Asymmetric Organocatalytic α-selective Vinylogous Michael Addition/isomerization for the Synthesis of Rauhut–Currier Type Products

Zhibing Weng,^{a,c} Ying Zhou,^{b,c} Xin Yue,^b Feng Jiang,^b and Wengang Guo*^b

^{*a*} School of Chemical and Pharmaceutical Engineering, Changzhou Vocational Institute of Engineering, Changzhou, China.

^b Schoolof Petrochemical Engineering, Jiangsu Key Laboratory of Advanced Catalytic Materials & Technology, Changzhou University, Changzhou, China.

Email: wgguo@cczu.edu.cn

^c These authors contributed equally to this work.

Contents

General InformationS	52
The Procedure for Synthesis of Vinylogous α-Keto EsterS3	3
The Procedure for the Vinylogous Michael Addition/Isomerization ReactionS3	3
The Procedure for the Transformation of 3p S4	4
Compound Characterization DataS6	,)
X-ray Crystal Structure of Compound 3b S12	2
Copy of NMR Spectrum and HPLC ChromatogramsS1	4

General Information

Unless otherwise noted, all reactions were carried out under argon atmosphere. TLC was performed with silica gel GF254 precoated on glass plates and spots were visualized with UV. ¹H and ¹³C spectra were recorded on a 400 MHz spectrometer (101 MHz for ¹³C NMR). The following abbreviations were used to designate chemical shift, multiplicities: s = singlet, d = doublet, t = triplet, q = quartert, m = multiplet, br = broad. All first-order splitting patterns were assigned on the basis of the appearance of the multiplet. Splitting patterns that could not be easily interpreted are designated as multiplet (m) or broad (br). Column chromatography was performed on silica gel (300-400 mesh). HPLC analysis was performed on Agilent HPLC 1100 or 1200 equipped with Daicel Chiralpak[®] AD-H, AS-H, or OD-H column. High resolution mass spectra for all the new compounds were done by an LTQ-Orbitrap instrument (ESI) (Thermo Fisher Scientific, USA). Dry THF and "PURIFICATION OF Et₂O were prepared based on the textbook LABORATORY CHEMICALS, FIFTH EDITION". Organocatalyst Α. allylmagnesium bromide (1.0 M solution in diethyl ether J&K Seal), and vinvlmagnesium bromide (1.0 M solution in THF J&K Seal) were purchased from J&K. Organocatalysts C, D, E, and F were purchased from Daicel Chiral Technologies (China) Co., LTD. Nitroalkenes 2a-2I, and 2g were purchased from Sigma-Aldrich or TCI. β -alkylnitroalkenes **2m**–**2p**. Dimethyl oxalate, diethyl oxalate, and Hoveyda-Grubbs II were purchased from the Energy Chemical. Other reagents and solvents were obtained from Tianjin Kemiou Chemical Reagent Co., Ltd. and directly used without further purification. The racemic products **3** were prepared under the catalysis of racemic organocatalyst F, which was obtained through the mixture of equal amount of (R,R)-**F** and (S,S)-**F** isomers.

The Procedure for Synthesis of Vinylogous α-Keto Ester 1

A solution of diethyl oxalate (4.38 g, 30 mmol) in anhydrous THF/Et₂O (1:1, 120 mL) under an argon atmosphere was cooled to -80 °C. Allylmagnesium bromide (30 mL of 1 M solution in diethyl ether) was added dropwise for 30 min, and the reaction was stirred for a further 30 min at the same temperature. A further small amount of allylmagnesium bromide (10 mL) was added, and the reaction was stirred for a further 10 min. The reaction was then warmed to -60 °C, and quenched by addition of 1 M H₂SO₄ (50 mL). The product was extracted into diethyl ether (3 x 50 mL), and the combined extracts were washed with brine (2 x 50 mL), dried (MgSO₄), and distilled under reduced pressure to give the desired product as yellow oil.

¹H NMR (400 MHz, CDCl₃) δ 5.93 (ddt, *J* = 17.1, 10.1, 6.8 Hz, 1H), 5.28 – 5.20 (m, 2H), 4.33 (q, *J* = 7.2 Hz, 2H), 3.61 (d, *J* = 6.8 Hz, 2H), 1.37 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 191.97, 160.89, 128.35, 120.41, 62.64, 43.74, 14.06; HRMS (ESI) m/z calcd for C₇H₁₀O₃, [M+H]⁺ 143.0708, found 143.0705.

The Procedure for the Preparation of Rauhut-Currier Type

Products

To a flame over-dried test-tube were added nitroalkene **2** (0.4 mmol), squaramide organocatalyst **F** (1.0 mol %, 0.004 mmol), and toluene (2.0 mL). The reaction mixture was stirred at -10 °C for 15 min, then vinylogous α -keto ester **1** (0.8 mmol, ~115 μ L) was added in one partition by using a pipettor. The reaction mixture was stirred at the same temperature until the full conversion of nitroalkene **2** (monitored by TLC analysis). After the completion of the reaction, the reaction was directly purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 5/1 as the eluent) to afford the pure product for NMR and HPLC analysis.

The Procedure for the Transformation of 3p

(1) To a solution of **3p** (0.4 mmol, 120 mg, 95% ee) and 2-vinylnaphthalene (1.2 mmol, 186 mg) in DCM (30 mL) was added Zhan catalyst-1B (5.0 mol %, 0.02 mmol, 16 mg) at room temperature under the protection of argon. The resulting mixture was stirred at reflux temperature. After 2 h, the mixture was filtered through Celite, concentrated under reduced pressure, and purified by silica gel column chromatography (petroleum ether/ethyl acetate = 5/1 as the eluent).

Waxy solid, 106 mg, 65% isolated yield based on **3p**.

¹H NMR (400 MHz, CDCl₃) δ 7.81 – 7.76 (m, 3H), 7.68 (s, 1H), 7.58 – 7.56 (m, 1H), 7.48 – 7.41 (m, 2H), 6.96 (q, *J* = 7.0 Hz, 1H), 6.54 (d, *J* = 15.8 Hz, 1H), 6.32 – 6.25 (m, 1H), 4.84 (dd, *J* = 12.6, 9.0 Hz, 1H), 4.61 (dd, *J* = 12.6, 5.8 Hz, 1H), 4.35 (q, *J* = 7.1 Hz, 2H), 3.63 – 3.55 (m, 1H), 2.27 (q, *J* = 7.0 Hz, 2H), 2.05 (d, *J* = 7.1 Hz, 3H), 1.91 – 1.84 (m, 1H), 1.73 – 1.65 (m, 1H), 1.55 – 1.41 (m, 2H), 1.37 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 189.07, 164.14, 152.17, 136.17, 135.08, 133.78, 132.85, 130.83, 130.23, 128.20, 127.94, 127.73, 126.28, 125.67, 125.60, 123.60, 77.60, 62.37, 36.86, 32.86, 29.57, 27.28, 15.53, 14.15.

HPLC analysis using Daicel CHIRALPAK OD-H column (*n*-Hexane/*i*PrOH = 85/15, 1.0 mL/min, $\lambda = 254$ nm, t_{major} = 27.9 min, t_{minor} = 35.0 min, 97% ee, $[\alpha]_D^{20} = 16.5$ (c = 0.8, in EtOAc))

HRMS (ESI) m/z calcd for C₂₄H₂₇NO₅, [M+H]⁺ 410.1967, found 410.1966.

(2) To a solution of **3p** (0.8 mmol, 120 mg) in DCM-H₂O (1:1, 8.0 mL) were added ruthenium (III) chloride (40 mol %, 0.32 mmol, 66.4 mg) and NalO₄ (1.5 eq, 1.20 mmol, 256 mg) at 25 °C. The solution was stirred for 3 h at the same temperature until the reaction was completed. The reaction was quenched by the addition of saturated aqueous $Na_2S_2O_3$ solution (10.0 mL). The solution was extracted with ethyl acetate, washed with brine, dried over Na_2SO_4 , filtered through Celite, and concentrated in vacuo to give the desired crude product. The desired product was obtained by silica gel column chromatography (PE/EA = 5/1 to 2/1 as the eluent).

0 O₂N CO₂Et

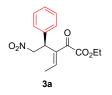
Light yellow oil, 94 mg, 41% isolated yield based on **3p**.

¹H NMR (400 MHz, CDCl₃) δ 9.73 (s, 1H), 6.95 (q, *J* = 7.1 Hz, 1H), 4.77 (dd, *J* = 12.7, 8.9 Hz, 1H), 4.59 (dd, *J* = 12.7, 6.0 Hz, 1H), 4.33 (q, *J* = 7.1 Hz, 2H), 3.59 – 3.52 (m, 1H), 2.45 (t, *J* = 7.0 Hz, 2H), 2.04 (d, *J* = 7.1 Hz, 3H), 1.85 – 1.75 (m, 1H), 1.66 – 1.51 (m, 3H), 1.36 (t, *J* = 7.1 Hz, 3H).

 ^{13}C NMR (101 MHz, CDCl₃) δ 201.39, 189.02, 164.01, 152.56, 135.73, 77.39, 62.41, 43.46, 36.78, 29.38, 19.85, 15.54, 14.11.

HRMS (ESI) m/z calcd for $C_{13}H_{19}NO_6$, [M+Na]⁺ 308.1110, found 308.1111. [α]_D²⁰ = 24.86 (c = 0.7, in EtOAc))

Compound Characterization Data



Light yellow liquid, 89.7 mg, 77% isolated yield based on **2a**.

¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.21 (m, 5H), 6.99 (q, *J* = 7.0 Hz, 1H), 5.23 – 5.10 (m, 2H), 4.87 (t, *J* = 7.6 Hz, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 2.12 (d, *J* = 7.0 Hz, 3H), 1.34 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.77, 164.00, 151.90, 137.30, 136.91, 129.08, 127.77, 127.67, 76.78, 62.45, 41.24, 15.63, 14.12.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 35.2$ min, $t_{minor} = 36.7$ min, 97% ee, $[\alpha]_D^{20} = 16.7$ (c = 0.8, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₁₇NO₅, [M+H]⁺ 292.1185, found 292.1181.



White solid, 118.5 mg, 80% isolated yield based on 2b.

¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, *J* = 8.4 Hz, 2H), 7.15 (d, *J* = 8.4 Hz, 2H), 7.00 (q, *J* = 7.1 Hz, 1H), 5.11 (d, *J* = 7.6 Hz, 2H), 4.81 (t, *J* = 7.6 Hz, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 2.12 (d, *J* = 7.1 Hz, 3H), 1.34 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.61, 163.79, 152.11, 136.54, 136.34, 132.18, 129.42, 121.81, 76.47, 62.56, 40.71, 15.69, 14.11.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 47.9$ min, $t_{minor} = 45.7$ min, 97% ee, $[\alpha]_D^{20} = 19.0$ (c = 1.1, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₁₆BrNO₅, [M+H]⁺ 370.0290, found 370.0292.

Light yellow oil, 114.0 mg, 77% isolated yield based on **2c**.

¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.38 (m, 2H), 7.22 – 7.16 (m, 2H), 7.03 (q, J = 7.1 Hz, 1H), 5.16 – 5.07 (m, 2H), 4.83 (t, J = 7.5 Hz, 1H), 4.32 (q, J = 7.1 Hz, 2H), 2.13 (d, J = 7.5 Hz, 3H), 1.34 (t, J = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.49, 163.76, 152.30, 139.51, 136.36, 131.00, 130.81, 130.60, 126.31, 123.04, 76.37, 62.57, 40.76, 15.74, 14.11.

HPLC analysis using Daicel CHIRALPAK AD-H column (n-Hexane/iPrOH =

97/3, 0.5 mL/min, λ = 220 nm, t_{major} = 37.9 min, t_{minor} = 34.4 min, 98% ee, $[\alpha]_D^{20}$ = 19.1 (c = 1.3, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₁₆BrNO₅, [M+H]⁺ 370.0290, found 370.0289.



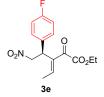
Light yellow oil, 103.7 mg, 70% isolated yield based on 2d.

¹H NMR (400 MHz, CDCl₃) δ 7.57 (d, *J* = 8.0 Hz, 1H), 7.41 (d, *J* = 8.0 Hz, 1H), 7.29 (t, *J* = 7.7 Hz, 1H), 7.15 (t, *J* = 7.7 Hz, 1H), 7.00 (q, *J* = 7.0 Hz, 1H), 5.30 – 5.15 (m, H), 4.90 (dd, *J* = 13.0, 5.7 Hz, 1H), 4.33 (q, *J* = 7.1 Hz, 2H), 2.08 (d, *J* = 7.0 Hz, 3H), 1.35 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 189.18, 163.86, 153.23, 135.89, 135.21, 133.48, 130.00, 129.55, 128.17, 124.33, 75.36, 62.50, 41.43, 16.26, 14.12.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 27.7$ min, $t_{minor} = 28.5$ min, 97% ee, $[\alpha]_D^{20} = 19.5$ (c = 1.0, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₁₆BrNO₅, [M+H]⁺ 370.0290, found 370.0279.



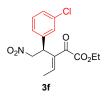
Light yellow oil, 85.4 mg, 69% isolated yield based on 2e.

¹H NMR (400 MHz, CDCl₃) δ 7.26 – 7.23 (m, 2H), 7.01 – 6.96 (m, 3H), 5.13 – 5.07 (m, 2H), 4.83 (t, *J* = 7.5 Hz, 1H), 4.31 (q, 7.5 Hz, 2H), 2.11 (d, *J* = 7.1 Hz, 3H), 1.33 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.76, 163.90, 162.14 (¹*J*_{C-F} = 247 Hz), 151.90, 136.82, 133.11 (⁴*J*_{C-F} = 3.1 Hz), 129.42 (³*J*_{C-F} = 8.2 Hz), 115.94 (²*J*_{C-F} = 21.6 Hz), 76.77, 62.49, 40.61, 15.57, 14.07.

HPLC analysis using Daicel CHIRALPAK OD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.8 mL/min, $\lambda = 220$ nm, $t_{major} = 49.8$ min, $t_{minor} = 41.6$ min, 97% ee, $[\alpha]_D^{20} = 18.9$ (c = 1.1, in EtOAc))

HRMS (ESI) m/z calcd for $C_{15}H_{16}FNO_5$, [M+H]⁺ 310.1091, found 310.1090.



Light yellow oil, 106.8 mg, 82% isolated yield based on 2f.

¹H NMR (400 MHz, CDCl₃) δ 7.28 – 7.24 (m, 3H), 7.17 – 7.16 (m, 1H), 7.03 (q, J = 7.1 Hz, 1H), 5.13 (d, J = 7.7 Hz, 2H), 4.84 (t, J = 7.7 Hz, 1H), 4.33 (q, J = 7.1 Hz, 2H), 2.13 (d, J = 7.1 Hz, 3H), 1.35 (t, J = 7.1 Hz, 3H).

 ^{13}C NMR (101 MHz, CDCl₃) δ 188.49, 163.76, 152.23, 139.24, 136.40, 134.87, 130.32, 128.07, 127.93, 125.83, 76.40, 62.56, 40.81, 15.72, 14.11.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 36.3$ min, $t_{minor} = 32.6$ min, 98% ee, $[\alpha]_D^{20} = 11.9$ (c = 0.8, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₁₆CINO₅, [M+H]⁺ 326.0795, found 326.0793.

Light yellow oil, 88.6 mg, 70% isolated yield based on 2g.

¹H NMR (400 MHz, CDCl₃) δ 7.58 (d, *J* = 8.4 Hz, 2H), 7.37 (d, *J* = 8.4 Hz, 2H), 7.04 (q, *J* = 7.1 Hz, 1H), 5.20 – 5.03 (m, 2H), 4.90 (t, *J* = 7.5 Hz, 1H), 4.29 (q, *J* = 7.1 Hz, 2H), 2.12 (d, *J* = 7.1 Hz, 3H), 1.30 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.41, 163.61, 152.82, 142.69, 136.04, 132.75, 128.48, 118.41, 111.68, 75.93, 62.64, 40.87, 15.78, 14.05.

HPLC analysis using Daicel CHIRALPAK OD-H column (*n*-Hexane/*i*PrOH = 85/15, 1.0 mL/min, $\lambda = 220$ nm, $t_{major} = 46.1$ min, >99% ee, $[\alpha]_D^{20} = 11.7$ (c = 0.7, in EtOAc))

HRMS (ESI) m/z calcd for C₁₆H₁₆N₂O₅, [M+H]⁺ 317.1137, found 317.1136.

Light yellow oil, 98.0 mg, 80% isolated yield based on **2h**.

¹H NMR (400 MHz, CDCl₃) δ 7.14 (q, *J* = 8.2 Hz, 4H), 6.97 (q, *J* = 7.1 Hz, 1H), 5.21 – 5.07 (m, 2H), 4.83 (t, *J* = 7.6 Hz, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 2.31 (s, 3H), 2.11 (d, *J* = 7.1 Hz, 3H), 1.35 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.83, 164.04, 151.70, 137.51, 137.06, 134.27, 129.74, 127.55, 76.91, 62.41, 40.97, 21.09, 15.60, 14.12.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, λ = 220 nm, t_{major} = 31.3 min, t_{minor} = 32.3, 97% ee, [α]_D²⁰ = 18.1 (c = 1.0, in EtOAc))

HRMS (ESI) m/z calcd for C₁₆H₁₉NO₅, [M+H]⁺ 306.1341, found 306.1329.

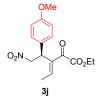
Light yellow oil, 92.0 mg, 69% isolated yield based on 2i.

¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.15 (m, 4H), 6.98 (q, *J* = 7.1 Hz, 1H), 5.23 – 5.06 (m, 2H), 4.84 (dd, *J* = 8.4, 6.9 Hz, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 2.87 (hept, *J* = 6.9 Hz, 1H), 2.11 (d, *J* = 7.1 Hz, 3H), 1.34 (t, *J* = 7.1 Hz, 3H), 1.22 (d, *J* = 6.9 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 188.85, 164.07, 151.73, 148.36, 137.02, 134.55, 127.61, 127.10, 76.92, 62.38, 40.98, 33.75, 23.94, 15.58, 14.10.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 98/2, 0.4 mL/min, $\lambda = 220$ nm, $t_{major} = 37.1$ min, $t_{minor} = 35.7$, 98% ee, $[\alpha]_D^{20} = 11.0$ (c = 0.5, in EtOAc))

HRMS (ESI) m/z calcd for C₁₈H₂₃NO₅, [M+H]⁺ 334.1654, found 334.1650.



Light yellow oil, 90.0 mg, 70% isolated yield based on 2j.

¹H NMR (400 MHz, CDCl₃) δ 7.19 (d, *J* = 8.6 Hz, 2H), 6.95 (q, *J* = 7.0 Hz, 1H), 6.84 (d, *J* = 8.6 Hz, 2H), 5.19 – 5.05 (m, 2H), 4.80 (t, *J* = 7.6 Hz, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 3.77 (s, 3H), 2.10 (d, *J* = 7.0 Hz, 3H), 1.34 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 188.90, 164.04, 159.04, 151.55, 137.13, 129.26, 128.87, 114.40, 77.03, 62.42, 55.35, 40.71, 15.57, 14.12.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 90/10, 0.8 mL/min, $\lambda = 220$ nm, $t_{major} = 20.2$ min, >99% ee, $[\alpha]_D^{20} = 10.4$ (c = 0.6, in EtOAc))

HRMS (ESI) m/z calcd for C₁₆H₁₉NO₆, [M+H]⁺ 322.1291, found 322.1289.

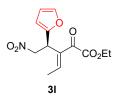
Light yellow oil, 103.5 mg, 87% isolated yield based on 2k.

¹H NMR (400 MHz, CDCl₃) δ 7.19 – 7.18 (m, 1H), 7.01 (q, *J* = 7.1 Hz, 1H), 6.95 – 6.91 (m, 2H), 5.21 – 5.06 (m, 3H), 4.33 (q, *J* = 7.1 Hz, 2H), 2.13 (d, *J* = 7.1 Hz, 3H), 1.35 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 188.19, 163.74, 151.94, 139.38, 136.34, 127.17, 125.92, 125.20, 77.19, 62.51, 36.96, 15.49, 14.12.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 38.7$ min, $t_{minor} = 37.5$ min, 98% ee, $[\alpha]_D^{20} = 10.5$ (c = 0.6, in EtOAc))

HRMS (ESI) m/z calcd for C₁₃H₁₅NO₅S, [M+H]⁺ 298.0749, found 298.0745.



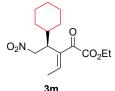
Light yellow oil, 86.6 mg, 77% isolated yield based on **2I**.

¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.30 (m, 1H), 7.05 (q, *J* = 7.1 Hz, 1H), 6.30 – 6.28 (m, 1H), 6.13 – 6.12 (m, 1H), 5.15 – 4.96 (m, 3H), 4.33 (q, *J* = 7.1 Hz, 2H), 2.09 (d, *J* = 7.1 Hz, 3H), 1.34 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 187.89, 163.84, 152.50, 149.96, 142.09, 134.40, 110.81, 107.15, 75.06, 62.49, 35.29, 15.51, 14.10.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 31.9$ min, $t_{minor} = 28.9$ min, 94% ee, $[\alpha]_D^{20} = 19.0$ (c = 1.2, in EtOAc))

HRMS (ESI) m/z calcd for C₁₃H₁₅NO₆, [M+H]⁺ 282.0978, found 282.0977.



Light yellow oil, 100.0 mg, 84% isolated yield based on 2m.

¹H NMR (400 MHz, CDCl₃) δ 6.92 (q, *J* = 7.0 Hz, 1H), 4.85 – 4.66 (m, 2H), 4.34 (q, *J* = 7.2 Hz, 2H), 3.25 (td, *J* = 9.9, 4.9 Hz, 1H), 1.99 (d, *J* = 7.0 Hz, 3H), 1.86 – 1.75 (m, 3H), 1.69 – 1.56 (m, 3H), 1.36 (t, *J* = 7.2 Hz, 3H), 1.30 – 0.95 (m, 4H), 0.84 – 0.75 (m, 1H).

 ^{13}C NMR (101 MHz, CDCl₃) δ 189.24, 164.28, 152.60, 136.14, 76.45, 62.30, 42.91, 37.16, 31.61, 31.44, 26.12, 26.05, 25.94, 15.58, 14.14.

HPLC analysis using Daicel CHIRALPAK OD-H column (*n*-Hexane/*i*PrOH = 98/2, 0.4 mL/min, $\lambda = 220$ nm, t_{major} = 25.4 min, t_{minor} = 27.1 min, >99% ee, $[\alpha]_D^{20} = 15.8$ (c = 0.9, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₂₃NO₅, [M+H]⁺ 298.1654, found 298.1651.

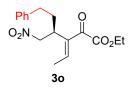
Light yellow oil, 72.1 mg, 60% isolated yield based on **2n**.

¹H NMR (400 MHz, CDCl₃) δ 6.90 (q, *J* = 7.0 Hz, 1H), 4.83 – 4.55 (m, 2H), 4.33 (q, *J* = 7.1 Hz, 2H), 3.56 – 3.48 (m, 1H), 2.01 (d, *J* = 7.0 Hz, 3H), 1.80 – 1.72 (m, 1H), 1.60 – 1.55 (m, 1H), 1.36 (t, *J* = 7.1 Hz, 3H), 1.24 (br, 8H), 0.86 (t, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 189.09, 164.17, 152.02, 136.28, 77.65, 62.31, 36.92, 31.63, 29.93, 29.03, 27.39, 22.60, 15.46, 14.12, 14.10.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 12.7$ min, $t_{minor} = 13.3$ min, 98% ee, $[\alpha]_D^{20} = 14.9$ (c = 0.7, in EtOAc))

HRMS (ESI) m/z calcd for C₁₅H₂₆NO₅, [M+H]⁺ 300.1811, found 300.1810.



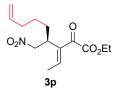
Light yellow oil, 112.4 mg, 88% isolated yield based on 20.

¹H NMR (400 MHz, CDCl₃) δ 7.30 – 7.27 (m, 2H), 7.22 – 7.18 (m, 1H), 7.15 – 7.13 (m, 2H), 6.93 (q, *J* = 7.1 Hz, 1H), 4.85 – 4.57 (m, 2H), 4.36 (q, *J* = 7.1 Hz, 2H), 3.58 – 3.50 (m, 1H), 2.68 – 2.61 (m, 1H), 2.55 – 2.47 (m, 1H), 2.19 – 2.09 (m, 1H), 2.03 – 1.94 (m, 1H), 1.83 (d, *J* = 7.1 Hz, 3H), 1.38 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 189.17, 164.16, 152.68, 140.41, 135.78, 128.66, 128.44, 126.39, 77.51, 62.41, 36.05, 33.23, 30.94, 15.35, 14.17.

HPLC analysis using Daicel CHIRALPAK AD-H column (*n*-Hexane/*i*PrOH = 97/3, 0.5 mL/min, $\lambda = 220$ nm, $t_{major} = 24.8$ min, $t_{minor} = 23.4$ min, 96% ee, $[\alpha]_D^{20} = 17.1$ (c = 0.8, in EtOAc))

HRMS (ESI) m/z calcd for C₁₇H₂₁NO₅, [M+H]⁺ 320.1498, found 320.1490.



Light yellow oil, 71.4 mg, 63% isolated yield based on **2p**.

¹H NMR (400 MHz, CDCl₃) δ 6.91 (q, *J* = 7.1 Hz, 1H), 5.72 (ddt, *J* = 16.9, 10.2, 6.7 Hz, 1H), 4.99 - 4.93 (m, 2H), 4.82 - 4.76 (m, 1H), 4.59 - 4.55 (m, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 3.56 - 3.49 (m, 1H), 2.05 - 2.00 (m, 5H), 1.82 - 1.72 (m, 1H), 1.62 - 1.55 (m, 1H), 1.39 - 1.24 (m, 5H).

¹³C NMR (101 MHz, CDCl₃) δ 189.05, 164.12, 152.12, 137.91, 136.14, 115.24, 77.56, 62.32, 36.77, 33.37, 29.34, 26.65, 15.46, 14.11.

HPLC analysis using Daicel CHIRALPAK OD-H column (*n*-Hexane/*i*PrOH = 99/1, 0.4 mL/min, $\lambda = 220$ nm, $t_{major} = 39.2$ min, $t_{minor} = 42.1$ min, 95% ee, $[\alpha]_D^{20} = 16.1$ (c = 1.0, in EtOAc))

HRMS (ESI) m/z calcd for C₁₄H₂₁NO₅, [M+Na]⁺ 306.1317, found 306.1319.

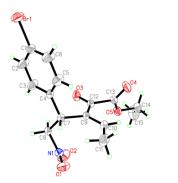
X-ray Crystal Structure of Compound 3b

Crystal data for compound 3b

Molecular formular	C ₁₅ H ₁₆ Br N O ₅
Fw	370.2
Crystal shape/color	Block/Colorless
Crystal size, mm	0.42×0.39×0.36
Crystal system	Triclinic
Space group	P-1
<i>a(</i> Å)	8.5887(8)
b(Å)	10.3821(10)
<i>c</i> (Å)	10.6032(8)
a (°)	96.304(7)
β(°)	112.149(8)
	105.649(8)
γ (°)	
V, Å3	819.19(13)
Z	2
Т, К	298(2)
m(Mo <i>K</i> a), mm ^{−1}	0.7107
$ ho_{calcd},~g~cm^{-3}$	1.501
Reflections/parameters	3617/201
Restraints	0
Observed reflections (<i>I</i> ³ 2 <i>s</i> (<i>I</i>))	2247
F(000)	376
$ au_{min}$	0.57498
\mathcal{T}_{max}	1.00000
Goodness of fit on <i>F</i> ²	1.063
$R_1, \omega r_2 \ (I \geq 2\sigma(I))^*$	0.0529,0.1111
$R_1, \omega R_2$ (all data)*	0.09994,0.1354
Largest peak and deepest hole (e Å-3)	0.605,-0.747

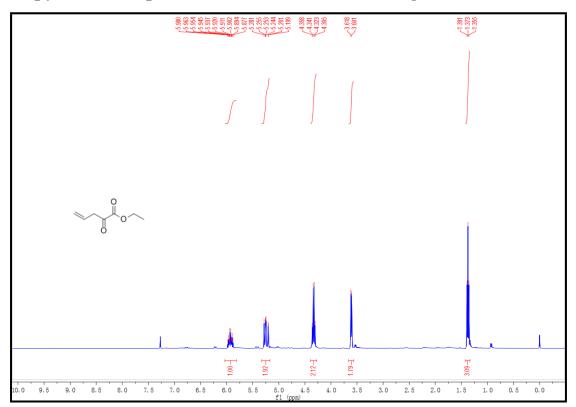
Br1-C1	1.893(4)
C1-C6	1.350(5)
C1-C2	1.361(6)
C2-C3	1.373(6)
C3-C4	1.371(5)
C4-C5	1.363(5)
C4-C7	1.528(5)
C5-C6	1.386(6)
C7-C9	1.523(4)
C7-C8	1.527(5)
C8-N1	1.490(5)
C9-C10	1.337(4)
C9-C12	1.461(5)
C10-C11	1.488(5)
C12-O3	1.212(4)
C12-C13	1.524(4)
C13-O4	1.196(4)
C13-O5	1.311(4)
C14-C15	1.454(7)
C14-O5	1.463(4)
N1-O2	1.204(5)
N1-O1	1.206(4)

Selected bond lengths (Å) and angles for 3b

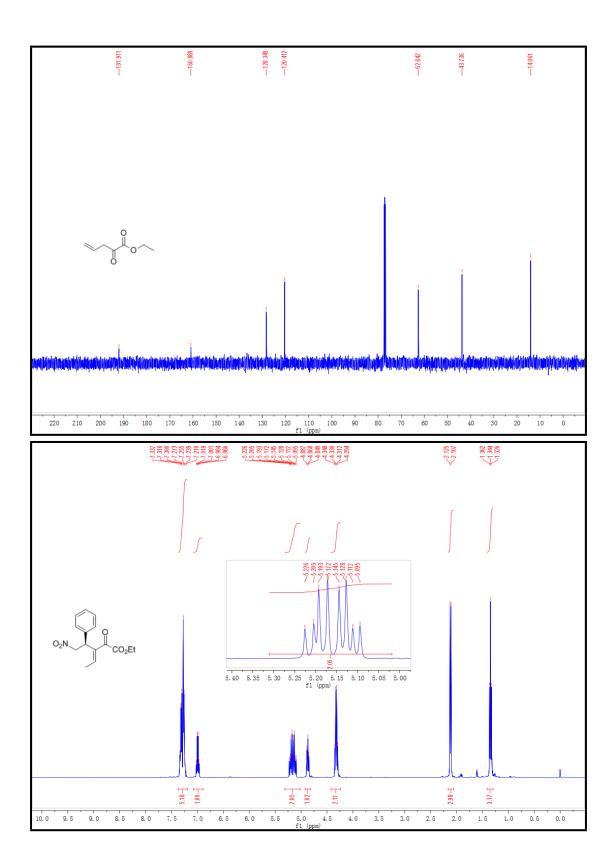


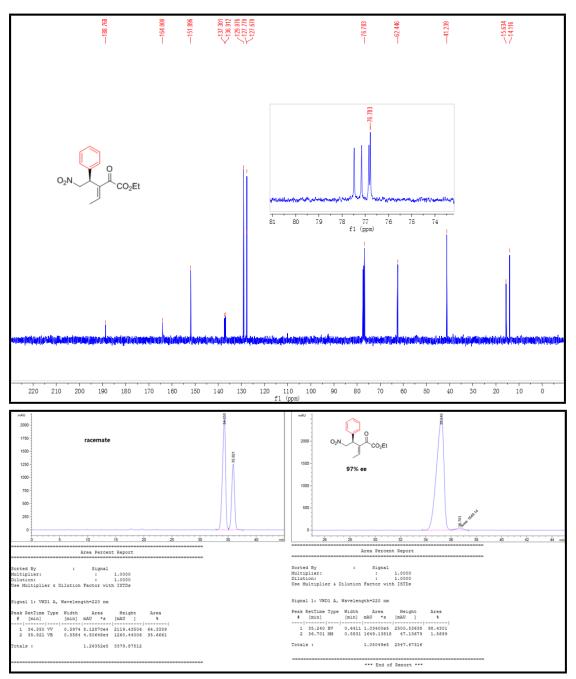


CCDC **1536616** contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from Cambridge Crystallographic Data Centre via <u>www.ccdc.cam.ac.uk/data request/cif</u>

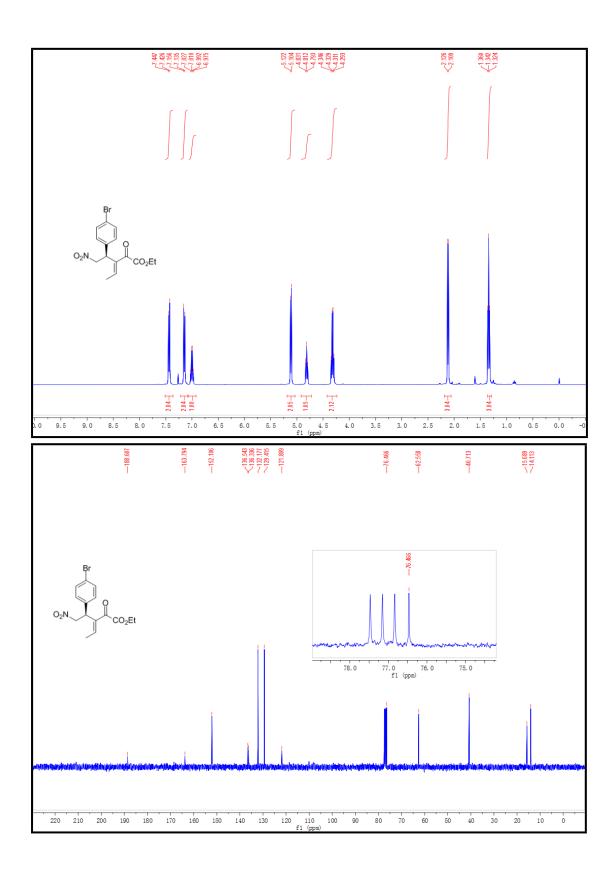


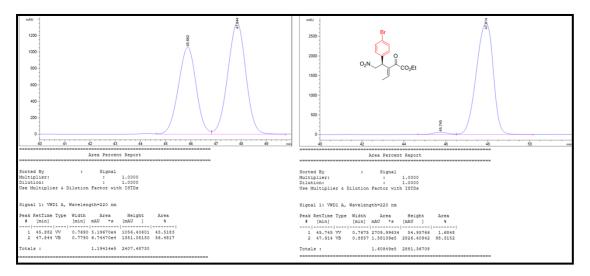
Copy of NMR Spectrum and HPLC Chromatograms



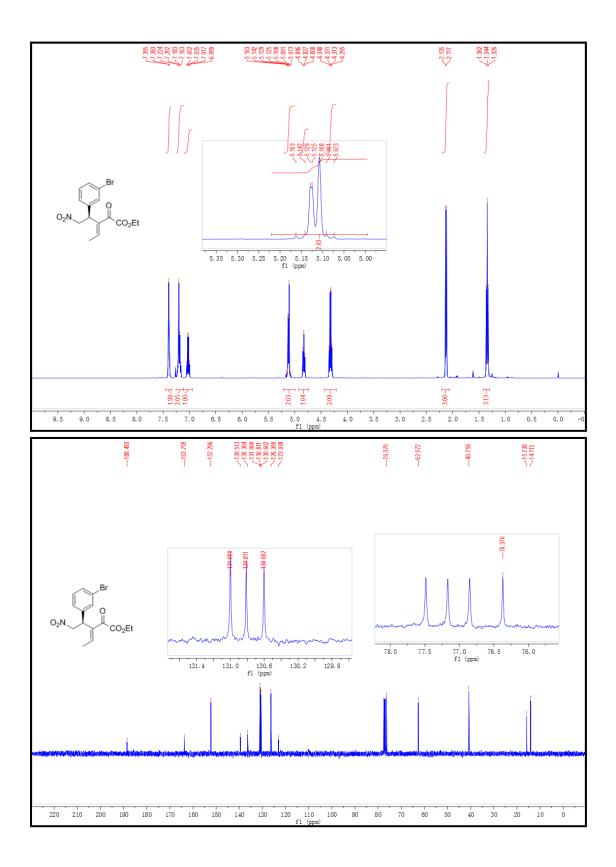


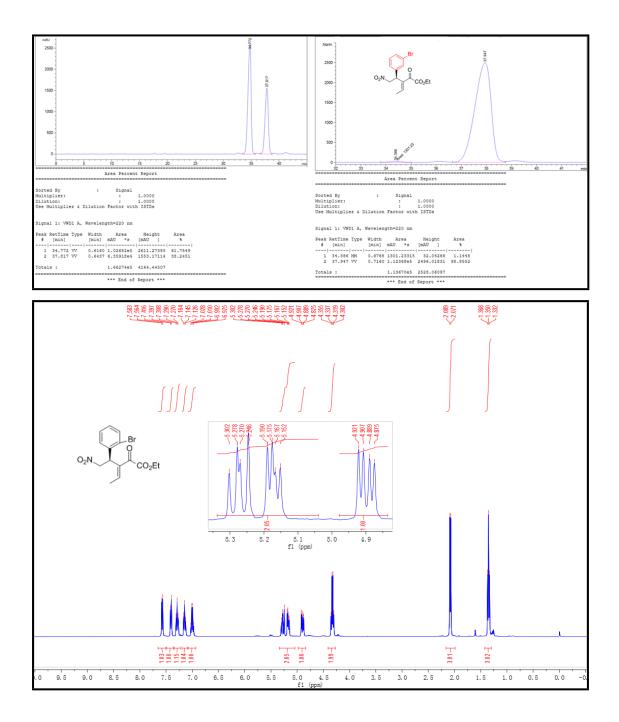
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.

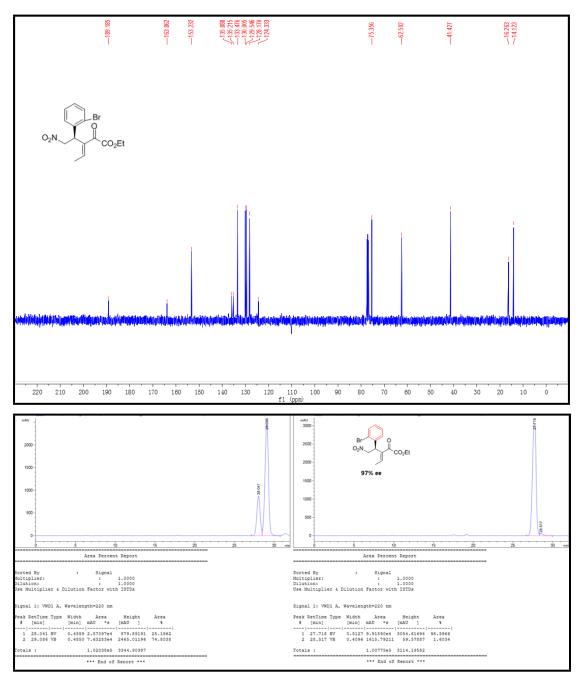




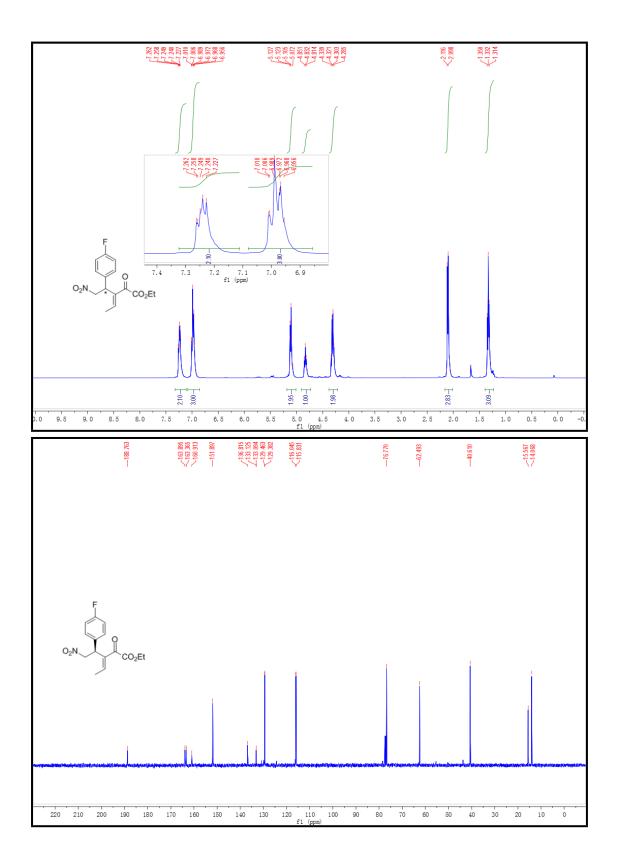
Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.



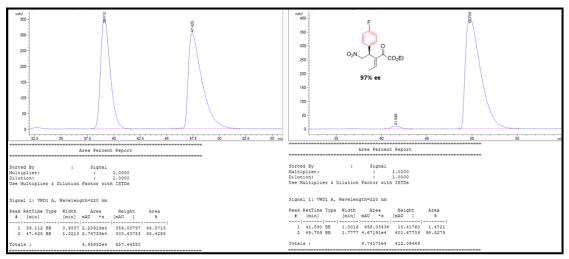




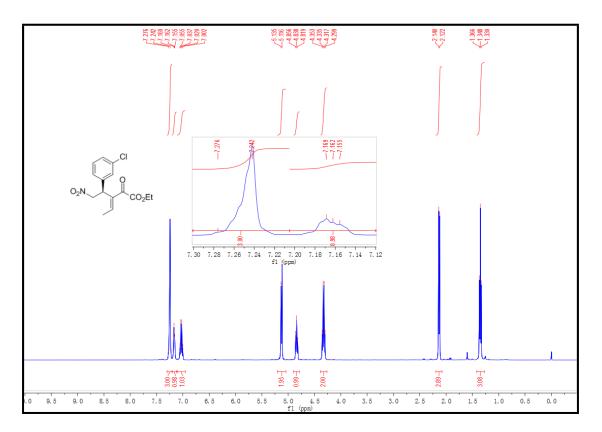
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.

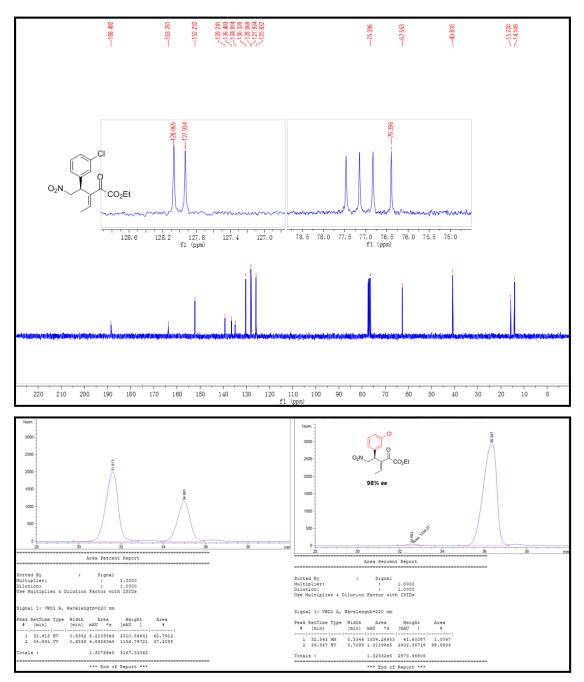


S22

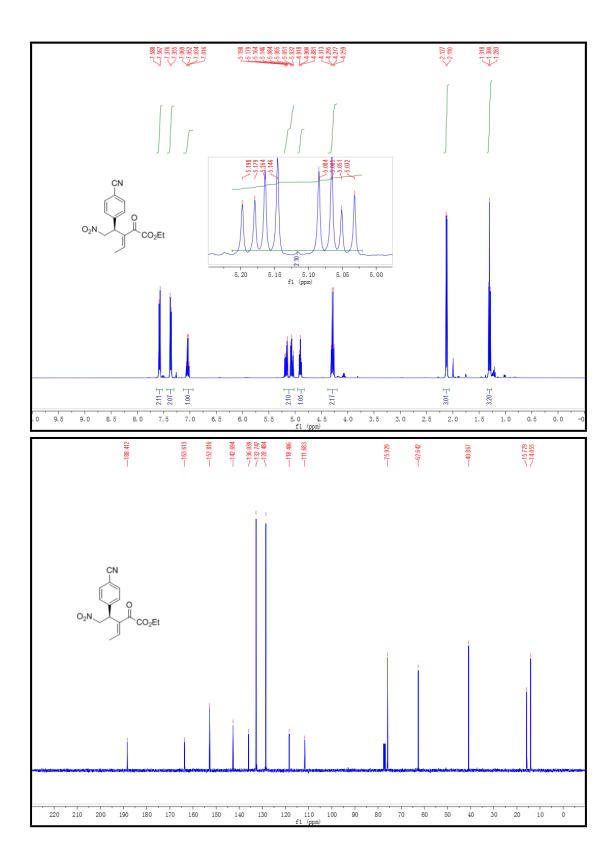


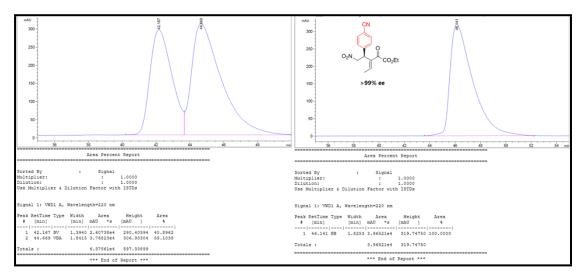
Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.



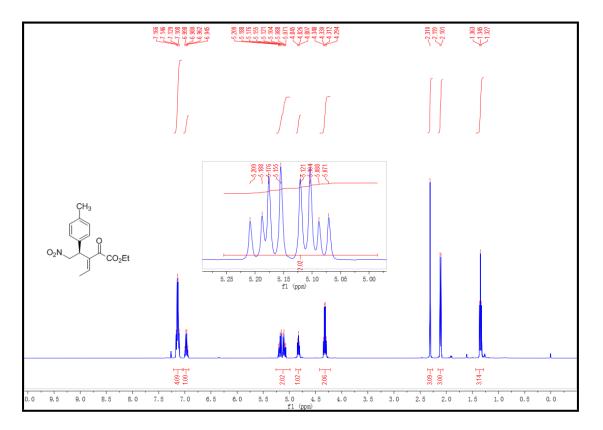


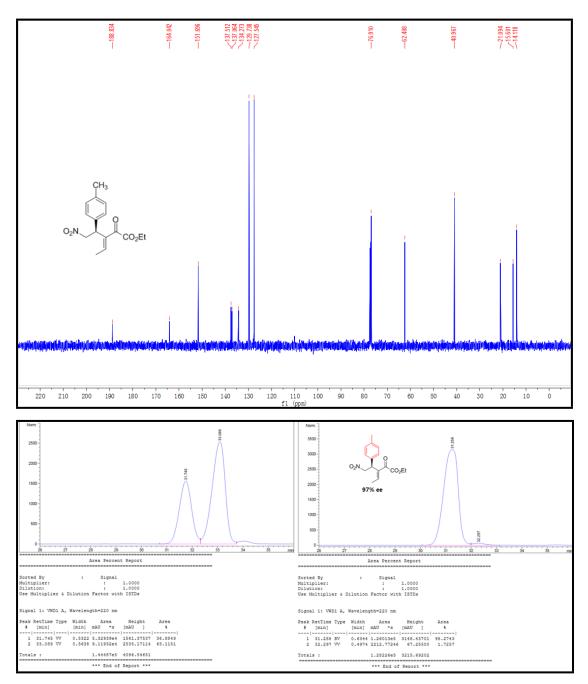
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.



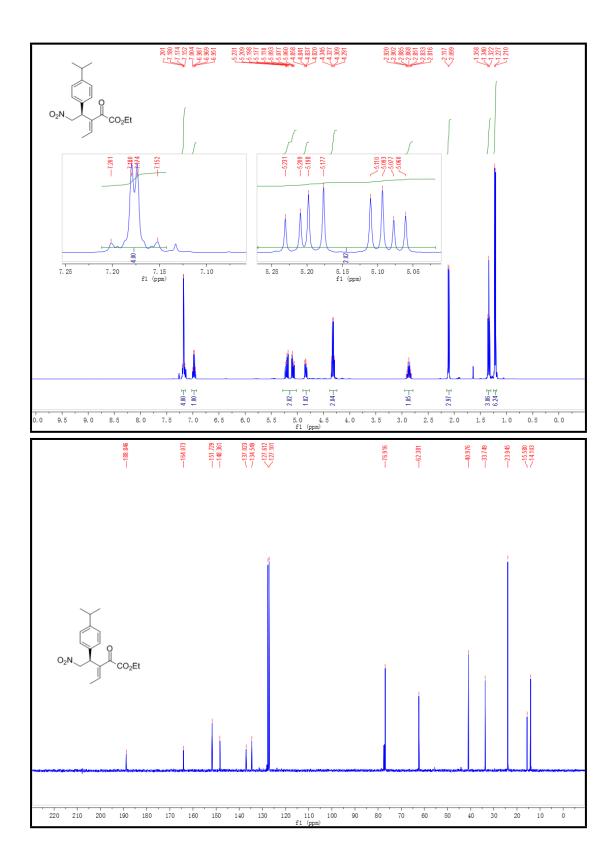


Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.

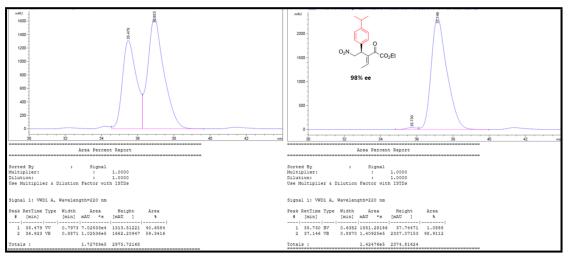




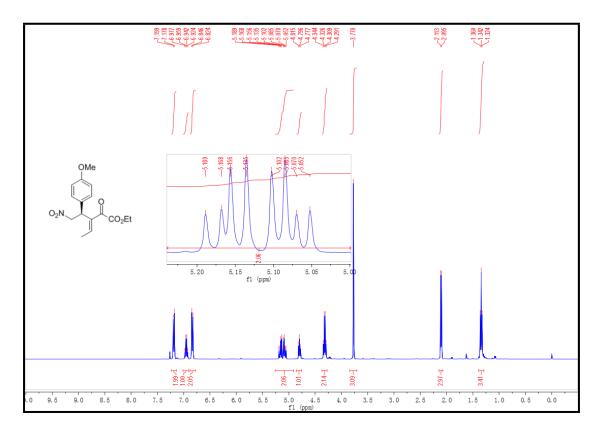
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.

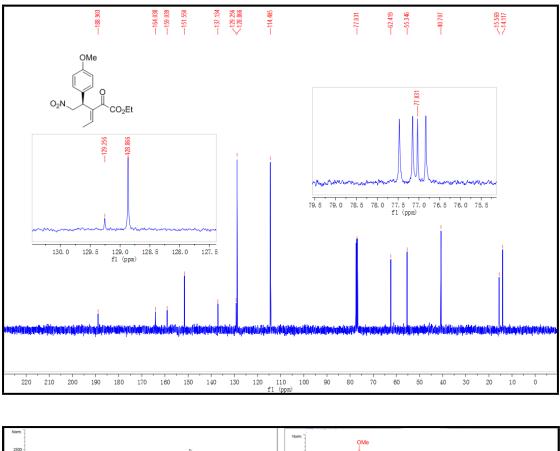


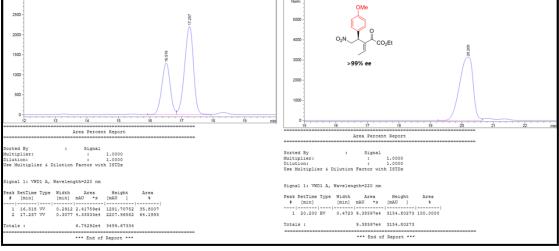
S28



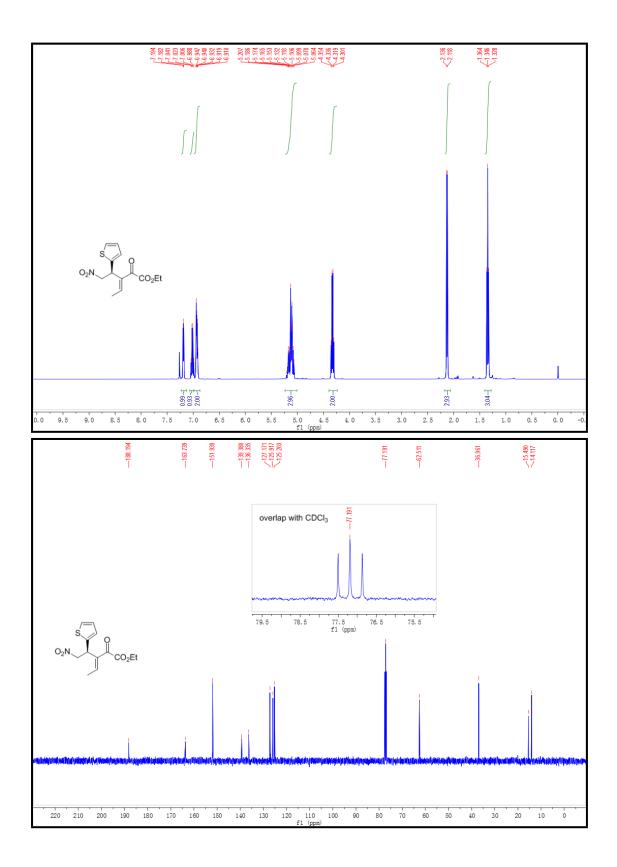
Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.

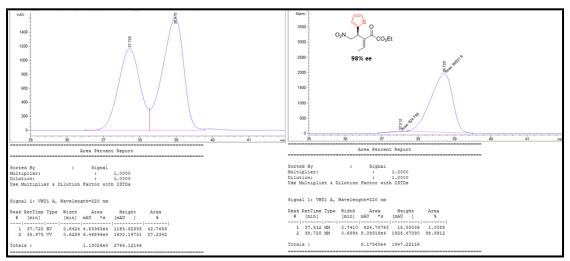




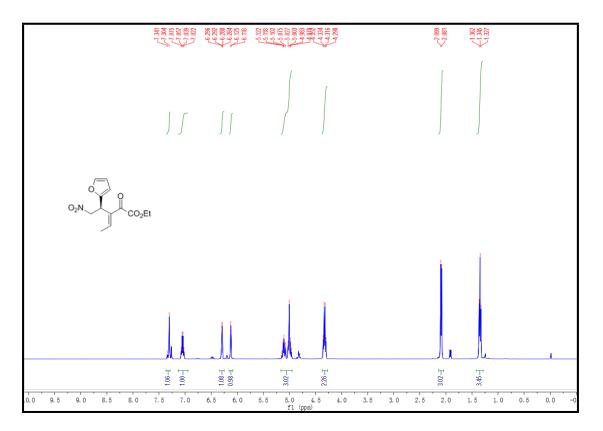


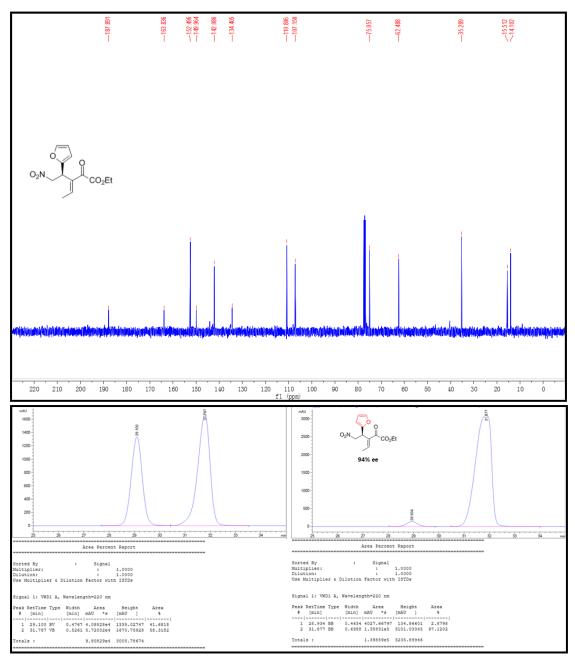
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (*S*,*S*)- \mathbf{F} and (*R*,*R*)- \mathbf{F} isomers.



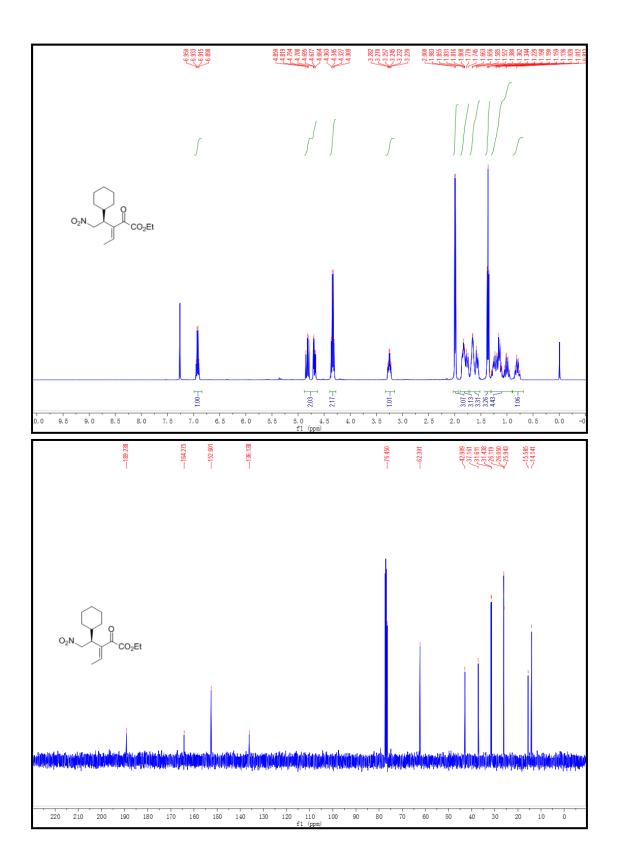


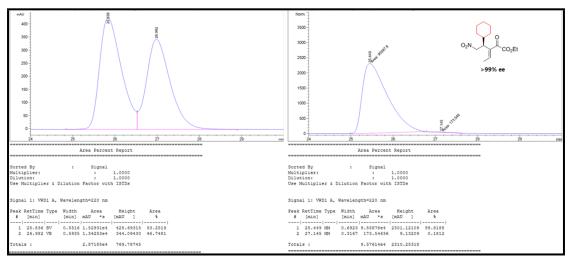
Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.



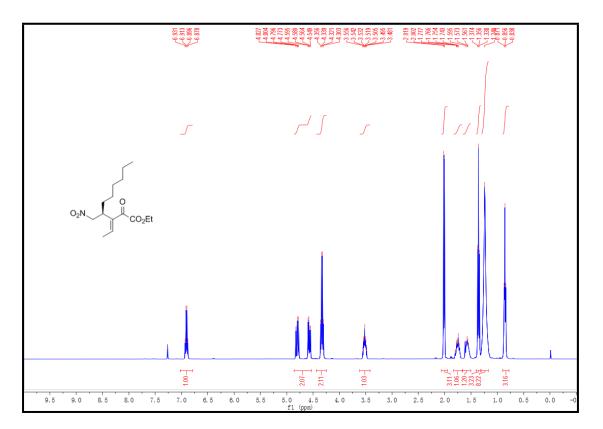


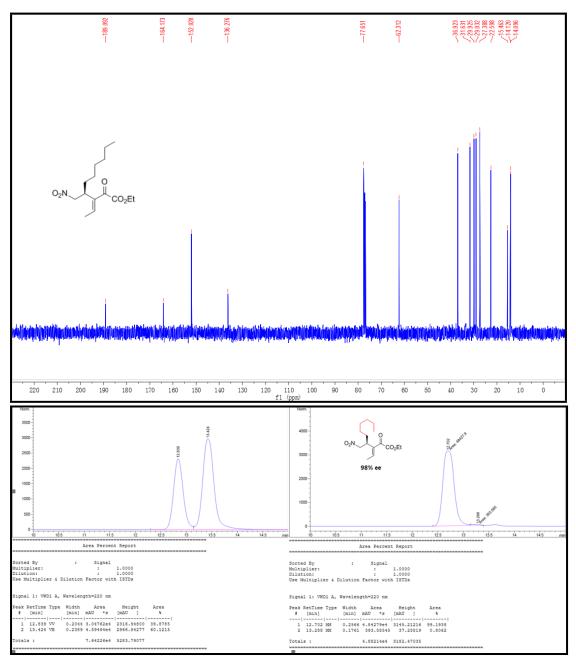
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.



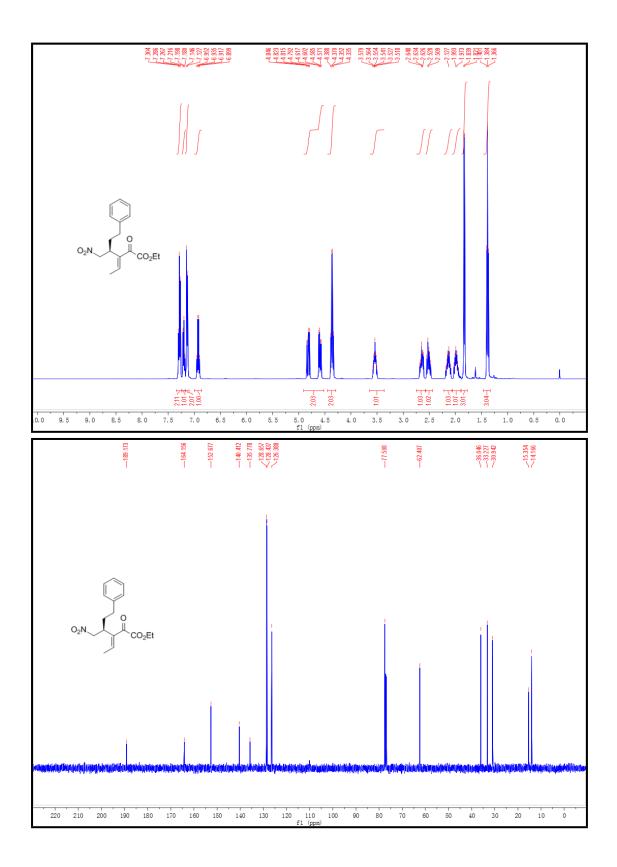


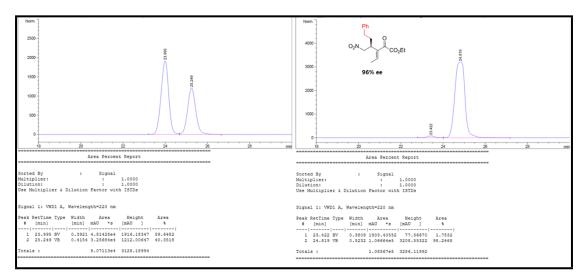
Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.



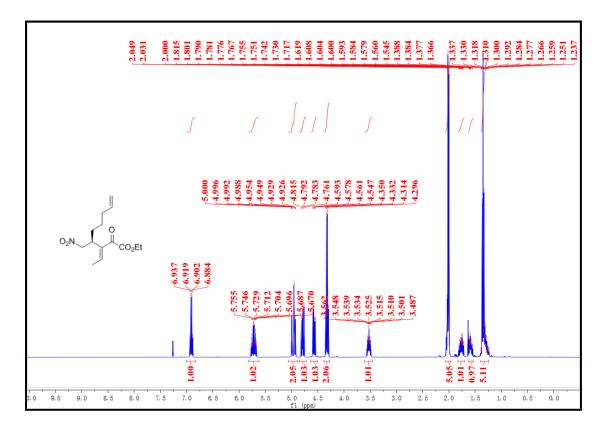


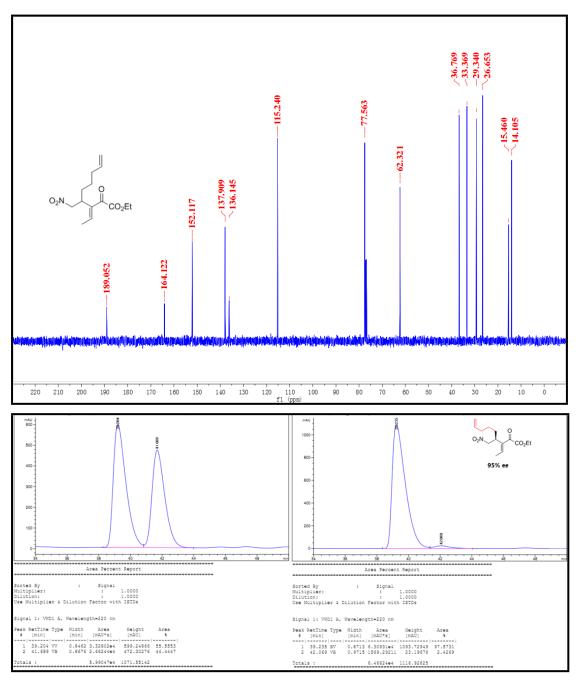
Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.



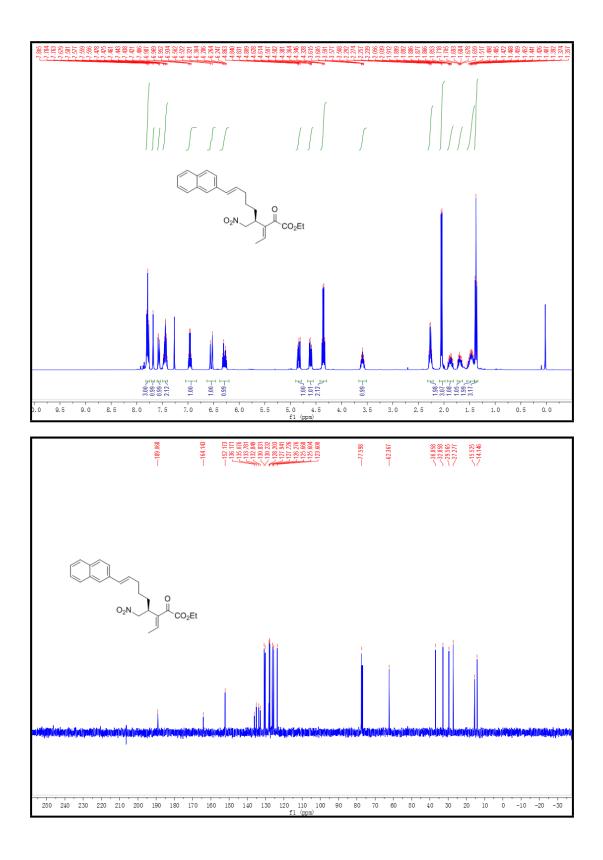


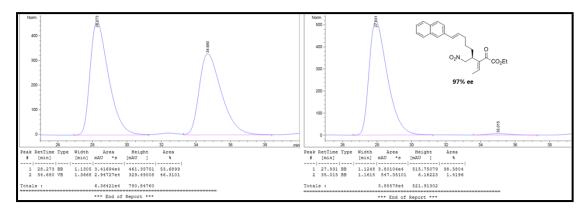
Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.





Note: the racemic product was prepared under the catalysis of racemic organocatalyst **F**, which was obtained through the mixture the equial amount of (S,S)-**F** and (R,R)-**F** isomers.





Note: the racemic product was prepared under the catalysis of racemic organocatalyst \mathbf{F} , which was obtained through the mixture the equial amount of (S,S)- \mathbf{F} and (R,R)- \mathbf{F} isomers.

