Supporting Information

Organocatalytic Atroposelective *N*-Alkylation: Divergent Synthesis of Axially Chiral Sulfonamides and Biaryl Amino Phenols

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I. Supplementary Methods

General information. All experiments were conducted under air atmosphere unless otherwise noted. ¹H and ¹³C NMR spectra were recorded on a Bruker AscendTM 400 (400 MHz) spectrometer. Chemical shifts were reported in parts per million (ppm), and the residual solvent peak was used as an internal reference: ¹H (chloroform δ 7.26; DMSO δ 2.50), ¹³C (chloroform δ 77.0; DMSO δ 39.5). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), coupling constants (Hz) and integration. For thin layer chromatography (TLC), Merck pre-coated TLC plates (Merck 60 F254) were used, and compounds were visualized with a UV light at 254nm. High resolution mass spectra (HRMS) were obtained on an Agilent 1290II-6545 spectrometer. Optical rotations were recorded on a Rudolph Research Analytical Autopol I automatic polarimeter. Enantiomeric excesses (ee) were determined by HPLC analysis on Agilent HPLC units, and Waters e2695; column of Chiralcel OD-H, Chiralpak AD-H, AS-H, ID or IE was used. Column chromatography was performed with silica gel (200-300 mesh ASTM). Unless otherwise noted, commercially available reagents purchased from Adamas-beta, TCI, or Energy Chemical and were used as received.

The 2,3-dienoate adducts 2^[1], MBH acetates 4^[2] and *rac*-**8**^[3] were synthesized in one step from commercially available materials by literature methods.

II. Optimization of axially chiral sulfonamides synthesis

Supplementary Table 1. Attempt of Synthesis of Axially Chiral Sulfonamides from Allenoates *via* Phosphine Catalysis^{a,b}



^aUnless noted otherwise, the reactions were performed with **1** (0.05 mmol), **2** (0.07 mmol, 1.4 equiv.), cat. (10 mol%), and Cs_2CO_3 (0.05 mmol, 1.0 equiv.) in toluene (0.5 mL) at 24 °C for 2 h. ^bThe *ee* value was determined by chiral HPLC.

To a Schlenk tube containing 1 (0.1 mmol), phosphine (10 mol%) and Cs_2CO_3 (0.1 mmol, 1.0 equiv.) were added toluene (0.5 mL) and dienoate **2a** (0.14 mmol, 1.4 equiv.). The reaction mixture was stirred at 24 °C for 2 h. Then, the *ee* value of **3a** was determined by chiral HPLC.

Supplementary Table 2. Base and Solvent Screening^a

O O Tol ^{-S} N ^{-H} Me Me 1a	+ =•=√ ^{OAc} CO ₂ Bn 2a (1.4 equiv.)	P1 (10 mol%) Base (1.0 equiv.) r.t., 2 h, Solvent $\overrightarrow{OTBDPS P1}$ \overrightarrow{PPh}_2 \overrightarrow{NHBoc}	O,O Tol ^{∕S} N Me Me Me 3a		
Entry	Base	Solvent	ee (%) ^b		
1	Cs ₂ CO ₃	PhMe	18		
2	K ₂ CO ₃	PhMe	12		
3	Na ₂ CO ₃	PhMe	13		
4	Li ₂ CO ₃	PhMe	-		
5	KHCO3	PhMe	10		
6	NaHCO ₃	PhMe	-		
7	K ₃ PO ₄	PhMe	7		
8	Na ₂ HPO ₄	PhMe	-		
9	Cs ₂ CO ₃	THF	0		
10	Cs ₂ CO ₃	HCCl ₃	0		
11	Cs ₂ CO ₃	Et ₂ O	9		
12	Cs ₂ CO ₃	EtOAc	3		
13	Cs ₂ CO ₃	CH ₃ CN	3		
14	Cs_2CO_3	PhCl	0		
15	Cs ₂ CO ₃	Hexanes	3		
16	Cs ₂ CO ₃	Acetone	0		
17	Cs ₂ CO ₃	PhCF ₃	0		

^aUnless noted otherwise, the reactions were performed with 1 (0.05 mmol), 2 (0.07 mmol, 1.4 equiv.), P1 (10 mol%), and Cs₂CO₃ (0.05 mmol, 1.0 equiv.) in toluene (0.5 mL) at 24 °C for 2 h. bThe ee value was determined by chiral HPLC.

To a Schlenk tube containing 1 (0.1 mmol), P1 (10 mol%) and base (0.1 mmol, 1.0 equiv.) were added toluene (0.5 mL) and dienoate 2a (0.14 mmol, 1.4 equiv.). The reaction mixture was stirred at 24 °C for 2 h. Then, the ee value of 3a was determined by chiral HPLC.

Supplementary Table 3. Optimization of the Reaction Conditions to Access



Axially Chiral Sulfonamides from Allenoates via Amine Catalysis^a

^aUnless noted otherwise, the reactions were performed with **1a** (0.05 mmol, 1.0 equiv.), **2a** (0.07 mmol, 1.4 equiv.), catalyst (10 mol%), and base (1.0 equiv.) in solvent (0.5 mL) at 24 to -50 °C for 12 h. ^bYield was detected by ¹H-NMR. ^cThe *ee* value was determined by chiral HPLC. ^dMesitylene (4 mL) was added.

Mesitylene

92

-50

90

15^d

Α

 Cs_2CO_3

To a Schlenk tube containing 1 (0.05 mmol), amine (10 mol%) and base (1.0 equiv.)

were added solvent (4 mL) and dienoate 2a (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at 24 to -50 °C for 12 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product 3a.

Supplementary Table 4. Optimization of the Reaction Conditions via Amine Catalysis^{a,b}



^aUnless noted otherwise, the reactions were performed with **1a** (0.05 mmol, 1.0 equiv.), **2a** (0.07 mmol, 1.4 equiv.), catalyst (10 mol%), in toluene (0.5 mL) at 24 °C for 2 h. ^bThe *ee* value was determined by chiral HPLC.

To a Schlenk tube containing 1 (0.05 mmol), amine (10 mol%) were added PhMe (0.5 mL) and dienoate 2a (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at 24 °C for 2 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product **3a**. The *ee* value of **3a** was determined by chiral HPLC.

Supplementary Table 5. Solvent Screening of the Reaction Conditions via Amine

Catalysis^{a,b}

O O Tol ^S N ^H Me Me 1a	I + $-OAc$ $\beta -ICD (10 mol\%)$ CO_2Bn $24 °C, 2 h, Solvent$ 2a (1.4 equiv.)		10 mol%) h, Solvent	O_O Tol ^{∕S} N CO₂Bn Me Me Me 3a	
	Entry	Solvent	ee (%)	-	
	1	DCM	47	-	
	2	THF	5		
	3	EtOAc	21		
	4	CH ₃ CN	33		
	5	DMSO	2		
	6	DMF	0		
	7	HCCl₃	49		
	8	PhCl	57		
	9	PhF	55		
	10	o-DCB	53		
	11	PhCF ₃	41		
	12	PhH	57		
	13	Hexanes	32		
	14	o-Xylene	41		
	15	<i>m</i> -Xylene	69		
	16	<i>p</i> -Xylene	67		
	17	PhEt	70		
	18	PhOMe	57		
	19	Mesitylene	72		
	20 ^c	Mesitylene	79		

^aUnless noted otherwise, the reactions were performed with **1a** (0.05 mmol, 1.0 equiv.), **2a** (0.07 mmol, 1.4 equiv.), catalyst (10 mol%), in toluene (0.5 mL) at 24 °C for 2 h. ^bThe *ee* value was determined by chiral HPLC. ^cMesitylene (4 mL) was added. To a Schlenk tube containing **1** (0.05 mmol), amine (10 mol%) were added PhMe (0.5 mL) and dienoate **2a** (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at 24 °C for 2 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product **3a**. The *ee* value of **3a** was determined by chiral HPLC.

Supplementary Table 6. Optimization of the Reaction Conditions to Access

NHTs Me Me	+	—OAc β-IC CO ₂ Bn Cs ₂ C Tem	CD (x mol%) O ₃ (1.0 equiv.) p., mesitylene	0,0 R ⁴ ·S [™] N CO ₂ Bn Me Me Me 5e	
Entry	Temp. (°C)	β-ICD (x mol%)	ee (%) ^b	Yield (%) ^c	
1	-50	5	91	95	
2	-40	5	91	96	
3	-30	5	91	95	
4	-20	5	87	95	
5	-30	3	91	93	
6 ^e	-30	2	91	95	
7 ^e	-30	1	91	96	
8 ^{d, f}	-30	1	91	94	

Axially Chiral Sulfonamides from MBH Acetate^a

^aReaction: **1** (0.1 mmol), **4** (0.14 mmol, 1.4 equiv.), β -ICD (x mol%), Cs₂CO₃ (0.1 mmol, 1.0 equiv.) in mesitylene (2 mL) at -30 to -50 °C for 24 h. ^bIsolated yield. ^cThe *ee* value was determined by chiral HPLC. ^dMesitylene (1 mL) was added. ^e48 h. ^f72 h.

To a Schlenk tube containing 1 (0.1 mmol), β -ICD (x mol%) and Cs₂CO₃ (0.1 mmol, 1.0 equiv.) were added mesitylene and MBH acetate 4e (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at 24 to -50 °C for 12 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product 5e.

Supplementary Table 7. Optimization of the Selective N-H Activation^a



^aUnless noted otherwise, the reactions were performed with **6a** (0.05 mmol), **4a** (0.07 mmol, 1.4 equiv.), β -ICD (10 mol%), Cs₂CO₃ (0.05 mmol, 1.0 equiv.) in mesitylene (3 mL) at -30 to -50 °C for 24 h. ^bIsolated yield. ^cThe *ee* value was determined by chiral HPLC.

To a Schlenk tube containing **6a** (0.05 mmol), β -ICD (10 mol%) and Cs₂CO₃ (0.05 mmol, 1.0 equiv.) were added mesitylene (3 mL) and MBH acetate **4c** (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at -30 to -50 °C for 24 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product **7a**.

Supplementary Table 8. Optimization of Kinetic Resolution of NOBINs^a

MeO Me Br	Me NHTs - OH	$+ = \sqrt{\begin{array}{c} -\text{OAc} & \beta - \text{ICD} \\ \\ \text{CO}_2^n \text{Bu} & \text{Base (0)} \\ \\ \text{Solve} \end{array}}$	M (3 mol%)).7 equiv.) ent, 0 °C Br	Me Me	N-Ts +	Br MeO	Me NHTs OH
(±) -8 (0.1	1 mmol)	4 (0.7 equiv.)		9		(<i>R</i>)-8
Entry	Base	Solvent	Temp. (^o C)	C ^b (%)	8a, ee ^c	9a, <i>ee^c</i>	s ^d
1	Cs_2CO_3	THF	0	54	71	60	8
2	Cs_2CO_3	CH ₃ CN	0	58	33	24	2
3	Cs_2CO_3	DCM	0	30	30	70	8
4	Cs_2CO_3	toluene	0	32	36	76	11
5	Cs_2CO_3	mesitylene	0	43	37	50	4
6	Cs_2CO_3	Chlorobenzene	0	51	73	70	12
7	K ₂ CO ₃	Chlorobenzene	0	43	60	81	16
8	Na ₂ CO ₃	Chlorobenzene	0	22	23	81	12
9	K ₂ CO ₃	Chlorobenzene	-10	40	52	80	14
10 ^e	K ₂ CO ₃	Chlorobenzene	0	55	77	94	27

^aUnless noted otherwise, the reactions were performed with **8a** (0.1 mmol), **4a** (0.07 mmol, 0.7 equiv.), β -ICD (3 mol%), Base (0.07 mmol, 0.7 equiv.) in Solvent (2 mL)

at 0 °C for 12 h. ^bConversion (C) = $ee_{8a}/(ee_{8a}+ee_{9a})$. ^cThe *ee* value was determined by chiral HPLC. ^dS = In[(1-Conv.)(1-ee_{8a})]/In(1-Conv.)(1+ee_{8a})]. ^ethe reactions were performed with **8a** (0.1 mmol), **4a** (0.035 mmol, 0.35 equiv.), β -ICD (3 mol%), Base (0.035 mmol, 0.35 equiv.) in Solvent (8 mL) at 0 °C for 36 h

To a Schlenk tube containing **6a** (0.05 mmol), β -ICD (10 mol%) and Cs₂CO₃ (0.05 mmol, 1.0 equiv.) were added mesitylene (3 mL) and MBH acetate **4c** (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at -30 to -50 °C for 24 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product **7a**.

III. General procedure and spectra data of axially chiral sulfonamides and biaryl amino phenols

Representative procedure for synthesis of 3:



To a Schlenk tube containing 1 (0.05 mmol), β -ICD (1.5 mg, 10 mol%) and Cs₂CO₃ (0.05 mmol, 1.0 equiv.) were added mesitylene (4 mL) and dienoate 2 (0.07 mmol, 1.4 equiv.). The reaction mixture was stirred at -50 °C for 24 hours to 7 days. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography with hexanes/ethyl acetate as the eluent to afford the product **3**.

Characterization of compounds 3:



3a (91% yield, Hexane-EtOAc = 10:1, Rf = 0.3). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.69 (d, J = 8.3 Hz, 2H), 7.44 – 7.37 (m, 1H), 7.28 – 7.14 (m, 7H), 6.86 (m, 1H), 5.03 (d, J = 12.5 Hz, 1H), 4.98 (d, J = 14.6 Hz, 1H), 4.95 (d, J = 12.5 Hz, 1H), 4.87 (d,

J = 14.6 Hz, 1H), 4.49 (s, 2H), 2.35 (s, 3H), 2.15 (s, 3H), 2.06 (s, 3H). ¹³C NMR (100 10/150

MHz, CDCl₃) δ 216.45, 165.7, 143.4, 141.9, 139.8, 138.9, 138.3, 137.4, 135.7, 132.1, 129.4, 128.4, 128.4, 128.1, 128.1, 101.4, 96.2, 79.3, 66.8, 48.7, 21.6, 20.4, 20.3. **HRMS (ESI)** m/z Calcd for [C₂₇H₂₆INO₄S, M + H]⁺: 588.0700; Found: 588.0703.

Optical Rotation: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 17.216$ min for minor isomer, $t_{R} = 20.148$ min for major isomer).





3b (92% yield, Hexane-EtOAc = 5:1, Rf = 0.4). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, *J* = 8.2 Hz, 2H), 7.51 (s, 1H), 7.28 (d, *J* = 8.2 Hz, 2H), 6.99 (s, 1H), 5.03 (d, *J* = 14.6 Hz, 1H), 4.93 (d, *J* = 14.6 Hz, 1H), 4.54 (s, 2H), 3.61 (s, 3H), 2.42 (s, 3H), 2.24

(s, 3H), 2.19 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ216.2, 166.2, 143.4, 141.9, 139.8, 138.9, 138.2, 137.3, 132.1, 129.3, 128.3, 101.4, 96.0, 79.2, 52.4, 48.7, 21.5, 20.3, 20.2. HRMS (ESI) m/z Calcd for [C₂₀H₂₂INO₄S, M + Na]⁺: 532.0050; Found: 532.0053.

Optical Rotation: $[\alpha]^{25}_{D}$ -35.0 (c = 1.0, CHCl₃). 85% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 12.906 min for minor isomer, t_{R} = 15.073 min for major isomer).





3c (85% yield, Hexane-EtOAc = 5:1, Rf = 0.4). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.77 (d, J = 8.2 Hz, 2H), 7.51 (s, 1H), 7.28 (d, J = 8.2 Hz, 2H), 6.99 (s, 1H), 5.04 (d, J = 14.5 Hz, 1H), 4.93 (d, J = 14.5 Hz, 1H), 4.64 – 4.46 (m, 2H), 4.13 – 3.96 (m, 2H),

2.41 (s, 3H), 2.23 (s, 3H), 2.20 (s, 3H), 1.14 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.2, 165.7, 143.4, 141.9, 139.8, 138.9, 138.2, 137.3, 132.0, 129.3, 128.3, 101.4, 96.23, 79.1, 61.3, 48.6, 21.5, 20.3, 20.2, 14.0. HRMS (ESI) m/z Calcd for $[C_{22}H_{24}INO_4S, M + Na]^+$: 548.0363; Found:548.0365.

Optical Rotation: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 15.301 min for minor isomer, t_{R} = 17.396 min for major isomer).





3d (95% yield, Hexane-EtOAc = 5:1, Rf = 0.4). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.77 (d, J = 8.2 Hz, 2H), 7.50 (s, 1H), 7.28 (d, J = 8.2 Hz, 2H), 6.99 (s, 1H), 5.03 (d, J = 14.4 Hz, 1H), 4.92 (d, J = 14.4 Hz, 1H), 4.90 - 4.84 (m, 1H), 4.64 - 4.46 (m, 2H), 2.42 (s, 3H), 2.23 (s, 3H), 2.20 (s, 3H), 1.14 (d, J = 6.3 Hz, 3H), 1.11 (d, J = 6.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.2, 165.3, 143.4, 142.1, 139.8, 138.9, 138.4, 137.2, 132.1, 129.4, 128.29, 101.3, 96.5, 78.9, 68.8, 48.5, 21.6, 21.5, 20.3, 20.3. HRMS (ESI) m/z Calcd for [C₂₂H₂₄INO₄S, M + Na]⁺: 548.0363; Found: 548.0368.

Optical Rotation: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AS-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 11.038$ min for minor isomer, $t_{R} = 13.114$ min for major isomer).





3e. (94% yield, Hexane-EtOAc = 5:1, Rf = 0.4). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.77 (d, J = 8.3 Hz, 2H), 7.51 (d, J = 0.9 Hz, 1H), 7.28 (d, J = 8.3 Hz, 2H), 6.99 (d, J = 0.9 Hz, 1H), 5.05 (d, J = 14.5 Hz, 1H), 4.94 (d, J = 14.5 Hz, 1H), 4.63 – 4.47 (m, 2H),

3.99 (qt, J = 10.8, 6.7 Hz, 2H), 2.42 (s, 3H), 2.23 (s, 3H), 2.20 (s, 3H), 1.52-1.45 (m, 6.8 Hz, 2H), 1.31-1.26 (m, J = 15.2, 7.5 Hz, 2H), 0.88 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.2, 165.8, 143.4, 141.9, 139.8, 138.9, 138.3, 137.4, 132.0, 129.4, 128.3, 101.4, 96.3, 79.1, 65.2, 48.6, 30.4, 21.6, 20.3, 20.2, 19.0, 13.7. HRMS (ESI) m/z Calcd for [C₂₄H₂₈INO₄S, M + Na]⁺: 576.0676; Found: 576.0680.

Optical Rotation: $[\alpha]^{25}_{D}$ -32.0 (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AS-H column, *n*-Hexane/*i*-PrOH = 98:2, flow rate = 1.0 mL/min, $\frac{13/150}{13/150}$



22.5

wavelength = 254 nm, $t_{\rm R}$ = 12.661 min for minor isomer, $t_{\rm R}$ = 15.323 min for major isomer).



17.5

PeakTable

15.0

2.5

1 Det.A Ch1/254nr

3f (94% yield, Hexane-EtOAc = 5:1, Rf = 0.4). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, *J* = 8.3 Hz, 2H), 7.50 (dd, *J* = 1.4, 0.5 Hz, 1H), 7.27 (d, *J* = 8.3 Hz, 2H), 6.99 (dd, *J* = 1.4, 0.5 Hz, 1H), 5.01 (d, *J* = 14.3 Hz, 1H), 4.87 (d, *J* = 14.3 Hz, 1H), 4.56 (d,

12.5

254nm Ret. Tir

10.0

1 Det.A Ch1/254nm

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17.5

PeakTable

Heigh

20.0

22.5

25.0

J = 14.1 Hz, 1H), 4.48 (d, J = 14.1 Hz, 1H), 2.41 (s, 3H), 2.23 (s, 3H), 2.23 (s, 3H), 1.32 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 216.3, 164.8, 143.3, 142.2, 139.8, 138.9, 138.5, 137.4, 132.1, 129.3, 128.3, 101.1, 97.5, 81.2, 78.6, 48.5, 27.7, 21.5, 20.3, 20.3. HRMS (ESI) m/z Calcd for [C₂₄H₂₈INO₄S, M + Na]⁺: 576.0676; Found: 576.0681.

Optical Rotation: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 11.029$ min for minor isomer, $t_{R} = 9.863$ min for major isomer).





3g (85% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, Rf = 0.5). syrup. ¹**H NMR** (400 MHz, CDCl₃) δ 7.66 (d, *J* = 8.0 Hz, 2H), 7.35 (s, 1H), 7.19-7.14 (m, 12H), 6.74 (s, 1H), 6.70 (s, 1H), 5.03 (d, *J* = 14.6 Hz, 1H), 4.92 (d, *J* = 14.6 Hz, 1H), 4.49

(s, 2H), 2.32 (s, 3H), 2.09 (s, 3H), 1.94 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.7, 164.8, 143.3, 141.9, 140.0, 139.7, 138.9, 138.3, 137.2, 132.0, 129.3, 128.3 127.8, 127.7, 127.2, 126.9, 101.3, 96.1, 79.1, 77.3, 48.5, 21.5, 20.3, 20.0. HRMS (ESI) m/z Calcd for [C₃₃H₃₀INO₄S, M + Na]⁺: 686.0832; Found: 686.0834.

Optical Rotation: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 14.295 min for minor isomer, t_{R} = 16.424 min for major isomer).





3h (92% yield, Hexane-EtOAc = 5:1, Rf = 0.3). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.63 (d, J = 7.8 Hz, 2H), 7.29 – 7.10 (m, 12H), 7.03 (s, 1H), 6.74 (s, 1H), 6.70 (s, 1H), 5.06 (d, J = 14.6 Hz, 1H), 4.92 (d, J = 14.6 Hz, 1H), 4.53 (d, J = 14.1

Hz, 1H), 4.40 (d, J = 14.1 Hz, 1H), 2.33 (s, 3H), 2.13 (s, 3H), 1.99 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.6, 164.9, 143.3, 142.8, 140.15, 139.7, 138.0, 133.8, 132.0, 131.2, 129.3, 128.4, 128.2, 127.9, 127.8, 127.2, 127.0, 125.0, 96.3, 79.3, 77.4, 48.3, 21.6, 20.7, 19.8. **HRMS (ESI)** m/z Calcd for [C₃₃H₃₀BrNO₄S, M + Na]⁺: 638.0971, 640.0956; Found: 638.0976, 640.0958.

Optical Rotation: $[\alpha]^{25}_{D}$ -8.5 (c = 0.5, CHCl₃). 93% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 12.527$ min for minor isomer, $t_{R} = 14.345$ min for major isomer).





3i (84% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, Rf = 0.5). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.74 (d, *J* = 7.8 Hz, 2H), 7.41 – 7.23 (m, 13H), 7.05-6.98 (m, 2H), 6.80 (s, 1H), 5.14 (d, *J* = 14.7 Hz, 1H), 4.98 (d, *J* = 14.7 Hz, 1H), 4.64 (d, *J*

= 14.1 Hz, 1H), 4.51 (d, J = 14.1 Hz, 1H), 2.42 (s, 3H), 2.13 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.5, 164.7, 143.5, 143.3, 140.0, 137.8, 136.4, 131.4, 130.2, 129.4, 129.3, 128.3, 128.1, 127.8, 127.7, 127.1, 126.9, 125.3, 96.1, 79.2, 77.4, 48.1, 21.5, 19.8. HRMS (ESI) m/z Calcd for [C₃₂H₂₈BrNO₄S, M + Na]⁺: 624.0815, 626.0799; Found: 625.0811, 626.0804.

Optical Rotation: $[\alpha]^{25}_{D}$ -10.6 (c = 0.5, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 11.580 min for minor isomer, t_{R} = 14.187 min for major isomer).





3j (91% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, Rf = 0.4). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.70 (d, *J* = 8.2 Hz, 2H), 7.33 – 7.21 (m, 12H), 7.12–7.03 (m, 2H), 7.01-6.99 (m, 1H), 6.79 (s, 1H), 5.13 (d, *J* = 14.6 Hz, 1H), 4.97 (d, *J* = 14.6

Hz, 1H), 4.66 (d, J = 14.0 Hz, 1H), 4.41 (d, J = 14.0 Hz, 1H), 2.41 (s, 3H), 2.13 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.4, 164.7, 143.4, 143.3, 140.0, 137.6, 135.0, 134.9, 129.6, 129.3, 129.1, 128.4, 128.0, 127.9, 127.8, 127.8, 127.1, 126.9, 96.1, 79.2, 77.4, 48.1, 21.5, 19.4. HRMS (ESI) m/z Calcd for $[C_{32}H_{28}CINO_4S, M + Na]^+$: 580.1320, 582.1304; Found: 580.1325, 582.1308.

Optical Rotation: $[\alpha]^{25}_{D}$ -5.4 (c = 0.25, CHCl₃). 84% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 11.567$ min for minor isomer, $t_{R} = 13.870$ min for major isomer).





3k (98% yield, Hexane-EtOAc = 5:1, Rf = 0.3). Syrup. ¹H **NMR** (400 MHz, CDCl₃) δ 7.85 - 7.67 (m, 3H), 7.37 - 7.22 (m, 13H), 6.80 (s, 1H), 5.15 (d, *J* = 14.8 Hz, 1H), 5.04 (d, J = 17/150 14.8 Hz, 1H), 4.58 (d, J = 14.2 Hz, 1H), 4.52 (d, J = 14.2 Hz, 1H), 2.56 (dq, J = 15.1, 7.5 Hz, 1H), 2.43 (s, 3H), 2.31 (dq, J = 15.0, 7.5 Hz, 1H), 0.97 (t, J = 7.5 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 216.8, 164.7, 149.8, 143.7, 140.1, 139.9, 138.7, 138.0, 132.1, 129.4, 128.4, 127.9, 127.8, 127.3, 127.0, 123.0, 102.0, 95.9, 79.4, 77.5, 48.8, 25.3, 21.6, 14.0. **HRMS (ESI)** m/z Calcd for [C₃₃H_{29Br}INO₄S, M + Na]⁺: 763.9938, 765,9922; Found: 763.9939, 769.9924.

Optical Rotation: $[\alpha]^{25}_{D}$ -15.3 (c = 0.5, CHCl₃). 75% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 12.590 min for minor isomer, t_{R} = 11.311 min for major isomer).





31 (72% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, R_f = 0.5). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.63 (dd, J = 6.6, 2.8 Hz, 1H), 7.55 (d, J = 8.2 Hz, 2H), 7.29-7.18 (m, 14H), 6.78 (s, 1H), 5.15 (d, J = 14.6 Hz, 1H), 5.03 (d, J = 14.6 Hz, 1H),

4.75 (d, *J* = 14.4 Hz, 1H), 4.66 (d, *J* = 14.4 Hz, 1H), 3.45 (s, 3H), 2.39 (s, 3H), 1.83 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.7, 166.6, 165.2, 142.9, 141.3, 140.2, 138.3, 136.5, 134.8, 132.4, 129.5, 129.3, 128.4, 128.3, 128.1, 127.8, 127.8, 127.5, 127.0, 96.9, 78.9, 51.8, 50.3, 21.5, 18.7. HRMS (ESI) m/z Calcd for [C₃₄H₃₁NO₆S, M + Na]⁺: 604.1764; Found: 604.1767.

Optical Rotation: $[\alpha]^{25}_{D}$ -45.0 (*c* = 1.0, CHCl₃). 82% *ee* (HPLC conditions: 18/150

Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{\rm R}$ = 26.212 min for minor isomer, $t_{\rm R}$ = 20.444 min for major isomer).





3m (98% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, $R_f = 0.5$). White solid. MP: 69-70 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.40 (s, 1H), 7.94 – 7.82 (m, 4H), 7.26-7.53 (m, 2H), 7.40 (d, J = 1.4 Hz, 1H), 7.25 – 7.19 (m,

10H), 6.85 - 6.79 (m, 1H), 6.75 (s, 1H), 5.08 (d, J = 14.7 Hz, 1H), 4.97 (d, J = 14.7 Hz, 1H), 4.63 (s, 2H), 2.16 (s, 3H), 1.99 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.8, 164.9, 142.0, 140.1, 140.1, 139.9, 139.0, 138.3, 137.1, 135.0, 132.3, 132.2, 129.8, 129.5, 129.4, 128.9, 128.6, 128.4, 127.9, 127.9, 127.8, 127.3, 127.0, 124.1, 101.4, 96.1, 79.3, 77.4, 48.8, 20.5, 20.2. HRMS (ESI) m/z Calcd for [C₃₆H₃₀INO₄S, M + Na]⁺: 722.0832; Found: 722.0833.

Optical Rotation: $[\alpha]^{25}_{D}$ -44.0 (c = 1.0, CHCl₃). 89% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 21.687$ min for minor isomer, $t_{R} = 29.458$ min for major isomer).





3n (90% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, R_f = 0.5). White solid. MP: 69-70 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.88-7.84 (m, 2H), 7.43 (s, 1H), 7.30 – 7.19 (m, 10H), 7.10 (t, *J* = 8.6 Hz, 2H), 6.85 (s, 1H), 6.78 (s, 1H),

5.08 (d, J = 14.7 Hz, 1H), 4.97 (d, J = 14.7 Hz, 1H), 4.59 (d, J = 14.2 Hz, 1H), 4.54 (d, J = 14.2 Hz, 1H), 2.19 (s, 3H), 2.04 (s, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 216.7, 165.2 (d, ${}^{1}J_{C-F} = 252.7$ Hz), 164.8, 141.9, 140.0, 140.0, 138.9, 137.3 (d, ${}^{4}J_{C-F} = 3.0$ Hz), 136.9, 132.2, 131.0 (d, ${}^{3}J_{C-F} = 9.3$ Hz), 128.3, 127.9, 127.8, 127.2, 126.9, 115.8 (d, ${}^{2}J_{C-F} = 22.4$ Hz), 101.0, 95.9, 79.2, 77.4, 48.8, 20.6, 20.0. ¹⁹**F NMR** (376 MHz, CDCl₃) δ -105.53. **HRMS (ESI)** m/z Calcd for [C₃₂H₂₇FINO₄S, M + Na]⁺: 690.0582; Found: 690.0587.

Optical Rotation: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 84% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 15.896$ min for minor isomer, $t_{R} = 17.224$ min for major isomer).





30 (98% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, $R_f = 0.5$). White solid. MP: 139-140 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.82 (d, J = 8.6 Hz, 2H), 7.45 (s, 1H), 7.43 (d, J = 8.6 Hz, 2H), 7.33 – 7.25 (m, 10H), 6.88 (s,

1H), 6.81 (s, 1H), 5.10 (d, J = 14.7 Hz, 1H), 4.99 (d, J = 14.7 Hz, 1H), 4.63 (d, J = 14.1 Hz, 1H), 4.56 (d, J = 14.1 Hz, 1H), 2.21 (s, 3H), 2.05 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.7, 164.8, 141.9, 140.0, 139.9, 139.7, 139.0, 138.9, 136.8, 132.2, 129.8, 128.9, 128.3, 127.9, 127.8, 127.3, 126.9, 101.1, 95.9, 77.4, 48.9, 20.4, 20.0. HRMS (ESI) m/z Calcd for [C₃₂H₂₇ClINO₄S, M + Na]⁺: 706.0286; Found: 706.0286.

Optical Rotation: $[\alpha]^{25}{}_{D}$ -33.0 (c = 1.0, CHCl₃). 88% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 18.899 min for minor isomer, t_{R} = 20.709 min for major isomer).





3p (92% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, $R_f = 0.4$). White solid. MP: 139-149 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, J = 8.5 Hz, 2H), 7.59 (d, J = 8.5 Hz, 2H), 7.45 (s, 1H), 7.34 – 7.22 (m, 10H), 6.88 (s, 1H),

6.81 (s, 1H), 5.09 (d, *J* = 14.7 Hz, 1H), 4.99 (d, *J* = 14.7 Hz, 1H), 4.63 (d, *J* = 14.1 Hz, 1H), 4.56 (d, *J* = 14.1 Hz, 1H), 2.21 (s, 3H), 2.05 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.7, 164.8, 141.9, 140.2, 140.0, 139.9, 138.9, 136.7, 132.2, 131.9, 129.9, 128.3, 127.9, 127.8, 127.5, 127.3, 126.9, 101.1, 95.8, 79.2, 77.4, 48.9, 20.4, 20.0.

HRMS (ESI) m/z Calcd for [C₃₂H₂₇BrINO₄S, M + Na]⁺: 749.9781, 751.9765; Found: 749.9785, 751.9761.

Optical Rotation: $[\alpha]^{25}_{D}$ -32.0 (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 23.170$ min for minor isomer, $t_{R} = 25.031$ min for major isomer).





3q (95% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, $R_f = 0.4$). White solid. MP: 139-140 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, J = 8.5 Hz, 2H), 7.59 (d, J = 8.5 Hz, 2H), 7.45 (s, 1H), 7.34 – 7.25 (m, 10H), 6.87 (s, 1H),

6.80 (s, 1H), 5.09 (d, J = 14.7 Hz, 1H), 4.98 (d, J = 14.7 Hz, 1H), 4.62 (d, J = 14.1 Hz, 1H), 4.55 (d, J = 14.1 Hz, 1H), 2.21 (s, 3H), 2.04 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.7, 164.8, 141.9, 140.9, 140.0, 139.9, 138.9, 137.9, 136.7, 132.2, 129.8, 128.3, 127.9, 127.8, 127.3, 126.9, 101.2, 100.0, 95.9, 79.2, 77.4, 48.9, 20.4, 20.0. HRMS (ESI) m/z Calcd for [C₃₂H₂₇I₂NO₄S, M + Na]⁺: 797.9642; Found: 797.9647.

Optical Rotation: $[\alpha]^{25}_{D}$ -13.0 (c = 1.0, CHCl₃). 85% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 24.642$ min for minor isomer, $t_{R} = 27.598$ min for major isomer).





3r (97% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, R_f = 0.4). Syrup. ¹**H NMR** (400 MHz, CDCl₃) δ 7.70 (d, J = 8.6 Hz, 2H), 7.39 (d, J = 8.6 Hz, 2H), 7.35 (s, 1H), 7.24 – 7.13 (m, 10H), 6.75 (s, 1H), 6.71 (s, 1H), 5.05 (d, J =

14.6 Hz, 1H), 4.94 (d, J = 14.6 Hz, 1H), 4.50 (s, 2H), 2.11 (s, 3H), 1.95 (s, 3H), 1.25 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 216.8, 164.8, 156.5, 142.0, 140.1, 140.0, 139.7, 138.9, 138.1, 137.2, 132.1, 128.3, 128.1, 127.8, 127.7, 127.2, 126.9, 125.7, 101.0, 96.3, 79.2, 77.4, 48.5, 35.1, 31.1, 20.3, 20.1. HRMS (ESI) m/z Calcd for $[C_{36}H_{36}INO_4S, M + Na]^+$: 728.1302; Found: 728.1306.

Optical Rotation: $[\alpha]^{25}_{D}$ -21.6 (c = 0.5, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 9.699 min for minor isomer, t_{R} = 11.065 min for major isomer).





3s (90% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, $R_f = 0.5$). White solid. MP: 138-139 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, J = 8.1 Hz, 2H), 7.72 (d, J = 8.1_{23/150} Hz, 2H), 7.45 (s, 1H), 7.35-7.26 (m, 10H), 6.89 (s, 1H), 6.80 (s, 1H), 5.09 (d, J = 14.8 Hz, 1H), 4.98 (d, J = 14.8 Hz, 1H), 4.66 (d, J = 14.1 Hz, 1H), 4.58 (d, J = 14.1 Hz, 1H), 2.22 (s, 3H), 2.05 (s, 3H). ¹³C **NMR** (100 MHz, CDCl₃) δ 216.8, 164.8, 144.5, 141.9, 140.2, 139.9, 139.9, 139.0, 136.5, 134.2 (q, ${}^{2}J_{C-F} = 32.8$ Hz), 132.3, 128.9, 128.3, 127.9, 127.8, 127.30, 126.9, 125.8 (q, ${}^{3}J_{C-F} = 3.3$ Hz), 123.4 (q, ${}^{1}J_{C-F} = 271.2$ Hz), 101.0, 95.8, 79.2, 77.5, 49.1, 20.4, 20.0. ¹⁹F **NMR** (376 MHz, CDCl₃) δ -62.85. **HRMS (ESI)** m/z Calcd for [C₃₃H₂₇F₃INO₄S, M + Na]⁺: 740.0550; Found: 740.0552.

Optical Rotation: $[\alpha]^{25}_{D}$ -28 (c = 1.0, CHCl₃). 85% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 16.342$ min for minor isomer, $t_{R} = 13.669$ min for major isomer).





3t (93% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, R_f = 0.5). White solid. MP: 77-78 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.93 (d, J = 8.3 Hz, 2H), 7.67 (d, J = 8.3 Hz, 2H), 7.62 (d, J = 7.5 Hz, 2H), 7.51 – 7.39 (m, 4H), 7.27-

7.22 (m, 10H), 6.86 (s, 1H), 6.80 (s, 1H), 5.13 (d, J = 14.7 Hz, 1H), 5.01 (d, J = 14.7 Hz, 1H), 4.63 (s, 2H), 2.20 (s, 3H), 2.06 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.8, 164.8, 145.4, 142.0, 140.0, 139.8, 139.8, 139.4, 138.9, 137.1, 132.1, 129.0, 128.8, 128.3, 128.3, 127.8, 127.7, 127.3, 127.3, 127.2, 126.9, 101.2, 96.1, 79.2, 77.4, 48.7, 20.4, 20.1. HRMS (ESI) m/z Calcd for [C₃₈H₃₂INO₄S, M + Na]⁺: 748.0989; Found: 748.0991.

Optical Rotation: $[\alpha]^{25}_{D}$ -31.0 (c = 1.0, CHCl₃). 95% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 37.085 min for minor isomer, t_{R} = 39.892 min for major isomer).





3u (83% yield). Syrup. ¹**H** NMR (400 MHz, CDCl₃) δ 7.68 (s, 1H), 7.66-7.64 (m, 1H), 7.43 (s, 1H), 7.36 – 7.31 (m, 2H), 7.30 – 7.22 (m, 10H), 6.83 (s, 1H), 6.77 (s, 1H), 5.12 (d, *J* = 14.6 Hz, 1H), 5.00 (d, *J* = 14.6 Hz, 1H), 4.57 (s, 2H), 2.37 (s,

3H), 2.18 (s, 3H), 2.02 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.8, 164.8, 142.0, 141.0, 140.0, 140.0, 139.8, 138.9, 138.9, 137.1, 133.4 132.1, 128.7, 128.6, 128.3, 127.8, 127.7, 127.2, 126.9, 125.3, 101.1, 96.1, 79.2, 77.3, 48.6, 21.3, 20.4, 20.1. HRMS (ESI) m/z Calcd for [C₃₃H₃₀INO₄S, M + Na]⁺: 686.0832; Found: 686.0837.

Optical Rotation: $[\alpha]^{25}_{D}$ -45.0 (c = 1.0, CHCl₃). 93% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 8.465$ min for minor isomer, $t_{R} = 9.646$ min for major isomer).





3v (90% yield, Hexane-EtOAc = 5:1, $R_f = 0.3$). White solid. MP: 134-135 °C. ¹**H** NMR (400 MHz, CDCl₃) δ 7.42 (s, 1H), 7.34 - 7.12 (m, 10H), 6.89 (s, 1H), 6.84 (s, 1H), 4.90 (d, J =14.7 Hz, 1H), 4.78 (d, J = 14.7 Hz, 1H), 4.64 (d, J = 14.2 Hz,

1H), 4.02 (d, J = 14.2 Hz, 1H), 3.26 (s, 3H), 2.22 (s, 3H), 2.16 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.6, 165.4, 140.9, 140.0, 134.0, 139.8, 138.4, 137.2, 132.5, 128.4, 128.4, 128.0, 127.9, 127.6, 126.9, 103.6, 95.5, 78.8, 77.6, 48.9, 42.7, 20.4, 19.9. HRMS (ESI) m/z Calcd for [C₂₇H₂₆INO₄S, M + Na]⁺: 610.0519; Found: 610.0524.

Optical Rotation: $[\alpha]^{25}_{D}$ -5.0 (c = 1.0, CHCl₃). 97% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 17.561$ min for minor isomer, $t_{R} = 11.438$ min for major isomer).





3w (97% yield, Hexane-EtOAc = 5:1, $R_f = 0.4$). White solid. MP: 114-115 °C. ¹**H** NMR (400 MHz, CDCl₃) δ 7.41 (s, 1H), 7.33 – 7.13 (m, 10H), 6.87 (s, 1H), 6.81 (s, 1H), 4.90 (d, J =14.7 Hz, 1H), 4.79 (d, J = 14.7 Hz, 1H), 4.61 (d, J = 14.0 Hz, 1H), 4.07 (d, J = 14.0 Hz, 1H), 3.48 (dq, J = 14.7, 7.4 Hz, 1H), 3.39 (dq, J = 14.7, 7.4 Hz, 1H), 2.23 (s, 3H), 2.14 (s, 3H), 1.35 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 216.8, 165.3, 141.2, 140.0, 139.9, 139.9, 138.4, 137.1, 132.5, 128.4, 128.4, 128.0, 127.9, 127.5, 126.9, 103.5, 95.6, 78.8, 77.5, 49.9, 48.9, 20.4, 20.0, 8.1. HRMS (ESI) m/z Calcd for [C₂₈H₂₈INO₄S, M + Na]⁺: 624.0676; Found: 624.0672.

Optical Rotation: $[\alpha]^{25}_{D}$ -4 (c = 1.0, CHCl₃). 97% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 16.322$ min for minor isomer, $t_{R} = 9.050$ min for major isomer).





3x (95% yield, Hexane-EtOAc = 5:1, $R_f = 0.3$). White solid. MP: 96-97 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.51 (d, J = 0.8 Hz, 1H), 7.38 – 7.23 (m, 10H), 6.91 (d, J = 0.8 Hz, 1H), 6.86 (s, 1H), 4.98 (s, 2H), 4.65 (d, J = 14.0 Hz, 1H), 4.37 (d, J = 14.0

Hz, 1H), 2.93-2.86 (m, 1H), 2.25 (s, 3H), 2.22 (s, 3H), 1.31 – 1.23 (m, 1H), 1.21 – 1,14 (m, 1H), 1.10 – 1.01 (m, 1H), 1.0 – 0.94 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 216.7, 165.0, 141.2, 140.0, 139.9, 139.8, 138.5, 137.2, 132.3, 128.4, 128.3, 127.9, 127.8, 127.3, 126.9, 103.1, 95.8, 78.9, 77.4, 48.8, 32.2, 20.4, 20.0, 7.2, 6.5. HRMS (ESI) m/z Calcd for [C₂₉H₂₈INO₄S, M + Na]⁺: 636.0676; Found: 636.0677.

Optical Rotation: $[\alpha]^{25}_{D}$ -3.0 (c = 1.0, CHCl₃). 99% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 14.665$ min for minor isomer, $t_{R} = 17.098$ min for major isomer).



Representative procedure for synthesis of 5:



To a Schlenk tube containing **6** (0.05 mmol), β -ICD (1.5 mg, 10 mol%) and Cs₂CO₃ (0.05 mmol, 1.0 equiv.) were added mesitylene (3 mL) and MBH acetate **4** (0.14 mmol, 1.4 equiv.). The reaction mixture was stirred at -50 °C for 24 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product **5**.



Characterization of compounds 5 5a. (94% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 77.75 (d, J = 8.2 Hz, 2H), 7.50 (s, 1H), 7.29 (d, J = 8.2 Hz, 2H), 6.98 (s, 1H), 6.27 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.57 (d, J =

14.2 Hz, 1H), 4.51 (d, J = 14.2 Hz, 1H), 3.55 (s, 3H), 2.42 (s, 3H), 2.22 (s, 3H), 2.16 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.5, 143.5, 141.8, 139.8, 139.0, 138.2, 137.1, 135.4, 132.2, 131.9, 129.4, 128.3, 100.8, 51.9, 50.0, 21.5, 20.3, 20.2. HRMS (ESI) m/z Calcd for [C₂₀H₂₂INO₄S, M + H]⁺: 500.0387; Found: 500.0391.

Optical Rotation: $[\alpha]^{25}_{D}$ - 32.0 (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 10.767$ min for minor isomer, $t_{R} = 9.470$ min for major isomer).





5b. (95% yield, Hexane-EtOAc = 5:1, R_f = 0.5). White solid. **MP:** 122-124 °C. ¹**H NMR** (400 MHz, CDCl₃) δ 7.74 (d, *J* = 8.2 Hz, 2H), 7.50 (s, 1H), 7.28 (d, *J* = 8.2 Hz, 2H), 6.98 (s, 1H), 6.29 (s, 1H), 5.80 (s, 1H), 4.58 (d, *J* = 14.2 Hz, 1H), 4.52 (d, *J* = 14.2 Hz,

1H), 4.13–3.92 (m, 2H), 2.42 (s, 3H), 2.22 (s, 3H), 2.18 (s, 3H), 1.12 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.1, 143.4, 142.1, 139.8, 139.0, 138.4, 137.3, 135.9, 132.2, 131.6, 129.4, 128.3, 100.6, 60.9, 49.9, 21.5, 20.3, 20.3, 13.8. HRMS (ESI) m/z Calcd for [C₂₁H₂₄INO₄S, M + Na]⁺: 536.0363; Found: 536.0366.

Optical Rotation: $[\alpha]^{25}_{D}$ - 24.0 (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 8.212 min for minor isomer, t_{R} = 8.861 min for major isomer).



5c. (97% yield, Hexane-EtOAc = 5:1, $R_f = 0.4$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.74 (d, J = 8.3 Hz, 2H), 7.49 (s, 1H), 7.28 (d, J = 8.3 Hz, 2H), 6.98 (s, 1H), 6.29 (d, J = 0.9 Hz, 1H), 5.83 (s, 1H), 4.58 (d, J = 14.3 Hz, 1H), 4.51 (d, J = 14.3 Hz, 1H),

4.04 - 3.85 (m, 2H), 2.41 (s, 3H), 2.22 (s, 3H), 2.17 (s, 3H), 1.50 - 1.41 (m, 2H), 1.28 (dq, J = 14.4, 7.3 Hz, 2H), 0.88 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.1, 143.4, 142.0, 139.7, 139.0, 138.3, 137.3, 135.9, 132.2, 131.6, 129.4, 128.2, 100.6, 64.8, 49.9, 30.3, 21.5, 20.3, 20.3, 19.0, 13.6. HRMS (ESI) m/z Calcd for [C₂₃H₂₈INO₄S, M + Na]⁺: 564.0676; Found: 564.0677.

0,0

Me

Tol⁵ Me CO₂ⁿBu

5c

Optical Rotation: $[\alpha]^{25}_{D}$ - 24.0 (c = 1.0, CHCl₃). >99% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 6.472$ min for minor isomer, $t_{R} = 7.548$ min for major isomer).





5d. (94% yield, Hexane-EtOAc = 5:1, R_f = 0.5). Syrup. ¹H NMR (400 MHz, CDCl₃) δ7.66 (d, *J* = 8.2 Hz, 2H), 7.42 (s, 1H), 7.20 (d, *J* = 8.2 Hz, 2H), 6.92 (s, 1H), 6.18 (d, *J* = 0.9 Hz, 1H), 5.73 (s, 1H), 4.50 (d, *J* = 14.3 Hz, 1H), 4.39 (d, *J* = 14.3 Hz, 1H), 2.35 (s,

3H), 2.15 (s, 3H), 2.14 (s, 3H), 1.24 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 165.1, 143.4, 142.4, 139.8, 139.0, 138.5, 137.2, 132.2, 131.2, 129.4, 128.3, 100.3, 81.0, 49.7, 27.7, 21.5, 20.5, 20.3. HRMS (ESI) m/z Calcd for [C₂₃H₂₈INO₄S, M + Na]⁺: 564.0676; Found: 564.0681.

Optical Rotation: $[\alpha]^{25}_{D}$ - 14.0 (c = 1.0, CHCl₃). 91% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 9.470$ min for minor isomer, $t_{R} = 10.767$ min for major isomer).



5e. (94% yield, Hexane-EtOAc = 5:1, $R_f = 0.4$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.64 (d, J = 8.3 Hz, 2H), 7.35 (d, J = 0.6Hz, 1H), 7.27 – 7.20 (m, 3H), 7.19 – 7.13 (m, 4H), 6.83 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.6 Hz, 1H), 6.25 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 4.98 (d, J = 0.8 Hz, 1H), 5.76 (s, 1H), 5.76 (s,

12.4 Hz, 1H), 4.88 (d, J = 12.4 Hz, 1H), 4.50 (d, J = 14.3 Hz, 1H), 4.45 (d, J = 14.3 Hz, 1H), 2.32 (s, 3H), 2.10 (s, 3H), 2.03 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 165.8, 143.4, 141.8, 139.7, 139.0, 138.1, 137.2, 135.6, 135.4, 132.1, 132.1, 129.4, 128.3, 128.2, 128.1, 128.1, 100.7, 66.6, 50.0, 21.5, 20.3, 20.2. HRMS (ESI) m/z Calcd for [C₂₆H₂₆INO₄S, M + Na]⁺: 598.0519; Found: 598.0523.

Optical Rotation: $[\alpha]^{25}_{D}$ - 24.0 (c = 1.0, CHCl₃). 91% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 12.137$ min for minor isomer, $t_{R} = 13.047$ min for major isomer).





5f. (92% yield, Hexane-EtOAc = 5:1, R_f = 0.4). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 7.72 (d, J = 7.8 Hz, 2H), 7.27 (d, J = 8.0 Hz, 2H), 7.17 (s, 1H), 6.97 (s, 1H), 6.27 (s, 1H), 5.80 (s, 1H), 4.61 (d,

J = 14.2 Hz, 1H), 4.44 (d, J = 14.2 Hz, 1H), 4.13 – 3.82 (m, 2H), 2.42 (s, 3H), 2.25 (s, 6H), 1.52–1.45 (m, 2H), 1.35 – 1.24 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.1, 143.4, 142.8, 139.6, 137.9, 136.0, 133.9, 132.1, 131.3, 131.2, 129.4, 128.1, 124.6, 64.8, 49.7, 30.4, 21.6, 20.6, 20.0, 19.1, 13.7. HRMS (ESI) m/z Calcd for [C₂₃H₂₈BrNO₄S, M + Na]⁺: 516.0815; Found: 516.0819.

Optical Rotation: $[\alpha]^{25}_{D} - 4.2$ (c = 1.0, CHCl₃). 83% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 95:5, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 10.953$ min for minor isomer, $t_{R} = 9.292$ min for major isomer).





5g. (82% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, J = 1.6 Hz, 1H), 7.73 (d, J = 8.2Hz, 2H), 7.52 (d, J = 1.6 Hz, 1H), 7.30 (d, J = 8.2 Hz, 2H), 6.33 (s, 1H), 5.88 (s, 1H), 4.56 (d, J = 14.3 Hz, 1H), 4.49 (d, J = 14.3

Hz, 1H), 4.00 (dt, J = 10.9, 6.8 Hz, 1H), 3.91 (dt, J = 10.9, 6.8 Hz, 1H), 2.43 (s, 3H), 2.18 (s, 3H), 1.50 – 1.41 (m, 2H), 1.33 – 1.26 (m, 2H), 0.90 (t, J = 7.3 Hz, 3H). ¹³C **NMR** (100 MHz, CDCl₃) δ 166.0, 146.1, 144.6, 143.8, 140.3, 140.2, 138.0, 135.7, 132.0, 129.6, 128.3, 101.9, 94.8, 65.0, 50.0, 30.3, 21.6, 20.2, 19.1, 13.7. **HRMS (ESI)** m/z Calcd for [C₂₂H₂₅I₂NO₄S, M + Na]⁺: 675.9486; Found: 675.9491.

Optical Rotation: $[\alpha]^{25}_{D} - 2.4$ (c = 0.5, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 95:5, flow rate = 1.0 mL/min, wavelength = 254 nm, t_R = 11.867 min for minor isomer, t_R = 9.781 min for major isomer).





5h. (80% yield, Hexane-EtOAc = 5:1, R_f = 0.5). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 7.81 (d, J = 1.8 Hz, 1H), 7.73 (d, J = 8.1 Hz, 2H), 7.41 (d, J = 1.8 Hz, 1H), 7.30 (d, J = 8.1 Hz, 2H), 6.34 (s, 1H), 5.86 (s, 1H), 4.54 (d, J = 14.3 Hz, 1H), 4.49 (d, J = 14.3 Hz, 1H)

1H), 4.03 - 3.96 (m, 1H), 3.96 - 3.88 (m, 1H), 2.66 (dq, J = 15.1, 7.5 Hz, 1H), 2.50 (dq, J = 15.1, 7.5 Hz, 1H), 2.44 (s, 3H), 1.50 - 1.43 (m, 2H), 1.33 - 1.25 (m, 2H), 1.14 (t, J = 7.5 Hz, 3H), 0.90 (t, J = 7.3 Hz, 3H).. ¹³C NMR (100 MHz, CDCl₃) δ 166.0, 162.5, 149.9, 143.8, 140.3, 138.7, 138.0, 135.5, 132.2, 129.6, 128.3, 123.0, 101.3, 65.0, 50.0, 30.4, 25.4, 21.6, 19.1, 14.2, 13.7. HRMS (ESI) m/z Calcd for [C₂₃H₂₇BrINO₄S, M + Na]⁺: 641.9781; Found: 641.9785.

Optical Rotation: $[\alpha]^{25}_{D} - 5.6$ (c = 1.0, CHCl₃). 89% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 95:5, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 8.678$ min for minor isomer, $t_{R} = 7.688$ min for major isomer)





5i. (95% yield, Hexane-EtOAc = 5:1, R_f = 0.5). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 7.73 (d, J = 8.2 Hz, 2H), 7.35 (d, J = 7.9 Hz, 1H), 7.28 (d, J = 8.2 Hz, 2H), 7.17 (d, J = 7.5 Hz, 1H), 7.04 (dd, J = 7.9, 7.5 Hz, 1H), 6.28 (s, 1H), 5.82 (s, 1H), 4.62 (d, J = 14.3)

Hz, 1H), 4.47 (d, J = 14.3 Hz, 1H), 4.00 (dt, J = 11.0, 6.6 Hz, 1H), 3.91 (dt, J = 11.0, 6.6 Hz, 1H), 2.43 (s, 3H), 2.30 (s, 3H), 1.52 – 1.43 (m, 2H), 1.34 – 1.25 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 166.1, 143.6, 143.5, 137.9, 136.7, 135.9, 131.6, 131.3, 130.4, 129.4, 129.4, 128.1, 125.0, 64.9, 49.7, 30.4, 21.6, 20.1, 19.1, 13.7. **HRMS (ESI)** m/z Calcd for [C₂₂H₂₆BrNO₄S, M + Na]⁺: 502.0658; Found: 502.0662.

Optical Rotation: $[\alpha]^{25}_{D}$ - 10.4 (c = 1.0, CHCl₃). 83% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 7.597$ min for minor isomer, $t_{R} = 6.677$ min for major isomer)





5j. (92% yield, Hexane-EtOAc = 5:1, $R_f = 0.4$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, J = 8.3 Hz, 2H), 7.25 (d, J = 8.3 Hz, $_{35/150}$ 2H), 7.18 – 7.02 (m, 3H), 6.23 (d, J = 1.1 Hz, 1H), 5.77 (s, 1H), 4.61 (d, J = 14.2 Hz, 1H), 4.37 (d, J = 14.2 Hz, 1H), 3.99 (dt, J = 10.9, 6.7 Hz, 1H), 3.91 (dt, J = 10.9, 6.7 Hz, 1H), 2.41 (s, 3H), 2.29 (s, 3H), 1.51 – 1.42 (m, 2H), 1.33 – 1.23 (m, 2H), 0.87 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.32, 143.77, 143.64, 137.95, 136.23, 135.47, 135.12, 131.31, 130.09, 129.69, 129.40, 128.40, 128.31, 65.15, 50.00, 30.68, 21.86, 20.06, 19.36, 13.95. HRMS (ESI) m/z Calcd for [C₂₂H₂₆ClNO₄S, M + Na]⁺: 458.1163; Found: 458.1168.

Optical Rotation: $[\alpha]^{25}_{D}$ - 2.6 (c = 1.0, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 5.660$ min for minor isomer, $t_{R} = 5.172$ min for major isomer).





5k. (87% yield, Hexane-EtOAc-Chlorobenzene = 7:1:1, $R_f = 0.6$). Syrup. ¹**H NMR** (400 MHz, CDCl₃) δ 7.64 (dd, J = 7.6, 1.4 Hz, 1H), 7.52 (d, J = 8.3 Hz, 2H), 7.31 (dd, J = 7.6, 1.4 Hz, 1H), 7.25 – 7.22 (m, 1H), 7.20 (d, J = 8.3 Hz, 2H), 6.31 (d, J = 1.3

Hz, 1H), 5.90 (s, 1H), 4.64 (d, J = 14.6 Hz, 1H), 4.58 (d, J = 14.6 Hz, 1H), 3.90 (t, J = 6.7 Hz, 2H), 3.54 (s, 3H), 2.38 (s, 3H), 1.94 (s, 3H), 1.41 – 1.32 (m, 2H), 1.26 – 1.16 (m, 2H), 0.82 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.8, 166.4, 143.0, 140.9, 138.1, 136.9, 135.0, 132.6, 131.1, 129.4, 128.1, 127.5, 64.7, 51.9, 51.8, 30.3, 21.5, 19.0, 18.9, 13.6. HRMS (ESI) m/z Calcd for [C₂₄H₂₉NO₆S, M + H]⁺: 460.1788; Found: 460.1783.
Optical Rotation: $[\alpha]^{25}_{D} - 0.4$ (c = 1.0, CHCl₃). 70% *ee* (HPLC conditions: Chiralpak AS-H column, *n*-Hexane/*i*-PrOH = 90:0, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 12.987$ min for minor isomer, $t_{R} = 12.167$ min for major isomer).





51. (97% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.54 (s, 1H), 7.03 (s, 1H), 6.22 (s, 1H), 5.45 (s, 1H), 4.71 (d, J = 14.1 Hz, 1H), 4.21 – 4.03 (m, 3H), 3.40 (s, 3H), 2.37 (s, 3H), 2.24 (s, 3H), 1.63 – 1.57 (m, 2H), 1.46 –

1.32 (m, 2H), 0.93 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.3, 140.9, 140.1, 138.6, 137.2, 135.2, 132.6, 131.7, 103.1, 65.1, 50.5, 42.8, 30.5, 20.4, 20.1, 19.2, 13.7. HRMS (ESI) m/z Calcd for [C₁₇H₂₄INO₄S, M + H]⁺: 466.0543; Found: 466.0547

Optical Rotation: $[\alpha]^{25}_{D} - 7.8$ (c = 1.0, CHCl₃). 98% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 7.17$ min for minor isomer, $t_{R} = 8.413$ min for major isomer).





5m. (92% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.54 (s, 1H), 7.03 (s, 1H), 6.22 (s, 1H), 5.49 (s, 1H), 4.69 (d, J = 14.0 Hz, 1H), 4.19 (d, J = 14.0 Hz, 1H), 4.16 – 3.96 (m, 2H), 3.75 – 3.46 (m, 2H), 2.38 (s, 3H), 2.24

(s, 3H), 1.61 - 1.54 (m, 2H), 1.47 (t, J = 7.4 Hz, 3H), 1.41 - 1.31 (m, 2H), 0.92 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.3, 141.2, 140.0, 138.6, 137.2, 135.4, 132.6, 131.8, 103.0, 65.1, 50.4, 50.0, 30.5, 20.3, 20.2, 19.2, 13.7, 8.1. HRMS (ESI) m/z Calcd for [C₁₈H₂₆INO₄S, M + Na]⁺: 502.0519; Found: 502.0523.

Optical Rotation: $[\alpha]^{25}_{D} - 4.6$ (c = 1.0, CHCl₃). 98% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 6.830$ min for minor isomer, $t_{R} = 5.341$ min for major isomer).





5n. (87% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.56 (s, 1H), 7.01 (s, 1H), 6.24 (s, 1H), 5.63 (s, 1H), 4.64 (d, J = 14.1 Hz, 1H), 4.42 (d, J = 14.1 Hz, 1H), 4.20 – 3.88 (m, 2H), 3.12 – 2.78 (m, 1H), 2.35 (s, 3H), 2.24 (s, 3H), 1.58 – 1.51 (m, 2H), 1.38 – 1.31 (m, 2H), 1.28 – 1.15 (m, 2H), 1.15 – 1.01 (m, 2H), 0.91 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.2, 141.4, 139.9, 138.7, 137.4, 135.7, 132.4, 131.6, 102.4, 65.0, 50.4, 32.4, 30.4, 20.3, 20.3, 19.1, 13.7, 7.2, 6.5. **HRMS (ESI)** m/z Calcd for [C₁₉H₂₆INO₄S, M + Na]⁺: 514.0519; Found: 514.0521.

Optical Rotation: $[\alpha]^{25}_{D} - 2.3$ (c = 1.0, CHCl₃). 96% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 6.928$ min for minor isomer, $t_{R} = 6.316$ min for major isomer).





50. (78% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.67–7.65 (m, 2H), 7.50 (s, 1H), 7.40–7.37 (m, 2H), 6.99 (s, 1H), 6.30 (s, 1H), 5.85 (s, 1H), 4.59 (d, J = 14.2 Hz, 1H), 4.52 (d, J = 14.2 Hz, 1H), 4.05 –

3.95 (m, 1H), 3.95 - 3.85 (m, 1H), 2.41 (s, 3H), 2.23 (s, 3H), 2.19 (s, 3H), 1.51 - 1.42 (m, 2H), 1.31 - 1.24 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.2, 142.2, 141.1, 139.8, 139.1, 139.0, 137.3, 136.0, 133.5, 132.2, 131.7, 128.7, 128.7, 125.4, 100.4, 64.9, 50.0, 30.4, 21.3, 20.4, 20.3, 19.1, 13.7. HRMS (ESI) m/z Calcd for [C₂₃H₂₈NO₄S, M + Na]⁺: 564.0676; Found: 564.0681.

Optical Rotation: $[\alpha]^{25}_{D} - 0.4$ (c = 1.0, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak AS-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, $_{39/150}$ wavelength = 254 nm, $t_{\rm R}$ = 5.553 min for minor isomer, $t_{\rm R}$ = 6.347 min for major isomer).





5p. (64% yield, Hexane-EtOAc = 5:1, R_f = 0.5). Syrup.
¹H NMR (400 MHz, CDCl₃) δ 7.79 (d, J = 8.7 Hz, 2H),
7.50 (s, 1H), 6.99 (s, 1H), 6.96 (d, J = 8.7 Hz, 2H), 6.29 (s,
1H), 5.83 (s, 1H), 4.57 (d, J = 14.3 Hz, 1H), 4.51 (d, J =

14.3 Hz, 1H), 4.00 (dt, J = 11.2, 6.8 Hz, 1H), 3.91 (dt, J = 11.2, 6.8 Hz, 1H), 3.87 (s, 3H), 2.23 (s, 3H), 2.20 (s, 3H), 1.53 – 1.41 (m, 2H), 1.34 – 1.24 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 166.2, 163.1, 142.1, 139.8, 139.1, 137.5, 136.0, 133.2, 132.3, 131.5, 130.4, 113.9, 100.5, 64.9, 55.6, 50.0, 30.4, 20.4, 20.3, 19.1, 13.7. **HRMS (ESI)** m/z Calcd for [C₂₃H₂₈INO₅S, M + H]⁺: 558.0806; Found: 558.0811.

Optical Rotation: $[\alpha]^{25}_{D} - 19.0$ (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 9.160$ min for minor isomer, $t_{R} = 9.902$ min for major isomer).





5q. (86% yield, Hexane-EtOAc = 5:1, R_f = 0.4). Syrup.
¹H NMR (400 MHz, CDCl₃) δ 8.34 (d, J = 8.5 Hz, 2H),
8.05 (d, J = 8.5 Hz, 2H), 7.49 (s, 1H), 7.03 (s, 1H), 6.31 (s, 1H),
5.78 (s, 1H), 4.65 (d, J = 14.1 Hz, 1H), 4.57 (d, J = 14.1 Hz, 1H),

14.1 Hz, 1H), 4.09 - 3.98 (m, 1H), 3.98 - 3.89 (m, 1H), 2.25 (s, 3H), 2.23 (s, 3H), 1.55 - 1.44 (m, 2H), 1.35 - 1.25 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 165.9, 150.1, 146.7, 142.3, 140.5, 139.2, 136.2, 135.3, 132.5, 132.2, 129.6, 124.1, 99.9, 65.0, 50.5, 30.4, 20.4, 20.4, 19.1, 13.6. HRMS (ESI) m/z Calcd for [C₂₂H₂₅IN₂O₆S, M + Na]⁺: 595.0370; Found: 595.0373.

Optical Rotation: $[\alpha]^{25}_{D} - 15.6$ (c = 1.0, CHCl₃). 89% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 7.914$ min for minor isomer, $t_{R} = 8.711$ min for major isomer).





5r. (97% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.87 (dd, J = 8.6, 5.2 Hz, 2H), 7.49 (s, 1H), 7.17 (t, J = 8.6 Hz, 2H), 7.00 (s, 1H), 6.30 (s, 1H), 5.80 (s, 1H), 4.60 (d, J = 14.2 Hz, 1H), 4.52 (d, J = 14.2 Hz, 1H), 4.09 – 3.97 (m, 1H), 3.97 – 3.88 (m, 1H), 2.23 (s, 3H), 2.21 (s, 3H), 1.52 – 1.43 (m, 2H), 1.34 – 1.25 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.4, 165.3 (${}^{1}J_{C-F} = 253.0$ Hz), 142.2, 140.1, 139.2, 137.4 (d, ${}^{4}J_{C-F} =$ 3.3 Hz), 137.0, 135.8, 132.4, 131.8, 131.0 (d, ${}^{3}J_{C-F} = 9.3$ Hz), 116.0 (d, ${}^{2}J_{C-F} = 22.5$ Hz), 100.2, 64.9, 50.2, 30.4, 20.4, 20.3, 19.1, 13.7. ¹⁹F NMR (376 MHz, CDCl₃) δ – 105.46. HRMS (ESI) m/z Calcd for [C₂₂H₂₅FINO₄S, M + Na]⁺: 568.0425; Found: 568.0429.

Optical Rotation: $[\alpha]^{25}_{D} - 30.0$ (c = 1.0, CHCl₃). 93% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 6.878$ min for minor isomer, $t_{R} = 6.110$ min for major isomer).





5s. (92% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.80 (d, J = 8.2 Hz, 2H), 7.50 (s, 1H), 7.47 (d, J = 8.2 Hz, 2H), 7.00 (s, 1H), 6.30 (s, 1H), 5.80 (s, 1H), 4.60 (d, J = 14.2 Hz, 1H), 4.52 (d, J = 14.2 Hz,

1H), 4.10 - 3.97 (m, 1H), 3.97 - 3.88 (m, 1H), 2.24 (s, 3H), 2.21 (s, 3H), 1.52 - 1.43 (m, 2H), 1.34 - 1.25 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.1, 142.2, 140.1, 139.7, 139.2, 139.2, 136.9, 135.7, 132.4, 131.9, 129.8, 129.1, 100.2, 65.0, 50.2, 30.4, 20.4, 20.3, 19.1, 13.7. HRMS (ESI) m/z Calcd for $[C_{22}H_{25}CIINO_4S, M + H]^+$: 584.0130; Found: 584.0132.

Optical Rotation: $[\alpha]^{25}_{D} - 15.0$ (c = 1.0, CHCl₃). 92% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 5.660$ min for minor isomer, $t_{R} = 5.172$ min for major isomer).





5t. (97% yield, Hexane-EtOAc = 5:1, $R_f = 0.4$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, J = 8.7 Hz, 2H), 7.63 (d, J = 8.7 Hz, 2H), 7.49 (s, 1H), 7.00 (s, 1H), 6.29 (d, J = 0.8 Hz, 1H), 5.79 (s, 1H), 4.59 (d, J = 14.2 Hz, 1H), 4.52 (d,

J = 14.2 Hz, 1H), 4.01 (dt, J = 10.9, 6.6 Hz, 1H), 3.92 (dt, J = 10.9, 6.6 Hz, 1H), 2.23 (s, 3H), 2.20 (s, 3H), 1.51 – 1.43 (m, 2H), 1.33 – 1.25 (m, 2H), 0.88 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.0, 142.1, 140.2, 140.0, 139.1, 136.8, 135.6, 132.3, 132.1, 131.8, 129.8, 127.7, 100.2, 64.9, 50.2, 30.3, 20.4, 20.3, 19.0, 13.6. HRMS (ESI) m/z Calcd for [C₂₂H₂₅BrINO₄S, M + Na]⁺: 627.9625; Found: 627.9630.

Optical Rotation: $[\alpha]^{25}_{D} - 25.5$ (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 6.173$ min for minor isomer, $t_{R} = 5.579$ min for major isomer).





5u. (98% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.84 (d, J = 8.5 Hz, 2H), 7.57 (d, J = 8.5 Hz, 2H), 7.49 (s, 1H), 6.99 (s, 1H), 6.29 (d, J = 0.8Hz, 1H), 5.79 (s, 1H), 4.59 (d, J = 14.2 Hz, 1H), 4.51 (d, J =

14.2 Hz, 1H), 4.00 (dt, J = 10.8, 6.8 Hz, 1H), 3.92 (dt, J = 10.8, 6.6 Hz, 1H), 2.22 (s, 3H), 2.20 (s, 3H), 1.50 – 1.43 (m, 2H), 1.33 – 1.24 (m, 2H), 0.88 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 166.0, 142.1, 140.8, 140.0, 139.1, 138.0, 136.8, 135.6, 132.3, 131.8, 129.7, 100.2, 100.1, 64.9, 50.2, 30.3, 20.4, 20.3, 19.0, 13.6. HRMS (ESI) m/z Calcd for [C₂₂H₂₅BrI₂NO₄S, M + Na]⁺: 675.9486; Found: 675.9491.

Optical Rotation: $[\alpha]^{25}_{D} - 25.7$ (c = 1.0, CHCl₃). 90% *ee* (HPLC conditions: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 85:25, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 6.114$ min for minor isomer, $t_{R} = 5.490$ min for major isomer).



Representative procedure for synthesis of 7:



To a Schlenk tube containing $6^{[4]}$ (0.05 mmol), β -ICD (1.5 mg, 10 mol%) and Cs₂CO₃ (0.05 mmol, 1.0 equiv.) were added mesitylene (3 mL) and MBH acetate 4c (0.14 mmol, 1.4 equiv.). The reaction mixture was stirred at -50 °C for 24 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography with hexanes/ethyl acetate as the eluent to afford the product 7.

Characterization of compounds of 7:



7a. (76% yield, Hexane-EtOAc = 3:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 9.15 (s, 1H), 8.07 (d, J = 8.2 Hz, 1H), 7.85 (d, J = 7.4 Hz, 2H), 7.55 (d, J = 8.2 Hz, 2H), 7.48-7.44 (m, 1H), 7.41-7.38 (m, 2H), 7.19-7.14 (m, 3H), 6.82 (d, J = 7.5 Hz, 1H), 6.03 (s,

1H), 5.53 (s, 1H), 4.72 (d, J = 14.0 Hz, 1H), 4.02 (d, J = 14.0 Hz, 1H), 3.72 – 3.54 (m, 2H), 2.31 (s, 3H), 1.60 (s, 3H), 1.33-1.26 (m, 2H), 1.15-1.06 (m, 2H), 0.75 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 165.6, 165.0, 144.2, 138.5, 138.3, 136.1, 135.2, 134.4, 131.7, 130.9, 129.7, 129.2, 128.5, 127.8, 127.8, 127.4, 127.1, 121.7, 64.8, 51.1, 30.2, 21.5, 18.9, 17.9, 13.6. **HRMS (ESI)** m/z Calcd for [C₂₉H₃₂N₂O₅S, M + Na]⁺: 543.1924; Found: 543.1929.

Optical Rotation: $[\alpha]^{25}_{D}$ 60.0 (c = 1.0, CHCl₃). 84% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 10.116$ min for minor isomer, $t_{R} = 7.849$ min for major isomer).

7b. (81% yield, Hexane-EtOAc = 3:1, R_f = 0.4). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 9.11 (s, 1H), 8.06 (d, J = 8.0 Hz, 1H), 7.74 (d, J = 8.2 Hz, 2H), 7.56 (d, J = 8.2 Hz, 2H), 7.20 (d, J = 7.8 Hz, 3H),
7.16 (d, J = 8.4 Hz, 2H), 6.81 (d, J = 7.5 Hz, 1H), 6.03 (s, 1H),

5.53 (s, 1H), 4.71 (d, J = 14.0 Hz, 1H), 4.02 (d, J = 14.0 Hz, 1H), 3.71-3.58 (m, 2H), 2.35 (s, 3H), 2.32 (s, 3H), 1.59 (s, 3H), 1.34 – 1.26 (m, 2H), 1.16-1.06 (m, 2H), 0.75 (t, J = 7.4 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 165.6, 165.0, 144.2, 142.2, 138.5, 138.4, 136.1, 135.2, 131.6, 131.0, 129.7, 129.2, 129.2, 127.8, 127.7, 127.4, 127.0, 121.7, 64.9, 51.1, 30.2, 21.6, 21.5, 18.9, 17.9, 13.6. **HRMS (ESI)** m/z Calcd for [C₃₀H₃₄N₂O₄S, M + Na]⁺: 557.2081; Found: 557.2085.

Optical Rotation: $[\alpha]^{25}_{D}$ 51.0 (c = 1.0, CHCl₃). 86% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 21.846$ min for minor isomer, $t_{R} = 14.998$ min for major isomer).

7c. (45% yield, Hexane-EtOAc = 3:1, $R_f = 0.4$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 9.15 (s, 1H), 8.12 (d, J = 8.0 Hz, 1H), 7.89 (d, J = 8.8 Hz, 2H), 7.63 (d, J = 8.2 Hz, 2H), 7.26 – 7.15 (m, 3H),

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6.96 (d, J = 8.8 Hz, 2H), 6.86 (d, J = 7.4 Hz, 1H), 6.10 (s, 1H), 5.58 (s, 1H), 4.79 (d, J = 14.0 Hz, 1H), 4.07 (d, J = 14.0 Hz, 1H), 3.86 (s, 3H), 3.76 (dt, J = 10.9, 6.8 Hz, 1H), 3.68 (dt, J = 10.9, 6.8 Hz, 1H), 2.39 (s, 3H), 1.65 (s, 3H), 1.42 – 1.31 (m, 2H), 1.18 (dq, J = 14.5, 7.3 Hz, 2H), 0.82 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) 8165.6, 164.6, 162.4, 144.2, 138.4, 136.1, 135.1, 130.9, 129.7, 129.3, 129.1, 127.8, 127.6, 126.8, 126.7, 121.6, 113.7, 64.8, 55.4, 51.0, 30.2, 21.5, 18.9, 17.9, 13.6. HRMS (ESI) m/z Calcd for [C₃₀H₃₄N₂O₆S, M + Na]⁺: 573.2030; Found: 573.2034.

Optical Rotation: $[\alpha]^{25}_D$ 34.0 (c = 1.0, CHCl₃). 76% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_R = 20.637$ min for minor isomer, $t_R = 12.800$ min for major isomer).

7d. (74% yield, Hexane-EtOAc = 3:1, R_f = 0.4). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 9.20 (s, 1H), 8.13 (d, J = 8.2 Hz, 1H), 7.85
(d, J = 8.5 Hz, 2H), 7.64 (d, J = 8.2 Hz, 2H), 7.48 (d, J = 8.5 Hz, 2H), 7.27 - 7.21 (m, 3H), 6.88 (d, J = 7.5 Hz, 1H), 6.10 (s, 1H),

5.58 (s, 1H), 4.78 (d, J = 14.0 Hz, 1H), 4.09 (d, J = 14.0 Hz, 1H), 3.74 (dt, J = 10.8, 6.8 Hz, 1H), 3.65 (dt, J = 10.8, 6.8 Hz, 1H), 2.39 (s, 3H), 1.66 (s, 3H), 1.42 – 1.28 (m, 11H), 1.17 (dq, J = 14.4, 7.4 Hz, 2H), 0.81 (t, J = 7.4 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 165.6, 164.9, 155.2, 144.1, 138.5, 138.4, 136.1, 135.1, 131.4, 131.0, 129.7, 129.1, 127.8, 127.7, 127.2, 127.0, 125.5, 121.7, 64.8, 51.0, 34.9, 31.1, 30.2, 21.5, 18.9, 17.9, 13.6. **HRMS (ESI)** m/z Calcd for [C₃₃H₄₀N₂O₅S, M + Na]⁺: 599.2550; Found: 599.2554.

Optical Rotation: $[\alpha]^{25}_{D}$ 45.0 (c = 1.0, CHCl₃). 89% *ee* (HPLC conditions: Chiralpak OZ-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 16.263$ min for minor isomer, $t_{R} = 10.780$ min for major isomer).

7e. (55% yield, Hexane-EtOAc = 3:1, R_f = 0.4). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 9.33 (s, 1H), 8.10 (d, J = 8.1 Hz, 1H), 8.03
(d, J = 8.1 Hz, 2H), 7.71 (d, J = 8.2 Hz, 2H), 7.57 (d, J = 8.2 Hz, 2H), 7.23 (d, J = 7.7 Hz, 1H), 7.18 (d, J = 8.1 Hz, 2H), 6.87 (d, J

= 7.5 Hz, 1H), 6.07 (s, 1H), 5.58 (s, 1H), 4.79 (d, J = 13.9 Hz, 1H), 3.99 (d, J = 13.9 Hz, 1H), 3.74 – 3.55 (m, 2H), 2.35 (s, 3H), 1.62 (s, 3H), 1.35 – 1.27 (m, 2H), 1.12 (dq, J = 14.4, 7.3 Hz, 2H), 0.77 (t, J = 7.3 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 165.6, 163.7, 144.3, 138.5, 138.0, 137.7, 135.9, 135.2, 133.3 (${}^{2}J_{C-F} = 32.5$ Hz), 131.1, 129.7, 129.3, 127.9, 127.9, 127.7, 127.5, 125.6 (${}^{3}J_{C-F} = 3.8$ Hz), 123.7 (${}^{1}J_{C-F} = 270.8$ Hz), 121.5, 64.9, 51.3, 30.2, 21.5, 18.9, 17.9, 13.5. ¹⁹**F NMR** (376 MHz, CDCl₃) 62.94. **HRMS (ESI)** m/z Calcd for [C₃₀H₃₁F₃N₂O₅S, M + Na]⁺: 599.2550; Found: 599.2554.

Optical Rotation: $[\alpha]^{25}_D$ 32.0 (c = 1.0, CHCl₃). 78% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_R = 13.026$ min for minor isomer, $t_R = 8.474$ min for major isomer).

Figure 50. HPLC Trace of 7e.

6f^[5]. (85% yield). Yellow solid. **MP:** 91-92 °C. ¹H NMR (400 MHz, DMSO) δ 12.12 (s, 1H), 11.04 (s, 1H), 9.49 (s, 1H), 7.29 (s, 1H), 3.16 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 154.7, 154.5, 126.6, 126.2, 125.3, 121.5, 115.1, 41.7. **HRMS (ESI)**

m/z Calcd for $[C_9H_7Cl_2N_3O_4S, M + Na]^+$: 345.9427, 347.9396; Found: 345.9428, 347.9400.

7g. (38% yield, Hexane-EtOAc = 2:1, $R_f = 0.4$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 8.01 – 7.90 (m, 2H), 7.89 – 7.81 (m, 2H), 7.59 (d, J = 8.2 Hz, 2H), 7.48 – 7.35 (m, 6H), 7.13 (d, J = 8.1 Hz, 2H), 7.01 (d, J = 8.1 Hz, 1H), 6.85 – 6.81 (m, 1H), 6.54 (d, J =

7.4 Hz, 1H), 6.42 (d, J = 10.6 Hz, 1H), 6.06 (s, 1H), 5.56 (s, 1H), 4.58 (d, J = 14.1 Hz, 1H), 4.04 (d, J = 14.1 Hz, 1H), 3.85 – 3.69 (m, 2H), 2.33 (s, 3H), 1.65 (s, 3H), 1.44 – 1.32 (m, 2H), 1.22 – 1.16 (m, 2H), 0.80 (s, 3H). ¹³C **NMR** (100 MHz, CDCl₃) δ 165.8, 144.0, 141.2, 138.8, 136.3, 135.5, 132.4 (${}^{1}J_{C-P} = 128.3$ Hz), 132.2 (${}^{2}J_{C-P} = 10.2$ Hz), 132.0 (${}^{1}J_{C-P} = 128.9$ Hz), 132.0 (${}^{3}J_{C-P} = 2.5$ Hz), 131.9 (${}^{3}J_{C-P} = 2.7$ Hz), 131.5 (${}^{2}J_{C-P} = 10$ Hz), 130.9, 129.7, 129.1, 128.5 (${}^{2}J_{C-P} = 24.7$ Hz), 128.6, 128.5, 127.9, 125.9 (${}^{3}J_{C-P} = 8.5$ Hz), 123.7, 116.5 (${}^{3}J_{C-P} = 4.7$ Hz), 65.0, 50.6, 30.3, 21.6, 19.0, 18.4, 13.7. ³¹P **NMR** (162 MHz, CDCl₃) 18.02 (d, J = 11.1 Hz). **HRMS (ESI)** m/z Calcd for [C₃₄H₃₇N₂O₅PS, M + Na]⁺: 639.2053; Found: 639.2058.

Optical Rotation: $[\alpha]^{25}_{D}$ 48.0 (c = 1.0, CHCl₃). 66% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254

nm, $t_{\rm R} = 9.723$ min for minor isomer, $t_{\rm R} = 12.157$ min for major isomer).

7h. (53% yield, Hexane-EtOAc = 5:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 9.11 (s, 1H), 8.21 (d, J = 8.2 Hz, 1H), 7.75 (d, J = 8.4 Hz, 2H), 7.64 (d, J = 8.2 Hz, 2H), 7.40 (d, J = 8.4 Hz, 2H), 7.27–7.22 (m, 4H), 7.21–7.13 (m, 7H), 7.12–7.04 (m,

2H), 6.83 (d, J = 7.3 Hz, 1H), 6.48 (s, 1H), 4.92 (d, J = 14.9 Hz, 1H), 4.86 (d, J = 14.9 Hz, 1H), 4.73 (d, J = 13.8 Hz, 1H), 4.19 (d, J = 13.8 Hz, 1H), 2.34 (s, 3H), 1.64 (s, 3H), 1.35 (s, 9H). ¹³**C NMR** (100 MHz, CDCl₃) δ 215.9, 164.8, 164.8, 155.0, 144.0, 140.0, 139.8, 138.9, 138.5, 136.2, 131.4, 129.7, 129.2, 128.3, 128.2, 127.7, 127.6, 127.3, 127.2, 127.0, 126.7, 126.5, 125.3, 121.0, 95.6, 79.9, 77.6, 49.1, 34.9, 31.1, 21.5, 18.1. **HRMS (ESI)** m/z Calcd for [C₄₃H₄₂IN₂O₅S, M+Na]⁺: 721.2707; Found: 721.2711.

Optical Rotation: $[\alpha]^{25}_{D}$ -7.2 (c = 0.5, CHCl₃). 16% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 16.569$ min for minor isomer, $t_{R} = 8.867$ min for major isomer).

7i. (46% yield, Hexane-EtOAc = 5:1, R_f = 0.5). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 9.12 (s, 1H), 8.19 (d, J = 8.0 Hz, 1H), 7.83
(d, J = 8.3 Hz, 2H), 7.61 (d, J = 8.3 Hz, 2H), 7.57 (d, J = 8.3 Hz, 2H), 7.29 (d, J = 8.0 Hz, 1H), 7.26 - 7.21 (m, 3H), 7.20 - 7.11

(m, 7H), 7.08 – 7.03 (m, 2H), 6.89 (d, J = 7.5 Hz, 1H), 6.47 (s, 1H), 4.97 (d, J = 14.9 Hz, 1H), 4.91 (d, J = 14.9 Hz, 1H), 4.84 (d, J = 13.6 Hz, 1H), 4.10 (d, J = 13.6 Hz, 1H), 2.32 (s, 3H), 1.71 (s, 3H). ¹³**C NMR** (100 MHz, CDCl₃) δ 216.1, 165.0, 163.5, 144.2, 139.9, 139.7, 138.9, 138.2, 137.4, 136.2, 132.9 (${}^{2}J_{C-F} = 32.3$ Hz), 129.7, 129.3, 128.4, 128.3, 127.9, 127.8, 127.7, 127.3, 127.2, 126.9, 126.2, 125.3 (${}^{3}J_{C-F} = 3.7$ Hz), 123.7 (${}^{1}J_{C-F} = 270.9$ Hz), 120.9, 95.4, 79.7, 77.7, 49.2, 21.5, 18.1. ¹⁹**F NMR** (376 MHz, CDCl₃) 62.83. **HRMS (ESI)** m/z Calcd for [C₄₀H₃₃F₃N₂O₅S, M + Na]⁺: 733.1954; Found: 733.1959.

Optical Rotation: $[\alpha]^{25}_{D}$ -6.6 (c = 0.5, CHCl₃). 42% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 12.534$ min for minor isomer, $t_{R} = 21.435$ min for major isomer).

Representative procedure for synthesis of 8:

To a Schlenk tube containing *rac*-8 (0.1 mmol), β -ICD (1.0 mg, 3 mol%) and K₂CO₃ (0.035 mmol, 0.35 equiv.) were added chlorobenzene (8 mL) and MBH acetate 4 (0.07 mmol, 0.7 equiv.). The reaction mixture was allowed to stir at 0 °C for 72 h. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography to afford the product 9 and unreacted starting material 8.

9a. (56% yield, Hexane-EtOAc = 5:1, R_f = 0.4). Syrup. ¹H
NMR (400 MHz, DMSO-d₆) δ 9.32 (s, 1H), 7.43 (d, J = 8.1
Hz, 2H), 7.31 (d, J = 8.1 Hz, 2H), 6.74 (s, 1H), 6.70 (s, 1H),
5.96 (s, 1H), 5.35 (s, 1H), 4.18 (d, J = 16.7 Hz, 1H), 3.96 (t, J
= 6.6 Hz, 2H), 3.89 (d, J = 16.7 Hz, 1H), 2.39 (s, 3H), 2.36 (s,

3H), 2.19 (s, 3H), 1.96 (s, 3H), 1.78 (s, 3H), 1.55 – 1.42 (m, 2H), 1.32 – 1.23 (m, 2H), 0.86 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 165.5, 156.1, 153.3, 143.5, 137.5, 137.3, 137.0, 136.2, 135.0, 134.3, 131.1, 130.4, 129.4, 129.4, 128.2, 127.8, 124.0, 116.8, 115.2, 64.1, 59.6, 50.3, 30.0, 23.7, 21.3, 21.0, 18.6, 15.7, 13.6, 13.1. HRMS (ESI) m/z Calcd for [C₃₂H₃₈BrNO₆S, M + Na]⁺: 666.1495, 668.1480; Found: 666.1497, 668.1481.

Optical Rotation: $[\alpha]^{25}_{D}$ 90.0 (c = 1.0, CHCl₃). 80% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 10.349$ min for minor isomer, $t_{R} = 5.710$ min for major isomer).

8a. (38% yield, Hexane-EtOAc = 2:1, R_f = 0.4). Syrup. ¹H NMR
(400 MHz, CDCl₃) δ 7.55 (d, J = 8.0 Hz, 2H), 7.41 (s, 1H), 7.22
(d, J = 8.0 Hz, 2H), 6.76 (s, 1H), 6.08 (s, 1H), 4.41 (s, 1H), 3.67
(s, 3H), 2.43 (s, 3H), 2.40 (s, 3H), 2.31 (s, 3H), 1.80 (s, 3H), 1.70
(s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 154.3, 151.6, 144.0,

140.3, 137.7, 136.3, 132.7, 131.8, 131.1, 129.7, 127.1, 124.1, 120.4, 120.0, 119.4, 115.7, 60.0, 24.2, 21.5, 20.4, 16.5, 13.1. **HRMS (ESI)** m/z Calcd for [C₂₄H₂₆BrNO₄S, M + Na]⁺: 526.0658, 528.0640; Found: 526.0661, 528.0644.

Optical Rotation: $[\alpha]^{25}_{D}$ -34.0 (c = 1.0, CHCl₃). 96% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 4.978 min for minor isomer, t_{R} = 8.937 min for major isomer).

9b. (58% yield, Hexane-EtOAc = 5:1, R_f = 0.5) Syrup. ¹H
NMR (400 MHz, DMSO-d₆) δ 9.75 (s, 1H), 7.84 (d, J = 8.9
Hz, 1H), 7.79 (t, J = 9.4 Hz, 1H), 7.38 (d, J = 8.9 Hz, 1H),
7.25 (d, J = 8.9 Hz, 2H), 7.20 - 6.97 (m, 4H), 6.88 (s, 1H),

5.84 (s, 1H), 5.13 (s, 1H), 4.14– 4.02 (m, 1H), 3.95 (t, J = 6.5 Hz, 2H), 3.83 (d, J = 16.7 Hz, 1H), 3.71 (s, 3H), 2.31 (s, 3H), 2.28 (s, 3H), 1.71 (s, 3H), 1.52 – 1.43 (m, 2H), 1.29– 1.22 (m, 2H), 0.86 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 165.6, 156.1, 151.9, 135.7, 135.2, 135.2, 135.0, 133.3, 131.8, 131.7, 129.6, 129.4, 128.9, 127.9, 127.7, 127.7, 127.6, 126.1, 126.1, 124.1, 122.7, 118.3, 117.1, 64.1, 59.5,

54.9, 49.6, 30.0, 18.6, 15.9, 13.6, 13.0. **HRMS (ESI)** m/z Calcd for [C₃₄H₃₆BrNO₆S, M + Na]⁺: 688.1339, 690.1324; Found: 688.1337, 690.1324.

Optical Rotation: $[\alpha]^{25}_D$ 48 (c = 1.0, CHCl₃). 80% *ee* (HPLC conditions: Chiralpak IE column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_R = 16.202$ min for minor isomer, $t_R = 19.509$ min for major isomer).

8b. (32% yield, Hexane-EtOAc = 3:1, R_f = 0.3). Syrup. ¹H
NMR (400 MHz, CDCl₃) δ 7.82 (d, J = 8.9 Hz, 1H), 7.68 (d, J = 8.7 Hz, 1H), 7.49 (s, 1H), 7.41 (dd, J = 8.7, 1.9 Hz, 1H), 7.35 (d, J = 8.3 Hz, 2H), 7.23 (d, J = 8.9 Hz, 1H), 7.09 (d, J = 8.1 Hz, 2H), 6.87 (d, J = 1.6 Hz, 1H), 6.07 (s, 1H), 4.98 (s, 1H),

3.72 (s, 3H), 2.38 (s, 3H), 2.36 (s, 3H), 1.74 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 154.7, 151.8, 143.9, 135.8, 133.8, 133.3, 132.6, 131.7, 130.9, 130.0, 129.6, 127.6, 127.3, 127.0, 125.4, 125.4, 122.0, 122.0, 121.3, 118.1, 113.3, 60.1, 21.7, 16.6, 13.3. **HRMS (ESI)** m/z Calcd for [C₂₆H₂₄BrNO₄S, M + Na]⁺: 548.0502, 550.0484; Found: 548.0504, 550.0479.

Optical Rotation: $[\alpha]^{25}_{D}$ -53 (c = 1.0, CHCl₃). 93% *ee* (HPLC conditions: Chiralpak IE column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 5.336$ min for minor isomer, $t_{R} = 8.541$ min for major isomer).

9c. (56% yield, Hexane-EtOAc = 4:1, R_f = 0.5). Syrup. ¹H
NMR (400 MHz, DMSO-d₆) δ 9.67 (br, 1H), 8.08 (d, J = 1.8 Hz, 1H), 7.81 (d, J = 8.9 Hz, 1H), 7.33 (d, J = 8.7 Hz, 1H), 7.26 (d, J = 8.9 Hz, 1H), 7.24 - 6.89 (m, 5H), 6.87 (s, 5H), 5.87 (s

1H), 5.88 (s, 1H), 5.21 (s, 1H), 4.12 (s, 1H), 3.95 (t, J = 6.4 Hz, 2H), 3.84 (d, J = 16.6 Hz, 1H), 3.70 (s, 3H), 2.31 (s, 3H), 2.26 (s, 3H), 1.69 (s, 3H), 1.54 – 1.42 (m, 2H), 1.31 – 1.22 (m, 2H), 0.86 (t, J = 7.4 Hz, 3H). ¹³**C NMR** (100 MHz, DMSO-d₆) δ 165.5, 156.2, 152.4, 143.3, 136.0, 135.3, 134.9, 134.9, 132.0, 131.9, 130.6, 129.8, 129.3, 129.1, 128.7, 128.4, 128.0, 128.0, 127.7, 127.1, 119.3, 117.4, 115.5, 64.1, 59.6, 50.3, 30.0, 21.0, 18.6, 15.8, 13.6, 13.0. **HRMS (ESI)** m/z Calcd for [C₃₄H₃₆BrNO₆S, M + Na]⁺: 688.1339, 690.1324; Found: 688.1338, 690.1321.

Optical Rotation: $[\alpha]^{25}_D$ 68 (c = 1.0, CHCl₃). 77% *ee* (HPLC conditions: Chiralpak IE column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_R = 13.233$ min for minor isomer, $t_R = 16.617$ min for major isomer).

8c. (32% yield, Hexane-EtOAc = 3:1, $R_f = 0.3$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.95 (d, J = 1.8 Hz, 1H), 7.75 (d, J = 8.955/150 Hz, 1H), 7.54 (s, 1H), 7.32 (d, J = 8.2 Hz, 2H), 7.23 (d, J = 8.9 Hz, 1H), 7.13 (dd, J = 8.9, 1.8 Hz, 1H), 7.05 (d, J = 8.2 Hz, 2H), 6.52 (d, J = 8.9 Hz, 1H), 6.02 (s, 1H), 4.96 (s, 1H), 3.72 (s, 3H), 2.39 (s, 3H), 2.38 (s, 3H), 1.72 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 154.7, 151.2, 144.0, 136.0, 133.2, 132.6, 131.7, 131.0, 130.5, 130.2, 129.9, 129.5, 127.0, 125.0, 122.4, 121.5, 118.8, 117.5, 114.2, 60.1, 21.6, 16.6, 13.2. HRMS (ESI) m/z Calcd for [C₂₆H₂₄BrNO₄S, M + Na]⁺: 548.0502, 550.0484; Found: 548.0505, 550.0489.

Optical Rotation: $[\alpha]^{25}_{D}$ -82 (c = 1.0, CHCl₃). 94% *ee* (HPLC conditions: Chiralpak IE column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 10.652$ min for minor isomer, $t_{R} = 8.992$ min for major isomer).

9d. (43% yield, Hexane-EtOAc = 5:1, R_f = 0.5). Syrup. ¹H
NMR (400 MHz, DMSO-d₆) δ 9.64 (s, 1H), 8.02 (d, J = 8.6
Hz, 2H), 7.96 - 7.85 (m, 2H), 7.52 (t, J = 7.5 Hz, 1H), 7.43
(d, J = 8.8 Hz, 1H), 7.34 - 7.24 (m, 3H), 7.23 - 7.05 (m, 10.5)

5H), 7.04 (d, J = 8.5 Hz, 1H), 6.91 (d, J = 8.2 Hz, 1H), 5.87 (s, 1H), 5.21 (s, 1H), 4.21 (d, J = 16.6 Hz, 1H), 3.96 (t, J = 5.3 Hz, 2H), 3.88 (d, J = 16.6 Hz, 1H), 2.30 (s, 3H), 1.51 – 1.40 (m, 2H), 1.28 – 1.19 (m, 2H), 0.83 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 165.4, 152.7, 143.5, 137.3, 135.9, 135.0, 134.0, 133.9, 133.5, 132.4, 129.7, 129.2, 128.6, 128.1, 127.9, 127.8, 127.7, 126.5, 126.5, 126.2, 126.0, 125.0, 122.7, 118.1, 115.5, 64.1, 49.7, 40.1, 38.9, 30.0, 20.9, 18.6, 13.6. HRMS (ESI) m/z Calcd for [C₃₅H₃₃NO₅S, M + Na]⁺: 602.1972; Found: 602.1969.

Optical Rotation: $[\alpha]^{25}_D$ 70 (c = 1.0, CHCl₃). 71% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_R = 10.883$ min for minor isomer, $t_R = 15.683$ min for major isomer).

8d. (51% yield, Hexane-EtOAc = 4:1, $R_f = 0.5$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 8.13 (d, J = 9.0 Hz, 1H), 8.04 – 7.93 (m, 2H), 7.88 (d, J = 8.2 Hz, 2H), 7.46 (d, J = 8.2 Hz, 2H), 7.41 (t, J = 7.5Hz, 1H), 7.35 (t, J = 7.5 Hz, 1H), 7.30 (d, J = 8.9 Hz, 1H), 7.25 (t,

J = 7.5 Hz, 1H), 7.12 – 7.07 (m, 3H), 7.04 (d, J = 8.5 Hz, 1H), 6.64 (d, J = 8.5 Hz, 1H), 6.51 (s, 1H), 4.69 (s, 1H), 2.34 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 151.8, 144.0, 135.8, 134.5, 132.9, 132.9, 131.5, 131.1, 130.5, 129.6, 129.2, 128.4, 128.2, 127.5, 127.4, 127.1, 1256, 125.2, 123.9, 123.7, 119.1, 118.4, 117.8, 111.9, 21.48. HRMS (ESI) m/z Calcd for [C₂₇H₂₁NO₃S, M+Na]⁺: 462.1134; Found: 462.1135.

Optical Rotation: $[\alpha]^{25}_{D}$ -47 (c = 1.0, CHCl₃). 79% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 9.539$ min for minor isomer, $t_{R} = 8.684$ min for major isomer).

9e. (60% yield, Hexane-EtOAc = 3:1, $R_f = 0.6$). Syrup. 1H NMR (400 MHz, DMSO) δ 9.55 (s, 1H), 7.81 – 7.71 (m, 2H), 7.25 – 7.13 (m, 3H), 7.06 (s, 1H), 6.97 (d, J =13.3 Hz, 1H), 6.03 (s, 1H), 5.56 (s, 1H), 4.18 – 3.75 (m, 4H), 3.61 (s, 3H), 2.54 – 2.29 (m, 3H), 2.25 (s, 3H), 1.61

(s, 3H), 1.49 - 1.36 (m, 2H), 1.26 - 1.17 (m, 2H), 0.80 (t, J = 7.4 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 165.6, 156.1, 151.9, 135.7, 135.2, 135.1, 133.3, 131.8, 131.7, 129.7, 129.4, 127.9, 127.7, 126.1, 124.2, 124.2, 122.7, 118.3, 117.1, 64.1, 59.5, 49.6, 39.9, 30.0, 18.6, 15.9, 13.6, 13.0. **HRMS (ESI)** m/z Calcd for [C₂₈H₃₃NO₆S, M + Na]⁺: 534.1921; Found: 534.1921.

Optical Rotation: $[\alpha]^{25}_D$ 26.0 (c = 1.0, CHCl₃). 54% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_R = 10.389$ min for minor isomer, $t_R = 9.564$ min for major isomer).

8e. (26% yield, Hexane-EtOAc = 5:1, $R_f = 0.2$). Syrup. ¹H NMR (400 MHz, CDCl₃) δ 7.87 – 7.83 (m, 2H), 7.48 (s, 1H), 7.40 – 7.31 (m, 2H), 7.24 (s, 1H), 7.13 – 7.04 (m, 1H), 5.77 (s, 1H), 5.22 (s, 1H), 3.75 (s, 3H), 2.69 (s, 3H), 2.40 (s, 3H), 1.86 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 155.0, 151.0, 133.23, 133.0, 132.4, 132.0, 131.1,

129.4, 128.7, 127.6, 124.2, 123.6, 123.1, 121.5, 117.7, 114.1, 60.1, 39.6, 16.5, 13.3. **HRMS (ESI)** m/z Calcd for [C₂₀H₂₁NO₄S, M + Na]⁺: 394.1083; Found: 394.1086. **Optical Rotation**: $[\alpha]^{25}_{D}$ -30.0 (c = 1.0, CHCl₃). 94% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_{R} = 9.005 min for minor isomer, t_{R} = 11.326 min for major isomer).

9f. (60% yield, Hexane-EtOAc = 5:1, R_f = 0.4) Syrup.
¹H NMR (400 MHz, DMSO-d₆) δ 10.11 (s, 1H), 8.55 (d, J = 1.4 Hz, 1H), 8.07 (d, J = 8.9 Hz, 1H), 7.70 (dd, J = 8.9, 1.6 Hz, 1H), 7.37 (d, J = 8.9 Hz, 1H), 7.15 (s, 1H),

7.09 (d, J = 7.0 Hz, 1H), 6.02 (s, 1H), 5.57 (s, 1H), 4.14 – 3.98 (m, 4H), 3.87 (s, 3H), 3.70 (s, 3H), 2.50 (s, 3H), 2.32 (s, 3H), 1.68 (s, 3H), 1.51 – 1.40 (m, 2H), 1.29 – 1.23 (m, 2H), 0.86 (t, J = 7.3 Hz, 3H). ¹³**C NMR** (100 MHz, DMSO-d₆) δ 166.5, 165.5, 156.1, 154.4, 135.8, 135.5, 135.3, 134.5, 131.7, 131.5, 131.2, 130.8, 130.1, 127.9, 127.8,126.8, 125.0, 123.7, 119.3, 117.4, 64.1, 59.6, 52.0, 49.8, 39.7, 30.0, 18.6, 15.9, 13.5, 13.0. **HRMS (ESI)** m/z Calcd for [C₃₀H₃₅NO₈S, M + Na]⁺: 592.1976; Found: 592.1978.

Optical Rotation: $[\alpha]^{25}_{D}$ 10.3 (c = 1.0, CHCl₃). 53% *ee* (HPLC conditions: Chiralpak IE column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 18.985$ min for minor isomer, $t_{R} = 22.803$ min for major isomer).

8f. (33% yield, Hexane-EtOAc = 2:1, R_f = 0.4) Syrup. ¹H
NMR (400 MHz, CDCl₃) δ 8.61 (d, J = 1.5 Hz, 1H), 8.00 (d, J = 8.8 Hz, 1H), 7.94 (dd, J = 8.8, 1.5 Hz, 1H), 7.49 (s, 1H), 7.35 (d, J = 8.8 Hz, 1H), 7.14 (d, J = 8.8 Hz, 1H), 5.75 (s, 1H), 5.53 (brs, 1H), 3.96 (s, 3H), 3.77 (s, 3H), 2.74 (s,

3H), 2.42 (s, 3H), 1.86 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 167.0, 155.1, 153.2, 135.0, 133.7, 133.0, 132.6, 132.0, 131.8, 128.4, 127.1, 125.8, 123.4, 122.8, 121.5, 118.8, 114.4, 60.2, 52.3, 39.8, 16.6, 13.3. HRMS (ESI) m/z Calcd for [C₂₂H₂₃NO₆S, M + Na]⁺: 452.1138; Found: 452.1139.

Optical Rotation: $[\alpha]^{25}{}_{D}$ -36.0 (c = 1.0, CHCl₃). 97% *ee* (HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wavelength = 254 nm, $t_{R} = 9.452$ min for minor isomer, $t_{R} = 7.949$ min for major isomer).

IV. Racemization experiment

$$\Delta G^{\dagger} = \frac{RT ln \ (\frac{K_B \cdot T}{hK_{enantiomerization}})}{K_{enantiomerization}}, K_{enantiomerization} = \frac{1}{2} K_{rac}$$

 K_B is the Boltzmann, $K_B = 1.38066E - 23$ J·K⁻¹. T is temperature. *h* is the Plance constant. h = 6.62608E-34 Js. *R* is the specific gas constant, R = 8.31451 J·K⁻¹mol⁻¹

Compound (0.1 mmol) was dissolved in mesitylene (1 mL) in a Schlenk tube. The tube was immersed in a pre-heated oil bath at 100-140 °C. At given interval of time, small samples (50 μ L) was removed via syringe and injected into the HPLC to measure the enantiomeric excess.

O O Tol ^S N CO ₂ CH Me Me Me	HPh₂ 140 °C mesitylene	O Tol ^S N Me Br Me Me
t (h)	ee (%)	ln(ee)
0	90	-0.083
1	65	-0.117
2	46	-0.151
3	35	-0.198
3 ^a	90	-0.083

 $^{a}T=110\ ^{o}C$

 $K_{rac} = 9E-5, K_{enantiomerization} = 4.5E-5, \Delta G^{\ddagger} = 32.65 \text{ Kcal/mol}$

 $K_{rac} = 1E-5, K_{enantiomerization} = 5E-6, \Delta G^{\dagger} = 34.02 \text{ Kcal/mol}$

t (h)	ee (%)	In(ee)
0	75	-0.288
1	71	-0.342
2	67	-0.400
3	62	-0.478
4	66	-0.562

 $K_{rac} = 2E-5, K_{enantiomerization} = 1E-5, \Delta G^{\ddagger} = 33.88 \text{ Kcal/mol}$

 $K_{rac} = 3E-5, K_{enantiomerization} = 1.5E-5, \Delta G^{\dagger} = 33.13 \text{ Kcal/mol}$

 $K_{rac} = 9E-5$, $K_{enantiomerization} = 4.5E-5$, $\Delta G^{\ddagger} = 32.65$ Kcal/mol

 $K_{rac} = 3E-5, K_{enantiomerization} = 1.5E-5, \Delta G^{\ddagger} = 31.05 \text{ Kcal/mol}$

t (h)	ee (%)	ln(ee)
0	78	-0.248
0.5	70	-0.357
1	64	-0.446
1.5	59	-0.528
2	50	-0.693
2.5	46	-0.777

 $K_{rac} = 6E-5, K_{enantiomerization} = 3E-5, \Delta G^{\ddagger} = 29.83 \text{ Kcal/mol}$

 $K_{rac} = 5E-5, K_{enantiomerization} = 2.5E-5, \Delta G^{\ddagger} = 29.85 \text{ Kcal/mol}$

t (h)	ee (%)	ln(ee)
0	66	-0.416
0.5	64	-0.446
1	62	-0.478
1.5	60	-0.511
2	57	-0.534

 $K_{rac} = 2E-5, K_{enantiomerization} = 1E-5, \Delta G^{\ddagger} = 31.36 \text{ Kcal/mol}$

V. Gram scale reaction

To a three-necked flask containing **1p** (650.3 mg, 2 mmol), β -ICD (60 mg, 0.2 mmol, 10 mol%) and Cs₂CO₃ (651.6 mg, 2 mmol, 1.0 equiv.) were added mesitylene (120 mL) and dienoate **2g** (773.7 mg, 2.4 mmol, 1.2 equiv.). The reaction mixture was stirred at -50 °C for 24 hours. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography (Hexane-EtOAc = 5:1) to afford the product **3v** (822 mg) in 70% yield with 97% *ee*.

To a three-necked flask containing **1a** (802.5 mg, 2 mmol), β -ICD (6 mg, 0.02 mmol, 1 mol%) and Cs₂CO₃ (651.6 mg, 2 mmol, 1.0 equiv.) were added mesitylene (20 mL) and MBH acetate **4c** (440.5 mg, 2.2 mmol, 1.1 equiv.). The reaction mixture was stirred at -30 °C for 72 hours. The solvent was removed by silica gel column chromatography and the residue was then purified by silica gel column chromatography (Hexane-EtOAc = 15:1) to afford the product **5c** (953 mg) in 88% yield with 91% *ee*.

VI. Further transformation

A solution of **3w** or **5c** (26.6 mg, 0.123 mmol, 15 mol%) and *m*-CPBA (85%, 37.3 mg, 0.218 mmol, 1.5 equiv.) in CH₂Cl₂ was stirred at 0 °C for 5 min, then 3-(1-hydroxy-2-naphthyl)propionic acid **SI-1** was added. After 36 h, the resulting mixture poured into aqueous Na₂S₂O₃ (5 mL) and aqueous NaHCO₃, and extracted with CH₂Cl₂. The organic layers were dried over anhydrous Na₂SO₄ and solvents were removed in vacuo. The residue was purified by column chromatography on silica gel (eluent: Hexane-EtOAc-CH₂Cl₂ = 8:1:1) to give **SI-2**.¹H NMR (CDCl₃, 400 MHz) 88.00 (d, *J* = 7.7 Hz, 1H), 7.62 (td, *J* = 7.5, 1.0 Hz, 1H), 7.40 (t, *J* = 7.5 Hz, 1H), 7.25 (d, *J* = 7.7 Hz, 1H), 6.65 (d, *J* = 9.9 Hz, 1H), 6.20 (d, *J* = 9.9 Hz, 1H), 2.90 (ddd, *J* = 9.7, 11.2, 17.6 Hz, 1H), 2.59 (ddd, *J* = 2.0, 9.6, 17.6 Hz, 1H), 2.41 (ddd, *J* = 2.0, 9.6, 13.2Hz, 1H), 2.18 (ddd, *J* = 9.8, 11.2, 13.2 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 196.5, 176.5, 136.8, 135.7, 132.3, 128.9, 127.9, 127.9, 127.7, 127.3, 83.4, 31.2, 26.5; **MS** (EI) m/z 214.06 (M⁺); HPLC condition: Chiralpak OD-H column, *n*-Hexane/*i*-PrOH = 85:15, flow rate = 1.0 mL/min, wavelength = 254 nm, *t*_R = 17.6 min for minor isomer, *t*_R = 21.1 min for major isomer).


Through a solution containing **3v** (108 mg, 0.2 mmol) in 2 mL of CH₂Cl₂ was bubbled an ozone-oxygen stream at -78 °C until the starting material was consumed. The resulting mixture poured into aqueous Na₂O₃S₂ (5 mL) and extracted with CH₂Cl₂. The organic layer is dried on anhydrous Na₂SO₄ and the solvent is removed in vacuum to obtain the product **10**. ¹H NMR (400 MHz, CDCl₃) δ 7.57 (s, 1H), 7.43 – 7.31 (m, 10H), 7.06 (s, 1H), 6.96 (s, 1H), 5.05 (d, *J* = 19.5 Hz, 1H), 4.91 (d, *J* = 19.5 Hz, 1H), 3.36 (s, 3H), 2.53 (s, 3H), 2.25 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 188.0, 159.1, 141.6, 141.0, 139.4, 139.3, 138.8, 133.1, 129.0, 128.8, 128.8, 127.6, 127.4, 101.2, 79.9, 58.9, 44.0, 20.6. HRMS (ESI) m/z Calcd for [C₂₅H₂₄INO₅S, M + Na]⁺: 600.0310; Found: 600.0311. [α]²⁵_D -13.3 (c = 1.0, CHCl₃). 97% *ee.* HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wavelength = 254 nm, t_R = 14.869 min for minor isomer, t_R = 18.558 min for major isomer).





A solution of **5c** (108 mg, 0.2 mmol) and DIBAL-H (0.4 mL, 0.6 mmol, 3 equiv.) in CH₂Cl₂ (0.6 mL) was stirred at -78 °C for 8 h. the resulting mixture poured into aqueous NH₄Cl (5 mL) and extracted with CH₂Cl₂. The organic layers were dried over anhydrous Na₂SO₄ and solvents were removed in vacuo. The residue was purified by column chromatography on silica gel (Hexane-EtOAc = 4:1) to give **11**. ¹H NMR (400 MHz, CDCl₃) δ 7.69 (d, *J* = 8.2 Hz, 2H), 7.51 (s, 1H), 7.28 (d, *J* = 8.1 Hz, 2H), 7.01 (s, 1H), 5.06 (s, 1H), 4.67 (s, 1H), 4.43 (d, *J* = 14.0 Hz, 1H), 4.29 (dd, *J* = 14.6, 2.7 Hz, 2H), 4.16 (d, *J* = 14.6 Hz, 1H), 2.50 (s, 1H), 2.42 (s, 3H), 2.23 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 143.7, 143.4, 142.2, 140.0, 139.3, 138.1, 136.9, 132.5, 129.6, 128.2, 117.6, 100.1, 64.3, 52.4, 21.5, 20.4, 20.3. HRMS (ESI) m/z Calcd for [C₁₉H₂₂INO₃S, M + Na]⁺: 494.0257; Found: 494.0258. [α]²⁵_D -22.5 (c = 1.0, CHCl₃). 86% *ee*. HPLC conditions: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 90:10, flow rate = 1.0 mL/min, wavelength = 254 nm, t_R = 16.564 min for minor isomer, t_R = 19.170 min for major isomer).



VII. Geometries of the enantiomeric transition states

The atroposelective model are depicted as follows: the deprotonated **1g** binds to catalyst β -ICD by hydrogen-bonding interaction between the S=O oxygen of **1g** and the OH of β -ICD; subsequently, the product **3g** was readily afforded by nucleophilic attack under the chiral environment; the orientation of the electronic abundant iodine atom in the favored TS-3g-major shows more favorable noncovalent interactions with hydrogens; hence placing the *ortho*-iodine inside gives rise to the favored transition state TS-3g-major, while placing the *ortho*-methyl inside results in the less favored TS-3g-minor.



TS-3g-major

TS-3g-minor

VIII. References

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IX. X-Ray crystallography analysis of compound 3v (CCDC-2078431)



Datablock: ga_210406a_a

Bond precision:	C-C = 0.0065	A	Waveleng	th=1.34138	
Cell:	a=9.5319(7) alpha=90	b=15.1834 beta=90	(11)	c=18.2181(14) gamma=90	
Temperature:	180 K				
	Calculated		Reporte	d	
Volume	2636.6(3)		2636.6(3)	
Space group	P 21 21 21		P 21 21	21	
Hall group	P 2ac 2ab		P 2ac 2	ab	
Moiety formula	C27 H26 I N 04	S	?		
Sum formula	C27 H26 I N 04	S	C27 H26	IN 04 S	
Mr	587.45		587.45		
Dx,g cm-3	1.480		1.480		
Z	4		4		
Mu (mm-1)	7.205		7.205		
F000	1184.0		1184.0		
F000'	1187.51				
h,k,lmax	11,19,22		11,19,2	2	
Nref	5475[3091]		5459		
Tmin, Tmax	0.374,0.487 0.430,0.			.752	
Thin'	0.178				
Correction meth	od- # Reported	T Limits: T	min=0.43	0 Tmax=0.752	
AbsCorr - MULTI	-SCAN				
Data completene	ss= 1.77/1.00	Theta (m	ax) = 57.	467	
R(reflections) =	0.0295(5355)	wR2(ref	lections) = 0.0723(5459	9)
S = 1.055	Npai	r= 318			

The following ALERTS were generated. Each ALERT has the format test-mame_ALERT_alert-type_elert-level. Click on the hyperlinks for more details of the test.

	A AVENUE A Makin of Maximum / Minimum Manidus] Paraita		Harmont			
PLATU	74 ALERT 2 C Racio of Maximum / Minimum Residual Density	2.94	neport			
PLAT	45 ALERT 2 C U(180) HIH Smaller than U(eq) Cl by	0.016	Ang			
PLATE	11 ALERT 3 C Missing PCF Refl Between Thmin & STh/L= 0.600	2	Report			
PLATE	AT ALERT 1 C The Fisch x 18 55 0 - Do a BASF/TWIN Refinement	PIGENC	Check			
	ert level G					
AIISM	01 ALERT 1 G Calculation of exptl absorpt correction mu					
	not performed for this radiation type.					
PLATO	02 ALERT 2 G Number of Distance or Angle Restraints on AtSite	3	Note			
PLATO	33 ALERT 4 G Plack x Value Deviates > 3.0 * signs from Hero .	0.066	Note			
PLATI	64 ALERT 4 G Nr. of Mefined C-H H-Atoms in Heavy-Atom Struct.	2	Note			
PLATI	72 ALERT 4 G The CIF-Rebedded .res File Contains DFIX Records	1	Report			
PLAT	12 ALERT 2 G Short Inter XY Contact 04C5	2.94	Ang.			
	-1/2+x, 3/2-y, 1-z =	4 466 Ches	:k			
PLATE	60 ALENT 3 G Number of Least-Squares Restraints	- 2	Note			
PLATE	#3 ALERT 1 G No Info/Value for atom sites solution primary .	Plassa	Do I			
PLATE	10 ALERT 3 G Missing # of FCF Reflection(s) Below Thets(Min).	1	Note			
PLATE	23 ALERT 2 G Number of OMIT Records in Embedded .res File	1	Note			
PLATE	78 ALERT 2 G Number C-C Bonds with Positive Residual Density.	2	Info			
0	ALERT level A = Most likely a serious problem - resolve or exp	lain				
0	ALERT level B = A potentially serious problem, consider carefu	illy				
4	ALERT level C = Check. Ensure it is not caused by an omission	or oversig	5/2			
11	ALXRT level G : General information/check it is not something	unexpected				
а	ALERT type 1 CIF construction/syntax error, inconsistent or mi	ssing data				
6	6 ALENT type 2 Indicator that the structure model may be wrong or deficient					
1	ALERT type 3 Indicator that the structure quality may be low					
	ALERT type 4 Improvement, methodology, query or suggestion					
3						

All these data can be obtained free of charge from Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/ data_request/ci.

X. NMR Spectra

Compound 3a



Compound 3b



Compound 3c



Compound 3d



Compound 3e



Compound 3f



Compound 3g



Compound 3h



Compound 3i





Compound 3k





90/150

Compound 3m



Compound 3n





Compound 30





Compound 3p





Compound 3q





Compound 3r





Compound 3s





Compound 3t







Compound 3v



Compound 3w



Compound 3x



Compound 5a



104/150

Compound 5b



Compound 5c







Compound 5e


Compound 5f



Compound 5g



Compound 5h



Compound 5i





113/150

Compound 5k







115/150







Compound 5n





117/150

Compound 50





Compound 5p



Compound 5q



Compound 5r





Compound 5s





Compound 5t





Compound 5u





Compound 7a





Compound 7b





Compound 7c











Compound 6f







132/150



Compound 7h





Compound 7i









Compound 8b





Compound 8c





Compound 8d







Compound 8f



0. UETUO -9.32 C7.29 6.73 6.70 -5.96 -5.35 74,15 74,15 74,15 75 3.4E+08 -3.2E+08 Me -3.0E+08 .CO₂ⁿBu MeO -2.8E+08 -2.6E+08 -2.4E+08 N<Ts Me -2.2E+08 OH Me -2.0E+08 -1.8E+08 Br -1.6E+08 Мe -1.4E+08 -1.2E+08 -1.0E+08 -8.0E+07 -6.0E+07 -4.0E+07 -2.0E+07 0.0E+00 1.03 ¥ 1.08-1 1.00H 2.06 ₹ 2.15 Æ 1.04-1 2.09 2.09 1.05 3.06 2.97 2.96 3.00 3.15 3.15 2.38 2.38 3.20 -2. 0E+07 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 fl (ppm) 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0

Compound 9a



142/150

Compound 9b



Compound 9c


Compound 9d



Compound 9e



Compound 9f



Compound SI-2



148/150

Compound 10



Compound 11

