Supporting Information

Activation of perfluoroalkyl iodides by anions: Extending the scope of halogen bond activation to C(sp³)-H amidation, C(sp²)-H iodination and perfluoroalkylation reactions

 $Yax in Wang, *^{\dagger ab} Zehui Cao, ^{\dagger a} Qin He, ^{\dagger a} Xin Huang, ^{c} Jiaxi Liu, ^{a} Helfried Neumann, *^{b} Gong Chen*^{c} and Matthias Beller, *^{b}$

^[a] College of Pharmacy, Nanjing University of Chinese Medicine, Nanjing 210023 (China).

Email: 300500@njucm.edu.cn, Yaxin.Wang@catalysis.de

^[b] Leibniz-Institut f
ür Katalyse e.V. an der Universit
ät Rostock, Albert-Einstein Stra
ße 29a, 18059 Rostock (Germany). E-mail: Matthias.Beller@catalysis.de

^[c] State Key Laboratory and Institute of Elemento-Organic Chemistry, Nankai University, Tianjin 300071 (China).

Content

1.	Reagents
2.	Instruments
3.	Synthesis of substrates
4.	Reaction optimization for the halogen-bond-promoted reaction of C(sp ³)-H amidation of N-(2-methoxybenzyl)-pyridine-2-sulfonamide
5.	General procedures and substrate scope of halogen-bond-promoted reactions for C(sp ³)-H amidationS7
6.	General procedures and substrate scope of halogen-bond-promoted reactions for C(sp ²)-H iodination
7.	Evaluation of anions promoters for synthesis of compound 44 under blue LED irradiation
8.	General procedures and substrate scope of phenanthridine and isoquinoline synthesis
9.	General procedures and substrate scope for addition of perfluorobutyl iodide to alkenes and alkynes
10.	General procedure and substrate scope for C-H perfluorobutylation of electron-rich arene and heteroare
11.	Gram-scale C(sp ³)-H amidation, C(sp ²)-H iodination and perfluoroalkylation reactions
12.	Mechanistic studies
13.	References
14.	¹ H NMR. ¹³ C NMR and ¹⁹ F NMR spectra

1. Reagents

All commercial materials were used as received unless otherwise noted. Dry solvents and deuterated solvents were purchased from J&K Chemical. Perfluoroalkyl iodides were purchased from Energy Chemical. Starting materials for this study were purchased from TCI or were synthesized according to reported procedures. Pd(OAc)₂ and Cs₂CO₃ (\geq 99%, Aladdin), K₂CO₃, Rb₂CO₃, AgOAc, Ag₂CO₃, KOH, KHCO₃, K₃PO₄, NaOCH₃, NaSCH₃, CsOH, NaO*t*Bu (99%, Energy Chemical), and NaH (60% dispersion in mineral oil, Energy Chemical) were used in the reactions of C(sp³)-H amidation or C(sp²)-H iodination and perfluoroalkylation.

TLC were performed on silica gel Huanghai HSGF254 plates and visualization of the developed chromatogram was performed by fluorescence quenching ($\lambda_{max} = 254$ nm). Flash chromatography was performed using silica gel (200-300 mesh) purchased from Qingdao Haiyang Chemical Co., China.

2. Instruments

NMR spectra were recorded on Bruker AVANCE AV 400 instruments and all NMR experiments were reported in units, parts per million (ppm), using residual solvent peaks as internal reference. Multiplicities are recorded as: s = singlet, d = doublet, t = triplet, dd = doublet of doublets, td = triplet of doublets, br = broad singlet, m = multiplet. High resolution ESI mass experiments were operated on a Waters LCT Premier instrument. All reactions were carried out in a 4 mL glass vial (Thermo SCIENTIFIC National B7999-2, made from superior quality 33 expansion borosilicate clear glass), sealed with a PTEF cap on bench top if necessary.

Lights: PHILIPS TORNADO 25W CFL

3. Synthesis of substrates

Compounds 1¹, 4-1¹, 13-1², 14-1², 15-1², isonitrile compounds (44-1 to 80-1)³, alkenes (86-1 to 89-1)³, alkynes (102-1 to 104-1)³, and biologically active molecules (90-1 to

Ethers and arenes:









Scheme S2. Substrates used in amidation and iodination



Scheme S3. Substrates used in perfluoroalkylation

100-1 and 108-1, 109-1)³ are known compounds and were synthesized following the reported procedures. Other compounds for this study are commercially available and used as received.

4. Reaction optimization for the halogen-bond-promoted reaction of C(sp³)-H amidation of N-(2-methoxybenzyl)-pyridine-2-sulfonamide

All screening reactions were carried out at a 0.2 mmol scale in a 4 mL glass vial (Thermo Scientific, National B7999-2) sealed with PTEF cap and stirred on bench top. The amidation reaction was discovered while aiming for a selective perfluoroalkylation of N-(2-methoxybenzyl)-pyridine-2-sulfonamide (see below).



Sulfonyl amide **1** (55.6 mg, 0.2 mmol, 1.0 equiv) and other specified reagents were dispersed in specific solvent and the resulting mixture was vigorously stirred at specified temperature with or without light irradiation for 12 hours. After removal of the solvent *in vacuo*, the resulting residue was dissolved in 1 mL of CDCl₃ along with $Cl_2CHCHCl_2$ (20 uL) as an internal standard for ¹H NMR analysis. The composition of the reaction mixture was analyzed based on the -(N)CH(O)- peak at 5.89 (t, J = 6.2 Hz, 1H) for compound **2**.

Table S1. Reaction optimization for the halogen-bond-promoted reaction of C(sp3)-H amidation of N-(2-methoxybenzyl)-pyridine-2-sulfonamide

entry reagent (equiv), reaction time, temp		solvent	2 %
1	C ₄ F ₉ I (2), Pd(OAc) ₂ (0.1), TEEDA (3), CFL (25 W), 60 °C, 12 h, Ar	THF	23
2	$C_4F_9I(2), Pd(OAc)_2(0.1), Cs_2CO_3(3), 60 \circ C, 12$	THF	62

	h, Ar,		
2	$C_4F_9I(2), Cs_2CO_3(3), Pd(OAc)_2(10 \text{ mol}\%),$	THE	62
3	AgOAc (3), 12 h, 60 °C, Ar, CFL (25 W)	IHF	(55 ^b)
4	C ₄ F ₉ I (2), Cs ₂ CO ₃ (3), 60 °C, 12 h, Ar	THF	65
4			(56 ^b)
5	C ₄ F ₉ I (2), Cs ₂ CO ₃ (3), 30 °C, 12 h, Ar	THF	<2
6	C ₄ F ₉ I (2), K ₂ CO ₃ (3), 60 °C, 12 h, Ar THF		10
7	C ₄ F ₉ I (2), Na ₂ CO ₃ (3), 60 °C, 12 h, Ar	THF	<2
8	C ₄ F ₉ I (2), Rb ₂ CO ₃ (3), 12 h, 60 °C, Ar	THF	56
9	C ₄ F ₉ I (2), Ag ₂ CO ₃ (3), 12 h, 60 °C, Ar	THF	<2
10	C ₄ F ₉ I (2), KHCO ₃ (3), 12 h, 60 °C, Ar	THF	10
11	C ₄ F ₉ I (2), K ₃ PO ₄ (3), 12 h, 60 °C, Ar THF		35
12	C ₄ F ₉ I (2), NaOCH ₃ (3), 60 °C, 12 h, Ar	THF	33
13	C ₄ F ₉ I (2), NaSCH ₃ (3), 12 h, 60 °C, Ar	THF	44
14	C ₄ F ₉ I (2), CsOH (3), 12 h, 60 °C, Ar	THF	46
15	C ₄ F ₉ I (2), KO <i>t</i> Bu (3), 60 °C, 12 h, Ar	THF	<2
16	C ₄ F ₉ I (2), KO <i>t</i> Bu (3), 30 °C, 12 h, Ar	THF	<2
17	C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 60 °C, Ar	THF	99
1/			(90 ^b)
10	C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, Ar	THE	99
18		IHF	(90 ^b)
19	C ₄ F ₉ I (2), NaO <i>t</i> Bu (2), 12 h, 30 °C, Ar	THF	81
20	C ₄ F ₉ I (2), NaO <i>t</i> Bu (1), 12 h, 30 °C, Ar	THF	58
21	C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, Ar, in	THE	99
21	darkness	ITT	(90 ^b)
22	C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, Air	THF	28
23	C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, O ₂	THF	26
24°	THF (10), C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, Ar	CH ₃ CN	<2
25°	THF (10), C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, Ar	PhCF ₃	67

26°	THF (10), C ₄ F ₉ I (2), NaO <i>t</i> Bu (3), 12 h, 30 °C, Ar	CCl ₄	71
27	TEEDA (3), CFL (25 W), 30 °C, 12 h, Ar	THF	21
29	Et ₃ N (3), CFL (25 W), 30 °C, 12 h, Ar	THF	18
28	Et ₃ N (3), 30 °C, 12 h, Ar, in darkness	THF	<2
30	C ₄ F ₉ I (2), NaH (3), 30 °C, 12 h, Ar	THF	<2
31	Bu ₄ NCl (3), 30 °C, 12 h, Ar	THF	<2
32	Bu ₄ NBr (3), 30 °C, 12 h, Ar	THF	<2
33	Bu ₄ NI (3), 30 °C, 12 h, Ar	THF	<2
34	Bu ₄ NCl (3), NaH (1), 30 °C, 12 h, Ar	THF	65
35	Bu ₄ NBr (3), NaH (1), 30 °C, 12 h, Ar	THF	63
37	Bu ₄ NI (3), NaH (1), 30 °C, 12 h, Ar	THF	64

Halogen-bond promoted amination of dioxane. All screening reactions were carried out on a 0.2 mmol scale at 0.1 M concentration; the reaction vial is purged with Ar. a) Yield are based on ¹H NMR analysis on 0.2 mmol scale in 4 mL glass vial. b) Isolated yield. c) 10 equiv THF were added.

- 5. General procedures and substrate scope of halogen-bond-promoted reactions for C(sp³)-H amidation
- 5.1 Halogen-bond-promoted reaction system for the alkyl ethers C(sp³)-H amidation



Scheme S4. Halogen-bond-promoted reaction system for the alkyl ethers C(sp³)-H amidation

Conditions A: Amide (0.2 mmol, 1.0 equiv), C_4F_9I (0.4 mmol, 2.0 equiv) and NaOtBu (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of ethers. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.



Compound **2** was isolated in 90% yield (crude ¹H NMR yield: 99%) following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 8.74 (d, *J* = 4.6 Hz, 1H), 7.97 (d, *J* = 7.8 Hz, 1H), 7.86 (td, *J* = 7.7, 1.5 Hz, 1H), 7.58 (d, *J* = 7.4 Hz, 1H), 7.50 – 7.39 (m, 1H), 7.21 (t, *J* = 7.2 Hz, 1H), 6.95 (t, *J* = 7.5 Hz, 1H), 6.80 (d, *J* = 8.1 Hz, 1H), 5.89 (t, *J* = 6.2 Hz, 1H), 4.67 (d, *J* = 18.2 Hz, 1H), 4.48 (d, *J* = 18.2 Hz, 1H), 3.84 – 3.74 (m, 4H), 3.71 – 3.61 (m, 1H), 2.11 – 1.92 (m, 1H), 1.83 – 1.66 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 158.05, 156.14, 150.08, 137.80, 128.33, 127.95, 126.74, 126.59, 122.61, 120.66, 109.68, 89.27, 68.52, 55.21, 41.76, 30.06, 24.87. HRMS Calcd for C₁₇H₂₀N₂O₄S [M+H⁺]: 349.1217; Found: 349.1218.



Compound **4** was isolated in 84% yield following the condition A. ¹**H** NMR (400 MHz, CDCl₃) δ 7.88 (d, *J* = 7.7 Hz, 2H), 7.56 (t, *J* = 7.3 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 2H), 5.32 (dd, *J* = 9.7, 2.5 Hz, 1H), 3.88 – 3.59 (m, 4H), 3.54 – 3.41 (m, 2H), 3.19 – 3.05 (m, 1H), 3.05 – 2.90 (m, 1H), 1.85 – 1.67 (m, 1H), 1.52 – 1.36 (m, 1H), 1.25 (s, 6H), 0.87 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 139.90, 132.79, 128.93, 127.76, 82.83, 69.21, 66.94, 65.74, 44.45, 31.87, 31.45, 26.62, 22.67, 14.11. HRMS Calcd for C₁₆H₂₆NO₄S [M+H⁺]: 328.1577; Found: 328.1578.



Compound **5** was isolated in 90% yield following the condition A. ¹**H** NMR (400 MHz, CDCl₃) δ 7.90 (d, *J* = 8.0 Hz, 2H), 7.57 – 7.51 (m, 1H), 7.51 – 7.44 (m, 2H), 5.78 (t, *J* = 5.9 Hz, 1H), 3.87 (q, *J* = 7.5 Hz, 1H), 3.73 (q, *J* = 7.4 Hz, 1H), 3.07 – 2.84 (m, 2H), 2.22 – 2.07 (m, 1H), 1.96 – 1.76 (m, 4H), 1.66 – 1.45 (m, 1H), 1.24 (s, 6H), 0.86 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.19, 132.48, 128.86, 127.72, 88.85, 68.11, 43.52, 31.47, 30.36, 26.94, 25.03, 22.67, 14.10. HRMS Calcd for C₁₆H₂₆NO₃S [M+H⁺]: 312.1628; Found: 312.1628.



Compound **6** was isolated in 91% yield following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 7.87 (d, J = 7.9 Hz, 2H), 7.58 – 7.49 (m, 1H), 7.46 (t, J = 7.5 Hz, 2H), 5.13 (d, J = 9.1 Hz, 1H), 3.85 (d, J = 11.7 Hz, 1H), 3.58 – 3.37 (m, 1H), 3.16 – 3.03 (m, 1H), 3.03 – 2.89 (m, 1H), 2.00 – 1.87 (m, 1H), 1.88 – 1.69 (m, 2H), 1.66 – 1.38 (m, 5H), 1.24 (s, 6H), 0.86 (t, J = 6.5 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.25, 132.49, 128.71, 127.86, 86.28, 68.06, 43.92, 31.67, 31.49, 31.08, 26.77, 25.06, 23.82, 22.71, 14.13. HRMS Calcd for C₁₇H₂₈NO₃S [M+H⁺]: 326.1784; Found: 326.1780.



Compound **7** was isolated in 85% yield following the condition A. ¹**H** NMR (400 MHz, CDCl₃) δ 7.82 – 7.75 (m, 2H), 7.57 – 7.51 (m, 1H), 7.51 – 7.44 (m, 2H), 5.23 (q, J = 6.0 Hz, 1H), 3.61 - 3.53 (m, 1H), 3.40 - 3.33 (m, 1H), 3.26 – 3.08 (m, 1H), 3.07 – 2.94 (m, 1H), 1.66 - 1.57 (m, 2H), 1.34 – 1.20 (m, 6H), 1.17 (d, J = 6.1 Hz, 3H), 1.12 (t, J = 7.0 Hz, 3H), 0.86 (t, J = 6.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 141.19, 132.43, 129.08, 126.92, 84.24, 63.12, 42.47, 31.49, 30.77, 26.95, 22.68, 19.97, 14.86, 14.10. HRMS Calcd for C₁₆H₂₈NO₃S [M+H⁺]: 314.1784; Found: 314.1787.



Compound **8** was isolated in 84% yield following the condition A. ¹**H** NMR (400 MHz, CDCl₃) δ 7.85 – 7.74 (m, 2H), 7.58 – 7.51 (m, 1H), 7.48 (t, *J* = 7.4 Hz, 2H), 4.99 (dd, *J* = 8.3, 4.3 Hz, 1H), 3.44 – 3.38 (m, 1H), 3.27 – 3.21 (m, 1H), 3.21 – 3.08 (m, 1H), 3.06 – 2.91 (m, 1H), 1.72 – 1.51 (m, 3H), 1.50 – 1.41 (m, 2H), 1.40 – 1.14 (m, 11H), 0.86 (dt, *J* = 12.0, 5.8 Hz, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 141.34, 132.41, 129.00, 127.06, 88.18, 68.04, 42.71, 36.49, 31.54, 31.50, 30.77, 27.02, 22.70, 19.44, 19.07, 14.12, 13.98, 13.83. **HRMS** Calcd for C₂₀H₃₆NO₃S [M+H⁺]: 370.2410; Found: 370.2415.



Compound **9** was isolated in 90% yield following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 7.97 – 7.81 (m, 2H), 7.56 – 7.48 (m, 1H), 7.48 – 7.39 (m, 2H), 4.75 (s, 2H), 3.24 – 2.95 (m, 2H), 1.71 – 1.41 (m, 2H), 1.31 – 1.22 (m, 6H), 1.14 (s, 9H), 0.85 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.44, 132.28, 128.49, 127.90, 72.91, 70.84, 45.88, 31.47, 28.40, 27.62, 26.35, 22.58, 14.06. HRMS Calcd for C₁₇H₃₀NO₃S [M+H⁺]: 328.1941; Found: 328.1940.



Compound **11**' was isolated in 59% yield and compound **11**'' was isolated in 34% yield following the condition A.

Compound **11**': ¹**H NMR** (400 MHz, CDCl₃) δ 7.91 – 7.82 (m, 2H), 7.58 – 7.52 (m, 1H), 7.51 – 7.44 (m, 2H), 5.15 (t, *J* = 6.1 Hz, 1H), 3.46 (dd, *J* = 10.2, 6.4 Hz, 1H), 3.30 (dd, *J* = 10.2, 5.9 Hz, 1H), 3.27 (s, 3H), 3.25 (s, 3H), 3.20 – 2.98 (m, 2H), 1.63 – 1.48 (m, 2H), 1.30 – 1.17 (m, 6H), 0.86 (t, *J* = 6.9 Hz, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 140.97, 132.50, 128.82, 127.36, 87.47, 72.52, 59.04, 55.95, 42.97, 31.38, 30.46, 26.81, 22.62, 14.06. **HRMS** Calcd for C₁₆H₂₈NO₄S [M+H⁺]: 330.1734; Found: 330.1734.

Compound **11**": ¹**H NMR** (400 MHz, CDCl₃) δ 7.85 – 7.78 (m, 2H), 7.59 – 7.53 (m, 1H), 7.52 – 7.44 (m, 2H), 4.83 (s, 2H), 3.68 – 3.61 (m, 2H), 3.54 – 3.47 (m, 2H), 3.37 (s, 3H), 3.14 – 3.18 (m, 2H), 1.58 – 1.48 (m, 2H), 1.27 – 1.16 (m, 6H), 0.85 (t, *J* = 6.9 Hz, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 140.63, 132.58, 129.04, 127.20, 78.80, 71.68, 67.25, 59.05, 47.23, 31.42, 28.52, 26.40, 22.57, 14.05. **HRMS** Calcd for C₁₆H₂₈NO₄S [M+H⁺]: 330.1734; Found: 330.1733.



Compound **12**' was isolated in 60% yield and compound **12**" was isolated in 18% yield following the condition A.

Compound **12**': ¹**H NMR** (400 MHz, CDCl₃) δ 7.89 (d, *J* = 7.6 Hz, 2H), 7.58 (t, *J* = 7.3 Hz, 1H), 7.51 (t, *J* = 7.5 Hz, 2H), 6.02 (dd, *J* = 6.4, 2.9 Hz, 1H), 5.15 (s, 1H), 4.72 (s, 1H), 3.99 (dd, *J* = 9.7, 2.8 Hz, 1H), 3.89 (dd, *J* = 9.6, 6.7 Hz, 1H), 3.07 – 2.99 (m, 1H), 2.99 – 2.87 (m, 1H), 1.88 – 1.69 (m, 1H), 1.65 – 1.44 (m, 1H), 1.24 (s, 6H), 0.86 (t, *J* = 6.6 Hz, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 139.65, 132.82, 129.04, 127.73, 95.81, 84.71, 68.57, 43.74, 31.42, 31.11, 26.92, 22.67, 14.12. **HRMS** Calcd for C₁₅H₂₃NO₄S [M+H⁺]: 314.1421; Found: 314.1424.

Compound **12**": ¹**H NMR** (400 MHz, CDCl₃) δ 7.86 (d, J = 7.4 Hz, 2H), 7.62 – 7.53 (m, 1H), 7.50 (t, J = 7.1 Hz, 2H), 6.47 (s, 1H), 4.07 – 3.98 (m, 2H), 3.98 – 3.88 (m, 2H), 3.15 – 2.87 (m, 2H), 1.76 – 1.57 (m, 2H), 1.23 (s, 6H), 0.86 (t, J = 5.7 Hz, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 140.29, 132.72, 129.04, 127.40, 107.19, 64.76, 42.77, 31.47, 30.78, 26.85, 22.67, 14.14. **HRMS** Calcd for C₁₅H₂₃NO₄S [M+H⁺]: 314.1421; Found: 314.1425.



Compound **13** was isolated in 51% yield following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 7.40 (d, *J* = 7.5 Hz, 2H), 7.34 (t, *J* = 7.5 Hz, 2H), 7.26 (t, *J* = 7.1 Hz, 1H), 5.69 (t, *J* = 6.8 Hz, 1H), 4.55 (d, *J* = 16.7 Hz, 1H), 4.25 (d, *J* = 16.7 Hz, 1H), 4.02 (q, *J* = 6.8 Hz, 1H), 3.80 (q, *J* = 7.3 Hz, 1H), 2.96 (s, 3H), 2.05 – 1.77 (m, 3H), 1.76 – 1.57 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 138.38, 128.68, 127.45, 127.08, 88.82, 68.42, 46.47, 39.52, 29.51, 24.92. **HRMS** Calcd for C₁₂H₁₈NO₃S [M+H⁺]: 256.1002; Found: 256.1000.



Compound **14** was isolated in 88% yield following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, *J* = 8.1 Hz, 2H), 7.40 (d, *J* = 7.5 Hz, 2H), 7.34 – 7.17 (m, 5H), 5.90 (t, *J* = 6.6 Hz, 1H), 4.41 (d, *J* = 17.0 Hz, 1H), 4.11 (d, *J* = 17.0 Hz, 1H), 3.86 (q, 7.0 Hz, 1H), 3.72 (q, *J* = 7.3 Hz, 1H), 2.41 (s, 3H), 2.05 – 1.90 (m, 1H), 1.83 – 1.57 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 143.41, 138.54, 136.90, 129.57, 128.53, 127.87, 127.28, 127.14, 88.92, 68.52, 46.03, 30.22, 24.85, 21.66. HRMS Calcd for C₁₈H₂₂NO₃S [M+H⁺]: 332.1315; Found: 332.1317.



Compound **15** was isolated in 33% yield following the condition A. ¹H **NMR** (400 MHz, CDCl₃) δ 8.64 (d, J = 2.8 Hz, 1H), 7.79 (t, J = 7.2 Hz, 1H), 7.70 (d, J = 7.4 Hz, 1H), 7.32 (t, J = 6.6 Hz, 2H), 7.20 (t, J = 7.1 Hz, 1H), 6.93 (t, J = 7.2 Hz, 1H), 6.85 (d, J = 7.9 Hz, 1H), 5.79 (s, 1H), 4.92 (d, J = 17.3 Hz, 1H), 4.54 (d, J = 17.2 Hz, 1H), 3.94 (q, J = 7.3 Hz, 1H), 3.86 (s, 3H), 3.76 – 3.58 (m, 1H), 2.07 (q, J = 8.4 Hz, 1H), 1.96 – 1.70 (m, 3H). ¹³C **NMR** (101 MHz, CDCl₃) δ 170.00, 156.38, 154.67, 148.61, 137.02, 127.54, 126.74, 126.29, 124.51, 123.83, 120.68, 109.85, 89.57, 68.28, 55.28, 39.21, 29.20, 25.47. **HRMS** Calcd for C₁₈H₂₁N₂O₃ [M+H⁺]: 313.1547; Found: 313.1547.



Compound **16** was isolated in 90% yield following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 7.87 – 7.78 (m, 2H), 7.75 – 7.66 (m, 2H), 6.03 (dd, *J* = 7.9, 4.9 Hz, 1H), 4.18 (q, *J* = 7.7 Hz, 1H), 3.94 (td, *J* = 7.8, 4.6 Hz, 1H), 2.60 – 2.47 (m, 1H), 2.43 – 2.32 (m, 1H), 2.32 – 2.21 (m, 1H), 2.08 – 1.91 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 167.96, 134.26, 132.03, 123.45, 80.98, 69.90, 29.21, 26.13. HRMS Calcd for C₁₂H₁₂NO₃ [M+H⁺]: 218.0812; Found: 218.0815.



Compound **17** was isolated in 59% yield (Crude ¹H-NMR yield: 78%) following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 8.02 (s, 1H), 7.86 – 7.76 (m, 1H), 7.50 – 7.41 (m, 1H), 7.29 (dt, *J* = 9.8, 5.1 Hz, 2H), 6.18 (t, *J* = 4.1 Hz, 1H), 4.19 (q, *J* = 7.3 Hz, 1H), 4.05 (q, *J* = 7.8 Hz, 1H), 2.53 – 2.30 (m, 2H), 2.24 – 1.99 (m, 2H).⁴



Compound **17**' was isolated in 20% yield following the condition A. ¹H NMR (400 MHz, CDCl₃) δ 7.71 (d, J = 7.4 Hz, 1H), 7.55 – 7.41 (m, 1H), 7.25 – 7.17 (m, 2H), 6.21 (t, J = 6.5 Hz, 1H), 4.39 (q, J = 15.2, 6.9 Hz, 1H), 4.09 (q, J = 13.7, 8.0 Hz, 1H), 2.49 – 2.20 (m, 4H).⁴



Compound **18** was isolated in 32% yield following the condition A. ¹H **NMR** (400 MHz, CDCl₃) δ 8.07 (d, *J* = 7.7 Hz, 2H), 7.51 (d, *J* = 8.3 Hz, 2H), 7.47 – 7.38 (m, 2H), 7.28 – 7.17 (m, 2H), 6.48 (t, *J* = 6.7 Hz, 1H), 4.49 – 4.30 (m, 1H), 4.13 – 4.00 (m, 1H), 2.56 – 2.44 (m, 1H), 2.42 – 2.28 (m, 2H), 2.28 – 2.17 (m, 1H).⁵



Compound **19**' was isolated in 35% yield and compound **19**" was isolated in 25% yield following the condition A.

Compound **19':** ¹**H NMR** (400 MHz, CDCl₃) δ 7.88 – 7.78 (m, 2H), 7.58 (d, *J* = 2.4 Hz, 1H), 7.40 (t, *J* = 7.6 Hz, 2H), 7.30 (t, *J* = 7.4 Hz, 1H), 6.58 (d, *J* = 2.4 Hz, 1H), 6.03 (dd, *J* = 6.6, 2.5 Hz, 1H), 4.24 - 4.12 (m, 1H), 4.01 (q, *J* = 7.4 Hz, 1H), 2.73 – 2.62 (m, 1H), 2.42 – 2.28 (m, 1H), 2.29 – 2.16 (m, 1H), 2.12 – 1.96 (m, 1H).⁶

Compound **19":** ¹**H NMR** (400 MHz, CDCl₃) δ 7.62 – 7.52 (m, 3H), 7.50 – 7.37 (m, 3H), 6.32 (d, *J* = 1.8 Hz, 1H), 5.97 (dd, *J* = 7.3, 3.3 Hz, 1H), 4.21 (q, *J* = 7.5 Hz, 1H),

4.03 - 3.90 (m, 1H), 2.84 – 2.68 (m, 1H), 2.56 – 2.40 (m, 1H), 2.31 – 2.15 (m, 1H), 2.11 – 1.93 (m, 1H).⁶



Compound **20**' was isolated in 80% yield and compound **20**'' was isolated in 9% yield following the condition A.

Compound **20':** ¹**H NMR** (400 MHz, CDCl₃) δ 8.05 (d, *J* = 8.4 Hz, 1H), 7.70 (d, *J* = 8.3 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 1H), 7.37 (t, *J* = 7.6 Hz, 1H), 6.50 (dd, *J* = 6.8, 2.3 Hz, 1H), 4.15 – 3.97 (m, 2H), 3.23 – 3.06 (m, 1H), 2.57 – 2.46 (m, 1H), 2.45 – 2.31 (m, 1H), 2.24 – 2.10 (m, 1H).⁶

Compound **20":** ¹**H NMR** (400 MHz, CDCl₃) δ 7.92 – 7.82 (m, 2H), 7.44 – 7.31 (m, 2H), 6.60 (dd, *J* = 6.4, 2.2 Hz, 1H), 4.40 – 4.28 (m, 1H), 4.21 – 4.07 (m, 1H), 2.85 – 2.67 (m, 1H), 2.61 – 2.42 (m, 2H), 2.21 – 2.09 (m, 1H).⁶

5.2 Halogen-bond-promoted reaction system for benzyl hydrocarbon C(sp³)-H amidation



Scheme S5. Halogen-bond-promoted reaction system for benzyl hydrocarbon C(sp³)-H amidation

General conditions B: Amide (0.2 mmol, 1.0 equiv), *i*-C₃F₇I (0.4 mmol, 2.0 equiv), arenes (2.0 mmol, 10.0 equiv) and NaO*t*Bu (0.6 mmol, 3.0 equiv) were dispersed in 1 mL of PhCF₃. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 60 °C for 24 hours. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash

chromatography to give the desired product.



Compound **21** was isolated in 79% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 7.5 Hz, 2H), 7.58 (t, *J* = 7.3 Hz, 1H), 7.52 (t, *J* = 7.5 Hz, 2H), 7.36 – 7.22 (m, 5H), 4.34 (s, 2H), 3.19 – 2.98 (m, 2H), 1.38 – 1.23 (m, 2H), 1.22 – 1.01 (m, 6H), 0.80 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.33, 136.57, 132.52, 129.19, 128.64, 128.36, 127.84, 127.21, 51.86, 48.13, 31.32, 27.92, 26.37, 22.52, 14.05. **HRMS** Calcd for C₁₉H₂₆NO₂S [M+H⁺]: 332.1679; Found: 332.1673.



Compound **22** was isolated in 33% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.93 – 7.80 (m, 2H), 7.63 – 7.55 (m, 1H), 7.55 – 7.47 (m, 2H), 7.33 – 7.18 (m, 5H), 5.19 (q, *J* = 7.0 Hz, 1H), 3.10 – 2.90 (m, 2H), 1.46 – 1.34 (m, 4H), 1.21 – 1.09 (m, 3H), 1.10 – 0.98 (m, 4H), 0.79 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 141.52, 140.34, 132.38, 129.15, 128.43, 127.73, 127.67, 127.20, 55.48, 44.48, 31.30, 30.76, 26.66, 22.56, 17.01, 14.07. HRMS Calcd for C₂₀H₂₈NO₂S [M+H⁺]: 346.1835; Found: 346.1839.



Compound **23** was isolated in 75% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.89 – 7.80 (m, 2H), 7.58 (t, *J* = 7.3 Hz, 1H), 7.52 (t, *J* = 7.4 Hz, 2H),

7.13 (q, J = 8.0 Hz, 4H), 4.30 (s, 2H), 3.20 – 2.95 (m, 2H), 2.33 (s, 3H), 1.39 – 1.25 (m, 2H), 1.22 – 1.01 (m, 6H), 0.80 (t, J = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.40, 137.51, 133.38, 132.45, 129.29, 129.15, 128.35, 127.18, 51.49, 47.84, 31.31, 27.85, 26.35, 22.51, 21.21, 14.04. **HRMS** Calcd for C₂₀H₂₈NO₂S [M+H⁺]: 346.1835; Found: 346.1836.



Compound **24** was isolated in 63% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.91 – 7.82 (m, 2H), 7.64 – 7.57 (m, 1H), 7.54 (t, *J* = 7.4 Hz, 2H), 7.23 – 7.08 (m, 4H), 4.32 (s, 2H), 3.13 – 2.92 (m, 2H), 2.37 (s, 3H), 1.27 – 0.96 (m, 8H), 0.79 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 139.58, 137.23, 133.90, 132.57, 130.68, 129.31, 129.19, 128.03, 127.34, 126.06, 50.73, 48.28, 31.26, 28.18, 26.37, 22.47, 19.34, 14.03. **HRMS** Calcd for C₂₀H₂₈NO₂S [M+H⁺]: 346.1835; Found: 346.1836.



Compound **25** was isolated in 67% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.90 – 7.77 (m, 2H), 7.63 – 7.55 (m, 1H), 7.52 (t, *J* = 7.4 Hz, 2H), 6.89 (s, 1H), 6.81 (s, 2H), 4.28 (s, 2H), 3.18 – 2.95 (m, 2H), 2.26 (s, 6H), 1.43 – 1.26 (m, 2H), 1.25 – 1.01 (m, 6H), 0.81 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.55, 138.12, 136.22, 132.43, 129.39, 129.13, 127.18, 126.12, 51.56, 47.82, 31.35, 27.78, 26.37, 22.54, 21.31, 14.06. **HRMS** Calcd for C₂₁H₃₀NO₂S [M+H⁺]: 360.1992; Found: 360.1991.



Compound **26** was isolated in 90% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.88 – 7.76 (m, 2H), 7.57 (t, *J* = 7.3 Hz, 1H), 7.51 (t, *J* = 7.4 Hz, 2H), 7.18 (d, *J* = 8.6 Hz, 2H), 6.83 (d, *J* = 8.6 Hz, 2H), 4.27 (s, 2H), 3.78 (s, 3H), 3.13 – 2.98 (m, 2H), 1.38 – 1.22 (m, 2H), 1.22 – 1.02 (m, 6H), 0.80 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.31, 140.41, 132.45, 129.72, 129.14, 128.43, 127.17, 113.99, 55.36, 51.25, 47.80, 31.31, 27.93, 26.36, 22.51, 14.03. **HRMS** Calcd for C₂₀H₂₈NO₃S [M+H⁺]: 362.1784; Found: 362.1782.



Compound **27** was isolated in 48% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.86 – 7.77 (m, 2H), 7.63 (d, *J* = 8.3 Hz, 2H), 7.60 – 7.56 (m, 1H), 7.52 (t, *J* = 7.4 Hz, 2H), 7.03 (d, *J* = 8.3 Hz, 2H), 4.27 (s, 2H), 3.17 – 2.94 (m, 2H), 1.37 – 1.22 (m, 2H), 1.21 – 1.02 (m, 6H), 0.80 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.08, 137.74, 136.52, 132.66, 130.21, 129.26, 127.18, 93.35, 51.43, 48.39, 31.32, 27.96, 26.36, 22.53, 14.07. HRMS Calcd for C₁₉H₂₅INO₂S [M+H⁺]: 458.0645; Found: 458.0640.



Compound **28** was isolated in 33% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, *J* = 7.4 Hz, 2H), 7.66 – 7.43 (m, 5H), 7.31 (t, *J* = 7.5 Hz, 1H), 7.14 (t, *J* = 7.5 Hz, 1H), 4.46 (s, 2H), 3.23 – 3.07 (m, 2H), 1.39 – 1.26 (m, 2H), 1.22 –

1.04 (m, 6H), 0.80 (t, J = 7.0 Hz, 3H). ¹³**C NMR** (101 MHz, CDCl₃) δ 140.05, 136.13, 132.75, 132.69, 130.19, 129.29, 129.18, 127.85, 127.26, 123.04, 51.77, 49.32, 31.36, 28.14, 26.47, 22.54, 14.06. **HRMS** Calcd for C₁₉H₂₅BrNO₂S [M+H⁺]: 410.0784; Found: 410.0784.



Compound **29** was isolated in 32% yield following the condition B. ¹H NMR (400 MHz, CDCl₃) δ 7.90 – 7.75 (m, 2H), 7.58 (t, *J* = 7.3 Hz, 1H), 7.55 – 7.42 (m, 3H), 7.31 – 7.20 (m, 1H), 7.12 (t, *J* = 7.2 Hz, 1H), 7.04 – 6.93 (m, 1H), 4.42 (s, 2H), 3.25 – 3.00 (m, 2H), 1.43 – 1.27 (m, 2H), 1.23 – 1.02 (m, 6H), 0.81 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 162.04, 159.59, 140.15, 132.59, 130.94, 130.90, 129.57, 129.49, 129.20, 127.22, 124.52, 124.49, 123.87, 123.73, 115.42, 115.20, 48.77, 44.87, 44.82, 31.35, 28.03, 26.40, 22.54, 14.06. ¹⁹F NMR (376 MHz, CDCl₃) δ -119.50. HRMS Calcd for C₁₉H₂₅BrNO₂S [M+Na⁺]: 372.1404; Found: 372.1406.

6. General procedure and substrate scope of halogen-bond-promoted reactions for C(sp²)-H iodination



Scheme S6. Halogen-bond-promoted reactions for C(sp²)-H iodination

Conditions C: Heteroarene (0.2 mmol, 1.0 equiv), C_4F_9I (0.4 mmol, 2.0 equiv) and NaO*t*Bu (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.

Conditions D: Heteroarene (0.2 mmol, 1.0 equiv), C_4F_9I (0.8 mmol, 4.0 equiv) and NaO*t*Bu (0.8 mmol, 4.0 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.



Compound **30** was isolated in 90% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 8.03 (d, *J* = 7.8 Hz, 1H), 7.88 – 7.78 (m, 1H), 7.50 – 7.34 (m, 2H).⁷



Compound **31** was isolated in 56% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 7.96 (dd, J = 8.9, 4.8 Hz, 1H), 7.52 (dd, J = 7.9, 2.3 Hz, 1H), 7.17 (td, J = 8.9, 2.3 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 162.07, 159.61, 151.15, 151.14, 140.20, 140.09, 123.63, 123.54, 115.30, 115.06, 107.10, 106.83, 104.56, 104.54. ¹⁹F NMR (376 MHz, CDCl₃) δ -114.83. HRMS Calcd for C₇H₄FINS [M+H⁺]: 279.9088; Found: 279.9080.



Compound **32** was isolated in 60% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 7.99 (s, 1H), 7.88 (d, J = 8.7 Hz, 1H), 7.55 (d, J = 8.7 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 153.28, 140.82, 130.07, 123.67, 123.14, 119.93, 106.16. HRMS Calcd for C₇H₄BrINS [M+H⁺]: 339.8287; Found: 339.8288.



Compound **33** was isolated in 90% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 7.88 (d, J = 9.0 Hz, 1H), 7.26 (d, J = 2.5 Hz, 1H), 7.02 (dd, J = 8.9, 2.4 Hz, 1H), 3.85 (s, 3H).⁷



Compound **34** was isolated in 92% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 2.32 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 151.07, 132.95, 95.09, 14.67, 11.22.⁷



Compound **35** was isolated in 85% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 7.75 – 7.68 (m, 1H), 7.34 (dd, J = 6.8, 1.9 Hz, 1H), 7.28 – 7.19 (m, 2H), 3.77 (s, 3H).⁷



Compound **36** was isolated in 13% yield (Crude ¹H-NMR yield: 80%) following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 3.60 (s, 3H), 2.19 (s, 3H), 2.17 (s, 3H).⁷



Compound **37** was isolated in 35% yield (Crude ¹H-NMR yield: 90%) following the condition C. ¹H NMR (400 MHz, DMSO) δ 3.60 (s, 3H). ¹³C NMR (101 MHz, DMSO) δ 143.23, 120.77, 95.37, 35.71. HRMS Calcd for C₄H₄ClIN₃O₂ [M+H⁺]: 287.9031; Found: 287.9035.



Compound **38** was isolated in 74% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 7.77 – 7.66 (m, 1H), 7.57 – 7.48 (m, 1H), 7.37 – 7.28 (m, 2H).⁷



Compound **39** was isolated in 84% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 7.64 – 7.56 (m, 2H), 7.42 (t, *J* = 7.5 Hz, 2H), 7.34 (t, *J* = 7.4 Hz, 1H), 7.28 (s, 1H).⁷



Compound **40** was isolated in 85% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 3.94 (s, 3H), 3.56 (s, 3H), 3.39 (s, 3H).⁷



Compound **41** was isolated in 87% yield following the condition D. ¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.39 (m, 2H), 7.25 – 7.17 (m, 2H), 2.22 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 158.04, 129.04, 124.23, 123.22, 122.81, 118.68, 110.97, 97.55, 10.60. HRMS Calcd for C₉H₈IO [M+H⁺]: 258.9614; Found: 258.9617.



Compound **42** was isolated in 84% yield following the condition C. ¹H NMR (400 MHz, CDCl₃) δ 6.94 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 138.92. HRMS Calcd for C₄H₃I₂S [M+H⁺]: 336.8039; Found: 336.8043.



Compound **43** was isolated in 86% yield following the condition D. ¹³C NMR (101 MHz, CDCl₃) δ 119.21, 102.04, 94.55, 87.80. HRMS Calcd for C₄HBrI₃O [M+H⁺]: 524.6339; Found: 524.6333.



Compound **43**' was isolated in 75% yield following the condition C. ¹H NMR (500 MHz, CDCl₃) δ 7.40 (s, 1H).⁷



Compound **43**" was isolated in 78% yield following the condition C. ¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, J = 1.0 Hz, 1H).⁷

7. Evaluation of anion promoters for synthesis of compound 44 under blue LED irradiation

All screening reactions were carried out at a 0.1 mmol scale in a 4 mL glass vial (Thermo Scientific, National B7999-2) sealed with PTEF cap and stirred on bench top. 4',5-Dichloro-2-isocyano-1,1'-biphenyl **44-1** (27.7 mg, 0.1 mmol, 1.0 equiv), C_4F_9I (103.8 mg, 0.3 mmol, 3.0 equiv) and inorganic salts were dispersed in THF (1 mL) and the resulting mixture was vigorously stirred at 30 °C with blue LED (25 W) irradiation for 36 hours. After removal of the solvent *in vacuo*, the resulting residue was dissolved in 1 mL of CDCl₃ along with Cl₂CHCHCl₂ (20 uL) as an internal standard for ¹H NMR analysis. The composition of reaction mixture was analyzed based on the Ar-H peak at $\delta 8.55 - 8.35$ (m, 3H) for compound **44**.

Procedure of phenanthridine synthesis using UV light (254 nm): **44-1** (27.7 mg, 0.1 mmol, 1.0 equiv), C₄F₉I (103.8 mg, 0.3 mmol, 3.0 equiv) and KOH (8.4 mg, 0.15 mmol,

1.5 equiv) were dispersed in 1 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C in **UV light (254 nm)** for 36 hours. Then, THF was removed under reduced pressure. The resulting residue was dissolved in 1 mL of CDCl₃ along with Cl₂CHCHCl₂ (20 μ L) as an external standard for ¹H NMR analysis.

Procedure of phenanthridine synthesis using sunlight: **44-1** (27.7 mg, 0.1 mmol, 1.0 equiv), $C_4F_9I(103.8 \text{ mg}, 0.3 \text{ mmol}, 3.0 \text{ equiv})$ and KOH (8.4 mg, 0.15 mmol, 1.5 equiv) were dispersed in 1 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C in **sunlight** for 9 hours. Then, THF was removed under reduced pressure. The resulting residue was dissolved in 1 mL of CDCl₃ along with Cl₂CHCHCl₂ (20 µL) as an external standard for ¹H NMR analysis.

CI	$\begin{array}{c} & CI \\ + & C_4 F_9 I \\ \hline N \\ C \\ 1 \end{array}$	anion promoter (1.5 equiv) THF, Ar, 30 °C blue LED ^a		CI N C ₄ F ₉
NaO <i>t</i> Bu	NaOH	NaH	Na ₂ CO ₃	NaNO ₃
72%	65%	53%	15%	12%
NaF	NaCl	NaBr	Nal	NaOAc
20%	15%	12%	<10%	<10%
KO <i>t</i> Bu 80%	KOH 95% (87%) ^b [CFL, 36 h 95% [UV 254nm ^c , 36 h 90% [sunlight ^d , 9 h]	LiO <i>t</i> Bu n] 65%]	LiOH 75%	CsOH 64%
<i>n</i> -Bu₄NF	<i>n</i> -Bu₄NCI	<i>n-</i> Bu ₄ NBr	<i>n</i> -Bu₄NI	<i>n</i> −Bu₄N(OH)
81%	38%	10%	<10%	85%

Scheme S7. Reaction optimization for the halogen-bond-promoted perfluoroalkylation reaction

a) Yields are based on ¹H-NMR analysis of crude reaction mixture on a 0.1 mmol scale; b) Isolated yield on 0.2 mmol scale. c) UV (254 nm): low-pressure Hg-vapor lamp, 25 W. d) Sunlight: direct irradiation for 6 h.

8. General procedures and substrate scope of phenanthridine and isoquinoline synthesis



Scheme S8. Halogen-bond-promoted reaction system for the synthesis of perfluoroalkylated phenanthridines and isoquinolines

Conditions E: 2-Isocyanobiaryl compound (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (0.6 mmol, 3.0 equiv) and KOH (0.3 mmol, 1.5 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of blue LED (25W) for 36 hours. Then, the mixture was concentrated in vacuo, and the residue was purified by silica gel flash chromatography to give the desired product.

Conditions F: 2-Isocyanobiaryl compound (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (1.2 mmol, 6.0 equiv) and KOH (0.6 mmol, 3.0 equiv) were dispersed in 1 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of blue LED (25W) for 36 h. Then, the mixture was concentrated in vacuo, and the residue was purified by silica gel flash chromatography to give the desired product.



Compound **44** was isolated in 87% yield (Crude ¹H-NMR yield: 95%) following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.55 – 8.35 (m, 3H), 8.20 – 8.13 (m, 1H), 7.87 –7.83 (m, 1H), 7.76 – 7.71 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 145.93 (t), 140.06, 136.38, 135.22, 132.83, 132.32, 131.38, 130.48, 125.65, 125.36, 124.37, 123.90, 121.71. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.99 (t, *J* = 10.6 Hz, 3F), -105.07 (dd, *J* = 21.2, 7.8 Hz, 2F), -119.87 – -119.94 (m, 2F), -122.56 – -125.30 (m, 2F). HRMS

Calcd for $C_{17}H_7Cl_{12}F_9N$ [M+H⁺]: 465.9806; Found: 465.9808. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **45** was isolated in 80% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.66 (d, J = 8.4 Hz, 1H), 8.43 (d, J = 8.4 Hz, 1H), 8.35 (s, 1H), 8.14 (d, J = 8.3 Hz, 1H), 7.87 (dd, J = 11.3, 4.0 Hz, 1H), 7.76 – 7.69 (m, 1H), 7.61 (dd, J = 8.3, 1.5 Hz, 1H), 2.65 (s, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -81.00 (t, J = 10.7 Hz, 3F), -104.53 – -105.38 (m, 2F), -119.80 – -119.87 (m, 2F), -123.70 – -123.77 (m, 2F).^{8b}



Compound **46** was isolated in 85% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.65 (d, J = 8.4 Hz, 1H), 8.45 (d, J = 8.4 Hz, 1H), 8.19 (d, J = 9.0 Hz, 1H), 7.94 – 7.88 (m, 2H), 7.76 (t, J = 7.7 Hz, 1H), 7.43 (dd, J = 9.0, 2.6 Hz, 1H), 4.06 (s, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.98 (t, J = 10.0 Hz, 3F), -104.85 (t, J = 12.4 Hz, 2F), -119.90 (dd, J = 6.6, 2.9 Hz, 2F), -123.75 – -123.82 (m, 2F).^{8b}



Compound **47** was isolated in 68% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.64 (d, J = 8.4 Hz, 1H), 8.58 (d, J = 2.1 Hz, 1H), 8.47 (d, J = 8.4 Hz, 1H), 8.20 (d, J = 8.7 Hz, 1H), 8.00 – 7.90 (m, 1H), 7.84 – 7.78 (m, 1H), 7.75 (dd, J = 8.7, 2.2 Hz, 1H). ¹⁹F NMR (471 MHz, CDCl₃) δ -70.36 – -90.67 (m, 3F), -105.06 (dd, J = 20.6, 7.4 Hz, 2F), -115.08 – -121.77 (m, 2F), -122.27 – -129.48 (m, 2F).^{8b}



Compound **48** was isolated in 50% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.51 (dd, J = 23.5, 8.5 Hz, 2H), 7.99 –7.93 (m, 2H), 7.88 – 7.77 (m, 1H), 7.35 – 7.20 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 163.70, 163.58, 161.41, 161.27, 161.20, 161.08, 158.79, 158.66, 146.32, 131.88, 129.48, 126.60, 123.51, 123.24, 105.40, 105.18, 105.12, 104.89, 103.23, 103.18, 103.00, 102.95. ¹⁹F NMR (471 MHz, CDCl₃) δ -81.08 (dd, J = 22.4, 10.7 Hz, 3F), -102.56 – -107.69 (m, 3F), -114.43 – -117.48 (m, 1F), -119.82 (dd, J = 11.8, 8.2 Hz, 2F), -122.70–-125.83 (m, 2F). HRMS Calcd for C₁₇H₇F₁₁N [M+H⁺]: 434.0397; Found: 434.0399. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **49** was isolated in 55% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.73 (dd, J = 9.2, 5.4 Hz, 1H), 8.61 – 8.52 (m, 1H), 8.36 – 8.21 (m, 1H), 8.09 (d, J = 10.1 Hz, 1H), 7.92 – 7.73 (m, 2H), 7.72 – 7.53 (m, 1H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.97 (t, J = 10.6 Hz, 3F), -105.64 (dd, J = 21.2, 7.8 Hz, 2F), -109.36 – -110.55 (m, 1F), -119.67 – -120.31 (m, 2F), -123.46 – -124.21 (m, 2F).^{8b}



Compound **50** was isolated in 81% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.71 – 8.46 (m, 2H), 8.41 (s, 1H), 8.32 – 8.14 (m, 1H), 7.94 – 7.68 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 145.68 (t), 141.69, 134.37, 132.41, 131.97, 131.40,

130.01, 129.76, 125.49, 124.34, 124.29, 123.69, 121.96. ¹⁹**F NMR** (471 MHz, CDCl₃) δ -80.98 (t, J = 10.6 Hz, 3F), -100.39 – -110.24 (m, 2F), -114.74 – -121.83 (m, 2F), -122.14 – -129.43 (m, 2F). **HRMS** Calcd for C₁₇H₈ClF₉N [M+H⁺]: 432.0196; Found: 432.0190. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **51** was isolated in 77% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.85 (d, J = 8.8 Hz, 1H), 8.74 (s, 1H), 8.67 – 8.60 (m, 1H), 8.36 – 8.26 (m, 1H), 8.12 (dd, J = 8.7, 1.2 Hz, 1H), 7.92 – 7.85 (m, 2H). ¹⁹F NMR (471 MHz, CDCl₃) δ -62.65 (s, 3F), -80.08 – -82.48 (m, 3F), -103.11 – -106.97 (m, 2F), -119.85 – -119.92 (m, 2F), -122.41 – -125.81 (m, 2F).³ⁱ



Compound **52** was isolated in 84% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.76 (d, *J* = 8.7 Hz, 1H), 8.66 (s, 1H), 8.61 (dd, *J* = 6.8, 2.8 Hz, 1H), 8.28 (dd, *J* = 6.0, 3.5 Hz, 1H), 8.15 (dd, *J* = 8.6, 1.6 Hz, 1H), 7.90 – 7.77 (m, 2H), 7.76 – 7.72 (m, 2H), 7.56 (t, *J* = 7.7 Hz, 2H), 7.47 (t, *J* = 7.4 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 146.83 (t), 141.75, 140.89, 140.00, 133.04, 131.30, 130.59, 129.62, 129.36, 129.32, 128.33, 127.59, 124.74, 124.09, 123.42, 123.27, 122.12. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.93 (t, *J* = 10.8 Hz, 3F), -103.93 – -105.33 (m, 2F), -118.06 – -120.81 (m, 2F), -123.49 (d, *J* = 8.7 Hz, 2F). HRMS Calcd for C₁₇H₈ClF₉N [M+H⁺]: 474.0899; Found: 474.0900. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **53** was isolated in 40% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 9.05 (s, 1H), 8.81 (d, *J* = 8.7 Hz, 1H), 8.67 (d, *J* = 7.9 Hz, 1H), 8.49 (dd, *J* = 8.7, 1.4 Hz, 1H), 8.31 (dd, *J* = 8.0, 1.0 Hz, 1H), 7.89 (dd, *J* = 8.0, 1.6 Hz, 2H), 2.79 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 196.93, 147.11 (t), 142.58, 136.90, 135.99, 131.44, 130.66, 130.03, 129.55, 127.40 (t), 124.17, 123.35, 122.73, 122.47, 26.62. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.93 (t, *J* = 10.7 Hz, 3F), -102.41 – -107.10 (m, 2F), -119.77 – -119.83 (m, 2F), -122.33 – -126.76 (m, 2F). HRMS Calcd for C₁₉H₁₁F₉NO [M+H⁺]: 440.0691; Found: 440.0697. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **54** was isolated in 30% yield following the conditions E. ¹**H** NMR (500 MHz, CDCl₃) δ 9.35 (s, 1H), 8.14 (d, *J* = 8.3 Hz, 1H), 8.09 (d, *J* = 8.7 Hz, 1H), 7.69 (t, *J* = 8.2 Hz, 1H), 7.62 (dd, *J* = 8.3, 1.6 Hz, 1H), 7.37 (d, *J* = 8.0 Hz, 1H), 4.17 (s, 3H), 2.66 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 158.53, 140.69, 139.56, 130.76, 130.36, 128.09, 127.68, 125.08, 124.65, 124.56, 118.20, 112.01, 56.02, 22.79. ¹⁹F NMR (471 MHz, CDCl₃) δ -81.06 (t, *J* = 10.8 Hz, 3F), -104.10 (t, *J* = 13.6 Hz, 2F), -119.21 (dd, *J* = 18.1, 7.8 Hz, 2F), -122.71 - -123.47 (m, 2F). HRMS Calcd for C₁₉H₁₃F₉NO [M+H⁺]: 442.0848; Found: 442.0845. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **55** was isolated in 35% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.71 (s, 1H), 8.16 (d, J = 8.3 Hz, 1H), 8.00 – 7.97 (m, 1H), 7.66 (d, J = 8.3 Hz, 1H), 7.50 – 7.34 (m, 1H), 2.66 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 162.68, 162.54, 161.82, 161.68, 160.09, 159.98, 159.33, 159.18, 144.37 (t), 141.25, 140.14, 131.33, 131.10, 126.02, 125.78, 125.07, 122.09, 120.41, 120.30, 108.23, 107.96, 107.68, 22.58. ¹⁹F NMR (471 MHz, CDCl₃) δ -81.03 (t, J = 10.7 Hz, 3F), -101.13 – 103.72 (m, 1F), -104.05 – -106.43 (m, 2F), -106.76 – -109.47 (m, 1F), -117.20 – -121.53 (m, 2F), -121.98 – -125.87 (m, 2F). HRMS Calcd for C₁₈H₉F₁₁N [M+H⁺]: 448.0554; Found: 448.0554. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **56** was isolated in 80% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.71 (d, J = 8.4 Hz, 1H), 8.61 (dd, J = 6.3, 3.1 Hz, 1H), 8.47 (d, J = 8.4 Hz, 1H), 8.31 – 8.25 (m, 1H), 7.91 (t, J = 7.7 Hz, 1H), 7.83 – 7.73 (m, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.99 (t, J = 10.7 Hz, 3F), -102.58 – -106.65 (m, 2F), -119.77 – -119.84 (m, 2F), -123.65 – -123.72 (m, 2F).^{8b}



Compound **57** was isolated in 70% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 9.37 (d, *J* = 8.3 Hz, 1H), 9.11 (dd, *J* = 4.3, 1.6 Hz, 1H), 8.55 (dd, *J* = 8.3, 1.6 Hz, 1H), 8.47 (d, *J* = 8.5 Hz, 1H), 8.01 (t, *J* = 7.7 Hz, 1H), 7.91 – 7.85 (m, 1H), 7.75 (dd, *J* = 8.3, 4.3 Hz, 1H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.95 (dd, *J* = 16.7, 6.2 Hz, 3F), -104.69 – -105.95 (m, 2F), -119.34 – -120.90 (m, 2F), -123.21 – -125.04 (m, 2F).^{8b}



Compound **58** was isolated in 56% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.12 – 8.02 (m, 2H), 7.92 – 7.88 (m, 1H), 7.67 – 7.60 (m, 1H), 7.54 – 7.48 (m, 1H), 7.14 (d, J = 3.0 Hz, 1H), 6.98 (dd, J = 4.1, 2.8 Hz, 1H). ¹⁹F NMR (471 MHz, CDCl₃) δ -79.69 – -82.52 (m, 3F), -112.89 – -114.33 (m, 2F), -121.08 – -122.88 (m, 2F), -124.64 – -126.52 (m, 2F).^{8b}



Compound **59** was isolated in 52% yield following the conditions E. ¹H NMR (400 MHz, CDCl₃) δ 8.53 (d, J = 8.4 Hz, 1H), 8.47 (d, J = 8.7 Hz, 1H), 8.11 (dd, J = 8.0, 1.4 Hz, 1H), 8.01 (d, J = 8.0 Hz, 1H), 7.77 – 7.71 (m, 1H), 7.65 – 7.58 (m, 1H), 7.54 – 7.41 (m, 3H). ¹⁹F NMR (376 MHz, CDCl₃) δ -78.47 – -83.51 (m, 3F), -110.94 – -115.05 (m, 2F), -120.61 – -122.93 (m, 2F), -124.01 – -126.61 (m, 2F).^{8b}



Compound **60** was isolated in 72% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.74 (d, J = 8.4 Hz, 1H), 8.65 – 8.61 (m, 1H), 8.47 (d, J = 8.3 Hz, 1H), 8.31 – 8.27 (m, 1H), 7.95 – 7.90 (m, 1H), 7.84 – 7.74 (m, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -78.48 – -79.64 (m, 3F), -105.51 – -106.41 (m, 2F), -123.26 – -124.07 (m, 2F).^{8b}



Compound **61** was isolated in 83% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.73 (d, J = 8.4 Hz, 1H), 8.65 – 8.56 (m, 1H), 8.47 (d, J = 8.4 Hz, 1H), 8.33 – 8.25 (m, 1H), 7.98 – 7.86 (m, 1H), 7.86 – 7.72 (m, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.04 – -81.63 (m, 3F), -104.48 – -105.62 (m, 2F), -118.46 – -119.35 (m, 2F), -119.92 (dd, J = 16.8, 6.3 Hz, 2F), -121.95 – -123.21 (m, 2F), -125.41 – -126.99 (m, 2F).^{8b}



Compound **62** was isolated in 85% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.73 (d, J = 8.4 Hz, 1H), 8.65 – 8.60 (m, 1H), 8.47 (d, J = 8.4 Hz, 1H), 8.33 – 8.26 (m, 1H), 7.95 – 7.89 (m, 1H), 7.84 – 7.74 (m, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.77 (t, J = 10.8 Hz, 3F), -104.66 – -105.26 (m, 2F), -119.03 (dd, J = 17.8, 14.2 Hz, 2F), -119.75 (s, 2F), -121.04 – -122.47 (m, 4F), -122.73 (d, J = 7.0 Hz, 2F), -125.79 – -126.73 (m, 2F).^{8b}



Compound **63** was isolated in 85% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.73 (d, J = 8.4 Hz, 1H), 8.67 – 8.57 (m, 1H), 8.47 (d, J = 8.4 Hz, 1H), 8.34 – 8.23 (m, 1H), 7.92 (t, J = 7.7 Hz, 1H), 7.85 – 7.67 (m, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.75 (dd, J = 53.9, 43.2 Hz, 3F), -104.31 – -105.78 (m, 2F), -119.03 (s, 2F), -119.65 (d, J = 100.8 Hz, 7F), -120.47 – -122.61 (m, 3F), -122.75 (s, 2F), -125.44 – -127.24(m, 2F).^{8b}



2-Isocyanobiaryl compound (0.2 mmol, 1.0 equiv) and KOH (0.3 mmol, 1.5 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with CF₃I for 5 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of blue LED (25W) for 36 hours. Then, the mixture was concentrated in vacuo, and the residue was purified by silica gel flash chromatography to give the compound **64** in 65% isolated yield. ¹H NMR (400 MHz, CDCl₃) δ 8.71 (d, *J* = 8.4 Hz, 1H), 8.64 – 8.59 (m, 1H), 8.43 – 8.36 (m, 1H), 8.34 – 8.26 (m, 1H), 7.98 – 7.90 (m, 1H), 7.86 – 7.74 (m, 3H). ¹⁹F NMR (376 MHz, CDCl₃) δ -63.44 (d, *J* = 2.8 Hz, 3F).^{8b}



Compound **65** was isolated in 80% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.75 (d, J = 8.4 Hz, 1H), 8.65 – 8.60 (m, 2H), 8.27 – 8.19 (m, 1H), 7.91 (t, J = 7.7 Hz, 1H), 7.83 – 7.73 (m, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -72.50 (d, J = 6.1 Hz, 6F), -175.04 – -175.26 (m, 1F).^{8b}



Compound **66** was isolated in 78% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.48 (d, J = 8.5 Hz, 1H), 7.83 – 7.70 (m, 3H), 7.57 – 7.48 (m, 3H), 7.41 – 7.31 (m, 2H), 3.71 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.61, 145.53 (t), 140.70, 137.28, 137.16, 135.03, 131.30, 129.91, 129.64, 128.73, 128.55, 127.66, 126.68, 125.09 (t), 52.69. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.93 (dd, J = 16.8, 5.9 Hz, 3F), -103.24 – -108.07 (m, 2F), -118.07 – -122.24 (m, 2F), -122.69 – -128.17 (m, 2F). HRMS

Calcd for $C_{21}H_{13}F_9NO_2$ [M+H⁺]: 482.0797; Found: 482.0799. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **67** was isolated in 54% yield following the conditions F. ¹H NMR (500 MHz, CDCl₃) δ 8.42 (d, *J* = 8.6 Hz, 1H), 8.26 (d, *J* = 8.5 Hz, 1H), 7.91 – 7.85 (m, 1H), 7.81 – 7.78 (m, 1H), 4.03 (s, 3H), 2.92 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 167.23, 144.09, 140.48, 137.43, 133.40, 131.21, 129.76, 126.30, 125.62, 125.22, 52.99, 14.92. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.40 – -81.57 (m, 3F), -104.86 – -105.97 (m, 2F), -119.67 – -121.08 (m, 2F), -123.93 – -124.93 (m, 2F). HRMS Calcd for C₁₆H₁₁F₉NO₂ [M+H⁺]: 420.0641; Found: 420.0643. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **68** was isolated in 40% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.50 – 8.42 (m, 2H), 7.82 (t, *J* = 7.5 Hz, 1H), 7.76 – 7.71 (m, 1H), 4.02 (s, 3H), 2.13 – 1.83 (m, 8H), 1.53 – 1.37 (m, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 168.59, 144.27, 142.25, 138.18, 136.60, 130.54, 128.92, 126.68, 125.83, 52.93, 31.75, 27.51, 26.18, 0.14. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.98 (t, *J* = 10.5 Hz, 3F), -103.61 – -106.65 (m, 2F), -120.18 (dd, *J*=13.7, 6.4 Hz, 2F), -122.56 – -125.98 (m, 2F). HRMS Calcd for C₂₁H₁₉F₉NO₂ [M+H⁺]: 488.1267; Found: 488.1260. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **69** was isolated in 60% yield following the conditions F. ¹H NMR (500 MHz, CDCl₃) δ 8.42 (d, *J* = 9.1 Hz, 2H), 7.85 – 7.80 (m, 1H), 7.77 – 7.72 (m, 1H), 4.02 (s, 3H), 3.84 – 3.78 (m, 1H), 1.59 (d, *J* = 7.2 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 168.43, 144.36 (t), 141.93, 139.52, 136.28, 130.55, 128.99, 126.77, 125.94, 125.59, 53.03, 29.86, 22.07. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.99 (t, *J* = 10.5 Hz, 3F), -103.98 – -105.64 (m, 2F), -119.32 – -121.11 (m, 2F), -123.06 – -125.34 (m, 2F). HRMS Calcd for C₁₈H₁₅F₉NO₂ [M+H⁺]: 448.0954; Found: 448.0956. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **70** was isolated in 70% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.66 (s, 1H), 7.97 (d, *J* = 9.0 Hz, 1H), 7.65 (s, 1H), 7.49 (dd, *J* = 9.0, 2.4 Hz, 1H), 4.04 (s, 3H), 3.99 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.62, 161.27, 144.51 (t), 138.43, 133.01, 130.79, 129.35, 127.26, 124.80, 103.20, 55.77, 53.05. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.48 – -81.39 (m, 3F), -106.34 – -107.16 (m, 2F), -120.03 – -121.25 (m, 2F), -124.13 – -125.37 (m, 2F). HRMS Calcd for C₁₈H₁₅F₉NO₂ [M+H⁺]: 436.0590; Found: 436.0599. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

$$MeO \xrightarrow{V} COOCH_3$$

$$MeO \xrightarrow{V} (CF_2)_4F$$
71

Compound **71** was isolated in 72% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.58 (s, 1H), 7.64 (s, 1H), 7.27 (s, 1H), 4.07 (s, 3H), 4.06 (s, 3H), 4.03 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.78, 153.64, 152.99, 143.72 (t), 139.16, 134.70, 125.90, 124.32, 106.65, 103.39, 56.44, 56.29, 53.04. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.90 (dd, *J* = 16.8, 6.3 Hz, 3F), -106.56 – -107.04 (m, 2F), -120.37 – 120.95 (m, 2F), -124.35 – -125.01 (m, 2F). HRMS Calcd for C₁₇H₁₃F₉NO₄ [M+H⁺]: 466.0695; Found: 466.0698. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **72** was isolated in 72% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.20 (s, 1H), 7.67 (d, *J* = 8.7 Hz, 1H), 7.54 (d, *J* = 8.7 Hz, 1H), 7.33 (d, *J* = 7.9 Hz, 2H), 7.22 (d, *J* = 7.9 Hz, 2H), 3.74 (s, 3H), 2.61 (s, 3H), 2.47 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.79, 144.50, 140.44, 139.87, 138.44, 137.31, 135.75, 133.42, 132.18, 129.50, 129.24, 127.52, 126.95, 123.82, 52.67, 22.48, 21.57. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.92 (dd, *J* = 16.8, 6.3 Hz,3F), -105.11 – -105.62 (m,2F), -120.23 – -120.29 (m, 2F), -123.94 – -124.62 (m,2F). HRMS Calcd for C₂₃H₁₇F₉NO₂ [M+H⁺]: 510.1110; Found: 510.1111. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.


Compound **73** was isolated in 85% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 7.72 – 7.66 (m, 2H), 7.35 (dd, J = 9.4, 2.4 Hz, 1H), 7.28 – 7.24 (m, 2H), 7.07 – 7.02 (m, 2H), 3.98 (s, 3H), 3.90 (s, 3H), 3.74 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.87, 160.19, 159.86, 143.24 (t), 139.19, 137.17, 133.09, 130.84, 129.28, 128.48, 127.24, 124.19, 114.00, 102.77 (t), 55.69, 55.42, 52.64. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.93 (t, J = 10.6 Hz, 3F), -105.29 – -107.26 (m, 2F), -119.13 – -121.37 (m, 2F), -124.34 – -124.42 (m, 2F). HRMS Calcd for C₂₃H₁₇F₉NO₄ [M+H⁺]: 542.1008; Found: 542.1011. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F⁻¹³C coupling and are not reported.



Compound **74** was isolated in 70% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.08 (d, *J* = 10.0 Hz, 1H), 7.76 (dd, *J* = 9.4, 5.6 Hz, 1H), 7.57 – 7.51 (m, 1H), 7.33 (dd, *J* = 8.5, 5.3 Hz, 2H), 7.24 (t, *J* = 8.7 Hz, 2H), 3.75 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.21, 164.11, 163.43, 162.13, 161.41, 145.18, 140.47, 136.31, 134.55, 131.43, 131.36, 130.56, 130.53, 130.48, 127.85, 127.78, 122.33, 122.13, 116.02, 115.84, 109.53, 109.34, 52.87. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.91 (t, *J* = 10.6 Hz, 3F), -104.92 – -105.50 (m, 1F), -106.09 – -106.57 (m, 2F), -111.84 – -112.76 (m, 1F), -120.13 – -120.84 (m, 2F), -124.24 – -125.04 (m, 2F). HRMS Calcd for C₂₁H₁₁F₁₁NO₂ [M+H⁺]: 518.0609; Found: 518.0610. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **75** was isolated in 65% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.37 (s, 1H), 8.21 (d, *J* = 9.1 Hz, 1H), 7.82 (dd, *J* = 9.1, 1.9 Hz, 1H), 4.03 (s, 3H), 2.92 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.86, 140.73, 136.39, 135.82, 133.48, 132.30, 126.96, 126.88, 124.60, 100.13, 53.08, 14.93. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.94 (t, *J* = 10.4 Hz, 3F), -105.38 – -105.90 (m,2F), -120.29 – -120.91 (m,2F), -124.31 – -124.38 (m, 2F). HRMS Calcd for C₁₆H₁₀ClF₉NO₂ [M+H⁺]: 454.0251; Found: 454.0257. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **76** was isolated in 78% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, J = 9.4 Hz, 1H), 7.63 (s, 1H), 7.49 (dd, J = 9.4, 2.4 Hz, 1H), 4.01 (s, 3H), 3.98 (s, 3H), 2.90 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 167.27, 160.11, 142.16 (t), 138.65, 133.87, 132.90, 128.15, 126.90, 124.07, 103.32, 55.66, 52.86, 14.89. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.98 (t, J = 10.6 Hz, 3F), -105.51 – -106.67 (m, 2F), -119.82 – -120.78 (m, 2F), -123.89 – -124.97 (m, 2F). HRMS Calcd for C₁₇H₁₃F₉NO₃ [M+H⁺]: 450.0746; Found: 450.0749. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **77** was isolated in 62% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 7.64 (s, 1H), 7.45 (s, 1H), 6.20 (s, 2H), 4.00 (s, 3H), 2.79 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 167.40, 151.65, 150.48, 142.04, 140.09, 136.44, 132.14, 124.31, 102.67, 101.67, 101.36, 52.91, 15.32. ¹⁹F NMR (471 MHz, CDCl₃) δ -79.49 –

-82.23 (m, 3F), -105.28 - -107.34 (m, 2F), -119.94 - -121.36 (m, 2F), -123.62 - -125.92 (m, 2F). **HRMS** Calcd for C₁₇H₁₁F₉NO₄ [M+H⁺]: 464.0539; Found: 464.0531. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **78** was isolated in 69% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.94 – 8.89 (m, 1H), 8.67 (s, 1H), 8.07 (d, *J* = 8.7 Hz, 1H), 8.01 – 7.97 (m, 1H), 7.85 – 7.74 (m, 3H), 4.06 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.07, 140.41, 139.50, 134.61, 133.80, 129.39, 129.26, 129.17, 128.94, 127.78, 127.19, 127.01, 126.60, 125.72, 53.08. ¹⁹F NMR (471 MHz, CDCl₃) δ -81.30 (t, *J* = 11.2 Hz, 3F), -102.40 (t, *J* = 13.7 Hz, 2F), -115.73 – -116.77 (m, 2F), -121.21 – -122.56 (m, 2F). HRMS Calcd for C₁₉H₁₁F₉NO₂ [M+H⁺]: 456.0641; Found: 456.0641. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **79** was isolated in 56% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.08 (d, J = 9.4 Hz, 1H), 7.64 (s, 1H), 7.38 (dd, J = 9.4, 2.5 Hz, 1H), 7.24 (t, J = 7.4 Hz, 2H), 7.17 (t, J = 7.3 Hz, 1H), 7.10 (d, J = 7.1 Hz, 2H), 4.79 (s, 2H), 3.95 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 167.07, 160.07, 143.14, 139.50, 139.30, 135.20, 132.79, 128.81, 128.72, 128.34, 127.64, 126.53, 124.39, 103.42, 55.67, 52.97, 33.98. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.94 (t, J = 10.5 Hz, 3F), -105.69 – -106.93 (m, 2F), -119.71 – -120.84 (m, 2F), -123.49 – -125.01 (m, 2F). HRMS Calcd for C₂₃H₁₇F₉NO₃ [M+H⁺]: 526.1059; Found: 526.1062. The ¹³C peaks of the perfluoroalkyl

moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **80** was isolated in 76% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 7.48 (s, 1H), 7.22 – 7.20 (m, 1H), 3.99 (s, 3H), 3.95 (s, 3H), 3.44 (t, *J* = 6.2 Hz, 2H), 3.09 (t, *J* = 6.1 Hz, 2H), 2.11 – 2.04 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 166.87, 160.05, 141.66 (t), 140.28, 136.98, 136.03, 130.59, 128.91, 121.69, 101.23, 55.55, 52.70, 30.76, 27.74, 22.45. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.98 (dd, *J* = 16.8, 6.3 Hz, 3F), -105.28 – -107.34 (m, 2F), -119.56 – -121.25 (m, 2F), -123.63 – 125.30 (m, 2F). HRMS Calcd for C₁₉H₁₅F₉NO₃ [M+H⁺]: 476.0903; Found: 476.0906. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **81** was isolated in 70% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.47 (d, J = 8.6 Hz, 1H), 7.82 – 7.71 (m, 3H), 7.56 – 7.50 (m, 3H), 7.39 – 7.34 (m, 2H), 3.71 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.63, 145.42 (t), 140.76, 137.28, 137.15, 135.04, 131.30, 129.89, 129.64, 128.72, 128.54, 127.65, 126.67, 125.11 (t), 52.71. ¹⁹F NMR (471 MHz, CDCl₃) δ -79.43 (t, J = 10.1 Hz, 3F), -105.81 – -107.08 (m, 2F), -123.71 – -124.74 (m, 2F). HRMS Calcd for C₂₀H₁₃F₇NO₂ [M+H⁺]: 432.0829; Found: 432.0823. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **82** was isolated in 77% yield following the conditions E. ¹**H** NMR (500 MHz, CDCl₃) δ 8.48 (d, J = 8.5 Hz, 1H), 7.84 – 7.70 (m, 3H), 7.60 – 7.50 (m, 3H), 7.41 – 7.34 (m, 2H), 3.72 (s, 3H). ¹³**C** NMR (126 MHz, CDCl₃) δ 166.62, 145.56 (t), 140.76, 137.31, 137.17, 135.05, 131.28, 129.91, 129.66, 128.73, 128.55, 127.66, 126.75, 125.12 (t), 52.68. ¹⁹**F** NMR (471 MHz, CDCl₃) δ -78.24 – -83.38 (m, 3F), -104.04 – 107.36 (m, 2F), -117.60 – -119.94 (m, 2F), -120.42 (d, J=90.4 Hz, 2F), -122.68 (s, 2F), -124.77 – -131.75 (m, 2F). **HRMS** Calcd for C₂₃H₁₃F₁₃NO₂ [M+H⁺]: 582.0733; Found: 582.0736. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **83** was isolated in 78% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.48 (d, J = 8.5 Hz, 1H), 7.80 – 7.71 (m, 3H), 7.55 – 7.51 (m, 3H), 7.38 – 7.34 (m, 2H), 3.71 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.62, 145.58, 140.77 (t), 137.32, 137.16, 135.06, 131.28, 129.91, 129.66, 128.73, 128.55, 127.66, 126.76, 125.13, 52.67. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.81 (t, J = 10.7 Hz, 3F), -105.35 – 105.80 (m,2F), -119.44 (dd, J = 23.7, 15.2 Hz, 2F), -120.34 (s,2F), -121.78 (dd, J = 37.7, 17.2 Hz,4F), -122.75 (s,2F), -125.80 – -126.39 (m,2F). HRMS Calcd for C₂₅H₁₃F₁₇NO₂ [M+H⁺]: 682.0669; Found: 682.0671. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **84** was isolated in 78% yield following the conditions E. ¹**H** NMR (500 MHz, CDCl₃) δ 8.48 (d, *J* = 8.5 Hz, 1H), 7.84 – 7.70 (m, 3H), 7.59 – 7.49 (m, 3H), 7.42 – 7.34 (m, 2H), 3.72 (d, *J* = 6.6 Hz, 3H). ¹³**C** NMR (126 MHz, CDCl₃) δ 166.63, 145.58, 140.76, 137.31, 137.17, 135.06, 131.28, 129.91, 129.66, 128.73, 128.55, 127.66, 126.76, 125.13, 52.68. ¹⁹**F** NMR (471 MHz, CDCl₃) δ -80.80 (t, *J* = 10.6 Hz, 3F), -105.17 – -106.07 (m, 2F), -119.44 (s, 3F), -120.33 (s, 2F), -121.73 (dd, *J* = 94.3, 59.3 Hz, 6F), -122.75 (s, 3F), -125.64 – -126.96 (m, 2F). HRMS Calcd for C₂₇H₁₃F₂₁NO₂ [M+H⁺]: 782.0605; Found: 782.0609. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **85** was isolated in 50% yield following the conditions E. ¹H NMR (500 MHz, CDCl₃) δ 8.61 (d, *J* = 8.5 Hz, 1H), 7.79 – 7.69 (m, 3H), 7.53 (dd, *J* = 8.3, 3.1 Hz, 3H), 7.36 (dd, *J* = 7.0, 2.2 Hz, 2H), 3.70 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.52, 144.73, 144.54, 140.10, 137.50, 136.40, 135.11, 131.11, 129.76, 129.73, 129.71, 128.66, 128.52, 127.84, 125.27, 125.10, 52.62. ¹⁹F NMR (471 MHz, CDCl₃) δ -72.76 (d, *J* = 6.3 Hz, 6F), -175.28 – -175.89 (m, 1F). HRMS Calcd for C₂₀H₁₃F₇NO₂ [M+H⁺]: 432.0829; Found: 432.0837. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

9. General procedure and substrate scope for addition of perfluorobutyl iodide to alkenes and alkynes



Scheme 9 Addition of perfluoroalkyl iodide to alkenes and alkynes

Conditions G: Substrate (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (0.6 mmol, 3.0 equiv) and KOH (0.3 mmol, 1.5 equiv) were dispersed in 2 mL of H₂O. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30°C under irradiation of blue LED (25W) for 20 h. Then, the reaction mixture was extracted with EtOAc (3 x 2 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography to afford fluorination compounds.



Compound **86** was isolated in 66% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.12 (d, J = 9.2 Hz, 2H), 6.88 (d, J = 9.0 Hz, 2H), 4.54 – 4.35 (m, 1H), 3.81 (s, 3H), 3.27 – 3.11 (m, 2H), 3.00 – 2.52 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 158.96, 130.79, 130.18, 114.13, 55.37, 46.39, 46.37, 40.65 (t), 20.27. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.57 – -81.63 (m, 3F), -111.57 – -114.63 (m, 2F), -124.56 (d, J = 10.0 Hz, 2F), -125.65 – -126.35 (m, 2F). HRMS Calcd for C₁₄H₁₃F₉IO [M+H⁺]: 494.9862; Found: 494.9860. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **87** was isolated in 85% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.86 (dd, J = 5.4, 3.1 Hz, 2H), 7.73 (dd, J = 5.5, 3.0 Hz, 2H), 4.33 – 4.27 (m, 1H), 3.95 – 3.79 (m, 2H), 3.03 – 2.79 (m, 2H), 2.34 – 2.19 (m, 2H). ¹³C NMR

 $(126 \text{ MHz}, \text{CDCl}_3) \delta 168.28, 134.27, 132.09, 123.55, 41.63 (t), 38.83, 38.43, 14.62.$ ¹⁹**F NMR** (471 MHz, CDCl₃) δ -80.03 – -81.92 (m, 3F), -111.02 – -112.47 (m, 1F), -113.56 – -116.40 (m, 1F), -124.54 (d, J = 10.5 Hz, 2F), -125.37 – -127.69 (m, 2F). **HRMS** Calcd for C₁₆H₁₂F₉INO₂ [M+H⁺]: 547.9764; Found: 547.9767. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **88** was isolated in 81% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.99 – 7.91 (m, 2H), 7.45 – 7.38 (m, 2H), 4.60 – 4.54 (m, 1H), 4.51 – 4.39 (m, 2H), 3.11 – 2.78 (m, 2H), 2.39 – 2.32 (m, 1H), 2.28 – 2.18 (m, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 165.51, 139.84, 131.08, 128.96, 128.34, 64.96, 41.98 (t), 39.01 (d), 15.00. ¹³C NMR (126 MHz, CDCl₃) (108.32 – 119.91, m) are signal peaks of perfloro-substituted carbons (-CF₂CF₂CF₃CF₃). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.23 – -81.85 (m, 3F), -110.12 – -112.48 (m, 1F), -113.66 – -116.02 (m, 1F), -124.48 (dd, *J* = 10.0, 3.7 Hz, 2F), -125.40 – -127.28 (m, 2F). HRMS Calcd for C₁₅H₁₂ClF₉IO₂ [M+H⁺]: 556.9421; Found: 556.9420.



Compound **89** was isolated in 79% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.40 – 7.35 (m, 2H), 6.79 – 6.75 (m, 2H), 4.43 – 4.36 (m, 1H), 3.97 (t, J = 5.6 Hz, 2H), 3.02 – 2.75 (m, 2H), 2.11 – 1.88 (m, 4H). ¹³C NMR (126 MHz, CDCl₃) δ 158.03, 132.43, 116.41, 113.16, 66.94, 41.80, 37.16, 29.59, 19.82. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.26 – -81.97 (m, 3F), -110.75 – -113.06 (m, 1F), -113.87 – -116.36 (m, 1F), -123.87 – -125.02 (m, 2F), -125.49 – -127.16 (m, 2F). HRMS Calcd for C₁₅H₁₄BrF₉IO [M+H⁺]: 586.9124; Found: 586.9130. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **90** was isolated in 73% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 8.12 (d, J = 2.1 Hz, 1H), 7.88 (d, J = 7.7 Hz, 1H), 7.55 (dd, J = 10.7, 4.1 Hz, 1H), 7.48 – 7.40 (m, 2H), 7.36 (d, J = 7.4 Hz, 1H), 7.02 (d, J = 8.4 Hz, 1H), 5.18 (s, 2H), 4.35 – 4.27 (m, 1H), 4.16 (td, J = 6.3, 1.8 Hz, 2H), 3.64 (s, 2H), 2.98 – 2.70 (m, 2H), 1.95 – 1.74 (m, 4H). ¹³C NMR (126 MHz, CDCl₃) δ 190.89, 171.41, 160.63, 140.53, 136.39, 135.68, 132.90, 132.56, 129.61, 129.36, 127.93, 127.80, 125.31, 121.21, 73.75, 63.73, 41.64, 40.34, 36.93, 28.99. ¹⁹F NMR (471 MHz, CDCl₃) δ -81.01 (s, 3F), -110.65 – -112.77 (m, 1F), -113.83 – -115.47 (m, 1F), -123.79 – -125.16 (m, 2F), -125.27 – -127.43 (m, 2F). HRMS Calcd for C₂₅H₂₁F₉IO₄ [M+H⁺]: 683.0335; Found: 683.0338. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **91** was isolated in 85% yield following the conditions G (dr = 1:1). The dr ratio of compound **91** is determined by ¹⁹F NMR (see ¹⁹F NMR analysis). ¹H NMR (500 MHz, CDCl₃) δ 7.10 (d, *J* = 8.5 Hz, 2H), 6.79 (d, *J* = 8.5 Hz, 2H), 4.30-4.20 (m, 3H), 2.93 – 2.79 (m, 2H), 2.75 – 2.60 (m, 1H), 1.95 – 1.85 (m, 2H), 1.79 – 1.69 (m, 4H), 1.66 – 1.55 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 174.25, 155.11, 155.10, 129.78, 128.22, 128.19, 118.47, 118.43, 79.22, 64.15, 64.14, 41.60, 36.73, 34.91, 28.91, 25.90, 25.63, 25.57, 25.44, 25.39, 19.32. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.75 – 81.41 (m, 3F), -111.31 – -113.11 (m, 1F), -113.89 – -115.96 (m, 1F), -124.52 (s, 2F), -125.64 – -126.26 (m, 2F). HRMS Calcd for C₂₂H₂₃Cl₂F₉IO₃ [M+H⁺]: 702.9920; Found: 702.9931. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **92** isolated in 60% yield following the conditions G(dr = 1:1).

The dr ratio of compound 92 is determined by ¹H NMR (see ¹H NMR analysis).

¹**H NMR** (500 MHz, CDCl₃) δ 4.66 – 4.62 (m, 1H), 4.41 (s, 1H), 4.37 (dd, J = 8.6, 4.0 Hz, 1H), 4.29 (t, J = 6.1 Hz, 2H), 3.49 (dd, J = 5.4, 3.2 Hz, 2H), 3.06 – 2.75 (m, 2H), 2.05 – 1.85 (m, 3H), 1.64 (s, 3H), 1.45 (d, J = 2.1 Hz, 3H). ¹³**C NMR** (126 MHz, CDCl₃) δ 170.78, 167.06, 65.27, 65.21, 63.43, 62.81, 61.28, 61.27, 38.50, 38.49, 36.73, 36.64, 31.58, 30.35, 29.84, 28.95, 28.92, 20.52, 19.24, 18.99, 18.75. ¹⁹F NMR (471 MHz, CDCl3) δ -80.20 – -81.92 (m, 3F), -109.68 – -112.83 (m, 1F), -114.88 (m, 1F), -123.14 – -124.98 (m, 2F), -125.36 – -128.12 (m, 2F). **HRMS** Calcd for $C_{17}H_{20}F_9INO_5S$ [M+H⁺]: 647.9958; Found: 647.9950. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **93** was isolated in 78% yield following the conditions G (dr = 1:1). The dr ratio of compound **93** is determined by ¹H NMR and HPLC (see ¹H NMR and HPLC analysis).

¹**H NMR** (500 MHz, CDCl₃) δ 5.86 (d, J = 3.7 Hz, 1H), 5.26 (s, 1H), 4.47 (d, J = 3.7 Hz, 1H), 4.35 – 4.29 (m, 1H), 4.22 – 4.18 (m, 2H), 4.10 – 4.06 (m, 1H), 4.03 – 3.99 (m, 1H), 2.98 – 2.68 (m, 2H), 2.38 – 2.30 (m, 2H), 1.86 – 1.71 (m, 2H), 1.66 – 1.62 (m, 2H), 1.46 (d, J = 56.8 Hz, 9H), 1.31 (t, J = 6.7 Hz, 13H). ¹³**C NMR** (126 MHz, CDCl₃) δ 172.45, 112.44, 109.46, 105.22, 83.57, 80.05, 76.01, 72.61, 67.45, 41.75 (t), 40.44, 34.38, 29.67, 29.30, 29.26, 29.12, 28.58, 26.95, 26.89, 26.35, 25.43, 24.98, 20.86. ¹⁹**F NMR** (471 MHz, CDCl₃) δ -80.74 – -81.40 (m, 3F), -111.47 – -112.88 (m, 1F), -113.65

- -115.78 (m, 1F), -124.29 - -124.84 (m, 2F), -125.92 (m, 2F). **HRMS** Calcd for C₂₇H₃₇F₉IO₇ [M+H⁺]: 771.1435; Found: 771.1437. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **94** was isolated in 87% yield following the conditions G (dr = 1:1). The dr ratio of compound **94** is determined by ¹H NMR (see ¹H NMR analysis). ¹H NMR (500 MHz, CDCl₃) δ 5.51 (d, *J* = 5.0 Hz, 1H), 4.59 (dd, *J* = 7.9, 2.5 Hz, 1H), 4.32 – 4.27 (m, 3H), 4.21 (dd, *J* = 7.9, 1.9 Hz, 1H), 4.14 (dd, *J* = 11.6, 7.8 Hz, 1H), 4.02 – 3.98 (m, 1H), 2.96 – 2.68 (m, 2H), 2.32 (t, *J* = 7.5 Hz, 2H), 1.84 – 1.69 (m, 2H), 1.59 (dd, *J* = 14.4, 7.1 Hz, 2H), 1.45 (d, *J* = 27.9 Hz, 7H), 1.33 – 1.26 (m, 15H). ¹³C NMR (126 MHz, CDCl₃) δ 173.77, 173.73, 109.73, 109.71, 108.82, 108.80, 96.42, 71.22, 70.84, 70.58, 66.16, 63.35, 41.86, 41.69, 41.53, 40.43, 34.26, 34.25, 29.79, 29.62, 29.23, 29.08, 28.53, 26.10, 26.08, 26.03, 26.02, 25.03, 24.57, 24.55, 20.70. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.59 – -81.74 (m, 3F), -112.09 (dd, *J* = 272.8, 33.6 Hz, 1F), -114.01 – -116.01 (m, 1F), -124.64 (s, 2F), -125.54 – -126.64 (m, 2F). HRMS Calcd for C₂₉H₄₅F₉IO₇ [M+H⁺]: 803.2061; Found: 803.2059. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **95** was isolated in 64% yield following the conditions G (dr = 1:1). The dr ratio of compound **95** is determined by ¹H NMR (see ¹H NMR analysis). ¹H NMR (500 MHz, CDCl₃) δ 7.20 (d, *J* = 8.6 Hz, 1H), 6.71 (dd, *J* = 8.6, 2.7 Hz, 1H), 6.65 (d, *J* = 2.6 Hz, 1H), 4.38 – 4.32 (m, 1H), 3.96 (t, *J* = 6.1 Hz, 2H), 3.01 – 2.74 (m, 4H), 2.50 (dd, J = 19.0, 8.6 Hz, 1H), 2.42 – 2.37 (m, 1H), 2.29 – 2.22 (m, 1H), 2.19 – 1.71 (m, 10H), 1.63 – 1.46 (m, 6H), 0.91 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 157.08, 137.89, 132.23, 126.46, 114.73, 112.24, 67.54, 50.57, 48.15, 44.13, 41.89, 41.73, 41.56, 40.16, 38.53, 36.01, 31.74, 29.78, 28.38, 26.70, 26.62, 26.07, 21.72, 20.35, 13.99. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.57 – -81.44 (m, 3F), -111.94 (dd, J = 271.5, 42.6 Hz, 1F), -114.73 (dd, J = 272.7, 36.6 Hz, 1F), -124.52 (d, J = 5.1 Hz, 2F), -125.27 – -127.51 (m, 2F). HRMS Calcd for C₂₈H₃₃F₉IO₂ [M+H⁺]: 699.1376; Found: 699.1379. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **96** was isolated in 78% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.79 (dd, J = 7.9, 1.4 Hz, 1H), 7.58 – 7.53 (m, 1H), 7.34 – 7.27 (m, 2H), 5.67 (s, 1H), 4.46 – 4.40 (m, 1H), 4.19 (t, J = 5.6 Hz, 2H), 3.07 – 2.78 (m, 2H), 2.26 – 2.05 (m, 4H). ¹³C NMR (126 MHz, CDCl₃) δ 165.57, 162.94, 153.47, 132.62, 124.07, 122.97, 116.96, 115.69, 90.78, 68.10, 41.71 (t), 36.76, 28.93, 19.26. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.57 – -81.44 (m, 3F), -110.81 – -112.44 (m, 1F), -113.84 – -115.98 (m, 1F), -124.45 (dd, J = 9.9, 4.0 Hz, 2F), -125.49 – -126.60 (m, 2F). HRMS Calcd for C₁₈H₁₅F₉IO₃ [M+H⁺]: 576.9917; Found: 576.9921. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **97** was isolated in 71% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.63 (d, J = 9.5 Hz, 1H), 7.36 (d, J = 8.6 Hz, 1H), 6.83 (dd, J = 8.6, 2.4 Hz, 1H), 6.79 (d, J = 2.3 Hz, 1H), 6.24 (d, J = 9.5 Hz, 1H), 4.43 – 4.36 (m, 1H), 4.10 – 4.04 (m, 2H), 3.03 – 2.75 (m, 2H), 2.14 – 1.93 (m, 4H). ¹³C NMR (126 MHz, CDCl₃)

δ 162.09, 161.31, 156.01, 143.50, 128.93, 113.35, 113.03, 112.81, 101.48, 67.33, 41.78 (t), 37.04, 29.43, 19.59. ¹⁹**F NMR** (471 MHz, CDCl₃) δ -80.62 – -81.42 (m, 3F), -111.82 (dd, *J* = 276.1, 40.1 Hz, 1F), -113.75 – -115.94 (m, 1F), -124.51 (d, *J* = 10.0 Hz, 2F), -125.51 – -126.32 (m, 2F). **HRMS** Calcd for C₁₈H₁₅F₉IO₃ [M+H⁺]: 576.9917; Found: 576.9921. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **98** was isolated in 71% yield following the conditions G. ¹H **NMR** (500 MHz, CDCl₃) δ 7.99 (s, 2H), 7.50 (d, J = 8.2 Hz, 1H), 7.41 (d, J = 7.7 Hz, 1H), 7.32 (dd, J = 8.1, 7.4 Hz, 1H), 7.24 (d, J = 7.4 Hz, 1H), 4.52 – 4.44 (m, 1H), 4.21 – 4.12 (m, 2H), 3.05 – 2.82 (m, 4H), 2.27 – 2.02 (m, 4H), 1.36 (t, J = 7.5 Hz, 3H). ¹³C **NMR** (126 MHz, CDCl₃) δ 188.15, 167.09, 156.77, 153.85, 137.40, 133.78, 126.53, 124.87, 124.02, 121.16, 118.72, 115.48, 111.30, 72.32, 41.88, 37.12, 30.61, 22.18, 20.04, 12.32. ¹⁹F **NMR** (471 MHz, CDCl₃) δ -80.49 – -81.55 (m, 3F), -111.19 – -112.44 (m, 1F), -113.83 – -115.30 (m, 1F), -123.85 – -124.74 (m, 2F), -125.08 – -126.56 (m, 2F). **HRMS** Calcd for C₂₆H₂₁Br₂F₉IO₃ [M+H⁺]: 836.8753; Found: 836.8756. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **99** was isolated in 70% yield following the conditions G (dr = 1:1). The spectra data are consistent with those reported in literature.^{8b}

The dr ratio of compound 99 is determined by ¹H NMR (see ¹H NMR analysis).

¹**H NMR** (400 MHz, CDCl₃) δ 8.62 (d, J = 3.8 Hz, 1H), 8.00 (d, J = 9.1 Hz, 1H), 7.54 (d, J = 3.8 Hz, 1H), 7.44-7.32 (m, 1H), 5.65 (s, 1H), 4.11-4.02 (m, 1H), 4.01-3.90 (m, 3H), 3.59 (s, 1H), 3.30-3.09 (m, 2H), 3.08-2.78 (m, 2H), 2.70 (d, J = 11.5 Hz, 1H), 2.59 (d, J = 10.1 Hz, 1H), 2.14 (s, 2H), 1.88 (s, 2H), 1.53 (s, 1H), 1.44-1.24 (m, 2H).



Compound **100** was isolated in 40% yield and compound **100'** was isolated in 32% yield following the conditions G.

Compound **100** (dr = 1:1): The dr ratio of compound **100** is determined by ¹H NMR (see ¹H NMR analysis).

¹**H NMR** (500 MHz, CDCl₃) δ 9.68 (s, 0.5H), 8.98 (d, J = 7.2 Hz, 0.5H), 7.37 (d, J = 7.7 Hz, 0.5H), 7.32 (d, J = 7.7 Hz, 0.5H), 7.22 – 6.96 (m, 2H), 4.35 – 4.29 (m, 1H), 4.25 – 4.20 (m, 1H), 4.17 – 4.11 (m, 1H), 4.08 – 4.02 (m, 1H), 3.98 – 3.90 (m, 1H), 2.98 – 2.73 (m, 8H), 2.17 (td, J = 14.1, 7.0 Hz, 1H), 2.02 (dt, J = 14.6, 7.4 Hz, 1H), 1.91 – 1.74 (m, 4H), 1.42 – 1.37 (m, 3H), 0.84 (dt, J=6.6, 3.6 Hz, 3H).^{8b}

Compound **100'** (dr = 1:1): The dr ratio of compound **100'** is determined by ¹⁹F NMR and HPLC (see ¹⁹F NMR and HPLC analysis).

¹**H NMR** (500 MHz, CDCl₃) δ 9.68 (s, 1H), 7.31 (d, J = 7.7 Hz, 1H), 7.09 (d, J = 7.7 Hz, 1H), 4.34 (dd, J = 8.2, 4.5 Hz, 1H), 4.24 (dd, J = 7.5, 3.8 Hz, 1H), 4.16 (dd, J = 11.2, 6.0 Hz, 1H), 4.02 (dd, J = 10.7, 5.7 Hz, 1H), 3.92 (dd, J = 7.4, 4.2 Hz, 1H), 3.12 – 2.74 (m, 8H), 2.23 – 2.13 (m, 1H), 2.03 – 1.77 (m, 5H), 1.40 (t, J = 7.6 Hz, 3H), 0.83 (t, J = 7.3 Hz, 3H). ¹³**C NMR** (126 MHz, CDCl₃) δ 173.35, 138.62, 135.40, 130.95, 123.51, 120.81, 119.37, 117.41, 114.21, 108.06, 74.48, 64.00, 63.89, 61.16, 43.17, 43.15, 41.76, 36.84, 36.79, 32.09, 31.59, 30.91, 30.36, 29.86, 29.52, 28.97, 28.93, 24.72, 24.19, 22.85, 19.35, 19.28, 14.26, 13.40, 7.59, 1.18, 0.13. ¹⁹**F NMR** (471 MHz, CDCl₃) δ -80.21 – -82.05 (m, 6F), -102.12 – -105.20 (m, 2F), -110.83 – -115.89 (m, 2F), -120.24

--121.50 (m, 2F), -124.51 (d, J=9.8 Hz, 2 F), -125.07 - -127.30 (m, 4F). **HRMS** Calcd for C₃₀H₂₉F₁₈INO₃ [M+H⁺]: 920.0899; Found: 920.0900. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.

Compound **101** was isolated in 70% yield following the conditions G (E:Z = 3:1). ¹**H NMR** (500 MHz, CDCl₃) δ 7.50 – 7.27 (m, 5H), 6.56 (dt, *J* = 40.0, 13.4 Hz, 1H). ¹³**C NMR** (**126 MHz, CDCl**₃) δ 141.46, 130.36, 129.44, 128.69, 128.37, 128.15, 127.22, 127.05, 126.99, 126.97, 126.87, 124.12. ¹⁹**F NMR** (471 MHz, CDCl₃) δ -80.31 – -81.46 (m, 3F), -104.35 – -106.41 (m, 2F), -123.81 (dd, *J* = 19.4, 9.7 Hz, 2F), -125.38 – -126.30 (m, 2F). **HRMS** Calcd for C₁₂H₇F₉I [M+H⁺]: 448.9443; Found: 448.9441. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **102** was isolated in 85% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.98 – 7.94 (m, 2H), 7.43 – 7.39 (m, 2H), 6.52 (t, J = 14.3 Hz, 1H), 4.53 (t, J = 6.1 Hz, 2H), 3.14 (t, J = 5.8 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 165.50, 139.82, 131.16, 129.42, 128.89, 128.28, 116.11, 63.54, 40.04. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.40 – -81.70 (m, 3F), -105.49 (dd, J = 29.5, 14.5 Hz, 2F), -124.07 (dd, J = 20.0, 9.8 Hz, 2F), -125.07 – -126.94 (m, 2F). HRMS Calcd for C₁₅H₁₀ClF₉IO₂ [M+H⁺]: 554.9265; Found: 554.9265. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **103** was isolated in 68% yield following the conditions G. ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.35 (m, 2H), 6.78 – 6.74 (m, 2H), 6.39 (t, J = 14.4 Hz, 1H), 3.96 (t, J = 6.1 Hz, 2H), 2.87 (t, J = 7.4 Hz, 2H), 2.12 – 2.04 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 157.93, 132.40, 127.45 (t), 121.21, 116.37, 113.17, 66.37, 38.11, 29.82. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.30 – -81.60 (m, 3F), -105.78 (dd, J = 29.8, 14.7 Hz, 2F), -124.13 (dd, J = 19.9, 9.8 Hz, 2F), -125.45 – -126.34 (m, 2F). HRMS Calcd for C₁₅H₁₂BrF₉IO [M+H⁺]: 584.8967; Found: 584.8969. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **104** was isolated in 75% yield following the conditions G. ¹**H** NMR (500 MHz, CDCl₃) δ 7.86 – 7.81 (m, 2H), 7.72 – 7.68 (m, 2H), 6.32 (t, *J* = 14.4 Hz, 1H), 3.70 (t, *J* = 7.0 Hz, 2H), 2.66 (t, *J* = 7.0 Hz, 2H), 1.73 – 1.61 (m, 4H). ¹⁹**F** NMR (471 MHz, CDCl₃) δ -80.35 – -81.74 (m, 3F), -104.62 – -106.40 (m, 2F), -123.85 – -124.74 (m, 2F), -125.30 – -126.61 (m, 2F).^{8b}

10. General procedure and substrate scope for C-H perfluorobutylation of electron-rich arenes and heteroarenes



Scheme 10 C-H perfluoroalkylation of aniline and heteroarenes

Conditions H: Substrate (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (0.6 mmol, 3.0 equiv) and KOH (0.3 mmol, 1.5 equiv) were dispersed in 2 mL of H_2O . The reaction vial was purged with Ar for 1 min then sealed with PTEF cap. The reaction mixture was vigorously stirred at 30°C under irradiation of blue LED (25W) for 20 h. Then, the

mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.

Conditions I: Substrate (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (0.6 mmol, 3.0 equiv) and KOH (0.3 mmol, 1.5 equiv) were dispersed in 2 mL of CH_2Cl_2 . The reaction vial was purged with Ar for 1 min then sealed with PTEF cap. The reaction mixture was vigorously stirred at 30°C under irradiation of blue LED (25W) for 20 h. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.



Compound **105** was isolated in 66% yield following the conditions H. ¹H NMR (500 MHz, CDCl₃) δ 7.99 (d, J = 9.0 Hz, 2H), 6.67 (d, J = 9.3 Hz, 2H), 3.12 (s, 6H); ¹⁹F NMR (471 MHz, CDCl₃) δ -80.08 – -81.95 (m, 3F), -107.18 – -108.84 (m, 1F), -110.27 (dd, J = 16.4, 13.8 Hz, 1F), -121.55 – -123.47 (m, 2F), -124.96 – -127.22 (m, 2F).^{8b}



Compound **106** was isolated in 57% yield following the conditions H. ¹**H** NMR (500 MHz, CDCl₃) δ 7.82 – 7.71 (m, 2H), 7.09 (d, J = 8.3 Hz, 2H), 6.89 – 6.81 (m, 1H), 6.75 (d, J = 2.8 Hz, 1H), 6.39 – 6.30 (m, 1H). ¹⁹**F** NMR (471 MHz, CDCl₃) δ -78.86 – 82.87 (m, 3F), -99.93 – -102.66 (m, 2F), -121.46 (d, J = 9.7 Hz, 2F), -125.86 (dd, J = 15.8, 11.3 Hz, 2F).^{8b}

Compound 107 was isolated in 54% yield following the conditions H. ¹H NMR (500

MHz, CDCl₃) δ 8.16 (s, 1H), 7.67 (d, J = 8.0 Hz, 1H), 7.40 (d, J = 8.3 Hz, 1H), 7.34 (t, J = 7.6 Hz, 1H), 7.21 (t, J = 7.5 Hz, 1H), 2.45 (s, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -80.66 - -81.46 (m, 3F), -108.36 - -109.16 (m, 2F), -122.25 - -123.45 (m, 2F), -125.53 - -126.44 (m, 2F).



Compound **108** was isolated in 60% yield following the conditions I. ¹H NMR (500 MHz, CDCl₃) δ 8.46 (s, 1H), 7.76 (d, *J* = 8.0 Hz, 1H), 7.40 (d, *J* = 8.2 Hz, 1H), 7.33 (t, *J* = 7.6 Hz, 1H), 7.21 (t, *J* = 7.5 Hz, 1H), 5.13 (d, *J* = 8.3 Hz, 1H), 4.66 (dd, *J* = 14.9, 7.1 Hz, 1H), 3.64 (s, 3H), 3.37 – 3.25 (m, 2H), 1.36 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 172.55, 155.07, 136.21, 127.69, 125.37, 121.10, 120.54, 115.61, 111.85, 80.00, 54.23, 52.39, 29.85, 28.31, 28.07. ¹⁹F NMR (471 MHz, CDCl₃) δ -80.31 – 81.74 (m, 3F), -105.29 – -109.79 (m, 2F), -122.76 (dd, *J* = 18.4, 7.9 Hz, 2F), -124.54 – -127.41 (m, 2F). HRMS Calcd for C₂₁H₂₂F₉N₂O₄ [M+H⁺]: 537.1430; Found: 537.1436. The ¹³C peaks of the perfluoroalkyl moiety is not visible due to ¹⁹F-¹³C coupling and are not reported.



Compound **109** was isolated in 65% yield following the conditions I. The spectra data are consistent with those reported in literature.^{8b} ¹**H NMR** (400 MHz, CDCl3) δ 8.51 (s, 1H), 7.81 (d, *J* = 7.6 Hz, 1H), 7.52 (d, *J* = 8.0 Hz, 2H), 7.40 (d, *J* = 8.2 Hz, 1H), 7.33 (t, *J* = 7.4 Hz, 1H), 7.21 (t, *J* = 7.4 Hz, 1H), 6.71 (d, *J* = 8.1 Hz, 2H), 6.08 (d, *J* = 6.7 Hz, 1H), 5.16 (s, 1H), 4.67 (q, *J* = 12.2, 1H), 4.50-4.30 (m, 1H), 3.58 (s, 3H), 3.28 (d, *J* = 5.2 Hz, 2H), 3.06-2.83 (m, 2H), 1.37 (s, 9H). ¹⁹**F NMR** (376 MHz, CDCl3) δ -79.81 – - 81.92 (m, 3F), -105.42 – -109.99 (m, 2F), -122.70 (d, *J* = 10.3 Hz, 2F), -124.60 – -

11. Gram-scale C(sp³)-H amidation, C(sp²)-H iodination and perfluoroalkylation reactions

a) Gram-scale C(sp³)-H amidation



b) Gram-scale C(sp²)-H iodination reactions



c) Gram-scale addition of R_f-I to alkene 88



Scheme S11. Gram-scale C(sp³)-H amidation, C(sp²)-H iodination and perfluoroalkylation reactions

Gram-scale C(sp³)-H amidation: Compound **4-1** (10.0 mmol, 1.0 equiv), C_4F_9I (20.0 mmol, 2.0 equiv) and NaO*t*Bu (30.0 mmol, 3.0 equiv) were dispersed in 100 mL of ethers. The reaction vial was then purged with Ar for 5 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product **4** in 80% isolated yield.

Gram-scale C(sp²)-H iodination: Compound **30-1** (10.0 mmol, 1.0 equiv), C₄F₉I (20.0 mmol, 2.0 equiv) and NaO*t*Bu (30.0 mmol, 3.0 equiv) were dispersed in 100 mL of THF. The reaction vial was then purged with Ar for 5 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Then, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash

chromatography to give the desired product **30** in 86% isolated yield.

Gram-scale perfluoroalkylation reactions: Compound **90-1** (10.0 mmol, 1.0 equiv), perfluoroalkyl iodide (30.0 mmol, 3.0 equiv) and KOH (15.0 mmol, 1.5 equiv) were dispersed in 100 mL of H₂O. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30°C under irradiation of blue LED (25W) for 20 h. Then, the reaction mixture was extracted with EtOAc (3 x 100 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography to afford fluorination compound **90** in 66 % isolated yield.

12. Mechanistic studies

12.1 Titration experiment of C₄F₉I with *t*BuONa⁸

¹⁹F NMR spectra of nine samples of mixtures of C₄F₉I and *t*BuONa in THF were recorded at 298 K. Fluorobenzene ($\delta_{CF3-Ph} = -63.72$) was used as internal standard. The formation of a halogen bonding complex between NaO*t*Bu and C₄F₉I is supported by ¹⁹F NMR titration experiments. The resonance corresponding to the F of the CF₂I group is shifted to the upfield region when NaO*t*Bu was added to the THF solution of C₄F₉I (Table S2 and Scheme S12).

$$C_4F_9I + tBuONa \xrightarrow{THF (1 mL)} C_4F_9I...OtBu + Na^+$$

$[C_4F_9I]/([C_4F_9I]+[tBuONa])$	$ \Delta \delta$
0.1	4.84
0.2	4.02
0.3	3.47
0.4	2.23
0.5	1.28
0.6	0.32
0.7	0.23
0.8	0.09

$$[C_4F_9I] + [tBuONa] = 0.25 M$$

0.7

Table S2. The ¹⁹F-NMR signals for the mixture of C4F9I with NaOtBu shifted downfield when the ratio of C4F9Ito NaOtBu increased



Scheme S12. The ¹⁹F-NMR signals for the mixture of C₄F₉I with NaO*t*Bu shifted downfield when the ratio of C₄F₉I to NaO*t*Bu increased

12.2Titration experiment of C₄F₉I with KOH⁸

¹⁹F NMR spectra of seven samples of mixtures of C_4F_9I and KOH in THF were recorded at 298 K. Fluorobenzene ($\delta_{F-Ph} = -113.07$) was used as internal standard. The formation of a halogen bonding complex between KOH and C_4F_9I is supported by ¹⁹F NMR titration experiments. The resonance corresponding to the F of the CF₂I group is shifted to the upfield region when KOH was added to the THF solution of C_4F_9I (Table S3 and Scheme S9).

$$C_4F_9I$$
 + KOH
THF (0.5 mL)
 \sim $C_4F_9I...OH$ + K +
rt, 10 min

```
[C<sub>4</sub>F<sub>9</sub>I] + [KOH] = 0.5 M
```

$[C_4F_9I] / ([C_4F_9I]+[KOH])$	$\Delta\delta$ (ppm)
0.3	0.90
0.4	0.72
0.5	0.65
0.6	0.57
0.7	0.46
0.8	0.26
0.9	0.17

Table S3. The ¹⁹F NMR signals for the mixture of C₄F₉I with KOH shifted downfield when the ratio of C₄F₉I to KOH increased



Scheme S13. The ¹⁹F NMR signals for the mixture of C₄F₉I with KOH shifted downfield when the ratio of C₄F₉I to KOH increased

Note: The binding constant of the halogen bond complex $(R_f - I - OtBu)$ or $(R_f - I - OH)$ could not be accurately measured because the NaO*t*Bu or KOH was not completely soluble in the solvent and the substrate R_f -I is converted to perfluorobutane over time.

12.3 Reaction of C₄F₉I with TEMPO

Perfluorobutyl iodide (0.1 mmol, 1.0 equiv) and *t*BuONa (0.2 mol, 2.0 equiv) were dispersed in 1.0 mL of THF in a 4 mL glass vial. The vial was purged with Ar for 1 min and the reaction mixture was stirred at 30 °C for 12 h. Fluorobenzene (10 μ L) was used as an internal standard ($\delta_{CF3-Ph} = -63.72$) for ¹⁹F NMR analysis. The composition of reaction mixture was identified based on the chemical shift of -OCF₂ group at -78.90 ppm for compound **110**.¹¹



Scheme S14. Reaction of C₄F₉I with TEMPO

12.4 Reaction of C₄F₉I with THF

Perfluorobutyl iodide (0.2 mmol, 1.0 equiv) and *t*BuONa (0.6 mol, 3.0 equiv) were dispersed in 2.0 mL of THF in a 4 mL glass vial. The vial was purged with Ar for 1 min and the reaction mixture was stirred at 30 °C for 12 h. Fluorobenzene (10 μ L) was used as an internal standard ($\delta_{CF3-Ph} = -63.72$) for ¹⁹F NMR analysis. The composition of reaction mixture was identified based on the chemical shift of -CF₂-group at -126.22 ppm for compound **111**.¹⁰

$$C_4F_9I + \bigcirc H \xrightarrow{\text{NaOtBu} (x \text{ mmol})} C_4F_9-H$$

0.1 mmol 1.0 mL 57% (crude ¹⁹E-NMR)



Scheme S15. Reaction of C₄F₉I with THF

12.5 Reaction of C₄F₉I with benzothiazole and benzotriazole

Benzothiazole (0.1 mmol, 1.0 equiv) and benzotriazole (0.1 mmol, 1.0 equiv), C₄F₉I (0.2 mmol, 2.0 equiv) and NaO*t*Bu (0.3 mmol, 3.0 equiv) were dispersed in 1 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Then the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired products, iodination product was isolated in 90% yield and amidation products were isolated in 83% yield (75%+8%).



Scheme S16. Reaction of C₄F₉I with benzothiazole and benzotriazole

12.6 Iodination reaction of benzothiazole with TEMPO

Benzothiazole (0.1 mmol, 1.0 equiv), TEMPO (0.3 mmol, 3.0 equiv), C₄F₉I (0.2 mmol,

2.0 equiv) and NaOtBu (0.3 mmol, 3.0 equiv) were dispersed in 1 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C for 12 hours. Iodination product was not observed by GCMS, which proved that the iodination process was involved in free radical intermediate.



Scheme S17. Iodination reaction of benzothiazole with BHT

13 References

- 1. M. York, R. A. Evans, Tetrahedron Lett., 2010, 51, 4677.
- (a) F. Shi, M. K. Tse, S. Zhou, M. Pohl, J. Radnik, S. Hubner, K. Jahnisch, A. Brückner, M. Beller, *J. Am. Chem. Soc.*, 2009, **131**, 1775. (b)D. C. JohnsonII, T. S. Widlanski, *Tetrahedron Lett.*, 2004, 45, 8483.
- (a) J. Rong, L. Deng, P. Tan, C. F. Ni, Y. C. Gu, J. B. Hu, *Angew. Chem., Int. Ed.,* 2016, **55**, 2743. (b) Z. He, M. Bae, J. Wu, T. F. Jamison, *Angew. Chem., Int. Ed.,* 2014, **53**, 14451. (c) M. Suzuki, K.-I. Nunami, K. Matsumoto, N. Yoneda, O. Kasuga, H. Yoshida, T. Yamaguchi, *Chem. Pharm. Bull.*, 1980, **28**, 2374. (d) B. Beck, G. Larbig, B. Mejat, M. Magnin-Lachaux, A. Picard, E. Herdtweck, A. Dömling, *Org. Lett.*, 2003, **5**, 1047. (e) A. Chighine, S. Crosignani, M. C. Marie-Claire Arnal, M. Bradley, B. Bruno Linclau, *J. Org. Chem.*, 2009, **74**, 4753. (f) T. Xu, C. W. Cheung, X. L. Hu, *Angew. Chem., Int. Ed.*, 2013, **53**, 4910. (g) B. S. L. Collins, M. G. Suero, M. J. Gaunt, *Angew. Chem., Int. Ed.*, 2013, **52**, 5799. (h) D. I. Rozkiewicz, D. Jańczewski, W. Verboom, B. J. Ravoo, D. N. Reinhoudt, *Angew. Chem., Int. Ed.*, 2006, **45**, 5292. (i) R. Sakamoto, H. Kashiwagi, S. Selvakumar, S. A. Moteki, K. Maruok, *Org. Biomol. Chem.*, 2016, **14**, 6417.
- 4. S. Pan, J. Liu, H. Li, Z. Wang, X. Guo, Z. Li, Org. Lett., 2010, 12, 1932.
- Q. Yang, P. Y. Choy, W. C. Fu, B. Fan, F. Y. Kwong, J. Org. Chem., 2015, 80, 11193.
- H. Aruri, U. Singh, S. Sharma, S. Gudup, M. Bhogal, S. Kumar, D. Singh, V. K. Gupta, R. Kant, R. A. Vishwakarma, P. P. Singh, *J. Org. Chem.*, 2015, 80, 1929.
- (a) Q. Shi, S. Zhang, J. Zhang, V. F. Oswald, A. Amassian, S. R. Marder, S. B. Blakey, *J. Am. Chem. Soc.*, 2016, **138**, 3946. (b) E. F. Flegeau, M. E. Popkin, M. F. Greaney, *Org. Lett.*, 2008, **10**, 2717. (c) J. Helgoual'ch, A. Seggio, F. Chevallier, M. Yonehara, E. Jeanneau, M. Uchiyama, F. Mongin, *J. Org. Chem.*, 2008, **73**, 177.
- (a) P. Job, Justus Liebigs Ann. Chem., 1928, 9, 113. (b) Y. X. Wang, J. H. Wang, G.-X. Li, G. He, G. Chen, Org. Lett., 2017, 19, 1442. (c) Y. Z. Cheng, S. Y. Yu,

Org. Lett., 2016, 18, 2962.

- 9. (a) H. Berkeley, J. Physic. Chem., 1963, 67, 846. (b) H. Ashbaugh, J. Physic. Chem., 1964, 68, 811. (c) R. Foster, C. A. Fyfe, Trans. Faraday Soc., 1965, 61, 1626. (d) R. Foster, C. A. Fyfe, J. Chem. Soc., Chem. Commun., 1965, 642. (e) M. G. Sarwar, B. Dragisic, L. J. Salsberg, C. Gouliaras, M. S. Taylor, J. Am. Chem. Soc., 2010, 132, 1646. (f) (e) M. G. Sarwar, B. Dragisic, E. Dimitrijevic, M. S. Taylor, Chem. Eur. J., 2013, 19, 2050.
- 10. (a) A. Foris, *Magn. Reson. Chem.*, 2004, 42, 534–555. (b) M. Hanack, J. Ullmann, *J. Org. Chem.*, 1989, 54, 1432-1435. (c) X. Wang, H. Song, S. Wang, J. Yang, H. Qin, X. Jiang, C. Zhang, *Tetrahedron*, 2016, 72, 7606-7612.
- 11. D.-F. Jiang, C. Liu, Y. Guo, J.-C. Xiao, Q. Chen, *Eur. J. Org. Chem.*, 2014, **2014**, 6303-6309.

14. ¹H-NMR, ¹³C-NMR spectra and ¹⁹F-NMR





¹H NMR of compound 4









¹H NMR of compound **7**











S72




S74



¹H NMR of compound **14**



¹³C NMR of compound **14**





¹H NMR of compound **15**



¹H NMR of compound **16**







S79



0.0 10.0





S81



¹³C NMR of compound **21**





20 10 (

0 190 180

170



















¹³C NMR of compound **27**



¹³C NMR of compound **28**







¹⁹F NMR of compound **29**



¹H NMR of compound **30** 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 fl (ppm) .0 9.5 9.0 8.5 -0 ¹H NMR of compound **31** ^{123,63}
^{124,63}
^{124,64}
^{124,64} 77.48 77.16 76.84 140 130 120 110 100 90 80 70 60 fl (ppm) 190 180 00 170 160 150 50 40 30 20 10 . (





¹⁹F NMR of compound **31**

-160

-170

-180

-190

-2



-100 -110 -120 -130 -140 -150 fl (ppm)

10

-50

-60

-70

-80

-90





¹³C NMR of compound **34**



36































S107
















S112











S116













¹⁹F NMR of compound **57**



¹⁹F NMR of compound **58**





S122



S123



S124





¹⁹F NMR of compound **62**





S126



















S132





¹⁹F NMR of compound **69**



S135









S138







S141





¹⁹F NMR of compound **75**



S144






S147



S148



S149

S153

¹³C NMR of compound 84

¹H NMR of compound **85**

S159

¹H NMR of compound **87**

S162

¹³C NMR of compound **89**

112.16 112.19 112.21 112.21 112.22 112.22 112.22 112.22 112.22 112.22 112.23 12

 $\xleftarrow{-81.07}{-81.06}$

¹⁹F NMR of compound **91**

The dr ratio of compound **91** is determined by ¹⁹F NMR:

222222222222222222222222222222222222222	128252344
199999999	22222269
TUTT	UUU
	11

	Retention time (min)	Relative area	dr = 1:1
1	1.945	44.58	
2	2.347	42.36	

.0 9.5 9.0 8.5 8.0

7.5

7.0 6.5 6.0 5.5 f1 (ppn) ¹H NMR of compound **94**

¥ 80.1 4.5 ₩₩₩ ₩ 8888 81111 4.0

5.0

500.21 500.21 1.5

2.0

2. 00

2.5

3.0

3.5

The dr ratio of compound **94** is determined by ¹H NMR:

	Chemical shift	Integral area	dr = 1:1
1	5.51 (s)	0.50	
2	5.52 (s)	0.49	

¹⁹F NMR of compound **95**

¹³C NMR of compound **97**





The dr ratio of compound **99** is determined by ¹H NMR:







The dr ratio of compound **100** is determined by ¹H NMR:







The dr ratio of compound **100'** is determined by HPLC:













S189







S192



S193







¹⁹F NMR of compound **108**