Supramolecular Cuboctahedra with Aggregation-Induced Emission Enhancement and External Binding Ability

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1. Schemes of preparation of ligands



Scheme S1. Synthesis route of ligand L1 and L2

2. Experimental section

General Procedures. Reagents and solvents were purchased from Energy Chemical, Bidepharm and used without purification. Thin layer chromatography (TLC) was performed on flexible sheets (Greagent) precoated with Al₂O₃ (IB-F) and SiO₂ (IB2-F) and visualized by UV light. Column chromatography was conducted using neutral Al₂O₃ (200-300 mesh) SiO₂ (200-300 mesh). ¹H, ¹³C, 2D COSY, 2D ¹H-¹H NOESY NMR and 2D DOSY NMR spectra were recorded on a Bruker NMR 400, 500, 600 MHz. Different NMR solvents were purchased from J&K scientific and Sigma/Aldrich. ESI-MS and TWIM-MS were recorded with a Waters Synapt G2-Si tandem mass spectrometer, using solutions of 0.01 mg sample in 1 mL of CHCl₃/CH₃OH (1:3, *v/v*) for ligands or 0.5 mg sample in 1 mL of DMF/MeOH (3:1, *v/v*) for complexes.

TWIM-MS. The TWIM-MS experiments were performed under the following conditions: ESI capillary voltage, 2 kV; sample cone voltage, 35 V; source offset, 42V; source temperature 150 °C; desolvation temperature, 250 °C; cone gas flow, 10 L/ h; desolvation gas flow, 700 L/h (N₂); source gas flow, 0 mL/min; trap gas flow, 3 mL/min;

helium cell gas flow, 120 mL/min; ion mobility (IM) cell gas flow, 30 mL/min; sample flow rate, 8 μ L/min; IM traveling wave height, 25 V; and IM traveling wave velocity, 1200 m/s. Q was set in rf-only mode to transmit all ions produced by ESI into the triwave region for the acquisition of TWIM-MS data. Data were collected and analyzed by using Mass Lynx 4.2 and Drift Scope 2.9.

gMS². Gradient tandem mass spectrometry was performed under the following conditions:17+ charged ions of complexes, 17+ charged ions of **S1 (S2)** \supset **G** were isolated by quadrupole for the following collision induced dissociation (CID), in which collision energy was gradually increased by changing the voltage of trap cell depended on different complexes.

TEM. Transmission electron microscopy tests were performed on the JEOL JEM-2100F equipment. The sample solutions were drop-casted on to a lacey carbon covered Cu grid (300 mesh, purchased from Beijing Zhongjingkeyi Technology Co., Ltd.) and the extra solution was absorbed by filter paper to avoid aggregation.

AFM. AFM imaging was performed on a Bruker Dimension Icon AFM system with ScanAsyst and the data was processed by NanoScope Analysis version 2.0 (Bruker Software, Inc.) The sample was diluted to a concentration of 10⁻⁶ M using DMF, dropped on freshly cleaved mica surface, and then dried in the air. Silicon cantilevers tip with spring constant of around 0.1 N/m was used for the experiments.

DLS. Dynamic light scattering (DLS) was carried out on a Nano-ZS90 instrument at room temperature. The sample was diluted to a concentration of 10⁻⁶ M using DMF.

Molecular Modeling. Energy minimization of the macrocycles was conducted with the Materials Studio version 6.0 program, using the Anneal and Geometry Optimization tasks in the Forcite module (Accelrys Software, Inc.). The counterions were omitted.

UV-vis absorption and fluorescence properties. UV-vis absorption spectra were recorded on a Thermo Fisher Scientific Evolution 201 spectrophotometer at room temperature (0.5 μ M in CHCl₃ or DMF) and were corrected with the background spectrum of the solvent. Fluorescence properties were performed on HITACHI Exciter F-4600 at room temperature. (1 μ M in CHCl₃ or DMF).

Single crystal X-ray diffraction. Structural confirmation of L1 and L2 were provided by X-ray crystallographic analysis. Single crystal X-ray diffraction data for L1 and L2 were collected on a XtaLAB Synergy diffractometer using a mirror monochromated Cu-K α radiation. Two suitable crystals of L1 and L2 were selected and texted on Bruker P4 diffractometer. These crystals were kept at 100.00 K corresponding to L1 and L2 during data collection. Using Olex2, the structures were solved with the SIR2004 [2] structure solution program using Direct Methods and refined with the XH [3] refinement package using CGLS minimisation. The structures were solved by direct methods and refined by full-matrix least-squares on F^2 with anisotropic displacement using the SHELXTL software package. Details on crystal data collection and refinement were summarized in Table S3. CCDC: 2132604 (L1), 2132525 (L2).

3. Synthesis of the compounds and supramolecules



Compound 1: To a stirred solution of thianthrene (2.2 g, 10.0 mmol) in CH₂Cl₂ (60 mL) at 0 °C, a solution of Br₂ (19.2 g, 120 mmol) in CH₂Cl₂ (10 mL) was added dropwise. After stirring at 25 °C for 48 h, the reaction mixture was washed with saturated Na₂SO₃ solution until colorless, dried over anhydrous MgSO₄, and then concentrated in vacuo to give the product as 3.20 g white solid (yield: 60%). ¹H NMR (500 MHz, CDCl₃, 300 K): $\delta = 7.23$ (s, 4 H, Ph-*H*^{*a*}). ¹³C NMR (100 MHz, CDCl₃, 300 K): $\delta = 138.91, 132.44, 125.27.^{1-2}$ ESI-TOF (*m*/*z*): Calcd. for [C₁₂H₄Br₄S₂+H⁺]=532.90, Found: 532.91.



L1: The mixture of compound 1 (595.9 mg, 1 mmol), 4'-(4-boronatophenyl)-2,2':6',2"terpyridine (2.12 g, 6 mmol), Na₂CO₃ (1.27 g, 12 mmol), and Pd(PPh₃)₂Cl₂ (140.2 mg, 0.2 mmol) in the solution of MeOH (80 mL), H₂O (120 ml) and toluene (200 mL) were refluxed for 96 h under N₂. After separating the toluene layer, the aqueous layer was extracted with CH₂Cl₂ (3×50 mL). The combined organic layers were dried with MgSO₄, and concentrated in vacuo. The residue was washed with methanol, collected and crystallized in CHCl₃ and methanol to give pure light blue solid: 896.0 mg (yield: 65%); ¹H NMR (500 MHz, CDCl₃, 298 K, ppm) 8.81 (s, 8H, tpy- $H^{3',5'}$), 8.67 (d, $J_{6,6"}$ -5,5"= 4.8 Hz, 8H, tpy- $H^{6,6"}$), 8.6 (d, $J_{3,3"-4,4"}$ = 8.5 Hz, 8H, tpy- $H^{3,3"}$), 7.85 (d, J_{a-b} = 8.4 Hz, 8H, Ph- H^{a}), 7.80 (m, 8H, tpy- $H^{4,4"}$), 7.73 (s, 4H, Ph- H^{c}), 7.37 (d, J_{b-a} = 8.7 Hz, 8H, Ph- H^{b}), 7.34 (m, 8H, tpy- $H^{5,5"}$). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 156.10, 156.05,

149.09, 149.03, 138.77, 138.55, 138.39, 136.77, 136.07, 129.00, 128.26, 128.22, 125.24, 123.78, 121.28, 118.84. ESI-TOF (m/z): Calcd. for [C₉₆H₆₀N₁₂S₂] = 1445.77, Found: 1445.78.



Compound 2: A solution of compound **1** and excess chromic oxide in acetic acid was heated at 100°C for 24 h. It was then cooled and the resulting white solid was collected by filtration. The compound is difficult to purify by the column as it is insoluble in most of the solvents and used without further purification for next step.³⁻⁴ ESI-TOF (m/z): Calcd. for [C₁₂H₄Br₄O₄S₂+Na⁺]=618.89, Found: 618.92.



L2: The mixture of compound **2** (537.5 mg, 1 mmol), 4'-(4-boronatophenyl)-2,2':6',2"terpyridine (1.91 g, 6 mmol), Na₂CO₃ (1.145 g, 12 mmol), and Pd(PPh₃)₂Cl₂ (208.1 mg, 0.2 mmol) in the solution of EtOH (80 mL), H₂O (120 mL) and toluene (200 mL) was refluxed for 48 h under Ar. After separating the toluene layer, the aqueous layer was extracted with CHCl₃ (150 mL, 3×50 mL). The combined organic layers were dried (MgSO₄), and concentrated in vacuo. The residue was washed with methanol, collected and crystallized in methanol to give pure light blue solid: 896.0 mg; ¹H NMR (500 MHz, CDCl₃, 298 K, ppm) 8.74 (s, 8H, tpy- $H^{3',5'}$), 8.68 (d, $J_{6,6"-5,5"}$ = 4.8 Hz, 8H, tpy- $H^{6,6"}$), 8.64 (d, $J_{3,3"-4,4"}$ = 8.5 Hz, 8H, tpy- $H^{3,3"}$), 8.44 (s, 4H, Ph- H^c), 7.90 (d, J_{a-b} = 8.4 Hz, 8H, Ph- H^a), 7.84 (m, 8H, tpy- $H^{4,4"}$), 7.38 (d, J_{b-a} = 8.7 Hz, 8H, Ph- H^b), 7.32 (m, 8H, tpy- $H^{5,5"}$). ¹³C NMR (125 MHz, CDCl₃, ppm): δ 156.10, 156.05, 149.09, 149.03, 138.77, 138.55, 138.39, 136.77, 136.07, 129.00, 128.26, 128.22, 125.24, 123.78, 121.28, 118.84. ESI-TOF (*m*/*z*): Calcd. for [C₉₆H₆₀N₁₂O₄S₂] =1509.71, Found: 1509.73.



S1: A methanol solution of Zn(NTf₂)₂ (37.63 mg, 0.06 mmol) was added slowly drop by drop into a stirred solution of **L1** (42.0 mg, 0.03 mmol) in CHCl₃ and MeOH (1:1, 20 mL). The mixture was stirred at 55°C for 8 h, then 10-fold excess LiNTf₂ was added for the anion exchange. The residue was filtered, washed with water (10 mL × 3) and MeOH (10 mL × 3), and dried *in vacuo* to give complex **3**, as pale yellow solid (76.8 mg, 98%), ESI-TOF (*m/z*): 2662.14 [M–11 NTf₂⁻]¹¹⁺ (calcd *m/z*: 2662.14), 2416.89 [M–12NTf₂⁻]¹²⁺ (calcd *m/z*: 2416.89), 2209.45 [M–13NTf₂⁻]¹³⁺ (calcd *m/z*: 2209.45), 2031.63 [M–14NTf₂⁻]¹⁴⁺ (calcd *m/z*: 2031.63), 1877.53 [M–15NTf₂⁻]¹⁵⁺ (calcd *m/z*: 1877.53), 1742.69 [M–16NTf₂⁻]¹⁶⁺ (calcd *m/z*: 1742.69), 1623.65 [M–17NTf₂⁻]¹⁷⁺ (calcd *m/z*: 1623.65), 1517.90 [M–18NTf₂⁻]¹⁸⁺ (calcd *m/z*: 1517.90), 1423.28 [M–19NTf₂⁻]¹⁹⁺ (calcd *m/z*: 1423.28), 1338.12 [M–20NTf₂⁻]²⁰⁺ (calcd *m/z*: 1338.12); ¹H NMR (500 MHz, CD₃CN/DMF-*d*₇ (v/v, 4:1), 298 K, ppm) 9.01 (s, 96H, tpy-*H*^{3',5'}), 8.73 (d, *J*_{3,3"-4,4"}= 8.5 Hz, 96H, tpy-*H*^{3,3"}), 8.18 (d, *J*_{a-b} = 8.1 Hz, 96H, Ph-*H*^a), 8.09 (m, 96H, tpy-*H*^{4,4"}), 8.01 (s, 48H, Ph-*H*^e), 7.85 (d, *J*_{6,6"-5,5"}= 4.5 Hz, 96H, tpy-*H*^{6,6"}), 7.66 (d, *J*_{b-a} = 8.6 Hz, 96H, Ph-*H*^b), 7.36 (m, 96H, tpy-*H*^{5,5"}).



S2: A methanol solution of $Zn(NTf_2)_2$ (22.88 mg, 0.04 mmol) was added slowly drop by drop into a stirred solution of **L2** (27.2 mg, 0.02 mmol) in CHCl₃ and MeOH (1:1,

20 mL). The mixture was stirred at 55°C for 8 h, then 10-fold excess LiNTf₂ was added for the anion exchange. The residue was filtered, washed with water (10 mL × 3) and MeOH (10 mL× 3), and dried *in vacuo* to give complex **3**, as light blue solid (48.7 mg, 98%), ESI-TOF (*m/z*): 2731.95 [M–11NTf₂⁻]¹¹⁺ (calcd *m/z*: 2731.95), 2840.95 [M–12NTf₂⁻]¹²⁺ (calcd *m/z*: 2840.95), 2268.50 [M–13NTf₂⁻]¹³⁺ (calcd *m/z*: 2268.50), 2086.48 [M–14NTf₂⁻]¹⁴⁺ (calcd *m/z*: 2086.48), 1928.71 [M–15NTf₂⁻]¹⁵⁺ (calcd *m/z*: 1928.71), 1790.68 [M–16NTf₂⁻]¹⁶⁺ (calcd *m/z*: 1790.68), 1668.82 [M–17NTf₂⁻]¹⁷⁺ (calcd *m/z*: 1668.82), 1560.56 [M–18NTf₂⁻]¹⁸⁺ (calcd *m/z*: 1560.56), 1643.68 [M–19NTf₂⁻]¹⁹⁺ (calcd *m/z*: 1643.68), 1376.51 [M–20NTf₂⁻]²⁰⁺ (calcd *m/z*: 1376.51); ¹H NMR (500 MHz, CD₃CN/DMF-*d*₇ (*v*/*v*, 4:1), 298 K, ppm) 9.02 (s, 96H, tpy-*H*^{3',5'}), 8.75 (d, *J*_{3,3"-4,4"}= 8.9 Hz, 96H, tpy- *H*^{3,3"}), 8.62 (s, 48H, Ph-*H*^e), 8.24 (d, *J*_{a-b} = 7.3 Hz, 96H, Ph-*H*^a), 8.10 (m, 96H, tpy-*H*^{4,4"}), 7.86 (d, *J*_{6,6"-5,5"}= 4.5 Hz, 96H, tpy-*H*^{6,6"}), 7.74 (d, J_{b-a} = 7.5 Hz, 96H, Ph-*H*^b), 7.38 (m, 96H, tpy-*H*^{5,5"}).

4. Synthesis of Porphyrin Zn



5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) porphyrin.⁵⁻⁷ In a three-necked round bottom flask, a mixture of the 3,4,5-trimethoxybenzaldehyde (2.00 g, 10.19 mmol), and pyrrole (0.67 g, 10.19 mmol), BF₃·Et₂O (498 µL, 4.08 mmol) were added to 500 mL dichloromethane and 5 mL ethanol. The mixture was stirred at room temperature for 120 mins then 2,3-dicloro-5,6-diciano-1,4-benzoquinona (DDQ) (2.28 g, 10.19 mmol) was added and kept the solution stirring for 4 h. In the end, triethylamine (5.7 mL, 40.76 mmol) was injected into the obtained solution. The mixture was concentrated to 80 mL under reduced pressure. The crude product was purified was subjected to column chromatography on SiO₂ with dichloromethane and concentrated under reduced pressure. Expected compound was obtained as a purple solid (373 mg, yield 15%) after washing with acetone and dried. ¹H NMR (400 MHz, 298 K, CDCl₃) δ 8.96 (s, 8H, H^β-pyrro), 7.47 (s, 8H, *Ph*), 4.18 (s, 12H, $-OCH_3^a$), 3.97 (s, 24H, $-OCH_3^b$), -2.78 (s, 2H, -NH). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 151.67, 138.04, 137.24, 120.33, 113.31, 60.92, 56.72, 31.12. ESI-TOF (*m*/*z*): Calcd. for [C₅₆H₅₄N₄O₁₂ + H⁺]: 976.06. Found:976.10.



5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) porphyrin zinc (Guest). In a round bottom flask, 5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) porphyrin (100 mg, 0.10 mmol) was dissolved in 60 mL chloroform and a solution of Zn(OAc)₂ (55 mg, 0.30

mmol) in 8 mL methanol was added. The reaction mixture was stirred at room temperature for 8 h. The crude product was concentrated under reduced pressure and the residue was washed with methanol and dried under vacuum to get the dark purple solid (97.9 mg, 92%). ¹H NMR (400 MHz, 298 K, DMSO-*d*₆) δ 8.94 (s, 8H, H^β-pyrro), 7.51 (s, 8H, *Ph*), 4.00 (s, 12H, $-OCH_3^a$), 3.90 (s, 24H, $-OCH_3^b$). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 151.35, 149.79, 138.77, 137.71, 132.10, 120.59, 113.25, 60.93, 56.67. ESI-TOF (*m/z*): Calcd. for [C₅₆H₅₂N₄O₁₂Zn]: 1038.43. Found:1038.46.

5. ¹H NMR, ¹³C NMR, 2D COSY NMR, 2D NOESY NMR, 2D DOSY NMR



Figure S1. ¹H NMR spectrum (500 MHz, 298 K) of L1 in CDCl₃.



Figure S2. ¹³C NMR spectrum (125 Hz, 298 K) of L1 in CDCl₃.



Figure S3. 2D ¹H-¹H COSY spectrum (500 MHz, 298 K) of L1 in CDCl₃.



Figure S4. ¹H NMR spectrum (400 MHz, 298 K) of S1 in DMF-*d*₇ and CD₃CN (1:4, *v/v*).



Figure S5. 2D ¹H-¹H COSY NMR spectrum (500 MHz, 298 K) of S1 in DMF- d_7 and CD₃CN (1:4, v/v).



Figure S6. 2D ¹H-¹H NOESY NMR spectrum (500 MHz, 298 K) of S1 in DMF-d₇ and CD₃CN (1:4,

v/v).



Figure S7. ¹H NMR spectrum (500 MHz, 298 K) of L2 in CDCl₃.



Figure S8. ¹³C NMR spectrum (125 MHz, 298 K) of L2 in CDCl₃.



Figure S9. 2D ¹H-¹H COSY NMR spectrum (500 MHz, 298 K) of L2 in CDCl₃.



Figure S10. ¹H NMR spectrum (400 MHz, 298 K) of S2 in DMF-*d*₇ and CD₃CN (1:4, *v/v*).



Figure S11. 2D ¹H-¹H COSY NMR spectrum (500 MHz, 298 K) of **S2** in DMF- d_7 and CD₃CN (1:4, v/v).



Figure S12. 2D ¹H-¹H NOESY NMR spectrum (500 MHz, 298 K) of S2 in DMF- d_7 and CD₃CN (1:4, v/v).



Figure S13. 2D ¹H-¹H DOSY NMR spectrum of S1 in DMF- d_7 and CD₃CN (1:4, v/v).

The sphere hydrodynamic radius can be estimated as follow according to the Stokes-Einstein Equation. Where D is the diffusion constant, k is the Boltzmann's constant, T is the temperature, μ is the viscosity of solvents, and R is the radius of the sphere-like particles:

$$D = \frac{kT}{6\pi\mu R}$$

 $D = 10^{-10.02} \text{ m}^2 \text{ s}^{-1}$ k = 1.38×10⁻²³ N m K⁻¹ T = 298 K $\mu = 4.22 \times 10^{-4} \text{ N m}^{-2} \text{ s} (\text{CD}_3\text{CN/DMF } (4:1, v/v)^8$ r = $\frac{kT}{6\pi\mu D}$ = 2.90×10⁻⁹ m = 2.72 nm

The radius of the spherical **S1** is 5.44 nm, which is consistent with the result of computer modeling.

Figure S14. 2D ¹H-¹H DOSY NMR spectrum of S2 in DMF- d_7 and CD₃CN (1:4, v/v).

The sphere hydrodynamic radius can be estimated as follow according to the Stokes-Einstein Equation. Where D is the diffusion constant, k is the Boltzmann's constant, T is the temperature, μ is the viscosity of solvents, and R is the radius of the sphere-like particles:

$$D = \frac{kT}{6\pi\mu R}$$

 $D = 10^{-10.05} \text{ m}^2 \text{ s}^{-1}$ $k = 1.38 \times 10^{-23} \text{ N m K}^{-1}$ T = 298 K $\mu = 4.22 \times 10^{-4} \text{ N m}^{-2} \text{ s (CD_3CN/DMF (4:1, v/v)^8)}$ $r = \frac{kT}{6\pi\mu D} = 2.90 \times 10^{-9} \text{ m} = 2.91 \text{ nm}$

The radius of the spherical S2 is 5.82 nm, which is consistent with the result of computer modeling.

Figure **S15**. ¹H NMR spectrum (400 MHz, 298 K) of 5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) porphyrin in DMSO-*d*₆.

Figure S16. ¹³C NMR spectrum (100 MHz, 298 K) of 5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) porphyrin in DMSO-*d*₆.

Figure S17. ¹H NMR spectrum (400 MHz, 298 K) of 5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) porphyrin zinc in DMSO-*d*₆.

Figure S18. ¹³C NMR spectrum (100 MHz, 298 K) of 5,10,15,20-tetrakis(3,4,5-trimethoxyphenyl) zinc porphyrin in DMSO-*d*₆.

Figure S19. 2D ¹H-¹H DOSY NMR spectrum of **S1** combing zinc porphyrin in DMF- d_7 and CD₃CN (1:4, v/v).

Figure S20. 2D ¹H-¹H DOSY NMR spectrum of **S2** combing zinc porphyrin in DMF- d_7 and CD₃CN (1:4, v/v).

6. ESI-MS spectra data of supramolecules (NTf₂⁻ as counterion)

Figure S21. TWIM-MS plot (m/z vs drift time) of S1 (a) and S2 (b).

Figure S22. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from S1 (NTf_2^- as counterion).

Figure S23. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from S2 (NTf₂⁻ as counterion).

Figure S24. gMS² of **S1** at m/z 1623.7 with different collision energies.

Figure S25. gMS² of S2 at m/z 1668.8 with different collision energies.

Figure S26. (a) ESI-MS (full spectra) of one porphyrin adducts $S1 \supset G$, (b) ESI-MS of free S1 (black), one zinc porphyrin adducts $S1 \supset G$ (green), two zinc porphyrin adducts $S1 \supset G_2$ (yellow), (c) TWIM-MS of free S1 (black), one zinc porphyrin adducts $S1 \supset G$ (green), two zinc porphyrin adducts $S1 \supset G$ (green).

Figure S27. (a) (b) ESI-MS of free S1 (S2) (black), one porphyrin adducts S1 (S2) \supset G (green), two porphyrin adducts S1 (S2) \supset G₂ (yellow), (c) (d) TWIM-MS of free S1 (S2) (black), one porphyrin adducts S1 (S2) \supset G (green), two porphyrin adducts S1 (S2) \supset G₂ (yellow).

Figure S28. (a) gMS² of one porphyrin adducts S1 \supset G (NTf₂⁻) at *m/z* 1681.4 with different collision energies; (b) gMS² of one zinc porphyrin adducts S1 \supset G (NTf₂⁻) at *m/z* 1684.7 with different collision energies.

Figure S29. gMS² of one porphyrin adducts S2 \supset G (NTf₂⁻) at *m*/*z* 1726.3 with different collision energies.

Figure S30. (a) ESI-MS of porphyrin adducts of S1 with different times. (a) 0 h, (b) 6 h, (c) 12 h, (d) 24 h (e) 7 days, (f) 15 days.

Figure S31. (a) ESI-MS of porphyrin adducts of S2 with different times. (a) 0 h, (b) 6 h, (c) 12 h, (d) 24 h (e) 7 days, (f) 15 days.

7. TEM images of supramolecules (NTf₂⁻ as counterion)

Figure S32. TEM images of S1 on the lacey carbon coated Cu grid.

Figure S33. TEM images of S2 on the lacey carbon coated Cu grid.

Figure S34. TEM images of nanofibers assembled by S1.

Figure S35. (a) (b) TEM images of nanofibers assembled by S2 (2 mg/ml in DMF solution under ethyl ether vapor); (c) the probable stacking approaches of S2 (stacking with square surfaces as contact surfaces).

Figure S36. The TEM images of the same S2 sample captured under different areas, which may reveal the possible growing processes of the nanofibers for S2.

Figure S37. (a) Element mapping of selected areas. (b) TEM-EDX spectrum of the nanowires of S2 collected on a lacey carbon covered Cu grid.⁹

Table S1. Atomic ratios of the elements in the nanowires of S2 collected on a lacey carbon covere
Cu grid by TEM-EDS

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element	k	Absorption	wt%	wt% Sigma	Element
С	2.74533	1.00	80.10	0.52	46.32
Ν	3.46969	1.00	4.57	0.48	8.31
0	2.00298	1.00	4.61	0.22	9.49
F	1.70760	1.00	0.79	0.12	16.91
S	0.99241	1.00	6.64	0.17	11.89
Zn	1.32169	1.00	3.28	0.15	7.08
Total amount:			100.00		

Figure S38. (a) Element mapping of selected areas. (b) TEM-EDX spectrum of the transferred nanowires collected on a lacey carbon covered Cu grid.

Table S2. Atomic ratios of the elements in the nanowires of S2 collected on a lacey carbo	n covered
Cu grid by TEM-EDS	

element	k	Absorption correction	wt%	wt% Sigma	Element analysis %
С	2.74533	1.00	84.95	0.50	45.24
Ν	3.46969	1.00	3.40	0.46	8.12
Ο	2.00298	1.00	5.09	0.21	11.59
F	1.70760	1.00	0.56	0.10	16.51
S	0.99241	1.00	3.36	0.12	11.61
Zn	1.32169	1.00	2.64	0.13	6.93
Total amount:			100.00		

8. Fluorescence emission measurement

Figure S39. Fluorescence spectrum ($\lambda_{ex} = 340$ nm, c = 1.0 μ M) of ligands L1, L2 and complexs S1, S2.

Figure S40. (a) (b) Fluorescence spectrum of **L1** and **L2** in different poor solvents (60%) ($\lambda_{ex} = 340$ nm, c = 1.0 μ M) (1,4 dioxane, ethyl acetate, methanol, tert-butanol, tetrahydrofuran, toluene); (c) (d) Photographs of florescence changes of **L1** and **L2**; (e) (f) Fluorescence spectrum of **S1** and **S2** in different poor solvents (60%) ($\lambda_{ex} = 340$ nm, c = 1.0 μ M) (1,4 dioxane, chloromethane, ethyl acetate, water, methanol, tert-butanol, tetrahydrofuran, toluene); (g) (h) Photographs of fluorescence changes of **S1** and **S2**.

Figure S41. Fluorescence spectrum of (a) L1 (λ ex = 340 nm, c = 1.0 μ M) at PMT Voltage 500 V and (b) L2 (λ ex = 340 nm, c = 1.0 μ M) at PMT Voltage 400 V with various methanol content.

Figure S42. Fluorescence spectrum of (a) **S1** ($\lambda_{ex} = 340$ nm, c = 1.0 μ M) at PMT Voltage 500 V and (b) **S2** ($\lambda_{ex} = 340$ nm, c = 1.0 μ M) at PMT Voltage 350 V with various H₂O content.

Figure S43. (a) (b) Fluorescence spectrum ($\lambda_{ex} = 340 \text{ nm}, c = 1.0 \mu\text{M}$); (c) (d) photographs of L1 in CHCl₃/1,4-dioxane and in CHCl₃/THF with various 1,4-dioxane or THF content.

Figure S44. (a) photographs of **S1** in DMF/THF with various THF content, (b) (a) photographs of **S1** in DMF/tert-butanol with various tert-butanol content.

Figure S45. (a) UV-vis (0.5 μ M in DMF) of free S2 and different amounts of porphyrin were added to S2; (b) Emission (0.5 μ M in DMF) of free S2 and different amounts of porphyrin were added to S2.

Figure S46. (a) UV-vis (0.5 μ M in DMF) of free S1 and different amounts of porphyrin were added to S1; (b) Emission (0.5 μ M in DMF) of free S1 and different amounts of porphyrin were added to S1.

Figure S47. (a) UV-vis (0.5 μ M in DMF) of free S1 and different amounts of zinc porphyrin were added to S1; (b) Emission (0.5 μ M in DMF) of free S1 and different amounts of zinc porphyrin were added to S1.

9. Crystallographic date of ligands

Figurer S48. The crystal structure photograph of (a) (b) S1 and (c) (d) S2.

Figure S49. The diffraction points of the S2 crystal.

Identification	Compound I 1	Compound L2		
code				
CCDC number	2132604	2132525		
Empirical	CocHeeNiaSo	$C_{96}H_{60}N_{12}O_4S_2$		
formula	C9611601N12S2			
Formula weight	1470.70	1504.73		
Temperature/K	100(2) K	100(2) K		
Crystal system	triclinic	monoclinic		
Space group	P-1	Cm		
	<i>a</i> = 13.8636(3) Å	<i>a</i> = 9.1813(8) Å		
	<i>b</i> = 15.9137(3) Å	<i>b</i> = 32.2736(14) Å		
Unit cell	c = 20.4987(4) Å	c = 40.651(3) (4) Å		
dimensions	$\alpha = 83.865(2)^{\circ}$	$\alpha = 90^{\circ}$		
	$\beta = 73.294(2)^{\circ}$	$\beta = 96.354(8)^{\circ}$		
	$\gamma = 85.019(2)^{\circ}$	$\gamma = 90^{\circ}$		
Volume	4299.28(16) Å	11971.5(15)		
Ζ	2	154		
Density	1.126 a/m^3	1.604 c/m^3		
(calculated)	1.150 g/m	1.004 g/m		
Absorption	$0.115 mm^{-1}$	0.767 mm^{-1}		
coefficient	0.115 11111	0.707 mm		
<i>F</i> (000)	1530.0	5852.0		
Crystal size	$0.2 \times 0.05 \times 0.05 \text{ mm}^3$	$0.4 \times 0.08 \times 0.08 \text{ mm}^3$		
Padiation	ΜοΚα	ΜοΚα		
Radiation	$(\lambda = 0.71073)$	$(\lambda = 0.71073)$		
Theta range for	1 321 to 62 16°	1 638 to 62 318°		
data collection	4.524 10 02.40	4.038 10 02.348		
	<i>−</i> 19 <i>≤h≤</i> 19,	-11≤ <i>h</i> ≤12,		
Index ranges	<i>−</i> 22≤ <i>k</i> ≤22,	-43≤k≤43,		
	-29 <i>≤l≤</i> 28	-57 <i>≤l</i> ≤54		
Reflections	70881	76442		
collected	/0001	/0442		

 Table S3. Crystallographic data of L1, L2

Independent	22440 [$R_{int} = 0.0470, R_{sigma} =$	28020 [$R_{int} = 0.1854, R_{sigma} =$
reflections	0.0635]	0.2404]
Data / restraints	22440/0/1004	28020//50/882
/ parameters	22440/0/1004	28020/650/883
Goodness-of-fit on F^2	1.358	2.242
Final R indexes	$R_1 = 0.1209,$	$R_1 = 0.2743,$
$[I \ge 2\sigma(I)]$	$wR_2 = 0.3490$	$wR_2 = 0.5503$
Final R indexes	$R_1 = 0.1655,$	$R_1 = 0.4547,$
[all data]	$wR_2 = 0.3759$	$wR_2 = 0.5918$
Largest diff. peak/hole	1.95/-0.41 e•Å ⁻³	3.03/-0.86 e•Å ⁻³
Flack parameter	-0.07(8)	-0.09(9)

Figure S50. ORTEP Drawing of L1. The thermal ellipsoids are drawn at 50% probability

Figure S51. ORTEP Drawing of L2. The thermal ellipsoids are drawn at 50% probability

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