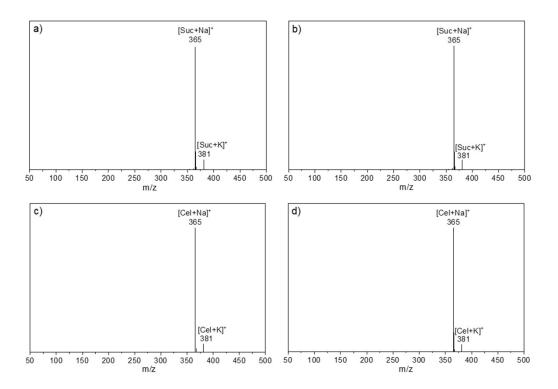
## - Supporting Information-

## Probing the Effects of Fructose Concentration on the Evolution of Humins during Fructose Dehydration

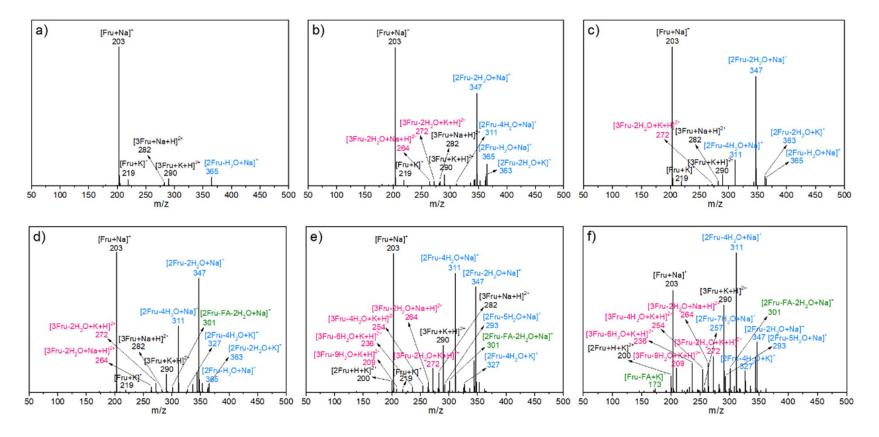
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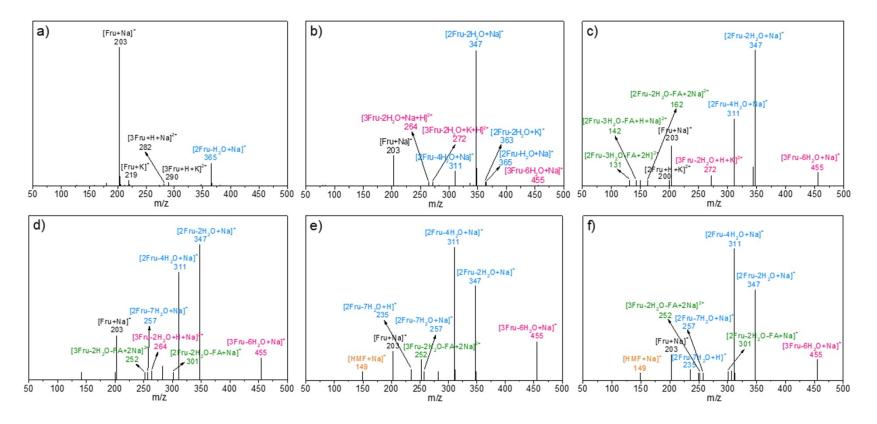
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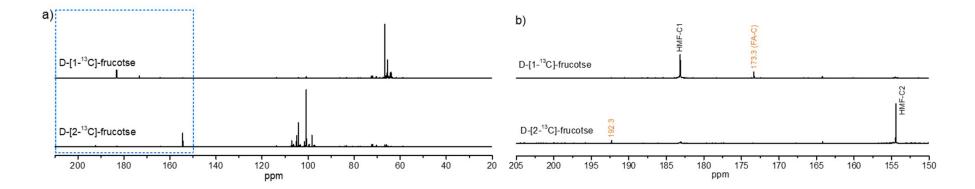
**Fig. S1** ESI-MS spectra of the aqueous solutions of a) 0.5 M sucrose, b) 2.0 M sucrose, c) 0.5 M cellobiose, and d) 2.0 M cellobiose.



**Fig. S2** The ESI-MS spectra of the reaction mixture before (a) and after (b ~ f) reacting fructose (18.0 *wt.*%) at 140 °C for different reaction time. b) 15 min, c) 30 min, d) 60 min, e) 90 min, and f) 120 min. Note: the peak with m/z of 209 could also be assigned to  $[2Fru+HMF-6H_2O+H+K]^{2+}$ ,  $[Fru+2HMF-3H_2O+H+K]^{2+}$ , or  $[3HMF+H+K]^{2+}$ ; the peak with m/z of 236 could also be assigned to  $[2Fru+HMF-3H_2O+H+K]^{2+}$ ; the peak with m/z of 254 could also be assigned to  $[2Fru+HMF-H_2O+H+K]^{2+}$ ; the peak with m/z of 264 could also be assigned to  $[2Fru+HMF+H_2O+H+K]^{2+}$ ; the peak with m/z of 272 could also be assigned to  $[2Fru+HMF+H_2O+H+K]^{2+}$ ; the peak with m/z of 272 could also be assigned to  $[2Fru+HMF+H_2O+H+K]^{2+}$ ; the peak with m/z of 301 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{2+}$ .

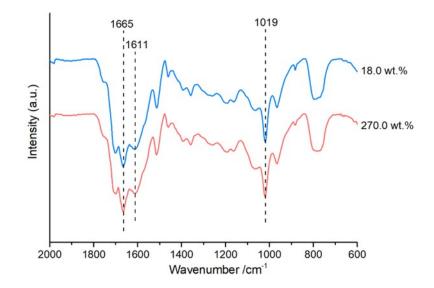


**Fig. S3** The ESI-MS spectra of the reaction mixture before (a) and after (b ~ f) reacting fructose (270.0 *wt.*%) at 140 °C for different reaction time. b) 15 min, c) 30 min, d) 60 min, e) 90 min, and f) 120 min. Note: the peak with m/z of 131 could also be assigned to  $[Fru-FA+HMF+2H]^{2+}$ ; the peak with m/z of 142 could also be assigned to  $[Fru-FA+HMF+H_2O+2Na]^{2+}$ ; the peak with m/z of 252 could also be assigned to  $[2Fru-FA+HMF+H_2O+2Na]^{2+}$ ; the peak with m/z of 264 could also be assigned to  $[2Fru+HMF+H_2O+H+Na]^{2+}$ ; the peak with m/z of 301 could also be assigned to  $[2Fru+HMF+H_2O+H+Na]^{2+}$ ; the peak with m/z of 301 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$  is the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be assigned to  $[2Fru+HMF+H_2O+Na]^{+}$ ; the peak with m/z of 455 could also be

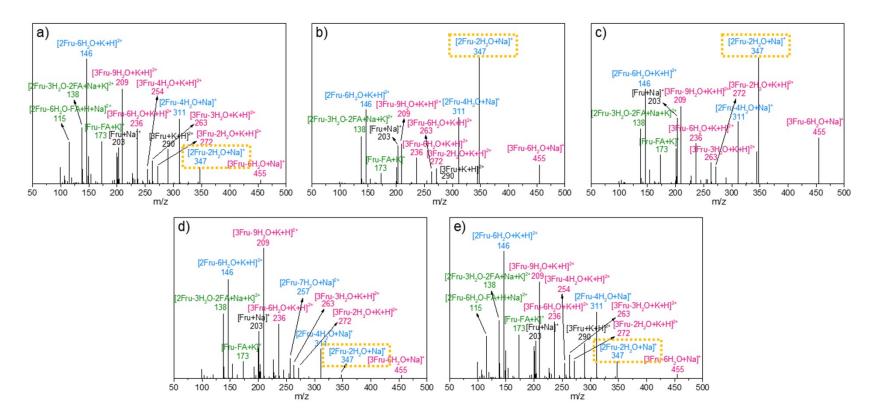


**Fig. S4** <sup>13</sup>C NMR spectra of the reaction mixture after reacting fructose or isotopically labelled fructose at 140 °C for 0.5 h. Reaction conditions: 720 mg substrate, 1 mL H<sub>2</sub>O, 0.05 M H<sub>2</sub>SO<sub>4</sub>, under a N<sub>2</sub> atmosphere. b) represented the enlarged spectra in the dotted regions of a).

The peaks in the range of 60 ~ 110 ppm were assigned to isotope-labelled fructose (Fig. S4a). The peak at 173.3 ppm in Fig. 4b was ascribed to carbon of FA formed by the C1-C2 cleavage of D-[ $1^{-13}$ C]-fructose. The peak at 192.3 ppm was assigned to the C11 species formed by condensation of degradative D-[ $2^{-13}$ C]-fructose with another fructose molecule.



**Figure S5** FT-IR spectra of the solid humins obtained after reacting fructose with different concentrations for a long reaction time. Reaction conditions: 1 mL of  $H_2O$ , 0.05 M of  $H_2SO_4$ , 140 °C, 2 h, under a  $N_2$  atmosphere.



**Fig. S6** ESI-MS spectra of the reaction mixture after reacting fructose (72.0 *wt.*%) at 140 °C in various solvents for different reaction time. a) H<sub>2</sub>O, 90 min, b) THF-H<sub>2</sub>O (*V*/*V*=7/3), 15 min, c) DIO-H<sub>2</sub>O (*V*/*V*=7/3), 15 min, d) THF-H<sub>2</sub>O (*V*/*V*=7/3), 30 min, and e) DIO-H<sub>2</sub>O (*V*/*V*=7/3), 30 min. The yellow dotted squares label the signal of DFAs. Note: the peak with m/z of 115 could also be assigned to [Fru-FA-3H<sub>2</sub>O+HMF+H+Na]<sup>2+</sup> or [2HMF-FA+H+Na]<sup>2+</sup>; the peak with m/z of 138 could also be assigned to [Fru+HMF-2FA+Na+K]<sup>2+</sup>; the peak with m/z of 209 could also be assigned to [2Fru+HMF-6H<sub>2</sub>O+H+K]<sup>2+</sup>, [Fru+2HMF-3H<sub>2</sub>O+H+K]<sup>2+</sup>, or [3HMF+H+K]<sup>2+</sup>; the peak with m/z of 236 could also be assigned to [2Fru+HMF-3H<sub>2</sub>O+H+K]<sup>2+</sup>; the peak with m/z of 254 could also be assigned to [2Fru+HMF-H<sub>2</sub>O+H+K]<sup>2+</sup>; the peak with m/z of 263 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 272 could also be assigned to [2Fru+HMF+H+K]<sup>2+</sup>; the peak with m/z of 455 could also be assigned to [2Fru+HMF-3H<sub>2</sub>O+Na]<sup>+</sup> or [Fru+2HMF+Na]<sup>+</sup>.

Fructose concentration (wt.%)	pH value
-	7.01
4.5	6.99
18.0	6.94
72.0	6.87
180.0	6.72
270.0	6.58
360.0	6.56

Table S1. The pH values of aqueous fructose solutions with various concentrations.<sup>[a]</sup>

<sup>[a]</sup> Different amount of fructose was dissolved in water and kept static for 5 h to ensure stable. The pH value was detected by a pen type pH meter (SX-620, Sanxin) at room temperature (20 °C).

r (cm⁻¹)	Assignments <sup>[a]</sup>	
781	deformation vibrations of skeleton & $\delta$ (C1-O-H) c	
811	υ <b>(O-C2-C3) of β-1,3-difuranose</b>	
	deformation vibrations of skeleton & $\delta$ (C6-O-H) c;	
821	υ(C2-C3-C4) & δ(C3-O-H) f;	
	stretch vibrations of skeleton & $\delta$ (C5-O-H) p	
870	δ(C2-O-H) & υ(C2-C3) c	
921	δ(C1H2) & υ(C2-O) p	
	deformation vibrations of skeleton & $\delta$ (C6-O-H) c;	
968	stretch vibrations of skeleton & $\delta$ (C2-O-H) & $\delta$ (C4-O-H) f;	
	<b>δ(C2-O-H) &amp; δ(C1H2OH) &amp;</b> υ(C5-C6-O) p	
	<b>υ(C1-O-H&amp;C6-O) c;</b>	
981	stretch vibrations of skeleton & $\delta$ (C6-O-H) & $\delta$ (C4-O-H) f;	
	<b>υ(C3-C4) &amp; v(C4-O-H) p</b>	
1017	υ(C5-O-C2) & δ(C6-O-H) & δ((C1)-O-H) f;	
	ບ <b>(C1-O)</b> p	
1036	δ(С5-О-Н) с;	
	υ(C1-O) & δ(C6)-O-H) f;	
	ບ(C5-O) p	
1064	υ(C1-O) & υ(C3-O) & δ(C6)-O-H) c;	
	υ(C5-O-C2) & δ(C5)-O-H) & δ(C1)-O-H) p	
1085	υ(C4-C5) & δ(C5-O-H) & δ((C1)-O-H) p	
	υ(C1-O) & υ(C3-O-H) & δ(C4)-O-H) c;	
1105	υ(C3-C4) & δ(C4)-O-H) & δ(C2)-O-H) f;	
	δ(C1)-O-H) p	
1158	δ(С6)-О-Н) с;	
	υ(C1-C2) & δ(C1-H) p	
1183	δ(С2)-О-Н) с;	
	δ(C6)-O-H) f;	
	δ(С3)-О-Н) & δ(С2)-О-Н) р	
	δ(С3)-О-Н) с;	
1252	δ(C4-O-H) f;	
	δ(C1-H) & ω(CH2) & ω(C4H) p;	

Table S2. The attribution of characteristic ATR-IR peaks in this study.<sup>1</sup>

<sup>[a]</sup>f = fructofuranose, p = fructopyranose, c = fructoketose,  $\upsilon$  = stretching vibration,  $\delta$  = deformation vibrations,  $\omega$  = wagging vibrations.

## References

1. X. Fu, Y. Hu, Y. Zhang, Y. Zhang, D. Tang, L. Zhu and C. Hu, ChemSusChem, 2020, 13, 501-512.