

## Electronic Supplementary Information

### Chiral Molecular Nanosilicas

Zhaohui Zong,<sup>a</sup> Aiyu Hao,<sup>a</sup> Pengyao Xing<sup>\*,a,b</sup> and Yanli Zhao<sup>\*b</sup>

<sup>a</sup>*Key Laboratory of Colloid and Interface Chemistry of Ministry of Education and School of Chemistry and Chemical Engineering, Shandong University, Jinan 250100, People's Republic of China.*

<sup>b</sup>*Division of Chemistry and Biological Chemistry, School of Physical and Mathematical Sciences, Nanyang Technological University, 21 Nanyang Link, Singapore 637371, Singapore.*

*Email: xingpengyao@sdu.edu.cn; zhaoyanli@ntu.edu.sg*

#### Experimental section

##### Materials

1,1'-Ferrocenedicarboxylic acid and all amino acid methyl ester hydrochlorides were purchased from HEOWNS Biochemical Technology Co., LTD, China. POSS-NH<sub>2</sub> was purchased from Xi'an Qiyue Biological Technology Co., Ltd. All reagents were used without further purification in this work. *L*-Configuration amino acids are used throughout this supporting information unless otherwise specified.

##### Characterizations

<sup>1</sup>H NMR spectra, <sup>13</sup>C NMR spectra and temperature-variable <sup>1</sup>H NMR spectra were collected by BRUKER AVANCE III HD 400. High-Resolution Mass Spectra (HR-MS) were performed on an Agilent Q-TOF 6510. Circular dichroism (CD) spectra, temperature-variable CD spectra and solid CD spectra were measured with an Applied Photophysics ChirascanV100 model. Vibrational circular dichroism (VCD) spectra were obtained by a JASCO FVS-6000. VCD testing parameters: data interval: 1.92847 cm<sup>-1</sup>; IR accumulation: 16; VCD accumulation: 6000;

resolution: 8 cm<sup>-1</sup>; scanning speed: 4 mm/sec; polarizer angle: -45°; cell length: 50 μm. Transmission electron microscope (TEM) images were measured on a HITACHI HT-7700. The samples for TEM detection were dropped in the copper grid and air-dried. Scanning Electron Microscopy (SEM) images were obtained by Carl Zeiss G300 FE-SEM System. The samples for SEM detection were dropped on the silicon wafers and air-dried. Atomic force microscopy (AFM) testing was conducted with a Bruker Bioscope Resolve and operated in tapping mode at ambient temperature. The AFM samples were dropped on the mica wafer and dried. Single crystal data were collected on a Rigaku XtaLAB Synergy. Powder X-ray diffraction (XRD) patterns were collected on a German Bruker/D8 Advanced diffractometer with Cu Kα radiation ( $\lambda = 0.15406$  nm, voltage 40 KV, current 40 mA). The samples for XRD were casted onto cover glasses (18 mm × 18 mm) and dried to form thin films.

### **Solid-state CD spectra**

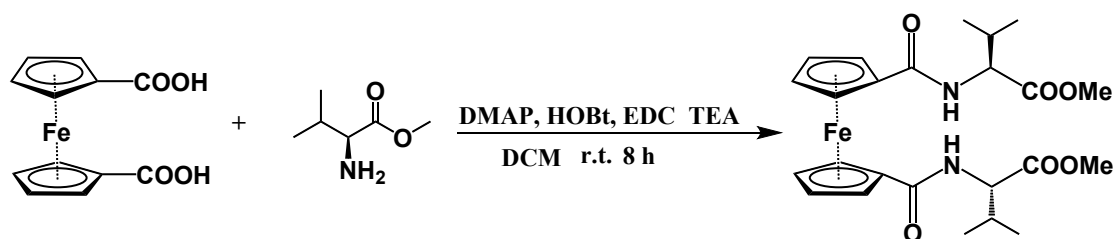
To obtain solid samples, building units were dissolved in CH<sub>2</sub>Cl<sub>2</sub>, followed by air-drying. The solid samples were then mixed in a mortar with KBr (5 mg solid sample in 200 mg KBr) under an infrared lamp. The ground mixtures were tableted into transparent tablets and tested.

### **Computational details**

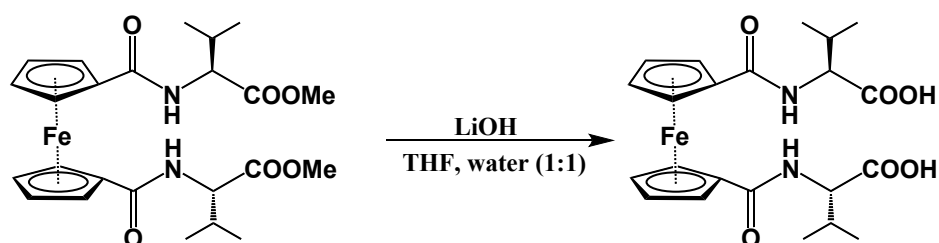
The geometric conformations of the molecules (Val-POSS, Met-POSS, Phe-POSS) used in the calculations were obtained from the corresponding single crystals. Then optimization to the energy-minimum structure in the gas phase was performed using Gaussian16 software with B3LYP/6-31g level of theory. Based on the optimized geometry, vibrational circular dichroism (VCD) spectra and electronic circular dichroism (ECD) spectra were calculated to obtain with basic set of B3LYP/6-311g. The geometric conformation of Glu-POSS was built in Gaussian View 06 and then optimized to the energy-minimum structure using Gaussian 16 software. The basis set used in the optimization process was B3LYP/6-31g. Vibrational circular dichroism (VCD) spectrum and electronic circular dichroism (ECD) spectrum were calculated based on the above pre-settings.

In the optimization process, the convergence criteria of optimization were as follows: Maximum Force 0.000015, RMS Force 0.000010, Maximum Displacement 0.000060, RMS

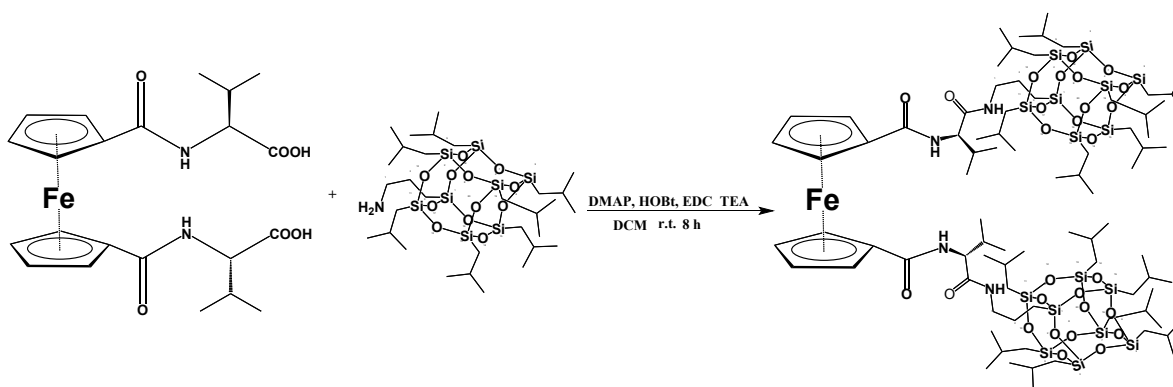
Displacement 0.000040. To ensure that the optimized geometry was at a minimum, all geometry optimizations were followed by a frequency calculation and only positive frequencies were obtained.



**Scheme S1.** Synthesis route of ferrocene diamino acid methyl ester.



**Scheme S2.** Synthesis route of ferrocene diamino acid.



**Scheme S3.** Synthesis route of Val-POSS.

## Ala-POSS

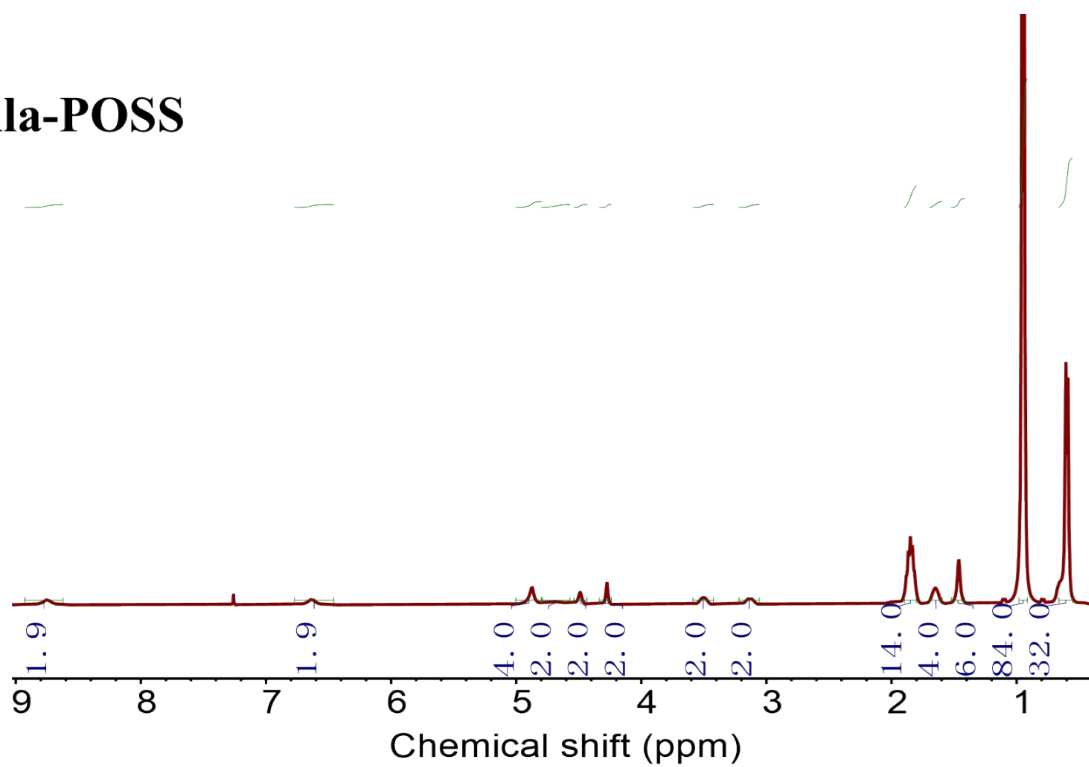


Figure S1. <sup>1</sup>H NMR spectrum of Ala-POSS.

## Val-POSS

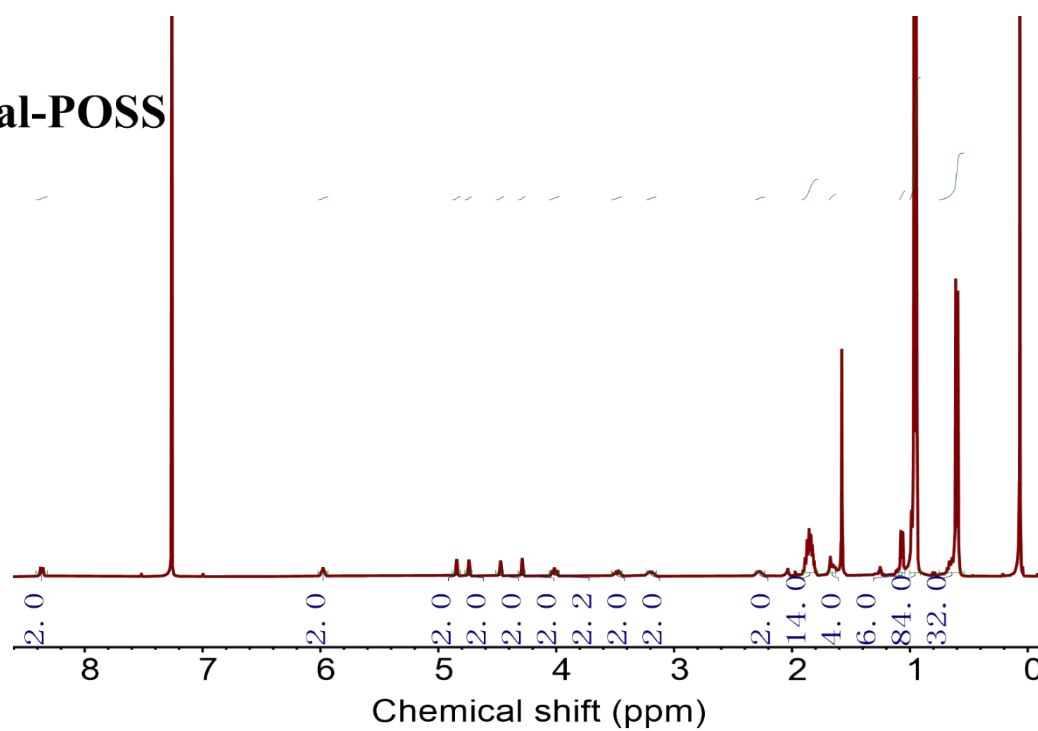


Figure S2. <sup>1</sup>H NMR spectrum of Val-POSS.

## D-Val-POSS

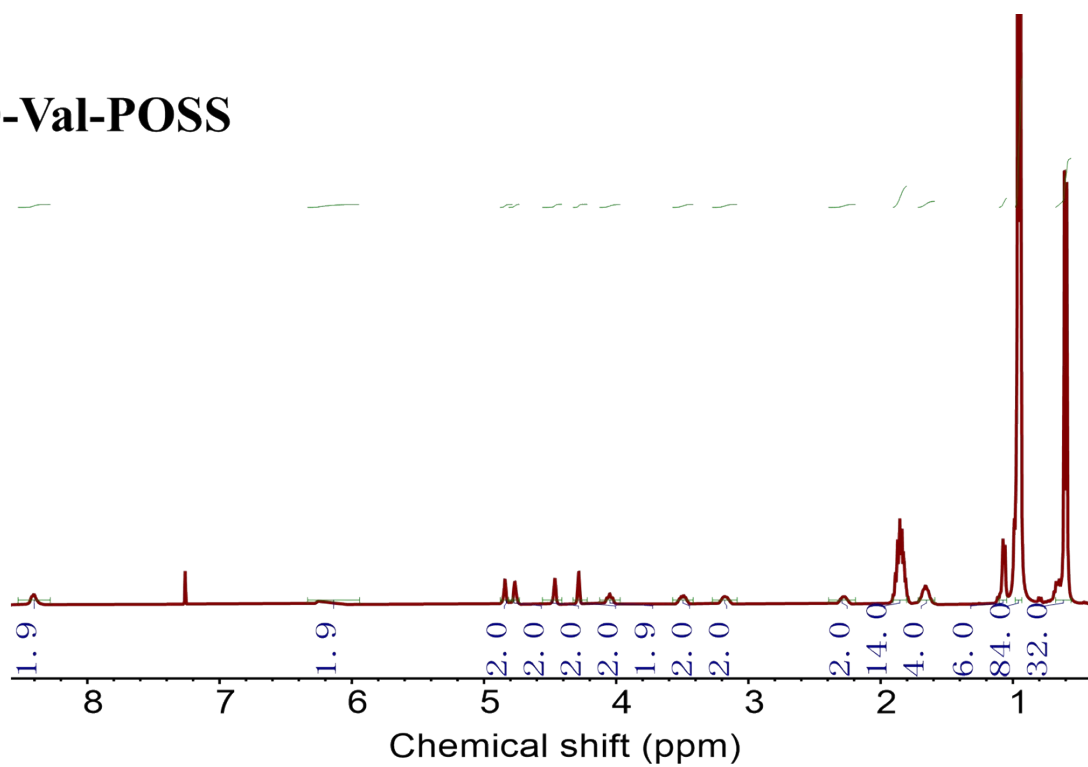


Figure S3. <sup>1</sup>H NMR spectrum of D-Val-POSS.

## Leu-POSS

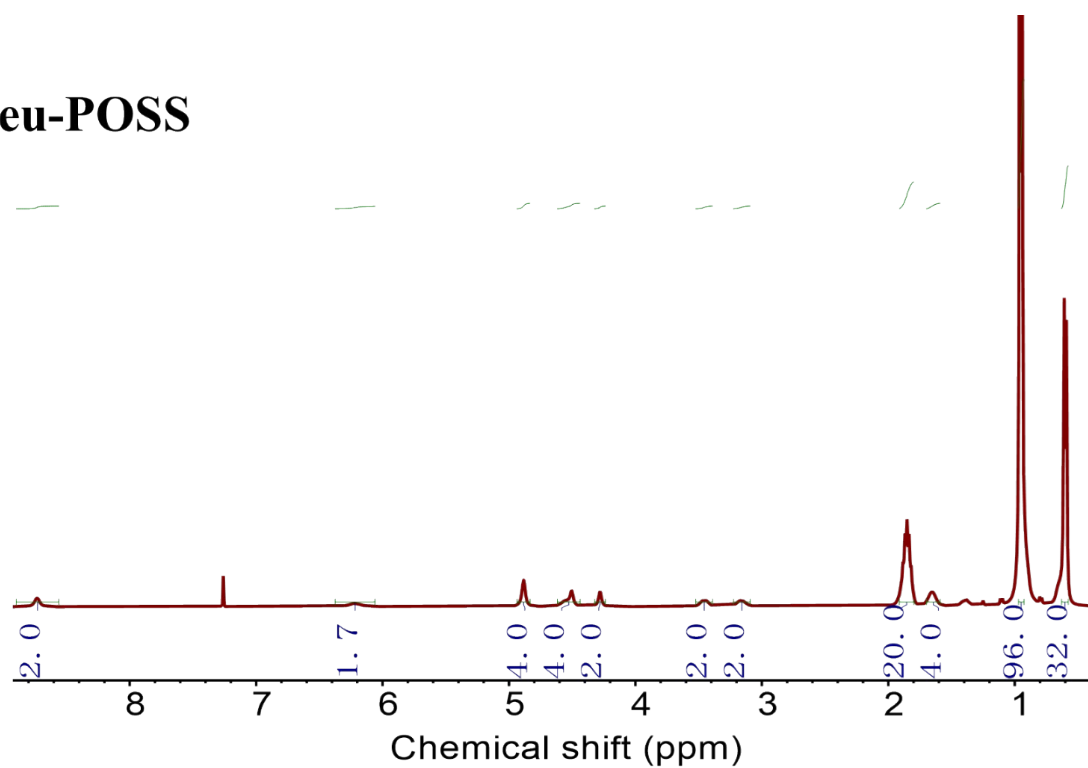


Figure S4. <sup>1</sup>H NMR spectrum of Leu-POSS.

## Pro-POSS

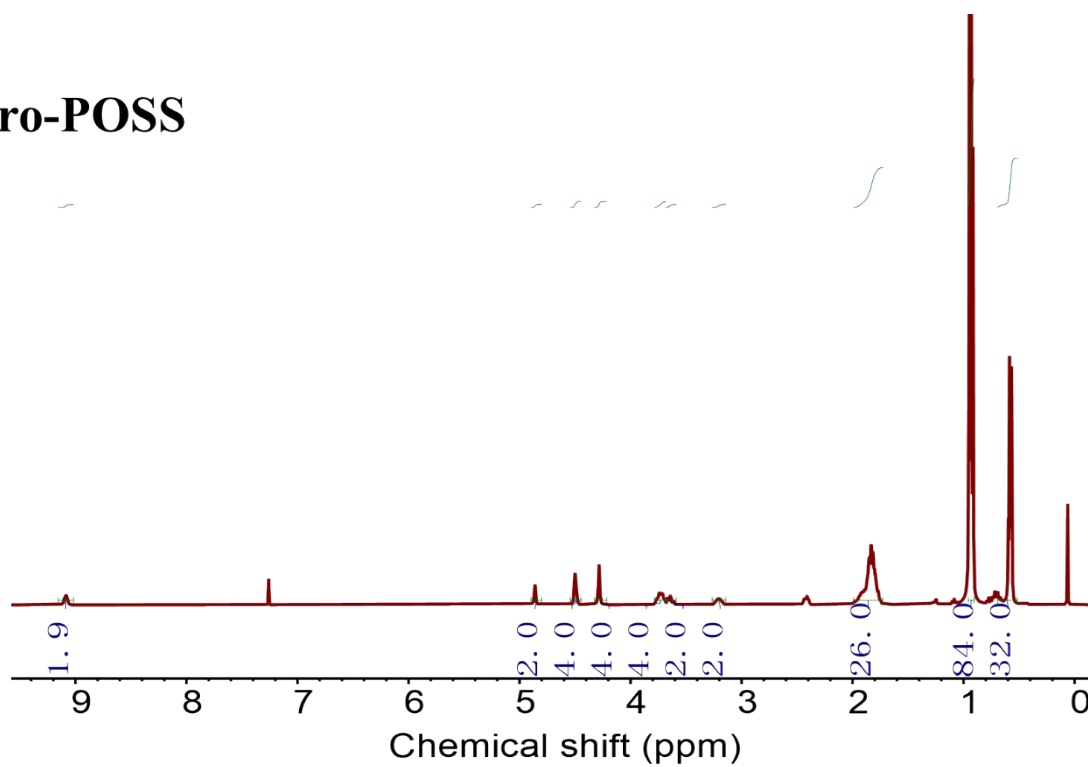


Figure S5. <sup>1</sup>H NMR spectrum of Pro-POSS.

## Met-POSS

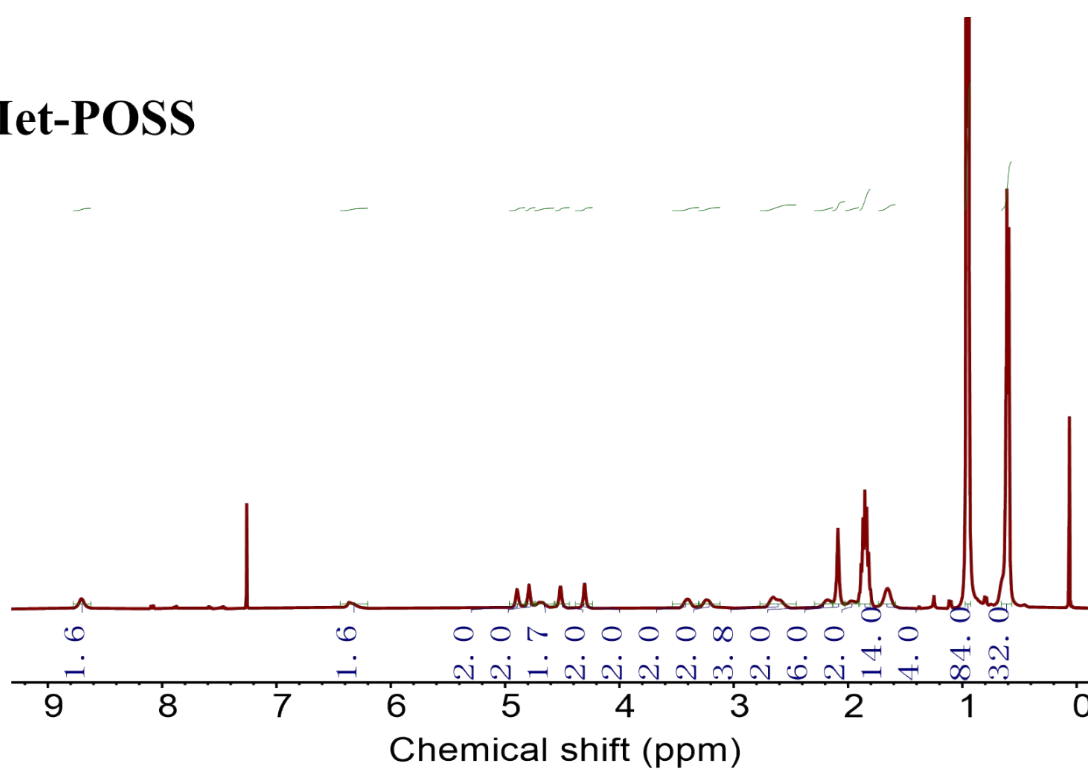


Figure S6. <sup>1</sup>H NMR spectrum of Met-POSS.

## Phe-POSS

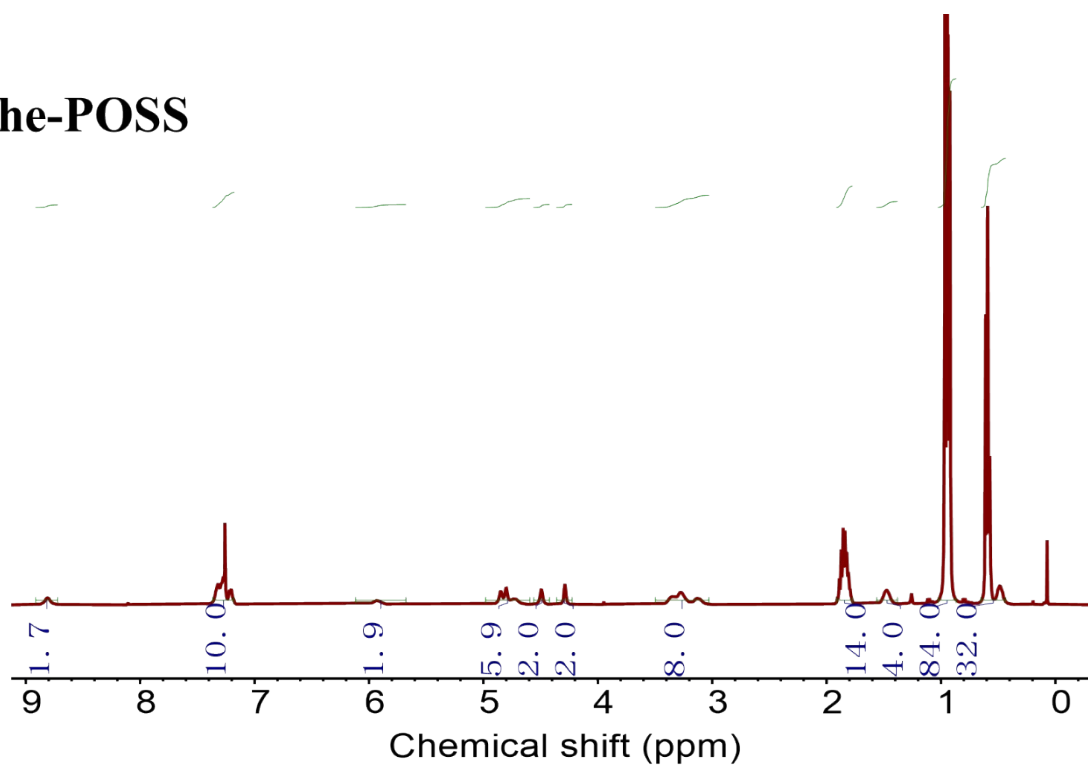


Figure S7. <sup>1</sup>H NMR spectrum of Phe-POSS.

## PGly-POSS

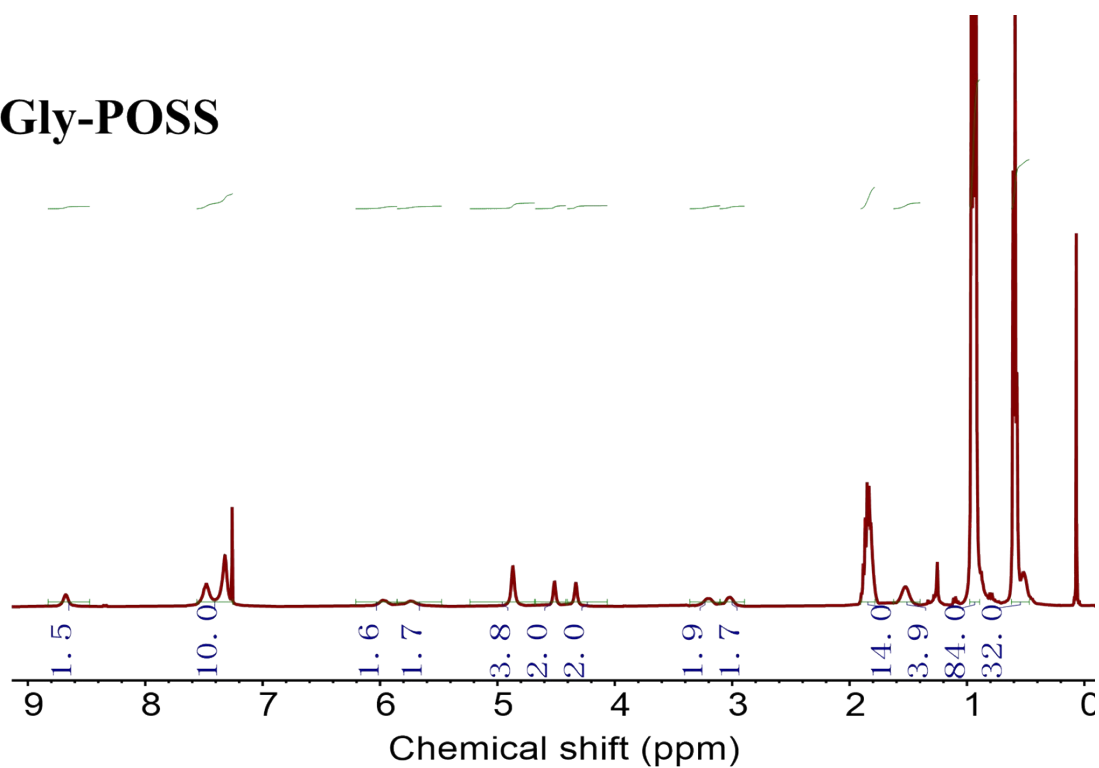
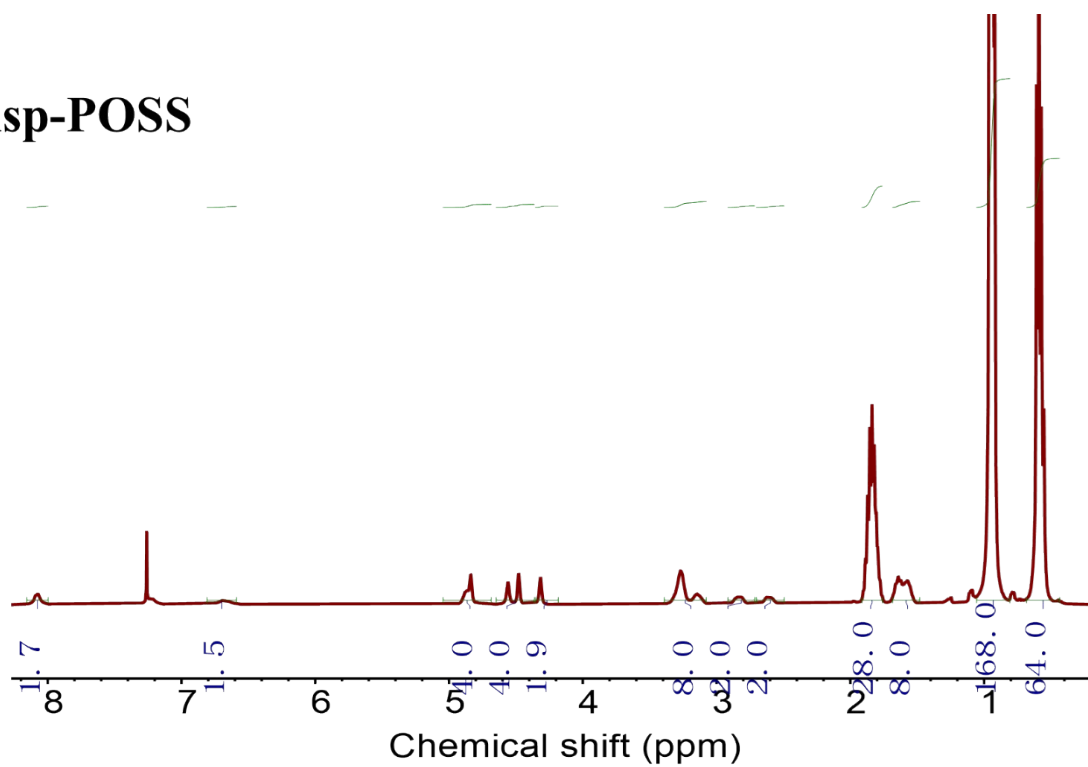


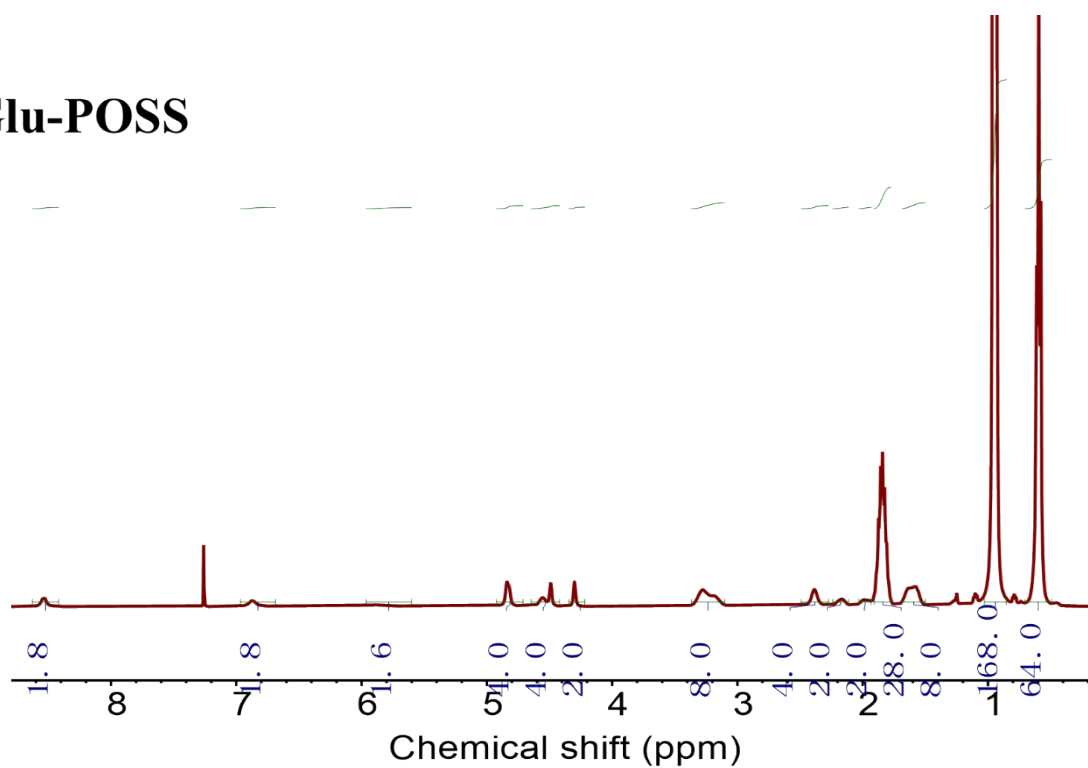
Figure S8. <sup>1</sup>H NMR spectrum of PGly-POSS.

## Asp-POSS



**Figure S9.** <sup>1</sup>H NMR spectrum of Asp-POSS.

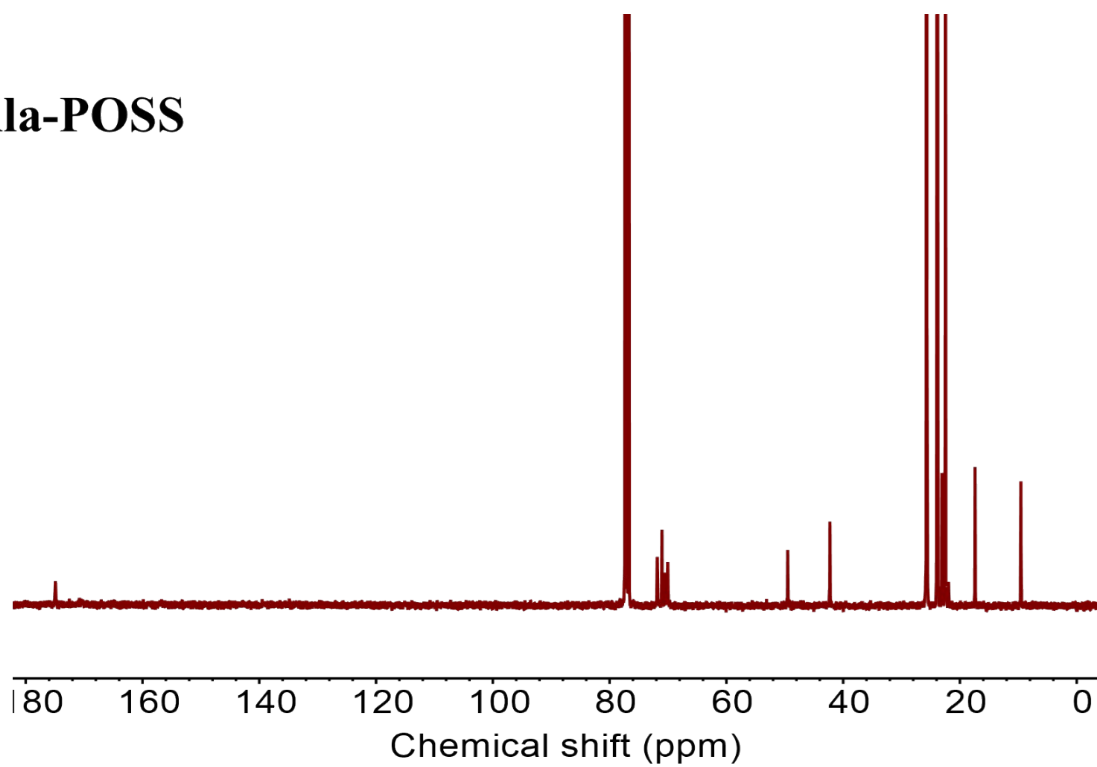
## Glu-POSS



**Figure S10.** <sup>1</sup>H NMR spectrum of Glu-POSS.

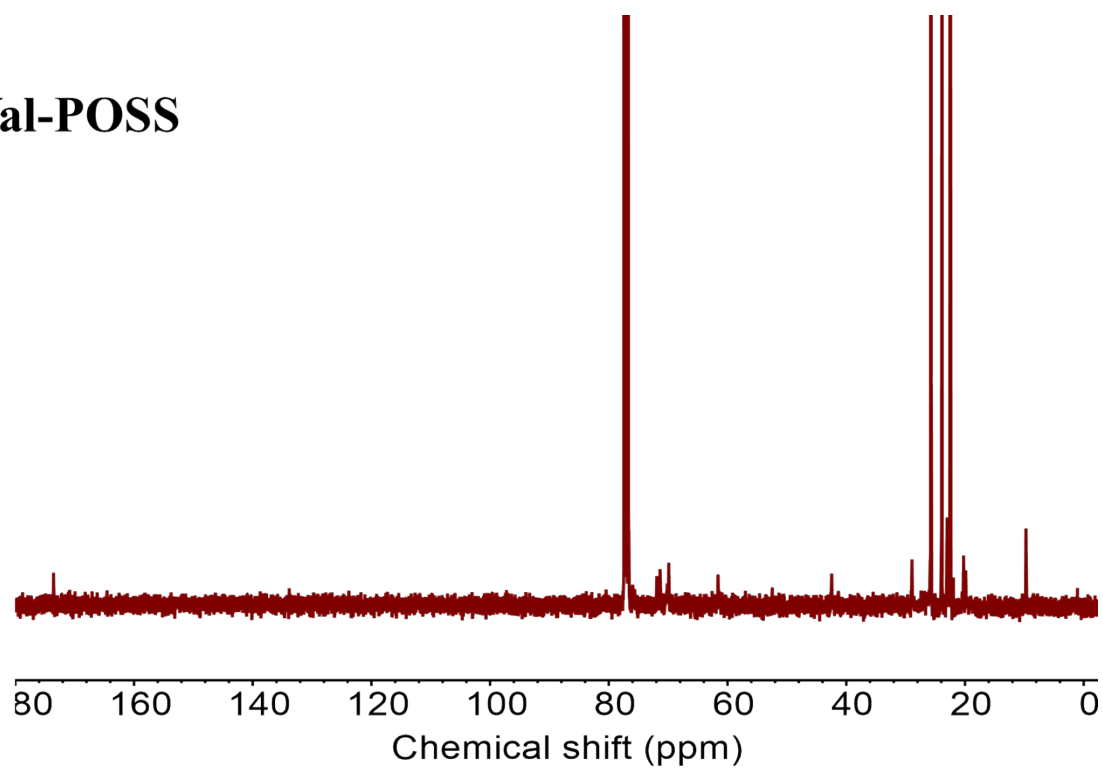


## Ala-POSS



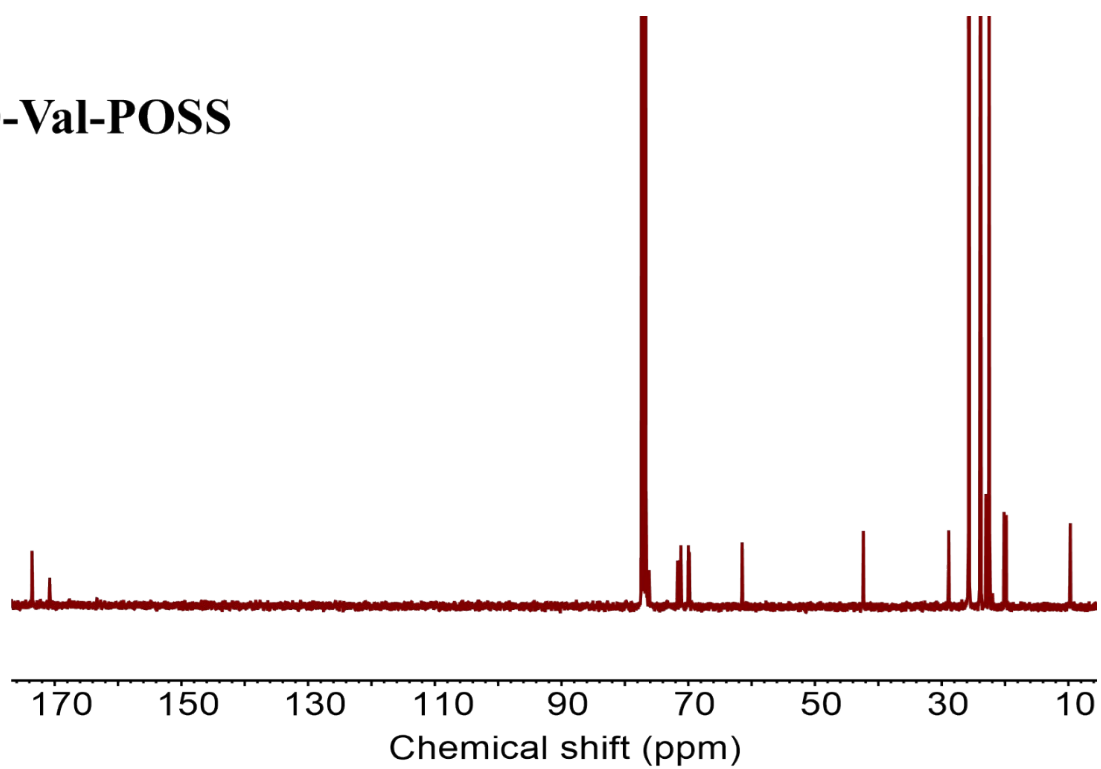
**Figure S11.**  $^{13}\text{C}$  NMR spectrum of Ala-POSS.

## Val-POSS



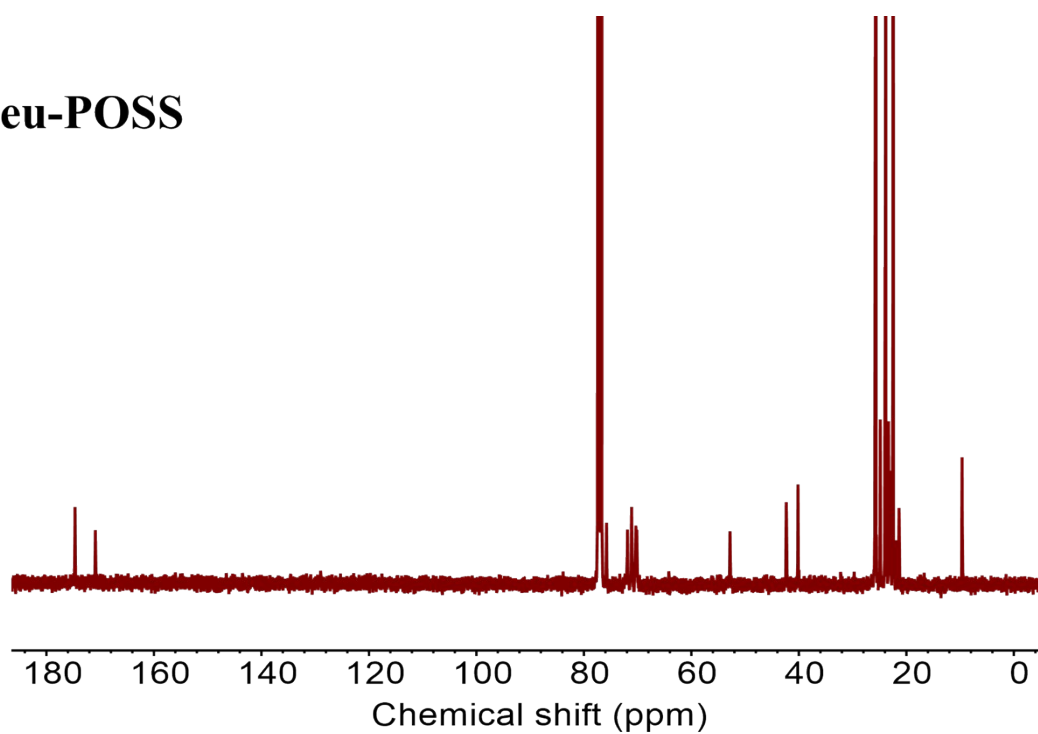
**Figure S12.**  $^{13}\text{C}$  NMR spectrum of Val-POSS.

## D-Val-POSS



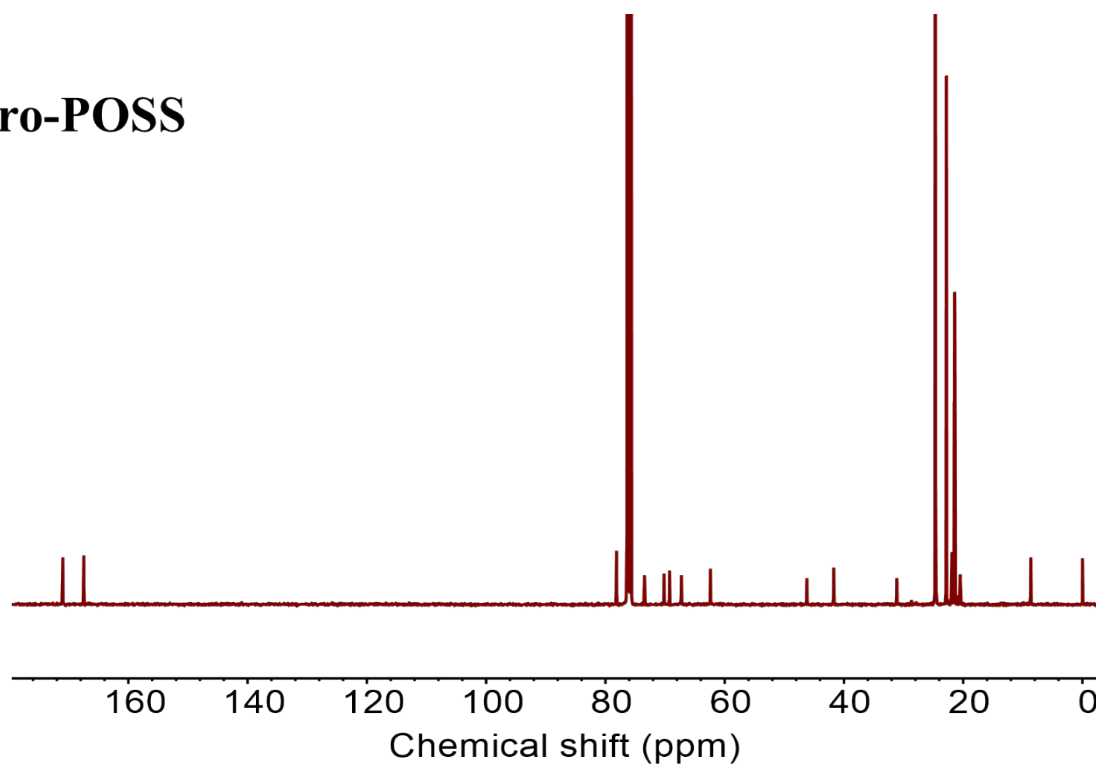
**Figure S13.**  $^{13}\text{C}$  NMR spectrum of D-Val-POSS.

## Leu-POSS



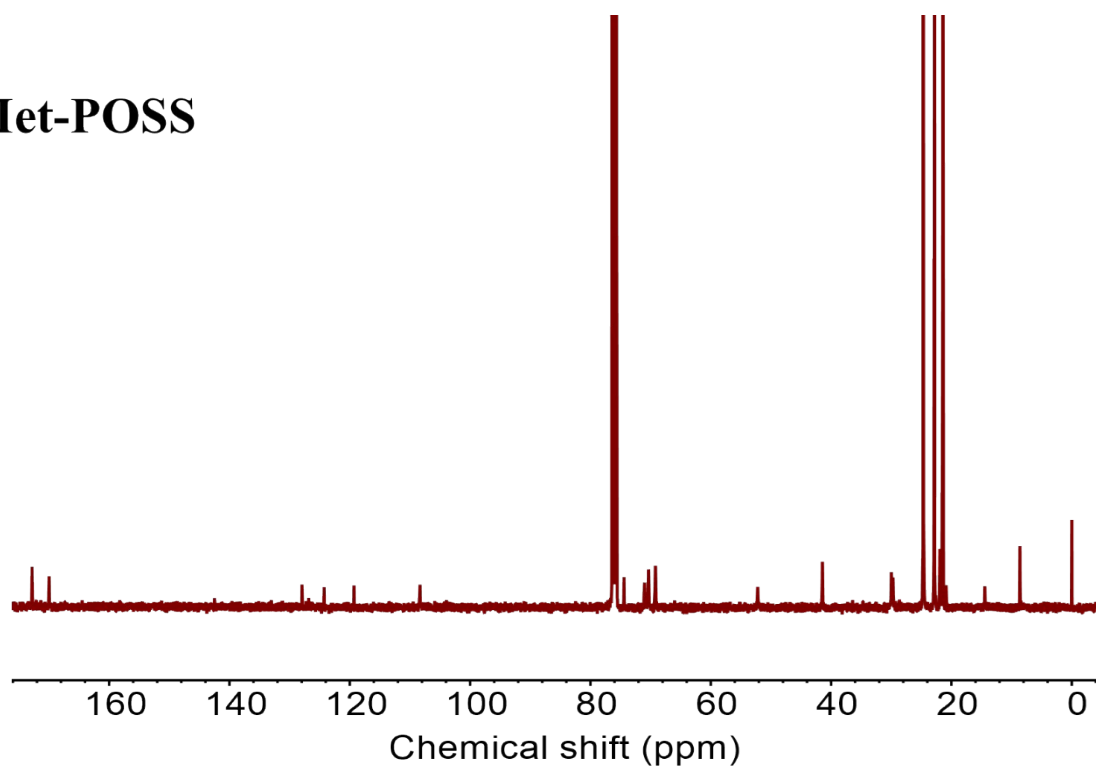
**Figure S14.**  $^{13}\text{C}$  NMR spectrum of Leu-POSS.

## Pro-POSS



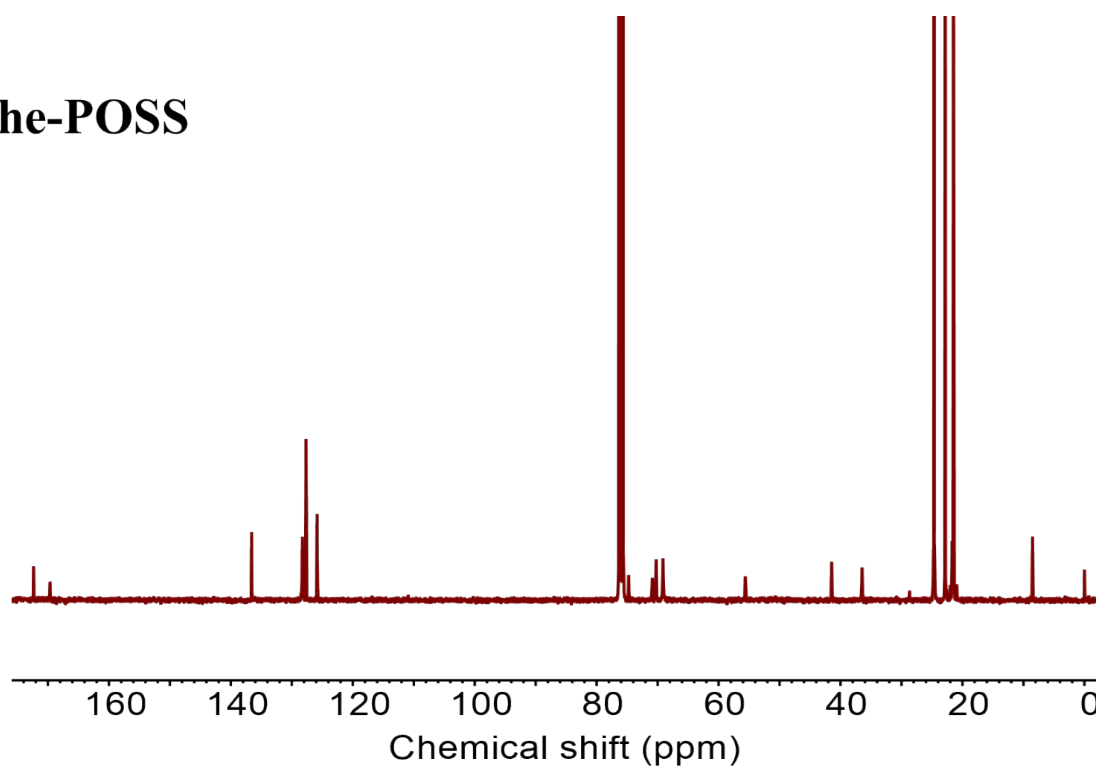
**Figure S15.**  $^{13}\text{C}$  NMR spectrum of Pro-POSS.

## Met-POSS



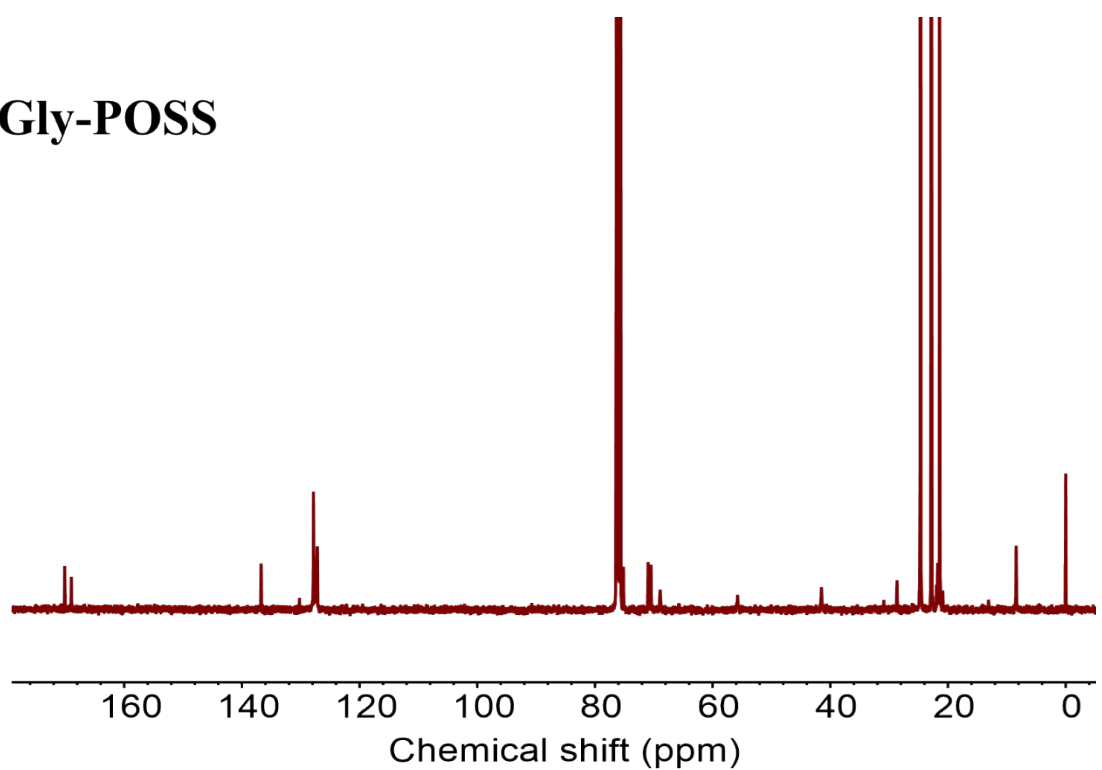
**Figure S16.**  $^{13}\text{C}$  NMR spectrum of Met-POSS.

## Phe-POSS



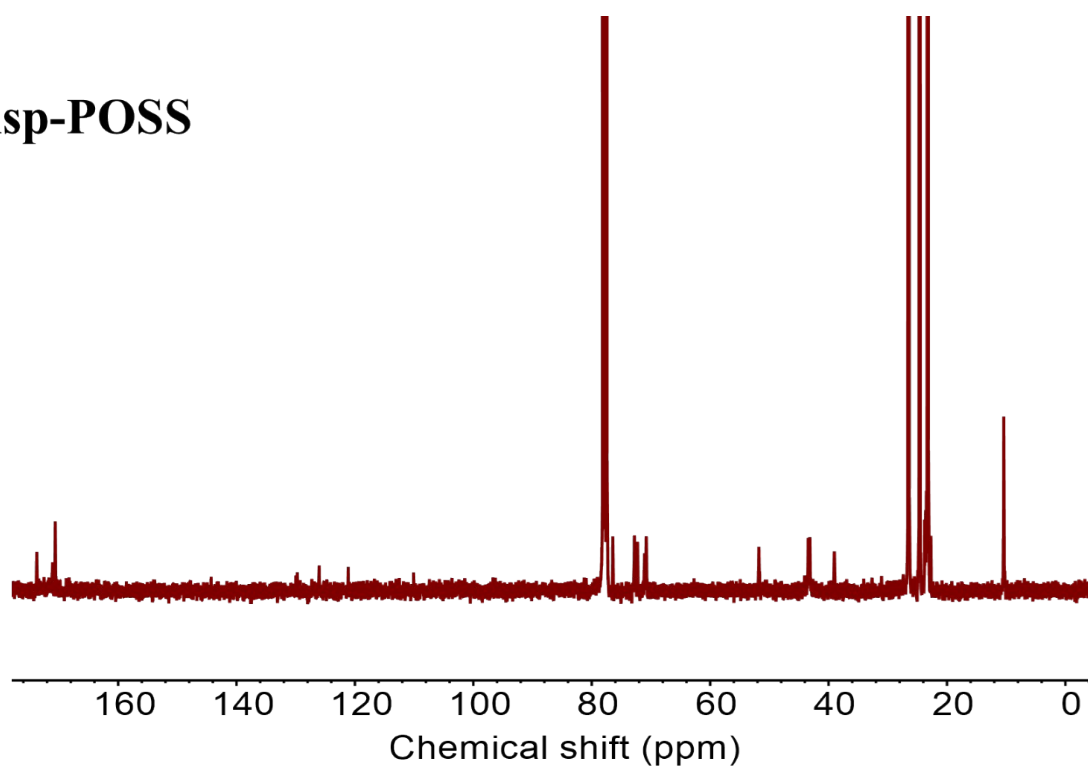
**Figure S17.**  $^{13}\text{C}$  NMR spectrum of Phe-POSS.

## PGly-POSS



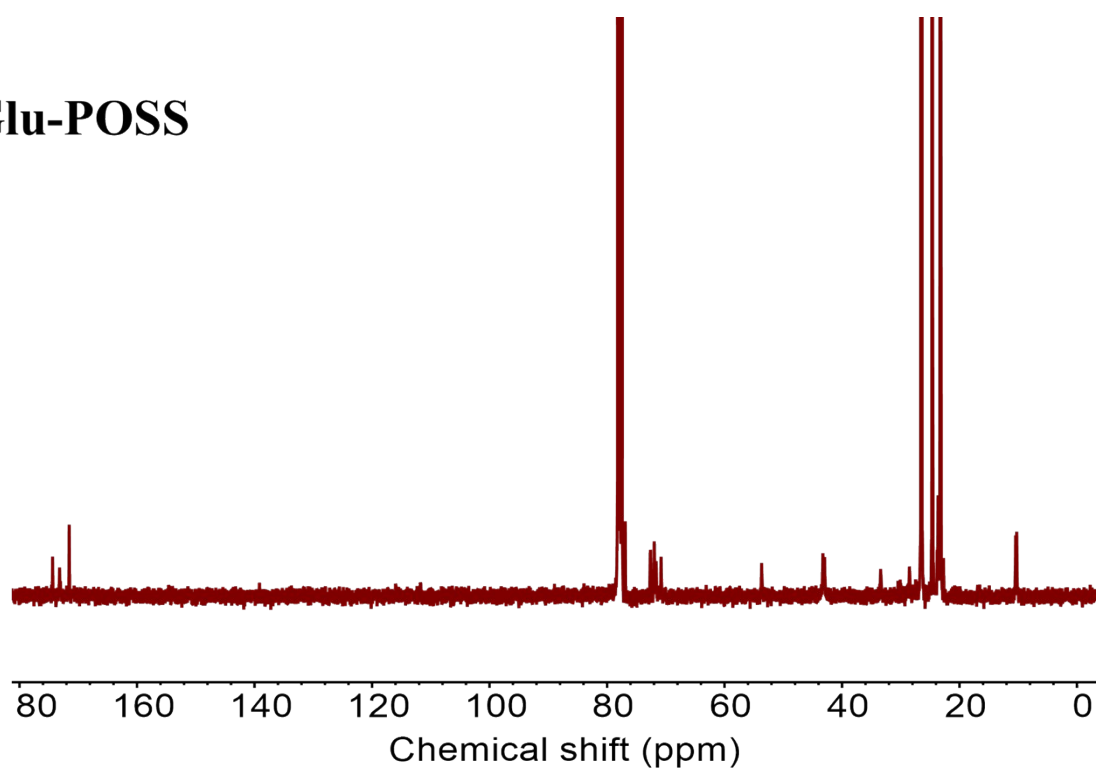
**Figure S18.**  $^{13}\text{C}$  NMR spectrum of PGly-POSS.

## Asp-POSS

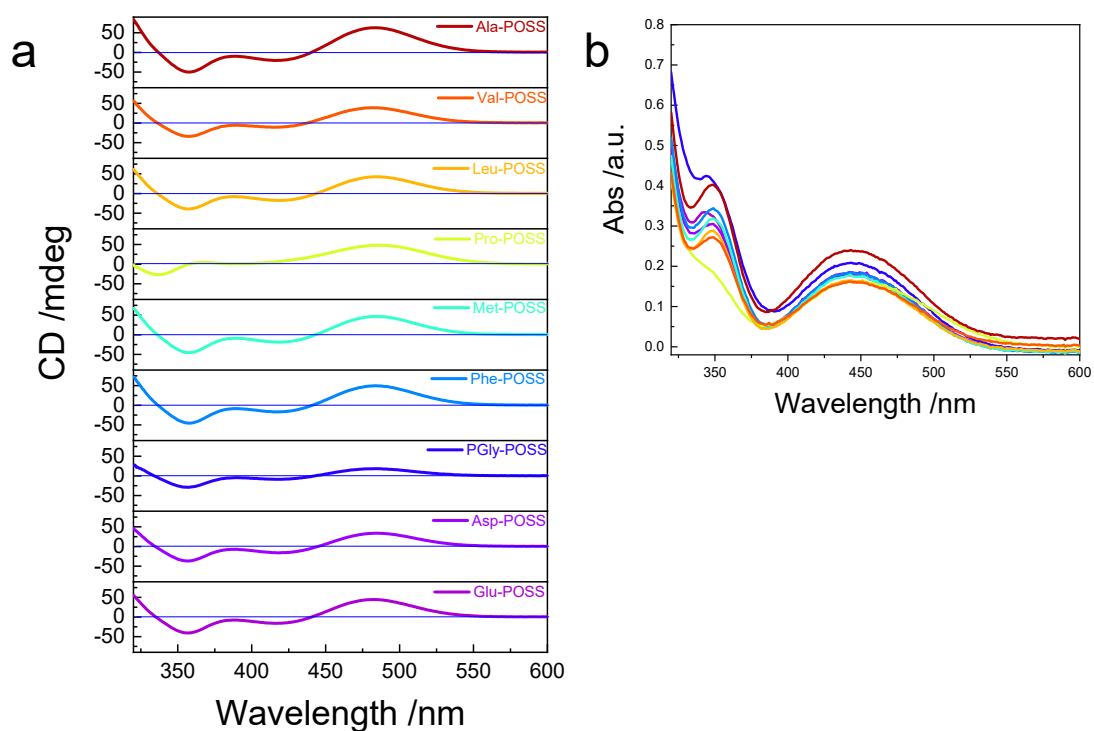


**Figure S19.** <sup>13</sup>C NMR spectrum of Asp-POSS.

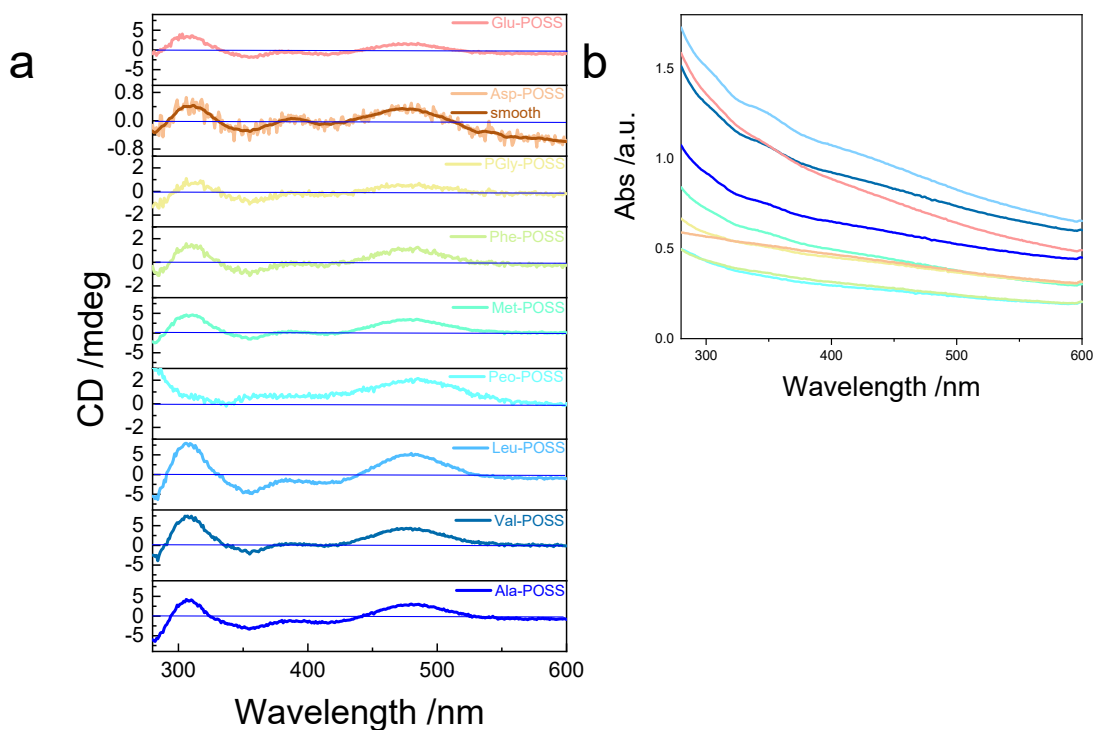
## Glu-POSS



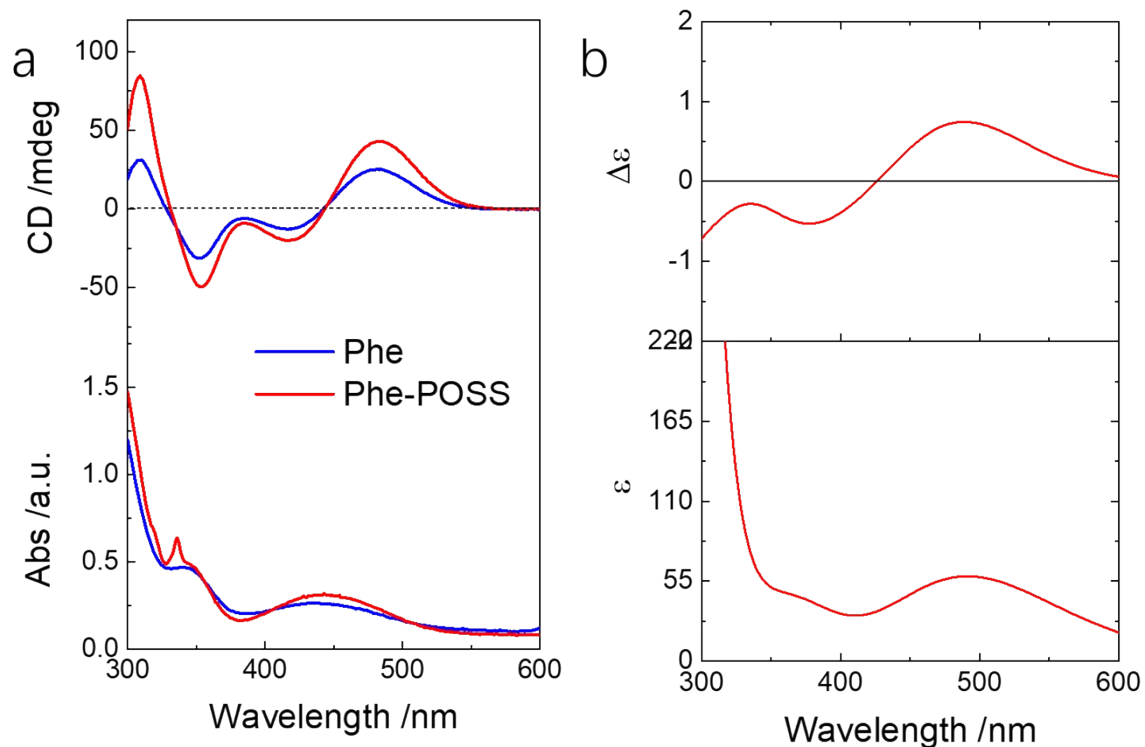
**Figure S20.** <sup>13</sup>C NMR spectrum of Glu-POSS.



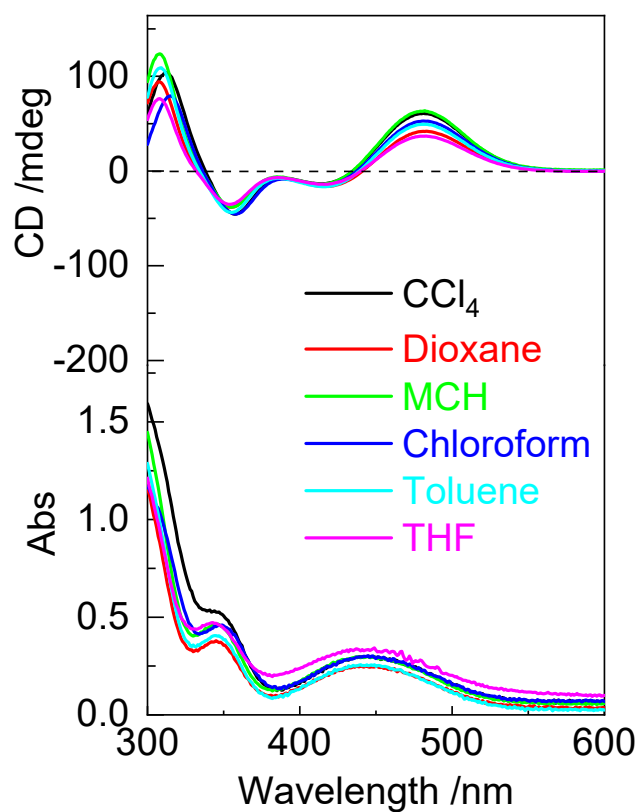
**Figure S21.** (a) CD spectra and (b) corresponding UV-vis spectra of POSS derivatives (1 mM, in  $\text{CHCl}_3$ )



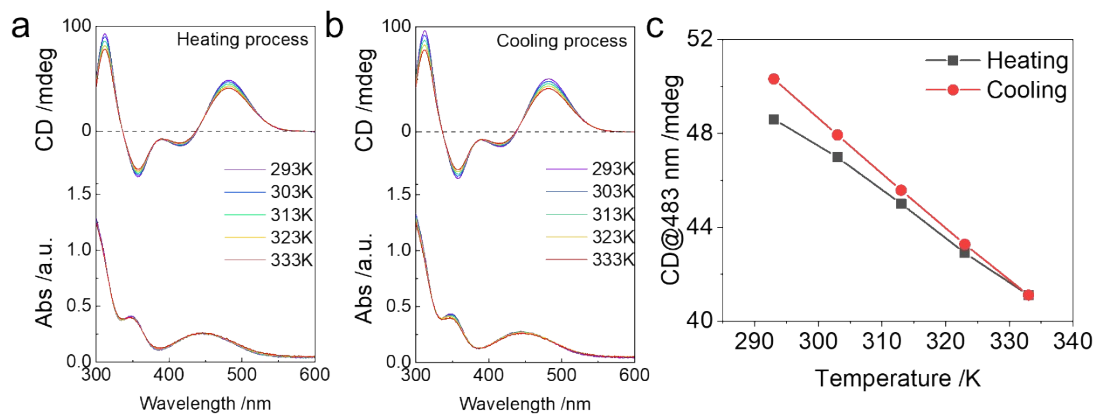
**Figure S22.** (a) Solid ECD spectra and (b) corresponding UV-vis spectra of POSS derivatives.



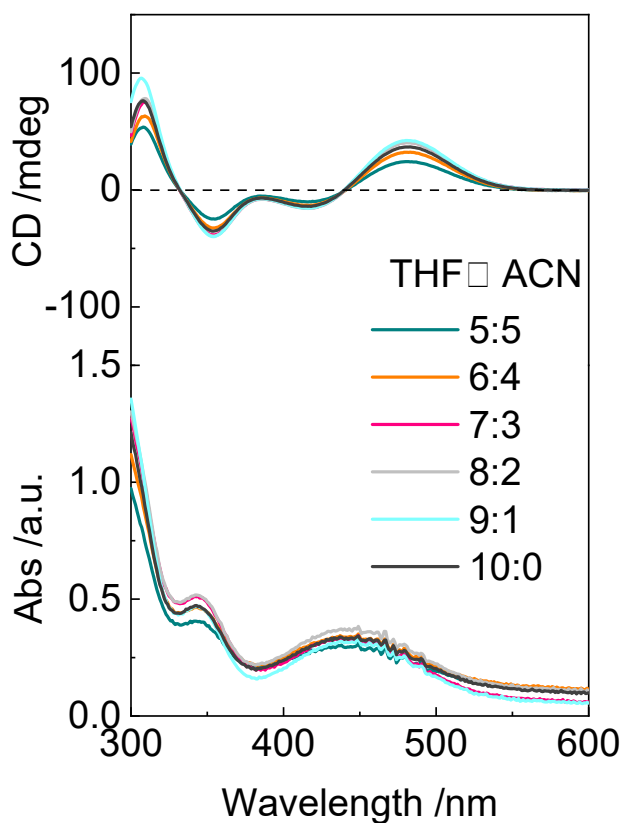
**Figure S23.** (a) Comparison of CD spectra between Phe (1,1'-ferrocenyl diphenylalanine) and Phe-POSS. (b) Calculated ECD spectra of Phe.



**Figure S24.** CD spectra of *L*-Val-POSS in different solvents (1 mM).

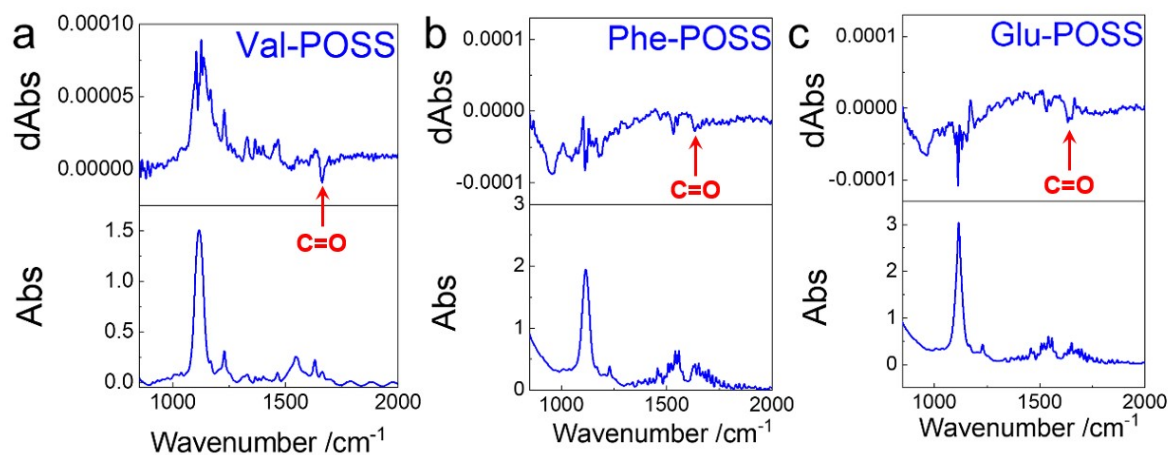


**Figure S25.** (a,b) Temperature-variable CD spectra of *L*-Val-POSS (1 mM) in the heating and cooling processes. (c) Corresponding CD intensity at 483 nm as a function of temperature.

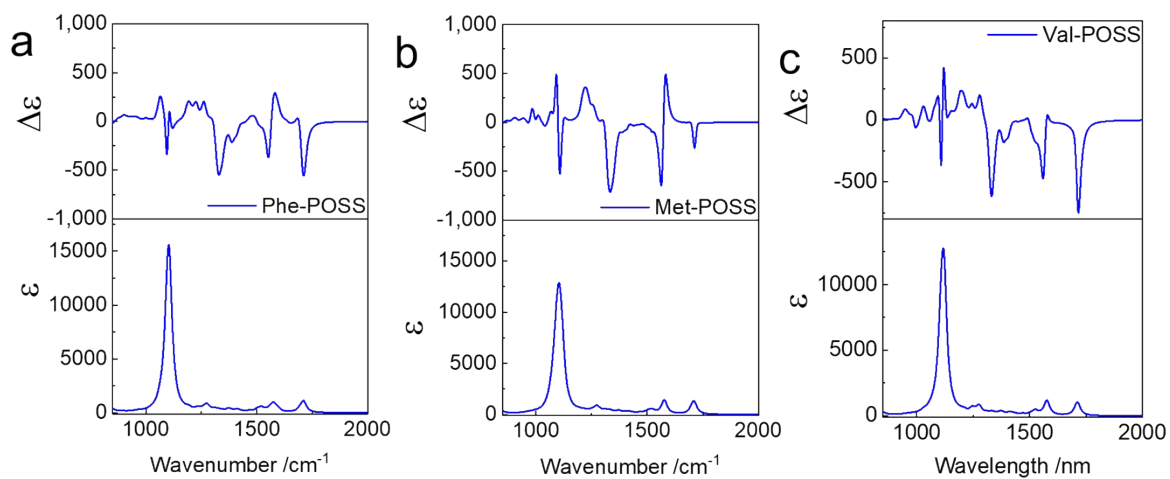


**Figure S26.** CD spectra of *L*-Val-POSS with different solvent ratios. When the fraction of ACN was higher than 50 vol%, precipitates shall be given. With increasing ACN fraction, no apparent correlation of CD signal and solvent fraction but irregularly changed Cotton effects were observed.

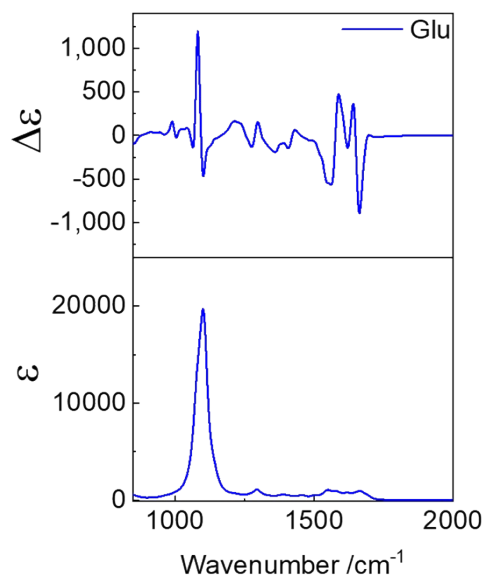




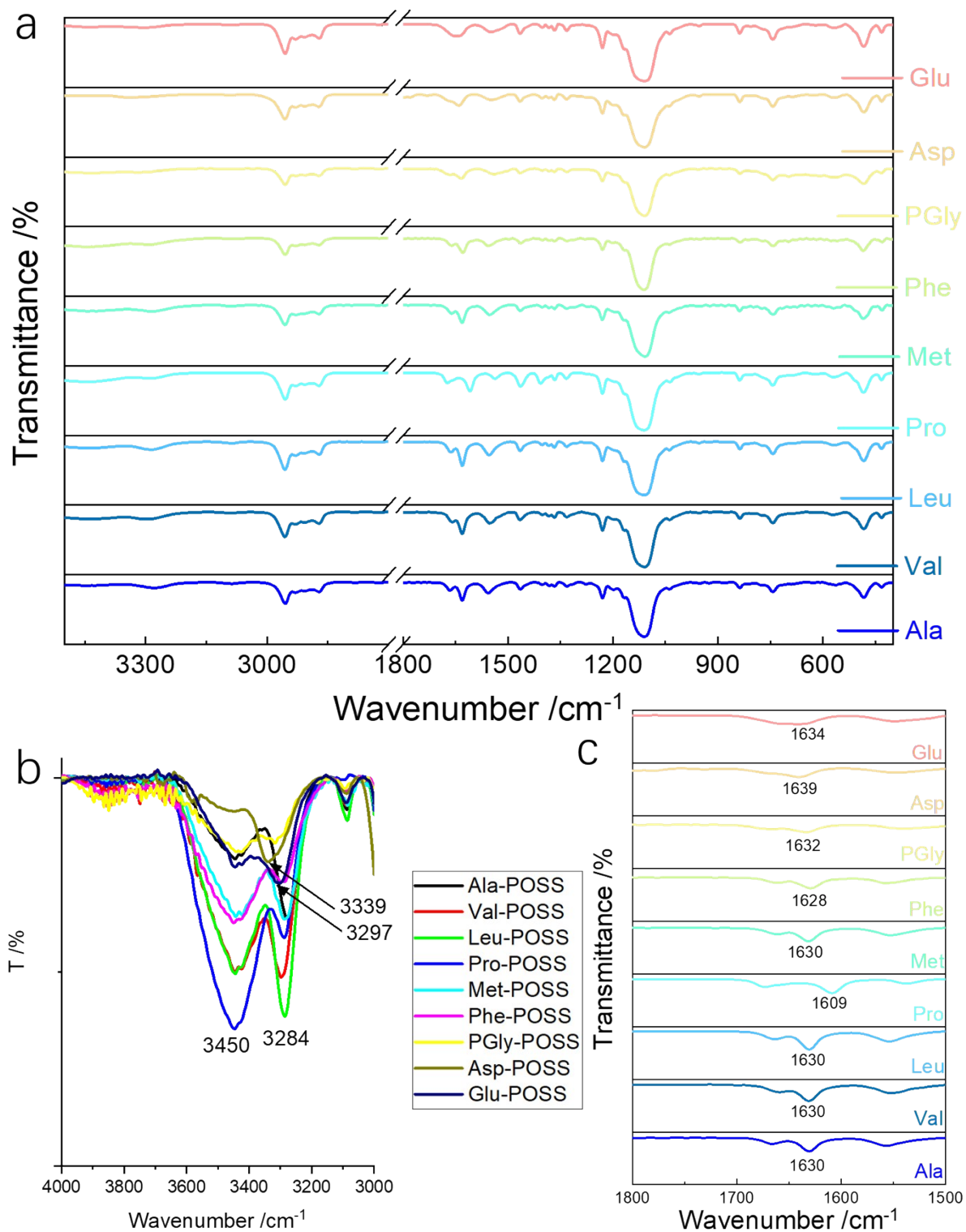
**Figure S27.** VCD spectra of (a) Val-POSS, (b) Phe-POSS, and (c) Glu-POSS (50 mg/mL in  $\text{CCl}_4$ ).



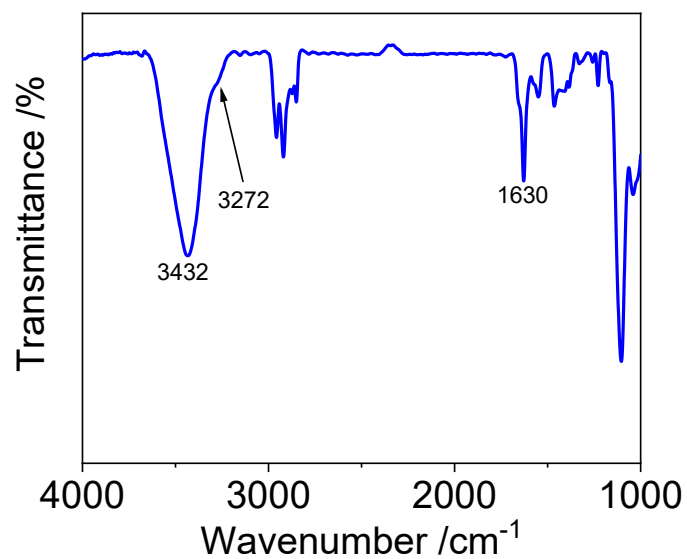
**Figure S28.** Simulated VCD spectra of (a) Phe-POSS, (b) Met-POSS and (c) Val-POSS (B3LYP/6-311g FWHM 15  $\text{cm}^{-1}$ ).



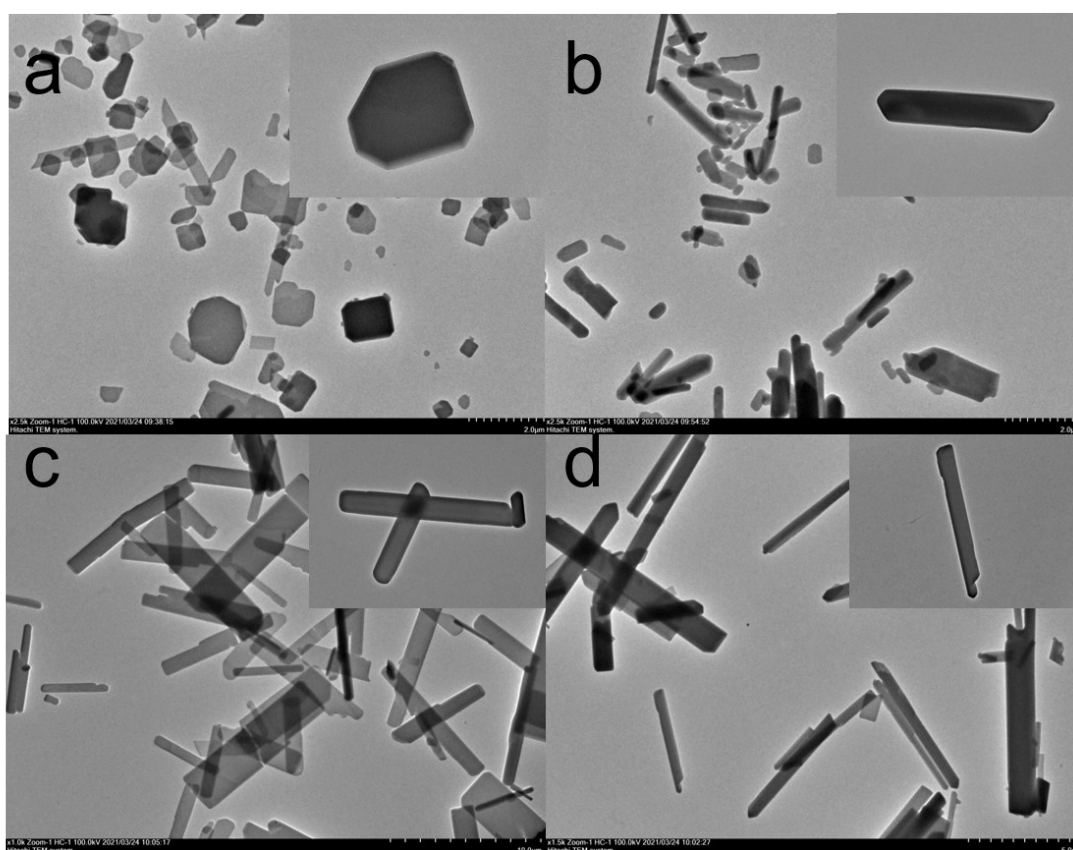
**Figure S29.** Simulated VCD spectra of Glu-POSS (B3LYP/6-31g FWHM 15 cm<sup>-1</sup>).



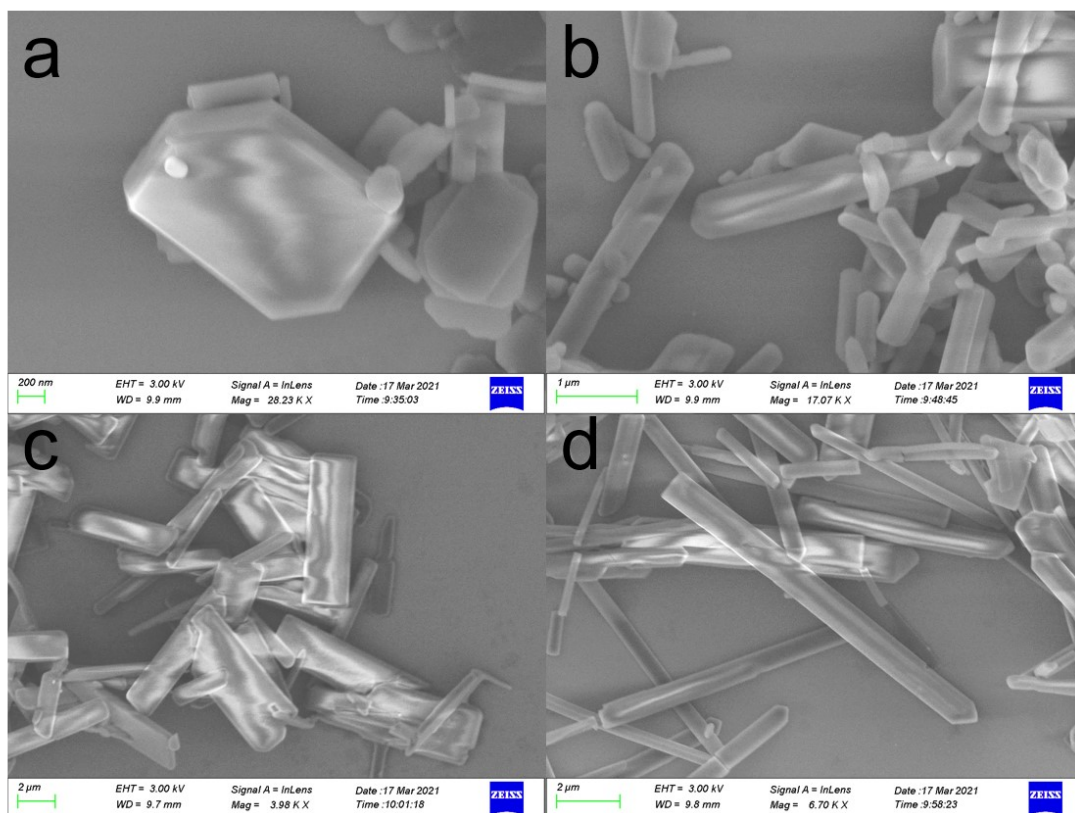
**Figure S30.** (a) FT-IR spectra of POSS derivatives. (b,c) Partial FT-IR spectra of each building units.



**Figure S31.** FT-IR spectrum of Val-POSS measured in liquid state (1 mM in CCl<sub>4</sub>).



**Figure S32.** TEM image of (a) Ala-POSS, (b) Leu-POSS, (c) Phe-POSS, and (d) Met-POSS (1 mM, THF/MeCN 1/9).



**Figure S33.** SEM image of (a) Ala-POSS, (b) Leu-POSS, (c) Phe-POSS, and (d) Met-POSS (1 mM, THF/MeCN 1/9).

**Table S1.** Crystal data of Phe-POSS.

<b>Deposition Number</b>	2096836
<b>Formula</b>	C <sub>92</sub> H <sub>166</sub> Fe N <sub>4</sub> O <sub>28</sub> Si <sub>16</sub>
<b>Temperature(K)</b>	293(2)
<b>Wavelength</b>	1.54184Å
<b>Crystal system</b>	orthorhombic
<b>Space group</b>	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
<b>a,b,c/Å</b>	<b>a</b> 16.2474(5) <b>b</b> 17.2899(8) <b>c</b> 46.6858(17)
<b>V, Å<sup>3</sup></b>	13114.8
<b>Cell angles</b>	$\alpha$ 90 $\beta$ 90 $\gamma$ 90
<b>Z, Z'</b>	Z: 4 Z': 0
<b>R-factor (%)</b>	13.09

**Table S2.** Crystal data of Val-POSS.

<b>Deposition Number</b>	2096837
<b>Formula</b>	C <sub>84</sub> H <sub>166</sub> Fe N <sub>4</sub> O <sub>28</sub> Si <sub>16</sub> ,2(C <sub>4</sub> H <sub>8</sub> O <sub>2</sub> )
<b>Temperature(K)</b>	173.00(10)
<b>Wavelength</b>	1.54184Å
<b>Crystal system</b>	monoclinic
<b>Space group</b>	P 2 <sub>1</sub>
<b>a,b,c/Å</b>	<b>a</b> 16.9205(5) <b>b</b> 17.5657(5) <b>c</b> 21.7093(5)
<b>V, Å<sup>3</sup></b>	6451.5
<b>Cell angles</b>	$\alpha$ 90 $\beta$ 90.980(2) $\gamma$ 90
<b>Z, Z'</b>	Z: 2 Z': 0
<b>R-factor (%)</b>	10.84

**Table S3.** Crystal data of Met-POSS.

<b>Deposition Number</b>	2096838
<b>Formula</b>	$C_{80} H_{156} Fe N_4 O_{27} S_2 Si_{16}$ , $C_{71} H_{136} Fe N_4 O_{28}$ $S Si_{16}$ , $C_2 H_6 S$
<b>Temperature(K)</b>	173.01(10)
<b>Wavelength</b>	1.54184Å
<b>Crystal system</b>	monoclinic
<b>Space group</b>	C2
<b>a,b,c/Å</b>	<b>a</b> 49.9932(14) <b>b</b> 17.2234(3) <b>c</b> 37.4731(9)
<b>V, Å<sup>3</sup></b>	26797.5
<b>Cell angles</b>	$\alpha$ 90 $\beta$ 123.849(3) $\gamma$ 90
<b>Z, Z'</b>	Z: 8 Z': 0
<b>R-factor (%)</b>	13.41