# **Electronic Supplementary Information**

# Anion-coordination-driven single-double helix switching and chiroptical molecular switch based on oligoureas

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#### S1. General Methods

All the reagents were obtained from commercial suppliers and used as received unless otherwise indicated. The solvents and other reagents were of reagent grade and purchased commercially. <sup>1</sup>H NMR spectra were obtained by using Bruker AVANCE III-400 MHz spectrometers under 298 K. <sup>1</sup>H NMR chemical shifts were reported relative to residual solvent peaks (2.50 *ppm* for DMSO-*d6*). The mass spectra of ligands and complexes were measured with a Bruker microTOF-Q II ESI-Q-TOF LC/MS/MS spectrometer. Circular Dichroism (CD) spectra and corresponding ultraviolet absorption spectra were recorded on a J-1500 spectropolarimeter (Jasco, Japan) at 298 K, using a 3 cm quartz cuvette. X-ray diffractions were recorded on a Bruker D8 Venture Photon II diffractometer and BL17B macromolecular crystallography beamline.

#### S2. Synthetic Procedures of Oligourea Ligands



Scheme S1. Synthetic scheme of preparing tetra-urea chiral ligand  $L^1$ .



**1a**: 1,2-Bis-(2-nitrophenyl-urea)-benzene. 1,2-diaminobenzene (1.0 g, 9.2 mmol) dissolved in tetrahydrofuran (THF, 200 mL) was added to a solution of *o*-nitro-phenylisocyanate (3.3 g, 20.0 mmol) in 80 mL THF. The precipitate thus formed was filtered

off and washed several times with toluene and diethyl ether, and then dried over vacuum to get

pure **1a** as a yellow solid, yield: 90 %. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm): δ 9.75 (s, 1H, Hb), 9.18 (s, 1H, Ha), 8.25 (d, *J* = 8.4 Hz,1H, H3), 8.06 (d, *J* = 8.4 Hz, 1H, H6), 7.70 (dd, *J* = 8.0 Hz, H4), 7.60 (dd, *J* = 8.0 Hz, 1H, H5), 7.21 (d, *J* = 7.6 Hz, 1H, H2), 7.14 (dd, *J* = 7.6 Hz, 1H, H1).



**1b**: 1,2-Bis-(2-aminophenyl-urea)-benzene. Hydrazine monohydrate (5.0 mL) was added dropwise to the suspension of **1a** (2.2 g, 5.1 mmol) and Pd/C (0.20 g, 10% cat.) in ethanol (200 mL) under stirring overnight. Whereafter, the solid was filtered off *via* suction filtration and then dissolved in a small amount of

dimethylformamide (DMF) and filtered through Celite to remove Pd/C. Diethyl ether (200 mL) was poured into the DMF solution after which the obtained precipitate was filtered off, washed several times with ethanol and diethyl ether and dried over vacuum, and finally to give **1b** as a white solid (1.3 g, 3.5 mmol, 69 %). <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  8.14 (s, 1H, Ha), 8.10 (s, 1H, Hb), 7.57 (dd, J = 6.0 Hz, 1H, H1), 7.32 (d, J = 8.0 Hz, 1H, H2), 7.06 (dd, J = 5.2 Hz, 1H, H5), 6.85 (dd, J = 7.2 Hz, 1H, H4), 6.75 (d, J = 8.0 Hz, 1H, H3), 6.57 (d, J = 8.0 Hz, 1H, H6), 4.81 (s, 2H, Hc).



Tetra-urea L<sup>1</sup>: (S)-(-)- $\alpha$ -methylbenzyl isocyanate (367 mg, 2.5 mmol) was added to a DMF (5 mL) solution of compound **1b** (376 mg, 1 mmol). After stringing overnight, the resulting precipitate was filtered off and washed several times with ethanol and diethyl ether. Then, the product was dried under vacuum to yield L<sup>4S</sup> as a white solid. (584 mg), yield: 87%. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  8.39 (s, 2H, Hc, Hb), 7.87 (s, 1H, Ha), 7.62 (m, *J* = 8.0 Hz, 2H, H3, H6), 7.43 (d, *J* = 8.0 Hz, 1H, H2),

7.32 (m, 4H, H9, H10), 7.22 (m, 1H, H11), 7.10 (m, 4H, Hd, H1, H4, H5), 4.80 (m, 1H, H7), 1.36 (d, J = 7.2 Hz, 3H, H8). <sup>13</sup>C NMR (400 MHz, DMSO-d6, ppm):  $\delta$  154.93 (CO), 154.21 (CO), 145.16 (C), 133.07 (C), 131.40 (C), 129.47 (C), 128.34 (CH), 126.69 (CH), 125.94 (CH), 125.03 (CH), 124.43 (CH), 124.14 (CH), 123.97 (CH), 122.65 (CH), 122.30 (CH), 48.33 (CH), 23.13 (CH<sub>3</sub>). ESI-MS: m/z, 100%, 693.2959 [M+Na]<sup>+</sup>; 42%, 709.2638 [M+K]<sup>+</sup>.



Scheme S2. Synthetic scheme of preparing pentakis-urea chiral ligand  $L^2$ .



**2a**: To a solution of 1,3-Bis(2-aminophenyl)urea (1.2 g, 5.0 mmol) in DMF (15 mL) in N<sub>2</sub> atmosphere, a solution of *o*-nitrophenylisocyanate (1.8 g, 11.5 mmol) in THF (7 mL) was added dropwise at 70 °C. After stirring at the same temperature for 1 h, the solvents were evaporated by reduced pressure and washed for

several times with ethanol and diethyl ether. A yellow solid (2.6 g) was produced as the **2a**, yield: 92%. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  9.75 (s, 1H, NHc), 9.21 (s, 1H, NHb), 9.51 (s, 1H, NHa), 8.24 (d, *J* = 8.4 Hz, 1H, H8), 8.05 (d, *J* = 8.0 Hz, 1H, H5), 7.73 (d, *J* = 8.0 Hz, 1H, H4), 7.65 (dd, *J* = 8.0 Hz, 1H, H6), 7.48 (d, *J* = 7.6 Hz, 1H, H1), 7.16 (m, 2H, H3, H7), 7.07 (dd, *J* = 7.6 Hz, 1H, H2).



**2b**: Hydrazine monohydrate (5.0 mL) was added dropwise to the suspension of **2a** (2.2 g, 4.3 mmol) and Pd/C (0.20 g, 10% cat.) in ethanol (200 mL) under stirring 2 h. Whereafter, the solid was filtered off *via* suction filtration and then dissolved in DMF (20 ml) and filtered through Celite to remove Pd/C. Diethyl ether (200 mL)

was poured into the DMF solution after which the obtained precipitate was filtered off, washed several times with ethanol and diethyl ether and dried over vacuum. Finally, a white solid (1.5 g) was produced as the **2b**, yield: 65%. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm): 8.45 (s, 1H, NHa),

8.16 (s, 1H, NHb), 8.15 (s, 1H, NHc), 7.60 (d, *J* = 6.4 Hz, 1H, H1), 7.55 (d, *J* = 2.4 Hz, 1H, H4), 7.30 (d, *J* = 5.2 Hz, 1H, H5), 7.06 (m, 2H, H2, H3), 6.84 (dd, *J* = 7.6 Hz, 1H, H7), 6.73 (d, *J* = 5.6 Hz, 1H, H8), 6.55 (dd, *J* = 4.8 Hz, 1H, H6), 4.82 (s, 2H, NHd).



Pentakis-urea  $L^2$ : (S)-(-)- $\alpha$ -methylbenzyl isocyanate (367 mg, 2.5 mmol) was added to a DMF (5 mL) solution of compound **1b** (510 mg, 1 mmol). After stringing overnight, the resulting precipitate was filtered off and washed several times with ethanol and diethyl ether. Then, the product was dried under vacuum to yield  $L^{5S}$  as a white solid (611 mg), yield: 76%. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  8.49 (s, 1H, Hb), 8.42 (br, 2H, Hc, Hd), 7.86 (s, 1H, Ha), 7.61 (m, 3H, H1, H4, H5), 7.40 (d, J = 8.0 Hz, 1H, H8), 7.32 (m,

4H, H11, H12), 7.06 (m, 6H, He, H2, H3, H6, H7, H13), 4.80 (m, 1H, H9), 1.35 (d, J = 7.6 Hz, 3H, H10). <sup>13</sup>C NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  154.93 (CO), 154.18 (CO), 154.11 (CO), 145.15 (C), 133.02 (C), 131.57 (C), 131.10 (C), 129.50 (C), 128.33 (CH), 126.68 (CH), 125.93 (CH), 124.95 (CH), 124.41 (CH), 124.35 (CH), 124.07 (CH), 123.98 (CH), 123.87 (CH), 122.68 (CH), 122.36 (CH), 48.82 (CH), 23.13 (CH<sub>3</sub>). ESI-MS: m/z, 100%, 827.3440 [M+Na]<sup>+</sup>; 39%, 805.3580 [M+H]<sup>+</sup>.



Scheme S3. Synthetic scheme of preparing hexa-urea chiral ligand  $L^3$ .



**3a**: To a solution of **1b** (1.9 g, 5.0 mmol) in DMF (25 mL) in N<sub>2</sub> atmosphere, a solution of *o*-nitro-phenylisocyanate (1.8 g, 11.5 mmol) in THF (7 mL) was added dropwise at 70 °C. After stirring at the same temperature for 1 h, the solvents were evaporated by reduced pressure and washed for several times with ethanol and diethyl ether. A yellow solid (3.2 g) was produced as the **3a**, yield: 90%. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  9.74 (s, 1H, NHd),

9.21 (s, 1H, NHc), 8.54 (s, 1H, NHb), 8.47 (s, 1H, NHa), 8.25 (d, *J* = 8.0 Hz, 1H, H10), 8.07 (d, *J* = 7.6 Hz, 1H, H7), 7.73 (d, *J* = 8.0 Hz, 1H, H6), 7.65 (t, *J* = 8.0 Hz, 1H, H8), 7.57 (d, *J* = 8.0 Hz, 1H, H3), 7.43 (d, *J* = 6.8 Hz, 1H, H2), 7.19 (dd, 1H, *J* = 8.0 Hz, 1H, H9), 7.11, (dd, *J* = 8.0 Hz, 1H, H4), 7.04 (m, 2H, H1, H5).



**3b**: Hydrazine monohydrate (5.0 mL) was added dropwise to the suspension of **1b** (2.2 g, 3.1 mmol) and Pd/C (0.20 g, 10% cat.) in ethanol (200 mL) under stirring 2 h. Whereafter, the solid was filtered off *via* suction filtration and then dissolved in DMF (20 ml) and filtered through Celite to remove Pd/C. Diethyl ether (200 mL) was poured into the DMF solution after which the obtained precipitate was filtered off, washed several times with ethanol and

diethyl ether and dried over vacuum. Finally, a white solid (1.3 g) was produced as the **2b**, yield: 65 %. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$ 8.48 (s, 1H, NHa), 8.46 (s, 1H, NHb), 8.15 (s, 1H, NHc), 8.14 (s, 1H, NHd), 7.59 (m, 2H, H2, H3), 7.52 (d, *J* = 7.2 Hz, 1H, H6), 7.29 (d, *J* = 7.2 Hz, 1H, H7), 7.04 (m, 3H, H1, H4, H9), 6.83 (t, *J* = 6.8 Hz, 1H, H5), 6.72 (d, *J* = 7.2 Hz, 1H, H10), 6.54 (t, *J* = 7.2 Hz, 1H, H8), 4.78 (s, 2H, NHe).



Hexa-urea  $L^3$ : (S)-(-)- $\alpha$ -methylbenzyl isocyanate (367 mg, 2.5 mmol) was added to a DMF (5 mL) solution of compound **3b** (645 mg, 1 mmol). After stringing overnight, the resulting precipitate was filtered off and washed several times with ethanol and diethyl ether. Then, the product was dried under vacuum to yield  $L^{6S}$  as a white solid (572 mg), yield: 61%. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  8.52 (s, 2H, He), 8.50 (s, 2H, Hd), 8.41 (s, 2H, Hc, Hb),

7.86 (s, 1H, Ha), 7.63 (m, 4H, H2, H3, H6, H7), 7.41 (d, J = 8.0 Hz, 1H, H10), 7.31 (m, 4H, H13, H14), 7.21 (m, 1H, H15), 7.12 (m, 6H, Hf, H1, H4, H5, H8, H9), 4.80 (m, 1H, H11), 1.36 (d, J = 6.8 Hz, 3H, H12). <sup>13</sup>C NMR (400 MHz, DMSO-*d6*, ppm):  $\delta$  154.93 (CO), 154.19 (CO), 154.10 (CO), 145.16 (C), 133.08 (C), 131.58 (C), 131.28 (C), 131.08 (C), 129.47 (C), 128.34 (CH), 126.69 (CH), 125.93 (CH), 125.01 (CH), 124.18 (CH), 124.07 (CH), 124.00 (CH), 123.88 (CH), 122.67 (CH), 122.34 (CH), 48.83 (CH), 21.13 (CH<sub>3</sub>). ESI-MS: m/z, 100%, 961.3899 [M+Na]<sup>+</sup>; 30%, 977.3658 [M+K]<sup>+</sup>.

#### **S3.** Preparations of Anion Coordination Driven Complexes

Crystal (TEA)<sub>2</sub>[ $L^{1}$ •Cl<sub>2</sub>]: (TEA)Cl solution (32 µL, 0.625 mol/L, in acetonitrile) was added to a suspension of  $L^{1}$  (6.70 mg, 10 mmol) in chloroform. After stirring overnight at room temperature, a colorless solution was obtained. Slow vapor diffusion of diethyl ether into this solution provided colorless crystals of (TEA)<sub>2</sub>[ $L^{1}$ •Cl<sub>2</sub>] within two weeks. The crystal was directly mounted on a diffractometer for data collections.

Crystal (TBA)<sub>2</sub>[ $L^2$ •Cl<sub>2</sub>]: (TBA)Cl solution (20 µL, 0.625 mol/L, in acetonitrile) was added to a suspension of  $L^2$  (5 mg, 6.2 mmol) in chloroform. After stirring overnight at room temperature, a clearly colorless solution was obtained. Slow vapor diffusion of diethyl ether into this solution provided yellow crystals of (TBA)<sub>2</sub>[ $L^2$ •Cl<sub>2</sub>] within two weeks. The crystal was directly mounted on a diffractometer for data collections.

Crystal  $(TMA)_3[L^{1}_2 \cdot PO_4]$ :  $(TMA)_3PO_4$  solution (8 µL, 0.625 mol/L, in water) was added to a suspension of L<sup>1</sup> (6.7 mg, 10 mmol) in mixed solution of acetone and acetonitrile. After stirring overnight at room temperature, a colorless solution was obtained. Slow vapor diffusion of diethyl ether into this solution provided colorless crystals of  $(TMA)_3[L^{1}_2 \cdot PO_4]$  within one week. The crystal was directly mounted on a diffractometer for data collections.

Crystal  $(TMA)_3[L^3 \cdot PO_4 \cdot (H_2O)_2]$ :  $(TMA)_3PO_4$  solution (8.6 µL, 0.625 mol/L, in water) was added to a suspension of  $L^3$  (5 mg, 5.3 mmol) in acetone. After stirring overnight at room temperature, a colorless solution was obtained. Slow vapor diffusion of diethyl ether into this solution provided colorless crystals of  $(TMA)_3[L^3 \cdot PO_4 \cdot (H_2O)_2]$  within two weeks.

According to general method of crystal growth, we respectively attempted to synthesize crystals of supermolecule complex  $[L^3 \cdot Cl_2]^{2-}$ ,  $[L^2_2 \cdot PO_4]^{3-}$  and  $[L^2 \cdot PO_4]^{3-}$  using the slow vapor diffusion of poor solvent. Unfortunately, crystals with better diffraction points have never been

obtained. However, we still used other detection methods (<sup>1</sup>H NMR, ESI-MS and CD spectrum) to prove the assembly configuration and structure information of complexes  $[L^3 \cdot Cl_2]^{2-}$ ,  $[L^2_2 \cdot PO_4]^{3-}$  and  $[L^2 \cdot PO_4]^{3-}$ .

## S4. X-ray Diffraction Data

 $(TBA)_{2}[L^{2} \cdot Cl_{2}], (TMA)_{3}[L^{1}_{2} \cdot PO_{4}]$ crystals X-ray diffraction data of and  $(TMA)_3[L^3 \cdot PO_4 \cdot (H_2O)_2]$  were detected on a Bruker D8 Venture Photon II diffractometer with graphite-monochromatic Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å) under 150 K. The diffraction data for crystal (TEA)<sub>2</sub>[L<sup>1</sup>•Cl<sub>2</sub>] were collected at the BL17B macromolecular crystallography beamline in Shanghai Synchrotron Facility ( $\lambda = 0.72929$  Å). An empirical absorption correction using SADABS was applied for all data. (SADABS v 2018. 1, Bruker AXS, Madison, WI, 2018.) The structures were solved by the dual methods *via* SHELXS program. (A short history of SHELX, G. M. Sheldrick, Acta Cryst. 2008, A64, 112-122). All structures were solved and refined to convergence by the full-matrix least-squares on  $F^2$  for all independent reflections by the use of the program SHELXL. Hydrogen atoms were included in idealized positions with thermal parameters equivalent to 1.2 times those of the atom to which they were attached. Thereinto, because the structure of crystal (TEA)<sub>2</sub>[L<sup>1</sup>•Cl<sub>2</sub>] is small and the light source of BL17B macromolecular crystallography beamline is too strong, the low-angle diffraction points were overexposure, which leads to the low completeness of the crystal structure. All the crystal structures have been deposited with the Cambridge Crystallographic Data Centre (CCDC), and their corresponding CCDC number are shown in Table S1 and S2.

Complexes	$(TEA)_2[L^1 \bullet Cl_2]$	$(TBA)_2[L^2 \cdot Cl_2]$
CCDC Number	2130743	2130744
Formula	$C_{54}H_{78}Cl_2N_{10}O_4$	C77H44Cl2N12O5
M	1002.16	1288.14
Crystal system	Monoclinic	Monoclinic
Space group	<i>C</i> 2	$P2_1$
<i>a</i> (Å)	20.7990(5)	11.9428(5)
<i>b</i> (Å)	14.2717(4)	25.5121(9)
<i>c</i> (Å)	20.0159(6)	12.5127(4)
$\alpha$ (deg)	90	90
$\beta$ (deg)	114.881(1)	92.975(1)
γ (deg)	90	90
$V(Å^3)$	5390.0(3)	3807.3(2)
Z	4	2
<i>T</i> (K)	150	150
<i>F</i> (000)	2152	1328
$D_{calc} (g \cdot cm^{-3})$	1.235	1.124
R(int)	0.0508	0.0596
Data/restraints/parameters	8941 / 1 / 641	13393 / 1 / 875
GOF	1.062	0.732
$R1 [I > 2\sigma(I)]$	0.0393	0.0425
$wR2 [I > \sigma(I)]$	0.1036	0.1156

Table S1. Crystal data and refinement details of complexes 1 and 2

Complexes	$(TMA)_3[L^1_2 \bullet PO_4]$	$(TMA)_3[L^3 \bullet PO_4 \bullet (H_2O)_2]$
CCDC Number	2130745	2130746
Formula	$C_{198}H_{239}N_{40}O_{31}P_2$	C67H96N15O13P
М	3737.22	3625.26
Crystal system	Triclinic	Monoclinic
Space group	<i>P</i> 1	$P2_1$
<i>a</i> (Å)	13.8456(13)	13.0070(12)
<i>b</i> (Å)	13.8504(12)	20.8259(16)
<i>c</i> (Å)	26.594(2)	13.9625(14)
$\alpha$ (deg)	98.630(3)°	90°
$\beta$ (deg)	90.112(3)°	106.566(3)°
$\gamma$ (deg)	101.910(3)°	90°
$V(Å^3)$	4930.7(8)	3625.2(6)
Ζ	1	2
<i>T</i> (K)	149.98	150
<i>F</i> (000)	1985	1444
$D_{calc} \left( \mathbf{g} \cdot \mathbf{cm}^{-3} \right)$	1.266	1.237
R(int)	0.1269	0.0670
Data/restraints/parameters	35990 / 306 / 2472	12829 / 1 / 867
GOF	1.001	1.038
$R1 \left[I > 2\sigma(I)\right]$	0.0726	0.0954
$wR2 [I > \sigma(I)]$	0.1712	0.2187

Table S2. Crystal data and refinement details of complexes 4 and 6

, 0	1	L 2	2	
$D-H \cdots A$	d(D-H)	$d(H \cdot \cdot \cdot A)$	$d(D \cdot \cdot \cdot A)$	∠(DHA)
N1-H1···Cl1	0.86	2.32	3.170(4)	167
$N2-H2 \cdot \cdot \cdot C11$	0.86	2.43	3.211(2)	151
N3-H3· · · Cl1	0.86	2.41	3.235(3)	161
$N4-H4 \cdot \cdot \cdot C11$	0.86	2.65	3.431(6)	150
N5-H5· · · Cl2	0.86	2.65	3.431(5)	150
N6-H6· · · Cl2	0.86	2.42	3.281(4)	161
$N7-H7 \cdot \cdot \cdot C12$	0.86	2.43	3.213(2)	151
N8-H8· · · Cl2	0.86	2.32	3.172(7)	167

Table S3. Hydrogen bond parameters [Å and °] in the crystal structure of (TEA)<sub>2</sub>[L<sup>1</sup>•Cl<sub>2</sub>].

Table S4. Hydrogen bond parameters [Å and °] in the crystal structure of (TBA)<sub>2</sub>[L<sup>2</sup>•Cl<sub>2</sub>].

$D-H \cdots A$	d(D-H)	$d(H \cdot \cdot \cdot A)$	$d(D \cdot \cdot \cdot A)$	$\angle$ (DHA)
N1-H1···Cl1	0.86	2.43	3.252(3)	161
N2-H2· · · Cl1	0.86	2.36	2.752(3)	108
N3-H3· · · Cl2	0.86	2.51	3.318(3)	157
$N4-H4 \cdot \cdot \cdot C12$	0.86	2.44	3.265(2)	162
N5-H5· · · Cl1	0.86	2.49	3.319(3)	163
N6-H6· · · C11	0.86	2.55	3.333(3)	153
$N7-H7 \cdot \cdot \cdot C12$	0.86	2.69	3.475(3)	153
N8-H8· · · Cl1	0.86	2.7	3.423(2)	143
N9-H9· · · Cl2	0.86	2.73	3.537(3)	158
N10-H10· · · Cl2	0.86	2.51	3.332(3)	160

Table S5. Hydrogen bond parameters [Å and °] in the crystal structure of  $(TMA)_3[L_2^1 \cdot PO_4]$ .

~ -		P m m m m m m m m			
	D-H· · · A	$\overline{d(D-H)}$	$d(\overline{H \cdot \cdot \cdot A})$	$\overline{d(D \cdot \cdot \cdot A)}$	∠(DHA)
	N1-H1A· · · O20	0.88	2.02	2.853(7)	157
	$N2-H2A \cdots O20$	0.88	2.24	3.021(7)	148
	N3-H3A· · · O17	0.88	2.11	2.865(7)	144
	N4-H4A· · · O17	0.88	2.09	2.708(7)	127
	N5-H5A· · · O17	0.88	2.00	2.845(7)	162
	N6-H6· · · O19	0.88	1.94	2.784(7)	160
	N7-H7A· · · O19	0.88	1.96	2.833(7)	169
	N8-H8· · · O18	0.88	2.10	2.899(7)	151
	N9-H9· · · O18	0.88	2.30	3.023(7)	140
	N10-H10· · · O18	0.88	1.90	2.726(7)	157
	N11-H11A· · · O18	0.88	2.20	2.986(7)	149
	N12-H12A· · · O20	0.88	1.84	2.717(7)	171
	N13-H13A· · · O20	0.88	2.17	2.853(7)	135
	N14-H14A· · · O19	0.88	1.85	2.717(7)	169
	N15-H15· · · O19	0.88	2.10	2.951(7)	162
	N16-H16· · · O17	0.88	2.02	2.884(7)	168

D-H···A	d(D-H)	$d(H \cdot \cdot \cdot A)$	$d(D \cdot \cdot \cdot A)$	∠(DHA)
N1-H1· · · O10	0.88	2.1	2.920(4)	154
N2-H2· · · O7	0.88	1.82	2.694(2)	171
N3-H3· · · O7	0.88	2.04	2.872(1)	157
N4-H4· · · O8	0.88	1.98	2.793(1)	153
N5-H5· · · O8	0.88	1.89	2.755(0)	167
N6-H6· · · O10	0.88	2.05	2.892(2)	159
N7-H7· · · O10	0.88	1.8	2.666(2)	166
N8-H8· · · O9	0.88	2.26	3.111(2)	162
N9-H9· · · O9	0.88	1.78	2.656(2)	174
N10-H10···O11	0.88	2.35	2.753(4)	153
N11-H11· · · O7	0.88	1.96	2.781(1)	155
N12-H12· · · O7	0.88	2.07	2.836(1)	144

Table S6. Hydrogen bond parameters [Å and °] in the crystal structure of (TMA)<sub>3</sub>[L<sup>3</sup>•PO<sub>4</sub>•(H<sub>2</sub>O)<sub>2</sub>].

Table S7. Hydrogen bonds (Å and deg) involved in chloride binding and  $Cl \cdot \cdot Cl$  separations in the crystal structures of complexes  $(TEA)_2[L^1 \cdot Cl_2]$  and  $(TBA)_2[L^2 \cdot Cl_2]$ .

Cl1				C12			
Complex	H- bond num ber	Average $d(N \cdots Cl)$ and $\angle(NHCl)$ (Å, deg)	H- bon d num ber	Average $d(N \cdots Cl)$ and $\angle$ (NHCl) (Å, deg)	Cl1· · · Cl2 separation[A°]		
$(TEA)_2[L^1 \cdot Cl_2]$	4	3.26, 157	4	3.28, 157	4.21		
$(TBA)_2[L^2 \cdot Cl_2]$	5	3.22, 145	5	3.39, 158	3.73		

Table S8. Hydrogen bonds (Å and deg) involved in phosphate binding in the crystal structures of complexes  $(TMA)_3[L^1_2 \cdot PO_4]$  and  $(TMA)_3[L^3 \cdot PO_4 \cdot (H_2O)_2]$ .

Complex	H-bond number	average $d(N \cdots O)$ and $\angle(NHO)$		
$(TMA)_3[L^1_2 \bullet PO_4]$	16	2.85, 154		
$(TMA)_3[L^3 \cdot PO_4 \cdot (H_2O)_2]$	12	2.81, 160		



*Figure S1.* (a) Double helix structure of phosphate-template  $[L_2 \cdot PO_4]^{3-}$  complex and (b) corresponding urea groups and hydrogen bond distribution around phosphate (the left and right ligands of double helix are shown in green and yellow, respectively). Selected intermolecular short-contacts ( $\pi$ - $\pi$  interaction) is shown.



*Figure S2.* Packing structures of crystals (a)  $(TEA)_2[L^1 \cdot Cl_2]$ , (b)  $(TBA)_2[L^2 \cdot Cl_2]$ , (c)  $(TMA)_3[L^1_2 \cdot PO_4]$  and (d)  $(TMA)_3[L^3 \cdot PO_4 \cdot (H_2O)_2]$  (chloride ions are shown as green sphere).

#### **S5. Studies of CD and UV-vis Experiments**



**Figure S3.** The corresponding anisotropy factors for the single helix of (a)  $[L \cdot Cl_2]^{2-}$  and (b)  $[L \cdot PO_4]^{3-}$  complexes in acetonitrile ([L] = 30  $\mu$ M). The anisotropy factor  $g = \Delta A/A = \theta[\text{mdeg}]/(32980 \times A)$ . (I. Dolamic, S. Knoppe, A. Dass and T. Bürgi, *Nat. Commun.*, **2012**, 3, 798.)



*Figure S4.* CD spectra of (a)  $L^1$ ,  $L^2$  and  $L^3$  ligands. CD titration of (b)  $L^1$ , (c)  $L^2$  and (d)  $L^3$  ([L] = 30  $\mu$ M, in 5% v/v DMSO/CH<sub>3</sub>CN) by adding 1.2 equivalent of phosphate anions gradually showing conversion in helical sense.



*Figure S5.* (a) CD spectra of  $[L^{1} \cdot Cl_2]^{2-}$  by adding 1 equivalent of phosphate anions gradually showing conversion in helical sense and (b) changes in CD spectra at wavelength of 244 nm, 258 nm and 280 nm. ( $[L^1] = 30 \mu M$ , CH<sub>3</sub>CN)



*Figure S6.* (a) UV-vis titration of  $L^1$  ([ $L^1$ ] = 30 µM) with Cl<sup>-</sup> in 5% v/v DMSO/CH<sub>3</sub>CN and (b) changes in UV-vis spectra at wavelength of 274 nm and 286 nm.



*Figure S7.* (a) UV-vis titration of  $L^2$  ([ $L^2$ ] = 30 µM) with Cl<sup>-</sup> in 5% v/v DMSO/CH<sub>3</sub>CN and (b) changes in UV-vis spectra at wavelength of 274 nm and 286 nm.



*Figure S8.* (a) UV-vis titration of  $L^3$  ([ $L^3$ ] = 30 µM) with Cl<sup>-</sup> in 5% v/v DMSO/CH<sub>3</sub>CN and (b) changes in UV-vis spectra at wavelength of 274 nm and 286 nm.



*Figure S9.* (a) UV-vis titration of  $L^1$  ([ $L^1$ ] = 30 µM) with PO<sub>4</sub><sup>3–</sup> in 5% v/v DMSO/CH<sub>3</sub>CN and (b) changes in UV-vis spectra at wavelength of 251 nm and 286 nm.



*Figure S10.* (a) UV-vis titration of  $L^2$  ([ $L^2$ ] = 30 µM) with PO<sub>4</sub><sup>3–</sup> in 5% v/v DMSO/CH<sub>3</sub>CN and (b) changes in UV-vis spectra at wavelength of 251 nm and 286 nm.



*Figure S11.* (a) UV-vis titration of  $L^2$  ([ $L^2$ ] = 30  $\mu$ M) with PO<sub>4</sub><sup>3–</sup> in 5% v/v DMSO/CH<sub>3</sub>CN and (b) changes in UV-vis spectra at wavelength of 251 nm and 286 nm.



*Figure S12.* (a) <sup>1</sup>H NMR titration (400 MHz, DMSO- $d_6$ , 298 K) of L<sup>1</sup> ([L<sup>1</sup>] = 0.6 mM) in the presence of various equiv. of chloride anions and (b) the corresponding binding constants were obtained from non-linear curve-fitting to a 1:1 binding model via the http://app.supramolecular.org/bindfit/view/dbcb92af-4e2f-4125-8090-3f3debdeb6d5.



*Figure S13.* (a) <sup>1</sup>H NMR titration (400 MHz, DMSO- $d_6$ , 298 K) of L<sup>2</sup> ([L<sup>2</sup>] = 0.6 mM) in the presence of various equiv. of chloride anions and (b) the corresponding binding constants were obtained from non-linear curve-fitting to a 1:1 binding model via the http://app.supramolecular.org/bindfit/view/83e937a4-7503-4b85-b5e5-82ced312fff2.



*Figure S14.* (a) <sup>1</sup>H NMR titration (400 MHz, DMSO- $d_6$ , 298 K) of L<sup>3</sup> ([L<sup>3</sup>] = 0.6 mM) in the presence of various equiv. of chloride anions and (b) the corresponding binding constants were obtained from non-linear curve-fitting to a 1:1 binding model via the http://app.supramolecular.org/bindfit/view/9e915224-125e-4066-9fa8-03657d6d3e63.



*Figure S15.* Partial <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>, 298 K) spectra of L<sup>1</sup> ([L] = 0.1 mM) in the presence of various equivalent of PO<sub>4</sub><sup>3–</sup> (black numbers indicate the equivalent of PO<sub>4</sub><sup>3–</sup> added; the signals of *MM* and *P* are shown in blue and red, respectively).



*Figure S16.* Partial <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, 298 K) spectra of  $L^2$  ([L] = 0.1 mM) in the presence of various equivalent of PO<sub>4</sub><sup>3-</sup> (black numbers indicate the equivalent of PO<sub>4</sub><sup>3-</sup> added; the signals of  $[L^2_2 \cdot PO_4]^{3-}$  and  $[L^2 \cdot PO_4]^{3-}$  complexes are shown in blue and red, respectively).



*Figure S17.* Partial <sup>1</sup>H NMR (400 MHz, DMSO-*d6*, 298 K) spectra of  $L^3$  ([L] = 0.1 mM) in the presence of various equivalent of PO<sub>4</sub><sup>3-</sup> (black numbers indicate the equivalent of PO<sub>4</sub><sup>3-</sup> added; the signals of [L<sup>3</sup>•PO<sub>4</sub>]<sup>3-</sup> complex are shown in red).



*Figure S18.* Stacked partial <sup>1</sup>H NMR spectra (400 MHz, 298 K, DMSO- $d_6$ ) for the oligourea ligands with their anion coordination complex of (a)  $[L^2 \cdot Cl_2]^{2-}$ , (b)  $[L^3 \cdot Cl_2]^{2-}$ , (c)  $[L^2 \cdot PO_4]^{3-}$ ,

and (d)  $[L^3 \cdot PO_4]^{3-}$ , respectively. Tetrabutylammonium chloride and tetramethylammonium phosphate salts were used.

NH Complex	На	Hb	Нс	Hd	Не	Hf
1	0.32	0.18	0.16	0.32		
2	0.28	0.33	0.1	0.07	0.28	
3	0.23	0.25	0.11	0.22	0.06	0.25

Table S9. Chemical shift of NH signals for complexes **1-3** in <sup>1</sup>H NMR.

Table S10. Chemical shift of NH signals for complexes **4-6** in <sup>1</sup>H NMR.

NH Complex	На	Hb	Нс	Hd	Не	Hf
4	2.60	3.30	2.85	1.69		
5	1.82	3.29	2.02	1.44	1.87	
6	1.61	1.98	1.72	1.84	1.49	2.11



*Figure S19.* High-resolution ESI-MS spectrum of (a-c) complexes of oligourea ligands with two equivalents of chloride anion.



*Figure S20.* High-resolution ESI-MS spectrum of the formation of (a) double helix  $[L^{1}_{2} \cdot PO_{4}]^{3-}$  and (b) single helix  $[L^{1} \cdot PO_{4}]^{3-}$ .



*Figure S21.* High-resolution ESI-MS spectrum of the formation of (a)  $[L^{2}_{2} \cdot PO_{4}]^{3-}$  and (b)  $[L^{2} \cdot PO_{4}]^{3-}$ .



*Figure S22.* High-resolution ESI-MS spectrum of the formation of single helix  $[L^3 \cdot PO_4]^{3-}$ .

#### **S7. Computational Analysis for the Helicity Bias**



*Figure S23.* Computationally optimized geometry (DFT, B3LYP, 6-31G\*) for the single helix of  $[L^1 \cdot Cl_2]^{2-}$  with (a) left-handed conformation M and (b) right-handed conformation P. The optimized structure indicates that the right-handed helix is more stable than the left-handed helix.



*Figure S24.* Computationally optimized geometry (DFT, B3LYP, 6-31G\*) for the double helices of  $[L^{1}_{2} \cdot PO_{4}]^{3-}$  with (a) left-handed conformation *MM* and (b) right-handed conformation *PP*. The results suggests that the left-handed double helices is more energetically favorable than the right-handed double helices. And the extra contact of C-H••• $\pi$  interaction between the strands of *MM* conformations could be driving force for the induced helicity bias.

# S8. <sup>1</sup>H and <sup>13</sup>C NMR Spectra





