## Supporting Information for:

# Synthesis of a novel carboxybetaine copolymer with different spacer lengths and inhibition of nonspecific protein adsorption on its polymer film 

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The supporting information are composed of 9 figures and 1 table in 11 pages

## List of abbreviations

AIBN: 2,2'-Azobis(isobutyronitrile)
anti-HSA: Antihuman serum albumin

DMAEMA/PDMAEMA: $N, N$-Dimethylaminoethyl methacrylate/ poly( $N, N$-dimethylaminoethyl methacrylate)

EDC: 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride

FBS: Fetal bovine serum

NHS: $N$-Hydroxysuccinimide

NMR: Nuclear magnetic resonance

RAFT: Reversible addition-fragmentation chain-transfer

SEC: Size exclusion chromatography

SPR: Surface plasmon resonance

TFA: Trifluoroacetic acid

XRR: X-ray reflectometry


Figure S1. SEC curves of PDMAEMA obtained with AIBN / RAFT agent initiating system in toluene at $60{ }^{\circ} \mathrm{C}$. $[\mathrm{DMAEMA}]_{0}=500 \mathrm{mM} .[D M A E M A]_{0} /[\text { RAFT agent }]_{0}=200$ (a), 100 (b), 50 (c), 25 (d). $[\text { AIBN }]_{0} /[\text { RAFT agent }]_{0}=0.3$.


Figure S2. SEC curves of P(CBMA2) obtained with VA-044 / RAFT agent initiating system in acetate buffer ( pH 5.2 ) at $37^{\circ}$ C. $[\mathrm{CBMA} 2]_{0}=500 \mathrm{mM} .[\mathrm{CBMA} 2]_{0} /[\mathrm{RAFT} \text { agent }]_{0}=200 .[\mathrm{VA}-044]_{0} /[\mathrm{RAFT}$ agent $]_{0}=0.3$.

Table S1. CBMA1/CBMA3 composition and unreacted DMAEMA unit ratio of P(CBMA1/CBMA3) and amount of coated $P(C B M A 1 / C B M A 3)$ on the surface of SPR sensor chips.

| PDMAEMA <br> (Theoretical chain length) | PCBMA1/PCBMA3 |  | Ratio of unreacted DMAEMA (\%) ${ }^{\text {b }}$ | Amount of coated P(CBMA1/CBMA3) ${ }^{c}\left(\mathrm{ng} \mathrm{cm}^{-2}\right)$ | Thickness of the coated $\mathrm{P}\left(\right.$ CBMA1/CBMA3) ${ }^{d}$ (nm) |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | Theoretical composition | Measured composition by ${ }^{1} \mathrm{H}$ NMR ${ }^{a}$ |  |  |  |
| 25 mer | 100:0 | 100:0 | 1 | $332 \pm 6$ |  |
|  | 80:20 | 83:17 | 4 | $296 \pm 16$ |  |
|  | 0:100 | 0:100 | 7 | $319 \pm 19$ |  |
| 50 mer | 100:0 | 100:0 | 4 | $313 \pm 18$ |  |
|  | 90:10 | 92:8 | 4 | $274 \pm 10$ |  |
|  | 80:20 | 79:21 | 9 | $293 \pm 12$ |  |
|  | 60:40 | 66:34 | 10 | $277 \pm 3$ |  |
|  | 0:100 | 0:100 | 30 | $305 \pm 9$ |  |
| 100 mer | 100:0 | 100:0 | 2 | $328 \pm 25$ |  |
|  | 80:20 | 80:20 | 6 | $311 \pm 9$ |  |
|  | 0:100 | 0:100 | 25 | $353 \pm 29$ |  |
| 200 mer | 100:0 | 100:0 | 1 | $368 \pm 6$ | 3.0 |
|  | 80:20 | 77:23 | 5 | $357 \pm 11$ | 3.5 |
|  | 60:40 | 63:37 | 8 | $249 \pm 24$ | 2.7 |
|  | 0:100 | 0:100 | 29 | $392 \pm 18$ | 2.9 |

a: Polymer composition was calculated using peak integration ratio of methyl protons on quaternary ammonium of CBMA1 (3.30~3.35 ppm) and CBMA3 (3.20~3.25 ppm) by ${ }^{1} \mathrm{H}$ NMR.
$b$ : The unreacted unit ratio was calculated using peak integration ratio of methyl protons on quaternary ammonium of protonated DMAEMA ( $\sim 3.0 \mathrm{ppm}$ ) unit as shown Figure $\mathrm{S} 1(\mathrm{C})$.
$c$ : Amount of coated $\mathrm{P}(\mathrm{CBMA1} / \mathrm{CBMA} 3)$ was measured by SPR.
$d$ : Measured by X-ray reflectometry (XRR, Malvern Panalytical, X'Pert PRO-MRD), X-ray: $\mathrm{CuK} \alpha, \lambda=$ $1.54 \AA$. The surface roughness of the SPR chips was evaluated by AFM before use based on ISO25178. The arithmetic mean height of the surface ( Sa ) and the root mean square height of the surface ( Sq ) of two different chips were 0.652 and 0.630 nm for Sa and 0.807 and 0.788 nm for Sq , respectively.


Figure S3. ${ }^{1} \mathrm{H}$ NMR spectra of PDMAEMA in $\mathrm{D}_{2} \mathrm{O}$ (A), 100 mM NaOD solution (B), 100 mM DCl solution (C).


Figure S4. ${ }^{13}$ C NMR spectra of PDMAEMA (A), quarternized PDMAEMA (B), and P(CBMA1/CBMA3) (C) in $\mathrm{CD}_{3} \mathrm{OD}$. Signals (a) at 162 ppm and ca. 120 ppm are assigned as remaining TFA, which was removed by dialysis before coating on SPR sensor chips and following measurements.


Figure S5. ${ }^{1} \mathrm{H}$ NMR spectra of PCBMA1 before (A) and after (B) RAFT group removal by TCEP/ethanolamine reduction.


Figure S6. Overall view of thermo-gravimetric analyses of carboxybetaine (co)polymers with different spacer lengths. (a) PCBMA1, (b) P(CBMA1/CBMA3) (80/20), (c) P(CBMA1/CBMA3) (60/40), (d) PCBMA3, and (e) PCBMA2 (control).


Figure S7. Nonspecific protein adsorption on P(CBMA1/CBMA3) (50 mer)-coated surface characterized by SPR. P(CBMA1/CBMA3) $=0 / 100$ (green), 60/40 (gray), 80/20 (orange), 100/0 (blue). Procedure: (a) injection of FBS for 540 sec , (b) end of injection. SPR response of $1000 \mathrm{RU}=100 \mathrm{ng} / \mathrm{cm}^{2}$. Lines of same color are results of different sensor chips coated with the same $\mathrm{P}(\mathrm{CBMA} 1 / \mathrm{CBMA} 3)$ components.


Figure S8. Nonspecific protein adsorption on P(CBMA1/CBMA3) (200 mer)-coated surface characterized by SPR. P(CBMA1/CBMA3) $=0 / 100$ (green), $60 / 40$ (gray), 80/20 (orange), 100/0 (blue). Procedure: (a) injection of FBS for 540 sec , (b) end of injection. SPR response of $1000 \mathrm{RU}=100 \mathrm{ng} / \mathrm{cm}^{2}$. Lines of same color are results of different sensor chips coated with the same $\mathrm{P}(\mathrm{CBMA} 1 / \mathrm{CBMA} 3)$ components.


Figure S9. Antibody immobilization on P(CBMA1/CBMA3)-coated surface via NHS/EDC activation characterized by SPR. P CBMA1/CBMA3) $=0 / 100$ (green), $60 / 40$ (gray), 80/20 (orange), 100/0 (blue). Procedure: (a) injection of NHS/EDC ( $0.05 \mathrm{M} / 0.2 \mathrm{M}$ ) solution for 450 sec , (b) end of injection of NHS/EDC, (c) anti-HSA antibody ( $0.1 \mathrm{mg} / \mathrm{mL}$ ) for 540 sec , (d) end of injection of anti-HSA antibody, (e) injection of 10 mM carbonate buffer ( pH 9.0 ) with 0.3 M NaCl , (f) end of injection of carbonate buffer. SPR response of $1000 \mathrm{RU}=100 \mathrm{ng} / \mathrm{cm}^{2}$. Lines of same color are results of different sensor chips coated with the same $\mathrm{P}(\mathrm{CBMA1/CBMA3)}$ components.

