, 540 cm⁻¹. ¹H NMR (400 MHz, DMSO-*d*₆): δ 4.66 (s, 6H, Ar–CH₂–N), 5.44 (s, 6H, O–CH₂–N), 6.65–6.77 (m, 3H, Ar), 6.83–6.91 (m, 3H, Ar), 7.04–7.21 (m, 12H, Ar), 7.33–7.82 (m, 9H, Ar) ppm. ¹³C NMR (100

MHz, CDCl₃): δ 50.1, 78.6, 87.1, 90.6, 115.1, 117.2, 117.3, 120.7, 121.1, 124.3, 126.8, 128.1, 133.0, 133.5, 148.4, 154.4 ppm. ESI-MS (*m/z*): calcd. 778.3064 ([C₅₄H₃₉N₃O₃ + H]⁺), found 778.3049.

Polymerization: typical procedure.

A powdery sample of **1** was fed into an aluminum sample pan. It was placed on a glass petri dish, and heated in a Nitto small electric furnace mini-BSI at 150–300 °C for 2 h under N₂ to obtain Poly(**1**).

Curing Kinetics.

The activation energies (E_a) for curing of **1–3** were determined by Ozawa–Flynn–Wall method,^{S7–S9} based on the DSC measurements at heating rates of 5, 10, 15, and 20 °C/min (Fig. S38–S40). The values of E_a were calculated using the following equation,

$$\ln(\theta) = -1.052E_{a}/RT_{p} + C$$

where β is the heating rate, T_p is the temperature at the exothermic peak maximum, R is the gas constant, and C is a constant. E_a can be obtained from the slope of the plot of ln(β) versus $1/T_p$. The E_a values for curing of **1**, **2** and **3** were 83.52, 133.75 and 67.14 kJ/mol, respectively. It is considered that molecules of **2** are densly packed compared with **1** and **3**, leading to low mobility, resulting in the high E_a . **Computations.** DFT calculations were performed with Gaussian 16,^{S10} ES64L-G16 Rev C.01, using ωB97X-D functional in conjunction with 6-31G* basis set, running on the supercomputer system, Academic Center for Computing and Media Studies, Kyoto University. Noncovalent interaction plot was performed using NCIPLOT.^{S11,S12} MD simulations were performed with LAMMPS 2020,^{S13} running on the supercomputer system, Fugaku provided by the RIKEN Center for Computational Science.

CO₂ adsorption. Unlike the corresponding benzoxazine monomers, polybenzoxazines poly(**1**)–poly(**3**) showed no sharp XRD peaks (Fig. S47). It is considered that poly(**1**)–poly(**3**) do not exhibit the regulated angstrom-order molecular alignment observed in the corresponding monomers, while the polymers formed micron/submicron pores that efficiently adsorb CO₂. Physisorption is an effective capture method because of the low energy consumption, high efficiency, easy regeneration and high cost-effectiveness.^{S14,S15}

Morphology. The samples of benzoxazines **1**, **2**, **3** and Pd-Bz were observed by FE-SEM after curing to obtain information on the surface morphology (Fig. S52).



Fig. S1 ¹H NMR spectrum of **1** measured at 400 MHz in CDCl₃ at 25 °C.



Fig. S2 ¹H NMR spectrum of **2** measured at 400 MHz in CDCl₃ at 25 °C.



Fig. S3 ¹H NMR spectrum of **6** measured at 400 MHz in CDCl₃ at 25 °C.



Fig. S4 ¹H NMR spectrum of **1** measured at 400 MHz in DMSO- d_6 at 25 °C.



Fig. S5 ¹³C NMR spectrum of **1** measured at 100 MHz in DMSO- d_6 at 25 °C.



Fig. S6 ¹H NMR spectrum of **2** measured at 400 MHz in CHCl₃ at 25 °C.



Fig. S7 13 C NMR spectrum of 2 measured at 175 MHz in CHCl₃ at 25 °C.



Fig. S8 ¹H NMR spectrum of **3** measured at 400 MHz in DMSO- d_6 at 25 °C.



Fig. S9 ¹³C NMR spectrum of **3** measured at 100 MHz in CDCl₃ at 25 °C.



Fig. S10 ¹H NMR spectrum of 1,3,5-triiodobenzene measured at 100 MHz in CDCl₃ at 25 °C.



Fig. S11 IR absorption spectrum (KBr pellet) of 1.



Fig. S12 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **1** at 150 °C.



Fig. S13 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **1** at 200 °C.



Fig. S14 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **1** at 250 °C.



Fig. S15 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **1** at 300 °C.



Fig. S16 IR absorption spectrum (KBr pellet) of 2.



Fig. S17 IR absorption spectrum (KBr pellet) of a resin obtained by curing of 2 at 150 °C.



Fig. S18 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **2** at 200 °C.



Fig. S19 IR absorption spectrum (KBr pellet) of a resin obtained by curing of 2 at 250 °C.



Fig. S20 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **2** at 300 °C.



Fig. S21 IR absorption spectrum (KBr pellet) of 3.



Fig. S22 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **3** at 150 °C.



Fig. S23 IR absorption spectrum (KBr pellet) of a resin obtained by curing of 3 at 200 °C.



Fig. S24 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **3** at 250 °C.



Fig. S25 IR absorption spectrum (KBr pellet) of a resin obtained by curing of **3** at 300 °C.



Fig. S26 ESI-TOF-MS of 1; (top) found, (bottom) calcd.



Fig. S27 ESI-TOF-MS of 2; (top) found, (bottom) calcd.



Fig. S28 ESI-TOF-MS of 3; (top) found, (bottom) calcd.



Fig. S29 IR absorption spectra of **1** and resins obtained by curing of **1** at 150–300 °C for 2 h. The peak at 1650 cm⁻¹ is assignable to C=N bond in the Schiff base structure formed during the ring-opening process.^{S16}



Fig. S30 DSC thermograms of **1** and resins obtained by curing of **1** at 150–300 °C for 2 h.



Fig. S31 TGA traces and DTG (dashed line) of **1** and resins obtained by curing of **1** at 150–300 °C for 2 h.



Fig. S32 IR absorption spectra of **2** and resins obtained by curing of **2** at 150–300 °C for 2 h. The peak at 1650 cm⁻¹ is assignable to C=N bond in the Schiff base structure formed during the ring-opening process.^{S16}



Fig. S33 DSC thermograms of **2** and resins obtained by curing of **2** at 150–300 °C for 2 h.



Fig. S34 TGA traces and DTG (dashed line) of **2** and resins obtained by curing of **2** at 150–300 °C for 2 h.



Fig. S35 IR absorption spectra of **3** and resins obtained by curing of **3** at 150–300 °C for 2 h. The peak at 1650 cm⁻¹ is assignable to C=N bond in the Schiff base structure formed during the ring-opening process.^{S16}



Fig. S36 DSC thermograms of **3** and resins obtained by curing of **3** at 150–300 °C for 2 h.



Fig. S37 TGA traces and DTG (dashed line) of **3** and resins obtained by curing of **3** at 150–300 °C for 2 h.



Fig. S38 (a) DSC thermograms of **1** at various heating rates. (b) Ozawa–Flynn–Wall plot for the data of (a). Correlation coefficient r = 0.9408.



Fig. S39 (a) DSC thermograms of **2** at various heating rates. (b) Ozawa–Flynn–Wall plot for the data of (a). Correlation coefficient r = 0.9964.



Fig. S40 (a) DSC thermograms of **3** at various heating rates. (b) Ozawa–Flynn–Wall plot for the data of (a). Correlation coefficient r = 0.9261.



Fig. S41 ¹³C CP/MAS NMR spectra of 2 (top) and resin obtained by curing of 2 at 300 °C (bottom).



Fig. S42 Deduced structures of 2 (left) and resin formed by curing of 2 (right).



Fig. S43 ¹³C CP/MAS NMR spectra of **3** (top) and resin obtained by curing of **3** at 300 °C (bottom).



Fig. S44 Deduced structures of 3 (left) and resin formed by curing of 3 (right)

Monomer



Tetramer Twisted (+10°)



Tetramer Twisted (+5°)



Tetramer Alternating



Tetramer Twisted (–5°)



 $\Delta E = 0$ kJ/mol (standard)



 $\Delta E/4 = -130.9 \text{ kJ/mol}$



 $\Delta E/4 = -131.0 \text{ kJ/mol}$



 $\Delta E/4 = -109.7 \text{ kJ/mol}$



 $\Delta E/4 = -132.6 \text{ kJ/mol}$



 $\Delta E/4 = -133.0 \text{ kJ/mol}$



Fig. S45 Initial and DFT (ω B97X-D/6-31G*)-optimized conformers of monomeric and tetrameric molecular models of **2**.



Fig. S46 Initial and DFT (ω B97X-D/6-31G*)-optimized conformers of monomeric and tetrameric molecular models of **3**.



Fig. S47 Powder XRD patterns of poly(1)-poly(3) with Cu-K α radiation.



Fig. S48 Snapshots of trajectories of MD simulation of **1–3** at 5 ns, 300 K using models consisting of 1000 molecules by LAMMPS 2020.^{S13} The densities of benzoxazines **1**, **2** and **3** were calculated to be 1.285, 1.313 and 1.264, respectively (average values of a period of 4–5 ns simulated for 1,000 ns). Also see trajectory movies in the Supporting Information.

Compound	Density	/ (g/cm³)
Compound	Observed ^a	MD simulated ^b
1	1.285	1.126
2	1.313	1.158
3	1.264	1.154

 Table S1
 Observed and MD simulated densities of 1–3

^{*a*} Average of 10 measurements by gas pycnometry at 26 °C. ^{*b*} Average value of a period of 4–5 ns simulated for 1,000 molecules at 300 K with LAMMPS 2020.^{S13}



Fig. S49 Chemical structures of Pd-Bz and poly(Pd-Bz).



Fig. S50 CO₂ adsorption isotherms for poly(**1**)–poly(**3**) and poly(Pd-Bz) measured at 298 K.



Fig. S51 N₂ adsorption isotherms for poly(1)–poly(3) and poly(Pd-Bz) measured at 77 K.

Table S2	Specific surface	area,	micropore	and	mesopore	volumes	of	poly(1)–poly(3)	and
poly(Pd-Bz)	determined usir	g N ₂							

Compound	Specific surface area ^a [m²/g]	Micropore volume [mL/g]	Mesopore volume [mL/g]
Poly(1)	0.3	0.000	_ <i>b</i>
Poly(2)	0.0	0.000	_ <i>b</i>
Poly(3)	0.0	0.000	_ b
Poly(Pd-Bz)	0.0	0.000	0.007

^{*a*} BET analysis. ^{*b*} Not determined.



Fig. S52 FE-SEM images. (a), (e): poly(**1**), (b), (f): poly(**2**), (c), (g): poly(**3**), (d), (h): poly(Pd-Bz).

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