

Supplementary material

Chronoampermetric Enzymatic Glucose Sensor Based on Doped Polyindole /Multi-walled Carbon Nanotube Composites Modified onto Screen-Printed Carbon Electrode as Portable Sensing Device for Diabetes

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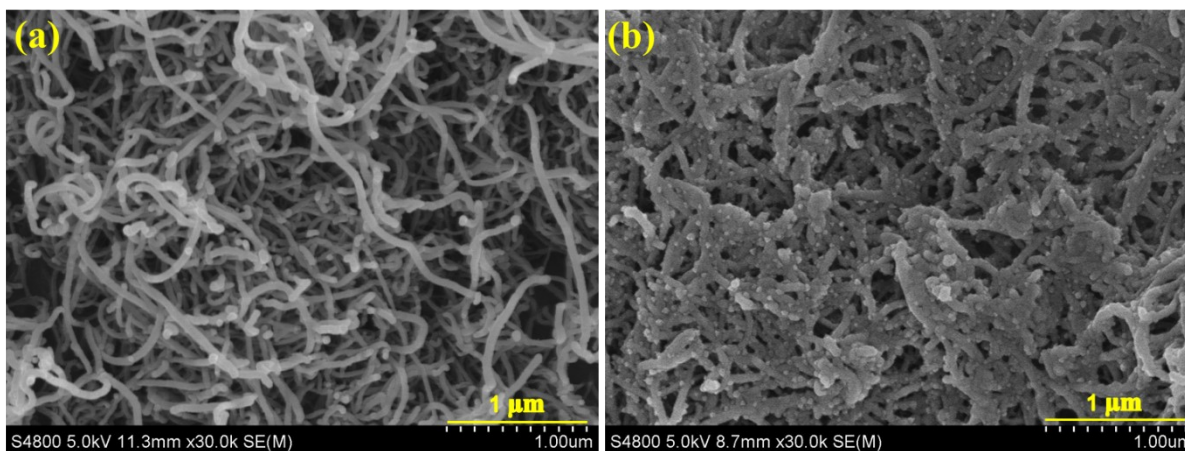


Fig. S1 Surface morphologies at the 30,000x magnification of: (a) MWCNT; and (b) 1.5%MWCNT-dPIIn.

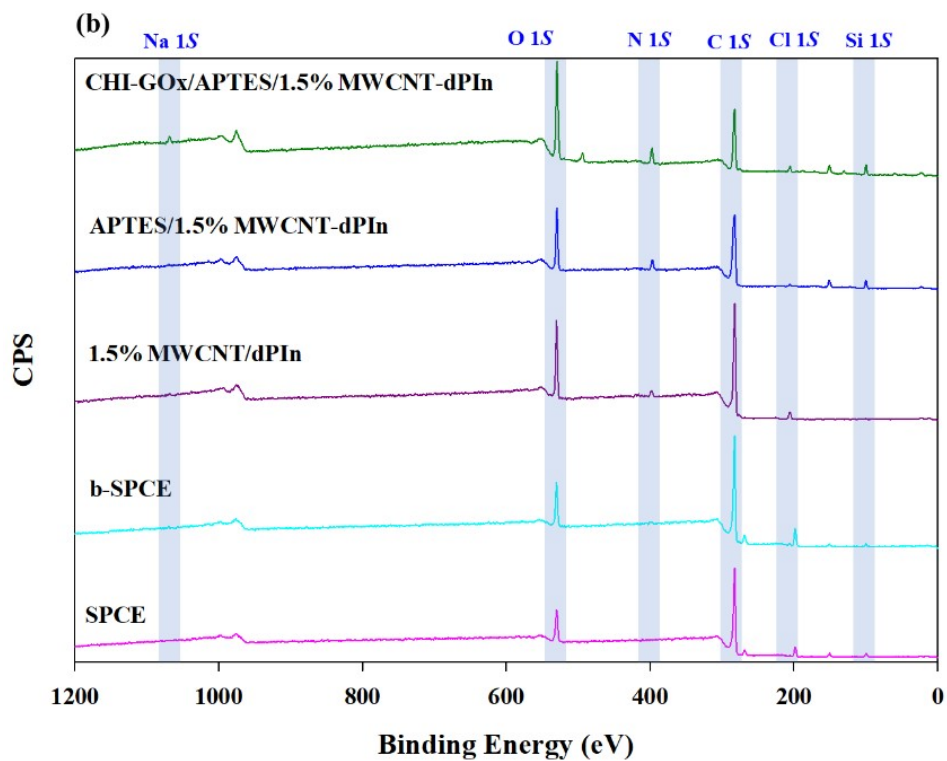
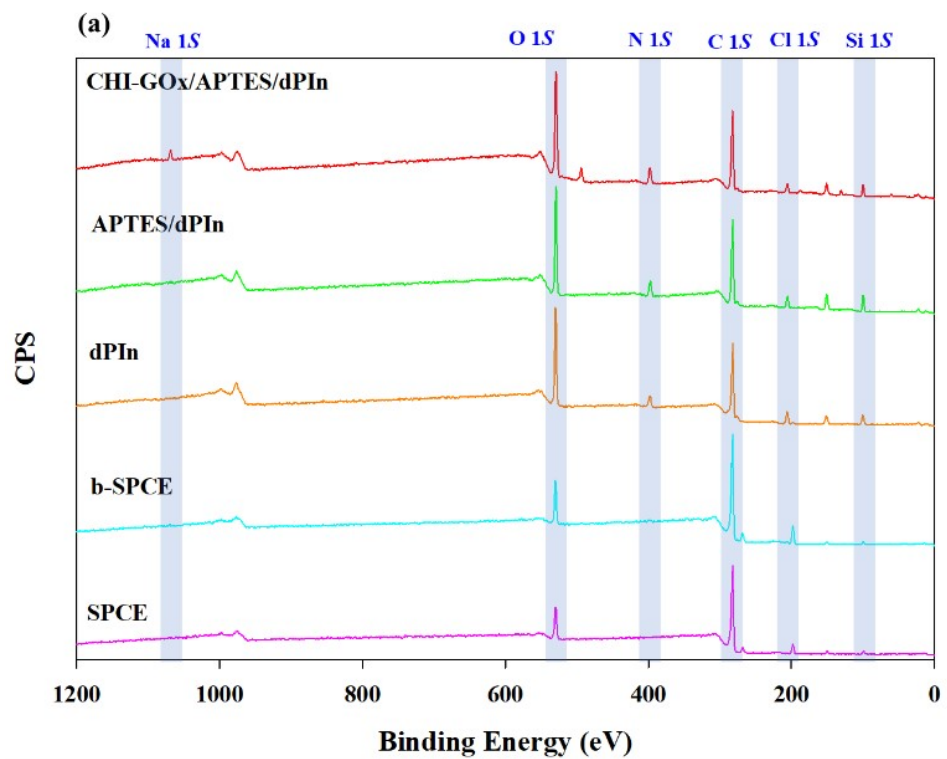
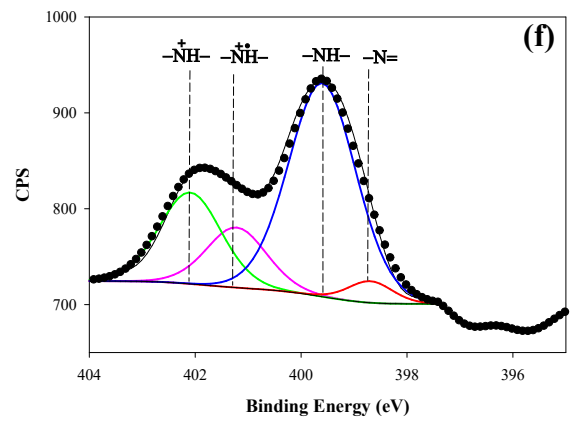
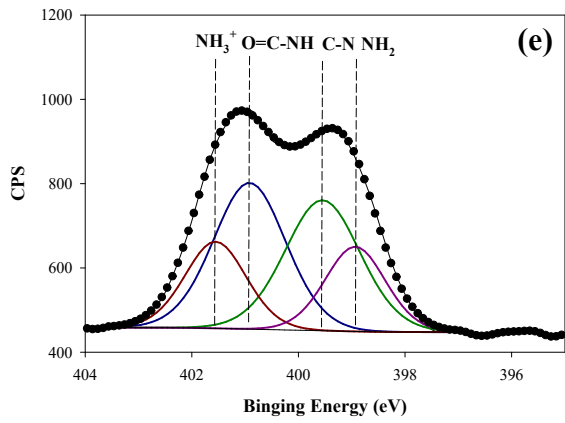
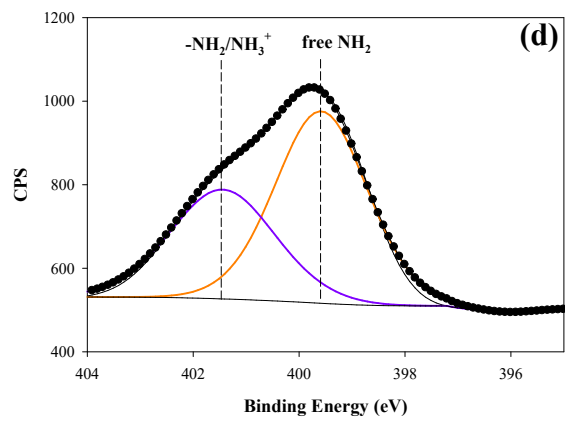
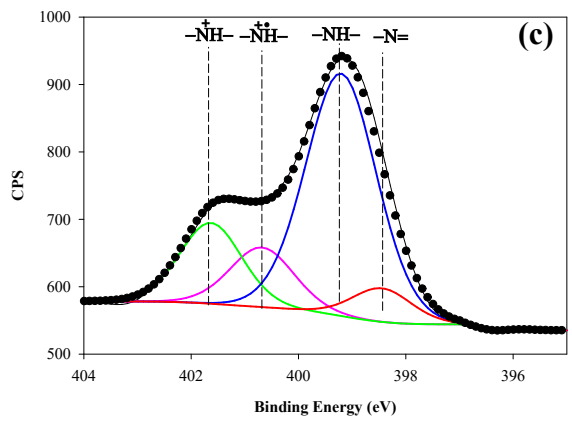
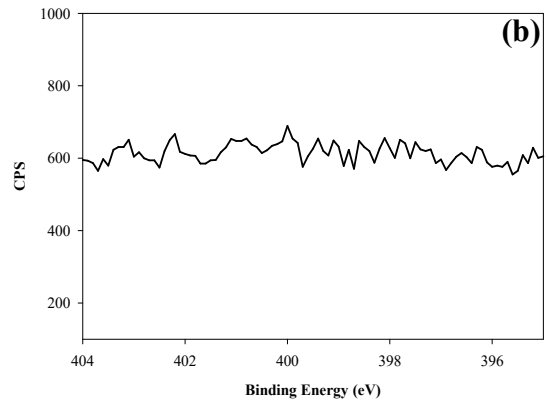
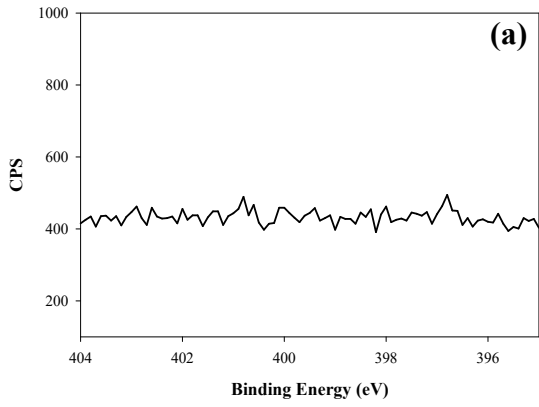


Fig. S2 Wide scan XPS spectra of various modified electrodes.



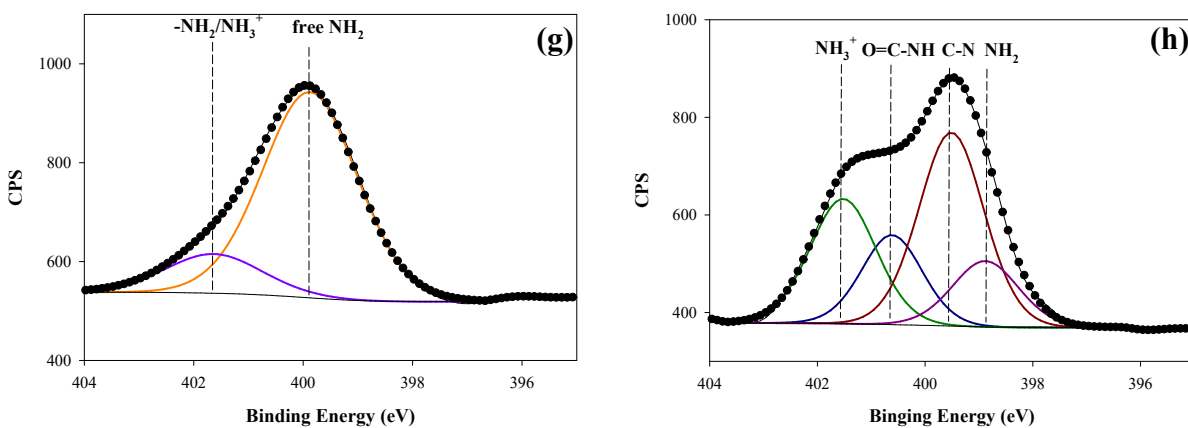


Figure S3 High-resolution spectra of N 1s of (a) SPCE; (b) b-SPCE; (c) dPIn; (d) APTES/dPIn; (e) CHI-GOx/ APTES/dPIn; (f) 1.5% MWCNT-dPIn; (g) APTES/1.5% MWCNT-dPIn; and (h) CHI-GOx/1.5% MWCNT-dPIn.

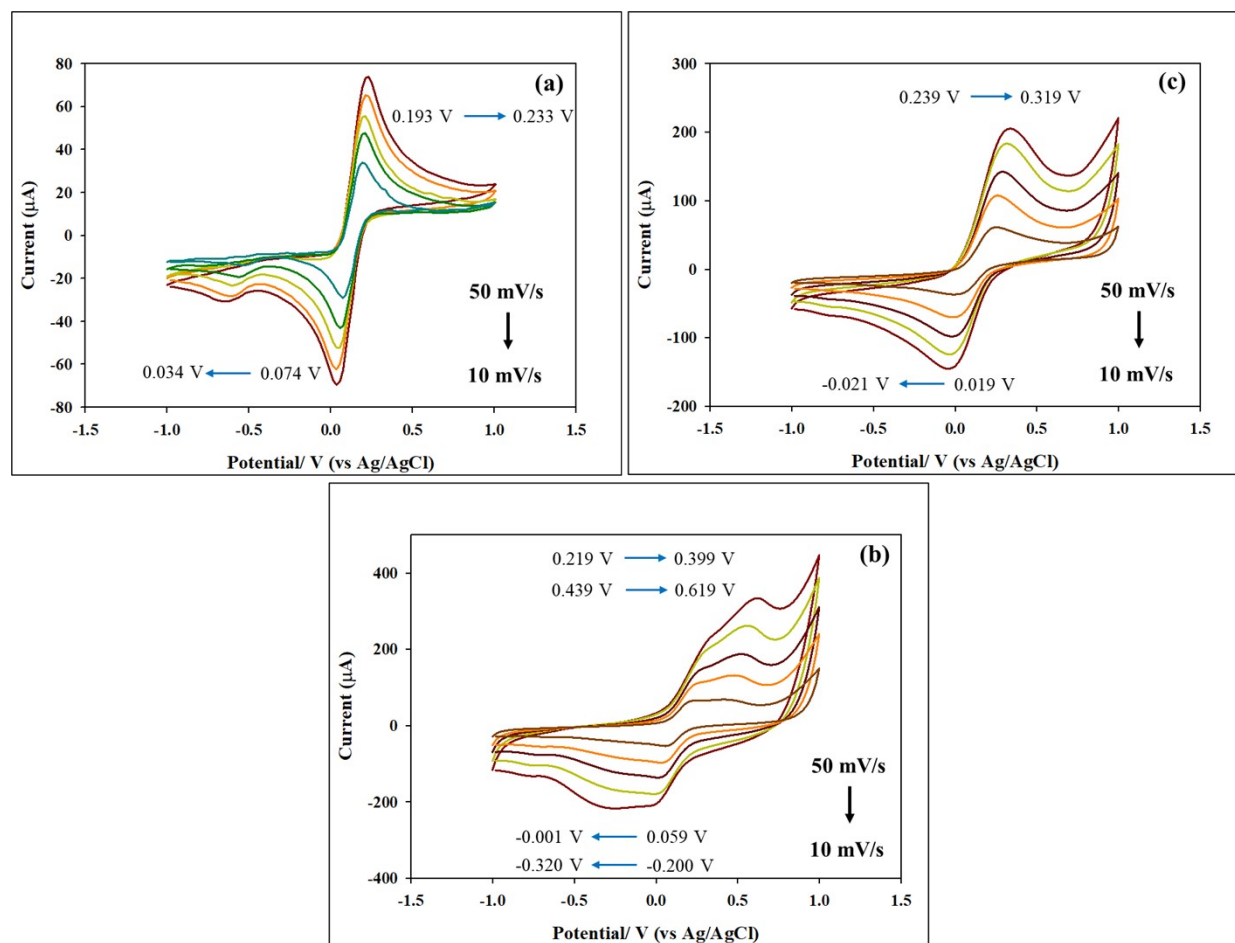


Fig. S4 Cyclic voltammograms of the modified electrodes in a solution of 5 mM of $K_3[Fe(CN)_6]/K_4[Fe(CN)_6]$ containing 0.1 M PBS solution (pH 7.4) and 0.1 M KCl: (a) SPCE; (b) dPIn; and (c) 1.5% MWCNT-dPIn.

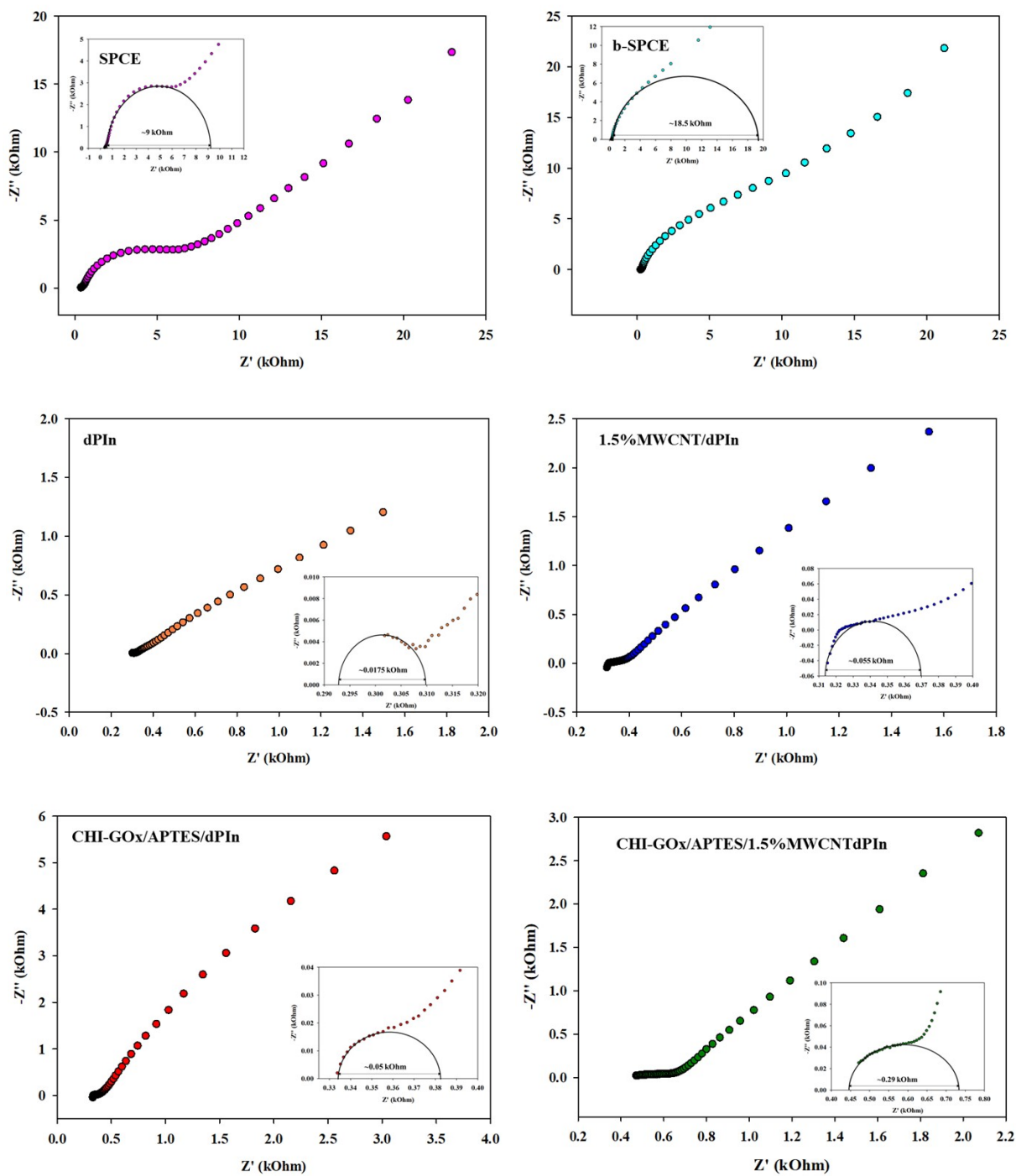


Fig. S5 Nyquist plots of various modified electrodes in a solution of 5 mM of $K_3[Fe(CN)_6]/K_4[Fe(CN)_6]$ containing 0.1 M PBS solution (pH 7.4) and 0.1 M KCl.

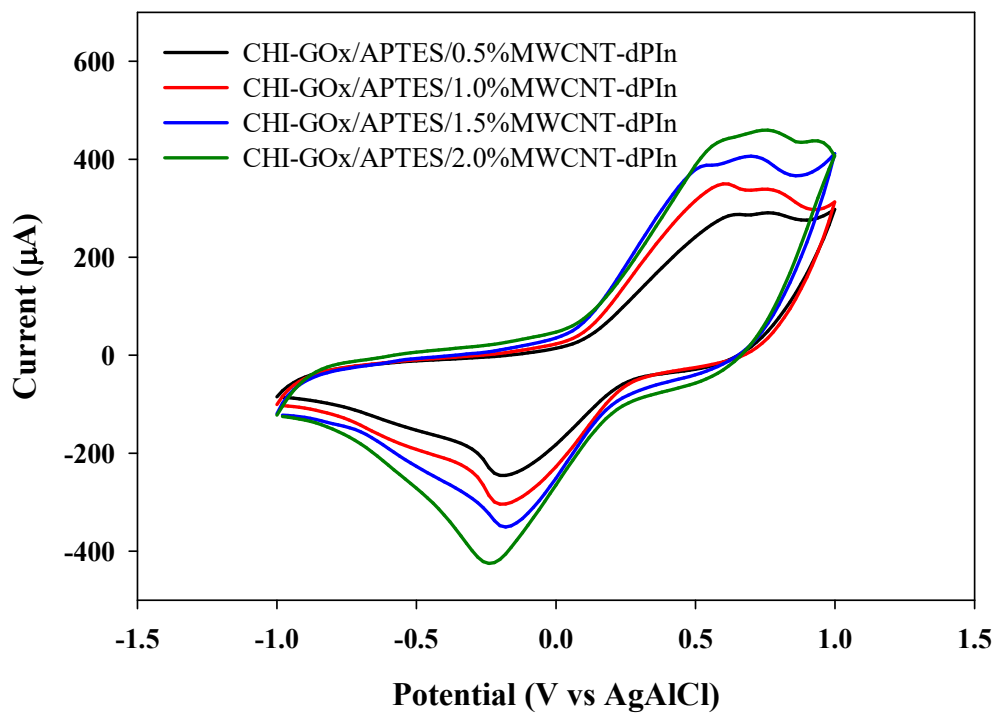


Fig. S6 Cyclic voltammograms of CHI-GOx immobilized on various APTES/MWCNTs-dPIn electrodes in a solution of 5 mM of $K_3[Fe(CN)_6]/K_4[Fe(CN)_6]$ containing 0.1 M PBS solution (pH 7.4) and 0.1 M KCl.



Fig. S7 Experimental setup of the portable enzymatic glucose sensor by chronoamperometric detection in glucose solution by PalmSens4 device connected with the modified electrode and linked to a laptop computer via Bluetooth or USB connector.

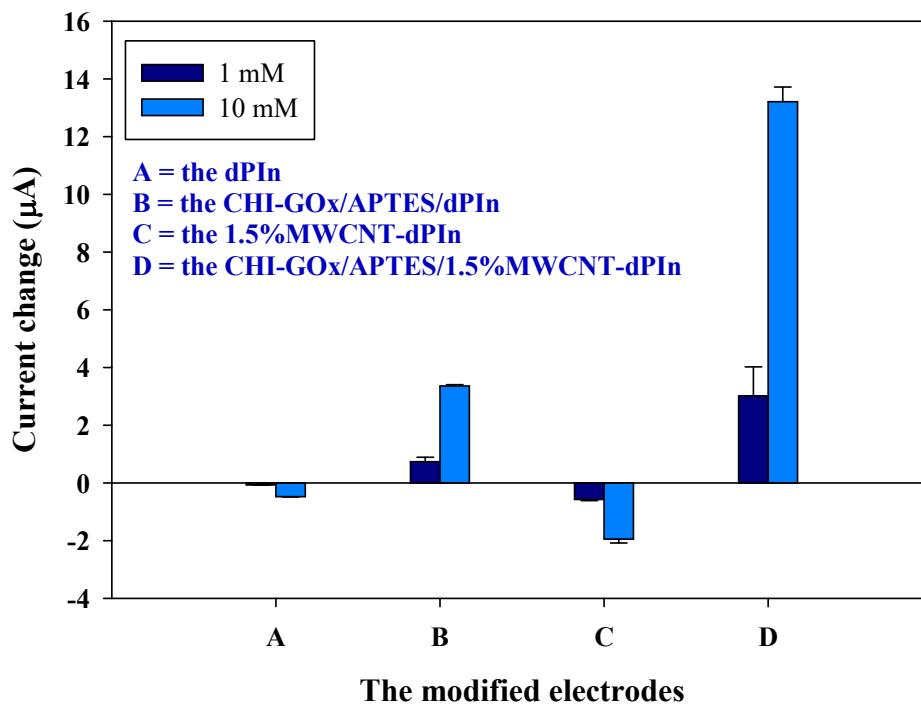


Fig. S8 Current changes ($\Delta I = I_{\text{glc}} - I_{\text{PBS}}$) at +0.6 V vs. Ag/AgCl in 1 mM and 10 mM glucose solutions of various modified electrodes.

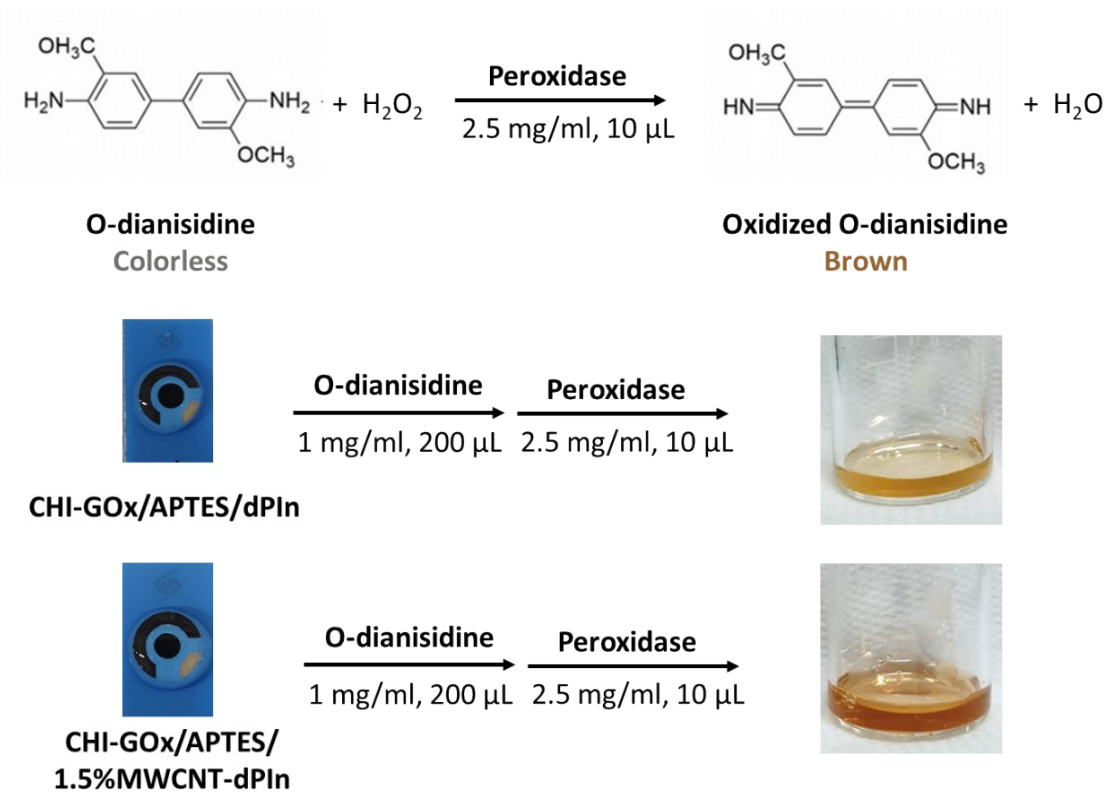


Fig. S9 Determination of H₂O₂ produced in glucose oxidation of the fabricated glucose sensors.

First, 200 μL of 1 mM glucose solution was dropped on the modified electrodes, CHI-GOx/APTES/dPIn and CHI-GOx/APTES/1.5%MWCNT-dPIn, the reaction time was 10 min. Then, the glucose solution reacted with 200 μL of o-dianisidine (1 mg/ml in 1 mL ethanol:49 mL 0.1 M PBS). Finally, 10 μL of the enzyme peroxidase (2.5 mg/ml) in 0.1 M PBS was dropped into the mixture solution. The color of the mixture solution suddenly changed from colorless to brown as shown in **Fig. S6**. This result confirms that H_2O_2 was generated by the interaction of glucose and GOx.

Calculations of sensitivity for CHI-GOx/APTES/dPIn and CHI-GOx/APTES/1.5%MWCNT- dPIn

- 1) The power law equation of the CHI-GOx/APTES/dPIn is $y = 0.8312x^{0.5251}$, $r^2 = 0.9805$, where y is the current change (ΔI) and x is the glucose concentration.

Thus, the power law equation is $\Delta I = 0.8312C^{0.5251}$

$$\Delta I = 0.8312 \cdot C^{0.5251}$$

$$\frac{d(\Delta I)}{dC} = \frac{0.8312 dC^{0.5251}}{dC}$$

$$\frac{d(\Delta I)}{dC} = 0.8312 \cdot 0.5251 C^{(0.5251-1)}$$

$$\text{At LOD, } C = 0.01 \text{ mM, } \frac{d(\Delta I)}{dC} = 3.9 \mu\text{A/mM}$$

The sensitivity was calculated from the slope (3.9 $\mu\text{A/mM}$) divided by the geometric surface area (0.07 cm^2). Thus, the sensitivity of the glucose sensor based on CHI-GOx/APTES/dPIn is 55.7 $\mu\text{A/mM} \cdot \text{cm}^2$.

- 2) The power law equation of the CHI-GOx/APTES/1.5%MWCNT- dPIn is $y = 3.0722x^{0.5667}$, $r^2 = 0.9927$, where y is the current change (ΔI) and x is the glucose concentration.

Thus, the power law equation is $\Delta I = 3.0722C^{0.5667}$

$$\Delta I = 3.0722 \cdot C^{0.5667}$$

$$\frac{d(\Delta I)}{dC} = \frac{3.0722 dC^{0.5667}}{dC}$$

$$\frac{d(\Delta I)}{dC} = 3.0722 \cdot 0.5667 C^{(0.5667-1)}$$

$$\text{At LOD, } C = 0.01 \text{ mM, } \frac{d(\Delta I)}{dC} = 12.8 \text{ } \mu\text{A/mM}$$

The sensitivity was calculated from the slope (12.8 $\mu\text{A/mM}$) divided by the geometric surface area (0.07 cm^2). Thus, the sensitivity of the glucose sensor based on CHI-GOx/APTES/1.5%MWCNT- dPIn is 182.9 $\mu\text{A/mM} \cdot \text{cm}^2$.